

1 A synthesis of senescence predictions for indeterminate growth,  
2 and support from multiple tests in wild lake trout

3  
4  
5 Craig F. Purchase<sup>1\*</sup>, Anna C. Rooke<sup>1§</sup>, Michael J. Gaudry<sup>2\$\$</sup>, Jason R.  
6 Treberg<sup>2,3</sup>, Elizabeth A. Mittell<sup>4</sup>, Michael B. Morrissey<sup>4</sup>, Michael D.  
7 Rennie<sup>5,6</sup>

8 <sup>1</sup> Department of Biology, Memorial University of Newfoundland, Canada

9 <sup>2</sup> Department of Biological Sciences, University of Manitoba, Canada

10 <sup>3</sup> Centre on Aging, University of Manitoba, Canada

11 <sup>4</sup> School of Biology, University of St. Andrews, United Kingdom

12 <sup>5</sup> Department of Biology, Lakehead University, Canada

13 <sup>6</sup> IISD Experimental Lakes Area, Canada

14 § Current address: Department of Biology, Queen's University, Canada

15 \$\$ Current address: Department of Molecular Biosciences, The Wenner-Gren Institute, Sweden

16 \* Corresponding author: [cfpurchase@mun.ca](mailto:cfpurchase@mun.ca)

17

18 **ABSTRACT**

19 Senescence, or the deterioration of functionality with age, varies widely across taxa in pattern and  
20 rate. Insights into why and how this variation occurs are hindered by the predominance of lab-  
21 focused research on short-lived model species with determinate growth. We synthesize  
22 evolutionary theories of senescence, highlight key information gaps, and clarify predictions for  
23 species with low mortality and variable degrees of indeterminate growth. Lake trout are an ideal  
24 species to evaluate predictions in the wild. We monitored individual males from two populations  
25 (1976-2017) longitudinally for changes in adult mortality (actuarial senescence) and body

26 condition (proxy for energy balance). A cross-sectional approach (2017) compared young (ages  
27 4-10 years) and old (18-37 years) adults for (1) phenotypic performance in body condition, and  
28 semen quality - which is related to fertility under sperm competition (reproductive senescence),  
29 and (2) relative telomere length (potential proxy for cellular senescence). Adult growth in these  
30 particular populations is constrained by a simplified food web, and our data support predictions  
31 of negligible senescence when maximum size is only slightly larger than maturation size.  
32 Negative senescence (aka reverse senescence) may occur in other lake trout populations where  
33 diet shifts allow maximum sizes to be much larger than maturation size.

34  
35 **KEYWORDS:** ageing, disposable soma, sperm senescence, life history theory, sexual  
36 selection, *Salvelinus namaycush*  
37

## 38 1. INTRODUCTION

39 Senescence is a decline in individual biological function with age, and is typically  
40 quantified as an increase in adult mortality rate or reduced ‘fertility’ [1], but can be applied to any  
41 decline in phenotypic performance. Tremendous variability exists among species in the shape  
42 (direction) and speed (rate) of senescence [2-5], and many authors seek to explain such patterns  
43 [e.g., 3, 6, 7]. The contention that the strength of selection declines with age is a common  
44 explanation of senescence [8]. The premise being that few individuals reach old age, and many  
45 have already reproduced at younger ages, therefore, selection cannot remove problematic traits  
46 that arise only at old age. An hypothesis that “low adult death rates should be associated with low  
47 rates of senescence, and high adult death rates with high rates of senescence” [9], has empirical  
48 support. However, the nuances of the hypothesis and its predictions are debated [6, 10, 11].  
49 Relative adult to juvenile mortality appears critical [6], but asymmetry between parent and  
50 offspring [7] can differ widely between determinate and indeterminate growers and  
51 generalizations can be problematic. An example with bivalves provides a useful illustration [see  
52 6, page 527], which would also apply to most fishes.

53 Our manuscript has three primary goals: 1) synthesize existing senescence theories,  
54 showing the importance of growth pattern, and highlight types of data needed to fill key voids, 2)

55 introduce lake trout (*Salvelinus namaycush*) as an ideal species to address senescence in the wild,  
56 3) present a case study of two lake trout populations with exceptional monitoring.

57

## 58 1.1 EVOLUTIONARY THEORIES OF SENESCENCE

59 Attempts to explain senescence are challenged by inconsistencies in terminology and in  
60 the hierarchy of how theories are grouped. Complicating things further, the major theories of  
61 senescence [7] are not mutually exclusive, and create similar predictions but for different reasons.  
62 Our interpretation (Figure 1) represents a modification from Maklakov and Chapman [8; their  
63 Figure 2]. The mutational accumulation theory (MAT, Figure 1), posits [12] that individuals  
64 senesce due to the accumulation of deleterious mutations through their lifetime, such that  
65 senescence is strictly maladaptive. Other theories (Figure 1) consider the notion of fitness  
66 optimization or life history tradeoffs, whereby declining performance with age may result from  
67 increased performance while young. The antagonistic pleiotropy hypothesis (APH, [9]) suggests  
68 senescence occurs when certain genes have positive effects in early life but negative effects later.  
69 The disposable soma hypothesis (DSH) proposes [13] energy allocated in reproduction is  
70 unavailable to maintain soma, resulting in deterioration. Many present APH and DSH as distinct,  
71 but we consider DSH to be a version of APH (Figure 1). More recently, optimization of function  
72 has been proposed; appearing as developmental function theory (DFT, [8]) and hyperfunction  
73 [14]. Conceptually this is similar to DSH but the proposed mechanism varies, being energy  
74 allocation for DSH (a tradeoff) and hyperfunctioning genes that lead to excessive biosynthesis  
75 and molecular turnover in mature individuals for DFT (which unlike [8] we consider as a putative  
76 constraint [sensu 15] – as opposed to a plasticity enabled tradeoff; Figure 1). How DFT might  
77 apply to indeterminate growers is unclear, as development never stops.

78

## 79 1.2 ATYPICAL PATTERNS OF SENESCENCE

80 Most empirical work on senescence has been framed in support of DSH [e.g, 8, 15, 16].  
81 However, there has been recent questioning of this [8, 15, 17], and new research that addresses  
82 some key gaps may be revealing. Examining unusual patterns of senescence [3, 7] may help  
83 illuminate why and how it occurs ([5]; Figure 1). Negligible senescence describes species with  
84 little or no deterioration with age [2, 18, 19], while negative (reverse) senescence [20] may occur  
85 when biological function increases with age. The tenet of this argument is that in all species,  
86 mature individuals have offspring that are smaller than themselves. As offspring grow, their  
87 ability to reproduce increases and their probability of mortality can decline. In species with  
88 determinate growth, this pattern stops at maturity. Indeterminate growers however continue to  
89 increase in size after maturity. If mortality declines and fertility increases with size (age), then  
90 there is increased selection against senescence in indeterminate versus determinate growers.

91 Across different conditions, an optimization model [20] concludes that the intrinsic  
92 growth pattern (determinate vs indeterminate) influences the shape (direction) of senescence,  
93 while mortality determines its rate. Predictions can be summarized as: (1) senescent conditions  
94 (classical ageing) occur when the size at maturity is close to the maximum size (determinate  
95 growth) with little scope for increasing fertility with age (e.g., mammals, birds, insects); (2)  
96 negative (reverse) senescence should occur when size at maturity is much less than maximum  
97 size (some indeterminate growers), and reproductive capacity increases with size; (3) negligible  
98 senescence (little ageing) is an arbitrary middle ground along this continuum and should occur  
99 when size at maturity is somewhat less than maximum size, but reproductive capacity increases  
100 with size (age). Support for this framework appears in a recent review [7].

101

### 102 1.3 DESIRABLE STUDY SYSTEMS TO FILL KEY VOIDS

103 Studies of senescence are heavily skewed towards a narrow range of conditions. A  
104 synthesis of the repeated calls [e.g., 16] to address knowledge gaps includes:

105 (1) A critical need for research focusing on species with indeterminate growth [1, 20-22],  
106 for example in certain plants [23], reptiles [24] and fishes [18, 19]. Most work on senescence has  
107 considered determinate growers (mammals, birds, insects), which may bias our view of ageing.

108 (2) A requirement to examine senescence in wild populations [1, 25, 26], which better  
109 encapsulate natural processes and influences of potential environmental covariates on senescence.  
110 Laboratory studies of model organisms lack this relevance.

111 (3) Research that combines both longitudinal and cross-sectional comparisons of age is  
112 valuable [e.g., 27]. Comparisons in fitness-related traits can be made among age classes (cross-  
113 sectional) or by following individuals through time (longitudinal; [23]). Because long-lived  
114 individuals may have inherent higher quality, their presence may bias cross-sectional  
115 comparisons, making longitudinal studies a desired approach [26, 28]. However, longitudinal  
116 studies are subject to other confounding variables (e.g., directional environmental change), and it  
117 can take decades to track new metrics if following future cohorts. Thus, studies reporting  
118 consistent conclusions across combined approaches may provide more robust tests of hypotheses.

119 (4) Examinations of wild populations not subject to confounding variables [29], such as  
120 immigration/emigration (which may influence estimates of adult mortality), anthropogenic  
121 effects (e.g., recent changes in mortality adding novel selective pressure), and adult diet shifts  
122 with increasing body size, which can have dramatic influence on reproduction (e.g., gape limited  
123 carnivorous reptiles shift diet and are a problem, filter feeding bivalves are not).

124 (5) Research using recognized cellular indices associated with senescence, like relative  
125 telomere length [30] and the influence of reactive oxidative species and their potential for  
126 oxidative stress or cellular damage [29] are needed [26, 31], particularly in wild ectotherms.  
127 Evolutionary literature on senescence ponders what happens (patterns), why it happens (or does  
128 not), but rarely addresses how it happens [8, 28, 31, 32]. Laboratory and model organism-based  
129 studies on the biochemical mechanisms, or at least correlates associated with aging and  
130 senescence, provide a framework that can be applied to study senescence in the wild.

131 (6) Research focusing on reproductive senescence [24, 26, 33]. Most studies [26] of  
132 senescence quantify it as change in adult mortality rates (actuarial senescence), yet invoking  
133 mortality as an explanation is circular [26, 34] being both a cause and consequence of  
134 senescence. Measures of reproductive senescence are free of this problem, as are other  
135 phenotypic traits.

136 (7) Senescence research that considers male individuals. Females have been the historical  
137 focus for senescence research [32, 33], but in most cases, males should senesce faster [8, 32, 35-  
138 39] thus offering larger effect sizes and greater power to answer key questions. This is most  
139 pronounced in species with intense sexual selection [36, 40] as increased reproductive effort may  
140 come at a cost to tissue maintenance, and mortality can be consistently higher on males due to  
141 conspicuous displays.

142 • **The special problem of sperm senescence**

143 Reproductive senescence includes senescence on the adult individual (such as ability to  
144 attract a mate), but additionally on gametes [31, 41-43]. Gamete senescence affects the fitness of  
145 the individual, but also its mate and offspring [43, 44]. However, separating effects of the parent,  
146 gamete, and offspring is difficult, especially in internal fertilizers. Egg senescence is rarely  
147 measured [33], but sperm senescence is gaining interest [43, 45]. Sperm senescence can be

148 considered in two phases [42, 45]: pre-meiotic (how the age of the male influences sperm) and  
149 post-meiotic (both before and after ejaculation). Sperm are particularly vulnerable to oxidative  
150 damage [31], and the male mutational bias [42, 46], has led to interest in human fertility and  
151 paternal effects. Male fitness is a function of mating opportunities, sperm performance and  
152 offspring viability [33, 44], which can be separated under experimental conditions [e.g., 47, 48].  
153 Older males generally produce sperm with reduced fertilization ability [27, 29, 33] and lead  
154 higher rates of developmental abnormalities among offspring [29].

155

## 156 2. LAKE TROUT

### 157 **Desirable attributes**

158 Lake trout present an ideal indeterminate growth model for studies of senescence in  
159 nature, with low adult mortality being a key attribute. They inhabit the hypolimnion of lakes [49],  
160 where there are functionally no predators on adults (contrasts greatly to marine predation on  
161 anadromous salmonids) and spawn on lake shoals at night [49, 50], where they are not exposed to  
162 terrestrial predators (unlike stream spawning salmonids).

163 Reproductive quality and investment can be accurately estimated from gametes. Lake  
164 trout do not typically migrate to spawn, show few secondary sexual characteristics, no sexual  
165 dimorphism, have no energetically costly courtship, and provide no parental care [49, 50].  
166 Fertility increases with size (age), as larger females produce more eggs. Males do not compete for  
167 territories [49, 50], but post-ejaculatory sexual selection [44] occurs due to sperm competition  
168 [50]. Larger (older) fish generally produce more sperm, and thus would gain paternity advantages  
169 (fertility) under a fair raffle system [51].

170 Variation in maximum body size across populations (variable realization of indeterminate  
171 growth) may be useful for testing predictions of negligible and negative senescence [20] within  
172 the same species. Lake trout are amongst the largest members of the Salmonidae family, but  
173 maximum body size varies greatly as a function of prey availability [52, 53]. Thousands of  
174 populations vary in life history traits that influence their fitness [54]. Inter-population  
175 comparisons could exploit environmental variation (something senescence literature has been  
176 asking for [e.g., 15, 16, 26, 28, 32]) in variables such as growing season, prey resources, and  
177 juvenile predators.

178

### 179 **Support for theories of ageing**

180 If senescence is optimized (Figure 1) between fitness benefits early in life at a cost to  
181 either hyperfunctioning genes (DFT) or somatic maintenance (DSH), then selection against a  
182 decline in performance with age is predicted to be relatively high in lake trout, as fitness potential  
183 increases dramatically with size (age), given adult mortality rates decline while fertility increases.  
184 We are unaware of any published data that can shed specific light on DFT in lake trout. However,  
185 low allocation in reproduction is predicted to plastically tradeoff with high investment in somatic  
186 maintenance under DSH [55]. Possibly supporting this, lake trout have relatively low secondary  
187 sexual characteristics/migration/courtship/fecundity (resulting in low annual reproductive effort)  
188 and a predictably high incidence of iteroparity [49]. Perhaps consequently, they can live to ages  
189 of >60 years [56], making them among the longest lived fishes, vertebrates, and animals on the  
190 planet. Using a variety of approaches, we sought to directly test the hypothesis that wild lake  
191 trout show little or no senescence [20].

192

### 193 **Case study of two populations**



194 Our study populations have several additional attributes making them valuable for testing  
195 hypotheses of senescence in the wild. Many potentially confounding variables can be ruled out,  
196 as the lakes are located at the IISD Experimental Lakes Area (Ontario, Canada), where  
197 recreational fishing is prohibited and there is no unquantifiable directed anthropogenic activity.  
198 Annual mark-recapture studies have been ongoing since 1976, enabling long-term monitoring of  
199 individuals. The lakes are very small (see methods) and all adults of various ages within a  
200 population experience similar environmental conditions. There are no piscivorous predators  
201 (except lake trout), adult trout are too large to be taken by loons (*Gavia immer*), but might  
202 occasionally be prey to otters (*Lontra canadensis*). Adult mortality is thus very low, whereas  
203 mortality of small juveniles is likely relatively high [sensu 6]. The lakes are connected in their  
204 surrounding watershed by very small streams, effectively eliminating immigration/emigration for  
205 this hypolimnetic species. Due to a simplified food-web [52, 57] adult trout in these two lakes do  
206 not switch diet as they age, and gain little body size after maturity (Figure 2a, and published  
207 growth curves [57]). This is critically important, as diet is known to affect gamete quality in  
208 fishes [e.g., 58] and would bias age (size) comparisons in most systems. Sampling over the  
209 course of 40+ years has shown that young and old adult male lake trout co-occur on the spawning  
210 shoals at the same time (Rennie, unpublished), thus our comparisons of age are not confounded  
211 by differential spawn timing.

212

### 213 3. METHODS

214 In polyandrous mating systems like lake trout, male “fertility” is influenced by the ability  
215 to achieve fertilizations under sperm competition [33, 44], a key component [45] being sperm  
216 swimming performance. We thus quantified male “fertility” by measuring sperm traits that

217 predict paternity. We also measured adult mortality estimates, body condition as a surrogate for  
218 general health [59, 60], and relative telomere length as a cellular-level marker of senescence [61-  
219 63]. Our study thus combines actuarial senescence, phenotypic measures of bodily function with  
220 age (including reproductive senescence), along with a potential biochemical senescence marker,  
221 providing a more holistic approach others have highlighted as being needed [e.g., 8].

222

### 223 3.1. LONGITUDINAL STUDY

224 At first capture, fish were tagged, measured (total length, mass) and sexed, with the  
225 leading fin ray of a pectoral fin removed for ageing [64]. Recaptures in subsequent years used tag  
226 identification to assign age. Fish over the entire duration of monitoring in Lake 224 (27.3 ha,  
227 1976–2017) were used, while from Lake 223 (26.4 ha) we restricted data to 1990-2017, to  
228 exclude the potential influence of an historical acidification experiment [57] – too few samples  
229 remained to track condition in Lake 223.

230

#### 231 **(A) Actuarial Senescence**

232 We estimated annual individual recapture and mortality probabilities using all adult males  
233 with known ages (Lake 223 = 385, Lake 224 = 422). To test for changes in adult mortality with  
234 age, we fitted a Cormack Jolly Seber model with a Bayesian framework (see Supplemental  
235 Methods). Recapture and mortality probabilities were modelled as logistic regression functions of  
236 age, which was treated as a continuous variable.

237

#### 238 **(B) Phenotypic performance senescence – body condition**

239 Length-based body condition was estimated as a percentage of standard weight [65]. Fish  
240 from Lake 224 that were recaptured at least 6 times during their adult life were used to determine  
241 if condition declined with age, and were analyzed with a mixed effects modelling framework  
242 (Supplemental Methods). Condition was evaluated as a function of fish age (fixed effect), and  
243 repeated measures on the same individuals (random slope), and the year sampled (random  
244 intercept).

245

## 246 3.2. CROSS-SECTIONAL STUDY

### 247 **(A) Fish collection**

248 We collected fish on spawning shoals at night from 11 to 16 October 2017 and sampled  
249 the next morning following previous procedures [66]. Ages of recaptured fish were determined in  
250 the field by cross-referencing a database of tag IDs. Younger adult trout were more abundant than  
251 older individuals. To avoid potential confounding variables associated with date of sampling  
252 (e.g., weather, transport time to laboratory), we grouped fish as either being young (ages 4–10) or  
253 old (18–37) and processed them in a ‘group design’ (i.e., the same number of young and old fish  
254 were sampled each day). We analyzed 15 groups in each lake (60 total; Supplemental Methods).

255

### 256 **(B) Sample collection**

257 Eggs were extruded from one female each day and later separated from ovarian fluid  
258 through a fine meshed net [67], which was used in sperm swimming performance trials [68], to  
259 avoid neutral sperm swimming environments when post-ejaculatory sexual selection occurs [29].  
260 From each male, blood was taken from the caudal peduncle and semen was expressed by gentle

261 abdominal massage. All samples were immediately immersed in ice, and transported to the lab  
262 for further processing (completed within 8 hours of collection).

263 Aliquots of blood and semen were removed from ice and centrifuged ( $5000 \times g$  at  $\sim 15^{\circ}\text{C}$   
264 for 5 mins). Prior to freezing in liquid nitrogen, plasma was separated from blood cells. A  
265 separate semen aliquot was centrifuged in hematocrit tubes, and spermatocrit was computed [69].  
266 This correlates with semen sperm density and often varies within individuals through a spawning  
267 season [e.g., 70].

268

### 269 **(C) Sperm swimming performance**

270 Details (Supplemental Methods) closely followed Purchase & Rooke [67]. Four technical  
271 replicates of sperm activation were obtained for each fish. We were able to get useful data within  
272 6 s of sperm/media mixing. Videos of swimming sperm were analyzed in 0.5 s increments using  
273 open source software [71]. We used sperm curvilinear swimming velocity (VCL;  $\mu\text{m/s}$ ) as a  
274 metric of male fertility, as it has been repeatedly shown to be correlated to paternity under sperm  
275 competition [72].

276

### 277 **(D) Relative telomere length**

278 We measured relative telomere length from DNA recovered from red blood cells and  
279 sperm pellets using a qPCR-based approach that produces a telomere repeat (T) to single gene (S)  
280 copy number ratio (T/S). The assay was performed with two single copy genes, *orexin* (*Ox*) and  
281 *follicle stimulating hormone beta subunit* (*FSH*), to verify consistency of T/S ratios  
282 (Supplemental Methods). Both genes (*Ox* and *FSH*) garnered congruent relative T/S ratios

283 (Pearson's correlation; blood:  $r = 0.67$ ,  $P < 0.0001$ , sperm:  $r = 0.72$ ,  $P < 0.0001$ ), thus only the  
284 results of  $Ox$  are presented.

285

## 286 **(E) 2017 cross-sectional statistical analyses**

287       Body condition, spermatocrit, and relative telomere length were evaluated as a function of  
288 fish age (young vs. old) crossed with lake of origin. Sperm swimming declines rapidly after  
289 activation, with most successful fertilizations occurring in the few seconds after release. As such,  
290 we quantified sperm swimming using two approaches. First, to assess maximum swimming  
291 speed, we measured sperm at 6 s post-activation as a function of fish age (continuous variable: 4–  
292 37 years) crossed with lake, including tag ID (random intercept) to account for the four technical  
293 replicates per male. We also tested for changes in sperm swimming speed over time post-  
294 activation (continuous: 6–30 s) crossed with age (young vs. old) and lake. Tag ID (random slope  
295 and intercept) and technical replicate (random slope and intercept) were included. In all cross-  
296 sectional analyses the interaction between age and lake was not significant ( $P > 0.23$ ), indicating  
297 that the effect of age was similar in both populations. We removed these non-significant  
298 interactions prior to reporting final model results.

299

## 300 **4. RESULTS**

### 301 **ACTUARIAL SENESCENCE**

302       Annual mortality probability estimates of adult male lake trout were low ( $< 0.20$ ) across  
303 all ages in both lakes, and suggest a modest increase with age (Figure 2b, c). This effect of age  
304 was clearer in Lake 224 compared to Lake 223 (99.8% and 80.5% of the posterior distributions of  
305 the slope parameter were positive, respectively).

306

## 307 **PHENOTYPIC PERFORMANCE SENESENCE**

### 308 **Longitudinal condition**

309 Accounting for random individual (194 fish, 1608 observations) and annual variation,  
310 there was a significant change in adult body condition with age in Lake 224 ( $t_{216.2} = -2.6$ ,  $P =$   
311  $0.009$ ; Figure 2d). The rate of decline was negligible at 1.4 units per decade, which is well within  
312 the variation among fish and years (most observations between 70-105 units).

313

### 314 **Cross-sectional condition (2017)**

315 Overall mean body condition in October 2017 was  $83.4 \pm 0.9\%$ , similar to historical  
316 records (Figure 2d). Body condition was similar in both lakes (Lake 224 – Lake 223 means  $\pm$   
317 SE:  $0.053 \pm 1.79\%$ ,  $t_{55} = 0.03$ ,  $P = 0.976$ ) and there was no difference between young and old  
318 trout (old – young means  $\pm$  SE:  $-0.146 \pm 1.79\%$ ,  $t_{55} = -0.082$ ,  $P = 0.935$ ; Figure 3a).

319

### 320 **Cross-sectional semen quality (2017)**

321 Although spermatocrit was significantly higher in Lake 223 in 2017 (Lake 224 – Lake  
322 223:  $-0.203 \pm 0.028\%$ ,  $t_{46.0} = -7.21$ ,  $P < 0.0001$ ), there was no difference between young and old  
323 fish (old – young:  $-0.019 \pm 0.028\%$ ,  $t_{46.0} = -0.70$ ,  $P = 0.49$ ; Figure 3b). Swimming speed at 6 s  
324 post-activation was not significantly different between lakes (Lake 224 – Lake 223:  $-15.0 \pm 8.1$   
325  $\mu\text{m/s}$ ,  $t_{56.9} = -1.84$ ,  $P = 0.071$ ), and was not related to fish age ( $0.71 \pm 0.45 \mu\text{m/s}$  per year,  $t_{56.9} =$   
326  $1.58$ ,  $P = 0.12$ ; Figure 4a). The rate of decline in swimming speed over time post-activation was  
327 faster in Lake 223 (rate difference, Lake 224 – Lake 223:  $0.97 \pm 0.37 \mu\text{m/s}$  per second post  
328 activation,  $t_{57.0} = 2.65$ ,  $P = 0.01$ ); however, there was no difference between young and old trout

329 (rate difference, old – young:  $-0.43 \pm 0.37 \mu\text{m/s}$  per second post activation,  $t_{57.0} = -1.18$ ,  $P =$   
330  $0.24$ ; Figure 4b).

331

## 332 **BIOCHEMICAL PROXY**

333 Relative telomere length in red blood cells was similar in both lakes (Lake 224 – Lake  
334 223:  $28 \pm 2308$ ,  $t_{56} = 0.012$ ,  $P = 0.99$ ), and in young and old individuals (old – young:  $251 \pm$   
335  $2308$ ,  $t_{56.0} = 0.11$ ,  $P = 0.914$ ; Figure 3c). Relative telomere length in sperm cells was significantly  
336 higher in Lake 224 (Lake 224 – Lake 223:  $8406 \pm 1673$ ,  $t_{57} = 5.03$ ,  $P < 0.0001$ ); however, there  
337 was no difference between young and old trout (old – young:  $-1196 \pm 1673$ ,  $t_{57} = -0.72$ ,  $P =$   
338  $0.48$ ; Figure 3d).

339

## 340 **5. DISCUSSION**

341 As a species, lake trout have evolved under indeterminate growth, and all individuals have  
342 this genetic potential. However, realized growth in lake trout varies depending on diet  
343 availability, with fish achieving enormous sizes in some lakes, but are stunted in others. We  
344 exploited this scenario to make age comparisons among male trout that were not confounded by  
345 diet differences. Adult trout in our study lakes have functionally determinate growth due to a  
346 simplified foodweb. Despite this, and as predicted by both DSH and DFT, they show little to no  
347 senescence in many traits measured here, and while we conclude that overall senescence is  
348 negligible in these populations, we argue that there may be negative (reverse) senescence in other  
349 populations that are not growth constrained.

350 Although we have no molecular data to underpin endorsement of DFT, the DSH is clearly  
351 supported in our lake trout model. That low adult mortality [relative to juveniles, 6] should be

352 associated with few negative effects of ageing [9, 11] in indeterminate growers [20] paints an  
353 incomplete picture of how selection across generations interacts with life history tradeoffs in  
354 individuals. Life history theory predicts that consistently low adult mortality *across generations*  
355 leads to low reproductive effort in a given year as a means of bet hedging reproductive success  
356 across many episodes/years [73]. Under the DSH, through phenotypically plastic allocation of  
357 resources, this low reproductive effort would result in high somatic maintenance and thus low  
358 senescence *within a generation* (individual). Connecting these concepts for the case of lake trout  
359 suggests that due to (1) the lack of adult predation in the growing and spawning environments  
360 they evolved under, adult mortality is consistently very low across generations (it is the lowest of  
361 any salmonid), (2) resulting in low reproductive effort in a given year (it is the lowest of any  
362 salmonid), and (3) through plasticity within-individuals, high somatic maintenance results in no,  
363 or limited, senescence, enabling full potential of long life (it is the highest of any salmonid) to  
364 hedge reproductive success with environmental stochasticity.

365       Observed variation in senescent patterns among species [5, 18, 19] suggests contrasting  
366 selection pressures as an ultimate cause. Indeterminate growth is predicted [20] to increase  
367 selection against senescence when adult individuals experience reduced mortality and increased  
368 fertility with age (increasing size). Testing this prediction in wild populations has been  
369 challenging due to often confounding variables. Lake trout from our particular populations enable  
370 unique opportunities to control such problems, including diet. However, a stable diet likely  
371 results in old fish having inferior performance than their inherent potential. Negative senescence  
372 is predicted when size at maturity is much smaller than maximum size [20]. In lake trout  
373 populations where adults can switch to larger or more energy dense prey as they grow, old fish  
374 achieve much larger sizes than young adults [52]. Very large adults would have high sperm  
375 quantity (predicted to win under sperm competition = fertility), and high sperm quality (predicted



376 to win under sperm competition = fertility) if under suitable diet (and females would have much  
377 more egg production). Such data would not only support negligible senescence, but also show the  
378 aptitude for negative (reverse) senescence in this species.

379 Our study suggests lake trout have at most negligible senescence, with the potential to  
380 exhibit negative (reverse) senescence in populations where adults can attain maximum sizes that  
381 are much larger than those at maturity, due to prey availability. These data provide support of  
382 evolutionary theories of ageing, from rarely studied long-lived indeterminate growing animals in  
383 the wild. Our data are unique in that they coalesce information on 1) actuarial senescence using  
384 mortality rates from mark-recapture, with 2) measures of phenotypic performance including  
385 reproductive senescence. Furthermore, blood and sperm cell telomere lengths did not decline  
386 with age, 3) indicating that, at least under the conditions of this study, telomere maintenance  
387 through adulthood may in part underpin the lack of apparent senescence. Our age comparisons  
388 combine longitudinal (same individuals across decades) with cross-sectional data (difference  
389 aged individuals at the same time), which is an infrequent approach. These age assessments are  
390 strengthened by the unique characteristics of the study populations that control for confounding  
391 variables that are profuse in most natural situations.

392 If our conclusions are accurate, one can make predictions that should be supported by  
393 other data. For instance, (1) old and young males should show equal paternity if tested under  
394 sperm competition in the lab, (2) pedigrees of wild populations should show equal average  
395 contributions of individual old and young males as fathers in a given year, and (3) laboratory  
396 studies should indicate no increase in abnormalities in offspring development from older vs  
397 young fathers. (4) If DFT is involved, relative adult to juvenile proxies for cellular hyperfunction,  
398 should be correlated with senescence, within and among genera of salmonids in relation to the  
399 degree of iteroparity and semelparity, with lake trout at one extreme. (5) Given the unusual

400 insensitivity of relative telomere length to aging in this species, further laboratory and field  
401 studies are needed to test if levels of other common molecular markers of senescence in this, and  
402 other long-lived ectotherms, may also fail to recapitulate the patterns expected from studies on  
403 endotherms or more typical laboratory model organisms. We encourage such studies to be  
404 undertaken where possible, along with comparisons across lake trout populations that vary in  
405 adult mortality and growth potential.

406

## 407 ACKNOWLEDGEMENTS

408 We thank staff of the IISD-ELA for collecting historical information from Lakes 223 and  
409 224, especially K. Mills, S. Chalanchuk and D. Allan. 2017 samples were collected with the  
410 assistance of L. Hrenchuk, C. Rogers, L. Hayhurst, A. Milling, C.V. Veen, and M. Fahmy. M.  
411 Fahmy also supported the microscope analyses. D. McLennan and K. Jeffries are thanked for  
412 assistance with developing and validating the telomere assay.

413

## 414 FUNDING

415 Funding provided by the Natural Sciences and Engineering Research Council of Canada to C.F.P,  
416 J.R.T and M.D.R., the Canada Research Chairs Program to J.R.T. and M.D.R, the Canada  
417 Foundation for Innovation and the Research and Development Corporation of Newfoundland  
418 Labrador to C.F.P., the University of Manitoba Research Grants Program to M.J.G. and J.R.T., a  
419 University Research Fellowship from the Royal Society (London) to M.B.M., a UK NERC  
420 Research Grant awarded to M.B.M., and IISD-ELA for Research Fellow support for M.D.R., and  
421 accommodation and food provided to C.F.P., A.C.R., J.R.T.

422

## 423 REFERENCES

- 424 1. Nussey D.H., Froy H., Lemaître J.F., Gaillard J.M., Austad S.N. 2013 Senescence in  
425 natural populations of animals: widespread evidence and its implications for bio-  
426 gerontology. *Ageing Res Rev* **12**(1), 214-225. (doi:10.1016/j.arr.2012.07.004).
- 427 2. Bernard C., Compagnoni A., Salguero-Gómez R. 2020 Testing Finch's hypothesis: the  
428 role of organismal modularity on the escape from actuarial senescence. *Functional Ecology*  
429 **34**(1), 88-106. (doi:10.1111/1365-2435.13486).
- 430 3. Jones O.R., Scheuerlein A., Salguero-Gomez R., Camarda C.G., Schaible R., Casper B.B.,  
431 Dahlgren J.P., Ehrlen J., Garcia M.B., Menges E.S., et al. 2014 Diversity of ageing across the  
432 tree of life. *Nature* **505**(7482), 169-173. (doi:10.1038/nature12789).
- 433 4. Baudisch A., Stott I. 2019 A pace and shape perspective on fertility. *Methods in*  
434 *Ecology and Evolution* **10**(11), 1941-1951. (doi:10.1111/2041-210x.13289).
- 435 5. Baudisch A., Vaupel J.W. 2012 Getting to the root of aging. *Science* **338**(6107), 618-  
436 619. (doi:10.1126/science.1226467).
- 437 6. Moorad J., Promislow D., Silvertown J. 2019 Evolutionary ecology of senescence and  
438 a reassessment of Williams' 'extrinsic mortality' hypothesis. *Trends Ecol Evol* **34**(6), 519-  
439 530. (doi:10.1016/j.tree.2019.02.006).
- 440 7. Roper M., Capdevila P., Salguero-Gómez R. 2021 Senescence: why and where  
441 selection gradients might not decline with age. *Proceedings of the Royal Society B: Biological*  
442 *Sciences* **288**(1955), 20210851. (doi:10.1098/rspb.2021.0851).
- 443 8. Maklakov A.A., Chapman T. 2019 Evolution of ageing as a tangle of trade-offs: energy  
444 versus function. *Proc Biol Sci* **286**(1911), 20191604. (doi:10.1098/rspb.2019.1604).
- 445 9. Williams G.C. 1957 Pleiotropy, natural selection, and the evolution of senescence.  
446 *Evolution* **11**(4), 398-411. (doi:10.1111/j.1558-5646.1957.tb02911.x).
- 447 10. da Silva J. 2018 Reports of the death of extrinsic mortality moulding senescence have  
448 been greatly exaggerated. *Evol Biol* **45**(2), 140-143. (doi:10.1007/s11692-018-9446-y).
- 449 11. Danko M.J., Burger O., Argasinski K., Kozłowski J. 2018 Extrinsic mortality can shape  
450 life-history traits, including senescence. *Evol Biol* **45**(4), 395-404. (doi:10.1007/s11692-  
451 018-9458-7).
- 452 12. Medawar P.B. 1952 *An unsolved problem of biology*. London, H.K. Lewis.
- 453 13. Kirkwood T.B.L. 1977 Evolution of aging. *Nature* **270**(5635), 301-304.  
454 (doi:10.1038/270301a0).
- 455 14. Blagosklonny M.V. 2006 Aging and immortality: quasi-programmed senescence and  
456 its pharmacologic inhibition. *Cell Cycle* **5**(18), 2087-2102. (doi:10.4161/cc.5.18.3288).
- 457 15. Cohen A.A., Coste C.F.D., Li X.-Y., Bourg S., Pavard S., Gaillard J.-M. 2020 Are trade-offs  
458 really the key drivers of ageing and life span? *Functional Ecology* **34**(1), 153-166.  
459 (doi:10.1111/1365-2435.13444).
- 460 16. Gaillard J.M., Lemaître J.F., Fox C. 2020 An integrative view of senescence in nature.  
461 *Functional Ecology* **34**(1), 4-16. (doi:10.1111/1365-2435.13506).
- 462 17. Lind M.I., Ravindran S., Sekajova Z., Carlsson H., Hinas A., Maklakov A.A. 2019  
463 Experimentally reduced insulin/IGF-1 signaling in adulthood extends lifespan of parents  
464 and improves Darwinian fitness of their offspring. *Evol Lett* **3**(2), 207-216.  
465 (doi:10.1002/evl3.108).

- 466 18. Finch C.E. 1998 Variations in senescence and longevity include the possibility of  
467 negligible senescence. *Journals of Gerontology Series a-Biological Sciences and Medical*  
468 *Sciences* **53**(4), B235-B239. (doi:10.1093/gerona/53A.4.B235).
- 469 19. Finch C.E. 2009 Update on slow aging and negligible senescence--a mini-review.  
470 *Gerontology* **55**(3), 307-313. (doi:10.1159/000215589).
- 471 20. Vaupel J.W., Baudisch A., Dolling M., Roach D.A., Gampe J. 2004 The case for negative  
472 senescence. *Theor Popul Biol* **65**(4), 339-351. (doi:10.1016/j.tpb.2003.12.003).
- 473 21. Fletcher Q.E., Selman C. 2015 Aging in the wild: insights from free-living and non-  
474 model organisms. *Exp Gerontol* **71**, 1-3. (doi:10.1016/j.exger.2015.09.015).
- 475 22. Olsson M., Shine R. 1996 Does reproductive success increase with age or with size in  
476 species with indeterminate growth? A case study using sand lizards (*Lacerta agilis*).  
477 *Oecologia* **105**(2), 175-178. (doi:10.1007/bf00328543).
- 478 23. Roach D.A., Smith E.F., Gaillard J.-M. 2020 Life-history trade-offs and senescence in  
479 plants. *Functional Ecology* **34**(1), 17-25. (doi:10.1111/1365-2435.13461).
- 480 24. Hoekstra L.A., Schwartz T.S., Sparkman A.M., Miller D.A.W., Bronikowski A.M.,  
481 Lemaître J.F. 2020 The untapped potential of reptile biodiversity for understanding how  
482 and why animals age. *Functional Ecology* **34**(1), 38-54. (doi:10.1111/1365-2435.13450).
- 483 25. Bonduriansky R., Brassil C.E. 2002 Rapid and costly ageing in wild male flies. *Nature*  
484 **420**(6914), 377-377. (doi:10.1038/420377a).
- 485 26. Monaghan P., Charmantier A., Nussey D.H., Ricklefs R.E. 2008 The evolutionary  
486 ecology of senescence. *Functional Ecology* **22**(3), 371-378. (doi:10.1111/j.1365-  
487 2435.2008.01418.x).
- 488 27. Johnson S.L., Zellhuber-McMillan S., Gillum J., Dunleavy J., Evans J.P., Nakagawa S.,  
489 Gemmell N.J. 2018 Evidence that fertility trades off with early offspring fitness as males age.  
490 *Proceedings of the Royal Society B-Biological Sciences* **285**(1871). (doi:20172174  
491 10.1098/rspb.2017.2174).
- 492 28. Roach D.A., Carey J.R. 2014 Population biology of aging in the wild. *Annual Review of*  
493 *Ecology, Evolution, and Systematics* **45**(1), 421-443. (doi:10.1146/annurev-ecolsys-120213-  
494 091730).
- 495 29. Johnson S.L., Gemmell N.J. 2012 Are old males still good males and can females tell  
496 the difference? Do hidden advantages of mating with old males off-set costs related to  
497 fertility, or are we missing something else? *Bioessays* **34**(7), 609-619.  
498 (doi:10.1002/bies.201100157).
- 499 30. Angelier F., Weimerskirch H., Barbraud C., Chastel O., Hopkins W. 2019 Is telomere  
500 length a molecular marker of individual quality? Insights from a long-lived bird. *Functional*  
501 *Ecology* **33**(6), 1076-1087. (doi:10.1111/1365-2435.13307).
- 502 31. Monaghan P., Metcalfe N.B. 2019 The deteriorating soma and the indispensable  
503 germline: gamete senescence and offspring fitness. *Proc Biol Sci* **286**(1917), 20192187.  
504 (doi:10.1098/rspb.2019.2187).
- 505 32. Lemaître J.F., Berger V., Bonenfant C., Douhard M., Gamelon M., Plard F., Gaillard J.M.  
506 2015 Early-late life trade-offs and the evolution of ageing in the wild. *Proc Biol Sci*  
507 **282**(1806), 20150209. (doi:10.1098/rspb.2015.0209).
- 508 33. Lemaître J.F., Gaillard J.M. 2017 Reproductive senescence: new perspectives in the  
509 wild. *Biol Rev Camb Philos Soc* **92**(4), 2182-2199. (doi:10.1111/brv.12328).
- 510 34. Graves B.M. 2007 Sexual selection effects on the evolution of senescence.  
511 *Evolutionary Ecology* **21**(5), 663-668. (doi:10.1007/s10682-006-9144-6).

- 512 35. Bartosch-Harlid A., Berlin S., Smith N.G.C., Moller A.P., Ellegren H. 2003 Life history  
513 and the male mutation bias. *Evolution* **57**(10), 2398-2406. (doi:10.1554/03-036).
- 514 36. Beirne C., Delahay R., Young A. 2015 Sex differences in senescence: the role of intra-  
515 sexual competition in early adulthood. *Proc Biol Sci* **282**(1811).  
516 (doi:10.1098/rspb.2015.1086).
- 517 37. Bonduriansky R., Maklakov A., Zajitschek F., Brooks R. 2008 Sexual selection, sexual  
518 conflict and the evolution of ageing and life span. *Functional Ecology* **22**(3), 443-453.  
519 (doi:10.1111/j.1365-2435.2008.01417.x).
- 520 38. Lemaitre J.F., Gaillard J.M., Pemberton J.M., Clutton-Brock T.H., Nussey D.H. 2014  
521 Early life expenditure in sexual competition is associated with increased reproductive  
522 senescence in male red deer. *Proc Biol Sci* **281**(1792). (doi:10.1098/rspb.2014.0792).
- 523 39. Metcalf C.J.E., Roth O., Graham A.L., Lemaître J.-F. 2020 Why leveraging sex  
524 differences in immune trade-offs may illuminate the evolution of senescence. *Functional*  
525 *Ecology* **34**(1), 129-140. (doi:10.1111/1365-2435.13458).
- 526 40. Grunst M.L., Grunst A.S., Formica V.A., Korody M.L., Betuel A.M., Barcelo-Serra M.,  
527 Gonser R.A., Tuttle E.M. 2018 Actuarial senescence in a dimorphic bird: different rates of  
528 ageing in morphs with discrete reproductive strategies. *Proceedings of the Royal Society B-*  
529 *Biological Sciences* **285**(1892). (doi:20182053  
530 10.1098/rspb.2018.2053).
- 531 41. Maklakov A.A., Immler S. 2016 The expensive germline and the evolution of ageing.  
532 *Current Biology* **26**(13), R577-R586. (doi:10.1016/j.cub.2016.04.012).
- 533 42. Pizzari T., Dean R., Pacey A., Moore H., Bonsall M.B. 2008 The evolutionary ecology of  
534 pre- and post-meiotic sperm senescence. *Trends Ecol Evol* **23**(3), 131-140.  
535 (doi:10.1016/j.tree.2007.12.003).
- 536 43. Reinhardt K., Turnell B. 2019 Sperm ageing: a complex business. *Functional Ecology*  
537 **33**(7), 1188-1189. (doi:10.1111/1365-2435.13350).
- 538 44. Purchase C.F., Evans J.P., Roncal J. 2021 Intergrating natural and sexual selection  
539 across the biphasic life cycle. *EcoEvoRxiv*. (doi:https://doi.org/10.32942/osf.io/eu3am).
- 540 45. Vega-Trejo R., Fox R.J., Iglesias-Carrasco M., Head M.L., Jennions M.D., Priest N. 2019  
541 The effects of male age, sperm age and mating history on ejaculate senescence. *Functional*  
542 *Ecology* **33**(7), 1267-1279. (doi:10.1111/1365-2435.13305).
- 543 46. Crow J.F. 2000 The origins patterns and implications of human spontaneous  
544 mutation. *Nature Reviews Genetics* **1**(1), 40-47. (doi:10.1038/35049558).
- 545 47. Gasparini C., Devigili A., Pilastro A. 2019 Sexual selection and ageing: interplay  
546 between pre- and post-copulatory traits senescence in the guppy. *Proceedings of the Royal*  
547 *Society B-Biological Sciences* **286**(1897). (doi:20182873  
548 10.1098/rspb.2018.2873).
- 549 48. Aich U., Head M.L., Fox R.J., Jennions M.D. 2021 Male age alone predicts paternity  
550 success under sperm competition when effects of age and past mating effort are  
551 experimentally separated. *Proceedings of the Royal Society B: Biological Sciences* **288**(1955),  
552 20210979. (doi:doi:10.1098/rspb.2021.0979).
- 553 49. Behnke R.J. 2002 *Trout and salmon of North America*. New York, The Free Press; 359  
554 p.
- 555 50. Esteve M., McLennan D.A., Gunn J.M. 2007 Lake trout (*Salvelinus namaycush*)  
556 spawning behaviour: the evolution of a new female strategy. *Environmental Biology of*  
557 *Fishes* **83**(1), 69-76. (doi:10.1007/s10641-007-9272-z).



- 558 51. Parker G.A. 1990 Sperm competition games - raffles and roles. *Proceedings of the*  
559 *Royal Society B-Biological Sciences* **242**(1304), 120-126. (doi:10.1098/rspb.1990.0114).
- 560 52. Cruz-Font L., Shuter B.J., Blanchfield P.J., Minns C.K., Rennie M.D. 2019 Life at the top:  
561 lake ecotype influences the foraging pattern, metabolic costs and life history of an apex fish  
562 predator. *Journal of Animal Ecology* **88**(5), 702-716. (doi:10.1111/1365-2656.12956).
- 563 53. Kennedy P.J., Bartley T.J., Gillis D.M., McCann K.S., Rennie M.D. 2018 Offshore prey  
564 densities facilitate similar life history and behavioral patterns in two distinct aquatic apex  
565 predators, northern pike and lake trout. *Transactions of the American Fisheries Society*  
566 **147**(5), 972-995. (doi:10.1002/tafs.10090).
- 567 54. Purchase C.F., Collins N.C., Shuter B.J. 2005 Sensitivity of maximum sustainable  
568 harvest rates to intra-specific life history variability of lake trout (*Salvelinus namaycush*)  
569 and walleye (*Sander vitreus*). *Fisheries Research* **72**(2-3), 141-148.  
570 (doi:10.1016/j.fishres.2004.11.006).
- 571 55. Kirkwood T.B., Melov S. 2011 On the programmed/non-programmed nature of  
572 ageing within the life history. *Curr Biol* **21**(18), R701-707. (doi:10.1016/j.cub.2011.07.020).
- 573 56. Campana S.E., Casselman J.M., Jones C.M. 2008 Bomb radiocarbon chronologies in the  
574 Arctic, with implications for the age validation of lake trout (*Salvelinus namaycush*) and  
575 other Arctic species. *Canadian Journal of Fisheries and Aquatic Sciences* **65**(4), 733-743.  
576 (doi:10.1139/f08-012).
- 577 57. Mills K.H., Chalanchuk S.M., Allan D.J. 2000 Recovery of fish populations in Lake 223  
578 from experimental acidification. *Canadian Journal of Fisheries and Aquatic Sciences* **57**(1),  
579 192-204. (doi:10.1139/cjfas-57-1-192).
- 580 58. Butts I.A.E., Hilmarisdóttir G.S., Zadmajid V., Gallego V., Støttrup J.G., Jacobsen C.,  
581 Krüger-Johnsen M., Politis S.N., Asturiano J.F., Holst L.K., et al. 2020 Dietary amino acids  
582 impact sperm performance traits for a catadromous fish, *Anguilla anguilla* reared in  
583 captivity. *Aquaculture* **518**, 734602. (doi:10.1016/j.aquaculture.2019.734602).
- 584 59. Blackwell B.G., Brown M.L., Willis D.W. 2000 Relative weight (Wr) status and current  
585 use in fisheries assessment and management. *Reviews in Fisheries Science* **8**(1), 1-44.  
586 (doi:10.1080/10641260091129161).
- 587 60. Hartman K.J., Margraf F.J. 2006 Relationships among condition indices, feeding and  
588 growth of wallye in Lake Erie. *Fisheries Management and Ecology* **13**, 121-130.
- 589 61. Carneiro M.C., de Castro I.P., Ferreira M.G. 2016 Telomeres in aging and disease:  
590 lessons from zebrafish. *Dis Model Mech* **9**(7), 737-748. (doi:10.1242/dmm.025130).
- 591 62. Hatakeyama H., Yamazaki H., Nakamura K.-I., Izumiyama-Shimomura N., Aida J.,  
592 Suzuki H., Tsuchida S., Matsuura M., Takubo K., Ishikawa N. 2016 Telomere attrition and  
593 restoration in the normal teleost *Oryzias latipes* are linked to growth rate and telomerase  
594 activity at each life stage. *Aging* **8**, 62-75.
- 595 63. Rollings N., Miller E., Olsson M. 2014 Telomeric attrition with age and temperature in  
596 Eastern mosquitofish (*Gambusia holbrooki*). *Naturwissenschaften* **101**(3), 241-244.  
597 (doi:10.1007/s00114-014-1142-x).
- 598 64. Mills K.H., Beamish R.J. 1980 Comparisons of fin-ray and scale age-determinations  
599 for lake whitefish (*Coregonus clupeaformis*) and their implications for estimates of growth  
600 and annual survival. *Canadian Journal of Fisheries and Aquatic Sciences* **37**(3), 534-544.  
601 (doi:10.1139/f80-068).

- 602 65. Piccolo J.J., Hubert W.A., Whaley R.A. 1993 Standard weight equation for lake trout.  
603 *North American Journal of Fisheries Management* **13**(2), 401-404. (doi:10.1577/1548-  
604 8675(1993)013<0401:sweflt>2.3.co;2).
- 605 66. Rennie M.D., Kennedy P.J., Mills K.H., Rodgers C.M.C., Charles C., Hrenchuk L.E.,  
606 Chalanchuk S., Blanchfield P.J., Paterson M.J., Podemski C.L. 2019 Impacts of freshwater  
607 aquaculture on fish communities: a whole-ecosystem experimental approach. *Freshwater*  
608 *Biology* **64**(5), 870-885. (doi:10.1111/fwb.13269).
- 609 67. Purchase C.F., Rooke A.C. 2020 Freezing ovarian fluid does not alter how it affects  
610 fish sperm swimming performance: creating a cryptic female choice 'spice rack' for use in  
611 split-ejaculate experimentation. *Journal of Fish Biology* **96**(3), 693-699.  
612 (doi:10.1111/jfb.14263).
- 613 68. Purchase C.F., Moreau D.T. 2012 Stressful environments induce novel phenotypic  
614 variation: hierarchical reaction norms for sperm performance of a pervasive invader. *Ecol*  
615 *Evol* **2**(10), 2567-2576. (doi:10.1002/ece3.364).
- 616 69. Purchase C.F., Butts I.A.E., Alonso-Fernández A., Trippel E.A. 2010 Thermal reaction  
617 norms in sperm performance of Atlantic cod (*Gadus morhua*). *Canadian Journal of Fisheries*  
618 *and Aquatic Sciences* **67**(3), 498-510. (doi:10.1139/f10-001).
- 619 70. Johnson K., Butts I.A.E., Wilson C.C., Pitcher T.E. 2013 Sperm quality of hatchery-  
620 reared lake trout throughout the spawning season. *North American Journal of Aquaculture*  
621 **75**(1), 102-108. (doi:10.1080/15222055.2012.711277).
- 622 71. Purchase C.F., Earle P.T. 2012 Modifications to the ImageJ computer assisted sperm  
623 analysis plugin greatly improve efficiency and fundamentally alter the scope of attainable  
624 data. *Journal of Applied Ichthyology* **28**(6), 1013-1016. (doi:10.1111/jai.12070).
- 625 72. Gage M.J.G., Macfarlane C.P., Yeates S., Ward R.G., Searle J.B., Parker G.A. 2004  
626 Spermatozoal traits and sperm competition in Atlantic salmon. *Current Biology* **14**(1), 44-  
627 47. (doi:10.1016/j.cub.2003.12.028).
- 628 73. Roff D.A. 2002 *Life history evolution*. Sunderland, Sinauer; 256 p.  
629  
630

## 631 FIGURE CAPTIONS

- 632
- 633 Figure 1: Our hierarchical conceptualization of the main evolutionary theories of senescence.  
634 Modified and expanded form Maklakov and Chapman [9].
- 635
- 636 Figure 2: Longitudinal data from individually tagged adult male lake trout. a) adults of age 9-  
637 years (back) and 37-years (front); b) and c) annual mortality probability as a function of  
638 continuous age in Lake 223 and Lake 224, respectively. The solid line is the mean predicted

639 probability, and the dashed lines are the 95% credible intervals; d) body condition relative to age  
640 from Lake 224. Each black line is a resampled fish (minimum six times) during October, 1976-  
641 2017.

642  
643 Figure 3: Phenotypic measures of young (pink: 4–10 years, n = 30) and old (blue: 18–37 years, n  
644 = 30) adult male lake trout from Lake 223 (n = 30) and Lake 224 (n = 30) in October 2017. Each  
645 point represents an individual trout. (a) Body condition, (b) spermatocrit,  
646 relative telomere length of (c) red blood cells, and (d) sperm cells. Telomere data presented using  
647 Ox reference gene, points represent average of three technical replicates per individual.

648  
649 Figure 4: Sperm swimming speed ( $\mu\text{m/s}$ ) of lake trout in October 2017. (a) velocity (VCL) at 6 s  
650 post-activation across age in years (black: Lake 223, n = 30; grey: Lake 224, n = 30), and (b)  
651 decline in sperm swimming velocity with time post-activation in young (pink: 4–10 years, n =  
652 30) and old (blue: 18–37 years, n = 30) fish. Points represent average of technical replicates for  
653 each fish, lines represent average among individuals from the same lake and age category. Error  
654 bars/bands represent  $\pm 1$  SE.



non-mutually exclusive theories

*Non-adaptive*

Mutational accumulation theory (MAT)

*Adaptive – based on optimization*

Antagonistic pleiotropy hypothesis (APH)

energy “tradeoff”  
mechanism

function “constraint”  
mechanism

Disposable soma  
hypothesis (DSH)

Developmental  
function theory (DFT)  
(hyperfunction)

Figure 1

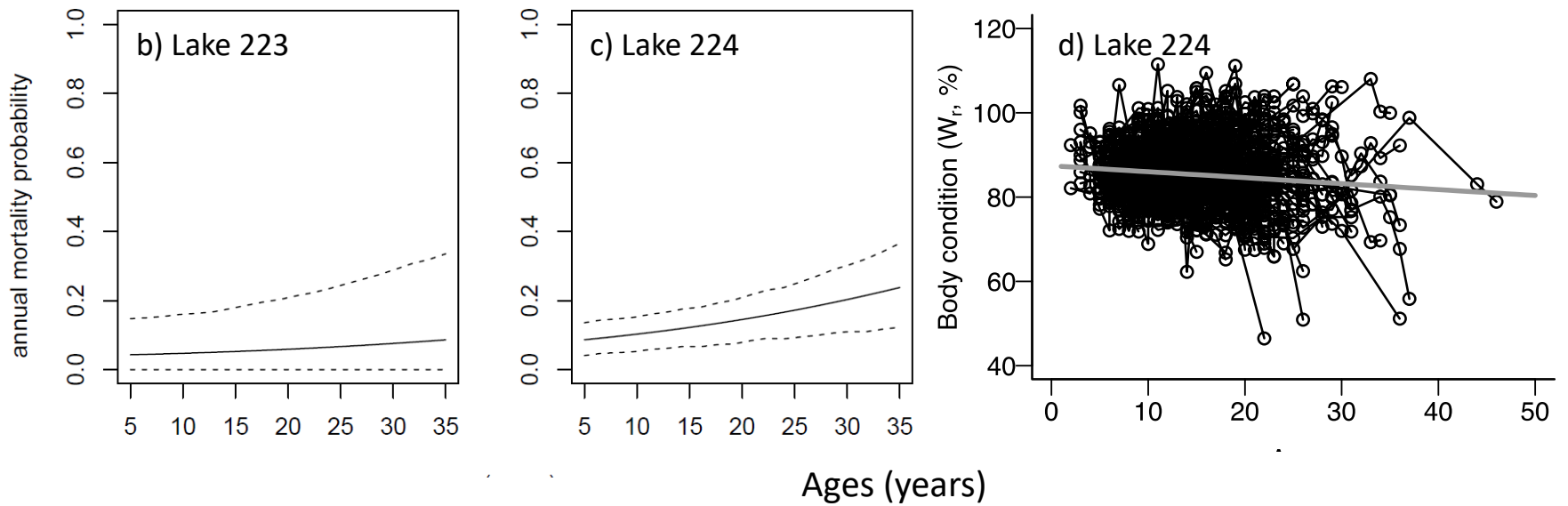
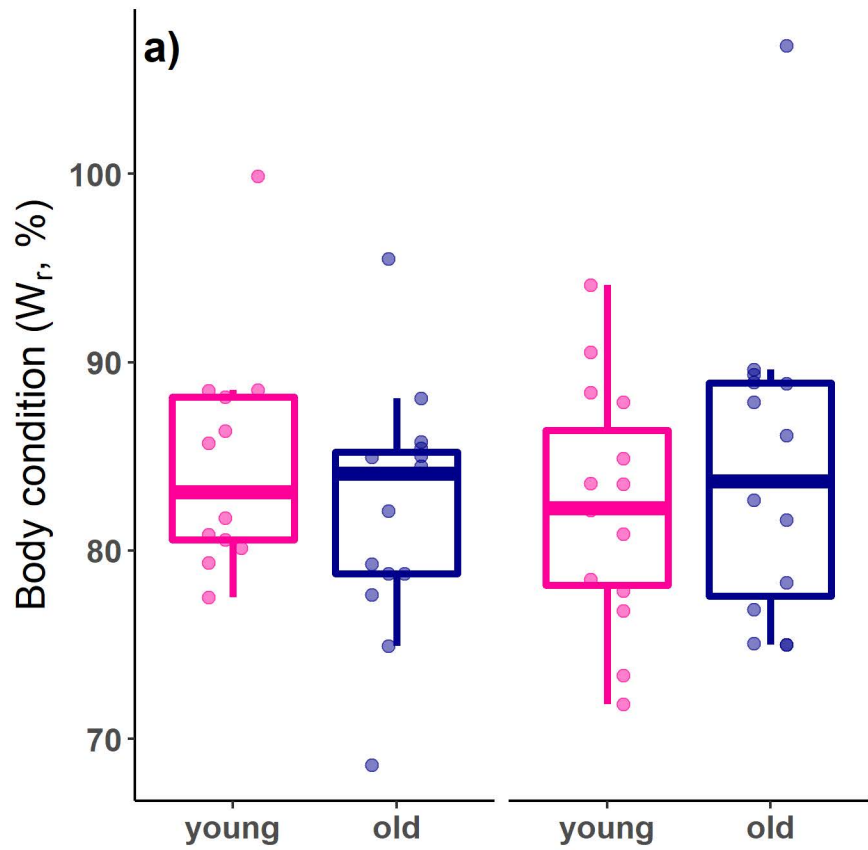


Figure 2: Longitudinal data

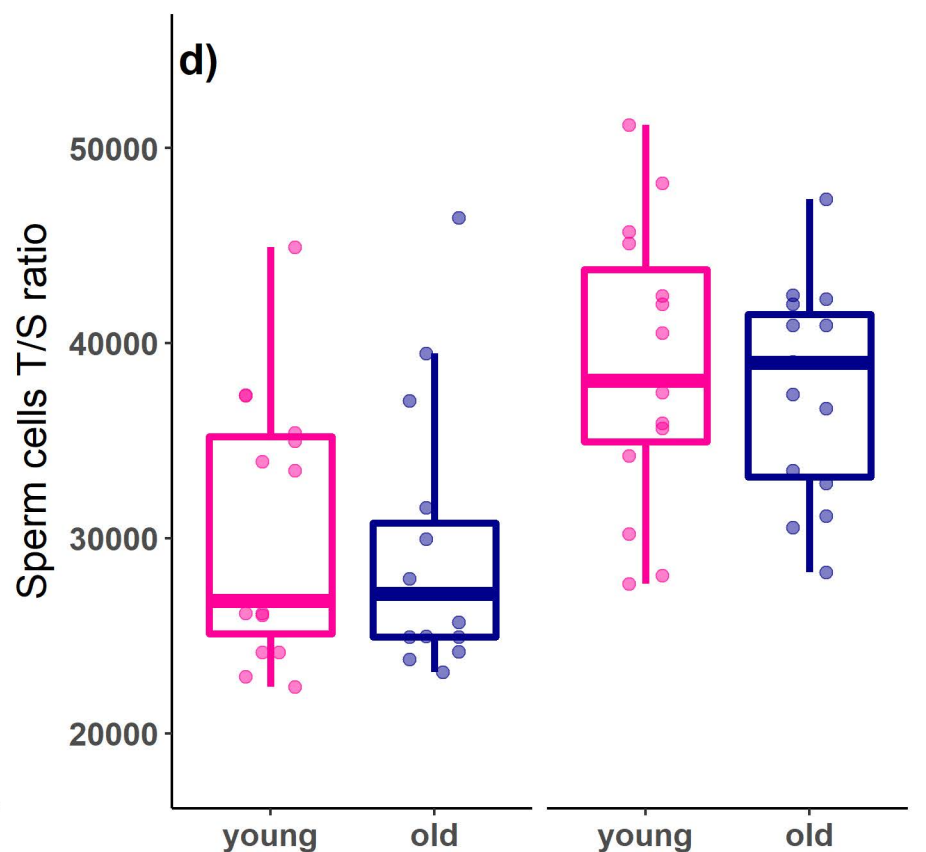
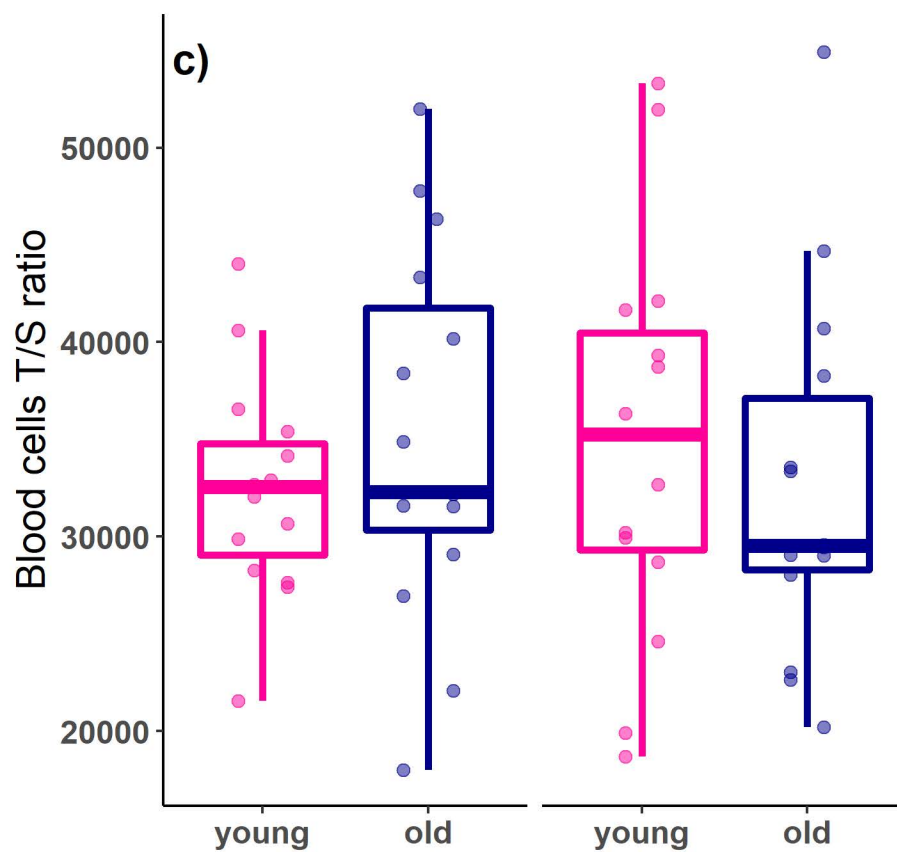
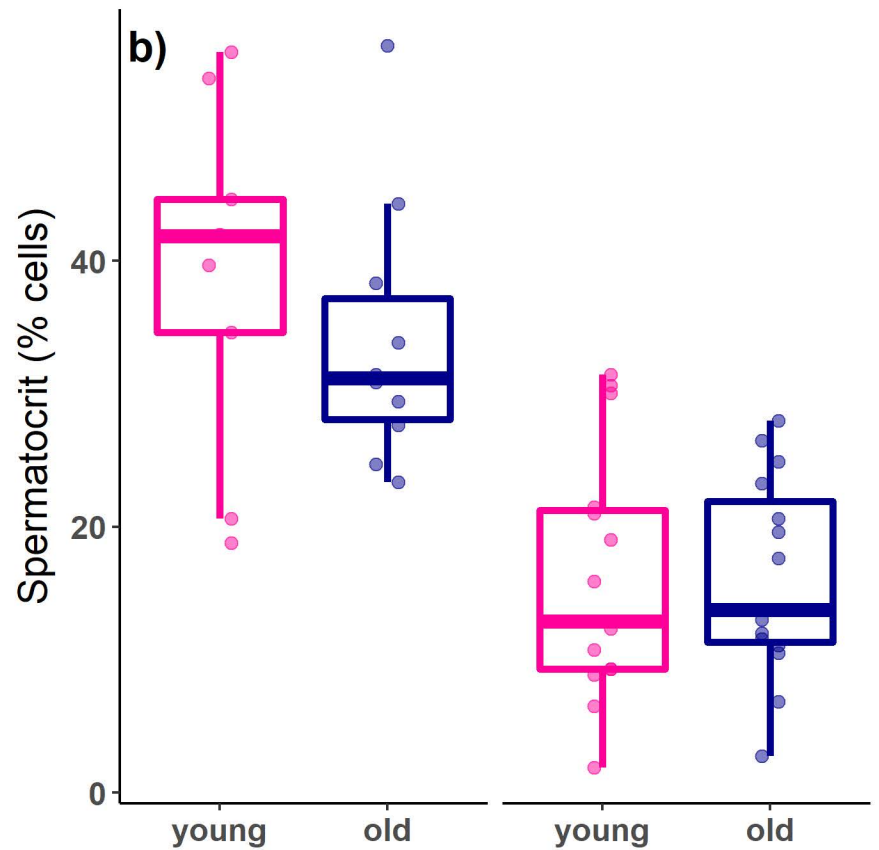
Lake 223

Lake 224

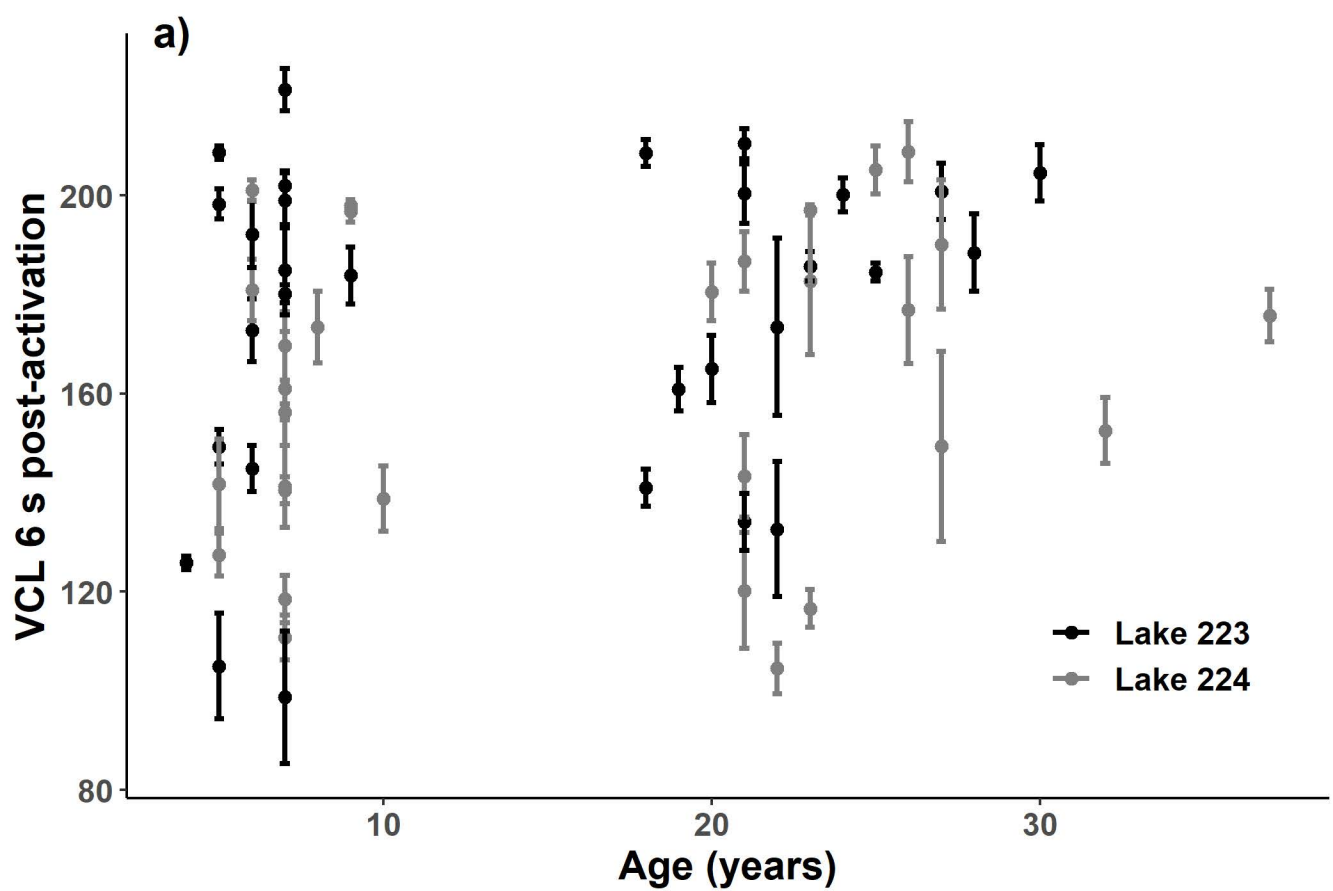


Lake 223

Lake 224

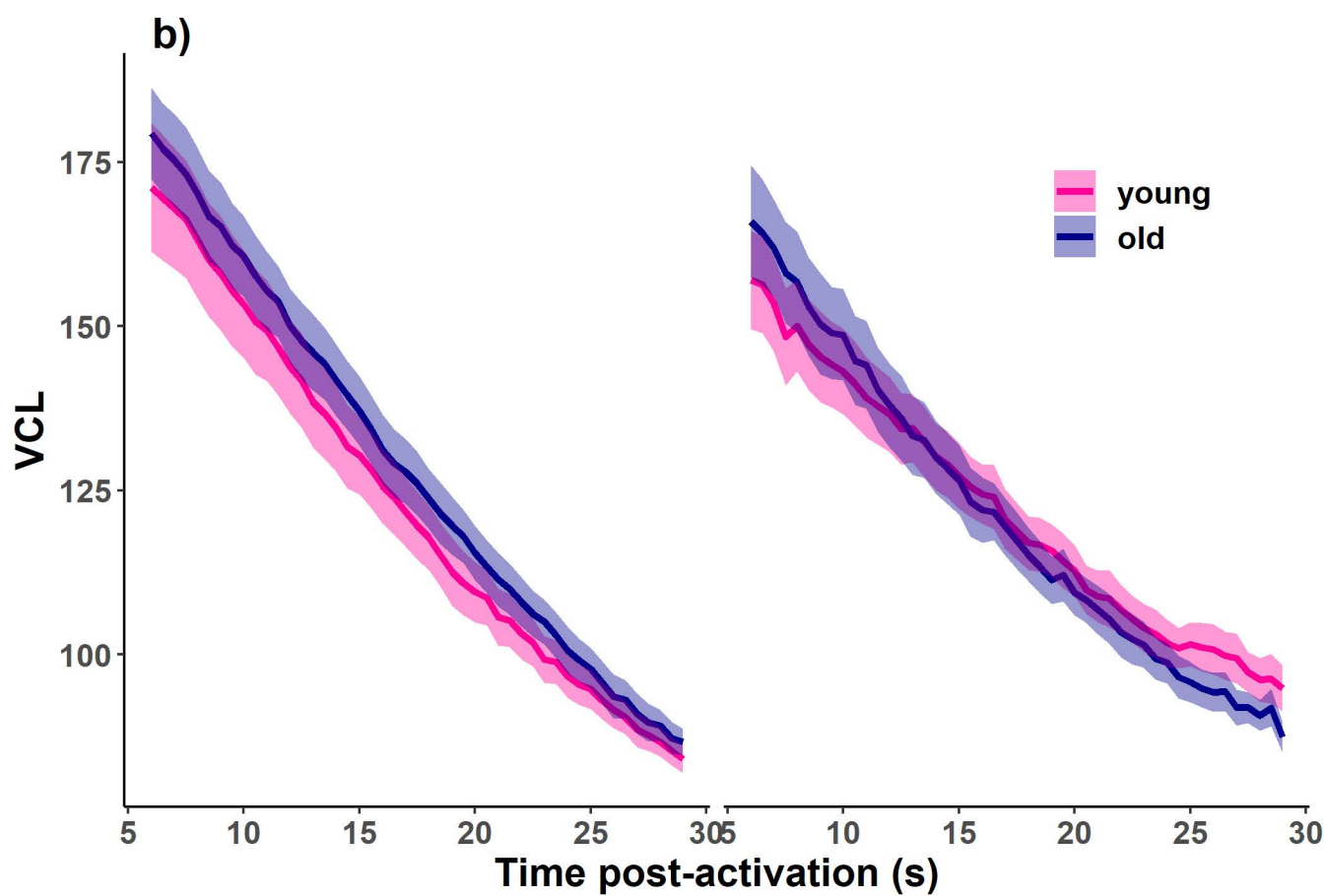


Age category



**Lake 223**

**Lake 224**



## **Supplementary Methods I – Longitudinal data**

### **Actuarial senescence**

Using all individual adult males with known ages (Lake 223 = 385, Lake 224 = 422), we treated age as a continuous variable and estimated the probability of recapture and the probability of survival (1 - mortality) of each individual adult male in each year (sampled from posterior distributions). As extrinsic adult mortality is low, and young/old adults experience the same conditions, any changes in mortality with age are assumed to be attributed to intrinsic processes.

In order to test for an increase in adult mortality with age, we fitted a Cormack Jolly Seber model (Lebreton *et al.*, 1992) with separate survival (1 – mortality) and capture probabilities as linear regression functions on the logistic scale. We fitted a first order autocorrelation structure for annual random effects on survival and recapture probability, as we expect that factors affecting these parameters (especially survival) are likely to be similar from year to year. The likelihood structure of the basic process model is thus defined as

$$g(\phi_{i,a,t}) = X_a \boldsymbol{\beta}_{\phi,a} + b_{\phi,t}$$

$$a_{i,a,t} \sim B(\phi_{i,a,t} \cdot a_{i,a,t-1})$$

where  $a_{i,a,t}$  is a latent Bernoulli variable indicating whether individual  $i$  of age  $a$  is alive at time  $t$ . An individual will survive from interval  $t - 1$  to interval  $t$  with probability  $\phi_{i,t}$ , only if it was alive at time  $t - 1$ , i.e., only if  $a_{i,a,t-1} = 1$ .  $\phi_{i,a,t}$  is modelled as a logistic regression with an effect of continuous age on survival contained in  $\boldsymbol{\beta}_{\phi,a}$ , and random effects of year,  $b_{\phi,t}$ . The temporal correlation structure of these year effects after the first year is defined by

$$b_{\phi,t} \sim N(\alpha_{\phi} b_{\phi,t-1}, \sigma_{d,\phi}^2)$$

where  $\alpha_\phi$  is the first-order autoregressive parameter describing the dependence of survival in one episode on the previous episode and  $\sigma_{d,\phi}^2$  is the variance of disturbances of the

autoregressive process. The stationary variance of such a process is  $\sigma_\phi^2 = \frac{\sigma_{d,\phi}^2}{1-\alpha_\phi^2}$ , and this

variance defines the distribution from which the random effect in the first year is drawn.

The observation model takes the form

$$g(p_{i,a,t}) = X_a \boldsymbol{\beta}_{p,a} + b_{p,t}$$
$$y_{i,a,t} \sim B(p_{i,a,t} \cdot a_{i,a,t} \cdot A_t)$$

where  $y_{i,a,t}$  is the observation (1 = captured, 0 = not captured) of individual  $i$  of age  $a$  at time  $t$ . Any individual not alive ( $a_{i,a,t} = 0$ ) cannot be observed, and those that are alive may be observed with probability  $p_{i,a,t}$ , provided that sampling was conducted in year  $t$  ( $A_t = 1$  if sampling was conducted;  $A_t = 0$  otherwise). As in the process (survival) part of the model, the probability of capture of live individuals is modelled as a logistic regression, with separate intercepts for each age, and an autoregressive structure for annual variation. The structure for annual variation is directly analogous to that described above for the survival part of the model.

The model was sampled by Gibbs sampling using jags (Plummer, 2010) in R version 3.6.2. We used diffuse normal priors on all fixed effects and autoregression parameters, and diffuse gamma priors on the precision (inverse of the variance) of the disturbances of the survival and capture parts of the model.

### *Additional parameters*

In addition, we tested models that included combinations of a quadratic term for the rate of mortality change and a parameter that varied the minimum mortality rate. However, these models did not converge well and were not numerically stable. This suggests that a



quadratic term did not fit our data, and therefore, we did not use these parameters in the final model described above. However, we have left these as options within our scripts for others to see how they were included, alongside the scripts for plots we used to check for convergence.

## Body condition

Length-based body condition was estimated as a percentage of standard weight (1993). Fish that were recaptured at least 6 times during their adult life were used to determine if condition declined with age, and were analyzed with a mixed effects modelling framework using the *lme4* package (Bates *et al.*, 2014) in R. Condition was evaluated as a function of fish age (fixed effect), and repeated measures on the same individuals (modelled as a random slope), and the year sampled (random intercept). Significance of fixed effects was assessed using the Satterthwaite approximation for degrees of freedom with the *lmerTest* package (Kuznetsova *et al.*, 2017). Random effects were retained if found to be significant in log-likelihood ratio tests using the *anova()* function in R. Assumptions of normality and homogeneity of variance were verified using model residuals. Only fish from Lake 224 were used in this analysis as exclusion of data prior to 1990 (Mills *et al.*, 2000) limited sample sizes in Lake 223.

## References

- Bates, D., Maechler, M., Bolker, B. & Walker, S. (2014). *lme4*: Linear mixed-effects models using eigen and S4., p. Retrieved from <https://cran.rproject.org/web/packages/lme4>.
- Kuznetsova, A., Brockhoff, P. B. & Christensen, R. H. B. (2017). *lmerTest* Package: tests in Linear Mixed Effects Models. *Journal of Statistical Software* **82**.

bioRxiv preprint doi: <https://doi.org/10.1101/2021.10.05.463025>; this version posted October 5, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under a [CC-BY 4.0 International license](#).

Lebreton, J.-D., Burnham, K. P., Clobert, J. & Anderson, D. R. (1992). Modeling survival and testing biological hypotheses using marked animals: a unified approach with case studies. *Ecological Monographs* **62**, 67-118.

Mills, K. H., Chalanchuk, S. M. & Allan, D. J. (2000). Recovery of fish populations in Lake 223 from experimental acidification. *Canadian Journal of Fisheries and Aquatic Sciences* **57**, 192-204.

Piccolo, J. J., Hubert, W. A. & Whaley, R. A. (1993). Standard weight equation for lake trout. *North American Journal of Fisheries Management* **13**, 401-404.

Plummer, M. (2010). Jags version 2.2.0 user manual.



## **Supplemental Methods II – fish used in 2017**

Table S1: Adult male lake trout sampled from spawning grounds in October 2017. Ages were known in years, and sampled as categories (young = 4–10, old = 18–37). Some data were missing on some fish.

Lake	Date sampled	Tag code	Age category	Age (years)	Total length (cm)	Weight (grams)
223	12-Oct	5-394577	young	5	413	523
223	12-Oct	5-401420	young	5	408	533
223	12-Oct	5-398162	young	6	432	594
223	12-Oct	434B567237	young	7	428	563
223	12-Oct	5-397652	young	7	461	818
223	12-Oct	5-048133	old	20	487	876
223	12-Oct	5-009735	old	21	473	845
223	12-Oct	5-065013	old	21	452	595
223	12-Oct	5-065079	old	22	463	728
223	12-Oct	5-065056	old	25	432	636
223	15-Oct	5-398312	young	5	410	509
223	15-Oct	5-009640	young	6	425	613
223	15-Oct	5-388070	young	7	432	612
223	15-Oct	5-013070	young	9	435	675
223	15-Oct	5-009760	old	18	332	715
223	15-Oct	5-065071	old	19	469	770
223	15-Oct	5-009723	old	23	437	638
223	15-Oct	5-065024	old	28	473	753
223	16-Oct	5-399348	young	7	451	690
223	16-Oct	5-009771	old	21	481	836
223	17-Oct	5-398523	young	4	403	529
223	17-Oct	5-401235	young	5	420	missing
223	17-Oct	5-013027	young	6	412	641
223	17-Oct	5-013198	young	7	425	590
223	17-Oct	5-397012	young	7	missing	missing
223	17-Oct	5-009787	old	18	437	664
223	17-Oct	5-009783	old	22	450	753
223	17-Oct	5-064922	old	24	453	738
223	17-Oct	5-009752	old	27	493	986
223	17-Oct	5-065058	old	30	459	775
224	14-Oct	5-398275	young	5	370	426
224	14-Oct	5-012925	young	7	420	562
224	14-Oct	5-012966	young	7	432	615
224	14-Oct	5-012970	young	7	409	554
224	14-Oct	5-012999	young	7	431	583

224	14-Oct	5-400985	young	7	400	528
224	14-Oct	5-013379	young	8	442	674
224	14-Oct	5-064585	old	21	436	593
224	14-Oct	5-046456	old	22	427	644
224	14-Oct	5-046426	old	23	463	824
224	14-Oct	5-064707	old	23	476	770
224	14-Oct	5-010046	old	27	441	712
224	14-Oct	5-064701	old	27	455	794
224	14-Oct	5-064994	old	32	453	722
224	16-Oct	5-395632	young	5	391	416
224	16-Oct	5-012848	young	6	432	583
224	16-Oct	5-402285	young	6	406	495
224	16-Oct	5-012909	young	7	430	648
224	16-Oct	5-013324	young	7	401	491
224	16-Oct	5-009816	young	9	447	601
224	16-Oct	5-012995	young	9	408	528
224	16-Oct	5-012828	young	10	440	583
224	16-Oct	5-010086	old	20	454	736
224	16-Oct	5-064608	old	21	491	1008
224	16-Oct	5-064742	old	21	466	718
224	16-Oct	5-046561	old	23	406	527
224	16-Oct	5-010204	old	25	494	1236
224	16-Oct	5-009882	old	26	461	724
224	16-Oct	5-064699	old	26	444	668
224	16-Oct	5-012804	old	37	434	570

### **Supplemental Methods III – sperm methods**

New sperm activation medium was made each day and contained 79.9% lake water from Lake 239 (site of field station), 20% ovarian fluid, and 0.1% bovine serum albumin, which reduces the likely of sperm sticking to the glass slides (e.g., Beirão *et al.*, 2014; Beirão *et al.*, 2015). Activation medium and a semen aliquot from each male were kept at 5°C in a temperature-controlled aluminum block next to the microscope. Semen was kept on ice until transfer to the block, and was assessed within 8 hours of stripping. 0.1 µl of semen from a given male was pipetted into the opening of a 2 chamber Cytonix Microtool slide, that was prechilled to 8°C (~ temperature of spawning) using a customized Physitemp TS-4 stage cooling system. This was followed quickly by 3.95 µl of activation media, which mixed with sperm as it filled the slide chamber. We were able to consistently adjust slide position and fine focus within 6 s of sperm/media mixing. Sperm swimming performance was captured at 100 frames per second using a Prosilica GE680 monochrome camera mounted to a Leica DM IL LED inverted microscope with a 20x phase-contrast lens. The entire procedure was repeated four times for each semen sample as a means of technical replication. Videos of swimming sperm were analyzed in 0.5s increments using the Computer Assisted Sperm Analysis (CASA) plugin for ImageJ (Wilson-Leedy and Ingermann, 2007), modified by Purchase and Earle (2012). We used sperm curvilinear swimming velocity (VCL; µm/s) as a metric of sperm quality, as it has been repeatedly shown to be correlated to paternity under sperm competition (e.g., Gage *et al.*, 2004; Evans *et al.*, 2013; Alonzo *et al.*, 2016).

- Alonzo, S. H., Stiver, K. A. & Marsh-Rollo, S. E. (2016). Ovarian fluid allows directional cryptic female choice despite external fertilization. *Nature communications* **7**, 12452.
- Beirão, J., Purchase, C. F., Wringe, B. F. & Fleming, I. A. (2014). Sperm plasticity to seawater temperatures in Atlantic cod *Gadus morhua* is affected more by population origin than individual environmental exposure. *Marine Ecology Progress Series* **495**, 263-274.
- Beirão, J., Purchase, C. F., Wringe, B. F. & Fleming, I. A. (2015). Inter-population ovarian fluid variation differentially modulates sperm motility in Atlantic cod *Gadus morhua*. *Journal of Fish Biology* **87**, 54-68.
- Evans, J. P., Rosengrave, P., Gasparini, C. & Gemmell, N. J. (2013). Delineating the roles of males and females in sperm competition. *Proceedings. Biological sciences* **280**, 20132047.
- Gage, M. J. G., Macfarlane, C. P., Yeates, S., Ward, R. G., Searle, J. B. & Parker, G. A. (2004). Spermatozoal traits and sperm competition in Atlantic salmon. *Current Biology* **14**, 44-47.
- Purchase, C. F. & Earle, P. T. (2012). Modifications to the ImageJ computer assisted sperm analysis plugin greatly improve efficiency and fundamentally alter the scope of attainable data. *Journal of Applied Ichthyology* **28**, 1013-1016.
- Wilson-Leedy, J. G. & Ingermann, R. L. (2007). Development of a novel CASA system based on open source software for characterization of zebrafish sperm motility parameters. *Theriogenology* **67**, 661-672.

## **Supplemental Methods IV – relative telomere length assay**

Relative telomere length has been shown to decline with age in several fishes (Rollings *et al.*, 2014; Carneiro *et al.*, 2016; Hatakeyama *et al.*, 2016), including a wild salmonid (McLennan *et al.*, 2017), and another long-lived species (Simide *et al.*, 2016), although ectotherms do not always show declining telomere length with age (Olsson *et al.*, 2018). We measured relative telomere length from DNA recovered from red blood cells and sperm pellets using a qPCR-based approach that produces a telomere repeat (T) to single gene (S) copy number ratio (T/S).

Genomic DNA (gDNA) extractions were performed with 10  $\mu$ l of RBCs or sperm pellets using a DNeasy Blood and Tissue kit (Qiagen) according to the manufacturer's directions. The recovered DNA was quantified using a Qubit DNA HS assay kit and Qubit 2.0 Fluorometer (Thermo Fisher Scientific) and subsequently diluted to 10 ng/ $\mu$ l. The qPCR-based approach developed by Cawthon (Cawthon, 2002), which produces a telomere repeat (T) to single gene (S) copy number ratio (T/S) for each DNA sample, was used to quantify relative telomere length. Telomere repeats were amplified with the universal primer pair Tel1b and Tel2b from Epel *et al.* (Epel *et al.*, 2004). *Ox* and *FSH* were both used as single copy genes to be able to verify consistency of T/S ratios depending on which single copy gene was targeted (see Supplemental Table I for primer sequences). Primers were designed in Geneious 9.1.8 (Biomatters Ltd.) from publicly available mRNA sequences (Genbank accession numbers HQ656804.1 and HM057170.1 for *Ox* and *FSH*, respectively). qPCRs were performed on separate 384-well plates for each primer pair using the QuantStudio 5 Real-Time PCR System (Thermo Fisher Scientific). Reactions were prepared in triplicate for each sample with 2x PowerUp SYBR Green Master Mix (Thermo Fisher Scientific), 10 ng DNA per reaction, and final concentrations of 800 nM for each primer. Thermocycling conditions for the telomere qPCR were 95°C for 2 min, and 27



- McLennan, D., Armstrong, J. D., Stewart, D. C., McKelvey, S., Boner, W., Monaghan, P., Metcalfe, N. B. & Williams, T. (2017). Shorter juvenile telomere length is associated with higher survival to spawning in migratory Atlantic salmon. *Functional Ecology* **31**, 2070-2079.
- Olsson, M., Wapstra, E. & Friesen, C. (2018). Ectothermic telomeres: it's time they came in from the cold. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences* **373**.
- Rollings, N., Miller, E. & Olsson, M. (2014). Telomeric attrition with age and temperature in Eastern mosquitofish (*Gambusia holbrooki*). *Die Naturwissenschaften* **101**, 241-244.
- Simide, R., Angelier, F., Gaillard, S. & Stier, A. (2016). Age and heat stress as determinants of telomere length in a long-lived fish, the Siberian sturgeon. *Physiological and biochemical zoology : PBZ* **89**, 441-447.