Strip1 regulates retinal ganglion cell survival by suppressing Jun-mediated apoptosis to promote retinal neural circuit formation

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Abstract

In the vertebrate retina, an interplay between retinal ganglion cells (RGCs), amacrine and bipolar cells establishes a synaptic layer called the inner plexiform layer (IPL). This circuit conveys signals from photoreceptors to visual centers in the brain. However, the molecular mechanisms involved in its development remain poorly understood. Striatin-interacting protein 1 (Strip1) is a core component of the STRIPAK complex, and it has shown emerging roles in embryonic morphogenesis. Here, we uncover the importance of Strip1 in inner retina development. Using zebrafish, we show that loss of Strip1 causes defects in IPL formation. In *strip1* mutants, RGCs undergo dramatic cell death shortly after birth. Amacrine and bipolar cells subsequently invade the degenerating RGC layer, leading to a disorganized IPL. Thus, Strip1 promotes IPL formation through RGC maintenance. Mechanistically, zebrafish Strip1 interacts with its STRIPAK partner, Striatin3, to promote RGC survival by suppressing Jun-mediated apoptosis. In addition to its function in RGC maintenance, Strip1 is required for RGC dendritic patterning, which likely contributes to proper IPL formation. Taken together, we propose that a series of Strip1-mediated regulatory events coordinates inner retinal circuit formation by maintaining RGCs during development, which ensures proper positioning and neurite patterning of inner retinal neurons.

Introduction

The retina has a highly organized neural circuit that consists of six major classes of neurons, which are assembled into three cellular layers with two synaptic or plexiform layers in between. This beautiful layered architecture is termed “retinal lamination” (Avanesov and Malicki, 2010, D’Orazi et al., 2014, Dowling, 1987). Lamination is conserved among vertebrates and critical for processing of visual information (Baden et al., 2020, Nassi and Callaway, 2009). During development, neurogenesis, cell migration and neurite patterning are spatially and temporally coordinated to form retinal lamination. Any defect in these events can disrupt retinal wiring and consequently compromise visual function (Amini et al., 2018). However, molecular mechanisms that govern retinal neural circuit formation are not fully understood.

The retinal neural circuit processes visual signals through two synaptic neuropils (Figure 1A). At the apical side, the outer plexiform layer (OPL) harbors synapses that transmit input from photoreceptors (PRs) in the outer nuclear layer (ONL) to bipolar (BP) and horizontal cells (HCs) in the inner nuclear layer (INL). At the basal side of the retina, the inner plexiform layer (IPL) is densely packed with synaptic connections formed between BPs and amacrine cells (ACs) in the INL and retinal ganglion cells (RGCs) in the ganglion cell layer (GCL). The retina contains one type of glial cells called Müller glia (MGs) which span the apico-basal axis of the retina (Hoon et al., 2014, Huberman et al., 2010).

RGCs are the first-born retinal neurons, which extend their axons to exit the eye cup and innervate visual centers in the brain (D'Souza and Lang, 2020, Robles et al., 2014). In mouse and zebrafish models, when RGCs are absent or exhibit defects in axon projections, vision is compromised (Kay et al., 2001, Moshiri et al., 2008, Rick et al., 2000). So, RGCs are indispensable for vision. RGC degeneration is often a secondary defect in optic neuropathies and the leading cause of blindness. Thus, tremendous efforts are being dedicated to decipher signaling pathways involved in RGC death (Maes et al., 2017, Munemasa and Kitaoka, 2012).

Strip1 is a recently identified protein with emerging functions in neuronal development. It was first described as one of the core components of the striatin-interacting phosphatases and kinases (STRIPAK) complex (Goudreault et al., 2009).

STRIPAK complex is an evolutionarily conserved supramolecular complex with diverse functions in cell proliferation, migration, vesicular transport, cardiac development, and cancer progression (He et al., 2010, Hwang and Pallas, 2014, La Marca et al., 2019, Madsen et al., 2015, Neisch et al., 2017, Shi et al., 2016). In addition, several STRIPAK components participate in dendritic development, axonal transport, and synapse assembly (Chen et al., 2012, Li et al., 2018, Schulte et al., 2010). In *Drosophila*, Strip (a homolog of mammalian Strip1/2) is essential for axon elongation by regulating early endosome trafficking and microtubule stabilization (Sakuma et al., 2014, Sakuma et al., 2015). In addition, Strip, together with other STRIPAK members, modulates synaptic bouton development and prevents ectopic retina formation (Neal et al., 2020, Sakuma et al., 2016). On the other hand, loss of mouse Strip1 causes early mesoderm migration defects leading to embryonic lethality (Bazzi et al., 2017, Zhang et al., 2021). Thus, the role of Strip1 in the vertebrate nervous system is largely unknown.

Here, we report an essential role for Strip1 in neural circuit formation of zebrafish retina. In zebrafish *strip1* mutants, retinal lamination, especially IPL formation, is disrupted. Loss of Strip1 causes RGC death shortly after birth. Cells in the INL subsequently infiltrate the degenerating GCL, leading to a disorganized IPL. Strip1 cell-autonomously promotes RGC survival; however, it is not required in INL cells for IPL formation. Therefore, Strip1-mediated RGC maintenance is required to establish the IPL. Mechanistically, Strip1 interacts with its STRIPAK partner, Striatin3, to promote RGC survival through the suppression of the Jun-mediated apoptotic pathway. We also found that Strip1 is necessary for RGC dendritic patterning, which likely promotes interaction between RGCs and ACs for IPL formation. Collectively, we demonstrate that Strip1 is crucial for RGC survival during development and thereby coordinates proper wiring of the inner retina.

Results

Strip1 is essential for inner retinal neural circuit development

To understand mechanisms of retinal neural circuit formation, we screened zebrafish retinal lamination-defective mutants (Masai et al., 2003) and identified the *rw147* mutant. At 4 days post-fertilization (dpf), *rw147* mutant embryos have small eyes, lower jaw atrophy and cardiac edema (Figure 1B). *rw147* mutants also show defects in retinal lamination, in which retinal layers, especially in the inner retina, fluctuate in a wave-like pattern (Figure 1C). The *rw147* mutation is lethal by 6 dpf due to cardiac edema. Mapping of the *rw147* mutation revealed a missense mutation in exon 7 of *striatin interacting protein 1* (*strip1*) gene of the *rw147* mutant genome (Figure 1D).

Next, we performed Crispr/cas9-mediated mutagenesis to generate a 10-base deletion mutant, *strip1*$^{crispr\Delta 10}$ (Figure 1–figure supplement 1A). *strip1*$^{crispr\Delta 10}$ mutants show similar morphology and retinal lamination defects to those of *strip1*^{rw147} mutants (Figure 1–figure supplement 1B-1D). Likewise, knockdown of Strip1 using translation-blocking morpholinos (MO-Strip1) phenocopied *strip1*^{rw147} mutants (Figure 1–figure supplement 1E and 1F). We verified the specificity of MO-Strip1 using a custom-made zebrafish Strip1 antibody that fails to detect a 93-kDa protein band corresponding to zebrafish Strip1 in the morphants (Figure 1–figure supplement 1G). Furthermore, we generated transgenic lines that express wild-type and *rw147* mutant forms of zebrafish Strip1 protein under the control of the heat shock promotor, *Tg[hsp:WT.Strip1-GFP]* and *Tg[hsp:Mut.Strip1-GFP],* respectively. Wild-type Strip1, but not the mutant form, rescued the retinal defects of *strip1*^{rw147} (Figure 1–figure supplement 1H and 1I). Taken together, the *strip1* mutation is the cause of retinal lamination defects.

Next, we examined Strip1 expression in wild-type retinas by labeling with the zebrafish Strip1 antibody. Strip1 was expressed in RGCs and ACs at 2 dpf (Figure 1E). *In situ* hybridization shows that *strip1* mRNA is maternally and zygotically expressed and by 2 dpf, expression becomes restricted to the eyes, optic tectum, and heart (Figure 1–figure supplement 2A and 2B). Like Strip1 protein, *strip1* mRNA was expressed in RGCs and ACs (Figure 1–figure supplement 2C). To visualize retinal neuropils, we performed whole-mount staining of the retina with anti-acetylated α-tubulin antibody. In wild-type retinas, IPL and OPL are evident at 3 dpf. In contrast, IPL shows abnormal

morphology, whereas OPL is relatively normal in *strip1*^{rw147} mutants (Figure 1F). We tracked IPL development using Bodipy TR stain. In wild-type retinas, a rudimentary IPL was formed as early as 52 hours post-fertilization (hpf); however, it was less defined in *strip1*^{rw147} mutants (Figure 1–figure supplement 2D). At 62 hpf, mutants exhibited a wave-like IPL. This temporal profile coincides with development of RGCs and ACs. Next, we visualized neurite morphology of RGCs, ACs, and BPs by transiently expressing fluorescent proteins under control of *ath5* (Masai et al., 2003), *ptf1a* (Jusuf and Harris, 2009), and *nyx* promoters (Schroeter et al., 2006) respectively. In wild-type siblings, RGCs and ACs normally extend their dendrites toward the IPL; however, *strip1*^{rw147} mutants show randomly directed dendritic patterns of RGCs and ACs (Figures 1G and 1H). In wild-type siblings, BPs normally extend their axons and dendrites toward IPL and OPL, respectively; however, BPs of *strip1*^{rw147} mutants show mis-routed axons and abnormal dendritic branching (Figure 1I). Thus, Strip1 is required for IPL formation and correct neurite patterning of RGCs, ACs, and BPs.

RGCs are reduced and INL cells infiltrate the GCL in *strip1* mutants

To examine how IPL is disrupted in *strip1* mutants, we combined *strip1*^{rw147} mutants with two transgenic lines, *Tg[ath5:GFP; ptf1a:mCherry-CAAX]*, to visualize RGCs and ACs. In *Tg[ath5:GFP]*, GFP is expressed strongly in RGCs and weakly in ACs and photoreceptors under control of the *ath5* enhancer (Masai et al., 2003). In *Tg[ptf1a:mCherry-CAAX]*, membrane-targeted mCherry is expressed in ACs and HCs under control of *ptf1a* promoter (Jusuf and Harris, 2009). Live imaging of 3-dpf retinas revealed that RGCs are severely reduced in *strip1*^{rw147} mutants (Figure 2A and 2B). On the other hand, there was no significant change in the number of ptf1a+ ACs between *strip1*^{rw147} mutants and wild-type siblings (Figure 2A and 2C). However, ptf1a+ ACs abnormally extended their dendrites to form an irregular pattern of IPL (Figure 2A). In wild-type retinas, the majority of ACs reside in the INL, except displaced ACs (Jusuf and Harris, 2009). However, in *strip1*^{rw147} mutants, a significant fraction of ptf1a+ ACs were abnormally located in the GCL (Figure 2A, bottom panels and 2D). Such abnormal positioning of ACs is correlated with the severity of IPL defects (Figure 2–figure supplement 1A). This phenotype is reminiscent of *ath5* mutants, *lakritz*, in which RGCs

fail to undergo neurogenesis, leading to infiltration of ACs into the GCL and transient IPL formation defects (Kay et al., 2001, Kay et al., 2004). We confirmed similar IPL defects in *ath5* morphant retinas at 3 dpf (Figure 2–figure supplement 1B), albeit weaker than those of *strip1*^{rw147} mutants.

Next, we visualized ACs using anti-Pax6 antibody, which strongly labels ACs and weakly labels RGCs (Macdonald and Wilson, 1997). In wild-type siblings, most strong Pax6+ cells were in the INL, and only 9.84±4.13% were in the GCL (Figure 2E, 2G). However, in *strip1*^{rw147} mutants, a significant percentage of Pax6+ cells (44.26±17.8%) was in the GCL (Figure 2E, 2G). The total number of Pax6+ cells did not differ between wild-type siblings and *strip1*^{rw147} mutants (Figure 2F). Next, we visualized BPs using anti-Prox1 antibody, which labels BPs and HCs (Jusuf and Harris, 2009). In wild-type, 100% of Prox1+ cells were in the INL (Figure 2H and 2J). However, 10.6±6.26% of Prox1+ cells were abnormally located in the GCL in *strip1*^{rw147} mutants (Figures 2H and 2J). The total number of Prox1+ cells did not differ between wild-type siblings and *strip1*^{rw147}mutants (Figure 2I). We performed labeling of double-cone and rod photoreceptors using zpr1 and zpr3 antibodies, respectively (Nishiwaki et al., 2008). Apart from occasional mildly disrupted areas, the photoreceptor cell layer was grossly intact, with no positioning defects (Figure 2–figure supplement 1C and 1D). Müller glia and proliferating cells at the ciliary marginal zone were visualized using anti-glutamine synthetase (GS) (Peterson et al., 2001) and anti-PCNA antibodies (Raymond et al., 2006), respectively. Both cell types showed grossly normal positioning in *strip1*^{rw147} mutants (Figure 2–figure supplement 1E and 1F). Thus, in the absence of Strip1, INL cells abnormally infiltrate the GCL and seem to replace the reduced RGCs.

Strip1 cell-autonomously promotes RGC survival

In zebrafish, RGC genesis starts in the ventronasal retina at 25 hpf, spreads into the entire retina by 36 hpf and is completed by 48 hpf (Avanesov and Malicki, 2010, Hu and Easter, 1999). The reduction of RGCs in *strip1* mutants could be due to compromised RGC genesis or RGC death after birth. To clarify which, we examined RGC genesis by monitoring ath5:GFP expression, and apoptosis by TUNEL. In *strip1*^{rw147} mutants, RGCs are normally produced at 36 hpf; however, apoptosis occurred in the GCL at 48 hpf

(Figure 3A). The number of apoptotic cells in GCL reached its highest level at 60 hpf, and apoptotic cells were eliminated by 96 hpf (Figure 3A and 3B). Accordingly, RGCs were significantly lower in *strip1*^{rw147} mutants than in wild-type siblings at 60 hpf and progressively reduced by 96 hpf (Figure 3C). In contrast, other retinal layers of *strip1*^{rw147} mutants showed slightly, but not significantly increased apoptosis at 72 hpf (Figure 3–figure supplement 1A), suggesting a specific function of Strip1 in RGC survival. In addition, despite the reduction in ath5:GFP+ area, the total presumptive GCL area, which was defined by retinal area between the lens and the IPL, was unchanged in *strip1*^{rw147} mutants throughout the stages (Figure 3–figure supplement 1B), suggesting that infiltrating INL cells replace the lost RGCs. We confirmed RGC death in *strip1*$^{crispr\Delta 10}$ mutants (Figure 3–figure supplement 1C and 1D). Interestingly, we observed apoptosis in the optic tectum of *strip1*^{rw147} mutants (Figure 3–figure supplement 1E and 1F), suggesting a common Strip1-dependent survival mechanism in the optic tectum. RGCs are the only retinal neurons whose axons project to the optic tectum. In *strip1*^{rw147} mutants, RGC axons appeared to exit from the eye cup and formed an optic chiasm at 3 dpf (Figure 3–figure supplement 1G). However, consistent with the reduction of RGCs, the optic nerve was thinner in *strip1*^{rw147} mutants than in wild-type siblings and showed elongation defects towards the optic tectum.

To determine whether Strip1 cell-autonomously promotes RGC survival, we conducted cell transplantation from *strip1*^{rw147} mutant donor cells into wild-type host embryos at the blastula stage. TUNEL of transplanted retinas at 60 hpf revealed that *strip1*^{rw147} mutant donor RGCs underwent apoptosis in wild-type host retinas (Figures 3D-3F). Thus, Strip1 is cell-autonomously required for survival of RGCs.

RGC death triggers abnormal positioning of ACs, leading to IPL disruption

ACs are proposed to be the main cell type responsible for IPL formation (Godinho et al., 2005, Huberman et al., 2010). To clarify how RGC death influences infiltration of ACs into GCL and IPL disruption, we performed time-lapse imaging of wild-type and *strip1*^{rw147} mutant retinas combined with the transgenic line *Tg[ath5:GFP; ptf1a:mCherry-CAAX]*. At 48 hpf, there were no apparent differences in position or morphology of RGCs and ACs between wild-type siblings and *strip1*^{rw147} mutants

(Figure 4A and Videos 1 and 2). In *strip1*^{rw147} mutants at 52 hpf, RGCs started to disappear, creating an empty spot in the GCL (Figure 4A, asterisks). However, ACs were still located in the INL. At 55 hpf, a rudimentary IPL was observed in the central retina of both wild-type siblings and *strip1*^{rw147} mutants. At 59 hpf, ACs started to invade the empty spaces in the GCL (Figure 4A, arrowheads). Infiltration of ACs into the GCL was more prominent at 62 hpf, resulting in a fluctuating IPL. Thus, loss of RGCs triggers infiltration of ACs into the GCL in *strip1*^{rw147} mutants.

To examine whether Strip1 is required in ACs for IPL formation, we performed cell transplantation using donor embryos carrying the transgene *Tg[ptf1a:mCherry-CAAX]* (Figure 4B). When mutant ACs were transplanted into wild-type host retinas, most donor ACs were normally positioned in the INL and extended dendrites toward the IPL, as in the case of wild-type donor ACs transplanted into a wild-type host retina (Figure 4C, 4D). Occasionally, three ACs extended 2 dendritic trees instead of 1 among 73 transplanted ACs; however, such dendritic mis-projection did not perturb IPL formation (Figure 4–figure supplement 1A). On the other hand, as with mutant donor ACs transplanted into mutant host retinas, when wild-type donor ACs were transplanted to mutant host retinas, they showed irregular neurite projection with many somas abnormally located toward the basal side, resulting in IPL formation defects (Figure 4C, 4D and Figure 4–figure supplement 1A). These data suggest a non-cell autonomous function of Strip in ACs for IPL formation.

Similarly, we conducted cell transplantation to assess the role of Strip1 in BPs. Mutant donor BPs labeled with the transgene *Tg[xfz43]* (Zhao et al., 2009) normally projected axons toward the IPL in wild-type host retinas, in the same fashion as wild-type donor BPs (Figure 4–figure supplement 1B-1D). Few transplanted columns of mutant donors showed extra lateral branching and excessive elongation of BP arbors (Figure 4–figure supplement 1D, arrows). However, such arbor defects did not disrupt the IPL. On the other hand, when wild-type donor BPs labeled with the transgene, *Tg[xfz3]*, were transplanted into a mutant host retina, wild-type donor bipolar axons failed to project toward the mutant host IPL, but rather seemed to be guided toward the wild-type donor IPL (Figure 4–figure supplement 1E-1G). Thus, Strip1 is not required in ACs or BPs for neurite projection to the IPL, although we do not exclude the possibility

that Strip1 is cell-autonomously required in a small subset of ACs and BPs to regulate dendritic branching and neurite extension. Taken together, it is likely that Strip1-mediated RGC maintenance is essential for proper neurite patterning of ACs and BPs and subsequent IPL formation.

Strip1 interacts with Striatin3 to regulate RGC survival

To identify which molecules interact with Strip1 to regulate RGC survival, we conducted a co-immunoprecipitation experiment coupled with mass spectrometry (Co-IP/MS). Head lysates of wild-type embryos combined with the transgenic line *Tg[hsp:WT.Strip1-GFP]* were used to pull-down wild-type Strip1 and its interacting partners. As a negative control, we used lysates from 2 other lines; *Tg[hsp:Mut.Strip1-GFP]* and *Tg[hsp:Gal4;UAS:GFP]*, to rule out proteins enriched by the mutant form of Strip1 or by GFP alone (Figure 5A and 5B). Six proteins were enriched only by the wild-type form of Strip1, 5 of which are components of the STRIPAK complex (Figures 5C, 5D, and Figure 5–figure supplement 1A and 1B). Since none of these components has been studied in zebrafish, we analyzed previously published single-cell RNA sequencing data on transcriptomes from zebrafish embryonic retinas at 2 dpf (Xu et al., 2020), and found that only *strip1* and *striatin3 (strn3)* mRNA are abundantly expressed in retinal cells (Figure 5–figure supplement 1C-1I). Next, we knocked down Strn3 using morpholino and observed a significant increase in apoptotic cells in the GCL at 60 hpf (Figure 5E and 5F). This leads to a significant reduction in RGCs at 60 and 76 hpf, as assessed by the *ath5:GFP* signal (Figures 5G-5I). Although *strn3* morphants showed a similar RGC loss to *strip1* mutants, it was weaker. IPL defects were also milder in *strn3* morphants than in *strip1*^{rw147} mutants at 76 hpf (Figure 5H). Taken together, Strip1 and Strn3 cooperatively regulate RGC survival.

Jun is a key mediator of RGC death in the absence of Strip1

To determine what kinds of molecules mediate RGC apoptosis in *strip1* mutants, we performed RNA sequencing on transcriptomes from 62-hpf eye cups of *strip1*^{rw147} mutants. Compared to wild-type siblings, *strip1* mutants had 131 significantly upregulated genes and 75 downregulated genes (Figure 6A, $\log_2$FC>|1|, and

FDR<0.05). Most downregulated genes were markers of RGCs, like *isl2b*, *pou4f3* (also known as *brn3c*) and *tbr1b,* which reflects the reduction in RGCs. Genes related to synaptic development and transmission were also downregulated (Figures 6A, 6B and Figure 6–figure supplement 1A). On the other hand, many of the significantly upregulated genes were related to apoptosis, oxidative phosphorylation, cellular response to stress, and the MAP kinase (MAPK) signaling pathway (Figure 6A, 6B and Figure 6–figure supplement 1B). Most reports of RGCs undergoing stress are in glaucoma and optic nerve injury models, where adult RGCs undergo cell death in response to injury (Bahr, 2000). So, we compared the transcriptomic profiles of *strip1* mutant eyes to those of adult zebrafish RGCs following optic nerve injury (Veldman et al., 2007) or adult eyes after optic nerve crush (McCurley and Callard, 2010). Indeed, there were several genes commonly upregulated in all three models, namely, *jun, atf3, gap43, stmn4, sox11b,* and *adcyap1b* (Figure 6–figure supplement 1C). These findings suggest that Strip1-deficient zebrafish RGCs share a similar stress response with adult RGCs following optic nerve injury.

In *strip1*^{rw147} mutants, *jun* was among the top upregulated stress response markers. Jun (the zebrafish homolog of mammalian c-Jun) is the canonical target of the Jun N-terminal kinase (JNK) pathway, which belongs to the MAPK super family. JNK/c-Jun signaling is a key regulator of stress-induced apoptosis (Dhanasekaran and Reddy, 2008, Ham et al., 2000). Activation of the JNK pathway involves phosphorylation events that end with c-Jun phosphorylation and transactivation, which in turn activates *c-jun* gene expression (Eilers et al., 1998). We stained *strip1*^{rw147} mutant retinas with anti-phosphorylated c-Jun (p-Jun) antibody. At 54 hpf, *strip1*^{rw147} mutants showed significantly elevated levels of p-Jun compared to wild-type siblings. This elevation is specifically localized in RGCs visualized with zn5 antibody (Figures 6C and 6D). Likewise, p-Jun was significantly elevated in RGCs of *strn3* morphants at 49 hpf compared to control injected embryos (Figures 6E and 6F). Next, we knocked down Jun using morpholino (MO-jun). At 60 hpf, apoptosis was significantly inhibited in the GCL of *strip1*^{rw147} mutants injected with MO-jun, compared to *strip1*^{rw147} mutants injected with a standard control morpholino (Figures 6G and 6H). Accordingly, at 76 hpf, RGCs were partially but significantly recovered in *strip1*^{rw147} mutants injected with MO-jun,

compared to *strip1*^{rw147} mutants injected with a standard control morpholino (Figures 6I and 6J). Taken together, Strip1 interacts with Strn3 to suppress Jun-mediated apoptotic signaling in RGCs.

Bcl2 rescues RGC survival in *strip1* mutants, but surviving RGCs do not project their dendrites to the IPL

The anti-apoptotic B-cell lymphoma 2 (Bcl2) is a key regulator of mitochondria-dependent apoptosis in neurons, including RGCs, both during survival and in response to injury (Anilkumar and Prehn, 2014, Bahr, 2000, Bonfanti et al., 1996, Maes et al., 2017). In addition, JNK/c-Jun activation induces neuronal apoptosis by modulating BCL2 family proteins (Guan et al., 2006, Hollville et al., 2019, Whitfield et al., 2001). Thus, we combined *strip1*^{rw147} mutants with the transgenic line *Tg[hsp:mCherry-Bcl2]*, which overexpresses mCherry-tagged Bcl2 protein under control of a heat shock promoter (Nishiwaki and Masai, 2020). Bcl2 overexpression significantly inhibited RGC apoptosis in *strip1*^{rw147} mutants (Figures 7A and 7B). Accordingly, at 78 hpf, RGCs were partially but significantly recovered in *strip1*^{rw147} mutants overexpressing Bcl2, compared to non-transgenic mutants (Figures 7C and 7D). Thus, loss of RGCs in *strip1*^{rw147} mutants depends on the mitochondria-mediated apoptotic pathway.

Surprisingly, *strip1*^{rw147} mutants overexpressing Bcl2 still displayed IPL defects. In this case, the IPL was not formed at the interface between surviving RGCs and ACs, but instead, thin IPL-like neuropil was ectopically formed in the intermediate position of presumptive ACs (Figure 7C). Thus, a fraction of presumptive ACs were abnormally located between surviving RGCs and IPL-like neuropil, although this AC fraction does not intermingle with surviving RGCs (Figure 7C, bottom panels, asterisks). In addition, surviving RGCs in *strip1*^{rw147} mutants apparently fail to project their dendrites to the IPL-like thin neuropil. Upon closer examination, few surviving RGCs successfully innervate the IPL, and such areas show less infiltration of ACs (Figure 7C, bottom panels, arrow heads). These data suggest an additional role of Strip1 in dendritic patterning of RGCs, which is likely to prevent ectopic IPL-like neuropil formation in the AC layer.

Discussion

Over the past decade, Strip1/Strip has emerged as an essential protein in embryonic development (Bazzi et al., 2017, La Marca et al., 2019, Neal et al., 2020, Sakuma et al., 2014, Sakuma et al., 2015). Using zebrafish, we demonstrate that Strip1 performs multiple functions in development of inner retinal neural circuit. First, we discovered a novel neuroprotective mechanism governed by Strip1, probably through the STRIPAK complex, to suppress Jun-mediated pro-apoptotic signaling in RGCs during development (Figure 8A). In addition, we demonstrate that Strip1-mediated RGC maintenance is essential for laminar positioning of other retinal neurons and structural integrity of the IPL.

RGCs are the most susceptible retinal neurons to cell death, both during development and in response to injury. Unlike zebrafish retina, in which only 1.06 % of RGCs die during development (Biehlmaier et al., 2001), around 50% of mammalian RGCs undergo apoptotic cell death (Bahr, 2000, Fawcett et al., 1984). Similarly, the survival rate of mouse RGCs following optic nerve injury (ONI) is only ~8% compared to a survival rate of ~75% of zebrafish RGCs (Li et al., 2020, Zou et al., 2013). This suggests that RGC survival signals are more active in zebrafish retina. Recently, many studies are seeking to identify such zebrafish-specific survival mechanisms, with the aim to develop therapy that can prevent death of mammalian RGCs (Chen et al., 2021). Interestingly, loss of zebrafish Strip1 causes an elevated apoptotic stress response profile in embryonic retinas with a degree of overlap with adult RGCs post-ONI. Indeed, 5 out of the 6 overlapping upregulated markers (*jun, atf3, stmn4, sox11b* and *adcyap1b)* are commonly upregulated in retinal transcriptomic studies of mammalian ONI models (Wang et al., 2021). Jun is the canonical target of JNK signaling and JNK/Jun activation is a major cause of axonal injury- or glaucoma-induced RGC death (Fernandes et al., 2012, Fernandes et al., 2013, Syc-Mazurek et al., 2017). Thus, Jun signaling appears to be a common mediator of RGC death among vertebrates. Our findings will open exciting new research avenues to determine whether Strip1-mediated Jun suppression can modulate proapoptotic signaling in adult RGCs of both zebrafish and higher vertebrates.

What is the molecular link between Strip1 and Jun suppression in RGCs? Our proteomic assays revealed that recruitment of many STRIPAK components is compromised in *strip1* mutants. Also, we demonstrate a functional interaction between Strip1 and Strn3 to regulate RGC survival. Recently, several studies on the human STRIPAK complex found that STRIP1 and STRN3 are organizing centers for the STRIPAK complex and that their mutant forms compromise complex assembly and function (Jeong et al., 2021, Tang et al., 2019). Modulation of JNK/Jun signaling by the STRIPAK complex is supported by several studies. MAP4 kinases activate the JNK signaling pathway and they are among kinase family members that are recruited and dephosphorylated by the STRIPAK complex (Fuller et al., 2021, Hwang and Pallas, 2014, Kim et al., 2020, Seo et al., 2020). Moreover, JNK signaling is activated in STRIP1/2 knockout human cell lines (Chen et al., 2019). Similarly, the interaction between Strip and CKa (*Drosophila* homolog of Striatins) suppresses JNK signaling in *Drosophila* testis (La Marca et al., 2019). Thus, it is likely that Strip1 and Strn3 function in the context of the STRIPAK complex to modulate JNK/Jun activity, thereby promoting RGC survival. To our knowledge, this study is the first *in vivo* evidence for a functional interaction between STRIPAK components and Jun signaling in vertebrates.

There are still many gaps in our knowledge of mechanisms underlying IPL development. Which molecular cues dictate the laminar positioning of inner retinal neurons? Which cell types are essential for IPL formation? It has been proposed that IPL development is a robust process. Upon genetic elimination of different inner retinal cells, the remaining cells manage to form an IPL-like neuropil (Randlett et al., 2013). However, it is widely agreed that ACs play the dominant role in IPL initiation. Elegant time-lapse experiments show that ACs project their neurites to form a proto-IPL (Chow et al., 2015, Godinho et al., 2005). This presumed IPL guides RGCs to extend dendritic arbors and stratify, although they are born earlier than ACs (Mumm et al., 2006). Other studies propose an active role for RGCs in shaping the developing IPL (Kay et al., 2004). We found that RGC death in *strip1* mutants is strongly linked to abnormal infiltration of ACs and BPs in the GCL and the perturbed IPL. This coincides with the phenotype of the *lakritz* mutant, in which similar defects occur when RGC genesis is inhibited (Kay et al., 2001, Kay et al., 2004). Knockout mice in which atypical Cadherin

Fat3 is absent in both RGCs and ACs, ACs invade the GCL abnormally. However, AC-specific knock out mice do not exhibit such positioning defects (Deans et al., 2011). Therefore, we re-introduce a model proposed by (Kay et al., 2004), in which both RGCs and ACs play distinct roles in shaping the developing IPL. In this model, RGCs provide positional cues for migrating ACs to initiate proper IPL program, whereas ACs subsequently project their dendritic plexus and establish the foundation for a proto-IPL.

So far, the RGC-dependent mechanisms that instruct the laminar positioning of ACs and IPL development remain unknown. However, Bcl2-rescued *strip1* mutants provide valuable new insights on such mechanisms. Bcl2-rescued mutants show defects in dendritic patterns of surviving RGCs, which are associated with an ectopic IPL formed within presumptive ACs. Thus, we propose that RGCs serve dual functions in IPL development (summarized in Figure 8B); (1) RGCs act as a physical barrier that prevents abnormal infiltration of ACs into the GCL and (2) RGCs show dendritic guidance cues that establish interactions between RGCs and ACs for a proper IPL program. It is unclear what guidance cues participate in this process. Possible candidates are N-Cadherin and Semaphorin-3 receptors, Neuropillin-1 (Nrp1) and PlexinA1. A hypomorphic allele of zebrafish *Ncad* mutants compromises IPL formation with abnormal neurite patterning of INL cells (Masai et al., 2003). In *Xenopus*, Nrp1 and PlexinA1 inhibition induced randomly oriented dendritic patterning of RGCs, similar to zebrafish *strip1* mutants (Kita et al., 2013). Future experiments will clarify whether these molecules participate in communication among RGCs and ACs to establish a proper IPL. Lastly, we show that Strip1 may function in neurite patterning of RGCs, and small subsets of ACs and BPs. This is supported by established roles of *Drosophila* Strip in dendritic branching and axon elongation (Sakuma et al., 2014). Future studies on cell-specific Strip1 knockout models will help understand Strip1 function in retinal neurite morphogenesis.

In summary, we demonstrate that a series of Strip1-mediated regulatory mechanisms constructs retinal neural circuit through RGC survival and neurite patterning of retinal neurons. Our findings provide valuable insights to mechanistic understanding of JNK/Jun-mediated apoptotic pathway and correct assembly of synaptic neural circuits in the brain. For medical perspective, our findings on a similar

stress response between zebrafish *strip1* mutants and mammalian ONI models pave the way for future research on potential Strip1-mediated therapeutic targets that could help mitigate RGC degeneration in glaucoma and optic neuropathies.

Author contributions

Conceptualization: MA, IM; Methodology: MA, IM; Software: MA; Validation: MA; Formal analysis: MA; Investigation: MA, YK, IM; Resources: MA, IM; Data curation: MA, IM; Writing - original draft: MA; Writing - review & editing: MA, IM; Visualization: MA, IM; Supervision: IM; Project administration: MA, IM; Funding acquisition: IM.

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Declaration of interests

The authors declare no competing interests.

Materials and methods

Key Resources Table				
Reagent type (species) or resource	**Designation**	**Source or reference**	**Identifiers**	**Additional information**
genetic reagent (*Danio rerio*)	*Tg[hsp:gal4]*^{kca4}	(Scheer et al., 2002)	ZDB-ALT-020918-6	
genetic reagent (*Danio rerio*)	*Tg[UAS:EGFP]*	(Koster and Fraser, 2001)	N/A	
genetic reagent (*Danio rerio*)	*Tg[UAS:MYFP]*	(Schroeter et al., 2006)	N/A	
genetic reagent (*Danio rerio*)	*Tg[ath5:GFP]* ^{rw021}	(Masai et al., 2003)	N/A	
genetic reagent (*Danio rerio*)	*Tg[ptf1a:mCherry-CAAX]*^{oki067}	This paper	N/A	
genetic reagent (*Danio rerio*)	*Tg[Gal4-VP16,UAS:EGFP]xfz43* or *xfz43*	(Zhao et al., 2009)	ZDB-ALT-100201-1	
genetic reagent (*Danio rerio*)	*Tg[Gal4-VP16,UAS:EGFP]xfz3* or *xfz3*	(Zhao et al., 2009)	ZDB-ALT-100201-2	
genetic reagent (*Danio rerio*)	*Tg[hs:mCherry-tagged Bcl2]*^{oki029}	(Nishiwaki and Masai, 2020)	ZDB-ALT-210524-5	
genetic reagent (*Danio rerio*)	*Tg[hsp:WT.Strip1-GFP]* ^{oki068}	This paper	N/A	
genetic reagent (*Danio rerio*)	*Tg[hsp:Mut.Strip1:GFP]* ^{oki069}	This paper	N/A	
genetic reagent (*Danio rerio*)	*strip1*^{rw147}	This paper	N/A	
genetic reagent (*Danio rerio*)	*strip1*$^{crispr\Delta 10}$ or *strip1*^{oki8}	This paper	N/A	
genetic reagent (*Danio rerio*)	*roy*	(D'Agati et al., 2017)	ZDB-GENE-040426-1168	
antibody	Mouse anti-acetylated α-tubulin	Sigma-Alrich	T6793	1:1000

antibody	Rabbit anti-Pax6	BioLegned	901301	1:500
antibody	Rabbit anti-Prox1	Genetex	GTX128354	1:500
antibody	Mouse anti-PCNA	Sigma-Aldrich	P8825	1:200
antibody	Mouse Zpr1	ZIRC	ZDB-ATB-081002-43	1:100
antibody	Mouse Zpr3	ZIRC	ZDB-ATB-081002-45	1:100
antibody	Mouse anti-glutamine synthetase	Sigma-Alrich	MAB302	1:150
antibody	Mouse Zn5	ZIRC	ZDB-ATB-081002-19	1:50
antibody	Rabbit anti-p-Jun	Cell Signal	9164S	1:100
antibody	Rabbit anti-Strip1	This paper	N/A	IF: 1:1000 WB: 1:500
antibody	Goat anti-rabbit Alexa 488 secondary antibody	Life Technologies	A11034	1:500
antibody	Goat anti-mouse Alexa 488 secondary antibody	Life Technologies	A11029	1:500
antibody	Goat anti-mouse Alexa 546 secondary antibody	Life Technologies	A11030	1:500
antibody	Goat anti-mouse Alexa 647 secondary antibody	Life Technologies	A21236	1:500
antibody	Anti-rabbit IgG, HRP-linked Antibody	Cell Signalling	7074	1:5000
antibody	Rabbit anti-GFP	Thermo Fisher Scientific	A11122	1:500
antibody	Rabbit anti β-actin	Abcam	AB8227	1:5000
recombinant DNA reagent	pTol2[ptf1a:mCherry-CAAX]	This paper	N/A	

recombinant DNA reagent	pG1[ptf1a:GFP]	Francesco Argenton Laboratory (Jusuf et al., 2012)	N/A	
recombinant DNA reagent	pTol2[hsp:WT.Strip1-GFP]	This paper	N/A	
recombinant DNA reagent	pTol2[hsp:Mut.Strip1:GFP]	This paper	N/A	
recombinant DNA reagent	pBluescript SK (+)	Stratagene	N/A	
recombinant DNA reagent	pT2AL200R150G	(Urasaki et al., 2006)	N/A	
recombinant DNA reagent	pB[ath5:Gal4-VP16]	This paper	N/A	
recombinant DNA reagent	pZNYX-GalVP16	Rachel Wong Laboratory (Schroeter et al., 2006)	N/A	
sequence-based reagent	Standard control MO (STD-MO)	GeneTools	N/A	5'-CCTCTTACCTCAGTTACAATTTATA-3'
sequence-based reagent	Strip1 morpholino (MO-strip1)	GeneTools	N/A	5'-TAGCACATAAACCGACACCGTCCAT-3'
sequence-based reagent	Ath5 morpholino (MO-ath5)	GeneTools	ZDB-MRPHLNO-100405-2	5'-TTCATGGCTCTTCAAAAAAGTCTCC-3'
sequence-based reagent	Striatin3 morpholino (MO-strn3)	GeneTools	N/A	5'-CCTGCTAGAAGTCGCCGATTGTTAC -3'
sequence-based reagent	Jun morpholino (MO-jun)	GeneTools	ZDB-MRPHLNO-080908-1	5' -CTTGGTAGACATAGAAGGCAAAGCG - 3'
peptide, recombinant protein	Cas9 protein	FASMAC	GE-006-S	

commercial assay or kit	JB-4® Embedding Kit	Polysciences	00226-1	
commercial assay or kit	In Situ Cell Death Detection Kit, TMR Red	Roche	12156792910	
commercial assay or kit	In Situ Cell Death Detection Kit, Fluorescein	Roche	11684795910	
commercial assay or kit	DIG RNA Labeling Kit	Roche	11277073910	
commercial assay or kit	GFP Trap Agarose	Chromotek	gta-20	
commercial assay or kit	Arcturus PicoPure RNA Isolation Kit	Thermo Fisher Scientific	KIT0204	
commercial assay or kit	NEB Next® Ultra™ II Directional RNA Library Prep Kit	New England BioLabs	E7760L	
chemical compound, drug	Acridine Orange (AO)	Nacalai tesque	1B-307	
chemical compound, drug	CellTrace™ BODIPY™ TR Methyl Ester	Thermo Fisher Scientific	C34556	
chemical compound, drug	Ethyl-3-aminobenzoate de methanesulfonate (Tricaine, MS-222)	Nacalai tesque	14805-82	
chemical compound, drug	PTU (N-Phenylthiourea)	Nacalai tesque	27429-22	
chemical compound, drug	Fast DiO solid	Thermo Fisher Scientific	D3898	
chemical compound, drug	Fast DiI solid	Thermo Fisher Scientific	D7756	
chemical compound, drug	TO-PRO™-3 Iodide (642/661)	Thermo Fisher Scientific	T3605	
chemical compound, drug	Hoechst 33342	Wako	346-07951	
chemical compound, drug	Toluidine Blue	Nacalai tesque	1B-481	

chemical compound, drug	Dextran, Tetramethylrhodamine	Thermo Fisher Scientific	D1817 LTJ	
chemical compound, drug	Dextran, Alexa Flour-488	Thermo Fisher Scientific	D22910	
chemical compound, drug	Dextran, Alexa Flour-647	Thermo Fisher Scientific	D22914	
chemical compound, drug	Dextran, Cascade Blue	Thermo Fisher Scientific	D1976	
software, algorithm	chopchop	chopchop	https://chopchop.cbu.uib.no	
software, algorithm	ImageJ (Fiji)	(Schneider et al., 2012)	https://imagej.nih.gov/ij/; RRID: SCR_003070	
software, algorithm	Imaris	Bitplane	http://www.bitplane.com/imaris; RRID: SCR_007370	
software, algorithm	Proteome Discoverer	Thermo	https://www.thermofisher.com/store/products/OPTON-30945#/OPTON-30945	
software, algorithm	STRING	(Szklarczyk et al., 2017)	https://string-db.org	
software, algorithm	Metascape	(Zhou et al., 2019)	https://metascape.org	
software, algorithm	R v3.4.2 , RStudio interface	(Team, 2016)	https://string-db.org	
software, algorithm	BioVenn	(Hulsen et al., 2008)	http://www.biovenn.nl/	
software, algorithm	Graphpad Prism v9.1.0.	Graphpad Prism	https://www.graphpad.com/scientific-software/prism/	

Ethics statement

Zebrafish (*Danio rerio*) were maintained on a 14:10 hour light: dark cycle at 28°C. Collected embryos were cultured in E3 embryo medium (5mM NaCl, 0.17mM KCl, 0.33mM $CaCl_2$, 0.33mM $MgSO_4$) containing 0.003% 1-phenyl-2-thiouera (PTU) to prevent pigmentation and 0.01% methylene blue to prevent fungal growth. All experiments were performed on zebrafish embryos between 36 hpf and 4 dpf prior to sexual differentiation. Therefore, sexes of the embryos could not be determined.

All zebrafish experiments were performed in accordance with the Animal Care and Use Program of Okinawa Institute of Science and Technology Graduate School (OIST), Japan, which is based on the Guide for the Care and Use of Laboratory Animals by the National Research Council of the National Academies. The OIST animal care facility has been accredited by the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC International). All experimental protocols were approved by the OIST Institutional Animal Care and Use Committee.

Transgenic fish lines

To visualize RGCs, the transgenic line *Tg[ath5:GFP]*^{rw021} *(Masai et al., 2003)* was used. In this line, GFP is expressed under control of the *ath5* (also referred to as *atoh7*) promoter. The transgenic lines *Tg(Gal4-VP16,UAS:EGFP)xfz43* or *xfz43* and *Tg(Gal4-VP16,UAS:EGFP)xfz3* or *xfz3* (Zhao et al., 2009) are enhancer trap lines and were both used to label distinct populations of BPs. To visualize ACs, the transgenic line *Tg[Ptf1a:mCherry-CAAX]*^{oki067} was generated by injecting the plasmid *pTol2[ptf1a:mCherry-CAAX]* into one-cell-stage fertilized eggs, together with Tol2 transposase mRNA.

For Bcl2 overexpression experiments, the line *Tg[hs:mCherry-tagged Bcl2]*^{oki029} (Nishiwaki and Masai, 2020) was employed, in which mCherry-tagged Bcl2 at the N-terminus is overexpressed under control of the heat shock promoter. For over-expression of wild-type and *rw147* mutant forms of zebrafish Strip1, the lines *Tg[hsp:WT.Strip1-GFP]*^{oki068} and *Tg[hsp:Mut.Strip1:GFP]* ^{oki069} were generated, respectively, to express GFP-tagged Strip1 at the C-terminus under control of the heat shock promoter. For line generation, the DNA constructs *pTol2[hsp:WT.Strip1-GFP]* and

pTol2[hsp:Mut.Strip1-GFP] were injected into one-cell-stage fertilized eggs together with Tol2 transposase mRNA. These injected F0 embryos were bred up to the adult stage and used to identify founder fish that produce F1 generation embryos showing stable GFP expression. Transgenic lines were established in F2 generation. The transgenic line *Tg[hsp:Gal4;UAS:EGFP]* was generated by combining *Tg[hsp:gal4]*^{kca4} (Scheer et al., 2002) with *Tg[UAS:EGFP] (Koster and Fraser, 2001)* to express EGFP under control of the heat shock promoter, while *Tg[UAS:MYFP]* expresses EYFP fused to the membrane targeting palmitoylation signal of gap43 under the control of 14XUAS E1b promoter (Schroeter et al., 2006). Some of the mentioned transgenic lines were combined with the mutant line *strip1*^{rw147}. The steps of plasmid construction are mentioned below in details.

Mutant line generation, mutant Identification, and genotyping

The *strip1*^{rw147} mutant line was generated from a mutagenesis screen (Masai et al., 2003) that used RIKEN Wako (RW) as a wild-type strain. Mutation mapping and subsequent experiments were carried out in the genetic background of WIK and Okinawa wildtype (oki), respectively. The *rw147* mutation was mapped on a genomic region in chromosome 22 flanked by two self-designed polymorphic markers; AL928817-12: (5'-TTCAACATCTGCTTTTCCTCCT-3' and 5'TCATGTCCCAGAAATCACACAT3') and zk253D23-4 (5'-CATTCTTCATTAAAGAGATCAGTGTGA-3' and 5'-AGTGATCACACACCCCCACT-3'). In addition, location of the *rw147* mutation was further restricted using another self-designed polymorphic marker Zk286J17-3 (5'- TTCACATTTACATTTTTCTGAACATTT-3' and 5'-CACACAGCCTTCTCTTGCAC -3') as no recombination was detected.

From 3 dpf, *strip1*^{rw147} homozygous mutants (*strip1*$^{-/-}$) are distinguished from wild-type siblings (*strip1*$^{+/+}$ or *strip1*$^{+/-}$) by external morphology since they exhibit cardiac edema, an abnormal lower jaw, and smaller eyes. From 54 to 72 hpf, *strip1*^{rw147} homozygous mutants are screened by Acridine Orange (AO) live staining to detect apoptotic cells (Casano et al., 2016) or Bodipy TR live staining to visualize lamination defects (Choi et al., 2010). Prior to 54 hpf, genotyping of *strip1*^{rw147} mutant embryos was performed by sequencing. The primer set, 5'-CGTGTGTTTTCAGGGTGTT-3' and 5'-

TCACCATCCCAAACAGCATA-3', was used. The 257-bp PCR amplicons were amplified using Phusion Hot Start II (Thermo Fisher Scientific) and sequenced for genotyping.

The *strip1*$^{crispr\Delta 10}$ (officially referred to as *strip1*^{oki8}) mutant line was generated using Crispr/Cas9 gene editing technology. The gRNA sequence, 5'-CCCGCGTCCGCCTCTGACCTCAT-3', was designed using chopchop (https://chopchop.cbu.uib.no) and it targets exon 9 of *strip1* gene. One-cell-stage embryos were injected with 200ng/µL gRNA and 500ng/µL Cas9 protein (FASMAC). F1 mutant founders were identified by sequencing. A 10-bp deletion was introduced at nucleotide 932 of the Strip1 coding sequence, resulting in a frameshift at amino acid 313 and a premature stop at amino acid 330. The primer set 5'-CGTTCCAAATCATTGAAACAGA-3' and 5'-TGTTTGTGATGTGTTGACCTTG-3' was used for genotyping. PCR amplicons were run on 15% polyacrylamide gel for identification of wild-type siblings and mutants.

All generated transgenic and mutant lines were combined with the zebrafish pigmentation mutant, *roy orbison (roy) (D'Agati et al., 2017)* to remove iridophores and enhance live imaging.

Molecular cloning

To generate pTol2[hsp:WT.Strip1-GFP] and pTol2[hsp:Mut.Strip1:GFP], a PCR strategy was used to amplify ~2.5 kb Strip1 cDNA from total cDNA of 4 dpf wild-type and *strip1*^{rw147} zebrafish embryos using the primers 5'-AGACTTGTGTCAGCGTGACGCGAG-3' and 5'-ACTCTAGCAAGTGTAGTGTTGTTGATG-3'. Then, using a Gibson Assembly Cloning Kit, a *strip1* cDNA fragment was cloned into a Tol2 transposon vector pT2AL200R150G (Urasaki et al., 2006) at the *XhoI* and *ClaI* sites with a heat-shock inducible promoter (hsp) at the N-terminus (Halloran et al., 2000) and a GFP tag at the C-terminus (separated by a linker sequence, CTCGAGGGAGGTGGAGGT). For pTol2[ptf1a:mCherry-CAAX] construction, pG1[ptf1a:GFP] was used as donor plasmid, which was kindly provided by the Francesco Argenton lab. A 5.5-kb fragment of the *ptf1a* promoter sequence was retrieved at *HindIII* and *SmaI* sites and inserted into a

pBluescript SK (+) (Stratagene) shuttle vector upstream of the membrane-targeting mCherry-CAAX sequence. Then, the ptf1a:mCherry-CAAX sequence was inserted into the *XhoI and BglII* sites of pT2AL200R150G. The pB[ath5:Gal4-VP16] plasmid was constructed by inserting a 6.6-kb fragment of the ath5 5'-enhancer/promoter region (including 5' UTR) into the *BamHI* site of the pB[Gal4-VP16] plasmid provided by Dr. R. Köster (Koster and Fraser, 2001).

In vivo **cell labeling**

Single-cell mosaic labeling to visualize the morphology of RGCs was done by injecting 20 ng/µL of pB[ath5:Gal4] into 1-cell-stage embryos from intercrosses of *strip1*^{rw147} heterozygous fish combined with *Tg[UAS:MYFP]* (Schroeter et al., 2006). Likewise, pZNYX-GalVP16, a kind gift from the Rachel Wong Laboratory, was injected to visualize ON-BPs (Schroeter et al., 2006). Single AC labeling was performed by injecting the DNA construct pG1[ptf1a:GFP] into 1-cell-stage embryos from intercrosses of *strip1*^{rw147} heterozygous fish at concentration 20 ng/µL (Jusuf et al., 2012).

Morpholino knockdown assay

Embryos produced by intercrosses of wild-type or *strip1*^{rw147} heterozygous fish were injected with antisense morpholino oligonucleotides at 1-cell stage. MO-strip1, MO-strn3, and MO-ath5 (Ranawat and Masai, 2021) were injected at a concentration of 250 µM, whereas MO-jun (Han et al., 2016) was injected at a concentration of 125 µM. For each morpholino experiment, the same concentration of the standard control morpholino (STD-MO) was used as a negative control. Detailed morpholino sequences are listed in the Key resources table.

DiI/DiO Injections

To trace the RGC axon projections into the optic tectum, 3-dpf embryos were fixed in 4% paraformaldehyde (PFA) and after washing with PBS several times, were injected with 2 mg/mL of the lipophilic dyes, DiI and DiO, in the area between the lens and retina. Large injection volumes were applied to label all RGCs. Embryos were incubated overnight at 4°C, and then mounted in 75% glycerol for confocal imaging.

Histological methods

Zebrafish embryos were embedded for JB4 plastic sectioning and toluidine blue counterstaining, as previously described (Sullivan-Brown et al., 2011). Immunolabeling of cryosections and paraffin sections were carried out according to standard protocols (Imai et al., 2010, Masai et al., 2003). An antigen retrieval step was performed on paraffin sections by heating in 10 mM citrate buffer pH 6.0 for 5 min at 121°C. TUNEL (terminal deoxynucleotidyl transferase dUTP nick end labeling) was performed using an In Situ Cell Death Detection Kit (Roche) according to the manufacturer's protocol. Nuclear staining was carried out using 1nM TOPRO3 or 1ng/mL Hoechst 33342.

Antibodies used in this study and their dilutions are as follows: anti-acetylated α-tubulin (1:1000), anti-Pax6 and anti-Prox1 (1:500), anti-PCNA (1:200), anti-glutamine synthetase (1:150), antibodies against zpr1 and zpr3 (1:100), zn5 antibody (1:50) and anti-p-Jun (1:100). Antibody against the peptide sequence of zebrafish Strip1 (amino acids 344-362: EKDPYKADDSHEDEEENDD) was generated using a synthetic peptide and used for immunostaining at 1:1,000. For adsorption control, purified antibody was preincubated with 3.6 μg/mL of corresponding blocking peptide for 1 h at room temperature. Secondary antibodies employed in this study were: Alexa488, 546 and 647 fluorophore-conjugated secondary antibodies used at a concentration of 1:500.

For whole-mount immunostaining against acetylated α-tubulin, 3-dpf embryos were fixed at room temperature for 3 h in 2% trichloroacetic acid (TCA). Then, embryos were washed in PBTr (Westerfield, 1995) (PO_4 buffer (0.1M, pH7.3)+0.1% Triton X-100) followed by permeabilization in 0.2% trypsin for 4 min at 4°C. After washing, a post-fixation step in 4% PFA for 5 min at 4°C was applied. Next, blocking was done in 10% goat serum in PBTr for 1 h at room temperature followed by incubation in mouse anti-acetylated α-tubulin in 1% goat serum/PBTr overnight at 4°C. After washing, embryos were incubated in secondary antibody diluted in 1% goat serum in PBTr overnight at 4°C. Whole-mount immunostaining against p-Jun was performed following standard protocols (Ungos et al., 2003). After staining, embryos were mounted in 75% glycerol for confocal imaging.

Whole-mount *in situ* hybridization was performed on wild-type zebrafish embryos at specific developmental stages as previously described (Xu et al., 1994). Hybridization was performed overnight at 65°C using *strip1* RNA probe at the concentration 2.5 ng/µL in hybridization buffer. *strip1* probe synthesis was performed according to standard protocols (Thisse and Thisse, 2008). Template regions were amplified from *strip1* cDNA using the primers, 5'-AATGCTGCCGAATAAAATGCGAG-3' and 5'-CCCAGAGTGAACAGGATGCTCT-3'. Antisense and sense probes were synthesized by *in vitro* transcription using a DIG RNA Labeling Kit (Roche). Following labeling, whole embryos were mounted in 75% glycerol for imaging. To visualize expression patterns in the retina, cryosections were prepared from whole mount embryos post-hybridization.

Live staining

To visualize lamination patterns, live staining of retinal landmarks was performed by incubating live zebrafish embryos in 100 nM solution of Bodipy TR methyl ester (Thermo Fisher Scientific) in E3 embryo rearing media for 1 h at room temperature following the manufacturer's protocol. To examine DNA condensation of apoptotic cells in the GCL, zebrafish embryos were incubated for 30 min in 5 µg/mL of AO stain dissolved in egg water. Following staining, embryos were extensively washed with egg water and observed using epifluorescence or imaged using confocal microscopy.

Overexpression experiments

For rescue experiments, the wild-type form of Strip1, the *rw147* mutant form of Strip1 or Bcl2 was overexpressed in *strip1^{rw14}* mutants by heat shock treatment using the transgenic lines *Tg[hsp:WT.Strip1-GFP]*, *Tg[hsp:Mut.Strip1:GFP]* and *Tg[hs:mCherry-tagged Bcl2]*, respectively. To perform heat shock, embryos from heterozygous intercrosses were incubated for 1 h at 39°C starting from 27-30 hpf and applied every 12 hours until the designated timepoints. For Co-IP/MS, heat shock was applied to embryos from intercrosses of wild-type zebrafish combined with *Tg[hsp:WT.Strip1-GFP]*, *Tg[hsp:Mut.Strip1:GFP] or Tg[hsp:Gal4;UAS:EGFP].* After screening for transgenic embryos, embryos were either fixed in 4% PFA for histological assays or processed for protein extraction.

Cell transplantation assays

Single-cell transplantations were performed at blastula stage, as previously described (Kemp et al., 2009). Genotypes of donor and host embryos were determined at 3-4 dpf based on morphological phenotype or they were genotyped at earlier time points by sequencing or AO live-staining of apoptotic cells. To trace transplanted donor cells in host retinas, 2-5% lysine-fixable dextran rhodamine, Alexa-488 dextran, Alexa-647 dextran or cascade blue dextran were injected in 1-2-cell-stage donor embryos, depending on study design. To assess the cell autonomy of Strip1 in RGC death, donor embryos from intercrosses of *strip1*^{rw147} heterozygous fish were transplanted into wild-type host embryos. Host embryos with successful retinal transplants were fixed in 4% PFA at 60 hpf and processed for TUNEL labeling. To assess the cell autonomy of Strip1 in AC or BP development, donor embryos from intercrosses of *strip1*^{rw147} heterozygous fish combined with *Tg[ptf1a:mCherry-CAAX]* or *Tg[Gal4-VP16,UAS:EGFP]xfz3/xfz43* were transplanted into embryos from intercrosses of *strip1*^{rw147} heterozygous fish, live imaging of host retinas with successful transplants was done at 3-4 dpf to assess the morphology of donor ACs labeled with mCherry or donor BPs labeled with EGFP. To visualize the retinal lamination phenotype, some hosts were stained with Bopidy TR live stain prior to imaging.

Microscopy

Imaging of toluidine blue-stained sections and retinal sections following *in situ* hybridization was performed using a Zeiss upright Axioplan2 equipped with an AxioCam HRC camera, while imaging of whole-mount *in situ* hybridization embryos was done using a Keyence BZ-X700. An inverted Zeiss LSM 780 was used to scan immunostained retinal sections with a 40x / 1.40 Plan-Apochromat Oil objective and whole-mount immunostained embryos using a 40x / 1.0 Plan-Apochromat water objective. Glycerol-mounted embryos were placed on glass-bottom depression slides for scanning. For live imaging, zebrafish embryos were anesthetized using 0.02% tricaine (3-amino benzoic acid ethyl ester) dissolved in E3 embryonic medium and mounted laterally in 1% low-melting agarose. Image acquisition of embryo retina was

carried out using an upright Zeiss LSM 710 with a 40x / 1.0 W Plan-Apochromat objective or an upright Fluoview FV3000 (Olympus) confocal microscope with a 40x / 0.8 water immersion objective.

To perform time-lapse imaging of retinal development, several embryos from intercrosses of *strip1*^{rw147} heterozygous fish carrying the transgenes *Tg[ath5:GFP;ptf1a:mCherry-CAAX]* were mounted simultaneously in a culture dish covered with E3 embryonic medium containing 0.003% PTU and 0.02% tricaine and overlayed with a thin layer of mineral oil to prevent evaporation of E3 medium to minimize embryotoxicity. Retinal z-stacks were acquired consecutively in 1-µm steps every ~2 hrs, starting at 48 hpf with undetermined genotypes. Scanning was done using the Multi Area Time Lapse (MATL) Software module of the FV3000 (Olympus) confocal microscope and a motorized XYZ-rotation stage.

All Images were processed using ImageJ (NIH, v2.1.0/1.53C), Imaris (Bitplane, v9.1.2) and Adobe Illustrator software. 3D rendering and analysis of time-lapse movies were performed on Imaris software. Whenever necessary, brightness and contrast display levels for the whole image were adjusted to aid visualization or decrease background noise.

Western Blotting and Co-immunoprecipitation (CO-IP)

Heads of non-injected or MO-strip1-injected wild-type embryos were dissected at 2 dpf in Leibovitz's L-15 ice cold medium and homogenized in lysis buffer (125mM NaCl, 50 mM Tris (pH 7.5), 0.5 mM EDTA (pH 8), 1% Triton X-100 and 1× cocktail protease inhibitors). Lysates were clarified by centrifugation at 10,000 g for 10 min at 4 °C. Equal amounts of denatured clarified lysates were run on 10% Mini-PROTEAN TGX gels for SDS-PAGE and transferred to PVDF membranes using Trans-Blot Turbo PVDF Transfer system. After blocking with 5% skim milk in 0.1% tween-20 in TBS, immunoblotting was performed using anti-Strip1 (1:500). HRP-linked rabbit IgG was used as a secondary antibody. Chemiluminescence signals were detected using a FUJI Las 4000 luminescence image analyzer.

For co-immunoprecipitation, wild-type embryos carrying the transgenes *Tg[hsp:WT.Strip1-GFP], Tg[hsp:Mut.Strip1-GFP]* or *Tg[hsp:Gal4;UAS:GFP]* were

exposed to 12-hr interval heat shocks starting at 27 hpf. At 2 dpf, lysates for each biological replicate were prepared from a pool of around 150 embryo heads in NP-40-based lysis buffer (150mM NaCl, 10 mM Tris (pH 7.5), 0.5 mM EDTA (pH 8), 0.5% NP-40 and 1× cocktail protease inhibitors), as described above. Immunoprecipitation was performed on clarified lysates using anti-GFP (GFP-Trap agarose beads, Chromotek) according to the manufacturer's protocol. Briefly, clarified lysates were diluted in wash buffer (150mM NaCl, 10 mM Tris (pH 7.5), 0.5 mM EDTA (pH 8) and 1× cocktail protease inhibitors) to reach 0.1% NP-40. Then, lysates were incubated with pre-equilibrated GFP-Trap beads for 1 h at 4°C. Afterwards, beads were collected by centrifugation and washed in wash buffer 5 times. To confirm that GFP-fused proteins were successfully pulled down, proteins were eluted from beads by boiling in 1x sample buffer for 5 mins. Then, 5% of pre-pulldown lysate (input) and 10% of the pulled-down proteins were run for western blotting as described above using anti-GFP (1:500).

Mass spectrometry (MS) and data analysis

To prepare protein samples for MS analysis, immunoprecipitated protein complexes were eluted from GFP-Trap beads using an on-bead trypsin-based digestion protocol according to manufacturer's protocol. Digestion was performed overnight at 32°C and under rotation at 400 rpm. Afterwards, digested peptides were cleaned and desalted using C_{18} stage tips, as previously described (Rappsilber et al., 2007). Eluted peptides were vacuum-dried and re-constituted in 1% acetic acid, 0.5% formic acid for MS analysis using an Orbitrap-Fusion Lumos mass spectrometer coupled to a Waters nanoACQUITY Liquid Chromatography System. Samples were trapped on a nanoACQUITY UPLC 2G-V/M Trap 5 µm Symmetry C_{18}, 180 µm × 20 mm column and analytical separation was performed on a nanoACQUITY UPLC HSS T3 1.8 µm, 75 µm × 150 mm column. Peptides were fractionated over a 60-min gradient from 1-32% acetonitrile with 0.1% formic acid. Solvent flow rate was 500 nL/min and column temperature was 40°C.

LC-MS raw data files were analyzed using Proteome Discoverer (PD, v.2.2, Thermo Fisher Scientific). The SEQUEST algorithm was used to match MS data to the *Danio rerio* (zebrafish) database downloaded from UniProt (July, 2021) and the common

Repository of Adventitious Protein (cRAP, https://www.thegpm.org/crap). Database search parameters included carbamidomethylation of cysteine as fixed modification and oxidation of methionine, deamidation of glutamine and asparagine as dynamic modifications. Trypsin was specified as a cleavage enzyme with up to 2 missed cleavages. Normalization was performed based on specific protein amount (Trypsin) and proteins were filtered based on a false discovery rate of $q < 0.05$. Abundance ratios were generated for wild-type compared to mutant and wild-type compared to GFP control. Enriched proteins with an abundance ratio ≥ 2 and adjusted p-value <0.05 were considered statistically significant. Search Tool for the Retrieval of Interacting Genes/Proteins (STRING, v11.0) was used to visualize the Strip1-interaction network with the enriched proteins and calculate protein-protein interaction (PPI) value (Szklarczyk et al., 2017).

RNA sequencing and analysis

Total RNA was isolated from 4 independent biological replicates of 60-64 hpf wild-type siblings and *strip*^{rw147} mutant eye cups using a PicoPure RNA Isolation Kit (Thermo Fisher Scientific) according to the manufacturer's instructions. Each biological replicate represented a pool of 20-30 eyes. All samples had RNA integrity number (RIN) values greater than 8.5. The purified RNA was used for Poly(A)-selected mRNA library preparation with a NEBNext® Ultra™ II Directional RNA Library Prep Kit for Illumina and sequenced on a NovaSeq6000 SP to generate 150 bp paired-end reads. Sequencing reads were quality checked using FastQC (Andrews, 2010) and trimmed with FastP (Chen et al., 2018). The resulting reads were mapped using hisat v2.1.0 (Kim et al., 2015) to the zebrafish reference genome (GRCz11). Mapped reads were counted using featureCounts, v1.6.2 (Liao et al., 2014) and differential gene expression analysis (Mutant vrs. Wildtype) was carried out on the counts files using the EdgeR package, v3.32.1 (Robinson et al., 2010) in RStudio, v1.4.1106 (Team, 2016). Genes with FDR <0.05 and $Log_2FC > |1|$ were considered statistically significant. EnhancedVolcano, v1.8.0 (Blighe et al., 2019) and pheatmap, v 1.0.12 (Kolde, 2012) packages were used to generate the volcano plot and heatmap, respectively. Gene Ontology analyses were performed using Metascape with *D. Rerio* as the input species and *M. Musculus* as the

analysis species (Zhou et al., 2019). To analyze published scRNA sequencing data of zebrafish retina at 48 hpf, raw count matrices were analyzed with the Seurat package, v4.0.1 (Satija et al., 2015), as previously described (Xu et al., 2020). Clustering results were visualized using Uniform Manifold Approximation and Projection (UMAP). The BioVenn web application was used to generate Venn diagrams to compare Upregulated DEGs in this study with published upregulated DEGs in optic nerve injury models (Hulsen et al., 2008).

Quantification and statistical analysis

To quantify RGC area, masks were generated for areas with strong ath5+ signals in the retina and quantified using the Color Threshold tool in ImageJ (Schneider et al., 2012). Afterwards, the retinal outline was defined using the lasso tool and retinal area was calculated. Data were represented as the percentage of ath5+ area to total retinal area. To calculate the area of p-Jun signal in RGCs, an ROI containing the GCL was defined based on zn5 antibody signal (antigen: alcama, previously referred to as DM-GRASP) to exclude noise at the retina boundary. Areas of p-Jun and zn5 signals were calculated as described above and data were represented as the percentage of p-Jun+ area to zn5+ area.

To quantify apoptotic cells, the number of TUNEL+ cells in GCL or retina was calculated manually within a single retinal section. To quantify the number of ptf1a+ cells that contribute to IPL formation (ACs), cells were manually counted in a unified area (8,500 μm^2) across all samples. Ptf1a+ cells that contributed to the OPL (presumably HCs) were excluded from quantification. To quantify the migration patterns of ptf1a+ cells that contribute to the IPL, cells located at the apical side relative to the IPL were assigned INL+, while cells located at the basal side of the IPL or near the lens were assigned GCL+. To determine the distribution of ptf1a+ cells within transplanted columns, ptf1a+ cells that contributed to the IPL were manually calculated in a z-stack and the distribution pattern was represented as the percentage of basally or apically located ptf1a+ cells to the total number of ptf1a+ cells. Numbers of strong Pax6+ and Prox1+ cells were calculated using the analyze particles tool in ImageJ and the distribution of cells (INL+ or GCL+) was assigned based on their location relative to the IPL, according

to the nuclear stain pattern. Distribution data were represented as the percentage of INL+ or GCL+ to the total number of Prox1+ or Pax6+ cells.

Statistical analysis was conducted using Graphpad Prism 9.1.0. Data are represented as means ±SD. Comparisons between two samples were done using the Mann Whitney U test or Student's t test with Welch's correction for normally distributed data. For multiple comparisons, 2-way ANOVA with the Tukey post hoc test was used. Details of statistical tests and number of samples used are in figures and figure legends. Significance level is indicated as $*p < 0.05$, $**p < 0.01$, $***p < 0.001$, $****p < 0.0001$, n.s. indicates not significant.

Data availability

Raw RNA-seq data files have been deposited in DDBJ under accession number DRA012640.

Mass spectrometry raw data and result files have been deposited in the ProteomeXchange Consortium (http://proteomecentral.proteomexchange.org) via the jPOST partner repository (https://jpostdb.org) (Okuda et al., 2017) under accession number PXD028131.

References

AMINI, R., ROCHA-MARTINS, M. & NORDEN, C. 2018. Neuronal Migration and Lamination in the Vertebrate Retina. *Frontiers in Neuroscience,* 11.

ANDREWS, S. 2010. FastQC: A Quality Control Tool for High Throughput Sequence Data.

ANILKUMAR, U. & PREHN, J. H. 2014. Anti-apoptotic BCL-2 family proteins in acute neural injury. *Front Cell Neurosci,* 8**,** 281.

AVANESOV, A. & MALICKI, J. 2010. Analysis of the retina in the zebrafish model. *Methods Cell Biol,* 100**,** 153-204.

BADEN, T., EULER, T. & BERENS, P. 2020. Understanding the retinal basis of vision across species. *Nat Rev Neurosci,* 21**,** 5-20.

BAHR, M. 2000. Live or let die - retinal ganglion cell death and survival during development and in the lesioned adult CNS. *Trends Neurosci,* 23**,** 483-90.

BAZZI, H., SOROKA, E., ALCORN, H. L. & ANDERSON, K. V. 2017. STRIP1, a core component of STRIPAK complexes, is essential for normal mesoderm migration in the mouse embryo. *Proc Natl Acad Sci U S A,* 114**,** E10928-E10936.

BIEHLMAIER, O., NEUHAUSS, S. C. & KOHLER, K. 2001. Onset and time course of apoptosis in the developing zebrafish retina. *Cell Tissue Res,* 306**,** 199-207.

BLIGHE, K., RANA, S. & LEWIS, M. 2019. EnhancedVolcano: Publication-ready volcano plots with enhanced colouring and labeling. *R package version,* 1.

BONFANTI, L., STRETTOI, E., CHIERZI, S., CENNI, M. C., LIU, X. H., MARTINOU, J. C., MAFFEI, L. & RABACCHI, S. A. 1996. Protection of retinal ganglion cells from natural and axotomy-induced cell death in neonatal transgenic mice overexpressing bcl-2. *J Neurosci,* 16**,** 4186-94.

CASANO, A. M., ALBERT, M. & PERI, F. 2016. Developmental Apoptosis Mediates Entry and Positioning of Microglia in the Zebrafish Brain. *Cell Rep,* 16**,** 897-906.

CHEN, R., XIE, R., MENG, Z., MA, S. & GUAN, K. L. 2019. STRIPAK integrates upstream signals to initiate the Hippo kinase cascade. *Nat Cell Biol,* 21**,** 1565-1577.

CHEN, S., LATHROP, K. L., KUWAJIMA, T. & GROSS, J. M. 2021. Retinal ganglion cell survival after severe optic nerve injury is modulated by crosstalk between JAK/STAT signaling and innate immune responses in the zebrafish retina. *bioRxiv***,** 2021.04.08.439090.

CHEN, S., ZHOU, Y., CHEN, Y. & GU, J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics,* 34**,** i884-i890.

CHEN, Y.-K., CHEN, C.-Y., HU, H.-T. & HSUEH, Y.-P. 2012. CTTNBP2, but not CTTNBP2NL, regulates dendritic spinogenesis and synaptic distribution of the striatin–PP2A complex. *Molecular biology of the cell,* 23**,** 4383-4392.

CHOI, J.-H., LAW, M.-Y., CHIEN, C.-B., LINK, B. A. & WONG, R. O. 2010. In vivo development of dendritic orientation in wild-type and mislocalized retinal ganglion cells. *Neural development,* 5**,** 29.

CHOW, R. W., ALMEIDA, A. D., RANDLETT, O., NORDEN, C. & HARRIS, W. A. 2015. Inhibitory neuron migration and IPL formation in the developing zebrafish retina. *Development,* 142**,** 2665-77.

D'AGATI, G., BELTRE, R., SESSA, A., BURGER, A., ZHOU, Y., MOSIMANN, C. & WHITE, R. M. 2017. A defect in the mitochondrial protein Mpv17 underlies the transparent casper zebrafish. *Dev Biol,* 430**,** 11-17.

D'SOUZA, S. & LANG, R. A. 2020. Retinal ganglion cell interactions shape the developing mammalian visual system. *Development,* 147.

D'ORAZI, F. D., SUZUKI, S. C. & WONG, R. O. 2014. Neuronal remodeling in retinal circuit assembly, disassembly, and reassembly. *Trends in neurosciences,* 37**,** 594-603.

DEANS, M. R., KROL, A., ABRAIRA, V. E., COPLEY, C. O., TUCKER, A. F. & GOODRICH, L. V. 2011. Control of neuronal morphology by the atypical cadherin Fat3. *Neuron,* 71**,** 820-32.

DHANASEKARAN, D. N. & REDDY, E. P. 2008. JNK signaling in apoptosis. *Oncogene,* 27**,** 6245-51.

DOWLING, J. E. 1987. *The retina: an approachable part of the brain,* Harvard University Press.

EILERS, A., WHITFIELD, J., BABIJ, C., RUBIN, L. L. & HAM, J. 1998. Role of the Jun kinase pathway in the regulation of c-Jun expression and apoptosis in sympathetic neurons. *J Neurosci,* 18**,** 1713-24.

FAWCETT, J. W., O'LEARY, D. D. & COWAN, W. M. 1984. Activity and the control of ganglion cell death in the rat retina. *Proc Natl Acad Sci U S A,* 81**,** 5589-93.

FERNANDES, K. A., HARDER, J. M., FORNAROLA, L. B., FREEMAN, R. S., CLARK, A. F., PANG, I. H., JOHN, S. W. & LIBBY, R. T. 2012. JNK2 and JNK3 are major regulators of axonal injury-induced retinal ganglion cell death. *Neurobiol Dis,* 46**,** 393-401.

FERNANDES, K. A., HARDER, J. M., KIM, J. & LIBBY, R. T. 2013. JUN regulates early transcriptional responses to axonal injury in retinal ganglion cells. *Exp Eye Res,* 112**,** 106-17.

FULLER, S. J., EDMUNDS, N. S., MCGUFFIN, L. J., HARDYMAN, M. A., CULL, J. J., ALHARBI, H. O., MEIJLES, D. N., SUGDEN, P. H. & CLERK, A. 2021. MAP4K4 expression in cardiomyocytes: multiple isoforms, multiple phosphorylations and interactions with striatins. *Biochem J,* 478**,** 2121-2143.

GODINHO, L., MUMM, J. S., WILLIAMS, P. R., SCHROETER, E. H., KOERBER, A., PARK, S. W., LEACH, S. D. & WONG, R. O. 2005. Targeting of amacrine cell neurites to appropriate synaptic laminae in the developing zebrafish retina. *Development,* 132**,** 5069-79.

GOUDREAULT, M., D'AMBROSIO, L. M., KEAN, M. J., MULLIN, M. J., LARSEN, B. G., SANCHEZ, A., CHAUDHRY, S., CHEN, G. I., SICHERI, F. & NESVIZHSKII, A. I. 2009. A PP2A phosphatase high density interaction network identifies a novel striatin-interacting phosphatase and kinase complex linked to the cerebral cavernous malformation 3 (CCM3) protein. *Molecular & Cellular Proteomics,* 8**,** 157-171.

GUAN, Q. H., PEI, D. S., ZONG, Y. Y., XU, T. L. & ZHANG, G. Y. 2006. Neuroprotection against ischemic brain injury by a small peptide inhibitor of c-Jun N-terminal kinase (JNK) via nuclear and non-nuclear pathways. *Neuroscience,* 139**,** 609-27.

HALLORAN, M. C., SATO-MAEDA, M., WARREN, J. T., SU, F., LELE, Z., KRONE, P. H., KUWADA, J. Y. & SHOJI, W. 2000. Laser-induced gene expression in specific cells of transgenic zebrafish. *Development,* 127**,** 1953-60.

HAM, J., EILERS, A., WHITFIELD, J., NEAME, S. J. & SHAH, B. 2000. c-Jun and the transcriptional control of neuronal apoptosis. *Biochem Pharmacol,* 60**,** 1015-21.

HAN, R., WANG, R., ZHAO, Q., HAN, Y., ZONG, S., MIAO, S., SONG, W. & WANG, L. 2016. Trim69 regulates zebrafish brain development by ap-1 pathway. *Sci Rep,* 6**,** 24034.

HE, Y., ZHANG, H., YU, L., GUNEL, M., BOGGON, T. J., CHEN, H. & MIN, W. 2010. Stabilization of VEGFR2 signaling by cerebral cavernous malformation 3 is critical for vascular development. *Sci Signal,* 3**,** ra26.

HOLLVILLE, E., ROMERO, S. E. & DESHMUKH, M. 2019. Apoptotic cell death regulation in neurons. *FEBS J,* 286**,** 3276-3298.

HOON, M., OKAWA, H., DELLA SANTINA, L. & WONG, R. O. 2014. Functional architecture of the retina: development and disease. *Progress in retinal and eye research,* 42**,** 44-84.

HU, M. & EASTER, S. S. 1999. Retinal neurogenesis: the formation of the initial central patch of postmitotic cells. *Dev Biol,* 207**,** 309-21.

HUBERMAN, A. D., CLANDININ, T. R. & BAIER, H. 2010. Molecular and cellular mechanisms of lamina-specific axon targeting. *Cold Spring Harb Perspect Biol,* 2**,** a001743.

HULSEN, T., DE VLIEG, J. & ALKEMA, W. 2008. BioVenn–a web application for the comparison and visualization of biological lists using area-proportional Venn diagrams. *BMC genomics,* 9**,** 1-6.

HWANG, J. & PALLAS, D. C. 2014. STRIPAK complexes: structure, biological function, and involvement in human diseases. *The international journal of biochemistry & cell biology,* 47**,** 118-148.

IMAI, F., YOSHIZAWA, A., FUJIMORI-TONOU, N., KAWAKAMI, K. & MASAI, I. 2010. The ubiquitin proteasome system is required for cell proliferation of the lens epithelium and for differentiation of lens fiber cells in zebrafish. *Development,* 137**,** 3257-68.

JEONG, B. C., BAE, S. J., NI, L., ZHANG, X., BAI, X. C. & LUO, X. 2021. Cryo-EM structure of the Hippo signaling integrator human STRIPAK. *Nat Struct Mol Biol,* 28**,** 290-299.

JUSUF, P. R., ALBADRI, S., PAOLINI, A., CURRIE, P. D., ARGENTON, F., HIGASHIJIMA, S., HARRIS, W. A. & POGGI, L. 2012. Biasing amacrine subtypes in the Atoh7 lineage through expression of Barhl2. *J Neurosci,* 32**,** 13929-44.

JUSUF, P. R. & HARRIS, W. A. 2009. Ptf1a is expressed transiently in all types of amacrine cells in the embryonic zebrafish retina. *Neural Dev,* 4**,** 34.

KAY, J. N., FINGER-BAIER, K. C., ROESER, T., STAUB, W. & BAIER, H. 2001. Retinal ganglion cell genesis requires lakritz, a Zebrafish atonal Homolog. *Neuron,* 30**,** 725-36.

KAY, J. N., ROESER, T., MUMM, J. S., GODINHO, L., MREJERU, A., WONG, R. O. & BAIER, H. 2004. Transient requirement for ganglion cells during assembly of retinal synaptic layers. *Development,* 131**,** 1331-42.

KEMP, H. A., CARMANY-RAMPEY, A. & MOENS, C. 2009. Generating chimeric zebrafish embryos by transplantation. *J Vis Exp.*

KIM, D., LANGMEAD, B. & SALZBERG, S. L. 2015. HISAT: a fast spliced aligner with low memory requirements. *Nat Methods,* 12**,** 357-60.

KIM, J. W., BERRIOS, C., KIM, M., SCHADE, A. E., ADELMANT, G., YEERNA, H., DAMATO, E., INIGUEZ, A. B., FLORENS, L., WASHBURN, M. P., STEGMAIER, K., GRAY, N. S., TAMAYO, P., GJOERUP, O., MARTO, J. A., DECAPRIO, J. & HAHN, W. C. 2020. STRIPAK directs PP2A activity toward MAP4K4 to promote oncogenic transformation of human cells. *Elife,* 9.

KITA, E. M., BERTOLESI, G. E., HEHR, C. L., JOHNSTON, J. & MCFARLANE, S. 2013. Neuropilin-1 biases dendrite polarization in the retina. *Development,* 140**,** 2933-41.

KOLDE, R. 2012. Pheatmap: pretty heatmaps. *R package version,* 1**,** 726.

KOSTER, R. W. & FRASER, S. E. 2001. Tracing transgene expression in living zebrafish embryos. *Dev Biol,* 233**,** 329-46.

LA MARCA, J. E., DIEPSTRATEN, S. T., HODGE, A. L., WANG, H., HART, A. H., RICHARDSON, H. E. & SOMERS, W. G. 2019. Strip and Cka negatively regulate JNK signalling during Drosophila spermatogenesis. *Development,* 146.

LI, D., MUSANTE, V., ZHOU, W.-L., PICCIOTTO, M. R. & NAIRN, A. C. 2018. Striatin-1 is a B subunit of protein phosphatase PP2A that regulates dendritic arborization and spine development in striatal neurons. *Journal of Biological Chemistry***,** jbc. RA117. 001519.

LI, L., HUANG, H., FANG, F., LIU, L., SUN, Y. & HU, Y. 2020. Longitudinal Morphological and Functional Assessment of RGC Neurodegeneration After Optic Nerve Crush in Mouse. *Front Cell Neurosci,* 14**,** 109.

LIAO, Y., SMYTH, G. K. & SHI, W. 2014. featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. *Bioinformatics,* 30**,** 923-30.

MACDONALD, R. & WILSON, S. W. 1997. Distribution of Pax6 protein during eye development suggests discrete roles in proliferative and differentiated visual cells. *Dev Genes Evol,* 206**,** 363-369.

MADSEN, C. D., HOOPER, S., TOZLUOGLU, M., BRUCKBAUER, A., FLETCHER, G., ERLER, J. T., BATES, P. A., THOMPSON, B. & SAHAI, E. 2015. STRIPAK components determine mode of cancer cell migration and metastasis. *Nat Cell Biol,* 17**,** 68-80.

MAES, M. E., SCHLAMP, C. L. & NICKELLS, R. W. 2017. BAX to basics: How the BCL2 gene family controls the death of retinal ganglion cells. *Prog Retin Eye Res,* 57**,** 1-25.

MASAI, I., LELE, Z., YAMAGUCHI, M., KOMORI, A., NAKATA, A., NISHIWAKI, Y., WADA, H., TANAKA, H., NOJIMA, Y. & HAMMERSCHMIDT, M. 2003. N-cadherin mediates retinal lamination, maintenance of forebrain compartments and patterning of retinal neurites. *Development,* 130**,** 2479-2494.

MCCURLEY, A. T. & CALLARD, G. V. 2010. Time Course Analysis of Gene Expression Patterns in Zebrafish Eye During Optic Nerve Regeneration. *J Exp Neurosci,* 2010**,** 17-33.

MOSHIRI, A., GONZALEZ, E., TAGAWA, K., MAEDA, H., WANG, M., FRISHMAN, L. J. & WANG, S. W. 2008. Near complete loss of retinal ganglion cells in the math5/brn3b double knockout elicits severe reductions of other cell types during retinal development. *Dev Biol,* 316**,** 214-27.

MUMM, J. S., WILLIAMS, P. R., GODINHO, L., KOERBER, A., PITTMAN, A. J., ROESER, T., CHIEN, C.-B., BAIER, H. & WONG, R. O. 2006. In vivo imaging reveals dendritic targeting of laminated afferents by zebrafish retinal ganglion cells. *Neuron,* 52**,** 609-621.

MUNEMASA, Y. & KITAOKA, Y. 2012. Molecular mechanisms of retinal ganglion cell degeneration in glaucoma and future prospects for cell body and axonal protection. *Front Cell Neurosci,* 6**,** 60.

NASSI, J. J. & CALLAWAY, E. M. 2009. Parallel processing strategies of the primate visual system. *Nat Rev Neurosci,* 10**,** 360-72.

NEAL, S. J., ZHOU, Q. & PIGNONI, F. 2020. STRIPAK-PP2A regulates Hippo-Yorkie signaling to suppress retinal fate in the Drosophila eye disc peripodial epithelium. *J Cell Sci,* 133.

NEISCH, A. L., NEUFELD, T. P. & HAYS, T. S. 2017. A STRIPAK complex mediates axonal transport of autophagosomes and dense core vesicles through PP2A regulation. *J Cell Biol,* 216**,** 441-461.

NISHIWAKI, Y., KOMORI, A., SAGARA, H., SUZUKI, E., MANABE, T., HOSOYA, T., NOJIMA, Y., WADA, H., TANAKA, H., OKAMOTO, H. & MASAI, I. 2008. Mutation of cGMP phosphodiesterase 6alpha'-subunit gene causes progressive degeneration of cone photoreceptors in zebrafish. *Mech Dev,* 125**,** 932-46.

NISHIWAKI, Y. & MASAI, I. 2020. beta-SNAP activity in the outer segment growth period is critical for preventing BNip1-dependent apoptosis in zebrafish photoreceptors. *Sci Rep,* 10**,** 17379.

OKUDA, S., WATANABE, Y., MORIYA, Y., KAWANO, S., YAMAMOTO, T., MATSUMOTO, M., TAKAMI, T., KOBAYASHI, D., ARAKI, N., YOSHIZAWA, A. C., TABATA, T., SUGIYAMA, N., GOTO, S. & ISHIHAMA, Y. 2017. jPOSTrepo: an international standard data repository for proteomes. *Nucleic Acids Res,* 45**,** D1107-d1111.

PETERSON, R. E., FADOOL, J. M., MCCLINTOCK, J. & LINSER, P. J. 2001. Muller cell differentiation in the zebrafish neural retina: evidence of distinct early and late stages in cell maturation. *J Comp Neurol,* 429**,** 530-40.

RANAWAT, N. & MASAI, I. 2021. Mechanisms underlying microglial colonization of developing neural retina in zebrafish. *bioRxiv***,** 2021.05.20.444912.

RANDLETT, O., MACDONALD, R. B., YOSHIMATSU, T., ALMEIDA, A. D., SUZUKI, S. C., WONG, R. O. & HARRIS, W. A. 2013. Cellular requirements for building a retinal neuropil. *Cell Rep,* 3**,** 282-90.

RAPPSILBER, J., MANN, M. & ISHIHAMA, Y. 2007. Protocol for micro-purification, enrichment, pre-fractionation and storage of peptides for proteomics using StageTips. *Nat Protoc,* 2**,** 1896-906.

RAYMOND, P. A., BARTHEL, L. K., BERNARDOS, R. L. & PERKOWSKI, J. J. 2006. Molecular characterization of retinal stem cells and their niches in adult zebrafish. *BMC Dev Biol,* 6**,** 36.

RICK, J. M., HORSCHKE, I. & NEUHAUSS, S. C. 2000. Optokinetic behavior is reversed in achiasmatic mutant zebrafish larvae. *Curr Biol,* 10**,** 595-8.

ROBINSON, M. D., MCCARTHY, D. J. & SMYTH, G. K. 2010. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics,* 26**,** 139-40.

ROBLES, E., LAURELL, E. & BAIER, H. 2014. The retinal projectome reveals brain-area-specific visual representations generated by ganglion cell diversity. *Curr Biol,* 24**,** 2085-2096.

SAKUMA, C., KAWAUCHI, T., HARAGUCHI, S., SHIKANAI, M., YAMAGUCHI, Y., GELFAND, V. I., LUO, L., MIURA, M. & CHIHARA, T. 2014. Drosophila Strip serves as a platform for early endosome organization during axon elongation. *Nat Commun,* 5**,** 5180.

SAKUMA, C., OKUMURA, M., UMEHARA, T., MIURA, M. & CHIHARA, T. 2015. A STRIPAK component Strip regulates neuronal morphogenesis by affecting microtubule stability. *Sci Rep,* 5**,** 17769.

SAKUMA, C., SAITO, Y., UMEHARA, T., KAMIMURA, K., MAEDA, N., MOSCA, T. J., MIURA, M. & CHIHARA, T. 2016. The Strip-Hippo Pathway Regulates Synaptic

Terminal Formation by Modulating Actin Organization at the Drosophila Neuromuscular Synapses. *Cell Rep,* 16**,** 2289-97.

SATIJA, R., FARRELL, J. A., GENNERT, D., SCHIER, A. F. & REGEV, A. 2015. Spatial reconstruction of single-cell gene expression data. *Nature biotechnology,* 33**,** 495-502.

SCHEER, N., RIEDL, I., WARREN, J. T., KUWADA, J. Y. & CAMPOS-ORTEGA, J. A. 2002. A quantitative analysis of the kinetics of Gal4 activator and effector gene expression in the zebrafish. *Mech Dev,* 112**,** 9-14.

SCHNEIDER, C. A., RASBAND, W. S. & ELICEIRI, K. W. 2012. NIH Image to ImageJ: 25 years of image analysis. *Nat Methods,* 9**,** 671-5.

SCHROETER, E. H., WONG, R. O. & GREGG, R. G. 2006. In vivo development of retinal ON-bipolar cell axonal terminals visualized in nyx:: MYFP transgenic zebrafish. *Visual neuroscience,* 23**,** 833-843.

SCHULTE, J., SEPP, K. J., JORQUERA, R. A., WU, C., SONG, Y., HONG, P. & LITTLETON, J. T. 2010. DMob4/Phocein regulates synapse formation, axonal transport, and microtubule organization. *Journal of Neuroscience,* 30**,** 5189-5203.

SEO, G., HAN, H., VARGAS, R. E., YANG, B., LI, X. & WANG, W. 2020. MAP4K Interactome Reveals STRN4 as a Key STRIPAK Complex Component in Hippo Pathway Regulation. *Cell Rep,* 32**,** 107860.

SHI, Z., JIAO, S. & ZHOU, Z. 2016. STRIPAK complexes in cell signaling and cancer. *Oncogene,* 35**,** 4549.

SULLIVAN-BROWN, J., BISHER, M. E. & BURDINE, R. D. 2011. Embedding, serial sectioning and staining of zebrafish embryos using JB-4 resin. *Nat Protoc,* 6**,** 46-55.

SYC-MAZUREK, S. B., FERNANDES, K. A. & LIBBY, R. T. 2017. JUN is important for ocular hypertension-induced retinal ganglion cell degeneration. *Cell Death Dis,* 8**,** e2945.

SZKLARCZYK, D., MORRIS, J. H., COOK, H., KUHN, M., WYDER, S., SIMONOVIC, M., SANTOS, A., DONCHEVA, N. T., ROTH, A., BORK, P., JENSEN, L. J. & VON MERING, C. 2017. The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible. *Nucleic Acids Res,* 45**,** D362-D368.

TANG, Y., CHEN, M., ZHOU, L., MA, J., LI, Y., ZHANG, H., SHI, Z., XU, Q., ZHANG, X., GAO, Z., ZHAO, Y., CHENG, Y., JIAO, S. & ZHOU, Z. 2019. Architecture, substructures, and dynamic assembly of STRIPAK complexes in Hippo signaling. *Cell Discov,* 5**,** 3.

TEAM, R. 2016. RStudio: Integrated development environment for R. Boston, MA.

THISSE, C. & THISSE, B. 2008. High-resolution in situ hybridization to whole-mount zebrafish embryos. *Nat Protoc,* 3**,** 59-69.

UNGOS, J. M., KARLSTROM, R. O. & RAIBLE, D. W. 2003. Hedgehog signaling is directly required for the development of zebrafish dorsal root ganglia neurons. *Development,* 130**,** 5351-62.

URASAKI, A., MORVAN, G. & KAWAKAMI, K. 2006. Functional dissection of the Tol2 transposable element identified the minimal cis-sequence and a highly repetitive sequence in the subterminal region essential for transposition. *Genetics,* 174**,** 639-49.

VELDMAN, M. B., BEMBEN, M. A., THOMPSON, R. C. & GOLDMAN, D. 2007. Gene expression analysis of zebrafish retinal ganglion cells during optic nerve regeneration identifies KLF6a and KLF7a as important regulators of axon regeneration. *Dev Biol,* 312**,** 596-612.

WANG, J., STRUEBING, F. L. & GEISERT, E. E. 2021. Commonalities of optic nerve injury and glaucoma-induced neurodegeneration: Insights from transcriptome-wide studies. *Exp Eye Res*, 207**,** 108571.

WESTERFIELD, M. 1995. *The zebrafish book: a guide for the laboratory use of zebrafish (Brachydanio rerio)*, University of Oregon press.

WHITFIELD, J., NEAME, S. J., PAQUET, L., BERNARD, O. & HAM, J. 2001. Dominant-negative c-Jun promotes neuronal survival by reducing BIM expression and inhibiting mitochondrial cytochrome c release. *Neuron*, 29**,** 629-43.

XU, B., TANG, X., JIN, M., ZHANG, H., DU, L., YU, S. & HE, J. 2020. Unifying developmental programs for embryonic and postembryonic neurogenesis in the zebrafish retina. *Development*, 147.

XU, Q., HOLDER, N., PATIENT, R. & WILSON, S. W. 1994. Spatially regulated expression of three receptor tyrosine kinase genes during gastrulation in the zebrafish. *Development*, 120**,** 287-99.

ZHANG, S., DONG, Y., QIANG, R., ZHANG, Y., ZHANG, X., CHEN, Y., JIANG, P., MA, X., WU, L., AI, J., GAO, X., WANG, P., CHEN, J. & CHAI, R. 2021. Characterization of Strip1 Expression in Mouse Cochlear Hair Cells. *Front Genet*, 12**,** 625867.

ZHAO, X.-F., ELLINGSEN, S. & FJOSE, A. 2009. Labelling and targeted ablation of specific bipolar cell types in the zebrafish retina. *BMC neuroscience*, 10**,** 107.

ZHOU, Y., ZHOU, B., PACHE, L., CHANG, M., KHODABAKHSHI, A. H., TANASEICHUK, O., BENNER, C. & CHANDA, S. K. 2019. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat Commun*, 10**,** 1523.

ZOU, S., TIAN, C., GE, S. & HU, B. 2013. Neurogenesis of retinal ganglion cells is not essential to visual functional recovery after optic nerve injury in adult zebrafish. *PLoS One*, 8**,** e57280.

Figure legends

Figure 1. Strip1 is essential for inner retinal neural circuit development.

(A) Zebrafish retinal neural circuit showing retinal neurons and synaptic layers. GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer; RGCs, retinal ganglion cells; ACs, amacrine cells; BPs, bipolar cells; HCs, horizontal cells; PRs, photoreceptors; MGs, Müller glia.

(B) Morphology of wild-type and *rw147* embryos at 4 dpf. Dotted lines demarcate the eye. An arrowhead indicates abnormal lower jaw. An asterisk indicates heart edema.

(C) Wild-type and *rw147* mutant retinas at 4 dpf. Red and yellow arrowheads indicate the IPL and OPL, respectively.

(D) A missense mutation occurs in *strip1* gene of *rw147* mutants leading to replacement of Leu195 with arginine.

(E) Wild-type retinas labeled with anti-Strip1 antibody (upper panels) and anti-Strip1 plus Strip1 blocking peptide as a negative control (lower panels). Nuclei are stained with Hoechst. Scale, 50 µm. Right panels show higher magnification of outlined areas. Scale, 10 µm.

(F) Whole-mount labeling of 3-dpf wild-type and *strip1*^{rw147} mutant retinas with anti-acetylated α-tubulin antibody. Bottom panels show higher magnification of outlined areas. Scale, 50 µm.

(G) Projection images of single RGCs at 2 dpf expressing *ath5:Gal4VP16; UAS:MYFP* in wild type and *strip1*^{rw147} mutants. Scale, 10 µm.

(H) Projection images of single ACs at 3 dpf expressing *ptf1a:GFP* in wild type and *strip1*^{rw147} mutants. Scale, 10 µm.

(I) Projection images of single BPs at 3 dpf expressing *nyx:Gal4VP16; UAS:MYFP* in wild type and *strip1*^{rw147} mutants. Scale, 10 µm.

Figure 2. RGCs are reduced and INL cells infiltrate the GCL in *strip1* mutants.

(A) Confocal sections of wild-type and *strip1*^{rw14} mutant retinas combined with the transgenic line *Tg[ath5:GFP; ptf1a:mCherry-CAAX]* to label RGCs and ACs. Middle panels represent higher magnification. Lower panels show the magenta channel. INL, inner nuclear layer; GCL, retinal ganglion cell layer. Scale, 50 µm (upper panels) and 10 µm (middle and lower panels).

(B) Percentage of ath5+ area relative to total retinal area. Student's t test with Welch's correction, n≥4.

(C) AC numbers per unified retinal area (8,500 µm^2). Student's t test with Welch's correction, n≥3.

(D) Distribution of ACs (GCL or INL) per unified retinal area (8,500 µm^2). Two-way ANOVA with the Tukey multiple comparison test, n≥3.

(E) Wild-type and *strip1*^{rw14} mutant retinas at 3 dpf labeled with anti-Pax6 which strongly labels ACs. Arrows indicate strong Pax6+ cells that infiltrate the GCL. Nuclei are stained with TOPRO. Scale, 50 µm.

(F) The number of strong Pax6+ cells per retina. Student's t test with Welch's correction, n=5.

(G) Percentage of strong Pax6+ cells (GCL+ or INL+) to the total number of strong Pax6+ cells. Two-way ANOVA with the Tukey multiple comparison test, n=5.

(H) Wild-type and *strip1*^{rw14} mutant retinas at 3 dpf labeled with anti-Prox1. Arrows indicate Prox1+ cells that infiltrate the GCL. Nuclei are stained with TOPRO. Scale, 50 µm.

(I) The number of Prox1+ cells per retina. Student's t test with Welch's correction, n=5.

(J) Percentage of Prox1+ cells (GCL+ or INL+) to the total number of Prox1+. Two-way ANOVA with the Tukey multiple comparison test, n=5.

For all graphs, data are represented as means ± SD. ns: not significant, *p < 0.05, **p < 0.01, and ****p < 0.0001.

Figure 3. Strip1 cell-autonomously promotes RGC survival.

(A) TUNEL of wild-type and *strip1*^{rw147} mutant retinas carrying the transgene *Tg[ath5:GFP]* to label RGCs. Nuclei are stained with Hoechst. Scale, 50 µm.

(B) The number of TUNEL+ cells in GCL. Two-way ANOVA with the Tukey multiple comparison test, n≥3.

(C) Percentage of ath5+ area relative to total retinal area. Two-way ANOVA with the Tukey multiple comparison test, n≥3.

(D) Cell transplantation design to evaluate the cell autonomy of Strip1 in RGC survival. Donor embryos from a *strip1*^{rw147} mutant background are labeled with dextran rhodamine and transplanted into host wild-type embryos. Hosts that show transplanted retinal columns at 60 hpf were subjected to TUNEL.

(E) 60-hpf host retinas stained with TUNEL FL to visualize apoptotic cells in wild type to wild type (upper panel) or *strip1*^{rw147} mutant to wild type (lower panel). Arrows indicate the presence of apoptotic donor cells. Scale, 10 μm.

(F) Percentage of TUNEL+ donor RGCs relative to total donor RGCs. Mann-Whitney U test, n=4.

For all graphs, data are represented as means□±□SD. *p□<□0.05, **p□<□0.01, and ****p□<□0.0001.

Figure 4. RGC death triggers abnormal positioning of ACs leading to IPL disruption.

(A) Time-lapse imaging of wild-type and *strip1*^{rw147} mutant retinas combined with the transgenic line *Tg[ath5:GFP; ptf1a:mCherry-CAAX]* to track ACs and RGCs during IPL formation. Asterisks denote empty areas in the GCL. Arrowheads represent infiltration of ACs into empty spaces in the GCL. Panels on the right show higher magnification of outlined areas. Scale, 50 μm.

(B) Cell transplantation design to evaluate the cell autonomy of Strip1 in AC-mediated IPL formation. Donor embryos are from intercross of *strip1*^{rw147} heterozygous fish combined with *Tg[ptf1a:mCherry-CAAX]* to label ACs. Host embryos are generated by non-transgenic intercross of *strip1*^{rw147} heterozygous fish. Donor cells are labeled with dextran Alexa-488 and transplanted into host embryos to make chimeric host retinas with donor-derived retinal columns.

(C) Confocal images of four combinations of transplantation outcomes: wild type to wild type, wild type to mutant, mutant to wild type and mutant to mutant.

Arrowheads represent abnormal positioning of ACs in basal side of IPL. Scale, 20 μm.

(D) Percentage of ACs (either at the apical or the basal side of the IPL) relative to total number of ACs within a transplanted column. Two-way ANOVA with the Tukey multiple comparison test, n≥4. Data are represented as means±SD. **p<0.01, and ****p<0.0001.

Figure 5. Strip1 interacts with Strn3 to regulate RGC survival.

(A) Design of co-immunoprecipitation coupled with mass spectrometry (Co-IP/MS) to identify zebrafish Strip1 interacting partners. Embryos carrying the transgenes *Tg[hsp:WT.Strip1-GFP], Tg[hsp:Mut.Strip1-GFP] or Tg[hsp:Gal4;UAS:GFP]* were used to pull down wild-type GFP-tagged Strip1, mutant GFP-tagged Strip1 or only GFP, respectively. Head lysates from 2-dpf zebrafish embryos were subjected to immunoprecipitation using GFP-Trap beads. Immunoprecipitates were digested and analyzed by mass spectrometry.

(B) Western blotting of whole head lysates (input) and immunoprecipitates (IP) using anti-GFP antibody. Red and black arrowheads indicate the expected band sizes for Strip1-GFP (120 kDa) and GFP (26 kDa), respectively.

(C) Venn Diagram comparing proteins significantly enriched in WT.Strip1-GFP relative to Control GFP (blue) and WT.Strip1-GFP relative to Mut.Strip1.GFP (magenta). Six proteins are commonly enriched in both groups, FC>2, p<0.05. n=3 for WT.Strip1-GFP and Mut. Strip1-GFP and n=2 for GFP-control.

(D) Five components of the STRIPAK complex found from six proteins commonly enriched in (C).

(E) TUNEL of 60-hpf retinas of *Tg[ath5:GFP]* transgenic embryos injected with standard MO and MO-Strn3. RGCs and apoptotic cells are labeled with *ath5:GFP* and TUNEL, respectively. Nuclei are stained with Hoechst (blue).

(F) The number of TUNEL+ cells in GCL. Mann-Whitney U test, n≥6.

(G) Percentage of ath5+ area relative to total retinal area. Student's t test with Welch's correction, n≥3.

(H) Confocal images of retinas of 76-hpf *Tg[ath5:GFP; ptf1a:mCherry-CAAX]* transgenic embryos injected with standard MO and MO-Strn3. *ath5:GFP* and *ptf1a:mCherry-CAAX]* label RGCs and ACs, respectively.

(I) Percentage of ath5+ area relative to total retinal area. Student's t test with Welch's correction, n=8.

Scale, 50 μm (E, H).

For all graphs, data are represented as means□±□SD. **p□<□0.01, ***p□<□0.001 and ****p□<□0.0001.

Figure 6. Jun is a key mediator of RGC death in the absence of Strip1.

(A) Volcano plot showing differentially expressed genes (DEGs) in *strip1*^{rw147} mutants compared to wild-type siblings. Colored points represent genes that are significantly upregulated (131 genes, red) or downregulated (75 genes, blue). Data are obtained from four independent collections of 62-hpf embryo eye cups. FDR<0.05, $Log_2FC > |1|$.

(B) Heatmap of expression values (z-score) representing selected DEGs in *strip1*^{rw147} mutants compared to wild-type siblings.

(C) Whole-mount labeling of 54-hpf wild-type and *strip1*^{rw147} mutant retinas with anti-phospho-Jun antibody and zn5 antibody, which label active Jun and RGCs, respectively.

(D) Percentage of phospho-Jun area relative to zn5 area at 54-58 hpf. Student's t test with Welch's correction, n=6.

(E) Whole-mount labeling of 49-hpf wild-type embryos injected with standard MO or MO-strn3 with anti-phospho-Jun antibody and zn5 antibody, respectively.

(F) Percentage of phospho-Jun area relative to zn5 area at 49 hpf. Student's t test with Welch's correction, n=5.

(G) TUNEL and zn5 antibody labeling of 60-hpf wild-type and *strip1*^{rw147} mutant retinas injected with standard MO and MO-Jun. Nuclei are stained with Hoechst.

(H) The number of TUNEL+ cells in GCL per retina. Two-way ANOVA with the Tukey multiple comparison test, n=6.

(I) Confocal images of 76-hpf wild-type and *strip1*^{rw14} mutant retinas injected with standard-MO and MO-Jun. Embryos carry the transgene *Tg[ath5:GFP]* to label RGCs and are stained with bodipy TR methyl ester to visualize retinal layers.

(J) Percentage of ath5+ area relative to total retinal area, Two-way ANOVA with the Tukey multiple comparison test, n≥3.

Scale, 50 μm (C, E, G, I).

For all graphs, data are represented as means□±□SD. ns: not significant, *p□<□0.05, **p□<□0.01 and ****p□<□0.0001.

Figure 7. Bcl2 rescues RGC death in *strip1* mutants, but surviving RGCs do not project their dendrites to the IPL.

(A) 60-hpf wild-type and *strip1*^{rw147} mutants combined with the transgenic line *Tg[hsp:mCherry-Bcl2]*. Non-transgenic embryos (Bcl2-, top panels) are compared to transgenic embryos (Bcl2+, bottom panels) after heat-shock treatment. Apoptotic cells are visualized by TUNEL FL and fluorescent signals from mCherry-Bcl2 are shown. Nuclei are stained with Hoechst.

(B) The number of TUNEL+ cells in GCL. Two-way ANOVA with the Tukey multiple comparison test, n=3.

(C) 78-hpf wild-type and *strip1*^{rw147} mutant retinas combined with the transgenic lines, *Tg[ath5:GFP]* and *Tg[hsp:mCherry-Bcl2]*. Non-transgenic embryos (Bcl2-, top panels) are compared to transgenic embryos (Bcl2+, bottom panels) after heat-shock treatment. RGCs are labeled with *ath5:GFP* and fluorescent signals from mCherry-Bcl2 are shown. Anti-acetylated α-tubulin labels the IPL. Nuclei are stained with Hoechst.

(D) Percentage of ath5+ area relative to retinal area. Two-way ANOVA with the Tukey multiple comparison test, n≥3.

Scale: 50 μm (A, C).

For all graphs, data are represented as means□±□SD. ns: not significant, *p□<□0.05, **p□<□0.01, ***p□<□0.001 and ****p□<□0.0001.

Figure 8. Summary of developmental and molecular events that underlie Strip1 function in inner retinal circuit formation.

(A) In wild-type retina, Strip1 suppresses Jun-mediated pro-apoptotic signals, probably through the STIPAK complex, to maintain RGCs during development. In absence of Strip1, Jun is activated within RGCs leading to severe degeneration of RGCs as early as 2 dpf. Subsequently, cells in the INL abnormally infiltrate the GCL leading to a disrupted IPL.

(B) Proposed model for Strip1 role within RGCs to regulate AC positioning and IPL formation. In wild type, Strip1 regulates (1) RGC survival to prevent AC infiltration, and (2) RGC dendritic patterning to promote RGC-AC interactions. In *strip1*^{rw147} mutants, both mechanisms are perturbed, leading to AC infiltration, increased AC-AC interactions, and IPL defects. In Bcl2-rescued *strip1*^{rw147} mutants, survived RGCs prevent AC infiltration. However, RGC dendritic defects lead to increased AC-AC interactions and ectopic IPL formation.

Supplementary materials legends

Figure 1–figure supplement 1. Retinal lamination defects in *strip1*^{rw147} mutants are phenocopied by either CRISPR- or MO-mediated knockdown of *strip1* and rescued by overexpression of Strip1.

(A) CRISPR-mediated, targeted mutagenesis of zebrafish *strip1* gene. A 10-bp deletion was introduced into exon 9, leading to a premature stop codon in *strip1* gene.

(B) Morphology of wild-type and *strip1*$^{crispr\Delta 10}$ embryos at 4 dpf. Dotted lines demarcate the eye outline. The arrowhead indicates an abnormal lower jaw. An asterisk indicates heart edema.

(C) Plastic sections of wild-type and *strip1*$^{crispr\Delta 10}$ mutant retinas stained with toluidine blue at 4 dpf.

(D) A polyacrylamide gel image showing PCR amplicons using primers that flank the 10-base deletion of *strip1*$^{crispr\Delta 10}$. Wild-type and *strip1*$^{crispr\Delta 10}$ mutant bands are expected to appear at 279 and 269 bps, respectively. Different band sizes were consistently obtained from progeny produced by pair-wise cross of *strip1*$^{crispr\Delta 10}$ heterozygous mutant fish.

(E) Morphology of non-injected and MO-strip1 injected wild-type embryos at 4 dpf. Dotted lines outline the eye. An arrowhead indicates the abnormal lower jaw. An asterisk indicates heart edema.

(F) Retinal plastic sections of non-injected and MO-strip1 injected wild-type embryos at 4 dpf.

(G) Western blotting of 2-dpf head lysates from non-injected and MO-strip1 injected wild-type embryos. Red and black arrows show expected band sizes for zebrafish Strip1 (93 kDa) and the loading control b-actin (42 kDa), respectively.

(H) Plastic sections of 4 dpf wild-type and *strip1*^{rw147} mutant retinas without transgene (top panels), and with the transgenes *Tg[hsp:WT.Strip1-GFP]* (middle panels) or *Tg[hsp:Mut.Strip1-GFP]* (bottom panels) post heat shock treatment. Over expression of GFP-tagged wild-type Strip1 rescues retinal defects in *strip1*^{rw147} mutants, while the GFP-tagged mutant form fails to rescue.

(I) A staggered bar graph, which represents the percentage of *strip1*^{rw147} mutants with a defective retina vs. mutants with a rescued retina in *strip1*^{rw147} mutants without transgenes, and with *Tg[hsp:WT.Strip1-GFP]* or *Tg [hsp:Mut.Strip1-GFP].*

Figure 1–figure supplement 2. *strip1* mRNA is expressed in developing retinas and is required for IPL formation.

(A) Expression patterns of *strip1* mRNA during embryonic development. *In situ* hybridization using a *strip1* antisense probe is shown at different developmental stages: 4-cell, blastula, 75% epiboly, 1 dpf, 2 dpf and 3 dpf. *In situ* hybridization using the sense probe at 1 dpf is shown as a negative control. *strip1* mRNA is ubiquitously expressed until gastrula stage. At 1-2 dpf, expression becomes restricted in the heart, optic tectum, and eyes. Ventral view of 2-dpf retina shows a stronger signal in GCL (left panels). OT, optic tectum; GCL, ganglion cell layer. Scale bar: 100 μm.

(B) Frontal cryosections of 2-dpf heads following *in situ* hybridization show that *strip1* mRNA is predominantly expressed in the inner retina and the optic tectum.

(C) Retinal cryosections show higher expression in RGCs and ACs. Right panels show higher magnification of outlined areas. RGCs, retinal ganglion cells; ACs, amacrine cells; HCs, horizontal cells; PRs, photoreceptors.

(D) Live confocal snapshots of the same wild-type and *strip1*^{rw147} mutant retinas at different developmental stages; 40, 52, 62 and 76 hpf stained with Bodipy TR. A primitive IPL is first observed in wild-type retinas as early as 52 hpf. However, it is not as prominent in *strip1*^{rw147} mutant retinas. By 62 hpf, *strip1*^{rw147} mutants display severe defects in IPL formation. Arrowheads indicate rudimentary IPL. Asterisks show IPL defects in *strip1*^{rw147} mutants. Scale bar: 50 µm.

Figure 2–figure supplement 1. ACs are abnormally positioned at the basal side of the IPL in *strip1* mutants and *ath5* morphants.

(A) Live confocal images of wild-type and *strip1*^{rw147} mutant retinas at 3 dpf combined with the transgenic line *Tg[ptf1a:mCherry-CAAX]*, which labels ACs. Middle and lower panels show higher magnification of outlined areas. Abnormal positioning of ACs in the GCL is more prominent in areas of strong IPL defects than in areas of weak lamination defects. Scale bar, 50 µm.

(B) Live confocal images of retinas of STD-MO and MO-ath5 injected wild-type embryos combined with the transgenic line *Tg[ath5:GFP; ptf1a:mCherry-CAAX]*, which labels RGCs and ACs. Middle and bottom panels show higher magnification images to show abnormal localization of ACs in GCL (arrowheads) and disrupted IPL. Scale bar, 50 µm (upper panels) and 20 µm (middle and lower panels).

(C-F) Labelling of 4-dpf wild-type and *strip1*^{rw147} mutant retinas with zpr3 (C), zpr1 (D), anti-glutamine synthetase (GS) (E) and anti-PCNA (F) antibodies, which visualize rod photoreceptors, double cone photoreceptors, Müller glia and proliferative cells in the ciliary marginal zone (CMZ), respectively. Both rods and cones are in ONL and are mostly intact in *strip1*^{rw147} mutants. Müller glia genesis and retinal stem cells and progenitor cells in the CMZ appear to be unaffected in *strip1*^{rw147} mutants. Scale bar, 50 µm.

Figure 3–figure supplement 1. *strip1* mutants show apoptosis in RGCs and optic tectum and elongation defects in retinal axons.

(A) The number of TUNEL+ in the GCL compared to the rest of the retina at 36, 48, 60, 72 and 96 hpf in *strip1*^{rw147} mutant retinas, for experiments shown in Figure 3A. Apoptosis gradually increases to reach its peak at 72 hpf in other retinal areas, but its number at 72 hpf is significantly lower than that of the GCL at 72 hpf. In addition, there is no significant difference in apoptotic cell number of other retinal areas between 72 hpf and other time points (36, 60, 96 hpf). Two-way ANOVA with the Tukey multiple comparison test, n≥3.

(B) Percentage of GCL area (area between the lens and IPL) relative to the total retinal area in wild-type and *strip1*^{rw147} mutant retinas at 36, 48, 60, 72 and 96 hpf, for experiments shown in Figure 3A. There is no significant difference between wild-type siblings and *strip1*^{rw147} mutants at any stage. Two-way ANOVA with the Tukey multiple comparison test, n≥3.

(C) Live confocal images of wild-type and *strip1*$^{crispr\Delta 10}$ mutant retinas stained with acridine orange (AO) to label apoptotic cells at 60 hpf. Scale bar, 50 µm.

(D) Percentage of AO area relative to total retina area in 60 hpf wild-type and *strip1*$^{crispr\Delta 10}$ mutant retinas. Mann Whitney U test, n=5.

(E) TUNEL of 60 hpf wild-type and *strip1*^{rw147} mutant heads combined with *Tg[ath5:GFP]* to label RGCs. All nuclei are counterstained with Hoechst. Apoptosis occurs markedly in RGCs and optic tectum in *strip1*^{rw147} mutants. Scale bar, 100 µm.

(F) The number of TUNEL+ in the optic tectum of wild-type and *strip1*^{rw147} mutants at 60 hpf. The number of TUNEL+ cells is significantly higher in *strip1*^{rw147} mutants than in wild-type siblings. Student's t test with Welch's correction, n=3.

(G) Dorsal view of the optic tectum of 3-dpf wild-type and *strip1*^{rw147} mutants. RGC axons are labeled using intraretinal injections of DiO or DiI. Axons of *strip1*^{rw147} appear to exit the optic disc normally and the optic chiasm is formed. However, the optic nerve is thinner in *strip1*^{rw147} mutants than in wild-type siblings. In addition, most axons fail to elongate properly and do not arborize within the optic tectum

(arrowheads) or very few axons elongate poorly to reach more posterior arborization fields within the optic tectum (asterisk). Scale bar: 50 µm.

For all graphs, data are represented as means ± SD. ns: not significant, *p < 0.05, and ***p < 0.001.

Figure 4–figure supplement 1. Strip1 is not required in ACs and BPs for their neurite projections to the IPL.

(A) 3D confocal live images showing neurite projection of ACs to the IPL in transplantation assays outlined in Figure 4B-C. Panels represent three combinations of transplantation experiments: from wild-type donor to wild-type host (left columns), from *strip1*^{rw147} mutant donor to wild-type host (middle columns), and from wild-type donor to *strip1*^{rw147} mutant host (right columns). Donor cells are labeled with dextran-Alexa-488 in green and ACs are labeled with *ptf1a:mCherry-CAAX* in magenta. Mutant donor ACs show a normal dendritic pattern and normal projection to the IPL in wild-type host retina, although on rare occasion we observed that mutant ACs display two dendritic trees (arrow in the bottom middle panel), but still contribute to a normal IPL. On the other hand, wild-type donor ACs transplanted into mutant host retinas show abnormal projection patterns and infiltrate the GCL (arrowheads in the right panels). Scale bar, 20 µm.

(B) Confocal image of 3-dpf live wild-type and *strip1*^{rw14} mutant retinas combined with the transgenic line, *xfz43*, to label subsets of BPs. Scale bar, 10 µm.

(C) Schematic drawings of cell transplantation design to evaluate cell autonomy of Strip1's role in BP development. *strip1*^{rw147} mutant donor cells carrying the transgene *xfz43* (green) are labeled with dextran-cascade blue or dextran-Alexa-flour 647 (blue) and transplanted into wild-type host embryos at blastula stage, leading to chimeric wild-type retina with *strip1*^{rw147} mutant retinal columns. Retinal lamination of the host retina is visualized with Bodipy TR (magenta).

(D) 3D confocal live images of four wild-type host retinas with *strip1*^{rw14} mutant donor retinal columns outlined in (C). *strip1*^{rw14} mutant donor BPs show normal neurite projection patterns to the IPL in wild-type host retina. Occasionally, bipolar axon terminals are abnormally extended laterally (arrow in the bottom panel); however, such defects are confined to a seemingly normal IPL. Scale bar, 10 µm.

(E) Confocal image of 3-dpf live wild-type and *strip1*^{rw14} mutant retinas combined with the transgenic line, *xfz3*, to label subsets of BPs. Scale bar, 10 μm.

(F) Schematic drawings of cell transplantation design to evaluate the cell autonomy of Strip1 role in BP development. Wild-type donor cells carrying the transgene *xfz3* (green) are labeled with dextran-rhodamine (magenta) and transplanted into *strip1*^{rw14} mutant host embryos at blastula stage, leading to chimeric *strip1*^{rw14} mutant retinas with wild-type retinal columns.

(G) 3D confocal live images of two different *strip1*^{rw147} mutant host retinas with wild-type donor retinal columns outlined in (F). Wild-type donor BPs show misguided neurite projections and excessive branching. Scale bar, 10 μm.

Figure 5–figure supplement 1. Components of the STRIPAK complex are highly enriched in the interactome of zebrafish Strip1 and their retinal expression, according to published scRNA-seq data.

(A) Schematic diagram showing the core components of the mammalian STRIPAK complex and some of its accessory molecules. Core STRIPAK components include serine/threonine-protein phosphatase catalytic subunit known as PP2Ac and the scaffolding subunit PP2Aa, together with striatins (STRN, STRN3 and STRN4), Mob3, STRIP1 or STRIP2, and a germinal center kinase GCKIII kinase bound via cerebral cavernous malformation 3 (Ccm3). Some molecules bind in a mutually exclusive pattern to the core components. Arrows show that sarcolemmal membrane-associated protein (SLMAP) and the suppressor of IKKe (SIKE) are not detected in STRIPAK complexes containing cortactin binding proteins (CTTNBP2/NL) and vice versa. (Figure adapted from Hwang J. and Pallas D.C., 2014).

(B) STRING protein-protein interaction (PPI) network analysis of six zebrafish Strip1 interacting partners significantly enriched from IP-MS analysis highlighted in Figure 5C. Network edges represent known and/or predicted functional interactions in the STRING database. Edge thickness reflects the combined STRING evidence score for each binary relationship. Thicker edges represent increased interaction evidence. PPI enrichment p-value <1.0e-16.

(C) UMAP plots showing different clusters of retinal cells at 48 hpf analyzed with Seuret R pipeline using data published by Xu B. et al, 2020. Twelve clusters are identified and categorized to different retinal cell types or sub-types. RPCs, retinal progenitor cells; RGCs, retinal ganglion cells; ACs, amacrine cells; BPs, bipolar cells; HCs, horizontal cells; PRs, photoreceptors; MGs, Müller glia.

(D-I) UMAP plots showing retinal mRNA expression patterns of *strip1* (D) and its interacting STRIPAK components, identified from Co-IP/MS; *cttnbp2* (E), *cttnbp2nlb* (F), *strn* (G), *strn3* (H) and *strn4* (I), Only *strip1* and *strn3* show high retinal expression levels at 48 hpf compared to other partners.

Figure 6–figure supplement 1. Gene ontology (GO) enrichment analysis of differentially expressed genes (DEGs) from RNA-seq of *strip1* mutants and comparison between upregulated DEGs in *strip1* mutant transcriptomes vs. transcriptomes of zebrafish RGCs under stress.

(A-B) List of enriched GO terms in DEGs downregulated (A) and upregulated (B) in *strip1*^{rw147} mutant relative to wild-type siblings as analyzed with Metascape, FDR<0.05 and Log_2FC > |1|. Downregulated genes show many GO terms related to the nervous system, RGCs, embryo morphogenesis, synaptic development, and transmission. Upregulated genes show terms related to stress response, cell death and MAPK signaling.

(C) Venn diagram showing overlap of upregulated DEGs in RNA sequencing datasets of *strip1*^{rw147} mutant relative to wild-type (FDR<0.05 and FC >1.5) in comparison with published upregulated DEGs in microarray data of adult zebrafish RGCs at 3 days post-optic nerve injury (ONI) (Veldman et al., 2007) and adult zebrafish eyes at 4 days-post optic nerve crush (ONX) (McCurley and Callard, 2010).

Legends of video files

Figure 4–video 1.

Development of ACs and IPL formation in wild-type sibling retina.

Time-lapse movie to track the development of ACs and RGCs in wild-type sibling combined with the transgenic line *Tg[ath5:GFP; ptf1a:mCherry-CAAX]* to label RGCs in green and ACs in magenta. Acquisition started ~48hpf. Movie on the right represents outlined areas for *ptf1a:mCherry-CAAX* channel.

Figure 4–video 2

Development of ACs and IPL formation in *strip1^{rw147}* mutant retina.

Time-lapse movie to track the development of ACs and RGCs in a *strip1^{rw147}* mutant combined with the transgenic line *Tg[ath5:GFP; ptf1a:mCherry-CAAX]* to label RGCs in green and ACs in magenta. Acquisition started ~48 hpf. The movie on the right represents outlined areas for the *ptf1a:mCherry-CAAX* channel. Asterisks depict the onset of severe RGC loss and arrows show areas of ectopic migration of ACs into the GCL.

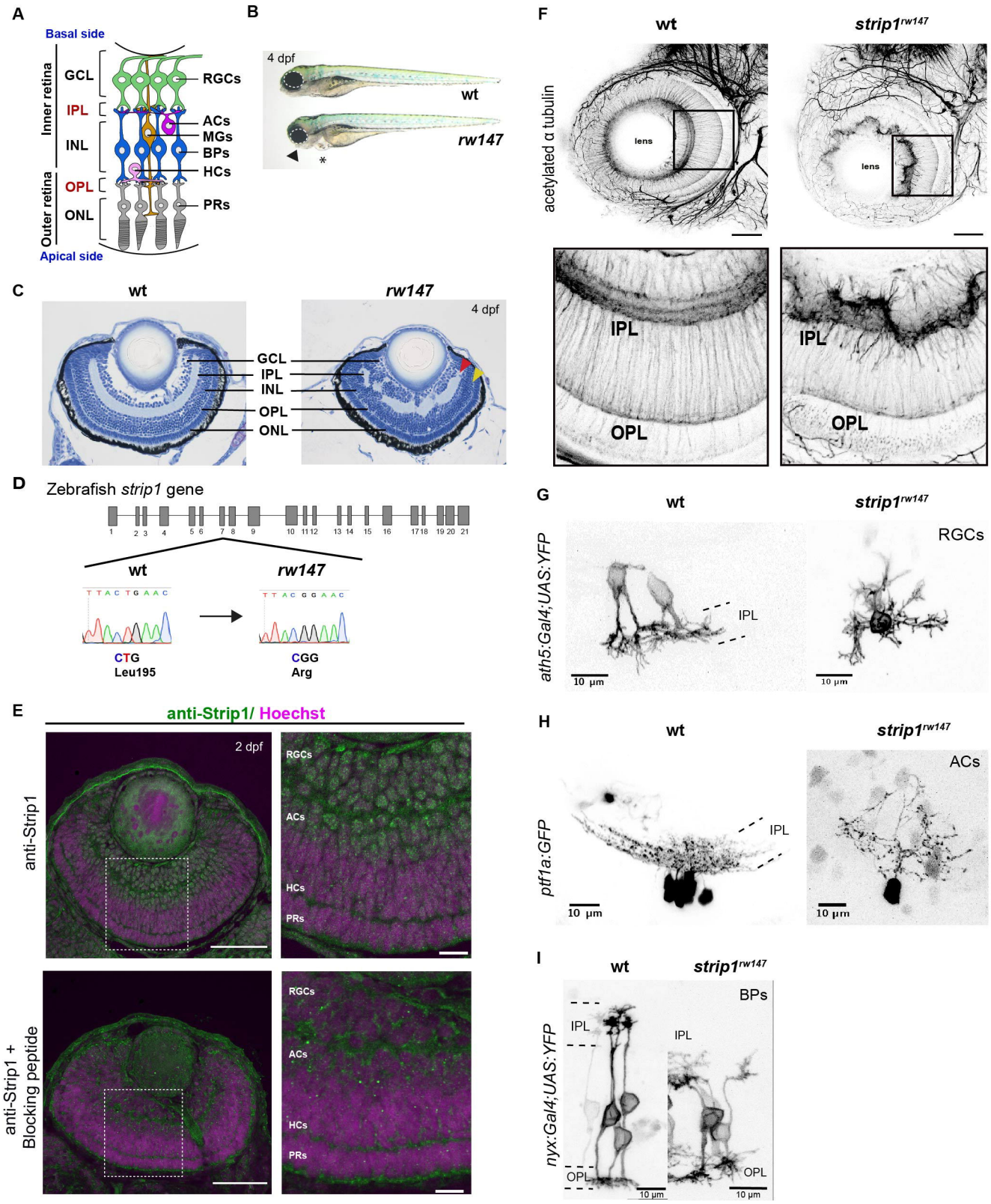

A
Basal side
Inner retina
GCL
IPL
INL
Outer retina
OPL
ONL
Apical side
RGCs
ACs
MGs
BPs
HCs
PRs
B
4 dpf
wt
rw147
C
wt
rw147
4 dpf
GCL
IPL
INL
OPL
ONL
D
Zebrafish strip1 gene
wt
rw147
CTG
Leu195
CGG
Arg
E
anti-Strip1/ Hoechst
2 dpf
anti-Strip1
anti-Strip1 +
Blocking peptide
RGCs
ACs
HCs
PRs
F
wt
strip1rw147
acetylated α tubulin
lens
IPL
OPL
G
wt
strip1rw147
ath5:Gal4;UAS:YFP
RGCs
IPL
10 µm
H
wt
strip1rw147
ptf1a:GFP
ACs
IPL
10 µm
I
wt
strip1rw147
nyx:Gal4;UAS:YFP
BPs
IPL
OPL
10 µm

Figure 1.

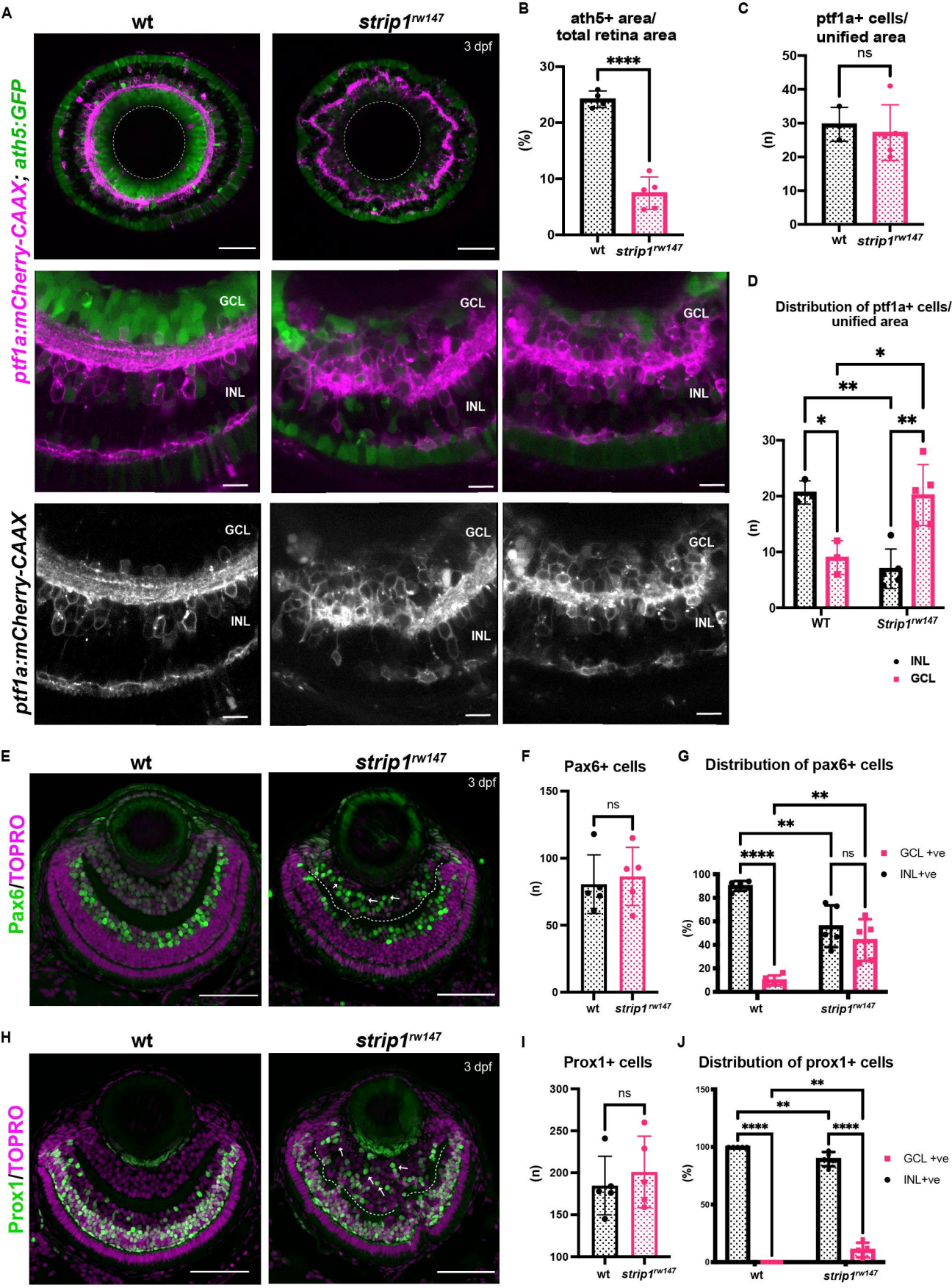

A
wt
strip1rw147
3 dpf
ptf1a:mCherry-CAAX; ath5:GFP
GCL
INL
ptf1a:mCherry-CAAX
B
ath5+ area/
total retina area
****
(%)
wt
strip1rw147
C
ptf1a+ cells/
unified area
ns
(n)
D
Distribution of ptf1a+ cells/
unified area
*
**
(n)
WT
Strip1rw147
INL
GCL
E
Pax6/TOPRO
F
Pax6+ cells
G
Distribution of pax6+ cells
GCL +ve
INL+ve
H
Prox1/TOPRO
I
Prox1+ cells
J
Distribution of prox1+ cells

Figure 2.

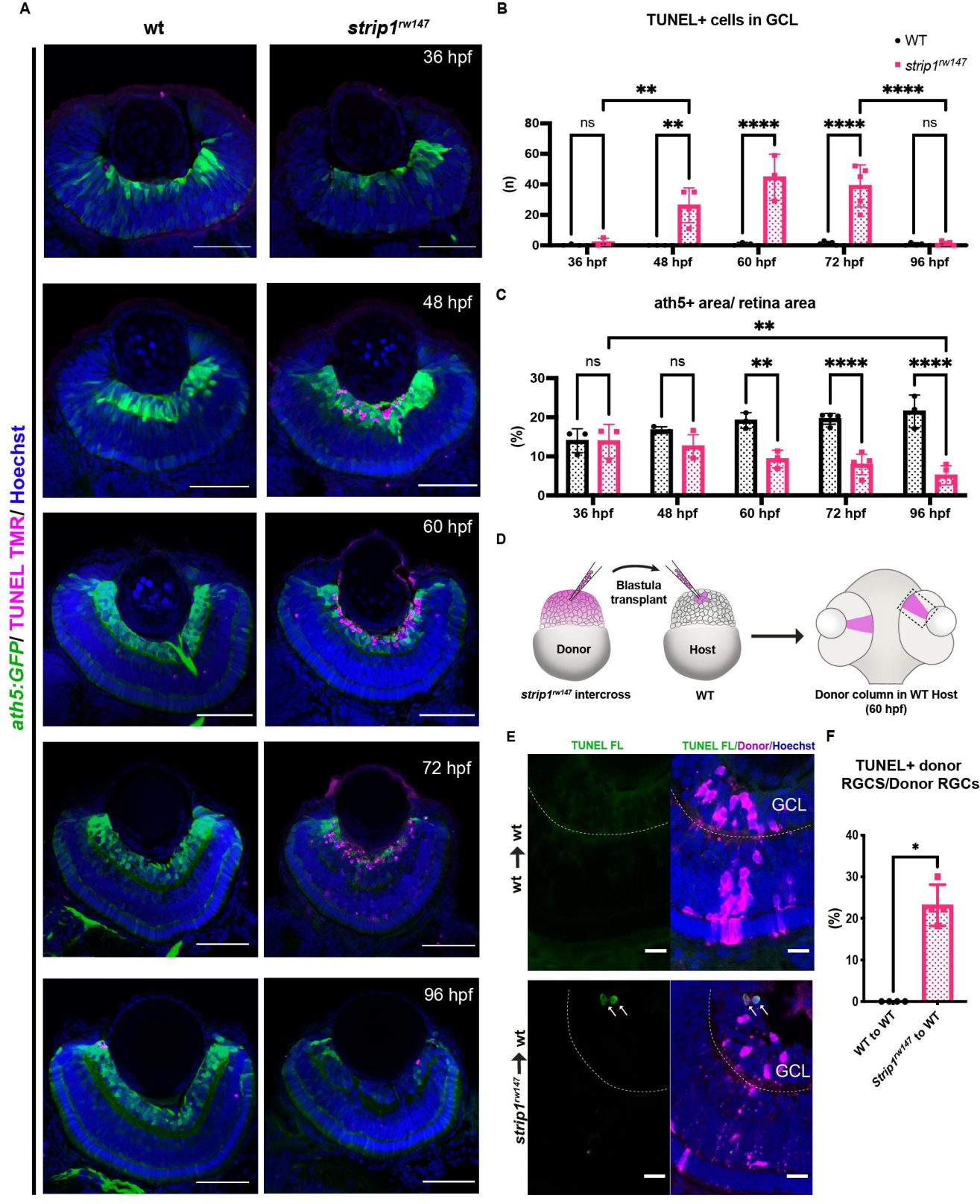

A
wt
strip1rw147
36 hpf
48 hpf
60 hpf
72 hpf
96 hpf
ath5:GFP/ TUNEL TMR/ Hoechst
B
TUNEL+ cells in GCL
WT
strip1rw147
(n)
ns
**
****
36 hpf
48 hpf
60 hpf
72 hpf
96 hpf
C
ath5+ area/ retina area
(%)
D
Blastula transplant
Donor
Host
strip1rw147 intercross
WT
Donor column in WT Host (60 hpf)
E
TUNEL FL
TUNEL FL/Donor/Hoechst
GCL
wt → wt
strip1rw147 → wt
F
TUNEL+ donor RGCS/Donor RGCs
*
WT to WT
Strip1rw147 to WT

Figure 3.

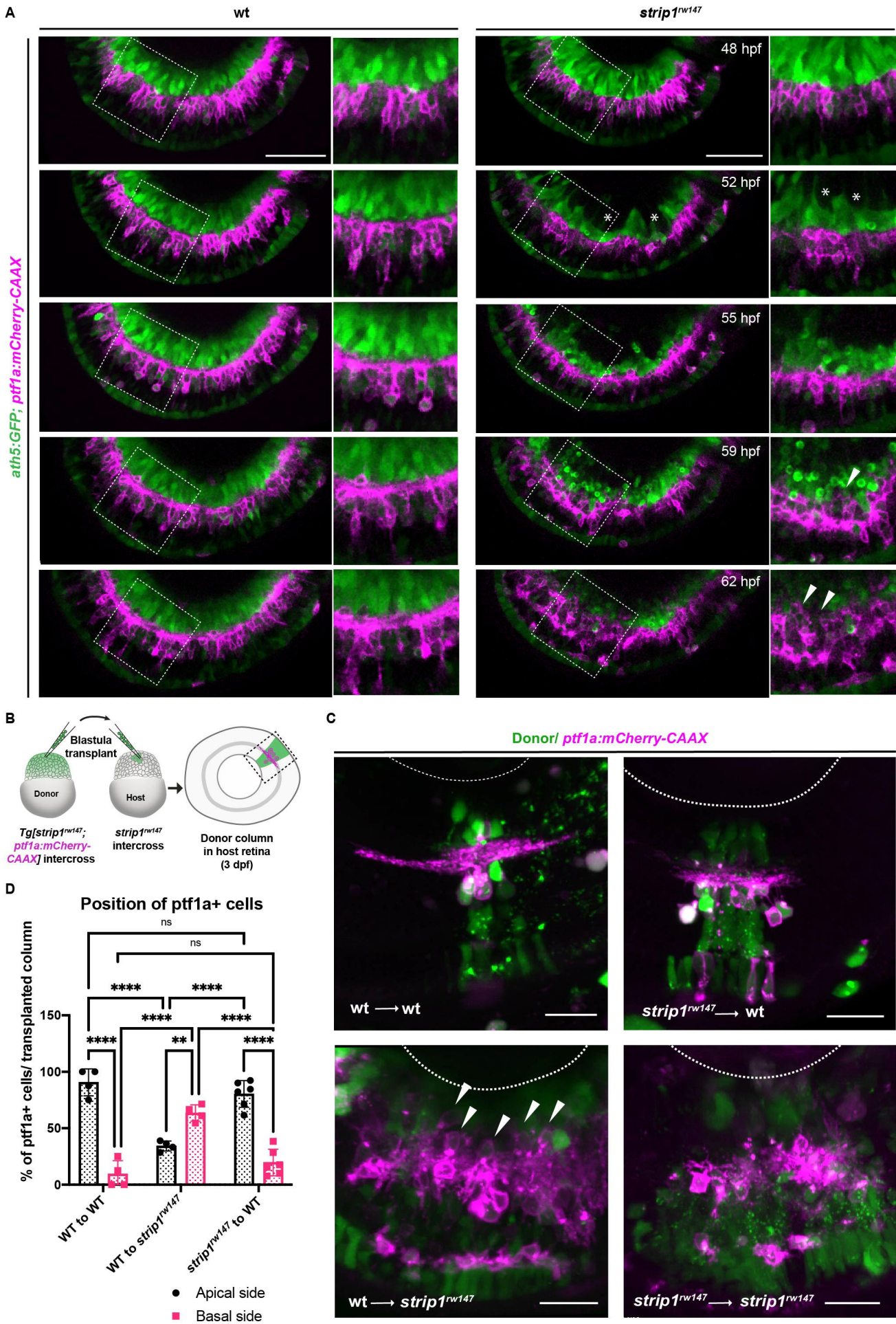
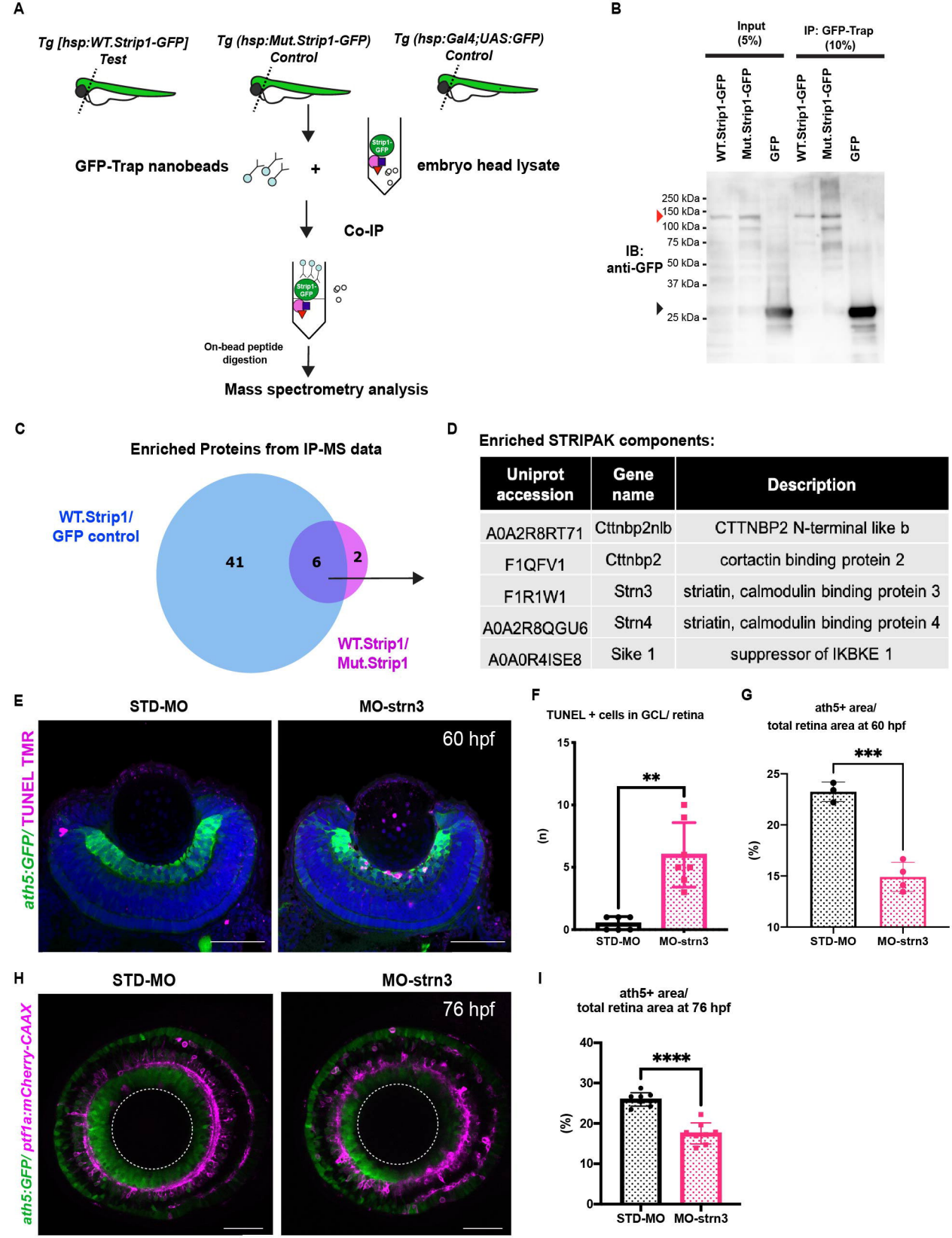

A
wt
strip1rw147
ath5:GFP; ptf1a:mCherry-CAAX
48 hpf
52 hpf
55 hpf
59 hpf
62 hpf
B
Blastula transplant
Donor
Host
Tg[strip1rw147; ptf1a:mCherry-CAAX] intercross
strip1rw147 intercross
Donor column in host retina (3 dpf)
C
Donor/ ptf1a:mCherry-CAAX
wt ⟶ wt
strip1rw147 ⟶ wt
wt ⟶ strip1rw147
strip1rw147 ⟶ strip1rw147
D
Position of ptf1a+ cells
% of ptf1a+ cells/ transplanted column
ns
ns
****
****
****
****
****
**
****
150
100
50
0
WT to WT
WT to strip1rw147
strip1rw147 to WT
Apical side
Basal side

Figure 4.



Uniprot accession	Gene name	Description
A0A2R8RT71	Cttnbp2nlb	CTTNBP2 N-terminal like b
F1QFV1	Cttnbp2	cortactin binding protein 2
F1R1W1	Strn3	striatin, calmodulin binding protein 3
A0A2R8QGU6	Strn4	striatin, calmodulin binding protein 4
A0A0R4ISE8	Sike 1	suppressor of IKBKE 1

Figure 5.

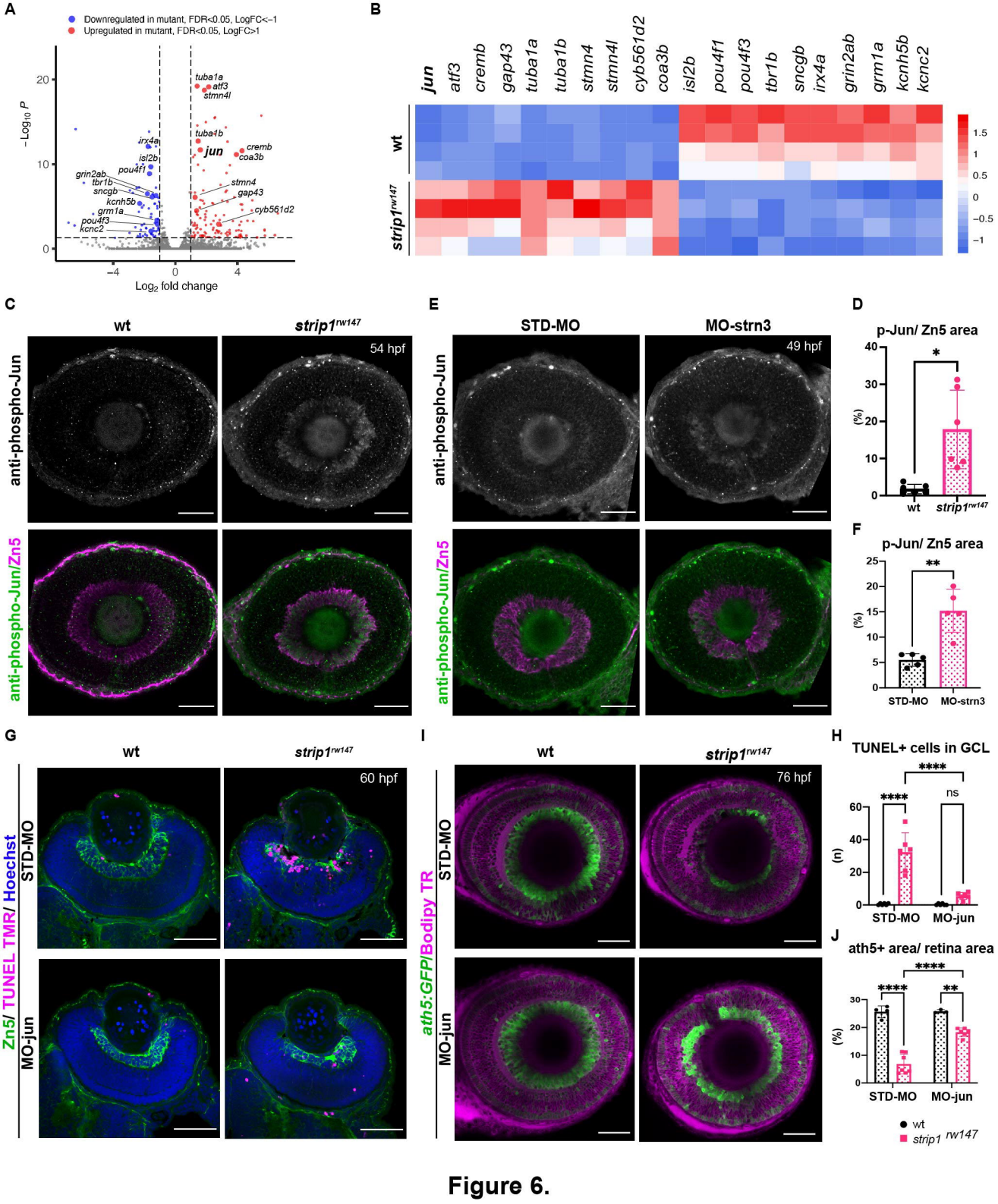

A
Downregulated in mutant, FDR<0.05, LogFC<-1
Upregulated in mutant, FDR<0.05, LogFC>1
tuba1a
atf3
stmn4l
tuba1b
jun
cremb
coa3b
irx4a
isl2b
pou4f1
grin2ab
tbr1b
sncgb
kcnh5b
grm1a
pou4f3
kcnc2
stmn4
gap43
cyb561d2
Log2 fold change
B
jun
atf3
cremb
gap43
tuba1a
tuba1b
stmn4
stmn4l
cyb561d2
coa3b
isl2b
pou4f1
pou4f3
tbr1b
sncgb
irx4a
grin2ab
grm1a
kcnh5b
kcnc2
wt
strip1rw147
C
wt
strip1rw147
54 hpf
anti-phospho-Jun
anti-phospho-Jun/Zn5
E
STD-MO
MO-strn3
49 hpf
D
p-Jun/ Zn5 area
(%)
wt
strip1rw147
F
p-Jun/ Zn5 area
STD-MO
MO-strn3
G
wt
strip1rw147
60 hpf
Zn5/ TUNEL TMR/ Hoechst
STD-MO
MO-jun
I
wt
strip1rw147
76 hpf
ath5:GFP/Bodipy TR
STD-MO
MO-jun
H
TUNEL+ cells in GCL
(n)
ns
STD-MO
MO-jun
J
ath5+ area/ retina area
(%)
STD-MO
MO-jun
wt
strip1 rw147

Figure 6.

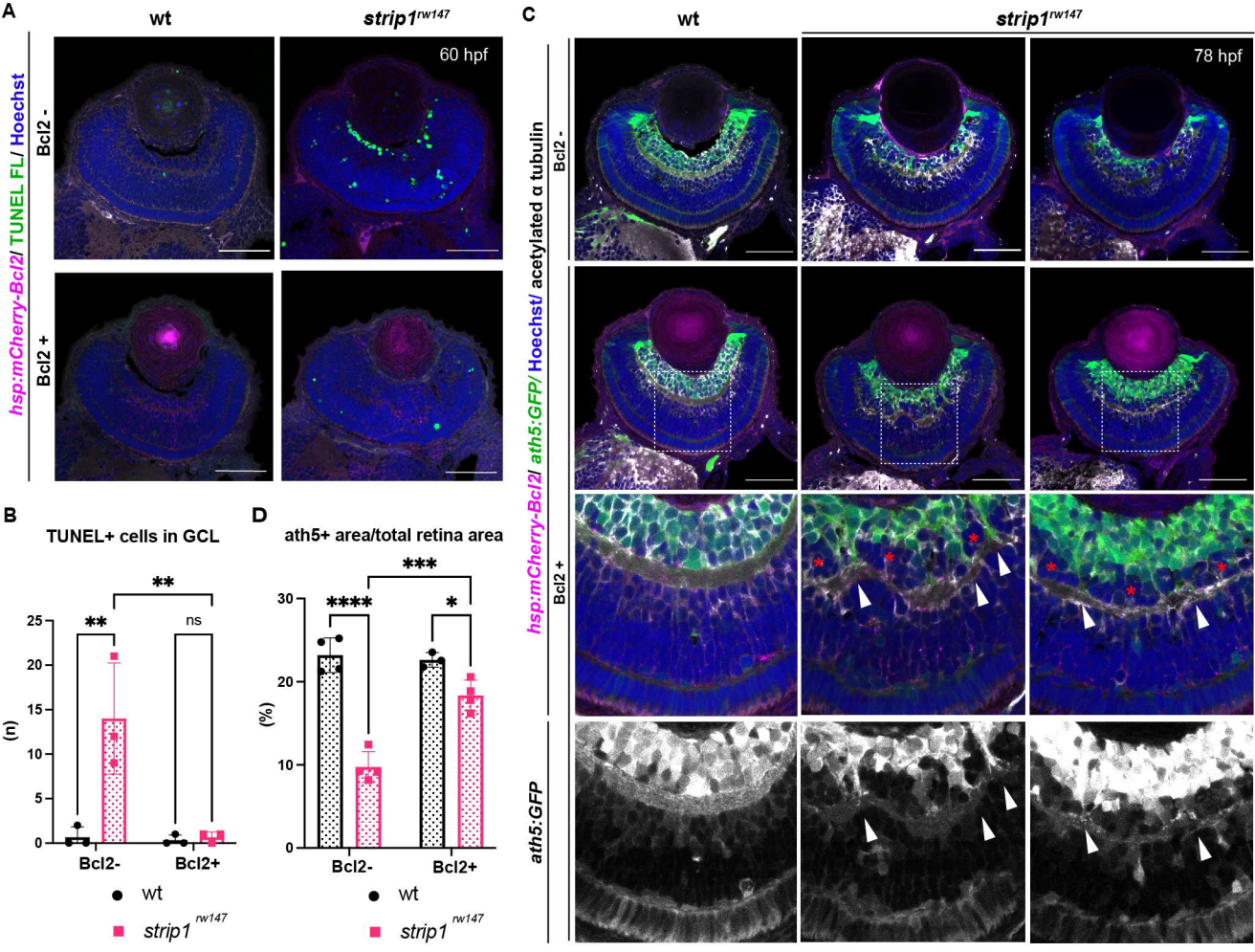

A
wt
strip1rw147
60 hpf
hsp:mCherry-Bcl2/ TUNEL FL/ Hoechst
Bcl2 -
Bcl2 +
B
TUNEL+ cells in GCL
(n)
**
**
ns
Bcl2-
Bcl2+
wt
strip1 rw147
D
ath5+ area/total retina area
(%)
***
****
*
Bcl2-
Bcl2+
wt
strip1 rw147
C
wt
strip1rw147
78 hpf
hsp:mCherry-Bcl2/ ath5:GFP/ Hoechst/ acetylated α tubulin
Bcl2 -
Bcl2 +
ath5:GFP

Figure 7.

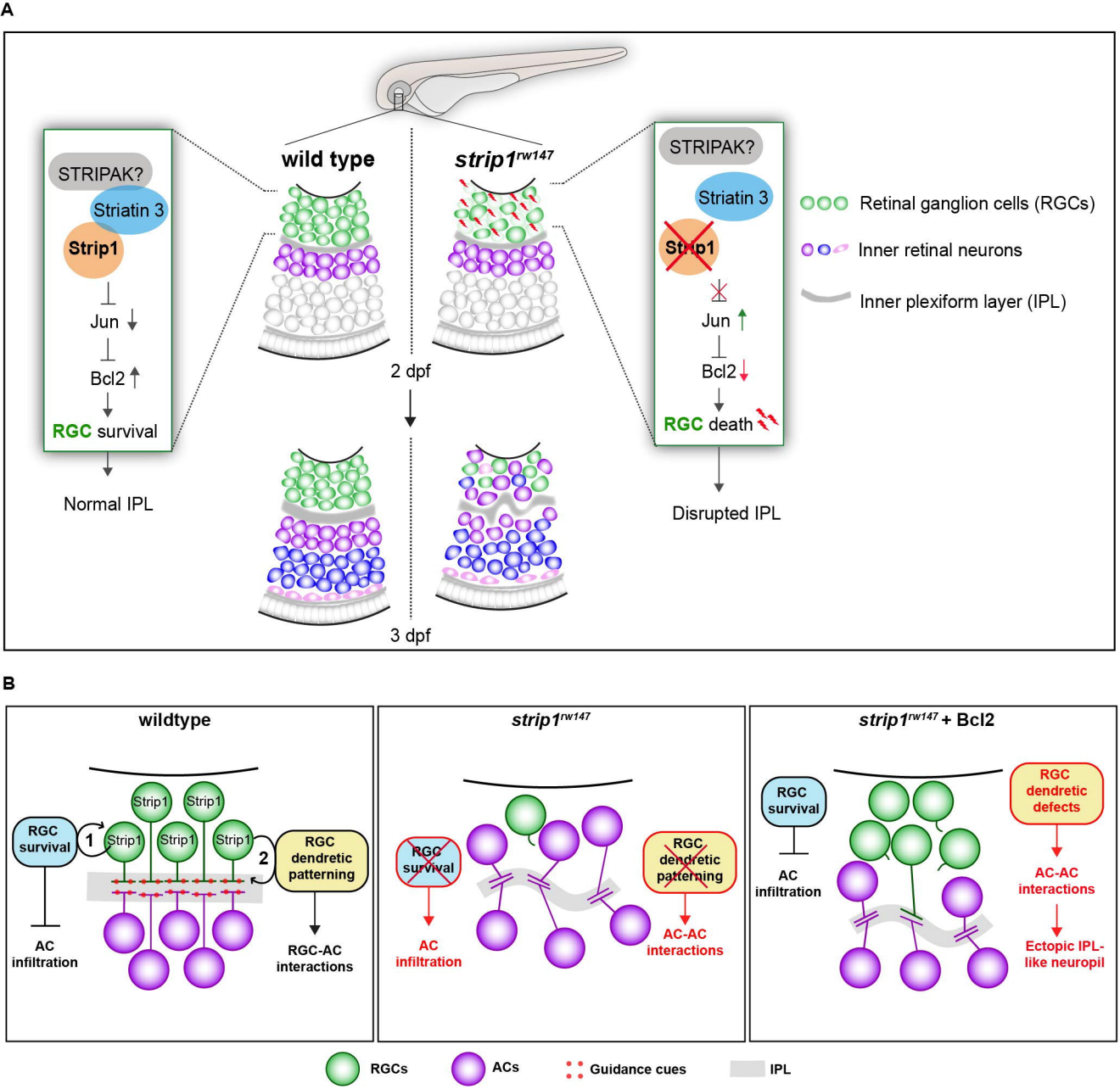

A
STRIPAK?
Striatin 3
Strip1
Jun
Bcl2
RGC survival
Normal IPL
wild type
strip1rw147
2 dpf
3 dpf
STRIPAK?
Striatin 3
Strip1
Jun
Bcl2
RGC death
Disrupted IPL
Retinal ganglion cells (RGCs)
Inner retinal neurons
Inner plexiform layer (IPL)
B
wildtype
RGC survival
1
Strip1
2
RGC dendretic patterning
AC infiltration
RGC-AC interactions
strip1rw147
RGC survival
AC infiltration
RGC dendretic patterning
AC-AC interactions
strip1rw147 + Bcl2
RGC survival
AC infiltration
RGC dendretic defects
AC-AC interactions
Ectopic IPL-like neuropil
RGCs
ACs
Guidance cues
IPL

Figure 8.