1	Phylogeny of the Formicoxenus genus-group (Hymenoptera: Formicidae) reveals isolated
2	lineages of Leptothorax acervorum in the Iberian Peninsula predating the Last Glacial
3	Maximum
4	
5	
6	Dario I. Ojeda ^{1*} , Max John ² , Robert L. Hammond ² , Riitta Savolainen ³ , Kari Vepsäläinen ⁴ and
7	Torstein Kvamme ⁵
8	
9	
10	¹ Department of Forest Genetics and Biodiversity. Norwegian Institute of Bioeconomy Research
11	(NIBIO), 1430 Ås, Norway
12	² Department of Genetics & Genome Biology. University of Leicester, University Road, Leicester,
13	LE1 7RH, UK
14	³ Department of Biosciences. University of Helsinki, P.O. Box 65, 00014, Finland
15	⁴ Retuperä Biological Station. 02880 Nuuksio, Finland
16	⁵ Department of Invertebrate Pests and Weeds in Forestry, Agriculture and Horticulture. Norwegian
17	Institute of Bioeconomy Research (NIBIO), 1430 Ås, Norway
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
28	
29	
30	
31	
32	* Corresponding author: <u>dario.alayon@nibio.no</u> , Tel: +47 47633227

Abstract

The Formicoxenus genus-group comprises six genera within the tribe Crematogastrini. The group is well known for repeated evolution of social parasitism among closely related taxa and cold-adapted species with large distribution ranges in the Nearctic and Palearctic regions. Previous analyses based on nuclear markers (ultraconserved elements, UCEs) and mitochondrial genes suggest close relationship between Formicoxenus Mayr, 1855, Leptothorax Mayr, 1855 and Harpagoxenus Forel, 1893. However, scant sampling has limited phylogenetic assessment of these genera. Also, previous phylogeographic analyses of L. acervorum (Fabricius, 1793) have been limited to its West-Palearctic range of distribution, which has provided a narrow view on recolonization, population structure and existing refugia of the species. Here, we inferred the phylogenenetic history of genera within the Formicoxenus genus-group and reconstructed the phylogeography of *L. acervorum* with more extensive sampling. We employed four datasets consisting of whole genomes and sequences of the COI. The topologies of previous nuclear and our inferences based on mitochondrial genomes were overall congruent. Further, Formicoxenus may not be monophyletic. We found several monophyletic lineages that do not correspond to the current species described within Leptothorax, especially in the Nearctic region. We identified a monophyletic L. acervorum lineage that comprises both Nearctic and Palearctic locations. The most recent expansion within L. acervorum probably occurred within the last 0.5 Ma with isolated populations predating the Last Glacial Maximum (LGM), which are localized in at least two refugial areas (Pyrenean and Northern plateau) in the Iberian Peninsula. The patterns recovered suggest a shared glacial refugium in the Iberian Peninsula with cold-adapted trees that currently share high-altitude environments in this region. Key words: Phylogenomics, mitogenomes, ant, Leptothorax, Formicoxenus, Last Glacial

Maximum refugia, biogeography, phylogeography.

71 **1. Introduction**

Several invertebrate groups have species with Holarctic distribution, including beetles 72 73 (Larson and Nilsson, 1985), Lepidoptera (Landry et al., 2013), spiders (Marusik and Koponen, 74 2005) and ants (Schär et al., 2018). Among ant species, only three species (*Camponotus* 75 herculeanus Linnaeus, 1758, Formica gagatoides Ruzsky, 1904 and Leptothorax acervorum 76 Fabricius, 1793) are known to have monophyletic lineages with a Holarctic distribution (Schär et 77 al., 2018). The latter of these ant species belongs to the tribe Crematogastrini Emery, 1914 within the subfamily Myrmicinae, with Crematogastrini comprising some 6,630 species (Blaimer et al., 78 79 2018; Ward et al., 2015). There, recent phylogenomic analyses based on ultraconserved elements (UCEs) have consistently recovered a monophyletic lineage of six genera (Vombisidris Bolton 80 81 1991, Gauromyrmex Menozzi, 1993, Harpagoxenus Forel, 1893, Formicoxenus Mayr, 1855, Temnothorax Mayr, 1861 and Leptothorax Mayr, 1855) referred to informally as the 82 83 Formicoxenus genus-group (Blaimer et al., 2018; Branstetter et al., 2017). These analyses have recovered a close relationship between Formicoxenus and Leptothorax. However, all these studies 84 85 have been focused on higher taxonomic relationships and few studies have included a comprehensive sampling of species within each genus (Heinze and Gratiashvili, 2015; Prebus, 86 2017; Schär et al., 2018). Among the *Formicoxenus* genus-group, *Leptothorax* is the second largest 87 genus with an estimated 20 species (AntWeb ver. 8.42, https://www.antweb.org, accessed 29 88 89 October 2020). The genus has a Holarctic distribution and it has been inferred to have originated in the Nearctic and dispersed in the Palearctic within the last 2 Ma (Schär et al., 2018). Relationships 90 91 within *Leptothorax* have not been fully resolved and previous phylogenetic analyses indicate the presence of multiple undescribed and non-monophyletic taxa, particularly in the Nearctic (Heinze 92 and Gratiashvili, 2015; Schär et al., 2018). At least seven species have been described in the 93 94 Palearctic region, most of them with limited distribution and only L. acervorum with a distribution both in the Nearctic and the Palearctic regions (Schär et al., 2018). 95

96 Taxa that span large geographic regions in both the Nearctic and Palearctic are expected to have experienced variable connectivity because of the fluctuating presence of the land bridge of 97 Beringia between Eurasia and North America (DeChaine, 2008). Also, climate oscillations during 98 the Quaternary (last 2 Ma), characterized by pronounced cycles between cold glacial (ca. 100,000 99 100 years) and warm interglacial periods (ca. 20,000 years) during the last 700,000 years, altered the 101 geographic distribution of species in the northern hemisphere (Nearctic and Palearctic) (Hewitt, 2000). During these glacial episodes, species ranges contracted to refugia in suitable areas in the 102 southern part of their distribution. As the climate warmed and glaciers retreated, species with a 103

temperate range of distribution expanded and reconnected. In contrast, the range of distribution for 104 105 boreal cold-adapted species likely reduced and fragmented (Hewitt, 1996; Petit et al., 2003). Evidence from several ant species have suggested the presence of several refugia during the 106 Pleistocene (2.58 - 0.012 Ma) in the southern Mediterranean peninsulas, the Caspian-Caucasus 107 region and further east in southern East Asia (Beibl et al., 2007; Goropashnaya et al., 2004; 108 Leppänen et al., 2013, 2011; Pusch et al., 2006; Schlick-Steiner et al., 2007). In addition, 109 populations of cold-adapted ant species could have also survived in more northerly refugia near the 110 permafrost (Leppänen et al., 2013, 2011). Indeed, L. acervorum is among the very few cold-111 112 adapted species that extend their distribution above the polar circle both in the Nearctic and the Palearctic (Berman et al., 2010; Heinze et al., 1998, 1996). In the Palearctic, this species occurs in 113 114 the boreal zone from the Atlantic Ocean to Japan, and in the mountains of southern Europe, the Caucasus, and the Tien-Shan and Pamir (Czechowski et al., 2012; Seifert, 2018). Thus, the 115 116 climatic fluctuations of the Quaternary have likely played a significant role in shaping its current 117 distribution, connectivity, and genetic diversity. Populations located near the permafrost and those 118 located on the southern range of its distribution were likely affected differently.

119 The most recent phylogenetic analysis of Leptothorax indicate that L. acervorum originated about 2 Ma, with the least diverged populations located in the Nearctic region. Within the 120 Palearctic, populations situated in the Iberian Peninsula were inferred to be the less divergent 121 122 among the specimens included (Schär et al., 2018), which might have been located in refugia 123 during the glacial cycles. In addition, more detailed analyses based on mitochondrial DNA (COI-124 3P region) and microsatellites (SSRs) have been used to infer the phylogeography and population structure of this species in the western part of its distribution (West Palearctic). These analyses 125 have found generally less population structure in *L. acervorum* compared to other species closely 126 127 related species within Leptothorax, e.g. L. muscorum (Nylander, 1846) and Harpagoxenus sublaevis (Nylander, 1849) (Brandt et al., 2007; Foitzik et al., 2009; Trettin et al., 2016), but also 128 129 evidence of divergent haplotypes have been found in the Pyrenees and Southern France (Trettin et al., 2016). Given the large distribution range across the Holarctic and the extensive variation in the 130 131 latitudinal range in western Europe (from the Iberian Peninsula to North Cape in Norway) (Heinze and Holldobler, 1994), approaches that combine analyses at different taxonomic levels with 132 133 extensive sampling are necessary to understand the phylogenetic relationships and evolutionary history of Leptothorax species. Here we present the most comprehensive sampling of members of 134 135 the *Formicoxenus* genus-group with an emphasis on the phylogenetic relationships within Leptothorax and the biogeography of L. acervorum across its range of distribution in the Holarctic 136

region. Our specific objectives are: 1) to infer relationships among the six genera of the

138 Formicoxenus genus-group using whole mitochondrial genomes and asses their correspondence

139 with previous topologies obtained with nuclear genes, 2) to clarify the relationships of *Leptothorax*

species and the timing of divergence of the Palearctic species, 3) to determine if the populations of

- 141 *L. acervorum* situated in the Iberian Peninsula survived in different refugia during the glacial
- 142 cycles of the Quaternary.
- 143

144 **2. Materials and methods**

145 2.1 Taxon sampling and datasets

The sampling strategy used in this study was developed to represent the 146 Formicoxenus genus-group at three different hierarchical levels. The first dataset consisted 147 of 49 specimens representing all six genera (Temnothorax, Leptothorax, Formicoxenus, 148 Harpagoxenus, Gauromyrmex and Vombisidris) currently recognized within this group 149 (Blaimer et al., 2015; Prebus, 2017), representing 17% of genera within the 150 Crematogastrini. We also included outgroups from Myrmicinae (all tribes), Dolichoderinae 151 152 and Ponerinae. In this data set we used whole mitochondrial genomes to explore the major relationships within Formicoxenus genus-group at the genus level. We used only one 153 154 representative specimen per species within each genus, except for L. acervorum, where we included multiple samples (Table S1). In the second dataset, we gathered specimens 155 156 representing eight out of the 20 Leptothorax species currently recognized (AntWeb ver. 8.42, https://www.antweb.org, accessed 29 October 2020), two Formicoxenus species and 157 H. sublaevis. In this dataset we sequenced the section of the mitochondrial cytochrome c 158 oxidase (COI-5P region, 658 bp) in 96 specimens (Table S2). The third dataset consisted of 159 113 specimens of L. acervorum across its distribution range in the Holarctic region, where 160 we sequenced the same gene region (COI-5P region, 658 bp) as the previous dataset. This 161 dataset was complemented with available sequences from public repositories (Table S3). 162 Finally, dataset four consisted of the mitochondrial region COI-trnL2-COII (2,293 bp) and 163 it was obtained from a total of 80 L. acervorum specimens from six populations in Spain 164 165 and the UK (Table S3).

- 166
- 167 2.2. Whole mitochondrial sequencing and assembly

Mitochondrial genomes were newly generated for six specimens of *L. acervorum*from six different populations in Spain and the UK (Spain: Valdelinares (V), Orihuela del

Tremedal (OT), Larra (L), Niela Refuge (NR), Pla de la Font (PF); UK: Santon Downham 170 (SD) (Table S3). A *de novo* mitochondrial genome was identified as part of a whole 171 genome sequencing project from a single adult male (PF population, sample: 172 PF18 15 M1) using 10x linked reads assembled with Supernova 2.1.1 (Weisenfeld et al., 173 2018). The scaffold containing the mtDNA genome was identified by a BLASTn query of 174 the assembled genome with two published *L. acervorum* mtDNA sequences (query 1: 175 COXI – tRNA - Leu - COXII: GenBank: KU245569 (Trettin et al., 2016); query 2: COB: 176 177 GenBank: HQ259995 (Gill et al., 2009). These two sequences, located ~6Kb apart in the canonical hymenopteran mtDNA genome, were used to minimize erroneous matches to 178 nuclear genomic scaffolds containing translocated mtDNA (NUMTs). Only two scaffolds 179 180 (102,807 and 104,071) showed convincing matches to both query sequences (E value = 0, bit scores > 1000). However, mapping re-sequenced samples (see below) showed scaffold 181 182 102,807 had 40 times higher coverage (200x-400x) than 104,071 (~5x-10x coverage) with the latter having similar coverage to the rest of the presumed nuclear genome. Furthermore, 183 184 scaffold 102,807 was 17Kb in length (the expected size of the mtDNA genome) whereas scaffold 104,071 was longer than expected at 24Kb. These lines of evidence clearly show 185 186 scaffold 102,807 contains the L. acervorum mtDNA genome whereas scaffold 104,071 is a transposition of mtDNA sequences to the nuclear genome (a NUMT). 187 To genotype single individuals in the six populations (V, OT, L, NR, PF, and SD), 188 short-read sequence data (Illumina HiSeq 2x150bp paired-end reads) were, after quality 189 control steps, aligned to the draft genome with Bowtie2 2.3.5 (Langmead and Salzberg, 190 2014) and processed with SAMtools (Li et al., 2009) to produce bam files. Bam files were 191 then subset to only include the identified mtDNA scaffold (scaffold: 102,807) with 192 SAMtools. These mtDNA alignments were converted to mpileup with BCFtools (--max-193 194 depth 1000) and BCFtools call used to produce vcf files. Vcf files were indexed and normalized and variants within 5bp of any indels removed with BCFtools. Finally, a fasta 195

196 file for each alignment was produced with BCFtools consensus (Supplementary

197 Information, Online Methods).

In addition, mitochondrial genomes of the taxa within the *Formicoxenus* genusgroup were extracted and assembled from ultra-conserved elements (UCE) libraries from
previous studies (Branstetter et al., 2017; Prebus, 2017) using MitoFinder (Allio et al.,
2020). Outgroup species within subfamilies Myrmicinae, Dolichoderinae and Ponerinae
were downloaded from Genbank, previously published in several studies (Cicconardi et al.,
2020; Du et al., 2019; Duan et al., 2016; Gotzek et al., 2010; Hasegawa et al., 2011; Liu et

al., 2016; Park et al., 2021, 2020b, 2020a, 2019; Rodovalho et al., 2014). SRA sequences
and the assembled mitochondrial genomes of *L. acervorum* are deposited in the NCBI
Bioproject PRJNA634471.

207

208 2.3 Phylogenomic analyses using whole mitochondrial genomes

All 49 whole mitochondrial genomes were aligned using MAFFT ver. 7.310 (Katoh and Kuma, 2002) with default parameters. Visual inspection and further adjustment were performed with AliView (Larsson, 2014) and summary statistics of the alignment were obtained with AMAS (Borowiec, 2016). Phylogenetic analysis was performed with maximum likelihood (ML) as implemented in IQ-TREE 1.6.1 (Nguyen et al., 2015) with ultrafast likelihood bootstrap with 1000 replicates. The final tree was visualized and edited with FigTree (Rambaut, 2016).

216

2.4 DNA extraction, PCR amplification and sequencing of cytochrome c oxidase (COI) 217 218 We collected either pupae or adults of workers, males, or queens from different 219 colonies of H. sublaevis, L. acervorum, L. kutteri (Buschinger, 1965) and L. muscorum 220 (Table S2). DNA was extracted from legs or whole specimens using the salt extraction method (Aljanabi and Martinez, 1997) and we sequenced the portion of the mitochondrial 221 COI using previous primers and PCR conditions (Folmer et al., 1994). The sequences 222 obtained were edited, visually inspected using Sequencher (Gene Codes), and aligned with 223 AliView (Larsson, 2014). COI sequences are deposited in the NCBI Bioproject 224 225

226

227 2.5 Phylogenetic analyses and dating estimation within Formicoxenus-Leptothorax based228 on the COI gene region

To infer the phylogenetic relationships within the Formicoxenus-Leptothorax, we 229 230 used the 5' region of COI (658 bp, ranging from 5442-6601 in the L. acervorum 231 mitochondrial genome assembly). First, we performed an explorative analysis based on a comprehensive sampling from this gene region using the sequences generated in this study 232 233 and available sequences from GenBank and BOLD: The Barcode of Life Data System 234 (www.barcodinglife.org). The matrix was aligned and manually edited with AliView 235 (Larsson, 2014), with summary statistics obtained with AMAS (Borowiec, 2016). This preliminary analysis was based in a total of 747 specimens of *Formicoxenus* (2 spp.), 236

237 Leptothorax (8 spp.) and specimens of H. sublaevis (outgroup). We used maximum

likelihood (ML) as implemented in IQ-TREE 1.6.1 (Nguyen et al., 2015) with ultrafast 238 239 likelihood bootstrap with 1000 replicates (Minh et al., 2013). Based on the results from this 240 analysis (data not shown), we selected representative specimens of the major Nearctic lineages identified (>80 bootstrap support) and all the specimens in the Palearctic lineage 241 of L. acervorum. This final dataset consisted of 96 specimens representing eight 242 Leptothorax spp., specimens labelled as Leptothorax sp., L. muscorum complex, 243 Leptothorax sp. AF CAN, two Formicoxenus spp. and H. sublaevis as an outgroup. This 244 dataset consisted of the newly generated sequences in this study and available sequences 245 from previous publications (Hebert et al., 2016; Prebus, 2017; Schär et al., 2018; Smith et 246 al., 2009; Stahlhut et al., 2013) (Table S2). The best nucleotide substitution model and ML 247 248 analysis were inferred with IQ-TREE 1.6.1. Clade support was assessed with ultrafast likelihood bootstrap with 1000 replicates. In addition, we also performed a Bayesian 249 250 inference (BI) as implemented in MrBayes 3.2.6 (Huelsenbeck and Ronquist, 2001; Ronquist et al., 2012) with four chains, two runs of 20 million generations with the 251 252 invgamma rate of variation, the GTR+ Γ model of nucleotide substitution and a sample 253 frequency of 1000. We used TRACER 1.7 (Rambaut et al., 2018) to verify whether effective 254 samples sizes (ESS values) were higher than 200 for all parameters. To estimate divergence times among the lineages in Formicoxenus-Leptothorax, we 255 256 used a simplified dataset representing the same number of species as above, but fewer specimens (63) of these two genera. We used BEAST 1.10.4 (Bouckaert et al., 2014; 257 Suchard et al., 2018) with a strict clock model and a constant population size under a 258 coalescence model. We employed the divergence time estimated in the Formicinae 259 260 (Blaimer et al., 2015) by placing a prior in the divergence estimate of *Harpagoxenus* and Formicoxenus-Leptothorax of 8.89 (13.89-3.89) Ma. We ran two independent runs of 50 261 262 million generations each, sampling values every 1,000 steps. Output files were analyzed with TRACER 1.7 to assess chain convergence and LogCombiner 1.10.4 was used to combine 263

independent runs. Finally, we used Treeannotator 1.10.4 to generate the maximum-clade-

- credibility tree. ML and BI consensus trees were visualized and edited with FigTree
- 266 (Rambaut, 2016).
- 267

268 2.6 *Phylogeography and genetic diversity of* Leptothorax acervorum

269 To gain further insights into the geographic distribution of genetic diversity of *L*.

acervorum across its Holarctic distribution, we first determined the number of haplotypes,

haplotype diversity (*Hd*) and defined haplotypes with DnaSP ver. 6.12 (Rozas et al., 2017).

Then, we reconstructed the haplotype network of all 113 specimens (Table S3) using the 272 273 COI-5P gene region (647 bp, dataset 3) with the statistical parsimony network using TCS 274 (Clement et al., 2002) as implemented in popart ver. 1.7 (Leigh and Bryant, 2015). Given that our interest was focused on the populations distributed in the Iberian Peninsula, we 275 explored in more detail five populations from this region and one population from England 276 (Table S3). In these analyses we used the mitochondrial region encompassing COI, trnL2 277 and COII comprising 2,293 bp (dataset 4). Haplotype (h), polymorphic sites and 278 nucleotide diversities (π) were calculated using the program DnaSP ver. 6.12. We 279 employed Fu's Fs and Tajima's D tests of selective neutrality to determine whether L. 280 acervorum populations from the Iberian Peninsula and England could have experienced 281 282 recent expansions. Fu's F is based in the infinite-site model and a population expansion increase of rare alleles in the population, leading to negative values (Fu, 1997). Tajima's D 283 284 uses the frequency of segregating nucleotide sites and the average number of nucleotide differences obtained from pairwise comparisons (Tajima, 1989). Deviations from neutrality 285 286 could indicate the effect of selection and/or population size changes. Population expansions 287 will increase rare alleles in the population, leading to values < 0, while population 288 contractions (bottlenecks) will increase intermediate variants in the population. COI, trnL2 and COII sequences are deposited in the NCBI Bioproject PRJNA634471, under 289 290 accessions XXXXXXXX-XXXXXXXX.

291

3. Results 292

3.1. Phylogenomic analyses of the Formicoxenus genus-group 293

294 The final alignment of the mitochondrial genomes consisted of 14,351 bp with 13.32 % 295 missing data, 69% of sites variable and 59% of sites parsimony informative (Supplementary 296 Information, Data S1). We recovered monophyletic lineages for all the tribes, except Attini Smith, 1858 within Myrmicinae, with most branches having moderate (>75%) to high (>85%) bootstrap 297 298 support. Our phylogenetic analysis recovered all six genera of the *Formicoxenus* genus-group as a 299 monophyletic lineages within Crematogastrini, with Formicoxenus as the most closely related genus to Leptothorax. The most closely related tribe was Solenopsidini Forel, 1893 (Fig.1). 300 301

- 302 3.2 Relationships within Leptothorax and divergence times of the Palearctic species

303 The alignment matrix of the COI-5P region (dataset 2) consisted of 658 bp with 200 variable sites (30.4 %), 148 parsimony informative sites (22.5%) and 1.42% of missing data. We 304 305 recovered the species of Formicoxenus on different lineages within Leptothorax, suggesting that

the former genus might not represent a monophyletic lineage. All the three Palearctic Leptothorax 306 307 species we included (L. muscorum, L. gredleri Mayr, 1855 and L. kutteri) represent monophyletic 308 lineages, whereas specimens assigned to L. muscorum from the Nearctic region represent several 309 undescribed taxa (Fig. 2 and Fig. S1). Similarly, we found non-monophyletic lineages for the other Nearctic species L. canadensis Provancher, 1887 and L. calderoni Creighton, 1950, but not for L. 310 311 *retractus* Francoeur, 1986. Our divergence estimate suggests that the stem age lineages of Palearctic taxa (L. gredleri, L. muscorum, L. kutteri and L. acervorum) ranges between 1-1.6 Ma 312 313 (Fig. 3). The crown age of the Palearctic lineage of L. acervorum was estimated at 0.56 Ma, with 314 specimens from the Iberian Peninsula ranging in age between 0.1 and 0.5 Ma. The most recent derived lineage (0.30 Ma) within L. acervorum comprises both specimens from the Nearctic and 315 316 Palearctic distribution, including specimens at high latitudes mainly from the West Nearctic (Fig. 4). 317

318 *3.3 Phylogeography of* L. acervorum *in the Holarctic region and genetic diversity in the*

319 *populations of the Iberian Peninsula*

Our analyses based on the COI-5P region (dataset 3, excluding gaps and missing data). 320 321 recovered 21 variable sites with 19 haplotypes (Hd = 0.826) among the 113 specimens examined 322 (Table S3). Only two populations from the Iberian Peninsula (Larra and Pla de la Font) shared haplotypes with the rest of the populations in the West Palearctic, East Palearctic and the Nearctic. 323 We also found a unique haplotype (H4) shared between the population in England and 324 Switzerland, and the presence of unique haplotypes in Bulgaria and Kamchatka. The most widely 325 distributed haplotype (H11) was shared across the entire geographic distributional range (Table 1), 326 and it likely represents the most recent expansion across the distribution of L. acervorum. The 327 328 haplotype network indicates that most populations in the Iberian Peninsula have been isolated from 329 the remaining range of distribution in the West Palearctic, with a recent expansion of the haplotype 330 H11 into this region (Fig. 5). Our examination of the genetic diversity in the populations in the Iberian Peninsula based on a larger segment of the mitochondrial COI-*trnL2*-COII (dataset 4) 331 suggests higher genetic diversity in Niela refuge and Larra than in the other Iberian populations 332 333 (Table 2).

334 4. Discussion

4.1 Congruence between phylogenomic inferences based on mitochondrial genomes and UCEs
 Recent analyses using large sequence datasets from UCEs have been employed to resolve
 relationships among ant subfamilies (Blaimer et al., 2015; Branstetter et al., 2017; Li et al., 2018;
 Longino and Branstetter, 2021). Phylogenomic inferences using mitochondrial genomes have also

been used in several ant subfamilies with mostly congruent topologies between nuclear (UCEs) 339 340 and mitochondrial genomes (Allio et al., 2020). Although we did not have a comprehensive mitochondrial representation within each of the six tribes currently recognized within Myrmicinae 341 (Borowiec et al., 2020), the topology we recovered from this analysis (Fig. 1) is congruent with the 342 known relationships among these tribes inferred from nuclear genes (Ward et al., 2015; 343 https://antwiki.org/wiki/Phylogeny of Myrmicinae). Crematogastrini harbors 40% of all 344 Myrmicinae species and 45% of the genera belong to this tribe (Blaimer et al., 2018). Recent 345 phylogenomic analyses based on UCEs (Blaimer et al., 2018; Prebus, 2017), as well as 346 347 mitochondrial genomes (Park et al., 2021) have been used to increase resolution among genera of of Crematogastrini. Analyses using UCEs have recovered eight clades with high support within 348 349 Crematogastrini, which have been treated as informal genus-groups. 350 The Formicoxenus genus group consists of six genera (Formicoxenus, Leptothorax,

351 *Vombisidris, Gauromyrmex, Harpagoxenus* and *Temnothorax*) and the relationships among the genera are relatively well established (Blaimer et al., 2018; Prebus, 2017), but less attention has 352 353 been paid to the Leptothorax genus group, which consists of Formicoxenus, Leptothorax and 354 Harpagoxenus. Species within these genera are prone to develop social parasitism, a set of 355 interrelated lifestyles where the parasitic species depend upon a free-living host to complete their life cycle (Beibl et al., 2005; Heinze, 1995). These three genera, together with Temnothorax, are 356 considered a hot spot for the evolution of social parasitism, where it has evolved at least five times 357 among closely related taxa (Beibl et al., 2005; Jongepier et al., 2021; Prebus, 2017). 358 Formicoxenus comprises about seven species of Nearctic or Palearctic distribution, with some 359 species ("guest ants") living in the nest of Formica Linnaeus, 1758, Myrmica Latreille, 1804 or 360 Manica Jurine, 1807 species. Workers of Formicoxenus species beg for food from their host 361 species, but rear their own brood in own chambers within the host nest (Franceur et al., 1985; 362 363 Heinze, 1995). Leptothorax contains some species that are workless social parasites that tolerate (inquilines, e.g., L. kutteri, L. pacis (Kutter, 1945) or kill the host queen (murder parasites, e.g., L. 364 goesswaldi Kutter, 1967, L. wilsoni Heinze, 1989) of other closely related Leptothorax species 365 366 (Foitzik et al., 2009; Heinze and Ortius, 1991). In contrast, Harpagoxenus taxa are slave-making (dulotic) ants whose queens kill or expel all adult residents after invading Leptothorax spp. (Brandt 367 368 et al., 2007; Fischer-Blass et al., 2006; Heinze and Ortius, 1991; Pusch et al., 2006). 369

- 370
- 371

4.2 Evidence on non-monophyletic taxa within Leptothorax and monophyletic lineage within L. acervorum

Mitochondrial genes (cytochrome b and cytochrome oxidase subunit 1) alone or in 374 combination with other nuclear markers have been previously used in phylogenetic inferences in 375 376 Leptothorax (Baur et al., 1996, 1995; Beibl et al., 2005; Heinze and Gratiashvili, 2015; Schär et al., 2018), but with limited representation of its species or without using other genera within the 377 378 Formicoxenus genus-group. Our more comprehensive sampling (42% of Leptothorax species, AntWeb ver. 8.42, https://www.antweb.org, accessed 29 October 2020 and multiple accessions of 379 380 L. acervorum) supports a close relationship between Formicoxenus and Leptothorax (Fig. 2), similar to previous analyses based on limited taxa of this group (Blaimer et al., 2018; Prebus, 381 382 2017; Ward et al., 2015). As it has been previously suggested (Heinze and Gratiashvili, 2015; Heinze and Ortius, 1991; Schär et al., 2018), some taxa within the genus *Leptothorax*, particularly 383 384 the Nearctic ones, represent species groups that deserve taxonomic adjustments. Our analyses suggest the presence of at least four Nearctic lineages (monophyletic groups with moderate to high 385 386 support) comprising taxa currently assigned to L. muscorum (Nearctic), L. canadensis, L. calderoni, L. sphagnicola and specimens assigned to the L. muscorum complex. This latter 387 complex seems to consists of a species group of three to four different taxa from the Nearctic that 388 display a set of similar morphological characters and chromosome numbers (Brown, 1955; Heinze, 389 1991, 1989; Loiselle et al., 1990). One of these lineages also includes a specimen of L. acervorum 390 (LT977594), which is more closely related to the Palearctic taxa L. muscorum, L. gredleri and L. 391 392 kutteri (Fig. 2). This latter lineage deserves further exploration, as it involves determining whether L. acervorum in the Nearctic represents a separate lineage from the remaining samples we 393 included. 394

In contrast, the Palearctic species *L. gredleri*, *L. muscorum* and the inquiline *L. kutteri* most likely represent monophyletic lineages (Fig. 2). The lineage of *L. acervorum* comprises both specimens from the Palearctic and Nearctic regions and our divergence age estimates suggest that this clade likely represents the most recent diversification event, within the last 0.5 Ma (Fig. 3). Despite the high support values observed for most of the lineages in the Palearctic region, the support values for the mutual relationships of the lineages were low, and therefore more informative regions will be necessary to determine their relationships.

- 403

404 *4.3 Evidence of isolated populations in the Iberian Peninsula with limited contribution to the most*405 *recent expansion of* L. acervorum

Leptothorax acervorum is one of the only three ant species with Holarctic distribution 406 (Schär et al., 2018); however, all current phylogeographic analyses of this species based on 407 alloenzymes, SSRs, mtDNA and nuclear markers have included only Palearctic specimens (Brandt 408 et al., 2007; Foitzik et al., 2009; Gill et al., 2009; Heinze et al., 1994; Stille and Stille, 1993; 409 Trettin et al., 2016), thus providing an incomplete picture of the patterns of recolonization and 410 411 populations structure. Previous phylogeographic analyses within L. acervorum species have been 412 based on the COI 3'P region and indicate substantial genetic diversity within the species (Brandt et 413 al., 2007; Foitzik et al., 2009; Trettin et al., 2016). The most recent analyses based on SSRs and 414 mtDNA (COI 3'P region) have found the existence of multiple refugia in SW-Europe, and evidence of spatial genetic structure across the sampled area (Trettin et al., 2016). Our most 415 416 extensive data set, including specimens we identified in the previous phylogenetic analyses (Fig. 2) 417 from the Nearctic region and several populations from the Iberian Peninsula (IP), suggests that the 418 IP populations represent the less derived lineages and that they might have experienced 419 fragmentation and isolation from the remaining Holarctic distribution (Fig. 5). Leptothorax 420 acervorum is a cold-adapted species that in the IP inhabits mostly mountainous pinewoods and pine-dominated forest (Pinus sylvestris) above 1500 m.a.s.l (Felke and Buschinger, 1999; Gill et 421 al., 2009). Our results indicate that all populations we included within the IP, except Larra and Pla 422 de la Font, seem to have been more isolated from the remaining range of distribution of this 423 species (Fig. 4), supporting previous evidence based on SSRs, which have found evidence of 424 425 bottlenecks and varying levels of connectivity in this area (Trettin et al., 2016). However, there are 426 only a few mutations separating even the most divergent haplotypes among these populations, but 427 these divergent haplotypes in the IP tend to be found in altitudinally restricted populations. In 428 contrast, the most common haplotype (H11) is found in locations were L. acervorum is not altitudinally restricted (Fig. 5) and there is greater connectivity of suitable habitat. The lack of 429 spatial genetic structure previously reported within L. acervorum using mtDNA (Brandt et al., 430 431 2007; Foitzik et al., 2009; Trettin et al., 2016) might be explained by the limited sampling outside the West-Palearctic regions in previous studies, as well as that this lineage represents the group 432 433 with the most recent expansion (Fig. 3). Additional sampling across the Holarctic distribution with 434 denser sampling among populations, together with the inclusion of additional markers, would be 435 required to further expand the phylogeographic signal we recovered in our analyses. Several refugia areas have been identified in the IP based on the ant species in this region 436

(Tinaut and Ruano, 2021), and our results suggests that only populations from the Pyrenean

437

13

refugia might have more recent connection with the rest of the West Palearctic range of 438 439 distribution (Fig. 4). In contrast, the populations located in the Cantabric and the Northern Plateu 440 (Tinaut and Ruano, 2021) were likely more isolated from the rest of the populations. Cold-adapted species (boreal) with wide distribution in the Palearctic could survive in periglacial areas during 441 the periods of maximum glacial expansion (e.g., during the LGM, 23-18 ka BP), expanding their 442 range into southern areas. During periods of postglacial warming, southern populations of these 443 species became isolated in mountainous regions (Schmitt, 2009; Schmitt and Varga, 2012), 444 surviving in southern refugia (Stewart et al., 2010). There is extensive evidence of the glacial-445 446 interglacial cycles during the Quaternary having influenced the individual genetic diversity and 447 population structure of plants and animals in the West Palearctic (Bennett et al., 1991; Morales-448 Barbero et al., 2018; Schmitt and Varga, 2012; Stewart et al., 2010), including the presence of several periglacial and southern refugia of cold-tolerant of *Myrmica* (Leppänen et al., 2013, 2011) 449 450 and Formica ant species (Goropashnaya et al., 2007, 2004). Emerging evidence seems to indicate 451 that these glacial-interglacial cycles could also have shared refugia; for example, the congruent 452 phylogeographic signal between *Myrmica* ants and *Betula* species (Leppänen et al., 2011; 453 Maliouchenko et al., 2007), the leaf beetle Gonioctena intermedia and its boreal-temperate host 454 trees Prunus padus and Sorbus aucuparia (Quinzin et al., 2017), and the similar patterns of 455 isolated populations in the Iberian Peninsula observed between L. acervorum (Trettin et al., 2016)

and *Pinus sylvestris* (Dering et al., 2017; Tyrmi et al., 2020).

457

458 Acknowledgements

The phylogenomic analyses based on whole mitochondrial genomes were performed on resources
provided by UNINETT Sigma2 - the National Infrastructure for High Performance Computing and
Data Storage in Norway. Funding was provided by NIBIO under the ForGeBiM project. MJ and

- 462 RLH's contribution were funded by BBSRC MIBTP PhD studentship to MJ supervised by RLH.
- 463 We thank Mari Mette Tollefsrud for drawing the map in Fig. 4.
- 464

465 Author Contribution

466 DIO, TK, JM and RB conceived the idea; DIO, KV, RS and JM performed the laboratory and

- 467 analyses; MJ and RLH assembled the *L. acervorum* mtDNA genome, identified the mtDNA
- 468 genomic scaffold and genotyped re-sequenced samples; DIO wrote the manuscript with
- 469 contributions from all authors.
- 470
- 471

References

473 474	Aljanabi, S.M., Martinez, I., 1997. Universal and rapid salt-extraction of high quality genomic DNA for PCR-based techniques. Nucleic Acids Res. 25, 4692–4693.
475	Allio, R., Schomaker-Bastos, A., Romiguier, J., Prosdocimi, F., Nabholz, B., Delsuc, F., 2020.
476	MitoFinder: Efficient automated large-scale extraction of mitogenomic data in target
477	enrichment phylogenomics. Mol. Ecol. Resour. 20, 892–905.
478	Baur, A., Chalwatzis, N., Buschinger, A., Zimmermann, F., 1995. Mitochondrial DNA sequences
479	reveal close relationships between social parasitic ants and their host species. Curr. Genet. 28,
480	242–247.
481 482 483 484	 Baur, A., Sanetra, M., Chalwatzis, N., Buschinger, A., Zimmermann, F., 1996. Sequence comparisons of the internal transcribed spacer region of ribosomal genes support close relationships between parasitic ants and their respective host species (Hymenoptera: Formicidae). Insectes Soc. 43, 53–67.
485	Beibl, J., Buschinger, A., Foitzik, S., Heinze, J., 2007. Phylogeny and phylogeography of the
486	Mediterranean species of the parasitic ant genus Chalepoxenus and its Temnothorax hosts.
487	Insectes Soc. 54, 189–199.
488	Beibl, J., Stuart, R.J., Heinze, J., Foitzik, S., 2005. Six origins of slavery in formicoxenine ants.
489	Insectes Soc. 52, 291–297.
490 491	Bennett, K.D., Tzedakis, P.C., Willis, K.J., 1991. Quaternary Refugia of North European Trees. J. Biogeogr. 18, 103–115.
492	Berman, D.I., Alfimov, A.V., Zhigulskaya, Z.A., Leirikh, A.N., 2010. Overwintering and cold-
493	hardiness of ants in the northeast of Asia. Pensoft, Sofia, Moscow.
494 495 496	 Blaimer, B.B., Brady, S.G., Schultz, T.R., Lloyd, M.W., Fisher, B.L., Ward, P.S., 2015. Phylogenomic methods outperform traditional multi-locus approaches in resolving deep evolutionary history: A case study of formicine ants. BMC Evol. Biol. 15, 1–14.
497	Blaimer, B.B., Ward, P.S., Schultz, T.R., Fisher, B.L., Brady, S.G., 2018. Paleotropical
498	diversification dominates the evolution of the hyperdiverse ant tribe Crematogastrini
499	(Hymenoptera: Formicidae). Insect Syst. Divers. 2.
500	Borowiec, M., Moreau, C., Rabeling, C., 2020. Ants: phylogeny and classification, in: Starr, C.

501 (Ed.), Encyclopedia of Social Insects. Springer, Switzerland, pp. 1–18.

- Borowiec, M.L., 2016. AMAS: a fast tool for alignment manipulation and computing of summary
 statistics. PeerJ 4, e1660.
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard, M.A., Rambaut,
 A., Drummond, A.J., 2014. BEAST 2: A software platform for Bayesian evolutionary
 analysis. PLoS Comput. Biol. 10, e1003537.
- Brandt, M., Fischer-Blass, B., Heinze, J., Foitzik, S., 2007. Population structure and the coevolution between social parasites and their hosts. Mol. Ecol. 16, 2063–2078.
- Branstetter, M.G., Longino, J.T., Ward, P.S., Faircloth, B.C., 2017. Enriching the ant tree of life:
 enhanced UCE bait set for genome-scale phylogenetics of ants and other Hymenoptera.

511 Methods Ecol. Evol. 8, 768–776.

- 512 Brown, W., 1955. The ant Leptothorax muscorum (Nylander) in North America. Entomol. News.
- Cicconardi, F., Krapf, P., D'Annessa, I., Gamisch, A., Wagner, H.C., Nguyen, A.D., Economo,
 E.P., Mikheyev, A.S., Guenard, B., Grabherr, R., Andesner, P., Wolfgang, A., Di Marino, D.,
 Steiner, F.M., Schlick-Steiner, B.C., 2020. Genomic signature of shifts in selection in a
 subalpine ant and its physiological adaptations. Mol. Biol. Evol. 37, 2211–2227.
- 517 Clement, M., Snell, Q., Walke, P., Posada, D., Crandall, K., 2002. TCS: estimating gene
 518 genealogies, in: Proceeding 16th International Parallel Distributed Processing Symposium. p.
 519 184.
- 520 Czechowski, W., Radchenko, A., Czechowska, W., Vepsäläinen, K., 2012. The ants of Poland
 521 with reference to the myrmecofauna of Europe. Muzeum i Instytut Zoologii Polskiej
 522 Akademii Nauk.
- 523 DeChaine, E.G., 2008. A bridge or a barrier? Beringia's influence on the distribution and diversity
 524 of tundra plants. Plant Ecol. Divers. 1, 197–207.
- Dering, M., Kosiński, P., Wyka, T.P., Pers–Kamczyc, E., Boratyński, A., Boratyńska, K., Reich,
 P.B., Romo, A., Zadworny, M., Żytkowiak, R., Oleksyn, J., 2017. Tertiary remnants and
 Holocene colonizers: Genetic structure and phylogeography of Scots pine reveal higher
 genetic diversity in young boreal than in relict Mediterranean populations and a dual
 colonization of Fennoscandia. Divers. Distrib. 23, 540–555.

- Du, Y., Song, X., Yu, H., Lu, Z., 2019. Complete mitochondrial genome sequence of Tapinoma
 melanocephalum (Hymenoptera: Formicidae). Mitochondrial DNA Part B Resour. 4, 3448–
 3449.
- Duan, X.Y., Peng, X.Y., Qian, Z.Q., 2016. The complete mitochondrial genomes of two globally
 invasive ants, the Argentine ant Linepithema humile and the little fire ant Wasmannia
 auropunctata. Conserv. Genet. Resour. 8, 275–277.
- Feldmeyer, B., Elsner, D., Alleman, A., Foitzik, S., 2017. Species-specific genes under selection
 characterize the co-evolution of slavemaker and host lifestyles. BMC Evol. Biol. 17, 1–11.
 https://doi.org/10.1186/s12862-017-1078-9
- Felke, M., Buschinger, A., 1999. Social organization, reproductive behavior and ecology of
 Leptothorax acervorum (Hymenoptera, Formicidae) from the Sierra de Albarracin in central
 Spain. Insectes Soc. 46, 84–91.
- Fischer-Blass, B., Heinze, J., Foitzik, S., 2006. Microsatellite analysis reveals strong but
 differential impact of a social parasite on its two host species. Mol. Ecol. 15, 863–872.
- Foitzik, S., Bauer, S., Laurent, S., Pennings, P.S., 2009. Genetic diversity, population structure and
 sex-biased dispersal in three co-evolving species. J. Evol. Biol. 22, 2470–2480.
- Folmer, O., Black, M., Hoeh, W., Lutz, R., Vrijenhoek, R., 1994. DNA primers for amplification
 of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Mol.
 Mar. Biol. Biotechnol. 3, 294–299.
- Franceur, A., Loiselle, R., Buschinger, A., 1985. Biosystématique de la tribu Leptothoracini
 (Formicidae, Hymenoptera). 1. Le genre Formicoxenus dans la région holarctique. Nat. Can.
 112, 343–403.
- Fu, X., 1997. Statistical tests of neutrality of mutations against population growth, hitchhiking and
 background selection. Genetics 147, 915–925.
- Gill, R.J., Arce, A., Keller, L., Hammond, R.L., 2009. Polymorphic social organization in an ant.
 Proc. R. Soc. B Biol. Sci. 276, 4423–4431.
- Goropashnaya, A., Fedorov, V., Seifert, B., Pamilo, P., 2007. Phylogeography and population
 structure in the ant Formica exzecta (Hymenopthera, Formicidae) across Eurasia as reflected
 by mitochondrial DNA variation and microsatellites. Ann. Zool. Fenn. 44, 462–474.

- Goropashnaya, A., Fedorov, V., Seifert, B., Pamilo, P., 2004. Limited phylogeographical structure
 across Eurasia in two red wood ant species Formica pratensis and F. lugubris (Hymenoptera,
 Formicidae). Mol. Ecol. 13, 1849–1858.
- Gotzek, D., Clarke, J., Shoemaker, D., 2010. Mitochondrial genome evolution in fire ants
 (Hymenoptera: Formicidae). BMC Evol. Biol. 10, 300.
- Guénard, B., Weiser, M.D., Gómez, K., Narula, N., Economo, E.P., 2017. The Global Ant
 Biodiversity Informatics (GABI) database: Synthesizing data on the geographic distribution
 of ant species (Hymenoptera: Formicidae). Myrmecological News 24, 83–89.
- Hasegawa, E., Kobayashi, K., Yagi, N., Tsuji, K., 2011. Complete mitochondrial genomes of
 normal and cheater morphs in the parthenogenetic ant Pristomyrmex punctatus
 (Hymenoptera: Formicidae). Myrmecological News 15, 85–90.
- Hebert, P.D.N., Ratnasingham, S., Zakharov, E. V., Telfer, A.C., Levesque-Beaudin, V., Milton,
 M.A., Pedersen, S., Jannetta, P., Dewaard, J.R., 2016. Counting animal species with DNA
 barcodes: Canadian insects. Philos. Trans. R. Soc. B Biol. Sci. 371, 20150333.
- Heinze, J., 1991. Biochemical studies on the relationship between socially parasitic ants and their
 hosts. Biochem. Syst. Ecol. 19, 195–206.

Heinze, J., 1989. A biochemical approach toward the systematics of the Leptothorax
"muscorum"Group in North America (Hymenoptera: Formicidae). Biochem. Syst. Ecol. 17,
595–601.

- Heinze, J., Foitzik, S., Kipyatkov, V.E., Lopatina, E.B., 1998. Latitudinal variation in cold
 hardiness and body size in the boreal ant species Leptothorax acervorum (Hymenoptera,
 Formicidae). Entomol. Gen. 22, 305–312.
- Heinze, J., Gratiashvili, N., 2015. High skew in the Caucasus: functional monogyny in the ant
 Leptothorax scanni. Insectes Soc. 62, 385–392.
- Heinze, J., Holldobler, B., 1994. Ants in the cold. Memorab. Zool. 48, 99–108.
- Heinze, J., Lipski, N., Schlehmeyer, K., Hölldobler, B., 1994. Colony structure and reproduction in
 the ant, Leptothorax acervorum. Behav. Ecol. 4, 359–367.
- Heinze, J., Ortius, D., 1991. Social organization of Leptothorax acervorum from Alaska
 (Hymenoptera: Formicidae). Psyche (New York) 98, 227–240.

- Heinze, J., Stahl, M., Hölldobler, B., 1996. Ecophysiology of hibernation in boreal Leptothorax
 ants (Hymenoptera: Formicidae). Écoscience 3, 429–435.
- Heinze, J. ürgen, 1995. The origin of workerless parasites in Leptothorax (s.str) (Hymenoptera:
 Formicidae). Psyche (Stuttg). 102, 195–214.
- Hewitt, G., 2000. The genetic legacy of the quaternary ice ages. Nature 405, 907–913.
- Hewitt, G.M., 1996. Some genetic consequences of ice ages, and their role in divergence and
 speciation. Biol. J. Linn. Soc. 58, 247–276.
- Huelsenbeck, J.P., Ronquist, F., 2001. MRBAYES: Bayesian inference of phylogeny.
 Bioinformatics 17, 754–755.
- Jongepier, E., Séguret, A., Labutin, A., Feldmeyer, B., Gstöttl, C., Foitzik, S., Heinze, J.,

Bornberg-Bauer, E., 2021. Convergent loss of chemoreceptors across independent origins of
slave-making in ants. bioRxiv 2021.05.11.443570.

- Katoh, M., Kuma, M., 2002. MAFFT: a novel method for rapid multiple sequence alignment based
 on fast Fourier transform. Nucleic Acids Res. 30, 3059–3066.
- Landry, J.F., Nazari, V., Dewaard, J.R., Mutanen, M., Lopez-Vaamonde, C., Huemer, P., Hebert,
 P.D.N., 2013. Shared but overlooked: 30 species of Holarctic Microlepidoptera revealed by
 DNA barcodes and morphology. Zootaxa 3749, 1–93.
- Langmead, B., Salzberg, S., 2014. Fast gapped-read alignment with Bowtie 2. Nat. Methods 9,
 357359.
- Larson, D.J., Nilsson, A.N., 1985. The holarctic species of agabus (sensu lato) leach (coleoptera:
 Dytiscidae). Can. Entomol. 117, 119–130.
- Larsson, A., 2014. AliView: A fast and lightweight alignment viewer and editor for large datasets.
 Bioinformatics 30, 3276–3278.
- Leigh, J.W., Bryant, D., 2015. POPART: Full-feature software for haplotype network construction.
 Methods Ecol. Evol. 6, 1110–1116.
- Leppänen, J., Vepsäläinen, K., Anthoni, H., Savolainen, R., 2013. Comparative phylogeography of
 the ants Myrmica ruginodis and Myrmica rubra. J. Biogeogr. 40, 479–491.
- Leppänen, J., Vepsäläinen, K., Savolainen, R., 2011. Phylogeography of the ant Myrmica rubra
 and its inquiline social parasite. Ecol. Evol. 1, 46–62.

617	Li	H	Handsaker,	B	Wysol	cer A	Fennell	T	Ruan	I	Homer N	Marth	G	Abecasis	G
01/	L/1,	11.,	rianusaner,	, D.	, ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	\mathbf{x}	, I CHICH	, I.	, ivuaii,		, 11011101, 14	, iviai un	U.,	, 11000asis,	U.,

- Durbin, R., 2009. The Sequence alignment/map (SAM) format and SAMtools. Bioinformatics
 25, 2078–2079.
- 620 Li, H., Sosa-Calvo, J., Horn, H.A., Pupo, M.T., Clardy, J., Rabeling, C., Schultz, T.R., Currie,
- 621 C.R., 2018. Convergent evolution of complex structures for ant-bacterial defensive symbiosis
 622 in fungus-farming ants. Proc. Natl. Acad. Sci. U. S. A. 115, 10720–10725.
- Liu, N., Duan, X.Y., Qian, Z.Q., Wang, X.Y., Li, X.L., Ding, M.Y., 2016. Characterization of the
 complete mitochondrial genome of the myrmicine ant Vollenhovia emeryi (Insecta:
 Hymenoptera: Formicidae). Conserv. Genet. Resour. 8, 211–214.
- Loiselle, R., Francoeur, A., Fischer, K., Buschinger, A., 1990. Variations and taxonomic

significance of the chomosome numbers in the Nearctic species of the genus Leptothorax
(s.s.) (Formicidae: Hymenoptera). Caryologica 43, 321–334.

- Longino, J.T., Branstetter, M.G., 2021. Integrating UCE phylogenomics with traditional taxonomy
 reveals a trove of New World Syscia species (Formicidae: Dorylinae). Insect Syst. Divers. 5,
 1–51.
- Maliouchenko, O., Palmé, A.E., Buonamici, A., Vendramin, G.G., Lascoux, M., 2007.
 Comparative phylogeography and population structure of European Betula species, with

particular focus on B. pendula and B. pubescens. J. Biogeogr. 34, 1601–1610.

- Marusik, Y.M., Koponen, S., 2005. A survey of spiders (Araneae) with holarctic distribution. J.
 Arachnol. 33, 300–305.
- Minh, B.Q., Nguyen, M.A.T., Von Haeseler, A., 2013. Ultrafast approximation for phylogenetic
 bootstrap. Mol. Biol. Evol. 30, 1188–1195.

Morales-Barbero, J., Martinez, P.A., Ferrer-Castán, D., Olalla-Tárraga, M., 2018. Quaternary
refugia are associated with higher speciation rates in mammalian faunas of the Western
Palaearctic. Ecography (Cop.). 41, 607–621.

- Nguyen, L.T., Schmidt, H.A., Von Haeseler, A., Minh, B.Q., 2015. IQ-TREE: A fast and effective
 stochastic algorithm for estimating maximum-likelihood phylogenies. Mol. Biol. Evol. 32,
 268–274.
- Park, Jonghyun, Kwon, W., Park, Jongsun, 2019. The complete mitochondrial genome of
 Ectomomyrmex javanus Mayr, 1867 (Hymenoptera: Formicidae). Mitochondrial DNA Part B

- 647 Resour. 4, 1636–1637.
- Park, Jonghyun, Xi, H., Park, Jongsun, 2021. Complete mitochondrial genome of the acrobat ant
 Crematogaster teranishii Santschi, 1930 (Formicidae; Hymenoptera). Mitochondrial DNA
 Part B Resour. 6, 593–595.
- Park, Jonghyun, Xi, H., Park, Jongsun, 2020a. The complete mitochondrial genome of
 Aphaenogaster famelica (Smith, 1874) (Hymenoptera: Formicidae). Mitochondrial DNA Part
 B 5, 492–494.
- Park, Jonghyun, Xi, H., Park, Jongsun, 2020b. The complete mitochondrial genome of Ochetellus
 glaber (Mayr, 1862) (Hymenoptera:Formicidae). Mitochondrial DNA Part B 5, 147–149.
- Petit, R., Aguinagalde, I., De Beaulieu, J., Bittkau, C., Brewer, S., Cheddadi, R., Ennos, R.,
- 657 Fineschi, S., Grivet, D., Lascoux, M., Mohanty, A., Müller-Starck, G., Demesure-Musch, B.,
- Palmé, A., Martín, J.P., Rendell, S., Vendramin, G.G., 2003. Glacial refugia: Hotspots but not
 melting pots of genetic diversity. Science (80-.). 300, 1563–1565.
- Prebus, M., 2017. Insights into the evolution, biogeography and natural history of the acorn ants,
 genus Temnothorax Mayr (Hymenoptera: Formicidae). BMC Evol. Biol. 17, 250.
- Pusch, K., Seifert, B., Foitzik, S., Heinze, J., 2006. Distribution and genetic divergence of two
 parapatric sibling ant species in Central Europe. Biol. J. Linn. Soc. 88, 223–234.
- Quinzin, M.C., Normand, S., Dellicour, S., Svenning, J.C., Mardulyn, P., 2017. Glacial survival of
 trophically linked boreal species in northern Europe. Proc. R. Soc. B Biol. Sci. 284.
- 666 Rambaut, A., 2016. FigTree ver. 1.4.3.
- Rambaut, A., Drummond, A.J., Xie, D., Baele, G., Suchard, M.A., 2018. Posterior summarisation
 in Bayesian phylogenetics using Tracer 1.7. Syst. Biol. 0, 1–5.
- Rodovalho, C.D.M., Lyra, M.L., Ferro, M., Bacci, M., 2014. The mitochondrial genome of the
 leaf-cutter ant Atta laevigata: A mitogenome with a large number of intergenic spacers. PLoS
 One 9, 1–9.
- Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D.L., Darling, A., Höhna, S., Larget, B., Liu,
 L., Suchard, M.A., Huelsenbeck, J.P., 2012. MrBayes 3.2: Efficient Bayesian phylogenetic
 inference and model choice across a large model space. Syst. Biol. 61, 539–542.
- 675 Rozas, J., Ferrer-Mata, A., Sanchez-DelBarrio, J.C., Guirao-Rico, S., Librado, P., Ramos-Onsins,

676 677	S.E., Sanchez-Gracia, A., 2017. DnaSP 6: DNA sequence polymorphism analysis of large data sets. Mol. Biol. Evol. 34, 3299–3302.
678 679 680	 Schär, S., Talavera, G., Espadaler, X., Rana, J.D., Andersen Andersen, A., Cover, S.P., Vila, R., 2018. Do Holarctic ant species exist? Trans-Beringian dispersal and homoplasy in the Formicidae. J. Biogeogr. 45, 1917–1928.
681 682 683	 Schlick-Steiner, B.C., Steiner, F.M., Sanetra, M., Seifert, B., Christian, E., Stauffer, C., 2007. Lineage specific evolution of an alternative social strategy in Tetramorium ants (Hymenoptera: Formicidae). Biol. J. Linn. Soc. 91, 247–255.
684 685	Schmitt, T., 2009. Biogeographical and evolutionary importance of the European high mountain systems. Front. Zool. 6, 1–10.
686 687	Schmitt, T., Varga, Z., 2012. Extra-Mediterranean refugia: The rule and not the exception? Front. Zool. 9, 1–12.
688	Seifert, B., 2018. The ants of central and northern Europe. Lutra, Germany.
689 690	Smith, M.A., Fernandez-Triana, J., Roughley, R., Hebert, P.D.N., 2009. DNA barcode accumulation curves for understudied taxa and areas. Mol. Ecol. Resour. 9, 208–216.
691 692 693 694	 Stahlhut, J.K., Fernández-Triana, J., Adamowicz, S.J., Buck, M., Goulet, H., Hebert, P.D.N., Huber, J.T., Merilo, M.T., Sheffield, C.S., Woodcock, T., Smith, M.A., 2013. DNA barcoding reveals diversity of Hymenoptera and the dominance of parasitoids in a sub-arctic environment. BMC Ecol. 13, 2.
695 696	Stewart, J.R., Lister, A.M., Barnes, I., Dalén, L., 2010. Refugia revisited: Individualistic responses of species in space and time. Proc. R. Soc. B Biol. Sci. 277, 661–671.
697 698 699	Stille, M., Stille, B., 1993. Intrapopulation nestclusters of maternal mtDNA lineages in the polygynous ant Leptothorax acervorum (Hymenoptera: Formicidae). Insect Mol. Biol. 1, 117–121.
700 701	Suchard, M.A., Lemey, P., Baele, G., Ayres, D.L., Drummond, A.J., Rambaut, A., 2018. Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. Virus Evol. 4, vey016.
702 703	Tajima, F., 1989. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics 123, 585–595.
704	Tinaut, A., Ruano, F., 2021. Biogeography of iberian ants (Hymenoptera: Formicidae). Diversity
	22

1-25.

706	Trettin, J., Agrawal, S., Heinze, J., 2016. Phylogeography of social polymorphism in a boreo-
707	montane ant. BMC Evol. Biol. 16, 1–14.
708	Tyrmi, J., Vuosku, J., Acosta, J.J., Li, Z., Sterck, L., Savolainen, O., Pyhäjärvi, T., 2020.
709	Genomics of clinal local adaptation in Pinus sylvestris under continuous environmental and
710	spatial genetic setting. G3 Genes Genomes Genet. 10, 2638–2696.
711	Ward, P.S., Brady, S.G., Fisher, B.L., Schultz, T.R., 2015. The evolution of myrmicine ants:
712	Phylogeny and biogeography of a hyperdiverse ant clade (Hymenoptera: Formicidae). Syst.
713	Entomol. 40, 61–81.
714	Weisenfeld, N.I., Kumar, V., Shah, P., Church, D.M., Jaffe, D.B., 2018. Direct determination of
715	diploid genome sequences. Genome Res. 28, 757–767.
716	
717	Supporting Information
718	Figures
719	Fig. S1. Consensus tree obtained with Bayesian inference as implemented in Mrbayes on the
720	dataset of COI-5P region. The black circle indicates the monophyletic lineage of L. acervorum
721	(plus specimens of L. muscorum complex and Leptothorax sp.) that were later used in the
722	phylogeographic analysis. Values next to the branches represent Bayesian support.
723	
724	Tables
725	Table S1. List of specimens used in the phylogenomic analysis of the Formicoxenus genus-group
726	using whole mitochondrial genomes. The specimens represent the six genera currently recognized
727	in the group and the outgroup species.
728	Table S2. List of specimens employed in the phylogenetic analyses of Formicoxenus -
729	Leptothorax using the cytochrome COI-5P region (658 bp).
730	Table S3. Specimens of <i>L. acervorum</i> used in the phylogeographic and genetic diversity analyses
731	across its distribution range in the Holarctic region.
732	Data S1. Final alignment of the whole mitochondrial genomes of ant species used in the
733	phylogenomic analysis of the Formicoxenus genus-group.
734	
735	Supplementary Methods. Methods for the whole mitochondrial genome assembly.
736	

737 Tables

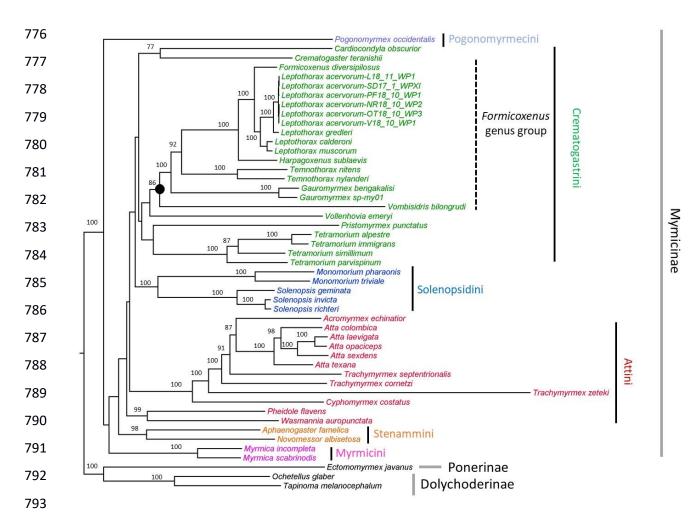
- **Table 1.** Details of the sampling localities included in the phylogeographic analysis of *L*.
- acervorum across its Holarctic distribution and the haplotypes observed in each population. N =
- number of individuals in each locality, BC = British Columbia, NWT = Northwest Territories.

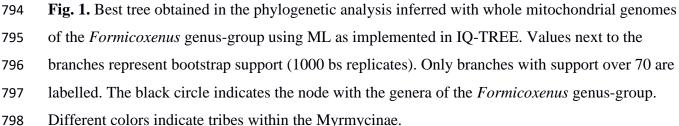
Country	Locality	Ν	Haplotypes
Spain	Cadí-Moixeró N.P.	1	H1
	Niela refuge	16	H14, H15, H18, H17, H19
	Valdelinares	14	H13, H12
	Orihuela de Tremedal	19	H19
	Aragon	1	H16
	Pla de la Font	12	H5
	Larra	12	H11, H8, H7
England	Santom Downham	7	H4, H11
Switzerland	Lausen, Wettingen	2	H4
Denmark	Nærum	1	H11
Norway	Østfold	1	H11
Sweden	Abisco	1	H11
Bulgaria	Panichishte	1	H9
Finland	Turku	1	H11
	Hikia	1	H11
	Lammi	1	H11
	Tvarminne	1	H11
Estonia	Liiva	1	H2
Russia	Kamchatka	2	H3
	Moscow	1	H10
Canada	Nearctic-West (Yukon, BC, NWT)	10	H11
	Nearctic-East (Manitoba)	5	H11

/31

- **Table 2.** Estimates of genetic diversity and neutrality tests obtained in the seven populations of *L*.
- *acervorum* analyzed in detail.

Population	N	Ncol	Polymorphic sites	PI	No. haplotyes	Hd	π	Tajima's D	Fu's F
Spain									
Larra	12	7	5	3	6	0.803	0.00072	-0.0093	-
									1.847
Niela refuge	16	13	16	15	6	0.817	0.00228	0.3364	2.118
Orihuela de	19	11	8	0	2	0.105	0.00037	-2.1619	2.452
Tremedal									
Pla de la	12	11	0	0	-	-	0	-	-
Font									
Valdelinares	14	12	1	1	2	0.363	0.00016	0.3244	0.643
England									
Santon	7	3	15	14	3	0.600	0.00340	1.1546	4.116
Dunham									





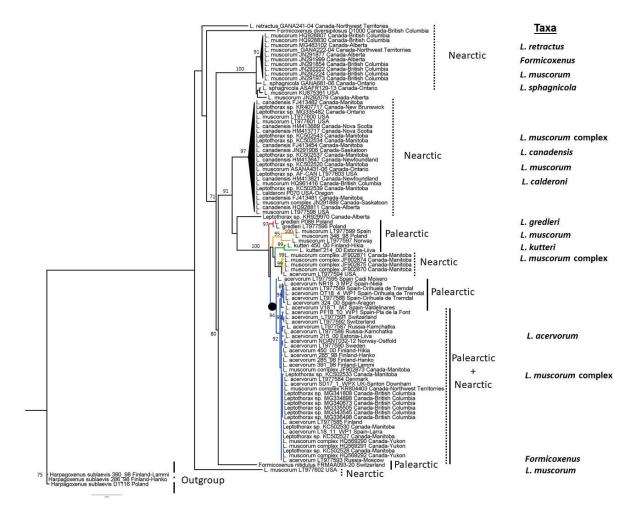


Fig. 2. Best tree recovered of the analysis of Formicoxenus-Leptothorax using ML as implemented in IQ-TREE using the dataset of COI-5P region. The different colours of the branches indicate monophyletic lineages recovered on the species with Palearctic distribution. The black circle indicates the monophyletic lineage of L. acervorum (plus specimens of L. muscorum complex and Leptothorax sp.) that were later used in the phylogeographic analysis. Species currently recognized in these genera are indicated next to the lineages (taxa). Values next to the branches represent bootstrap support (1000 bs replicates). Only branches with >70 bootstrap support are labelled.

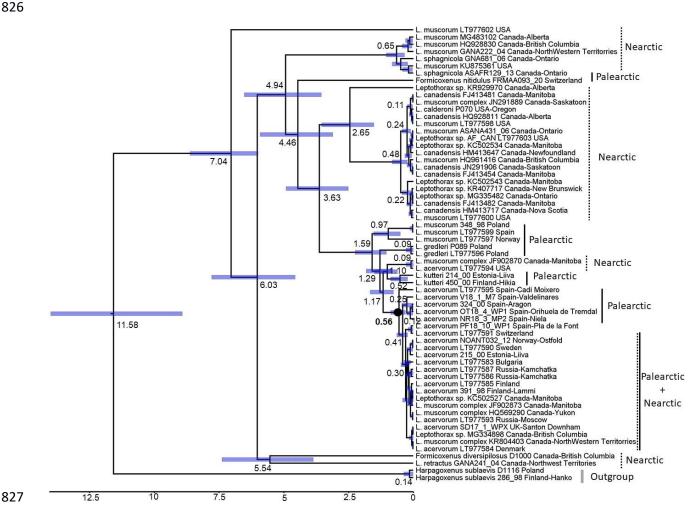


Fig. 3. Chronogram of the divergence times estimated in the *Formicoxenus - Leptothorax* lineages
obtained with Beast. The black circle indicates the monophyletic lineage of *L. acervorum*. Values
next to the branches indicate stem ages with blue columns displaying 95% confidence intervals.

- _ . . .

bioRxiv preprint doi: https://doi.org/10.1101/2021.11.05.467305; this version posted November 5, 2021. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-NC-ND 4.0 International license.

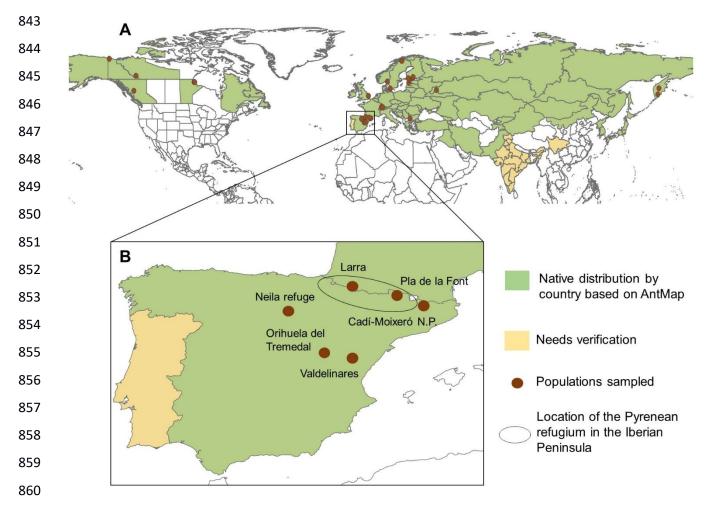


Fig. 4. A) Distribution map of *L. acervorum* across the Holarctic region based on the Global Ant
Biodiversity Informatics (GABI) database (Guénard et al., 2017). Locations of the different
populations included in the phylogeographic analysis are indicated with red dots. B) Close-up of
the populations located in the Iberian Peninsula, indicated with blue circles those populations that
contributed to the most recent expansion in the Holarctic. The black oval indicates the location of
the Pyrenean refugium (Tinaut and Ruano, 2021). N.P. = National Park.

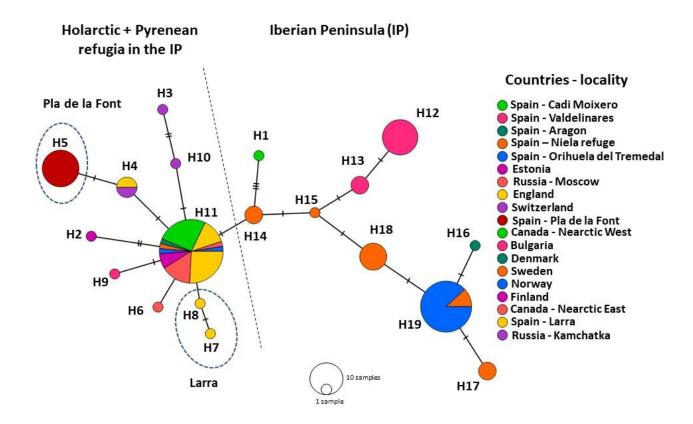


Fig. 5. Haplotype network of *L. acervorum* across its Holarctic distribution range. Codes next to
the circle indicate the haplotype classification and their distribution. Hatch marks represent
mutation differences among the haplogroups. The dash line separates most of the Iberian Peninsula
populations from the rest of distribution. Pla de la Font and Larra (in dash circles) are both located
in the Pyrenean refugia.