1

Epistasis at the SARS-CoV-2 RBD Interface and the Propitiously Boring Implications for Vaccine Escape

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Abstract

At the time of this writing, December 2021, potential emergence of vaccine escape variants of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a grave global concern. The interface between the receptor-binding domain (RBD) of SARS-CoV-2 spike (S) protein and the host receptor (ACE2) overlap with the binding site of principal neutralizing antibodies (NAb), limiting the repertoire of viable mutations. Nonetheless, variants with multiple mutations in the RBD have rose to dominance. Non-additive, epistatic relationships among RBD mutations are apparent, and assessing the impact of such epistasis on the mutational landscape is crucial. Epistasis can substantially increase the risk of vaccine escape and cannot be completely characterized through the study of the wild type (WT) alone. We employed protein structure modeling using Rosetta to compare the effects of all single mutants at the RBD-NAb and RBD-ACE2 interfaces for the WT, Delta, Gamma, and Omicron variants. Overall, epistasis at the RBD interface appears to be limited and the effects of most multiple mutations are additive. Epistasis at the Delta variant interface weakly stabilizes NAb interaction relative to ACE2 interaction, whereas in the Gamma variant, epistasis more substantially destabilizes NAb interaction. Although a small, systematic trend towards NAb destabilization not observed for Delta or Gamma was detected for Omicron, and despite bearing significantly more RBD mutations, the epistatic landscape of the Omicron variant closely resembles that of Gamma. These results suggest that, although Omicron poses new risks not observed with Delta, structural constraints on the RBD hamper continued evolution towards more complete vaccine escape. The modest ensemble of mutations relative to the WT that are currently known to reduce vaccine efficacy is likely to comprise the majority of all possible escape mutations for future variants, predicting continued efficacy of the existing vaccines.

Significance

Emergence of vaccine escape variants of SARS-CoV-2 is arguably the most pressing problem during the COVID-19 pandemic as vaccines are distributed worldwide. We employed a computational approach to assess the risk of antibody escape resulting from mutations in the receptor-binding domain of the spike protein of the wild type

SARS-CoV-2 virus as well as the Delta, Gamma, and Omicron variants. At the time of
writing, December, 2021, Omicron is poised to replace Delta as the dominant variant
worldwide. The efficacy of the existing vaccines against Omicron could be substantially
reduced relative to the WT and the potential for vaccine escape is of grave concern. Our
results suggest that although Omicron poses new evolutionary risks not observed for
the Delta variant, structural constraints on the RBD make continued evolution towards
more complete vaccine escape unlikely. The modest set of escape-enhancing
mutations already identified for the wild type likely include the majority of all possible
mutations with this effect.

Introduction

When severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), first emerged as a global public health concern early in 2020, there was considerable debate regarding whether the low mutation rate of the virus and the relatively inflexible receptor-binding domain (RBD) of the antigenic spike (S) protein would admit robust host adaptation(1, 2). By 2021, it became clear that SARS-CoV-2 has access to a broad mutational repertoire enabling extensive diversification(3) and that without vaccination, SARS-CoV-2 would likely result in substantial global disease burden for a protracted period(4, 5). The development of multiple, effective vaccines against SARS-CoV-2(6) make it possible to dramatically reduce this burden. However, at the time of writing, December, 2021, the majority of the global population remains unvaccinated as the Omicron variant is poised to replace the Delta variant as the dominant strain worldwide. Existing vaccine efficacy against the Omicron variant might be substantially reduced relative to the WT https://www.cdc.gov/coronavirus/2019-ncov/science/science-briefs/scientific-brief-omicron-variant.html, and the potential for continued evolution towards more complete vaccine escape(7) is a major global concern https://www.cdc.gov/coronavirus/2019-ncov/variants/variant-info.html.

The interface between the receptor-binding domain (RBD) of the S protein and the host receptor (ACE2) largely overlaps with the binding sites for the most potent neutralizing antibodies (NAb)(8, 9), limiting the scope of viable mutations. Nevertheless, multiple variants containing single mutations in the RBD that, to different extents, reduce NAb binding have begun to circulate(8-10). Moreover, variants with multiple mutations in the RBD have risen to dominance outcompeting the wild type (WT, identical to Wuhan-Hu-1) and single mutants (see below). These dynamics could result from non-additive, epistatic interactions among the mutated sites(10, 11) or simply from additive effects of multiple mutations(11). The effects of all single mutations in the RBD relative to the WT have been studied, and several mutations producing partial antibody escape have been identified (8, 12). Epistasis among RBD mutations has the potential to substantially increase the risk of escape variant emergence and cannot be characterized through the study of the WT alone.

Using the Rosetta software suite https://rosettacommons.org(13), we estimated and

compared the effects of all single non-synonymous mutants at the RBD-NAb and RBD-

ACE2 interfaces for the WT as well as Delta (452R, 478K), Gamma (417T, 484K,

501Y), and Omicron (339D, 371L, 373P, 375F, 417N, 440K, 446S, 477N, 478K, 484A,

493R, 496S, 498R, 501Y, 505H) variants. The Delta and Gamma variants were

dominant in different regions of the world with rising frequencies as of Summer,

2021(14, 15). Delta rose to global dominance in the following months and, at the time of

writing, Omicron is rapidly growing in frequency and is expected to become the next

globally dominant strain. We establish the distribution of RBD mutations on the plane

bounded by the costs of ACE2 and NAb binding and classify the direction and

magnitude of epistatic interactions between variant mutations and the broader

mutational repertoire. The results reveal only weak epistasis which, although more

pronounced for the Gamma and Omicron variants than for the Delta variant, suggests

limited potential for continued evolution towards more complete vaccine escape.

Results

Rationale

Epistatic interactions among mutations in the RBD of SARS-CoV-2 S protein are of

interest and concern because they might substantially increase the risk of vaccine

escape. Mutations in the RBD subtly change the shapes of the interfaces between RBD

and ACE2, and between RBD and NAb (Figure 1A). While the structure of the WT RBD-

ACE2 interface is highly similar to that of the RBD-NAb interface (see below), a single

mutation in the RBD can result in distinct shape changes in both interfaces. These

changes can be depicted by the position of each mutant on the plane bounded by the

receptor binding cost and the antibody cost (Figure 1B). The cost is the increase

(positive cost) or decrease (negative cost) in the ACE2 or NAb binding affinity relative to

the WT. The four quadrants of this plane (Figure 1B) represent four broad categories of

mutations. Mutations in the top, right quadrant are strongly destabilizing relative to both

ACE2 and the NAb. The bottom, right quadrant contains mutants that strongly

destabilize the interaction with ACE2 but not with the NAb. Most mutants in these

quadrants are not evolutionarily viable. Mutants in the bottom, left quadrant stabilize or

only weakly destabilize both interfaces. These mutations may or may not provide a

selective advantage to the virus depending on the fraction of the host population that

has been vaccinated or has recovered from prior infection. The top, left quadrant

contains mutations that strongly destabilize the interaction with NAb but not with ACE2

and therefore are most likely to admit vaccine escape.

Single-mutant vaccine escape candidates for the wild type RBD

Starting with the two crystal structures of interest, the RBD in complex with ACE2

https://www.rcsb.org/structure/6M0J(16) and the RBD in complex with the NAb CV30,

https://www.rcsb.org/structure/6XE1, we generated a representative ensemble of 50

native conformations per complex following standard Rosetta protocols (see Methods

and Discussion for details). Although NAb that bind epitopes, which do not overlap with

the RBD have been identified(8), at the time of writing, the antibodies most critical for

assessing the risk of vaccine escape appear to overlap with the RBD(9) and are well

represented by CV30. Regions important for antibody binding are known to overlap

broadly among human coronaviruses(17). We then identified the RBD residues at the

interface for each conformation and, in all conformations, introduced all single amino

acid substitutions at these sites. For the WT and Delta variant, 52 residues (Table 1)

were identified at the interface of at least one conformation for either complex. Four

additional residues were identified for the Gamma and/or Omicron variants. Sites 480

and 488 are connected by a disulfide bond and were found to be unsuitable for

substitution.

All 19 substitutions in each of the remaining 54 sites were investigated, with the

exception of WT reversion for the variants. Note that substitutions made in the WT are

all single mutants; those in the Delta variant are triple mutants (given that this variant

contains two mutations in the RBD); those in the Gamma variant are quadruple

mutants; and those in the Omicron variant each encompass 16 mutations in total. We

chose to examine all variants containing Delta/Gamma/Omicron substitutions and an

additional substitution at the interface rather than probing a randomly selected,

representative ensemble of multiple mutants relative to the WT because the mutations

present in the Delta/Gamma/Omicron variants are known to be of biological and

epidemiological relevance and the space of all such multiple mutants is prohibitively

large. Altogether, this analysis produced more than 400k structures which necessitated

the development of a computationally efficient, and therefore simplified, protocol. To

address this need, mutants were introduced into each conformation without repacking of

adjacent sidechains or backbone minimization. This minimalist approach yielded

favorable comparisons to the available experimental data (see below). However,

generally, substitutions might introduce steric or charge clashes within the

conformations in which the mutations were introduced (without repacking and

minimization). Inference of the relative change in binding affinity for the ACE2 and NAb

complexes is limited for such mutations. However, we observed a favorable comparison

to experimental data in this respect as well, whereby few experimentally predicted

escape mutations (relative to the WT) fall into this, inference-limited, category (see

below).

Structure stability was estimated by the total score, S, in arbitrary units produced by the

empirically-driven Rosetta Energy Function 2015(18) (labelled REU for "Rosetta Energy

Units", https://new.rosettacommons.org/docs/latest/rosetta_basics/Units-in-Rosetta).

The total score was calculated for each of the 50 conformations of the NAb and ACE2

complexes generated, and the mean value was assessed with, S_M^C, and without, S_{WT}^C,

the mutation. The receptor cost and antibody cost were estimated as $[S_M^C - S_{WT}^C]_{ACE2}$

and $[S_M^C - S_{WT}^C]_{NAb}$, respectively. The interface free energy (ΔG) was also more directly

approximated by the difference between the total score of the unbound state and the

complex, $S^C - S^U$. The effect of the mutation on this value ($\Delta\Delta G$) was reported for both

complexes, $(S_M^C - S_M^U) - (S_{WT}^C - S_{WT}^U)$. Figure 1C shows the distribution of RBD

mutations on the plane bounded by the ACE2 and NAb binding costs and putative NAb

escape candidates, for which $[S_M^C - S_{WT}^C]_{NAb} - [S_M^C - S_{WT}^C]_{ACE2} > 1$ or $\Delta\Delta G_{NAb}\text{-}\Delta\Delta G_{ACE2}>1$

and $[S_M^C - S_{WT}^C]_{ACE2} < 13$. The threshold value of 13 was selected to remove from

consideration mutations that likely produce steric or charge clashes in the structure; few

experimentally validated escape candidates were observed above this value (see

below).

Mutants showed strong clustering along the diagonal (identity line: receptor cost is

equal to antibody cost), indicating that most mutations similarly affected the WT RBD-

ACE2 and RBD-NAb complexes. Mutations in the top, left quadrant of the plane, which

corresponds to strong destabilization of the interaction with NAb but not with ACE2, are

the strongest candidates for vaccine escape, followed by those in the bottom, left

quadrant, which includes weakly destabilizing mutations. The selective advantage (or

lack thereof) of mutations in this quadrant depends on the fraction of the host population

that has been vaccinated or has recovered from prior infection (see Discussion). In a

fully vaccinated population, mutations that substantially reduce infectivity through the

destabilization of receptor binding, while also destabilizing the interaction with NAb,

could still provide a selective advantage. In particular, multiple mutations in 6 sites (417,

477, 484, 491, 493, 499) were found to substantially destabilize the RBD-NAb complex

relative to the RBD-ACE2 complex (Figure S1). Additionally, we identified site 453 to

harbor mutations that simultaneously stabilize the RBD-ACE2 complex and destabilize

the RBD-NAb complex. These observations are broadly consistent with the results of

deep mutational scanning(8, 9, 19-22). Therefore, we conservatively considered all

mutations, for which the antibody cost exceeded the receptor cost, to be viable escape

candidates.

Single-mutant vaccine escape candidates for the Gamma, Delta, and Omicron

variant RBDs and predicted epistatic interactions

Having charted the ACE2-binding and NAb-binding landscapes for the WT RBD, we

sought to identify the most prominent combinations of RBD mutations circulating over

the course of the pandemic. As of June, 16th, 2021, there were 53 countries, from which

more than 1,000 SARS-CoV-2 isolates were contributed to the GISAID(23) database

(see Data Availability). For each of these locations, we randomly selected 1,000 isolates

and reported the frequency of each combination of RBD mutations among the 53,000

selected isolates over time. Figure 2A displays this region-normalized global prevalence of the 10 most common combinations of RBD mutations. The 6 RBD single-mutants (501Y/Alpha Variant, 477N, 439K, 484K, 478K, and 459F) began emerging between July and November 2020. All these single mutants were eventually displaced by 4 RBD multiple mutants (452R|478K/Delta Variant, 417T|484K|501Y/Gamma Variant, 417N|484K|501Y/Beta Variant, and 346K|484K|501Y), which began emerging in November, 2020, with the exception of Beta, which according to our analysis, first appeared in July. By March, 2021, the WT had become less prevalent than the Alpha, Gamma, and Delta variants. We pursued further analysis for the Delta, Gamma, and the recently identified Omicron variant RBDs given their high and rising global prevalence. The complexes were prepared starting from the WT crystal structures and treated identically to the WT. We also discuss how similar results may be obtained starting directly with the native conformations approximated for the WT, substantially reducing the computational burden (see Methods).

The rapid emergence and subsequent displacement of RBD single mutants might in part result from epistasis among the RBD variant residues or could be due to purely additive interactions. The most prominent trend was the displacement of the single mutant, 501Y (Alpha variant) by the Beta and Gamma variants (both also containing 501Y). Residue 501Y has been shown to substantially increase the binding affinity with ACE2 which, however, is reduced with the addition of mutation 417N in the Beta variant(11). In contrast, 417N severely reduces the neutralizing activity of a variety of NAb(24). These observations imply that mutations in site 417 provide a selective advantage through destabilization of the NAb complex, but given the large overlap between the RBD-NAb and RBD-ACE2 interfaces, maintenance of sufficient infectivity requires a compensatory mutation, such as 501Y, that stabilizes the RBD-ACE2 complex. Note that the emergence of these variants preceded widespread vaccination and that, although the competition between antibody and receptor binding is present even during an infection of a naïve host, evolutionary pressures are likely to shift with increasing rates of vaccination and prior infection (see below). At the time of writing, the origin of the Omicron variant , and thus the evolutionary pressures that led to its

emergence remain unknown (see discussion); however, despite bearing many more

RBD mutations, the epistatic landscape at the interface is highly similar between the

Gamma and Omicron variants (see below).

Examination of the interface footprints, defined as the ensemble of sites predicted to lie

at the interface of at least one of the 50 conformations for each complex, for the 8

complexes of interest, demonstrates that the RBD makes a greater number of contacts

with NAb than with ACE2 within the same range of sites, 403-506 (Figure 2B). The

footprints of the WT and Delta variant interfaces in both the RBD-ACE2 and the RBD-

NAb complexes are identical. The WT/Delta RBD-ACE2 footprint consists of 37 sites

whereas the WT/Delta RBD-NAb footprint includes 51 sites (Table 1). The Gamma

RBD-ACE2 footprint consists of 41 sites including all those in the WT/Delta footprint,

with the single exception of site 484, and five additional sites. The Gamma RBD-NAb

footprint consists of 53 sites including all those in the WT/Delta interface and, in

addition, sites 408 and 480. The Omicron RBD-ACE2 footprint consists of 37 sites with

three WT sites missing (including 484) and three additions. The Omicron RBD-NAb

footprint includes 50 sites with one WT site missing.

Sites in the RBD-NAb footprint that are not shared by the RBD-ACE2 footprint might

provide routes for the emergence of vaccine escape variants. However, because the

RBD-ACE2 footprint is smaller than the RBD-NAb interface, the former is more sensitive

to perturbation than the latter, for example, from mutations in site 417, which is part of

the footprint of all 8 complexes. Notably, site 484 is absent from the Gamma and

Omicron RBD-ACE2 footprints, but remains in the respective RBD-NAb footprints. Also

of note, site 446, which is mutated in Omicron, is present in the RBD-NAb footprint but

not the RBD-ACE2 footprint for this variant. Consistent with the differences in the

footprints, we found the Omicron and, to a lesser extent, Gamma variant RBD

conformations in complex with ACE2 to be significantly different from that of the WT and

Delta variants, which could not be differentiated from one another. All variant RBD

conformations in complex with the NAb were found to be significantly different from that

of the WT. For Delta and Gamma, this difference was modest and smaller in magnitude

than the variability among the RBD-NAb conformations, whereas Omicron showed a

more pronounced difference from the WT (Figure 2C).

Classical multidimensional scaling (CMDS) applied to the pairwise interface RMSDs

among all RBD-ACE2 and RBD-NAb complexes showed that the Gamma and Omicron

RBD-ACE2 structures lie on a shared continuum of conformational change relative to

the WT, with the Omicron conformations being closer to Gamma than to the WT. In

contrast, Omicron, Gamma, and to a lesser extent, Delta RBD-NAb structures all

represent distinct conformational changes relative to the WT (Figures 2D and S2).

Despite these differences, the effects of mutations in the RBD were found to be

principally additive in all variants, that is, there seems to be little epistasis.

When a second mutation, M_j, is introduced in addition to a prior mutation, M_i (Figure

3A), the resulting conformational change can be additive so that the effect of the two

mutations is the sum of the effects of the two individual mutations. In this case, the

position of the double-mutant $M_{i,j}$ on the plane defined by the receptor cost and antibody

cost relative to the single mutant, M_i, will be the same as that of the single-mutant, M_j,

relative to the WT. If the conformational change is non-additive, representing an

epistatic relationship, the resulting trends can be classified by their impact on potential

vaccine escape. Such trends could be escape-neutral when the ensemble of candidate

vaccine escape mutations differs from that for the WT, but the number of such

candidates is the same; escape-minimizing when the antibody cost is on average

reduced relative to the receptor cost across all mutations for the mutant vs the WT; or

escape-exacerbating where the antibody cost is on average increased.

The landscape of mutants predicted to enhance vaccine escape for the Delta variant

was almost identical to that of the WT but differed significantly from the Gamma and

Omicron landscapes (Figure 3B). These trends are summarized in Table 2, which

tabulates all non-shared candidates. There are 15(13) escape candidates in the WT that

were not predicted to enhance escape for Delta and 6(2) candidates in Delta but not WT

(values in parentheses are mutations with $[S_M^C - S_{WT}^C]_{ACE2} < 13$, regardless of whether

or not the mutation is a candidate, a threshold chosen to mitigate potential artifacts

caused by steric or charge clashes). In contrast, in the case of Gamma, there were

32(28) candidates identified in WT but not Gamma, and 86(66) candidates identified in

Gamma but not the WT. Omicron demonstrated intermediate behavior with 67(59)

candidates identified in WT but not Omicron, and 75(48) candidates identified in

Omicron but not the WT. Thus, we identified 9(11) fewer escape candidates for Delta

compared to the WT, but 54(38) additional candidates for Gamma, and either 8

additional candidates or 11 fewer candidates (ignoring potential steric/charge clashes in

the WT) for Omicron.

Consistent with the more dramatic conformational change observed in the RBD-ACE2

complex relative to the RBD-NAb complex, the non-additive effects observed in the

Gamma variant appear to predominantly result from the decreased sensitivity of the

RBD-ACE2 interface to mutation. This conclusion is compatible with the available

experimental results. Figure S3 shows the distribution of the receptor cost, $[S_M^C -$

$S_{WT}^C]_{ACE2}$, for two categories of mutations and for each of the three receptor complexes:

those at the interface that have been experimentally demonstrated to reduce

neutralizing activity of antibodies COV2-2050 and COV2-2479 in the WT(8), and all

others (included in the same experimental study) at the interface. As discussed above,

the upper bound for the receptor cost, $[S_M^C - S_{WT}^C]_{ACE2}$, is lower for mutations predicted

to reduce NAb activity than for other mutations. However, the Gamma candidate

ensemble exhibits a reduced median receptor cost. In other words, in the Gamma

variant, mutations that are predicted to reduce NAb activity are also less likely than

other mutations to reduce the receptor binding affinity relative to the WT and Delta

variant. The three residues within the RBD-ACE2 footprint of the Omicron variant not

present in the WT overlap with those of the Gamma variant (404, 439, and 499) and,

although not statistically significant, the decreased cost of receptor binding was

observed for Omicron as well. The Omicron variant additionally displayed a modest

reduction in median receptor cost for the broader category of experimentally studied

mutations, suggesting greater flexibility at the receptor interface (Figure S3).

Figure 4 summarizes the magnitude of the increased risk of vaccine-escape for each

mutation at the RBD interface (given $[S_M^C - S_{WT}^C]_{ACE2} < 13$) for each variant relative to

the WT. Epistasis increasing the risk of vaccine-escape is primarily apparent in three

regions of the RBD interface: site 417, site 477, and site 494 together with the

surrounding neighborhood. The trend in site 417 was observed only in the Gamma and

Omicron variants, which already contain mutations at that site, showing that further

changes to this site could result in enhanced vaccine escape. However, the

epidemiological implications of this finding are limited considering that mutations in site

417 are likely to pose a risk of vaccine escape in most variants. The enhanced escape

associated with mutations in site 477 for all variants relative to the WT, together with the

early spread of 477N and the presence of 477N in the Omicron variant, suggest that this

site could play an important role in host adaptation. Most prominently, mutations in site

494 and the surrounding neighborhood are likely to enhance vaccine escape in all

variants. Indeed, 494P has both been found in circulation and experimentally

demonstrated to reduce antibody neutralization capacity of convalescent sera(25).

In addition to these apparent differences among the ensembles of candidate vaccine-

escape mutations, we observed sites that harbored no candidates, but nevertheless

displayed signatures of increased risk of vaccine escape for Gamma and/or Omicron.

The two most notable trends were observed in sites 408 and 504 (Figure S4). All but

one substitution in site 408 enhance vaccine-escape for Gamma and Omicron, but

surprisingly, all have the opposite effect in Delta. Similarly, all substitutions at site 504

substantially enhance vaccine-escape in Gamma and Omicron but exert a modest

opposite effect in Delta. However, these mutations are not considered candidates in our

analysis because, even for Gamma and Omicron, they destabilize the ACE2 interaction

to a greater extent than the interaction with NAb. Additionally, $[S_M^C - S_{WT}^C]_{ACE2} \gg 13$ for

substitutions at site 504, which limits confidence in the assessment of trends at this site.

Importantly, few differences were observed between the epistatic landscapes of the

Gamma and Omicron variants, mainly in sites 417, 439, 499 (see Figure 4), and 446

(see Figure S4). Sites 417 (reduced escape-related epistasis) and 446 (increased

escape-related epistasis) are already mutated in Omicron and the signatures in sites

439 and 499 are modest relative to the consistent trend observed in site 494.

Nonetheless, it is important to acknowledge a small, systematic bias towards NAb

destabilization within the Omicron variant that is not observed within Gamma or Delta

(Figure 4, inset and Figure S5). As demonstrated above, unlike the RBD-ACE2

complex, which displays a shared continuum of conformational change relative to the

WT, RBD-NAb structures all represent distinct conformational changes (Figures 2D and

S2) with the greatest change observed for Omicron. Most mutations, which modestly

destabilize the WT RBD-NAb complex ($[S_M^C - S_{WT}^C]_{NAb} < 10$), are slightly more

destabilizing in the Omicron RBD-NAb complex compared to the other variants. As

discussed above, only a subset of these mutations are considered candidates for

antibody escape, which depends on the properties of the mutation within the RBD-

ACE2 complex as well. The principal escape signatures are conserved between the

Gamma and Omicron variants, with fewer escape candidates identified overall for the

Omicron variant. However, this systematic destabilization of the Omicron RBD-NAb

complex suggests, in principle, additional avenues towards vaccine escape.

Discussion

Here we report the results of a computational study predicting the effects of all single

mutants at the RBD-NAb and RBD-ACE2 interfaces for the WT as well as the Delta,

Gamma, and Omicron variants of SARS-CoV-2 on receptor and antibody binding. For

the WT, we found multiple mutations in 6 sites (417, 477, 484, 491, 493, 499) that are

predicted to significantly destabilize the RBD-NAb complex relative to the RBD-ACE2

complex and appear to pose a risk of vaccine escape, which is broadly consistent with

the results of deep mutational scanning(8, 9, 19-22). Overall, most mutations at the

interface were found to similarly effect the WT and all variants, indicating limited

epistasis at the interface. Non-additive, epistatic interactions predicted to increase the

risk of vaccine-escape were apparent, however, at sites 477 and 494 as well as in the

surrounding neighborhood. This trend is particularly prominent in the Gamma variant so

that, across all sites at the interface, we predicted 22% more escape candidate

mutations for the Gamma variant than for the WT. In contrast, there is little apparent

epistasis in the Delta variant, and across all sites at the interface, we predicted 4%

fewer candidate mutations compared to WT. Despite harboring many more RBD

mutations and displaying a small, systematic trend towards NAb complex

destabilization, the Omicron variant demonstrated intermediate behavior with only 3%

more candidate escape mutations compared to the WT.

Epistasis is a major if not the principal driver of protein evolution(26). Compensatory

mutations are particularly strong epistatic interactions that can result in

chemotherapeutic(27) or antimicrobial(28) drug resistance, and are commonly observed

throughout species evolution(29). In a completely susceptible population, mutation

501Y, which appears to substantially increase infectivity(11), is expected to evolve

under positive selection. As a population gains immunity, through prior exposure and/or

vaccination, selective pressures rapidly change to promote the emergence of resistant

variants(7). Under these conditions, 501Y and other mutations, which increase

infectivity might primarily play the role of compensators for mutations destabilizing NAb

interactions, such as 417N or 417T. As global vaccinations rise, due to changing

evolutionary pressures, it can be expected that more mutations emerge that destabilize

the interactions of the RBD with both NAb and ACE2, thus resulting in (partial) escape

variants that, however, also have reduced infectivity. However, variants such as

Gamma that carry both an antibody destabilizing mutation and a compensatory

mutation have the potential to undercut this trend.

At the time of writing, the origins of the Omicron variant are unclear and is suspected to

have evolved over an extended period of time within an immunocompromised

individual(s) or an animal reservoir(30). Such environments could present selective

pressures distinct from the broader human host population, but the changing

competition between receptor and antibody binding described above is likely to be

conserved. Despite the more pronounced conformational change, the average behavior

of the conformational ensemble selected to represent the Omicron variant was found to

be less destabilizing for the receptor compared to Gamma but highly destabilizing for

the antibody, indicating that our model for Omicron is likely to be at least as accurate if

not more accurate than those for the Gamma and Delta variants (Figures S12-13).

Above all else, we find it important to highlight three points. 1) The epistatic landscapes

of the Gamma and Omicron variants are highly similar (Figure 4). 2) We find fewer, not

more, escape-exacerbating mutations in Omicron compared to Gamma (Table 2). 3)

These features persist in spite of the fact that RBD-ACE2 structures represent a shared

continuum of conformational change relative to the WT, with the Omicron conformations

being closer to Gamma than the WT (Figures 2D and S2), indicating that the magnitude

of conformational change does not trivially correlate with the propensity for exacerbated

antibody escape. The neutralizing activity of existing vaccine elicited NAb against the

Omicron variant is likely to be substantially reduced(31), but multi-dose vaccination is

expected to recover efficacy(32). Our results emphasize that, although the adaptive

repertoire of SARS-CoV-2 may be robust, structural constraints on the RBD make

continued evolution towards more complete vaccine escape unlikely, suggesting

continued efficacy of the existing vaccines.

The work presented here is strictly computational, and although we demonstrate

agreement with experimental results where possible, many features not captured by the

models presented (involving protein expression, docking, and other factors) could

modulate antigen-receptor and/or antigen-antibody binding. Furthermore, although we

explore many conformations for both the RBD-ACE2 and RBD-NAb interfaces, we start

from a single crystal structure for each. We believe the conformational ensembles

selected to represent each complex are diverse enough to accurately reflect the relative

destabilization of the NAb and ACE2 complexes across the spectrum of RBD interface

mutations, which is the primary concern. However, this conformational diversity makes it

difficult to demonstrate stabilizing interactions, which are typically much weaker than

destabilizing ones(12). Although multiple low-energy conformations were resolved for all

variants and the WT, the average behavior of the conformational ensemble selected to

represent the Delta variant relative to the WT was found to be weakly destabilizing for

NAb and neutral for ACE2, whereas in the case of Gamma, it was weakly destabilizing

for both complexes. This is unlikely to accurately reflect the relative binding affinities

between these variants and the WT given the enhanced infectivity of both variants,

particularly Delta(14). However, it should be recognized that the relationship between

the measures of interface stability we report and viral life history traits (infectivity,

immune activity, etc.) is complex.

Although we believe we proposed sensible thresholds for determining which structures

can be analyzed with high confidence and the biological implications of the relative

destabilization of the NAb vs ACE2 are overt, the effects studied in this work do not

represent the entire diversity of possible host adaptation. It is incompletely understood

at the time of writing why the Delta variant appears to replicate faster than the WT(14)

and substitutions outside the Spike protein could play key roles in immune

modulation(33, 34). Even less is known about the Omicron variant. A targeted

exploration of the lowest-energy conformations achievable for each variant might yield

better agreement with the known properties of these variants and, in particular, reveal

stabilizing RBD-ACE2 interactions. However, this would likely come at the cost of

generalizability and decrease the power of our approach to predict relative

destabilization of interface mutations between NAb and ACE2 complexes.

We also emphasize that we limit our study to the RBD of the S protein. In principle,

epitopes located outside the RBD and far from the interface could play an important role

in the emergence of vaccine escape variants by decreasing the similarity between the

interaction with the antigen and receptor, and that between the antigen and an

antibody(35). However, if such effects were dominant, the satisfactory agreement

between our predictions, deep mutational scanning, and the observed frequency of

mutations within the RBD among the circulating variants, presumably, would not be

recovered. Nevertheless, many routes of adaptive evolution are potentially available to

this virus so that agreement with prior results is not a guarantee of predictive success.

Although we believe our results strongly suggest a limited repertoire of escape-

mediating mutations within the RBD, the possibility should be considered that mutations

outside the RBD have the potential to increase this repertoire.

Finally, we note that the space available for neutral evolution, even within the RBD

alone, is large. In principle, this makes possible the acquisition of many RBD mutations,

some combinations of which might exhibit substantially greater escape-exacerbating

epistatic effects than the variant substitutions explored in this work. However, this

appears unlikely considering the large number of mutations observed in the Omicron

variant, 15. We believe this variant is an excellent test case to probe the limits of

epistatic potential at the RBD interface.

Conclusions

We employed a computational approach to study the effects of all single mutations at

the RBD-NAb and RBD-ACE2 interfaces for the WT as well as the Delta, Gamma, and

Omicron variants of SARS-CoV-2. Overall, little epistasis at the RBD interface was

detected, with additive effects on the binding affinities observed for most pairs of

mutations. In the Delta variant, the observed non-additive trends weakly stabilize the

interaction of the RBD with the NAb relative to the interaction with ACE2, whereas in the

Gamma variant, epistasis is predicted to more substantially destabilize interaction with

the NAb relative to ACE2. The epistatic landscape of the Omicron variant closely

resembles that of Gamma, with an additional small systematic bias towards NAb

destabilization, but with fewer predicted escape candidates overall. These results

suggest that, although the Omicron variant poses new risks not observed for Delta,

including the evolution towards greater NAb destabilization, structural constraints on the

RBD make the continued evolution towards more complete antibody escape unlikely.

The modest ensemble of mutations relative to the WT that are currently known to

reduce vaccine efficacy is likely to comprise the majority of all possible escape

mutations for future variants, predicting continued efficacy of the existing vaccines.

Methods

Selection of Crystal Structures

In this work, we considered a single crystal structure of the SARS-CoV-2 spike protein
Receptor Binding Domain (RBD) in complex with the human receptor, Angiotensin
Converting Enzyme 2 (ACE2), PDB:6M0J(16). While there are likely multiple mutations

outside of the RBD which significantly affect binding characteristics of the spike protein,
as has been demonstrated for site 614(36), the structure of the RBD itself is unlikely to
be substantially modified by such mutations. Thus, while only able to reveal a subset of
mutations of interest, the focused study of RBD complexes presented here remains
biologically realistic.

Similarly, we consider a single crystal structure of the RBD in complex with a
neutralizing antibody (NAb),CV30 PDB:6XE1(37). CV30 was recognized early on as the
most potent NAb observed within the sera of a SARS-CoV-2 positive donor while the
majority of antibodies were found to target non-neutralizing epitopes outside of the
RBD(38). Subsequently, it became clear that antibodies targeting epitopes outside the
RBD, particularly those targeting the N-terminal domain (NTD), likely play a protective
role and mutations reducing the affinity of these antibodies may be epidemiologically
significant(35, 39). As acknowledged above regarding the selection of the spike crystal
structure, while our focus on CV30 is only able to reveal a subset of mutations of
interest, the mutations discussed in this work predicted to affect CV30 binding affinity
are likely highly epidemiologically relevant.

Construction of Representative Ensembles of Interface Conformations

Protein crystal structures may differ substantially from the native conformations(40).
Throughout this work, we utilize the Rosetta(13) software suite to approximate both wild
type (WT) and mutant conformations of the receptor and NAb complexes. All protocols
used throughout were implemented using the RosettaScripts(41) package and the XML
files used along with the associated executed command lines are made available in
Table S1. Approximation of the native conformational ensemble may be separated into
two steps, identifying the optimal side chain conformation (repacking) and moving the
protein backbone (minimization), to minimize the energy function applied. This may be
accomplished using the Rosetta Relax application which iteratively applies each of
these two steps.

Beginning with the crystal structure, we iteratively applied the FastRelaxMover using
default parameters (with the exception of disabling design) for up 12 iterations(15 for
Omicron) and up to 1000 repeats. We found the total score to be insensitive to
additional applications of FastRelax after 5 iterations on average. Each resulting
structure was scored using the InterfaceAnalyzerMover, repacking the unbound state
but not the bound state (as the input complex has already been optimized).

This protocol returns the total score, S, in arbitrary units produced by the empirically-
driven Rosetta Energy Function 2015(18) (labelled REU for "Rosetta Energy Units",
https://new.rosettacommons.org/docs/latest/rosetta_basics/Units-in-Rosetta), as well as
dG separated which is the difference in the total score between the bound (complex)
and unbound state, S^C-S^U, derived from separating the binding partners. This protocol

may also be used to identify the residues within the complex which constitute the
interface.

Such an ensemble of structures often forms an "energy funnel"(42) where the root mean
square distance (RMSD) between superimposed backbone carbon atoms of each
structure and the structure with the lowest total score or the lowest dG separated is
positively correlated with the total score of that structure. In order to evaluate whether
such a funnel exists for these ensembles, we identified the structure with the lowest dG
separated and 90 residues predicted to be interface residues within the RBD in at least
one conformation in addition to the (+/-) 3 adjacent amino acid neighborhood of each
such residue (sites 400-424, 440-464, and 470-509 in the spike protein for WT;
additional sites 436-439 for Delta; additional sites 434-439 for Gamma; additional sites
434-439 and 510 for Omicron).

Figure S6 displays dG separated vs the interface RMSD for both complexes for the WT.
Few structures appear in the lower left corner of each plot from only one or two (NAb or
ACE2 respectively) independent "trajectories" of iterative FastRelax application
beginning with the crystal structure. Furthermore, while the lowest dG separated is more
than 30% greater in magnitude than the highest for both complexes, the interface
RMSD between any complex and the minimum dG separated complex is less than an
angstrom. These findings suggest selecting the single minimum dG separated
conformation for either complex is unlikely to constitute a realistic model of the native
interface and may in fact represent an unrealistic, entropically disfavored state(43).

Figures S7-9 display dG separated vs the interface RMSD for both complexes for the
Delta, Gamma, and Omicron variants. The distribution of conformations for the NAb is
similar to the WT for both Delta and Gamma; however, the minimum dG separated
obtained for Gamma is higher than that for the WT suggestive of antibody
destabilization for this variant. dG separated for Omicron is substantially higher
indicating significant destabilization. For Delta and Omicron, the distribution of
conformations for ACE2 more resembles a funnel suggestive of receptor stabilization for
these variants. For Gamma, there appear to be 2-3 distinct, equally low energy
conformations for ACE2 which makes the interpretation of the energy landscape more
challenging. For consistency, representative conformations were selected according to
the same protocol for all variants.

The construction of an ensemble of representative conformations is desired so that the
average behavior of such an ensemble is likely to reflect that of the native complex.
Ideally all available structures would be statistically weighted and included in this
ensemble; however, this is computationally intractable. Instead we selected 50
conformations for each complex as follows (see Figures S10-13): 1) The conformation
corresponding to the minimum total score after any iteration of FastRelax from each
independent "trajectory" beginning with the crystal structure was selected. 2) The 10
structures with the lowest total score were removed and the 10 structures with the
lowest dG separated were removed (these are not identical structures and as discussed

above, may represent unrealistic, entropically disfavored conformations). 3) The
remaining structures were ranked by total score, r_S, and dG separated, r_G, and the 50
structures with the lowest composite rank, r_S+r_G, were selected. The PDB files for these
structures are made available (see Data Availability). We believe the equally-weighted
average behavior of these conformations, which are stable with well-resolved interfaces,
constitute a reasonable model of the native complexes.

Analysis of Single Mutants Relative to WT, Delta, and Gamma

Given the 100 structures selected as described above (50 for each complex), a more
restrictive list of 52 residues were predicted to lie at the interface of at least one
structure for WT/Delta (spike protein sites 403-406,409,414-417,419-421,445-447,
449,453,455-461,473-478,484-498,500-506), 56 for Gamma
(WT/Delta+408,439,480,499), and 53 for Omicron (WT/Delta+439,499,-445). We
introduced all of the 19 possible mutations at each interface site (all 56 identified for any
variant) for these 100 structures for the WT, Delta, Gamma, and Omicron variants with
the exception of WT reversion for the variants. Mutations were introduced using the
PackRotamersMover with design specified by the input resfile (see Data Availability for
example). Only the targeted residue was modified: side chain conformations for all other
residues were fixed and no backbone minimization was applied before filtering for
candidates of interest as described below.

Introducing the mutation in this way without repacking and minimization results in many
unrealistic, high-energy conformations which are difficult or impossible to interpret
without further optimization. The benefit is speed. This procedure can be executed in
under 30 seconds per structure while global repacking and minimization may take
hours. An alternative approach is to apply backbone minimization and sidechain
repacking within a local region of the structure centered at the modified site as
implemented within Rosetta through the Flex ddG protocol(44). This approach has been
used to accurately predict mutations conferring increased infectivity for SARS-CoV-
2(45) and rationally design NAb against the antigen(46, 47) but is significantly slower
than introducing mutations without, even local, optimization. With or without local
optimization prior to filtering, global repacking and minimization may be desired to
confirm top candidates. While our approach may introduce a higher false-negative rate
than what one would achieve applying local optimization prior to filtering, it maximizes
the breadth of candidate mutations considered and is able to recapitulate experimental
results.

The total score and dG separated of the mutant structures were computed. Unlike in
earlier steps where the unbound state was repacked during the dG separated
calculation, no repacking was conducted for the reasons described above. Each mutant
structure was matched with its initial WT/variant conformation and the change in dG
separated, $(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)$, and total score, $S_M^C-S_{WT}^C$, were calculated. The
average of each value taken over the ensemble of the 50 RBD-ACE2 conformations

was compared with experimentally determined ACE2 binding affinity for single RBD mutants(12). While the change in dG separated is technically the ddG for the mutant and may be expected to correspond to binding affinity, we find $(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)$ over this ensemble is not generally predictive of ACE2 affinity as $(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)$ is approximately zero for many mutants (Figure S14). This may be expected without repacking and minimization. Nonetheless, when nonzero, the change in dG separated compares favorably with ACE2 affinity (see below).

The change in total score is less sensitive and an adequate predictor of affinity (Figure S15) within the range $S_M^C-S_{WT}^C<5$ such that the minimum relative binding affinity decreases with increasing change in total score. Additional validation that $S_M^C-S_{WT}^C$ is a reasonable predictor of binding affinity, below some threshold, may be obtained through demonstrating $S_M^C-S_{WT}^C$ for each mutant is highly correlated between the RBD-ACE2 complex and the RBD-NAb complex. Furthermore, when the difference between the change in dG separated for the NAb complex and the dG separated for the ACE2 complex, $[(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)]_{NAb}-[(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)]_{ACE2}$, is positive, mutants lie above the identity line (y==x). The reverse is true for mutants falling below the identity line (Figures 1C/3B). In other words, on average, mutants in the RBD-ACE2 complex behave similarly to mutants in the RBD-NAb complex (which is what one would expect for realistic models of interfaces with an overlapping footprint). When differences do appear, they correspond to interactions with the RBD which are specific to the binding partner (ACE2/NAb) and are correctly reflected by the change in dG separated.

Directly Assessing Additive Effects Vs Epistasis

In the main text, we demonstrate that most mutations appear in a similar position of the receptor cost / antibody cost phase space for the WT and Gamma/Delta variants. Consequently, the ensemble of vaccine escape candidates is largely conserved. Whether epistatic effects due to multiple mutations are present or if the impact of multiple mutations is simply additive can be more directly assessed for each complex by plotting the change in the total score after introducing each mutation in the variant against the change in total score after introducing each mutation in the WT. Figure S5 displays the change in total score for each interface mutation relative to the WT and all variants for both ACE2 and the NAb. The minimum value (less 1) is subtracted from each distribution. The effect of each mutation at the interface in the WT is highly correlated for the Delta variant. This correlation is observed for the Gamma and Omicron variants as well with more significant variation as expected. There is additionally a small systematic bias towards greater NAb destabilization for mutations with small changes in the total score for the Omicron variant. As discussed in the main text, while this epistatic signature is not observed for the Gamma variant, this trend did not significantly alter the landscape of escape-exacerbating mutations.

Alternative Construction of Variant Ensembles

Beginning with the conformational ensemble constructed for the WT, we introduced the variant mutations 452R|478K for the Delta variant, 417T|484K|501Y for the Gamma variant, and 339D|371L|373P|375F|417N|440K|446S|477N|478K|484A|493R|496S| 498R|501Y|505H for the Omicron variant and completed 5 rounds of the FastRelaxMover (15 for the Omicron variant) using default parameters (with the exception of disabling design) as described above. The resulting values for the total score and dG separated were comparable to that of the ensembles constructed for the variants beginning with the WT crystal structure (Figures S16-18); however, some structures displayed footprints more similar to the WT than those of the reference ensemble constructed as described in the main text. Thus, this computationally cheap alternative may be utilized for rapid evaluation; but the more comprehensive exploration of the conformational space described in the main is likely desired in many circumstances.

Data Availability

The data has been deposited through Zenodo(48) (https://doi.org/10.5281/zenodo.5297698) including GISAID acknowledgements. Previously published data were used for this work: GISAID(23). Data is additionally made available through FTP: https://ftp.ncbi.nih.gov/pub/wolf/_suppl/SARSstruct21/

Author contributions

NDR, and GF collected data; NDR, YIW, GF, PF, FZ, and EVK analyzed data; NDR and EVK wrote the manuscript that was edited and approved by all authors.

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Figure legends

Figure 1. Landscape of Vaccine Escape Mutants for the WT RBD

A. Cartoon depicting unique conformational changes to the RBD (blue) in complex with ACE2 (orange) and the NAb (green) associated with the same mutation. **B.** Cartoon depicting the landscape of vaccine escape mutations (the plane of receptor cost vs antibody cost). **C.** Landscape of vaccine escape mutations for the WT RBD. Circles with a black outline are NAb escape candidates. Color indicates propensity for escape as measured by $\varDelta\varDelta G$.

Figure 2. Dominant Trends in Circulating RBD Mutations

A. Region-normalized global prevalence of the top 10 most common combinations of RBD mutations over time. Lines are solid up to peak prevalence and dashed afterwards. Shading indicates confidence intervals. **B.** Structural comparison of the complexes of ACE2 and NAb with RBD for WT, Delta, Gamma, and Omicron variants. *Top*: WT footprints including the residues with an interaction within 4A of the partner. *Bottom:* Visualization of the Delta, Gamma, and Omicron variant interfaces. Mutations are labeled and represented as sticks. WT structures are superimposed for the RBD of each variant: WT, orange; variant, olive **C.** Interface RMSD for ACE2 and NAb complexes relative to an arbitrary WT conformation (over spike protein sites 403-406,408-409,414-417,419-421,439,445-447,449,453,455-461,473-478,480,484-506). Asterisks denote p-values less than 0.02 for a Wilcoxon Rank Sum Test. **D.** CMDS applied to the pairwise RMSDs among all RBD-ACE2 and RBD-NAb complexes. 2D visualizations of 3D projections are displayed (see Figure S2 for 3D).

Figure 3. Epistasis within the RBD.

A. Cartoon illustrating additive and non-additive (epistatic) interactions between mutations. From top to bottom: additive, escape-neutral, escape-minimizing, and escape-exacerbating. *Mi, Mj, Mij* denote the effects of the single and double mutants. **B.** *Top:* Landscape of vaccine escape mutations for the variant RBDs. Coloring, as in Figure 1C, indicates propensity for escape as measured by $\varDelta\varDelta G$. Circles with a black

outline denote NAb escape candidates. Dashed lines highlight differences among

variants. *Bottom:* Landscape of vaccine escape mutations for the WT RBD. Black points

are candidates for both WT and variant; gray points are not candidates for either WT or

variant; green points are only candidates for WT; red points are only candidates for the

variant.

Figure 4. Landscape of Epistatic Effects Supporting Enhanced Vaccine Escape

Non-additive escape-exacerbating motifs in Delta (top), Gamma (middle), and Omicron

(bottom) variants. The size of each letter corresponds to the increased likelihood of

vaccine escape for the substitution in the variant relative to the WT. *Inset*: Total score

change ($[S_M^C - S_{WT}^C]_{NAb}$) induced by mutation in the Gamma (black) and Omicron

variants (red) vs the WT (identity line overlaid). The minimum value (less 1) is

subtracted from each distribution for normalization (see Figure S5 for more).

Table 1: Receptor-binding and antibody-binding interface footprints in the RBD

WT RBD-ACE2 Footprint	403, 405, 417, 445-447, 449, 453, 455-456, 473-478, 484-491, 493-498, 500-506
Delta RBD-ACE2 Footprint	Same as WT
Gamma RBD-ACE2 Footprint	WT + 404, 406, 408, 439, 499 - 484
Omicron RBD-ACE2 Footprint	WT + 404, 439, 499 - 445, 484, 491
WT RBD-NAb Footprint	403-406, 409, 414-417, 419-421, 446-447, 449, 453, 455-461, 473-478, 484-498, 500-506
Delta RBD-NAb Footprint	Same as WT
Gamma RBD-NAb Footprint	WT + 408, 480
Omicron RBD-NAb Footprint	WT - 446

880 **Table 2: Numbers of antibody-escape candidates**

	Total		WT not Variant	Variant not WT	Difference
WT	241	Δ	15(13)	6(2)	-9(-11)
Delta	232	Γ	32(28)	86(66)	54(38)
Gamma	295	O	67(59)	75(48)	8(-11)
Omicron	249				

883 Values in parentheses are tabulated mutations with $[S_M^C - S_{WT}^C]_{ACE2} < 13$, for both the

884 WT and variant.

A WT M_i

ACE2
RBD

RBD NAb

B

Antibody Cost

Escape | Not Viable

Neutral/Escape | Not Viable

Receptor Cost

C $[(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)]_{NAb} - [(S_M^C-S_M^U)-(S_{WT}^C-S_{WT}^U)]_{ACE2}$ **[REU]**

$[S_M^C-S_{WT}^C]_{NAb}$ [REU]

$[S_M^C-S_{WT}^C]_{ACE2}$ [REU]