Pre- and post-oviposition behavioural strategies to protect eggs against

extreme winter cold in an insect with maternal care

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ABSTRACT

Depositing eggs in an area with adequate temperature is often crucial for mothers and their

offspring, as the eggs are immobile and therefore cannot avoid exposure to sub-optimal

temperatures. However, the importance of temperature on oviposition site selection is less

clear when mothers can avoid these potential adverse effects by both moving their eggs after

oviposition and providing other forms of egg care. In this study, we addressed this question in

the European earwig, an insect in which mothers care for the eggs during several months in

winter, frequently moving them during this period. We set up 60 females from two random

natural populations (as this species often exhibits population-specific life-history traits and

behaviours) under controlled thermal gradients, and recorded the temperature at which they

built their nests, tested whether they moved their eggs after an experimental temperature

change, and measured the effects on egg development and hatching rate. Our results

demonstrate that females indeed select oviposition sites according to temperature, and can

move their eggs to reach warmer temperatures. We also show that these warmer

temperatures are necessary to ensure egg hatching. Although this set of behavioural

thermoregulations is present in the two tested populations, we found a population-specific

modality of expression. These included the range of temperatures explored before

oviposition, temperature selected at oviposition and dynamics of egg transport following a

temperature change. Overall, our study sheds light on a new post-oviposition strategy in

female insects that overwinter with their eggs for coping with temperature changes. More

generally, it also reveals that egg care and/or egg transport do not prevent behavioural

thermoregulation via oviposition site selection and highlights the diversity of behaviours that

insects can adopt to enhance their tolerance to global climate change.

Keywords: Dermaptera, Egg brooding, Overwinter, Parental care, Thermal preferences

INTRODUCTION

Oviposition site selection shapes the fitness of most oviparous species (Thompson 1988;

Refsnider and Janzen 2010; Meunier et al. 2022). This is because choosing the right place to

deposit eggs typically provides direct and indirect benefits to egg-laying females, their current

eggs, and their future juveniles. This behaviour may first limit the high risk of predation on

adult females that is inherent to their lack of mobility during oviposition and favour their direct

access to specific food sources necessary for oviposition. For instance, females of the water

strider *Aquarius paludum insularis* (Motoschulsky, 1866) avoid ovipositing in sites containing

a predator attacking adults only (Hirayama and Kasuya 2013), while females of the orange tip

butterfly *Anthocharis cardamines* (Linnaeus, 1758) deposit eggs on plants that have high

nutritional value for adults but poor nutritional value for their larvae (Courtney 1981).

Oviposition site selection can also provide direct benefits to eggs by limiting the risks of

predators finding the eggs or eggs drying out. In the aquatic beetles *Hydroporus incognitus*

Sharp, 1869 and *H. nigrita* (Fabricius, 1792), for example, females select waters where no fish

can predate on their eggs (Brodin et al. 2006), while in the damselfly *Lestes macrostigmata*

(Eversmann, 1836) females prefer to lay their eggs on plants growing in the deeper parts of

temporary ponds to minimize the risk of future egg desiccation (Lambret et al. 2018). Finally,

oviposition site selection can help future offspring by favouring proximity to suitable habitats

and providing direct access to resources necessary for juveniles. This is the case in the

sandpaper frog *Lechriodus fletcher* (Boulenger, 1890) and several lady beetle species, where

females choose oviposition sites that contain the largest quantities of nutritive resources for

their future larvae (Sicsú et al. 2020; Gould et al. 2021).

The effect of temperature on egg development and survival is another potential driver

of oviposition site selection by females. Exposure to extreme temperatures can indeed

damage living organisms of all ages through alterations in their physiology, immunity and

behaviour, which may overall reduce their fitness and/or lead to premature death (Hance et

al. 2007; Dillon et al. 2009; Fey et al. 2015; Filazzola et al. 2021). These effects can be

particularly strong in eggs because they are immobile and thus unable to escape from

environmental temperatures, their shell often provides limited thermal protection, and the

development and survival of embryos (contained in eggs) are generally sensitive to subtle

changes in surrounding temperatures (Wang et al. 2010; Nicolai et al. 2013; Mortola and

Gaonac'h-Lovejoy 2016; Cordero et al. 2018; Yang et al. 2018). As a result, females of many

species select oviposition sites according to optimal temperatures for their eggs, such as in

the toad-headed agama lizard *Phrynocephalus przewalskii* Strauch, 1876 (Li et al. 2018), the

solitary red mason bee *Osmia bicornis* (Linnaeus, 1758) (Ostap-Chec et al. 2021), or the flat-

rock spiders *Hemicloea major* (Koch, 1875) (Pike et al. 2012).

By contrast, the importance of temperature for oviposition site selection becomes less

clear when mothers can transport their eggs from one location to another, as it may allow

them to secondarily adjust the temperature of their eggs throughout development. This

transport, included in a broader phenomenon called egg brooding in insects (Machado and

Trumbo 2018), is known to allow parents to limit the risk of egg predations or promote egg

oxygenation in aquatic species. For instance in the golden egg bug *Phyllomorpha laciniata*

(Villers, 1789), females lay their eggs on conspecifics whose mobility improves the avoidance

of egg parasitoids (Carrasco and Kaitala 2009). Similarly, in the water bug *Abedus herbeti*

Hidalgo, 1935 females lay their eggs on the males' back, which then move (with the eggs) to

ensure that they receive a proper level of oxygenation (Smith 1997). However, whether active

egg transport (e.g. parents actively moving their eggs from one location to another) could be

an adaptive behaviour by which mothers adjust the thermal needs of the embryo during

development remains unexplored. Yet, this process could operate in the European earwig

Forficula auricularia Linnaeus, 1758. In this complex of cryptic species (Wirth et al. 1998;

González-Miguéns et al. 2020) that can be found on almost all continents (Lamb and

Wellington 1975; Guillet, Josselin, et al. 2000; Quarrell et al. 2018; Hill et al. 2019), females

usually lay their eggs just before or during winter, and then remain with their eggs several

weeks or months until they hatch (Lamb 1976; Gingras and Tourneur 2001). During this period,

females provide extensive forms of egg care including, for instance, grooming behaviours to

remove pathogens, the application of chemical compounds on eggshells to improve resistance

against desiccation (Liu et al. 1997; Boos et al. 2014), and fierce protection against predators

(Thesing et al. 2015; Van Meyel et al. 2019). Moreover, females are frequently observed

transporting their eggs from one location to another by holding them individually between

their mouthparts (Diehl and Meunier 2018; Meunier et al. 2020).

Recent results and observations suggest that earwig eggs could benefit from

temperature-dependent oviposition site selection and temperature-dependent maternal

transport during development. First, the duration of egg development in winter varies from

three weeks (e.g. in Southern Europe) to several months (e.g. in North America) (Ratz et al.

2016; Tourneur 2018). This suggests that eggs could be exposed to extremely low (and

damaging) temperatures for a very long time if females do not select the oviposition site

accordingly and/or do not transport the eggs during development. Second, a recent study

shows that prolonging egg exposure to cold (5°C) for 15-day during winter delays the hatching

date and development of juveniles to adulthood (which typically takes two months in *F.*

auricularia), leads to the production of lighter adult females, and shapes the basal immunity

of these females: it increases their overall phenoloxidase activity and reduces the number of

haemocytes in females facing a changing social environment (Körner et al. 2018). Because

these traits are likely to affect negatively and/or positively the fitness of the resulting adults

(Koch and Meunier 2014), temperature-dependent oviposition site selection and egg

transport during development could be adaptive strategies in *F. auricularia* mothers. Finally,

several laboratory breeding trials indicate that eggs of some populations need to be exposed

to near-zero temperatures to trigger embryo development and that subsequently the

temperature needs to be increased to continue this development, while others do not (e.g.

Wirth et al. 1998; Meunier et al. 2012; Ratz et al. 2016; Tourneur and Meunier 2020).

In this study, we investigated whether European earwig females select an oviposition

site based on environmental temperature, and then move their eggs depending on egg age

and experimental changes in environmental temperature. Because previous studies point out

that the European earwig may show population-specific life-history traits and behaviours (Ratz

et al. 2016; Tourneur 2017; Tourneur 2018; Tourneur and Meunier 2020), we also tested

whether temperature-dependent oviposition site selection and egg transport vary between

two (randomly selected) populations naturally and experimentally sharing comparable

climatic conditions. We set up 60 females from two Canadian populations in two experimental

devices (Fig 1) allowing different thermal gradients and then recorded 1) the range of

temperatures explored by each female during the 15 days preceding oviposition, 2) the

temperature at which females laid their eggs, 3) whether and how mothers transported their

clutch along three thermal gradients throughout egg development and finally, 4) how the

temperature of these gradients affected juvenile production. Overall, our results reveal that

environmental temperature shapes female exploration before oviposition, oviposition site

selection, egg transport during development and the production of juveniles. They also show

that both oviposition site selection and the dynamic of egg transport during development

varies between the tested populations.

MATERIAL AND METHODS

Earwig sampling and experimental process

The experiment involved a total of 60 *F. auricularia* females field sampled in Harvey station

(67°00'52.2"W, 45°38'23.6"N; New Brunswick, Canada; HNB) and St John's (47°34'42.6"N,

52°44'37.0"W; Newfoundland and Labrador, Canada; SJNL) in September 2020 and then

maintained in plastic containers until October 2020. These two populations were selected as

independent random units on the basis that they were 2500 km apart by land (1000 km by

sea) and had comparable climatic conditions (Fig 2). Subsequent genetic analyses revealed

that HNB individuals belonged to the *F. auricularia* genetic clade 'A' and SJNL females to the

F. auricularia genetic clade 'B' (see details in the supplementary material and discussion; Wirth

et al. 1998, González-Miguéns et al. 2020). The field sampled females were maintained in

groups with males from the same population (sampled at the same time as the females) to

allow the completion of the gregarious phase of the life-cycle, which lasts several months

during which females mate with multiple partners (Sandrin et al. 2015) and express simple

social behaviours (Costa 2006; Weiß et al. 2014). The plastic containers were lined with wet

sand, contained two small shelters, and were maintained at room temperature under natural

day : light. In October 2020, we isolated each female as this is the period when they usually

leave their group to search for a future nesting site (Lamb 1976). To this end, we transferred

36 of 60 females (18 from HNB and 18 from SJNL) to the middle of 36 aluminium rails (Fig 1C;

1.8 x 1. 8 x 72 cm = height x width x length) covered with a layer of wet sand and closed with

a plastic cover for subsequent measurement of the temperature range explored before and

at oviposition. We isolated the remaining 24 females (12 from HNB and 12 from SJNL) in Petri

dishes (diameter 10 cm) covered with moist sand and kept in complete darkness until their

use in the measurement of egg transport after oviposition.

To determine the temperature range explored by each female before egg production

and the temperature chosen to deposit eggs, we transferred the 36 aluminium rails containing

one female each on thermoelectric plates (AHP-1200CPV- Thermoelectric Cooling America

Corporation. 4049W Schubert Avenue. Chicago IL. USA) with temperatures linearly ranging

from 0°C to 20°C (Fig 1A and S1). We then measured the distance between each female and

the coldest edge of its rail every day from the time they were placed in the rails until they

produced eggs (including the day of oviposition). Because the date of oviposition greatly varies

within populations (up to several months between the first and last oviposition; e.g. Tourneur

and Meunier 2020), we standardised our measurement to the temperature range explored by

each female 15 days before its own oviposition. To facilitate distance measurement, we

divided each rail into 12 zones of 6 cm length and defined the distance between a female and

the coldest edge as the centre of the zone she was in. In the very few cases where females

were observed between two zones, we assigned females' location to the colder of the two

zones. From field sampling to oviposition, we fed the 36 females twice a week with fresh

carrots placed on a soaked cotton pad, plus an artificial diet composed of 1/3 dry power of

egg yolk, 1/3 bee collected pollen (Community Apiaries. 576 Plymouth road, Richmond Corner,

New-Brunswick. Canada) and 1/3 cricket powder (Entomo Farms. 31 industrial drive Norwood,

Ontario. Canada). We then did not feed females from the day of oviposition to egg hatching,

as earwig mothers typically stop foraging during this period (Kölliker 2007).

We then tested whether mothers moved their eggs depending on experimental

changes in environmental temperature and/or egg age during the 15 weeks following

oviposition. Three days after each female has laid its eggs, we transferred the mother and all

its eggs to a new (shorter) aluminium rail (1.8 x 1.8 x 66 cm), which was deposited into a

climate cabinet (Fig 1B) providing three non-linear ranges of temperature: warm (0.9°C to

7.0°C), intermediate (-3.6°C to 2.7°C) or cold (-4.5°C to 1.6°C) (Fig 1B and S1). These three

thermal gradients encompass the above-ground natural range of temperatures of the two

populations during the time females were maintained in our laboratory, i.e. during the natural

period of egg care (Gingras and Tourneur 2001; Fig 2). To experimentally change the

temperature at which females and eggs were maintained at oviposition, we deposited each

female and its eggs in the middle of the new rail (Fig 1C), i.e. at either 5.2°C (warm), 1.2°C

(intermediate) or 0.1°C (cold). We then measured the distance (in cm) between the centre of

the pile of eggs and the coldest edge of the rail once a week during the 15 following weeks.

This measurement of egg transport involved the 36 females used in the measurements of

temperature range explored before egg production (see above), and the 24 females (12 from

HNB and 12 from SJNL) previously maintained in Petri dishes until oviposition (60 females

total). We had not been able to use these 24 females in the electrothermal plates due to the

lack of space in the units. Overall, we thus tested 10 HNB and 10 SJNL females in the warm,

10 HNB and 10 SJNL females in the intermediate and 10 HNB and 10 SJNL females in the cold

range. In the climate cabinet, the rails were lined with a layer of wet sand and closed with a

plastic cover to ensure complete darkness. They were slightly shorter than those used in the

thermoelectric plates for reasons of fit.

We finally tested whether the temperatures at which eggs were maintained during the

15 weeks following oviposition affect egg development and hatching rate. Fifteen weeks after

oviposition, we transferred the 60 (short) rails containing the mothers and their eggs (i.e. 5

rails per thermal range and population) to thermoelectric plates with temperatures linearly

ranging from 0°C to 19°C (and not 20°C due to the shorter rails). This transfer allowed females

to have access to a wider thermal range, as expected in Spring – the moment at which eggs

typically hatch in *F. auricularia*. We then checked each female daily to record the date of the

first egg hatching, the location of the clutch at hatching (based on the distance between the

centre of the clutch and the coldest edge of the rail) and the number of newly hatched

juveniles. All distance recordings were made very gently so that the females were not

disturbed during the observation.

We recorded the thermal gradients present in the different types of aluminium rails to

the nearest 0.1°C using 4 channel K type Thermometers (SD. Amazon. CA) connected to four

probes located in the sand either at 2.0, 25.5, 49.0 and 72.0 cm (long aluminium rails) or 1.5,

22.5, 43.5 and 64.5 cm (short aluminium rails) of the coldest edge of the rails. The recordings

occurred every hour during the entire experiment on either four aluminium rails evenly

distributed over the thermoelectric plates or six aluminium rails evenly distributed among the

trails and thermal constraints in the climate cabinet. There were no edge effects on

temperatures. We then used these recordings to compute linear (thermoelectric plates) and

non-linear (climate cabinet) equations linking distance to temperature (Fig S1), which we then

used to obtain the temperature of the location of the females and their eggs.

Statistical analyses

We used a series of eight non-parametric Exact Mann-Whitney Rank Sum tests correcting for

tied observations to test the effect of population on the amplitude of temperatures at which

females were observed before oviposition, the warmest and coldest temperatures reached by

females before oviposition, the location of females at oviposition, the date of oviposition, the

number of eggs produced, the number of weeks until egg hatching and the temperature of

the area of egg hatching. Changes in the location of the clutch after oviposition were then

analysed using parametric Linear Mixed-effects models (LME) in which the week (1 to 15),

population (SJNL and HNB), range of temperature (warm, intermediate and cold) and the

interaction between these three factors were entered as fixed explanatory factors, while the

ID of each female was used as a random factor to correct for multiple measurements. To

interpret the significant interaction between the three fixed explanatory factors, we divided

the data set according to temperature range (i.e. in three subsets), in which we ran another

series of LMEs in which we used week, population and the interaction as fixed explanatory

factors, and the female ID as a random factor. When the interaction between weeks and

population were significant, we conducted post hoc pairwise comparisons between the

temperature of the location where the eggs were initially transferred (i.e. 5.2, 1.18 and 0.11°C)

and the temperature of the location where the eggs were observed each week using a series

of one sample Exact Mann-Whitney Rank Sum tests. To correct for multiple testing, the P-

values of this series of pairwise comparisons were adjusted using the False Discovery Rate

(FDR) method (Benjamini and Hochberg 1995). Finally, the likelihood to produce at least one

juvenile and the hatching rate were tested using two Generalized linear models with binomial

error distribution and corrected for overdispersion. In these models, the population, the range

of temperatures and their interaction were entered as explanatory factors, while we used

either the presence of at least one hatched juvenile as a binary response variable (1 or 0) or

the hatching rate (the number of eggs that hatched divided by the total number of eggs

produced) as a continuous response variable. We checked that the assumptions of the LMEs

were met using the DHARMa package (Hartig 2020). The analyses were conducted with the

software R v4.1.1 (R Core Team, 2017) loaded with the packages *car* (Fox and Weisberg 2019),

exactRankTests (Hothorn and Hornik 2021), *emmeans* (Lenth 2021) and *DHARMa* (Hartig

2020). The R-script and data are available in an open data repository (see below).

RESULTS

Earwig females explored non-random ranges of temperature during the 15 days preceding

oviposition (Fig 3). The maximum temperature range was greater for the more artic SJNL than

the HNB females (median = 12.2 and 7.8°C, respectively; Fig 3A; W = 53, P < 0.001). The

warmest temperature at which females were observed was higher in SJNL than HNB females

(median = 19.6 and 12.7°C, respectively; Fig 3B; W = 2, P < 0.001), whereas the coldest location

was the same for SJNL and HNB females (median = 5.7 and 4.0°C, respectively; Fig 3C; W =

135.5, P = 0.404).

Earwig females chose their oviposition site according to the environmental

temperature (Fig 3). The temperature of the oviposition site was higher in SJNL compared to

HNB females (median = 16.1 and 9.2°C, respectively; Fig 3D; W = 17.5, P < 0.001). Moreover,

HNB females produced their eggs earlier in the season (median = 12 and 60 days after first egg

production in all females tested, respectively; Fig 3E; W = 38, P < 0.001) and laid overall more

eggs (median = 68 and 46, respectively; Fig 3F; W = 261, P < 0.001) than SJNL females.

During the 15 weeks following oviposition, earwig mothers moved their eggs

depending on experimental changes in environmental temperature, egg age and population

(Fig 4; Triple interaction between weeks, population and range of temperature: Likelihood

ratio χ^2_5= 33.13, P < 0.001). Egg location depended on an interaction between week and

population in the warmest range of temperatures (LR χ^2= 12.43, P < 0.001), which reveals that

SJNL females moved their eggs towards warmer locations about 7 weeks after oviposition,

whereas HNB females did not specifically target warmer locations (Fig 4). Egg location also

depended on an interaction between week and population in both the intermediate and the

coldest range of temperatures (LR χ^2= 16.53, P < 0.001 and χ^2= 6.27, P = 0.012, respectively),

which shows that both SJNL and HNB females moved their eggs towards warmer locations but

that this move started six weeks earlier in SJNL compared to HNB populations (Fig 4).

Interestingly, the mothers did not only move their eggs on the sand but built new nests each

time they moved their eggs.

Finally, the temperatures at which eggs were maintained during the 15 weeks

following oviposition affected both egg development and hatching rate. The likelihood to

produce juveniles (i.e. that at least one egg hatched) and the egg hatching rate were overall

higher for HNB compared to SJNL females (Fig 5A; 53% versus 20%; LR χ^2= 8.52, P = 0.004 and

Fig 5C; 30% versus 12%, LR χ^2= 6.20, P = 0.013, respectively), overall higher in females

previously maintained under the warmest range of temperatures (Fig 5B; 60% versus 25% and

25%; LR χ^2= 8.09, P = 0.018 and Fig 5D; 36% versus 12% and 15%, LR χ^2=7.99, P = 0.018,

respectively), and not affected by the interaction between these two factors (LR χ^2= 3.49, P =

0.175 and LR χ^2=0.61, P = 0.737, respectively). When they hatched, the temperature of the

place at which the first juveniles were observed was the same for both populations (Fig 5C; W

= 60, P = 0.396), while the eggs took overall more time to develop in HNB compared to SJNL

females (Fig 5D; W = 80.5, P = 0.012).

DISCUSSION

While temperature often drives oviposition site selection to ensure that the lack of egg

mobility does not result in exposure to sub-optimal or extreme temperatures (Pike et al. 2012;

Li et al. 2018; Ostap-Chec et al. 2021), the role of temperature on oviposition site selection is

less clear when mothers can transport the eggs after oviposition. Here, we show in the

European earwig that this capability does not prevent females from selecting oviposition sites

according to temperature and that egg transport can indeed help mothers to adjust egg

temperature after oviposition. Although this set of behavioural thermoregulations is present

in the two tested populations, we found that their modality of expression varied between the

two tested populations: St John's (Newfoundland and Labrador, Canada) females explore a

greater range of temperatures before oviposition, lay their eggs in warmer areas, move their

eggs quicker toward warm locations when suddenly exposed to cold, but were overall less

likely to produce juveniles under our experimental conditions than mothers from Harvey

station (New Brunswick, Canada).

Earwig females of both populations chose oviposition sites with temperatures above

10°C, which is in stark contrast to the sub-zero temperatures measured above ground in these

two locations during the natural oviposition period (Fig 2). This finding highlights the fact that

some ectotherms, such as earwigs, cannot only develop physiological mechanisms to

withstand their own freezing during wintering (Toxopeus and Sinclair 2018), but may also

prefer to find places exhibiting a large thermal difference from the ground temperature to

establish nests and deposit their eggs (Leather et al. 1993). Multiple strategies have been

reported in insects to achieve this goal, among which digging nests or burrows to obtain

efficient isolation from above-ground temperatures (Davis et al. 2015; Huang et al. 2020),

hiding under rocks and nesting into trunks to use the thermal inertia of the substrate as a

shelter (Brower et al. 2009; Trájer et al. 2014), and nesting close to human constructions (e.g.

underground pipeline, houses, building walls, etc) to benefit from their constant source of

heat during winter (Labrie et al. 2008; Trájer et al. 2014). These three strategies are also likely

to be adopted by the European earwig, as earwig adults are frequently found in human

habitations, underground burrows and under rock and trunks during winter (Goodacre 1997;

Gingras and Tourneur 2001; Kölliker and Vancassel 2007; Binns et al. 2021). Moreover,

another study suggests that the nest proximity to human constructions could be an effective

overwintering strategy for Canadian populations of earwigs (Goodacre 1997).

When experimentally exposed to temperatures below 5.5°C after oviposition, earwig

mothers of both populations transported their eggs to warmer locations. Interestingly, these

eggs were less likely to hatch when mothers were experimentally prevented from reaching

warmer locations, i.e. when mother and eggs were maintained in the cold and intermediate

temperature ranges. These results overall support the hypothesis that egg transport is an

adaptive post-oviposition behaviour by which earwig mothers protect eggs against extreme

cold and/or adjust the thermal needs of their embryos. This discovery sheds light on a new

strategy in female insects that overwinter with their eggs for coping with temperature changes

(Lee and Dellinger 1991; Sinclair et al. 2003), and emphasizes that the potential costs

associated with building a nest and finding a burrow in another location during winter do not

prevent such egg transport (Danks 2002). It now calls for future studies on the physiological

costs of egg transport for females at a time when they typically stop their foraging activity

(Kölliker 2007)(but see Van Meyel and Meunier 2020), and on the impact of temperature

variation during egg development (see Figure 4) on hatching success and offspring quality.

More generally, the egg transport capability of earwig females combined with their

oviposition site selection based on temperature could explain, at least in part, how such an

insect with long-overwintering eggs has been able to invade extremely cold climates (Guillet,

Josselin, et al. 2000; Quarrell et al. 2018; Hill et al. 2019; Tourneur and Meunier 2020).

Interestingly, St John's females transported their eggs much earlier than Harvey station

females. This suggests that the cold tolerance of eggs in the first few weeks after egg laying is

less effective at St John's than at Harvey station, either due to population-specific egg quality,

as reported in numerous oviparous species (Jing and Kang 2003; Stålhandske et al. 2015), or

population-specific egg development time, as egg sensitivity to cold often increases when they

are closer to hatching (e.g. Gray, 2009). In line with the last scenario, egg development time

greatly varies between natural populations of the European earwig (Tourneur and Gingras

1992; Meunier et al. 2012; Ratz et al. 2016; Tourneur and Meunier 2020), and our data show

that eggs produced by St John's females indeed take less time to hatch compared to eggs

produced by Harvey station females when reared in similar conditions. At a more general

level, it is typically expected that a population-specific cold tolerance of eggs should lead to

population-specific timing of oviposition, the less cold-tolerant eggs being laid later in winter

than the more cold-tolerant eggs (Tourneur and Meunier 2020). This is again in line with our

results: St John's females laid eggs about one month after Harvey station females. Overall, our

results thus demonstrate that winter temperatures are an important factor in the pre- and

post-oviposition strategies of earwig females.

Somewhat surprisingly, our study finally reveals that the modality of expression of the

reported strategies to protect eggs against severe winter cold varies between the two studied

populations. Although our experimental design does not allow us to conclude robustly on the

reasons for this variation, we propose three potential explanations. First, this could be due to

local adaptation to environmental conditions. In line with this explanation, previous studies

reported that multiple traits can vary between populations of the European earwig, such as

the number of clutches produced by females, clutch size, juvenile quality, the timing of egg

production, duration of egg development and female body mass (Ratz et al. 2016; Tourneur

2017; Tourneur 2018; Tourneur and Meunier 2020). However, these studies often compared

populations with contrasting climatic conditions, which was not the case between St John's

and Harvey stations (Fig 2). Second, the reported variation could be due to population-specific

differences in the phenology of females at field sampling. However, this is unlikely to explain

our results, as the females were sampled late in the breeding season and our setup control

for this potential variation by standardising the measurements around the natural day of

oviposition for each female. Finally, the behavioural variation reported between the two

tested populations could be due to the presence of different genetic clades. In line with this

hypothesis, our genetic analyses reveal that females of Harvey station belong to the genetic

clade 'A' and females of St John's to the genetic clade 'B' (this was surprising, as this is the first

time that the clade 'B' is found in Canada outside British Columbia). The European earwig is a

complex of cryptic species, for which genetic divergence and reproductive isolation are well

established (Wirth et al. 1998; Guillet, Josselin, et al. 2000; González-Miguéns et al. 2020), but

the specificity of their life-history traits remains largely unexplored. To date, the only known

species-specific trait refers to their reproduction, with females 'A' producing one clutch and

females 'B' producing two clutches (Wirth et al. 1998; Tourneur 2018). However, other studies

demonstrate that the number of clutches produced by a female can vary within *F. auricularia*

species (Tourneur and Gingras 1992; Ratz et al. 2016) depending on numerous parameters

acting during and after the early life of an earwig female (Meunier et al. 2012; Meunier and

Kölliker 2012; Wong and Kölliker 2014; Tourneur 2018). If this third hypothesis is true, our

results may thus have shed light on the first behavioural difference between 'A' and 'B'

females. Nevertheless, better understanding what drives population-specific dynamics of

maternal strategies to protect eggs against cold needs additional studies involving, for

instance, several populations of 'A' and 'B' females and/or population transplants.

Whereas ectotherms typically have very limited to no physiological capacities to limit

the costs of exposure to extreme temperatures via internal regulation (Stevenson 1985), some

have evolved forms of behavioural thermoregulation to avoid locations with extreme

temperatures (Lee Jr. 1991; Terrien 2011). Our study demonstrates that ectotherm females of

the European earwig adopt a broad set of behavioural thermoregulations when tending their

eggs overwinter. In particular, they select oviposition sites exhibiting specific temperatures

and transport their eggs to warmer locations when experimentally exposed to cold. While this

set of behavioural thermoregulations is present in the two studied populations, its modality

of expression varied between Harvey station and St John's – either as a result of local

adaptation or as species-specific traits among the complex of species composing the European

earwig (Wirth et al. 1998; Guillet, Josselin, et al. 2000; González-Miguéns et al. 2020). Overall,

our findings emphasize that earwig females have evolved behavioural strategies to mitigate

the risks inherent to tending eggs during several months and extreme winter cold. More

generally, they highlight the diversity of behaviours that insects can adopt to cope with

extreme temperatures, and could favour their tolerance to the effects of moderate climate

change.

DATA ACCESSIBILITY

397 The complete data set and R script are archived and freely available in the open data

398 repository Zenodo (https://doi.org/10.5281/zenodo.6338078).

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CONFLICT OF INTEREST

402   The authors of this preprint declare that they have no financial conflict of interest with the

403   content of this article. J Meunier is one of the PCI Zool recommenders.

REFERENCES

Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool.
J Mol Biol. 215(3):403–410. doi:10.1016/S0022-2836(05)80360-2.

Benjamini Y, Hochberg Y. 1995. Controlling the false discovery rate: a practical and powerful
approach to multiple testing. J R Stat Soc Ser B. 57(1):289–300. doi:10.2307/2346101.

Binns M, Hoffmann AA, Helden M, Heddle T, Hill MP, Macfadyen S, Nash MA, Umina PA.
2021. Lifecycle of the invasive omnivore, *Forficula auricularia*, in Australian grain growing
environments. Pest Manag Sci. 77(4):1818–1828. doi:10.1002/ps.6206.
https://onlinelibrary.wiley.com/doi/10.1002/ps.6206.

Boos S, Meunier J, Pichon S, Kölliker M. 2014. Maternal care provides antifungal protection
to eggs in the European earwig. Behav Ecol. 25(4):754–761. doi:10.1093/beheco/aru046.
http://www.beheco.oxfordjournals.org/cgi/doi/10.1093/beheco/aru046.

Brodin T, Johansson F, Bergsten J. 2006. Predator related oviposition site selection of aquatic
beetles (*Hydroporus* spp.) and effects on offspring life-history. Freshw Biol. 51(7):1277–1285.
doi:10.1111/j.1365-2427.2006.01563.x.

Brower LP, Williams EH, Slayback DA, Fink LS, Ramirez IM, Zubieta RR, Limon Garcia MI, Gier
P, Lear JA, Van hook T. 2009. Oyamel fir forest trunks provide thermal advantages for
overwintering monarch butterflies in Mexico. Insect Conserv Divers. 2(3):163–175.
doi:10.1111/j.1752-4598.2009.00052.x. https://onlinelibrary.wiley.com/doi/10.1111/j.1752-
4598.2009.00052.x.

Carrasco D, Kaitala A. 2009. Egg-laying tactic in *Phyllomorpha laciniata* in the presence of

parasitoids. Entomol Exp Appl. 131(3):300–307. doi:10.1111/j.1570-7458.2009.00857.x.

Cordero GA, Telemeco RS, Gangloff EJ. 2018. Reptile embryos are not capable of behavioral
thermoregulation in the egg. Evol Dev. 20(1):40–47. doi:10.1111/ede.12244.

Costa JT. 2006. Dermaptera. Earwig mothers. In: The other insect societies. Cambridge,
Massachusetts & London, England: The Belknap Press of Harvard University Press. p. 49–80.

Courtney SP. 1981. Coevolution of pierid butterflies and their cruciferous foodplants - III.
Anthocharis cardamines (L.) survival, development and oviposition on different hostplants.
Oecologia. 51(1):91–96. doi:10.1007/BF00344658.

Danks H V. 2002. Modification of adverse conditions by insects. Oikos. 99(1):10–24.
doi:10.1034/j.1600-0706.2002.990102.x. [accessed 2019 Sep 3].
https://onlinelibrary.wiley.com/doi/pdf/10.1034/j.1600-0706.2002.990102.x.

Davis JM, Coogan LE, Papaj DR. 2015. Big maggots dig deeper: size-dependent larval
dispersal in flies. Oecologia. 179(1):55–62. doi:10.1007/s00442-015-3314-6.
http://dx.doi.org/10.1007/s00442-015-3314-6.

Diehl JM, Meunier J. 2018. Surrounding pathogens shape maternal egg care but not egg
production in the European earwig. Behav Ecol. 29(1):128–136. doi:10.1093/beheco/arx140.
http://academic.oup.com/beheco/article/doi/10.1093/beheco/arx140/4560300.

Dillon ME, Wang G, Garrity PA, Huey RB. 2009. Thermal preference in Drosophila. J Therm
Biol. 34(3):109–119. doi:10.1016/j.jtherbio.2008.11.007.

Fey SB, Siepielski AM, Nusslé S, Cervantes-Yoshida K, Hwan JL, Huber ER, Fey MJ, Catenazzi
A, Carlson SM. 2015. Recent shifts in the occurrence, cause, and magnitude of animal mass
mortality events. Proc Natl Acad Sci U S A. 112(4):1083–1088.
doi:10.1073/pnas.1414894112.

Filazzola A, Matter SF, MacIvor JS. 2021. The direct and indirect effects of extreme climate
events on insects. Sci Total Environ. 769:145161. doi:10.1016/j.scitotenv.2021.145161.
https://doi.org/10.1016/j.scitotenv.2021.145161.

Fox J, Weisberg S. 2019. An R Companion to Applied Regression. Sage, Thousand Oaks CA,
3rd edition. URL http://z.umn.edu/carbook. http://z.umn.edu/carbook.

Gingras J, Tourneur J-C. 2001. Timing of adult mortality, oviposition, and hatching during the
underground phase of *Forficula auricularia* (Dermaptera: Forficulidae). Can Entomol.
133:269–278.

González-Miguéns R, Muñoz-Nozal E, Jiménez-Ruiz Y, Mas-Peinado P, Ghanavi HR, García-
París M. 2020. Speciation patterns in the *Forficula auricularia* species complex: cryptic and
not so cryptic taxa across the western Palaearctic region. Zool J Linn Soc. 190(3):788–823.
doi:10.1093/zoolinnean/zlaa070. https://academic.oup.com/zoolinnean/advance-
article/doi/10.1093/zoolinnean/zlaa070/5894910.

Goodacre S. 1997. The overwintering biology of the European earwig *Forficula auricularia*
Linneaus (Insecta: Dermaptera: Forficulinae). [Ottawa]: Carleton University.

Gould J, Clulow J, Rippon P, Doody JS, Clulow S. 2021. Complex trade-offs in oviposition site
selection in a cannibalistic frog. Anim Behav. 175:75–86.
doi:10.1016/j.anbehav.2021.02.021. https://doi.org/10.1016/j.anbehav.2021.02.021.

Gray DR. 2009. Age-dependent postdiapause development in the gypsy moth (Lepidoptera:
Lymantriidae) life stage model. Environ Entomol. 38(1):18–25. doi:10.1603/022.038.0104.

Guillet S, Guiller A, Deunff J, Vancassel M. 2000. Analysis of a contact zone in the *Forficula*
auricularia L. (Dermaptera: Forficulidae) species complex in the Pyrenean Mountains.
Heredity (Edinb). 85(5):444–449. doi:10.1046/j.1365-2540.2000.00775.x.
http://www.nature.com/doifinder/10.1046/j.1365-2540.2000.00775.x.

Guillet S, Josselin N, Vancassel M. 2000. Multiple introductions of the *Forficula auricularia*
species complex (Dermaptera: Forficulidae) in eastern North America. Can Entomol.
132(1):49–57. doi:10.4039/Ent13249-1.

Hajibabaei M, Janzen DH, Burns JM, Hallwachs W, Hebert PDN. 2006. DNA barcodes
distinguish species of tropical Lepidoptera. Proc Natl Acad Sci U S A. 103(4):968–971.
doi:10.1073/pnas.0510466103.

Hance T, Van Baaren J, Vernon P, Boivin G. 2007. Impact of extreme temperatures on
parasitoids in a climate change perspective. Annu Rev Entomol. 52:107–126.
doi:10.1146/annurev.ento.52.110405.091333.

Hartig F. 2020. DHARMa: Residual diagnostic for hierarchical (Multi-level / mixed) regression
models. http://florianhartig.github.io/DHARMa/.

Hill MP, Binns M, Umina PA, Hoffmann AA, Macfadyen S. 2019. Climate, human influence
and the distribution limits of the invasive European earwig, *Forficula auricularia*, in Australia.
Pest Manag Sci. 75(1):134–143. doi:10.1002/ps.5192.
http://doi.wiley.com/10.1002/ps.5192.

Hirayama H, Kasuya E. 2013. Effect of adult females' predation risk on oviposition site
selection in a water strider. Entomol Exp Appl. 149(3):250–255. doi:10.1111/eea.12124.

Hothorn T, Hornik K. 2021. Exact distributions for rank and permutation tests. R package
version v0.8-34.

Huang J, Li G, Lei H, Fan C, Tian C, Chen Q, Huang B, Li H, Lu Z, Feng H. 2020. Low-
temperature derived temporal change in the vertical distribution of *Sesamia inferens* larvae
in winter, with links to its latitudinal distribution. PLoS One. 15(7 July):1–15.
doi:10.1371/journal.pone.0236174. http://dx.doi.org/10.1371/journal.pone.0236174.

Jing XH, Kang L. 2003. Geographical variation in egg cold hardiness: A study on the
adaptation strategies of the migratory locust *Locusta migratoria* L. Ecol Entomol. 28(2-):151–
158. doi:10.1046/j.1365-2311.2003.00497.x.

Koch LK, Meunier J. 2014. Mother and offspring fitness in an insect with maternal care:
phenotypic trade-offs between egg number, egg mass and egg care. BMC Evol Biol.
14(1):125. doi:10.1186/1471-2148-14-125.
http://www.ncbi.nlm.nih.gov/pubmed/24913927.

Kölliker M. 2007. Benefits and costs of earwig (*Forficula auricularia*) family life. Behav Ecol
Sociobiol. 61(9):1489–1497. doi:10.1007/s00265-007-0381-7.
http://link.springer.com/10.1007/s00265-007-0381-7.

Kölliker M, Vancassel M. 2007. Maternal attendance and the maintenance of family groups
in common earwigs (*Forficula auricularia*): a field experiment. Ecol Entomol. 32(1):24–27.
doi:10.1111/j.1365-2311.2006.00831.x.

Körner M, Foitzik S, Meunier J. 2018. Extended winters entail long-term costs for insect
offspring reared in an overwinter burrow. J Therm Biol. 74:116–122.
doi:10.1016/j.jtherbio.2018.03.021.
http://linkinghub.elsevier.com/retrieve/pii/S0306456517304321.

Labrie G, Coderre D, Lucas É. 2008. Overwintering strategy of multicolored Asian lady beetle
(Coleoptera: Coccinellidae): Cold-free space as a factor of invasive success. Ann Entomol Soc
Am. 101(5):860–866. doi:10.1603/0013-8746(2008)101[860:OSOMAL]2.0.CO;2.

Lamb RJ. 1976. Parental behavior in the dermaptera with special reference to *Forficula
auricularia* (Dermaptera: Forficulidae). Can Entomol. 108(6):609–619.
doi:10.4039/ent108609-6.

Lamb RJ, Wellington WG. 1975. Life history and population characteristics of the european
earwig, *Forficula auricularia* (Dermaptera: forficulidae), at Vancouver, British columbia. Can J
Entomol. 107:819–824.

Lambret P, Rutter I, Grillas P, Stoks R. 2018. Oviposition plant choice maximizes offspring
fitness in an aquatic predatory insect. Hydrobiologia. 823(1):1–12. doi:10.1007/s10750-018-
3663-3. https://doi.org/10.1007/s10750-018-3663-3.

Leather SR, Walters KFA, Bale JS. 1993. The ecology of insect overwintering. Cambridge
University Press.
https://www.cambridge.org/core/product/identifier/9780511525834/type/book.

Lee Jr. R E. 1991. Principles of insect low temperature tolerance. In: Lee Jr. Richard E.,
Denlinger DL, editors. Insects at Low Temperature. Boston, MA: Springer US. p. 17–43.

Lee RE, Dellinger DL. 1991. Insects at low temperatures. New York, USA: Chapman and Hall.

Lenth R V. 2021. emmeans: Estimated Marginal Means, aka Least-Squares Means. R package
version 1.5.5-1. https://cran.r-project.org/package=emmeans.

Li S, Hao X, Wang Y, Sun B, Bi J, Zhang Y, Janzen FJ, Du W. 2018. Female lizards choose warm,
moist nests that improve embryonic survivorship and offspring fitness. Wilson R, editor.
Funct Ecol. 32(2):416–423. doi:10.1111/1365-2435.12995.
https://onlinelibrary.wiley.com/doi/10.1111/1365-2435.12995.

Liu Z, Vancassel M, Quris R. 1997. Female feeding aid in common earwig *Forficula auricularia*
L. Entomol Sin. 4(1):67–73. doi:10.1111/j.1744-7917.1997.tb00074.x.

Machado G, Trumbo ST. 2018. Parental care. In: Cordoba-aguilar A, Gonzalez-Tokman D,
Gonzalez-santoyo I, editors. Insect behavior: from mechanisms to ecological and
evolutionary consequences. Oxford: Oxford University Press. p. 203–218.

Meunier J, Dufour J, Van Meyel S, Rault M, Lécureuil C. 2020. Sublethal exposure to
deltamethrin impairs maternal egg care in the European earwig *Forficula auricularia*.
Chemosphere. 258:127383. doi:10.1016/j.chemosphere.2020.127383.
https://linkinghub.elsevier.com/retrieve/pii/S0045653520315769.

Meunier J, Kölliker M. 2012. Parental antagonism and parent-offspring co-adaptation
interact to shape family life. Proc R Soc London B Biol Sci. 279(1744):3981–8.
doi:10.1098/rspb.2012.1416.
http://rspb.royalsocietypublishing.org/cgi/doi/10.1098/rspb.2012.1416.

Meunier J, Körner M, Kramer J. 2022. Parental care. In: Omkar, Mishra G, editors.

Reproductive Strategies in Insects. Boca Raton: CRC Press. p. 337–348.
https://www.taylorfrancis.com/books/9781003043195.

Meunier J, Wong JWY, Gómez Y, Kuttler S, Röllin L, Stucki D, Kölliker M. 2012. One clutch or two clutches? Fitness correlates of coexisting alternative female life-histories in the European earwig. Evol Ecol. 26:669–682. doi:10.1007/s10682-011-9510-x. http://www.springerlink.com/index/10.1007/s10682-011-9510-x.

Van Meyel S, Devers S, Meunier J. 2019. Love them all: mothers provide care to foreign eggs in the European earwig *Forficula auricularia*. Behav Ecol. 30(3):756–762. doi:10.1093/beheco/arz012. https://academic.oup.com/beheco/article/30/3/756/5311762.

Van Meyel S, Meunier J. 2020. Filial egg cannibalism in the European earwig: its determinants and implications in the evolution of maternal egg care. Anim Behav. 164:155–162. doi:10.1016/j.anbehav.2020.04.001. https://linkinghub.elsevier.com/retrieve/pii/S0003347220300920.

Mortola JP, Gaonac'h-Lovejoy V. 2016. The cooling time of fertile chicken eggs at different stages of incubation. J Therm Biol. 55:7–13. doi:10.1016/j.jtherbio.2015.10.009. http://dx.doi.org/10.1016/j.jtherbio.2015.10.009.

Nicolai A, Vernon P, Lenz R, Le Lannic J, Briand V, Charrier M. 2013. Well wrapped eggs: Effects of egg shell structure on heat resistance and hatchling mass in the invasive land snail *Cornu aspersum*. J Exp Zool Part A Ecol Genet Physiol. 319(2):63–73. doi:10.1002/jez.1767.

Ostap-Chec M, Kierat J, Kuszewska K, Woyciechowski M. 2021. Red mason bee (*Osmia bicornis*) thermal preferences for nest sites and their effects on offspring survival. Apidologie. 52(3):707–719. doi:10.1111/j.1365- 2435.2011.01946.x.

Pike DA, Webb JK, Shine R, Ť DAP, Webb JK, Shine R. 2012. Hot mothers, cool eggs : nest-site selection by thermal optima egg-guarding spiders accommodates conflicting. Funct Ecol. 26(2):469–475. doi:65-2435.201 1.01946.x Hot.

Quarrell SR, Arabi J, Suwalski A, Veuille M, Wirth T, Allen GR. 2018. The invasion biology of the invasive earwig, *Forficula auricularia* in Australasian ecosystems. Biol Invasions. 20(6):1553–1565. doi:10.1007/s10530-017-1646-3. http://link.springer.com/10.1007/s10530-017-1646-3.

Ratz T, Kramer J, Veuille M, Meunier J. 2016. The population determines whether and how life-history traits vary between reproductive events in an insect with maternal care. Oecologia. 182(2):443–452. doi:10.1007/s00442-016-3685-3. http://link.springer.com/10.1007/s00442-016-3685-3.

Refsnider JM, Janzen FJ. 2010. Putting eggs in one basket: Ecological and evolutionary hypotheses for variation in oviposition-site choice. Annu Rev Ecol Evol Syst. 41(1):39–57. doi:10.1146/annurev-ecolsys-102209-144712. http://www.annualreviews.org/doi/10.1146/annurev-ecolsys-102209-144712.

Sandrin L, Meunier J, Raveh S, Walser J-C, Kölliker M. 2015. Multiple paternity and mating group size in the European earwig, *Forficula auricularia*. Ecol Entomol. 40(2):159–166. doi:10.1111/een.12171. http://doi.wiley.com/10.1111/een.12171.

Sicsú PR, Macedo RH, Sujii ER. 2020. Lady beetle oviposition site choices: Maternal effects on offspring performance. Florida Entomol. 103(2):228–235. doi:10.1653/024.103.0212.

Sinclair BJ, Vernon P, Klok CJ, Chown SL. 2003. Insects at low temperatures: An ecological
perspective. Trends Ecol Evol. 18(5):257–262. doi:10.1016/S0169-5347(03)00014-4.

Smith RL. 1997. Evolution of paternal care in the giant water bugs (Heteroptera:
Belostomatidae). In: Choe JC, Crespi BJ, editors. The evolution of social behavior in insects
and arachnids. Cambridge: Cambridge University Press. p. 116–149.

Stålhandske S, Lehmann P, Pruisscher P, Leimar O. 2015. Effect of winter cold duration on
spring phenology of the orange tip butterfly, *Anthocharis cardamines*. Ecol Evol. 5(23):5509–
5520. doi:10.1002/ece3.1773.

Stevenson RD. 1985. The relative importance of behavioral and physiological adjustments
controlling body temperature in terrestrial ectotherms. Am Nat. 126(3):362–386.

Terrien J. 2011. Behavioral thermoregulation in mammals: a review. Front Biosci. 16(1):1428.
doi:10.2741/3797. https://fbscience.com/Landmark/articles/10.2741/3797.

Thesing J, Kramer J, Koch LK, Meunier J. 2015. Short-term benefits, but transgenerational
costs of maternal loss in an insect with facultative maternal care. Proc R Soc London B Biol
Sci. 282(1817):20151617. doi:10.1098/rspb.2015.1617.
http://rspb.royalsocietypublishing.org/lookup/doi/10.1098/rspb.2015.1617.

Thompson JN. 1988. Evolutionary ecology of the relationship between oviposition
preference and performance of offspring in phytophagous insects. Entomol Exp Appl.
47(1):3–14. doi:10.1111/j.1570-7458.1988.tb02275.x.

Tourneur J-C. 2017. Epigeal phase of the biological cycle of *Forficula auricularia* Linnaeus
(Dermaptera: Forficulidae) in eastern Canada. Can Entomol. 149(5):600–606.
doi:10.4039/tce.2017.33.

Tourneur J-C. 2018. Factors affecting the egg-laying pattern of *Forficula auricularia*
(Dermaptera: Forficulidae) in three climatologically different zones of North America. Can
Entomol. 150(4):511–519. doi:10.4039/tce.2018.24.

Tourneur J-C, Gingras J. 1992. Egg laying in a northeastern North American (Montréal,
Québec) population of *Forficula auricularia* L. (Dermaptera: Forficulidae). Can Entomol.
124:1055–1061.

Tourneur J-C, Meunier J. 2020. Variations in seasonal (not mean) temperatures drive rapid
adaptations to novel environments at a continent scale. Ecology. 101(4):e02973.
doi:10.1002/ecy.2973. https://onlinelibrary.wiley.com/doi/abs/10.1002/ecy.2973.

Toxopeus J, Sinclair BJ. 2018. Mechanisms underlying insect freeze tolerance. Biol Rev.
93(4):1891–1914. doi:10.1111/brv.12425.

Trájer A, Mlinárik L, Juhász P, Bede-Fazekas Á. 2014. The combined impact of urban heat
island, thermal bridge effect of buildings and future climate change on the potential
overwintering of *Phlebotomus* species in a central European metropolis. Appl Ecol Environ
Res. 12(4):887–908. doi:10.15666/aeer/1204_887908.

Wang SF, Sun SC, Okazaki RK. 2010. Comparative study on thermotolerance of Artemia
resting eggs from Qinghai-Xizang Plateau, China. Aquaculture. 307(1–2):141–149.
doi:10.1016/j.aquaculture.2010.07.009.
http://dx.doi.org/10.1016/j.aquaculture.2010.07.009.

Weiß C, Kramer J, Holländer K, Meunier J. 2014. Influences of relatedness, food deprivation,

and sex on adult behaviors in the group-living insect *Forficula auricularia*. Ethology. 120:923–
932. doi:10.1111/eth.12261. http://doi.wiley.com/10.1111/eth.12261.

Wirth T, Guellec R Le, Vancassel M, Veuille M. 1998. Molecular and reproductive
characterization of sibling species in the european earwig (*Forficula auricularia*). Evolution.
52(1):260.

Wong JWY, Kölliker M. 2014. Effects of food restriction across stages of juvenile and early
adult development on body weight, survival and adult life history. J Evol Biol. 27(11):2420–
30. doi:10.1111/jeb.12484.

Yang C, Huang Q, Wang L, Du W-G, Liang W, Møller AP. 2018. Keeping eggs warm: thermal
and developmental advantages for parasitic cuckoos of laying unusually thick-shelled eggs.
Sci Nat. 105(1–2):10. doi:10.1007/s00114-017-1532-y.
http://link.springer.com/10.1007/s00114-017-1532-y.

Figure 1 – Schema of the thermal setups used in the experiment. (A) Schema of one of the two

thermoelectric plates with temperatures linearly ranging from 0°C to 20°C and insulated on

every side with thick foam (blue) to ensure complete darkness and thermal isolation. Each

plate received 16 rails, each containing one female until they laid eggs and then from the 15th

week after oviposition until egg hatching. (B) Schema of the homemade climate cabinet build

from a small vertical freezer whose door had been removed and the apparatus covered with

plywood enclosure (blue=foam) to ensure complete darkness and thermal isolation, and a

vertically sliding front door to allow removal. The cabinet was subdivided into three parts to

obtain warm (0.9°C to 7.0°C), cold (-4.5°C to 1.6°C) and intermediate (-3.6°C to 2.7°C)

temperature ranges. This cabinet hosted 60 females (20 per temperature range) and their

eggs for 15 weeks after oviposition. (C) Picture of a female with its eggs in an experimental

rail.

Figure 2 – Locations of Harvey Station (HNB, Canada; Blue) and St Johns (SJNL, Canada;

Orange), and their monthly variation of temperatures recorded from 1970 to 2000. Dots

depict mean monthly temperatures, with whiskers extending from minimum to maximum

mean temperatures. These data were extracted from the WorldClim database.

Figure 4 – Effect of population and temperature range on egg location during the 15 weeks

following oviposition. Brackets indicate thermal ranges. Dashed grey lines indicate the

location of mothers and eggs when the experiment was set up. The maximum temperature of

each area is indicated with a red line, while its minimum temperature is provided between

brackets. Grey dots are jittered experimental values for each clutch of eggs. Coloured dots are

mean values ± SE per week. Exact Mann-Whitney tests to compare values of each week to the

initial temperature: **P<0.01, *P<0.05. P-values corrected for multiple comparisons. HNB:

Harvey Station. SJNL: St Johns.

Figure 5 – Effect of population and temperature range on (A,B) the percentage of females with

at least one hatched egg and (C,D) the egg hatching rate. Effect of population on the location

of the eggs at the time of hatching (E) and the number of weeks between oviposition and egg

hatching (F). Figures E and F involved females from the warmest range of temperature only,

as their numbers were too limited in the other range of temperatures. Box plots depict median

(middle bar) and interquartile range (light bar), with whiskers extending to 1.5 times the

interquartile range and dots representing jittered experimental values. ns: P>0.05; *P<0.05;

**P<0.01; · p = 0.07. Different letters indicate P<0.05. HNB: Harvey Station. SJNL: St Johns.

SUPPLEMENTARY MATERIAL

Analyses of the genetic clades of *F. auricularia*

To determine whether HNB and SJNL females belonged to the same genetic clade (species) of the European earwig (Wirth et al. 1998; González-Miguéns et al. 2020), we analysed the Cytochrome Oxidase I (COI) gene of 6 females per population. This number of females is enough to robustly assess the origin of an entire population, as previous work demonstrated that the two species do not co-exist in the same population (Wirth et al. 1998; Guillet, Guiller, et al. 2000). Genomic DNA was extracted from whole individuals with the NucleoMag® Tissue kit (Macherey Nagel) following manufacturer instructions. The COI gene (658 bp) was amplified from each individual using routine barcoding primers LepF and LepR (Hajibabaei et al. 2006). PCR amplifications was performed with the DreamTaq® PCR Master Mix Kit (Thermo Scientific) using an ESCO Swift Maxi® thermocycler with an initial denaturation step at 95°C (2 min) followed by 35 cycles at 95°C (45 sec), 52°C (60 sec) and 72°C (90 sec) and finally an extension step at 72°C (10 min). PCR products were purified, and Sanger sequenced in both direction using an ABI 3730XL sequencing system (Thermo Fisher Scientific) at Eurofins Genomics Company. Sequences obtained were corrected using Geneious® 9.1.8. The species status of each female was identified from NCBI databases using the BLAST tool (Altschul et al. 1990). The BLAST results were ranked by percent identity and the reference sequences with at least 100% identity to our sequences were used to assign the species status. All sequences obtained in this study have been submitted to GenBank; their accession numbers are OL512959 to OL512964 for the 6 HNB females and OL512965 to OL512970 for the 6 SJNL females. The COI analyses revealed that all (6/6) HNB females belonged to the species

"*Forficula auricularia* A" and all (6/6) SJNL females belonged to the species "*Forficula*

auricularia B".

Figure S1 – Temperatures measured throughout the experiment in the thermoelectric plates

(Blue circles) and climate cabinet (Orange squares, Green diamonds, and Grey triangles) as a

function of distance from the coldest edges. In the thermal bridges, the linear regression

connecting the measurements is y = 0.2889x - 0.3452. In the climate cabinet, the polynomial

regressions connecting the measurements differed between the warmest (Orange square; y =

$-0.0012x^2$ + 0.1714x + 0.8747), intermediate (Blue circle; y = $-0.0015x^2$ + 0.1955x - 3.643) and

coldest (Grey triangles; y = $-0.0014x^2$ + 0.1842x - 4.445) temperature ranges.

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