Evolutionary history limits species' ability to match color sensitivity to available habitat light

Matthew J. Murphy^{1*†} and Erica L. Westerman^{1‡§}

¹Department of Biological Sciences, University of Arkansas, Fayetteville, AR 72701

*Author for correspondence: mjm052@uark.edu

[†] ORCiD: 0000-0002-2683-3348

[‡]ewesterm@uark.edu

[§] ORCiD: 0000-0002-3575-8298

1 Abstract

2 The spectrum of light that an animal sees – from ultraviolet to far red light – is governed by the 3 number and wavelength sensitivity of a family of retinal proteins called opsins. It has been 4 hypothesized that the spectrum of light available in an environment influences the range of 5 colors that a species has evolved to see. However, invertebrates and vertebrates use 6 phylogenetically distinct opsins in their retinae, and it remains unclear whether these distinct 7 opsins influence what animals see, or how they adapt to their light environments. Systematically 8 utilizing published visual sensitivity data from across animal phyla, we found that terrestrial 9 animals are more sensitive to shorter and longer wavelengths of light than aquatic animals, and 10 that invertebrates are more sensitive to shorter wavelengths of light than vertebrates. Controlling 11 for phylogeny removes the effects of habitat and lineage on visual sensitivity. Closed and open 12 habitat terrestrial species have similar spectral sensitivities when comparing across the Metazoa, 13 and deep water animals are more sensitive to shorter wavelengths of light than shallow water 14 animals. Our results suggest that animals do adapt to their light environment, however the 15 invertebrate-vertebrate evolutionary divergence has limited the degree to which animals can 16 perform visual tuning.

17 *Keywords:* visual tuning, light environment, ciliary opsin, rhabdomeric opsin, λ_{max}

18

19 1. Introduction

20 Animals use vision for many tasks, including finding prey, avoiding toxic animals and plants, 21 identifying predators, assessing mate quality, and navigating their environment [1–5]. In many 22 cases, the objects of interest to the animal need to be distinguished from the background [6,7]. For 23 example, food that does not contrast with the background is harder for foragers to detect than food 24 that does contrast with the background [8–12]. Signals that contrast with background colours and 25 patterns are also used for mating displays [13–18]. Furthermore, many species' body colour 26 patterns have evolved to be simultaneously cryptic to predators while conspicuous to intended 27 receivers [19–21]. Finally, contrasting colours can improve animals' ability to learn the meaning 28 of signals, as when chicks learn more quickly to avoid bitter, aposematically coloured food 29 [2,3,22].

30

An animal's ability to detect visual information depends upon the colour and amount of light in its habitat, otherwise known as the *light environment* [6,23,24]. For example, red and blue light are filtered out by chloroplasts, lending forests and estuarine environments a yellow-green cast [24– 26]. Likewise, the water column progressively filters red and UV light [23,24]. Animals' signalling behaviours, choice of microhabitat, and visual physiology are thus expected to co-evolve to suit their light environment [6].

37

38 Sighted species' photoreceptors (the light-absorbing neurons which enable vision) are theorized to
39 have undergone adaptation to best absorb the light most often present in their environments [23].

40 This process, called *visual tuning*, is made possible by both filtering pigments [27–35] and 41 differences in the amino acid sequence or 3-dimensional shape of photosensitive proteins called 42 opsins [23]. Visual tuning has been found to shift wavelengths of maximum sensitivity in species 43 as diverse as birds, fish, and mammals [23,36,45,46,37-44]. Although the effect of light 44 environment on vision has been extensively studied in fish [47,48], a systematic study of visual 45 tuning in terrestrial animals has not yet occurred; neither have terrestrial animals been 46 systematically compared to aquatic species. Both aquatic and terrestrial animals are found in a 47 variety of light environments, and multiple phyla have independently made the water-to-land 48 habitat transition. Additionally, studies of animals which transition from aquatic larvae to 49 terrestrial adults have found that these species change their visual pigment expression patterns in 50 a manner that matches their changing light environment [49–53]. Understanding whether 51 phylogeny constrains the extent of visual tuning, particularly during these water-to-land 52 transitions, is critical for understanding the evolutionary ecology of animal vision.

53

54 If opsin tuning faces phylogenetic constraints, the evolutionary history of animal vision may have 55 shaped the degree to which different phyla have adapted to their light environments. The types of 56 opsins differ between chordates and other phyla [54]. Chordates use c-opsins in cilia-bearing 57 photoreceptors to transduce photons into vision, while non-chordate animals use r-opsins in 58 rhabdomere-bearing photoreceptors; no animals have been identified that use both c- and r-type 59 photoreceptors for vision [55]. Phylogenetic analyses reveal that c- and r-opsins diverged 400 60 million years ago and were likely both present in the urbilaterian, with r-opsins closely related to 61 the melanopsins used by chordates for non-visual tasks [55]. The r-opsin/ c-opsin divergence may 62 have given rise to different degrees of tuning between chordates and non-chordates.

63

64	The diverse habitats in which animals live, combined with the long evolutionary history of visual
65	pigments, leads to several questions. First, have transitions from aquatic to terrestrial habitats
66	influenced the spectra of light that animals can see? And, are differences in the spectra that animals
67	can see associated with the c-opsin/ r-opsin divergence? Second, do animals that live in visual
68	environments that filter red and blue light, such as closed-canopy forests and estuarine habitats,
69	see colours more similarly to each other than to open terrestrial or freshwater aquatic
70	environments, in which colours are less (if at all) strongly filtered? And, if there is an effect of
71	habitat greenness, is this effect outweighed by phylogeny?
72	
73	To answer these questions, we performed a phylogenetically weighted systematic analysis of the
74	maximum and minimum wavelength of visual sensitivity, as well as the range of visual sensitivity,
75	across animals.

76

77 (2) Materials and method

78Paper selection:
We conducted Google Scholar searches in October 2017 and January 2018. Our79first search used the search pattern "visual pigment" OR opsin OR "absorbance spectrum" " λ 80max" -human -man -men -woman -women - "Homo sapiens" -disease -regeneration. We81conducted a second Google Scholar search using the search pattern visual pigment, opsin82sensitivity, absorbance spectrum. For both searches, we excluded citations and patents.

We reviewed candidate articles using a three-step process. First, we screened by title and abstract to identify original research articles and review papers that examined animal visual physiology. We then screened articles to determine if they used microspectrophotometry, electrophysiology, pigment extraction, or *in vitro* mRNA expression followed by spectrophotometry, and that they measured visual sensitivity or visual pigment absorption from at least two animals. Finally, we only kept articles which used animals that were wild-caught or reared in full-spectrum light conditions, to avoid any effects of artificial lighting on visual sensitivity [56,57].

91

For review articles, we determined whether the authors had included measurements of the mean wavelength of peak sensitivity (λ_{max}) of some population in the article's figures or tables. We downloaded the corresponding primary sources and filtered them using the process described above.

96

97 <u>*Visual pigment sensitivity data*</u>: We recorded the following data for each species of each paper that 98 passed our filters: 1) mean wavelength of peak sensitivity (λ_{max}) for each visual pigment measured; 99 2) number of animals measured (*n*); 3) standard deviation of the mean λ_{max} (SD) (when available); 100 and 4) where animals were caught (when available). We calculated sampling error for visual 101 pigments when possible.

102

Habitat data: We used standardized data sources to classify each species by habitat. Sources
 included field guides [58–60], public databases (BugGuide, <bugguide.net>, Butterflies and Moths
 of North America, <butterfliesandmoths.org>, FishBase <fishbase.org>, SealifeBase

106 <sealifebase.org>, IUCN Redlist <iucnredlist.org>) and online encyclopaedias including Animal 107 Diversity Web (<animaldiversity.org>) and Encyclopedia of Life (<eol.org>). After first 108 classifying species as terrestrial or aquatic, we then defined terrestrial sub-habitats: rainforest, 109 forest, woodland, shrubland, grassland, and desert. We recategorized these habitats into three 110 habitat types based on canopy density. Rainforest and temperate forest were designated as "closed" 111 habitats. Woodland was considered to have "intermediate" canopy density [25]. Shrubland, 112 grassland, and desert were classified as "open" habitats.

113

114 Aquatic habitats included river, stream, pond, lake, coastal, estuarine, open-water marine, bottom-115 dwelling marine, abyssopelagic, abyssodemersal, bathypelagic, and bathydemersal habitats. We 116 recategorized these habitats into two habitat types based on salinity. River, stream, pond, and lake 117 habitats were considered "freshwater" habitats; while coastal, estuarine, open-water marine, and bottom-living marine habitats were "marine" habitats. Animals considered "coastal" were those 118 119 described as living in water along the coast, near shore, or in estuaries. We also recategorized these 120 habitats into two habitat types based on whether light was abundant or not. Abyssopelagic, 121 abyssodemersal, bathypelagic, and bathydemersal habitats receive little or no sunlight due to their 122 depth in the water column and were considered "lightless" habitats. Species that were considered 123 by our sources as deep-water species were also considered species that lived in "lightless" habitats. 124 All other habitats were considered "lit" habitats. Finally, we used FishBase, SealifeBase or field 125 guides to identify the minimum and maximum depths for each species. We then used these data to 126 calculate average depth per species ($D_{average} = (D_{max} + D_{min}) * 2^{-1}$).

127

128 <u>Phylogenetic control:</u>

129 To control for the effect of evolutionary relatedness on visual sensitivity we built a phylogenetic 130 tree of all animals in our analysis (see the electronic supplementary material: figure S1). We used 131 the function thrs match names in the R package rotl [61] to acquire data from the Open Tree of 132 Life database (<tree.opentreeoflife.org>) for each of the species represented in our regression, and 133 to generate a phylogenetic tree using default arguments and excluded species flagged as *incertae* 134 *cedis*; i.e., with uncertain phylogenetic position (25 species) and species which had no sequencing 135 data in the Open Tree of Life database (6 species). We created an induced subtree with the resulting 136 data using the function tol induced subtree in the package *rotl*.

137

Trees produced using *rotl* are unrooted, without branch lengths, and sometimes with unresolved polytomies. We used the R packages *phytools* [62] and *ape* [63] to resolve these issues. We used the root function in *ape* to root the tree using *Saccharomyces cerevisiae* (ottid: 5262624) from <tree.opentreeoflife.org>) as the outgroup. We computed branch lengths using the compute.brlen function in *ape* with default arguments. Finally, we randomly resolved polytomies using the multi2di function in *ape* with default parameters. Subtrees of the primary tree were constructed as needed using the drop.tips function in *ape*.

145

146 <u>Statistical Analyses:</u>

147 To determine whether longest λ_{max} , shortest λ_{max} , and range of λ_{max} followed the normal 148 distribution, we used the shapiro.test function in R. To determine whether the variances of longest

149 λ_{max} , shortest λ_{max} , and range of λ_{max} differed between broad habitat type (aquatic or terrestrial) or 150 lineage (invertebrates or vertebrates) we used levene.test function in R.

151

152 To determine whether there was an effect of broad habitat type or lineage on the longest λ_{max} , 153 shortest λ_{max} , and range of λ_{max} , we used the glm function in R to construct generalized linear 154 models with the formula $\lambda_{max} \sim$ broad habitat * lineage.

155

To determine whether phylogeny could explain extant differences in longest λ_{max} , shortest λ_{max} , and range of λ_{max} between broad habitat type or lineage, we constructed phylogenetically controlled linear models using the phylolm function in the *phylolm* [64] package for R with the formula $\lambda_{max} \sim$ broad habitat * lineage with a bootstrap of 100. Since we had to exclude 31 species from our phylogenetic tree, we first ran the glm described above with the trimmed data set, and then compared those results to the results of our phylolm models. For these models, we used the overall phylogenetic tree and our trimmed dataset.

163

We then subset our overall dataset for terrestrial animals and aquatic animals, re-tested for normal distributions and variances, and conducted a set of statistical analyses specific to terrestrial or aquatic animals. To determine whether there was an effect of terrestrial habitat type (closed, intermediate, or open) or lineage on longest λ_{max} , shortest λ_{max} , and range of λ_{max} , we used the glm function with the formula $\lambda_{max} \sim$ terrestrial habitat type * lineage. However, since there were only 2 species that were intermediate habitat specialists, and they had similar visual spectra to those in closed canopies, we combined closed and intermediate habitat treatments into a single treatment,

171 closed_intermediate, and re-ran the generalized linear models described above using the new172 habitat treatment levels (closed intermediate vs open).

173

174 We examined the effects of depth and habitat on visual sensitivities of aquatic animals. To determine whether there were effects of minimum, maximum, or average depth of habitat on 175 176 longest λ_{max} , shortest λ_{max} , and range of λ_{max} among aquatic species, we constructed linear models 177 using the lm command with the formula $\lambda_{max} \sim$ depth. To determine whether phylogeny could 178 explain extant differences in minimum, maximum, or average depth of habitat among longest λ_{max} , 179 shortest λ_{max} , and range of λ_{max} among aquatic species, we first re-ran the linear models without 180 the species excluded from our phylogenetic tree, and then constructed phylogenetically controlled 181 linear models using the phylolm function in the *phylolm* package for R with the formula $\lambda_{max} \sim$ 182 depth with a bootstrap of 100. For these models, we used a subtree of our overall phylogenetic tree 183 (see above), which omitted all terrestrial species.

184

185 Finally, we subset our overall dataset for open terrestrial animals and non-deep water aquatic 186 animals (both freshwater and coastal) and conducted the following analyses. To assess whether 187 the visual systems of animals in open terrestrial habitats were more similar to the visual systems 188 of animals in open water habitats (coastal or freshwater) than those of closed terrestrial habitats 189 we compared the longest λ_{max} , shortest λ_{max} , and range of λ_{max} , of species in coastal-aquatic, 190 freshwater-aquatic, terrestrial-closed, and terrestrial-open habitats using the kruskal.wallis 191 function in R. Following this, we performed a pairwise (Steel-Dwass) test using the dscfAllPairs 192 function in the R package PCMCRplus [65]. To determine whether phylogeny could explain extant 193 differences in longest λ_{max} , shortest λ_{max} , and range of λ_{max} between animals living in these four 194 habitat types we re-ran the above analyses only using species in our phylogenetic tree, and then 195 constructed phylogenetically controlled linear models using the phylolm function in the *phylolm* 196 package for R with using the following formula with a bootstrap of 100: $\lambda_{max} \sim \text{coastal-aquatic} +$ 197 freshwater-aquatic + terrestrial-closed and $\lambda_{max} \sim \text{coastal-aquatic} +$ freshwater-aquatic + terrestrial-198 open.

199

200 (3) Results

Our dataset included 1,114 opsins from 446 species, extracted from a total of 156 articles (See the electronic supplementary materials: table S1). Of these, 868 opsins were recorded from 355 aquatic species, and 246 opsins were recorded from 91 terrestrial species. Our data were not normally distributed (Shapiro-Wilk test: longest λ_{max} : p < 0.05, W = 0.94; shortest λ_{max} : p < 0.05, W = 0.83; range of λ_{max} : p < 0.05, W = 0.70). Shortest, but neither longest nor range of λ_{max} were found to have equal variances when compared across broad habitat and lineage (Levene's test: longest λ_{max} : p = 0.16, t = 1.74; shortest λ_{max} : p < 0.05, t = 25.28; range of λ_{max} : p < 0.05, t = 21.62).

208

209 (a) <u>Terrestrial species were maximally sensitive to longer wavelengths of light than aquatic</u> 210 <u>species</u>

211 Terrestrial species were maximally sensitive to longer wavelengths of light than aquatic species,

- independent of opsin type (GLM, n=433: habitat $p = 3.83*10^{-8}$, t = 5.600, lineage p = 0.309, t = -
- 213 1.019; interaction: p = 0.595, t = -0.532; λ_{max} longest long-wavelength terrestrial species: 535±41.6

nm, aquatic species: 506±30.6 nm, invertebrate species: 513±38.9 nm, vertebrate species:
512±33.0 nm) (figure 1a).

216

217 (b) <u>Terrestrial species saw shorter wavelengths of light than aquatic species</u>

218 Terrestrial species were maximally sensitive to shorter wavelengths of light than aquatic species, 219 but there was a significant interaction between habitat and lineage: aquatic vertebrate species were 220 more sensitive to short wavelengths than aquatic invertebrate species, but terrestrial invertebrate 221 species were more sensitive to short wavelengths that terrestrial vertebrate species. Additionally, 222 invertebrates trended towards seeing short wavelengths of light (GLM, n = 433: habitat p = 0.045, t = -2.012, lineage p = 0.051, t = 1.960, interaction: p = $2.34*10^{-3}$; t = -3.061; λ_{max} shortest short-223 224 wavelength terrestrial species: 442 ± 79.2 nm, aquatic species: 476 ± 39.3 nm; invertebrate species: 225 466±70.2 nm, vertebrate species: 471±41.3 nm) (figure 1b).

226

(c) <u>Terrestrial species and invertebrates saw a larger range of colours than aquatic species</u> and vertebrates

Terrestrial species saw a larger range of colours than aquatic species. In addition, there was a significant interaction between habitat and lineage: aquatic invertebrates saw a narrower range of colours than aquatic vertebrates, but terrestrial invertebrates saw a broader range of colours than terrestrial vertebrates (GLM, n = 443: habitat p = $2.51*10^{-6}$, t = 4.772, lineage p = 0.03, t = -2.184, interaction: p = 0.00261; t =2.232; λ_{max} range terrestrial species: 92 ± 85.6 nm, aquatic species: 30.9 ± 51.6 nm; invertebrate species: 47.2 ± 80.4 nm, vertebrate species: 40.7 ± 56.7 nm) (figure 1c).

235 (d) <u>Accounting for phylogeny removes the effect of habitat and lineage on visual pigment</u> 236 sensitivity

When we ran our analyses with only the subset of species included in our phylogenetic tree, we found that we lost the effect of lineage on the range of visual pigment sensitivity but did not lose the effect of lineage on the longest or shortest wavelengths of maximum sensitivity (see the electronic supplementary materials: tables S2 and S3). With this in mind, we controlled for phylogeny. Controlling for phylogeny removed the effect of habitat and lineage on longest λ_{max} , shortest λ_{max} and range of λ_{max} (table 1).

Table 1. No effect of habitat or lineage on visual sensitivity following phylogenetic control.										
	Variable	Longest λ_{max}			She	ortest λ_m	ax	λ_{max} Range		
		р	t	SE	р	t	SE	р	t	SE
	Habitat	0.257	-1.13	11.4	0.439	0.774	17.0	0.226	-1.211	21.7
	Lineage	0.953	0.058	12.5	0.940	0.075	18.5	0.978	-0.028	23.7
Habit	at * Lineage	0.474	-0.72	20.3	0.214	-1.25	30.1	0.552	0.596	38.6

243

244 (e) *Forest-woodland and open habitat species have similar spectral sensitivities*

245 There was no effect of tree canopy openness on λ_{max} , shortest λ_{max} , and range of λ_{max} (table 2; see

the electronic supplementary materials: figure S2, table S4 and table S5).

Table 2. Effect of habitat greenness on shortest λ_{max} . P-values are above diagonal; q-values are						
below diagonal. Numbe	below diagonal. Numbers in bold are statistically significant.					
p p		Forest +		Open	Mean	SD
q	Coastal	Intermediate	Freshwater	Terrestrial	(nm)	(nm)
Coastal		5.75*10 ⁻⁷	0.903	3.92*10⁻⁶	254.4	55.2
Forest + Intermediate	-7.54		2.62*10 ⁻⁵	0.24	470.4	68
Freshwater	-0.970	-6.49		1.92*10 ⁻⁵	261.1	56.5
Open Terrestrial	-8.26	-2.64	-7.034		423.5	82.1

(f) <u>Average and maximum depth, but not minimum depth, influenced sensitivity to blue but</u> not red light

250 Species living at deeper average depth had longer shortest λ_{max} than shallow-living species (p = 251 0.046, t = 2.03; see the electronic supplementary material: figure S3a). Additionally, species 252 living at a deeper maximum depth had shortest λ_{max} that were longer than shallow-living species 253 shortest λ_{max} (p = 0.033, t = 2.18; electronic supplementary material: figure S4). Average depth 254 and minimum depth did not affect species' longest λ_{max} or range of λ_{max} , and there was no effect 255 of maximum depth on spectral sensitivity (electronic supplementary material: figures S3b-c and 256 S5). Controlling for phylogeny removes the effect of average depth on shortest λ_{max} (phylolm: t = $-4.688*10^{-1}$, p = 0.641). 257

258

(g) <u>Animals in coastal and freshwater habitats saw shorter wavelengths while animals in</u> <u>forest+intermediate or open-canopy habitats saw longer wavelengths</u>

We found that coastal animals' and freshwater animals' shortest λ_{max} were shorter than both forest+intermediate animals' shortest λ_{max} and open terrestrial animals' shortest λ_{max} (omnibus test: $p = 1.359*10^{-12}$, $\chi^2 = 53.296$, df = 3, pair-wise comparisons: table 2; figure 2). We also found that forest+intermediate animals' longest λ_{max} and open terrestrial animals' longest λ_{max} were longer than freshwater animals' and coastal animals' longest λ_{max} (omnibus test: $p = 1.19*10^{-14}$, $\chi^2 = 67.92$,

$$df = 3$$
; pairwise comparisons: table 3 and figure 2).

268 (h) Open terrestrial animals had a broader visual range than coastal animals

- 269 We found that open terrestrial animals had a larger range (longest λ_{max} shortest λ_{max}) than coastal
- animals, but all other habitat groups were statistically similar (omnibus test: p = 0.03366, $\chi^2 =$
- 271 8.6931, df = 3; pairwise comparisons: table 4).

Table 3. Effect of habitat greenness on longest λ_{max} . P-values are above diagonal; q-values are below diagonal. Numbers in bold are statistically significant.

p p		Forest +		Open	Mean	SD
q	Coastal	Intermediate	Freshwater	Terrestrial	(nm)	(nm)
Coastal		1.72*10 ⁻⁷	0.77	6.23*10 ⁻¹⁰	297.3	72.4
<i>Forest</i> + <i>Intermediate</i>	-7.82		7.54*10-6	0.22	547.1	17.7
Freshwater	-2.40	-6.85		2.77*10 ⁻⁵	314.3	73.8
Open Terrestrial	-9.14	-2.71	-7.73		534.1	49.1

272

- 273 (i) <u>Accounting for lineage removed the effects of water and dissolved particles on visual</u>
- 274 *pigment sensitivity*
- 275 Controlling for phylogeny removed the effect of habitat greenness on shortest λ_{max} , longest λ_{max} ,
- and λ_{max} range (table 5), even when we accounted for the species absent from our phylogenetic
- 277 tree (see the electronic supplementary material: tables S6 S8).

Table 4. Effect of habitat greenness on λ_{max} range. P-values are above diagonal; q-values are below diagonal. Numbers in bold are statistically significant.pForest +OpenMeanSD

p p		rorest +		Open	Mean	SD
q	Coastal	Intermediate	Freshwater	Terrestrial	(nm)	(nm)
Coastal		0.239	0.411	0.0497	42.88	70.2
Forest + Intermediate	-2.65		0.827	0.756	76.63	72.6
Freshwater	-2.18	-1.21		0.561	53.26	69.3
Open Terrestrial	-3.64	-1.40	-1.84		110.6	98.9

Table 5. Loss of effect of habitat greenness on visual sensitivity following phylogenetic						
correction. Comparisons shown are those	correction. Comparisons shown are those found significant in tables $2 - 4$.					
Comparison	Dependent variable	t	р	SE		
Coastal vs forest + intermediate	Shortest λ_{max}	-0.576	0.566	253.78		
Coastal vs open terrestrial	Shortest λ_{max}	-0.490	0.625	251.87		
Freshwater vs open terrestrial	Shortest λ_{max}	-0.445	0.658	254.04		
Coastal vs open terrestrial	Longest λ_{max}	-1.37	0.174	164.14		
Forest + intermediate vs freshwater	Longest λ_{max}	-1.74	0.0847	166.79		
Freshwater vs open terrestrial	Longest λ_{max}	-1.69	0.0941	165.55		
Open terrestrial vs coastal	λ_{max} Range	-0.397	0.692	255.87		

279

280

281 (4) Discussion

282

283 (a) The transition from aquatic to terrestrial habitats has influenced animal vision

284 We found that terrestrial species see longer long-wavelength light and a larger range of colours 285 overall compared to aquatic species. Few other studies have broadly investigated the effect of 286 animals' evolutionary transitions between aquatic and terrestrial habitats on colour vision. 287 However, transitions from aquatic to terrestrial life stages that lead to the development of different 288 visual abilities can indicate whether differences between terrestrial and aquatic lifestyles 289 themselves necessitate different strategies for perceiving the world [49]. Such studies have been 290 conducted within single species: for example, in several species of dragonflies, adults have short 291 wavelength-shifted vision, express more visual pigments than larvae, and have dorsal eye regions 292 specialized to detect shorter wavelengths of light refracted from the sky [49,50]. Similar types of 293 visual shifts have been observed in southern leopard frogs [66]. Just as animal development 294 favours the expression of environmentally matched opsins over an intra-generational timescale, 295 our results suggest that evolutionary adaptation favours the use of environmentally matched opsins 296 over an inter-generational timescale.

297

298	The results of our terrestrial vs. aquatic models are congruent with the visual tuning hypothesis,
299	that animal visual systems undergo adaptation to best detect the light most often present in their
300	environments [23]. Terrestrial animals are exposed to a dynamic range of colours that changes
301	throughout the day, including both short and long-wavelength light, as well as ultraviolet light in
302	large forest gaps and open environments [25,26]. By contrast, aquatic animals, which we found to
303	be less sensitive to long-wavelength and ultraviolet light, live in environments that are exposed to
304	relatively less long-wavelength and ultraviolet light [24]. Absent phylogenetic controls, our
305	regressions suggest that animals are likely to be maximally sensitive to colours most often present
306	in their environment, and insensitive to colours likely to be absent.

307

308 (b) Canopy coverage does not influence visual tuning

We found that animals which live in densely forested environments do not differ in their visual sensitivities from animals that live in open, prairie-like habitats. Although the forest light environment directly beneath the canopy is dominated by middle wavelengths (i.e., greens and yellows) under most conditions [25,26], spatial and temporal variations in forest light's spectral qualities may require forest animals to possess visual sensitivities similar to those of animals living in open habitats.

315

Additionally, animals may choose to use light microhabitats which are suitable to their current visual physiology. Endler and Théry observed that forest birds use areas in which they are most conspicuous to advertise to potential mates [67]. Some species also modify their habitats to improve the visibility of their visual displays. For example, male golden-collared manakins clean the arenas they use to court females; the background of a cleaned arena contrasts better with male manakins' plumage than the background of the forest surrounding the arena [68]. Arena cleaning also seems to improve white-bearded manakins' ability to detect predators [69]. In such cases, evolution may be driving site preferences which match vision rather than driving vision to match site preferences, a complete reversal of the mechanism being investigated in our study.

325

326 (c) The ciliary/rhabdomeric opsin divergence may impact the colours that animals can see

327 We found that animals that use rhabdomeric opsins for vision see a broader range of wavelengths 328 of light than animals that use ciliary opsins for vision. Many animals that use rhabdomeric 329 photoreceptors for vision, especially arthropods, have opsins that are maximally sensitive to 330 ultraviolet light [31,70–75]. By contrast, comparatively few animals that use ciliary photoreceptors 331 for vision have opsins that are maximally sensitive to UV light, although several species of birds 332 and fish are sensitive to ultraviolet light [40,76–78]. Additionally, many mammals that utilize high 333 acuity colour vision and whose short wavelength sensitive photoreceptors are sensitive to UV light, 334 have corneas that selectively filter UV, inhibiting their ability to see those wavelengths [79,80]. 335 Both ciliary and rhabdomeric opsins are thought to have been present in the urbilaterian, the 336 common ancestor of all modern animals save sponges, cnidarians, placozoans, and ctenophores 337 [55]. The emergence in chordates of ciliary opsins for vision rather than photoentrainment 338 represents a singular event, one that may have also heralded differences in visual perception 339 associated with reduced sensitivity to short wavelengths of light.

341 (d) Phylogeny outweighs the effect of habitat

342 We found that the effects of habitat upon the spectrum of light animals can see were reduced once 343 we controlled for phylogenetic history. These findings differ from those of studies looking at 344 individual animal clades. For example, a 2018 survey of ray-finned fish found that species living 345 at depth have reduced chromacy even after controlling for phylogeny [47]. Similarly, a historic 346 study of cottoid fish in Lake Baikal found that there was a correlation between λ_{max} and habitat 347 depth [81]. While studies of marine mammals found that species that forage near the surface have 348 visual pigments that resemble those of terrestrial mammals while those that foraged at depth had 349 visual pigments with amino acid substitutions that shifted the λ_{max} towards shorter wavelengths 350 [82]. We found that, when expanding to include multiple clades - both chordates and non-351 chordates – a similar pattern emerged: terrestrial species had broader sensitivity to light and more 352 sensitivity to long wavelengths of light compared to aquatic animals. However, these effects are 353 lost once we account for phylogeny. This loss of an effect might be because the historical 354 divergence between the visual pigments used by vertebrates and invertebrates is an important 355 limiting factor on the degree to which visual pigments can accommodate for light environment, 356 something that would not be detected in analyses limited to vertebrates.

357

The effect of the c-opsin/r-opsin divergence is lost once we account for phylogeny in our analyses, but since this transition happened once and maps onto the metazoan phylogenetic tree, this loss of effect might be expected. Outside of this transition, opsin evolutionary history such as mutation biases may account for the effect of phylogeny on visual ability in our analysis. Retinal is covalently bonded to opsin *via* a Schiff base and the charge of the amino acid residues near the Schiff base influence the ability of retinal to change conformation and λ_{max} of the associated opsin [83,84], which has been experimentally confirmed using directed mutagenesis [37,85,86]. Future research should consider whether there are inherent differences in the electronic charge of the binding pocket between ciliary and rhabdomeric type opsins. Additionally, studies examining whether non-opsin means of visual tuning, including the differential absorption of light by screening pigments, differ between animals which use ciliary and rhabdomeric opsins and which live in the same light environment may prove particularly illuminating.

370

(e) Conclusions

372 Here we used visual sensitivity data from nearly 450 animal species and 3 phyla to conduct a systematic survey of the effects of habitat light on the colours animals can see. We found that 373 374 terrestrial animals and aquatic animals possess different ranges of spectral sensitivity from each 375 other, but that evolutionary processes such as the c-opsin/r-opsin divergence may have limited 376 chordates' ability to tune their opsins to short-wavelength light. Additionally, the eyes of animals 377 living in terrestrial habitats are not specifically tuned to forest canopy cover. Future research 378 should consider whether inherent differences between chordate and non-chordate opsin amino acid 379 sequences, or downstream neural signalling, are responsible for the evolutionary limitations to 380 visual tuning.

381 Acknowledgments

382	We would like to thank Adam Siepielski for training in systematic analysis, Jeremy Beaulieu for
383	advice on conducting phylogenetically weighted regressions, and Nagayasu Nakanishi for his
384	valuable feedback on early versions of this project and manuscript.

385

- 386 Data Accessibility:
- 387 Data pertaining to this study are available in the supplementary materials or available at Dryad
- 388 repository <u>https://doi.org/10.5061/dryad.47d7wm3fc</u>. Code pertaining to this study is available at
- 389 GitHub repository: <u>https://github.com/mjosmurphy/opsin-evolutionary-ecology</u>.

390

- 391 Funding
- 392 This project was partially supported by the University of Arkansas and by an Arkansas Biosciences
- 393 Institute grant to ELW.

395 **<u>REFERENCES</u>**

396	1.	Uetz GW, Roberts JA, Taylor PW. 2009 Multimodal communication and mate choice in
397		wolf spiders: female response to multimodal versus unimodal signals. Anim. Behav. 78,
398		299–305. (doi:10.1016/j.anbehav.2009.04.023)
399	2.	Thurman TJ, Seymoure BM. 2015 A bird's eye view of two mimetic tropical butterflies:
400		coloration matches predator's sensitivity. J. Zool. 298, 159–168. (doi:10.1111/jzo.12305)
401	3.	Lindstrôm L, Rowe C, Guilford T. 2001 Pyrazine odour makes visually conspicuous prey
402		aversive. Proc. R. Soc. B 268, 159-162. (doi:10.1098/rspb.2000.1344)
403	4.	Endler JA. 1978 A Predator's View of Animal Color Patterns. In Evolutionary Biology,
404		Vol. 11 (eds MK Hecht, WC Steere, B Wallace), pp. 319-364.
405	5.	Cheroske AG, Cronin TW, Durham MF, Caldwell RL. 2009 Adaptive signalling
406		behaviour in stomatopods under varying light conditions. Mar. Freshw. Behav. Physiol.
407		42, 219–232. (doi:https://doi.org/10.1080/10236240903169222)
408	6.	Endler JA. 1992 Signals, signal conditions, and the direction of evolution. Am. Nat. 139,
409		S125-S153. (doi:https://doi.org/10.1086/285308)
410	7.	Guilford T, Dawkins MS. 1991 Receiver psychology and the evolution of animal signals.
411		Anim. Behav. 42, 1-14. (doi:https://doi.org/10.1016/S0003-3472(05)80600-1)
412	8.	Majerus MEN, Brunton CFA, Stalker J. 2000 A bird's eye view of the peppered moth. J.
413		Evol. Biol. 13, 155–159. (doi:https://doi.org/10.1046/j.1420-9101.2000.00170.x)
414	9.	Cook LM, Grant BS, Saccheri IJ. 2012 Selective bird predation on the peppered moth: the
415		last experiment of Michael Majerus. Biol. Lett. 8, 609-612.

416 (doi:https://doi.org/10.1098/rsbl.2011.1136)

417	10.	Johnsen S, Sosik HM. 2003 Cryptic coloration and mirrored sides as camouflage
418		strategies in near-surface pelagic habitats: Implications for foraging and predator
419		avoidance. Limnol. Oceanogr. 48, 1277-1288. (doi:10.4319/lo.2003.48.3.1277)
420	11.	Johnsen S. 2005 The red and the black: Bioluminescence and the color of animals in the
421		deep sea. Integr. Comp. Biol. 45, 234–246. (doi:10.1093/icb/45.2.234)
422	12.	Harmon JP, Losey E. 1998 The role of vision and color in the close proximity foraging
423		behavior of four coccinellid species. Oecologia 115, 287-292.
424		(doi:https://doi.org/10.1007/s004420050518)
425	13.	Endler JA, Day LB. 2006 Ornament colour selection, visual contrast and the shape of
426		colour preference functions in great bowerbirds, Chlamydera nuchalis. Anim. Behav. 72,
427		1405–1416. (doi:10.1016/j.anbehav.2006.05.005)
428	14.	Heindl M, Winkler H. 2003 Female Canaries (Serinus canaria) Associate more with
429		Males that Contrast Strongly against the Background. <i>Ethology</i> 109 , 259–271.
430		(doi:http://dx.doi.org/10.1046/j.1439-0310.2003.00869.x)
431	15.	Fleishman LJ, Wadman CS, Maximov KJ. 2020 The interacting effects of total light
432		intensity and chromatic contrast on visual signal visibility in an Anolis lizard. Anim.
433		Behav. 167, 263–273. (doi:10.1016/j.anbehav.2020.07.008)
434	16.	Gomez D, Théry M. 2004 Influence of ambient light on the evolution of colour signals:
435		Comparative analysis of a Neotropical rainforest bird community. <i>Ecol. Lett.</i> 7 , 279–284.
436		(doi:10.1111/j.1461-0248.2004.00584.x)

437	17.	Pincebourde S, Théry M, Feer F. 2008 Dusk light environment optimizes visual
438		perception of conspecifics in a crepuscular horned beetle. Behav. Ecol. 29, 627-634.
439		(doi:10.1093/beheco/arn024)
440	18.	Endler JA, Théry M. 1996 Interacting effects of lek placement, display behavior, ambient
441		light, and color patterns in three neotropical forest-dwelling birds. Am. Nat. 148, 421-452.
442		(doi:10.1086/285934)
443	19.	Håstad O, Victorsson J, Ödeen A. 2005 Differences in color vision make passerines less
444		conspicuous in the eyes of their predators. Proc. Natl. Acad. Sci. 102, 6391-6394.
445		(doi:https://doi.org/10.1073/pnas.0409228102)
446	20.	Cole GL, Endler JA. 2016 Male courtship decisions are influenced by light environment
447		and female receptivity. Proc. R. Soc. B 283, 20160861.
448		(doi:http://dx.doi.org/10.1098/rspb.2016.0861)
449	21.	Marshall KLA, Stevens M. 2014 Wall lizards display conspicuous signals to conspecifics
449 450	21.	Marshall KLA, Stevens M. 2014 Wall lizards display conspicuous signals to conspecifics and reduce detection by avian predators. <i>Behav. Ecol.</i> 25 , 1325–1337.
	21.	
450	21.	and reduce detection by avian predators. <i>Behav. Ecol.</i> 25 , 1325–1337.
450 451		and reduce detection by avian predators. <i>Behav. Ecol.</i> 25 , 1325–1337. (doi:10.1093/beheco/aru126)
450 451 452		and reduce detection by avian predators. <i>Behav. Ecol.</i> 25 , 1325–1337. (doi:10.1093/beheco/aru126) Sherratt TN, Beatty CD. 2003 The Evolution of Warning Signals as Reliable Indicators of
450 451 452 453	22.	 and reduce detection by avian predators. <i>Behav. Ecol.</i> 25, 1325–1337. (doi:10.1093/beheco/aru126) Sherratt TN, Beatty CD. 2003 The Evolution of Warning Signals as Reliable Indicators of Prey Defense. <i>Am. Nat.</i> 162, 377–389. (doi:10.1086/378047)
450 451 452 453 454	22.	 and reduce detection by avian predators. <i>Behav. Ecol.</i> 25, 1325–1337. (doi:10.1093/beheco/aru126) Sherratt TN, Beatty CD. 2003 The Evolution of Warning Signals as Reliable Indicators of Prey Defense. <i>Am. Nat.</i> 162, 377–389. (doi:10.1086/378047) Cronin TW, Johnsen S, Marshall NJ, Warrant EJ. 2014 <i>Visual Ecology</i>. Princeton, NJ:

458 (doi:https://doi.org/10.2307/2937121)

- 459 26. Federer CA, Tanner CB. 1966 Spectral Distribution of Light in the Forest. *Ecology* 47,
 460 555–560. (doi:https://doi.org/10.2307/1933932)
- 461 27. Arikawa K, Stavenga D. 1997 Random array of colour filters in the eyes of butterflies. J.

462 *Exp. Biol.* **200**, 2501–256. (doi:https://doi.org/10.1242/jeb.200.19.2501)

- 463 28. Arikawa K, Scholten DGW, Kinoshita M, Stavenga DG, Arikawa K, Scholten DGW,
- 464 Kinoshita M. 1999 Tuning of Photoreceptor Spectral Sensitivities by Red and Yellow
- 465 Pigments in the Butterfly Papilio xuthus Tuning of Photoreceptor Spectral Sensitivities by
- 466 Red and Yellow Pigments in the Butterfly Papilio xuthus. *Zoolog. Sci.* **16**, 17–24.
- 467 (doi:https://doi.org/10.2108/zsj.16.17)
- 468 29. Stavenga DG, Arikawa K. 2006 Evolution of color and vision of butterflies. *Arthropod*469 *Struct. Dev.* 35, 307–318. (doi:10.1016/j.asd.2006.08.011)
- 470 30. Stavenga DG. 2002 Reflections on colourful ommatidia of butterfly eyes. J. Exp. Biol.

471 **205**, 1077–1085. (doi:https://doi.org/10.1242/jeb.205.8.1077)

472 31. Cronin TW, Marshall NJ. 1989 Multiple spectral classes of photoreceptors in the retinas

473 of gonodactyloid stomatopod crustaceans. J. Comp. Physiol. A 166, 261–275.

- 474 (doi:10.1007/BF00193471)
- 475 32. Ogawa Y, Awata H, Wakakuwa M, Kinoshita M, Stavenga DG, Arikawa K. 2012
- 476 Coexpression of three middle wavelength-absorbing visual pigments in sexually
- 477 dimorphic photoreceptors of the butterfly *Colias erate*. J. Comp. Physiol. A Neuroethol.
- 478 Sensory, Neural, Behav. Physiol. 198, 857–867. (doi:10.1007/s00359-012-0756-8)

479	33.	Hart NS, Partridge JC, Cuthill IC. 1999 Visual pigments, cone oil droplets, ocular media
480		and predicted spectral sensitivity in the domestic turkey (Meleagris gallopavo). Vision
481		Res. 39, 3321-3328. (doi:https://doi.org/10.1016/S0042-6989(99)00071-1)
482	34.	Hart NS. 2004 Microspectrophotometry of visual pigments and oil droplets in a marine
483		bird, the wedge-tailed shearwater Puffinus pacificus: topographic variations in
484		photoreceptor spectral characteristics. J. Exp. Biol. 207, 1229-1240.
485		(doi:10.1242/jeb.00857)
486	35.	Beason RC, Loew ER. 2008 Visual pigment and oil droplet characteristics of the bobolink
487		(Dolichonyx oryzivorus), a new world migratory bird. Vision Res. 48, 1-8.
488		(doi:10.1016/j.visres.2007.10.006)
489	36.	Yokoyama S, Radlwimmer FB, Blow NS. 2000 Ultraviolet pigments in birds evolved
490		from violet pigments by a single amino acid change. Proc. Natl. Acad. Sci. 97, 7366-
491		7371. (doi:https://doi.org/10.1073/pnas.97.13.7366)
492	37.	Yokoyama R, Knox BE, Yokoyama S. 1995 Rhodopsin from the fish, Astyanax: Role of
493		tyrosine 261 in the red shift. Investig. Ophthalmol. Vis. Sci. 36, 939-945.
494	38.	Carvalho LS, Cowing JA, Wilkie SE, Bowmaker JK, Hunt DM. 2007 The molecular
495		evolution of avian ultraviolet- and violet-sensitive visual pigments. Mol. Biol. Evol. 24,
496		1843–1852. (doi:10.1093/molbev/msm109)
497	39.	Butlin R et al. 2012 What do we need to know about speciation? Trends Ecol. Evol. 27,
498		27-39. (doi:10.1016/j.tree.2011.09.002)
499	40.	Van Hazel I, Sabouhanian A, Day L, Endler JA, Chang BS. 2013 Functional

500	characterization of	f spectral tuni	ng mechanisms	in the great	bowerbird shor	t-wavelength

- 501 sensitive visual pigment (SWS1), and the origins of UV/violet vision in passerines and
- 502 parrots. *BMC Evol. Biol.* **13**. (doi:10.1186/1471-2148-13-250)
- 503 41. Townson SM, Chang BSW, Salcedo E, Chadwell L V, Pierce NE, Britt SG. 1998
- 504 Honeybee Blue- and Ultraviolet-Sensitive Opsins: Cloning, Heterologous Expression in
- 505 *Drosophila*, and Physiological Characterization. **18**, 2412–2422.
- 506 (doi:https://doi.org/10.1523/JNEUROSCI.18-07-02412.1998)
- 507 42. Yokoyama S. 2002 Molecular bases of color vision in vertebrates. Genes Genet. Syst. 74,
- 508 189–199. (doi:10.1266/ggs.74.189)
- 43. Yokoyama S, Bernhard Radlwimmer F. 2001 The molecular genetics and evolution of red
 and green color vision in vertebrates. *Genetics* 158, 1697–1710.
- 511 44. Frentiu FD, Briscoe AD. 2008 A butterfly eye's view of birds. *BioEssays* 30, 1151–1162.
 512 (doi:10.1002/bies.20828)
- 513 45. Hunt DM, Wilkie SE, Bowmaker JK, Poopalasundaram S. 2001 Vision in the ultraviolet.
 514 *Cell. Mol. Life Sci.* 58, 1583–1598. (doi:10.1007/PL00000798)
- 515 46. Fasick JI, Robinson PR. 2000 Spectral-tuning mechanisms of marine mammal rhodopsins
 516 and correlations with foraging depth. *Vis. Neurosci.* 17, 781–788.
- 517 (doi:10.1017/S095252380017511X)
- 518 47. Schweikert LE, Fitak RR, Caves EM, Sutton TT, Johnsen S. 2018 Spectral sensitivity in
- 519 ray-finned fishes: diversity, ecology, and shared descent. J. Exp. Biol., jeb.189761.

520 (doi:10.1242/jeb.189761)

521	48.	Partridge JC, Shand J, Archer SN, van Groningen-Luyben WAH. 1989 Interspecific
522		variation in the visual pigments of deep-sea fishes. J. Comp. Physiol. A 164, 513-529.
523		(doi:https://doi.org/10.1007/BF00610445)
524	49.	Chou A, Lin C, Cronin TW. 2020 Visual metamorphoses in insects and malacostracans:
525		Transitions between an aquatic and terrestrial life. Arthropod Struct. Dev. 59, 100974.
526		(doi:10.1016/j.asd.2020.100974)
527	50.	Futahashi R, Kawahara-miki R, Kinoshita M, Yoshitake K, Yajima S. 2015 Extraordinary
528		diversity of visual opsin genes in dragonflies. (doi:10.1073/pnas.1424670112)
529	51.	Almudi I et al. 2020 Genomic adaptations to aquatic and aerial life in mayflies and the
530		origin of insect wings. Nat. Commun. 11, 1-11. (doi:10.1038/s41467-020-16284-8)
531	52.	Schott RK, Bell RC, Loew ER, Thomas KN, Gower DJ, Streicher JW, Fujita MK. 2021
532		Genomic and Spectral Visual Adaptation in Southern Leopard Frogs during the
533		Ontogenetic Transition from Aquatic to Terrestrial Light Environments. bioRxiv Prepr.
534		(doi:https://doi.org/10.1101/2021.02.19.432049)
535	53.	Labhart T, Nilsson DE. 1995 The dorsal eye of the dragonfly Sympetrum: specializations
536		for prey detection against the blue sky. J. Comp. Physiol. A 176, 437-453.
537		(doi:10.1007/BF00196410)
538	54.	Porter ML, Blasic JR, Bok MJ, Cameron EG, Pringle T, Cronin TW, Robinson PR. 2011
539		Shedding new light on opsin evolution. Proc. R. Soc. B Biol. Sci. 279, 3-14.
540		(doi:10.1098/rspb.2011.1819)
541	55.	Arendt D, Wittbrodt J. 2001 Reconstructing the eyes of Urbilateria. 1563.

542 (doi:10.1098/rstb.2001.0971)

- 543 56. Ullmann JFP, Gallagher T, Hart NS, Barnes AC, Smullen RP, Collin SP, Temple SE.
- 544 2011 Tank color increases growth, and alters color preference and spectral sensitivity, in
- 545 barramundi (*Lates calcarifer*). *Aquaculture* **322–323**, 235–240.
- 546 (doi:10.1016/j.aquaculture.2011.10.005)
- 547 57. Fuller RC, Claricoates KM. 2011 Rapid light-induced shifts in opsin expression: Finding
- 548 new opsins, discerning mechanisms of change, and implications for visual sensitivity.

549 *Mol. Ecol.* **20**, 3321–3335. (doi:10.1111/j.1365-294X.2011.05180.x)

- 550 58. Nowak RM. 1983 *Walker's Mammals of the World*. 4th edn. Baltimore: Johns Hopkins
 551 University Press.
- 552 59. Opler PA, Malikul V. 1999 *A Field Guide to Eastern Butterflies*. Houghton Mifflin
 553 Harcourt.
- 554 60. Opler PA. 1999 A Field Guide to Butterflies of Western North America. Houghton Mifflin
 555 Harcourt.
- 556 61. Hinchliff C *et al.* 2015 Synthesis of phylogeny and taxonomy into a comprehensive tree of
 557 life. *Proc. Natl. Acad. Sci.* 112, 12764–12769.
- 558 (doi:https://doi.org/10.1073/pnas.1423041112)
- 62. Revell LJ. 2012 phytools: An R package for phylogenetic comparative biology (and other
 things). *Methods Ecol. Evol.* 3. (doi:10.1111/j.2041-210X.2011.00169.x)
- 561 63. Paradis E, Schliep K. 2018 ape 5.0: an environment for modern phylogenetics and
 562 evolutionary analyses in R. *Bioinformatics* 35, 526–528.

563 ((doi:https://doi.org/10.1093/bioinforr	matics/bty633)

- 64. Ho L, Ane C. 2014 A linear-time algorithm for Gaussian and non-Gaussian trait evolution
 models. *Syst. Biol.* 63, 397–408. (doi:https://doi.org/10.1093/sysbio/syu005)
- 566 65. Pohlert T. 2020 PMCMRplus: Calculate Pairwise Multiple Comparisons of Mean Rank
 567 Sums Extended.
- 568 66. Schott RK, Bell RC, Loew ER, Thomas KN, Gower DJ, Streicher JW, Fujita MK. 2021
- 569 Genomic and Spectral Visual Adaptation in Southern Leopard Frogs during the
- 570 Ontogenetic Transition from Aquatic to Terrestrial Light Environments. *bioRxiv Prepr*.
- 571 (doi:https://doi.org/10.1101/2021.02.19.432049)
- 572 67. Endler J, Théry M. 1996 Interacting effects of lek placement, display behavior, ambient
- 573 light, and color patterns in three Neotropical forest-dwelling birds. Am. Nat. 148, 421–

574 452. (doi:https://doi.org/10.1086/285934)

- 575 68. Uy JAC, Endler JA. 2004 Modification of the visual background increases the
- 576 conspicuousness of golden-collared manakin displays. *Behav. Ecol.* **15**, 1003–1010.
- 577 (doi:10.1093/beheco/arh106)
- 69. Cestari C, Aurélio Pizo M. 2014 Court cleaning behavior of the White-Bearded Manakin
 (*Manacus manacus*) and a test of the anti-predation hypothesis. *Wilson J. Ornithol.* 126,
- 580 98–104. (doi:https://doi.org/10.1676/13-032.1)
- 581 70. Yang E, Osorio D. 1991 Spectral sensitivities of photoreceptors and lamina monopolar
- 582 cells in the dragonfly, *Hemicordula tau. J. Comp. Physiol. A* **169**, 663–669.
- 583 (doi:https://doi.org/10.1007/BF00194895)

584	71.	Bybee SM, Yuan F, Ramstetter MD, Llorente-Bousquets J, Reed RD, Osorio D, Briscoe
585		AD. 2012 UV Photoreceptors and UV-Yellow Wing Pigments in Heliconius Butterflies
586		Allow a Color Signal to Serve both Mimicry and Intraspecific Communication. Am. Nat.
587		179, 38–51. (doi:10.1086/663192)
588	72.	Briscoe AD. 2008 Reconstructing the ancestral butterfly eye: focus on the opsins. J. Exp.
589		Biol. 211, 1805–1813. (doi:10.1242/jeb.013045)
590	73.	Everett A, Tong X, Briscoe AD, Monteiro A. 2012 Phenotypic plasticity in opsin
591		expression in a butterfly compound eye complements sex role reversal. BMC Evol. Biol.
592		12 . (doi:10.1186/1471-2148-12-232)
593	74.	Arikawa K, Stavenga DG. 2014 Insect photopigments: photoreceptor spectral sensitivities
594		and visual adaptations. In Evolution of Visual and Non-Visual Pigments (eds DM Hunt,
595		MW Hankins, SP Collin, NJ Marshall), pp. 1–276. (doi:10.1007/978-1-4614-4355-1)
596	75.	Arikawa K. 2005 Sexual Dimorphism of Short-Wavelength Photoreceptors in the Small
597		White Butterfly, Pieris rapae crucivora. J. Neurosci. 25, 5935–5942.
598		(doi:10.1523/JNEUROSCI.1364-05.2005)
599	76.	Carvalho LS, Knott B, Berg ML, Bennett ATD, Hunt DM. 2011 Ultraviolet-sensitive
600		vision in long-lived birds. Proc. R. Soc. B Biol. Sci. 278, 107-114.
601		(doi:10.1098/rspb.2010.1100)
602	77.	Losey GS, Cronin TW, Goldsmith TH, Hyde D, Marshall NJ, McFarland WN. 1999 The
603		UV visual world of fishes: a review. J. Fish Biol. 54, 921–943.
604		(doi:https://doi.org/10.1111/j.1095-8649.1999.tb00848.x)

605	78.	Marshall NJ, Jennings K, McFarland WN, Loew ER, Losey GS. 2003 Visual Biology of
606		Hawaiian Coral Reef Fishes. II. Colors of Hawaiian Coral Reef Fish. Copeia 2003, 455-
607		466. (doi:10.1643/01-055)
608	79.	Douglas RH, Jeffery G. 2014 The spectral transmission of ocular media suggests
609		ultraviolet sensitivity is widespread among mammals. Proc. R. Soc. B Biol. Sci. 281,
610		20132995-20132995. (doi:10.1098/rspb.2013.2995)
611	80.	Hogg C, Neveu M, Stokkan K-A, Folkow L, Cottrill P, Douglas R, Hunt DM, Jeffery G.
612		2011 Arctic reindeer extend their visual range into the ultraviolet. J. Exp. Biol. 214, 2014-
613		2019. (doi:10.1242/jeb.053553)
614	81.	Bowmaker JK, Govardovskii VI, Shukolyukov SA, L.V. Zueva J, Hunt DM, Sideleva VG,
615		Smirnova OG. 1994 Visual pigments and the photic environment: The cottoid fish of Lake
616		Baikal. Vision Res. 34, 591-605. (doi:10.1016/0042-6989(94)90015-9)
617	82.	Levenson DH, Ponganis PJ, Crognale MA, Deegan JF, Dizon A, Jacobs GH. 2006 Visual
618		pigments of marine carnivores: Pinnipeds, polar bear, and sea otter. J. Comp. Physiol. A
619		Neuroethol. Sensory, Neural, Behav. Physiol. 192, 833-843. (doi:10.1007/s00359-006-
620		0121-x)
621	83.	Sekharan S, Katayama K, Kandori H, Morokuma K. 2012 Color vision: 'OH-site' rule for
622		seeing red and green. J. Am. Chem. Soc. 134, 10706-10712. (doi:10.1021/ja304820p)
623	84.	Honig B, Greenberg AD, Dinur U, Ebrey TG. 1976 Visual-Pigment Spectra: Implications
624		of the Protonation of the Retinal Schiff Base. Biochemistry 15, 4593-4599.
625		(doi:10.1021/bi00666a008)

626	85.	Davies WL, Collin SP, Hunt DM. 2009 Adaptive gene loss reflects differences in the
627		visual ecology of basal vertebrates. Mol. Biol. Evol. 26, 1803-1809.
628		(doi:10.1093/molbev/msp089)
629	86.	Hiramatsu C, Radlwimmer FB, Yokoyama S, Kawamura S. 2004 Mutagenesis and
630		reconstitution of middle-to-long-wave-sensitive visual pigments of New World monkeys
631		for testing the tuning effect of residues at sites 229 and 233. Vision Res. 44, 2225–2231.
632		(doi:10.1016/j.visres.2004.04.008)

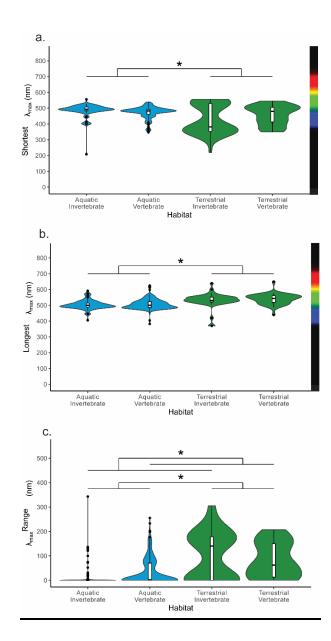


Figure 1. Effect of coarse habitat and lineage on mean visual pigment sensitivity, prior to phylogenetic control. A. Shortest opsin: Aquatic invertebrates: n = 78, $\mu = 485.3$ nm; Aquatic vertebrates: $n = 273 \ \mu = 473.0$ nm; Terrestrial invertebrates: n = 43, $\mu = 430.1$ nm; Terrestrial vertebrates: n = 39, $\mu = 456.1$ nm. B. Longest opsin: Aquatic invertebrates: n = 78, $\mu = 503.3$ nm; Aquatic vertebrates: n = 273, $\mu = 507.6$ nm; Terrestrial invertebrates: n = 43, $\mu = 530.3$ nm; Terrestrial vertebrates: n = 39, $\mu = 539.1$ nm. C. Opsin range: Aquatic invertebrates: n = 78, $\mu = 18.0$ nm; Aquatic vertebrates: n = 273, $\mu = 34.6$ nm; Terrestrial invertebrates: n = 43, $\mu = 100.2$ nm; Terrestrial vertebrates: n = 39, $\mu = 83.0$ nm. *: p < 0.05.

635

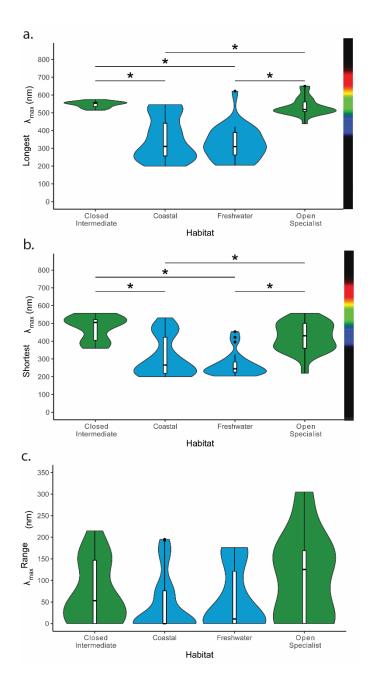


Figure 2. Effect of habitat greenness on visual sensitivities. A. Longest opsin: forest + intermediate: n = 14, $\mu = 547.1$ nm, sd = 17.71 nm; coastal: n = 40, $\mu = 297.3$ nm, sd = 72.42 nm; freshwater: n = 23, $\mu = 314.3$ nm, sd = 73.77 nm; open terrestrial: n = 22, $\mu = 534.1$ nm, sd = 49.09 nm; B. Shortest opsin: forest + intermediate: n = 14, $\mu = 470.4$ nm, sd = 60.00 nm; coastal: n = 40, $\mu = 254.4$ nm, sd = 55.16 nm; freshwater: n = 23, $\mu = 261.1$ nm, sd = 56.52 nm; open terrestrial: n = 22, $\mu = 423.4$ nm, sd = 82.06 nm C. Opsin range: forest + intermediate: n = 14, $\mu = 76.63$ nm, sd = 72.61 nm; coastal: n = 40, $\mu = 742.88$ nm, sd = 70.15 nm; freshwater: n = 23, $\mu = 53.26$ nm, sd = 69.30 nm; open: n = 22, $\mu = 110.6$ nm, sd = 98.82 nm. *: p < 0.05.