Discovery of imidazole-based inhibitors of *P. falciparum* cGMP-dependent protein kinase

Rammohan R. Yadav Bheemanaboina⁺, Mariana Laureano de Souza^{\$}, Mariana Lozano Gonzalez⁺, Shams Ul Mahmood⁺, Tyler Eck⁺, Tamara Kreiss⁺, Samantha O. Aylor[#], Alison Roth[#], Patricia Lee[#], Brandon S. Pybus[#], Dennis J. Colussi[%], Wayne E. Childers[%], John Gordon[%], John J. Siekierka⁺, Purnima Bhanot^{\$*}, David P. Rotella^{+*}

AUTHOR ADDRESS +-Department of Chemistry and Biochemistry and Sokol Institute of Pharmaceutical Life Sciences Montclair State University, Montclair NJ 07043; %-Moulder Center for Drug Discovery Research, Temple University, Philadelphia PA, 19140; \$- Rutgers New Jersey Medical School, Department of Microbiology, Biochemistry and Molecular Genetics, 225 Warren Street, Newark NJ 07103; #-Department of Drug Discovery. Experimental Therapeutics Branch, Walter Reed Army Institute of Research, 503 Robert Grant Avenue, Silver Spring MD 20910.

KEYWORDS: malaria, protein kinase inhibitor, PfPKG inhibitor, cellular activity, P. falciparum drug target.

ABSTRACT: The discovery of new targets for treatment of malaria and in particular those aimed at the pre-erythrocytic stage in the life cycle, advanced with the demonstration that orally administered inhibitors of *Plasmodium falciparum* cGMP-dependent protein kinase (PfPKG) could clear infection in a murine model. This enthusiasm was tempered by unsatisfactory safety and/or pharmaco-kinetic issues found with these chemotypes. To address the urgent need for new scaffolds, this manuscript presents initial structure-activity relationships in an imidazole scaffold at four positions, representative *in vitro* ADME, hERG characterization and cell-based anti-parasitic activity. This series of PfPKG inhibitors has good *in vitro* PfPKG potency, low hERG activity and cell-based anti-parasitic activity against multiple *Plasmodium* species that appears to correlate with *in vitro* potency.

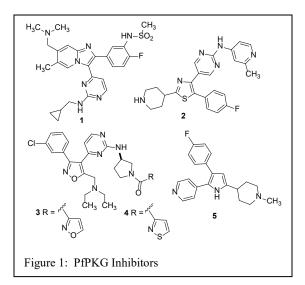
The emergence of artemisinin-resistant *Plasmodium falciparum* in Africa^{1,2} and the slowing decline in deaths from malaria³ signal the need to identify new targets for prophylaxis and treatment.⁴ The pre-erythrocytic portion of the life cycle is an attractive and comparatively under explored point for therapeutic attack because of the very low parasite burden compared to other life cycle stages. Drugs that target these stages are an essential component of the anti-malarial effort because a decrease in liver infection by sporozoites significantly reduces severity and incidence of malaria.⁶ There is a comparative paucity of candidates in this area of anti-malarial drug development.^{5,7}

To address this issue, *Plasmodium falciparum* cGMPdependent protein kinase (PfPKG) is of particular interest because it is essential in pre-erythrocytic, asexual and sexual stages of the parasite.^{8,9,10} Baker and co-workers described the discovery and optimization of an orally bioavailable, potent, selective small molecule inhibitor (**1**, Figure 1). This imidazopyridine cleared infection at a dose of 10 mg/kg orally in a SCID mouse model.^{11,12,13}

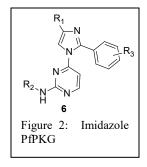
A subsequent report that unspecified examples in this series were Ames positive and limited progression of the scaffold.¹⁴ The Baker group disclosed trisubstituted thiazoles such as **2** (Figure 1) that exhibited rapid killing of *P. falciparum* in culture.¹⁵ Interestingly this desirable property was independent of PfPKG inhibition. Proteomic experiments suggested that inhibition of a serine/arginine protein kinase SRPK2 was a key contributor to rapid parasite killing, comparable to artesunate, a recognized standard. Examples in this series of thiazoles, including **2**, showed single digit micromolar hERG activity and/or *in vitro* metabolic instability limiting their use in more advanced studies.

Important pharmaceutical property and safety issues can be addressed by identifying new chemical matter to provide novel candidates to address this unmet need. We previously reported the discovery of an isoxazole chemotype, exemplified by **3** and **4** (Figure 1).¹⁶ These compounds showed enzymatic potency comparable (IC₅₀s ~20 nM) to known pyrrole **5**¹⁷ *in vitro* against PfPKG, and were not active against human PKG or the T618Q mutant PfPKG¹⁸ at 10 μ M. Parasite expressing this mutant enzyme demonstrate lower sensitivity to PfPKG inhibitors such as **5** but retain enzymatic activity and the ability to proceed through the life cycle ¹⁸.

In the course of the focused screen that identified isoxazole hits, we also identified an imidazole scaffold 6 (Figure 2) that was active in a PfPKG screening assay. We were attracted to the comparatively low molecular weight of this chemotype



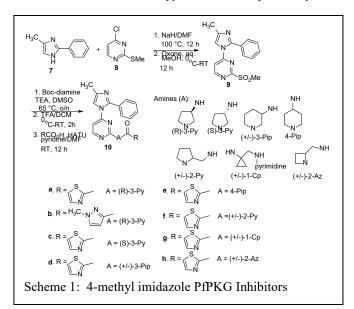
and the potential for optimization at multiple positions that could result in improved properties. Additionally, the synthesis of this class of compounds was significantly shorter than the isoxazole series.



Based on previous experience, we chose to focus first on exploration of the amino substituent (R_2) on the pyrimidine. The synthesis of this set of derivatives is outlined in Scheme 1. Commercially available 4-methyl-2-phenyl imidazole was deprotonated with sodium hydride then treated with 4-chloro-2-methythiopyrimidine in dry DMF at 60-70°C to arylate the

imidazole nitrogen, followed by oxone-mediated conversion to the sulfone. Displacement of the sulfone with a variety of diamines, Boc-deprotection and acylation with preferred carboxylic acids as described previously¹⁶ afforded the target amides **10a-h**.

It is evident from the data in Table 1 that the (R)-3aminopyrrolidine linker is strongly preferred compared to the other cyclic amine variations, a cyclopropyl alkyl diamine and the S-enantiomer of 3-aminopyrrolidine. We previously ob-



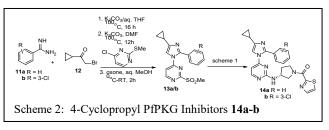
served this stereoselective effect in the isoxazole class of PfPKG inhibitors.¹⁶ In this case, all alternative amine linkers af-

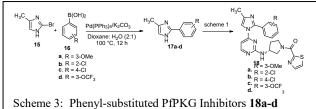
Cpd	IC50 (nM)/% inh @ 1 μM	Cpd	IC50 (nM)/% inh @ 1 μM
10a	320	10e	5%
10b	1000	10f	3%
10c	4%	10g	6%
10d	4%	10h	2%
5	22		
	1: <i>in vitro</i> Pf nethyl imidaz		

forded inactive PfPKG inhibitors whereas with the isoxazole scaffold-selected examples retained some activity.16 This suggests distinct and specific SAR at this point in the structure. An additional difference in SAR for the imidazole scaffold is the more pronounced

potency difference between the 2-thiazolyl and 1-N-methyl-3pyrazolyl amides, with a strong preference for the former. In this group, the most potent derivative (**10a**, PfPKG IC₅₀ 320 nM) underwent further evaluation to provide baseline data for this chemotype on selectivity for human PKG and the T618Q mutant PfPKG. We observed that **10a** had excellent selectivity versus these related PKGs (3-10% inhibition @ 10μ M).

Assessment of *in vitro* ADME characteristics revealed that **10a** was metabolically unstable in human (HLM) and murine (MLM) liver microsomes (half life less than 2 minutes) with excellent water solubility (200 μ M) and moderate CYP3A4 inhibition (IC₅₀ 0.94 μ M). We elected to address the metabolic stability issue first by replacing the 4-methyl group and substituting the aromatic ring because we viewed these as potential site(s) for oxidative metabolism. The 4-methyl group was replaced with a cyclopropyl ring and in view of existing isoxazole SAR with phenyl substituents¹⁶, chose to target an unsubstituted and 3-chlorophenyl derivative. The synthesis, outlined in Scheme 2, condensed appropriate chlorobenzamidines with cyclopropyl bromomethyl ketone **12** to afford 4-cyclopropyl-2-phenyl imidazoles **13a** and **13b** in good yield. Following the steps outlined in Scheme 1, the targets **14a** and **14b** were





obtained in a straightforward manner. In parallel, using the 4-

Cpd	IC ₅₀ (nM)/% inhib @ 1 μM
14a	100
14b	60
18 a	86
18b	47%
18c	1160
18d	150
Table 2:	in vitro
PfPKG	inhibition
14a/b-18a-d	

methylimidazole template, we explored a sampling of aryl substituents on the benzene ring to explore this feature of SAR. The synthesis of these analogs was accomplished as shown in Scheme 3. Suzuki coupling between an appropriate boronic acid and commercially available 2-bromo-4-methyl imidazole afforded the corresponding 2-phenyl derivatives 17a-d that were processed as described in Scheme 1 to afford the respective 2- and 4-chlorophenyl thiazolyl analogs 18b and 18c, respectively as well as the 3-methoxy and 3-trifluoromethoxy targets

18a and **18d**. In both sets of analogs, we elected to use only the optimal 2-thiazolyl amide to provide the best comparison for activity versus **10a-h**.

Evaluation of these compounds as PfPKG inhibitors revealed that the cyclopropyl group in **14a** provides a three-fold improvement in potency compared to **10a** (Table 2) and that 3chloro substitution provides a small additional benefit in **14b**.

Among the group of phenyl substituents evaluated, 3-chloro is preferred to its regioisomers **18b** and **18c**, similar to our previous observations. The other 3-substituted derivatives examined in this small set are comparable (**18a**, **18d**) to **14a** or less potent (**18b**, **18c**).

Cpd	HLM t _{1/2} (min)	MLM t _{1/2} (min)	H2O sol (µM)	3A4 IC50 (μM)	2C9 IC50 (µM)	2D6 IC50 (µM)	hERG % inh. @ 10 µM
14a	4.8	< 2	92	0.29	>10	>10	1
14b	3.7	<2	28	0.19	2.8	>10	9
Table	3: <i>in vit</i>	ro ADM	E Chara	acterizat	tion 14a	-b	

ined for inhibition of human PKG and *P. falciparum* mutant T618Q, and as observed previously, demonstrated excellent selectivity against these two kinases with no inhibition at 10 μ M. These results led us to examine **14a** and **14b** in more detail with a focus on cellular activity and *in vitro* ADME. These imidazole-based PfPKG inhibitors have moderate to good water solubility, and submicromolar inhibition of CYP3A4 (Table 2). Both compounds show poor metabolic stability and metabolite ID studies are underway to guide solutions to this important issue. The positive data in Table 3 shows neither **14a** nor **14b** have measurable hERG activity, unlike the thiazole example in Figure 1.

Cpd	% decr infn ± SD @ 2 μM	% decr infn ± SD @ 10 μΜ	P value
3	52±15	55±21	>0.05
4	0	22±38	>0.05
10a	78±20	89±7	< 0.001
14b	94±3	93±3	< 0.001
5	91±8	94±4	< 0.001
	4: <i>P. bergl</i> vity assay	<i>hei</i> sporozoi	te screening

Using HepG2-P. the berghei Luc (PbLuc) sporozoite infectivity assay and 5 as a positive control, we chose to evaluate 3, 4, 10a and 14b to investigate a correlation between enzymatic and cellular activity. We were encouraged by

the strong activity displayed the imidazoles (Table 4). The more efficacious imidazole, **14b**, like **5**, does not show a dose response (>90% at 2 and 10 μ M) and exhibits comparable activity to **5**. Although it is not definite from the limited concentrations employed in this assay, **14b** appears to be more effica-

cious compared to the **10a** in this initial screen, suggesting a potential correlation between *in vitro* enzymatic inhibition and cellular efficacy. This positive data contrasts with the poor activity of isoxazoles **3** and **4**.

A more quantitative examination of **14a** and

Cpd	EC50 (µM) P. falcipa- rum 3D7	EC50 (μM) <i>PbLuc</i> hepatic stage
5	0.15	0.3
14a	7.6	20
14b	3.3	5.7
	<i>P. falciparum</i> b ei sporozoite EC	U

14b against asexual blood stages (3D7) and PbLuc sporozoite-HepG2 infectivity revealed distinct differences between 5, 14a and 14b (Table 5). This data shows that 5 is more effective in these cellular assays compared to imidazoles 14a and 14b. We note that the approximate 20-fold EC_{50} difference in the PbLuc HepG2 assay between the more efficacious 5 and 14b supports

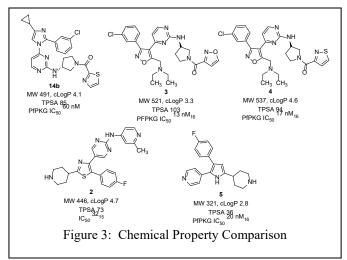
the observation above of a correlation between *in vitro* PfPKG potency and cellular activity in the imidazole series. The modest activity exhibited by **14a** and **14b** in the asexual blood stage assay is not surprising since the asynchronous asexual replication of *P. falciparum* makes PfPKG inhibition less effective at this stage of the life cycle.⁸

This encouraging data led us to examine these imidazoles against additional *Plasmodium* species in other cellbased assays to more completely characterize this series and provide a baseline for future work. Imidazoles **10a** and **14b** were investigated in a dose-response assay that evaluated *P. cynomolgi* infectivity in prophylactic and radical cure modes. These two examples were selected to provide additional evidence to support the hypothesis of a correlation between *in vitro* PfPKG potency and cellular efficacy against other *Plasmodium* species. The prophylactic assay is a measure of the ability of *P. cynomolgi* to form hepatic schizonts or hypnozoites. The radical cure mode evaluates activity against existing schizonts and hypnozoites. The data in Table 6 show that **14b** is more active

	Prophylactic Mode			
Compound	Schizont IC ₅₀ (µM)	Hypnozoite IC ₅₀ (µM)	Toxicity (µM)	
14b	0.93	9.88	>20	
10a	7.91	> 20	>20	
Maduramicin	0.01	0.02	5.78	
Tafenoquine	0.19	0.14	13.82	
	Radical Cure Mode		e	
C I	Schizont IC50	Hypnozoite IC50	Toxicity	
Compound	(μM)	μM)	(µM)	
Compound 14b	- • •	- • •	(µM)	
•	(µM)	(μM)		
14b	(µM) 4.62	(μM) > 20	> 20	

than **10a** in the prophylactic mode with a sub-micromolar IC_{50} , and interestingly has a modest effect in the radical cure mode against schizonts, comparable to tafenoquine. This data is consistent with a dose response effect for both compounds and indicates **14b** is more efficacious than **10a** in cell-based anti-parasitic assays. Efficacy in the prophylactic schizont model is important because it demonstrates potential for interrupting the parasite's liver development post sporozoite invasion of hepatocytes. Reducing liver infection by human-infective sporozoites is known to reduce severity and incidence of malaria.⁶ The data in Table 6also show these two compounds, unlike the controls, are comparatively non-toxic to host cells.

When comparing these new PfPKG inhibitors to known inhibitors (Figure 3), it is not surprising that enzymatic potency, while important, is not the sole determinant for cellular



activity. It is encouraging to note the cellular activity of imidazoles 14a and 14b, in contrast to the more potent isoxazoles. One observation based on data in this paper is that molecular weight contributes to cellular activity; a comparison of 3, 4, 14b and 5 shows that the two highest molecular weight compounds show at best weak activity in cells. There does not appear to be a direct relationship between cLogP and cellular activity (cf. 5, 4 and 14b).

We have shown that **5** and the two isoxazoles are competitive PfPKG inhibitors that bind in the ATP pocket of the enzyme¹⁹. Studies are ongoing with imidazoles such as **14b** to determine to mode of action for this class of compounds, along with investigations into obtaining what would be the initial report of an inhibitor bound to *P. falciparum* PKG.

In conclusion, in response to the need to identify novel chemotypes as PfPKG inhibitors that lack the safety issues associated with many known scaffolds, we report the discovery and initial characterization of a new imidazole-based chemotype with good *in vitro* PfPKG inhibition, and promising cellular activity that includes a correlation between *in vitro* enzymatic activity and efficacy, lacks the hERG issues associated with other chemotypes and does not have any structural alerts associated with genotoxicity. Initial structure-activity relationships are distinct from known PfPKG inhibitors and while the ADME profile of lead **14b** has weaknesses that are not unusual in early leads, the positive aspects of the imidazole series provide the impetus to address the pharmaceutical property issues that currently exist. Those efforts are ongoing and will be reported in due course.

ASSOCIATED CONTENT

Supporting Information

Full experimental details on the synthesis and characterization of compounds, *in vitro* enzyme assay, cellular parasite infectivity and *in vitro* ADME assays are provided along with the manuscript in review cited as reference 19.

The Supporting Information is available free of charge on the ACS Publications website.

Chemistry-synthesis and characterization PDF

Biology: *in vitro* PfPKG assays, cellular parasite infectivity, *in vitro* ADME PDF

AUTHOR INFORMATION

Corresponding Authors

* Contact information: <u>rotellad@montclair.edu</u>; <u>bhanotpu@njms.rutgers.edu</u>

Present Addresses

[†]If an author's address is different than the one given in the affiliation line, this information may be included here.

Author Contributions

All authors have given approval to the final version of the manuscript.

Funding Sources

This research was supported by the Sokol Institute for Pharmaceutical Life Sciences (JJS and DPR), by NIH RO1-AI-133633-01 (JJS, PB and DPR) and by the Military Infectious Disease Research Program Q0480_19_WR_CS_OC for BSP and PJL

ACKNOWLEDGMENT

We acknowledge the Entomology Branch and Veterinary Medicine Branch AFRIMS, with special thanks to Ratawan Ubalee and team for the production of *P. cynomolgi*-infected mosquitoes.

ABBREVIATIONS

PfPKG-*Plasmodium falciparum* cGMP-dependent protein kinase; PbLuc-*Plasmodium berghei* luciferase; SAR-structure-activity relationship; ATP-adenosine triphosphate; ADME-absorption, distribution, metabolism, elimination; CYP3A4-cytochrome P450 3A4; hERG-human ether-a-go-go related gene.

REFERENCES

- Uwimana, A.; Legrand, E.; Stokes, B. H.; Ndikumana, J.M.; Warsame, M.; Umulisa, N.; Ngamije, D.; Munyaneza, T.; Mazarati, J. B.; Munguti, K.; Campagne, P.; Criscuolo, A.; Ariey, F.; Murindahabi, M.; Ringwald, P.; Fidock, D.A.; Mbituyumuremyi, A.; Menard, D. Emergence and clonal expansion of in vitro artemisininresistant Plasmodium falciparum kelch13 R561H mutant parasites in Rwanda. *Nat Med.* 2020, *26*, 1602-8.
- Uwimana, A.: Umulisa, N.; Venkatesan, M.; Svigel, S. S.; Zhou, Z.; Munyaneza, T.; Habimana, R.M.; Rucogoza, A.; Moriarty, L.F.; Sandford, R.; Piercefield, E.; Goldman, I.; Ezema, B.; Talundzic, E.; Pacheco, M.A.; Escalante, A.A.; Ngamije, D.; Mangala, J.N.; Kabera, M.; Munguti, K.; Murindahabi, M.; Brieger, W.; Musanabaganwa, C.; Mutesa, L.; Udhayakuma, V.; Mbituyumuremyi, A.; Halsey, E.S.; Lucchi N.W. Association of Plasmodium falciparum kelch13 R561H genotypes with delayed parasite clearance in Rwanda: an open-label, single-arm, multicentre, therapeutic efficacy study. *Lancet Infect. Dis.* 2021, 21, 1120-1128.
- 3. World Health Organization World Malaria Report 2018.
- Ashley, E.A.; Dhorda, M.; Fairhurst, R.M.; Amaratunga, C.; Lim, P.; Suon, S.; Sreng, S.; Anderson, J.M.; Mao, S.; Sam, B.; Sopha, C.; Chuor, C.M.; Nguon, C.; Sovannaroth, S.; Pukrittayakamee, S.; Jittamala, P.; Chotivanich, K.; Chutasmit, K.; Suchatsoonthorn, C.; Runcharoen, R.; Hien, T.T.; Thuy-Nhien, N.T.; Thanh, N.V.; Phu, N.H.; Htut, Y.; Han, K.T.; Aye, K.H.; Mokuolu, O.A.; Olaosebikan, R.R.; Folaranmi, O.O.; Mayxay, M.; Khanthavong, M., Hongvanthong, B., Newton, P.N., Onyamboko, M.A.; Fanello, C.I.; Tshefu, A.K.; Mishra,

N.; Valecha, N.; Phyo, A.P.; Nosten, F.; Yi, P.; Tripura, R.; Borrmann, S.; Bashraheil, M.; Peshu, J.; Faiz, M.A.; Ghose, A.; Hossain, M.A.; Samad, R.; Rahman, M.R.; Hasan, M.M.; Islam, A.; Miotto, O.; Amato, R.; MacInnis, B.; Stalker, J.; Kwiatkowski, D.P.; Bozdech, Z.; Jeeyapant, A.; Cheah, P.Y.; Sakulthaew, T.; Chalk, J.; Intharabut, B.; Silamut, K.; Lee, S.J.; Vihokhern, B.; Kunasol, C.; Imwong, M.; Tarning, J.; Taylor, W.J.; Yeung, S.; Woodrow, C.J.; Flegg, J.A.; Das, D.; Smith, J.; Venkatesan, M.; Plowe, C.V.; Stepniewska, K.; Guerin, P.J.; Dondorp, A.M.; Day, N.P.; White, N.J.; Tracking Resistance to Artemisinin C. Spread of artemisinin resistance in Plasmodium falciparum malaria. *New Engl. J. Med.* **2014**, *371*, 411-23.

- Burrows, J.N.; Duparc, S.; Gutteridge, W.E.; Hooft van Huijsduijnen, R.; Kaszubska, W.; Macintyre, F.; Mazzuri, S.; Mohrle, J.J.; Wells, T.N.C. New developments in antimalarial target candidate and product profiles. *Malaria J.* 2017, 16, 26.
- Alonso, P.L.; Sacarlal, J.; Aponte, J.J.; Leach, A.; Macete, E.; Aide, P.; Sigauque, B.; Milman, J.; Mandomando, I.; Bassat, Q.; Guinovart, C.; Espasa, M.; Corachan, S.; Lievens, M.; Navia, M.M.; Dubois, M.C.; Menendez, C.; Dubovsky, F.; Cohen, J.; Thompson, R.; Ballou, W.R.; Duration of protection with RTS,S/AS02A malaria vaccine in prevention of Plasmodium falciparum disease in Mozambican children: single-blind extended follow-up of a randomised controlled trial. *Lancet* 2005, *366*, 2012-8.
- Wells, T.N.; Hooft van Huijsduijnen, R.; Van Voorhis, W.C.; Malaria medicines: a glass half full? *Nat. Rev. Drug Discov.* 2015, 14, 424-42.
- Hopp CS, Bowyer PW, Baker DA. The role of cGMP signalling in regulating life cycle progression of Plasmodium. Microbes Infect. 2012;14(10):831-7
- Alam, M.M.; Solyakov, L.; Bottrill A.R.; Flueck, C.; Siddiqui, F.A.; Singh, S.; Mistry, S.; Viskaduraki, M.; Lee, K.; Hopp, C.S.; Chitnis, C.E.; Doerig, C.; Moon, R.W.; Green, J.L.; Holder, A.A.; Baker, D.A.; Tobin, A.B.; Phosphoproteomics reveals malaria parasite Protein Kinase G as a signalling hub regulating egress and invasion. *Nat. Commun.* 2015, *6*, 7285.
- Brochet, M.; Collins, M.O.; Smith, T.K.; Thompson, E.; Sebastian, S.; Volkmann, K.; Schwach, F.; Chappell, L.; Gomes, A.R.; Berriman, M.; Rayner, J.C.; Baker, D.A.; Choudhary, J.; Billker, O.; Phosphoinositide metabolism links cGMP-dependent protein kinase G to essential Ca²⁺ signals at key decision points in the life cycle of malaria parasites. *PLoS Biol.* **2014**, *12*, e1001806.
- 11. Arendse, L. B.; Wyllie, S.; Chibale, K.; Gilbert, I. H. Plasmodium kinases as potential drug targets for malaria: challenges and opportunities. *ACS Infect. Dis.* **2021**, *7*, 518-534.

12. Vanaerschot, M.; Murithi, J. M.; Pasaje, C.; Ghidelli-Disse, S.; Dwomoh, L.; Bird, M.; Spottiswoode, N.; Mittal, N.; Arendse, L. B.; Owen, E. S.; Wicht, K. J.; Siciliano, G.; Bösche, M.; Yeo, T.; Kumar, S. T. R.; Mok, S.; Carpenter, E.; Giddins, M. J.; Sanz, O.; Ottilie, S.; Alano, P.; Chibale, K.; Llinás, M.; Uhlemann, A. C.; Delves, M.; Tobin, A.; Doerig, C.; Winzeler, E.; Lee, M. C. S.; Niles, J.; Fidock, D. A. Inhibition of the resistance-refractory *P. falciparum* kinase PKG delivers prophylactic, blood stage and transmission-blocking antiplasmodial activity. *Cell Chem. Biol.* **2020**, *27*, 806-816.

13. Baker, D.A.; Stewart, L.B.; Large, J.M.; Boyer, P.W.; Ansell, K.H.; Jiménez, M.B.; El Bakkouri, M.; Birchall, K.; Dechering, K.J.; Bouloc, N.S.; Coombs, P.J.; Whalley, D.; Harding, D.J.; Smiljanic-Hurley, E.; Weldon, M.C.; Walker, E.M.; Dessens, J.T.; Lafuente, M.J.; Sanz, L.M; Gamo, F.-J.; Ferrer, S.B.; Hui, R.; Bousema, T.; Angulo-Barturén, I.; Merritt, A.T.; Croft, S.L.; Gutteridge, W.E.; Kettleborough, C.A.; Osborne, S.A.; A potent series targeting the malarial cGMP-dependent protein kinase clears infection and blocks transmission, *Nat. Commun.* **2017**, *8*, 430-439.

14. Penzo, M.; de las Heras-Dueña, L.; Mata-Cantero, L.; Diaz-Hernandez, B.; Vazquez-Muñiz, M.-J.;Ghidelli-Disse, S.; Drewes, G.; Fernández-Álvaro, E.; Baker, D.A., High-throughput screening of the *Plasmodium falciparum* cGMP-dependent protein kinase identified a thiazole scaffold which kills erythrocytic and sexual stage parasites, *Sci. Reports*, **2019**, *9*, 7005-7018.

15. Matralis, A.N.; Malik, A.; Penzo, M.; Moreo, I.; Almela, M.J.; Camino, I.; Crespo, B.; Saadeddin, A.; Ghidelli-Disse, S.; Rueda, L.; Calderon, F.; Osborne, S.A.; Drewes, G.; Böesche, M.; Fernández-Álvaro, E; Hernando, J.I.M.; Baker, D.A. Development of chemical entities endowed with potent, fast-killing properties against *Plasmodium falciparum* malaria parasites, *J. Med. Chem.* **2019**, *62*, 9217-9235.

16. Mahmood, S.U.; Chang, H.; Tummalapalli, S.R.; Chakrasali, R.; Bheemanaboina, R.R.Y.; Kreiss, T.; Chojnowski, A.; Eck. T.; Siekierka, J.J.; Rotella, D.P. Discovery of isoxazolylbased inhibitors of *Plasmodium falciparum* cGMP-dependent protein kinase, *RSC Med. Chem.* **2020**, *11*, 98-101.

17. Biftu, T.; Feng, D.; Ponpipom, M.; Girotra, N.; Liang, G.-B.; Qian, X.; Bugianesi, R.; Simeone, J.; Chang, L.; Gurnett, A.; Liberator, P.; Dulski, P.; Leavitt, P.S.; Crumley, T.; Misura, A.; Murphy, T.; Rattray, S.; Samaras, S.; Tamas, T.; Mathew, J.; Brown, C.; Thompson, D.; Schmatz, D.; Fisher, M.; Wyvratt,M.; Synthesis and SAR of 2,3-diarylpyrrole inhibitors of parasite cGMP-dependent protein kinase as novel anticoccidial agents, *Bioorg. Med. Chem. Lett.* **2005**, *15*, 3296-3301.

18. Govindasamy, K.; Jebiwott, S.; Jaiyan, D.K.; Davidow, A.; Ojo, K.K.; Van Voorhis, W.C.; Brochet, M.; Billker, O.; Bhanot, P., Invasion of hepatocytes by sporozoites requires cGMP-dependent protein kinase and calcium dependent protein kinase 4, *Mol. Microbiol.* **2016**, *102*, 349-363.

19. Manuscript under review-see supplementary information.

Authors are required to submit a graphic entry for the Table of Contents (TOC) that, in conjunction with the manuscript title, should give the reader a representative idea of one of the following: A key structure, reaction, equation, concept, or theorem, etc., that is discussed in the manuscript. Consult the journal's Instructions for Authors for TOC graphic specifications.

