1 **State-dependent representations of mixtures by the olfactory bulb**
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9 **Abstract**
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Sensory systems are often tasked to analyse complex signals from the environment, separating
relevant from irrelevant parts. This process of decomposing signals is challenging when a mixture of
signals does not equal the sum of its parts, leading to an unpredictable corruption of signal patterns.
In olfaction, nonlinear summation is prevalent at various stages of sensory processing. Here, we
investigate how the olfactory system deals with binary mixtures of odours under different brain states,
using two-photon imaging of olfactory bulb (OB) output neurons. Unlike previous studies using
anaesthetised animals, we found that mixture summation is more linear in the early phase of evoked
responses in awake, head-fixed mice performing an odour detection task, due to dampened responses.
Despite this, and responses being more variable, decoding analyses indicated that the data from
behaving mice was well discriminable. Curiously, the time course of decoding accuracy did not
correlate strictly with the linearity of summation. Further, a comparison with naïve mice indicated
that learning to accurately perform the mixture detection task is not accompanied by more linear
mixture summation. Finally, using a simulation, we demonstrate that, while saturating sublinearity
tends to degrade the discriminability, the extent of the impairment may depend on other factors,
including pattern decorrelation. Altogether, our results demonstrate that the mixture representation
in the primary olfactory area is state-dependent, but the analytical perception may not strictly
correlate with linearity in summation.

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31 **Introduction**
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As animals in nature navigate through their environment in order to find food, mates, and to avoid
dangers, their sensory systems are tasked to detect and recognise signals of interest despite a
background of interfering signals. This figure-ground segregation is a ubiquitous task for many, if not
all, sensory systems. In the visual system, for example, segmentation of spatial patterns of light allows
animals to recognise objects despite some parts being obscured (Marr, 1982). In the auditory system,
spectral combinations of sound waves are recognized and strung together over time to form a stream,
allowing animals to recognise social calls from specific individuals among other noises (Bregman,
1990). In olfaction, too, animals face challenges in identifying an odour of interest in the presence of
other molecules (Laing and Francis, 1989; Rokni et al., 2014).
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Olfactory stimuli are first detected by a large family of olfactory receptors residing in the nasal
epithelium (Buck and Axel, 1991). Due to a broad ligand-receptor binding (Araneda et al., 2000; del

Mármol et al., 2021; Malnic et al., 1999), each odour molecule may activate a number of olfactory
receptor types, leading to combinatorial representations (Malnic et al., 1999). As a result, when
several compounds are present in a given mixture, they can activate overlapping sets of olfactory
receptors, causing complex pharmacological interactions. For example, molecules may bind a
common receptor, which, depending on the efficacy, can lead to antagonism (Cruz and Lowe, 2013;
Kurahashi et al., 1994; Oka et al.; Reddy et al., 2018; Singh et al., 2019) or enhancement (Xu et al.,
2020). Recent large-scale studies demonstrate that this is a widespread phenomenon (Inagaki et al.,
2020; Xu et al., 2020; Zak et al., 2020), which means that neural responses to mixtures often do not
equal the sums of responses to the individual odours. In addition to the interactions at the periphery,
there are many forms of nonlinear summation at multiple stages of olfactory processing, including the
saturation of neural responses (Firestein et al., 1993; Wachowiak and Cohen, 2001) and inhibitory
interactions in the olfactory bulb (Economo et al., 2016). Widespread suppressive interactions are also
observed in downstream areas, including the piriform cortex (Penker et al., 2020; Stettler and Axel,
2009).
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Nonlinear summation of signals in some brain areas may be desirable when specific combinations of
signals carry special meanings (Agmon-Snir et al., 1998; Jacob et al., 2008). However, in primary
sensory areas, it is sometimes considered information limiting (Laughlin, 1989). For olfaction, this is
thought to limit the analytical ability – whether a mixture *can* be perceived in terms of the constituent
qualities (Bell et al., 1987; Jinks and Laing, 1999; Laing and Glemarec, 1992). Nonlinear summations,
or "interactions", do not occur for all odour mixtures (Fletcher, 2011; Gupta et al., 2015; Tabor et al.,
2004), but occur prevalently when the background and target activation patterns overlap. This
happens when a mixture contains many components (Mathis et al., 2016), as well as when the
background odours are structurally related to the target odour (Cruz and Lowe, 2013; Fletcher, 2011;
Jinks and Laing, 1999; Kay et al., 2003; Mathis et al., 2016; Tabor et al., 2004). Nonlinear summation
poses a difficulty because it may distort a pattern of interest brought by non-uniform addition of
unpredictable background patterns, although more recent studies suggest beneficial effects of
antagonism, for example by reducing saturation-related loss of information (Reddy et al., 2018). Due
to this difficulty, some studies have suggested that the olfactory system may not decompose mixture
representations into component parts, but may solve the task by learning task-specific boundaries
instead (Mathis et al., 2016; Wilson and Stevenson, 2003).
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The question therefore remains: how does the mammalian olfactory system deal with nonlinear
summation of responses? To investigate this, we used binary mixtures of odours to investigate mixture
representations in mice performing tasks. While temporal structures caused by turbulence may be
used to segregate odours of interest from the background (Ackels et al., 2021; Hopfield, 1991), we
tackle the case when temporal information from the environment is not available. We demonstrate,
by comparing the mixture responses using various paradigms, that the property of mixture summation
depends on the brain state.
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Results

Olfactory figure-ground segregation is most difficult when the target and background odours evoke overlapping activity patterns (Rokni et al., 2014). To study this task using binary mixtures, we first characterized how our target odour relates to other odours in our panel. The target odour was ethyl butyrate and the rest of the odours in our set comprised a range of small esters structurally similar to ethyl butyrate, as well as non-esters (Fig. 1A). According to Rokni et al., the masking index, which measures the amount of overlap in response patterns between the target odour and background odours, correlates well with behavioural performances (Rokni et al., 2014).

To measure the masking indices from single odour responses, we obtained activity patterns from the OB output neurons, the mitral and tufted (M/T) cells. Using a two-photon microscope, we imaged from the glomerular layer of the olfactory bulb in *Tbet-Cre::Ai95D* mice, which express the calcium indicator GCaMP6f (Dana et al., 2019) in M/T cells (Haddad et al., 2013). To reproduce previous results, these initial imaging experiments took place in mice under anaesthesia (Fig. 1B). We studied the degree of overlap between ethyl butyrate and other odour responses by presenting single odours in a randomized order. An analysis of glomerular activity patterns revealed that methyl butyrate responses overlap the most with the ethyl butyrate response patterns, followed by closely related esters (Fig. 1C-E).

Previous studies from anaesthetised animals showed that neural responses to odour mixtures exhibit widespread nonlinear summation (Inagaki et al., 2020; Oka et al.; Reddy et al., 2018; Singh et al., 2019; Xu et al., 2020). In particular, suppressive interactions become dominant among large responses, due to the saturation effect (Mathis et al., 2016). This pattern is observed at many stages of olfactory processing, including in the olfactory bulb (Economo et al., 2016; Fletcher, 2011) and anterior piriform cortex (Penker et al., 2020), but it depends on the complexity of mixtures, as well as odorant choices (Fletcher, 2011; Gupta et al., 2015; Rokni and Murthy, 2014; Tabor et al., 2004). We therefore characterized the property of binary mixture summation using our odour set.

We first confirmed that our olfactometer is capable of presenting stable concentrations of odours for mixtures. We used a photoionisation detector to ensure that, when two odours are mixed, ionisation levels sum linearly (Fig. 2A-C). Then, to assess how the OB output represents binary mixtures, we imaged the individual somata of M/T cells in *Tbet-cre::Ai95D* mice (Fig. 2D). A typical session consisted of about 40 trials, to minimise time-dependent effects. Single odours and their binary combinations were presented in a semi-random order. Since ethyl butyrate, methyl butyrate, and the mixture of these two odours are of particular importance in this study, these three trial types appeared every 10 trials (Fig. 2E; see methods). To assess how the mixture of ethyl butyrate and methyl butyrate is represented, the amplitudes of the mixture responses were compared against those of linear sums of single odour responses (Fig. 2F,G). As reported before, a large proportion of M/T cells exhibited nonlinear summation (39.0% of M/T cells showed a deviation from linearity greater than 2; 71/182 ROIs, 7 fields of view, 4 mice; see methods) with a large fraction showing sublinear summation (35.7%, 65/182 ROIs).

Given that nonlinear summation is so widespread in the OB output, how well can mice analyse binary mixtures to accurately detect the presence of the target odour at the behavioural level? To assess this,

we used a Go/No-Go paradigm for head-fixed mice. The rewarded stimulus (S+ odour) contained ethyl
butyrate, either as a single odour or as a component of binary mixtures (Fig 3A). To train mice on this
task efficiently, after habituation, the head-fixed mice were first trained to discriminate ethyl butyrate
against other single odours (Fig 3A). The mice learned to perform this task well, reaching an accuracy
of 80% within 200 trials on average (Fig. 3 B,C; number of trials to reach 80% accuracy = 194.3 ± 21.9;
n = 7 mice). Subsequently, these mice were trained to detect the presence of ethyl butyrate in binary
mixtures (Fig. 3B,C). The mice performed the mixture task at a high accuracy from the beginning (90.2
± 1.3 % of trials with correct responses in the first session; n = 7 mice). However, the mistakes they
made were odour-specific, in that they tended to lick more indiscriminately on methyl butyrate-
containing trials regardless of the presence of ethyl butyrate (Fig 3D,E; mean lick preference index for
MB trials = 0.74 ± 0.04 vs. 0.93 for 5 other background odours, p = 0.0035, 1-way ANOVA; n = 7 mice).
However, with training, the performance on methyl butyrate trials, too, became accurate,
demonstrating that mice can learn to accurately detect the target odour in binary mixtures even when
the background odour is similar.
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To understand why mice were able to acquire the binary mixture task so easily, we analysed how M/T
cells represent binary olfactory mixtures while the trained mice accurately performed the task.
GCaMP6f fluorescence was measured from M/T cell somata. Data from the behaving mice was
compared to the case when the same mice were later anaesthetised with ketamine and xylazine (Fig
4A-D). This revealed that the M/T cells in behaving mice tended to sum mixture responses more
linearly than when the mice were anaesthetised (Fig. 4D). To quantify this data, we expressed a
deviation from the linear sum as a fraction of the predicted, linear sum of component responses
("fractional deviation"; Fig. 4E). Data from awake, behaving mice showed a clear shift in the
distribution of this index, compared to the anaesthetised case. In the anaesthetised case, a steep slope
in the cumulative histogram occurs at a negative value (median fractional deviation = -0.39), indicating
that the majority of the OB output neurons exhibit sublinear summation. On the other hand, M/T cells
from behaving mice showed a broader distribution centred around 0 (median fractional deviation = -
0.12; p = 3.83 x 10^{-10}, two-sample Komogorov-Smirnov test comparing the behaving vs. anaesthetised
data; n = 202 ROIs and 103 ROIs, respectively). The broader distribution for the awake, behaving
condition likely reflects a greater variability in this condition. Consistent with this, adding a Gaussian
noise to the data from anaesthetised mice made the distribution broader, but without affecting the
median (Fig. 4F). Using this index to compare the two conditions, we found that the difference was
most striking in the first second after the odour onset ("early phase"; Fig. 4G). Further, we observed
that the responses in the somata are more linear at an early phase compared to the apical dendrites
(Fig. 4 – supplement 1). These observations together may indicate that dampened responses in the
early phase of somatic responses may underlie the reduced mixture suppression, although some
contribution may come from more decorrelated responses at the somata (Fig. 4 – supplement 1).
Overall, the results indicate that the property of mixture summation is more linear in the awake,
behaving mice, and that linearisation is not imprinted permanently as a result of learning.
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While the above result demonstrates that mixtures sum more linearly in awake, behaving mice, M/TC
responses are also significantly smaller and more variable in this condition. This is quantified in an
analysis of trial-by-trial response similarity, which measures the Pearson correlation coefficient
between population responses from individual trials (Fig. 5A-C). This showed that, in the
anaesthetised mice, the M/TC response patterns are highly correlated, both within class (comparison

between trials with ethyl butyrate) and across classes (Fig. 5A,B). In the awake, behaving mice, as
expected, responses were less consistent, indicated by the lower within-class correlation (Fig. 5A,B).
However, because across-class patterns were substantially less correlated (Fig. 5A,B), S+ responses
may be more discriminable in the behaving mice.
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The ability to reliably encode the presence of ethyl butyrate was first quantified based on within-class
vs. across-class correlation, by counting the proportion of trials where within-class correlation was
higher than across-class (Bridgeford et al., 2021). This indicated that M/T cells from anaesthetised
mice showed better discriminability initially, but later, M/T cells in the behaving mice performed as
well as the anaesthetised mice (Fig. 5C). Note a similar result is obtained when trials are sorted by the
presence of methyl butyrate (Fig. 5 supplement 1). While useful, the Pearson correlation coefficient
takes into account all ROIs, irrespective of how informative they are in discriminating patterns, and
may not reflect the true ability of a population of neurons to encode odours. To address this, we
trained support vector machines (SVMs) to discriminate responses evoked by odours containing ethyl
butyrate (S+) vs. odours without ethyl butyrate (S-). We used 80% of randomly selected trials from
each session for training, and the remaining 20% of the trials to test the performance (Fig. 5D). This
time, M/T cells from the two conditions performed similarly for the first 1 second, performing above
chance soon after the odour onset (earliest time where accuracy is significantly above 0.5 = 0.67 s for
the behaving case, and 1.12 s for anaesthetised data; t-test at significance level of 0.05; n = 13 fields
of view, 6 mice for behaving case; 8 fields of view, 4 mice for the anaesthetised case; Fig. 5D). Further,
the data from behaving mice outperformed the anaesthetised case in the later phase (median
accuracy for 2-3 s after odour onset = 0.58 vs. 0.70 for anaesthetised vs. behaving mice, respectively;
p = 0.027, Mann-Whitney U test for equal medians). We also wished to assess how mixture responses
relate to single odour patterns. To do so, we trained SVMs using single odour responses (EB vs. other
single odours) and tested the performance on mixture responses (Fig. 5E). Again, the data from
behaving mice outperformed the data from anaesthetised mice in the late phase although the
difference was not statistically significant (median accuracy for 2-3 s after odour onset = 0.55 vs. 0.64
for anaesthetised vs. behaving mice, respectively; p = 0.065, Mann-Whitney U test for equal medians).
This may suggest that, over time, mixture responses come to resemble the target component odour
pattern more in the awake, behaving mice than in the anaesthetised mice. Curiously, the time course
of decoder accuracy did not strictly depend on the linearity of mixture summation (Fig. 6 – figure
supplement 2). Overall, the result suggests that, despite the substantial differences in the amplitudes
and variability of responses, M/T cells in behaving mice are able to encode the presence of the target
odour well.
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To what extent is the mixture representation in the olfactory bulb affected by learning? Previous
studies suggest that prior exposures and familiarity to odours affect the ability to analyse odour
mixtures (Grabska-Barwińska et al., 2017; Poupon et al., 2018). To address this, we assessed how ethyl
butyrate and methyl butyrate mixtures are represented in naïve but awake, head-fixed mice. Since
different levels of motivation, with accompanying changes in the sniff patterns, affect the olfactory
system (Carey and Wachowiak, 2011; Jordan et al., 2018), we made one group naïve mice engaged
and a second group of mice disengaged, by changing the reward contingency (Fig. 6A). That is, we
presented the same sets of odour stimuli, but instead of associating specific odours with reward, the
water reward was either delivered after odour onset but on randomly selected trials to engaged the
mice, or about 15 seconds before the odour onset on all trials in order to disengage the mice (Fig. 6A;

n = 17 sessions, 6 mice and 14 sessions, 4 mice). The level of engagement was confirmed by the speed
of inhalations during odour presentations (Fig. 6B) and the number of anticipatory licks generated (Fig.
6 - figure supplement 1).
224
In all awake conditions, we found the ethyl butyrate and methyl butyrate responses to sum equally
linearly (Fig. 6 - figure supplement 1), indicating that the linearity of summation does not depend on
the behavioural states or learning. However, the performance of SVMs on these datasets showed a
state-dependence. While the general ability to discriminate S+ vs. S- odours was comparable across
the states, the ability to analyse mixtures in terms of the constituents - SVMs trained on single odours
and tested on mixture patterns – was particularly poor for the disengaged mice (Fig. 6C-F). Since the
linearity of summation was comparable across the three groups (Fig.6 – figure supplement 1), this,
again, suggests that the ability to correctly decode the presence of odour does not strictly correlate
with the linearity of mixture summation. In all cases, the decoder performed most accurately in the
late phase. We hypothesized that this extra time may reflect an involvement of recurrent interactions
with the piriform cortex, which is thought to store learned representations and feed back to the
olfactory bulb (Chapuis and Wilson, 2012; Grabska-Barwińska et al., 2017). Our preliminary result from
muscimol infusion in the ipsilateral piriform cortex, however, suggests that this may not be the case
(Fig. 6 supplement 3).
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Under what circumstance does discriminability of odour mixtures decouple from linearity in mixture
summation? To study the effect of saturating sublinearity in isolation, we made a simulation as follows.
Linear sums of responses were constructed by adding component responses obtained from the
imaging sessions. In one test, these sums, with added noise, were passed through SVMs that had been
trained with component responses. In another test, the linearly summed responses with noise were
transformed with a normalising function (Mathis et al., 2016; Penker et al., 2020), which adds
sublinearity in an amplitude-dependent manner (Fig. 7B; see methods). Then, these signals were
passed through the same SVMs to assess how discriminable the activity patterns were. When the data
from the anaesthetised mice was used, normalising sublinearity was particularly detrimental around
2 seconds after the odour onset (Fig. 7C), qualitatively reproducing the transient decrease in the
accuracy seen with the observed mixture responses (Fig. 5E). In contrast, with the data from the
behaving mice, normalising sublinearity did not have as significant an effect on the mixture
discriminability, even at the later stage when sublinear summation becomes more widespread (Fig.
7D,E). Thus, the functional consequence of nonlinear summation may depend on specific
circumstances, for example on how separable, or decorrelated, the activity patterns are. Overall, our
result indicates that mixture responses in the olfactory bulb are highly state-dependent and evolve
over time (Fig. 6F).
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Discussion

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Segmentation and extraction of relevant information from mixtures of signals are important and
perpetual tasks for sensory systems. Nonlinear summation of signals is generally thought to pose
difficulty in demixing component signals. Here, we demonstrate that the signal summation in the
primary olfactory area of the mouse is significantly more linear in awake mice. This is notable, because
many previous studies reported that suppressive mixture interactions are widespread in the central

nervous system, based on anaesthetised preparations (Bell et al., 1987; Cruz and Lowe, 2013; Penker
et al., 2020; Stettler and Axel, 2009).

However, our analysis also indicates that the ability to discriminate binary mixture responses does not
correlate strictly with linearity in summation. First, the behavioural improvement was not
accompanied by more linear summation. Second, the time course of decoder accuracy does not follow
the linearity of summation. Indeed, in the data from behaving mice, the decoder performance peaked
when mixture responses were most sublinear. This raises a question: is sublinear summation in
mixtures detrimental, or beneficial to discriminating mixtures of odours? Our simple simulation
indicates that, generally, a sublinear mixture summation due to saturating influences limits the
discriminability of mixture responses. Thus, dampened responses in awake animals bring some
advantage by maintaining the mixture responses in the linear range. However, in the behaving mice,
even when the responses became larger at a later phase, the saturating influence was less detrimental
to discrimination. Here, decorrelated response patterns may play a more crucial role, as it is known to
enhance classifier or behavioural performances (Bhattacharjee et al., 2019; Friedrich and Wiechert,
2014; Gschwend et al., 2015; Padmanabhan and Urban, 2010). While the behavioural task described
in this study is simple and animals make accurate decisions within 1 s (771 ± 97 ms) of stimulus onset,
the slower mechanisms described here may be important when larger and more reliable responses
are required for accurate decisions. So, in addition to sampling time (Rinberg et al., 2006), this
phenomenon may be a part of mechanisms needed to solve more difficult tasks accurately (Abraham
et al., 2010; Wilson et al., 2017).

Mechanistically, the ability to accurately analyse, or discriminate between, mixtures based on the
knowledge of component is hypothesised to involve the piriform cortex, through learned
representations (Chapuis and Wilson, 2012; Grabska-Barwińska et al., 2017). For tufted cells, however,
the relevant sources of feedback could be other brain regions, such as the anterior olfactory nucleus
(Chae et al., 2021). It will be an intriguing future investigation to identify the source and mechanisms
of state-dependent mixture representations.

Perception of olfactory mixtures has long fascinated investigators. Mixtures of odours often have
qualities that are different from those of the individual components. Accurate recognition of
components is particularly hard for human subjects. For untrained subjects, olfactory mixtures that
contain about 30 components tend to smell alike (Weiss et al., 2012). Even highly trained people like
perfumers can only accurately identify individual components if unfamiliar mixtures contained no
more than 5 components (Poupon et al., 2018). These demonstrate an ultimate limit in the analytical
perception of olfactory mixtures. In addition, in non-expert humans and rodents alike, when the task
is to look for a particular aroma in the mixture, the tendency is to falsely report the presence of the
target odour in target-odour detection tasks (Laing and Glemarec, 1992; Rokni et al., 2014). In all cases,
an extensive training for specific odours can improve the ability to detect the target odour, as seen in
the case for sommeliers who routinely analyse key components in wines, which can contain several-
hundred component mixtures (Ilc et al., 2016). Thus, while mixture perception is highly context
specific (Rokni and Murthy, 2014), training in specific odours seems to be key to improving on mixture
analysis. With more complex mixtures, mechanisms within the olfactory bulb may reach a limit, and
also is unlikely to be the only mechanism that is used to solve the task. Elucidating what roles the

primary sensory areas plays is a crucial step towards a mechanistic understanding of complex sensory
processing.
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313 **Acknowledgement**
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Methods

Animals

All animal experiments have been approved by the OIST Graduate University's Animal Care and Use
Committee (Protocol 2016-151 and 2020-310). Tbet-Cre (Haddad et al., 2013) and B6J.Cg-
Gt(ROSA)26Sor$^{tm95.1(CAG-GCaMP6f)Hze}$/MwarJ, also known as Ai95D (Madisen et al., 2015), were originally
obtained from Jackson Laboratory (stock numbers 024507 and 028865, respectively). Tbet-Cre::Ai95D
mice were generated by crossing homozygous Tbet-Cre and Ai95D mice resulting in heterozygous
animals used for imaging experiments. C57Bl6J mice were purchased from Japan CLEA (Shizuoka,
Japan) and were acclimatized to the OIST facility for at least 1 week before they were used for
experiments. All mice used in this study were adult male (8 – 11 weeks old at the time of surgery).

Olfactometry

A custom-made flow-dilution olfactometer was used to present odours. Briefly,
custom Labview codes were used to control to control solenoid valves, and flow controller (C1005-
4S2-2L-N2, FCON, Japan) was used to regulate the rate of air flow. A pair of normally closed solenoid
valves was assigned per odorant and used to odorise the air. These solenoid valves were attached to
a manifold, such that a set of 8 pairs had access to the common stream of air. To generate binary
mixtures of odours while keeping concentrations stable, one odour from each manifold (lines A &
B) was used. For single odour presentations, odour was mixed with air passed through an empty
canister. The odorised air was directed towards the animal only when the solenoid valve closest to
the animal (final valve) opened. Final valve was opened for a short time (0.1 s for experiments
involving behavioural analysis only, and 0.5 s for all imaging experiments, to accommodate for slow
respirations during anaesthesia), to avoid adaptation related to temporal filtering for high sniff
frequencies associated with long odour pulses (Verhagen et al., 2007). Odours were presented at 1-
5% of the saturated vapour. Total air flow, which is a sum of odorised air and the dilution air, was
approximately 2 L/min, which was matched by the air that normally flows towards the animal. Inter-
trial interval was approximately 40 seconds to ensure that the flow controllers have stabilised before
each odour presentation. All odorants were from Tokyo Chemical Industry (Tokyo, Japan), apart for
ethyl butyrate (W242705), which was from Sigma-Aldrich. Product number were: T0247 (Ethyl tiglate),
V0005 (Methyl valerate), A0061 (Acetophenone), A0500 (Methyl anthranilate), B0757 (Butyl butyrate),
B0763 (Methyl butyrate), S0015 (Methyl salicylate), S0004 (Salicylaldehyde), T0248 (Methyl tiglate),
A0232 (Eugenol). Purity of all odorants was at least 98 % at the time of purchase. Stock odorants were
stored at room temperature in a cabinet filled with N_2 and away from light.

Surgery

Head plate implantation: All recovery surgery was conducted in an aseptic condition. 8-11week old
male C57Bl6/J mice were deeply anaesthetised with isoflurane. The body temprature was kept at
36.5 °C using a heating blanket with a DC controller (FHC, Bowdoin, USA). To attach a custom head
plate about 1 cm in width weighing a few grams, the skin over the parietal bones was excised and the
soft tissue underneath was cleaned, exposing the skull. The exposed skull was gently scarred with a
dental drill, cleaned, dried, and coated with cyanoacrylate (Histoacryl, B.Braun, Hessen, Germany)
before placing the headplate and fixing with dental cement (Kulzer, Hanau, Germany). For optical
window implantation, adult, male *Tbet-Cre::Ai95D* mice were deeply anaesthetised, and underwent a
window implantation procedure as previously described (Koldaeva et al., 2019). Briefly, after exposing

the frontal bone, a craniotomy about 1 mm in diameter was made over the left olfactory bulb and on
the exposed dorsal surface, a cut piece of coverslip that snugly fit in the craniotomy, on the edge of
the drilled bone, was gently pressed down and sealed with a cyanoacrylate and fixed with dental
cement. Mice were recovered in a warm chamber, returned to their cages, and given carprofen
subcutaneously (5 mg/kg) for 3 consecutive days.
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Habituation and behavioural measurements
Water restriction began two weeks after surgery. Mice went through 3 days of habituation to head
fixation, one session per day for approximately 30 minutes, until mice learned to lick vigorously for
water reward. Respiration pattern was measured by sensing the air flow just outside the right nostril
by placing a flow sensor (AWM3100V, Honeywell, North Carolina, USA), and the data was acquired at
1 kHz. Lick responses were measured using an IR beam sensor (PM-F25, Panasonic, Osaka, Japan) that
was part of the water port. Nasal flow, an analog signal indicating the odours used, lick signal, a copy
of the final valve and water valve timing were acquired using a data acquisition interface (Power1401,
CED, Cambridge, UK).
380
Discrimination training
After habituation, the head-fixed mice were trained to associate a water reward with a target odour
(ethyl butyrate). The reward was two droplets of water (10 μl each), that arrived 3 seconds after the
onset of the final valve opening. The mice underwent single odour discrimination training first, until
they generated anticipatory licks in response to ethyl butyrate presentations, and correctly refrained
from licking in response to other single odours. Once the overall accuracy was above 80% in at least
one behavioural session, the mice went through the mixture detection task. A typical training session
comprised roughly 100 trials, lasting about 1 hour. Rewarded trials comprised a third of all trials. Two-
photon imaging took place once the mice performed at 80% accuracy or above.
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Random association paradigm
After habituation, the head-fixed mice were presented with the same odour mixture stimuli as those
that underwent the discrimination training. The water reward was delivered on randomly selected
trials, 3 seconds after the onset of the final valve opening. One behavioural session was used to
accustom the mice to the odours. Two-photon imaging commenced from the second behavioural
session.
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Disengagement paradigm
After habituation, the head-fixed mice were presented with the same odour mixture stimuli as above.
The water reward was delivered every trial, 15 seconds before the onset of the final valve opening,
arriving in the middle of the inter-trial interval, which was 40 seconds to ensure thorough purging to
clear the lines, as well as to stabilize the flow controllers. Two-photon imaging commenced from the
second behavioural session.
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Odour mixture trial composition
Binary mixtures have been chosen due to the smaller number of possible odour combinations
compared to more complex mixtures. However, even with 11 odours, there is a limit in the number of
trials each head-fixed mouse can sample in a given session. We therefore decided to focus on the EB
+ MB mixtures. However, it was crucial that, EB, MB, and EB + MB mixture are not presented too

frequently, to avoid possible adaptation, as well as mice becoming over familiar, especially in the EB-
detection task, where the goal was not to train mice to remember specific odour combinations. Thus,
we used a compromise paradigm where EB, MB, and EB + MB appeared every 10 trials.
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Data analysis
The data was analysed offline using custom Matlab codes. To calculate the accuracy, the number of
licks during 3 seconds from the final valve onset was measured for each trial. Threshold for an
anticipatory lick was set to 2, thus the correct response for rewarded trials was 2 or more beam breaks,
and the correct response for unrewarded trials was 1 lick or less during the response time window. To
calculate the learning curve, the accuracy was expressed as the proportion of correct trials in a
given block of 50 trials. *Time to inhalation peak*: to calculate the speed of inhalation, onset of
inhalation and peak of inhalation was detected using Spike2 (CED, Cambridge, UK), using the built-in
event detection functions. Briefly, inhalation peaks were detected using the "rising peak" function.
These events were used to search backwards in time for the inhalation onset when the flow signal
crossed a threshold value. *Lick preference index*: To measure how well mice discriminated rewarded
vs. unrewarded mixtures, anticipatory licking patterns for the two types of trials were compared using
the following formula:
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428 $$\text{Lick preference index} = (\text{Lick}_{rewarded} - \text{Lick}_{unrewarded})/(\text{Lick}_{rewarded} + \text{Lick}_{unrewarded})$$
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430 Where $\text{Lick}_{rewarded}$ corresponds to the average number of anticipatory licks on rewarded trials, and
431 $\text{Lick}_{unrewarded}$ corresponds to that for unrewarded trials.
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Imaging
A custom-made two-photon microscope (INSS, UK) with a resonant scanner was used to observe
fluorescence from the olfactory bulb of *Tbet-Cre::Ai95D* mice *in vivo*. 3D co-ordinates for imaged field
of view were recorded relative to the location of a reference, blood vessel pattern on the
surface. Unless otherwise stated, imaging from somata was done relatively superficially, just below
the glomeruli, thus mainly comprised TCs, which use firing rate modulation to represent odours
(Fukunaga et al., 2012). Fields of view for glomerular and somatic levels were 512 μm x 512 μm, and
256 μm x 256 μm, respectively, and overlapped for awake and anaesthetised conditions, but fewer
sessions took place under anaesthesia. In each trial, 400 image frames were acquired at 30 frames per
second, with 200 frames before the final valve opening to obtain steady baseline. Unless otherwise
stated, the time window analysed for the odour-evoked responses was the first 1 second since the
onset of final valve opening. For imaging under ketamine/xylazine anaesthesia (100 mg.kg^{-1}/20 mg.kg^{-1}
intraperitoneally), mice were kept on a warm blanket (FHC, Bowdoin, USA) to maintain the body
temperature at 36 °C.
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Ipsilateral muscimol infusion in the anterior piriform cortex
Animals were unilaterally implanted with a 26-gauge cannula (10 mm length; C315GS-4/SPC, Plastics
One). Cannula was inserted through a small craniotomy at a stereotactic coordinate [AP, 2.2 mm; ML,
2.4 mm relative to bregma] and advanced over 6.1 mm at 45 degree to target the anterior piriform
cortex, and fixed with dental cement (Kulzer, Hanau, Germany). For infusion, 500 nL of 2mM muscimol
(M1523, Sigma-Aldrich, Missouri, USA) was injected using a Hamilton microsyringe, at a rate of 100
nL/min approximately 10 minutes before the imaging started.

Post-hoc verification of infusion cannula placement: Animals were infused with 500nl DiI (Invitrogen,
V22885), immediately transcardially perfused with phosphate buffer (225.7 mM NaH$_2$PO$_4$, 774.0 mM
Na2HPO$_4$. pH 7.4) and PFA (4%, dissolved in phosphate buffer. The heads (including the implants) were
postfixed in PFA at 4°C for at 48 – 96 hours before extraction of the brains. Coronal sections of 100
μm thickness were cut on a vibratome (5100 mz-Plus, Campden Instruments, Leicestershire, UK) and
counterstained using DAPI (D9542, Sigma-Aldrich). Images were acquired using a Leica SP8 confocal
microscope using a 10X (NA 0.40 Plan-Apochromat, Leica) objective. Images were taken at a resolution
of 1024 x 1024 pixels per field of view (1163.64 x 1163.64 μm).

Image analysis

Transients were extracted as follows. Regions of interest (ROI) were manually delineated using an
average frame from each imaging session. For glomeruli, ROIs were delineated by tracing the outer
rim of congregated labelled processes surrounded by a dark region. Pixel values within each ROI were
averaged to obtain a time series. Imaging sessions with motion artefacts and drifts were removed
from analyses. For each transient, the baseline period was defined as 2 seconds preceding the final
valve opening. Relative fluorescence change (ΔF/F) was calculated with respect to this baseline. Odour
response period was 1 second (30 frames) starting at the onset of the final valve opening, unless
otherwise stated. Awake mice tended to adjust sniff patterns, so the final valve opening was not
triggered by nasal flow. Masking index for the glomerular level was calculated as previously described
(Rokni et al., 2014). Briefly, responses to single odours were obtained from glomeruli of anaesthetised
mice. In each field of view, only the glomeruli responsive to the target odour (EB) were analysed. To
obtain glomeruli evoked by the target odour, the evoked response amplitudes were converted into z-
scores. Glomeruli that responded to ethyl butyrate with z-scores higher than 2 were considered. The
masking index was the average overlap in the evoked response, where the maximum value for each
glomerulus was 1. For each odour, responses were averaged over 3 or more trials. Mean and standard
error of the mean are shown in figures, unless otherwise stated. Boxplots were constructed using the
Matlab function *boxplot*, and show the median, 25th and 75th percentiles, and extreme data points not
considered outliers. Outliers are shown with red crosses. The experiments were not done blindly, since
the stimulus-reward contingency was visible to the experimenter. However, the olfactometer
performance, age and sex of the mice, and analyses codes used were the same for all conditions.

Fractional deviation from linearity was calculated as $(R_{observed} - R_{linear\ sum})/| R_{linear\ sum} |$, where $R_{observed}$ is
the mean response amplitude for observed mixture (e.g., response to EB + MB mixture), $R_{linear\ sum}$ is
the trial average linear sum of component responses (e.g., EB response + MB response. Median
fractional deviation was obtained from each field of view. For testing the effect of trial-by-trial
variability, Gaussian noise was generated using the Matlab function *normrnd* with the mean set to 0,
and added to responses from each trial, before the fractional deviation was calculated.

Deviation from linearity was the difference between an observed mixture response amplitude and a
linear sum of components normalized by the joint standard deviation: $(R_{observed} - R_{linear\ sum})/(s.e.m._{observed} + s.e.m._{linearsum})$

Discriminability index based on correlation coefficient was calculated in the same way as *Dicr*,
described in (Bridgeford et al., 2021), except that the distance measure used was 1 − Pearson's
correlation coefficient, instead of the Euclidian distance.

Decoding analysis: Support vector machines were trained using the Matlab function *fitclinear* with
linear model intercept ("*FitBias*") set to 0. Test datasets were used to predict the trial types using the
Matlab function *predict*. Signals were averaged over 30 frames (~1 second) for each time point to
obtain the time course.

Simulating the effect of normalising sublinearity on discriminability : Support vector machines (SVMs)
were trained as above, using the single odour responses only. Then simulated mixture responses were
constructed as follows. Single odour responses were averaged over trials. Combinations of two
averaged responses were added linearly, and uncorrelated Gaussian noise (mean = 0) was added. The
noise term comprised two components: fixed, baseline standard deviation (0.2 for anaesthetised data,
and 2 for awake, behaving case) and standard deviation that scaled with response amplitude, by taking
the slope of a linear regression of the observed data. In one test, linear sum with added noise was
used as inputs to the above SVMs. In a second set of tests, these simulated responses were further
passed through a normalising sublinearity and tested on the same SVMs. The equation of
normalization (Penker et al., 2020) was:

$$R_j^* = R_{max}\left(\frac{2}{1 + e^{-s.R_j}} - 1\right)$$

where R_j represents the linear-sum amplitude of the jth neuron, R_j^* is the normalized response
amplitude of jth neuron. Parameters s and R_{max} were obtained by fitting the data to observed
mixture responses (on data from behaving animals at t = 2 seconds after odour onset). R_{max} was 6
and s was 0.2.

Figure 1: *Masking indices of odours used with respect to the ethyl butyrate pattern*

(**A**) Odours in the panel with abbreviations used in the rest of the manuscript. Ethyl butyrate was the target

odour for behavioural experiments. (**B**) Two-photon imaging of GCaMP6f signals from the apical dendrites

of M/T cells in *Tbet-Cre::Ai95D* mice under ketamine and xylazine anaesthesia. Scale bar = 0.1 mm. (**C**)

Example GCaMP6f transients expressed as a change in fluorescence (ΔF/F). Scale bar = 1 ΔF/F. Gray = Odour

presentation (0.5 s). (**D**) Example of evoked responses. Manually delineated ROIs are shown with

fluorescence change evoked by odours, indicated with the corresponding colour map. The amplitude

indicated is the average change during 1 second from the final valve opening. (**E**) Masking indices for all

odours in the panel, with EB as the target. N = 4 fields of view, 4 mice. Mean and SEM of 3 trials or more

shown.

Figure 2: *Mixture suppression dominates under anaesthesia*

(**A-C**) *Validation of linear mixing by the olfactometer.* (**A**) Binary mixtures are generated by mixing odorized air from two streams each equipped with a mass flow controller and presented as a stimulus when a five-way valve ("final valve") is actuated. To present single odours, odorized air from one line was mixed with air that passes through a blank canister in the other line. A photoionisation detector (PID) was used for calibration. (**B**) Example PID measurements for single odours ("Odour A" and "Odour B"), and their mixtures. Linear sum of the components (light blue trace) is shown superimposed with the observed PID signal (black). (**C**) Observed amplitudes for mixtures vs. linear sum of component amplitudes. Light blue line = the observed mixture amplitude equals the linear sum of components. (**D-G**) *Investigation of mixture summation by M/T cells under anaesthesia*. (**D**) Top: Schematic of two-photon GCaMP6f imaging from somata of M/T cells in naïve *Tbet-Cre::Ai95D* mice under ketamine/xylazine anaesthesia. Bottom: Example field of view. Scale bar = 50 μm. (**E**) Example session structure. Single EB and MB trials, as well as EB+MB mixture trials, are indicated with colour codes. (**F**) Transients from 3 example ROIs as highlighted in **D**. Linear sum (blue trace) was constructed by linearly summing averages of single EB and MB responses. Gray bar represents time of odour presentation (0.5 s). (**G**) Scatter plot of observed mixture response amplitude against linear sum of components. N = 183 ROIs, 7 fields of view, 4 mice.

Figure 3: *Mice can learn to accurately analyse difficult binary mixtures*

(**A**) Behavioural paradigm: Go/No-Go task with head-fixed mice. EB-containing olfactory stimulus was the rewarded stimulus. (**B**) After habituation, mice learned to discriminate ethyl butyrate against other single odours in the panel. Once proficient, mice learned to detect the presence of ethyl butyrate in binary mixtures. 30% of stimuli in the mixture stage were single odours. (**C**) Behavioural performance for all mice (n = 7 mice). Mean and SEM shown. (**D**) Odour-specific accuracy shown for the 1st (left) and last day of mixture training (right). Green shades indicate high accuracy. Top row corresponds to rewarded trials, and bottom 6 rows correspond to unrewarded trials. Average accuracy from all animals shown (n = 7 mice). (**E**) Lick preference index measures licks that occur preferentially on rewarded trials for a given background odour. A lick preference value of 1 occurs when all anticipatory licks were observed in rewarded trials only. Gray points = data from individual animals (n = 7 mice); Mean and SEM shown in black.

Figure 4: *Mixture summation is more linear in awake, behaving mice.*

(**A**) Schematic of experimental setup. Imaging of GCaMP6f signals from the somata in Tbet-Cre::Ai95D mice performing the mixture detection task. On the last day, imaging took place under ketamine/xylazine anaesthesia (Anae). (**B**) Example field of view showing the same neuron from two imaging sessions. (**C**) Relative fluorescence change evoked by EB, MB and their mixture for the two conditions, for the neuron indicated by arrow in (**B**). Grey bar = odour presentation (0.5 s). (**D**) Scatter plot of observed EB + MB mixture response amplitude against linear sum of component odour responses (average of 20 frames). Indicated time is relative to odour onset. (**E**) Cumulative histograms of fractional deviation from linear sum, for data from anaesthetised mice (blue) and trained mice performing mixture detection task (black). N = 202 ROIs from 13 fields of view, 6 mice for behaving case; 103 ROIs from 8 imaging sessions, 4 mice for the anaesthetised case. (**F**) Effect of Gaussian noise on the fractional deviation distribution. Colours correspond to data with different amount of noise added. (**G**) Time-course of median fractional deviation for the two datasets. Median deviation was obtained from each field of view; thick line shows median for all fields of view, along with 25th and 75th percentiles (shaded areas). See also **Figure 4 – figure supplement 1.**

Figure 5: *Encoding the presence of the target odour is better in behaving mice.*

(A) Correlation matrices for data from anaesthetised mice (top) and awake, behaving mice (bottom),

similarity of evoked responses between individual trials, for two time points, t1 and t2 as indicated in panel

(B). S+ trials were trials with ethyl butyrate either as single odour or in binary mixtures. (B) Average

correlation coefficient within S+ trials (magenta), within S- trials (cyan), or across trials (green) for M/T cells

responses from anaesthetised mice (top panel) and awake, behaving mice (bottom panel). (**C**)

Discriminability index, which measures the proportion of trials in which the within-S+ correlation was

higher than the across class, was compared at each time point for data from anaesthetised mice (blue

trace) and awake, behaving mice (black). See also **Figure 5 – figure supplement 1**. (**D**) SVM for

discriminating EB vs. non-EB responses, trained on M/TC somatic responses from randomly selected 80%

of trials, and tested on 20% of trials for anaesthetised (blue) and behaving (black) conditions. Both training

data and test data contained random selection of single odour responses (hollow rectangles in scheme)

and mixture responses (grey) rectangles. (**E**) SVM trained with single odour responses, tested on mixture

responses.

Figure 6: *Mixture representation depends on behavioural states.*

(**A**) Reward contingencies used to achieve different behavioural states in awake mice. Top: Mice were trained to discriminate between odours with EB vs. odours without EB. Middle: Naïve mice received water reward 3 seconds after the onset of odour, on randomly selected trials. Bottom: Naïve mice received water every trial, 15 seconds before odour onset. (**B**) Sniff patterns associated with trained and behaving mice (black), naïve and engaged mice (orange) and naïve and disengaged mice (light blue). Speed of inhalation during odour presentations shown. (**C**) Comparison of SVM performance using data from behaving mice (black) vs. naïve engaged mice (left panel, orange), and vs. naïve disengaged mice (right pane, light blue). SVM was trained to discriminate responses to S+ vs. S- odours using randomly selected 80% of trials and tested on the remaining 20% of the trials. (**D**) Decoder performance for the results in (**C**) plotted using approximate z-values obtained from Matlab implementation of Wilcoxon rank sum test. (**E**) As in (**C**), but SVM was trained using responses to single odours, and tested on mixture responses. (**F**) As in (**D**) but for the results in (**E**). n = 13 fields of view, 6 mice for behaving, 17 fields of view, 6 mice for naïve, engaged, and 14 sessions, 4 mice for naïve, disengaged mice. See **Figure 6 – figure supplement 1** for linearity of summation.

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Figure 7: *Simulation indicates data from behaving mice is less susceptible to normalizing sublinearity*
(**A**) Simulation approach. For each field of view, component responses were averaged across trials and
summed to obtain simulated mixture responses. Subsequently, uncorrelated Gaussian noise was added,
and the summed responses were further transformed using a normalisation model (Mathis et al., 2016;
Penker et al., 2020). These were then used as test inputs on SVMs that had been trained with single odour
responses. (**B**) Parameter extraction. Scatter plot shows observed EB + MB mixture responses against linear
sums of component responses (black dots). Red line shows the model fit used in the simulation. Data was
from behaving mice, at 2 seconds after the odour onset. (**C**) Comparison of SVM performance using data
from anaesthetised mice. Black line corresponds to linear sum + noise as inputs; Green line corresponds to
the results with additional normalising sublinearity. Mean ± SEM shown. (**D**) Same as in (**C**) but for data
from awake, behaving mice. (**E**) Comparison of SVM performance. SVM accuracy for simulated mixture
responses with sublinearity expressed as a fraction of accuracy obtained with simulated linear sum. p =
0.03 for early phase (0 - 1.5 s) and $8.13*10^{-4}$ for late phase (1.5 - 3 s after odour onset); paired t-test. (**F**)
Schematic of the finding: possible mechanisms of mixture representation in behaving mice may evolve over
time. In awake, behaving mice, responses evoked in the OB output neurons are dampened early on,
remaining largely in the linear range of the normalization curve. Over time, the responses become larger,
making sublinearity more widespread, but a pattern decorrelation may make evoked mixture responses
still discriminable.
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Supplementary figures

Figure 4 – figure supplement 1: *Subcellular dependence on mixture summation properties*

(**A**) Two-photon imaging of the OB output at three depths in Tbet-Cre::Ai95D mice performing the mixture task. Top: Imaging from the apical dendrites at the glomerular level. Middle: Imaging from the TC somata at the external plexiform layer. Bottom: Imaging from MC somata at the mitral cell layer. Scale bar = 50 μm. (**B**) Evoked responses in the apical dendrites (top), TC somata (middle), and MC somata (bottom) by ethyl butyrate on its own (left column), methyl butyrate on its own (middle column) and by the mixture of ethyl butyrate and methyl butyrate (right). (**C**) Median fractional deviation for all fields of view (n = 7 fields of view, 7 mice for apical dendrites; 13 fields of view, 6 mice for TC somata; 5 fields of view, 3 mice for MC somata). Thick lines correspond to the median, and shaded areas show the 25th - 75th percentile range. (**D**) Boxplot summary of median fractional deviations for the imaged structures (not significant). (**E**) Average fluorescence change for all apical dendrites at the glomeruli (black trace), TC somata (green), and MC somata (red). Mean ± s.e.m. shown. (**F**) Pearson's correlation coefficient between EB responses and MB responses from each field of view, averaged. Mean ± SEM shown.

Figure 5 – figure supplement 1: *Discriminability of MB responses*
(**A**) Correlation matrices were obtained in the same way as in Figure 5A, except that the trials are sorted
by whether the stimuli contained methyl butyrate. (**B**) Discriminability index over time, where within-odour
comparisons are between trials where methyl butyrate was presented.

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Figure 6 – **figure supplement 1**: *Further characterization of awake behavioural states.*
(**A**) Lick raster plots for example mice; left, trained mice performing the mixture task; middle, naïve
engaged mice; right, naïve disengaged mice. Vertical grey bars indicate the time of odour presentation (0.5
s). Trials with ethyl butyrate presentations are ordered below the black horizontal bar. (**B**) Average number
of anticipatory licks generated by mice in each group, on trials with EB vs. trials without EB. Each data point
corresponds to each imaging session. (**C**) Time course of linearity in summation; median fractional
deviation from each field of view is summarized here with median (thick line), and 25th and 75th percentile
range (shaded areas).
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Figure 6 – figure supplement 2: *Relationship between discriminability and linearity of mixture*
summation.
Comparison of discriminability against the linearity of summation for all behavioural states. The average
ability (fraction of trials correct) of SVMs (trained on single odour responses) to discriminate EB mixtures
against non-EB mixtures is shown against the median fractional deviation from linear sum at the
corresponding time. The time elapsed from the odour onset is colour coded.
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Figure 6 – figure supplement 2: *Mixture representation in TCs is not affected by ipsilateral piriform*
inactivation.

(**A**) To assess if the learning-dependent improvement in the decoder performance requires an involvement
of the piriform cortex, we infused muscimol (500 nl, 2 mM) in the ipsilateral, anterior piriform cortex during
imaging, while trained mice performed the mixture task. A cannula was implanted at an angle, in a manner
similar to (Otazu et al., 2015) in order to infuse muscimol. (**B**) When the mice were trained to perform the
task well, the imaging sessions commenced. Muscimol and control sessions alternated, where co-ordinates
for the imaged fields of view were recorded and could be revisited, so that imaging could be obtained from
the same location under the control and muscimol conditions. (**C**) Example matched fields of view for
control session (left) and muscimol session (right). Arrow heads point to the same tufted cells imaged on
different days. (**D**) Example confocal image of a coronal section of the brain, showing a track left by an
implanted cannula (white dotted line) in the left piriform cortex. Blue = DAPI fluorescence; red =
fluorescence from DiI, which was injected before perfusion. (**E**) Behavioural performance for control and
muscimol sessions. Sessions with the common fields of view are paired. P = 0.069, paired t-test (n = 6
sessions, 3 mice). (**F**) SVM performance in the ability to discriminate S+ odours (EB present) vs. S- odours
(EB absent), for control (black) and muscimol (green) datasets. Mean ± s.e.m. shown. (**G**) Same as (**F**),
except that the training set comprised responses from single odour trials, and the test set comprised
mixture responses.
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References

Abraham, N.M., Egger, V., Shimshek, D.R., Renden, R., Fukunaga, I., Sprengel, R., Seeburg, P.H., Klugmann, M., Margrie, T.W., Schaefer, A.T.*, et al.* (2010). Synaptic Inhibition in the Olfactory Bulb Accelerates Odor Discrimination in Mice. Neuron *65*, 399-411.

Ackels, T., Erskine, A., Dasgupta, D., Marin, A.C., Warner, T.P.A., Tootoonian, S., Fukunaga, I., Harris, J.J., and Schaefer, A.T. (2021). Fast odour dynamics are encoded in the olfactory system and guide behaviour. Nature *593*, 558-563.

Agmon-Snir, H., Carr, C.E., and Rinzel, J. (1998). The role of dendrites in auditory coincidence detection. Nature *393*, 268-272.

Araneda, R.C., Kini, A.D., and Firestein, S. (2000). The molecular receptive range of an odorant receptor. Nature Neuroscience *3*, 1248-1255.

Bell, G.A., Laing, D.G., and Panhuber, H. (1987). Odour mixture suppression: evidence for a peripheral mechanism in human and rat. Brain Research *426*, 8-18.

Bhattacharjee, A.S., Konakamchi, S., Turaev, D., Vincis, R., Nunes, D., Dingankar, A.A., Spors, H., Carleton, A., Kuner, T., and Abraham, N.M. (2019). Similarity and Strength of Glomerular Odor Representations Define a Neural Metric of Sniff-Invariant Discrimination Time. Cell Reports *28*, 2966-2978.e2965.

Bregman, A.S. (1990). Auditory Scene Analysis: The Perceptual Organization of Sound (The MIT Press).

Bridgeford, E.W., Wang, S., Wang, Z., Xu, T., Craddock, C., Dey, J., Kiar, G., Gray-Roncal, W., Colantuoni, C., Douville, C.*, et al.* (2021). Eliminating accidental deviations to minimize generalization error and maximize replicability: Applications in connectomics and genomics. PLOS Computational Biology *17*, e1009279.

Buck, L., and Axel, R. (1991). A novel multigene family may encode odorant receptors: A molecular basis for odor recognition. Cell *65*, 175-187.

Carey, R.M., and Wachowiak, M. (2011). Effect of Sniffing on the Temporal Structure of Mitral/Tufted Cell Output from the Olfactory Bulb. The Journal of Neuroscience *31*, 10615.

Chae, H., Banerjee, A., and Albeanu, D.F. (2021). A non-canonical feedforward pathway for computing odor identity. bioRxiv, 2020.2009.2028.317248.

Chapuis, J., and Wilson, D.A. (2012). Bidirectional plasticity of cortical pattern recognition and behavioral sensory acuity. Nature Neuroscience *15*, 155-161.

Cruz, G., and Lowe, G. (2013). Neural coding of binary mixtures in a structurally related odorant pair. Scientific Reports *3*, 1220.

Dana, H., Sun, Y., Mohar, B., Hulse, B.K., Kerlin, A.M., Hasseman, J.P., Tsegaye, G., Tsang, A.,
Wong, A., Patel, R., *et al.* (2019). High-performance calcium sensors for imaging activity in
neuronal populations and microcompartments. Nature Methods *16*, 649-657.

del Mármol, J., Yedlin, M.A., and Ruta, V. (2021). The structural basis of odorant recognition
in insect olfactory receptors. Nature *597*, 126-131.

Economo, M.N., Hansen, K.R., and Wachowiak, M. (2016). Control of Mitral/Tufted Cell
Output by Selective Inhibition among Olfactory Bulb Glomeruli. Neuron *91*, 397-411.

Firestein, S., Picco, C., and Menini, A. (1993). The relation between stimulus and response in
olfactory receptor cells of the tiger salamander. The Journal of Physiology *468*, 1-10.

Fletcher, M.L. (2011). Analytical Processing of Binary Mixture Information by Olfactory Bulb
Glomeruli. PLOS ONE *6*, e29360.

Friedrich, R.W., and Wiechert, M.T. (2014). Neuronal circuits and computations: Pattern
decorrelation in the olfactory bulb. FEBS Letters *588*, 2504-2513.

Fukunaga, I., Berning, M., Kollo, M., Schmaltz, A., and Schaefer, Andreas T. (2012). Two
Distinct Channels of Olfactory Bulb Output. Neuron *75*, 320-329.

Grabska-Barwińska, A., Barthelmé, S., Beck, J., Mainen, Z.F., Pouget, A., and Latham, P.E.
(2017). A probabilistic approach to demixing odors. Nature Neuroscience *20*, 98-106.

Gschwend, O., Abraham, N.M., Lagier, S., Begnaud, F., Rodriguez, I., and Carleton, A. (2015).
Neuronal pattern separation in the olfactory bulb improves odor discrimination learning.
Nature Neuroscience *18*, 1474-1482.

Gupta, P., Albeanu, D.F., and Bhalla, U.S. (2015). Olfactory bulb coding of odors, mixtures and
sniffs is a linear sum of odor time profiles. Nature Neuroscience *18*, 272-281.

Haddad, R., Lanjuin, A., Madisen, L., Zeng, H., Murthy, V.N., and Uchida, N. (2013). Olfactory
cortical neurons read out a relative time code in the olfactory bulb. Nature Neuroscience *16*,
949.

Hopfield, J.J. (1991). Olfactory computation and object perception. Proceedings of the
National Academy of Sciences *88*, 6462.

Ilc, T., Werck-Reichhart, D., and Navrot, N. (2016). Meta-Analysis of the Core Aroma
Components of Grape and Wine Aroma. Frontiers in Plant Science *7*, 1472.

Inagaki, S., Iwata, R., Iwamoto, M., and Imai, T. (2020). Widespread Inhibition, Antagonism,
and Synergy in Mouse Olfactory Sensory Neurons In Vivo. Cell Reports *31*, 107814.

Jacob, V., Le Cam, J., Ego-Stengel, V., and Shulz, D.E. (2008). Emergent Properties of Tactile
Scenes Selectively Activate Barrel Cortex Neurons. Neuron *60*, 1112-1125.
805
806 Jinks, A., and Laing, D.G. (1999). A Limit in the Processing of Components in Odour Mixtures.
807 Perception *28*, 395-404.
808
809 Jordan, R., Fukunaga, I., Kollo, M., and Schaefer, A.T. (2018). Active Sampling State
810 Dynamically Enhances Olfactory Bulb Odor Representation. Neuron *98*, 1214-1228.e1215.
811
812 Kay, L.M., Lowry, C.A., and Jacobs, H.A. (2003). Receptor contributions to configural and
813 elemental odor mixture perception. Behavioral Neuroscience *117*, 1108-1114.
814
815 Koldaeva, A., Schaefer, A.T., and Fukunaga, I. (2019). Rapid task-dependent tuning of the
816 mouse olfactory bulb. eLife *8*, e43558.
817
818 Kurahashi, T., Lowe, G., and Gold, G.H. (1994). Suppression of odorant responses by odorants
819 in olfactory receptor cells. Science *265*, 118.
820
821 Laing, D.G., and Francis, G.W. (1989). The capacity of humans to identify odors in mixtures.
822 Physiology & Behavior *46*, 809-814.
823
824 Laing, D.G., and Glemarec, A. (1992). Selective attention and the perceptual analysis of odor
825 mixtures. Physiology & Behavior *52*, 1047-1053.
826
827 Laughlin, S.B. (1989). The role of sensory adaptation in the retina. Journal of Experimental
828 Biology *146*, 39-62.
829
830 Madisen, L., Garner, Aleena R., Shimaoka, D., Chuong, Amy S., Klapoetke, Nathan C., Li, L.,
831 van der Bourg, A., Niino, Y., Egolf, L., Monetti, C.*, et al.* (2015). Transgenic Mice for
832 Intersectional Targeting of Neural Sensors and Effectors with High Specificity and
833 Performance. Neuron *85*, 942-958.
834
835 Malnic, B., Hirono, J., Sato, T., and Buck, L.B. (1999). Combinatorial Receptor Codes for Odors.
836 Cell *96*, 713-723.
837
838 Marr, D. (1982). Vision: A Computational Investigation into the Human Representation and
839 Processing of Visual Information (W.H.Freeman and Company).
840
841 Mathis, A., Rokni, D., Kapoor, V., Bethge, M., and Murthy, V.N. (2016). Reading Out Olfactory
842 Receptors: Feedforward Circuits Detect Odors in Mixtures without Demixing. Neuron *91*,
843 1110-1123.
844
845 Oka, Y., Omura M Fau - Kataoka, H., Kataoka H Fau - Touhara, K., and Touhara, K. Olfactory
846 receptor antagonism between odorants.
847
848 Otazu, Gonzalo H., Chae, H., Davis, Martin B., and Albeanu, Dinu F. (2015). Cortical Feedback
849 Decorrelates Olfactory Bulb Output in Awake Mice. Neuron *86*, 1461-1477.

851 Padmanabhan, K., and Urban, N.N. (2010). Intrinsic biophysical diversity decorrelates
852 neuronal firing while increasing information content. Nature Neuroscience *13*, 1276-1282.
853

854 Penker, S., Licht, T., Hofer, K.T., and Rokni, D. (2020). Mixture Coding and Segmentation in the
855 Anterior Piriform Cortex. Frontiers in Systems Neuroscience *14*, 89.
856

857 Poupon, D., Fernandez, P., Boisvert, S.A., Migneault-Bouchard, C., and Frasnelli, J. (2018). Can
858 the identification of odorants within a mixture be trained? Chemical Senses *43*, 721-726.
859

860 Reddy, G., Zak, J.D., Vergassola, M., and Murthy, V.N. (2018). Antagonism in olfactory
861 receptor neurons and its implications for the perception of odor mixtures. eLife *7*, e34958.
862

863 Rinberg, D., Koulakov, A., and Gelperin, A. (2006). Speed-Accuracy Tradeoff in Olfaction.
864 Neuron *51*, 351-358.
865

866 Rokni, D., Hemmelder, V., Kapoor, V., and Murthy, V.N. (2014). An olfactory cocktail party:
867 figure-ground segregation of odorants in rodents. Nature Neuroscience *17*, 1225-1232.
868

869 Rokni, D., and Murthy, V.N. (2014). Analysis and Synthesis in Olfaction. ACS Chemical
870 Neuroscience *5*, 870-872.
871

872 Singh, V., Murphy, N.R., Balasubramanian, V., and Mainland, J.D. (2019). Competitive binding
873 predicts nonlinear responses of olfactory receptors to complex mixtures. Proceedings of the
874 National Academy of Sciences *116*, 9598.
875

876 Stettler, D.D., and Axel, R. (2009). Representations of Odor in the Piriform Cortex. Neuron *63*,
877 854-864.
878

879 Tabor, R., Yaksi, E., Weislogel, J.-M., and Friedrich, R.W. (2004). Processing of Odor Mixtures
880 in the Zebrafish Olfactory Bulb. The Journal of Neuroscience *24*, 6611.
881

882 Verhagen, J.V., Wesson, D.W., Netoff, T.I., White, J.A., and Wachowiak, M. (2007). Sniffing
883 controls an adaptive filter of sensory input to the olfactory bulb. Nature Neuroscience *10*,
884 631-639.
885

886 Wachowiak, M., and Cohen, L.B. (2001). Representation of Odorants by Receptor Neuron
887 Input to the Mouse Olfactory Bulb. Neuron *32*, 723-735.
888

889 Weiss, T., Snitz, K., Yablonka, A., Khan, R.M., Gafsou, D., Schneidman, E., and Sobel, N. (2012).
890 Perceptual convergence of multi-component mixtures in olfaction implies an olfactory white.
891 Proceedings of the National Academy of Sciences *109*, 19959.
892

893 Wilson, C.D., Serrano, G.O., Koulakov, A.A., and Rinberg, D. (2017). A primacy code for odor
894 identity. Nature Communications *8*, 1477.
895

896   Wilson, D.A., and Stevenson, R.J. (2003). The fundamental role of memory in olfactory
897   perception. Trends in Neurosciences *26*, 243-247.
898
899   Xu, L., Li, W., Voleti, V., Zou, D.-J., Hillman, E.M.C., and Firestein, S. (2020). Widespread
900   receptor-driven modulation in peripheral olfactory coding. Science *368*, eaaz5390.
901
902   Zak, J.D., Reddy, G., Vergassola, M., and Murthy, V.N. (2020). Antagonistic odor interactions
903   in olfactory sensory neurons are widespread in freely breathing mice. Nature
904   Communications *11*, 3350.
905
906