Active Translation Control of CD4 T Cell Activation by Regulatory T Cells

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SUMMARY

Increased protein synthesis is a hallmark of lymphocyte activation. Regulatory T cells (Tregs) suppress the activation and subsequent effector functions of CD4 effector T cells (Teff). Molecular mechanisms that enforce suppression on CD4 Teff cell function are unclear. Control of CD4 Teff cell activation by Tregs has largely been defined at the transcriptional level, which does not reflect changes in post-transcriptional control. We found that Tregs suppressed activation-induced global protein synthesis in CD4 Teff cells prior to cell division. We analyzed genome-wide changes in the transcriptome and translatome of activated CD4 Teff cells using two independent approaches. We show that mRNAs encoding for the protein synthesis machinery are regulated at the level of translation in activated Teff cells. Strikingly, Tregs suppressed global protein synthesis of CD4 Teff cells by specifically inhibiting mRNAs of the translation machinery at the level of mTORC1-mediated translation control. Lastly, we found that the RNA helicase eIF4A inhibitor rocaglamide A (RocA) can suppress CD4 Teff activation *in vitro* to alleviate inflammatory CD4 Teff activation caused by acute Treg depletion *in vivo*. These data provide evidence that peripheral tolerance is enforced by Tregs, mediated by IL-10, through mRNA translational control in CD4 Teff cells. Therefore, therapeutic targeting of the protein synthesis machinery can mitigate inflammatory responses invoked by Treg loss of function.

INTRODUCTION

Most self-reactive T cells are eliminated in the thymus through the process of central tolerance. However, a small percentage of cells escape to the periphery, where they have the potential to promote autoimmunity. These cells are normally held in check by a population of CD4 T cells referred to as regulatory T cells (Tregs). Tregs are essential to maintain immune homeostasis, and the transcription factor FOXP3 has been shown to be central to the development and function of Tregs. Mutations in FOXP3 gene in mice and human patients with IPEX syndrome develop a common set of autoimmune symptoms(Bennett et al., 2001; Ochs et al., 2007; Patel, 2001; Wildin et al., 2001). Mutations in the FOXP3 gene and the autoimmune phenotype is linked to a loss of Tregs or their function(Bacchetta et al., 2018; Ochs et al., 2007). Tregs have the ability to potently suppress CD4 effector T cells (Teff) either directly or through the modulation of antigen presenting cells (mainly DCs) to ultimately suppress activation, proliferation, and subsequent effector functions of Teff cells(Josefowicz et al., 2012; Tang and Bluestone, 2008; Vignali et al., 2008). Several mechanisms have been proposed for Treg-mediated suppression, including release of suppressive cytokines (e.g., TGFβ, IL-10, IL-35) and expression of inhibitory receptors (e.g., CTLA-4, PD-1, TIGIT)(Shevach, 2009; Sojka et al., 2008). Although a block in proliferation and effector T cell function have been the hallmarks of Treg mediated suppression, the molecular changes in target Teff cells following Treg encounter still remains unclear. This is especially true for the first 24 hours prior to the onset of proliferation, when the biosynthetic capacity of the cell is greatly expanded(So et al., 2016; Wang et al., 2011).

Upon activation, resting T cells undergo a rapid biosynthetic and metabolic reprogramming in preparation for cell division(Manfrini et al., 2020; Ricciardi et al., 2018; Wang et al., 2011; Wolf et al., 2020). Included in this reprogramming is an increase in translational activity and capacity(Araki et al., 2017; Bjur et al., 2013). In this study, we show that Tregs suppress activation of Teffs by enforcing a global inhibition of mRNA translation. We assessed the genome-wide changes in transcriptome and translatome in activated CD4 Teff cells and identified translation control of mRNAs encoding components of the protein synthesis machinery. In the first 24 hours of Teff cell activation a set of mRNAs encoding proteins involved in the translational machinery, and whose transcription was unchanged, are shifted to polysomes. We found that Tregs specifically inhibit the shift of these mRNAs to polysomes by suppressing mammalian target of rapamycin complex 1 (mTORC1) signaling. In support of these findings, we provide new evidence that direct targeting of protein synthesis using rocaglamide A (RocA), a RNA helicase eIF4A inhibitor, inflammatory CD4 Teff activation caused by *in vivo* Treg loss can be alleviated. In summary, we provide a novel mechanism of Treg-mediated suppression of CD4 Teff activation through inhibition of protein synthesis machinery mRNAs at the post-transcriptional level and that such biological mechanism can be therapeutically targeted using small molecule inhibitors.

RESULTS

Tregs control protein synthetic capacity of activated CD4 Teff cells.

The 24-48 hours following CD4 Teff cell activation are critical for subsequent proliferation and expansion. This is a period where cellular biomass is accumulated through expansion of global protein synthetic capacity in preparation for cell division. We reasoned that this time period would be a target for Treg-mediated suppression in order to inhibit cell activation prior to proliferation. Overall protein

synthesis rate can be quantified at the single-cell level by pulsing cells with the tRNA-analog puromycin (PMY) and intracellular staining for PMY. We co-cultured CD4 Tconv cells ($CD4^{+}CD25^{-}$) and congenically marked Tregs (CD4+Foxp3+) at varying ratios with anti-CD3/CD28 coated beads and pulsed the culture with PMY. CD4 Tconv cells co-cultured with Tregs exhibited marked inhibition of proliferation in a dose-dependent manner (Figure S1A). Interestingly, Tconv cells co-cultured with Tregs clearly showed significantly less PMY incorporation in a Treg-dose-dependent manner before the onset of proliferation (Figure 1A). The pattern of suppression was not bimodal indicating that protein synthesis rate of all responding Tconv cells were modulated by Tregs but not completely compared to cycloheximide (CHX) treatment (Figure 1B). Since anti-CD3/CD28 beads were used to activate both populations of cells in the co-culture system, the downregulation of protein synthesis by Tregs is independent of antigen-presenting cells (APC). The Treg-mediated translational inhibition was observed as early as 6 hours post-activation, well before any metabolic changes occur in T cells (Figure 1C). Suppression was primarily due to early prevention of T cell activation as we found no difference in the ability of CD4 Teff cells to downregulate CD62L with or without Tregs (Figure S1B). Thus, suppression of global protein synthesis in CD4 Teff cells by Tregs could not be attributed to dampening or cold inhibition of general T cell activation. To test if similar regulatory pathways were operative in human T cells, we established *ex vivo* Treg suppression assays using PMY incorporation as the readout. We used *in vitro*-expanded Tregs from a single donor, and Tconv (defined as CD4+CD45RA+CD127+CD25-) from 5 individual healthy donors. We found that 24 hours of stimulation resulted in a significant increase in PMY staining, both in percentage of cells labeled and the MFI of PMY staining. In the cultures containing Tregs PMY incorporation was significantly reduced, both in total incorporation and in the percentage of cells that incorporated PMY (Figure 1D). These data demonstrate that Treg-mediated inhibition of activation-induced translation is a conserved function.

Next, to assess the role of Tregs in controlling protein synthesis in CD4 T cells *in vivo*, we acutely depleted Tregs through diphtheria toxin (DT) treatment of *Foxp3*^{DTR} mice(Kim et al., 2007). Within 3 days post initial DT-induced depletion of Tregs (2 consecutive DT injections on day 0 and 1), we observed rapid appearance of a CD4 T cell population with significantly elevated incorporation of puromycin *ex vivo* compared to the PBS treated control mice, suggesting activation of the autoreactive CD4 T cell pool in the periphery (Figure 1E). When CD4 T cells from spleen and lymph nodes of DT-treated *Foxp3*^{DTR} mice were purified and stimulated *ex vivo*, they proliferated with faster kinetics and were significantly larger in size indicating an aberrantly enhanced protein synthesis capacity in Teff cells activated *in vivo* attributed to Treg loss (Figure S1C). These data suggest that Tregs are both sufficient and necessary to suppress the rapid upregulation of protein synthesis in activated CD4 T cells both *in vitro* and *in vivo*.

To uncover the underlying mechanism of Treg-mediated translational inhibition we examined signaling pathways downstream of TCR stimulation in activated CD4 T cells. Specifically, we examined the mammalian target of rapamycin (mTOR) signaling pathway as it has been shown to be critical for coordinating cell growth and proliferation in lymphocytes through the eukaryotic translation initiation factor 4E (eIF4E) in translation initiation(Wardman et al., 2016). mTOR exists in 2 multi-protein complexes, mTOR complex 1 (mTORC1) and mTOR complex 2 (mTORC2)(Jhanwar-Uniyal et al., 2019; Zoncu et al., 2011). To assess mTORC1 signaling, we examined phosphorylation of ribosomal protein S6 (rpS6: S240/244) and eIF4E-binding proteins (p4EBP1/2: T37/46) at their respective

mTORC1 specific phosphosites in activated CD4 T cells. mTORC2 signaling was assessed by phosphorylation of AKT at S473. As expected, both mTORC1 and mTORC2 signaling increased in activated CD4 T cells. Strikingly, Tregs significantly suppressed mTORC1 signaling (rpS6 and p4EBP1/2). However, mTORC2 signaling (AKT S473) remained intact, as did the PDK1- and PI3K-dependent phosphosite T308 which is more proximal to TCR engagement (Figure 1F). Using Nur77-GFP reporter Teffector cells, we see no differences in Teff cells co-cultured with Tregs indicating no change in proximal TCR signaling (Figure S1D). These findings are consistent with previous data showing that genetic or chemical inhibition of mTORC1 significantly inhibits lymphocyte proliferation in a 4EBP/eIF4E- dependent manner to control translation initiation in various cell types including lymphocytes (So et al., 2016; Thoreen et al., 2012).

Translatome capture by RiboTag and novel method called Simple Polysome Efficient Extraction and Distribution (SPEED).

Based on our observation that Tregs suppress protein synthesis in Teffs, we investigated if Tregs affect global mRNA translation using genome-wide approaches to interrogate changes in both the transcriptome and translatome of CD4 Tconv cells. First, we used the RiboTag system to capture translatome changes in CD4 T cells. This system utilizes an HA epitope-tagged ribosomal protein large subunit L22 (eL22) that allows for immunoprecipitation of proteins and mRNA associated with ribosomes (Ribo-IP)(Sanz et al., 2009). RiboTag CD4 T cells were generated using *Cd4-Cre* mice (*eL22*$^{Ribo f/f}$*Cd4Cre*: T-Ribo) (Figure S4A-B).

While the data using the Ribo-Tag system showed that Tregs can control mRNA-ribosome association in Teff cells, it does not distinguish between mRNAs with low ribosome occupancy (monosomes) from those associated with polysomes (Supplement text 1, Figure S4). The transition of mRNAs from monosome to polysome and back is a critical aspect of translation control. To resolve and distinguish mRNAs bound to polysomes from monosome-associated mRNAs we optimized the classical polysome profiling approach for low input cytosolic lysates (Figure 3A). We reasoned that by assessing the quantity and quality of total RNA extracted from fraction, we could determine ribosome positions since total RNA is mainly composed of ribosomal RNA (rRNA). Using lysates prepared from as few as 500,000 to 1 million activated CD4 T cells, we found that total RNA extracted from each fraction and plotted as a percent distribution plot closely resembled a classical A254nm polysome trace obtained using >20 million activated CD4 T cells (Figure 3B). We have termed this a 'Simple Polysome Efficient Extraction and Distribution' (SPEED) plot to distinguish it from the traditional polysome traces requiring greater cellular input (Figure 3C). Furthermore, qualitative analysis of the extracted RNA using Bioanalyzer gave us information as to the position of the intact 80S monosome in the gradient (Figure 3C). The SPEED plots generated from unstimulated CD4 T cells showed most of the total RNA to be enriched in the monosome fraction (Figure 3C), confirming our observations that CD4 T cells prior to activation have low translational activity (Figure 1A). Upon stimulation, nearly 50% of the monosomes shifted towards the heavier sucrose fractions indicating the assembly of polysomes and increased protein synthesis (Figure 3C). To ensure SPEED plots were a faithful representation of the mRNA translational status of a cell, we took advantage of the translation initiation inhibitor harringtonine, which only interferes with initiating ribosomes and allows elongating ribosomes to run-off(Fresno et al., 1977). Activated CD4 Teff cells were treated with homoharringtonine (HHT) for 10 min to allow run-off elongation of ribosomes before cytosolic lysate preparation. Remarkably, SPEED plot from HHT treated activated CD4 Teff cells resembled unstimulated CD4 T cells with majority of ribosomes enriched in the

monosome fraction (Figure 3C). Lastly, the distribution of beta-actin (*Actb*) mRNA was analyzed using quantitative PCR (qPCR) from each fraction. We chose *Actb* as it is routinely used as a housekeeping control mRNA as it has no apparent *cis*-regulatory sequence motif in its 5' untranslated region (UTR) and is highly translated. Despite low polysome levels in unstimulated CD4 Teff cells, we found *Actb* mRNA to be abundantly enriched in the polysome fractions. Activated CD4 Teff cells also translated *Actb* mRNA with high efficiency. As expected for a highly translated mRNA, HHT treatment led to a complete shift in *Actb* mRNA towards the lighter sucrose fractions indicating successful ribosome run-off (Figure 3D). In summary, our SPEED technique faithfully captured the translational status of cellular lysates from low biological input, making it ideal to assess translatome of primary immune cells and most importantly, in Treg suppressed CD4 Teff cells.

To determine the identity of the mRNAs that shift between monosomes and polysomes following activation we performed SPEED on CD4 T cells activated for 24h with CD3/CD28 beads. A portion (10%) of cellular lysate was used to isolate total RNA, while the remainder was subjected to SPEED analysis with monosome and polysome fractions collected and RNA isolated and sequenced (Figures 4A, S4 and S5). We found approximately 380 genes whose overall RNA level was unchanged but whose mRNAs were shifted from monosomes to polysomes following activation (Figures 3B and C). Interestingly, gene ontology analysis of these genes showed that the vast majority fell into functional categories involving ribosome biogenesis and mRNA translation, consistent with preparation for subsequent cell division following activation (Figures 3D and E). It is important to note that the genes that encode these mRNAs are considered to be housekeeping genes whose expression is very high and largely unchanged by cell activation. The data presented here demonstrates that while the RNA level of transcripts encoding the transcriptional machinery is unchanged, their translation is likely increased following activation due to their shift onto polyribosomes.

Treg sensitive mRNAs are enriched for terminal oligopyrimidine (TOP) motif.

We next determined the fate of these mRNAs in Teff cells stimulated in the presence of Tregs. We examined the mRNAs that showed increased translation efficiency (TE) in stimulated vs. resting cells and calculated their TE in CD4 T cells stimulated in the presence of Tregs (Stim+Treg). The set of mRNAs showing increased TE in stimulated cells were specifically reduced in their TE when the cells were stimulated in the presence of Tregs (Figure 3C). These data are consistent with our hypothesis that Treg-mediated translational control targets mRNAs involved in preparing the cell for subsequent proliferation, with ribosome biogenesis and translational activity being critically important for this process. The finding that stimulation in the presence of Tregs reversed the positive TE changes in transcripts encoding proteins involved in mRNA translation is consistent with data showing that overall protein synthesis is suppressed by Tregs and our observations using the RiboTag system (Figures 1 and 2). To identify the mechanism(s) that underlie Treg-specific translation control, we first examined *cis*-regulatory elements shared by the 5'UTRs among Treg-sensitive mRNAs identified by SPEED (Supp Table 1). We searched for novel sequence-specific motifs within target UTRs in these Treg sensitive genes by retrieving murine 5'UTR sequences from Ensembl BioMart database (Ensembl Genes78, Mus musculus) and using Multiple Em for Motif Elicitation (MEME), to uncover common *cis*-regulatory elements(Cunningham et al., 2014; Leppek et al., 2018; Truitt et al., 2015). We found an oligopyrimidine tract enriched in the 5'UTR in Treg sensitive mRNAs (Figure 4A). The terminal oligopyrimidine (TOP) motif is a well-characterized motif which regulates key mRNAs encoding the translational machinery.

The TOP motif is known to be present in the 5'UTR of ribosomal proteins (RPs) and other genes required for mRNA translation(Meyuhas et al., 1987). Interestingly, TOP motif containing RP transcripts are regulated by the well-known signaling kinase mammalian target of rapamycin (mTORC1)(Schneider et al., 2013; Thoreen et al., 2012). We further validated select genes that were differentially regulated in SPEED RNA-seq. We observed *Rps10*, *Rpl14*, *Dhx34*, *eIF3e*, and *Rpl8,* mRNAs shift to polysomes upon CD4 T cell stimulation and back to monosomes following activation in the presence of Tregs (Figure 4B). This shift does not reflect a change in the RNA level of these genes, but rather is a change in the translation of these mRNAs. As predicted, the mRNA distribution shifted back to the monosome fraction when the CD4 T cells were stimulated in the presence of Tregs. As a control, *ActB* mRNA is unchanged by Tregs. These data confirm global changes in the translatome induced by Tregs to control CD4 T cell activation by blocking the translation of a subset of mRNAs, thereby blunting the ability of these cells to respond appropriately to stimulation.

IL-10 from Tregs block protein synthesis in Teffector cells

Since the activation of CD4 T cells in the presence of Tregs resulted in a specific inhibition of mTORC1 activation, as indicated by the lack of ribosomal protein S6 and 4E-BP1/2 phosphorylation, we hypothesized that Tregs suppress the immediate biosynthetic response to antigen-specific activation through regulation of mTORC1-mediated translational control. To understand the pathways leading to the inhibition of mTORC1 activation in stimulated CD4 T cells by Tregs, we examined the role of soluble factors known to be produced by Tregs, using recombinant proteins and neutralizing antibodies specific for anti-inflammatory factors. While blocking IL-2 mediated signaling in stimulated T cells with IL-2 or CD25 antibodies showed a slight reduction in pS6, we did not observe significant effects on 4E-BP1 or puromycin incorporation, a surrogate of protein synthesis (Figure 5A). Treatment with recombinant TGF-β failed to suppress the mTORC1 pathway or protein synthesis (Figure 5B). However, treatment with recombinant IL-10 significantly reduced pS6, p4E-BP1, and puromycin incorporation in stimulated CD4 T cells (Figure 5C). Furthermore, we also observed inhibition of Treg mediated suppression of mTORC1 signaling and protein synthesis by αIL10 antibody treatment (Figure 5D). We saw a similar effect when Tregs were cocultured with stimulated IL-10Rb$^{-/-}$ CD4 T cells (Figure 5E). These data suggest that Tregs use IL-10 to disrupt mTORC1 signaling, affecting mRNA translation in stimulated CD4 T cells.

Acute inflammation due to Treg deficiency rescued by an mRNA translation inhibitor

While we show that mRNA translation suppression by Tregs has a direct effect on cell activation and proliferation, we wanted to investigate if this is an important mechanism by which Tregs exert their function downstream of mTORC1. To test this, we used a translation inhibitor Rocaglamide A (RocA,1*H*-2,3,3a,8b-tetrahyrocyclopenta[*b*]benzofuran), which has been shown to reduce overall protein synthesis but exhibit preferential inhibition of a specific subset of mRNAs in cell culture models(Iwasaki et al., 2019). RocA is a secondary metabolite from the plant genus *Aglaia* with anti-tumor and anti-inflammatory properties (Ebada et al., 2011; Li-Weber, 2015) (Bordeleau et al., 2008; Cencic et al., 2009; Li-Weber, 2015). RocA affects protein synthesis by binding to eIF4A, a DEAD-box RNA helicase, that is part of the eIF4F mRNA initiation complex (Rogers et al., 2002). RocA binds to eIF4A and clamps it on poly-purine sequences in the 5' untranslated regions of mRNAs, thereby preferentially inhibiting these set of mRNAs at the level of translation control where stable RocA-eIF4A-mRNA complexes are formed to block 43S pre-initiation complex scanning (Ernst et al., 2020; Iwasaki et al., 2019). While RocA has been shown to inhibit overall protein synthesis through the translational blockade of a subset of mRNAs

(Iwasaki et al., 2016), its effect on CD4 T cell activation and proliferation is unclear. To address this, we measured protein synthesis rate in activated CD4 T cells 24h post RocA treatment using the puromycylation assay (Schmidt et al., 2009; Seedhom et al., 2016). RocA-treated cells showed a dose-dependent decrease in puromycin incorporation, suggesting inhibition of protein synthesis. We also found that RocA treatment inhibited cell-proliferation in a dose-dependent manner at sub nanomolar concentrations (Figures 6A, B). Surprisingly, early T cell activation genes such as IL-2 and CD25 (IL-2Rα) were unaffected both at the mRNA and protein level by RocA, suggesting that early TCR signaling dependent on NFAT is not how RocA inhibits T cell proliferation. Furthermore, this supports the notion that translational blockade through RocA impacts subset of mRNAs even in T cells. As expected, when cells were treated with the calcineurin inhibitor Tacrolimus (FK506), T cell proliferation was inhibited with corresponding downregulation of both IL-2 and CD25 (IL-2Rα) (Figure 6C). These results suggest that while both RocA and FK506 inhibit CD4 T cell activation and proliferation, they do so through a different mechanism. While our finding is different from a previous study suggesting that RocA inhibits NFAT activity in human CD4 T cells (Proksch et al., 2005), we propose at higher concentrations (50-100 fold) RocA could affect transcription regulation.

Inflammation mediated by Treg deficiency rescued by an mRNA translation inhibitor.

The data presented above demonstrated that RocA-mediated translation inhibition could suppress CD4 T cell proliferation *ex vivo*. To extend these studies we determined whether RocA administration could ameliorate inflammation-induced CD4 T cell activation *in vivo*. First, we assessed the effect of RocA on the general health of the mice and immune homeostasis by treating mice every other day for a week with RocA (0.5 and 1mg/kg i.p.) (Figure S6). We found no differences in the overall weight with RocA treatment, nor in various cell populations within the $CD45^+$ compartment, including CD4+ T cell subsets indicating no deleterious effects on immune homeostasis (Figure S6). Thus, short-term *in vivo* treatment with RocA was well tolerated by the mice as we did not observe any significant changes in the general health of these animals.

We next asked whether inflammatory responses were limited by RocA treatment in the absence of Tregs. For these studies we used the $Foxp3^{DTR}$ mice, treated with DT, to deplete Tregs as described in Figure 1D. To test the ability of RocA to affect acute inflammation we dosed $Foxp3^{DTR}$ mice with RocA (0.2 mg/kg i.p.) or DMSO (vehicle) on days -1 and -2, and then treated with DT (400 ng i.p.) on days 0, 2, 4, and 6, followed by sacrifice and analysis on day 8 (Figure 6E). A hallmark feature of Treg depletion is a rapid loss of body weight, which is seen in the vehicle-treated mice on day 7 with an average weight loss of 20% of initial body weight (Figure 6F). We found that the RocA-treated mice showed significantly less weight loss, averaging between 85-90% of starting weight, suggesting that RocA treatment was blunting the inflammation caused by Treg depletion. Consistent with this we found that numbers of splenic CD4 T cells producing IL-17A, IFNγ, and both were significantly reduced in RocA-treated mice (Figure 6G). Especially important was the reduction in IFNγ/IL-17A co-producers as this subset has been found to be pathogenic in inflammatory settings (Doodes et al., 2010; Duhen et al., 2013; Kaiser et al., 2016; Lee et al., 2009). These data demonstrate that RocA-mediated direct translation inhibition can ameliorate the inflammation seen in mice lacking functional Tregs, supporting our findings that Treg-mediated immune tolerance on CD4 T cells is through an active translation control on the subset of mRNAs required for protein synthesis itself.

Lastly, we also found suppression of proliferation in primary human T cells treated with RocA and stimulated with αCD3/CD28 (Figure 6H). Similar to what was seen in mouse CD4 T cells, there was no difference in the surface expression of CD25 (Figure 6H). We also observed significant suppression of IFNγ producing cells with RocA treatment compared to the vehicle control post PMA/ionomycin stimulation (Figure 6I). Altogether, here we show a novel mechanism of Treg-mediated CD4 T cell activation which acts through active mRNA translation control to downregulate the protein dosage of protein synthesis machinery components in activated CD4 T cells. Finally, we show in a physiologically relevant setting that direct inhibition of protein synthesis using a small molecule inhibitor (RocA) can have therapeutic efficacy in alleviating unwanted inflammatory CD4 T cell activation.

DISCUSSION

Regulatory T cells possess the ability to potently suppress effector CD4 T cell responses and maintain proper immune homeostasis. Several mechanisms have been proposed for Treg-mediated suppression, all predominately focused on inhibiting aspects of effector T cell proliferation. Despite this emphasis, it is still unclear how target cell gene expression is modulated by Tregs. One key hallmark of CD4 T cell activation is the significant upregulation of global protein synthesis capacity and rate. This raises an important question as to how Tregs mediate the suppression of proliferation and activation of Teff cells. While the Treg-mediated suppression mechanisms have been heavily studied from the Treg-centered perspective yet the molecular events that happen in effector CD4 T upon Treg encounter have remained unclear(Akkaya and Shevach, 2020; Sojka et al., 2008; Vignali, 2012). To address this, we determined the effect of Tregs on post-transcriptional gene regulation in Teff cells, focusing on protein biosynthesis. We observed that Teff cells rapidly increased their global protein synthetic rate following TCR-mediated activation, and that Tregs were capable of suppressing this increase in translation at 24 hours of activation. Interestingly, we found this process is inhibited by Tregs to shut down the biosynthetic ramp-up, as early as 6 hours following T cell activation, required for subsequent cell division. Similarly, an acute depletion of Tregs *in vivo* led to the robust upregulation of protein synthesis rate in peripheral CD4 T cells. These data suggest that an important aspect of Treg-mediated suppression is inhibition of the ability of Teff cells to increase their biomass prior to cell division. The rapid increase in protein synthesis following acute loss of Tregs demonstrates a need for Tregs to hold in check the aberrant increase in protein synthesis of autoreactive T cells in the periphery. This is a highly efficient mode of regulation, inhibiting the ramp-up of biosynthesis that proceeds cell division, thereby stopping proliferation before it has begun.

We used two complementary approaches to uncover the mechanism by which Tregs suppress protein synthesis in Teff cells. We found that, upon activation, Teff cells displayed a rapid redistribution of mRNAs from monosome to polysomes, leading to increased translation. This redistribution was independent of acute changes in the transcription of the mRNAs. The vast majority of these mRNAs encoded for proteins important for protein biosynthesis, including ribosomal proteins, RNA-binding proteins, elongation, and splicing factors, suggesting that this process was involved in increasing the biomass prior to cell division. Importantly, we found that activation in the presence of Tregs inhibited the redistribution of these mRNAs, causing them to remain either free or associated with monosomes. As these changes in ribosome occupancy were independent of gene transcription and occurred within 24

hours of stimulation, they would be undetectable by the usual methods employed to study Treg-mediated suppression-gene expression changes and cell division. Interestingly, the mRNAs affected by Tregs were enriched in 5' UTR terminal oligopyrimidine (TOP) motifs, from monosomes to polysomes in the absence of increased transcription. This motif is enriched in 5'UTRs of mRNA including ribosomal proteins and elongation factors that are core proteins of the translational machinery. This is a motif recognized by the RNA binding protein LARP1 and regulates the association of these mRNAs with stress granules(Mattijssen et al., 2021; Philippe et al., 2020). The presence of this motif on Treg-affected mRNAs suggests that Tregs may regulate the trafficking of these mRNAs from ribosomes to stress granules, thus controlling their translation.

From the perspective of the requirements for efficient Teff cell activation and clonal expansion, it is essential that activated Teff cells synthesize new proteins and increase their biomass. We observed that this process is specifically regulated by mTORC1 signaling. While it is clear that the mTOR pathway is critical for coordinating cell growth and proliferation in lymphocytes(Howie et al., 2014; So et al., 2016), we observe that Tregs specifically block mTORC1 signaling required for mRNA translation. We found that activation in the presence of Tregs led to an inhibition of S6 and 4E-BP1/2 phosphorylation, both downstream targets of mTorc1. Blockade of IL-10 signaling in the Teff cells relieved the inhibition of protein synthesis, while the addition of IL-10 during Teff stimulation (in the absence of Tregs) resulted in a reduction of global translation. Thus, IL-10 signaling is involved in Treg-mediated trans-inhibition of protein synthesis. Previous studies have shown divergent effects of IL-10 signaling on mTORC1 activity. In macrophages, IL-10 acts in a paracrine or autocrine fashion to inhibit mTORC1 activation following LPS stimulation(Baseler et al., 2016; Ip et al., 2017). The outcome was a metabolic reprogramming of the macrophages to promote oxidative phosphorylation. However, in human NK cells, IL-10 regulated metabolic changes that enhanced cellular function (e.g., IFNγ production), and these changes were mediated via activation of mTORC1(Wang et al., 2021). While these studies show opposite effects of IL-10 on cellular activity, they focus on changes in metabolism mediated through regulation of mTOR signaling. However, these studies do not address the role of IL-10 on protein synthesis.

Finally, to determine whether inhibition of translation was an effective method of regulating cellular activation we treated Treg-depleted mice with the translation inhibitor RocA. We choose RocA as it is a specific inhibitor of mRNA translation and well-tolerated *in vivo*. CD4 T cells in mice with acute Treg depletion display an immediate increase in global protein synthesis. Consistent with our hypothesis that inhibition of mRNA translation is an effective means of maintaining tolerance, treatment of these mice with RocA abrogated enhanced protein synthesis and inhibited induction of cytokine expression. Taken as a whole, our data show that Tregs suppress Teff cell activation, and subsequent proliferation and cytokine production, through inhibition of mTORC1, leading to suppression of protein translation through sequestration of specific mRNAs from polysomes. Our study provides a novel mechanism for the induction and maintenance of peripheral tolerance and identifies Treg derived IL-10 as one of the critical the regulators of mRNA translation affecting proliferation and activation of effector T-cells.

In this study we show that peripheral tolerance is enforced by Tregs through mRNA translational control in CD4 Teff cells. Regulatory T cells possess the ability to potently suppress effector CD4 T cell responses and maintain proper immune homeostasis. Despite the many mechanisms of suppression that have been proposed, it is still unclear how target cell gene programming is modulated by Tregs. One key

hallmark of CD4 T cell activation is the significant upregulation of global protein synthesis capacity and rate. The major unanswered question is the mechanism through which Tregs mediate suppression that blocks proliferation and activation of Teff cells. Molecular events mediating Treg-mediated suppression of effector CD4 T have remained unclear. To address this, we interrogated if post-transcriptional gene regulation played a role in activated Teff cells undergoing Treg-mediated suppression. We observed that the Tregs were both necessary and sufficient to suppress this global rate of protein synthesis in activated Teff cells. Importantly, we found this process is inhibited by Tregs to shut down the biosynthetic ramp-up seen in the first 24 hours following T cell activation required for subsequent cell division. This was based on our observation that the addition of Tregs to Teff cells *in vitro* was sufficient to suppress overall protein synthesis rates in activated Teff cells. As mRNA translation is energetically demanding, an mTOR dependent switch to aerobic glycolysis is well known(Huang et al., 2020; Makowski et al., 2020). Recent study has shown key enzymes in the pathway are translationally regulated important for metabolic reprogramming of activated cell(Ricciardi et al., 2018). These data suggest that translation remodeling occurs earlier than metabolic reprogramming in these cells. We did not detect any mRNAs linked to metabolic processes as we probed for early events of translation remodeling. Similarly, acute depletion of Tregs *in vivo* led to the acute upregulation of protein synthesis rate in peripheral CD4 T cells. This suggests that there is a need for Tregs to hold in check the aberrant increase in protein synthesis of autoreactive T cells in the periphery. Regardless of the different mechanisms that Tregs may utilize for its suppression on Teff cells, a common theme is that changes in intracellular signaling events in Teff cells result in inhibition of proliferation. From the perspective of the requirements for efficient Teff cell activation and clonal expansion, it is essential that activated Teff cells synthesize new proteins. We observed that this process is specifically regulated by mTORC1 signaling and Tregs specifically block mTORC1 signaling required for mRNA translation.

Materials and Methods

Mice

RiboTag (B6J.129(Cg)-*Rpl22*$^{tm1.1Psam}$/SjJ: Stock# 02997 from Jackson Labs) mice were a gift from Dr. Stanley McKnight (University of Washington(Sanz et al., 2009). These mice were either bred with *Cd4Cre* mice (B6.Cg-Tg(Cd4-cre)1Cwi/BfluJ: Stock# 022071 from Jackson Labs) for mature T cell-specific RiboTag mice or *CMVCre* mice (B6.C-Tg(CMV-cre)1Cgn/J: Stock# 006054) for germline whole-body RiboTag mice. For Treg-depletion studies, *Foxp3-DTR* mice (B6.129(Cg)-*Foxp3*$^{tm3(DTR/GFP)Ayr}$/J: Stock# 016958 from Jackson Labs) were used. For Treg isolation and expansion *in vivo*, *Foxp3*$^{YFP\text{-}Cre}$ mice (B6.129(Cg)-*Foxp3*$^{tm4(YFP/icre)Ayr}$/J: Stock#016959 from Jackson Labs) that have been crossed to B6 Cd45.1 mice (B6.SJL-*Ptprc*^{a}*Pepc*^{b}/BoyJ: Stock# 002014 from Jackson Labs) in house were used. For the studies of IL-10 in Treg-mediated suppression of protein synthesis, 7 weeks old female of IL-10RB$^{-/-}$ (B6.129S2-Il10rbtm1Agt/J: Stock# 005027 from Jackson Labs) mice were used.

In vivo treatments

Foxp3^{DTR} mice were intraperitoneally injected with 100 ng of Diphtheria toxin (DT) at indicated days. For treatment of rocaglamide (RocA), mice were intraperitoneally administrated with indicated concentrations of RocA.

Murine T cell isolation, culture and suppression assays

Bulk CD4 T cells were isolated from indicated mice and stained with antibodies against CD25 and CD4 with the exception of CD4 T cells from *Foxp3*$^{YFP\text{-}Cre}$ mice since Foxp3+ Tregs were identified using YFP fluorescence from these mice. CD4 Teff cells devoid of Tregs (CD4+CD25-) were sorted using the BD FACS Aria sorter. Unless noted, all isolated T cells were cultured in T cell media (RPMI-1640, 10% Fetal Bovine Serum, 2mM GlutaMAX^{tm}-I, 100U/mL Penicillin-Streptomycin, 55µM 2-Mercaptoethanol, 1mM Sodium Pyruvate, 1X Non-Essential Amino Acids, 10mM HEPES). For Treg suppression assays, indicated numbers of tTregs (CD4+YFP+) and 1 x 10^5 CD4 Teff cells (CD4+CD25-) were cultured in round-bottom 96-well plate for the indicated times with 4 x 10^4 anti-CD3/CD28 coated magnetic beads (Teff:beads = 2.5:1). To control for T cell density, control cultures were cultured with either the same number of total Teff cells as in the Teff:Treg co-culture conditions or in some cases, CD4+YFP-CD45.1+ Teff cells sorted from *Foxp3*$^{YFP\text{-}Cre}$ mice were used. To isolate ribosome-associated mRNAs from only RiboTag CD4 Teff cells (CD45.2+) after Treg co-culture, CD45.1+ Tregs were positively depleted using CD45.1-FITC and anti-FITC microbeads and running the cell mixture through LD columns (all reagents from Miltenyi Biotech). Routinely, the purity of CD45.2+ Teff cells after purification was 99.9-100% based on flow cytometry. Ice-cold PBS and MACS buffer (0.5% Bovine Serum Albumin fraction V, 2mM EDTA) were supplemented with 100µg/ml cycloheximide (CHX) (Sigma, C4859) to stall elongating ribosomes throughout all isolation processes. To test proximal TCR signaling, in vitro Treg suppression assay was performed as described with modifications. CD4 Teff cells were isolated from Nur77-GFP mice and Tregs were sorted from *Foxp3*$^{YFP\text{-}Cre}$ mice. Prior to co-culture, CD4 Teff cells were further stained with CellTraceViolet to assess proliferative status and ensure proximal TCR signaling assessed by Nur77-GFP was prior to proliferation of these cells. Unstimulated Nur77-GFP CD4 Teff cells and Tregs from wild-type C57/B6 mice served as a negative for GFP signal.

Ex-vivo human Treg suppression assay

We used in vitro-expanded Tregs from a single donor, and Teff (defined as CD4+CD45RA+) from 5 individual healthy donors for Treg suppression assay. Established protocols were used for Treg expansion (Long et al., 2017; Putnam et al., 2009) and Teff ($CD4^+CD45RA^+$) cells were purified PBMC using the naïve human T cell isolation kit from Miltenyi. To determine the effect of Tregs on overall translation in Teff we cultured Teff (10^4 cells) in absence or presence of Tregs (1:4 Treg:Teff) for 24 hours with anti-CD3/CD28 beads (at a ratio of 1:28 [bead:Teff]). Puromycin (10 µg/ml) was added for the last 15 minutes of culture, and then the cells were permeabilized and stained with Alexa647-coupled anti-puromycin antibody and analyzed by flow cytometry. Controls included unstimulated Teff cells also labeled with puromycin, and stimulated Teff given CHX at the same time as puromycin (CHX results in a complete translation blockade, so this controls for non-specific uptake of puromycin).

RiboTag immunoprecipitation (Ribo-IP) and RNA isolation

RiboTag CD4 Teff cells (or control CD4 Teff cells: RiboTag mice with no *Cd4Cre* expression) were washed once with ice-cold PBS with CHX and lysed in 100µl of Polysome Lysis Buffer (PLB) (25mM Tris-HCl pH 7.5 (Ambion, AM9850G), 150mM KCl (Ambion, AM9640G), 15mM MgCl2 (Ambion, AM9530G), 1mM DTT (Sigma, 646563), 1% NP-40 (ThermoFisher, 28324), 100µg/ml CHX, 100U/ml SUPERaseIn RNAse Inhibitor (Ambion, AM2694), 25U/ml TurboDNAse (Ambion, AM2238), and Complete Protease Inhibitor EDTA-free (Sigma, 11836170001) in nuclease-free water (ThermoFisher, 10977015). Lysates were incubated on ice for 5 min and nuclei were removed by centrifugations at 200g, 5min at 4C and 13,000g, 5min at 4C. A 10% aliquot (10µl) of the lysate was harvested and kept in Trizol for total RNA extraction and the remaining lysate volume was adjusted to 400µl with PLB for even rotation with the antibodies and beads for Ribo-IP in the subsequent steps. For each sample, 3µl of anti-HA antibody (Abcam, ab9110) was added and samples were allowed to rotate at 4C for 4h in a tube rotator (ThermoFisher, 88881001) at 20 rpm speed. During rotation, 40µl of Dynabeads™ Protein G (ThermoFisher, 10004D) were prepared by washing two times in ice-cold PLB. After the anti-HA binding step, Dynabeads were added to each sample for an additional 4h rotation at 4C (speed 20rpm). Next, samples were magnetized on the DynaMag-2 magnet (ThermoFisher, 12321D) and washed four times with 500µl of Polysome High Salt Wash Buffer (PHSWB) (25mM Tris-HCl pH 7.5, 300mM KCl, 15mM MgCl2, 1mM DTT, 1% NP-40, 100µg/ml CHX, 100U/ml SUPERaseIn RNAse Inhibitor, Complete Protease Inhibitor EDTA-free). Usually, before the first wash, 10% of the lysate was saved to routinely check for Ribo-IP efficiency between samples by immunoblotting. For RNA extraction, Dynabeads after the final wash were resuspended in 100µl of PHSWB and homogenized in 350µl of RLT buffer containing 40mM freshly added DTT (Qiagen RNeasy micro kit, 74004) by vigorous vortexing. Samples were left at room temperature for 10min before magnetizing the beads on the magnet and proceeding with the RNA extraction following the RNeasy micro kit protocol. At the last step, total RNA was eluted in 11ul of nuclease-free water and RNA integrity and quantity was assessed using the Bioanalyzer 2100 total RNA pico kit. RNA integrity number (RIN) values for all samples were >8.6 (SFig. 2) and next-generation sequencing compatible cDNA libraries were generated according to the SMARTseq v4 Ultra-Low Input kit (Clontech, 634888).

RiboTag immunoprecipitation (Ribo-IP) and protein isolation
Ribo-IP was performed identical to the RNA isolation method (above). After the final wash in PHSWB, Dynabeads were resuspended in 1x LDS sample buffer and heated at 65C for 10min. Samples were magnetized and loaded onto 12% NuPAGE Bis-Tris gel and proteins were separated by SDS-PAGE. Separated proteins were visualized using the Colloidal Blue Staining kit (ThermoFisher, LC6025) and the entire lane corresponding to each sample were excised

Low input sucrose gradient polysome fractionation
Sucrose gradients (15-60%) were prepared in SW55Ti rotor-compatible Ultra-Clear ultracentrifuge tubes (Beckman Coulter, 344057). Briefly, a 2M (68.5%) sucrose solution in nuclease-free water and a 10X sucrose buffer (250mM Tris-HCl pH 7.5, 1.5M KCl, 150mM MgCl2, 10mM DTT, 1mg/ml CHX, Complete protease inhibitor EDTA-free (2 Tablets per 50ml), 20U/ml SUPERaseIn RNAse Inhibitor) was prepared. Using these two solutions, sucrose solutions with 1X sucrose buffer and different concentrations (60%, 45%, 30%, 15%) were prepared. Each solution was added from the bottom of the tubes in the following order and quantity (60%: 750µl, 45%: 1.5ml, 30%: 1.5ml, 15%: 750µl). For each addition, tubes were kept in the -80C for at least 15min to freeze the sucrose solutions before adding the next sucrose solution. All tubes were sealed with parafilm and kept at -80C forever. Tubes were allowed to thaw at 4C for 12-16h before the fractionation. Samples were prepared in PLB buffer similar to the Ribo-IP method and total RNA was quantified using RiboGreen in the low-range assay (1ng/mL – 50ng/mL). All samples were adjusted with PLB to at least 500ng of total RNA and layered carefully on top of thawed sucrose gradient tubes. Tubes were ultracentrifuged at 35,000rpm in a SW55Ti rotor using the L8-70M ultracentrifuge (acceleration: default, deceleration: 0). Separated lysates were fractionated from top to bottom with an Auto-Densi Flow fractionator (Labconco) with continuous reading of absorbance at 254nm (A254nm) using a UA-6 UV/VIS detector (Teledyne Isco). A total of 16-18 fractions (250-300µl) were fractionated with the Foxy R1 fractionator in 2ml Eppendorf tubes and kept on ice. For digital conversion of the A254nm signal, we attached the LabQuest Mini data-collection interface (Vernier, LQ-MINI) to the UA-6 detector. For RNA extraction, we used 3 volumes of Trizol LS to each sucrose fractions and vortexed vigorously. Each fractions were spiked-in with *in vitro* transcribed firefly Luciferase RNA (uncapped) to assess RNA extraction efficiency between fractions. Samples were either kept in -80C at this point or proceeded with standard Trizol-mediated RNA extraction protocol or the Direct-zol 96 kit (Zymo Research, R2054) was used following the on-column DNA digestion protocol. Total RNA quantity from each fraction was measured using RiboGreen in the low-range assay to generate ribosome traces. The Bioanalyzer 2100 RNA pico kit was also used to assess the starting point of the intact 80S monosome peak (containing both 28S and 18S rRNA bands). Equal volumes of RNA from fractions corresponding to polysomes (3 fractions after the 80S monosome peak) were pooled and digested with RQ1 DNase (Promega) and cleaned-up using RNA Clean&Concentrator-5 kit (Zymo Research, R1013). Finally, RNA quantity (RiboGreen) and RNA quality (Bioanalyzer) were measured again before proceeding with cDNA libraries construction using the SMARTseq v4 Ultra-Low Input kit (Clontech).

RNA-seq bioinformatics methods
Reads were aligned by STAR (Dobin et al., 2012) 2.4.2a to GRCm38.91, and genes that were not expressed in at least 20% of samples were filtered out. The reads were TPM normalized and modeled using the software LIMMA[(Ritchie et al., 2015)]. The LIMMA model included a variable for each of the four

mice and categorized samples by fractionation (input, polysomal, or sub-polysomal), whether they were stimulated, and their treatment (HHT, T-regulatory cells, or none). Combinations of those conditions resulted in 12 categories total. Translation efficiency was computed as the log fold-change in TPM between the polysomal and sub-polysomal fraction per condition, both at the individual mouse and aggregate levels. Differential translation efficiency was computed in LIMMA as differences between translation efficiencies between conditions. Gene set enrichment statistics were computed from LIMMA-computed log fold-changes and the fgsea (Korotkevich et al., 2021) package.

Antibody neutralization assays

Tregs were sorted from CD45.1 Foxp3-yfp-cre mice as CD4+Foxp3(YFP)+ cells, subjected to labeled with CFSE (Thermo). Indicated number of Tregs were stimulated with anti-CD3 and anti-CD28 antibody-conjugated beads for overnight prior to suppression assay. Naïve CD4+T cells were isolated from WT mice or IL-10Rb-deficient mice using Mojo sort (biolegend), followed by staining with cell trace blue (Thermo). Indicated number of naïve T cells was added into the stimulated Treg cells for suppression assay. In some experiments, those naïve T cells were pre-incubated with 10 mg/mL of neutralizing antibody for IL-2R (3C7)(Biolegend), TGF-β (1D11)(BioXcell) and/or IL-10Rb (1B1.3A)(BioXcell) for 30 min before adding Tregs and stimulation beads.

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Figure Legends

Figure 1. Tregs suppress global protein synthesis in CD4 Teff cells. (A) Purified Foxp3-CD4+ Teff cells were stimulated with anti-CD3/CD28 beads with or without indicated ratio of FACS sorted Foxp3-YFP+ Tregs from *Foxp3*$^{YFP\text{-}Cre}$ mice (Treg:Teff = 1:2). For cultures with CD4 Teff cells only, cell number was doubled to match total cell number in the culture (2x). Cells were pulsed with puromycin (PMY) for 15min before harvest and intracellular staining for incorporated puromycin using a fluorophore-conjugated anti-PMY antibody. (B) PMY signal and median fluorescence intensity (MFI) was quantified. Cycloheximide (CHX) was added to 24h stimulated CD4 Teff cells 5min prior to PMY pulse to act as a negative control and set the baseline protein synthesis signal. (C) Purified Foxp3-CD4+ Teff cells were stimulated with anti-CD3/CD28 beads with or without indicated ratio of FACS sorted Foxp3-YFP+ Tregs from *Foxp3*$^{YFP\text{-}Cre}$ mice (Treg:Teff = 1:1) for indicated times, followed by analysis of PMY incorporation as (A). (D) *In vitro*-expanded Tregs from a single donor, and Teff (defined as $CD4^{+}CD45RA^{+}$) from 5 individual healthy donors were used for human Treg suppression assay. Cells were co-cultured and stimulated with anti-CD3/CD28 beads for 24h and PMY staining was performed as in (A). (E) *Foxp3-DTR* mice were injected with diphtheria toxin (DT) for two consecutive days and splenocytes were harvested on day 2. Protein synthesis in CD4+ T cells was measured by *ex vivo* puromycin pulsing of splenocytes and gating on CD4+ T cells. (F) CD4 Teff cells stimulated in the absence and presence of Tregs were identified by congenic markers and intracellular signaling molecules were assessed by phospho-flow cytometry. One-way ANOVA was applied for all comparisons. $*P<0.05$, $**P<0.01$, $***P<0.001$. All experiments were repeated at least twice.

Figure 2. SPEED analysis as a novel polysome profiling approach. (A) Polysome profiling approach using sucrose gradients to physically stratify cellular cytosolic lysates. Classically, continuous absorbance at 254nm (A254nm) is used to assess monosome and polysome positions. SPEED utilizes analysis of quantity and quality of total RNA from each fraction making it amenable for ultra-low biological input that is below the detection limit of A254nm reading. (B) Indicated numbers of bulk CD4 T cells were unstimulated or stimulated with anti-CD3/CD28 beads for 24h and subjected for classical polysome profiling using A254nm reading to obtain polysome traces. (C) Same samples from (B) but equivalent to 500,000-1 million activated CD4 T cells and 2-3 million unstimulated CD4 T cells were subjected for polysome fractionation. No A254nm traces were obtained. Total RNA was extracted from each sucrose fractions and quantified to plot total RNA percent distribution across fractions. Total RNA quality was analyzed using Bioanalyzer (bottom Bioanalyzer results; only fractions #1-11 were analyzed since each RNA pico chip can analyze 11 samples at a time). (D) Equal volume of RNA from each fraction was reverse transcribed into cDNA and *Actb* mRNA levels were quantified using qPCR. The percent *Actb* mRNA across fractions was quantified and plotted. Red dashed line indicates fraction #12 in each sample.

Figure 3. Tregs specifically suppress mRNAs related to the translational machinery via active translation control. (A) Scheme to capture polysome and subpolysome-associated mRNAs for next-generation sequencing. (B) Translation efficiency (TE) was calculated as polysomal/subpolysomal enrichment for four conditions; 1) Unstimulated CD4 Teff cells, 2) 24h stimulated CD4 Teff cells, 3) 24h stimulated CD4 Teff cells co-cultured with Tregs, 4) 24h stimulated CD4 Teff cells treated with harringtonine for 30min. Histograms of $\log_2$ translation efficiency (TE) for all genes were computed for the four conditions. Genes whose $\log_2$ TE were different than 0 at ≤ 5% false-discovery rate (FDR) are highlighted in red. (C) Heatmap of TE in stimulated CD4 Teff cells and stimulated CD4 Teff cells co-cultured with Tregs. (D) Volcano plot of stimulated CD4 Teff cells compared to unstimulated CD4 Teff cells. 3D. Volcano plot for the differential TE of genes across stimulated and unstimulated samples. Genes. Genes whose $\log_2$ TE were different than 0 at ≤ 5% false-discovery rate (FDR) are highlighted in red. Row-normalized $\log_2$ TE for genes whose $\log_2$ TE were significantly different at ≤ 5% FDR between stimulated and unstimulated samples across all four mice. Genes highlighted in red are members of the Reactome translation pathway. (E) Barcode plot ranks genes by their log fold-change of TE between Treg-exposed stimulated cells and stimulated cells. Genes highlighted in black are genes within the Reactome Translation gene set identified to have increased their TE in the stimulated cells vs. unstimulated cell comparison. P-values are computed using the fgsea(1) package that implements Gene Set Enrichment Analysis (GSEA) statistics.

Figure 4. Tregs suppress translation of TOP motif containing mRNAs. (A) Motif discovery of Treg-regulated mRNAs enriches for TOP motif. (B) mRNAs regulated by Tregs in Fig. 3 were validated using SPEED polysome qPCR. Actb mRNA served as a control. The above experiments were repeated twice.

Figure 5. IL-10 from Tregs block protein synthesis in T effector cells. (A-D) CD4 Teff cells were harvested with or without anti-CD3/CD28 beads in the presence or absence of Tregs for 24 h, subjected for analyzing puromycin incorporation and phosphorylation of S6 and 4E-BP1. (A-C) Indicated recombinant cytokines or neutralizing antibodies for cytokines were added into the culture of CD4 Teff

cells stimulated with anti-CD3/CD28 beads. (D) The neutralizing antibody for IL-10 receptor was added to co-culture of CD4 Teff cells and Tregs. (E) CD4 Teff cells were isolated from WT mice (black circle) and IL-10Rb-deficient mice (red circle) were harvested with or without anti-CD3/CD28 beads in the presence or absence of Tregs for 24hr, subjected for analyzing puromycin incorporation and phosphorylation of S6 and 4E-BP1. One-way ANOVA was applied for all comparisons. *$P<0.05$, **$P<0.01$, ***$P<0.001$. The experiments were repeated at least twice (A-D) and once (E).

Figure 6. Acute inflammation due to Treg deficiency rescued by an mRNA translation inhibitor. (A-C) Mouse naïve CD4+ T cells were isolated from spleen of naïve C57BL/6 mice, subjected to CFSE. The cells were stimulated with anti-mouse CD3 and anti-mouse CD28-conjugated beads in the presence of indicated concentration of Roc A, FK506, or vehicle indicated as αCD3/CD28 in A, Stim in B, or without beads indicated as Media in A for 24hr (A, C) or 72hr (B). The cells were stained for anti-CD4 (A and C) or anti-CD4 and anti-CD25 (B) for flowcytometric analysis. The data of frequency of CD25 in A was summarized in C. murine IL-2 was assayed by ELISA from supernatants collected after 24 hours after stimulation (C). (D) Incorporation of puromycin analyzed by flowcytometry post-Roc A treatment. (E-G) Foxp3-DTR mice were treated with diphtheria toxin (DT) every other day and 0.2 mg/kg of Roc A everyday started from 2 days before DT treatment, followed by isolation of splenocytes at day 8. The cells were stained with anti-CD4, anti-TCR-β, and anti-CD44 followed by restimulation with PMA and Ionophore for 6 h. The cells were stained by anti-IL-17A and anti-IFN-γ, followed by flowcytometric analysis (E). The bodyweight of mice at day 8 (F). The number of Th1 cells (IFN- γ +CD44hiCD4+TCR-β+) and pathogenic Th17 cells (IFN-γ+IL-17A+CD44^{hi}CD4+TCR-β+) (G). (H) Human naïve CD4+ T cells isolated from frozen PBMC, subjected to CFSE. The cells were stimulated with anti-human CD3 and anti-human CD28-conjugated beads in the presence of 10nM of Roc A or vehicle or without the beads indicated as unstim. After 3 days, cells were stained with anti-human CD25 and anti-CD4, subjected to flow cytometry for checking the frequency of divided cells and CD25+ cells. (I) Human memory Th1 cells isolated as CD4+CD25-CD45RA-CXCR3+CCR6- cells from frozen PBMC were incubated in the presence of 10 nM of Roc A or vehicle for 2 days, followed by restimulation with PMA and Ionophore for 6 h. The cells were stained by anti-CD4, subjected to cytoplasmic staining for IFNγ and IL-13. The frequency of IFN-γ+ cells is summarized in the dot plots. One-way ANOVA was used for C, D and H; Two-tailed *t*-test (unpaired) was applied for F, G and I. *$P<0.05$, **$P<0.01$, ***$P<0.001$. The above experiments were repeated at least twice.

Figure 7. A model depicting Treg mediated disruption of signaling of mTORC1 that leads to the suppression of protein synthesis of mRNAs enriched for TOP motifs.

Supplementary Figures

Supplementary Figure 1. (A) *In vitro* Treg suppression assay using different doses of purified Foxp3-YFP+ Tregs and co-culturing with CFSE labeled YFP-CD4+ Teff cells. Data is presented in two ways. Numerating the undivided CFSE peak to show percent undivided cells. Percent suppression was calculated by measuring cells that have divided at least once. (B) CD62L expression was measured in unstimulated and stimulated CD4 Teff cells with or without Tregs at 24h post stimulation. (C) Bulk CD4+ T cells were purified on day 2 from control C57/B6 mice and Foxp3-DTR mice treated with DT for two consecutive days. Cells were labeled with CFSE and activated using anti-CD3/CD28 beads. Cell size (forward scatter) and proliferation (CFSE dilution) was measured at 72h post stimulation. (D) CD4 Teff cells from Nur77-GFP mice labeled with CellTraceViolet were either activated with anti-CD3/CD28 beads alone or with titrating amounts of sorted Tregs. Some conditions received increasing concentrations of RocA for eIF4A inhibition. Proximal TCR signaling was assessed using GFP in CD4 Teff cells by gating on CellTraceViolet+ population. Each datapoint represents a biological replicate (CD4 Teff cells from individual Nur77-GFP mice).

Supplementary Figure 2. (A) Non-recombined RiboTag mice (Rpl22Tag) and CMV-Cre mediated whole-body recombined RiboTag mice ($Rpl22^{HA/+}$) were compared for the thymic cellularity and splenic CD4, CD8 T cells and Foxp3+CD25+ Tregs. (B) Bulk CD4 T cells from T-Ribo mice were activated and subjected for fractionation on a 15-60% sucrose gradient. Proteins were precipitated from sucrose fractions and immunoblotted using anti-HA, anti-RPS6 and anti-Tubulin. Samples from $RPL22^{\Delta T/+}$ show highly specific enrichment for ribosomal proteins (9- 52 kDa range) as well as high molecular weight proteins that represent ribosome- associated proteins (confirmed by MS analysis). Absolute spectral counts for all RPs identified (68 out of 80). (C) Purified CD4+ T cell samples from $RPL22^{\Delta T/+}$ were subjected for Ribo-IP along with control T cells to show background immunoprecipitation.

Supplementary Figure 3. (A) CD45.1 congenically labeled Foxp3-YFP+ Tregs and Foxp3-YFP- Teff cells were co-cultured with CD45.2 CD4 Teff cells from T-Ribo mice. After 24h of stimulation, CD45.1+ cells were positively selected using magnetic activated cell sorting (MACS) using CD45.1-FITC microbeads. Pre- and post-MACS CD45.2+ percentage is plotted for all experiments. (B) Suppression of CD4 Teff cells by Tregs was measured using CFSE 72h after stimulation. (C) Specificity of Ribo-IP was tested using CD4 Teff cells from T-Ribo mice and *Cre*- control mice. After Ribo-IP, captured total RNA was measured using Bioanalyzer (RNA pico kit).

Supplementary Figure 4. Treg modulated translatome in CD4 Teff cells using the RiboTag approach. (A) Schematic of the RiboTag model. Exon4 of *eL22* is replaced by a HA-epitope sequence containing downstream Exon4 in a *Cre*-dependent manner (*Cd4Cre* was used: T-Ribo mice). Bulk CD4 T cells were purified from *Cd4Cre* positive and negative mice and immunoblotted for anti-HA. (B) Schematic of the translatome capture using the RiboTag for RNA-seq (C) Principal component analysis of Ribo-IP RNA samples and total input RNA samples. (D) Ribosome load (RL: Ribo-IP signal/Total input signal) was analyzed by calculating Ribo-IP mRNA signals to its corresponding total input mRNA signal in each of the condition. Density of RLs are plotted as a histogram with low RL mRNAs (number in red bracket) and high RL mRNAs (number in blue bracket). 1.5-fold cutoff was used to highlight mRNAs with

significant RLs (red open bars) in relation to non-significant mRNAs (gray filled bars). Number of mRNAs with differential ribosome load are quantified as a histogram further supporting the idea that the RiboTag approach mainly discovers ribosome de-enriched mRNAs (negative ribosome load) as all ribosomes are captured. (D) Ribosome load was calculated for each condition and mRNAs with significant changes in ribosome (E)Venn-diagram shows that mRNAs with differential ribosome load for each condition is largely unique with minimal overlap. (F) Heatmap of Log2TMM value of mitochondrial genes and their enrichment in Ribo-IP samples. Mitochondrial genes are translated by mitoribosomes and are completely de-enriched in Ribo-IP, which further validates Ribo-IP capturing cytosolic ribosomes. (G) mRNAs showing further de-enrichment from total ribosomes (Ribo-IP) after Treg co-culture. One-way ANOVA was applied was used for all comparisons between two groups. *$P<0.05$, **$P<0.01$.

Supplementary Figure 5. (A) Scatterplot for each sample of quality control metrics percent alignment and median covariance of coverage, colored by fraction and treatment. Percent alignment refers to the percent of reads that can be aligned to the reference genome. Median covariance of coverage refers to the median coefficient of variation (mean/standard deviation) of gene sequence coverage for the 1000 most highly expressed transcripts. All samples pass quality control thresholds of 75% percent alignment and 0.55 median covariance of coverage. (B) Same as panel A, zoomed into the region of the graph where the samples reside. (C) Bar graph of the number of millions of reads sequenced from each sample.

Supplementary Figure 6. (A) Scatterplots showing principal components 1 and 2 for each sample across the four treatment conditions and two fractions. Principal components were computed separately for each treatment condition. (B) Scatterplots showing translation efficiency ($\log_2$ fold-change of gene expression between the polysomal fraction and the subpolysomal fraction) and $-\log_{10}$ false-discovery rate for each stimulation condition. Translation efficiency false-discovery was computed using the software package LIMMA, and the false-discovery rate refers to Benjamini-Hochberg-corrected p-values for the hypotheses for each gene that the $\log_2$ fold-change between the fractions does not equal 0. Genes are colored red if their $\log_2$ translation efficiencies are less than -1 and false-discovery rates are less than or equal to 5%. Genes are colored blue if their $\log_2$ translation efficiencies are greater than 1 and false-discovery rates are greater than or equal to 5%.

Supplementary Figure 7. Mice were treated with 0.5 or 1 mg/kg of Roc A or vehicle everyday for 7 days, followed by isolation of bone marrow and spleen at day 8. (B) Total body weight was monitored everyday post treatment. C. Incorporation of puromycin analyzed by flowcytometry post-Roc A treatment. C-F. Bone marrow cells and splenocytes were stained for neutrophils (Ly6G+), eosinophils (SiglecF+CD11c-), basophils (ckit-CD200R3+FceRI+), Monocyte (CD11b+Ly6c+), naïve CD4+T (CD4+CD3+CD44lowFoxp3-), memory CD4+T (CD4+CD3+CD44hiFoxp3-), Treg (CD4+CD3+Foxp3+), CD8T(CD8+CD3+) and B (CD19+) cells in indicated fractions, subjected to flow cytometry.

Supplementary Table 1. A list of 149 genes, identified by SPEED, whose TE increased after stimulation at 5% FDR, and the logFC of the TE changes. These genes were examined for *cis*-regulatory elements shared by the 5'UTRs among Treg-sensitive mRNAs.

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Figure 1: Tregs suppress global protein synthesis in CD4 Teff cells.

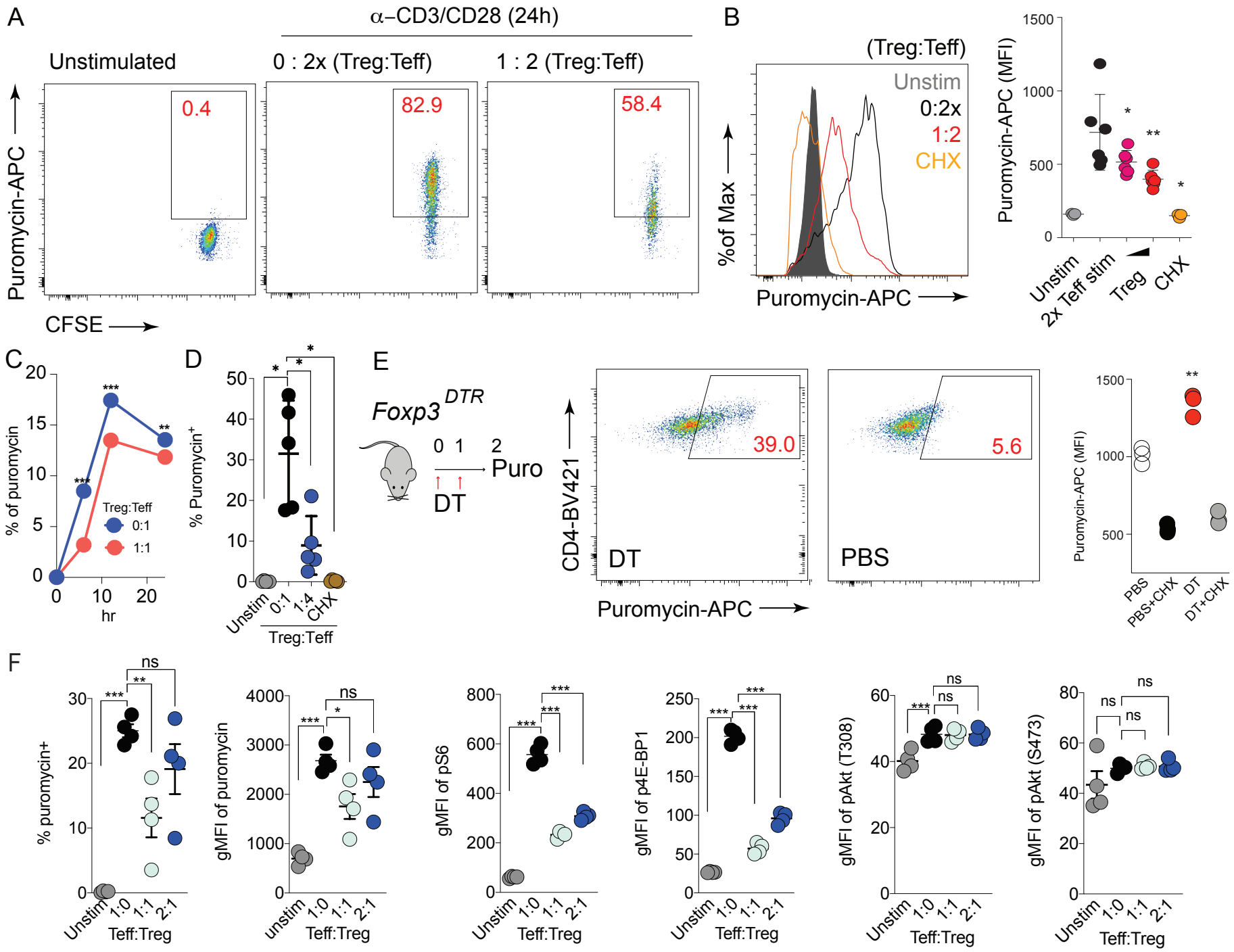


Figure 2: SPEED analysis as a novel polysome profiling approach.

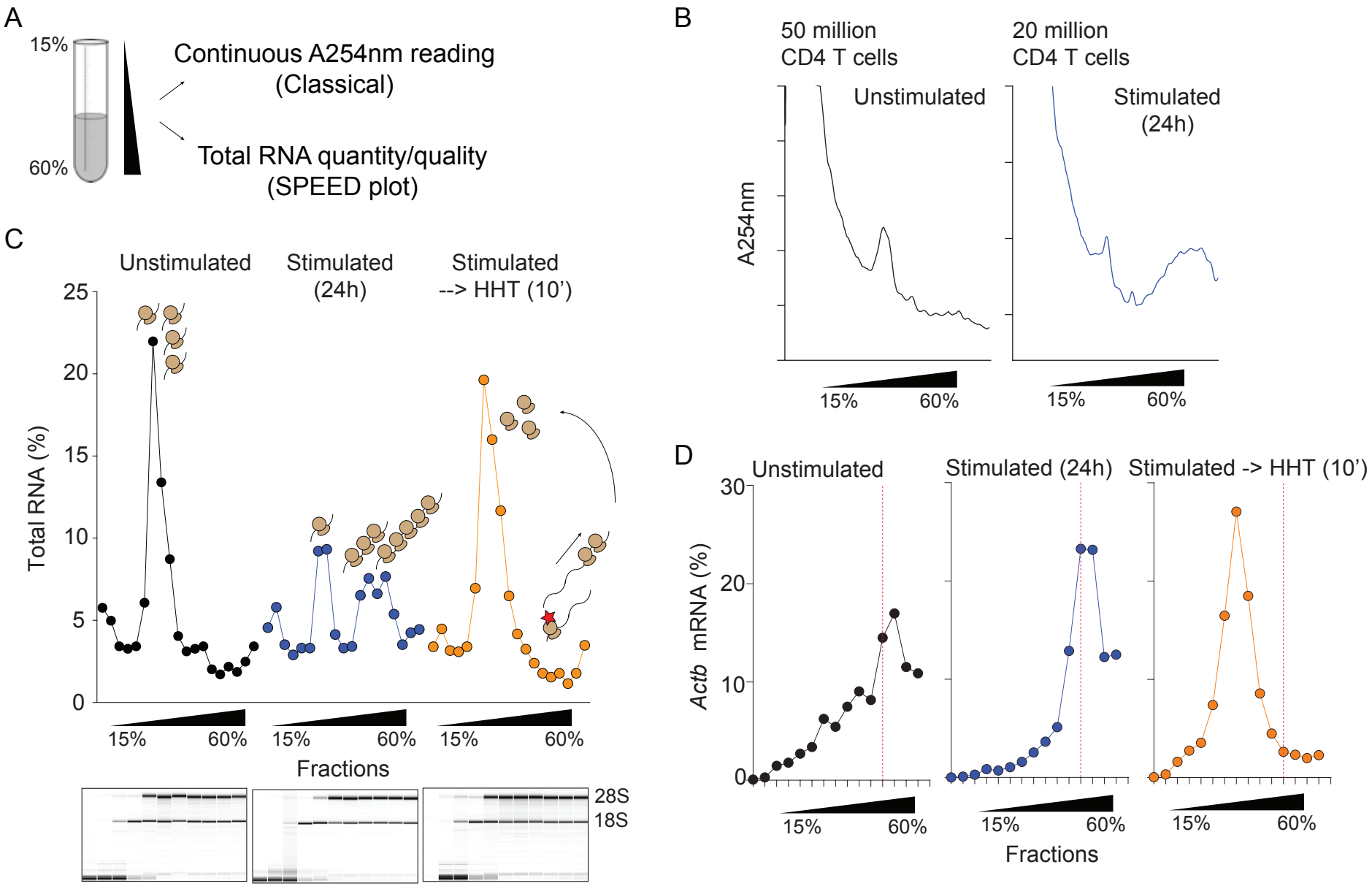
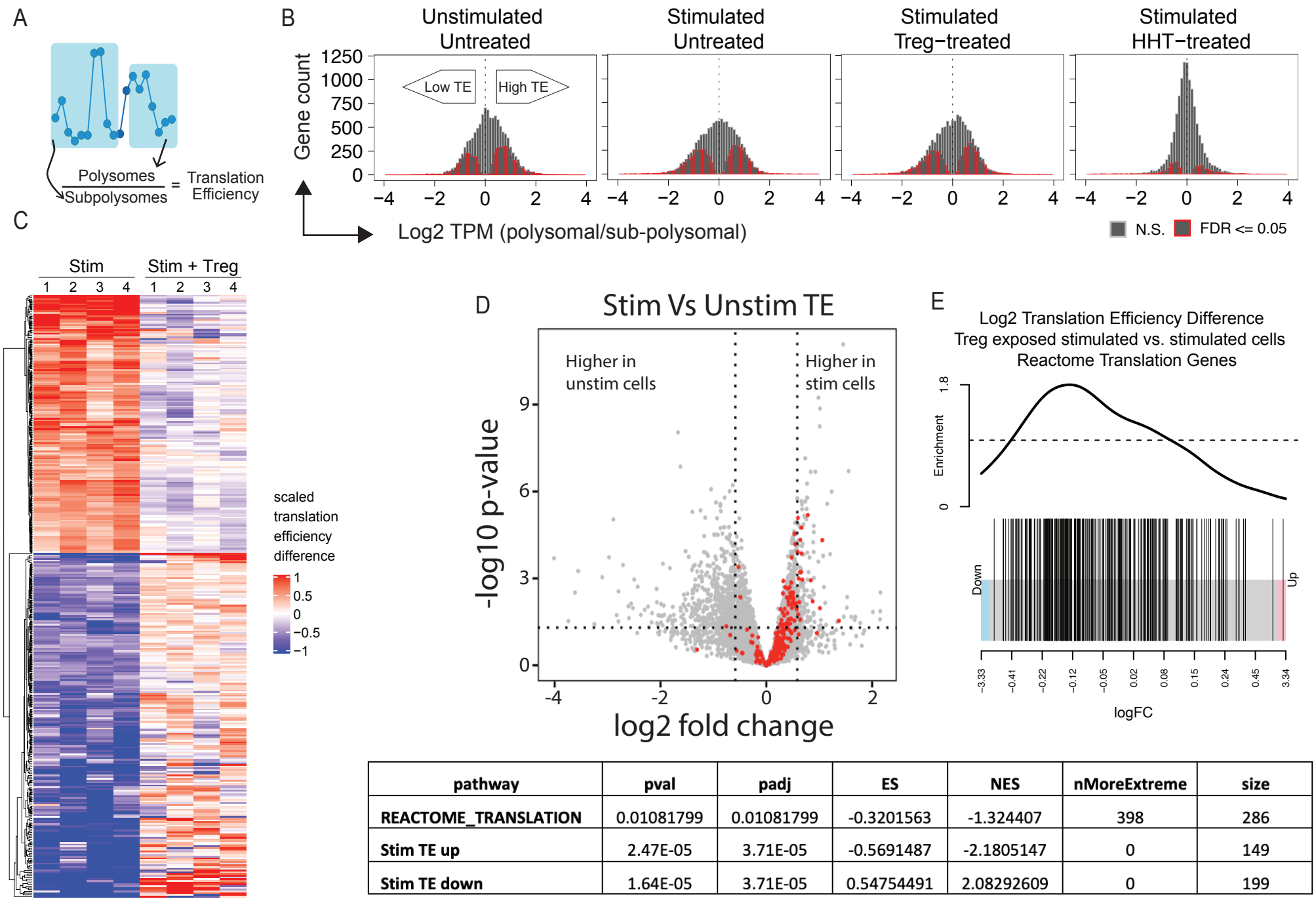


Figure 3: Tregs specifically suppress mRNAs related to the translational machinery via active translation control.



pathway	pval	padj	ES	NES	nMoreExtreme	size
REACTOME_TRANSLATION	0.01081799	0.01081799	-0.3201563	-1.324407	398	286
Stim TE up	2.47E-05	3.71E-05	-0.5691487	-2.1805147	0	149
Stim TE down	1.64E-05	3.71E-05	0.54754491	2.08292609	0	199

Figure 4. Tregs suppress translation of TOP motif containing mRNAs.

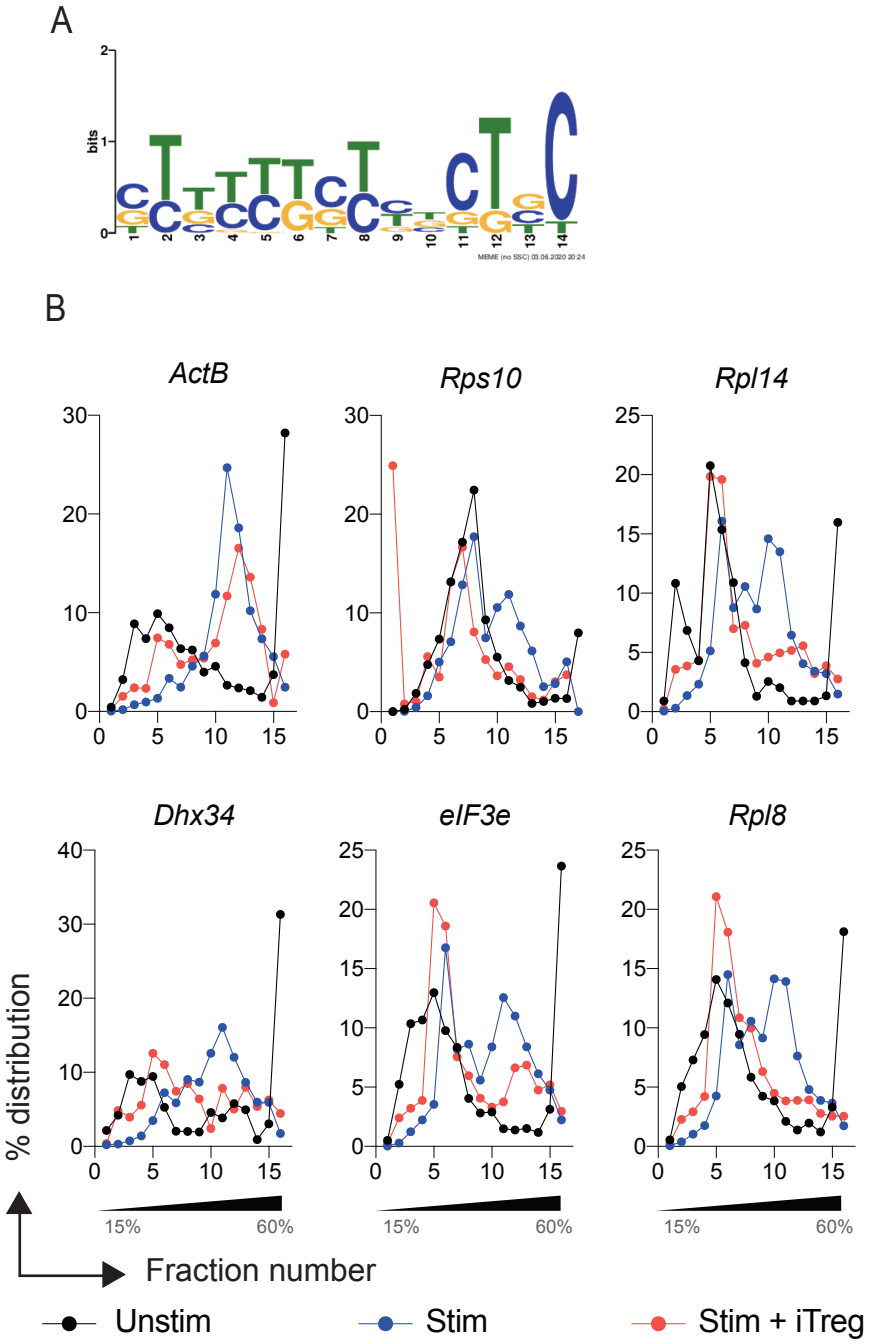


Figure 5. Tregs suppress translation of TOP motif containing mRNAs.

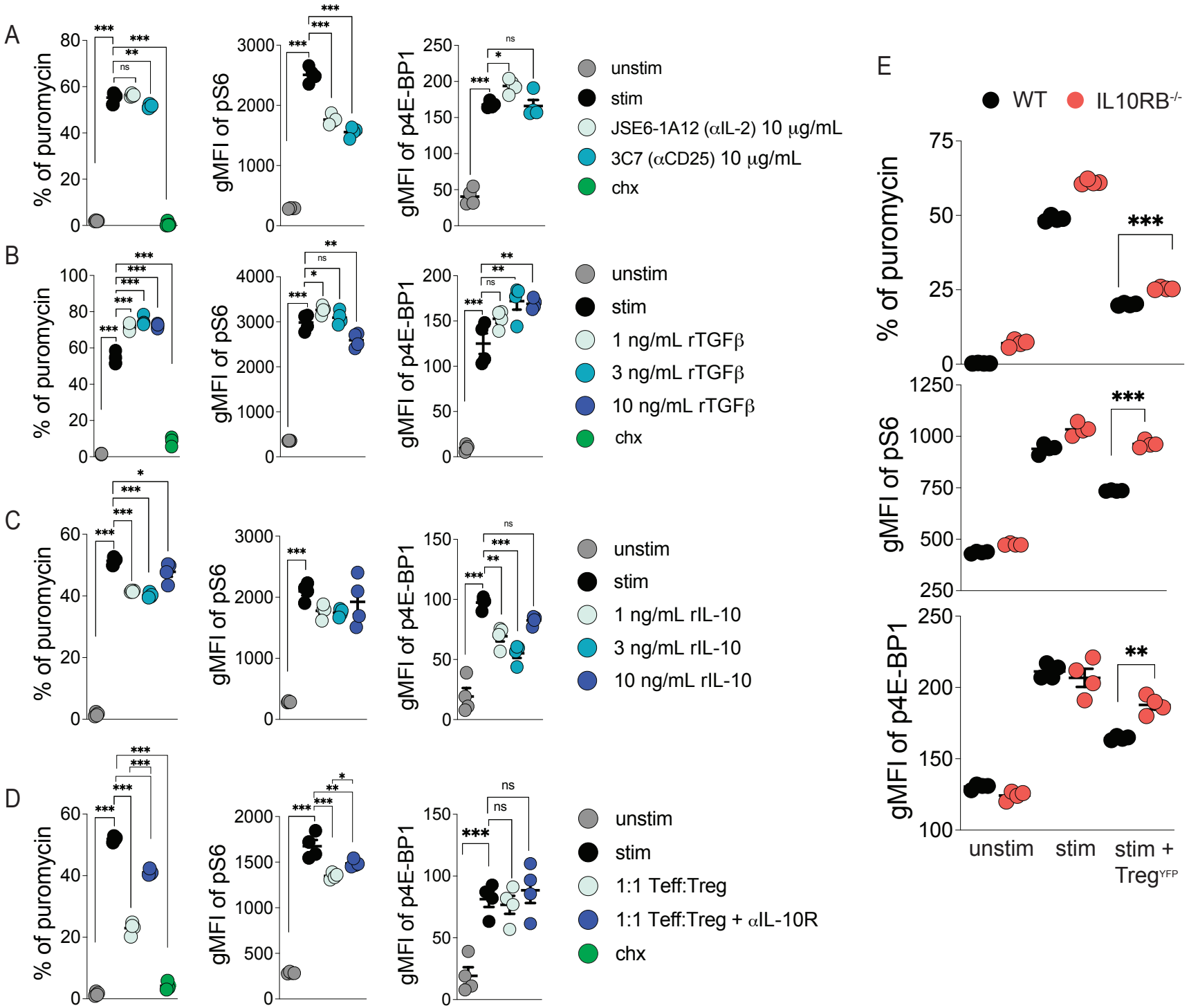


Figure 6. Acute inflammation due to Treg deficiency rescued by an mRNA translation inhibitor.

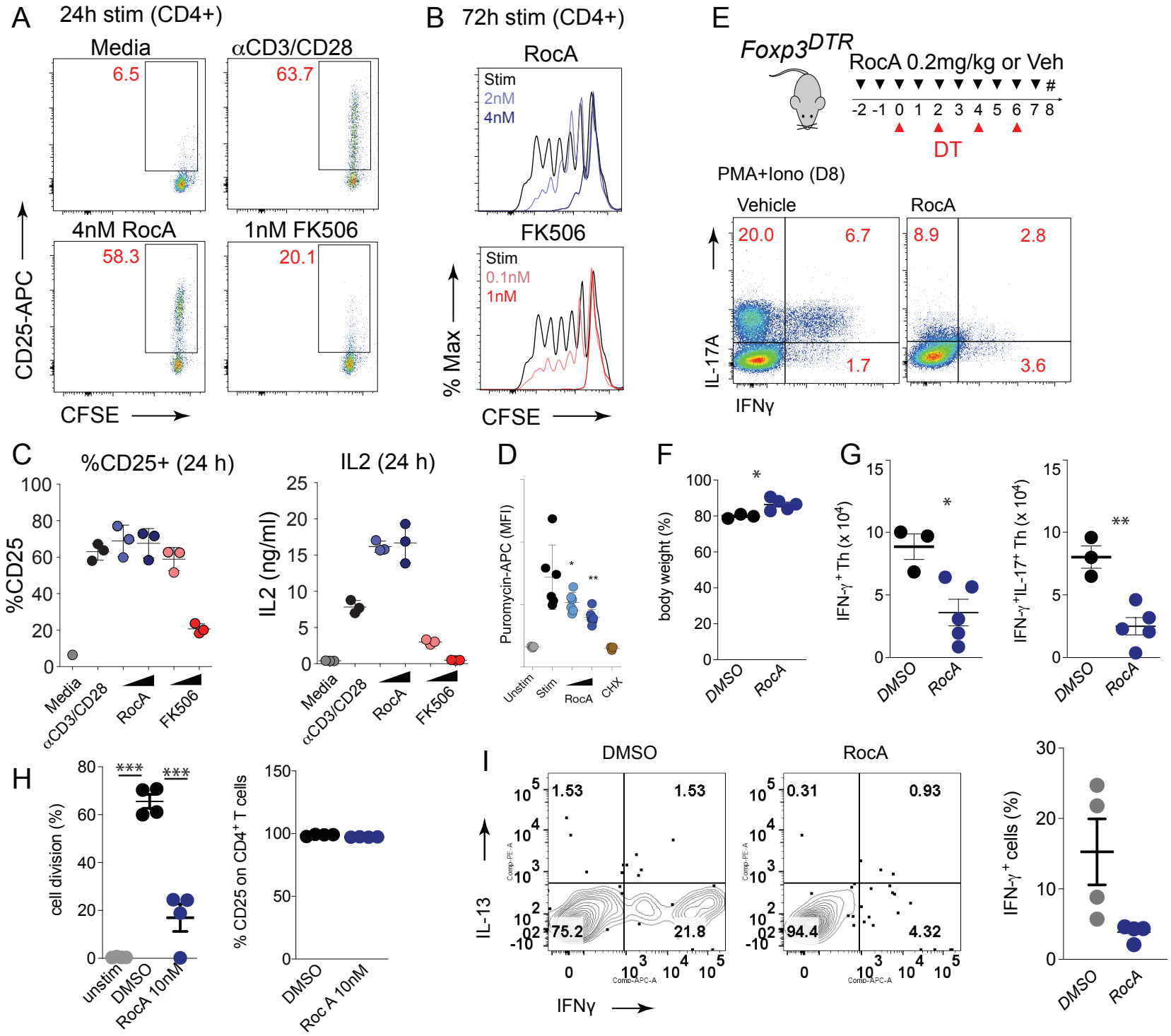
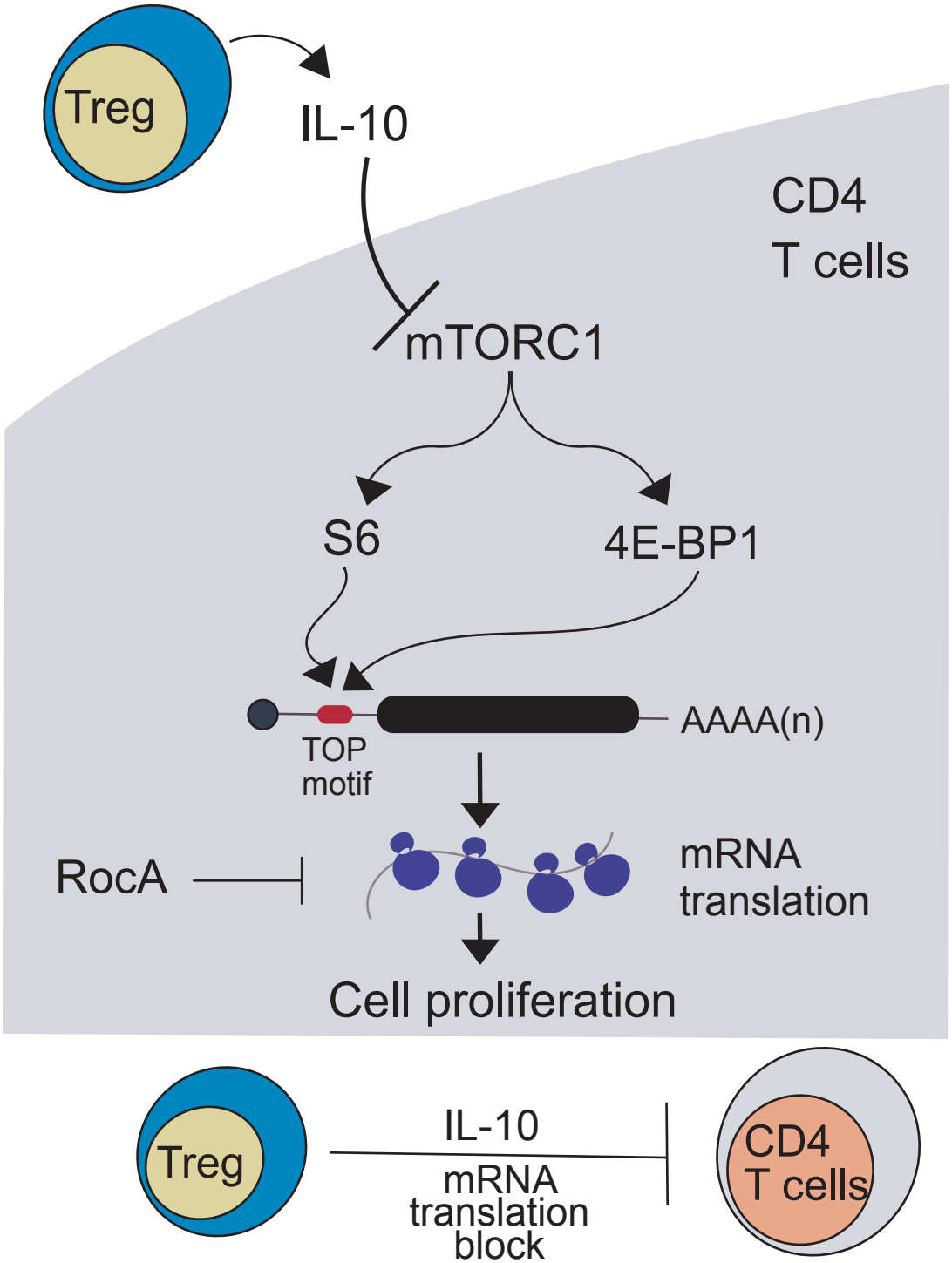
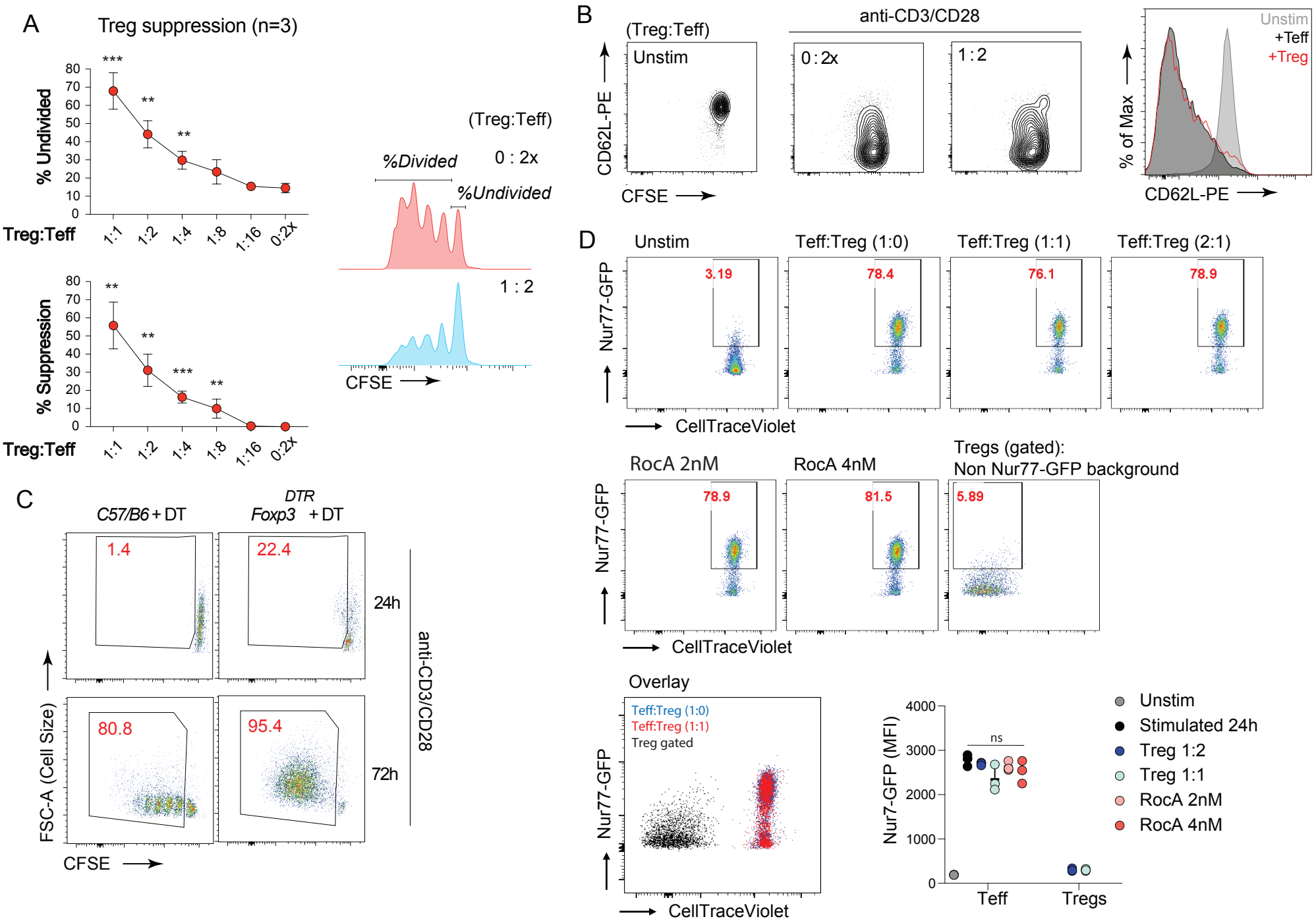


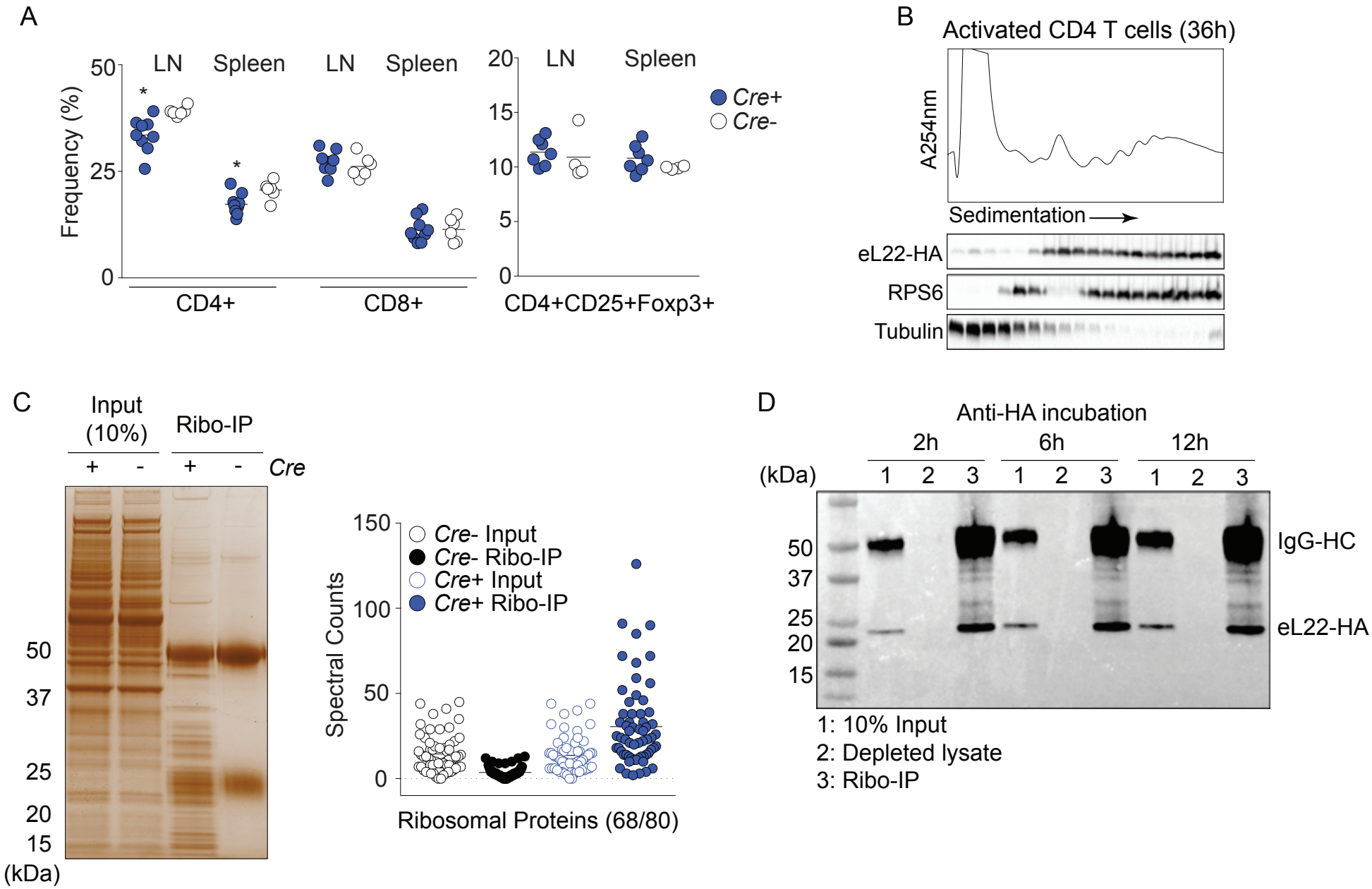
Figure 7. A model depicting Treg mediated disruption of signaling of mTORC1 that leads to the suppression of protein synthesis of mRNAs enriched for TOP motifs.



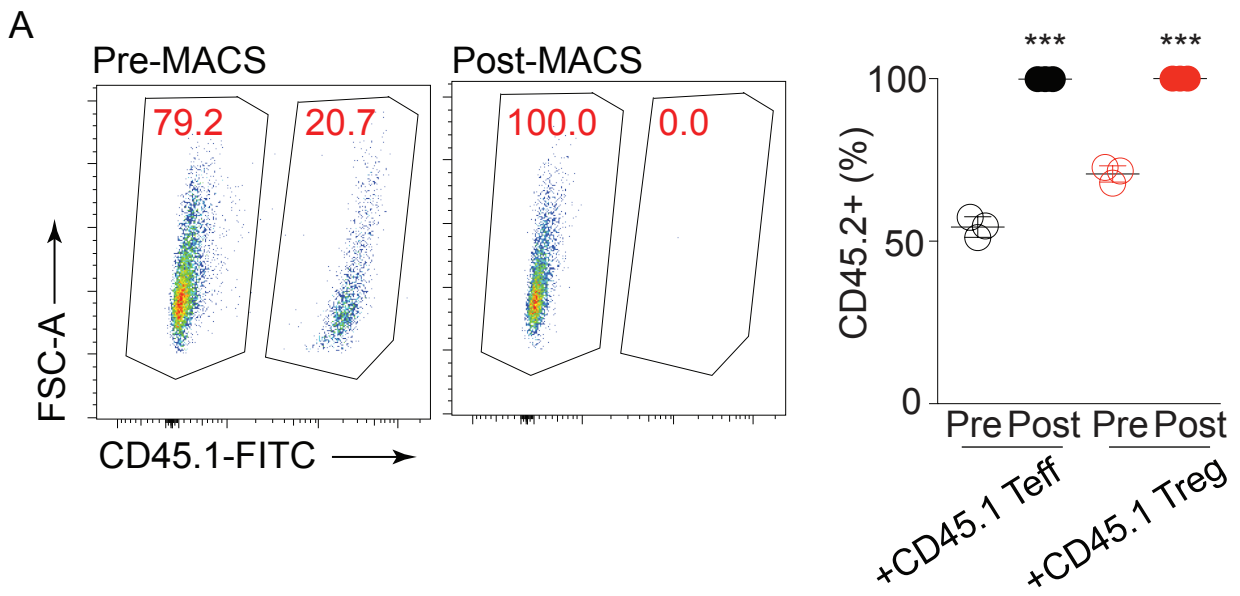
S Fig 1

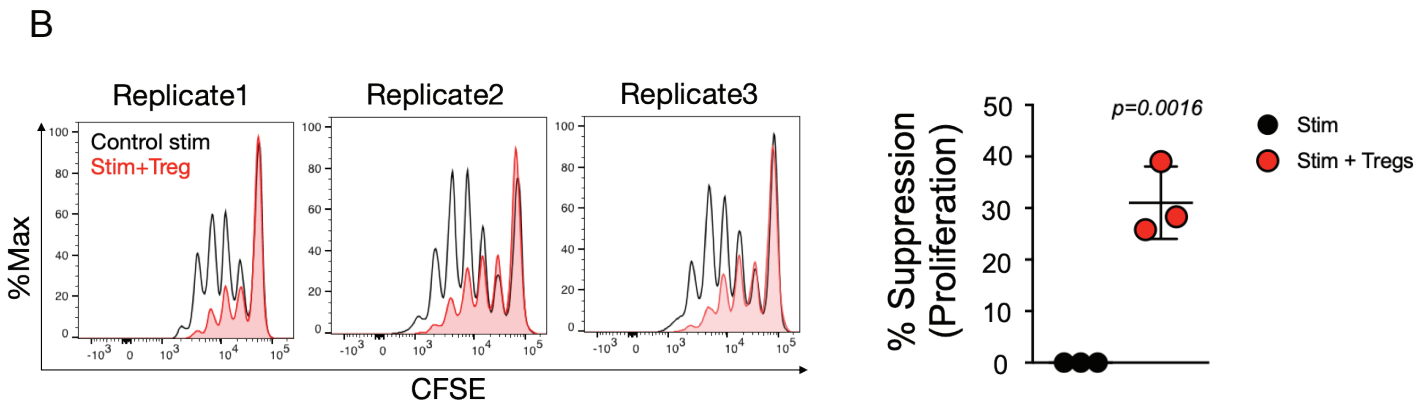

A
Treg suppression (n=3)
% Undivided
Treg:Teff
1:1
1:2
1:4
1:8
1:16
0:2x
% Suppression
(Treg:Teff)
%Divided
0 : 2x
%Undivided
1 : 2
CFSE
B
(Treg:Teff)
anti-CD3/CD28
Unstim
0 : 2x
1 : 2
CD62L-PE
CFSE
% of Max
Unstim
+Teff
+Treg
C
C57/B6 + DT
DTR
Foxp3 + DT
1.4
22.4
24h
80.8
95.4
72h
anti-CD3/CD28
FSC-A (Cell Size)
D
Unstim
Teff:Treg (1:0)
Teff:Treg (1:1)
Teff:Treg (2:1)
3.19
78.4
76.1
78.9
Nur77-GFP
CellTraceViolet
RocA 2nM
RocA 4nM
Tregs (gated):
Non Nur77-GFP background
78.9
81.5
5.89
Overlay
Teff:Treg (1:0)
Teff:Treg (1:1)
Treg gated
Nur7-GFP (MFI)
ns
Teff
Tregs
Unstim
Stimulated 24h
Treg 1:2
Treg 1:1
RocA 2nM
RocA 4nM

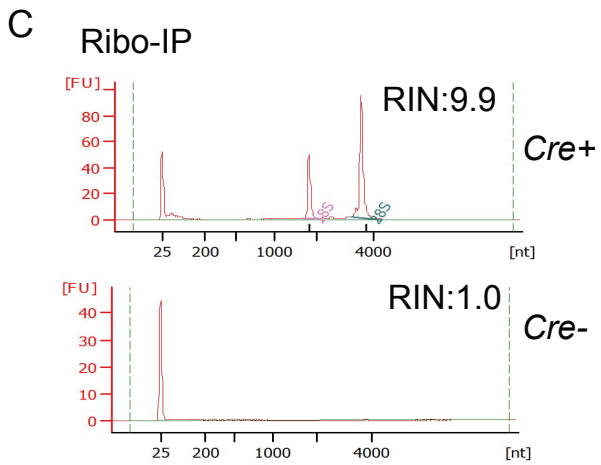
S Fig2


A
LN
Spleen
LN
Spleen
LN
Spleen
Frequency (%)
50
25
0
20
15
10
5
0
*
*
Cre+
Cre-
CD4+
CD8+
CD4+CD25+Foxp3+
B
Activated CD4 T cells (36h)
A254nm
Sedimentation
eL22-HA
RPS6
Tubulin
C
Input (10%)
Ribo-IP
+
-
+
-
Cre
50
37
25
20
15
(kDa)
150
100
50
0
Spectral Counts
Cre- Input
Cre- Ribo-IP
Cre+ Input
Cre+ Ribo-IP
Ribosomal Proteins (68/80)
D
Anti-HA incubation
2h
6h
12h
(kDa)
1
2
3
1
2
3
1
2
3
50
37
25
20
15
IgG-HC
eL22-HA
1: 10% Input
2: Depleted lysate
3: Ribo-IP

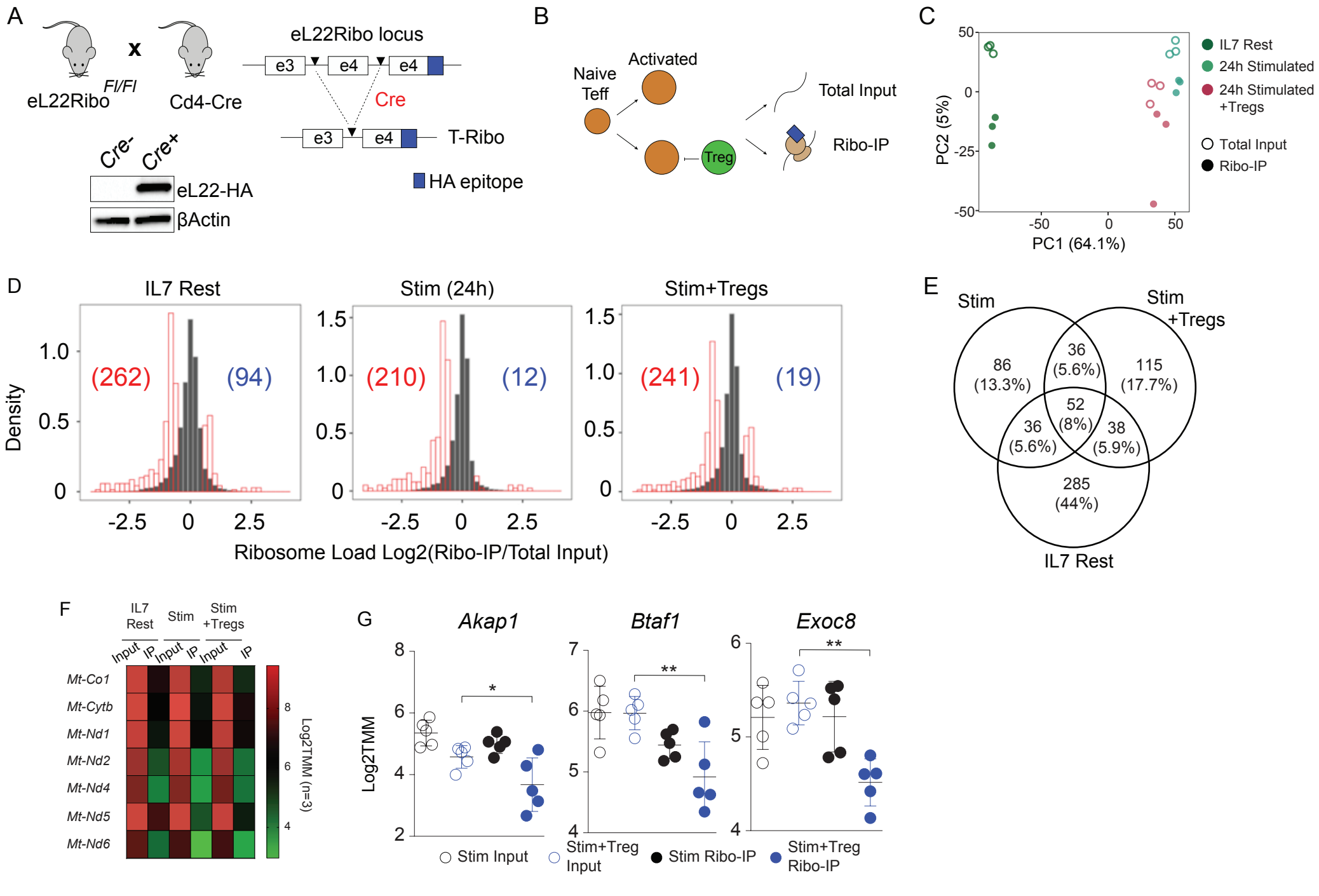
S Fig 3


A
Pre-MACS
Post-MACS
79.2
20.7
100.0
0.0
FSC-A
CD45.1-FITC
CD45.2+ (%)
100
50
0
***
***
Pre Post Pre Post
+CD45.1 Teff
+CD45.1 Treg

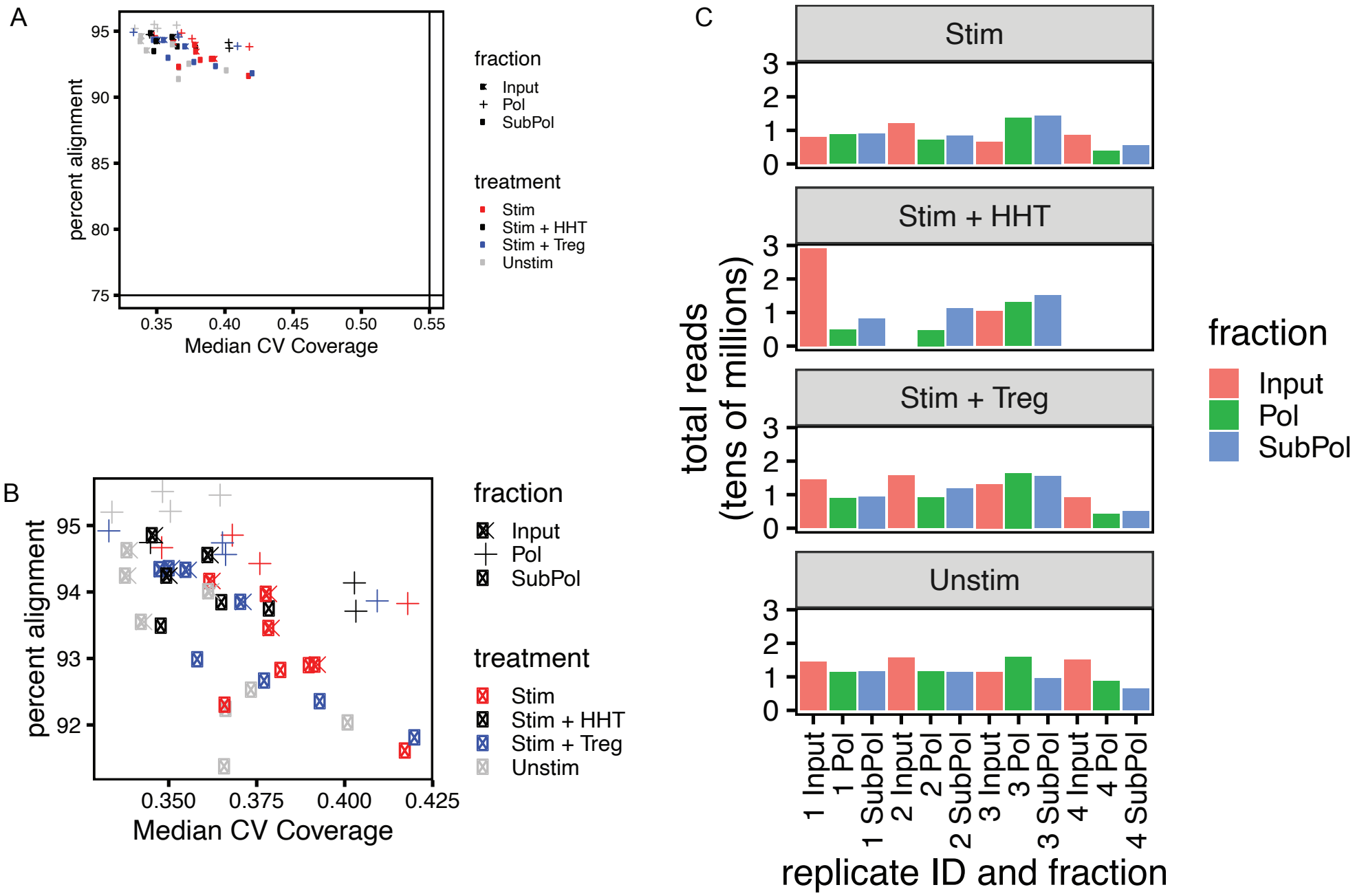

B
Replicate1
Replicate2
Replicate3
Control stim
Stim+Treg
%Max
CFSE
% Suppression (Proliferation)
p=0.0016
Stim
Stim + Tregs


C
Ribo-IP
RIN:9.9
Cre+
RIN:1.0
Cre-
[FU]
[nt]

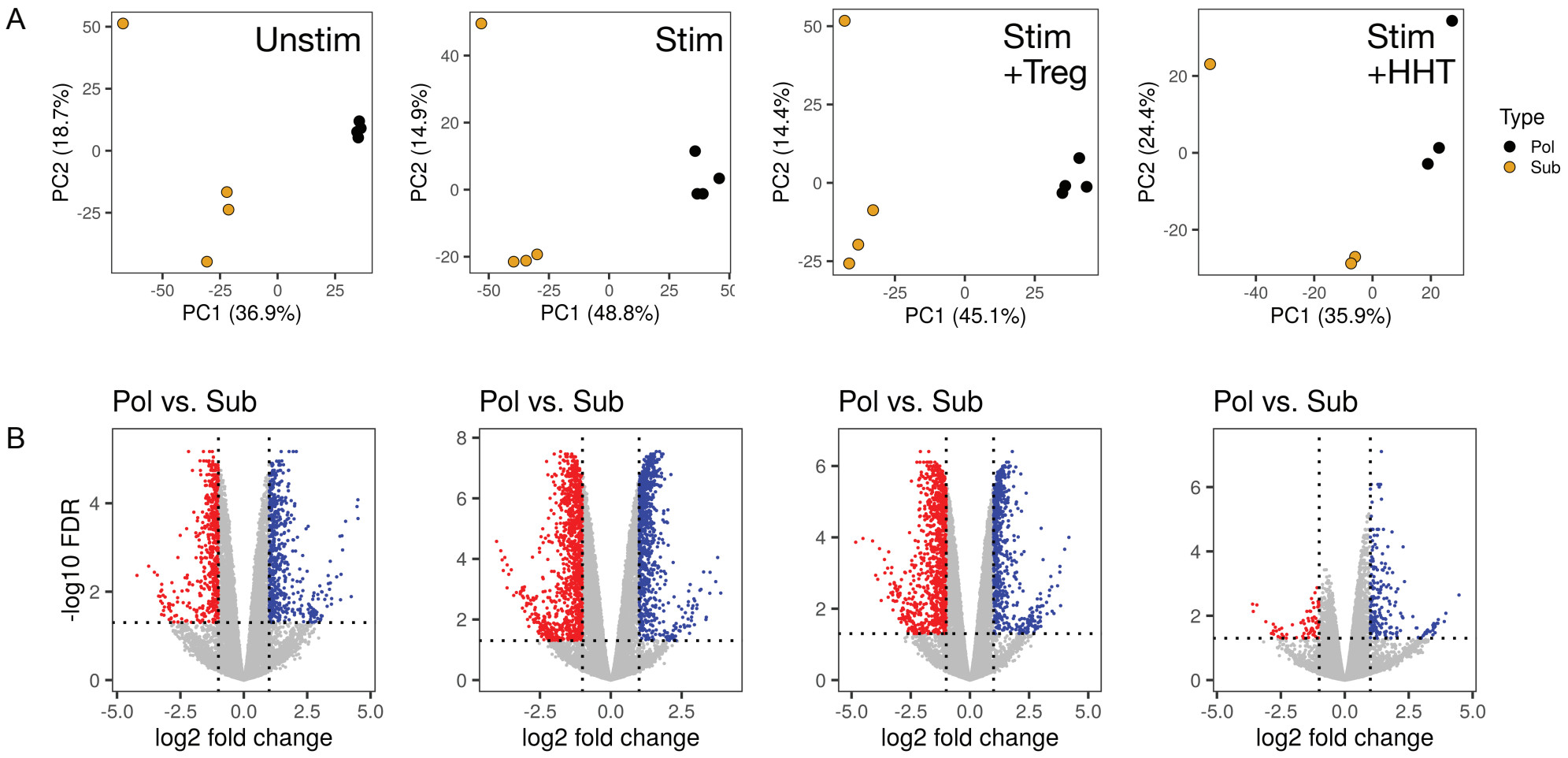
S Fig 4


A
eL22Ribo Fl/Fl
x
Cd4-Cre
eL22Ribo locus
e3
e4
e4
Cre
e3
e4
T-Ribo
HA epitope
Cre-
Cre+
eL22-HA
βActin
B
Naive Teff
Activated
Treg
Total Input
Ribo-IP
C
IL7 Rest
24h Stimulated
24h Stimulated +Tregs
Total Input
Ribo-IP
PC2 (5%)
PC1 (64.1%)
D
IL7 Rest
Stim (24h)
Stim+Tregs
(262)
(94)
(210)
(12)
(241)
(19)
Density
Ribosome Load Log2(Ribo-IP/Total Input)
E
Stim
Stim +Tregs
IL7 Rest
86 (13.3%)
36 (5.6%)
115 (17.7%)
52 (8%)
36 (5.6%)
38 (5.9%)
285 (44%)
F
IL7 Rest
Stim
Stim +Tregs
Input
IP
Mt-Co1
Mt-Cytb
Mt-Nd1
Mt-Nd2
Mt-Nd4
Mt-Nd5
Mt-Nd6
Log2TMM (n=3)
G
Akap1
Btaf1
Exoc8
Log2TMM
Stim Input
Stim+Treg Input
Stim Ribo-IP
Stim+Treg Ribo-IP

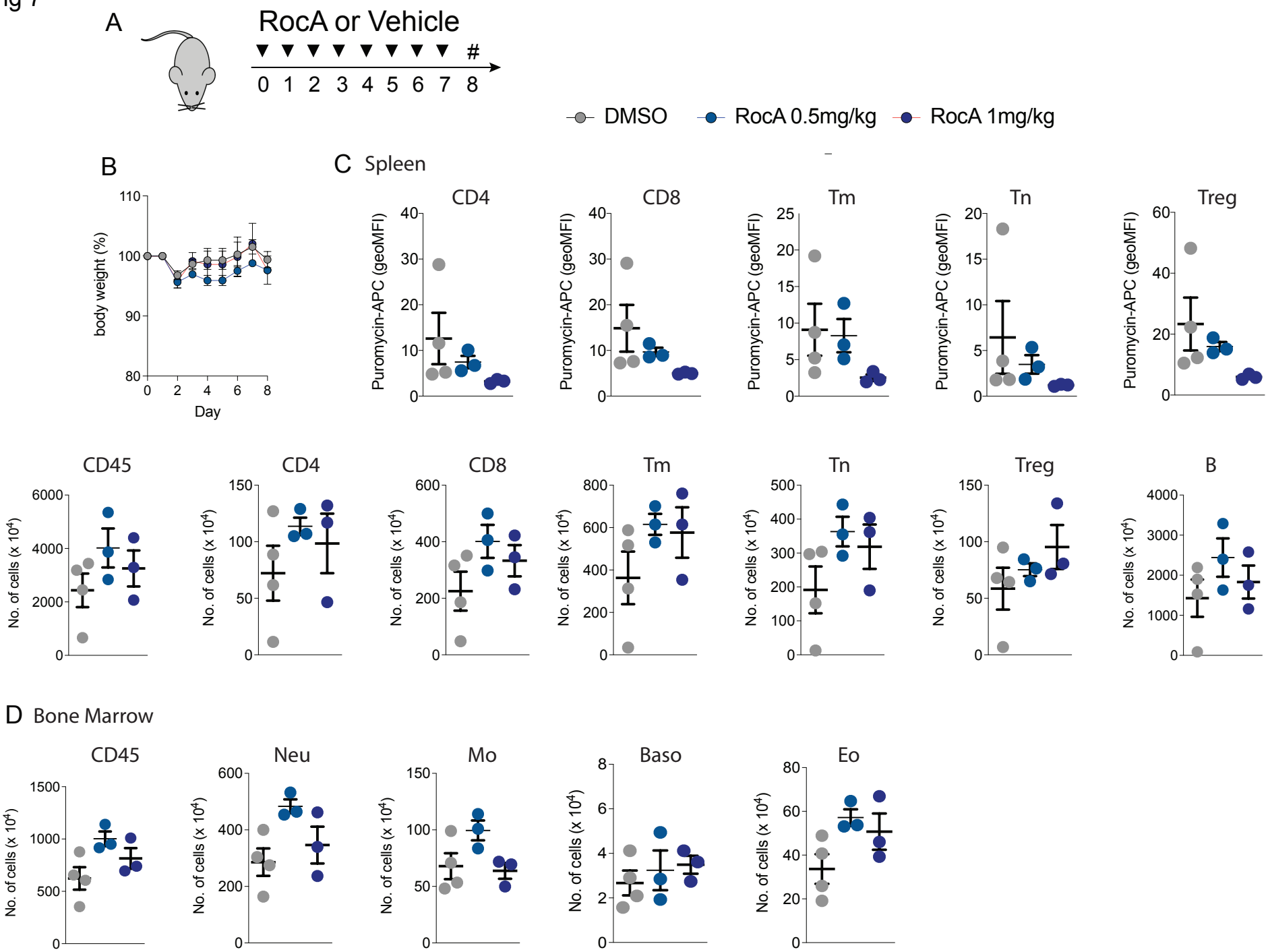
S Fig 5


A
percent alignment
95
90
85
80
75
0.35
0.40
0.45
0.50
0.55
Median CV Coverage
fraction
Input
Pol
SubPol
treatment
Stim
Stim + HHT
Stim + Treg
Unstim
B
percent alignment
95
94
93
92
0.350
0.375
0.400
0.425
Median CV Coverage
fraction
Input
Pol
SubPol
treatment
Stim
Stim + HHT
Stim + Treg
Unstim
C
Stim
Stim + HHT
Stim + Treg
Unstim
total reads
(tens of millions)
0
1
2
3
1 Input
1 Pol
1 SubPol
2 Input
2 Pol
2 SubPol
3 Input
3 Pol
3 SubPol
4 Input
4 Pol
4 SubPol
replicate ID and fraction
fraction
Input
Pol
SubPol

S Fig 6


A
Unstim
Stim
Stim +Treg
Stim +HHT
PC2 (18.7%)
PC1 (36.9%)
PC2 (14.9%)
PC1 (48.8%)
PC2 (14.4%)
PC1 (45.1%)
PC2 (24.4%)
PC1 (35.9%)
Type
Pol
Sub
B
Pol vs. Sub
-log10 FDR
log2 fold change

S Fig 7


A
RocA or Vehicle
0 1 2 3 4 5 6 7 8
#
DMSO
RocA 0.5mg/kg
RocA 1mg/kg
B
body weight (%)
Day
C Spleen
CD4
CD8
Tm
Tn
Treg
Puromycin-APC (geoMFI)
CD45
CD4
CD8
Tm
Tn
Treg
B
No. of cells (x 10^4)
D Bone Marrow
CD45
Neu
Mo
Baso
Eo
No. of cells (x 10^4)

Supplement text 1

Translatome capture by RiboTag.

Mature CD4 Teff cells with an HA-epitope tag on the ribosomal large subunit protein L22 (eL22; known as RiboTag) were generated using *Cd4-Cre* mice (*eL22*$^{Ribo f/f}$*Cd4Cre*: T-Ribo) where exon 4 of eL22 is substituted with an identical exon containing HA epitope sequences (Fig 2A). No obvious defects were observed in steady-state mature T cell frequencies in the spleen and lymph nodes with only a mild reduction in CD4+ T cell frequency (Fig S2A). Purified CD4 T cells from T-Ribo (Cd4-Cre+) expressed eL22-HA protein that co-sedimented with polysome fractions upon activation indicating normal ribosome incorporation (Fig S2B). Immunoprecipitation of eL22-HA (termed Ribo-IP) followed by mass spectrometry also identified most ribosomal proteins (RPs: 68/80) suggesting eL22-HA as a *bona fide* RP assembled as an intact ribosome (Fig S2C). Ribo-IP was highly efficient with complete depletion of eL22-HA with as little as 2h of antibody incubation (Fig S2D).

The RiboTag system allowed profiling of both the steady-state mRNA levels and the ribosome-bound mRNA levels from the same cytosolic lysate from CD4 Teff cells (Fig 2B). CD4 Teff cells from T-Ribo mice were stimulated alone or co-cultured with isolated Tregs and stimulated with anti-CD3/CD28 coated beads for 24h, a timepoint before the onset of proliferation. For resting conditions, we used CD4 T cells cultured in IL-7, which promotes survival but not activation, to match the timepoint for lysate preparation. Cytoplasmic lysates were prepared from pure CD4 Teffs after magnetic activated cell sorting (MACS) to remove either congenically marked CD4 Teff cells or Tregs where we obtained >99.8% purity (Fig S3A). A small aliquot from the lysate (10%) was taken for total input RNA extraction and Ribo-IP was performed on the remaining lysates to extract ribosome-bound mRNA. We also confirmed that Tregs faithfully suppressed CD4 Teff cell proliferation 3 days after stimulation compared to control stimulated Teff cells (Fig S3B). Ribosome-bound mRNA and total RNA input for each condition were subjected to RNA-seq. mRNA pulldown by Ribo-IP was highly specific as control activated CD4 Teff cells from *Cre*- mice (n=2) showed negligible traces of RNA based on Bioanalyzer analysis (Fig S3C). Principal component analysis (PCA) analysis separated the samples by the first component (64.1% of variance) according to their activation status (IL7 rested vs activated) and suppressed CD4 Teff cells separated from activated CD4 Teff cells by the second component (5% of variance) (Fig S4C). Importantly, total RNA input samples were clearly separated from their corresponding Ribo-IP samples by the second component indicating discrepancies in the total RNA input and its corresponding Ribo-IP mRNA. mRNAs of the mitochondrial DNA (mtDNA) are normally translated by mitoribosomes and not eL22 containing cytosolic ribosomes. In all conditions, we observed a significant de-enrichment of mitochondrial derived mRNAs in the Ribo-IP samples, internally validating that the Ribo-IP faithfully captures mRNAs only bound to the cytosolic ribosome (Fig S4F).

As total input samples were distinct from their corresponding Ribo-IP samples, we measured 'ribosome load' (RL) by calculating the ratio of Ribo-IP signal to its total RNA input signal. In all conditions, RL values were significantly biased towards the negative direction indicating the majority of mRNAs have less ribosome occupancy compared to their total input levels (Fig S4D). We attribute this bias to the fact that Ribo-IP signals represent mRNAs bound to all ribosomes, even including monosomes, thereby efficiently distinguishing free mRNAs from ribosome-bound mRNAs but not necessarily resolving polysome-bound mRNAs. One question arising from this is whether mRNAs with low RLs found in all conditions have low translation efficiencies due to their intrinsic properties in T cells or whether they are

dynamically regulated depending on the stimuli. We found very low overlap (<8%) between the differential RL mRNAs in all our conditions supporting the notion that the stimuli dictated the outcome of ribosome binding of a given mRNA in CD4 Teff cells (Fig S4E).

We assessed whether there were mRNAs with significantly lower RLs in the Treg suppressed Teff cells compared to the stimulated control. A distinct set of mRNAs including *Akap1*, *Btaf1*, and *Exoc8* displayed lower RL only in suppressed Teff cells with no change in total RNA input levels (Fig SG). Suppressed mRNAs comprised protein coding genes not studied in the context of T cell activation, perhaps due to no apparent changes in the RNA level alone. A kinase anchor protein 1 (*Akap1*) and its family members have been extensively studied in the regulation of cAMP signaling by spatial and temporal regulation of the components in this pathway. However, no definitive role for this protein has been identified in the context of CD4 T cell activation. BTAF1 (*Btaf1:* TATA-binding protein-associated factor 172), a co-factor of B-TFIID has recently been shown to be a direct target of miR-132, a miRNA upregulated during CD4 T cell activation. In the context of CD4 T cell activation, BTAF1 has been shown to be a context-dependent activator of ribosomal protein gene transcription and forced expression of miR-132 results in global protein synthesis shutdown through BTAF1 silencing. The exocyst complex component 8 (*Exoc8*) is a member of the octameric exocyst complex involved in membrane trafficking as well as efficient cytokine secretion in activated T cells. In summary, the RiboTag approach revealed context dependent changes in gene expression beyond transcriptome changes and suggested an active regulation of ribosome dissociation of mRNAs in CD4 Teff cells by Tregs for the first time. We were able to identify mRNAs transcriptionally unaffected by activation in the presence of Tregs, but whose association with ribosomes was inhibited (Fig S4G). These data support a role for Treg-mediated regulation of mRNA translation.

	logFC	AveExpr	t	P.Value	adj.P.Val	B	mgi_symbol
ENSMUSG00000007850	1.452705465	6.90514408	9.670263598	8.37E-12	9.41E-08	15.8065461	Hnrnph1
ENSMUSG00000026003	0.98579119	7.62000787	8.217608565	5.80E-10	3.26E-06	12.56564947	Acadl
ENSMUSG00000024097	1.016468763	8.94969103	7.930151142	1.38E-09	5.18E-06	11.868448	Srsf7
ENSMUSG00000022858	1.009681492	6.41853076	7.467585597	5.69E-09	1.60E-05	10.02167782	Tra2b
ENSMUSG00000021218	0.73561758	8.70454643	6.654320507	7.16E-08	0.000134098	8.073005656	Gdi2
ENSMUSG00000029817	1.557007491	6.66848205	6.331517803	1.98E-07	0.000236076	6.945128554	Tra2a
ENSMUSG00000038299	0.926291039	7.6128867	6.315681511	2.08E-07	0.000236076	7.043274241	Wdr36
ENSMUSG00000030934	0.840628651	8.36793726	6.312507651	2.10E-07	0.000236076	7.039582708	Oat
ENSMUSG00000022186	0.770088838	7.98865936	6.014881064	5.38E-07	0.000549654	6.155690568	Oxct1
ENSMUSG00000021428	1.23562732	7.14282707	5.8135716	1.02E-06	0.00076197	5.551561545	Riok1
ENSMUSG00000026234	0.772953994	8.14352396	5.6639805	1.63E-06	0.001019085	5.09298276	Ncl
ENSMUSG00000022300	0.584131698	8.47764336	5.646288592	1.73E-06	0.001020983	5.015065136	Dcaf13
ENSMUSG00000028790	0.663325744	6.84075493	5.574478931	2.17E-06	0.001173979	4.823981214	Khdrbs1
ENSMUSG00000015837	0.737153452	8.10981975	5.570433668	2.19E-06	0.001173979	4.793024859	Sqstm1
ENSMUSG00000025190	1.1512838	7.95376434	5.509762152	2.66E-06	0.001244116	4.616939093	Got1
ENSMUSG00000056851	0.675847891	7.15198435	5.496852376	2.77E-06	0.001244116	4.607194401	Pcbp2
ENSMUSG00000030560	0.762388559	6.90098995	5.378450321	4.02E-06	0.001632896	4.259276828	Ctsc
ENSMUSG00000028330	0.677190353	7.95233827	5.374726368	4.07E-06	0.001632896	4.221588482	Ncbp1
ENSMUSG00000055762	0.783824471	6.76796003	5.225935847	6.50E-06	0.002282407	3.777295542	Eef1d
ENSMUSG00000004789	0.716576197	8.06460372	5.202998056	6.99E-06	0.00233642	3.705666202	Dlst
ENSMUSG00000001525	0.727071908	9.14726002	5.179067786	7.53E-06	0.002417881	3.598111261	Tubb5
ENSMUSG00000024360	0.597908447	8.26516528	5.152472878	8.19E-06	0.002432597	3.525963425	Etf1
ENSMUSG00000024975	0.754636722	7.96863182	5.123049193	8.98E-06	0.002522347	3.446657309	Pdcd4
ENSMUSG00000030884	0.570868515	8.17005382	5.052642402	1.12E-05	0.002995334	3.246696042	Uqcrc2
ENSMUSG00000026260	0.544485297	8.40690526	5.02802777	1.21E-05	0.003160004	3.160411278	Ndufa10
ENSMUSG00000020361	0.704062471	7.73062556	4.991224036	1.36E-05	0.003351167	3.109258072	Hspa4
ENSMUSG00000001016	0.608241392	8.13503831	4.98768173	1.37E-05	0.003351167	3.068797488	Ilf2
ENSMUSG00000038510	0.896138181	7.70566318	4.952115845	1.53E-05	0.003665291	3.008031677	Rpf2
ENSMUSG00000019179	0.635278652	9.11454946	4.911226748	1.74E-05	0.003890526	2.787317165	Mdh2
ENSMUSG00000015672	0.664658489	8.39904344	4.906823799	1.77E-05	0.003890526	2.783312229	Mrpl32

ENSMUSG00000022336	0.617783361	9.09580155	4.86757586	2.00E-05	0.004152356	2.661295677	Eif3e
ENSMUSG00000060373	0.525744431	8.24858231	4.814231626	2.36E-05	0.004812786	2.506596729	Hnrnpc
ENSMUSG00000079111	0.813996178	7.79214496	4.800921921	2.46E-05	0.004926427	2.539486047	Kdelr2
ENSMUSG00000031715	0.897340719	5.96874637	4.791857973	2.53E-05	0.004978191	2.450713856	Smarca5
ENSMUSG00000053565	0.530976072	9.6074959	4.750815209	2.87E-05	0.005462669	2.341480088	Eif3k
ENSMUSG00000021832	0.556180466	8.76452888	4.742774231	2.94E-05	0.00550721	2.283146083	Psmc6
ENSMUSG00000020708	0.5046479	9.14988387	4.675706777	3.62E-05	0.005980462	2.087934611	Psmc5
ENSMUSG00000005625	0.500966439	8.09176807	4.654180928	3.87E-05	0.006289241	2.054628773	Psmd4
ENSMUSG00000064023	0.506522511	7.68403189	4.617437191	4.33E-05	0.00676142	1.955555418	Klk8
ENSMUSG00000025794	0.641642326	8.1989659	4.593984143	4.66E-05	0.007071752	1.87863647	Rpl14
ENSMUSG00000035202	1.057772713	6.50116884	4.584950685	4.79E-05	0.00716537	1.944816052	Lars2
ENSMUSG00000001783	0.478244045	8.3917028	4.57546983	4.93E-05	0.00716537	1.815113226	Rtcb
ENSMUSG00000047514	0.602442179	7.24014323	4.572611467	4.97E-05	0.00716537	1.850537263	Tspyl1
ENSMUSG00000016541	0.530998893	8.58354601	4.541460749	5.47E-05	0.007688453	1.700527025	Atxn10
ENSMUSG00000001767	0.885148311	6.217693	4.530413032	5.66E-05	0.007855629	1.801617167	Crnkl1
ENSMUSG00000020612	0.504320895	8.53295621	4.523038915	5.79E-05	0.007937511	1.640371706	Prkar1a
ENSMUSG00000018326	0.484013903	8.67153626	4.505671374	6.11E-05	0.008172624	1.583258024	Ywhab
ENSMUSG00000001962	0.685877503	7.10377649	4.496997486	6.27E-05	0.008197717	1.679578144	Fam50a
ENSMUSG00000001380	0.54583261	8.38689755	4.482592729	6.56E-05	0.008469211	1.559982406	Hars
ENSMUSG00000038462	0.497613447	9.22116356	4.473856674	6.73E-05	0.008600014	1.504785068	Uqcrfs1
ENSMUSG00000040028	0.602477276	7.11689312	4.451429784	7.21E-05	0.009084323	1.574343171	Elavl1
ENSMUSG00000033793	0.674124483	7.83084463	4.431867601	7.66E-05	0.009353373	1.43361175	Atp6v1h
ENSMUSG00000055044	0.56245614	6.53590316	4.426887132	7.78E-05	0.009394645	1.523037486	Pdlim1
ENSMUSG00000037916	0.568827616	7.9625517	4.35217982	9.76E-05	0.01119509	1.227028901	Ndufv1
ENSMUSG00000025066	0.522301986	8.80876721	4.302474037	0.000113544	0.012638069	0.991126782	Sfr1
ENSMUSG00000027433	0.577157298	8.13001897	4.295863033	0.000115842	0.012638069	1.018555223	Xrn2
ENSMUSG00000007739	0.461178408	8.33419346	4.287350538	0.00011887	0.012843635	0.974744677	Cct4
ENSMUSG00000020069	0.895767788	6.04132415	4.278635442	0.000122049	0.012856671	1.095162808	Hnrnph3
ENSMUSG00000027170	0.588901934	7.87588804	4.274984634	0.000123406	0.012856671	0.991110731	Eif3m
ENSMUSG00000041459	0.775827289	6.30279505	4.261257252	0.00012864	0.013141148	1.045687306	Tardbp
ENSMUSG00000063884	0.664128116	7.79591832	4.239784593	0.000137266	0.013555147	0.909081998	Ptcd3

ENSMUSG00000005846	0.473423974	8.89081698	4.239178221	0.000137518	0.013555147	0.809032869	Rsl1d1
ENSMUSG00000042590	1.206631851	6.50448663	4.220945002	0.000145299	0.014019529	0.956754485	Ipo11
ENSMUSG00000047879	0.67389615	6.82130773	4.21941351	0.000145972	0.014019529	0.950243337	Usp14
ENSMUSG00000039828	0.707294798	7.16843271	4.210507418	0.000149945	0.014159133	0.865313229	Wdr70
ENSMUSG00000007564	0.501923587	8.42762787	4.198845115	0.000155309	0.014350323	0.707283802	Ppp2r1a
ENSMUSG00000071172	0.5419848	7.24453192	4.19779481	0.000155801	0.014350323	0.833486924	Srsf3
ENSMUSG00000022024	0.45005592	8.12982002	4.187810207	0.000160557	0.014637099	0.700766906	Sugt1
ENSMUSG00000026377	0.468703025	8.51671519	4.185823182	0.00016152	0.014637099	0.678862559	Nifk
ENSMUSG00000026575	1.075709934	7.12446427	4.174948861	0.000166892	0.014715343	0.831284128	Nme7
ENSMUSG00000031701	0.408652827	8.53106947	4.16387302	0.000172543	0.014800506	0.597479788	Dnaja2
ENSMUSG00000022312	0.477200682	9.18841857	4.133304815	0.000189127	0.01536346	0.527804841	Eif3h
ENSMUSG00000004018	0.816469135	6.17763262	4.131692294	0.000190044	0.01536346	0.682322676	Fancl
ENSMUSG00000052253	0.539580237	7.33972185	4.099393924	0.000209347	0.016533629	0.500557885	Zfp622
ENSMUSG00000029169	0.506186457	8.44678662	4.097710813	0.000210404	0.016533629	0.41891431	Dhx15
ENSMUSG00000078920	0.516677536	8.94102912	4.074287279	0.000225662	0.017488013	0.33953031	Ifi47
ENSMUSG00000070697	0.551595722	8.307165	4.021746888	0.000263915	0.019091048	0.224187012	Utp3
ENSMUSG00000031731	0.982814777	6.14568901	4.021676386	0.00026397	0.019091048	0.423699378	Ap1g1
ENSMUSG00000025742	0.550941235	7.323234	4.001134758	0.000280588	0.019955472	0.278575898	Prps2
ENSMUSG00000031320	0.723100928	6.56641085	3.989840437	0.000290154	0.020251305	0.334738166	Rps4x
ENSMUSG00000025337	0.533188669	7.62183042	3.985034296	0.00029432	0.020298011	0.167755405	Sbds
ENSMUSG00000014195	0.434839468	7.06664405	3.958312697	0.000318568	0.021564771	0.174348408	Dnajc7
ENSMUSG00000007891	0.461934514	7.70308641	3.924970179	0.00035156	0.022967898	-0.032425437	Ctsd
ENSMUSG00000068882	0.531333297	8.31899707	3.895829127	0.000383095	0.024184473	-0.121199477	Ssb
ENSMUSG00000029422	0.627342641	6.11913237	3.87280679	0.000409935	0.025032287	0.026210616	Rsrc2
ENSMUSG00000018446	0.628272621	7.91276715	3.870998635	0.000412118	0.025032287	-0.074395047	C1qbp
ENSMUSG00000025287	0.527510232	7.56674985	3.849723927	0.000438674	0.025747882	-0.135480259	Acot9
ENSMUSG00000031708	0.422546449	9.09853232	3.845936718	0.000443572	0.025747882	-0.293453334	Tecr
ENSMUSG00000031672	0.650158104	6.91057442	3.845207005	0.000444522	0.025747882	-0.04120348	Got2
ENSMUSG00000027455	0.559335682	6.41766652	3.827564914	0.000468095	0.026597382	-0.087045321	Nsfl1c
ENSMUSG00000026753	0.520113349	7.37478302	3.804452891	0.000500818	0.027632597	-0.289614225	Ppp6c
ENSMUSG00000003970	0.599233238	10.210691	3.800252536	0.000506999	0.027656034	-0.329235345	Rpl8

ENSMUSG00000001774	0.65527451	8.06265364	3.78931761	0.000523437	0.028414776	-0.419480357	Chordc1
ENSMUSG00000043931	0.476825487	8.71621878	3.780986936	0.000536305	0.028819061	-0.477232676	Gimap7
ENSMUSG00000008333	0.643423041	6.8193216	3.777980133	0.000541025	0.028819061	-0.238834585	Snrpb2
ENSMUSG00000030798	0.505844111	7.22041593	3.775450918	0.000545026	0.028888926	-0.387825543	Cd37
ENSMUSG00000003429	0.658523721	9.21718406	3.740937031	0.000602568	0.030468068	-0.545751119	Rps11
ENSMUSG00000037022	1.429261215	4.13198304	3.739577911	0.000604951	0.030468068	-0.483800356	Mmaa
ENSMUSG00000020664	0.798223252	7.59507404	3.738698439	0.000606497	0.030468068	-0.452200297	Dld
ENSMUSG00000028187	0.513983066	8.55795011	3.737498087	0.000608614	0.030468068	-0.598444726	Rpf1
ENSMUSG00000026727	0.654636221	5.55691335	3.736676631	0.000610066	0.030468068	-0.325539063	Rsu1
ENSMUSG00000062203	0.636367355	6.95600969	3.734183041	0.000614497	0.030553536	-0.330599182	Gspt1
ENSMUSG00000091649	0.482749213	8.37218129	3.718086143	0.000643853	0.031593806	-0.567019808	Phf11b
ENSMUSG00000020372	0.638344526	8.21325616	3.712550282	0.000654259	0.031964835	-0.618610098	Rack1
ENSMUSG00000041645	0.495840593	8.15595142	3.704968873	0.000668774	0.032253288	-0.660628582	Ddx24
ENSMUSG00000022247	0.541878153	7.88262391	3.701635867	0.000675253	0.032426592	-0.614110386	Brix1
ENSMUSG00000014769	0.485536429	9.45083188	3.684258023	0.000710029	0.033599346	-0.712381771	Psmb1
ENSMUSG00000059208	0.453216169	8.22818485	3.675459368	0.00072829	0.034211775	-0.715940759	Hnrnpm
ENSMUSG00000029486	0.858419839	5.82399622	3.661384441	0.000758447	0.034928985	-0.533580228	Mrpl1
ENSMUSG00000041028	0.510577662	8.77446311	3.651036533	0.000781384	0.035548222	-0.832866627	Ghitm
ENSMUSG00000022052	0.507541622	7.47163506	3.64310802	0.000799409	0.036058084	-0.756040941	Ppp2r2a
ENSMUSG00000027248	0.423380511	8.82644235	3.640381289	0.0008057	0.036070336	-0.860121999	Pdia3
ENSMUSG00000057113	0.642632239	8.10770452	3.632665529	0.000823762	0.036570446	-0.797050125	Npm1
ENSMUSG00000041278	0.422910803	7.7573784	3.606305008	0.000888462	0.037514775	-0.872439897	Ttc1
ENSMUSG00000003421	0.492663509	7.20372668	3.597822764	0.000910302	0.037606865	-0.788996388	Nosip
ENSMUSG00000095567	0.400051313	8.56874016	3.581665818	0.000953343	0.038955322	-1.005111033	Noc2l
ENSMUSG00000022905	0.898284412	6.01877168	3.577400515	0.000965028	0.039289927	-0.753499121	Kpna1
ENSMUSG00000032458	0.519633323	7.79657371	3.560170523	0.00101365	0.040434724	-0.938806591	Copb2
ENSMUSG00000062006	0.611162599	7.55924468	3.559794549	0.001014736	0.040434724	-0.915345781	Rpl34
ENSMUSG00000051695	0.43885489	8.98156134	3.551553424	0.001038842	0.040956315	-1.099284823	Pcbp1
ENSMUSG00000024191	1.231564931	4.85132548	3.551119529	0.001040126	0.040956315	-1.051371456	Bnip1
ENSMUSG00000036427	0.488283041	7.73501084	3.550350847	0.001042405	0.040956315	-0.958039569	Gpi1
ENSMUSG00000028249	0.418889314	7.21901382	3.547862045	0.001049817	0.041103805	-0.988117964	Sdcbp

ENSMUSG00000037204	0.463494881	8.06022019	3.54522422	0.001057728	0.041269746	-1.098487153	Atg101
ENSMUSG00000018189	0.480486713	8.46620946	3.541091168	0.001070238	0.041570649	-1.068428469	Uchl5
ENSMUSG00000036693	0.776914837	7.21014124	3.531528157	0.001099732	0.041894151	-0.923471344	Nop14
ENSMUSG00000052146	0.680689692	5.98588746	3.530306219	0.001103557	0.041894151	-0.833319137	Rps10
ENSMUSG00000011114	0.408895814	7.11749723	3.526776563	0.001114676	0.042173792	-0.987450098	Tbrg1
ENSMUSG00000025651	0.51145494	9.49403245	3.514313062	0.001154807	0.043111506	-1.173488165	Uqcrc1
ENSMUSG00000004771	0.417711493	7.26062015	3.511564722	0.001163841	0.043245513	-1.060006004	Rab11a
ENSMUSG00000027374	0.841344929	5.13205109	3.510882684	0.001166093	0.043245513	-0.97077027	Mrps5
ENSMUSG00000054079	0.482114542	7.17985991	3.5045578	0.001187182	0.043453942	-1.019329602	Utp18
ENSMUSG00000032096	0.448571694	7.07916683	3.497326881	0.001211739	0.04406575	-1.043523036	Arcn1
ENSMUSG00000021282	0.320886241	8.38745085	3.494164902	0.001222631	0.044318397	-1.247051779	Eif5
ENSMUSG00000024188	0.821053708	5.91539543	3.492659102	0.001227851	0.04436449	-0.927425943	Luc7l
ENSMUSG00000031807	0.458796279	6.80981096	3.475799805	0.00128777	0.045648823	-1.008804009	Pgls
ENSMUSG00000016921	0.389636884	8.15231783	3.473165879	0.001297382	0.045738865	-1.234988006	Srsf6
ENSMUSG00000022827	1.085870804	4.60698662	3.472873848	0.001298451	0.045738865	-1.185214393	Rabl3
ENSMUSG00000024404	0.509165188	7.32127404	3.469466179	0.001310998	0.045905955	-1.209944962	Riok3
ENSMUSG00000039356	0.457560019	7.98022237	3.456574676	0.001359522	0.046818517	-1.224704706	Exosc2
ENSMUSG00000025525	0.658332659	6.21572746	3.455796573	0.001362505	0.046818517	-1.011523416	Apool
ENSMUSG00000063457	0.470968762	8.9261147	3.447239647	0.001395729	0.047240384	-1.369942805	Rps15
ENSMUSG00000033161	0.708056613	7.15885683	3.444997465	0.001404562	0.047254692	-1.195988141	Atp1a1
ENSMUSG00000029328	0.522764503	7.30768527	3.440996754	0.001420457	0.047526889	-1.185691314	Hnrnpdl
ENSMUSG00000005683	0.404213931	8.58553601	3.436082796	0.001440215	0.04773952	-1.390395751	Cs
ENSMUSG00000031304	0.523071467	8.28879398	3.432188177	0.001456062	0.047981716	-1.402642687	Il2rg
ENSMUSG00000049760	0.384488856	8.80461198	3.418395329	0.001513536	0.049154927	-1.446809412	2410015M20Rik
ENSMUSG00000020589	0.680224388	5.84157261	3.411387322	0.001543564	0.049962878	-1.139370591	Fam49a
ENSMUSG00000024539	0.412927436	7.60766941	3.410523107	0.001547306	0.049962878	-1.362275157	Ptpn2

Supplementary Table 2: Reagent identifiers

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
Anti-Puromycin, clone 12D10, AlexaFluor647 Conjugate	EMD Milipore	MABE343-AF647
Anti-HA tag antibody-ChIP Grade	Abcam	Ab9110
S6 Ribosomal Protein (5G10) Rabbit antibody	Cell Signaling Technology	2217L
CD4-BV421 (GK1.5)	BioLegend	100438
CD62L-PE (MEL-14)	BioLegend	104408
CD45.1-FITC (A20)	BioLegend	110706
Alpha-Tubulin	Cell Signaling Technology	
Anti-IL-10R, clone 1B1.3A	BioXcell	BE0050
Anti-IFNg, XMG1.2, PerCP/Cyanine5.5 conjugate	Biolegend	505822
Anti-IL-17A, TC11-18H10.1, PE conjugate	Biolegend	506904
Anti-human IFN-g, 4S.B3,APC conjugate	Biolegend	502512
Anti-IL-13, JES10-5A2, PE-conjugate	Biolegend	501903
Anti-IL-2, JSE6-1A12	Biolegend	503706
Anti-IL-2R, 3C7	biolegend	101926
Biological samples		
Human PBMC	Benaroya Research Institute Biorepository	
Chemicals, peptides, and recombinant proteins		
Cycloheximide, ready-solution	Sigma	C4859-1ML
Rocaglamide (Roc-A)	MedChemExpress	HY-19356
Recombinant Human IL-2 (carrier-free)	BioLegend	589106
Recombinant Mouse TGFb1 (carrier-free)	BioLegend	763104
Puromycin ready-made solution	Sigma	P9620-10ML
Tacrolimus (FK506)	MedChemExpress	HY-13756
Sapanisertib (MLN0128)	MedChemExpress	HY-13328
CellTrace CFSE Cell Proliferation Kit	ThermoFisher	C34554
CellTrace Blue cell proliferation kit	ThermoFisher	C34568
Recombinant Mouse IL-10 (carrier-free)	Biolegend	575804
Recombinant Mouse TGF-β1 (carrier-free)	Biolegend	763104
Critical commercial assays		
Agilent RNA 6000 Pico Kit	Agilent	5067-1513
SMARTseq v4 Ultra-low input kit	Clontech	634888
RNeasy micro kit	Qiagen	74004
Colloidal Blue Staining Kit	ThermoFisher	LC6025
Direct-zol 96 kit	Zymo Research	R2054
Quant-iT RiboGreen RNA Assay kit	ThermoFisher	R11490
Pan Human T Cell Isolation Kit	Miltenyi	130-096-535
MojoSort Mouse CD4 T cell Isolation Kit	BioLegend	480033
Dynabeads Protein G	ThermoFisher	10003D

Oligonucleotides		
Actb	ThermoFisher	Mm02619580_g1
Rps10	ThermoFisher	Mm02391992_g1
Rpl14	ThermoFisher	Mm00782569_s1
Dhx34	ThermoFisher	Mm01268294_g1
eIF3e	ThermoFisher	Mm01700222_g1
Rpl8	ThermoFisher	Mm00657299_g1
Deposited data		
Analysis Code	This paper	https://github.com/BenaroyaResearch/P232RiboSeq
Raw Data	This Paper	GEO ID:GSE171789
Experimental models: organisms/strains		
B6J.129(Cg)-*Rpl22*$^{tm1.1Psam}$/SjJ	Jackson Laboratories	02997
B6.Cg-Tg(Cd4-cre)1Cwi/BfluJ	Jackson Laboratories	022071
B6.C-Tg(CMV-cre)1Cgn/J	Jackson Laboratories	006054
B6.129(Cg)-*Foxp3*$^{tm3(DTR/GFP)Ayr}$/J	Jackson Laboratories	016958
B6.129(Cg)-*Foxp3*$^{tm4(YFP/icre)Ayr}$/J	Jackson Laboratories	016959
B6.SJL-*Ptprc*^{a}*Pepc*^{b}/BoyJ	Jackson Laboratories	002014
IL-10RB-/- (B6.129S2-Il10rbtm1Agt/J	Jackson Laboratories	005027
Software and algorithms		
STAR 2.4.2a read aligner	Dobin et al. 2013	https://github.com/alexdobin/STAR
LIMMA	Ritchie et al. 2015	https://www.bioconductor.org/packages/release/bioc/html/limma.html
fgsea	Korotkevich et al. 2019	https://www.biorxiv.org/content/10.1101/060012v3
Htseq-count	Anders et al. 2014	https://github.com/htseq/htseq