On the hybrid origin of the C$_2$ *Salsola divaricata* agg. (Amaranthaceae) from C$_3$ and C$_4$ parental lineages

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Abstract

- C_2 photosynthesis is characterized by recapturing photorespiratory CO_2 by RuBisCO in Kranz-like cells and is therefore physiologically intermediate between C_3 and C_4 photosynthesis. C_2 can be interpreted as an evolutionary precursor of C_4 and/or as the result of hybridization between a C_3 and C_4 lineage.

- We compared the expression of photosynthetic traits among populations of the *Salsola divaricata* agg. (C_2) from humid subtropical to arid habitats on the coasts of the Canary Islands and Morocco and subjected them to salt and drought treatments. We screened for enhanced C_4-like expression of traits related to habitat or treatment. We estimated species trees with a transcriptome dataset of Salsoleae and explored patterns of gene tree discordance. With phylogenetic networks and hybridization analyses we tested for hybrid origin of the *Salsola divaricata* agg.

- We observed distinct independent variation of photosynthetic traits within and among populations and no clear evidence for selection towards C_4-like trait expression in more stressful habitats or treatments. We found reticulation and gene tree incongruence in Salsoleae supporting a putative hybrid origin of the *Salsola divaricata* agg.

- C_2 photosynthesis in the *Salsola divaricata* agg. combines traits inherited from its C_3 and C_4 parental lineages and seems evolutionarily stable, possibly well adapted to a wide climatic amplitude.

Keywords: C_2 photosynthesis, carbon isotope values, CO_2 compensation point, Chenopodiaceae, phosphoenolpyruvate carboxylase, *Salsola divaricata* agg., phylogenomics, hybridization events.

Introduction

In current models of C_4 evolution, the C_3-C_4 intermediate phenotypes (including C_2 plants) are interpreted as a transitional evolutionary link between the ancestral C_3 photosynthesis pathway and the derived C_4 pathway and showcase the complexity of the numerous structural, genetic, and functional changes necessary to establish a functioning C_4 pathway (Bräutigam & Gowik, 2016; Schlüter & Weber, 2016). There are about 50 known species with intermediate C_3-C_4 traits, some of which do not share a common ancestor with one of the more than 60 independent C_4 lineages, and some of which represent lineages up to 20 or even 30 million years in their crown age (Sage *et al*., 2018). The fact that we observe these intermediate phenotypes in nature repeatedly implies that they can be evolutionary stable (Lundgren, 2020). Their phenotypic diversity has been classified into four photosynthetic categories of C_3-C_4 intermediacy: proto-Kranz, C_2 type I, C_2 type II and C_4-like (Sage *et al*., 2014). C_2 photosynthesis has been repeatedly interpreted as a crucial stepping stone towards C_4 photosynthesis (Edwards, 2019). In C_2 species the photorespiratory enzyme glycine decarboxylase (GDC) is restricted to the bundle sheath or Kranz-like cells and mitochondria are absent or distinctly reduced in the mesophyll cells. This induces a photorespiratory glycine shuttle to the bundle sheath cells where the glycine is then processed by GDC and the photorespiratory CO_2 is recaptured by RuBisCO (Schulze *et al*., 2016). Bräutigam & Gowik (2016) proposed that once the photorespiratory CO_2 pump is active, establishing the C_4 cycle is inevitable. According to this model, a strong selective pressure towards an increase of phosphoenolpyruvate carboxylase (PEPC) activity in C_2 species should be expected under carbon deficient conditions (Bräutigam & Gowik, 2016).

A possible alternative, but not mutually exclusive hypothesis, is the origin of C_3-C_4 intermediates through hybridization (Monson *et al*., 1984). Experimental hybrids of C_3 and C_4 species reveal similar phenotypes as naturally occurring C_3-C_4 intermediate species and a segregating of photosynthetic traits in F2+ generations (Björkman *et al*., 1969; Holaday *et al*., 1985; Cameron *et al*., 1989; Brown *et al*., 1993; Brown & Bouton, 1993; Oakley *et al*., 2014). Furthermore, conflicting topologies in molecular phylogenetic studies, that includes C_3-C_4 intermediate species, indicate possible hybridization events in the evolutionary history of these lineages (reviewed in Kadereit *et al*., 2017). Hybridization at some point in the history of a C_4 lineage might have perturbed the sequence of events depending on the photosynthetic phenotypes of the parental lineages. To investigate the possibility of a hybrid origin of a C_3-C_4 intermediate lineage, detailed phylogenomic studies are needed. For example, evidence of

recent and ancient events of reticulate evolution in a phylogenomic study of *Flaveria* revealed

a potential major role of hybridization in the evolution of C_4 photosynthesis in that genus

(Morales-Briones and Kadereit, 2022).

Here we conducted a study of a C_2 species aggregate at the population level and

looked at C_4-adaptive traits to search for empirical evidence for the expected evolutionary

shifts towards more C_4-like traits under stressful conditions according to the model proposed

by Bräutigam & Gowik (2016). At the same time, we investigated the origin of this aggregate

using a phylogenomic approach. The C_2 species we chose for our study are *Salsola divaricata*

Moq. (Amaranthaceae, subfamily Salicornioideae, tribe Salsoleae; Schüssler *et al*., 2017;

Morales-Briones *et al*., 2021) and its close relatives *S. verticillata* Schousb., *S.*

gymnomaschala Maire and *S. deschaseauxiana* Litard. & Maire (hereafter called the *S.*

divaricata agg.). The distribution area of the *Salsola divaricata* agg. on the Canary Islands

and the coasts of Western Morocco, spans a considerable climatic gradient with an arid

climate in the east and a mesic mediterranean climate in the west (García-Herrera *et al*.,

2001). All four species of the *Salsola divaricata* agg. are salt tolerant, perennial shrubs with

terete succulent leaves which are morphologically difficult to distinguish (Padrón Mederos,

2012) and seem mainly geographically defined. They all have a Kranz-like salsoloid leaf

anatomy (Voznesenskaya *et al*., 2013; Schüssler *et al*., 2017). *Salsola divaricata* was

categorized as a C_2 type I which is characterized by the photorespiratory CO_2 pump through

GDC being confined to the bundle sheath cells while having C_3-like activity of C_4 enzymes

(e.g., PEPC and NADP-ME). Their CO_2 compensation points around 30 μmol mol^{-1} are

intermediate to C_3 and C_4 species (Schüssler et al., 2017). The *Salsola divaricata* agg.

belongs to Salsoleae, a tribe rich in C_4 species, but also with several C_3 species and several

C_3-C_4 intermediate species. The *Salsola divaricata* agg. forms a monophyletic, well-

supported subclade nested in a clade of C_3-C_4 intermediate and C_3 species with uncertain

placement (Schüssler *et al*., 2017).

The two aims of this study were, to test whether the lineage is stable in its C_2 state or

is shifting towards stronger C_4 physiology under more stressful climatic conditions on the

eastern Canary Island and Morocco, and to test for a hybrid origin of the *Salsola divaricata*

agg. To achieve these aims we studied C_4 adaptive traits at the population level in the 26

populations of the *S. divaricata* agg. distributed from La Gomera in the west to Morocco in

the east and searched for more C_4-like phenotypes in the more arid eastern parts of the

distribution area. We also analyzed 991 loci from a transcriptome data set comprising C_3, C_4

and C_2 species of the Salsoleae and explored incongruences to determine if a hybridization

event is plausible in the lineage of the *S. divaricata* agg.

Fig. 1: Map of the collection locations of seeds and vouchers for the *S. divaricata* agg. (See also **Supporting**
Information Table S1 for voucher information). In total we analyzed 26 populations of the *S. divaricata* agg.
on the Canary Islands and Morocco. Species name abbreviation and population number: Sd = *Salsola divaricata*,
Sg = *S. gymnomaschala*, Sv = *S. verticillata*, Sx = *S. deschaseauxiana;* P*xxx* = population number. Note:
Morphological studies on the species complex (Brullo, 1982; Fennane and Ibn Tattou, 1998) consider *S.*
verticillata and *S. deschaseauxiana* as conspecific, while *S. gymnomaschala* seems more like *S. divaricata* s.s.

Materials and Methods

Plant Material

Seeds and vouchers of 26 populations of the *S. divaricata* agg. were collected in 2013 and

2014 in the Canary Islands and Morocco (**Supporting Information Table S1**; **Fig. 1**) and

grown in a greenhouse at the Botanical Garden Mainz with multiple individuals per

population. Most individuals of a population share a mother plant, when this is not the case

different mother plants are indicated with roman numbers in the individual names (LS No.).

Plants were grown in custom mixed soil in standardized clay pots. Day/night cycles of

14h/10h were artificially maintained with natural light and supplementary light providing a

light intensity of 200–400 µmol m^{-2} s^{-1}. The minimum temperature at night was 18 ºC and

daytime temperatures ranged between 25 to 35 °C in summer and 20 to 25 °C in winter. All

plants were watered once a week in the winter and twice a week in the summer. The

individuals included in the salt treatment were watered once a week and every two weeks

with 2% saltwater (20 g NaCl per liter; 34.3 mM) from June 2015 to April 2021. Individuals

in the dry treatment were only watered every two weeks from June 2015 to March 2017. The

analyses of the photosynthesis traits began in 2016. **Supporting Information Table S1** lists

the populations included in this study as well as the collection and voucher information.

Carbon isotope measurements

In C_3 plants Rubisco discriminates against assimilating atmospheric $^{13}CO_2$. C_4 plants exhibit

much lower discrimination against $^{13}CO_2$ through its conversion to bicarbonate and

assimilation by PEPC. Thus, calculation of $\delta^{13}C$ values can be used to detect C_4 cycle activity

where more negative values indicate higher discrimination against fixing $^{13}CO_2$ (values for C_3

plants are -22 to -32 ‰ and C_4 plants -9 to -16 ‰, Sage 2016). We measured samples of

dried leaves collected in the field from the same mother plants from which seeds were

collected. We compared their $\delta^{13}C$ values with samples taken from the greenhouse cultivated

offspring. We also determined and compared $\delta^{13}C$ values between and within greenhouse

cultivated populations of *S. divaricata*. To determine carbon isotope composition, a standard

procedure with PDB (Pee Dee Belemnite) limestone as the carbon isotope standard was used

(Bender *et al.*, 1973). Leaf samples were dried in silica gel, and then 1–2 mg was placed in a

tin capsule and combusted in an EuroVector elemental analyser (EuroVector, Italy). After

separating the resulting N_2 and CO_2 gasses by gas chromatography, they were fed into the

IsoPrime^{TM} isotope ratio mass spectrometer (IRMS; GV Instruments Ltd. (Micromass Ltd.,

United Kingdom) for determination of $^{13}C/^{12}C$ ratios (R). $\delta^{13}C$ values were determined

according to $\delta^{13}C(‰)=1000(R_{sample}/R_{standard})^{-1}$, where R is $^{13}C/^{12}C$. Measurements were taken

at Washington State University and at the Geology Department of the University of Mainz.

CO_2 compensation point measurements

The CO_2 compensation point (Γ), the concentration where there is no net uptake of CO_2 in the

light, is a trait that is often used to identify C_4 species due to their relatively low values

(usually less than 4 μmol CO_2 mol^{-1}) compared to C_3 species. We measured individuals of

several populations per island of all species in the *S. divaricata* agg. (n = 53; 1 to 3 technical

replicates for each individual). The C_3 *S. webbii* and C_4 *S. oppositifolia* were measured for

comparison. CO_2 compensation points were determined using the portable gas exchange

measurement system GFS-3000 (Heinz Walz GmbH, Germany) with the standard measuring

head 3010-S equipped with a standard leaf area cuvette and a LED-Array (for details of the

cuvette settings see **Supporting Information Material & Methods S1**). As a general

calculation parameter in GFS-Win, we used leaf area which was measured using Easy Leaf

Area v1.02 or Image J 1.49d (Wayne Rasband, National Institutes of Health, USA) followed

by a calculation according to the approximately cylindrical leaf shape. To determine the CO_2

compensation point, net rates of CO_2 assimilation were plotted against intercellular CO_2

concentration.

PEPC activity measurements

Phosphoenolpyruvate carboxylase (PEPC) is a key enzyme in the C_4 pathway. It fixes

atmospheric CO_2 after its conversion to HCO_3^- by carbonic anhydrase into a four-carbon

compound which gave this photosynthesis pathway its name (Reyna-Llorens & Hibberd,

2017). C_4 plants show high levels of PEPC activity compared to C_3 plants and an increased

activity is seen as an important stage in evolving from an intermediate to a full C_4

photosynthesis pathway (Sage *et al.*, 2012). PEPC activity measurements were conducted

using the Tecan infinite M1000 (Tecan Trading AG, Männedorf, Switzerland) and a Greiner

UV-Star 96-well plate (Sigma-Aldrich Chemie GmbH, Munich, Germany). For details of

sample preparation see **Supporting Information Material & Methods S1**.

The assay of PEPC was initiated by adding leaf extract and 3.9 mM

phosphoenolpyruvate (PEP). The chlorophyll content of the leaf extract was measured

according to Wintermans and De Mots (1965) in 96% ethanol. PEPC activity (*a*) was

calculated based on the linear part of the progress curve where $a = SU[\mu\text{mol ml}^{-1} \text{min}^{-1}]V_{\text{assay}}[\text{ml}]/(1000 V_{\text{enzyme}}[\text{ml}]c_{Chla+b}[\text{mg ml}^{-1}])$.

SU is the substrate turnover rate, V_{assay} is the total assay volume, V_{enzyme} is the volume of the

leaf extract and c_{Chla+b} is the concentration of chlorophyll in the leaf extract. Statistical

analyses were conducted in R v3.1.2 (R Core Team, 2014).

Transcriptome processing and nuclear phylogenetic analyses

To evaluate the potential hybrid origin of the *S. divaricata* agg., we assembled a dataset

comprising 13 transcriptomes representing 12 C_3, C_4 and C_2 species of Salsoleae. The data set

included ten publicly available transcriptomes and three newly sequenced species on an

Illumina HiSeq 2500 platform at the University of Minnesota Genomics Center (paired-end

125 bp). For RNA extraction protocol and library preparation refer to Morales-Briones *et al.*

(2021). Additionally, we sampled three closely related Amaranthaceae species as outgroups.

Table 1 lists all samples with their photosynthesis type, ploidy if known and SRA accession

number.

Raw read processing, transcriptome assembly, low-quality and chimeric transcript

removal, transcript clustering into putative genes, translation, final coding sequences (CDS)

redundancy assessment, and homology and orthology inference were carried out following

Morales-Briones et al. (2021; for details see **Supporting Information Material & Methods**

S1). Briefly, an all-by-all BLASTN search was performed, and putative homologs groups

were clustered using MCL (van Dongen 2000). Homolog trees were built using RAxML

(Stamatakis 2014) and spurious tips were removed with TreeShrink (Mai and Mirarab 2018).

Monophyletic and paraphyletic tips that belonged to the same taxon were removed, leaving

one tip with the most aligned characters to obtain final homologs. Orthology inference was

carried out following the 'monophyletic outgroup' approach from Yang and Smith (2014),

keeping only ortholog groups with all 17 taxa present.

We applied concatenation and coalescent-based methods for phylogenetic

reconstruction. For the concatenation approach, we prepared a supermatrix by keeping only

ortholog alignments with at least 300 bp. We estimated a maximum likelihood (ML) tree with

RAxML v8.2.11 (Stamatakis, 2014) using a partition by gene scheme. Clade support was

assessed with 100 rapid bootstrap (BS) replicates. To estimate a species tree that is

statistically consistent with the multi-species coalescent (MSC), individual gene trees were

used to infer a species tree using ASTRAL-III v5.6.3 (Zhang *et al.*, 2018) using local

posterior probabilities (LPP; Sayyari & Mirarab, 2016) to assess clade support. To examine

nuclear gene tree discordance, we first calculated the Internode Certainty All score (ICA;

Salichos *et al.*, 2014). Also, we calculated the number of concordant and conflicting

bipartitions on each node of the species trees. We calculated both the ICA scores and conflict

analyses with Phyparts (Smith *et al.*, 2015). Additionally, to distinguish strong conflict from

weakly supported branches, we evaluated tree conflict and branch support with Quartet

Sampling (QS; Pease *et al.*, 2018) using 1,000 replicates (for details see **Supporting**

Information Material & Methods S1).

Assessment of hybridization

To detect possible hybridization, first we inferred species networks under a maximum

pseudo-likelihood (Yu & Nakhleh, 2015) approach using PhyloNet v3.6.9. (Than *et al.*,

2008). To estimate the best number of hybridizations and test whether the species network fits

our gene trees better than a bifurcating tree, we performed model selection using the bias-

corrected Akaike information criterion (Sugiura, 1978) and the Bayesian information criterion

(Schwarz, 1978). We also used HyDe (Blischak *et al.*, 2018) to estimate the amount of

admixture (γ) in putative hybrid lineages. We tested all triples combinations and significance

was assessed with a Bonferroni correction (for details see **Supporting Information Material**

& Methods S1).

Plastome assembly and phylogenetic analysis

To investigate phylogenetic signals from plastid sequences, *de novo* assemblies were carried

out with the Fast-Plast v1.2.6 pipeline (https://github.com/mrmckain/Fast-Plast) using the

filtered organelle reads obtained from the transcriptome raw read processing. A ML tree was

inferred with IQ-TREE v1.6.1 (Nguyen *et al.*, 2015) using the automated model selection

(Kalyaanamoorthy *et al.*, 2017) and 200 standard non-parametric BS replicates for branch

support (for details see **Supporting Information Material & Methods S1**). Additionally, we

used QS with 1,000 replicates, to detect potential plastome conflict in the backbone as seen in

other groups of Amaranthaceae s.l. (Morales-Briones *et al.*, 2021).

Analysis of photosynthetic gene trees

We inferred trees for 50 genes that are important in photosynthesis to determine if the genes in

Salsola divaricata agg. show a tendency to group with either a C_3 or a C_4 species. We

downloaded coding and protein sequences from *Arabidopsis thaliana* as baits from the TAIR

website (https://www.arabidopsis.org/tools/bulk/sequences/index.jsp). These loci were

determined based on Lauterbach *et al.* (2017a, b) and included two RuBisCO subunit genes,

four genes coding for proteins of the glyoxylate cycle, 17 genes encoding photorespiratory

proteins, and 25 genes coding for C_4-associated proteins (Lauterbach *et al.*, 2017b) as well as

two C_4-associated transcription factors (Lauterbach *et al.*, 2017a). Gene annotation and

pathway assignment can be found in **Supporting Information Table S3**. We examined

whether the *S. divaricata* agg. was sister to the C_3 species *S. montana* or the C_4 clade. When

there were several gene copies or weak bootstrap support, we additionally noted whether

species of the *S. divaricata* agg. were in a clade with C_3 or C_4 species (for details see

Supporting Information Material & Methods S1).

Results

Carbon isotope measurements

Salsola divaricata plants growing in the wild on the Canary Islands generally had higher carbon isotope (δ^{13}C) values (-21 to -28; mean -24.44, n = 30) than those grown from seeds under greenhouse conditions (-22 to -34; mean -27.77, n = 165). Overall, the δ^{13}C values of the *S. divaricata* agg. showed a distinct range but not a single individual (stressed or not stressed) came close to a C$_4$-like δ^{13}C value of > -16 (**Fig. 2**). The control and drought treatment showed no significant difference of δ^{13}C values among populations (Kruskal-Wallis chi-squared = 5.1081 and 8.61, df = 5, n = 165 and 36, p-value = 0.40 and 0.13, respectively). In the salt treatment there was a significant difference of δ^{13}C values among populations (Kruskal-Wallis chi-squared = 15.47, df = 5, n = 36, p-value = 0.009), which was due to significantly higher δ^{13}C values in salt treated individuals of populations 191 compared to 182 and 194. We tested for differences of carbon isotope values between treatments within populations of the *Salsola divaricata* agg. and found a significant difference for populations 182, 191 and 198 (**Fig. 3**). However, there was no significant difference among the three treatments for populations 184, 188 and 194 (**Fig. 3**). After grouping δ^{13}C values according to island, significant differences were found among islands (ANOVA: df = 6, F value = 4.08, p-value = 0.001) due to differences between La Gomera and the mainland (SW Morocco), as well as between Lanzarote, Tenerife, and the mainland, respectively. All other differences were not statistically significant. We found that the populations of *S. verticillata*, *S. deschaseauxiana* and *S. gymnomaschala* on the Moroccan mainland often had higher δ^{13}C values than the populations of *S. divaricata* on the Canary Islands (the non-stressed control groups that are regularly watered, **Supporting Information Fig. S1**). However, due to very different sample sizes further statistical tests including all populations were inconclusive.

Overall, these results showed variation in δ^{13}C values among populations associated with source population and growth conditions; but all were well within the range typical of C2 and C$_3$ species. The significantly higher δ^{13}C values of salt treated individuals indicated that salinity rather than drought affects the δ^{13}C in *Salsola divaricata* agg. and could be responsible for the occasional observation of relatively high δ^{13}C values of our samples collected in the wild close to the shoreline.

318 **Fig. 2:** Distribution of δ¹³C (‰) of 195 samples of the *Salsola divaricata* agg., of these 30 were collected from
319 plants growing in the wild and 165 from plants in cultivation at the Botanical Garden Mainz. This includes 36
320 individuals in dry treatment and 36 in salt treatment, all others are in the control group. See **Table S1** for
321 information on populations included.

Fig. 3: Comparison of carbon isotope values ($\delta^{13}C$ [‰]) between treatments within populations (P) of the *Salsola divaricata* agg. Significant differences were found for populations 182 from La Gomera (a; Kruskal-Wallis chi-squared = 7.94, df = 2, n = 18, p-value = 0.02), 191 from Fuerteventura (d; Kruskal-Wallis chi-squared = 10.84, df = 2, n = 18, p-value = 0.004), and 198 from Gran Canaria (f; Kruskal-Wallis chi-squared = 8.84, df = 2, n = 18, p-value = 0.01). No significant differences were found between the three treatments for 184 from Lanzarote (b; Kruskal-Wallis chi-squared = 2.33, df = 2, n = 18, p-value = 0.31), 188 from La Graciosa (c; Kruskal-Wallis chi-squared = 0.78, df = 2, n = 18, p-value = 0.68), and 194 from Fuerteventura (e; Kruskal-Wallis chi-squared = 2.27, df = 2, n = 18, p-value = 0.32).

CO$_2$ compensation point measurements

The CO_2 compensation points (Γ) of the four species of the *Salsola divaricata* agg. lained between those of the C$_3$ (*Salsola webbii*) and C$_4$ species (*Salsola oppositifolia*) as expected (between 20 and 30 µmol CO_2 mol^{-1}, **Fig. 4a**). The mean values of *S. deschaseauxiana* and *S. verticillata* were slightly lower than those of *S. divaricata* and *S. gymnomaschala*. When looking at individual measurements we again observed a broad range from 17 to 40 µmol CO_2 mol^{-1} with one outlier at 9 µmol CO_2 mol^{-1} (**Supporting Information Fig. S2**). Populations growing on the more arid Islands such as Lanzarote and Fuerteventura or in Morocco did not show lower CO_2 compensation points. Due to varying values within each

population (**Supporting Information Fig. S2**) and the relatively small sample sizes (one to

two individuals per population) we did not conduct further statistical analysis.

Fig. 4: a) CO_2 compensation points (Γ in $\mu mol\ CO_2\ mol^{-1}$) of the separate species of the C_2 *Salsola divaricata* agg. (Sd_C_2 = *Salsola divaricata*, mean 25.14 $\mu mol\ CO_2\ mol^{-1}$, n = 21; Sg_C_2 = *S. gymnomaschala*, mean 26.75 $\mu mol\ CO_2\ mol^{-1}$, n = 4; Sv_C_2 = *S. verticillata*, mean 22.21 $\mu mol\ CO_2\ mol^{-1}$, n = 4; Sx_C_2 = *S. deschaseauxiana*, mean 19.56 $\mu mol\ CO_2\ mol^{-1}$, n = 4) compared to the values of C_3 species *Salsola webbii* (Sw_C_3, 66.3 $\mu mol\ CO_2\ mol^{-1}$, n = 2) and C_4 species *Salsola oppositifolia* (So_C_4, 2.35 $\mu mol\ CO_2\ mol^{-1}$, n = 3). We measured 1–3 technical replicates per individual. **b)** PEPC activity [$\mu mol\ min^{-1}\ mg_{chlorophyll}^{-1}$] of the separate species of the C_2 *Salsola divaricata* agg. (*Salsola divaricata*, mean activity 33 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$, n = 82; *S. gymnomaschala*, mean 46.8 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$, n = 10; *S. verticillata*, mean 37.4 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$, n = 7; *S. deschaseauxiana*, mean 27.6 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$, n = 6) compared to C_3 *S. webbii* (mean 30 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$, n = 1) and C_4 *S. oppositifolia* (mean 216 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$, n = 2).

PEPC activity measurements

First, we compared the PEPC activity of the separate species of the *Salsola divaricata* agg. to

the values of the closely related C_3 species *Salsola webbii* and C_4 species *Salsola*

oppositifolia. C_3 and C_4 values were as expected clearly distinguishable (**Fig. 4b**). The values

of the C_2 species were closer to those of the C_3 species and much lower than the C_4 species

(**Fig. 4b**). We used a Kruskal-Wallis test to determine if statistically significant differences

exist between C_2 species (Kruskal-Wallis chi-squared = 9.94, df = 3, p-value = 0.019). This

significant difference was observed due to the values of *S. divaricata* on the Canary Islands

having a median of 30.05 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$ and *S. gymnomaschala* in Morocco having

a median of 48.32 $\mu mol\ min^{-1}\ mg_{Chlorophyll}^{-1}$. Due to the high level of variation within species

we took a closer look at 14 populations of *S. divaricata* (n = 5–7 per population) and

observed a significant difference between them (Kruskal-Wallis chi-squared = 34.64, df = 13,

p-value = 0.001, see also **Fig. 5a**). When grouped according to island, we found a significant

difference (Kruskal-Wallis chi-squared = 22.199, df = 5, p-value = 0.0005) which is due to

the difference in values between populations on Gran Canaria and Lanzarote, as well as

populations on La Graciosa and Fuerteventura and on La Graciosa and Gran Canaria (**Fig.**

5b). We did not find a significant difference between treatments in the greenhouse in *S.*

divaricata (Kruskal-Wallis chi-squared = 1.21, df = 2, p-value = 0.55). Again, we observe a

high variation of PEPC activity values within populations and islands.

Linear regression to test for correlation between PEPC activity and carbon isotope

($\delta^{13}C$) values (Multiple R² = 0.008, F-statistic = 0.35 , df = 44, p-value = 0.56), between

PEPC activity and CO_2 compensation point (Γ) (Multiple R² = 0.04, F-statistic = 1.19, df =

29, p-value = 0.28), and between $\delta^{13}C$ values and Γ (Multiple R² = 0.017, F-statistic = 0.46,

df = 27, p-value = 0.5) recovered no significant correlation among the three.

Fig. 5: Boxplots of PEPC activity [μmol min⁻¹ mg$_{chlorophyll}$⁻¹] of *Salsola divaricata* Moq. **a)** Values grouped by
population (P). **b)** Values grouped by island; F = Fuerteventura, GC = Gran Canaria, L = Lanzarote, LGo = La
Gomera, LGra = La Graciosa, T = Tenerife.

Phylogenomic analyses

The raw reads for the newly generated transcriptomes are available from the NCBI Sequence

Read Archive (**Table 1**). The 991 final orthologs had alignments of lengths between 315 and

6621 bp. The concatenated matrix comprised 1,427,449 aligned columns with overall matrix

occupancy of 86% (see **Supporting Information Table S2** per details).

The ASTRAL species tree and the concatenated RAxML tree (**Supporting**

Information Fig. S3) had similar topologies with most nodes receiving the maximum support

(BS = 100%; LPP = 1). The only difference between the two topologies is that *Anabasis*

articulata and *Salsola soda* were sisters with maximum support in the RAxML tree but

formed a grade with lower support (LPP = 0.74) in the ASTRAL tree. Concordance analyses

(**Supporting Information Fig. S3** and **S4**) showed high support for the monophyly of

Salsoleae and the *S. divaricata* agg., while signaling conflict for the remaining taxa. The

placement of *S. genistoides* had low support with only 261 (of 794) concordant gene trees

(ICA = -0.45) and a QS score (0.22/0/0.98) suggesting an alternative topology. Monophyly of

the *S. divaricata* agg. clade composed of *S. divaricata* + *S. verticillata* had strong support

with 808 (of 913) concordant gene trees (ICA = 0.71) and maximum QS support (1/–/1; i.e.,

all sampled quartets supported that branch). However, the two samples of *S. divaricata*

showed signals of alternative topologies with only 332 (of 900) gene trees supporting their

sister relationship (ICA = 0.56) and QS counter support (-0.25/0/1). The sister relationship of

S. divaricata agg. and the C4 remaining Salsoleae had low support with only 178 (of 705)

concordant gene trees (ICA = 0.29) and low QS support (0.085/0/1 [ASTRAL]; (0.11/0/1)

[RAxML]) suggesting an alternative topology. The placement of the C4 *Anabasis articulata*

and *S. soda,* which varied between RAxML and ASTRAL trees, also had low gene tree and

QS support with signals of alternative topologies. The clade composed of *S. oppositifolia*,

Halogeton, *Hammada*, and *Holoxylon* was supported only by 280 (of 692l ICA = 0.24), but

had maximum QS support, while within the clade all relationships had low gene tree and QS

support signaling alternative topologies.

The final plastid alignment had 80,903 characters with a matrix occupancy of 70%.

The plastid tree largely differed from the nuclear topologies and had most nodes with

maximum support (BS = 100; **Fig. 6** and **S5**). *Caroxylon vermiculatum, Kali collina,* and *S.*

genistoides form a well-supported clade (BS = 100; QS = 0.56/0/1) that is sister to all

Salsoleae. *S. webbii* and *Anabasis articulata* form a grade with strong support (BS =100; QS

= 0.6/0/0.99 [*S. webii*]; QS = 1/–/1 [*A. articulata*]). The remaining species form an

unsupported clade (BS = 0.47; QS = -0.0096/0.88/0.68) composed of two subclades. The first

subclade (BS = 93; QS = 0.19/0.91/0.78) is composed of *S. verticillata* + *S. montana* (BS =

100; QS = 0.61/0/1) and the two samples of *S. divaricata* (BS = 100; QS = 1/–/0.77). The

second subclade (BS = 100; QS = 0.59/0/0.95) has *Salsola soda* as the sister the clade (BS

=100; QS = 1/–/1) composed of *S. oppositifolia*, *Halogeton*, *Hammada*, and *Haloxylon.*

Fig. 6: a) Maximum likelihood phylogeny of Salsoloideae inferred from the RAxML analysis of the concatenated 991-nuclear gene supermatrix from the 'monophyletic outgroup' (MO) orthologs. Bootstrap support and local posterior probabilities (BS/LPP) are shown above branches. Nodes with full support (BS = 100/LPP = 1) are not shown. Em dashes (—) denote alternative topology compared to the ASTRAL tree (**Supporting Information Fig. S3**). Quartet Sampling (QS) scores are shown below branches. QS score: Quartet concordance (QC)/Quartet differential (QC)/Quartet informativeness QI). Full QS support (1/–/1) not shown. Pie charts represent the proportion of gene trees that support that clade (blue), the proportion that support the main alternative bifurcation (green), the proportion that support the remaining alternatives (red), and the proportion (conflict or support) that have < 50% bootstrap support (gray). Branch lengths are proportional to the number of substitutions per site (scale bar on the bottom). b) Cladogram of Salsoloideae inferred from IQ-TREE analysis of concatenated complete and partial plastomes. QS scores and BS support are shown above and below branches, respectively. See Supporting Information Fig. S5 for phylogram. c) Best maximum pseudo-likelihood species network inferred with PhyloNet. Red and blue curved branches indicate the minor and major edges, respectively of hybrid nodes. Numbers next to curved branches indicate inheritance probabilities for each hybrid node.

Assessment of hybridization

Model selection preferred a network as a better model than any bifurcating tree (**Table 2**). A

network with three reticulation events was the best model overall (PhyloNet-4-hybridization

search; AICc = 37875.93; BIC = 38054.302; **Fig. 6c**). The next model was a significantly

worse model (also a network with three reticulation events [PhyloNet-3-hybridization search]

with ΔAICc and ΔBIC of 753.208. The *S. divaricata* agg. clade was shown as the product of a

hybridization event between *S. soda* (C_4) (inheritance probability $p_i = 0.346$) and *S. montana*

(C_3) ($p_i = 0.654$). Then the C_4 clade *S. oppositifolia* + *Halogeton* + *Hammada* + *Haloxylon*

was the product of reticulation between *S. soda* (C_4) ($p_i = 0.846$) and the stem lineage of the

C2 *S. divaricata* agg. ($p_i = 0.154$). These two clades were also recovered as hybrids in the

networks with one, two and three reticulation events with similar parental lineages and

inheritance probabilities (**Supporting Information Fig S6**). The third event was a deeper

reticulation within Salsoleae which showed that most species of the group (excluding

Caroxylon) were a product of an ancient hybridization event between *S. genistoides* ($p_i =$

0.212) and the sister lineage of *S. genistoides* + *S. webbii* ($p_i = 0.788$). The HyDe analysis

resulted in 1,680 hybridization tests, of which 298 triples were significant (**Supporting**

Information Table S3, **Fig. S7**). The significant triples showed that 12 out of 13 individuals

of Salsoleae are involved in multiple hybridization events, which is mostly in agreement with

the nested reticulation events detected with Phylonet. All sampled accessions of the *S.*

divaricata aggr. showed similar hybridization patterns (**Fig. S7;** i.e. same potential parental

lineages and admixture parameter [γ]) consistent with a single ancient hybrid origin of *S.*

divaricata agg. as shown in the Phylonet analyses. Overall, the admixture parameter (γ)

ranged from 0.013 to 0.986 (average 0.458). These results indicate multiple, complex

hybridizations within Salsoleae. Additionally, Hyde also identified *Eokochia saxicola* as a

hybrid (**Supporting Information Table S3**, **Fig. S7**).

Analysis of individual genes involved in photosynthesis or photorespiration

The 50 genes ranged from 7 to 65 sequences due to variation in the number of accessions and

gene copies (**Supporting Information Table S4**). Summarizing all genes, *Salsola divaricata*

agg. appeared 22 times as sister to C_3 *S. montana* and 26 times as sister to C_4 species (**Fig 7;**

Supporting Information Table S4). *Salsola divaricata* agg. was not detected in three genes

(Isocitrate lyase, RuBisCO subunits). When multiple copies per gene were recovered for *S.*

divaricata agg, they were often non-monophyletic, but instead separated in clades with copies

from the C_3 and/or C_4 species. In these 23 gene trees, sequences of the *S. divaricata* agg.

appeared 21 times with C_3 and 26 times with C_4 species. Summarizing the results for genes

typical for C_4 pathways, the *S. divaricata* agg. was 27 times sister to and/or nested in a C_3

clade and 32 times sister to and/or nested in a C_4 clade). Taking into consideration bootstrap

support (BS $\geq$ 70), the numbers reduced to 18 times sister to and/or nested in a C_3 clade and

23 times sister to and/or nested in a C_4 clade for C_4 genes. For the two C_4-associated

transcription factors, *S. divaricata* agg. was sister to the C_4 clade for SHORTROOT with

strong support (BS = 86). The sister relationship to the C_4 clade was moderately supported for

BEL1-like homeodomain 7 (BS = 57 in clade with C_4). For the genes assigned to the

Glyoxylate cycle *S. divaricata* agg. had multiple copies that were grouped with either C_3 or

C_4 with high support (BS > 80). The gene for Isocitrate lyase was missing in the *S. divaricata*

agg. and *S. montana*. *Salsola divaricata* agg. is also missing in the alignments for the

RuBisCO subunits. For photorespiration-associated genes many species, including those of *S.*

divaricata agg. had multiple copies, even though during the transcriptome processing putative

genes were inferred to remove isoforms and assembly artifacts. Copies of *S. divaricata* agg.

were nested within C_3 clades with strong support (9 times, BS > 70). Affinity to the C_4 clade

is well supported in 13 of 19 cases, in which they appear as sister or nested within the clade.

Just like the *S. divaricata* agg., the C_4 clade is also not monophyletic in most cases. Clade

assignment for all 50 genes is listed in **Supporting Information Table S4** and the individual

trees are shown in the **Supporting Information Fig S8**.

Fig. 7. Summary of the phylogenetic affinity of the *Salsola divaricata* agg. in analyzed photosynthetic gene trees. Red and green colored boxes represent phylogenetic affinity with C_3 or C_4 species in the specified gene, respectively. Gray boxes denote no affinity. Gene with only grey boxes denote that the gene was not detected in *Salsola divaricata* agg. transcriptomes we sequenced. For detailed phylogenetic affinity see **Supporting Information Table S4 and Fig S8**.

Discussion

Hybridization experiments with parental lineages exhibiting different photosynthetic types have shown the transferability of C_4 traits (Apel et al., 1988). For a few examples, it has been suggested that this might also occur in nature (Dunning et al., 2017) which could mean an

immediate adaptive advantage of the hybrid lineage in more stressful environments. A

phylogenomic study of *Flaveria* revealed a major role of hybridization and transgressive

segregation in C_4 evolution (Morales-Briones and Kadereit, 2022). In this study we aimed to

infer the phylogenetic origin of C_3-C_4 intermediacy of the *Salsola divaricata* agg. and

possible phenotypic variation across a steep climate gradient. *Salsola divaricata* is currently

classified as a C_2 Type I according to Sage *et al.* (2014). According to the model of C_4

evolution an increase of PEPC activity should be expected under more stressful (more carbon

deficient) conditions in the more arid areas (Lanzarote, Fuerteventura, Morocco) with a

transition to the C_2 Type II or even C_4-like intermediate type, resulting in a reduction of

photorespiration (Sage *et al.*, 2014; Bräuigan & Gowik, 2026). Alternatively, C_2

photosynthesis in *Salsola divaricata* agg. may represent a stable condition across mesic and

arid growing conditions as presently observed in this study.

Carbon isotope measurements depict a stress reaction in the *S. divaricata* agg.; but

without occurrence of a more C_4-like photosynthesis in populations growing in drier

environments

Although samples collected in the wild had slightly higher $\delta^{13}C$ values than those sampled in

our greenhouse (stressed or control), there is still a clear difference to typical values of C_4

species. None of the samples showed a $\delta^{13}C$ value typical for a C_4-like or a C_4 plant, which

would be around -9 to -16 ‰ (Sage, 2016); instead, all samples had values falling within the

range typical for C_3 species, which would be around -22 to -32 ‰ (Sage, 2016). In C_3 plants

the $\delta^{13}C$ values of leaves depend on several factors including stomatal conductance and

capacity to capture CO_2 in photosynthesis, which is dependent on light intensity, growth

temperature, photochemistry, and capacity for carbon assimilation. The plants growing in the

greenhouse are under less stress for potential water loss than field conditions due to lower

light intensity, and temperature being more moderate than some field conditions. The results

suggest a greater stomatal limitation on photosynthesis in the field grown plants, which

results in a low intercellular CO_2 concentration. This will change the discrimination against

the heavier C isotope since RuBisCO will capture more $^{13}CO_2$ resulting in slightly higher

$\delta^{13}C$ values (von Caemmerer, 1992).

We observed variation in the $\delta^{13}C$ values. Populations on the Moroccan mainland

often have higher values than the populations on the Canary Islands; but there are many

exceptions in populations on the more arid Islands of Fuerteventura and La Graciosa as well

as on the more humid Islands of Gran Canaria, Tenerife and La Gomera. There does not seem

to be a clear pattern correlated with the general climatic conditions on the Islands. Therefore,

the observed variation is likely due to plasticity or local, microclimatic differences. In any

case, there are no indications of populations exhibiting C_4-like photosynthesis based on the

$\delta^{13}C$ values.

Salsola divaricata plants from all populations grown for several years under salt stress in the

greenhouse showed clear morphological differences in comparison to the control plants. They

had more succulent leaves, did not grow to the same height as their counterparts in the control

group and most individuals in the salt group flowered earlier. In three of the six tested

populations, La Gomera (182), Fuerteventura (191), and Gran Canaria (198), this long-term

salt stress was reflected in a significantly higher $\delta^{13}C$ value. The three other populations from

Lanzarote, La Graciosa and Fuerteventura had slightly, but not significantly higher, values for

the salt treatment compared to the control group. This suggests *Salsola divaricata* can adapt

to salt stress by increasing water storage and decreasing stomatal conductance and it explains

some of the $\delta^{13}C$ value variation found in the wild as salinity certainly varies among sites.

C_3-C_4 intermediate CO_2 compensation points for *S. divaricata* agg. supports a stable C_2-

state

The CO_2 compensation points of the *S. divaricata* agg. varied but they are well within the

range of other C_2 species (Sage *et al.*, 2014 and ref. therein) and they were significantly lower

than the values of C_3 species *S. webbii* and significantly higher than those of C_4 species *S.*

oppositifolia. There is no evidence for constantly lower Γ values in populations that

originated from more arid regions, although individuals of *S. verticillata* and *S.*

deschaseauxiana mostly showed somewhat lower values, many individuals from similar

climatic environments (e.g., *S. gymnomaschala* or individuals of *S. divaricata* from

Fuerteventura or Lanzarote) showed higher values which skews a possible relation of climate

and lower Γ values. Sage *et al.* (2014) suggested a Γ value below 10 µmol CO_2 mol^{-1}

characterizes the transition from C_2 Type I to C_2 Type II and represents an evolutionary

progression towards C_4. An accessory C_4 metabolic cycle is present in C_2 Type II plants

and was so far only observed in several species of *Flaveria and Mollugo verticillata*

(Edwards & Ku, 1987; Sage et al., 2018). None of the Γ measurements of 53 individuals of

the *Salsola divaricata* agg. showed an indication of an accessory C_4 metabolic cycle and a

significantly higher carboxylation by PEPC, therefore indicating a stable C_2-state for *S.*

divaricata agg.

So far, the analyzed traits seem to be independent between populations and islands

and show substantial variation. On Lanzarote for example, populations of *S. divaricata* have

comparably high Γ values and medium to low carbon isotope values compared to other

populations.

PEPC activity has little or no correlation with Γ values or carbon isotope values in *S.*

***divaricata* agg.**

The PEPC activity of the *S. divaricata* agg. was similar to the C_3 species *S. webbii*. Even

though there was a high standard deviation within C_4 *S. oppositifolia*, none of the C_2

individuals comes even close to a C_4-like value. The activity of PEPC for *S. verticillata* and *S.*

deschaseauxiana was slightly lower than those for *S. divaricata*; therefore, their slightly

lower Γ values and slightly higher carbon isotope values are not due to increased CO_2 fixation

by an optimized PEPC. Thus, S. *verticillata* and *S. deschaseauxiana* do not seem to be closer

to a more C_4-like state than *S. divaricata* or *S. gymnomaschala*.

The PEPC activity in the *S. divaricata* agg. is highly variable and showed only a weak

correlation with carbon isotope values and CO_2 compensation points, as discussed above for

S. verticillata. We would expect a higher PEPC activity in the salt treatment or the more arid

locations e.g., Lanzarote or Fuerteventura, however that was not the case. Although there

were statistically significant differences between islands, it is not possible to predict a value

for PEPC activity given the location since there is generally a large overlap of values. The

differences between populations could not be explained by the climatic context of the parent

plant in the wild and they were all treated the same in the greenhouse. We could only detect a

very weak correlation between PEPC activity, CO_2 compensation points and carbon isotope

values, supporting a rather independent distribution of the traits and their high phenotypic

variation.

The independent variation in three photosynthetic traits (carbon isotope value, CO_2

compensation point and PEPC activity) does not support a directed selection towards C_2 Type

II or C_4-like photosynthesis in any of the populations studied, not even those growing in

semi-desert conditions. In contrast, our trait observations are more in line with the results of

hybridization experiments. Summarizing the physiology of advanced generations of

interspecific C_3 and C_4 hybrids, Brown & Bouton (1993) stated that the 'correlation among

photosynthetic traits were low' which indicates 'a high degree of independence, both genetic

and physiological, among C_4 traits.' Like in our study, the hybridization experiment of C_3 and

C_4 *Atriplex* (Oakley *et al.*, 2014) showed a distinct range of Γ values (in nine F_2 individuals

from 25-45 µmol CO_2 mol^{-1}; and one C_4-like value). Brown & Bouton (1993) hypothesize

that some of the photosynthetic traits measured can be more C_4-like in a still functioning C_3

metabolism, such as Γ, while others just mirror the physiological C_3 default condition, such as

enzyme activity. This together with the phylogenomic results and photosynthetic gene tree

analyses (see below) supports the hypothesis of a hybrid origin of the *Salsola divaricata* agg.

and subsequent stabilizing selection forming an evolutionary stable C_2 lineage.

First phylogenomic evidence of a hybrid origin of the *S. divaricata* agg.

The Salsoleae species tree showed a high level of conflict including the *S. divaricata* agg,

despite high bootstrap support. The assessment of hybridization events using PhyloNet and

HyDe showed multiple hybridization events within Salsoleae in which the *Salsola divaricata*

agg. was likely involved. The incomplete sampling of Salsoleae does not allow us to identify

the exact parental lineages; but a hybrid origin of the *Salsola divaricata* agg. from C_3 and C_4

parental lineages was highly supported.

The network that was recovered most often showed an ancient hybridization event

involving an ancestor of C_4 *Salsola soda* and the C_3 proto-kranz *Salsola montana* giving rise

to the *Salsola divaricata* agg. clade. This is the first phylogenomic evidence that a C_3-C_4

intermediate species is the result of (ancient) hybridization between a C_4 and a C_3 lineage.

The second reticulation event later indicates a hybridization event involving the ancestor of

Salsola divaricata agg. and the C_4 *Salsola soda* giving rise to the C_4 clade formed by *Salsola*

oppositifolia, *Halogeton*, *Hammada*, and *Haloxylon*. This might indicate that this relative old

C_4 lineage (Salsoleae ~ 21–22 MYA; Morales-Briones et al., 2021), could have inherited C_4

or antecedent traits from a parental lineage giving them an adaptive advantage that might

have influenced their photosynthesis pathway, e.g., adaptations in photosynthesis or

photorespiration associated enzymes or Kranz-cells. This is in line with recent findings in the

genus *Flaveria* (Morales-Briones and Kadereit, 2022) where ancient hybridization was found

to be a step towards C_4 evolution and likely promoted the rapid acquisition of C_4 traits. We

also found hybridization events in the two clades in the other networks with similar parental

lineages, supporting the evidence that these lineages were involved in hybridization events.

This could indicate either an incomplete reproductive barrier in the early lineages and

recurrent backcrossing or incidences of horizontal gene transfer. We must keep in mind

though that our sampling is limited and that other lineages not sampled in the C_4 clade might

also play a role (see Schüssler *et al.*, 2017 for a plastid tree of the Salsoloideae).

Phylogenetic analysis of photosynthetic genes supports the hybrid origin hypothesis and

suggests candidate genes for further study

A closer look of 50 gene trees from genes or gene families involved in photosynthesis or

photorespiration revealed the same overall pattern, namely incongruence regarding the

position of the *Salsola divaricata* agg., which in most cases either groups with the C_3 *S.*

montana (also the case in the plastid dataset) or with the C_4 species. This pattern could be

explained by hybridization of parental C_3 and C_4 lineages giving rise to an allopolyploid *S.*

divaricata agg. with an intermediate expression of photosynthetic traits. The Kranz-like

salsoloid leaf type of the *S. divaricata* agg. (Schüssler *et al.*, 2017: Fig. 7), for example, might

occur as in the case of the gene SHORTROOT, a C_4 associated transcription factor which

influences the development of Kranz cells (Kelly *et al.*, 2017, Slewinski *et al.* 2014), with

expression of the parental C_4 copy. Studying transcriptome profiles of Salsoleae and

Camphorosmeae leaves, respectively, Lauterbach *et al.* (2017a) and Siadjeu *et al.* (2021)

revealed a high expression of SHORTROOT in C_2 species. In the case of PEPC, two copies

were recovered from our transcriptomes and for both the *S. divaricata* agg. group with the C_3

species supporting the C_3-like PEPC activity data found in this study.

Conclusions

The phylogenetic signal in our trees, gene trees and networks indicate hybridization events

gave rise to the *Salsola divaricata* agg. Furthermore, the distinct variation of three

photosynthetic traits within a C_3-C_4 intermediate range without indication of selection

towards a more C_4-like expression in this relatively old lineage supports the interpretation of

an evolutionarily stable C_2 lineage of hybrid origin. Overall, the photosynthetic traits seem

plastic and genetically fixed stress reactions are not prominent among the populations studied.

The *Salsola divaricata* agg. is well adapted to a broad range of climatic conditions including

mesic to arid coastal habitats and to salinity. Part of the success of the species aggregate

might be the photorespiratory pump acquired through hybridization with a C_4 lineage in the

past. However, a broader sampled phylogenomic study is needed to further narrow down the

parental lineages. We propose that reticulation events might play an important role in the multiple convergent acquisition of complex traits such as C_2 or C_4 photosynthesis as a possible fast track.

Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author Contributions

GK designed the study. DT and GE (carbon isotope values) measured the trait data and DT analyzed them. DT, DMB and YY conducted the phylogenomic analyses. DMB prepared the figures and DT the map. DT, GK and DMB wrote the first draft. All authors contributed to the discussion of the results and the final version of the manuscript.

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Data Availability Statement

Transcriptome data generated for this study can be found in the NCBI Sequence Read Archive (SRA) (see **Table 1** for SRA accession numbers).

References

Apel P, Bauwe H, Bassüner B, Maass I. 1988. Photosynthetic properties of *Flaveria cronquistii*, *F. palmeri*, and hybrids between them. Biochemie und Physiologie der Pflanzen **183**: 291–299.

Bender MM, Rouhani I, Vines HM, Black CC. 1973. 13C/12C Ratio Changes in Crassulacean Acid Metabolism Plants. *Plant Physiology* **52**: 427–430. doi:10.1104/pp.52.5.427

Björkman O, Gauhl E, Nobs MA. 1969. Comparative studies of *Atriplex* Species with and without beta-Carboxylation photosynthesis and their first-generation hybrid. *Yearbook 68 of the Carnegie Institution of Washington*, 620–633.

Blischak PD, Chifman J, Wolfe AD, Kubatko LS. 2018. HyDe: A Python Package for Genome-Scale Hybridization Detection (D Posada, Ed.). *Systematic Biology* **67**: 821–829. doi:10.1093/sysbio/syy023

Bräutigam A, Gowik U. 2016. Photorespiration connects C_3 and C_4 photosynthesis. *Journal of Experimental Botany* **67**: 2953–2962. doi:10.1093/jxb/erw056.

Brown R, Bouton JH. 1993. Physiology and Genetics of Interspecific Hybrids Between Photosynthetic Types. *Annual Review of Plant Physiology and Plant Molecular Biology* **44**: 435–456. doi:10.1146/annurev.pp.44.060193.002251

Brown RH, Byrd GT, Bouton JH, Bassett CL. 1993. Photosynthetic Characteristics of Segregates from Hybrids between *Flaveria brownii* (C_4 Like) and *Flaveria linearis* (C_3-C_4). *Plant Physiology* **101**: 825–831. doi: 10.1104/pp.101.3.825

Brullo AS. 1982. Notes on the genus *Salsola* (Chenopodiaceae). 1. The *Salsola oppositifolia* and *S*. *longifolia* groups. *Willdenowia* **12**: 241–247. http://www.jstor.org/stable/3995936

Cameron RG, Bassett CL, Bouton JH, Brown RH. 1989. Transfer of C_4 Photosynthetic Characters through Hybridization of *Flaveria* Species. *Plant Physiology* **90**: 1538–1545. doi: 10.1104/pp.90.4.1538

Chen R, Huangfu L, Lu Y, Fang H, Xu Y, Li P, Zhou Y, Xu C, Huang J, Yang Z. 2021. Adaptive innovation of green plants by horizontal gene transfer. *Biotechnology Advances* **46**: 107671. doi:10.1016/j.biotechadv.2020.107671.

Dunning LT, Lundgren, MR, Moreno-Villena JJ, Namaganda M, Edwards EJ, Nosil P,

Osborne CP, Christin PA. 2017. Introgression and repeated co-option facilitated the
recurrent emergence of C$_4$ photosynthesis among close relatives. *Evolution* **71**: 1541–
1555.

Edwards E. 2019. Evolutionary trajectories, accessibility and other metaphors: the case of C$_4$
and CAM photosynthesis. *New Phytologist* **223**: 1742-1755. doi.org/10.1111/nph.15851

Edwards GE, Ku MSB. 1987. Biochemistry of C$_3$-C$_4$ intermediates. In: Hatch MD,
Boardman NK, eds. *The Biochemistry of Plants*, *Volume 10 Photosynthesis*. New York,
USA: Academic Press, 275-325.

Fennane M, Ibn Tattou M. **1998**. Catalogue des plantes vasculaires rares, menacées ou
endémiques du Maroc. *Bocconea* **8**: 1–243.

García-Herrera R, Gallego D, Hernández E, Gimeno L, Ribera P, Calvo N. **2003**.
Precipitation trends in the Canary Islands. *International Journal of Climatology* **23**: 235–
241. doi:10.1175/1520-0442(2001)014<3889:IOTNAO>2.0.CO;2.

Holaday AS, Talkmitt S, Doohan ME. **1985**. Anatomical and enzymic studies of leaves of a
C$_3$ × C$_4$ *Flaveria* F$_1$ hybrid exhibiting reduced photorespiration. *Plant Science* **41**: 31–
39. doi:10.1016/0168-9452(85)90062-7.

Kadereit G, Bohley K, Lauterbach M, Tefarikis DT, Kadereit JW. **2017**. C$_3$–C$_4$
intermediates may be of hybrid origin – a reminder. *New Phytologist* **215**: 70–76.
doi:10.1111/nph.14567.

Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS. **2017**.
ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature Methods*
14: 587–589. doi:10.1038/nmeth.4285.

Kelly S, Covshoff S, Wanchana S, Thakur V, Quick P, Wang Y, Ludwig M,
Bruskiewich R, Fernie A, Sage RF, *et al*. 2017. Wide sampling of natural diversity
identifies novel molecular signatures of C$_4$ photosynthesis. *bioRxiv*.
doi:https://doi.org/10.1101/163097.

Lauterbach M, Billakurthi K, Hankeln TM, Westhoff P, Gowik U, Kadereit G. 2017a.
Organ-specific gene expression profiling of cotyledons and leaves of C$_4$ *Salsola*
oppositifolia and insights into regulation of C$_2$ and C$_4$ photosynthesis in Salsoleae. In:
Lauterbach M. *C$_4$ photosynthesis in Salsoleae: Ontogenetic gene expression profiling of*

closely related C₃, C₃-C₄ and C₄ species using RNA-Seq. PhD thesis, Mainz, Germany.

Lauterbach M, Billakurthi K, Kadereit G, Ludwig M, Westhoff P, Gowik U. 2017. C_3

cotyledons are followed by C_4 leaves: Intra-individual transcriptome analysis of *Salsola*

soda (Chenopodiaceae). *Journal of Experimental Botany* **68**: 161–176.

doi:10.1093/jxb/erw343.

Lundgren MR. 2020. C_2 photosynthesis: a promising route towards crop improvement? *New*

Phytologist **228**: 1734–1740. doi:10.1111/nph.16494.

Monson RK, Edwards GE. 1984. C_3-C_4 Intermediate Photosynthesis in Plants. *BioScience*

34: 563–574. doi:10.2307/1309599

Morales-Briones DF, Kadereit G, Tefarikis DT, Moore MJ, Smith SA, Brockington SF,

Timoneda A, Yim WC, Cushman JC, Yang Y. 2021. Disentangling Sources of Gene

Tree Discordance in Phylogenomic Datasets: Testing Ancient Hybridizations in

Amaranthaceae s.l. *Systematic Biology* **70**: 219–235. doi:10.1093/sysbio/syaa066.

Morales-Briones DF, Kadereit G. 2022. Exploring the possible role of hybridization in the

evolution of photosynthetic pathways in *Flaveria* (Asteraceae), the prime model of C4

photosynthesis evolution. *bioRxiv*: 2022.01.31.478436.

Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. 2015. IQ-TREE: A Fast and

Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies.

Molecular Biology and Evolution **32**: 268–274. doi:10.1093/molbev/msu300.

Oakley JC, Sultmanis S, Stinson CR, Sage TL, Sage RF. 2014. Comparative studies of C_3

and C_4 *Atriplex* hybrids in the genomics era: physiological assessments. *Journal of*

Experimental Botany **65**: 3637–3647. doi:10.1093/jxb/eru106.

Padrón Mederos MA. 2012. *Caracterización morfológica y genética de especies arbustivas*

de los géneros Salsola L. y Suaeda Forssk. ex J.F. Gmel. en las Islas Canarias. PhD

thesis, Universidad de La Laguna, San Cristóbal de La Laguna, Santa Cruz de Tenerife,

España.

Pease JB, Brown JW, Walker JF, Hinchliff CE, Smith SA. 2018. Quartet Sampling

distinguishes lack of support from conflicting support in the green plant tree of life.

American Journal of Botany **105**: 385–403. doi:10.1002/ajb2.1016.

Reyna-Llorens I, Hibberd JM. 2017. Recruitment of pre-existing networks during the

evolution of C$_4$ photosynthesis. *Philosophical Transactions of the Royal Society B:*

Biological Sciences **372**: 20160386. http://dx.doi.org/10.1098/rstb.2016.0386

Sage RF. **2004**. The evolution of C$_4$ photosynthesis. *New Phytologist* **161**: 341–370.

doi:10.1046/j.1469-8137.2004.00974.x.

Sage RF, Christin PA, Edwards EJ. **2011**. The C$_4$ plant lineages of planet Earth. *Journal of*

Experimental Botany **62**: 3155–3169. doi:10.1093/jxb/err048.

Sage RF, Khoshravesh R, Sage TL. **2014**. From proto-Kranz to C$_4$ Kranz: building the

bridge to C$_4$ photosynthesis. *Journal of experimental botany* **65**: 3341–3356.

doi:10.1093/jxb/eru180.

Sage RF. **2016**. Tracking the evolutionary rise of C$_4$ metabolism. *Journal of Experimental*

Botany **67**: 2919–2922. doi:10.1093/jxb/erw137

Sage RF, Monson RK, Ehleringer JR, Adachi S, Pearcy RW. **2018**. Some like it hot: the

physiological ecology of C$_4$ plant evolution. *Oecologia* **187**: 941–966.

doi:10.1007/s00442-018-4191-6.

Sage RF, Sage TL, Kocacinar F. **2012**. Photorespiration and the Evolution of C$_4$

Photosynthesis. *Annual Review of Plant Biology* **63**: 19–47. doi:10.1146/annurev-

arplant-042811-105511.

Salichos L, Stamatakis A, Rokas A. **2014**. Novel Information Theory-Based Measures for

Quantifying Incongruence among Phylogenetic Trees. *Molecular Biology and Evolution*

31: 1261–1271. doi:10.1093/molbev/msu061.

Sayyari E, Mirarab S. **2016**. Fast Coalescent-Based Computation of Local Branch Support

from Quartet Frequencies. *Molecular Biology and Evolution* **33**: 1654–1668.

doi:10.1093/molbev/msw079.

Schlüter U, Weber APM. **2016**. The Road to C$_4$ Photosynthesis: Evolution of a Complex

Trait via Intermediary States. *Plant and Cell Physiology* **57**: 881–889.

doi:10.1093/pcp/pcw009.

Schulze S, Westhoff P, Gowik U. **2016**. Glycine decarboxylase in C$_3$, C$_4$ and C$_3$-C$_4$

intermediate species. *Current Opinion in Plant Biology* **31**: 29–35.

doi:10.1016/j.pbi.2016.03.011.

Schüssler C, Freitag H, Koteyeva N, Schmidt D, Edwards G, Voznesenskaya E,

Kadereit G. **2017**. Molecular phylogeny and forms of photosynthesis in tribe Salsoleae

(Chenopodiaceae). *Journal of Experimental Botany* **68**: 207–223.

doi:10.1093/jxb/erw432.

Schwarz G. **1978**. Estimating the Dimension of a Model. *The Annals of Statistics* **6**: 461–

464. doi:10.1214/aos/1176344136.

Siadjeu C, Lauterbach M, Kadereit G. **2021**. Insights into regulation of C_2 and C_4

photosynthesis in Amaranthaceae/Chenopodiaceae using RNA-Seq. *bioRxiv*:

2021.09.14.460237. doi:10.1101/2021.09.14.460237.

Slewinski TL, Anderson AA, Price S, Withee JR, Gallagher K, Turgeon R. **2014**. Short-

Root1 Plays a Role in the Development of Vascular Tissue and Kranz Anatomy in

Maize Leaves. *Molecular Plant* **7**: 1388–1392. doi:10.1093/mp/ssu036.

Smith SA, Moore MJ, Brown JW, Yang Y. **2015**. Analysis of phylogenomic datasets

reveals conflict, concordance, and gene duplications with examples from animals and

plants. *BMC Evolutionary Biology* **15**: 150. doi:10.1186/s12862-015-0423-0.

Stamatakis A. **2014**. RAxML version 8: a tool for phylogenetic analysis and post-analysis of

large phylogenies. *Bioinformatics* **30**: 1312–1313. doi:10.1093/bioinformatics/btu033.

Sugiura N. **1978**. Further analysts of the data by akaike's information criterion and the finite

corrections. *Communications in Statistics - Theory and Methods* **7**: 13–26.

doi:10.1080/03610927808827599.

Than C, Ruths D, Nakhleh L. **2008**. PhyloNet: A software package for analyzing and

reconstructing reticulate evolutionary relationships. *BMC Bioinformatics* **9**: 1–16.

doi:10.1186/1471-2105-9-322.

von Caemmerer S. **1992**. Carbon isotope discrimination in C_3-C_4 intermediates. *Plant, Cell*

and Environment **15**: 1063–1072. doi:10.1111/j.1365-3040.1992.tb01656.x.

Voznesenskaya E V, Koteyeva NK, Akhani H, Roalson EH, Edwards GE. **2013**.

Structural and physiological analyses in Salsoleae (Chenopodiaceae) indicate multiple

transitions among C_3, intermediate, and C_4 photosynthesis. *Journal of Experimental*

Botany **64**: 3583–3604. doi:10.1093/jxb/ert191.

Yang Y, Smith SA. **2014**. Orthology inference in nonmodel organisms using transcriptomes

and low-coverage genomes: Improving accuracy and matrix occupancy for

865  phylogenomics. *Molecular Biology and Evolution* **31**: 3081–3092.

866  doi:10.1093/molbev/msu245.

867 **Yu Y, Nakhleh L**. **2015**. A maximum pseudo-likelihood approach for phylogenetic networks.

868  *BMC Genomics* **16**: S10. doi:10.1186/1471-2164-16-S10-S10.

869 **Zhang C, Rabiee M, Sayyari E, Mirarab S. 2018.** ASTRAL-III: polynomial time species

870  tree reconstruction from partially resolved gene trees. *BMC bioinformatics* **19**: 15–30.

871  doi:10.1186/s12859-018-2129-y.

The following Supporting Information is available for this article:

Fig. S1: Carbon isotope values ($\delta13C[‰]$) of different populations (P) of the *Salsola*
divaricata agg. on the Canary Islands and Morocco.

Fig. S2: CO2 compensation points (Γ in µmol CO2 mol-1) of populations (P) of the *Salsola*
divaricata agg.

Fig. S3: a) Cladogram of Salsoloideae inferred from the Maximum likelihood analyses of the
concatenated 991-nuclear gene supermatrix. b) Cladogram of Salsoloideae inferred from the
ASTRAL analyses of 991 nuclear genes trees.

Fig. S4: A) RAxML cladogram of Salsoloideae inferred from the concatenated 991-nuclear
gene supermatrix. B) ASTRAL Cladogram of Salsoloideae inferred from 991 nuclear genes
trees.

Fig S5: Maximum likelihood phylogeny of Salsoloideae s.l. inferred from the IQ-TREE
analysis of complete and partial plastomes.

Fig. S6: Maximum pseudo-likelihood species network inferred with PhyloNet with up to five
hybridization events.

Fig. S7: HyDe boxplot of the distribution of the admixture parameters (γ) from the 298
significant tests.

Fig. S8: Maximum likelihood cladograms of the 50 photosynthetic gene trees. Bootstrap
support (BS) values are shown above the branches.

Table S1: Populations (named by living collection number in the Botanical Garden Mainz;
see Fig. 1) included in the carbon isotope measurements, CO2 compensation point
measurements and PEPC activity measurements, including sampling location and collector.

Table S2: Gene and character occupancy of the concatenated matrix.

Table S3: HyDe results from the 298 significant tests.

Table S4: Fifty trees of photosynthesis pathway-associated genes characteristic.

Materials and Methods S1: More detailed description of Plant material, methods of CO_2
compensation point, PEPC activity measurement, Transcriptome processing and nuclear
phylogenetic analyses, Assessment of hybridization, Plastome assembly and phylogenetic
analysis, and Analysis of photosynthetic gene trees.

Tables

Table 1: List of the 17 samples in the transcriptome phylogeny; with photosynthesis type, ploidy (Kew C value database 2018) and NCBI Sequence Read Archive (SRA) accession number.

Species	Tribe	Ploidy	Photosynthesis type	SRA accession No.
Anabasis articulata Choul. ex Pomel	Salsoleae	NA	C_4	SRR6435311
Halogeton glomeratus (M. Bieb.) C.A. Mey.	Salsoleae	2x	C_4	SRR1503502
Haloxylon ammodendron (C.A. Mey.) Bunge	Salsoleae	2x	C_4 (C_3 cotyledon)	SRR1697346
Hammada scoparia (Pomel) Iljin	Salsoleae	2x	C_4 (C_3 cotyledon)	ERR2060287
Kali collina Akhani & Roalson	Salsoleae	2x	C_4	SRR6435349
Salsola divaricata Moq. (Pop.184)	Salsoleae	4x	C_2	ERR2060298
Salsola divaricata Moq. (Pop.198)	Salsoleae	4x	C_2	ERR2060294
Salsola genistoides Juss. ex Poir.	Salsoleae	4x	C_3	SRR14783909
Salsola montana Litv.	Salsoleae	NA	C_3 proto-kranz	SRR14783908
Salsola oppositifolia Desf.	Salsoleae	8x	C_4	ERR2060302
Salsola soda L.	Salsoleae	2x	C_4 (C_3 cotyledon)	SRR3544552
Salsola verticillata Schousb. (Pop.171)	Salsoleae	4x	C_2	SRR14783907
Salsola webbii Moq.	Salsoleae	4x	C_3	ERR2060308
Caroxylon vermiculatum (L.) Akhani & Roalson	Caroxyloneae	NA	C_4	SRR6435345
Bassia scoparia (L.) A.J. Scott	Camphorosmeae	2x	C_4	ERR364385
Eokochia saxicola (Guss.) Freitag & G.Kadereit	Camphorosmeae	NA	C_3	SRR6435348
Beta vulgaris subsp. vulgaris L.	Beteae	2x	C_3	SRX335625

927 **Table 2**: Model selection between bifurcating trees and networks. The best model (lowest score) is the network
928 with three hybridization events from PhyloNet (best MPL run allowing four hybridization events). k = No.
929 branch lengths + hybridization probabilities [(2n-3)+(2h)]; h = number of hybridization events. All trees and
930 networks had 17 taxa (n). Networks and bifurcating trees log likelihood values were inferred with 991 loci.

tree/network	ln L (log likelihood)	k	h[1]	AICc	ΔAICc	BIC	ΔBIC
ASTRAL-nuclear	-19754.54543	31	N/A	39573.130	1697.190	39722.951	1668.649
RAxML-nuclear	-19741.89558	31	N/A	39547.830	1671.891	39697.651	1643.349
IQTREE-platome	-22836.08408	31	N/A	45736.207	7860.268	45886.028	7831.726
PhyloNet. 1-hybridization search (network 1)	-19451.79848	31	1	38967.636	1091.696	39117.457	1063.155
PhyloNet. 1-hybridization search (network 2)	-19605.00833	33	1	39278.323	1402.383	39437.674	1383.372
PhyloNet. 1-hybridization search (network 3)	-19624.56237	33	1	39317.431	1441.491	39476.782	1422.480
PhyloNet. 2-hybridization search	-19435.04197	35	2	38942.674	1066.734	39111.539	1057.237
PhyloNet. 3-hybridization search	-19276.12872	37	3	38629.147	753.208	38807.510	753.208
PhyloNet. 4-hybridization search	-18899.52481	37	3	37875.940	0.000	38054.302	0.000
PhyloNet. 5-hybridization search	-19283.48635	41	5	38652.512	38652.512	38849.820	38849.820

931 [1]The number of reticulation events in the best networks of a search is not necessarily the same number of
932 reticulation events allowed in the search.