Interplay between Brownian motion and cross-linking kinetics controls bundling dynamics in actin networks

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Abstract

Morphology changes in cross-linked actin networks are important in cell motility, division, and cargo transport. Here we study the transition from a weakly cross-linked network of actin filaments to a heavily cross-linked network of actin bundles through microscopic Brownian dynamics simulations. We show that this transition occurs in two phases: first, a composite bundle network of small and highly aligned bundles evolves from cross linking of individual filaments; second, small bundles coalesce into the clustered bundle state. We demonstrate that Brownian motion speeds up the first phase of this process at a faster rate than the second. We quantify the time to reach the composite bundle phase and show that it is a strong function of mesh size only when the concentration of cross links is small, and that it remains roughly constant if we decrease the relative ratio of cross linkers as we increase the actin concentration. Finally, we examine the dependence of the bundling timescale on filament length, finding that shorter filaments bundle faster because they diffuse faster.

1 Introduction

The structure and mechanical properties of eukaryotic cells are largely controlled by the actin cytoskeleton, which contains a network of actin filaments interconnected by protein cross linkers (CLs) [1, 4]. Changes in cell mechanical properties, from more viscous to more elastic, relate to corresponding cytoskeletal morphology changes, from a weakly cross-linked network of actin filaments to a network of clustered bundles [27, 21]. The formation of a clustered bundle state

has previously been observed in actin suspensions with CLs such as filamin [31, 14], scruin [10], and α-actinin [22, 35]. In all of these systems, increasing the concentration of the cross-linking protein progressively transitions the steady state from a homogeneous meshwork, where filaments are distributed isotropically, through a composite bundle phase, where bundles are composed of only a few filaments, to the clustered bundle phase, where bundles can be separated by distances as large as 100 μm [31, 22].

Usually, the bundled network steady state is the result of a balance between cross-linking and other mechanisms that break up bundles. Indeed, in our previous work [27], we introduced actin filament turnover (to model (de)polymerization) and found that the steady state network morphology is the result of a competition between actin bundling and actin turnover. In particular, we observed either a homogeneous filament meshwork or network of bundles embedded in the filament meshwork, depending on the relationship between the turnover time and the timescale of filament bundling. In this paper, we will disable filament turnover and study how the timescale of bundling, which we define approximately as the time to reach the composite bundle state, is affected by the underlying microscopic parameters and the Brownian motion of the filaments. Quantifying this timescale is important because its competition with filament turnover rate determines the steady state network structure.

While it was observed over 30 years ago [14] that Brownian motion drives bundle formation, its precise mechanism for doing so remains unclear. For instance Hou et al. [14], speculated that rotational diffusion aids in bundling, as filaments that are linked at one location rotate until other locations can be linked together, resulting in a bundle. More recently, it was shown that bundling is most efficient in a fluid-like environment, where actin filaments can diffuse more readily [8, 9]. At minimum, these studies imply that bundling is more difficult without Brownian motion, but could actin filaments still arrange into bundles without it?

The importance of Brownian motion in bundling can be seen in experiments where filament length varies, or when polymerization and bundling are initiated simultaneously. In this case, shorter filaments form a more stable clustered bundle phase [17, 9], with the shortest filaments organizing into spindle-type structures [24, 36]. In systems where polymerization and bundling happen simultaneously, it has been shown that the formation of the clustered bundle phase can be prevented via an increase in the actin polymerization rate [8]. Mean field theory and simulations show that the slow down in bundling at high polymerization rates could be driven by a combination of steric interactions and the Brownian motion of the fibers being constrained by cross linkers [9].

It remains unclear, however, to what extent the attenuation of bundling is driven by sterics vs. cross linking, and even if a composite bundle phase can form if the length of the filaments is larger than the initial mesh size.

An underlying assumption in conceptual explanations of bundling is that sufficient CL is available to cross link filaments once they move closer together. The literature is conflicted, however, on exactly how much CL is sufficient. For instance, in the same experimental system of filamin and actin, some authors report a constant ratio of CL to actin necessary for bundling [14, 35], while others report that the relative amount of CL necessary for bundling decreases as actin concentration increases [31]. There is also a nontrivial effect of temperature on the amount of CL required for bundling; with higher CL-to-actin ratios bundling can occur at lower temperatures [33]. Experimental investigation of the precise amount of CL necessary for the clustered bundle state to form is difficult since the observation of bundles is a qualitative phenomenon with a subjective definition, and therefore varies based on the tools used. Simulations can provide a more definitive analysis of how bundling depends on CL concentration.

In this paper, we use agent-based simulations to quantify the evolution of the clustered bundle phase from a homogeneous meshwork of filaments and examine the role of Brownian motion therein. We begin in Section 2 by describing our computational methods [26, 27]. In Section 3, we demonstrate how a composite bundle state, and subsequently a clustered bundle state, evolve from a homogeneous meshwork, similar to what is observed in experimental networks [21]. We introduce a timescale, τ_c, that quantifies the time to reach the composite bundle state, and show that the dynamics on shorter, but not so much on longer, timescales are accelerated by Brownian motion. We show that the bundling timescale is limited by filament diffusion for smaller CL concentrations, while for larger CL concentrations this diffusion has a minor effect. We find that the relative CL-to-actin ratio required to achieve the same bundling time decreases with increasing actin network mesh size. Finally, we show that the diffusion effect explains the faster bundling for shorter filaments. We discuss some remaining questions, and possible extensions of our model necessary to answer them, in Section 4.

2   Methods

We begin with a review of the kinematics of inextensible fibers, slender body hydrodynamics, and our model of dynamic cross linking [26, 27] in Section 2.1. We then discuss new material pertinent

to the simulation of actin bundles, including how we modify our algorithm to simulate rigid fibers (Section 2.2) and to account for their translational and rotational diffusion (Section 2.3). Once we introduce thermal motion, a consistent model also requires us to keep the CL dynamics in detailed balance, i.e., that the binding and unbinding dynamics are in equilibrium with respect to the Gibbs-Boltzmann distribution. We account for this in Section 2.4 via a simple change to the rates of CL binding. Finally, in Section 2.5, we discuss how we use a time-splitting algorithm to evolve the system in time. While the CL binding and unbinding dynamics and filament evolution are treated in a first-order-accurate manner, we use a higher-order integrator for the Brownian term that can more accurately preserve fluctuation-dissipation balance.

2.1 Dynamic cross linking of semiflexible, inextensible fibers

This section reviews our algorithm for simulating the dynamic cross linking of semiflexible fibers [26, 27], beginning with the kinematics of inextensible fibers and slender body hydrodynamics [26], and concluding with our model of dynamic cross linking [27].

2.1.1 Semiflexible, inextensible fibers

We represent the centerline of each fiber by the Chebyshev interpolant $\boldsymbol{X}(s)$, where $s \in [0, L]$ is arclength and L is the fiber length. Likewise, the corresponding fiber tangent vector is represented by $\boldsymbol{\tau}(s) = \boldsymbol{X}_s(s)$. Because the fibers are inextensible, the tangent vector should have unit length for all time, $\boldsymbol{\tau}(s,t) \cdot \boldsymbol{\tau}(s,t) = 1$, for all s and t. Differentiating this constraint with respect to time, we obtain $\boldsymbol{\tau}_t \cdot \boldsymbol{\tau} = 0$, so that the velocity of the filament centerline can be parameterized as [26]

$$\boldsymbol{U}(s) = \boldsymbol{X}_t(s) = \overline{\boldsymbol{U}} + \int_0^s \left(\alpha_1(s')\boldsymbol{n}_1(s') + \alpha_2(s')\boldsymbol{n}_2(s') \right) \, ds' := \left(\mathcal{K}[\boldsymbol{X}]\boldsymbol{\alpha} \right)(s), \tag{1}$$

where $\boldsymbol{\tau}(s), \boldsymbol{n}_1(s), \boldsymbol{n}_2(s)$ are an orthonormal coordinate system at each s, and $\alpha_1(s)$ and $\alpha_2(s)$ are two unknown functions. Equation (1) defines a continuum kinematic operator $\mathcal{K}$ that parameterizes the space of inextensible motions [26, Sec. 3].

To close the system and solve for $\boldsymbol{\alpha} = \left\{ \alpha_1, \alpha_2, \overline{\boldsymbol{U}} \right\}$, we need to state the forces acting on the fiber centerline. To enforce the inextensibility constraint, we introduce a Lagrange multiplier force density $\boldsymbol{\lambda}(s,t)$. In addition to the constraint force, the fibers are also subject to a bending force with density $\boldsymbol{f}^\kappa[\boldsymbol{X}] = -\kappa\boldsymbol{X}_{ssss}$, where κ is the bending stiffness, and an external force density that comes from any attached cross links, which we denote by $\boldsymbol{f}^{(\text{CL})}$. The total force density at every instant in time is therefore $\boldsymbol{f} = \boldsymbol{\lambda} + \boldsymbol{f}^\kappa + \boldsymbol{f}^{(\text{CL})}$. Introducing the hydrodynamic mobility operator

$\boldsymbol{\mathcal{M}}[\boldsymbol{X}]$ that gives velocity from force (density), the evolution equation of the fiber centerline can be written as

$$\mathcal{K}[\boldsymbol{X}]\boldsymbol{\alpha} = \boldsymbol{\mathcal{M}}\left[\boldsymbol{X}\right]\left(\boldsymbol{f}^{\kappa}\left[\boldsymbol{X}\right] + \boldsymbol{\lambda} + \boldsymbol{f}^{(\text{CL})}\right), \tag{2}$$

$$\mathcal{K}^{*}[\boldsymbol{X}]\boldsymbol{\lambda} = \boldsymbol{0},$$

subject to the "free fiber" boundary conditions [34]

$$\boldsymbol{X}_{ss}\left(s = 0, t\right) = \boldsymbol{X}_{sss}\left(s = 0, t\right) = \boldsymbol{0}, \tag{3}$$

$$\boldsymbol{X}_{ss}\left(s = L, t\right) = \boldsymbol{X}_{sss}\left(s = L, t\right) = \boldsymbol{0}.$$

We solve (2) for the kinematic coefficients $\boldsymbol{\alpha}$ and constraint forces $\boldsymbol{\lambda}$. The adjoint condition $\mathcal{K}^{*}\boldsymbol{\lambda} = \boldsymbol{0}$ closes the system of equations, and encodes the principle of virtual work that constraint forces $\boldsymbol{\lambda}$ do no work for any inextensible motion of the fiber centerline [26, Sec. 3.4]. We still have to discuss the evaluation of $\boldsymbol{\mathcal{M}}\left[\boldsymbol{X}\right]$ and $\boldsymbol{f}^{(\text{CL})}$, which we do next.

2.1.2 Mobility evaluation

In previous work [26, 27], we utilized three different approaches to evaluate the mobility operator $\boldsymbol{\mathcal{M}}[\boldsymbol{X}]$. All of these approaches are based on traditional slender body theories [18, 16], which relate the velocity of a slender filament in Stokes flow to the force density exerted on its centerline. The total velocity at a point on the filament can be broken into three parts: that from force concentrated near the point (the "local drag" part, which dominates as the fiber becomes infinitely slender), that from the rest of the filament (intra-fiber hydrodynamics), and that from forcing on other filaments (through hydrodynamic interactions mediated by the fluid medium). The first two of these are simple to evaluate, given that they can be computed on each filament separately, but the third is expensive to compute because it involves all-to-all interactions through the fluid.

Because we have already studied the role of nonlocal hydrodynamic interactions in previous work [27], where we found that the time required to reach a particular bundled state changes by at most $10-20\%$, in this paper we will ignore hydrodynamic interactions between distinct filaments and evaluate the mobility by including only the local drag part and intra-fiber hydrodynamics. Specifically, the mobility operator on each fiber is given by nonlocal slender body theory [18, 16]

$$(8\pi\mu)\boldsymbol{U}(s) = (8\pi\mu)\left(\boldsymbol{\mathcal{M}}[\boldsymbol{X}]\boldsymbol{f}\right)(s) = \left(c(s)(\boldsymbol{I} + \boldsymbol{\tau}(s)\boldsymbol{\tau}^{T}(s)) + (\boldsymbol{I} - 3\boldsymbol{\tau}(s)\boldsymbol{\tau}^{T}(s))\right)\boldsymbol{f}(s) \tag{4}$$

$$+ \int_{0}^{L}\left(\left(\frac{\boldsymbol{I} + \hat{\boldsymbol{r}}(s, s')\hat{\boldsymbol{r}}^{T}(s, s')}{r(s, s')}\right)\boldsymbol{f}(s') - \left(\frac{\boldsymbol{I} + \boldsymbol{\tau}(s)\boldsymbol{\tau}^{T}(s)}{|s - s'|}\right)\boldsymbol{f}(s)\right)ds',$$

where $\boldsymbol{r}(s, s') = \boldsymbol{X}(s) - \boldsymbol{X}(s')$, $r = \|\boldsymbol{r}\|$, $\hat{\boldsymbol{r}} = \boldsymbol{r}/r$, and $c(s)$ is a local drag coefficient which has a logarithmic dependence on the fiber radius a. Away from the fiber endpoints, we use the classical result [18]

$$c(s) = \ln\left(\frac{4(L-s)s}{a^2}\right). \tag{5}$$

Near the endpoints, we regularize (5) over a distance $\delta = 0.1L$ as discussed in [26, Section 2.1]. The choice of mobility (4) allows us to simulate the evolution of bundles faster, and prevents possible numerical problems that could result when evaluating the nonlocal flows induced by hundreds of filaments in a bundle on each other [27].

2.1.3 Evaluation of $\boldsymbol{f}^{(\mathbf{CL})}$

We use a stochastic simulation algorithm to update the locations of the dynamic cross linkers. At each time step, theis algorithm, which we discuss in the next section, gives the fiber indices i and j that are linked by each link, as well as the arclength coordinates s_i^* on fiber i and s_j^* on fiber j where the link is bound. Letting K_c be the link stiffness (units force/length) and ℓ the CL rest length, we define the force density on fiber i due to the CL as

$$\boldsymbol{f}_i^{(\mathrm{CL})}(s) = -K_c \left(1 - \frac{\ell}{\left\|\boldsymbol{X}^{(i)}(s_i^*) - \boldsymbol{X}^{(j)}(s_j^*)\right\|}\right) \delta_h(s - s_i^*) \int_0^L \left(\boldsymbol{X}^{(i)}(s) - \boldsymbol{X}^{(j)}(s')\right) \delta_h(s' - s_j^*) \, ds', \tag{6}$$

where δ_h is a Gaussian density with standard deviation σ. While $\sigma \to 0$ corresponds to a standard spring point force, we use a finite σ to preserve smoothness for our spectral numerical method. For $N = 16$ points per fiber, which we use throughout this paper, we use $\sigma/L = 0.1$ [26]. As discussed in [27], this model is an approximation to the complex elasticity of α-actinin, and is based on experimental observations that the torsional stiffness of the α-actinin-actin bond does not influence the dynamics of that bond [5].

2.1.4 Dynamic cross-linking

Our model of dynamic cross linking is discussed in detail in [27, Sec. 2.2]. Briefly, we discretize each fiber into N_u uniformly-spaced "binding sites" with distance $\Delta s_u = L/(N_u - 1)$ between the sites. We make the assumption that the diffusion of individual CLs is sufficiently fast that it can be coarse-grained into a single binding rate k_{on} with units $1/(\text{length}\times\text{time})$. This means that a CL end can bind to a single discrete fiber binding site with rate $k_{\mathrm{on}}\Delta s_u$ per second. In the absence

of Brownian motion, as in [27], when one end of the CL is bound, the second end can bind to a nearby fiber with rate $k_{\text{on,s}}$. By "nearby" we mean a binding site on a distinct fiber that is within a distance interval

$$(\ell - \delta l, \ell + \delta l), \quad \text{where} \quad \delta \ell = \sqrt{\frac{kT}{K_c}} \tag{7}$$

from the first bound end, where ℓ is the CL rest length and $\delta \ell$ is a measure of the fluctuations in spring length.

Each of the binding reactions has a reverse reaction: a CL with both ends bound can have one end unbind, leaving one end bound, with rate $k_{\text{off,s}}$, and a CL with one end bound can unbind with rate k_{off} to have zero ends bound. There are thus four possible reactions, which we simulate stochastically using a version of the standard Stochastic simulation / Gillespie algorithm [11, 2]. The details of our implementation can be found in [27].

In the clustered bundle states that we simulate here, the number of links attached to a given site can grow without bound. To prevent this, we introduce a CL width $c_w = 20$ nm [29] and set the maximum number of bound CLs at each site to $\lceil \Delta s_u / c_w \rceil$. We implement this in the stochastic simulation algorithm using rejection: if a binding event is selected and the binding site is full, we simply move on to the next possible event.

2.2 Modifications for rigid fibers

In order to straightforwardly account for thermal fluctuations, we will consider the case when the fibers are rigid, so that the only possible fluctuations are translational and rotational diffusion. To simulate rigid fibers, we simply modify the kinematic operators $\mathcal{K}[\boldsymbol{X}]$ and $\mathcal{K}^*[\boldsymbol{X}]$ in (1) and (2). For rigid fibers, we introduce $\boldsymbol{\alpha} \equiv \boldsymbol{V} = \{\boldsymbol{U}_c, \boldsymbol{\Omega}\}$ to parameterize the space of rigid body motions, where $\boldsymbol{U}_c = d\boldsymbol{X}_c/dt$ is the translational velocity of the fiber center $\boldsymbol{X}_c = \boldsymbol{X}(L/2)$, and $\boldsymbol{\Omega}$ is the angular velocity of the fiber about its center. This gives the fiber velocity

$$\boldsymbol{U}(s) = (\mathcal{K}_r \boldsymbol{V})(s) = \boldsymbol{U}_c + \boldsymbol{\Omega} \times (\boldsymbol{X} - \boldsymbol{X}_c), \tag{8}$$

which reduces the constraint of virtual work to the fact that $\boldsymbol{\lambda}$ produces no net force and torque,

$$\mathcal{K}_r^* \boldsymbol{\lambda} = \begin{pmatrix} \int_0^L \boldsymbol{\lambda}(s)\, ds \\ \int_0^L (\boldsymbol{X}(s) - \boldsymbol{X}_c) \times \boldsymbol{\lambda}(s)\, ds \end{pmatrix} = \boldsymbol{0}. \tag{9}$$

We then solve the system (2) with $\mathcal{K}_r$ and $\mathcal{K}_r^*$ replacing $\mathcal{K}$ and $\mathcal{K}^*$. In the Supplementary Text, we show how to easily generalize our discretization for inextensible fibers [26] to straight rigid fibers

by restricting the number of Chebyshev modes included in the kinematic operator $\mathcal{K}$ to only the first one.

Because the fibers are rigid, we can formulate the hydrodynamic mobility as a 6×6 mobility matrix $\boldsymbol{N}[\boldsymbol{X}]$ which computes the fiber motion due to a total force $\boldsymbol{F}$ and torque $\boldsymbol{T}$,

$$\boldsymbol{V} = \begin{pmatrix} \boldsymbol{N}_{\mathrm{tt}} & \boldsymbol{N}_{\mathrm{tr}} \\ \boldsymbol{N}_{\mathrm{rt}} & \boldsymbol{N}_{\mathrm{rr}} \end{pmatrix} \begin{pmatrix} \boldsymbol{F} \\ \boldsymbol{T} \end{pmatrix} = \boldsymbol{N} \begin{pmatrix} \boldsymbol{F} \\ \boldsymbol{T} \end{pmatrix}. \tag{10}$$

When the fibers are straight, as in this work, and we measure the mobility about the geometric center of the fiber, the cross translation-rotation and rotation-translation mobilities vanish, $\boldsymbol{N}_{\mathrm{tr}} = \boldsymbol{N}_{\mathrm{rt}} = \boldsymbol{0}$. We recall that in this work we neglect hydrodynamic interactions between fibers, so the mobility matrix $\boldsymbol{N}$ can be computed for each fiber separately.

The mobility $\boldsymbol{N}$ can be obtained numerically from the slender body mobility matrix $\boldsymbol{M}$ (see [26, Sec. 4.2] for the discretization) via the Schur complement [32, 37]

$$\boldsymbol{N} = \left(\boldsymbol{K}_r^* \boldsymbol{M}^{-1} \boldsymbol{K}_r \right)^{\dagger}. \tag{11}$$

Note that the pseudo-inverse is required because applying a torque about the axis of a straight fiber produces no net motion (other than twisting, which we do not account for here). For straight fibers with constant tangent vector $\boldsymbol{\tau}$, by symmetry, the mobility $\boldsymbol{N}$ must be of the form

$$\boldsymbol{N}_{\mathrm{tt}} = \frac{1}{\mu L} \left(\alpha(\epsilon) \boldsymbol{I} + \beta(\epsilon) \boldsymbol{\tau}\boldsymbol{\tau} \right), \qquad \boldsymbol{N}_{\mathrm{rr}} = \frac{\gamma(\epsilon)}{\mu L^3} \left(\boldsymbol{I} - \boldsymbol{\tau}\boldsymbol{\tau} \right). \tag{12}$$

In Supplementary Table S1, we tabulate the coefficients α, β, and γ for biologically-relevant ϵ. See also [39] for semi-analytical approximations.

2.3 Thermal fluctuations with rigid fibers

For Brownian dynamics simulations, we need to solve the overdamped Ito Langevin equation

$$\begin{aligned} \frac{\partial \boldsymbol{X}}{\partial t} &= \boldsymbol{\mathcal{M}} \left(\boldsymbol{\lambda} + \boldsymbol{f}^{(\mathrm{CL})} \right) + \sqrt{2 k_B T} \, \boldsymbol{\mathcal{K}}_r \boldsymbol{N}^{1/2} \boldsymbol{\mathcal{W}}, \\ &= \boldsymbol{\mathcal{K}}_r \boldsymbol{N} \boldsymbol{\mathcal{K}}_r^* \boldsymbol{f}^{(\mathrm{CL})} + \sqrt{2 k_B T} \, \boldsymbol{\mathcal{K}}_r \boldsymbol{N}^{1/2} \boldsymbol{\mathcal{W}} \end{aligned} \tag{13}$$

where $\boldsymbol{\mathcal{W}}(t)$ is a vector of six i.i.d white-noise processes, and $\boldsymbol{N}^{1/2} \left(\boldsymbol{N}^{1/2} \right)^T = \boldsymbol{N}$. The last equality, which puts the overdamped Langevin equation into the more traditional symmetric form, follows from the fact that the deterministic velocity can be written using (9) and (10) as

$$\boldsymbol{U} = \boldsymbol{\mathcal{M}} \left(\boldsymbol{\lambda} + \boldsymbol{f}^{(\mathrm{CL})} \right) = \boldsymbol{\mathcal{K}}_r \boldsymbol{V} = \boldsymbol{\mathcal{K}}_r \boldsymbol{N} \boldsymbol{\mathcal{K}}_r^* \left(\boldsymbol{\lambda} + \boldsymbol{f}^{(\mathrm{CL})} \right) = \boldsymbol{\mathcal{K}}_r \boldsymbol{N} \boldsymbol{\mathcal{K}}_r^* \boldsymbol{f}^{(\mathrm{CL})}. \tag{14}$$

Note that because the fiber mobility is measured around the obvious geometric center, there is no stochastic drift term in the resulting Ito overdamped Langevin equation (13) [25, 6]. In our Brownian dynamics simulations with straight fibers, we use the precomputed values of α, β, and γ in (12) to generate $\boldsymbol{N}_{\mathrm{tt}}^{1/2}$ and $\boldsymbol{N}_{\mathrm{rr}}^{1/2}$ from the fiber tangent vector $\boldsymbol{\tau}$ according to (S10).

The random displacement of a fiber over a time interval τ can be sampled the following way,

1. Draw a vector $\boldsymbol{W}$ of six i.i.d. standard Gaussian variates and sample the rigid velocity

$$
\begin{pmatrix} \boldsymbol{U}_c \\ \boldsymbol{\Omega} \end{pmatrix} = \sqrt{\frac{2k_B T}{\tau}} \boldsymbol{N}^{1/2} \boldsymbol{W}. \tag{15}
$$

2. Update the fiber by translating its center by $\boldsymbol{U}_c \tau$ and rotating the fiber about its center by an oriented angle $\boldsymbol{\Omega}\tau$.

Note that for straight fibers one can simplify the formulation of Brownian dynamics; the formulation presented here applies to curved rigid fibers as well.

2.4 Keeping the CL dynamics in detailed balance

When we account for thermal translation and rotation of the fibers, we also want to be sure that the binding and unbinding of the links is consistent with detailed balance, which is not the case for the constant rates we introduced in Section 2.1.4. Let $\boldsymbol{C}$ denote a configuration of C links (list of fiber pair connections), and $\boldsymbol{x}$ denote the configuration of fibers (binding site positions). The desired Gibbs-Boltzmann equilibrium distribution is

$$
P_{\mathrm{eq}}(\boldsymbol{C}, \boldsymbol{x}) = \zeta(\boldsymbol{C}) \prod_{k \in \boldsymbol{C}} \exp\left(-\frac{K_c}{2} \frac{(\ell_k - \ell)^2}{k_B T}\right), \tag{16}
$$

where ℓ_k is the length of link k, and $\zeta(\boldsymbol{C})$ determines the probability to observe the cross-linking configuration $\boldsymbol{C}$. Now consider a transition to/from a state $\boldsymbol{C}'$ with one added link C', which has length ℓ_k'. Then at equilibrium the transition between the two states must obey

$$
P_{\mathrm{eq}}(\boldsymbol{C}, \boldsymbol{x}) k_{\mathrm{on,s}}(\ell_k') = P_{\mathrm{eq}}(\boldsymbol{C}', \boldsymbol{x}) k_{\mathrm{off,s}}(\ell_k'). \tag{17}
$$

Substituting P_{eq} from (16) into (17), we obtain the constraint of detailed balance

$$
\frac{k_{\mathrm{on,s}}(\ell_k')}{k_{\mathrm{off,s}}(\ell_k')} = \frac{\zeta(\boldsymbol{C}')}{\zeta(\boldsymbol{C})} \exp\left(-\frac{K_c}{2} \frac{(\ell_k' - \ell)^2}{k_B T}\right). \tag{18}
$$

To satisfy (19) for every choice of C and C' with binding and unbinding rates that only depend on spring length and not C and C', we must have

$$\frac{k_{\text{on,s}}\left(\ell'_k\right)}{k_{\text{off,s}}\left(\ell'_k\right)} = \frac{k^0_{\text{on,s}}}{k^0_{\text{off,s}}} \exp\left(-\frac{K_c}{2}\frac{(\ell'_k - \ell)^2}{k_B T}\right), \tag{19}$$

where $k^0_{\text{on,s}}$ and $k^0_{\text{off,s}}$ are the transition rates for a link at rest length.

To satisfy (19), we maintain a constant $k_{\text{off,s}}\left(\ell'_k\right) = k^0_{\text{off,s}}$ and set

$$k_{\text{on,s}}\left(\ell'_k\right) = k^0_{\text{on,s}} \exp\left(-\frac{K_c}{2}\frac{(\ell'_k - \ell)^2}{k_B T}\right). \tag{20}$$

Other choices are possible; for example, the rate of unbinding can depend on the stretch [3, 7]. To efficiently search for possible binding pairs, we approximate the set of all binding combinations by setting the maximum link stretch in (7) to be two standard deviations of the Gaussian (20), i.e., $\delta\ell = 2\sqrt{k_B T / K_c}$.

2.5 Temporal integration

We employ a time splitting approach to evolve the cross-linked actin network. At each time step, we have three processes to simulate: the thermal diffusion of the fibers, the binding and unbinding of the dynamic CLs, and the deterministic evolution of the fiber positions. The last two of these steps are laid out in full in [27, Sec. 2.6], where we employ Lie splitting to first process binding and unbinding events, and then use the method developed in [26] to evolve the fiber positions in an inextensible (or rigid, see Supplementary Section 6.1) way. Here we use the first-order accurate, backward Euler version of the deterministic fiber update which is discussed in [27, Sec. 2.6].

It remains to determine how we will treat the Brownian update. We use a second-order Strang-type splitting scheme, where during each time step of duration Δt we:

1. Randomly displace and rotate the fibers over a time interval $\tau = \Delta t / 2$ using the algorithm in Section 2.3.

2. Update the cross link attachments (using the stochastic simulation algorithm) and perform a deterministic fiber update, both over a time interval Δt, using the method of [27, Sec. 2.6].

3. Randomly displace and rotate the fibers over a time interval $\tau = \Delta t / 2$ using the algorithm in Section 2.3.

2.6 Network statistics

We quantify the evolution of the cross-linked actin network by examining the connectivity of the fibers in two ways. First, given the total number of CLs in the system C, we compute an average link density per fiber via the formula "Link density" $= 2C/(LF)$. Second, we map the network to a connected graph to study how the structure evolves in time [24]. We define a "bundle" as a connected groups of filaments, where a connection between two fibers is a pair of links with anchoring locations at least $d_{\text{bund}} = L/4$ apart on each fiber [27]. Then the bundle density is the number of bundles per unit volume, "Bundle density" $= B/L_d^3$, where B is the number of bundles and L_d is the length of the simulation cell.

For a bundle of b filaments, we define an orientation parameter as the maximum eigenvalue of the matrix [24]

$$Q = \frac{1}{bL} \sum_{i=1}^{b} \int \boldsymbol{\tau}^{(i)}(s) \left(\boldsymbol{\tau}^{(i)}(s) \right)^T ds. \tag{21}$$

The orientation parameter takes values in $[1/3, 1]$, with 1 being the value for a group of straight fibers with the same tangent vector. Given information about the bundles, we compute an average bundle orientation parameter by taking an average over bundles with at least two filaments, weighted by the number of filaments in each bundle.

Throughout this paper, we will quantify the concentration of fibers in terms of the initial mesh size [30] of the suspension, $\ell_m = \sqrt{L_d^3/(FL)}$ (parameters are defined in Table 1). Note that this estimate for ℓ_m applies to non-bundled (disordered) suspensions of fibers, so really when we use ℓ_m we mean the *initial* mesh size, prior to the bundling process beginning.

3 Results

We begin this section by discussing the kinetics of bundling for *non-Brownian, semiflexible* fibers, and establish that semiflexible fibers with a persistence length similar to that of actin [12] can be well-approximated by rigid filaments. We then show that rigid, Brownian fibers have similar kinetic behavior, except that the Brownian motion (translational and rotational diffusion) speeds up the timescale of bundling.

After these preliminaries, we use our simulations to clarify and explain some of the experimental results on the dynamic formation of cross-linked actin bundles. First, we show how the timescale needed to reach the composite bundle state depends on the fiber concentration (initial mesh size)

Parameter	Definition	Value	Unit	Notes
a	Fiber radius	4	nm	[13]
L	Fiber length	$0.5, 1$	μm	[15, 28]
L_d	Simulated volume's extent	$2 - 4$	μm	Cubic unit cell
μ	Cytoplasm viscosity	0.1	Pa·s	$100\times$ water (cytoplasm) [23]
κ	Fiber bending stiffness	0.07	pN·μm^2	17 μm persistence length [12]
K_c	CL spring stiffness	10	pN/μm	[20]
ℓ	CL rest length	50	nm	[29]
k_{on}	CL first end binding rate	5	$1/(\mu$m·s)	[27]
$k_{\text{on,s}}$	CL second end binding rate	50	$1/(\mu$m·s)	$k_{\text{on,s}} \gg k_{\text{on}}$, not measured
k_{off}	CL (one end bound) unbinding rate	1	1/s	[19, 38]
$k_{\text{off,s}}$	CL (both ends bound) unbinding rate	1	1/s	$k_{\text{off,s}} = k_{\text{off}}$
c_w	Actin binding site width	20	nm	[29]
$k_B T$	Thermal energy	4×10^{-3}	pN·μm	25°C
N	Number of Chebyshev points	16		[26, Sec.6.3.1]
Δs_u	Binding site spacing	0.026	μm	[27]
Δt	Time step size	10^{-4}	s	Limited by K_c

Table 1: Simulation parameters.

and concentration of CLs (which controls k_{on} in our model). We show that the bundling process is slower when the actin or CL concentration is lower, but that bundling can still occur at low actin concentration, provided there are enough CLs available to bundle the fibers, and that the relative amount of CLs needed for a fixed bundling time decreases as actin concentration increases. Second, we show that the experimental result that bundling occurs faster for shorter fibers [17, 8] can only be reproduced in systems where we consider translational and rotational diffusion. Unless otherwise noted, we will use the simulation parameters listed in Table 1. As discussed in [27], the cross-linking parameters are chosen to mimic α-actinin, although we will compare our results to systems with different CLs, such as filamin [31].

Figure 1: Bundling dynamics on small and large scales. Top: a small-scale bundling process with three filaments and snapshots taken at times $t = 0, 2, 4, 6, 8$, and 10 s. Bottom: Snapshots of the bundling process taken (from left to right and top down) at $t = 5, 10, 20$, and 40 seconds for semiflexible fibers with stiffness $\kappa = 0.07$ pN$\cdot\mu$m^2. Fibers in the same bundle are colored with the same color. The two networks at the middle are before the coalescence transition time $\tau_c \approx 16$ s, while the two networks at the bottom are after the coalescence time.

3.1 Kinetics of bundling for semiflexible fibers without fluctuations

We begin with simulations in a system of initial mesh size $\ell_m = 0.2$ μm, which translates to $F = 200$ filaments in an $L_d = 2$ domain, $F = 675$ filaments in an $L_d = 3$ domain, and $F = 1600$ filaments in an $L_d = 4$ domain. In Supplementary Fig. S1, we show that the statistics of the bundling process are insensitive to the domain size up to the point where there is mass coalescence of almost all the fibers in the simulation cell. For this reason, we will consider results from only one set of simulations, the one with $F = 675$ filaments and $L_d = 3$.

We initialize the set of F filaments with random locations and orientations, then during each time step we evolve the fibers by updating the dynamic CLs and then updating the fiber positions in sequential order. Figure 1 shows how the bundling process evolves in small and large systems. On the microscopic scale, filaments that are initially not parallel are linked by CLs, which pull them closer together and allow more links to bind. The binding of additional links leads to the alignment of filaments. Note that the key to the bundling process is the flexibility of the CL, in particular rapid thermal fluctuation of the CL length, which is present implicitly in our model. Because the CLs are small, fluctuations in their length occur on a time scale which is much faster than other characteristic time scales, and so we do not model the fluctuations explicitly. The combination of the CLs' elasticity and length fluctuations is crucial, as the length fluctuations effectively allow the CLs to "find" the neighboring fibers and bind them, whereupon the elasticity of the CL aligns the fibers, making further cross linking faster.

This process plays out on a larger scale in snapshots from the simulations, shown in Fig. 1 at $t = 5, 10, 20$, and 40 seconds. The initial phase of bundling (first two snapshots) is characterized by bundles of a few straight, aligned filaments, which is similar to the experimentally-observed composite bundle state [21] and the three-filament bundle shown at the top of Fig. 1. Later times (bottom two frames) show coalescence of these smaller bundles into larger bundles, with some curvature appearing in the fibers in the final frame. By $t = 40$ seconds, there are only a few bundles made of coalesced smaller bundles, and the network resembles the experimentally-observed clustered bundle state [21].

To quantify our observations, in Fig. 2 we plot the mean link density ($2C/(LF)$, see Section 2.6), bundle density (B/L_d^3), percentage of fibers in bundles, mean bundle alignment parameter, and mean and maximum bundle size throughout the bundling process for three values of fiber stiffness: $\kappa = 0.07$ (the value for actin), $\kappa = 0.007$ (fibers ten-fold less stiff), and $\kappa = \infty$ (rigid

Figure 2: Statistics for the bundling process, where we compare the base parameters ($\kappa = 0.07$ pN·μm^2, blue) with the systems with smaller bending stiffness ($\kappa = 0.007$, orange) and rigid fibers (yellow, $\kappa \to \infty$). After $\tau_c \approx 16$ s, the bundling dynamics for the less stiff fibers are significantly faster. Fibers with similar bending stiffness to actin are well-approximated by rigid fibers.

filaments). In all systems, we see the number of links per fiber grow in time to approach the maximum of $\lceil \Delta s_u / c_w \rceil \times L / s_u = 80$, while the bundle density in all systems exhibits a peak around a critical time $\tau_c \approx 16$ seconds. At this time, the other panels of Fig. 2 tell us that 60% of the fibers are already in bundles, which have a mean alignment parameter larger than 0.9. Figure S2 gives a more precise look at the composition of the bundles, which are the same for the three values of stiffness when $t \leq \tau_c$: at $t = \tau_c$, most ($> 50\%$) of the fibers are in bundles of size 11 or less, with a small percentage ($< 10\%$) in bundles of size $10 - 20$, and the other 40% of the filaments not in bundles at all. Thus, a time τ_c into the bundling process, most of the fibers are in small, highly aligned bundles, as we see in the snapshots in Fig. 1, and the dynamics up to this point are roughly independent of the fiber stiffness. Based on Fig. 1, we can also think of τ_c as the time required to reach the composite bundle state.

After the coalesence time, we see a transition to coalescence of bundles, and the flexibility of the fibers comes into play. Figure 2 shows that the number of bundles is declining and the mean bundle alignment is dropping for $t > \tau_c$, which implies that bundles are forming with non-aligned fibers. The mean and maximum bundle sizes also start to grow, which again means that small bundles are coming together to form the larger ones we see in the bottom row of Fig. 1. Figure S2 shows that by $t = 60$ seconds, at least 75% of the fibers are in bundles of size 30 or larger. It is in this stage where the flexibility of the fibers can become important: when $\kappa = 0.007$ (fibers ten-fold less stiff than actin), coalescence of bundles occurs faster than in systems with $\kappa = 0.07$ or systems with rigid fibers, since in the former case the fibers are more compliant and can be linked together more easily by deforming. That said, when $\kappa = 0.07$ (persistence length 17 μm), Figs. 2 and S2 show that the dynamics throughout the bundling process are well-approximated by rigid filaments. This analysis is of course limited by the fiber length we've chosen: in particular, we have shown that, in the absence of Brownian motion, rigid filaments are a good approximation to semiflexible actin filaments for fibers of length ≤ 1 μm, which are most common in vivo. The approximation will be worse as the filament length gets larger. Henceforth, we will consider rigid fibers only.

3.2 Thermal fluctuations speed up the bundling process

We now consider simulations with rigid fibers, for which we can account for translational and rotational diffusion using standard Brownian dynamics methods [25], while maintaining detailed balance in the cross-linking kinetics, as discussed in Section 2.4. An important quantity in this case is the time for a fiber to diffuse across a mesh size. In our initial set-up, the fibers are spaced

approximately ℓ_m apart, and they first must find each other in order to cross link and begin the bundling process. The theoretical translational diffusion coefficient of a straight fiber, derived in [25], can be written in terms of the 3×3 translational mobility matrix $\boldsymbol{N}_{\mathrm{tt}}$ for rigid body motions as

$$D_t = \frac{k_B T}{3} \mathrm{tr}\left(\boldsymbol{N}_{\mathrm{tt}}\right) \approx \frac{k_B T}{3\mu L} 1.67, \tag{22}$$

where the last equality gives the result for a fiber aspect ratio of $\epsilon = 0.004$, which we obtain from slender body theory with intra-fiber hydrodynamics (see Table S1). The mean square displacement of the fiber center is $\langle r^2(t) \rangle = 6 D_t t$. Substituting the parameters in Table 1, we obtain $6D_t \approx 0.13$ $\mu\mathrm{m}^2/\mathrm{s}$, and thus the time to diffuse a mesh size is given by $\tau_m = \ell_m^2 / \left(6 D_t\right) \approx \ell_m^2 / 0.13$ s. Since diffusion promotes mixing of the suspension and gives more opportunities for cross linking, our expectation is that thermal fluctuations should speed up the transition from the homogeneous meshwork to the composite bundle phase, where bundles are made of a few fibers which must be close enough together to cross link. This assumes that the CL concentration is large enough for links to bind as soon as fibers are close enough together; we will analyze this assumption in Section 3.3.

To understand how thermal diffusion affects the bundling process, in Fig. 3 we plot the statistics both with (orange) and without (blue) thermal fluctuations. We see that the entire process is faster with diffusion, as we might expect, but the degree of acceleration changes before and after τ_c. Before τ_c, the process with diffusion is significantly faster than without; for instance, it takes about 3 seconds for 50% of the fibers to be in bundles with diffusion, while without diffusion it takes 12 seconds (Fig. 3 inset), which is a difference of a factor of 4. Indeed, the critical bundling time $\tau_c \approx 4$ seconds with diffusion, while we have already seen $\tau_c \approx 16$ seconds without diffusion, so that the difference is again a factor of 4. For $t > \tau_c$, when bundles start to coalesce, the difference is only a factor of 2; an exponential fit to the decaying bundle density gives a constant of 20 seconds for simulations with diffusion and 40 seconds for simulations without diffusion. Thus, thermal fluctuations make more of a difference when fibers are less constrained by cross-links, as we would expect. Sure enough, Fig. S3 (left, blue curve) shows that the mean squared displacement for simulations with Brownian motion decays exponentially to a constant, meaning that at times larger than τ_c the Brownian motion is inhibited by cross linking, and therefore becomes less important.

To show that the network morphology has not changed when we add thermal movement, in Fig. S4 we show the networks at $t \approx \tau_c$ without and with thermal fluctuations. The composite bundle

Figure 3: Statistics for the bundling process with and without thermal fluctuations. The blue lines show the results without thermal movement, while the orange lines show the results with translational and rotational diffusion. Here we use $\Delta t = 10^{-4}$; we have verified that the statistical noise exceeds the time stepping error for this time step size. The peak in the bundle density occurs at $\tau_c \approx 16$ for systems without diffusion, while for systems with diffusion it occurs at $\tau_c \approx 4$ seconds.

network morphology at τ_c is similar between the two, which demonstrates that fluctuations speed up the pace of bundling without changing the types of bundles that evolve.

3.3   Dependence of bundling timescale on actin and CL concentration

Our conclusion that thermal fluctuations significantly accelerate the initial phase of the bundling process is dependent on having a sufficient concentration of CLs. While thermal fluctuations undoubtedly increase the frequency of fibers coming close enough together for cross linking, the bundling process still must be initiated via binding of a CL. Consequently, in this section we consider a range of values of mesh size (actin concentration) and k_{on} (CL attachment rate, which is proportional to CL concentration), to get a more complete picture of how the critical bundling time τ_c depends on these parameters. In particular, we will consider mesh sizes $\ell_m = 0.2$ ($F = 675$, $L_d = 3$ μm), 0.4 ($F = 400$, $L_d = 4$ μm), and 0.8 μm ($F = 338$, $L_d = 6$ μm), and single-end binding rates $k_{\text{on}} = 1.25$, 5 (the base value), and 20 $(\mu\text{m·s})^{-1}$. By changing the rate at which a single CL end binds to a fiber, we effectively vary the CL concentration.

Figure 4 shows the resulting evolution of the bundle density for the nine different systems, as well as the resulting critical bundling time τ_c. For systems with large k_{on}, where binding is essentially instantaneous once filaments come close enough together, $\tau_c \approx 3$ s for the small enough mesh sizes of $\ell_m = 0.2$ and 0.4 μm. Once the mesh size increases to 0.8 μm, the bundling time increases, but only to about 4.5 s (see inset of Fig. 4). Thus τ_c is not a strong function of mesh size for larger k_{on}, which implies that the process for large k_{on} is primarily limited by cross-linking dynamics (alignment of filaments), with diffusion (across the mesh size) playing only a secondary role.

Let us now consider the case of slower k_{on}. In this case, filaments could come close enough to link together, but diffuse away before a CL can actually bind them. As a result of this, the bundling process is slowed, and in fact the peak bundle density drops (orange curves in Fig. 4 peak at significantly lower values for $\ell_m = 0.2$ and 0.4) or disappears all together over the timescale investigated ($\ell_m = 0.8$). Indeed, as shown in Fig. 5, networks with smaller k_{on} (lower CL concentration) contain larger bundles at $t = \tau_c$ than those with larger k_{on} (higher concentration).

For smaller k_{on}, the bundling process is limited by a combination of CL binding and filament diffusion, in the sense that filaments must be close enough for binding to occur *and* the binding event must occur in the short time window that the filaments are together. In fact, diffusion is actually a hindrance to bundling in some sense, since fibers that are close to each other diffuse away

Figure 4: Bundling time scales for a range of initial mesh sizes ℓ_m and binding rate k_{on}. The first three frames show the trajectory of the bundle density for the different mesh sizes, where blue lines denote our base value of $k_{\mathrm{on}} = k_{\mathrm{on}}^{(0)} = 5$ $(\mu\mathrm{m}\cdot\mathrm{s})^{-1}$, orange lines denote $k_{\mathrm{on}}/k_{\mathrm{on}}^{(0)} = 1/4$, and yellow lines $k_{\mathrm{on}}/k_{\mathrm{on}}^{(0)} = 4$. The bottom right frame shows the dependence of the critical bundling time τ_c on ℓ_m and k_{on}.

Figure 5: Snapshots of the network at $t = \tau_c$ with initial mesh size $\ell_m = 0.4\ \mu$m ($F = 400$ filaments of length $L = 1$ in a domain of size $L_d = 4$) with $k_{\mathrm{on}}/k_{\mathrm{on}}^{(0)} = 1/4$ (left, $\tau_c \approx 17$), $k_{\mathrm{on}}/k_{\mathrm{on}}^{(0)} = 1$ (middle, $\tau_c \approx 5$), and $k_{\mathrm{on}}/k_{\mathrm{on}}^{(0)} = 4$ (right, $\tau_c \approx 2.5$). A smaller k_{on} (smaller CL concentration) gives fewer but larger bundles at $t = \tau_c$, as well as a smaller percentage of fibers in bundles.

before a CL can bind them together. The scaling of τ_c at small k_{on} (left column of the bottom right panel in Fig. 4) is reminiscent of a diffusion-limited process, as it increases from 9 s to 17 s to 56 s as the mesh size doubles, scaling approximately as ℓ_m^2 as the mesh size increases.

We note that a roughly constant bundling time can be achieved by decreasing k_{on} as the mesh size decreases (moving from the top right to the bottom left of the bottom right panel in Fig. 4). This implies that the relative concentration of CL required for a particular bundled state decreases with the mesh size, as has been found experimentally [31]. When the mesh size is smaller, the filaments are in contact for longer, and so it is less important that a CL be available immediately to bind them together. By contrast, filaments in larger-mesh-size systems are only in contact for a brief time, so relatively more CLs are necessary to ensure that these filaments are linked when they come into contact with each other.

3.4 Brownian motion is responsible for faster bundling with shorter filaments

We will now explore the dependence of the critical bundling time τ_c on the fiber length. Experimentally, it has been shown that shorter filaments bundle faster [17, 8], but it is still unclear whether this is due to thermal movements, cross-linking kinetics, or some combination of both. In this section, we show that the experimental results can only be reproduced if we consider thermal movements, so that cross-linking kinetics are not responsible for the speedup in bundling. We use a fixed mesh size of $\ell_m = 0.2\ \mu$m, which translates to $F = 675$ filaments of length $L = 1\ \mu$m in a

domain of size $L_d = 3$ μm, and $F = 400$ filaments of length $L = 0.5$ μm in a domain of size $L_d = 2$ μm.

In Fig. 6, we show how the link density, bundle density, and percentage of fibers in bundles evolve for the two different filament lengths both (a) without and (b) with actin diffusion. In Fig. 6(a), we see that in the absence of Brownian motion the behavior in the two systems is similar, with the peak bundle density occurring in both cases around $\tau_c \approx 15$ seconds. Furthermore, there is only a mild difference in the percentage of fibers in bundles over time. The mean bundle size is at most twice larger for the system with shorter filaments, but we would expect this since the filaments are twice as short and there are twice as many of them if ℓ_m is fixed.

In Section 3.2, we showed that Brownian motion speeds up the bundling process by promoting mixing and more near-contacts of filaments. In particular, we saw that the time for a filament with length $L = 1$ μm to diffuse a mesh size of $\ell_m = 0.2$ μm is $\tau_m \approx 0.30$ s, so that filaments can find each other rapidly and begin the bundling process. In the case of filaments with $L = 0.5$ μm, our thermal diffusion coefficient (22) scales log-linearly with the fiber length, so that it takes $\ell_m = 0.17$ s to diffuse a mesh size of $\ell_m = 0.2$ μm. We might expect, therefore, that at least the initial stages of the bundling process will be sped up by a factor of 2.

Figure 6(b) shows that this is indeed the case. For $\ell_m = 0.2$ μm, the bundle density peak occurs around $\tau_c \approx 2$ seconds when $L = 0.5$ μm, while with $L = 1$ μm it occurs around $\tau_c \approx 4$ seconds, so it appears that bundling time with thermal motion scales linearly with filament length, which is in (approximate) accordance with the scaling of the translational diffusion coefficient. The faster bundling behavior also manifests itself in the link density and percentage of fibers in bundles, where we see that systems with shorter filaments reach a number of links or percentage of fibers about twice as fast. For instance, 80% of the fibers are in bundles by $t \approx 4$ seconds in the $L = 0.5$ μm case, while with $L = 1$ μm the 80% mark is not reached until about $t \approx 8$ seconds.

4 Discussion

We used numerical simulations to investigate the kinetics of bundling in cross-linked actin suspensions. After validating that semiflexible actin fibers can be approximated as rigid in non-Brownian suspensions, we treated actin fibers as Brownian rigid, straight, slender rods, in accordance with a number of other simulation studies [9, 24]. We coarse grained the diffusion and binding and unbinding of α-actinin cross-linkers (CLs) into four microscopic rates: $k_{\mathrm{on}}, k_{\mathrm{on,s}}, k_{\mathrm{off}},$ and $k_{\mathrm{off,s}}$. This

(a) No diffusion

(b) With diffusion

Figure 6: Effect of changing filament length for rigid fibers with and without Brownian motion, with constant initial mesh size $\ell_m = 0.2$ μm. (a) Without fiber diffusion, we show the statistics for filaments of length $L = 0.5$ μm (orange) and $L = 1$ μm (blue), where we observe dynamics occurring on a similar timescale, especially in the initial phase ($t \leq \tau_c \approx 20$) of bundling. (b) When we add fiber diffusion, the bundling process for $L = 0.5$ μm (orange) is significantly faster than $L = 1$ μm (blue), because filaments can diffuse faster.

enabled the simulation of a gel with about 700 actin fibers and as many as 50 CLs bound to each fiber.

We found that, even without thermal movements, actin filaments can still bundle, as filaments that are initially close enough are linked together at small patches with CLs. These CLs pull fibers together and align them, thereby allowing more CLs to bind to other sections of the fibers. What results initially, for times smaller than the critical bundling coalescence time τ_c, is a collection of bundles with a few highly aligned filaments, also called a composite bundle phase [21]. For times larger than τ_c, these bundles coalesce into larger bundles using a similar mechanism as that for individual fibers, and a clustered bundle phase forms. Our critical bundling timescale τ_c thus describes the initial time at which networks transition from the composite bundle phase to the clustered bundle phase. In networks with fiber turnover, a clustered bundle steady state is only possible if the turnover time is much larger than τ_c [27].

We quantified the role of diffusion throughout the bundling process, finding that it has a larger impact in the initial stages of bundling, when the filaments are not severely constrained by CLs and can move freely to find each other. We associated this stage with $t < \tau_c$, and showed that adding thermal fluctuations decreases τ_c from 16 seconds to 4 seconds. We showed that the stage when bundles coalesce ($t > \tau_c$) is less affected by thermal diffusion (sped up by a factor of 2), since at that stage the filaments are constrained by CLs, which are more involved in bundle coalescence. This complements the observation in [8, 36] that bundling occurs faster in a fluid-like environment, where filaments can move freely prior to kinetic arrest.

At first glance, the order of magnitude of τ_c that we obtained seems shorter than the characteristic bundling time obtained experimentally, which is generally reported to be on the order of minutes [29]. The comparison is difficult, however, since experimental times generally include polymerization, and the bundling timescale in experiments is defined by the onset of the clustered bundle steady state, which is much later than the composite bundle phase where we define τ_c. Nevertheless, the most instructive comparison is between our work and [8, Fig. 4], which shows experimentally that the addition of 10% nucleates (which speeds up the polymerization process) gives a saturated bundled state after 100 seconds of polymerization and bundling, where the bundles are made of at least $15 - 30$ filaments and are spaced some $10 - 20$ μm apart. Given this observation, and the fact that bundling slows down over time, it is not difficult to imagine that the transition from the homogeneous state to the composite bundle phase could take place on the order of $5 - 10$ seconds after cross linkers are added to a system of (polymerized and capped) actin filaments.

While diffusion of fibers speeds up the bundling process, we showed that it must be combined with a sufficient concentration of CLs for rapid bundling to occur. In particular, we showed that a high concentration of CLs (high CL binding rate) can induce bundling for filaments of any mesh size, with a critical bundling time τ_c that depends only weakly on the mesh size. By contrast, when the concentration of CLs is small, bundling is more difficult for any fixed mesh size, and gets near impossible as the mesh size increases, as near-fiber contacts become less frequent. This is in accordance with a number of experimental papers [31, 17] which find that bundling requires a critical CL concentration. In addition, because the fibers are in contact for a short time at larger mesh sizes, the system must be saturated with CLs for bundling to proceed at a reasonable rate. This saturation is less important at smaller mesh sizes, where fiber pairs come into contact more frequently. This explains why experiments have shown that a lesser CL to actin concentration ratio is needed for bundling as the concentration of actin fibers increases [31, Fig. 3].

As already mentioned, one the drawbacks of some experimental studies is the sensitivity of the bundling time to the rate of actin polymerization. For example, it is shown in [8, Fig. 4(d)] that polymerization kinetics make an order of magnitude difference in the bundling kinetics. While simultaneous polymerization and bundling also occurs in vivo, our study here allowed us to divorce bundling and polymerization by focusing on a fixed filament length. By doing this, we showed that shorter filaments bundle faster exclusively because they can diffuse faster, because without thermal fluctuations we saw no difference in the bundling kinetics between short and long filaments. This clarifies why shorter actin filaments are able to associate more rapidly into bundles without the presence of a background actin mesh [17, 36].

There are, of course, other timescales that we could have examined in the bundling process. For instance, Figs. S3 and S5 show that the timescale for slow down of the fibers' diffusivity, measured by the decay of their mean square displacement, is related to, but certainly not the same as, the critical bundling time τ_c. Our choice to focus on the timescale τ_c was motivated by our observation in previous work [27, Sec. 3.1] that the steady state morphology of cross-linked actin networks is driven by a competition between bundling (which occurs on timescale τ_c) and filament turnover (which occurs on timescale τ_f). While it is intuitively obvious that increasing the turnover timescale τ_f will produce a steady state with more bundles, it is fair to ask whether the ratio τ_f/τ_c alone controls the steady state morphology, or if some other microscopic parameters come into play. In Fig. S6, we show that for turnover times $\tau_f = \tau_c/2$, τ_c, and $2\tau_c$, the gel evolves to a steady state where the bundle density and percentage of fibers in bundles depend primarily on the ratio τ_f/τ_c,

for either Brownian or non-Brownian fibers (recall that τ_c differs by a factor of 4 for these two cases). Snapshots in Fig. S7 show little qualitative difference between the network morphology of the Brownian and non-Brownian steady states for a fixed τ_f/τ_c. Thus, for a fixed turnover time τ_f, the steady state morphology is controlled by τ_c, which is the timescale we studied in detail here.

Our study here used rigid filaments and coarse grained the dynamics of CL diffusion and binding. While we showed that *non-Brownian* semiflexible actin filaments can be approximated by rigid ones, we have not accounted for the transverse bending fluctuations in actin filaments. In some sense, softening the stiffness of the cross linkers, which gives a wider range of binding distances than might otherwise be possible, qualitatively accounts for this, but we plan to develop a numerical method that includes bending fluctuations in the future. We also hope to place our model of cross-linker dynamics on more rigorous footing by comparing it to a model that actually tracks the diffusion, binding, and unbinding of individual CLs. Finally, in this work it was too difficult for us to simulate the experimental steady state clustered bundled morphologies, since we simulated actin filament lengths of 1 μm and the observed steady states have bundles separated by hundreds of microns [31, 22]. More efficient, GPU-based, simulation techniques might enable the efficient simulation of even larger systems.

5   Acknowledgements

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Code and input files for the simulations are available at `https://github.com/stochasticHydroTools/SlenderBody`. Supplemental movies are available at `https://cims.nyu.edu/~om759/BundlingVideos`. All of our simulations were run on the NYU HPC Greene Supercomputer cluster.

References

[1] Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, and Peter Walter. *Molecular biology of the cell*. Garland Science, 2002.

[2] David F Anderson. A modified next reaction method for simulating chemical systems with time dependent propensities and delays. *The Journal of chemical physics*, 127(21):214107, 2007.

[3] Tamara Carla Bidone, Wonyeong Jung, Daniel Maruri, Carlos Borau, Roger D Kamm, and Taeyoon Kim. Morphological transformation and force generation of active cytoskeletal networks. *PLoS computational biology*, 13(1):e1005277, 2017.

[4] Loïc Chaubet, Abdullah R Chaudhary, Hossein K Heris, Allen J Ehrlicher, and Adam G Hendricks. Dynamic actin cross-linking governs the cytoplasm's transition to fluid-like behavior. *Molecular biology of the cell*, 31(16):1744–1752, 2020.

[5] David S Courson and Ronald S Rock. Actin cross-link assembly and disassembly mechanics for α-actinin and fascin. *Journal of Biological Chemistry*, 285(34):26350–26357, 2010.

[6] Steven Delong, Florencio Balboa Usabiaga, and Aleksandar Donev. Brownian dynamics of confined rigid bodies. *The Journal of chemical physics*, 143(14):144107, 2015.

[7] Denis James Evans, Debra Joy Searles, and Stephen Rodney Williams. *Fundamentals of classical statistical thermodynamics: dissipation, relaxation, and fluctuation theorems*. John Wiley & Sons, 2016.

[8] Tobias T Falzone, Martin Lenz, David R Kovar, and Margaret L Gardel. Assembly kinetics determine the architecture of α-actinin crosslinked f-actin networks. *Nature communications*, 3(1):1–9, 2012.

[9] Giulia Foffano, Nicolas Levernier, and Martin Lenz. The dynamics of filament assembly define cytoskeletal network morphology. *Nature communications*, 7(1):1–8, 2016.

[10] ML Gardel, Jennifer Hyunjong Shin, FC MacKintosh, L Mahadevan, P Matsudaira, and DA Weitz. Elastic behavior of cross-linked and bundled actin networks. *Science*, 304(5675):1301–1305, 2004.

[11] Daniel T Gillespie. Stochastic simulation of chemical kinetics. *Annu. Rev. Phys. Chem.*, 58:35–55, 2007.

[12] Frederick Gittes, Brian Mickey, Jilda Nettleton, and Jonathon Howard. Flexural rigidity of microtubules and actin filaments measured from thermal fluctuations in shape. *The Journal of cell biology*, 120(4):923–934, 1993.

[13] Enrico Grazi. What is the diameter of the actin filament? *FEBS letters*, 405(3):249–252, 1997.

[14] Li Hou, Katherine Luby-Phelps, and Frederick Lanni. Brownian motion of inert tracer macromolecules in polymerized and spontaneously bundled mixtures of actin and filamin. *The Journal of cell biology*, 110(5):1645–1654, 1990.

[15] Paul A Janmey, Soren Hvidt, J Käs, Dietmar Lerche, Anthony Maggs, Erich Sackmann, Manfred Schliwa, and Thomas P Stossel. The mechanical properties of actin gels. elastic modulus and filament motions. *Journal of Biological Chemistry*, 269(51):32503–32513, 1994.

[16] Robert E Johnson. An improved slender-body theory for stokes flow. *Journal of Fluid Mechanics*, 99(2):411–431, 1980.

[17] KE Kasza, CP Broedersz, GH Koenderink, YC Lin, W Messner, EA Millman, F Nakamura, TP Stossel, FC MacKintosh, and DA Weitz. Actin filament length tunes elasticity of flexibly cross-linked actin networks. *Biophysical journal*, 99(4):1091–1100, 2010.

[18] Joseph B Keller and Sol I Rubinow. Slender-body theory for slow viscous flow. *Journal of Fluid Mechanics*, 75(4):705–714, 1976.

[19] Philip A Kuhlman, Jacqueline Ellis, David R Critchley, and Clive R Bagshaw. The kinetics of the interaction between the actin-binding domain of α-actinin and f-actin. *FEBS letters*, 339(3):297–301, 1994.

[20] Shimin Le, Xian Hu, Mingxi Yao, Hu Chen, Miao Yu, Xiaochun Xu, Naotaka Nakazawa, Felix M Margadant, Michael P Sheetz, and Jie Yan. Mechanotransmission and mechanosensing of human alpha-actinin 1. *Cell reports*, 21(10):2714–2723, 2017.

[21] Oliver Lieleg, Mireille MAE Claessens, and Andreas R Bausch. Structure and dynamics of cross-linked actin networks. *Soft Matter*, 6(2):218–225, 2010.

[22] Oliver Lieleg, Kurt M Schmoller, Christian J Cyron, Yuxia Luan, Wolfgang A Wall, and Andreas R Bausch. Structural polymorphism in heterogeneous cytoskeletal networks. *Soft Matter*, 5(9):1796–1803, 2009.

[23] Katherine Luby-Phelps. Cytoarchitecture and physical properties of cytoplasm: volume, viscosity, diffusion, intracellular surface area. In *International review of cytology*, volume 192, pages 189–221. Elsevier, 1999.

[24] Rui Ma and Julien Berro. Structural organization and energy storage in crosslinked actin assemblies. *PLoS computational biology*, 14(5):e1006150, 2018.

[25] Masato Makino and Masao Doi. Brownian motion of a particle of general shape in newtonian fluid. *Journal of the Physical Society of Japan*, 73(10):2739–2745, 2004.

[26] Ondrej Maxian, Alex Mogilner, and Aleksandar Donev. Integral-based spectral method for inextensible slender fibers in stokes flow. *Physical Review Fluids*, 6(1):014102, 2021.

[27] Ondrej Maxian, Raul P Peláez, Alex Mogilner, and Aleksandar Donev. Simulations of dynamically cross-linked actin networks: morphology, rheology, and hydrodynamic interactions. *bioRxiv*, 2021.

[28] James L McGrath, Eric A Osborn, Yanik S Tardy, C Forbes Dewey, and John H Hartwig. Regulation of the actin cycle in vivo by actin filament severing. *Proceedings of the National Academy of Sciences*, 97(12):6532–6537, 2000.

[29] Rudolf K Meyer and Ueli Aebi. Bundling of actin filaments by alpha-actinin depends on its molecular length. *The Journal of cell biology*, 110(6):2013–2024, 1990.

[30] David C Morse. Viscoelasticity of concentrated isotropic solutions of semiflexible polymers. 1. model and stress tensor. *Macromolecules*, 31(20):7030–7043, 1998.

[31] KM Schmoller, O Lieleg, and AR Bausch. Structural and viscoelastic properties of actin/filamin networks: cross-linked versus bundled networks. *Biophysical journal*, 97(1):83–89, 2009.

[32] Brennan Sprinkle, Florencio Balboa Usabiaga, Neelesh A Patankar, and Aleksandar Donev. Large scale brownian dynamics of confined suspensions of rigid particles. *The Journal of chemical physics*, 147(24):244103, 2017.

[33] Marcus Tempel, Gerhard Isenberg, and Erich Sackmann. Temperature-induced sol-gel transition and microgel formation in α-actinin cross-linked actin networks: a rheological study. *Physical Review E*, 54(2):1802, 1996.

[34] Anna-Karin Tornberg and Michael J Shelley. Simulating the dynamics and interactions of flexible fibers in stokes flows. *Journal of Computational Physics*, 196(1):8–40, 2004.

[35] Daniel H Wachsstock, WH Schwartz, and Thomas D Pollard. Affinity of alpha-actinin for actin determines the structure and mechanical properties of actin filament gels. *Biophysical journal*, 65(1):205, 1993.

[36] Kimberly L Weirich, Shiladitya Banerjee, Kinjal Dasbiswas, Thomas A Witten, Suriyanarayanan Vaikuntanathan, and Margaret L Gardel. Liquid behavior of cross-linked actin bundles. *Proceedings of the National Academy of Sciences*, 114(9):2131–2136, 2017.

[37] Timothy A Westwood, Blaise Delmotte, and Eric E Keaveny. A generalised drift-correcting time integration scheme for brownian suspensions of rigid particles with arbitrary shape. *arXiv preprint arXiv:2106.00449*, 2021.

[38] Jingyuan Xu, Denis Wirtz, and Thomas D Pollard. Dynamic cross-linking by α-actinin determines the mechanical properties of actin filament networks. *Journal of Biological Chemistry*, 273(16):9570–9576, 1998.

[39] KM Zero and R Pecora. Rotational and translational diffusion in semidilute solutions of rigid-rod macromolecules. *Macromolecules*, 15(1):87–93, 1982.

6 Supplemental text

6.1 Rigid fibers as a special case of inextensible fibers

In [26], we discretized the inextensible system (2) by discretizing each fiber with N Chebyshev collocation points and representing the functions $\alpha_1(s)$ and $\alpha_2(s)$ by their Chebyshev coefficients, $\alpha_j(s) = \sum_{k=0}^{N-2} \alpha_{jk} T_k(s)$. Our goal here is to show that setting $\alpha_j(s) = \alpha_j = $ const. instead gives the straight rigid fiber kinematic operators (8) and (9). For convenience, we first restate the kinematic equations for inextensible fibers, which are Equations (41) and (44) in [26],

$$(\mathcal{K}[\boldsymbol{X}]\boldsymbol{\alpha})(s) = \overline{\boldsymbol{U}} + \int_0^s \sum_{j=1}^2 \sum_k \alpha_{jk} T_k(s') \boldsymbol{n}_j(\boldsymbol{\tau}(s')) \, ds' \tag{26.41}$$

$$\mathcal{K}^*[\boldsymbol{X}]\boldsymbol{\lambda} := \begin{pmatrix} \int_0^L \left(\int_0^s T_k(s') \boldsymbol{n}_1(\boldsymbol{\tau}(s')) \, ds' \right) \cdot \boldsymbol{\lambda}(s) \, ds \\ \int_0^L \left(\int_0^s T_k(s') \boldsymbol{n}_2(\boldsymbol{\tau}(s')) \, ds' \right) \cdot \boldsymbol{\lambda}(s) \, ds \\ \int_0^L \boldsymbol{\lambda}(s) \, ds \end{pmatrix} = \begin{pmatrix} 0 \\ 0 \\ \boldsymbol{0} \end{pmatrix}, \tag{26.44}$$

where the first two components of (26.44) hold for all k. Let us denote by $\mathcal{K}_s$ the operator $\mathcal{K}$ in the case of straight fibers with $k = 0$ being the only included Chebyshev mode, and likewise for $\mathcal{K}^*$. Then, since the fibers are straight, the orthonormal frame $(\boldsymbol{\tau}, \boldsymbol{n}_1, \boldsymbol{n}_2)$ is constant along the fiber, and thus $\mathcal{K}_s$ and $\mathcal{K}_s^*$ simplify to

$$\mathcal{K}_s \boldsymbol{\alpha} = \overline{\boldsymbol{U}} + \alpha_1 \boldsymbol{n}_1 s + \alpha_2 \boldsymbol{n}_2 s \tag{S1}$$

$$\mathcal{K}_s^* \boldsymbol{\lambda} = \begin{pmatrix} \boldsymbol{n}_1 \cdot \int_0^L s \boldsymbol{\lambda}(s) \, ds \\ \boldsymbol{n}_2 \cdot \int_0^L s \boldsymbol{\lambda}(s) \, ds \\ \int_0^L \boldsymbol{\lambda}(s) \, ds \end{pmatrix}. \tag{S2}$$

We now want to show that $\mathcal{K}_s$ and $\mathcal{K}_r$ (defined in (8)) parameterize the same linear space of rigid motions. To do this, let us write $\boldsymbol{X}$ in (8) as an integral of the tangent vector

$$\mathcal{K}_r \boldsymbol{\alpha} = \boldsymbol{U}_c + \boldsymbol{\Omega} \times (\boldsymbol{X}_0 + s\boldsymbol{\tau} - \boldsymbol{X}_c) \tag{S3}$$

$$= \boldsymbol{U}_c + \boldsymbol{\Omega} \times (\boldsymbol{X}_0 - \boldsymbol{X}_c) + s\boldsymbol{\Omega} \times \boldsymbol{\tau} \tag{S4}$$

$$= \widehat{\boldsymbol{U}}_c + s\left((\boldsymbol{\Omega} \cdot \boldsymbol{\tau})\boldsymbol{\tau} + (\boldsymbol{\Omega} \cdot \boldsymbol{n}_1)\boldsymbol{n}_1 + (\boldsymbol{\Omega} \cdot \boldsymbol{n}_2)\boldsymbol{n}_2\right) \times \boldsymbol{\tau} \tag{S5}$$

$$= \widehat{\boldsymbol{U}}_c + s\left(-(\boldsymbol{\Omega} \cdot \boldsymbol{n}_1)\boldsymbol{n}_2 + (\boldsymbol{\Omega} \cdot \boldsymbol{n}_2)\boldsymbol{n}_1\right). \tag{S6}$$

This is exactly the form of $\mathcal{K}_s$ in (S1) with $\alpha_1 = -\boldsymbol{\Omega} \cdot \boldsymbol{n}_2$ and $\alpha_2 = -\boldsymbol{\Omega} \cdot \boldsymbol{n}_1$. So $\mathcal{K}_r$ and $\mathcal{K}_s$ parameterize the same space.

To complete the equivalence, we now show that $\mathcal{K}_s^* \boldsymbol{\lambda} = \boldsymbol{0}$ iff $\mathcal{K}_r^* \boldsymbol{\lambda} = \boldsymbol{0}$. Obviously, $\int_0^L \boldsymbol{\lambda}(s)\, ds = \boldsymbol{0}$ in both cases, so we only have to deal with the torque constraint. If we use the fact that $\int_0^L \boldsymbol{\lambda}(s)\, ds = \boldsymbol{0}$, we can write the second component of (9) as

$$\mathcal{K}_r^* \boldsymbol{\lambda} = \int_0^L \left(\boldsymbol{X}(s) - \boldsymbol{X}_c \right) \times \boldsymbol{\lambda}(s)\, ds = \int_0^L \boldsymbol{X}(s) \times \boldsymbol{\lambda}(s)\, ds = \int_0^L s\boldsymbol{\tau} \times \boldsymbol{\lambda}(s)\, ds \tag{S7}$$

$$= \int_0^L s\boldsymbol{\tau} \times \left((\boldsymbol{\tau} \cdot \boldsymbol{\lambda}(s))\, \boldsymbol{\tau} + (\boldsymbol{n}_1 \cdot \boldsymbol{\lambda}(s))\, \boldsymbol{n}_1 + (\boldsymbol{n}_2 \cdot \boldsymbol{\lambda}(s))\, \boldsymbol{n}_2 \right)\, ds \tag{S8}$$

$$= -\boldsymbol{n}_1 \left(\boldsymbol{n}_2 \cdot \int_0^L s\boldsymbol{\lambda}(s)\, ds \right) + \boldsymbol{n}_2 \left(\boldsymbol{n}_1 \cdot \int_0^L s\boldsymbol{\lambda}(s)\, ds \right) \tag{S9}$$

And now, since $\boldsymbol{n}_1$ and $\boldsymbol{n}_2$ are orthogonal and nonzero, we see from (S2) that $\mathcal{K}_s^* \boldsymbol{\lambda} = \boldsymbol{0} \leftrightarrow \mathcal{K}_r^* \boldsymbol{\lambda} = \boldsymbol{0}$. This shows that we can implement rigid fibers using the same algorithms as in [26], except we just need to keep a single ($k = 0$) Chebyshev polynomial. Note that the value of κ we use does not matter except for numerical stability since the fibers stay straight for all time, and so we set $\kappa = 0$.

6.2 Form and coefficients of the rigid body mobility matrix

Because of the symmetry of the fiber, the mobility matrix and its "square root" for a single fiber can be written in the form

$$\boldsymbol{N}_{\text{tt}} = \frac{1}{\mu L} \left(\alpha(\epsilon)\boldsymbol{I} + \beta(\epsilon)\boldsymbol{\tau}\boldsymbol{\tau} \right), \qquad \boldsymbol{N}_{\text{tt}}^{1/2} = \frac{1}{\sqrt{\mu L}} \left(\sqrt{\alpha}\boldsymbol{I} + \left(-\sqrt{\alpha} + \sqrt{\alpha + \beta} \right) \boldsymbol{\tau}\boldsymbol{\tau} \right), \tag{S10}$$

$$\boldsymbol{N}_{\text{rr}} = \frac{\gamma(\epsilon)}{\mu L^3} \left(\boldsymbol{I} - \boldsymbol{\tau}\boldsymbol{\tau} \right), \qquad \boldsymbol{N}_{\text{rr}}^{1/2} = \sqrt{\frac{\gamma}{\mu L^3}} \left(\boldsymbol{I} - \boldsymbol{\tau}\boldsymbol{\tau} \right).$$

Notice that the rotational mobility has a null space of the tangent vector $\boldsymbol{\tau}$. The dimensionless coefficients $\alpha, \beta,$ and γ are given for various ϵ in Table S1.

Figure S1: Link density, bundle density, % of fibers in bundles, mean bundle alignment, mean bundle size, and maximum bundle size, in a network of semiflexible ($\kappa = 0.07$ pN·μm^2) non-Brownian filaments with initial mesh size $\ell_m = 0.2$ μm. We show curves with different domain sizes (in μm) to establish that the statistics are repeatable in larger systems. The only statistic which is not repeatable is the maximum bundle size after many of the filaments have collapsed into one bundle ($t \gtrsim 40$ s).

Figure S2: Bundle sizes over time in the system with $F = 675$ semiflexible filaments in a domain of size $L_d = 3$ μm ($\tau_c \approx 16$). We show the percentage of fibers that are in bundles of various sizes over time for $\kappa = 0.07$ pN$\cdot \mu$m^2 (blue), $\kappa = 0.007$ pN$\cdot \mu$m^2 (orange), and rigid fibers (yellow). For times $t = 25, 40$, and 60 seconds, the x coordinate reflects the center of a histogram bin with logarithmically-scaled width. At $t = 5$ s, 25% of the filaments are in bundles of sizes 2 or 3, while most of the other fibers are not in bundles. At $t = \tau_c = 16$ s, about 50% of the fibers are in bundles of size 10 or less, with a small percentage in larger bundles, and the rest not in bundles at all (this is the composite bundle phase). For semiflexible fibers with $\kappa = 0.07$ and rigid fibers, about 75% of the fibers are in bundles of size 30 or larger by $t = 60$, while for fibers with $\kappa = 0.007$ the entire suspension has coalesced together by $t = 40$ seconds.

ϵ	$\alpha^{(\text{FP})}$	$\beta^{(\text{FP})}$	$\gamma^{(\text{FP})}$
0.01	0.3841	0.2230	3.4263
0.008	0.4020	0.2413	3.6412
0.006	0.4251	0.2646	3.9179
0.005	0.4396	0.2793	4.0931
0.004	0.4575	0.2973	4.3073
0.002	0.5129	0.3530	4.9721
0.001	0.5682	0.4085	5.6361

Table S1: Mobility coefficients for the 6×6 rigid body mobility matrix $\boldsymbol{N}$ defined in (S10) for straight fibers. The numerical estimate of the matrix $\boldsymbol{N}$ is related to the slender body mobility $\boldsymbol{M}$ in (11), which is computed using intra-fiber hydrodynamics as discussed in Section 2.1.2.

Figure S3: Square root of average mean square displacement of the fibers' centers over a time span of 0.05 seconds for simulations with Brownian dynamics and varying k_{on} (colors) and mesh size (from small mesh size to large going from left to right), all normalized by the value for a freely diffusing fiber. We normalize time by τ_c. While all of the curves show significant decay on the timescale τ_c, it is clear that τ_c is not the only timescale in the problem, since curves with small k_{on} are qualitatively different. This is not a surprise, since we saw in the main text that the bundling process with small k_{on} is more biased towards large bundles (see Fig. 5).

Figure S4: Representative snapshots of a network of rigid fibers without (left) and with (right) Brownian motion. Both snapshots are taken at $t = \tau_c$, which is 16 seconds for simulations without fluctuations and 4 seconds for simulations with fluctuations. The networks are qualitatively the same.

Figure S5: Decay of fibers' centers' displacement for different fiber lengths and mesh sizes. We show (the square root of) the average MSD of the fibers' centers over a time span of 0.05 seconds, normalized by the free space diffusion for fibers of the same length. We show mesh size $\ell_m = 0.2$ μm (blue), 0.4 μm (red), and 0.8 μm (black). Lighter colors are for filament length $L = 0.5$ μm, darker are $L = 1$ μm.

Figure S6: Steady state morphologies for systems with turnover. We introduce filament turnover with mean filament lifetime τ_f (see [27, Sec. 2.3] for implementation details) and observe the steady state bundle density (left) and percentage of fibers in bundles (right) for $\tau_f/\tau_c = 1/2$ (black), 1 (green), and 2 (red). Using both non-Brownian (lighter colors, $\tau_c \approx 16$ s) and Brownian (darker colors, $\tau_c \approx 4$ s) filaments, we show that the steady state bundling statistics depend primarily on the ratio τ_f/τ_c, and not the absolute value of either quantity.

Figure S7: Steady states with fiber turnover for (top to bottom) $\tau_f = \tau_c/2$, $\tau_f = \tau_c$, and $\tau_f = 2\tau_c$ for a system with $L_d = 2$ and $\ell_m = 0.2$. The left column is for non-Brownian fibers ($\tau_c = 16$ seconds) and the right column is for Brownian ones ($\tau_c = 4$ seconds). There is little qualitative difference between the left and right columns, which indicates that the network morphology is controlled primarily by the ratio τ_f/τ_c.