Computational and experimental studies of *plasmodium falciparum* protein *Pf*AMA1 domain-II loop dynamics: implications in *Pf*AMA1-*Pf*RON2 binding event

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Keywords

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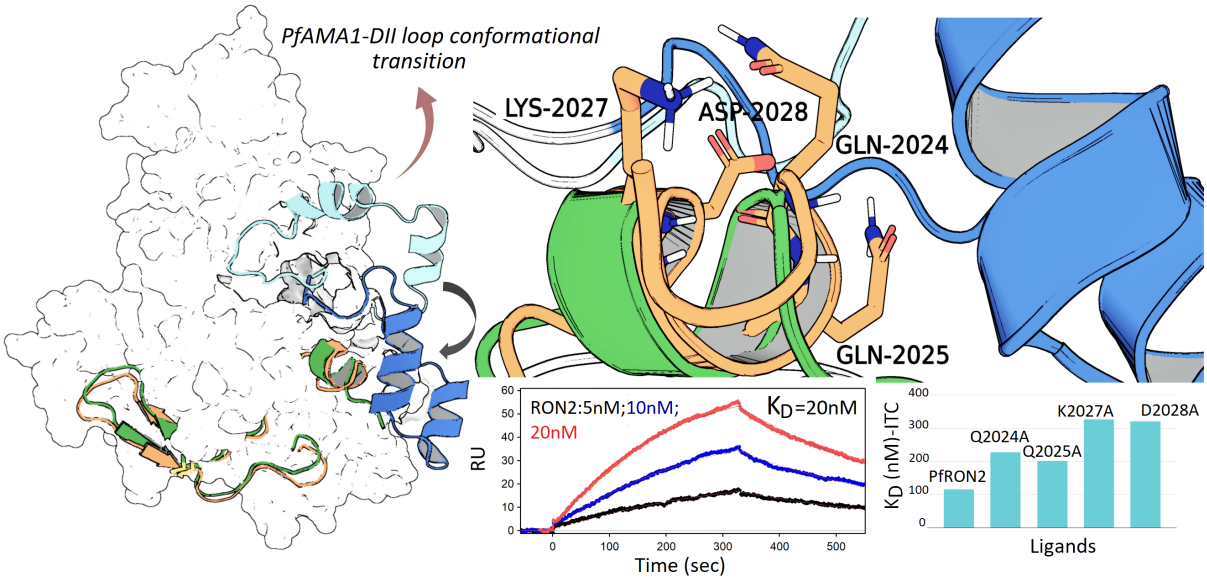
Abstract

Protein-protein interactions are interesting targets for various drug discovery campaigns. One such promising and therapeutically pertinent protein-protein complex is *Pf*AMA1-*Pf*RON2, which is involved in malarial parasite invasion into human red blood cells. A thorough understanding of the interactions between these macromolecular binding partners is absolutely necessary to design better therapeutics to fight against the age-old disease affecting mostly under-developed nations. Although crystal structures of several *Pf*AMA1-*Pf*RON2 complexes have been solved to understand the molecular interactions between these two proteins, the mechanistic aspects of the domain II loop-*Pf*RON2 association is far from clear. The current work investigates a crucial part of the recognition event; i.e., how the domain II loop of *Pf*AMA1 exerts its effect on the alpha helix of the *Pf*RON2, thus influencing the overall kinetics of this intricate recognition phenomenon. To this end, we have conducted thorough computational investigation of the dynamics and free energetics of domain II loop closing processes using molecular dynamics simulation. The computational results are validated by systematic alanine substitutions of the *Pf*RON2 peptide helix. The subsequent evaluation of the binding affinity of Ala-substituted *Pf*RON2 peptide ligands by surface plasmon resonance (SPR) and isothermal titration calorimetry (ITC) provides a rank of the relative importance of the residues in context. Our combined (computational and experimental) investigation has revealed that the domain II loop of *Pf*AMA1 is in fact responsible for arresting the *Pf*RON2 molecule from egress, K2027 and D2028 of *Pf*RON2 being the determinant residues for the capturing event. Our study provides a comprehensive understanding of the molecular recognition event between *Pf*AMA1 and *Pf*RON2, specifically in the post binding stage, which potentially can be utilized for drug discovery against malaria.

Graphical abstract



Introduction

Malaria remains the dreaded disease with high mortality rates, mostly affecting populace of the economically challenged countries of the world. Despite the availability of approved therapeutic intervention strategies mortality caused by this disease poses a global health and economic concern, mostly due to continuous emergence of drug resistant strains (1, 2). One of the primal considerations for anti-malarial drug discovery is a thorough understanding of the malaria parasite invasion biology. It is known that members of the phylum Apicomplexa, which include the malaria parasite *Plasmodium sp.*, exhibit common features in their invasion strategies despite their diverse host cell specificities and life cycle characteristics. Mechanistically, as pointed out by various research groups earlier, the formation of a moving junction between the membranes of the invading apicomplexan parasites and host cells is an essential step during the invasion process (3, 4). The moving junction in *Plasmodium sp.* comprises of two key parasite proteins; the surface protein apical membrane antigen 1 (AMA1) and its receptor, the rhoptry neck protein 2 (RON2). Reported crystal structures of the AMA1 have revealed that a partially mobile loop, termed domain-II (DII) loop, occupies part of a deep groove of the domain I in the absence of its receptor RON2 apparently safeguarding the binding groove from any other possible polygamous recognition event (5, 6). During the moving junction formation, the DII loop gets replaced by the extracellular domain of RON2 (*Pf*RON$_{2021-2059}$) letting the DII loop dangle freely as the RON2 interacts tightly within the binding pocket predominantly defined by an evolutionarily conserved hydrophobic trough. Atomistic details of *Pf*AMA1-*Pf*RON2$_{2021-2059}$ recognition event, particularly, the role of the DII loop post RON2 binding, energetics related to it at large and the role of individual amino acid residues in specific, remain rather elusive till date. Boulanger and co-workers, in one of their papers (4), provided significant clues pertaining to *Pf*AMA1-*Pf*RON2 interactions upon presence versus absence of the DII loop. They proposed that the DII loop exerts influential molecular contributions towards the kinetic locking of the *Pf*AMA1-*Pf*RON2$_{2021-2059}$ complex. The 2-fold larger affinity and the 18-fold increased half-life of the *Pf*AMA1- *Pf*RON2$_{2021-2059}$ complex in presence of the DII loop compared to the complex with truncated DII loop was attributed to the observed slower dissociation kinetics due to the restrained release of the *Pf*RON2 peptide in the former case (4). This report prompted us to investigate the atomistic details pertinent to the arrest of *Pf*RON2$_{2021-2059}$ by the DII loop. An overall understanding of this process remained difficult to address, as all AMA1-RON2 complex crystal structures available in the Protein Data Bank show missing electron densities (Lys351-Ala387) reiterating the intrinsic disorderness of the DII loop (7). Although a loop-closed crystal structure has been solved recently [PDB:6N87, (8)], it is still far from clear how the DII loop interacts with specific parts of the *Pf*RON2$_{2021-2059}$ peptide or how the interim dynamics of the loop contributes to the overall stability of the complex. In the present work, we have addressed the questions mentioned above using atomistic molecular dynamics (MD) simulations (both unbiased and umbrella sampled, summing up to approximately 1.8 microsecond) coupled with biophysical experiments with systematic alanine substitution on the terminal helix of the *Pf*RON2$_{2021-2059}$ peptide. We have used surface plasmon resonance (SPR) and isothermal titration calorimetry (ITC) to determine the binding affinity of chemically synthesized Ala-substituted *Pf*RON2$_{2021-2059}$ analogues with AMA1 which helped us to experimentally verify the roles of key amino acid residues, originally identified by molecular simulations, that drive this recognition event.

Results

Loop dynamics

Multiple repeats of atomistic simulations revealed that the DII loop reaches out towards the *Pf*RON2 peptide ligand (fig-1A) in such a way that specific residues of the terminal helix of the *Pf*RON2 ligand interact with the loop as well as other adjacent residues of the AMA1 binding pocket. However, we focused only on the DII-loop-RON2 helix interactions to comprehend and quantify contributions from individual amino acids of the *Pf*RON2 helix which play dominant roles in this inter-molecular recognition event. We have computed distance profile between Ser373 of the DII loop and Gln2024 of the RON2 helix to quantify the extent of conformational fluctuations (fig-1B). Our study found evidences of intermittent association and dissociation of the RON2 helix and the DII loop hinting at the role of DII loop in arresting the *Pf*RON2$_{2021\text{-}2059}$ binding to *Pf*AMA1.

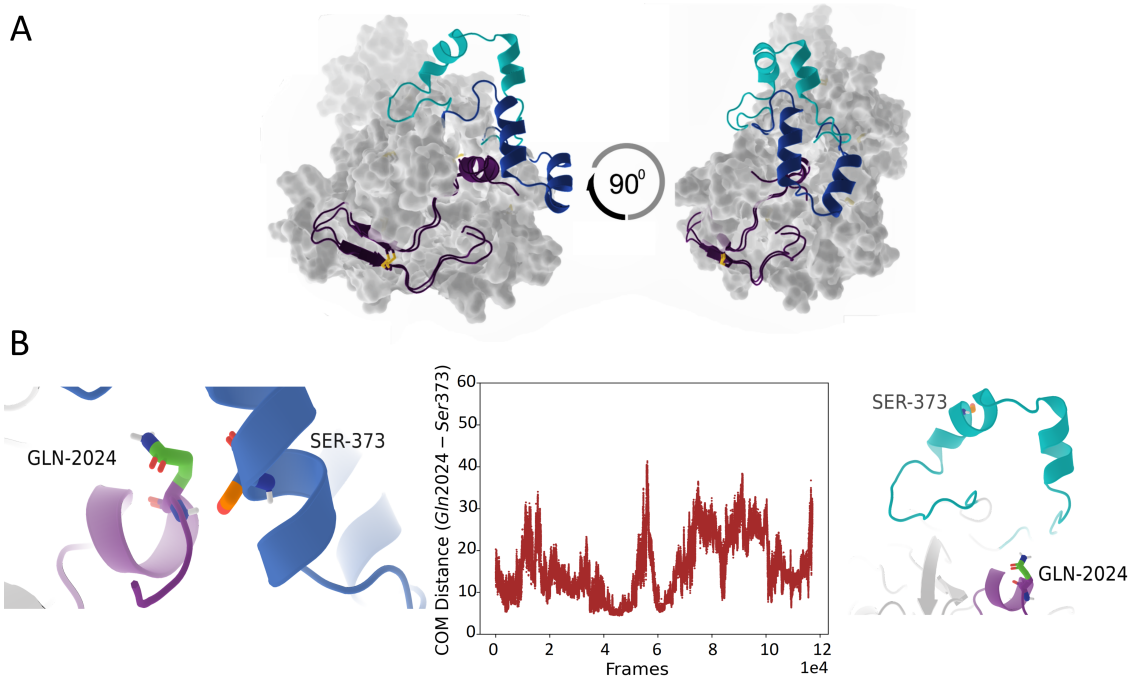


Figure 1. A. Loop closing event captured by molecular dynamics simulations. AMA1 is shown in surface representation; RON2 in purple; DII loop in cyan (open conformation) and deep blue (closed conformation). B. Centre of mass (COM) distance profile of Ser373 and Gln2024 in closed and open conformation.

In the closed conformation we found that the Ile2022 formed hydrophobic contacts, intermittently, with the surrounding residues many of which are located at the DII loop. Most important residues making hydrophobic contacts with Ile2022 are Met374, Ile375, Phe385 and Ala378 (fig-2). The evidence of the formation of a cryptic hydrophobic pocket is also reflected in the distance profiles of individual residues with Ile2022 (SI-fig-1).

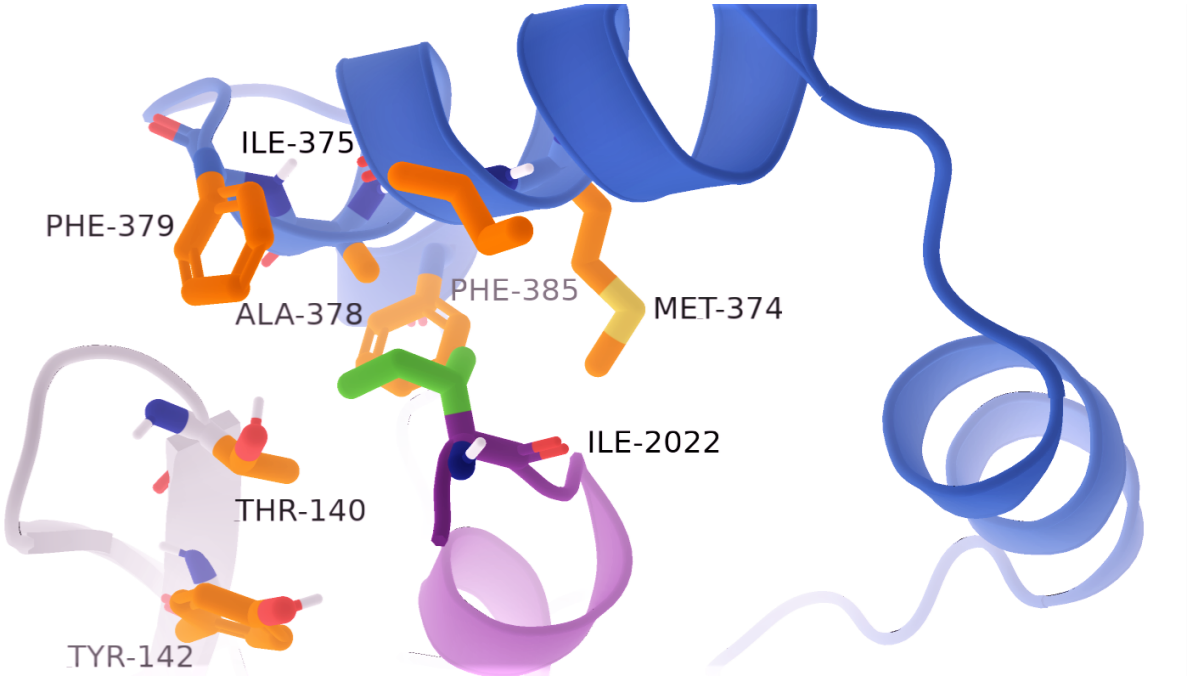


Figure 2. Hydrophobic pocket formed between Ile2022 and specific residues of AMA1(RON2: purple, DII loop closed conformation: deep blue)

Most important interactions are, however, hydrogen bonds evolved with Gln2024, Gln2025, Lys2027 and Asp2028 which interact with various residues from the DII loop and adjacent areas of AMA1 (fig-3, detailed interactions have been shown in SI-fig-2).

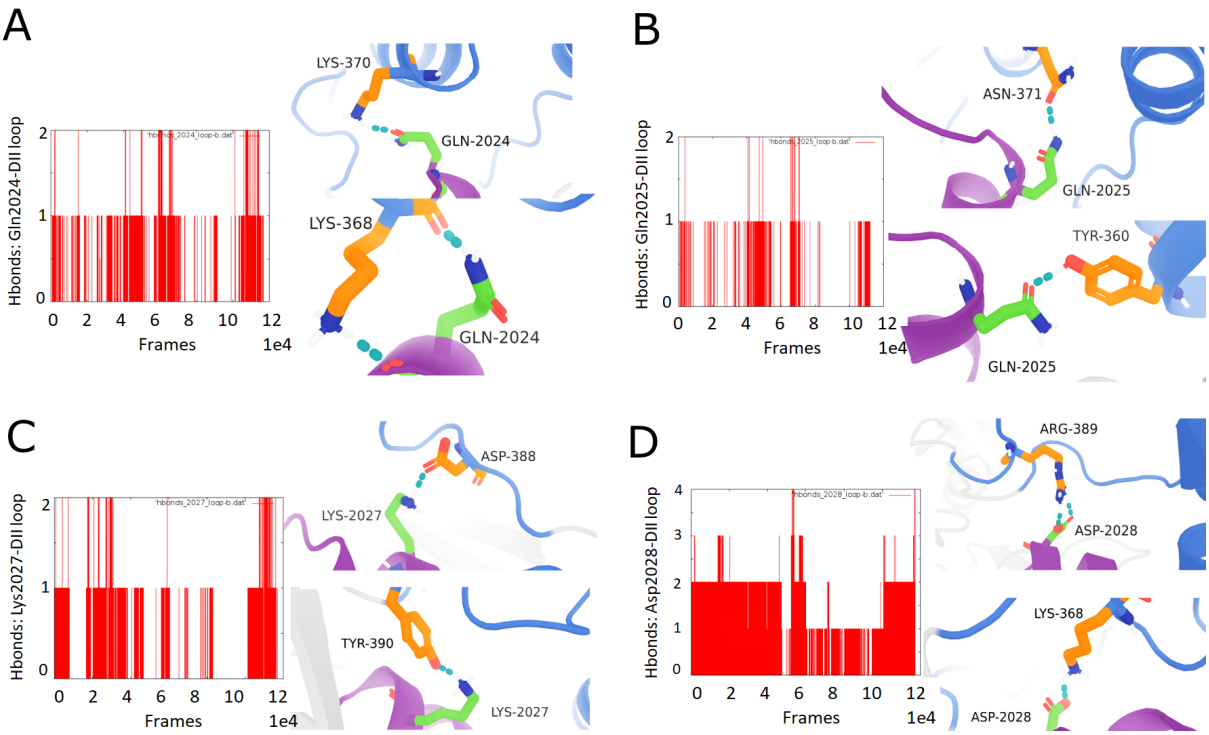


Figure 3. Hydrogen bonds formed between Gln2024 (A), Gln2025 (B), Lys2027 (C), Asp2028 (D) with representative residues of DII loop (detailed interactions are provided in SI-fig-2) (RON2: purple, DII loop closed conformation: deep blue, hydrogen bonds: teal)

The occurrences of association-dissociation process of DII loop with RON2 peptide which we observed in our MD simulations prompted us for a more quantitative characterization. Towards this end, we envisioned that a meaningful free energetic insight on the DII loop closing event and its associated interactions with the alpha helix of $RON2_{2021-2059}$ peptide can be achieved by simulating a free energy profile along an appropriate reaction coordinate. For this purpose, we used centre of mass distance between Ser373 and Gln2024 as the reaction coordinate. The free energy curve (see method section for details) indicated the formation of a favourable free energy basin when the residues mentioned in the collective variable came relatively closer (1.5Å) to each other and led to a state which we have defined as the closed conformation (fig-4). These results unambiguously stated that the DII loop possess high propensity of conforming to a state where it closes down towards AMA1 binding pocket and eventually arrests RON2 peptide from escaping.

To further assess the status of the free energies of loop-closing upon mutation of specific amino acids, we performed independent free-energy calculations on a set of systems in each of which a single residue has been mutated to alanine. By doing a scan along a wide range of values of the chosen reaction coordinate (centre of mass distance between each of the residues Gln2024Ala, Gln2025Ala, Lys2027Ala and Asp2028Ala individually with Ser373), we obtained a measure of free energy of loop closing upon these mutations. A close inspection of these free energy curves revealed that the free-energy minima in all four mutants would appear at a relatively larger distance than that of wild-type, indicating that the mutations would result in a relatively open conformation compared to wild type system (fig-5). A possible reason of relatively open conformation in these mutants than the wild-type is mostly due to the lack of crucial hydrogen bonds in the former (fig-3). Additionally, we have observed strikingly different free energy profiles specially for K2027A and D2028A in which the open conformations are more stabilised than loop-closed conformation. Our visual inspection of the simulation trajectories suggests that these two mutations make an unfavourable environment for the DII loop to close, due to absence of suitable contacts with the alpha helix of RON2 peptide. As revealed in the aforementioned paragraphs, further experiments with ITC and SPR in the later sections also reiterate the relative importance of these residues.

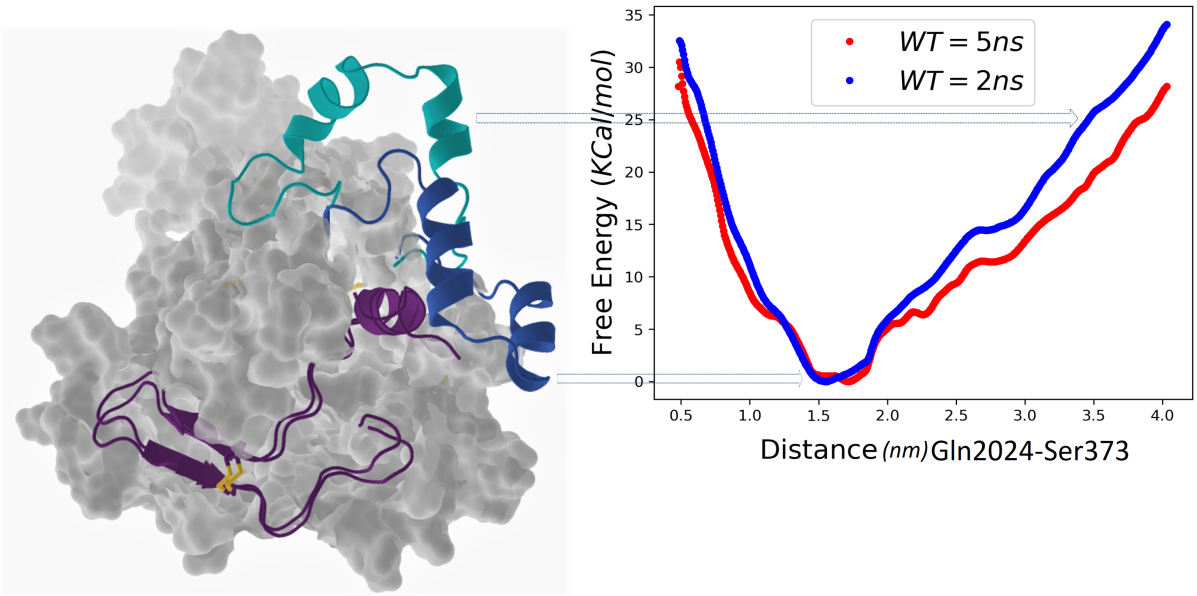


Figure 4. DII loop closing event. (Left) AMA1 represented in surface; RON2 in purple, D-II loop in cyan (open conformation) and deep blue (closed conformation). (Right) Free energy plot (5ns: red and 2ns: blue)

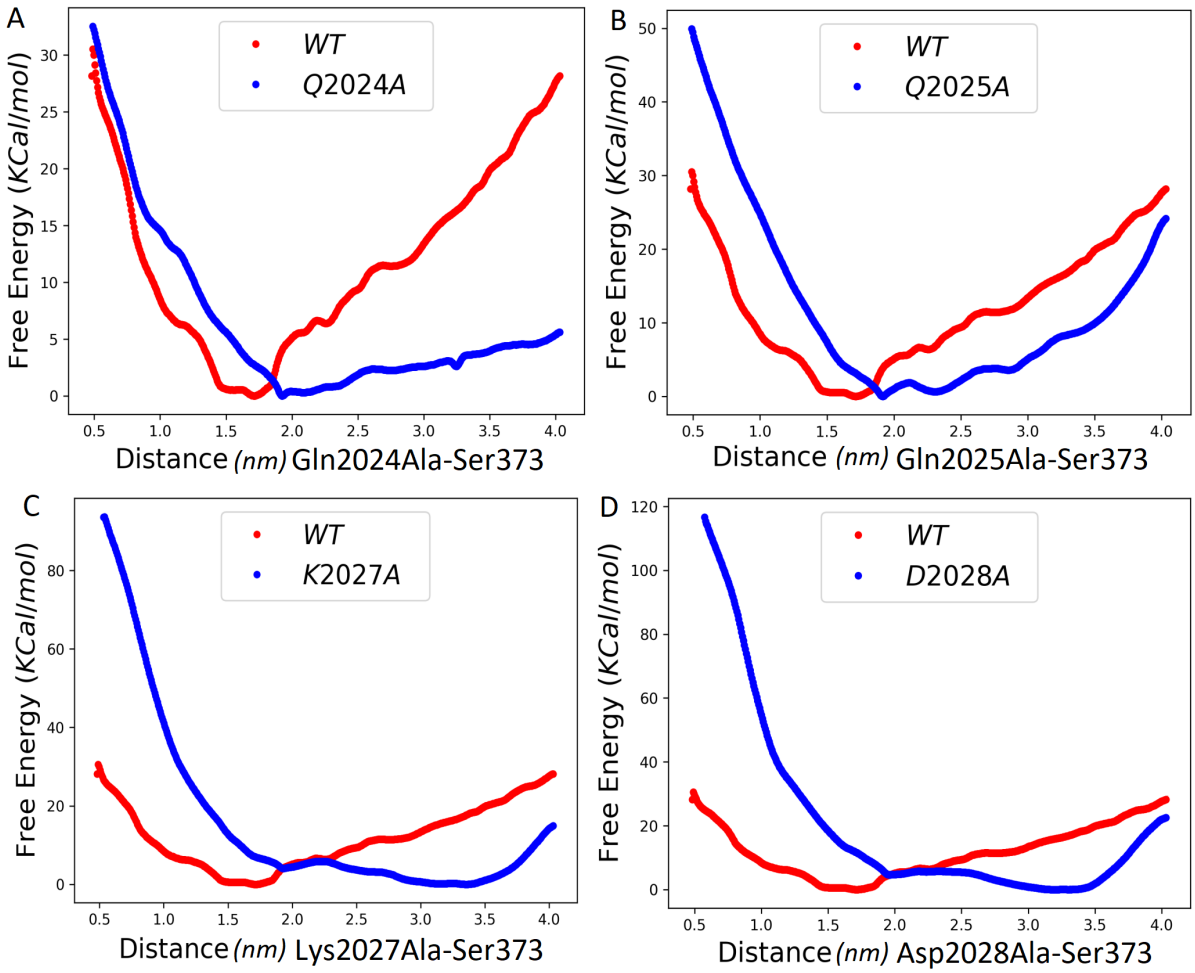


Figure 5. Free energy surfaces of Ala-substituted *Pf*RON2 analogues (A) Q2024A, (B) Q2025A, (C) K2027A and (D) D2028A compared with wild type *Pf*RON2 (wild type: red and mutated: blue)

PfAMA1 expression from E. coli and in vitro refolding

Although *Pf*AMA1 contains three distinct domains, studies have revealed that domain I and II are responsible for the binding with the extracellular domain of *Pf*RON2(9). In order to understand the interaction of the *Pf*RON2 with domain II loop, we set out to express domain I and domain II of *Pf*AMA1 protein in *Escherichia coli.* We expressed the protein {*Pf*AMA1(DI+DII)} in BL21 (DE3) RIL cells with an N-terminal His_6-tag; so that the protein can be purified by immobilized metal affinity chromatography after extracting from the bacterial cell. The protein got expressed in the inclusion body of *E. coli.* After extraction from inclusion bodies, the reduced polypeptide was purified using Ni-affinity column followed by reverse phase HPLC and lyophilized. The lyophilized polypeptide was then refolded in vitro as described earlier (10). The final refolded protein was purified by size exclusion column chromatography to get rid of the other soluble misfolded proteins formed during the course of refolding. The freshly purified folded protein was used for SPR and ITC experiments to compare the binding affinity of the *Pf*RON2$_{2021\text{-}2059}$ peptides and its Ala-substituted analogues.

*Chemical synthesis of the Pf*RON2$_{2021\text{-}2059}$ *peptide and its analogues*

To check the contribution of some of the key amino acid residues located in the *Pf*RON2 helix region in binding with the *Pf*AMA1 loop, we chemically synthesized the parent *Pf*RON2$_{2021\text{-}2059}$ peptide as well as several other peptides where some of the key residues, which were

computationally predicted to be responsible for binding with the DII loop region, were replaced by an alanine residue, one at a time. All peptides were synthesized by step-wise Fmoc SPPS using an automated peptide synthesizer. Lyophilized crude peptides were allowed to fold under air oxidation condition in 2M Gu.HCl, 100 mM Tris, pH 8 for 15h at room temperature to form one disulfide bond. Folded peptides were finally purified using reverse phase HPLC and lyophilized (fig 6)(10).

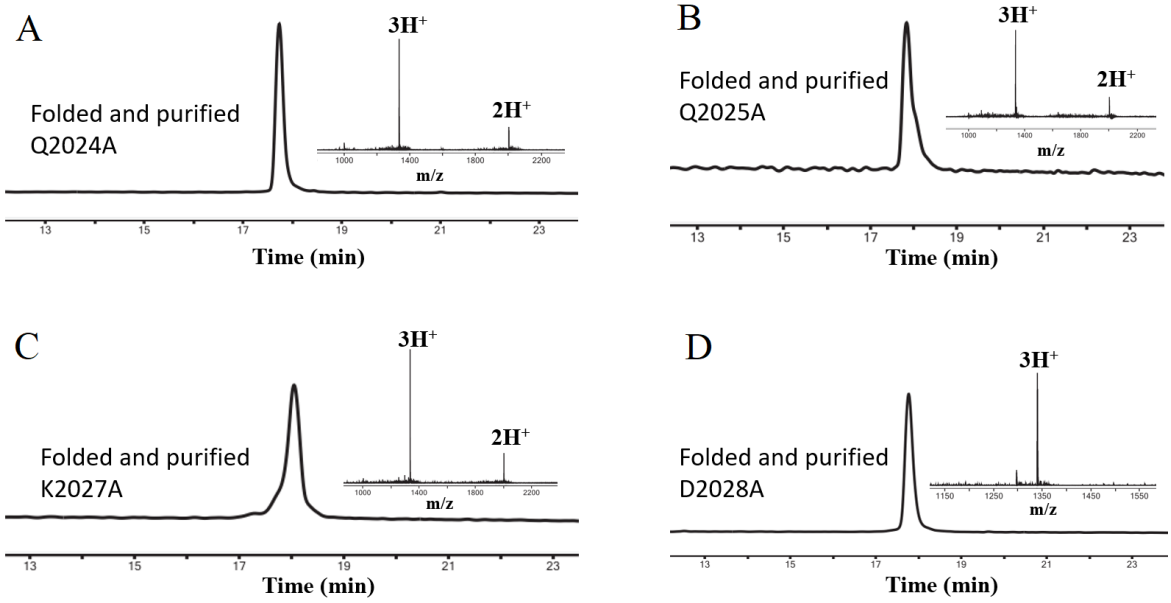


Figure 6. LC-MS chromatogram of the folded and purified *Pf*RON2 analogues. (A) Q2024A (calculated mass: 4003.97 Da, observed mass (most abundant isotopologue): 4003.99 ± 0.02 Da); (B) Q2025A (calculated mass: 4003.97 Da, observed mass (most abundant isotopologue): 4004.00 ± 0.02 Da); (C) K2027A (calculated mass: 4003.93 Da, observed mass (most abundant isotopologue): 4003.94 ± 0.01 Da); (D) D2028A (calculated mass: 4017.00 Da, observed mass (most abundant isotopologue): 4017.01 ± 0.02 Da).

Binding study using SPR and ITC

Our goal was to evaluate the binding affinity of the Ala-substituted analogues of *Pf*RON2$_{2021-2059}$ in order to identify their relative importance in binding with the DII loop. Therefore, we compared the binding profiles of the four peptide analogues (Q2024A, Q2025A, K2027A and D2028A) with *Pf*AMA1 with respect to *Pf*RON2$_{2021-2059}$ peptide by SPR and ITC.

In SPR, we observed decrease in the association rate of RON2 analogues towards AMA1, while at the same time increase in the dissociation rate of the ligands from the AMA1-RON2 complex (Table 1). As a result, we observed a significant leap in the K_D value of the Ala-substituted analogues when compared with the native RON2 ligand (K_D= 21.88 nM), indicating the weaker binding with AMA1 upon substitution in the RON2 helix. We observed a consistent trend in the K_D value between the substituted amino acid residue in SPR. We observed a K_D value of 898.56 nM and 227.63 nM for K2027A and D2028A analogues, respectively, indicating they are the two most important residues for the binding event; whereas, a K_D of 140.99 nM and 74.77 nM for Q2024A and Q2025A, respectively (K_D values reported here are the average of three repetitions, fig-7 and Table-1). A similar trend in ΔG values was also evident from ITC indicating that the complex formed by the substituted RON2 peptide analogues with AMA1 are lesser stable than that of AMA1-RON2 (fig-8, Table-1). The K_D values of the analogues obtained from ITC also showed similar trend as SPR. which reveals that K2027 and D2028 are the most important residues in the binding phenomena followed by

Q2024 and Q2025. The observed K_D values from ITC for Q2024A, Q2025A, K2027A and D2028A, were 228 nM, 201.6 nM, 326.8 nM and 320.6 nM, respectively, whereas K_D^{RON2} was 115.2 nM as shown in fig-8 and Table-1 (average of two repetitions).

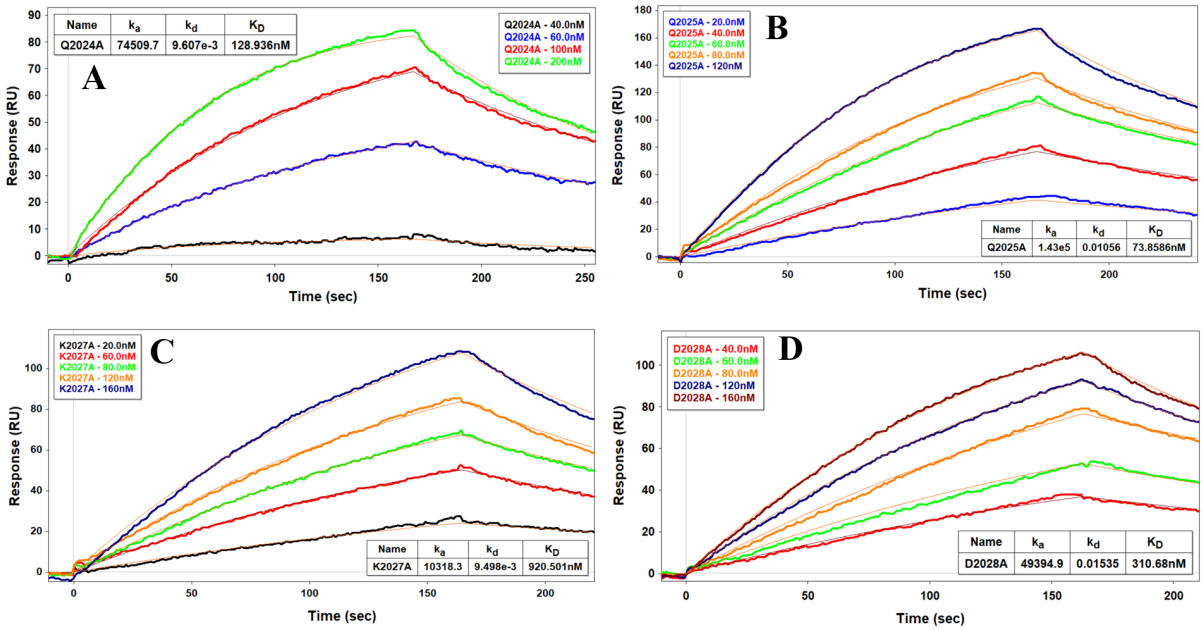


Figure 7. Representative SPR sensorgrams (one of the three repeats) of all alanine substituted peptides with *Pf*AMA1(DI+DII). (A) Q2024A (B) Q2025A (C) K2027A (D) D2028A.

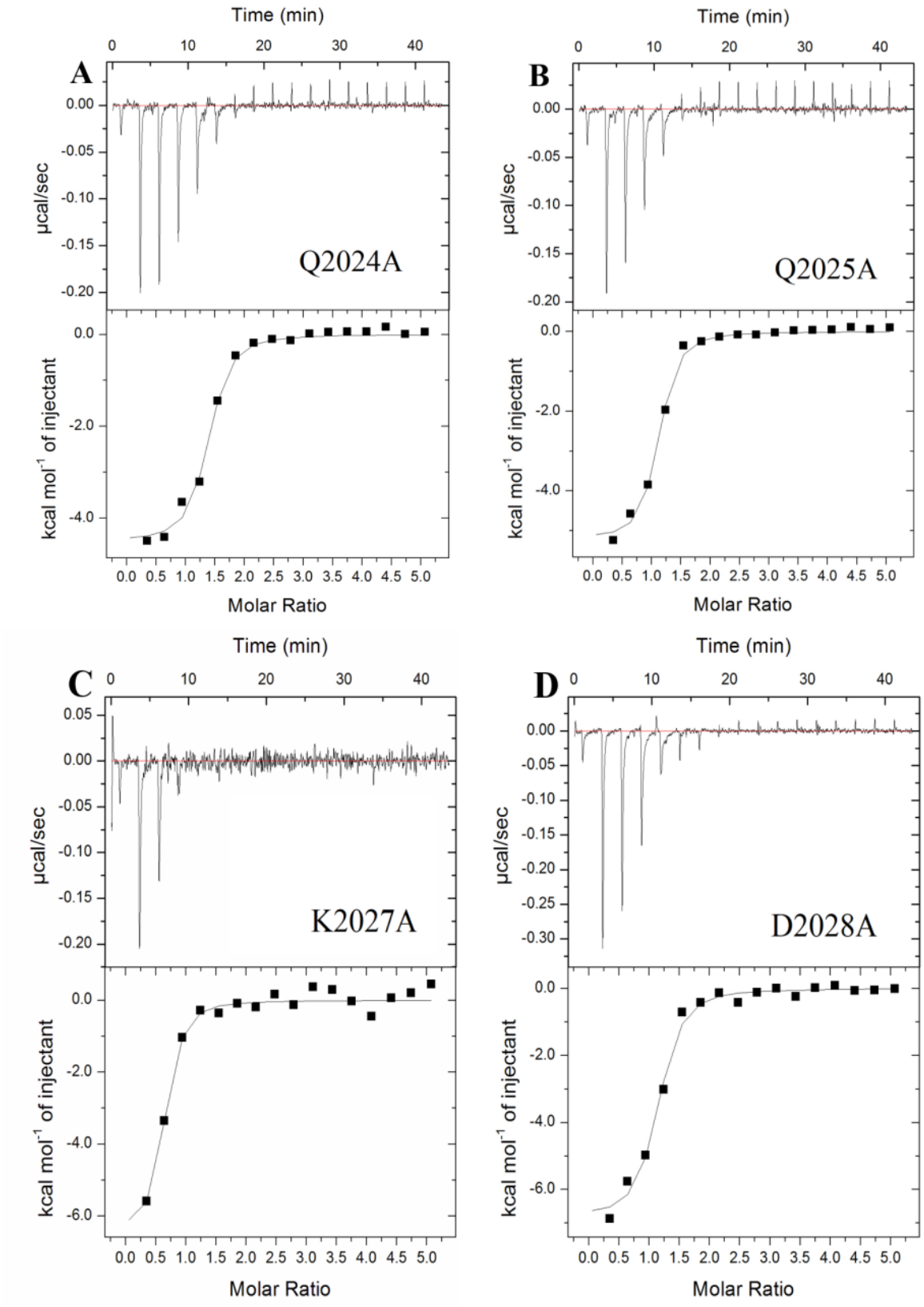


Figure 8. Binding profile of *Pf*AMA1 with all alanine substituted peptides in ITC experiment (representative one of the two repetitions). (A) Q2024A (B) Q2025A (C) K2027A (D) D2028A.

Table 1. SPR (average of three repetitions; error represents the standard deviation of three experiments) and ITC (average of two repetitions) data comparing the binding affinity and thermodynamic parameters (one of the two repetitions; error represents the fitting error of the particular experiment) of the alanine substituted peptides with respect to *Pf*RON2.

Ligand	K_D (nM) (SPR)	K_a (M^{-1}. Sec^{-1}) (SPR)	K_d (Sec^{-1}) (SPR)	K_D (nM) (ITC)	Molar ratio (ITC)	ΔH (kcal/mol)	-TΔS (kcal/mol)	ΔG (kcal/mol)
*Pf*RON2	21.88 ± 1.89 (10)	1.486E+05 ± 2.839E+04	3.21E-03 ± 0.0004	115.2 ± 21.7	1.48 ± 0.02	-4.122	-5.46	-9.58
Q2024A	140.99 ± 15.11	6.686E+04 ± 6.071E+03	9.34E-03 ± 0.0003	228.0 ± 43.7	1.28 ± 0.02	-4.507	-4.45	-8.96
Q2025A	74.77 ± 4.85	1.485E+05 ± 2.903E+04	0.011 ± 0.002	201.6 ± 26.2	1.01 ± 0.02	-5.227	-3.85	-9.08
K2027A	898.56 ± 17.09	2.416E+04 ± 1.218E+04	0.0216 ± 0.0108	326.8 ± 52.1	0.55 ± 0.04	-6.578	-2.18	-8.758
D2028A	227.63 ± 64.46	6.546E+04 ± 1.187E+04	0.014 ± 0.0016	320.6 ± 55.3	1.05 ± 0.03	-6.852	-1.91	-8.762

Discussion

Multiple repeats of atomistic simulations led us to infer that the DII loop, in effect, reaches out to the ectodomain of *Pf*RON2 in such a manner that the specific residues of the terminal helix of the *Pf*RON2$_{2021-2059}$ interacts strongly with both DII loop and adjacent residues leading to the macromolecular arrest, which stops the egress of the bound *Pf*RON2 from the *Pf*AMA1 pocket. We have been able to decipher residue pairs which are responsible for the formation of free energetically favourable basins as a consequence of the closing of the DII loop after *Pf*AMA1-*Pf*RON2 binding event. As observed during MD simulations, Ile2022 makes hydrophobic contacts, intermittently, with Met374, Ile375, Phe385 and Ala378 leading to the formation of cryptic hydrophobic pocket. However, the most important interactions are originated from Gln2024, Gln2025, Lys2027 and Asp2028 of the *Pf*RON2$_{2021-2059}$ peptide which are involved in hydrogen bonding interactions with various residues from the DII loop and adjacent areas of the AMA1 (fig-3). Converged free energy calculations eventually led us to reach to a meaningful atomistic insight into the mechanism of DII loop closing and its interactions with the alpha helix of the *Pf*RON2 peptide. To this end, we analysed several observables from umbrella sampling simulations. As collective variable, we used centre of mass distance between two amino acids (Ser373 and Gln2024) as hinted from the unbiased simulations. The free energy surface (FES) plot indicated formation of a favourable free energy basin when the residues mentioned in the collective variable came relatively closer, a state which we have defined as the closed conformation (fig-4). To verify the importance of selected few most interactive residues, predicted from our computation data, we systematically

substitute them with alanine. We observed that the loop closing was free energetically unfavourable with the chemically synthesized Ala-analogues, as the free energy dividend for bringing the loop closer from far off was more positive in Ala-substituted peptides (fig-5). According to our calculations, Lys2027 and Asp2028 were the most important residues from *Pf*RON2$_{2021-2059}$ peptide contributing most in this intra-molecular recognition event. This claim was justified by both free energy simulations as well as the experimental data obtained from binding studies of the *Pf*RON2$_{2021-2059}$ and its analogue peptides with *Pf*AMA1 by ITC and SPR experiments. In ITC, the average K_D was observed to shift from 115.2 nM (*Pf*RON2$_{2021-2059}$) to 326.8 nM and 320.6 nM in case of K2027A and D2028A, respectively (Table-1). However, in SPR experiment this K_D shift was observed from 21.88 nM (*Pf*RON2$_{2021-2059}$) to 898.56 nM and 227.63 nM for K2027A and D2028A, respectively (Table-1). These results led us to believe that Lys2027 and Asp2028 are the most important residues that interact with the DII loop during the recognition event. This fact is further illustrated in fig-5 (C and D) where the free energy minima is found to be appearing far away compared to *Pf*RON2$_{2021-2059}$ (fig-4), which clearly explained the reluctance to let the event of loop closing happen upon substitutions at these positions. Similar trends were also noticed in case of other substitutions such as Q2024A and Q2025A where K_D obtained from ITC was found to shift to 228.0 nM and 201.6 nM, respectively, as compared to 115.2 nM for *Pf*RON2$_{2021-2059}$ and also the K_D obtained from SPR was found to shift to 140.99 nM and 74.77 nM respectively as compared to *Pf*RON2$_{2021-2059}$ which is 21.88 nM (Table-1). The combined results from molecular dynamics and systematic alanine substitution experiments further confirm that hydrogen bonds are accountable for the loop closing event to a larger extent than the hydrophobic transience exhibited by Ile2022 along with feeble hydrogen bonds contributed from its main chain.

Moreover, recent literature report (4) using ITC studies states that the free energy of binding of *Pf*RON2$_{2021-2059}$ (ΔG=-10.6 kcal/mol) with *Pf*AMA1 in presence of DII loop is more favourable than with the AMA1 in absence of DII loop (ΔG = -9.6 kcal/mol), whilst witnessing a significant leap in K_D (92.6±6.4 nM in the case of truncated DII loop versus 42.9 ±11.4 nM where the DII loop is present). Although the DII loop thermodynamically stabilizes the binding of *Pf*RON2$_{2021-2059}$ by a 2-fold larger affinity, it was observed by the authors to possess a particularly influential role on the kinetics by restraining the release of the *Pf*RON2 peptide and increasing the complex half-life by 18-fold with a value of 900s (15 min) at 25°C for *Pf*AMA1- *Pf*RON2$_{2021-2059}$ complex compared to 48s for ΔDII-*Pf*AMA1-*Pf*RON2$_{2021-2059}$ complex (4). Contextually, peptide *Pf*RON2 (KDIGAGPVASCFTTRMSPPQQICLNSVVN) which also targets AMA1 but lacks a terminal helix blocked invasion by preventing *Pf*AMA1 interactions with the *Pf*RON2 complex (K_D=520±74 nM; PDB-3SRI) also supports our understanding, with caveats though, as the free energy of binding of *Pf*RON2sp2 has not been reported so far(7). In another recent work, pertaining to the design of β-hairpin peptides (Cys2037-Cys2049 patch), which completely excludes the α-helix motif also showed diminished functional activity (K_D=57±12 μM, (11)). Therefore, it was very evident that both DII loop and alpha helix from RON2 are absolutely necessary for an effective binding to happen. In the current study, while investigating the quantitative role of individual amino acids in RON2 helix where the helix residues are systematically altered with alanine keeping the beta-loop completely intact, computational insights showed us that the primary mode of contact of RON2 with AMA1 is significantly unperturbed. However, substitutions at the particularly designated positions, as hinted from multiple repeats of molecular simulation studies, led to the disruption of interactions of the RON2 helix with AMA1. In case of these Ala-analogues of *Pf*RON2$_{2021-2059,}$ the structural perturbations caused by Ala substitutions led to a faster egress of RON2 from the AMA1 binding pocket. This fact was further bolstered from the multiple replicas of binding assay

measured by SPR where we have observed quicker dissociation rates in case of the alanine substituted *Pf*RON2$_{2021-2059}$ analogous compared to the native one. It was observed that K2027A (k_d = 0.0216 min^{-1}) and D2028A (k_d = 0.014 min^{-1}) dissociate faster followed by Q2024A (k_d = 0.00934 min^{-1}) and Q2025A (k_d = 0.011 min^{-1}); whereas in case of native *Pf*RON2$_{2021-2059}$ the dissociation rate was least (k_d = 0.00321 min^{-1}) among all five cases (Table 1). This observation indicates the suitability of wild type residues in those particular positions as a decisive metric towards arrest of *Pf*RON2$_{2021-2059}$ to the AMA1 binding pocket. Moreover, repeated ITC experiments on the substituted *Pf*RON2$_{2021-2059}$ peptides showed there is a significant change in the free energy of binding against *Pf*AMA1(DI+DII), making the substitutions unfavourable for the AMA1-RON2 complex formation. The free energy of binding decreases the most in case of K2027A (ΔG = -8.758 kcal/mol) and D2028A (ΔG = -8.762 kcal/mol) compared to *Pf*RON2$_{2021-2059}$ (ΔG = -9.58 kcal/mol) further indicating these are the two most important residues. We have also observed a jump in the free energy of binding in case of the other two substitutions as well (ΔG^{Q2024A} = -8.96 kcal/mol, ΔG^{Q2025A} = -9.08 kcal/mol) with respect to the native *Pf*RON2$_{2021-2059,}$ although relative importance is lesser than that of K2027A and D2028A.

Conclusions

A fundamental understanding of the mode of interactions between the two important *Plasmodium falciparum* protein partners like AMA1-RON2 is absolutely necessary to design potential therapeutics against the dreaded disease like malaria. To that end, the current work undertook both computational as well as experimental binding studies with systematically substituted alanine analogues of *Pf*RON2$_{2021-2059}$ in order to understand in detail how *Pf*RON2$_{2021-2059}$ interacts with AMA1, with special emphasis on the role of DII loop in the due process of the recognition event. From the detailed analysis of the computational as well as the experimental data, we have found a direct correlation between the DII loop and the alpha helix of the *Pf*RON2$_{2021-2059}$ that influence each other leading to the formation of a stable complex. We have not only identified key binding residues of the alpha helix of the *Pf*RON2$_{2021-2059}$ but also many other important molecular interactions responsible for carrying out this binding event. The current report also reiterates previous findings on specific kinetic aspects, like egress rates, which are found to be significantly perturbed by the Ala substitutions of specific residues on the N-terminal alpha helix of the *Pf*RON2$_{2021-2059}$. In summary, the current work provides novel insights into key interactions between *Pf*AMA1 and *Pf*RON2 proteins which will further assist in designing potential RON2 based AMA1 inhibitors.

Experimental Procedures

MD simulations and the potential of mean force (PMF) calculations using umbrella sampling

We performed MD simulations using the GROMACS code (version 2019)(12, 13). For simulations we used the CHARMM36 force field (14) with TIP3P water model (15). Charmm-gui(16) web-server was used to prepare the systems including the construction of the loop. Coordinates from PDB:3ZWZ were used as a starting structure for simulations. A cubic box type with a distance of 1.0 nm between the complex and the box edge was defined to set the minimum distance of at least 2 nm between any two periodic images of the solute. Then, the box was solvated with TIP3P water and the system was neutralized with potassium chloride counter ions in a concentration of 0.15 mol/lit. The steepest descent algorithm (17) was applied to optimize the system during a 100 ps run time. A position restraint of 1000 kJ mol^{-1} nm^{-2} was applied to fix the positions of both ligand and protein atoms and the whole system was

equilibrated under the NVT ensemble through 100 ps time during which the V-rescale thermostat was used to regulate the temperature close to 310 K. Following the NVT step, the system was equilibrated under the NPT ensemble through 100-ps time in such a way that the pressure of the system was stabilized to 1 atm. The production MD of 200 ns (seven repeats) was performed on the well equilibrated system at the temperature of 310 K and the pressure of 1 atm. The Particle-mesh Ewald (PME) algorithm(18) was used to calculate the long-range electrostatic contributions and the length of all covalent bonds was constrained using the LINCS algorithm(19), a faster algorithm in comparison to the SHAKE algorithm(20). Upon termination of the simulation, the protein was backed and cantered in the box along with the removing of periodic boundary condition from the trajectory.

The potential of mean force for dynamics of the DII loop movement with reference to RON2 peptide was constructed using umbrella sampling MD simulations. Umbrella sampling (21) is a very popular technique for PMF calculation to study binding-unbinding processes related to protein-ligand and protein-protein interactions (22–24). The binding free energy can also be extracted from the obtained PMF. The umbrella sampling is a non-Boltzmann sampling where an extra term is added to the hamiltonian for sampling to explore the collective variable (CV) space. In practice, CV space is divided into a series of windows and an external potential, usually harmonic, is applied to keep the distribution peaked within a region of the order parameter. As CV, we used centre of mass distance between two amino acids (Ser373 and Gln2024) as deduced from the unbiased simulations. 50 umbrella windows were created along this distance at 0.5 Å intervals with a force constant (k) of 500kcal/mol/$Å^2$ until it indicated an overlapping histogram of positional distributions. The umbrella sampling calculations were repeated for three cycles for all the systems for 2 ns and 5 ns for convergence in the wild type system and 5 ns for the alanine substituted analogues. Finally, we used weighted histogram analysis method (WHAM)(25, 26) which combines the individual PMFs optimally to weigh the individual distributions to obtain the free energy profile.

Expression, purification and refolding of PfAMA1(DI+DII)

The expression, purification and refolding of the *Pf*AMA1(DI+DII) was performed following the published procedure (10). In brief, the expression plasmid encoding the sequence of 3D7 *Pf*AMA1(DI+DII) (104-438) was obtained commercially from GenScript (NI, USA) having N-terminal hexa-His tag along with one TEV-protease cleavage site. The protein was expressed in *Escherichia coli* BL21 (DE3) RIL cells and purified through Ni-affinity column followed by reverse phase HPLC purification and lyophilization. Lyophilized protein was then refolded and purified by size exclusion chromatography as described earlier (10). Protein containing fractions were pooled and used for the binding assay by SPR and ITC.

Synthesis and oxidative folding of RON2 and other alanine substituted peptides

The 39-mer extracellular domain of RON2 (Asp2021-Ser2059, *Pf*RON2$_{2021-2059}$) was synthesized on 2-chlorotritylchloride resin using Fmoc protected amino acids by solid phase peptide synthesis (SPPS) at a single stretch using an automated peptide synthesizer (Tribute UV-IR, Protein Technologies Inc. USA). After cleaving the peptide from the solid resin support, the crude peptide was dissolved in 6M guanidine hydrochloride (Gu.HCl), centrifuged, filtered with 0.2 micron filter and slowly diluted with Tris buffer to get the ultimate folding condition of 2M Gu.HCl and 100 mM Tris (pH 8). To form the disulfide bond, the peptide was incubated under air oxidation condition for 15h. The folded peptide was purified by reverse phase HPLC and lyophilized. The other Ala substituted analogues Q2024A, Q2025A, K2027A and D2028A) were also synthesized adapting the same protocol described above followed by oxidative disulfide formation, purification by reverse phase HPLC and lyophilization. All the lyophilized peptides were reconstituted in running buffer (10 mM

phosphate buffer saline, pH 7.4, 0.005% Tween-20, 40 µM EDTA) for SPR experiments and in elution buffer (20 mM Tris pH 7.8, 100 mM NaCl) of size exclusion chromatography for ITC experiments to study the binding against *Pf*AMA1(DI+DII).

Binding affinity of peptides with AMA1 using surface plasmon resonance

BI-4500AP SPR instrument was used to measure the binding of the *Pf*RON2$_{2021-2059}$ peptides and its alanine substituted analogues with *Pf*AMA1. 2.4 µM freshly refolded His-tagged AMA1 protein was immobilized at pH 7.8 over a Nickel-NTA chip while 10 mM phosphate buffer saline, pH 7.4 with 0.005% Tween-20 was used as the running buffer. Three flow cells were used for the immobilization of the AMA1 protein using 'G-inject' function to get gradually decreasing or increasing level of immobilization. The reference flow-cell was created by injecting only the running buffer. Before passing through the analytes over the chip, 40 µM EDTA was added in the running buffer to minimize non-specific interactions. A series of concentrations of a particular analyte dissolved in the running buffer (10 mM Phosphate buffer saline, pH 7.4 with 0.005% Tween-20 and 40 µM EDTA) were then passed through all the four flow-cells at a constant flow rate of 30 µl/min to obtain the association curve. After 360 sec (for *Pf*RON2$_{2021-2059}$) and 150 sec (for Ala substituted peptides due to their weaker binding propensity), the injection of the peptide analytes was stopped and the chip was allowed to regenerate in the running buffer at the same flow rate to capture the dissociation event. The final sensorgram was generated by subtracting the reference flow cell from the ligand flow cell and the resulting data was fitted with 1:1 Langmuir adsorption binding isotherm using the BI-data analysis software. For all analytes, the binding experiments were performed in three repeats using the same batch of refolded *Pf*AMA1(DI+DII) protein and reported as an average of the three data.

Binding affinity of peptides with AMA1 using isothermal titration calorimetry

MicroCaliTC200 instrument was used to check the binding affinity of the peptides with the *Pf*AMA1 protein. 200 µl of the 10 µM *Pf*AMA1 was loaded into the ITC sample cell and the protein was titrated with 2 µl of the ligand (peptide) for total 17 injections. Ligands were dissolved in 20 mM Tris and 100 mM NaCl (pH 7.8), the same buffer in which the AMA1 protein was eluted. In optimized conditions, *Pf*RON2$_{2021-2059}$ concentration was kept as 200 µM while the other Ala substituted peptide concentrations were kept at 300 µM due to their weaker binding affinity towards *Pf*AMA1 protein as compared to *Pf*RON2$_{2021-2059}$. Heat of dilution data was recorded in Tris-NaCl buffer for each peptide and final data was obtained by subtracting this data from the raw data. The data fit was obtained using Origin software (Microcal) and dissociation constant was calculated for each ligand using one site model. The binding study for each peptide was repeated twice for reproducibility against freshly refolded *Pf*AMA1(DI+DII) and K_D value was reported as average of the two.

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