A biologically accurate model of directional hearing in the parasitoid fly *Ormia ochracea*

Max R. Mikel-Stites^{a,b,c}, Mary K. Salcedo^a, John J. Socha^a, Paul E. Marek^d, and Anne E. Staples^{a,b,c}

^a Department of Biomedical Engineering and Mechanics, Virginia Tech, Blacksburg, VA, 24061, USA; ^bEngineering Mechanics program, Virginia Tech, Blacksburg, VA, 24061, USA; ^cDepartment of Mathematics, Virginia Tech, Blacksburg, VA, 24061, USA; ^dDepartment of Entomology, Virginia Tech, Blacksburg, VA, 24061, USA;

This manuscript was compiled on October 5, 2021

Although most binaural organisms localize sound sources using 1 neurological structures to amplify the sounds they hear, some an-2 imals use mechanically coupled hearing organs instead. One of 3 these animals, the parasitoid fly Ormia ochracea, has astoundingly accurate sound localization abilities and can locate objects in the az-5 imuthal plane with a precision of 2°, equal to that of humans. This 6 is accomplished despite an intertympanal distance of only 0.5 mm, which is less than 1/100th of the wavelength of the sound emitted by the crickets that it parasitizes. In 1995, Miles et al. developed 9 a model of hearing mechanics in O. ochracea, which works well for 10 incoming sound angles of less than $\pm 30^\circ$, but suffers from reduced 11 accuracy (up to 60% error) at higher angles. Even with this limita-12 tion, it has served as the basis for multiple bio-inspired microphone 13 designs for decades. Here, we present critical improvements to the 14 classic O. ochracea hearing model based on information from 3D re-15 constructions of O. ochracea's tympana. The 3D images reveal that 16 the tympanal organ has curved lateral faces in addition to the flat 17 front-facing prosternal membranes represented in the Miles model. 18 To mimic these faces, we incorporated spatially-varying spring and 19 damper coefficients that respond asymmetrically to incident sound 20 waves, making a new quasi-two-dimensional (q2D) model. The q2D 21 model has high accuracy (average errors of less than 10%) for the en-22 tire range of incoming sound angles. This improved biomechanical 23 hearing model can inform the development of new technologies and 24 may help to play a key role in developing improved hearing aids. 25

acoustics | binaural hearing | biomechanics | mathematical modeling | sensory modeling | *Ormia ochracea*| parasitoidism | bioinspiration

he ability to localize sound allows animals to avoid preda-2 tors and assists them in finding mates and capturing prey. Binaural organisms, those with two ears, locate sound-3 emitting objects by comparing the intensity and timing of 4 incident sound waves arriving at their two hearing organs (Fig. 5 1A). Sound localization in binaural organisms is commonly described using two metrics: 1) interaural time delay (ITD), 7 the difference in time it takes sound to reach the two hearing organs, and 2) interaurual amplitude difference (IAD), the difference in sound amplitude between the two organs (Fig. 10 1A) (1). 11

In vertebrates, ITD is calculated in the superior olivary 12 nucleus of the brain stem and IAD is calculated in the infe-13 14 rior colliculus in the midbrain nucleus (2). By comparison, many invertebrates lack significant neural investment in cen-15 tral processing and rely heavily on mechanical structures to 16 pre-process sensory signals (3-5). The particulars of sound 17 localization are complex and vary widely among animals, as 18 the ITD and IAD ranges experienced by binaural animals 19 demonstrate (Fig. 1B) and the biophysics of sound localiza-20 tion for specific species are often too complex to be modeled 21 well by simple analytical models. However, simplistic insect 22

hearing models can be used to extract key principles of binaural hearing without complex physiological modeling of neural processes. 25

For the tachinid fly Ormia ochracea, the ability to hear 26 its host plays a key role in its reproductive cycle and over-27 all fitness. As a parasitoid, O. ochracea listens to chirping 28 male crickets and follows the sound back to the source, where 29 female O. ochracea then deposit their larvae (6). Gravid O. 30 ochracea females will remain in an area for extended peri-31 ods of time in response to cricket chirping sounds, even if 32 no cricket is present (7). Given its small size, if O. ochracea 33 relied exclusively on the distance between its prosternal tym-34 panal membranes (Fig. 1C), the ITD it experienced would 35 be at the nanosecond scale or below (far too small to reli-36 ably perceive differences in azimuthal angle between sound 37 source locations), and there would be no practical difference 38 in sound amplitude between the two membranes (IAD). To 39 solve this scaling problem without resorting to bulky neurolog-40 ical investment, O. ochracea have two mechanically coupled 41 membranous tympana directly beneath their head (Fig. 1C). 42 These coupled tympana are composed of a pair of prosternal 43 membranes, joined together by an intertympanal bridge (8), 44 and are significantly larger in female Ormia (Fig. 1C). This 45 distinctive mechanical coupling serves to increase the ITD and 46

Significance Statement

The ability to identify the location of sound sources is critical to organismal survival and for technologies that minimize unwanted background noise, such as directional microphones for hearing aids. Because of its exceptional auditory system, the parasitoid fly *Ormia ochracea* has served as an important model for binaural hearing and a source of bioinspiration for building tiny directional microphones with outsized sound localization abilities. Here, we performed 3D imaging of the fly's tympanal organs and used the morphological information to improve the current model for hearing in *O. ochracea*. This model greatly expands the range of biological accuracy from $\pm 30^{\circ}$ to all incoming sound angles, providing a new avenue for studies of binaural hearing and further inspiration for fly-inspired technologies.

Author contributions: M.R.M.-S. developed the modified model; M.R.M.-S. and A.E.S. designed and wrote the code; M.K.S. and J.J.S. performed 3D imaging data collection and analysis; M.R.M.-S. produced the hearing model data; M.R.M.-S. and A.E.S. analyzed the hearing model data; M.R.M.-S. and M.K.S. made the figures; P.E.M. performed photographic work; M.R.M.-S. wrote the paper, and M.R.M.-S., M.K.S, A.E.S, J.J.S, and P.E.M. edited the paper.

The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

²E-mail: staplesa@vt.edu

IAD perceived by the fly. We will refer to the increased ITD 47 and IAD as mITD (mechanical ITD) and mIAD (mechanical 48 IAD), respectively. These mechanically amplified values allow 49 it to successful localize chirping crickets. Since O. ochracea 50 51 are active at or after dusk, they also use their hearing to avoid 52 predation by bats, exhibiting a startle response while in flight to bat sonar frequency sound, similar to preying mantises 53 (9, 10). As such, the ability to accurately and quickly locate 54 a source of incoming sound at high levels of lateral angular 55 resolution is a significant advantage, especially in potentially 56 noisy environments (11), suggesting that O. ochracea may be 57 a good source of bioinspiration to tackle the so-called cocktail 58 party problem (12) (isolating sounds in a noisy environment) 59 for directional microphones and hearing aids. 60

Accurate models of binaural hearing in animals are gener-61 ally highly complex. Because of the mechanical nature of its 62 acoustic sensing organ, O. ochracea is one of the few excep-63 tions, and it has been the focus of numerous studies featuring 64 its uniquely "simple" hearing organs and how they function 65 (8, 9, 13-19). To investigate the biomechanical mechanisms 66 that underlie O. ochracea's unusual hearing abilities, Miles, 67 Robert, and Hoy developed mechanical and mathematical mod-68 els of the ormiine's coupled tympana in 1995 (15). The authors 69 validated their model against experimental data, recording 70 tympanal membrane positions and velocities, and consequently 71 72 mIAD and mITD, as a function of the incident sound pressure, intensity, and angle. The Miles model becomes analytically 73 solvable under the assumptions of continuous sinusoidal input 74 and symmetric model parameters, in addition to being numer-75 ically solvable without requiring the assumptions of symmetry 76 or continuity. The model allowed Miles et al. to demonstrate 77 that O. ochracea's impressive sound localization abilities are 78 due to the pre-processing performed by their structurally cou-79 pled tympana, which mechanically amplify the ITD and IAD 80 experienced by the fly. 81

In addition to providing a physiological explanation for O. 82 ochracea's localization prowess, the Miles model also accurately 83 predicted mITD for all incoming sound angles and mIAD for 84 angles below $\pm 30^{\circ}$ in a sample *O. ochracea* population. Both 85 the measured and predicted mITD indicated that O. ochracea 86 possesses an mITD comparable to the ITD of an animal closer 87 in size to a rat (Fig. 1B). Later experiments successfully 88 determined that O. ochracea has a sound localization precision 89 in the azimuthal plane of 2° (20, 21), a precision comparable 90 to that of humans. This high precision, together with the 91 relative simplicity of the model and the easily reproducible 92 93 structure of the hearing mechanism used by O. ochracea led to a new stream of research in ochracea-inspired designs for 94 directional microphones and hearing aids (22–28). Despite its 95 utility, the model contains a number of simplifications that 96 limit its biological accuracy. 97

The Miles model is a lumped-element model that primar-98 ily considers the dynamics of the intertympanal bridge and 99 the front of the tympanal membranes (Fig. 1C), modeling 100 each membrane as a flat plate with a purely one-dimensional 101 amplitude response. The model's spring and damper coeffi-102 cients were adjusted until the model response approximated 103 the experimental responses in recently deceased O. ochracea 104 specimens measured using laser-vibrometry. Although the 105 model is relatively accurate for mITD in a narrow range of 106 incident sound angles, it displays significant errors in mIAD 107

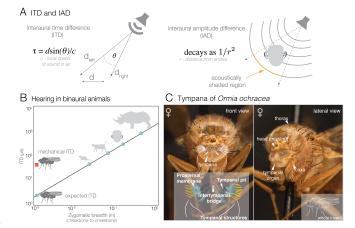


Fig. 1. Hearing in binaural animals and the fly Ormia ochracea. (A) Physical meaning of ITD and IAD. ITD is the time delay between sound reaching one sensory organ relative to the other, defined by the equation $\tau = dsin(\theta)/c$, where d is the distance between the hearing organs and \boldsymbol{c} is the speed of sound in air. IAD is the difference in response amplitude between the left and right sensory organs, due to acoustic shading or signal decay. (B) Approximate ITD values in representative animals (O. ochracea, rat, cat, human, and rhinoceros) for a sound source at a 45° angle from the midline, calculated using the formula in (A), with the zyogmatic breadth (cheekbone-to-cheekbone distance) used as an approximate measure of interaural distance for mammals. Data from (29-32). (C) Female O. ochracea post-decapitation, showing location of prosternal membrane, tympanal pits, and intertympanal bridge. key physical features in the modeling of its binaural hearing.

for incident sound angles larger than approximately $\pm 30^{\circ}$ from the midline of the fly, and mITD becomes increasingly inaccurate at angles above approximately $\pm 40^{\circ}$. This inaccuracy 110 across large angles limits the model's power for explaining 111 binaural hearing in *O. ochracea* and its potential for inspiring 112 new hearing-based engineering. 113

Previous scanning electron microscopy images of O. 114 ochracea tympana had indicated that a certain degree of dy-115 namic curvature and morphological complexity was present 116 (8). It was excluded, however, primarily to avoid increasing 117 model complexity. We hypothesized that inclusion of 3D fea-118 tures could improve model accuracy and extend the effective 119 range of the model, allowing it to predict more accurate tym-120 panal displacements for incoming sound at high angles when 121 compared to experimental data. To identify the sources of 122 inaccuracy in the model, we investigated the detailed morpho-123 logical structures involved. Using 3D reconstructions of O. 124 ochracea's tympana as a guide, we modified the original model 125 by adding terms that simulate the mechanics of the hearing 126 organ in the lateral plane. We represent these lateral mechan-127 ics mathematically via a spatially-dependent asymmetry in 128 the model spring and damper coefficients. 129

Materials and Methods

Synchrotron x-ray imaging of the ormiine tympanal organ. To 131 examine the 3D nature of Ormia ochracea's tympanal mor-132 phology, we performed tomographic imaging of preserved O. 133 ochracea specimens using the synchrotron x-rays at the Ad-134 vanced Photon Source at Argonne National Laboratory. Two 135 O. ochracea dried specimens were borrowed from the Virginia 136 Tech Insect Collection. The specimens were placed in slender 137 tubes made of polyimide, and the ventral thorax was imaged 138 using beamline 2-BM. Each specimen was imaged using the 139

108

109

130

t40 beamline's fast 2D phase contrast imaging, giving stacks of t41 images along the z-axis at intervals of 1.72 μ m.

Raw microtomographic images were cropped and downsampled using FIJI (33) and segmented in SlicerMorph, (34) an imaging extension of 3D Slicer. To segment, features of the tympana were highlighted and then rendered in 3D for applicable measurements. The three-dimensional scans are available upon request.

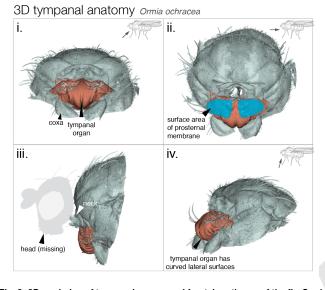


Fig. 2. 3D rendering of tympanal organs and frontal prothorax of the fly Ormia ochracea. Tympanal membranes highlighted in blue (ii.), with supporting structures highlighted in peach. Orientation of image relative to *O. ochracea* body indicated in schematics at top right of images. 3D images made in SlicerMorph software. (34)

Previous model. The previous model of binaural hearing in 148 O. ochracea includes two components: a mechanical model of 149 the anatomy and a corresponding mathematical model. The 150 mechanical model (15) treats the tympanal structure as a 151 pair of beams pinned at a central pivot, with lumped-mass 152 approximations of the two sides of the hearing organ located 153 at the ends of the beams (Fig. 3A,B). The beams are anchored 154 to the substrate at their distal ends with a pair of symmetric 155 spring-damper elements, and to each other with a third spring-156 damper element (Fig. 3B). Pressure forces from incident sound 157 waves are applied to the point masses via a forcing function 158 composed of the product of the incident pressure magnitude, 159 the inward-facing unit normal vector, and the tympanal surface 160 area, A (see the Supplemental Material for numerical values 161 used in this study). A time delay is applied between the left 162 and right sides based on the angle θ the incoming sound wave 163 has relative to the midline of the fly, with 0° defined as straight 164 ahead (15). 165

The mathematical model is a set of coupled ordinary differential equations that are the equations of motion for the mechanical model. It treats the incident acoustic pressure acting on the tympanal membranes as two point forces, $f_1(t)$ and $f_2(t)$, acting on the point masses representing the tympanal membranes and associated structures. The dependent variable in the problem is $\mathbf{x}(t)$, which represents the one-dimensional response of each tympanum. The model can be written as:

$$\begin{bmatrix} k_1 + k_3 & k_3 \\ k_3 & k_2 + k_3 \end{bmatrix} \mathbf{x} + \begin{bmatrix} c_1 + c_3 & c_3 \\ c_3 & c_2 + c_3 \end{bmatrix} \dot{\mathbf{x}} + \begin{bmatrix} m & 0 \\ 0 & m \end{bmatrix} \ddot{\mathbf{x}} = \mathbf{f},$$
[1] 174

$$\mathbf{f} = \begin{bmatrix} f(t)\\ f(t+\delta t) \end{bmatrix}$$
[2] 176

where $\mathbf{x} = (x_1(t), x_2(t))$ is the unknown response vector con-177 taining the vertical displacement of the left and rightmost 178 tips of the beams in Figure 3B, which represent the two sides 179 of the intertympanal cuticular bridge, the applied force is 180 $\mathbf{f} = (f_1(t), f_2(t)), \text{ and } ()$ represents differentiation with re-181 spect to time, t. The parameters k_i and c_i are spring stiffness 182 and damper constants, respectively, and the parameter m183 is the effective mass of all the moving parts of the auditory 184 system (15).

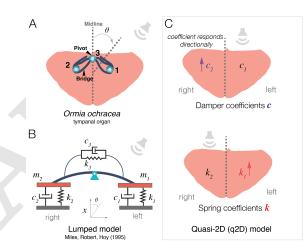


Fig. 3. Modeling of binaural hearing in the fly Ormia ochracea. (A) Schematic of the coupled tympanal membranes of *O. ochracea* (peach-colored, labeled 1 and 2), connected by the cuticular bridge (blue) with sound incident at θ degrees. (B) The hearing system can be represented as a pair of coupled beams joined and anchored by a set of springs and dampers (adapted from Liu *et al.* (35). (C) The q2D model has an asymmetric response: the spring and damper coefficients on the contraleral (opposite) side from the sound source increase as a function of incident sound angle, while the coefficients on the ipsialateral side remain constant.

Q2D model modifications based on ormiine morphology. In Miles *et al.*'s analysis of their model, the ormiine hearing structure is assumed to be left-right symmetrical, and the spring and damper coefficients on the right and left sides are identical and constant for all incident sound angles, with $k_1 =$ $k_2 = k$ and $c_1 = c_2 = c$, independent of the values of k_3 and c_3 .

In order to add a realistic degree of sensitivity to the angle 193 of the incoming sound, we modified the spring and damper 194 parameters to incorporate aspects of the 3D morphology of 195 the fly's hearing organ. Specifically, we did this by treating 196 the magnitude of k and c as functions of the incoming sound 197 angle. The functions were structured such that for an incident 198 sound angle above $\pm 30^{\circ}$, the k and c values corresponding 199 to the contralateral tympanum are increased compared to 200 those for the ipsilateral tympanum, mimicking the presence of 201 lateral sides on the tympana, which can both shield the rest 202 [3]

[4]

[5]

of the structure and be more responsive to laterally oriented
incoming sounds (Fig. 3C). We provided the following quasitwo-dimensional modification to the Miles model of ormiine
hearing:

or
$$k(\theta) = \begin{cases} k_0 & \text{if } \theta < |30^\circ| \\ \alpha |\theta| k_0 + \beta & \text{if } |55^\circ| > \theta \ge |30^\circ| \\ k_f & \text{if } \theta \ge |55^\circ| \end{cases}$$

$$\alpha = \frac{k_f - k_0}{25^\circ}$$

209
210
$$\beta = \frac{2.6k_0}{25^\circ}$$

where k_0 and k_f are the minimum and maximum values that 211 the spring stiffness coefficients can take on, respectively. The 212 form of the modified spring coefficient function, two constant 213 segments with a linear ramp between $|30|^{\circ}$ and $|55|^{\circ}$ (Fig. 3C, 214 Fig. 4A), was informed by the lateralization behavior observed 215 in O. ochracea (20) and the analysis of an O. ochracea-inspired 216 sensor (35). These works indicated the presence of two separate 217 behavioral regimes, a localization regime from 0° to $\leq |30|^{\circ}$ and 218 a lateralization regime at higher angles. This choice is further 219 supported by the accuracy of the fit to experimental data 220 for sound incident at $\geq |30|^{\circ}$ (Fig. 4B,C), and physically 221 represents a degree of elastic response to incoming sound 222 223 waves in the lateral direction .

The constants in equations 3-5 were chosen to provide 224 the best fit to the available behavioral data (15): mITD and 225 mIAD derived from laser-vibrometry measurements of tympa-226 nal membrane vibrations in O. ochracea specimens in response 227 to a 6 kHz sound source, as a function of incident sound an-228 gle. The coefficients are only modified on the contralateral 229 side and remain constant for the side on which the sound 230 source is located. As the incident sound angle approaches 231 $\pm 90^{\circ}$ relative to the fly's head, the spring coefficient for the 232 contralateral side increases from k_0 , and approaches k_f accord-233 ing to Equation 3. For example, for sound incident from 30° . 234 the spring and damper coefficients for the left side, k_1 and c_1 , 235 would change and k_2 and c_2 would remain unchanged. We 236 assume the total tympanal surface area, A, is fixed, and we 237 use previously established values (15) throughout this work 238 $(A = 0.288 \times 10^{-6} m^2)$. The increases in the spring and damper 239 coefficients, normalized relative to their nominal values (k_0) , 240 are visible in Figure 4A. 241

MATLAB's ODE45 function was used to integrate equations 1-2 and a custom peak-finding algorithm was implemented to calculate mITD and mIAD. Further computational details and a link to representative code samples can be found in the Supplemental Material.

247 Results

2

The tympana of O. ochracea protrude anteriorly from un-248 249 derneath the cervix (fly's neck), with distinct lateral faces and sharp curvature (Fig. 2). Figure 2 shows 3D surface 250 renderings of O. ochracea tympanal membranes in teal, with 251 the supporting structures highlighted in peach. The organs 252 are far from the simple two-dimensional surfaces most often 253 depicted in the literature (14, 15, 28, 35). These new 3D 254 models motivated our modifications to include aspects of ac-255 tual morphology. The confirmation of significant lateral-facing 256 portions of the tympana led to the modifications present in 257

the q2D model (equations 3-5), which account for the lateral tymapanal response to acoustic stimuli.

Values of mITD and mIAD, calculated from the q2D and 260 Miles models, are shown in Figure 4B as a function of incident 261 sound angle, and are compared to experimental measurements 262 in recently sacrificed O. ochracea specimens (15). Both models 263 are identical for incident sound angles less than $\pm 30^{\circ}$, so 264 the results are identical within that range (Fig. 4C, grav 265 box). When we included the lateral response through the 266 new $k(\theta)$ and $c(\theta)$ functions, the gap between experimental 267 measurements and model results in both mIAD and mITD 268 narrowed significantly for 6 kHz signal input (Fig. 4B,C), with 269 the q2D model having average error of approximately 6% and 270 a peak error of approximately 28% in mITD, and an average 271 error of approximately 7% and a peak error of approximately 272 10% in mIAD. These results additionally confirm that aspects 273 of mechanics in two dimensions are important elements of 274 ormiine hearing. 275

Discussion

In this paper, we present the results of 3D X-ray synchrotron 277 imaging of the mechanically-coupled tympana in the para-278 sitoid fly, Ormia ochracea, and our subsequent modification 279 to the classic mathematical model of hearing in O. ochracea 280 inspired by those results. The tympanal organ was confirmed 281 to be highly 3D, with significant lateral-facing membranes, 282 in contrast to the commonly simplified representation of the 283 membranes as flat, front-facing plates. 284

Detailed knowledge of the hearing organ's morphology al-285 lowed us to update the classic 1995 one-dimensional mathe-286 matical model into a quasi-two-dimensional model of ormiine 287 hearing that mimics the tympanal organ response in the lateral 288 direction. Our updated q2D model has significantly improved 289 fidelity to available experimental data (15) compared to the 290 Miles model, both in the mechanical interaural time delay 291 (mITD) and in the mechanical interaural amplitude differ-292 ence (mIAD) (Fig.4B,C). When compared to the Miles model, 293 the new q2D model exhibits maximum errors (relative to ex-294 perimental values) reduced by approximately 50% and 85% 295 respectively. This strongly supports the premise that there are 296 important aspects of the mechanics of Ormia hearing aside 297 from the response of the front-facing tympanal membranes, 298 and that the entirety of the hearing organ structures are sen-290 sitive to the angle of incoming sound, a feature that was not 300 included in the Miles model. 30

Prior to our study, the original Miles model was the only 302 existing model of ICE (internally coupled ears)-based hearing 303 in ormine flies (36). This is one of the first attempts to update 304 the foundational Miles model for hearing in O. ochracea. Our 305 model may be further refined by incorporating additional me-306 chanical behaviors of the tympana, such as tympanal deflection 307 in the lateral direction or a representation of the tympanal 308 response in the vertical direction. It could also be improved by 309 simple analytic modifications to expand the model's capabili-310 ties without impacting its tractability, such as using functions 311 that are more flexible than simple linear ramps for the spring 312 and damper coefficients. For example, in our g2D model, the 313 "bump" visible near $\pm 45^{\circ}$ in mIAD in Figure 4B and the uptick 314 at the same point in mITD may be a result of the values for 315 either the springs, dampers, or the ratio between the two, 316 being slightly too high at that point. It is also important to 317

276

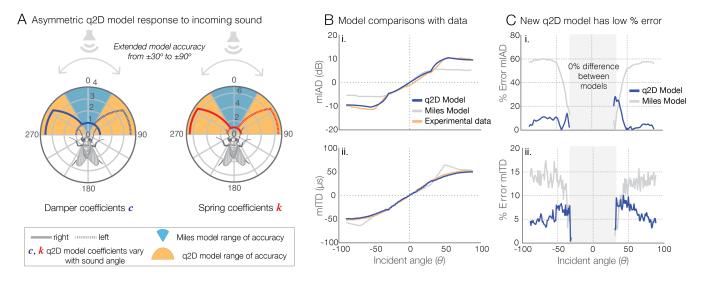


Fig. 4. Model modifications and comparison between the Miles model, experimental data, and q2D model. (A) The modified quasi-two-dimensional (q2D) model shows improved range of accuracy in its response to incident sound. In the q2D model, the normalized damper (red) and spring (blue) coefficients are functions of the incoming sound angle. The improved q2D model responds accurately within $\pm 90^\circ$, compared to $\pm 30^\circ$ for the Miles model. Experimental and model results (B) and error (C) in mITD and mIAD for the standard one-dimensional (Miles model) and q2D models. In (B), mITD and mIAD were calculated from the q2D and Miles models as a function of incident sound angle for a frequency of 6000 Hz, and compared with laser-vibrometry measurements from recently deceased *O. ochracea* specimens (15). The significant divergence from behavioral data present in the Miles model outside $\pm 30^\circ$, particularly for mIAD, is rectified in the modified q2D model. In (C), the gray box indicates errors below $\pm 30^\circ$, which are not considered because the q2D and Miles models are identical for these ranges. The errors for the q2D model peak close to $\pm 30^\circ$, then decrease as the incident sound angle is increased.

note that this work and the Miles model both rely on tuning 318 the coefficients so that the model outputs better match the ex-319 perimental response to sinusoidal input (2 kHz for the original 320 1995 work and 6 kHz for the work here). Although the model's 321 performance was not observed to degrade at other frequencies 322 that we checked, the degree of improvement (relative to the 323 6 kHz experimental data) was far less significant for other 324 frequencies. The model's reduced performance at frequencies 325 other than those tuned specifically for crickets could poten-326 tially be resolved by introducing other morphological features 327 in the form of frequency-dependent functions, in a similar way 328 as we have introduced spatially-dependent functions here. 329

Our model demonstrates that the mechanics of hearing in O. 330 ochracea are dependent on the complex tympanal morphology 331 present in the animal, especially with respect to mIAD, and in-332 dicates that this morphology serves a specific angle-dependent 333 role in responding to incoming sound waves. The inclusion 334 of angle-dependent behavior in the spring and damper coef-335 ficients provides a more accurate understanding of how the 336 insect receives sound. Previous work has demonstrated that 337 O. ochracea engages in different behaviors depending on the 338 relative angle of incoming sound (15, 20, 35, 37), with two 339 distinct response patterns. In the first, from 0° to $\pm 30^{\circ}$, the 340 fly makes relatively narrow adjustments to localize the origin 341 of the sound (localization). In the other, at angles exceed-342 ing approximately $\pm 30^{\circ}$, the fly makes significantly larger 343 adjustments, more akin to determining the side from which 344 the sound originates (lateralization). Our results show that 345 this difference in response is not strictly a result of behavioral 346 differences, but is paired with a difference in physiological 347 responses to incoming sound. 348

Furthermore, there is growing evidence that some *O*. *ochracea* are involved in an evolutionary arms race with their host species (38, 39), and that they are capable of differentiating between different cricket host species based on their 352 acoustic signalling, exhibiting preference towards local popu-353 lations (40). Consequently, the mechanical parameters for the 354 model may depend heavily not only on the geographic origin 355 of O. ochracea samples, but also when collection occurred. 356 The degree of tuning to host-searching behavior, as opposed 357 to predator-avoidance behavior, also remains unaddressed ex-358 perimentally, despite the startle responses when in flight and 359 subjected to sound consistent with bat sonar frequencies (9). 360 O. ochracea also exhibits a sorting behavior (being able to 361 rapidly categorize sounds as belonging to a predator or not) in 362 response to predator-consistent sound sources, as opposed to 363 host or neutral sound sources (9). O. ochracea is also only one 364 of many Ormia species, which parasitize a diverse range hosts, 365 and display different behavioral responses to the acoustic sig-366 nalling of their hosts (7). Only O. ochracea has been examined 367 in sufficient detail to develop a mechanical model with accu-368 rate parameters; consequently, it may be worth investigating 369 the mechanics of other ormine species (7, 41), and developing 370 mechanical models similar to the q2D model presented here. 371 It may also be worth revisiting the hearing organs in Emble-372 masoma, another group of parasitoid flies, which represent 373 a case of convergent evolution in a distantly related family, 374 Sarcophagidae (42, 43). 375

O. ochracea's hearing system has repeatedly served as a 376 source of inspiration for bio-inspired designs for directional 377 microphones and hearing aids (22-28, 35). Including the 378 angle-dependent behavior of the expanded q2D model in fu-379 ture Ormia-inspired device designs may also provide significant 380 avenues for improvement in device performance, or may ex-381 pand the functionality of devices like acoustic sensors through 382 miniaturization and tunable frequency sensitivities. Currently, 383 work is being undertaken to explore the inclusion of lateral 384 faces on a directional microphone to further study the role that 385

these elements play and to attempt to develop a novel practical 386 application. However, there are numerous avenues for explo-387

- ration remaining, both experimental and theoretical. These 388
- include the development of improved bio-inspired technology 389
- 390 by incorporating higher-dimensional features and parameter
- 391 variations in the mechanical system, studying the behavior
- of the model at frequencies commensurate with bat sonar, 392
- and investigating the role that mechanical differences play in 393
- O. ochracea's hearing when addressing acoustic preferences. 394
- Finally, our expanded q2D model is the first mathematical 395
- model of hearing in an binaural fly that is accurate for all mea-396
- sured incident sound angles. It demonstrates the importance 397
- of incorporating higher-dimensional model elements consis-398
- tent with observed physiology, furthering our understanding 399

of binaural and insect hearing. 400

ACKNOWLEDGMENTS. The authors thank the Virginia Tech 401 Insect Collection for lending the Ormia ochracea samples for 402 imaging, and Pavel Shevchenko for assistance in imaging at 2-BM 403 at Argonne National Laboratory. This material is based upon work 404 supported by the National Science Foundation under Grant number 405 2014181. 406

407 408

409

- 1. LROP R.S., XII, On our perception of sound direction. The London, Edinburgh, Dublin Philos Mag. J. Sci. 13, 214-232 (1907).
- 2 LA Jeffress, A place theory of sound localization. J. Comp. Physiol. Psychol. 41, 35 (1948). 410 3. G Menda, et al., The long and short of hearing in the mosquito Aedes aegypti. Curr. Biol. 29, 411 709-714 (2019). 412
- JFC Windmill, JC Jackson, EJ Tuck, D Robert, Keeping up with bats: dynamic auditory tuning 413 414 in a moth. Curr. Biol. 16, 2418-2423 (2006).
- P Brownell, RD Farley, Detection of vibrations in sand by tarsal sense organs of the nocturnal 5. 415 scorpion, Paruroctonus mesaensis. J. Comp. Physiol. 131, 23-30 (1979). 416
- WH Cade, Acoustically orienting parasitoids: fly phonotaxis to cricket song. Science 190. 417 6.
- 1312-1313 (1975). 418 7. TJ Walker, Phonotaxis in female Ormia ochracea (Diptera: Tachinidae), a parasitoid of field 419
- crickets. J. Insect Behav. 6 (1993). 420 D Robert, M Read, R Hoy, The tympanal hearing organ of the parasitoid fly Ormia ochracea 421 (Diptera, Tachinidae, Ormiini), Cell Tissue Res, 275, 63-78 (1994),
- 422 423 9. M Rosen, EC Levin, RR Hoy, The cost of assuming the life history of a host: acoustic startle
- in the parasitoid fly Ormia ochracea, J. Exp. Biol. 212, 4056-4064 (2009). 424 10. DD Yager, Predator detection and evasion by flying insects. Curr. Opin. Neurobiol. 22, 201-425
- 426 207 (2012) N Lee, DO Elias, AC Mason, A precedence effect resolves phantom sound source illusions 427 11. in the parasitoid fly Ormia ochracea, Proc. Natl. Acad. Sci. 106, 6357-6362 (2009)
- 428 429 12. MA Bee, C Micheyl, The cocktail party problem: what is it? how can it be solved? and why
- 430 should animal behaviorists study it? J. Comp. Psychol. 122, 235 (2008). 431 13. D Robert, J Amoroso, RR Hoy, The evolutionary convergence of hearing in a parasitoid fly and its cricket host. Science 258, 1135-1137 (1992). 432
- 433 14. M Akcakaya, A Nehorai, Performance analysis of the Ormia ochracea's coupled ears. The J. 434 Acoust. Soc. Am. 124, 2100-2105 (2008)
- 435 15 RN Miles, D Robert, RR Hoy, Mechanically coupled ears for directional hearing in the para 436 sitoid fly Ormia ochracea. The J. Acoust. Soc. Am. 98, 3059-3070 (1995).
- 437 16 WH Cade, M Ciceran, AM Murray, Temporal patterns of parasitoid fly (Ormia ochracea) at-438 traction to field cricket song (gryllus integer). Can. J. Zool. 74, 393-395 (1996).
- 439 17. D Robert, RN Miles, R Hoy, Tympanal mechanics in the parasitoid fly Ormia ochracea: inter-440
- tympanal coupling during mechanical vibration. J. Comp. Physiol. A 183, 443-452 (1998). 441 18. D Robert, U Willi, The histological architecture of the auditory organs in the parasitoid fly Ormia ochracea. Cell Tissue Res. 301, 447-457 (2000) 442
- 443 ML Oshinsky, RR Hoy, Physiology of the auditory afferents in an acoustic parasitoid fly. J. 19. 444 Neurosci. 22, 7254-7263 (2002).
- AC Mason, ML Oshinsky, RR Hoy, Hyperacute directional hearing in a microscale auditory 445 20. system. Nature 410, 686-690 (2001). 446
- 447 21. AC Mason, N Lee, ML Oshinsky, The start of phonotactic walking in the fly Ormia ochracea 448 a kinematic study. J. Exp. Biol. 208, 4699-4708 (2005).
- 449 22. RN Miles, R Hoy, The development of a biologically-inspired directional microphone for hear 450 ing aids. Audiol. Neurotol. 11, 86-94 (2006).
- R Bauer, et al., Influence of microphone housing on the directional response of piezoelectric 451 23. 452 mems microphones inspired by Ormia ochracea. IEEE Sensors J. 17, 5529-5536 (2017).
- 453 Y Zhang, et al., A low-frequency dual-band operational microphone mimicking the hearing 454 property of Ormia ochracea. J. Microelectromechanical Syst. 27, 667-676 (2018).
- 455 25. M Touse, J Sinibaldi, G Karunasiri, Mems directional sound sensor with simultaneous detec 456 tion of two frequency bands in SENSORS, 2010 IEEE. (IEEE), pp. 2422-2425 (2010).
- 457 26. C Gibbons, RN Miles, Design of a biomimetic directional microphone diaphragm in Proceedings of IMECE. (ASME, State University of New York at Binghamton, Department of Mechan-458 459 ical Engineering), pp. 173-179 (2000)
- 27. A Rahaman, B Kim, Sound source localization by Ormia ochracea inspired low-noise piezo 460 electric mems directional microphone. Sci. Reports 10, 1-10 (2020). 461

28. BN Miles, et al., A low-noise differential microphone inspired by the ears of the parasitoid fly 462 Ormia ochracea. The J. Acoust. Soc. Am. 125, 2013-2026 (2009)

463

464

465

466

467

468

469

470

494

- 29. C Susanne, A Guidotti, R Hauspie, Age changes of skull dimensions. Anthropol. Anzeiger, 31-36 (1985).
- 30. CP Groves, The skulls of asian rhinoceroses; wild and captive. Zoo Biol. 1, 251-261 (1982). 31. DJ Tollin, K Koka, Postnatal development of sound pressure transformations by the head and
- pinnae of the cat; monaural characteristics. The J. Acoust. Soc. Am. 125, 980-994 (2009). 32. C Katsaros, B Berg, S Kiliaridis, Influence of masticatory muscle function on transverse skull
- dimensions in the growing rat. J. Orofac. Orthop. der Kieferorthopädie 63, 5-13 (2002)
- 33 J Schindelin, et al., Fiji: an open-source platform for biological-image analysis. Nat. Methods 471 9,676-682 (2012) 472
- 34. S Rolfe, et al., Slicermorph: An open and extensible platform to retrieve, visualize and analyse 473 3d morphology. Methods Ecol. Evol. n/a (2021). 474
- 35. H Liu, L Currano, D Gee, T Helms, M Yu, Understanding and mimicking the dual optimality of 475 the fly ear. Sci. Reports 3, 1-6 (2013) 476
- JL van Hemmen, J Christensen-Dalsgaard, CE Carr, PM Narins, Animals and ice: meaning, 477 36. origin, and diversity (2016) 478
- 37 P Muller, D Robert, A shot in the dark: the silent quest of a free-flying phonotactic fly. J. Exp. 479 Biol. 204, 1039-1052 (2001). 480
- 38 M Zuk, JT Rotenberry, RM Tinghitella, Silent night: adaptive disappearance of a sexual signal 481 in a parasitized population of field crickets. Biol. Lett. 2, 521-524 (2006). 482
- 39 S Pascoal, et al., Rapid convergent evolution in wild crickets. Curr. Biol. 24, 1369-1374 483 (2014)484
- DA Gray, C Banuelos, SE Walker, WH Cade, M Zuk, Behavioural specialization among pop-40. 485 ulations of the acoustically orienting parasitoid fly Ormia ochracea utilizing different cricket 486 species as hosts. Animal Behav. 73, 99-104 (2007). 487
- T Walker, S Wineriter, Hosts of a phonotactic parasitoid and levels of parasitism (Dipteration 41. 488 Tachinidae: Ormia ochracea). Fla. Entomol., 554-559 (1991). 489
- 42. R Lakes-Harlan, H Stölting, A Stumpner, Convergent evolution of insect hearing organs from 490 a preadaptive structure. Proc. Royal Soc. London. Ser. B: Biol. Sci. 266, 1161–1167 (1999). 491
- 43. D Robert, RN Miles, R Hoy, Tympanal hearing in the sarcophagid parasitoid fly Emblema-492 soma sp.: the biomechanics of directional hearing. J. Exp. Biol. 202, 1865-1876 (1999). 493

Abbreviations

The following abbreviations are used in this manuscript: 495

- ITD Interaural Time Delay
- IAD Interaural Amplitude Difference (sometimes called the Interaural Intensity Difference (
- mITD Mechanical Interaural Time Delay
- mIAD Mechanical Interaural Amplitude Difference