In vitro analysis of the effects of acetaminophen, ibuprofen and
aspirin on motility of female adult *Onchocerca volvulus* worms

Kenneth B. Otabil [1,2,3], Rhoda K. Antwi[3], Prince Nyarko[3], Theophilus N. Babae[1,3], Blessing
Ankrah[1], Emmanuel J. Bart-Plange[1,3], Rexford Kyei[3], Joseph Ameyaw[5], Joseph G. Bamfo[6],
Samuel F. Gyasi[2,3], Henk DFH Schallig[3]

[1]The NeTroDis Research Group, Centre for Research in Applied Biology, School of Sciences,
University of Energy and Natural Resources, Sunyani, Bono Region, Ghana
[2]Centre for Research in Applied Biology, School of Sciences, University of Energy and Natural
Resources, Sunyani, Bono Region, Ghana
[3]Department of Basic and Applied Biology, School of Sciences, University of Energy and
Natural Resources, Sunyani, Bono Region, Ghana
[4]Amsterdam University Medical Centres, Academic Medical Centre, University of
Amsterdam, Department of Medical Microbiology, Experimental Parasitology Unit,
Amsterdam, The Netherlands;
[5]Office of the Medical Superintendent, Happy Family Hospital, Nkoranza, Bono East Region,
Ghana
[6]Tain District Hospital, Nsawkaw, Bono East Region, Ghana

Correspondence should be addressed to Kenneth B. Otabil; kenneth.otabil@uenr.edu.gh

19 Abstract

In view of the very ambitious global timelines for elimination of onchocerciasis in 2030, the
search for alternative antifilarials cannot depend on drug development from scratch, and
repurposed drugs offer cheaper and faster alternatives. Previous studies had demonstrated the
presence and potential expression of amidase, an enzyme that can be targeted by repurposed
analgesics, *in Onchocerca volvulus worms*. The aim of this study was to determine the effects
of acetaminophen, ibuprofen and aspirin on the motility of adult *O. volvulus* worms. In total,
thirty (30) female *O. volvulus* worms were exposed to acetaminophen, ibuprofen and aspirin
in concentrations of either 5mg/ml, 2.5mg/ml, 1.25mg/ml or 0.63mg/ml control in duplicates.
Worm motility was observed and recorded using the WormAssay software and a darkfield
imaging apparatus starting on day 2 of incubation and ending on day 8. Acetaminophen,
ibuprofen and aspirin inhibited *O. volvulus* motility by 2-fold compared to control in the first
24 hours of drug exposure. However, an extended exposure of the worms to these test drugs
rather improved the motility of the worms. The study has demonstrated that a 24-hour exposure
of *O. volvulus* worms to the analgesic drugs studied, results in significant inhibition in worm
motility compared with the control group, but extended duration of exposure led to an
enhancement in motility of the previously immobile worms. This finding supports the idea that
aspirin and ibuprofen may have some longevity enhancing properties. Further research on the
utility of these analgesics as possible anti-filarial drugs is thus warranted.

38 Introduction

39 Onchocerciasis (river blindness) is a neglected tropical disease (NTD), caused by the parasite
40 *Onchocerca volvulus* and transmitted by blackflies [1]. Currently, the disease is still a major
41 public health concern even after years of both vector control and Mass Drug Administration
42 (MDA) with ivermectin [2]. It is a chronic cutaneous and ocular disease characterized by skin

nodules, severe itching and ocular lesions that can progress to partial or total blindness [3]. Globally, about 21 million people are infected and a further 200 million people at risk of contracting the disease according to the 2017 Global Burden of Disease Study [4]. Sub-Saharan Africa (SSA) bears approximately 99% of the disease burden [5]. In Ghana, recent updates demonstrated that 15 of its 16 regions (120 districts) are endemic for onchocerciasis, with more than 7 million people at risk of contracting the disease [6].

The key strategy for onchocerciasis control has been the use of ivermectin for the past 10 years as an effective microfilaricidal drug with no significant macrofilaricidal effects. However, some worrying reports from studies in West Africa have provided evidence of the development of sub-optimal response of *O. volvulus* parasites to ivermectin [7]–[9]. Aside this challenge, as ivermectin is purely microfilaricidal and does not kill adult worms [10], control programmes have had to administer the drug for many years in order to interrupt the transmission of the disease in endemic communities [11]. A major breakthrough in anti-filarial therapy was the discovery of anti-*Wolbachia* antibiotics such as doxycycline, which demonstrably had adulticidal effects [12]. However, logistical challenges associated with mass administration of antibiotics prevents the deployment of these antibiotics at the community level (Hoerauf, 2008; Hugues, Nana-Djeunga et al., 2014; Debrah et al., 2015). As a result of these difficulties, coupled with the suspension of the World Health Organization (WHO) sponsored Onchocerciasis Control Programmes in endemic communities since 2002, there seems to be a resurgence of the disease in some tropical foci [14].

It is against this background that the global health community, led by the WHO through a recently published NTD roadmap for 2021–2030 [15] has set elimination of transmission (EOT) targets for onchocerciasis, with 12 countries verified for EOT by 2030. Nevertheless, in some highly endemic foci, it might be impossible to achieve elimination of the disease with a sole reliance on ivermectin [16]. Therefore, the search for a microfilaricide and alternative drugs against onchocerciasis remains a top research priority [17].

The discovery and development of new drugs requires a lot of time (10–15 years), complex and expensive processes, and the success rate has been low, only 2.01% [18]. This makes it less advantageous for current onchocerciasis control efforts to depend on development of new drugs from scratch in view of the ambitious EOT timelines of 2030. In this regard, drug repurposing may be the best strategy to discover effective adulticidal drugs from existing approved drugs since these have already passed preclinical and clinical stages [19], [20] and this stands out as an attractive option to find alternative antifilarial options for onchocerciasis.

A major discovery was made by Cotton *et al.,* [21] when they sequenced the entire genome of *O. volvulus* parasites and its *Wolbachia* endosymbiont. They provided a list of gene families that had key functions, which could potentially be exploited as targets for repurposed drugs against the parasite [21]. The investigators also identified enzymes (and their gene targets) that are potentially chokepoint enzymes and may thus be essential for *O. volvulus* survival [21]. One of the top 16 *O. volvulus* gene targets identified was the OVOC4110 gene which potentially codes for amidase, which may be critical in metabolic pathways of *O. volvulus* [21]. The authors opined that analgesics such as acetaminophen may block the action of amidase and consequently disrupt critical metabolic pathways in the worm [21]. As yet, no study has confirmed the expression of amidase in *O. volvulus* worms or the effects of these analgesics on adult *O. volvulus* worms, to the best of our knowledge. For the present study, we investigated the effects of acetaminophen, ibuprofen and aspirin on adult *O. volvulus* worms with the goal

of assessing the potential of these analgesics as repurposed, macrofilaricidal drugs to aid in the
control of onchocerciasis.

Acetaminophen (paracetamol), an analgesic and antipyretic, is the most widely used over-the-
counter medication in the most countries [22]. The mechanism of action of acetaminophen is
complex and includes the effects of both the peripheral (Cyclo-oxygenase inhibition), and
central (Cyclo-oxygenase, serotonergic descending neuronal pathway, L-arginine/Nitric
Oxide pathway, cannabinoid system) antinociception processes and Redox mechanisms [22].
Though, some studies have reported some antimicrobial effects of acetaminophen (AL-Janabi,
2010), the direct activity of acetaminophen against helminths in general and *O. volvulus* in
particular has not been demonstrated by any study till date, to the best of our knowledge. Cotton
et al. (2016) however speculated that acetaminophen might bind to amidases in *O. volvulus* and
potentially cause the death of the worms, hence its choice in the present study.

Ibuprofen is also a medication in the nonsteroidal anti-inflammatory drug (NSAID) class that
is used for treating pain, fever, and inflammation [24]. Like other NSAIDs, it works by
inhibiting the production of prostaglandins by decreasing the activity of the enzyme
cyclooxygenase [25] though it might be a weaker anti-inflammatory drug than other NSAIDs
[25]. The drug was chosen for the present study because aside being an analgesic, some
antimicrobial activity has been reported by previous authors [23], [26]. However, till date no
study has fully investigated the direct effects of ibuprofen on *O. volvulus* survival.

Aspirin (acetylsalicylic acid, ASA), an analgesic, antipyretic and anti-inflammatory drug, has
emerged as an interesting option for repurposing drugs against parasites [18]. ASA may also
be used for secondary prevention of stroke and acute cardiac events. ASA acts on the two
cyclooxygenase isoforms (Cyclo-oxygenase 1and 2) via acetylation of serine 532 in the active
site; inhibits the action of both enzymes and prevents the formation of prostaglandins from
arachidonic acid [27]. Epidemiological and clinical studies has shown that ASA also reduces
the incidence of epithelial tumours by the acetylation of multiple proteins including
transcription factors, cytoskeleton proteins, stress response proteins (including the heat shock
proteins, HSPs), membrane proteins, among others [28], [29], suggesting other mechanisms
and molecular targets. Some studies have evidenced antiparasitic activity, for instance, Singh
and Rathaur [30] demonstrated that ASA improved the antifilarial activity of
diethylcarbamazine [30]. Aspirin also demonstrably affects dynamics of the actin cytoskeleton
and decreases amebic movement in *Entamoeba histolytica (López-Contreras et al., 2013)*.
The aim of the study therefore was to determine the effects of acetaminophen, ibuprofen and
aspirin on the survivability of adult *O. volvulus* worms. This was important because there is
the need to quickly discover and develop effective macrofilaricidal alternatives to complement
ivermectin if global elimination targets of onchocerciasis are to be achieved.

Materials and Methods

Patient recruitment and nodulectomies
Onchocerca nodules were obtained from 11 participants who had consented to participate in
this study by signing/thumbprinting a written, informed consent form. The study was approved
by the Institutional Ethics Committee of the Kintampo Health Research Centre in Ghana
(Approval number: KHRCIEC/2018-18). The participants were recruited from 7
onchocerciasis endemic communities during a cross-sectional, epidemiological field survey in
November 2020 in the Bono Region of Ghana. A total of 560 individuals from all 7
communities were screened for cutaneous nodules (onchocercoma) by palpation [32] by the

medical team and eligible participants with palpable nodules, with no condition requiring long-
term medication, were recruited and scheduled for nodulectomies. Using local anaesthesia,
palpable nodules were surgically removed from the patients by the medical team. Participants
were then transported to their homes and wound care provided at the nearest health care facility
until the wound was healed under the supervision of the medical team. The excised nodules
were preserved in 0.85% normal saline in 20 ml plastic vials, placed on ice, transported to the
laboratory and stored at room temperature on the laboratory bench.
147
Collagenase digestion of excised nodule
In order to isolate the adult worms contained in the nodules, the latter were digested using the
collagenase technique [33]. Briefly, each nodule was incubated for 48 hours at 37°C
(depending on the nodule's weight) in 5ml of Gibco's high glucose Dulbecco's Modified
Eagle's Medium (DMEM, Gibco, 12100061) containing type I collagenase (Sigma Aldrich,
SCR103) at a final concentration of 2.25 mg/ml. The product of digestion (the worm mass and
digested human tissues constituting the nodule) was placed in a Petri dish containing 15 ml of
DMEM, sodium bicarbonate (Dainess, Ghana), and supplemented with Penicillin-
Streptomycin (Gibco, 15140122) at a final concentration of 2 mg/ml. Individual worms were
isolated under a dissecting microscope using entomological forceps and needles. Each entire
and living worm (dead or calcified and incomplete or broken worms were counted but
discarded from the further process) was then examined for sex. Entire male worms were
individually frozen for further studies. Thirty (30) female worms were examined for activity in
order to include them in the live worm assays after 24 hours of separation from nodules. Males
were excluded from this study because initial culture of the male worms indicated an unusually
fast decay in DMEM.
164
Drug preparation
Stock solutions of acetaminophen, ibuprofen, aspirin were prepared by grinding 500 mg of the
commercially available drugs [Paracetamol (M&G Pharmaceuticals, Ghana); Ibuprofen (Letap
Pharmaceuticals, Ghana); Aspirin (M & A Pharmaceuticals, UK)] in a breakable crucible and
dissolved in 50 ml of DMEM solution in a 250 ml beaker. Subsequently, concentrations of 5
mg/ml, 2.5 mg/ml, 1.25 mg/ml and 0.63 mg/ml were prepared using two-fold serial dilution
following a protocol developed by Al-Janabi [23]. The concentration of acetaminophen,
ibuprofen and aspirin in micromolars (mM) were; Acetaminophen (5 mg/ml=33.08mM, 2.5
mg/ml=16.54 mM, 1.25 mg/ml = 8.27 mM and 0.63 mg/ml = 4.17 mM), Ibuprofen (5 mg/ml
= 24.24 mM, 2.5 mg/ml = 12.12 mM, 1.25 mg/ml = 6.06 mM and 0.63 mg/ml = 3.06 mM),
Aspirin (5 mg/ml = 27.75 mM, 2.5 mg/ml = 13.88 mM, 1.25 mg/ml = 6.94 mM and 0.63 mg/ml
= 3.50 mM). The drug dilutions were stored at 4° C in a refrigerator for 24 hours prior to usage
in the O. volvulus assays.
178
O. volvulus worm culture nodule
The worms were cultured following established protocols [34] with a few modifications. The
experiment was set up by adding a single female adult *O. volvulus* worm to each well of 6-well
plates containing 5 ml of DMEM +10% fetal bovine serum (Gibco, A3840002) + 100 u/ml
penicillin + 100 ug/ml streptomycin + sodium bicarbonate (2 g/1). The worms were then
incubated at 37 °c in a gas phase of 5 % CO_2 in air for 24 hours prior to the addition of the
analgesics to adapt them to the culture medium. After 24 hours of incubation in the above
medium, 1 ml of each concentration of analgesics (5, 2.5, 1.25 and 0.63 mg/ml) was pipetted
into a previously appropriately labelled wells containing the pre-incubated *O. volvulus* worms
in the medium. Two (2) wells with DMEM but no analgesics added were used as the control
for this study. The worms were again incubated at 37 °C in a gas phase of 5% CO_2 in air for 8

days with motility reading performed daily starting after 24 hours. Close examination of worms was performed to identify signs of disintegration and the medium changed every other day as colour changes indicated depletion of glucose in the culture medium.

The WormAssay and the darkfield imaging apparatus

The WormAssay software previously developed by Marcellino et al. (2012) for antifilarial screening of *Brugia malayi* was repurposed in the present study for *O. volvulus* [36]. The WormAssay is an inexpensive system for quantifying parasite movement based on worm motility. The apparatus uses a commodity video camera, computer and a newly developed free and open source software application [35] to provide quantitative measurements of parasite motility on entire plates. The application is designed to process multiple wells simultaneously without user interaction and this automatically identifies each well in the plate and labels the output data accordingly. This system can be used to assay large parasites such as filarial nematodes as well as other macroparasites. WormAssay's automation of the video capture step and lack of need for any interaction with the computer software during scoring differentiates it from other existing motion-based schemes. For the video capture system, a dark field macroscopic imaging apparatus based on the design of Marcellino et al. (2012) was developed with slight modifications.

Measurement of motility using the WormAssay software and darkfield imaging system

Before the addition of the test drugs, the motility of the female worms to be used were observed by competent laboratory technicians (parasitologists) and judged to be of sufficient motility to include in the assays. Using the WormAssay and the capture device, each 6-well microplate was observed and scored for motility. One (1) minute video recordings using the Lucas-Kande Optical Flow algorithm were taken of each plate and mean optical flow movement units for each worm were converted to percent motility by normalization in Graphpad Prism. The effective concentration to reduce mean motility levels to 50% (EC50) of control values were also determined using Graphpad Prism. The full profile of % mean motility scores (MMS) were illustrated graphically and statistical comparisons made.

Statistical analyses

Statistical analysis was performed using Graphpad Prism (version 8.2.1). In plotting the dose response curves, all movement units were normalized and the EC50 (the concentration of a drug that gives half-maximal response was calculated for acetaminophen, ibuprofen and aspirin) were determined. For the graphical plots, the mean of the normalized movement units was plotted against the concentration or the day depending on the type of plot. Differences in mean motility scores between groups were calculated using the Mann-Whitney test with differences considered to be statistically significant at the 95 % confidence interval if $P < 0.05$. The standard deviations (SD) of the mean motility scores were determined and subsequently converted to % SD for consistency.

Results and Discussion

Results

The results of the effects of acetaminophen, ibuprofen and aspirin on the survival of adult female *O. volvulus* worms are presented in Figures 1-5. The indicator of survival of *O. volvulus* female worms in this study was the percentage mean motility scores (% MMS).

Effects of acetaminophen, ibuprofen and aspirin on motility of adult worms

The results of the % MMS of the *O. volvulus* after a 24-hour exposure to the various concentrations of analgesics are presented in Figure 1. It can be seen in Figure 1 that at the 0.63 mg/ml dose of acetaminophen, the % MMS was 41% (SD=11.6%). The value of % MMS continued to increase as the concentration of acetaminophen increased [1.25 mg/ml (63%, SD=100%), then 2.5 mg/ml (68%, SD=18%)]. The only concentration of acetaminophen which performed best in terms of reducing motility of the worms was the 5 mg/ml, where the % MMS decreased to 5% (SD=0%). The EC50 (the concentration of a drug that gave half-maximal response) value of acetaminophen in this study was 8.74 mg/ml (57.82 mM).

The results show that at the lowest concentration of ibuprofen (0.63 mg/ml), the % MMS of worms in the ibuprofen group was 0% (SD=7%) compared to 100% (SD=100%) in the control group. For ibuprofen concentrations of 1.25 mg/ml, 2.5 mg/ml and 5 mg/ml, the % MMS of the female adult worms were respectively at 20 % (SD=0%), 23 % (SD=1%) and 24 % (SD=17%). The EC50 for ibuprofen was 0.65mg/ml (3.15 mM).

The findings on the effects of aspirin on the worms showed that at the startup aspirin concentration (0.63 mg/ml), 30 % (SD=61 %) of the worms were markedly motile within the aspirin group compared to 100 % in the control group. The lowest % MMS for worms in the aspirin group (0%, SD=0 %) was observed at an aspirin concentration of 1.25 mg/ml. However, at the 5 mg/ml concentration of aspirin, 41 % (SD=55 %) of the female adult worms in the group were still motile. The EC50 of aspirin in this study was 0.26 mg/ml (1.44mM).

Figure 1: Dose response curve of the effects of Acetaminophen, Ibuprofen and Aspirin on female adult *O. volvulus* worms after 24 hours of incubation

Response of O. volvulus parasites to varying concentrations of the test drugs during 8 days of culture

The following Figures (2-5) presents the results of the effects of varying concentrations of the test drugs (control, 5 mg, 2.5mg, 1.25mg/ml and 0.63 mg/ml) on female adult worms in culture for 8 days. Figure 2 summarizes the response of the of *O. volvulus* worms to 5 mg/ml concentrations of acetaminophen, ibuprofen, aspirin and control. The findings demonstrate that at 24 hours of culture (day 2), all test drugs including control had no effects on the motility of the worms as the % MMS of female adult worms in each group was 100 %. A similar observation was made on day 3 as all test drugs again had similar effects on worm motility. For day 4, the study observed that acetaminophen had the lowest effect on worm motility with % MMS of 21% (SD = 1.6%), with this trend continuing even to the 8th day of culture where 55% (SD=5.7%) of worms were still highly motile despite exposure to 5mg/ml of acetaminophen. Generally, the differences in % MMS between the aspirin and control were statistically significant (P = 0.026; Mann-Whitney test). There was however no statistically significant difference in % MMS between acetaminophen and control (P = 0.052; Mann-Whitney test) and ibuprofen and control (P = 0.127; Mann-Whitney test).

Figure 2: Comparison of 5 mg concentration of Acetaminophen, Ibuprofen, Aspirin on the motility of *O. volvulus* over the course of 8 days of culture

A summary of the effects of the 2.5 mg/ml concentration of acetaminophen, ibuprofen and aspirin on the motility of female adult *O. volvulus* are presented in Figure 3. After 24 hours (day 2) of exposure of female adult *O. volvulus* to the test drugs, aspirin had the highest effect of reducing the % MMS, but even then, still had a % MMS of about 59% (SD = 2.2%). For the remaining days of culture (except on day 7), acetaminophen and ibuprofen outperformed aspirin. For instance, on day 3 of culture, the % MMS of the acetaminophen and ibuprofen group were only 5 % (SD = 56%) and 11% (SD = 16%) of worms in the, respectively, were motile compared to 30% of the worms in the aspirin group. It is interesting to note that on this day, the control group performed better than all the test drugs in terms of reduction in % MMS. There was a statistically significant difference between ibuprofen and control (P = 0.037; Mann-Whitney test) as well as between aspirin and control (P = 0.026; Mann-Whitney test). However, the difference between acetaminophen and control for this concentration group was not statistically significant (P = 0.104; Mann-Whitney test).

295
296 **Figure 3: Comparison of 2.5mg concentration of Acetaminophen, Ibuprofen, Aspirin on**
297 **the motility of *O. volvulus* over the course of 8 days of culture**
298
For the 1.25 mg /ml concentrations of the test drugs, aspirin performed best on day 2 (after 24
hours of exposure to the drug) though again, it was subsequently outperformed by all other test
drugs including control (Figure 4). The only exception was on day 6, when no motility was
observed for the aspirin group (0 % MMS, SD = 0 %) and day 8, when it performed better than
ibuprofen. Surprisingly however, the control group continued to demonstrate very low motility
throughout the culture period. For this drug concentration, there were statistically significant
differences between acetaminophen and control (P = 0.026; Mann-Whitney test), ibuprofen
and control (P = 0.037; Mann-Whitney test) but not between aspirin and control (P = 0.071;
Mann-Whitney test).
308

309
310 **Figure 4: Comparison of 1.25 mg concentration of Acetaminophen, Ibuprofen, Aspirin**
311 **on the motility of *O. volvulus* over the course of 8 days of culture**

312
The findings on the response of *O. volvulus* adult females to 0.63 mg/ml concentration of the
various test agents as well as control is presented in Figure 5. On day 2, the best performing
drug in terms of reduction in % MMS was ibuprofen, but even then, the % MMS was 70%
(SD=7%) indicating a high worm motility in the group. Generally, however, acetaminophen
performed better at this concentration when compared with aspirin and ibuprofen. Again, the
control group continued to show relatively lower motility scores when compared with the
acetaminophen, ibuprofen and aspirin. These differences in % MMS was statistically
significant for ibuprofen versus control (P = 0.026; Mann-Whitney test) but not for aspirin
versus control (P = 0.052; Mann-Whitney test) and acetaminophen versus control (P = 0.104;
Mann-Whitney test).
323

Figure 5: Comparison of 0.63 mg/ml concentration of Acetaminophen, Ibuprofen, Aspirin on the motility of *O. volvulus* over the course of 8 days of culture

Discussion

In response to the need to develop alternative drugs to augment ivermectin and based on empirical evidence from the sequencing of the genome of *O. volvulus* by Cotton et al.(2016), the present study tested the hypothesis that analgesics such as acetaminophen, ibuprofen and aspirin has adulticidal effects on adult female *O. volvulus* worms.

For acetaminophen, this study demonstrated that a the 24-hour exposure of the worms to 5 mg/ml acetaminophen completely immobilized the worms suggesting a potential anti-filarial effect. However, the summary of results of the various concentrations of acetaminophen over the course of the 8 days incubation revealed that all concentrations from this drug improved the motility of the female adult worms and hence did not indicate any anti-filarial effect. In fact, the 1.25 mg/ml concentration of acetaminophen significantly improved worm motility. This finding is contradictory to the suggestion by Cotton et al. (2016) that analgesics, such as acetaminophen, could have adulticidal effects on *O. volvulus* by blocking amidase, a potentially choke point enzyme. The ability of acetaminophen to enhance parasite motility and survival demonstrated in this study may not be surprising as a previous study on *Plasmodium falciparum* also demonstrated that the drug may prolong parasite clearance and had no direct antiparasitic effects [37]. Though the foregoing study was on malaria parasites, the results from the current study suggest a similar worm enhancing effect. It is therefore justified that prior to our study, acetaminophen had only been utilized as an antipyretic in the treatment of helminthic infections, with no direct effects on parasite intended. However, as the present study did not investigate the combination of acetaminophen and antifilarials, it is still possible that such a combination, in the right concentration, might be deleterious to *O. volvulus* parasite survival. This may not be far-fetched as some investigators have demonstrated the antimicrobial ability of acetaminophen on bacteria and fungi at different concentrations [23], [26]. Taken together, it is posited that there is a possible inhibitory effect of acetaminophen which is both dose and

time dependent as the 24-hour exposure of worms to 5mg/ml acetaminophen completely immobilized them, but more research is needed in this area.

This study also investigated the effects of ibuprofen on the survivability of adult female *O. volvulus* worms. The findings showed that ibuprofen reduced the motility of the parasites at different concentrations within the first 24 hours of culture and this represented a two-fold increase in inhibitory activity over that of control. This finding is novel as this study represents the very first report of the inhibitory effects of ibuprofen on *O. volvulus* or on helminths in general. The antimicrobial effects of ibuprofen have been demonstrated previously for bacteria such as *Acinetobacter baumannii, Bacillus spp*., *Helicobactor pylori*, *Klebsiella spp*, etc as well as for fungi including *Candida albicans*, *Epidermophyton floccosum*, *Penicillium expansum* etc [26]. A rather surprising finding was the fact that summary of results of long-term (8 days) of exposure of worms to ibuprofen improved the motility of the worms. The improvement in worm motility by ibuprofen was statistically significant at all concentrations with the exception of the 5 mg/ml concentration when compared to the control group. This finding was particularly interesting because based on a previous study by Cotton et al.(2016), the current study had hypothesized that ibuprofen, being an analgesic, binds to amidase which is probably a chokepoint enzyme, leading to worm death. Thus, the reason for this observed significant improvement in motility of female adult worms following a relatively long-term exposure to ibuprofen is not known though the idea that ibuprofen may improve the longevity of microbes is not new [38]. He et al. (2014) discovered that the common drug ibuprofen increases the lifespan of yeast, worms and flies (specifically *Saccharomyces cerevisiae*, *Caenorhabditis elegans* and *Drosophila melanogaster* respectively), indicative of conserved longevity effects. They opined that in budding yeast, ibuprofen's pro-longevity action is independent of its known anti-inflammatory role [38]. They also revealed that the critical function of ibuprofen in longevity is to inhibit the uptake of aromatic amino acids, by destabilizing the high-affinity tryptophan permease. Nevertheless, in consideration of the differences in effects of ibuprofen in the short-term and after extended exposure in this study, it is suggested that effect of ibuprofen just like acetaminophen is both dose and time dependent. The findings from the present study therefore begs for more research in this area to unravel further details on the effects of ibuprofen on *O. volvulus* worms and its potential role either as a treatment option or as a supplement in culturing worms in the lab.

The study further investigated the effects of aspirin on female adult *O. volvulus* worms. The findings showed that in the short-term, aspirin also demonstrated a 2-fold inhibition of the motility of the worms compared to the control group. This finding was not surprising as aspirin is a known nonsteroidal anti-inflammatory drug which can inhibit prostaglandin H synthase and also induces apoptosis [30]. In fact, Singh and Rathaur (2010) working with *Setaria cervi,* a bovine filarial parasite, demonstrated that exposure of the worms to a combination of diethylcarbamazine plus aspirin at a concentration of 100 mM irreversibly paralyzed adult worms as well as microfilariae within 2 hours of exposure. Evidence from their study also showed that the worms might have been killed due to mitochondrial mediated apoptosis [30]. In another study by Ochoa-Maganda et al. (2020), aspirin showed a direct anti-giardial activity and affected the adhesion and growth of trophozoites in a time-dose-dependent manner [18]. However, for the present study, what was surprising in the aspirin group was the observation that over the course of the entire period of incubation, aspirin seemed to enhance the motility of the female adult worms. The reason for this is not known though aspirin has also long been a subject of research on pro-longevity in humans, thereby warranting further research in the area.

As observed in this study, the general trend is that at the onset of incubation, acetaminophen, ibuprofen and aspirin, demonstrated some inhibitory effects on female adult worms. However, after an extended period of exposure, the drugs improved the motility of the worms indicating that the effects of these drugs are both dose and time dependent. It is speculated, that at the beginning of the introduction of the drugs, the analgesics might bind to amidase [21] and inhibit the worms within 24 hours duration of exposure. It is however possible, that amidase might not be a chokepoint enzyme. Hence, the worms could switch to alternative metabolic pathways [17] which does not require amidase and might thus increase their survivability *in vitro*, thereby accounting for the observed enhancement in worm motility scores in this study.

It was further observed that the control group which had only medium showed the worst motility score from day 3 onward. This was quite unexpected and the reasons are not immediately apparent. It is possible that the addition of feeder cells such as monkey kidney cells (LLXC-NK2-cells) in the assay would have significantly improved the survival of worms in the control [39]. However, Townson and colleagues (1986) in their work demonstrated the survival of *O. volvulus* worm *in vitro* for over 15 days even without feeder cells. More so, in the present study, the worms in the drug arms survived better even in the absence of feeder cells, hence the reasons for the low survival in the control group after day 3 cannot be due to the absence of these feeder cells alone. It is speculated that the additional filing materials in the drugs could contribute to the worm's metabolism and thus improve worm motility compared to control. Hence, it is recommended that future investigations utilize pure research grade drug powder to eliminate possible confounders in the tablets. Nevertheless, the present study provides the foundation for further studies on the potential repurposing of these drugs for the treatment of onchocerciasis.

Conclusions

The study provides, for the first time, evidence on the direct effects of acetaminophen, ibuprofen and aspirin on the motility of female adult *O. volvulus* worms. The study has demonstrated that a 24-hour exposure of these parasitic worms to the study analgesic drugs results in significant inhibition in worm motility compared with the control group. Longer duration of exposure of the worms to the different concentrations of the study drugs however led to a rather surprising enhancement in motility of the previously immobile worms. There is therefore the need for more research to determine the utility of these analgesics as possible anti-filarial drugs or supplements to improve *in vitro* culturing of *O. volvulus* worms for other studies.

Data Availability

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Conflicts of Interest

The author(s) declare(s) that there is no conflict of interest regarding the publication of this paper.

Funding Statement

This research received no specific grant from any funding agency, commercial or not-for-profit sectors.

Acknowledgments

We thank all the staff and students of the Department of Basic and Applied Biology who provided support in one way or the other to ensure the successful implementation of this research. We also appreciate the efforts of the directors and staff of the Wenchi Municipal Health Directorate, Nsawkaw District and the Subinso Health Centre for their enormous contributions towards the study. Our appreciation also goes to the chiefs and people of the study communities who partnered with us in this study.

Supplementary Materials

Not applicable

References

[1]   WHO, "WHO | Onchocerciasis elimination mapping of endemic countries is key to defeating river blindness," *WHO*, 2019, Accessed: Apr. 14, 2019. [Online]. Available: https://www.who.int/neglected_diseases/news/Onchocerciasis-elimination-mapping-of-endemic-countries-is-key/en/.

[2]   K. B. Otabil *et al.*, "Prevalence of onchocerciasis and associated clinical manifestations in selected hypoendemic communities in Ghana following long-term administration of ivermectin," *BMC Infect. Dis.*, vol. 19, no. 1, p. 431, Dec. 2019, doi: 10.1186/s12879-019-4076-2.

[3]   E. V Noormahomed, K. Akrami, and C. Mascaró-lazcano, "Onchocerciasis , an undiagnosed disease in Mozambique : identifying research opportunities," *Parasit. Vectors*, pp. 1–8, 2016, doi: 10.1186/s13071-016-1468-7.

[4]   WHO, "Onchocerciasis Factsheet," 2020. https://www.who.int/news-room/fact-sheets/detail/onchocerciasis (accessed Jul. 30, 2020).

[5]   G. Gebrezgabiher, Z. Mekonnen, D. Yewhalaw, and A. Hailu, "Reaching the last mile: Main challenges relating to and recommendations to accelerate onchocerciasis elimination in Africa," *Infectious Diseases of Poverty*, vol. 8, no. 1. 2019, doi: 10.1186/s40249-019-0567-z.

[6]   K. Marfo, "Update of Oncho Program Status," 2018. [Online]. Available: https://dndi.org/wp-content/uploads/2018/05/Marfo_CountryProgressGhana_UgandaMeeting2018.pdf.

[7]   K. Awadzi *et al.*, "An investigation of persistent microfilaridermias despite multiple treatments with ivermectin, in two onchocerciasis-endemic foci in Ghana," *Ann. Trop. Med. Parasitol.*, vol. 98, no. 3, pp. 231–249, Apr. 2004, doi: 10.1179/000349804225003253.

[8]   M. Y. Osei-Atweneboana, D. A. Boakye, K. Awadzi, J. O. Gyapong, and R. K. Prichard, "Genotypic analysis of β-tubulin in Onchocerca volvulus from communities and individuals showing poor parasitological response to ivermectin treatment," *Int. J. Parasitol. Drugs Drug Resist.*, vol. 2, pp. 20–28, Dec. 2012, doi: 10.1016/j.ijpddr.2012.01.005.

[9]   A. Eisenbarth, M. D. Achukwi, and A. Renz, "Ongoing transmission of Onchocera volvulus after 25 years of annual mass ivermectin treatments in the Vina Du Nord River Valley, in North Cameroon," *PLoS Negl Trop Dis*, vol. 10, no. 2, 2016.

[10]   A. T. Abegunde, R. M. Ahuja, and N. J. Okafor, "Doxycycline plus ivermectin versus ivermectin alone for treatment of patients with onchocerciasis," *Cochrane Database of Systematic Reviews*, vol. 2016, no. 1. 2016, doi: 10.1002/14651858.CD011146.pub2.

[11] A. Y. Debrah *et al.*, "Doxycycline Leads to Sterility and Enhanced Killing of Female Onchocerca
volvulus Worms in an Area with Persistent Microfilaridermia after Repeated Ivermectin Treatment: A
Randomized, Placebo-Controlled, Double-Blind Trial," *Clin. Infect. Dis.*, vol. 61, no. 4, pp. 517–526,
2015, doi: 10.1093/cid/civ363.

[12] A. Hoerauf, "Filariasis : new drugs and new opportunities for lymphatic filariasis and onchocerciasis,"
2008, doi: 10.1097/QCO.0b013e328315cde7.

[13] H. Nana-Djeunga *et al.*, "Reproductive Status of Onchocerca volvulus after Ivermectin Treatment in an
Ivermectin-Naïve and a Frequently Treated Population from Cameroon," *PLoS Negl. Trop. Dis.*, vol. 8,
no. 4, p. e2824, Apr. 2014, doi: 10.1371/journal.pntd.0002824.

[14] WHO, "The Diseases," *Non-invasive Vent. Weaning Princ. Pract.*, pp. 193–194, 2015, doi:
10.1201/b13434-25.

[15] World Health Organization, "Ending the neglect to attain the Sustainable Developmetn Goals - A road
map for neglected tropical diseases 2021–2030," no. February 2019, pp. 1–13, 2020, [Online].
Available: http://apps.who.int/bookorders.

[16] M. G. Basáñez, S. D. S. Pion, T. S. Churcher, L. P. Breitling, M. P. Little, and M. Boussinesq, "River
blindness: A success story under threat?," *PLoS Medicine*, vol. 3, no. 9. pp. 1454–1460, 2006, doi:
10.1371/journal.pmed.0030371.

[17] M. A. Seidu, A. A. Rashid, G. K. Richard, and T. Yao, "Carbohydrate Catabolism in Adult Onchocerca
volvulus : An Carbohydrate Catabolism in Adult Onchocerca volvulus : An Immunohistochemical
Study," no. May, 2016, doi: 10.11648/j.ajbls.20160403.13.

[18] V. Y. Ochoa-Maganda *et al.*, "Antigiardial activity of acetylsalicylic acid is associated with
overexpression of hsp70 and membrane transporters," *Pharmaceuticals*, vol. 13, no. 12, pp. 1–16, 2020,
doi: 10.3390/ph13120440.

[19] J. A. DiMasi, "Risks in new drug development: Approval success rates for investigational drugs,"
Clinical Pharmacology and Therapeutics, vol. 69, no. 5. Clin Pharmacol Ther, pp. 297–307, May 2001,
doi: 10.1067/mcp.2001.115446.

[20] J. A. DiMasi, R. W. Hansen, and H. G. Grabowski, "The price of innovation: New estimates of drug
development costs," *J. Health Econ.*, vol. 22, no. 2, pp. 151–185, 2003, doi: 10.1016/S0167-
6296(02)00126-1.

[21] J. A. Cotton *et al.*, "The genome of Onchocerca volvulus, agent of river blindness," *Nat. Microbiol.*, vol.
2, no. November 2016, pp. 1–12, 2016, doi: 10.1038/nmicrobiol.2016.216.

[22] M. J. Benista and J. Z. Nowak, "PARACETAMOL : MECHANISM OF ACTION , APPLICATIONS
AND SAFETY CONCERN," vol. 71, no. 1, pp. 11–23, 2014.

[23] H. AL-Janabi, "In Vitro antibacterial activity of ibuprofen and acetaminophen," *J. Glob. Infect. Dis.*,
vol. 2, no. 2, p. 105, May 2010, doi: 10.4103/0974-777x.62880.

[24] N. M. Davies, "Clinical Pharmacokinetics of Ibuprofen The First 30 Years," vol. 34, no. 2, pp. 101–154,
1998.

[25] C. Gunaydin and S. S. Bilge, "Effects of nonsteroidal anti-inflammatory drugs at the molecular level,"
Eurasian J. Med., vol. 50, no. 2, pp. 116–121, 2018, doi: 10.5152/eurasianjmed.2018.0010.

[26] P. Zimmermann and N. Curtis, "Antimicrobial Effects of Antipyretics.," *Antimicrob. Agents
Chemother.*, vol. 61, no. 4, pp. e02268-16, Apr. 2017, doi: 10.1128/AAC.02268-16.

[27] J. Lei, Y. Zhou, D. Xie, and Y. Zhang, "Mechanistic insights into a classic wonder drug-aspirin," *J. Am.
Chem. Soc.*, vol. 137, no. 1, pp. 70–73, Jan. 2015, doi: 10.1021/ja5112964.

[28] E. Flossmann and P. M. Rothwell, "Effect of aspirin on long-term risk of colorectal cancer: consistent
evidence from randomised and observational studies," *Lancet*, vol. 369, no. 9573, pp. 1603–1613, May
2007, doi: 10.1016/S0140-6736(07)60747-8.

[29] A. T. Chan and E. L. Giovannucci, "Primary Prevention of Colorectal Cancer," *Gastroenterology*, vol.
138, no. 6, p. 2029, 2010, doi: 10.1053/j.gastro.2010.01.057.

[30] A. Singh and S. Rathaur, "Combination of DEC plus aspirin induced mitochondrial mediated apoptosis
in filarial parasite Setaria cervi," *Biochimie*, vol. 92, no. 7, pp. 894–900, Jul. 2010, doi:
10.1016/j.biochi.2010.03.018.

[31] L.-C. L *et al.*, "Exploring the possible role of lysine acetylation on Entamoeba histolytica virulence: a

focus on the dynamics of the actin cytoskeleton," *Biomed Res. Int.*, vol. 2013, 2013, doi:
10.1155/2013/757392.

[32]   H. P. Duerr, G. Raddatz, and M. Eichner, "Diagnostic value of nodule palpation in onchocerciasis,"
Trans. R. Soc. Trop. Med. Hyg., vol. 102, no. 2, pp. 148–154, Feb. 2008, doi:
10.1016/j.trstmh.2007.10.009.

[33]   H. C. Nana-Djeunga *et al.*, "Reproductive Status of Onchocerca volvulus after Ivermectin Treatment in
an Ivermectin-Naïve and a Frequently Treated Population from Cameroon," *PLoS Negl. Trop. Dis.*, vol.
8, no. 4, p. e2824, Apr. 2014, doi: 10.1371/journal.pntd.0002824.

[34]   S. Townson, A. R. Dobinson, J. Townsend, J. Siemienska, and G. Zea-Flares, "The effects of ivermectin
used in combination with other known antiparasitic drugs on adult Onchocerca gutturosa and O.
volvulus in vitro," *Trans. R. Soc. Trop. Med. Hyg.*, vol. 84, no. 3, pp. 411–416, 1990, doi:
10.1016/0035-9203(90)90340-K.

[35]   C. Marcellino, J. Gut, K. C. Lim, R. Singh, J. McKerrow, and J. Sakanari, "WormAssay: A novel
computer application for whole-plate motion-based screening of macroscopic parasites," *PLoS Negl.
Trop. Dis.*, vol. 6, no. 1, Jan. 2012, doi: 10.1371/journal.pntd.0001494.

[36]   G. S. Bedi, "Comparative study of kallikrein-like serine proteinases from rat submandibular glands,"
Prep. Biochem. Biotechnol., vol. 26, no. 2, pp. 85–104, 1996, doi: 10.1080/10826069608000056.

[37]   C. H. Brandts, M. Ndjavé, W. Graninger, and P. G. Kremsner, "Effect of paracetamol on parasite
clearance time in Plasmodium falciparum malaria," *Lancet*, vol. 350, no. 9079, pp. 704–709, Sep. 1997,
doi: 10.1016/S0140-6736(97)02255-1.

[38]   C. He *et al.*, "Enhanced Longevity by Ibuprofen, Conserved in Multiple Species, Occurs in Yeast
through Inhibition of Tryptophan Import," *PLoS Genet.*, vol. 10, no. 12, p. e1004860, Dec. 2014, doi:
10.1371/journal.pgen.1004860.

[39]   S. Townson, C. Connelly, and R. Muller, "Optimization of culture conditions for the maintenance of
Onchocerca gutturosa adult worms in vitro," pp. 323–330, 1986.

[40]   K. B. Otabil *et al.*, "In vitro analysis of the effects of acetaminophen , ibuprofen and aspirin on motility
of female adult Onchocerca volvulus worms," no. October, 2021.