Microglia-specific promoter activities of human *HexB* gene

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Abstract

Adeno-associated virus (AAV)-mediated genetic targeting of microglia remains challenging. Overcoming this hurdle is essential for the eradication of HIV reservoirs in the central nervous system (CNS) wherein microglia harbor the main HIV reservoirs. Here, we characterized the minimal/native promoter of the *HexB* gene, which is known to be specifically and stably expressed in the microglia under homeostasis and pathological conditions. Dual reporter and serial deletion assays identified the critical role of the natural 5' untranslated region (-97 bp related to the first ATG) in driving transcriptional activity of mouse *HexB* gene. The native promoter region of mouse, human and monkey *HexB* located at -135, -134 and -170 bp to the first ATG, respectively. These promoters were highly active and specific in microglia with strong cross-species transcriptional activity, but had no activities in primary astrocytes. To further exploit microglia specific promoters, we identified a 135 bp *CD68* gene promoter, similar to 134 bp *HexB* gene, was highly active in microglia but not in astrocytes. These data suggest that the novel microglia-specific *HexB* promoter can be an ideal candidate for specific microglia-targeting AAV gene therapy, which could be developed for HIV proviral eradication in the CNS.

Summary

It is hard to overstate the importance of gene therapy that can remove viral genes from human cells. A cure for HIV would mean a lifestyle free of treatment for patients who must constantly ensure that they are maintaining a strict regimen of ART indefinitely. In order to develop a cure using AAV delivery of HIV eradicator, DNA must meet the viral size limitations, and the genes must be expressed appropriately. Previous studies have identified microglia-specific *HexB* gene that shows stable expression during neural homeostasis and pathogenesis. Our study identified the essential *HexB* gene promoter (134 bp) as a strong candidate for AAV gene therapy to specifically target the brain microglia, the main cellular reservoirs of HIV in the CNS. Our studies continue to move us closer to identifying target-specific gene therapy for NeuroAIDS.

Introduction

Treatment for HIV-1 (HIV) has become increasingly effective in the past few decades since the epidemic swept the United States. While antiretroviral therapy (ART) decreases viral counts to undetectable levels and improves the quality of life for those affected, there is no cure for HIV/AIDS (1-3). Upon cessation of ART, viral loads will rebound to baseline or higher than baseline infection levels (3). In the brain, ART drugs have limited bioavailability and may not efficiently cross the blood-brain-barrier (BBB) (4), which may be associated with the failure in preventing the initial seeding of HIV in the central nervous system (CNS). This further leads to the establishment of the latent reservoirs in the brain and contributes to the HIV-Associated Neurocognitive Disorders (HAND), which persists in people living with HIV (PLWH) even with ART. In order to eradicate the disease as a whole, cellular HIV reservoirs hosting replication-competent HIV in the brain must be targeted by HIV cure strategies such as gene editing to excise disease-causing genetic material of HIV.

While there is evidence that many cell types harbor HIV proviral DNA and/or RNA in CNS (5, 6), such as brain myeloid cells (BMCs), astrocytes, neural stem cells (7-10), oligodendrocytes and CD4+ T cells (11-15); it remains to be determined which cell type(s) is the major HIV reservoir in CNS. As one major cell type of BMCs, the resident microglia act as the first line of defense against pathogens in CNS (15, 16). They are long-lived (17) and can self-renew *in vivo* (18-23), which may allow the persistence of HIV infection and serve for HIV reservoirs in the CNS. Blood-borne monocyte-derived macrophages (MDMs) are another source of BMCs (about 5-10%) (24, 25). They are in perivascular regions, terminally differentiated and frequently replenished by hematopoietic progenitors. However, with the relative short lifespan (~ a few months) and lack of self-renewing potential, it is unclear whether CNS MDMs can support long-lasting HIV infection. Mounting evidence supports the notion that the long-lasting HIV reservoirs establish in the CNS, where the brain microglia may constitute the major cellular CNS reservoirs (11, 14, 15, 26-33). In addition, it has been shown that microglia activation and HIV-associated neuronal damage are linked to HAND developed in up to half of PLWH (34-37). Therefore,

BMCs, especially microglia, are one of the major targets for HIV curative strategies that would permit ART-free remission and prevent CNS dysfunction.

Discovery of a proper microglia-specific promoter is needed for the development of gene editing tools specific for the eradication of HIV CNS reservoirs. An early study showed very weak expression of hexosaminidase subunit beta (HexB) in human brain (38). Recent RNAscope and single-cell RNA sequencing (scRNA seq) analysis determined *HexB* is exclusively expressed in mouse brain microglia but not monocyte/macrophages and other neural cells (39, 40). Importantly, this novel microglia-specific gene retains its expression under various pathological conditions, while many other microglia core genes are substantially downregulated (39, 40). This indicates that *HexB* could be an excellent option for microglia-specific targeting gene therapy for CNS injury and diseases. The insert size limitation in most gene therapy vectors has been a challenge for packaging capacity and transduction efficiency. In this study, we determined whether the *HexB* promoter is specific to microglia and whether its activity is species dependent. We also optimized the essential part of the native *HexB* promoter that has the potential to be used in the AAV gene therapy for brain HIV eradication.

Materials and Methods

Construction of HexB Promoter Vectors

NEBuilder® *HiFi* DNA Assembly cloning kit (NEB, Cat# E5520S) was used to clone *HexB* gene promoters of various sizes and species into a promoterless AAV plasmid vector containing gaussia dura luciferase and GFP (LG) dual reporter. First, PCR from genomic DNA was used to generate HexB promoter inserts. The three main sources of promoter DNA were human induced pluripotent stem cells (IPS), mouse embryonic stem cells (ES), and monkey LC30 cells. PCR was completed with Phusion High-Fidelity PCR Master Mix kit (Thermo Fisher, F531). At the same time, the promoterless AAV vector was digested with the appropriate restriction enzymes (Thermo Scientific FastDigest) to allow for insertion of the *HexB* promoter. The PCR products were purified from an agarose gel, and the digestion product was purified using Monarch PCR & DNA Cleanup Kit (NEB, T1030S). Following the preparation of both the backbone vector and insert, the two

components were ligated with the NEB HiFi assembly system, using a 1:10 ratio of backbone to insert. The assembled vector was then transformed into NEB stable competent cells and incubated overnight. In creating most vectors, colony PCR was used to confirm that the correct promoter size was inserted prior to sequencing. The correct clones were verified by restriction enzyme digestion and Sanger sequencing as well as functional measures.

Functional Testing of HexB Promoter in Cell Culture

The first experiments to determine the activity of the *HexB* promoters were performed in HEK293T cells using standard PeiMax transfection method. Then the promoter activities were tested in C20 human microglia and SIM-A9 mouse microglia cell lines using Glial-Mag kit (OZ Bioscience, Cat # GL002500). The cells were cultured under standard conditions and seeded into 96-well plates at a density of 3.0×10^4/100 µl. HEK293T cells were cultured in DMEM with 10% FBS. C20 cells were cultured in a DMEM with 5% FBS medium. SIM-A9 cells were cultured in DMEM with 10% FBS and 5% horse serum. After growing overnight, the cells were transfected with indicated AAV-HexB vectors. Transfections were completed in quadruplicates where each well received 100 ng of promoter reporter DNA and 20 ng of pSEAP2 control vector for transfection normalization. Empty promoterless LG vector was used as a negative control (empty vector). The CMV-driven pRRL-Flag-LG or pcDNA3-GFP vectors were used as the positive controls. At 24-96 hours post transfection, the cells were assessed for expression *via* fluorescence microscopy of GFP and gaussia luciferase assay.

After preliminary testing in microglial lines, transfections were performed using primary human microglia cells (Celprogen) and primary microglia cells isolated from fresh brain tissue of rhesus macaques. Primary human and NHP microglia were cultured in DMEM/F12 (Gibco) supplemented with 10% defined fetal bovine serum (HyClone), recombinant human M-CSF (R&D Systems), 100 µg/mL streptomycin, 100 U/mL penicillin, 2 mM L-glutamine, 3 mM sodium pyruvate and 10 mM HEPES. Primary human and NHP astrocytes were cultured in astrocyte medium (AM, ScienCell Research Laboratories) supplemented with FBS, Penicillin/Streptomycin and Astrocyte Growth Supplement (AGS) (Sciencell Research Laboratories). All cells were cultured in 37°C

incubator with 5% CO2. Cell transfections were carried out in 96-well plate with 1.0x10^4 target cells where each well received 100 ng DNA with a mix of Lipofectamine 3000 (Invitrogen) in Opti-MEM.

Luciferase Assays and GFP microscopy

Approximately 50 µl of culture supernatants were harvested 24-72 hours post transfection. Prior to luciferase assay, supernatant from cells was combined with an equal volume of coelenterazine (CTZ) substrate that was diluted 1:50 with CTZ dilution buffer. Prior to measurement, samples were allowed to incubate for 5 min at room temperature. Luciferase activity was measured by fluorescence plate reader (5 second integration) per manufacturer's instructions (NanoLight Technology). SEAP activity in the culture supernatants were measured with QUANTI-Blue™ Solution (Invivogen) following the manufacture's protocol. The data were analyzed in comparison with control empty vector transfection after SEAP normalization. At the end of transfection, the GFP+ cells were also analyzed by fluorescence microscopy to determine the gene transcription driven by promoters. The relative luciferase values=luciferase values in cells transfected by promoter-driven reporter - luciferase values in cells transfected with the empty vector where the negative values were normalized as zero.

Statistical analysis

Quantification of fold changes in promoter groups compared with corresponding promoterless or other promoter groups was performed. Statistical analysis was performed using Prism GraphPad 9.1. Significance at *$P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ and **** $P < 0.0001$ was determined using a two-tailed student's *t*-test between two groups or by one-way ANOVA for multiple comparisons. Data were presented as mean ± SE. The size and type of individual samples were indicated and specified in the figure legends.

Results

Validation of mouse *HexB* gene promoter activity with dual reporter assay

Given AAV gene therapy is promising in clinical trials, we selected AAV transfer vector for future application to AAV packaging and gene delivery. We have established dual

reporter assay using gaussia dura luciferase (gdLuc) and destabilized GFP (dsGFP) expression (thereafter referred as LG) that offer several benefits. It can measure the dynamic quantification of secreted luciferase in culture media with highly sensitive gdLuc assay. The transcriptional activity can also be determined by dsGFP expression with either fluorescence microscopy or flow cytometry (**Fig. 1A**). Both measures are useful for a fast and high-throughput analysis of *HexB* promoter activity. Based on previous reports on the optimal promoter activity of mouse *HexB* gene (38), we initially evaluated the activity of *HexB* promoter region -330 bp upstream of ATG (mHexB-330) (**Fig. 1A**). We selected HEK293T cells as the test platform because they are highly transfectable. We found that the mHexB-330 robustly facilitated dgLuc reporter expression by 20~22 fold 48 hours after transfection (**Fig. 1B-C**), which is consistent with the previous report using chloroamphenicol acetyltransferase reporter assay in NIH-3T3 cells (38). Fluorescent microcopy further confirmed the dramatic increase of dsGFP expression in mHexB-330 transfected cells (**Fig. 1C**). When 5'-UTR region was deleted, no transcription activity was observed, which validated the critical role of 5'-UTR (-97) in driving the promoter activity of HexB-330 (**Fig. 1B**) (38).

Cloning and characterization of essential promoters for human and rhesus macaque *HexB*

Given the high level of sequence conservation across species in the promoter region of *HexB* genes (41), we cloned the essential/minimal promoter region of human *HexB* for the potential application in human proviral eradication in future. As expected, the shorter mouse promoter mHexB-135 (135 bp) retained robust activity, despite moderately weaker than mHexB-330 (**Fig. 2**). Similarly, both human 134 bp *HexB* (hHexB-134) and 183 bp *HexB* (hHexB183) promoters have robust promoter activity, while the shorter promoter (HexB-134) retained high activity in driving gene transcription (**Fig. 2**). However, when a 68 bp core mouse *HexB* promoter (mHexB-Core68) was transfected, no transcription activity was observed compared with empty vector. This was also true for the human *HexB* promoter, where the 93 bp core (hHexB-Core93) and 43 bp core (HexB-core43) promoters did not have any transcriptional activities (**Fig. 2**), indicating the promoter regulatory region (40 bp) at -134 bp to -93 bp is required for human *HexB* promoter activity,

consistent with a previous report in human fibroblasts (42). To expand the potential application to SIV gene therapy in rhesus macaques, we cloned rhesus macaque 170 bp *HexB* (mkHexB-170) promoter (**Fig. 2**), which showed a similar activity to the human HexB-183 promoter. These data indicated that the essential and minimal/native *HexB* promoters are sufficient to drive gene transcription.

***HexB* promoter is active in microglia cell line derived from different species.**

HexB is exclusively expressed in microglia in the CNS *in vivo* (40). Thus, we determined whether mouse *HexB* promoter is active in microglia cell line derived from mouse brain. After transfection, we found that mHexB-330 but not mHexB135 promoter induced luciferase expression in SIM-A9 mouse microglial cells (**Fig. 3A**). Interestingly enough, hHexB-134, and to a less extent mkHexB-170, also was able to drive gene expression in mouse microglia. Surprisingly, the longer human HexB promoter, hHexB-183 was less effective to activate transcription, which was similar to the core promoters, hHexB-Core93 and hHexB-Core43.

To see whether there is cross-species activity of *HexB* promoters, we transfected these reporters into human microglia cell line C20. As shown in **Fig. 3B**, mouse HexB promoters (mHexB-330 and mHexB-135) lost their transcriptional capacity. In contrast, both human (hHexB-183) and monkey *HexB* (mkHexB-170) promoters highly induced transcriptional activity, which was lost when the core promoters were transfected into C20 cells. These data showed that human and monkey *HexB* promoters are active in driving gene transcription in both of the microglial cell lines originated from different species, suggesting that *HexB* promoter activity retains a high level of cross-species interaction. It was interesting that the transcription was similarly driven by human or rhesus macaque *HexB* promoters, this may reflect the similar genetic background between human and rhesus macaques. Since mHexB-135 failed to induce transcription in either of the above microglia cell line models (**Fig. 3**), we decided to focus on mHexB-330 promoter in the following studies.

***HexB* promoter is highly active in driving gene transcription in human and rhesus macaque primary microglia cells, but not in primary astrocytes.**

While SIM-A9 was derived from cortical tissues collected from postnatal mouse pups and spontaneously immortalized weeks after *in vitro* passaging (ATCC)(43), C20 microglia cell line model was originated from primary human microglia transformed by SV40 T antigen and human TERT(44). In order to generate an AAV vector for microglia targeting in a preclinical or clinical setting in the future, it is essential to test its transcriptional activity in the primary human microglia, but not in transformed human microglia. Thus, we examined the promoter activities of these *HexB* promoters in primary microglia cells. We first transfected HexB-driven dual LG reporters into the primary human microglial cells, where a CMV-driven pcDNA3-GFP and CMV-driven dual LG plasmids were included as positive transfection controls. We found that all these *HexB* promoters highly induced GFP expression (**Fig. 4A**). When further quantitated by luciferase assays, we discovered that the transcriptional activities of these *HexB* promoters were strong and comparable with each other (**Fig. 4B**), further supporting the cross-species activity observed in SIM-A9 and C20 (**Fig. 3)**. Surprisingly, when tested in the primary astrocytes, extremely low to no transcription activities were observed in the cells transfected in any species of the *HexB* promoters (**Fig. 4C-D**), either measured by GFP microscopy or by luciferase activity. Also, CMV-driven gene transcription in astrocytes was generally much lower than in microglia, which may be due to its lower transfection efficiency in astrocytes or astrocytes failed to efficiently support *HexB* promoter activity (**Fig. 4A-D**). It was notable that no luciferase activities were detectable in cells transfected with CMV-driven pcDNA3-GFP plasmids because pcDNA3-GFP does not contain luciferase gene (**Fig. 4A-D**). Similar patterns were observed in primary microglia and astrocytes derived from nonhuman primates (NHPs), i.e. rhesus macaques (**Fig. 4E-H**), where all these *HexB* promoters actively induces gene expression in microglia cells, but not in astrocytes. Taken together, these data support a notion that *HexB* promoter is able to drive gene transcription in a microglia-specific and possibly cross-species manner.

Efficient transcription potency of minimal and essential promoter of human *HexB* gene in microglia cells.

Essential *HexB* promoter with a minimal size is important for AAV gene therapy in targeting microglia. In the immortalized HEK293T cell model, we discovered that the core

HexB promoters had low to no transcription activities (**Fig. 2**) compared with their native mHexB-330, hHexB-134/183, or mkHexB-170 promoter, regardless of its origination from mouse (mHexB-Core68) or from human (hHexB-Core93/Core43). We have validated these observations in mouse or human microglia cell line, SIM-A1 and C20 (**Fig. 3A-B**). Similar to the HEK293T model cells, we found that hHexB-core promoters lacked transcription when the functional 5'-UTR were absent (**Fig. 3A-B**). This was similarly observed in the primary microglia cells derived from human or monkey brains (**Fig. 5A-D**). In primary astrocytes, except CMV-Flag-LG positive control transfection, no transcription was detected in either naïve or core HexB promoters-transfected cells (**Fig. 5E-H**). These data suggest that, considering the packaging size limits (~4.8 kb) in AAV packaging, the essential and small-sized human *HexB* promoter hHexB-134 may be an ideal candidate for AAV gene therapy targeting human microglia.

Transcription activity of *HexB* promoter in comparison to other known microglial gene promoters.

It has been shown that promoter from microglia markers, such as CD68, were able to drive gene transcription in mouse microglia, which promotes microglia-specific gene targeting *via* a novel capsid-modified AAV6 vector (45). However, it is not clear whether they have a similar promoter activity in the human microglia, particularly in the primary microglia cells. Since *HexB* promoter displayed unique transcription activity preferred in in microglia, it prompted us to compare their capacities of driving gene transcription. We cloned the well-established human *CD68* (695 bp, hCD68-695) promoter and its shorter version of 135 bp promoter (hCD68-135) (16, 39, 44) into our dual LG reporter vector. Initially, we discovered that hCD68-695 showed low transcription activity in HEK293T cells (**Fig. 2**), but strong promoter activities in SIM-A9 and C20 cell lines after transfection (**Fig. 3**). Surprisingly, its short version of counterpart hCD68-135 was also highly active in both microglia cell line models. Since human *HexB* promoter is more relevant to our human cell study, we set to examine the activities of these human microglia-specific promoters in primary microglia. In human primary microglial cells, all these promoters were highly active in gene transcription including hCD68-695 (**Fig. 6A and C**), which was expected. In rhesus macaque primary microglia, hHexB-183 has less promoter activity

while hHexB-134, hCD68-135, and hCD68-695 retained high transcription capacity (**Fig. 6B, D**). When tested in the human or rhesus macaque primary astrocytes, no luciferase activity were detectable in any of these microglia-specific promoters (**Fig. 6E-H**). CMV-Flag LG vector served as positive transfection controls (**Fig. 6E-H**). Again, CMV-driven gene transcription in astrocytes was also lower than in microglia due to lower transfection efficiency (**Fig. 6E-H**). Considering the transcriptional activity and optimal size for AAV packaging, these analyses indicate that hHexB-134 promoter can be further developed to serve as a candidate microglia-specific promoter for AAV gene therapy. Our data also expanded the previous findings of CD68 promoter as an appropriate promoter by identifying a new and much short version of human CD68 (hCD68-135), ideally for gene targeting in microglia cells in the future.

Discussion

With AAV delivery of gene editors as one of the foremost technologies in development for gene therapy of latent HIV reservoirs, finding an effective gene editing system is essential. This is particularly relevant to the HIV cure study targeting CNS reservoirs, as the current "shock and kill" strategy may induce high levels of neuron damage or other side effects (46, 47). One way is to develop an effective AAV serotype which can pass through BBB and/or deliver into cellular HIV reservoirs in the brain, such as microglia cells (45). Alternatively, finding an effective promoter to express gene editor specifically in the brain microglia is paramount to progress. Considering the size limitations for AAV packaging, it is even more important to identify a promoter that is both small and transcriptionally active in microglia. Combination of both approaches may achieve an effective HIV gene editing in the brain *via* not only penetrating BBB but also specific targeting into microglia, one of the major cellular reservoirs in the CNS (13, 48-50).

Microglia-specific gene promoters have been studied in the mouse microglia cell models using mouse myeloid cell-specific promoter including *CD68* promoter (16, 45); however, it is not clear whether it is effective in human primary microglial cells. While CD68 is considered as surface marker of myeloid cells, it remains to be determined whether it is specific to human microglia, considering the heterogeneity of myeloid cells in tissues. Fortunately, using scRNA-seq technology, recent studies have identified

several new microglia biomarkers, which have proven specific to brain microglia, including *TMEM119* and *HexB* (39, 40, 44). Among them, *HexB* expression is stable in the microglia *in vivo* even during the neuropathogenic condition, superior to other known microglia biomarkers (40, 44). In this study, we have exploited the promoter of this new biomarker for human microglia cells, HexB. By carrying multiple iterations of cloning, we were able to generate an array of *HexB* promoter vectors of varying size and derived from different species. We found that *HexB* promoter is specific to microglia cells, possibly in a cross-species manner. Our data support that *HexB* promoter may be appropriate to develop into a new gene editing system for a microglia specific targeting (**Figs. 3, 4 and 6**).

It was believed that longer endogenous promoters showed higher levels of gene transcription in human genome (51). Our studies confirmed that this is crucial in determining if *HexB* promoter is able to induce the expression of our dual LG reporter in primary human microglia cells (**Figs. 2 and 5**). In the initial study, mHexB-330, hHexB-183 and mkHexB-170 promoters showed the highest expression of luciferase in a model transformed cell line of HEK293T (**Fig. 2**). When comparing the human promoters of sizes 183 bp, 134 bp, 93 bp, and 43 bp, the distinction between the latter two and the former two was clear: the core promoters (93bp and 43bp) were ineffective in boosting luciferase expression over the promoterless vector (Empty) while the longer promoter that includes 5'-UTR was more active to induce transcription. It has been demonstrated that the essential sequences for *HexB* promoter activity are localized in the region between 150 bp and 90 bp upstream of the ATG codon (38). The pair of promoters containing 5'-UTR (134 bp and 183 bp), however, has proved to augment transcription activity, indicating that this 5'-UTR region is vital for the human *HexB* promoter activity in the induction of its target gene transcription. Therefore, the native promoter of *HexB* comprising of core promoter and 5'-UTR is essential and minimal in driving its transcription.

We also investigated *CD68* promoters. Initially, we found that 695 bp human *CD68* promoter, hCD68-695, was highly active in SIM-A9 mouse microglia and C20 human microglial cell lines. This was also true in the primary human or rhesus macaque microglia (**Figs. 3 and 6**). Surprisingly, the much shorter version of human *CD68* promoter, hCD68-135, was also active in transcription, although less active than hCD68-695, in microglia (**Figs. 3 and 6**). In comparison to hCD68 promoters, human *HexB* promoters, hHexB-

183 and hHexB-134, strongly induced both GFP and luciferase expression in the primary human microglia, which was comparable to the short version of hCD68 promoter hCD68-135, however, slightly lower than the much longer version of hCD68 promoter hCD68-695 (**Figs. 6A and C**).

While our data validated the previous observations of *CD68* promoter activity in mouse microglia, the size of *CD68* promoter is relatively big (695 bp), compared with *HexB* promoter (134 bp) or the short version of hCD68, hCD68-135, which may limit its application in the future. In another report, *F4/80* (667 bp), *CD68* (543 bp) and *CD11b* (496 bp) promoters have been shown some activities in AAV targeting in microglia cells. However, their promoter sizes are relative big (equal or >500 bp) (52) (**Figs. 6A and C**). These observations support that HexB-135 and CD68-135 promoters warrant further investigations for AAV microglia targeting.

Our study also showed that the dual LG reporter assay is powerful in the discovery of novel cell type-specific promoters. Given that this analysis was carried out in the 96-well plate format, it has the potential to be developed into a high throughput assay to screen promoter activity in cell type-dependent and possibly species-dependent manners, which is useful for the gene editing studies in the future.

Taken together, we characterized *HexB* gene promoter as a novel tool suitable for microglia specific targeting, which could be exploited for the development of novel gene editing vectors for the cure of HIV reservoirs in the CNS in the future.

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AUTHOR CONTRIBUTIONS

WH and GJ conceived the study. SS, KE, BB, SS, JT, ZW, XW and HW constructed the plasmids and performed the studies in HEK293, SIM-A9 and C20 microglia models. LMW and YT performed studies involved in human and rhesus macaque primary microglia and astrocytes. BL, DMM and JVGM provided necessary advice during the discussion of the

study. GJ and WH wrote the manuscript with multiple revisions. All authors read and approved the manuscript for submission.

DECLARATION OF INTERESTS

The authors declare no conflict of interest.

Figure Legends

Figure 1. LG dual reporter assays to measure *HexB* promoter activity. (A) Diagram for LG dual reporter assays driven by *HexB* promoter and representative nucleotide numbers for mouse HexB. (B) HEK293T cells were transfected with mouse 330 bp *HexB* promoter-driven dual LG reporter (mHexb-330) and no *HexB* 5'UTR-driven dual LG reporter (mHexB-no UTR) plasmid in 96-well plate, where Empty promoterless LG reporter vector served as negative control. At 48-72 hours after transfection, gdLuc levels in culture media were measured to determine the *HexB* promoter activities. (C) Representative images were taken to show GFP expression in HEK293T cells 72 hours after transfection. ***, $p<0.001$ analyzed with student's *t*-test compared with empty vector transfection control (n=4).

Figure 2. Activity of microglia-related promoters, including *HexB* and *CD68*, in the induction of gene transcription in HEK293T cells. (A) Different lengths of mouse, human or monkey *HexB* promoter-driven dual-LG reporters were transfected into HEK293T cells in 96-well plate. At 72 hours after transfection, gdLuc levels in culture media were measured to determine the *HexB* promoter activities. Cells transfected with plasmids driven by other microglia-related promoters, such as *CD68*, were also included, and activities of transcription were similarly measured by luciferase levels in the supernatants. (B) Representative images were shown after transfection. ****, $p<0.0001$, compared with empty vector transfection controls (One-way Annova, n=4). #, $p<0.05$; ##, $p<0.01$; ###, $p<0.001$, compared with indicated groups (*t*-test, n=4). m, mouse; h, human; mk, monkey.

Figure 3. Transcription activities of *HexB* promoters in mouse SIM-A9 and human C20 microglia cells. *HexB* promoter-driven dual LG reporters were transfected into mouse SIM-A9 (A) or human C20 (B) microglial cells for 72 hours. Then, the supernatants were collected and subjected for luciferase assays. *, $p<0.05$; **, $p<0.01$; compared with empty vector transfection control (One-way Annova, n=4). #, $p<0.05$; ##, $p<0.01$, compared with cells with indicated groups (*t*-test, n=4). m, mouse; h, human; mk, monkey.

Figure 4. *HexB* promoter induces reporter transcription mainly in primary microglia but not in astrocytes. Mouse, human or monkey *HexB* promoter-driven dual-LG reporters were transfected into human primary microglia (A, B) or human primary astrocytes (C, D) in 96-well plate. After transfection, GFP+ cells were observed with microscopy (A and C), and gdLuc levels (B and D) in culture media were measured to determine the *HexB* promoter activities. Similarly, *HexB* promoter-driven dual-LG reporters were transfected into primary rhesus macaque microglia (E, F) or primary rhesus macaque astrocytes (G, H) in 96-well plate. After transfection, GFP+ cells were observed with microscopy (E and G, 10x magnification), and gdLuc levels in culture media (F and H) were measured to determine the *HexB* promoter activities. *, $p<0.05$; **, $p<0.01$; ***, $p<0.001$; ****, $p<0.0001$, compared with empty vector transfection controls (One-way Annova, n=3).

Figure 5. Core *HexB* promoter fails to drive transcription in primary microglia. Similar to Fig. 4, dual LG reporter driven by hHexB-183, hHexB-134 or hHexB core promoters were transfected into human (A, C) or monkey (B, D) primary microglial cells in 96-well plate. These plasmids were similarly transfected in human (E and G) or monkey (F and H) primary astrocytes. At 48 hours post-transfection, GFP+ cells (10x magnification) and gdLuc levels were measured to determine the *HexB* promoter activities. **, $p<0.01$; ***, $p<0.001$; ****, $p<0.0001$, compared with empty vector transfection controls (One-way Annova, n=3).

Figure 6. Human *HexB* promoter induces gene transcription in comparison with other microglia gene promoters in primary human microglia and astrocytes.

Microglia-specific promoter-driven dual-LG reporters were transfected into human primary microglia (A and C) or human primary astrocytes (E and G) in 96-well plate. After transfection, GFP+ cells were observed with microscopy (A and E), and gdLuc levels (C and G) in culture media were measured to determine the *HexB* promoter activities. Similarly, promoter-driven dual-LG reporter were transfected into primary rhesus macaque microglia (B and D) or primary rhesus macaque astrocytes (F and H) in 96-well plate. GFP+ cells were observed with microscopy (B and F, 10x magnification), and gdLuc levels in culture media (D and H) were measured to determine the *HexB* promoter activities. *, $p<0.05$; **, $p<0.01$; ***, $p<0.001$; ****, $p<0.0001$; compared with empty vector transfection controls (One-way Annova, n=3). #, $p<0.05$; ##, $p<0.01$ (*t-test*, n=3).

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Fig. 1

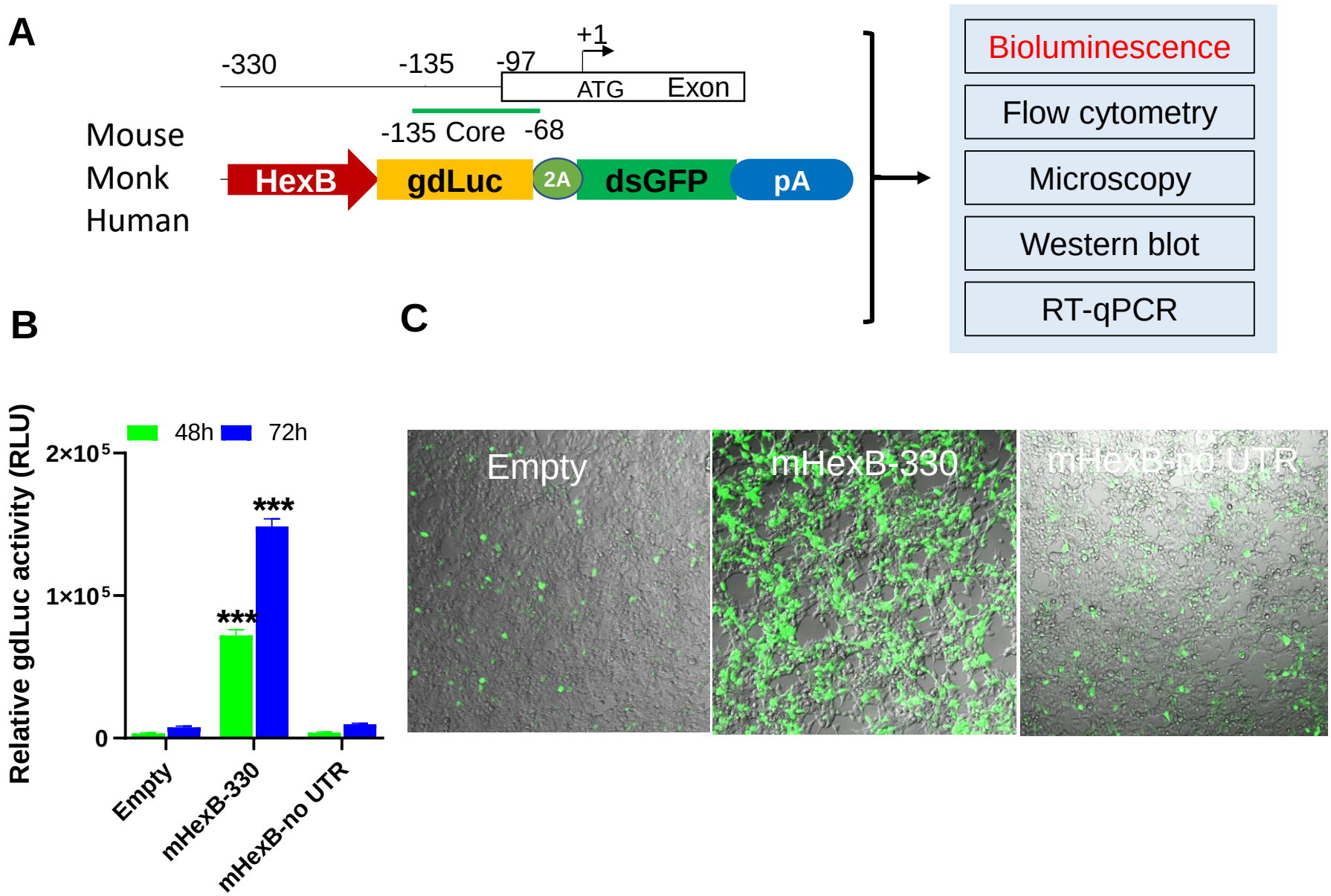
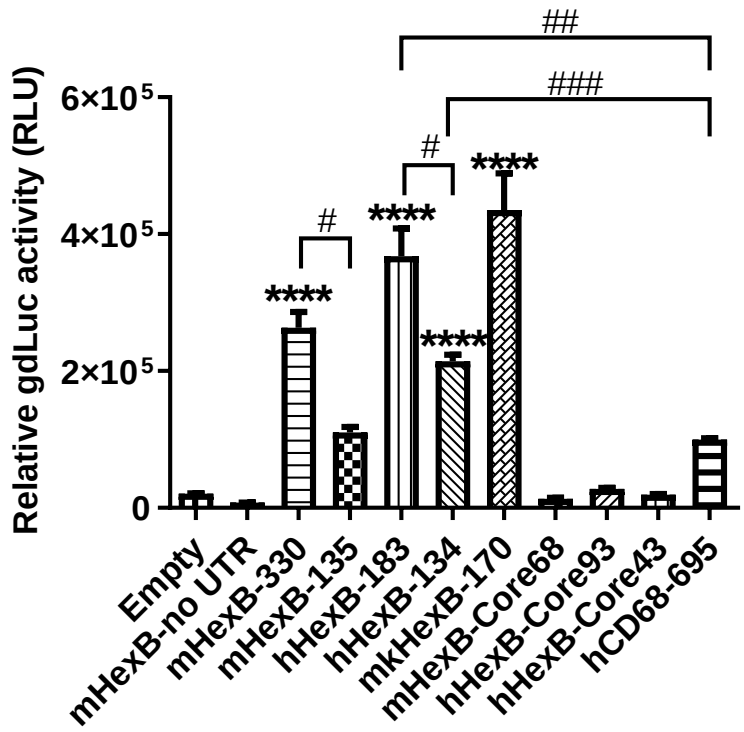

A
-330
-135
-97
+1
ATG
Exon
-135 Core
-68
Mouse
Monk
Human
HexB
gdLuc
2A
dsGFP
pA
Bioluminescence
Flow cytometry
Microscopy
Western blot
RT-qPCR
B
48h
72h
Relative gdLuc activity (RLU)
2×10⁵
1×10⁵
0
***
***
Empty
mHexB-330
mHexB-no UTR
C
Empty
mHexB-330
mHexB-no UTR

Fig. 2

A


Relative gdLuc activity (RLU)
6×10⁵
4×10⁵
2×10⁵
0
##
###
#
#
****
****
****
****
Empty
mHexB-no UTR
mHexB-330
mHexB-135
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hHexB-134
mkHexB-170
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hHexB-Core93
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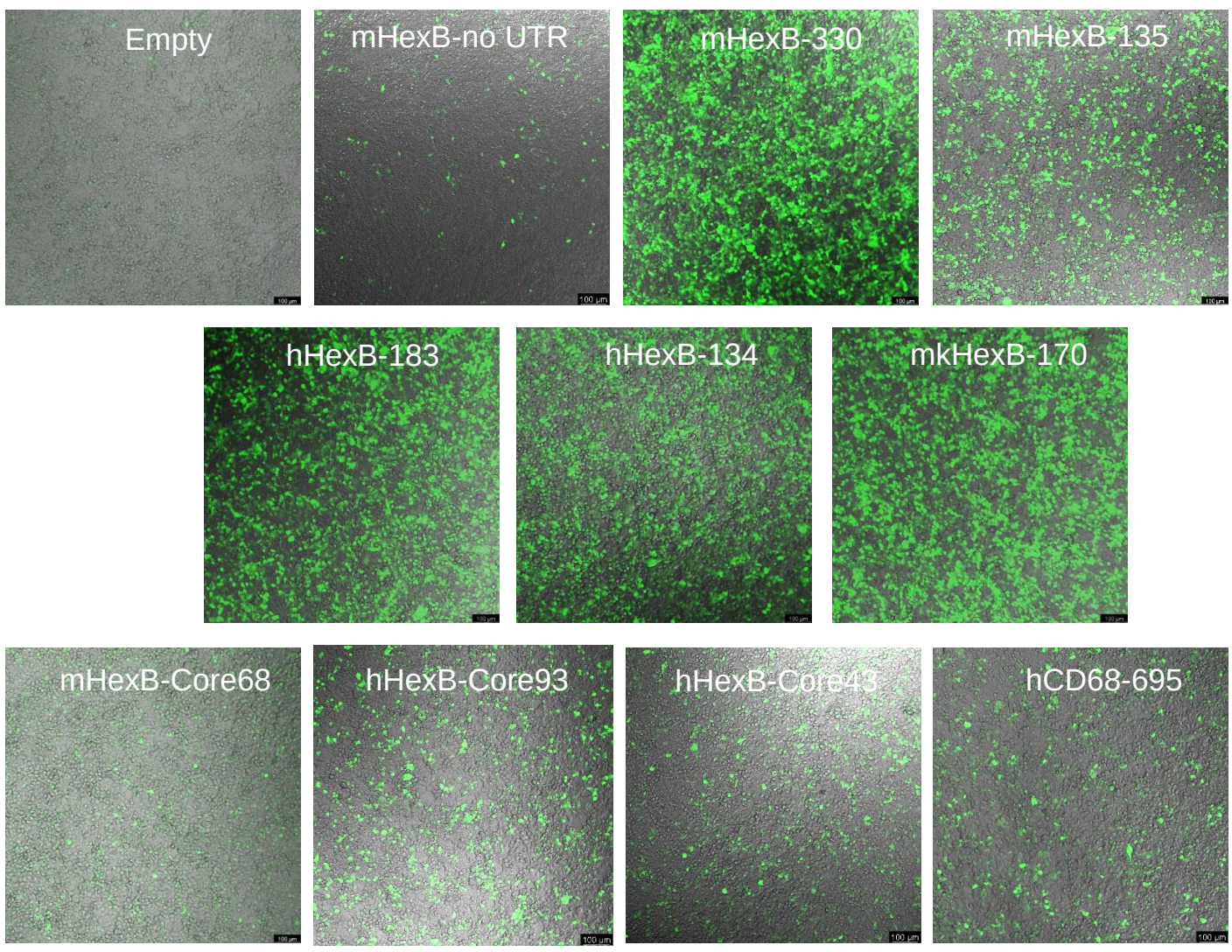
hCD68-695

B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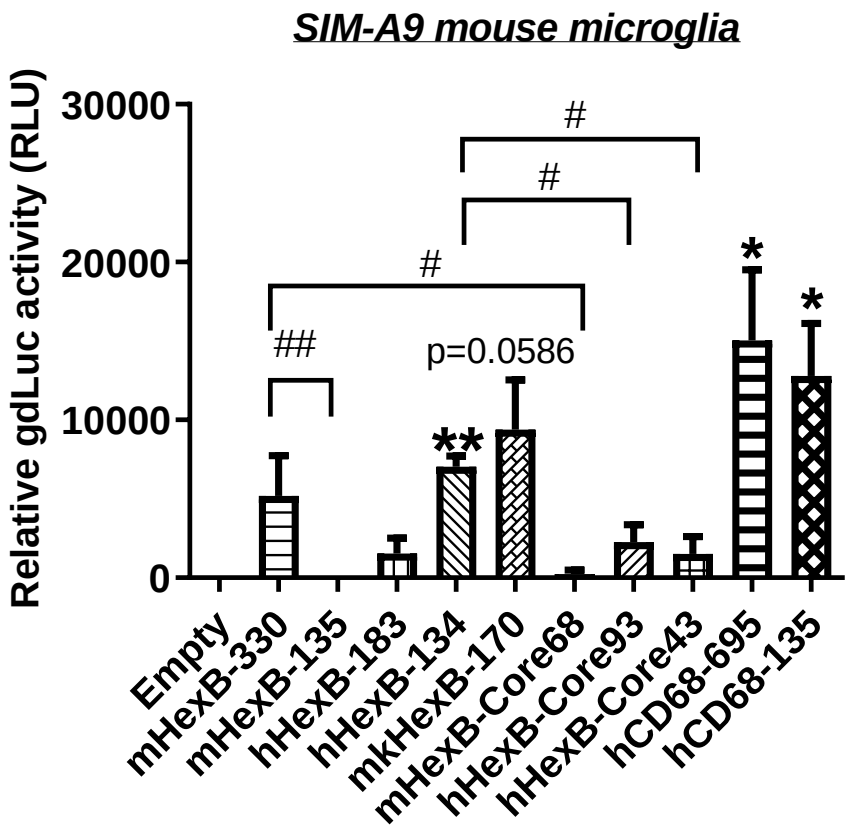
hHexB-Core93
hHexB-Core43
hCD68-695

Fig. 3

A


SIM-A9 mouse microglia
Relative gdLuc activity (RLU)
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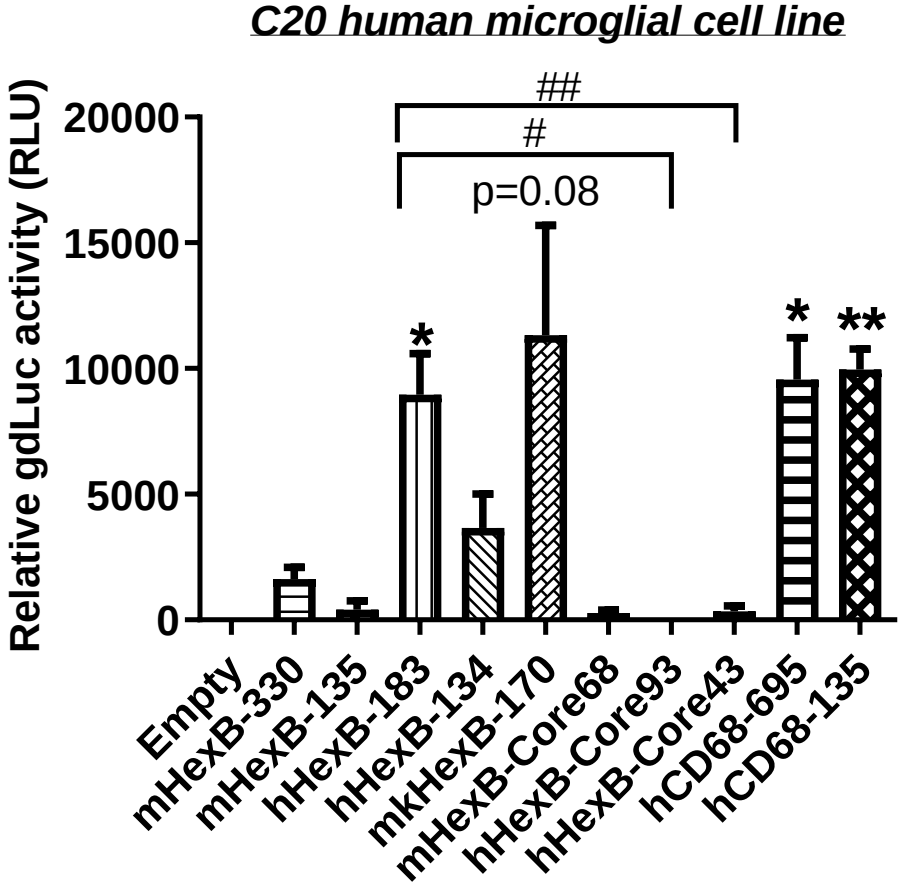
20000
10000
0
#
#
#
##
p=0.0586
**
*
*
Empty
mHexB-330
mHexB-135
hHexB-183
hHexB-134
mkHexB-170
mHexB-Core68
hHexB-Core93
hHexB-Core43
hCD68-695
hCD68-135

B


C20 human microglial cell line
Relative gdLuc activity (RLU)
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15000
10000
5000
0
##
#
p=0.08
*
*
**
Empty
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mHexB-135
hHexB-183
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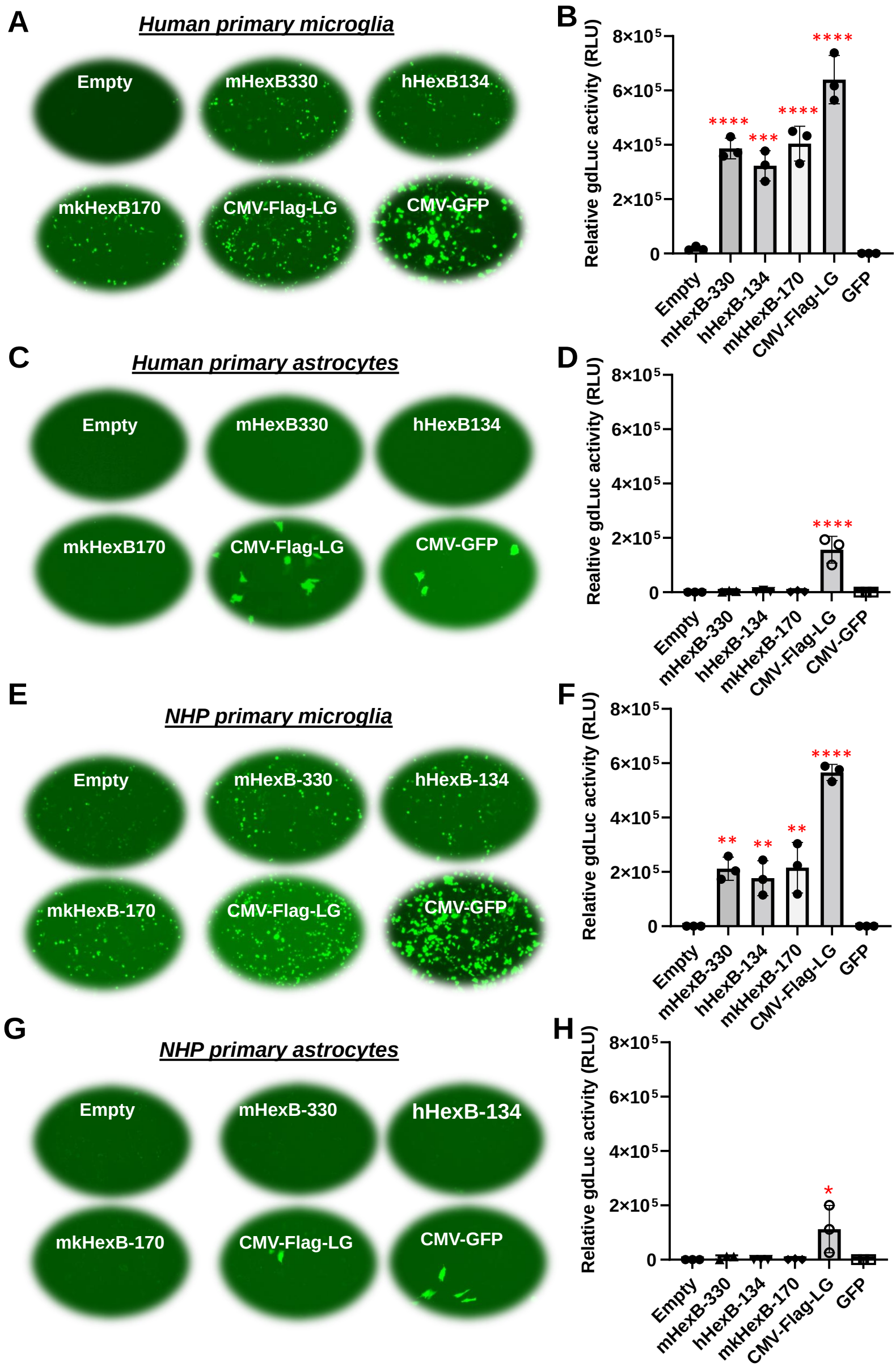
hHexB-Core93
hHexB-Core43
hCD68-695
hCD68-135

Fig. 4


A
Human primary microglia
Empty
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hHexB134
mkHexB170
CMV-Flag-LG
CMV-GFP
B
Relative gdLuc activity (RLU)
8×10⁵
6×10⁵
4×10⁵
2×10⁵
0
****
***
****
****
Empty
mHexB-330
hHexB-134
mkHexB-170
CMV-Flag-LG
GFP
C
Human primary astrocytes
Empty
mHexB330
hHexB134
mkHexB170
CMV-Flag-LG
CMV-GFP
D
Realtive gdLuc activity (RLU)
8×10⁵
6×10⁵
4×10⁵
2×10⁵
0
****
Empty
mHexB-330
hHexB-134
mkHexB-170
CMV-Flag-LG
CMV-GFP
E
NHP primary microglia
Empty
mHexB-330
hHexB-134
mkHexB-170
CMV-Flag-LG
CMV-GFP
F
Relative gdLuc activity (RLU)
8×10⁵
6×10⁵
4×10⁵
2×10⁵
0
**
**
**
****
Empty
mHexB-330
hHexB-134
mkHexB-170
CMV-Flag-LG
GFP
G
NHP primary astrocytes
Empty
mHexB-330
hHexB-134
mkHexB-170
CMV-Flag-LG
CMV-GFP
H
Relative gdLuc activity (RLU)
8×10⁵
6×10⁵
4×10⁵
2×10⁵
0
*
Empty
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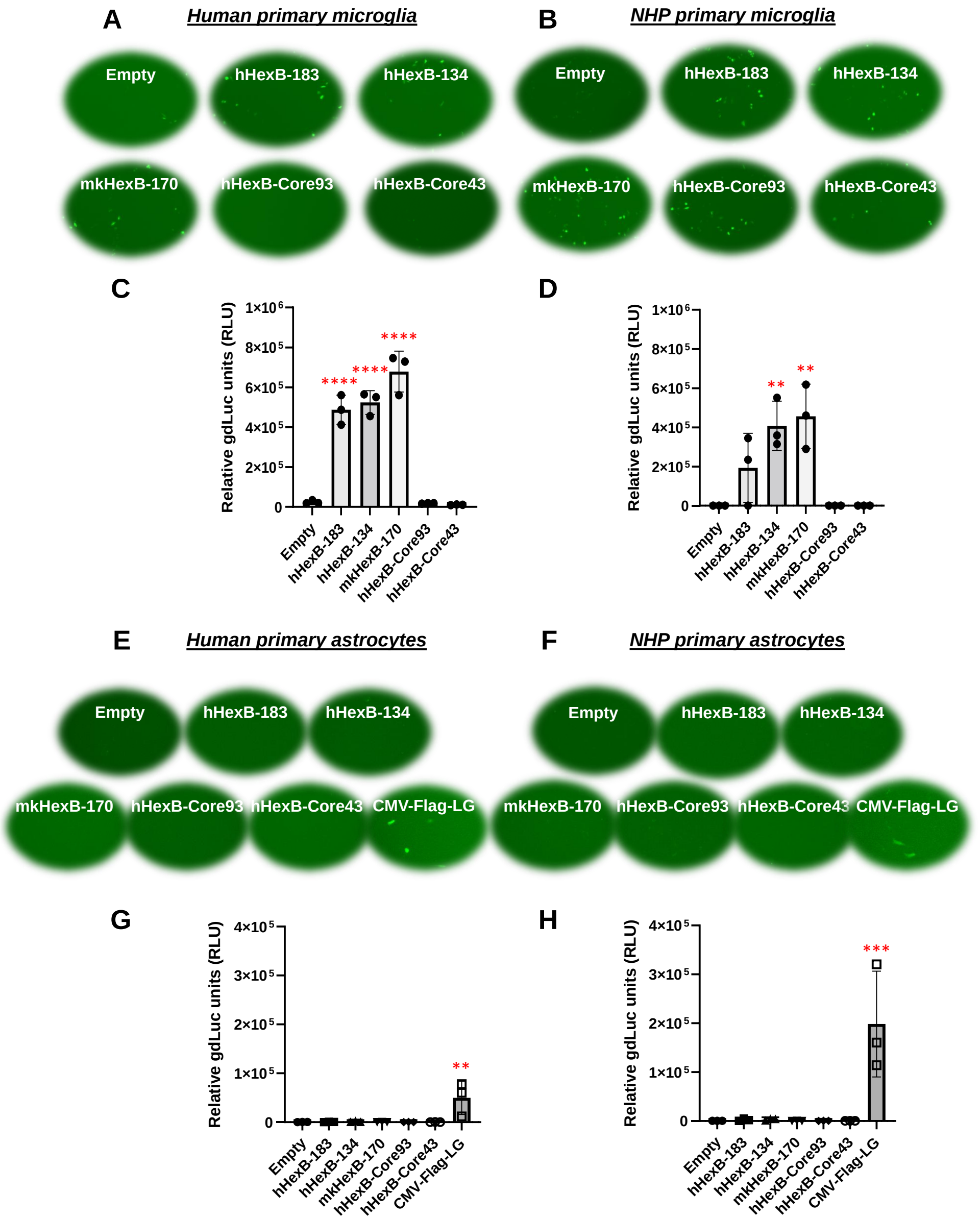
hHexB-134
mkHexB-170
CMV-Flag-LG
GFP

Fig. 5


A
Human primary microglia
Empty
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hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
B
NHP primary microglia
Empty
hHexB-183
hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
C
Relative gdLuc units (RLU)
1×10^6
8×10^5
6×10^5
4×10^5
2×10^5
0
****
****
****
Empty
hHexB-183
hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
D
Relative gdLuc units (RLU)
1×10^6
8×10^5
6×10^5
4×10^5
2×10^5
0
**
**
Empty
hHexB-183
hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
E
Human primary astrocytes
Empty
hHexB-183
hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
CMV-Flag-LG
F
NHP primary astrocytes
Empty
hHexB-183
hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
CMV-Flag-LG
G
Relative gdLuc units (RLU)
4×10^5
3×10^5
2×10^5
1×10^5
0
**
Empty
hHexB-183
hHexB-134
mkHexB-170
hHexB-Core93
hHexB-Core43
CMV-Flag-LG
H
Relative gdLuc units (RLU)
4×10^5
3×10^5
2×10^5
1×10^5
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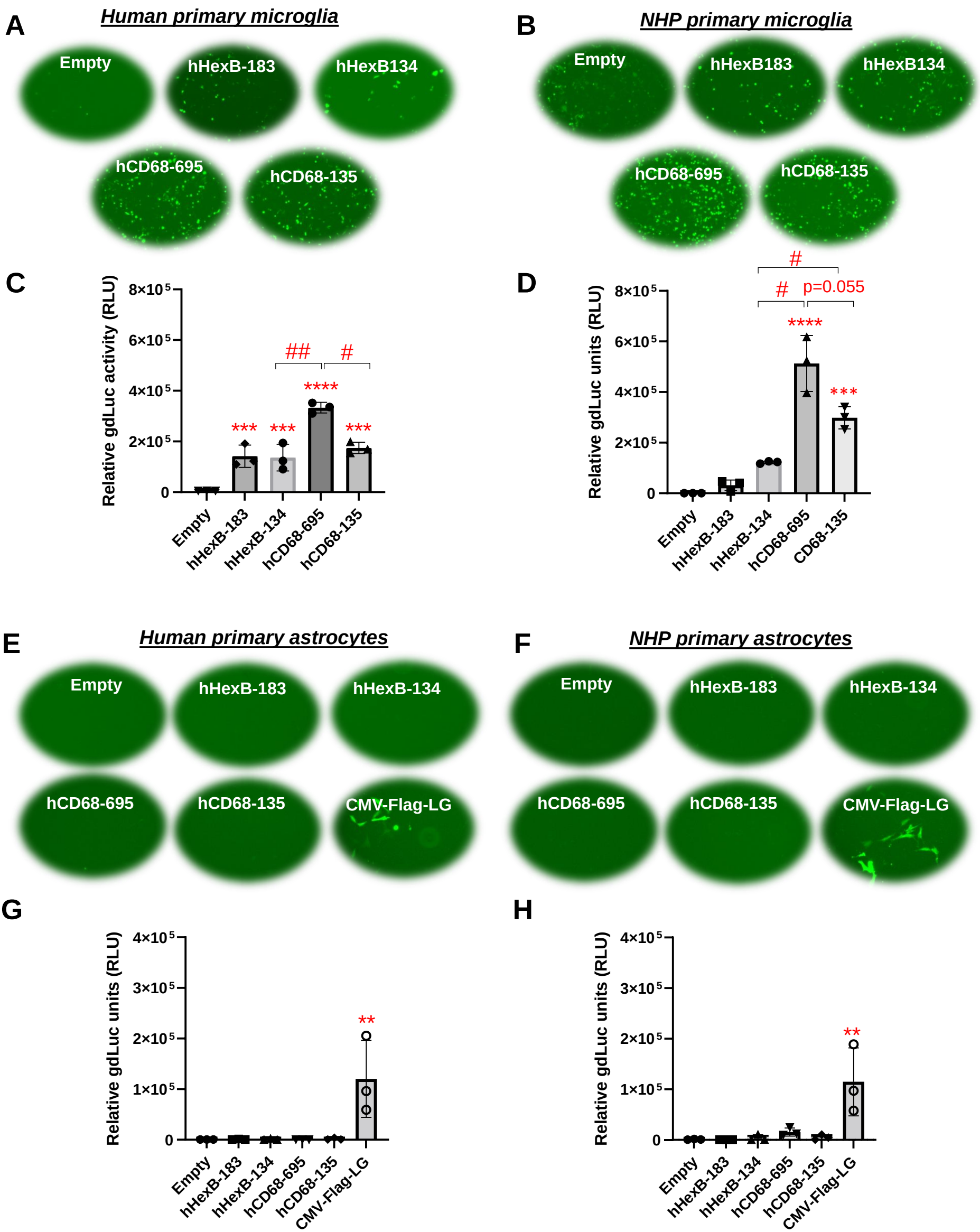
hHexB-Core93
hHexB-Core43
CMV-Flag-LG

Fig. 6


A
Human primary microglia
Empty
hHexB-183
hHexB134
hCD68-695
hCD68-135
B
NHP primary microglia
Empty
hHexB183
hHexB134
hCD68-695
hCD68-135
C
Relative gdLuc activity (RLU)
8×10⁵
6×10⁵
4×10⁵
2×10⁵
0
***
***
****
***
##
#
Empty
hHexB-183
hHexB-134
hCD68-695
hCD68-135
D
Relative gdLuc units (RLU)
8×10⁵
6×10⁵
4×10⁵
2×10⁵
0
#
#
p=0.055
****
***
Empty
hHexB-183
hHexB-134
hCD68-695
CD68-135
E
Human primary astrocytes
Empty
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hHexB-134
hCD68-695
hCD68-135
CMV-Flag-LG
F
NHP primary astrocytes
Empty
hHexB-183
hHexB-134
hCD68-695
hCD68-135
CMV-Flag-LG
G
Relative gdLuc units (RLU)
4×10⁵
3×10⁵
2×10⁵
1×10⁵
0
**
Empty
hHexB-183
hHexB-134
hCD68-695
hCD68-135
CMV-Flag-LG
H
Relative gdLuc units (RLU)
4×10⁵
3×10⁵
2×10⁵
1×10⁵
0
**
Empty
hHexB-183
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hCD68-695
hCD68-135
CMV-Flag-LG