Identification of a promiscuous conserved CTL epitope within the SARS-CoV-2 spike protein

Sheng Jiang[a,b], Shuting Wu[a], Gan Zhao[c], Yue He[c], Xinrong Guo[d], Zhiyu Zhang[c], Jiawang Hou[c], Yuan Ding[c], Alex Cheng[c] and Bin Wang[a,b,c,*]

[a]Key Laboratory of Medical Molecular Virology (MOE/NHC/CAMS), School of Basic Medical Sciences, Shanghai Medical College(SHMC), Fudan University & [b]National Clinical Research Center for Aging and Medicine, Huashan Hospital, Fudan University, Shanghai, China.

[c]Advaccine Biopharmaceutics (Suzhou) Co. LTD, Jiangsu Province, China.

[d]Colby College, Waterville, Maine, USA

*Corresponding Author: Bin Wang, School of Basic Medical Sciences, Fudan University, 131 Dong An Road, 409 Fuxing Building, Shanghai, China 200032. E-mail: bwang3@fudan.edu.cn

ABSTRACT

The COVID-19 disease caused by infection with SARS-CoV-2 and its variants is devastating to the global public health and economy. To date, over a hundred COVID-19 vaccines are known to be under development and the few that have been approved to fight the disease are using the spike protein as the primary target antigen. Although virus neutralizing epitopes are mainly located within the RBD of the spike protein, the presence of T cell epitopes, particularly the CTL epitopes that are likely to be needed for killing infected cells, has received comparatively little attention. In this study, we predicted several potential T cell epitopes with web-based analytic tools, and narrowed them down from several potential MHC-I and MHC-II epitopes by ELIspot and cytolytic assays to a conserved MHC-I epitope. The epitope is highly conserved in current viral variants and compatible with presentation by most HLA alleles worldwide. In conclusion, we identified a CTL epitope suitable for evaluating the CD8+ T cell-mediated cellular response and potentially for addition into future COVID-19 vaccine candidates to maximize CTL responses against SARS-CoV-2.

INTRODUCTION

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) was first identified in Wuhan in early 2020, spread at unprecedented speed, and became a disaster to human beings worldwide[1]. Effective vaccines, antiviral drugs, and treatments have high priorities to defend against such challenges. SARS-CoV-2 has four main structural proteins: the envelope, membrane, nucleocapsid, and spike protein that can be considered for inclusion in vaccines. The spike protein has a receptor-binding domain (RBD) that specifically binds to human angiotensin-converting enzyme 2 (hACE2) as a receptor and mediates virus entry into the host cell[2,3]. Neutralizing antibodies recognizing the RBD can block the spike protein from binding to the hACE2 and inhibit virus entry[4,5]. Therefore, spike protein has been the primary choice as the immunogen in candidate vaccines.

Although protection against disease via vaccine-induced neutralizing antibodies has been demonstrated, the elimination of SARS-CoV-2 infection within the host is also essential. The numbers of mild and asymptomatic cases are rising dramatically in recent years and such cases remain infective, prolonging viral dissemination. To eliminate the viral infection, induction of a potent antigen-specific CD8+ T cell response by vaccination is probably critical. To activate a viral-specific CD8+ T cell response, the vaccine must contain highly active major histocompatibility complex class I (MHC-I) epitopes that can be presented by MHC-I molecules to interact with CD8+ T cell receptors (TCR). The potentiation of viral-specific CD8+ T cell responses is dependent upon the high affinity and avidity of MHC-I and TCR binding. There is a lack of information on the CD8+ T cell-recognized epitopes within the spike antigen; consequently, only overlapping peptide pools that covered the whole region of spike antigen have been used routinely to evaluate cell-mediated immunity (CMI) of a vaccine candidate[6,7,8]. However, a few reports had suggested that CTL epitopes are present within the spike protein[9], but no detailed mapping information had been reported or potential sequences discovered. The identification of those CD8+ T cell epitopes would provide an important tool to evaluate the T cell immunity in vaccinated individuals or patients and was undertaken here.

This study utilized web-based tools to analyze the potentials for transportation associated with antigen processing (TAP) in the human MHC-I epitopes that were predicted by the Immune Epitope Database analysis (IEDB) resource[10] to be present in peptide pools covering the n-terminal domain (NTD) and receptor-binding domain (RBD) of the spike protein. We demonstrated that peptide 2 (YYVGYLQPRTFLLKY), although it did not give the highest score in the web-based analysis of immunogenicity, was the best epitope for inducing a robust antigen-specific IFN-γ producing CD8+ T response as defined by ELIspot assay. This epitope sequence is also highly conserved among currently discovered SARS-CoV-2 variants.

RESULTS

Strong CD8+ CTL epitope activity is embedded in an overlapping peptide pool 1 that covers the NTD and RBD region of the spike protein

When Balb/c mice were immunized twice with the pGX9501 DNA vaccine expressing the spike protein of SARS-CoV-2, a higher level of IFN-γ expression by splenocytes was often seen by the ELIspot assay when the cells were stimulated in vitro with spike peptide pool 1 (Table 1) compared with pool 2 (Figure 1A). In addition, when an in vivo CTL assay was done with identically immunized animals, the same peptide pool 1 gave a strong CTL response in vivo (Figure 1B), suggesting that MHC-I epitope(s) were present within pool 1.

Screening and identification of an MHC-I epitope in peptide pool 1.

We placed the entire 41 peptide sequences from peptide pool 1 into the Immune Epitope Database analysis (IEDB) (http://www.iedb.org/) to seek critical epitopes. An evaluation method was established by integrating MHC-I binding prediction, MHC-I immunogenicity, and MHC natural processing (MHC-NP) prediction from three H-2d MHC-I alleles to improve the accuracy of prediction results (Table 2). In the H-2D^{d} allele, peptide 2 showed good MHC-I binding ability, immunogenicity, and TAP ability. In the H-2K^{d} allele, peptide 12 showed the strongest immunogenicity, and peptide 2 presented the most potent MHC-I binding ability and TAP ability among all peptides. In the H-2L^{d} allele, both peptide 2 and peptide 11 showed the strongest immunogenicity, while peptide 12 emerged as having the most potent TAP ability (Figure 2A&2B). Consequently, we used peptide 2, peptide 11, and peptide 12 to conduct an ELIspot assay of T cell IFN-γ response.

As shown in Figure 3A, peptide 2 from pool 1 presented the best stimulation to induce the IFN-γ secretion compared to the other two selected peptides. Thus, peptide 2, consisting of 15 amino acids, stimulated CD8+ T cells via MHC-I or/and CD4+ T cells via MHC-II. To identify which T cell type was stimulated by peptide 2, purified CD4+ T cells or CD8+ T cells were used (sFigure 1). Peptide 2 stimulated CD8+ T cells but not CD4+ cells, indicating that it can only be presented by

MHC-I (Figure 3B). To further investigate its sequence specificity, we mutated several predicted anchor amino acids of peptide 2 according to the preferences of the H-2d MHC-I allele[11-14]. The mutated peptide 2 had a low MHC-I binding score in the IEDB prediction (Table 3) and showed a significantly reduced ability to stimulate IFN-γ secretion by CD8+ T cells (Figure 3C).

Analysis of peptide 2 epitope conservation and HLA distribution

We used the peptide 2 sequence, YYVGYLQPRTFLLKY (amino acid 264-278), to compare with current posted SARS-CoV-2 variants including variant-of-interest (VOI) and variant-of-concern (VOC) published by WHO and observed that this sequence is highly conserved (Figure 4A), and located at the end of the NTD of Spike protein and upstream of the RBD (Figure 4B). The highly conserved epitopic sequence provides a useful tool for evaluating the CD8+ T cell-mediated responses to vaccination in animals and humans.

Since MHC-I-biased expression patterns in different populations globally are diversified and variable, the peptide 2 sequence that can be recognized by one population may not be recognized by others. To investigate if peptide 2 sequence could be recognized by most populations worldwide, we performed an HLA allele analysis for different regions to assess binding to one or more of the 27 prevalent MHC-I molecules, including HLA-A (01:01, 02:01, 02:03, 02:06, 03:01, 11:01, 23:01, 24:02, 26:01, 30:01, 30:02, 31:01, 32:01, 33:01, 68:01, 68:02), and HLA-B (07:02, 08:01, 15:01, 35:01, 40:01, 44:02, 44:03, 51:01, 53:01, 57:01, 58:01), as shown in Figure 4C & Table 4. The frequency by HLAs was calculated by the online analysis tool at http://www.allelefrequencies.net/. We also evaluated the MHC-I binding ability, immunogenicity, and TAP potential of peptide 2 on different HLA alleles by IEDB (Figure 4D & Table 5). The results indicated that peptide 2 could be recognizable by the HLA-A*02:01 allele (most in Europe and America), HLA-B*08:01 allele (in Europe and Australia), HLA-A*23:01 allele (in North Africa and Sub-Saharan Africa), HLA-A*02:03 allele (in Southeast Asia), HLA-A*24:02 allele (in Oceania), HLA-A*02:06 allele (in North America, North-East Asia, and Oceania), HLA-A*33:01 allele (in China and Pakistan), HLA-B*35:01 allele (in Oceania), and HLA-A*03:01 allele (in Europe). These findings suggest that peptide 2 could be well recognized by the most frequent HLA alleles of the worldwide population

and can therefore be considered to be promiscuous.

DISCUSSION

In this study, we have defined and characterized a potential CTL epitope conserved and established a BALB/c mice model to evaluate CTL responses to the spike protein of SARS-CoV-2. Furthermore, we found the peptide 2 may be well recognized by HLA alleles in most populations worldwide.

In recent studies, CD8+ T cell immunity was found to make major contributions to the protective efficacy of SARS-CoV-2 vaccines[15-17]. Additionally, lymphopenia was more accentuated in symptomatic COVID-19 patients with pneumonia than in those without pneumonia, consistent with T cell immunity playing a protective role in pre-existing immunity against SARS-CoV-2[17-19]. However, the role of T cell immunity in the pathology of COVID-19 has not been clarified[20] and awaits further definition of T cell epitopes and their functions. Engineered improvements in the peptide epitopes recognized by CTL have been an essential feature of the advancements made in evaluating the cellular immunity induced by vaccines. Our work has provided a tool to monitor virus-specific CD8+ T cells and assess the contribution of CTLs to the control and the elimination of the virus.

The SARS-CoV-2 virus was found to mutate rapidly, the development of vaccines that could protect people from different variants was seen to be urgent. The neutralizing antibodies induced by vaccines were found to have variable efficacies against the different SARS-CoV-2 variants and efficacies declined over time, whereas the protection represented by $CD8^+$ T cell immunity remained unchanged[15]. We showed here that one particular peptide from the spike protein, peptide 2 is highly conserved among variants and could be well presented by MHC-1 of all of the HLA alleles that are most frequent worldwide. Thus, the conserved peptide 2 was found suitable for developing peptide vaccines to induce cellular immunity against different variants of SARS-CoV-2, including variant-of-interest (VOI) and variant-of-concern (VOC). A recent study confirmed that peptide 2 probably has a strong cell-mediated immunological function in man; a 9-mer (YLQPRTFLL) peptide that was overlapped by peptide 2 was able to induce a high level of IFN-γ

129 expression from PBMCs of patients who had recovered from COVID-19 and carried the HLA-
130 A*02:01 allele[21].

131 In conclusion, our study utilized web-based tools to predict human MHC-I epitopes although
132 peptide 2 (YYVGYLQPRTFLLKY) did not give the strongest total TAP score in the prediction, the
133 simulations indicated particularly robust antigen-specific IFN-γ-expressing CD8+ T responses
134 overall compared to the other predicted epitope sequences. This epitope sequence is highly
135 conservative among currently known SARS-CoV-2 variants and recognizable by most world
136 populations. This critical MHC-I epitope sequence is located at the end of NTD of the spike protein;
137 it can be used to assess CMI induced by a COVID-19 vaccination; it may be strategically
138 incorporated into vaccine designs to enhance the prospect of viral elimination by vaccination.

139 MATERIALS AND METHODS

140 Mice

141 Female BALB/c mice (6-8 weeks of age) were purchased from Beijing Vital Laboratory
142 Animal Technology Co., Ltd. (Beijing, China) and Shanghai Jiesjie Laboratory Animal Co., Ltd.
143 (Shanghai, China), and were kept in SPF conditions. All animal experiments were approved by the
144 Experimental Animals Committee of SHMC, and all methods were carried out in accordance with
145 relevant guidelines and regulations. This study was carried out in compliance with the ARRIVE
146 guidelines. All mice was sacrificed after experiment under euthanasia with Isoflurane overdose.

147 Peptide pool derived from SARS-CoV-2 Spike protein

148 The spike receptor-binding domain (RBD) peptide pool (SARS-CoV-2 spike protein 258-
149 518aa) published previously[7] was used for the study (Table 1).

150 Immunization

151 The mice were injected twice, with a two-week interval, via the intramuscular route (i.m.)
152 with 25 μg pGX9501 expressing a synthetic, optimized sequence of the SARS-CoV-2 full-length
153 spike glycoprotein[7] and then electroporation was applied with the Cellectro2000 device. Serum
154 samples and spleens were collected 14 days after the second immunization.

155 **Cytotoxic lymphocyte (CTL) killing ability**

156 A single-cell suspension of splenocytes from naïve syngeneic mice was diluted to
157 $1.5*10^8$/ml in RPMI1640 containing 10% FBS and 2% penicillin and streptomycin and pulsed
158 at 37°C with or without 5 ug/ml peptides as described previously[22]. After 4 h, eflour450
159 (eBioscience, 65-0842-85) at 5 mM (high concentration) was used to label peptide-pulsed cells
160 at room temperature in the dark. The non-peptide-pulsed cells were labeled with a low
161 concentration of eflour450 at 0.5 mM. After being rinsed three times with PBS, $4*10^6$ labeled
162 and peptide-pulsed cells and an equal number of labeled non-peptide-pulsed cells were
163 adoptively transferred by tail vein injections into mice that had previously been immunized.
164 Six hours later, the percentage of labeled cells in spleens was detected with LSRFortessa flow
165 cytometry (BD) and analyzed by FlowJo (TreeStar). The following formula calculated the
166 specific cell lysis: Specific cell lysis ability = (1-(percentage of cells incubated with
167 peptide/percentage of cells incubated without peptide)) *100%.

168 **IFN-γ ELIspot**

169 Spleens were collected from mice individually into RPMI1640 media supplemented with 10%
170 FBS (Gibco) and penicillin/streptomycin and processed into single-cell suspensions. ELIspot assays
171 were performed using Mouse IFN-γ ELIspot plates (Dakewei Biotech Co., Ltd, 2210006). The
172 ELIspot plates were repeat washed 5 times at RT with 100 μL of PBS per well and incubated with
173 200 μL of RPMI1640 10% FBS (R10) for 10 min before cell plating. Two hundred fifty thousand
174 mouse splenocytes, CD4+, or CD8+ T cells were plated into each well and stimulated for 16 h with
175 15-mer peptides from the SARS CoV-2 Spike peptide pools that overlapped by nine amino acids as
176 previously described[7]. Each peptide was present at a final concentration of 1 μg in 100 μl R10 per
177 well. The spots were developed based on the manufacturer's instructions. R10 and cell stimulation
178 cocktails (Invitrogen) were used for negative and positive controls, respectively. Spots were scanned
179 and quantified by AID ELIspot READER (AID, Germany). Spot-forming units (SFU) per million
180 cells were calculated after subtracting the negative control wells.

Statistical Analysis

The statistical analysis methods and sample sizes (n) are specified in the results section or figure legends for all quantitative data. All values are reported as means ± sem with the indicated sample size. No samples are excluded from the analysis. All relevant statistical tests are two-sided. P values less than 0.05 were considered statistically significant. All animal studies were performed with randomized animal selection. Statistics were performed using GraphPad Prism 7 software. In all data, *$p<0.05$, **$p<0.01$, ***$p<0.001$, and ****$p<0.0001$.

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AUTHOR CONTRIBUTIONS: BW and SJ conceived, designed the study and drafted manuscript. BW supervised the study. SJ, SW, XG, JH, YD and ZZ performed the experiments. SJ, YH, GZ and AC analyzed the data. All authors reviewed the manuscript.

Tables

Table 1. Overlapping Peptide Pool 1.

Peptide number	Sequence	Start	End	aa
1	TAGAAAYYVGYLQPR	258	272	15
2	YYVGYLQPRTFLLKY	264	278	15
3	QPRTFLLKYNENGTI	270	284	15
4	LKYNENGTITDAVDC	276	290	15
5	GTITDAVDCALDPLS	282	296	15
6	VDCALDPLSETKCTL	288	302	15
7	PLSETKCTLKSFTVE	294	308	15
8	CTLKSFTVEKGIYQT	300	314	15
9	TVEKGIYQTSNFRVQ	306	320	15
10	YQTSNFRVQPTESIV	312	326	15
11	RVQPTESIVRFPNIT	318	332	15
12	SIVRFPNITNLCPFG	324	338	15
13	NITNLCPFGEVFNAT	330	344	15
14	PFGEVFNATRFASVY	336	350	15
15	NATRFASVYAWNRKR	342	356	15
16	SVYAWNRKRISNCVA	348	362	15
17	RKRISNCVADYSVLY	354	368	15
18	CVADYSVLYNSASFS	360	374	15
19	VLYNSASFSTFKCYG	366	380	15
20	SFSTFKCYGVSPTKL	372	386	15
21	CYGVSPTKLNDLCFT	378	392	15
22	TKLNDLCFTNVYADS	384	398	15
23	CFTNVYADSFVIRGD	390	404	15
24	ADSFVIRGDEVRQIA	396	410	15
25	RGDEVRQIAPGQTGK	402	416	15
26	QIAPGQTGKIADYNY	408	422	15
27	TGKIADYNYKLPDDF	414	428	15
28	YNYKLPDDFTGCVIA	420	434	15
29	DDFTGCVIAWNSNNL	426	440	15
30	VIAWNSNNLDSKVGG	432	446	15
31	NNLDSKVGGNYNYLY	438	452	15
32	VGGNYNYLYRLFRKS	444	458	15
33	YLYRLFRKSNLKPFE	450	464	15
34	RKSNLKPFERDISTE	456	470	15
35	PFERDISTEIYQAGS	462	476	15

36	STEIYQAGSTPCNGV	468	482	15
37	AGSTPCNGVEGFNCY	474	488	15
38	NGVEGFNCYFPLQSY	480	494	15
39	NCYFPLQSYGFQPTN	486	500	15
40	QSYGFQPTNGVGYQP	492	506	15
41	PTNGVGYQPYRVVVL	498	512	15

Notes:

1. The peptides covered the entire sequence of 258-512 amino acids in the spike protein;
2. The peptides were synthesized with an average 15 amino acid length and with nine amino acids overlapping each other.

Table 2. MHC-I epitope analysis for Overlapping Peptide Pool 1.

allele	peptide number	MHC-I binding	immunogenicity	Proteasome Score	TAP Score	MHC Score	Processing Score	TAP Total Score
H-2-Dd	21	0.01	0.03612	1.44	1.12	-3.49	2.57	-0.92
H-2-Dd	41	0.04	0.13706	1.74	0.4	-2.56	2.14	-0.42
H-2-Dd	2	0.05	0.1573	1.36	1.07	-3.26	2.43	-0.83
H-2-Dd	26	0.13	-0.02676	1.38	1.27	-4.51	2.65	-1.86
H-2-Dd	9	0.19	-0.11058	1.27	0.99	-3.48	2.26	-1.22
H-2-Dd	19	0.25	0.03263	1.33	1.04	-3.45	2.37	-1.08
H-2-Dd	12	0.29	0.1431	1.05	1.29	-3.9	2.35	-1.55
H-2-Dd	11	0.49	0.1386	1.36	1.21	-4.02	2.57	-1.45
H-2-Dd	32	0.62	0.0966	1.19	1.17	-3.83	2.36	-1.46
H-2-Dd	37	0.65	0.12191	1.38	1.09	-3.76	2.47	-1.29
H-2-Dd	14	0.67	0.08562	1.39	0.82	-3.76	2.21	-1.55
H-2-Dd	31	0.7	0.023	1.31	1.15	-4.17	2.46	-1.71
H-2-Dd	20	0.7	-0.31841	1.47	1.3	-4.6	2.77	-1.83
H-2-Dd	27	0.89	-0.11289	1.4	1.21	-4.54	2.61	-1.92
H-2-Dd	28	1.1	-0.19576	0.98	1.12	-4.22	2.1	-2.12
H-2-Dd	24	1.2	0.12947	1.27	0.71	-4.63	1.98	-2.64
H-2-Dd	25	1.2	-0.11559	1.43	0.15	-4.39	1.58	-2.81
H-2-Dd	18	1.2	-0.22309	1.33	1.17	-4.05	2.5	-1.55
H-2-Dd	40	1.3	0.1256	1.38	1.34	-4.5	2.71	-1.79
H-2-Dd	1	1.3	0.06158	1.24	1.23	-4.32	2.47	-1.85
H-2-Dd	33	1.5	-0.21085	1.03	1.24	-4.23	2.27	-1.96
H-2-Dd	29	1.6	0.05792	1.52	0.5	-4.17	2.02	-2.15
H-2-Dd	30	1.6	0.05792	1.35	0.46	-4.17	1.8	-2.36
H-2-Dd	39	1.6	-0.15021	1.25	1.15	-3.84	2.41	-1.44
H-2-Dd	10	2	0.01977	1.1	0.24	-4.23	1.34	-2.89
H-2-Kd	2	0.01	0.06572	1.24	0.48	-1.78	1.72	-0.06
H-2-Kd	41	0.05	0.04196	1.74	0.46	-3.43	2.2	-1.22
H-2-Kd	9	0.06	0.12441	1	0.23	-2.55	1.23	-1.32
H-2-Kd	4	0.07	0.28634	1.34	0.37	-2.34	1.71	-0.63
H-2-Kd	3	0.07	0.05892	1.16	0.37	-2.34	1.54	-0.8
H-2-Kd	20	0.08	0.25644	1.45	0.49	-2.65	1.93	-0.71

H-2-Kd	21	0.08	0.05832	1.75	0.36	-2.65	2.12	-0.53
H-2-Kd	16	0.11	0.16858	1.36	0.4	-2.32	1.77	-0.55
H-2-Kd	40	0.19	0.0905	1.38	1.32	-3.9	2.69	-1.21
H-2-Kd	23	0.26	0.0573	1.31	0.44	-3.18	1.75	-1.43
H-2-Kd	24	0.61	-0.0378	1.07	0.2	-3.02	1.27	-1.75
H-2-Kd	19	0.68	0.0279	1.33	1.2	-3.27	2.52	-0.75
H-2-Kd	10	0.79	0.034	1.1	0.23	-3.35	1.33	-2.02
H-2-Kd	12	0.92	0.34063	1.45	0.59	-3.13	2.03	-1.1
H-2-Kd	1	0.96	-0.04018	1.24	1.29	-4.51	2.53	-1.98
H-2-Kd	32	0.99	0.13255	1.19	1.18	-3.56	2.37	-1.19
H-2-Kd	39	1.1	0.0801	1.25	1.31	-3.16	2.56	-0.6
H-2-Kd	31	1.2	0.1811	1.48	0.48	-3.32	1.96	-1.36
H-2-Kd	29	2	0.07062	1.52	0.5	-3.5	2.02	-1.48
H-2-Kd	30	2	0.07062	1.35	0.46	-3.5	1.8	-1.7
H-2-Kd	35	2	-0.15381	1.42	1.16	-4.52	2.57	-1.95
H-2-Ld	11	0.06	0.30371	1.36	0.98	-3.5	2.34	-1.16
H-2-Ld	41	0.12	-0.07228	1.74	0.35	-3.54	2.09	-1.45
H-2-Ld	21	0.2	0.05832	1.44	0.99	-3.65	2.44	-1.21
H-2-Ld	39	0.27	-0.19696	1.25	0.91	-3	2.16	-0.84
H-2-Ld	12	0.28	0.09851	1.05	0.94	-2.75	2	-0.75
H-2-Ld	37	0.44	0.0801	1.38	0.99	-4.01	2.37	-1.65
H-2-Ld	2	0.54	0.28634	1.41	1.19	-3.97	2.6	-1.38
H-2-Ld	3	0.57	-0.08994	1.52	1.13	-3.97	2.65	-1.32
H-2-Ld	27	0.77	0.0573	1.4	1.21	-4.39	2.61	-1.78
H-2-Ld	28	0.77	0.03448	0.98	1.12	-4.18	2.1	-2.07
H-2-Ld	19	0.95	0.11915	1.53	1.42	-4.32	2.95	-1.37
H-2-Ld	18	1.1	-0.22669	1.33	1.17	-3.82	2.5	-1.32
H-2-Ld	34	1.9	0.1811	0.91	1.16	-4.58	2.07	-2.5
H-2-Ld	9	1.9	-0.08994	1.27	0.99	-4.13	2.26	-1.87
H-2-Ld	38	2	-0.19696	0.99	0.28	-2.8	1.27	-1.53

Notes:

1. MHC-I binding score was between 0 and 2. <0.5 strong binder, 0.5-2 weak binder, >2 non-binder.
2. A high Immunogenicity score indicates that the degree of the peptide conformity to sequence preference was good.
3. The higher the TAP total score, the higher the likelihood that the peptide will be presented after being swallowed by DCs.

Table 3. Prediction scores of peptide 2 and mutated peptide 2

	Sequence	Score	MHC-I binding
Peptide 2	YYVGYLQPRTFLLKY	0.91863	0.01
Peptide 2 mut	YEVGELQDRTFELKY	0.025669	>2

Notes:

1. MHC-I binding score was between 0 and 2, where, the <0.5 represents a strong binder, 0.5-2 weak binder, >2 non-binder.
2. Underlined amino acids were the mutated residues in the Peptide 2 mut.

Table 4. Geographic Distribution of HLA allele

Continent	Allele	Frequency	Allele	Frequency
Australia	HLA-A*24:02	0.2	HLA-B*07:02	0.08
	HLA-A*02:01	0.11	HLA-B*40:01	0.07
	HLA-A*11:01	0.08		
Europe	HLA-A*02:01	0.26	HLA-B*07:02	0.08
	HLA-A*01:01	0.12	HLA-B*08:01	0.07
	HLA-A*03:01	0.12	HLA-B*51:01	0.07
	HLA-A*24:02	0.1		
North Africa	HLA-A*02:01	0.12	HLA-B*51:01	0.07
			HLA-B*08:01	0.05
			HLA-B*35:01	0.05
North America	HLA-A*02:01	0.2	HLA-B*35:01	0.08
	HLA-A*24:02	0.12	HLA-B*07:02	0.07
			HLA-B*08:01	0.05
			HLA-B*15:01	0.05
			HLA-B*44:03	0.05
			HLA-B*51:01	0.05
North-East Asia	HLA-A*24:02	0.22	HLA-B*51:01	0.08
	HLA-A*02:01	0.14	HLA-B*35:01	0.07
			HLA-B*15:01	0.07
			HLA-B*44:03	0.06
			HLA-B*07:02	0.05
Oceania	HLA-A*24:02	0.3	HLA-B*35:01	0.15
	HLA-A*11:01	0.15		
South and Central America	HLA-A*02:01	0.2	HLA-B*40:01	0.25
	HLA-A*24:02	0.14		
South Asia	HLA-A*11:01	0.13		
	HLA-A*01:01	0.1		
	HLA-A*24:02	0.1		
South-East Asia	HLA-A*11:01	0.2	HLA-B*40:01	0.1
	HLA-A*24:02	0.15	HLA-B*58:01	0.06
Sub-Saharan Africa	HLA-A*23:01	0.11	HLA-B*07:02	0.06
	HLA-A*02:01	0.1	HLA-B*08:01	0.05
Western Asia	HLA-A*02:01	0.15		
	HLA-A*01:01	0.1		

Notes

1. Frequency means the proportion of HLA allele present in the survey population. This table only shows the alleles for which frequency was higher than 0.05 in the continent.

Table 5. Human MHC-I epitope analysis for peptide 2

Allele	Peptide number	MHC-I binding	Immunogenicity	Proteasome Score	TAP Score	MHC Score	Processing Score	TAP Total Score
HLA-A*02:01	2	0.02	0.18048	1.39	0.39	-0.66	1.78	1.12
HLA-B*08:01	2	0.02	0.14268	1.39	0.39	-1.31	1.78	0.47
HLA-A*23:01	2	0.03	0.19092	1.36	1.13	-1.38	2.5	1.12
HLA-A*02:03	2	0.03	0.18064	1.39	0.39	-0.91	1.78	0.87
HLA-A*24:02	2	0.04	0.19092	1.36	1.13	-1.67	2.5	0.83
HLA-A*02:06	2	0.04	0.18064	1.39	0.39	-0.96	1.78	0.82
HLA-A*33:01	2	0.09	0.18048	0.76	0.68	-1.13	1.43	0.3
HLA-B*35:01	2	0.1	0.18064	1.41	1.19	-2.35	2.6	0.25
HLA-A*03:01	2	0.12	0.18064	0.89	0.18	-1.35	1.06	-0.29
HLA-A*32:01	2	0.21	0.18064	1.39	0.39	-2.21	1.78	-0.43
HLA-B*53:01	2	0.21	0.18064	1.41	1.19	-3.4	2.6	-0.8
HLA-A*01:01	2	0.23	0.18424	1.41	1.24	-2.91	2.65	-0.26
HLA-B*15:01	2	0.29	0.18064	1.41	1.28	-1.79	2.69	0.89
HLA-A*30:02	2	0.35	0.19092	1.41	1.28	-2.29	2.69	0.4
HLA-A*31:01	2	0.45	0.18064	0.76	0.68	-1.9	1.43	-0.47
HLA-A*26:01	2	0.52	0.18048	1.41	1.28	-4.14	2.69	-1.45
HLA-A*11:01	2	0.53	0.18064	0.89	0.18	-1.63	1.06	-0.57
HLA-B*58:01	2	0.55	0.18064	1.36	1.06	-3.33	2.42	-0.9
HLA-B*57:01	2	0.56	0.18064	1.36	1.07	-3.39	2.43	-0.95
HLA-B*51:01	2	0.65	0.18064	1.41	1.28	-4.36	2.69	-1.67
HLA-B*07:02	2	0.72	0.18064	1.41	1.19	-3.75	2.6	-1.15
HLA-A*30:01	2	0.73	0.18424	1.41	1.28	-3.73	2.69	-1.05
HLA-A*68:02	2	1.4	0.18064	1.39	0.39	-3.11	1.78	-1.33
HLA-B*44:03	2	1.6	0.18424	1.41	1.28	-4.01	2.69	-1.33
HLA-B*44:02	2	1.9	0.18424	1.41	1.28	-4.16	2.69	-1.47
HLA-A*68:01	2	2.4	0.18064	0.76	0.68	-2.33	1.43	-0.9
HLA-B*40:01	2	3.1	0.18048	1.41	1.28	-4.44	2.69	-1.75

Notes

4. MHC-I binding score was between 0 and 2. <0.5 strong binder, 0.5-2 weak binder, >2 non-binder.
5. The high Immunogenicity score means the degree of the peptide conformity to sequence preference was good.
6. The higher the TAP total score, the higher the likelihood that the peptide will be presented after being swallowed by DCs.

Figure Legends

Figure 1. Peptide pool 1 Induced strong T cell responses in BALB/c mice post-administration of pGX9501

Balb/c mice (n = 5/group) were immunized twice two weeks apart with 25 μg pGX9501or pVAX1 (empty vector). T cell responses were analyzed on day 14 after the second injection. (A) Splenocytes were harvested and T cell responses were measured by IFN-γ ELIspot after stimulation for 20 h with overlapping peptide pools 1 or 2. (B) Antigen-specific cytotoxic lymphocyte (CTL) killing activity was evaluated by an in vivo CTL assay. Target cells at $4*10^6$/ml from naïve mice were peptide-pulsed with pool 1 then labeled with a high concentration of eFlour450 in vitro Control cells were non-peptide-pulsed cells and labeled with a low concentration of eFluor450.The cells were mixed and transferred i.v. into immunized mice. After 5 h, splenocytes were harvested and the intensity of eFlour450 peptide labeled target cells was compared with the non-peptide-labeled negative control cells by flow cytometry.

Figure 2. IEDB prediction scores of peptides in pool 1

CTL epitope peptides were screened by integrating MHC-I binding prediction, MHC-I immunogenicity (A), and MHC-NP (B) prediction from three H-2d MHC-I alleles.

Figure 3. Peptide 2 is identified as the CD8+ CTL epitope

BALB/c mice were immunized with the pGX9501. Splenocytes were obtained and specific T-cell induction was analyzed with IFN-γ ELIspot, using stimulation with the indicated peptides. (A) Splenocytes, (B) CD4+, and (C) CD8+ T cells. .

Figure 4. Conserved Sequence & MHC-I HLA Analysis of peptide 2

(A) The sequence of peptide 2 was highly conserved in the 11 virus variants that have been identified as the variants of interest and the variants of concern, as published by WHO. (B) The position of the peptide in the stereoscopic structure of the spike protein. (C) The global distribution of HLA alleles. (D) Analysis of peptide 2 by integration of MHC-I binding prediction, MHC-I immunogenicity, and MHC-NP prediction from the HLA alleles.

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Figure 1

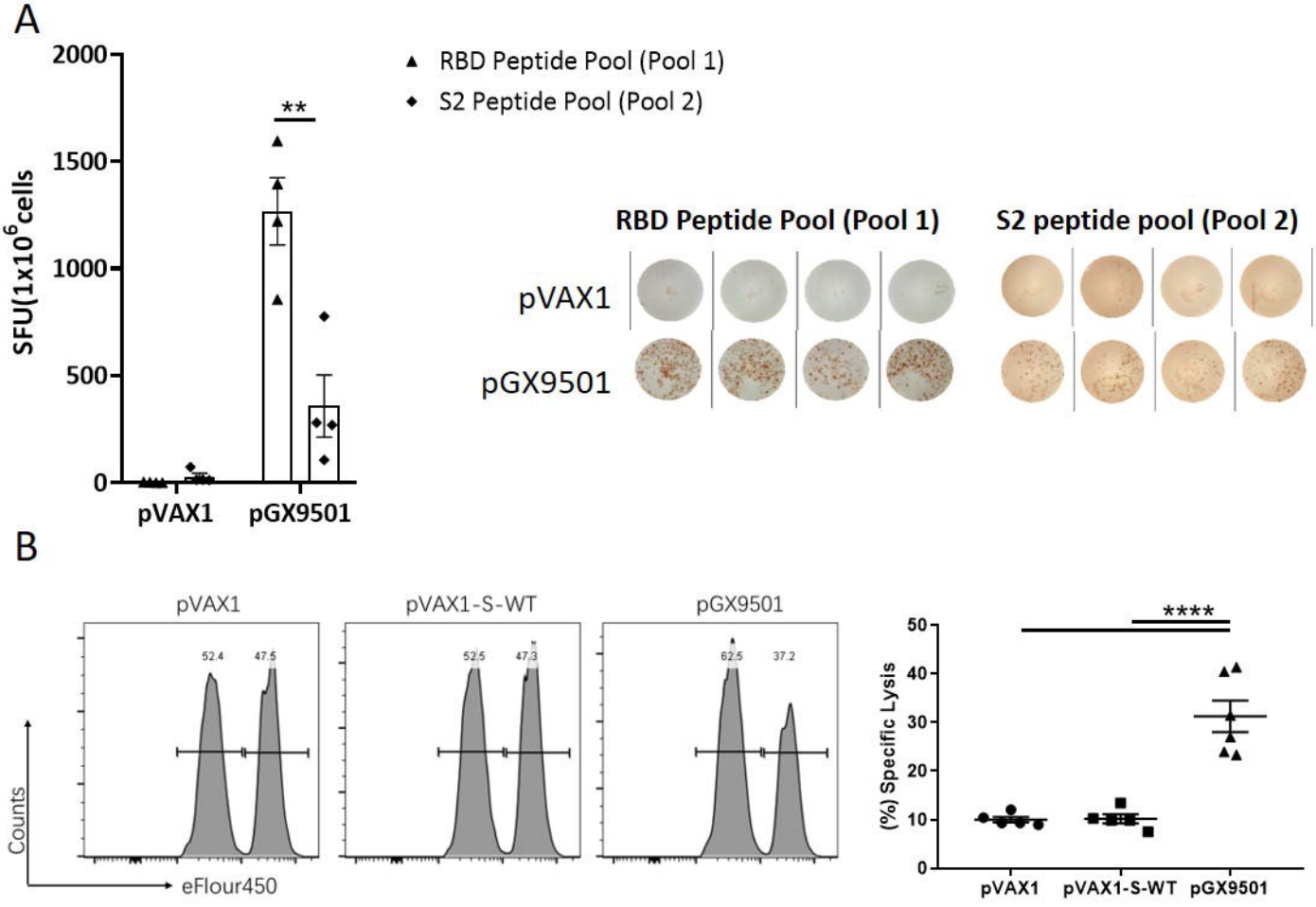
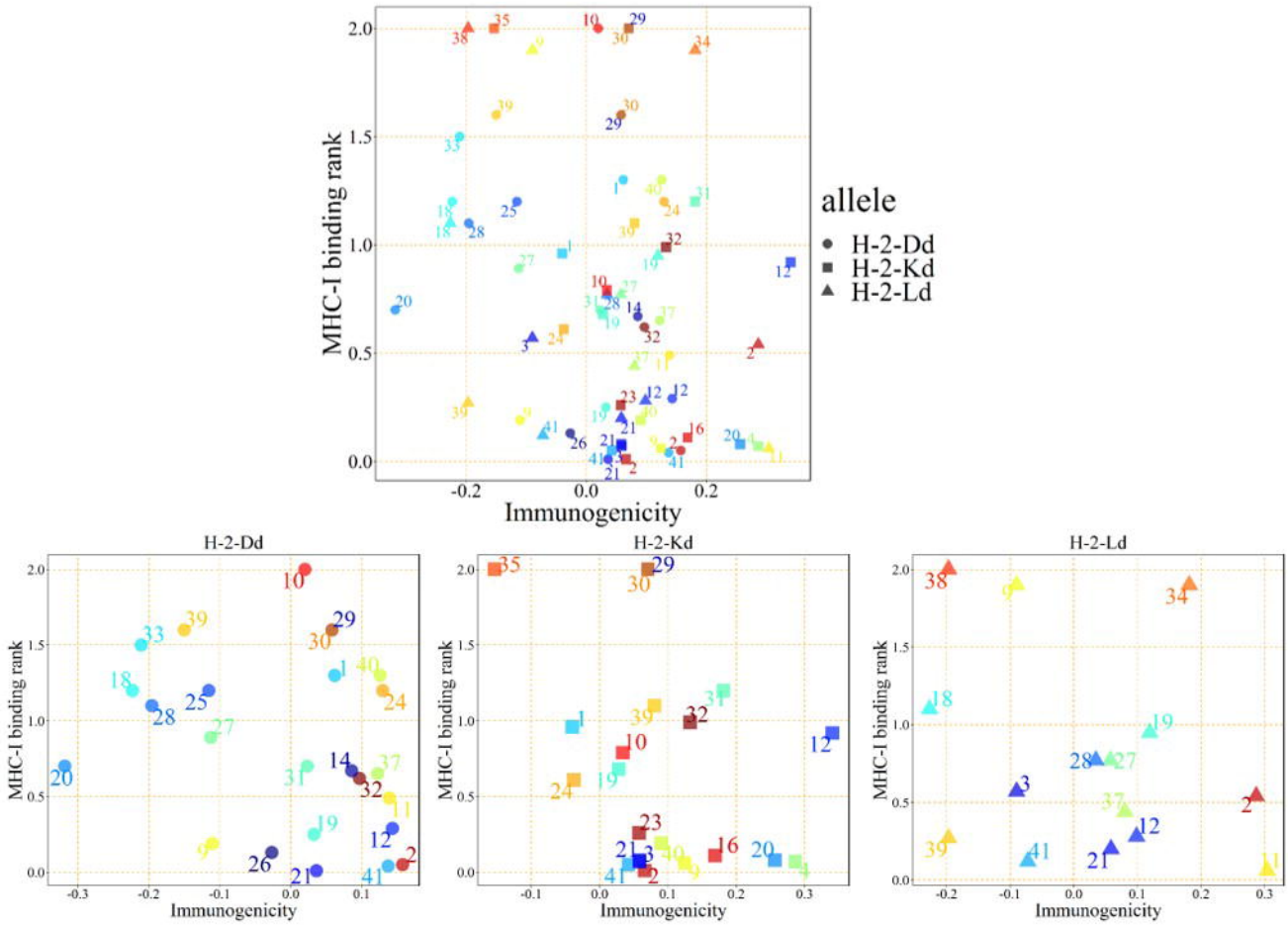

A
2000
1500
1000
500
0
SFU(1x10^6cells
**
pVAX1
pGX9501
RBD Peptide Pool (Pool 1)
S2 Peptide Pool (Pool 2)
RBD Peptide Pool (Pool 1)
S2 peptide pool (Pool 2)
pVAX1
pGX9501
B
pVAX1
pVAX1-S-WT
pGX9501
52.4
47.5
52.5
47.3
62.5
37.2
Counts
eFlour450
****
50
40
30
20
10
0
(%) Specific Lysis
pVAX1
pVAX1-S-WT
pGX9501

Figure 2

A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Immunogenicity
allele
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H-2-Kd
H-2-Ld
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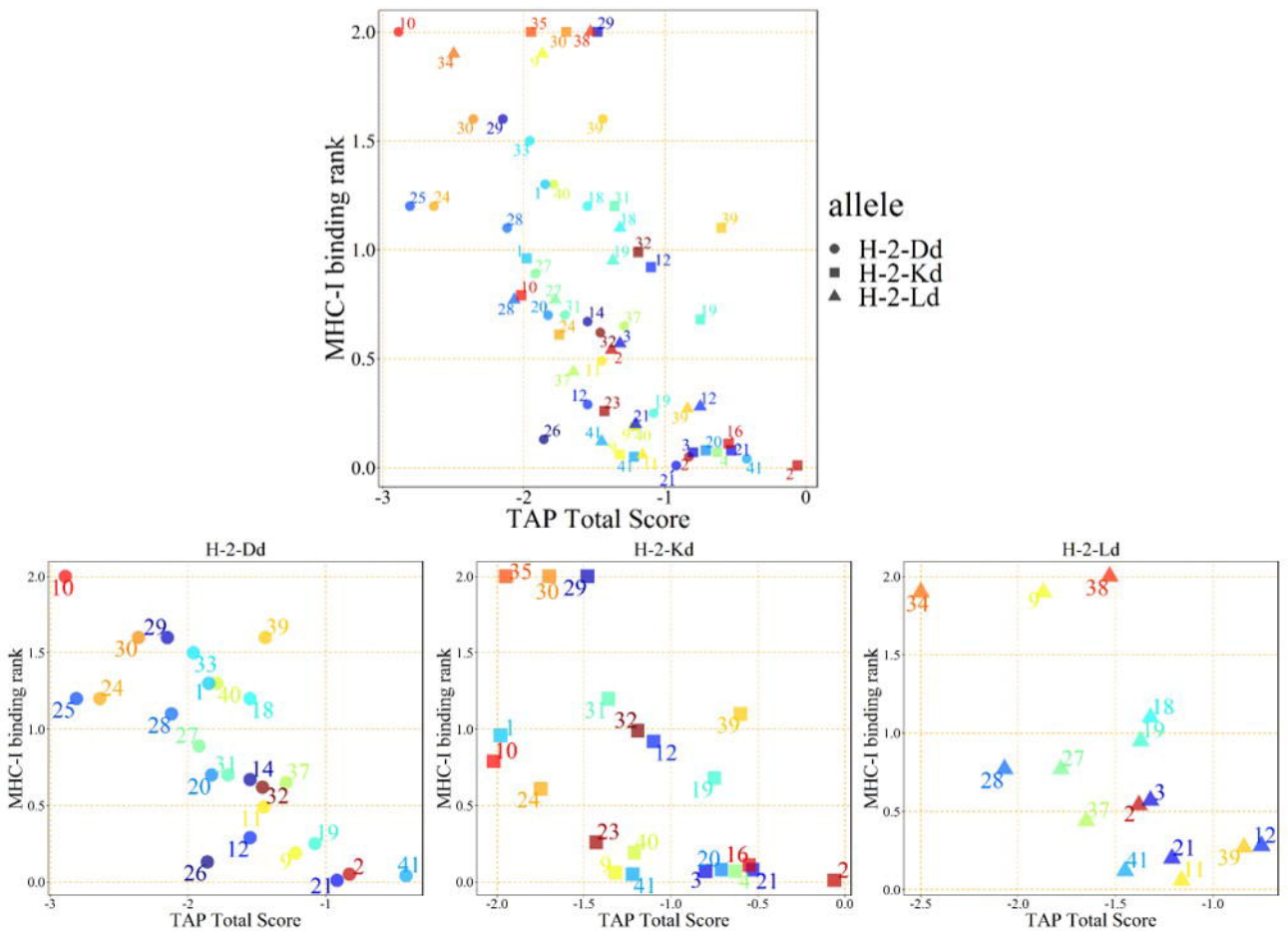
H-2-Kd
H-2-Ld

B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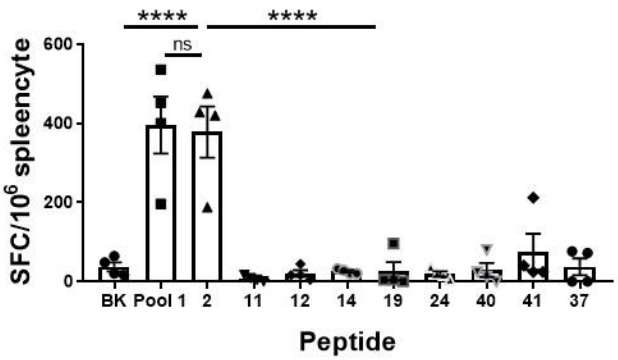
TAP Total Score
allele
H-2-Dd
H-2-Kd
H-2-Ld
H-2-Dd
H-2-Kd
H-2-Ld

Figure 3

A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ns
****
SFC/10^6 spleencyte
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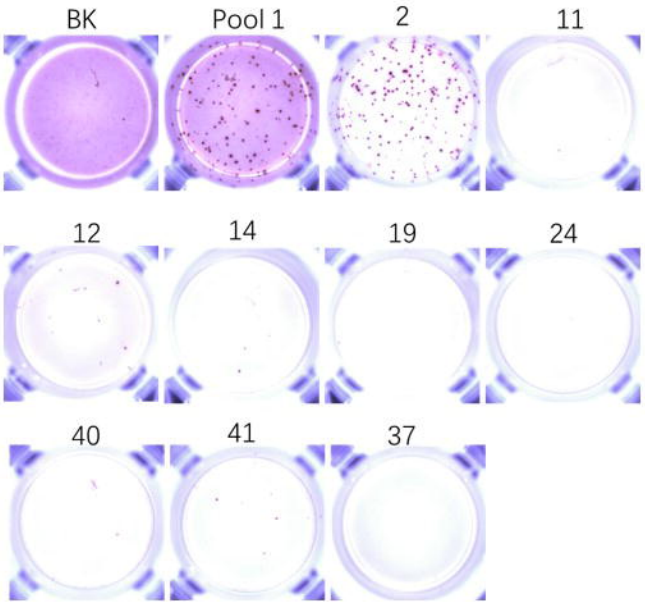
400
200
0
BK
Pool 1
2
11
12
14
19
24
40
41
37
Peptide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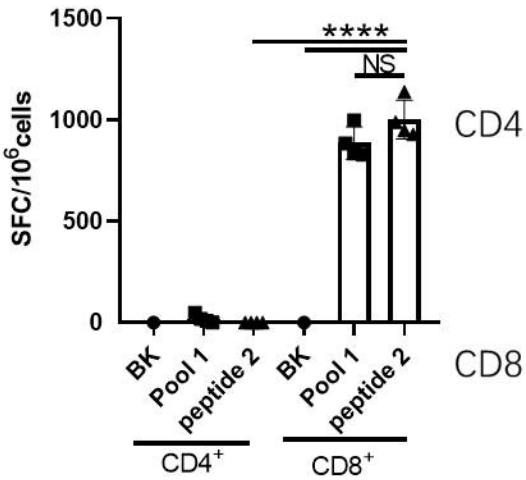
2
11
12
14
19
24
40
41
37

B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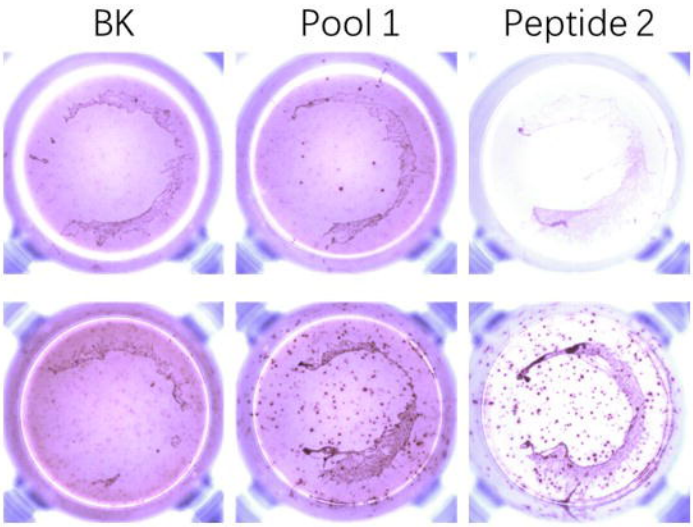
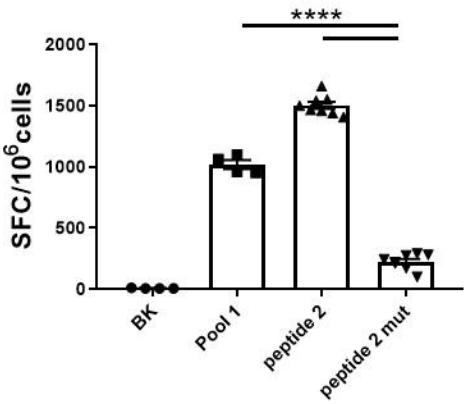
1000
500
0
SFC/10^6cells
****
NS
BK
Pool 1
peptide 2
BK
Pool 1
peptide 2
CD4+
CD8+
CD4
CD8


BK
Pool 1
Peptide 2

C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1500
1000
500
0
SFC/10^6cells
BK
Pool 1
peptide 2
peptide 2 mut

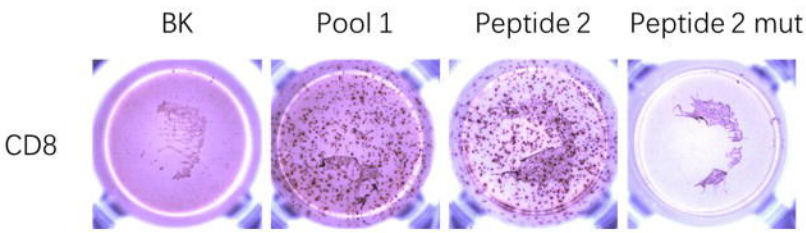
BK
Pool 1
Peptide 2
Peptide 2 mut
CD8

Figure 4

A

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	ε	B.1.427	QWT66198
	ζ	P.2	QWT11210
	η	B.1.525	QVU61512
	ι	B.1.526	QWT57870
	κ	B.1.617.1	QWT51828
	λ	C.37	QUR41526
	μ	B1.621	UAC19622.1
variant of concern,VOC	α	B1.1.7	QWT51566
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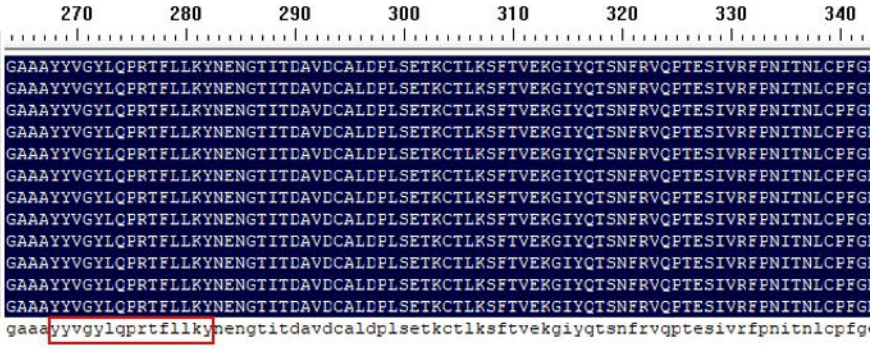

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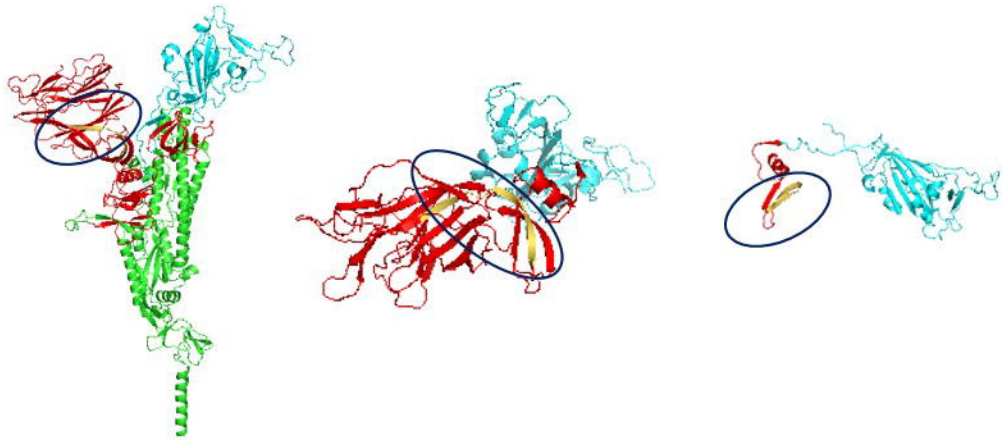

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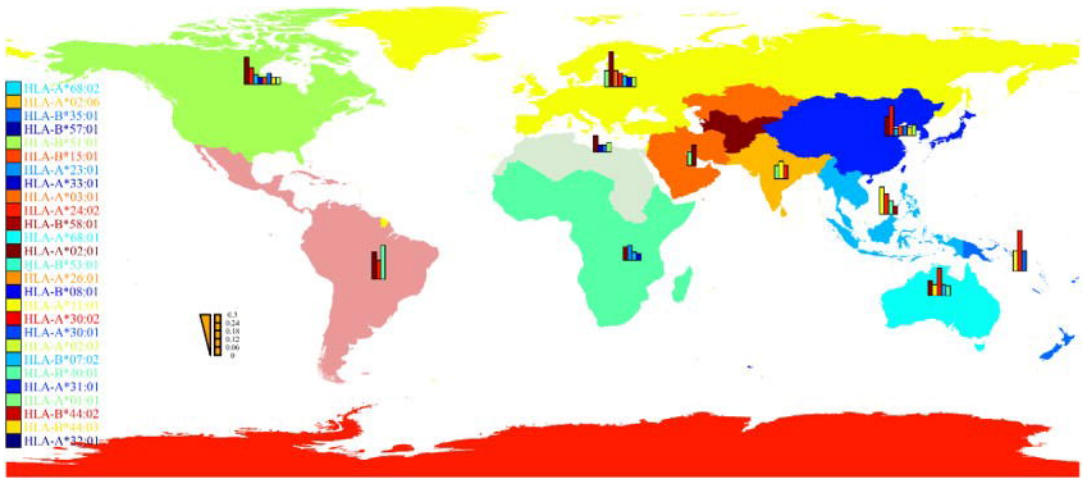
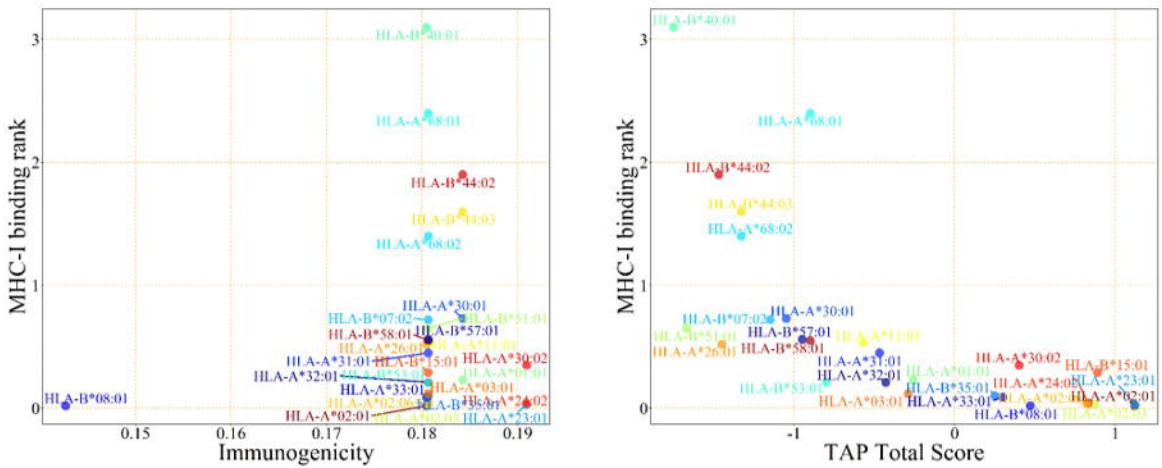
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