

1 Comparison of subjective peripheral sensation, F-waves, and somatosensory evoked potentials in response
2 to a unilateral pinch task measured on the contractile and non-contractile sides

3 Short title: Neurophysiological parameters during a unilateral pinch task

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18

19 **Abstract**

20 Depression of sensory input during voluntary muscle contractions has been demonstrated using
21 electrophysiological methods in both animals and humans. However, the association between
22 electrophysiological responses of the sensory system and subjective peripheral sensation (SPS) during a
23 voluntary muscle contraction remains unclear. Our aim in this study was to describe the changes in SPS,
24 spinal α -motoneuron excitability (F-wave to M-wave amplitude), and somatosensory evoked potentials
25 (SEPs) during a unilateral pinch-grip task. Outcome variables were measured on the side ipsilateral and
26 contralateral to the muscle contraction, and at rest (control). Participants were 8 healthy men, 20.9 ± 0.8
27 years of age. The isometric pinch-grip task was performed at 30% of the maximum voluntary isometric
28 force measured for the right and left hand separately. The appearance rate of the F-wave during the task
29 was significantly higher for the ipsilateral (right) hand than for the contralateral (left) hand and control
30 condition. Although there was no difference in F-wave latency between hands and the control condition, the
31 amplitude of the F-wave was significantly higher for the ipsilateral (right) hand than for the contralateral
32 (left) hand and the control condition. There was no difference in the amplitude of the SEP at N20. However,
33 the amplitude at P25 was significantly lower for the ipsilateral (right) hand than for the contralateral (left)
34 hand and the control condition. The accuracy rate of detecting tactile stimulation, evaluated for 20
35 repetitions using a Semmes–Weinstein monofilament at the sensory threshold for each participant, was
36 significantly lower during the pinch-grip task for both the ipsilateral (right) and contralateral (left) hand
37 compared to the control condition. Overall, our findings show that SPS and neurophysiological parameters
38 were not modulated in parallel during the task, with changes in subjective sensation preceding changes in
39 electrophysiological indices during the motor task. Our findings provide basic information on sensory-
40 motor coordination.

41

42 Keywords: somatosensory evoked potential, F-wave, subjective peripheral sensation

43

44 **Introduction**

45 When peripheral nerves are electrically stimulated, the ascending afferent input is projected to the
46 somatosensory cerebral cortex via the spinal cord and the resulting cortical somatosensory evoked
47 potentials (SEPs) can be recorded. During voluntary muscle contraction, sensory information induced by
48 electrostimulation of the nerves supplying the contracting muscle is inhibited and the amplitude of SEPs
49 decreases [1–4]. This suppression of the sensory potential is known as “gating.” The amount of gating
50 during voluntary movement is dependent on the difficulty of the movement [5] and is observed during
51 both the pre-movement and movement phases [6,7]. Based on these findings, the main functional role of
52 gating is to eliminate unnecessary sensory information during the execution of purposeful voluntary
53 movements.

54 Studies using the H-reflex [8] and F-wave [9,10] have shown an increase in the excitability of spinal
55 α -motoneurons innervating active muscles during voluntary muscle contraction of the upper and lower
56 limbs. The excitability of spinal α -motoneurons has also been shown to increase with contractions of distal
57 [11] and contralateral [9] muscles. SEP gating has also been observed in the primary sensory cortex on
58 the side contralateral to active muscle contraction [12], although there is no consensus on this finding [13].
59 Moreover, although depression of sensory input during voluntary muscle contraction has been
60 demonstrated using electrophysiological methods in both animal and human studies, the association
61 between the electrophysiological response of the sensory system and subjective peripheral sensation (SPS)
62 during an active muscle contraction remains unclear.

63 In a previous study, we reported a reduction in cutaneous sensation on the dorsal surface of the hand
64 during an isometric pinch-grip task under submaximal conditions compared to a no-motion (rest) condition
65 [10]. However, it is not clear whether this response occurred locally only in the hand on the side of the
66 contraction or would also be observed on the non-contracting side. Therefore, our aim in this study was to
67 evaluate the changes in SPS, spinal α -motoneuron excitability, and SEPs on the side ipsilateral and
68 contralateral to an active contraction of a hand muscle, the right abductor pollicis brevis (APB).

69 **Materials and Methods**

70 *Participants and statement of ethics*

71 The study group included 8 healthy adult men (mean \pm SD age, 20.9 \pm 0.8 years; height, 170.0 \pm 4.5 cm;
72 and weight, 66.3 \pm 9.5 kg) with no history of neurophysiological diseases. Our study was conducted in
73 accordance with the principles of the Declaration of Helsinki. Informed consent was obtained from all
74 participants. The study was approved by the Ethical Review Board of Kibi International University (No.
75 20-41).

76 *Electrical stimulation protocol*

77 A square wave pulse, 0.2 ms in duration, was applied to the median nerve in the area of the carpal tunnel
78 of the right hand, using a surface electrostimulation apparatus (NM-420S, Nihon Kohden, Japan), to
79 stimulate the APB. The stimulation electrodes were placed 20 mm apart, with the cathode proximal and
80 anode distal. The stimulation electrodes were secured using a fixation band to regulate the pressure applied
81 to the electrodes. The minimum intensity of electrostimulation to induce an M-wave in the APB was
82 confirmed.

83 *F-wave recording and analysis*

84 Surface electromyogram (EMG) during electrostimulation was recorded over the right APB (side of
85 stimulation at the carpal tunnel) using Ag/AgCl bipolar electrodes (5 mm diameter, 20 mm interelectrode
86 distance; Nihon Kohden, Japan). The recording electrode was applied over the muscle belly of the APB,
87 with the reference electrode placed over the first proximal phalanx. The position of both electrodes was
88 fixed with surgical tape. Standard skin preparation was used prior for electrode placement: the site was
89 cleaned with rubbing alcohol and the skin abraded using sandpaper to achieve a skin resistance of <5 k Ω .
90 An EMG/evoked potential testing device (Neuropack MEB-9404, Nihon Kohden, Japan) was used. F-
91 waves were recorded using a band-pass filter (1.5–3 kHz), at a sampling frequency of 10 kHz. To obtain
92 the maximum M-wave, the intensity of electrostimulation was set to 1.2x the amplitude recorded for the
93 M-wave appearance in the right APB. The stimulation frequency was set to 1 Hz and was applied for

94 approximately 30 s. The following parameters of the F-wave were calculated during electrostimulation:
95 appearance rate (%), latency (ms), and amplitude of the F/M ratio (%). The appearance rate was the number
96 of F-waves observed on the monitor (threshold, 500 μ V/D) from the total of the 30 possible waves
97 generated by the electrostimulation. Latency was quantified as the average time from electrostimulation
98 to F-wave initiation. The amplitude of the F-wave was expressed as the ratio of the average peak-to-peak
99 amplitude of the F-wave to the maximum M-wave amplitude.

100 *SEP recording and analysis*

101 Based on the international 10–20 system, the SEPs were recorded from the somatosensory area of the
102 right upper arm (C3', 2 cm posterior to C3) on the side ipsilateral to the electrostimulation. The reference
103 electrode was placed at the point Fz. Electrodes were attached to the skin surface using a conductive paste,
104 with a skin resistance of <5 k Ω after standard preparation. The electrostimulation intensity was set to just
105 above the motor threshold, with a stimulation rate of 3 Hz. An EMG/evoked potential testing device
106 (Neuropack MEB-9404, Nihon Kohden, Japan) was used and SEP waveforms were recorded using a band-
107 pass filter (20 Hz to 10 kHz), at a sampling frequency of 10 kHz, with 200 responses averaged. SEP
108 waveforms were evaluated for 100 ms, both at the time of electrostimulation and 100 ms after stimulation.
109 Epochs with artifacts due to eye movement or blinking ($> \pm 6$ μ V from baseline) were excluded
110 automatically prior to averaging. A plate electrode was used to record the evoked electroencephalogram
111 (Ag/AgCl electrode, NE-132B (Φ , 10 mm), Nihon Kohden, Japan). The peak-to-peak amplitude of the
112 SPE at N20 and N20-P25, from baseline, which are early components after electrostimulation, were
113 analyzed.

114 *SPS measurement*

115 Prior to the experiment, the SPS threshold on the dorsal surface of the right hand was measured using
116 the Semmes-Weinstein monofilament test (SAKAI Medical Co., Ltd., Tokyo, Japan) [10]. After
117 establishing their peripheral sensory thresholds, participants reported the presence or absence of peripheral
118 cutaneous stimulation during SPS measurements. Monofilament stimulation was performed using

119 gradually thicker filaments, starting from thin filaments of 0.008 g. Confirmation tests were repeated
120 approximately five times for each intensity and the filament thickness that could be sensed correctly at a
121 rate of approximately 100% was defined as the SPS threshold. The experimenter lowered the filament
122 vertically onto the hand, removed it, and returned it to its original position within 1 s. The stimulation
123 interval of the filament was random and was repeated 20 times. Participants were instructed to give verbal
124 cues when they sensed filament stimulation and the accuracy rate of detection was calculated. All
125 measurements were performed by the same experimenter. The filament stimulation site was marked to
126 avoid experimenter error resulting in random deviation in measurement due to a large stimulation site.

127 *Experimental procedure*

128 Participants were seated in a chair with both arms placed on armrests, with eyes open to perform a
129 pinch-grip task. When the pinch-grip task was performed with the right hand, the left arm and hand were
130 maintained in a neutral position (no-motion, rest, condition). When the pinch-grip task was performed
131 using the left hand, the right arm and hand were positioned in the neutral position. Prior to task
132 performance, the maximum voluntary isometric force (MVIF) was measured separately for the right and
133 left hand, from which the target force level was calculated. To calculate the MVIF, participants held the
134 pinch force meter with the thumb and index finger and were asked to exert their maximum pinch force
135 and to hold this force for 5 s. The peak force measured over this 5 s epoch was defined as the MVIF. After
136 a sufficient rest period (≥ 10 min), the experimental task was performed. Participants were asked to
137 maintain a 30% MVIF for a duration of 2 min, with the target pinch force to be exerted displayed visually
138 on a computer screen placed 1 m in front of participants. The task was performed with both the right hand
139 (ipsilateral to the side of recording) and the left hand (contralateral to the side of recording). In the control
140 condition, no pinch force was exerted. The sequence of conditions was randomly selected across
141 participants.

142 *Statistics*

143 All values are presented as the mean \pm standard deviation. Differences in measured parameters between

144 conditions were evaluated using a repeated-measures analysis of variance. The sphericity of the data was
145 evaluated using Mauchly's test, with Greenhouse-Geisser-corrected significance values being used when
146 sphericity was not met. Post-hoc analysis was performed using Tukey's test for multiple comparisons. The
147 statistical significance level was set at 5% ($P < 0.05$) for all analyses. All analyses were performed using
148 GraphPad Prism (version 8.3.1 for Machintosh).

149

150 **Results**

151 A typical example of the M- and F-waves is shown in Fig 1A. The appearance rate of the F-wave (Fig
152 1B) was significantly higher for the side ipsilateral to the SEP recording (right hand, $85.1 \pm 11.3\%$) than
153 for the contralateral side (left hand, $36.4 \pm 19.8\%$) and control ($30.5 \pm 14.3\%$) condition ($F_{1.811, 12.68} = 39.78$,
154 $P < 0.01$). There were no significant differences in the F-wave latency (Fig 1C) between the control
155 condition (27.0 ± 1.8 ms), the ipsilateral side (right hand, 26.3 ± 1.5 ms), and contralateral side (left hand,
156 25.7 ± 3.1 ms; $F_{1.523, 10.66} = 1.049$, $P = 0.36$). Similar to the appearance rate, the F-wave amplitude (Fig 1D)
157 was significantly higher for the ipsilateral side (right hand, $7.4 \pm 4.4\%$) than for the contralateral side (left
158 hand, $3.2 \pm 1.4\%$) and control condition ($2.7 \pm 1.0\%$; $F_{1.060, 7.419} = 8.206$, $P = 0.02$).

159

160 **Fig 1.** A typical example of an induced electromyogram waveform (10 stimulations) from a single
161 participant in each condition. (A) The waveforms for the M-waves and F-waves are shown on the left and
162 right side of the central thick line, respectively, with the amplitude scaling being 10 X higher for the F-
163 than M-wave, for the control (rest), active contraction (right) side, and contralateral (left) side; (B) average
164 appearance rate of the F-wave appearance; (C) F-wave latency, and (D) F-wave amplitude. *, $P < 0.05$

165

166 A typical example of SEP waveforms is shown in Fig 2A. There were no differences in the SEP
167 amplitude at N20 (Fig 2B) between the control condition (3.41 ± 1.11 μV), ipsilateral side (right hand,
168 2.73 ± 1.16 μV) and contralateral side (left hand, 3.33 ± 0.87 μV ; $F_{1.247, 8.726} = 3.222$, $P = 0.10$). However, the

169 SEP amplitude at P25 (Fig 2C) was significantly lower for the ipsilateral side (right hand, $4.65 \pm 1.27 \mu\text{V}$)
170 than for the contralateral side (left hand, $6.36 \pm 1.52 \mu\text{V}$) and the control condition ($6.42 \pm 1.22 \mu\text{V}$;
171 $F_{1,478,10,34} = 14.63$, $P = 0.001$).

172

173 **Fig 2. (A).** A typical example of somatosensory evoked potentials (SEPs) waveforms from a single
174 participant (for 200 stimulations) for the control (rest) condition (solid black line), in the active contraction
175 (right) side (black dashed line), and contralateral (left) side (gray solid line). Changes in the average SEP
176 amplitude **(B)** at N20 and **(C)** P25. *, $P < 0.05$

177

178 The accuracy rate for the 20 repetitions of monofilament stimulations was significantly lower for the
179 ipsilateral (right) hand ($61.9 \pm 21.9\%$) and the contralateral (left) hand ($61.9 \pm 11.3\%$) than for the control
180 condition ($84.4 \pm 11.2\%$; $F_{1,297,9,080} = 8.158$, $P = 0.01$; Fig 3).

181

182 **Fig 3.** Changes in the average accuracy rate of detection for the 20 repetition monofilament test of all
183 participants for the control (rest) condition, the active contraction (right) side, and the contralateral (left)
184 side. *, $P < 0.05$

185

186 Discussion

187 A novel observation of our study was that the pinch-grip task significantly reduced the SPS with active
188 contraction of the right hand in both the ipsilateral (right) and contralateral (left) side compared to the
189 control condition, although a significant increase in the appearance rate and amplitude of the F- wave and
190 a significant decrease in the amplitude of the SEP (P25) were observed only on the ipsilateral side.

191 F-waves are muscle action potentials recorded when electrostimulation to a peripheral nerve causes
192 retrograde conduction within the axon of an α -motoneuron, followed by subsequent anterograde
193 conduction through the automatic firing of the α -motoneuron in the anterior horn of the spinal cord. In our

194 study, the F-wave appearance rate for the right APB increased during the isometric pinch-grip task
195 performed by the right hand (ipsilateral to the cortical recording side). The F-wave appearance rate
196 indicates the number of motor units participating in the waveform [14] and naturally varies, even at rest.
197 This natural variation suggests that sensory input influences the recruitment of α -motoneurons. This
198 variation in the F-wave appearance rate increased during the isometric pinch-grip task in the ipsilateral
199 (right) hand, which might reflect a suppression of activity within the corticospinal tract, which converges
200 on the spinal anterior motor nerve, and inhibition via higher (cortical) control systems. This regulatory
201 effect was not observed for contractions performed using the contralateral (left) hand, with no increase in
202 the appearance rate of the F-wave for the left hand. This finding is different from previous reports of
203 similar modulation of the F-wave on both the ipsilateral and contralateral side, with this difference likely
204 reflecting differences in the motor task performed. While we used a pinch-grip task, previous studies used
205 a hand-grip task, with the greater force generated by the hand-grip than the pinch-grip task increasing the
206 firing rate and recruitment of α -motoneurons [15–17]. We do note that another study reported an increased
207 responsiveness of neurons in the primary motor cortex for a precision (pinch-grip) rather than gross (power
208 grip) motor task [18]. The increased muscle recruitment during a power grip, compared to a pinch-grip,
209 task complicates the information measured from the upper motor centers due to the integrated processes
210 of the central nervous system. These integrated processes exert an inhibitory effect on the α -motoneurons
211 of the spinal anterior horn for muscles on the contralateral (non-contraction) side.

212 The source of the SEP at N20 is considered to be the 3b area of the primary somatosensory cortex,
213 representing the stage when the sensory stimulation reaches the primary sensory cortex via the thalamus
214 [19]. The source of the SEP at P25 is considered to be higher than the 3b area [20]. Therefore, the
215 submaximal isometric pinch-grip task performed with the right hand in our study caused suppression of
216 the ipsilateral somatosensory input at a higher level than the 3b area. Previous studies on SEP gating during
217 voluntary movement have reported an absence of gating in components corresponding to N20, which is
218 consistent with findings from previous studies. For these reasons, although the electrophysiological input

219 that is projected to the primary somatosensory area is the same for a given amount of physical stimulation
220 (regardless of the presence or absence of the motor task), this electrophysiological input is suppressed
221 during the subsequent more complex phase of information processing. Additionally, as the amplitude of
222 all components of the SEPs was not different between the left (contralateral) hand and the control (rest)
223 condition, it appears that the electrophysiological sensory input is projected from the periphery to the
224 primary somatosensory cortex, without influence from the contralateral muscle contraction.

225 In our study, we used the accuracy rate for cutaneous stimulation at the sensory threshold in the right
226 hand as an index of SPS. The accuracy rate for cutaneous stimulation was reduced by submaximal
227 isometric muscle contraction in both the ipsilateral (right) and contralateral (left) hands, compared to the
228 control (rest) condition. This decrease in accuracy rate resulted from an increased difficulty in recognizing
229 the sensory stimulation. There would be a need to develop a test in which the sensory information can be
230 correctly recognized during a voluntary muscle contraction. Compared to the resting state, transient
231 exercise has been shown to reduce skin temperature sensation [21], with the kinetic threshold also being
232 lower during active muscle contraction than at rest [22]. Therefore, the tactile threshold of the skin surface
233 may increase in the presence of a motor output, such as voluntary muscle contraction. Furthermore, the
234 sensory thresholds with a muscle contraction may vary to a certain degree, both increasing or decreasing
235 in amplitude. This phenomenon is also observed during muscle contraction on the contralateral side of the
236 filament stimulation. These results suggest that localized muscle contraction modulates the SPS even in
237 areas that are not related to a muscle contraction or movement.

238 The primary limitation to generalization of our findings is that the F-wave, SEP, and SPS measurements
239 were conducted in separate sessions and not measured simultaneously in real time. As such, identification
240 of the electrophysiological parameters corresponding to correct and incorrect results on the SPS test was
241 not possible. Real-time measurement of F-waves, SEP, and SPS in future research would clarify the
242 association between sensory-motor processes and subjective sensory changes in future studies.

243

244 **Conclusion**

245 Overall, our findings show that SPS and neurophysiological parameters were not modulated in parallel
246 during the task, with changes in subjective sensation preceding changes in physiological indices during
247 the motor task. Our findings provide basic information on sensory-motor coordination.

248

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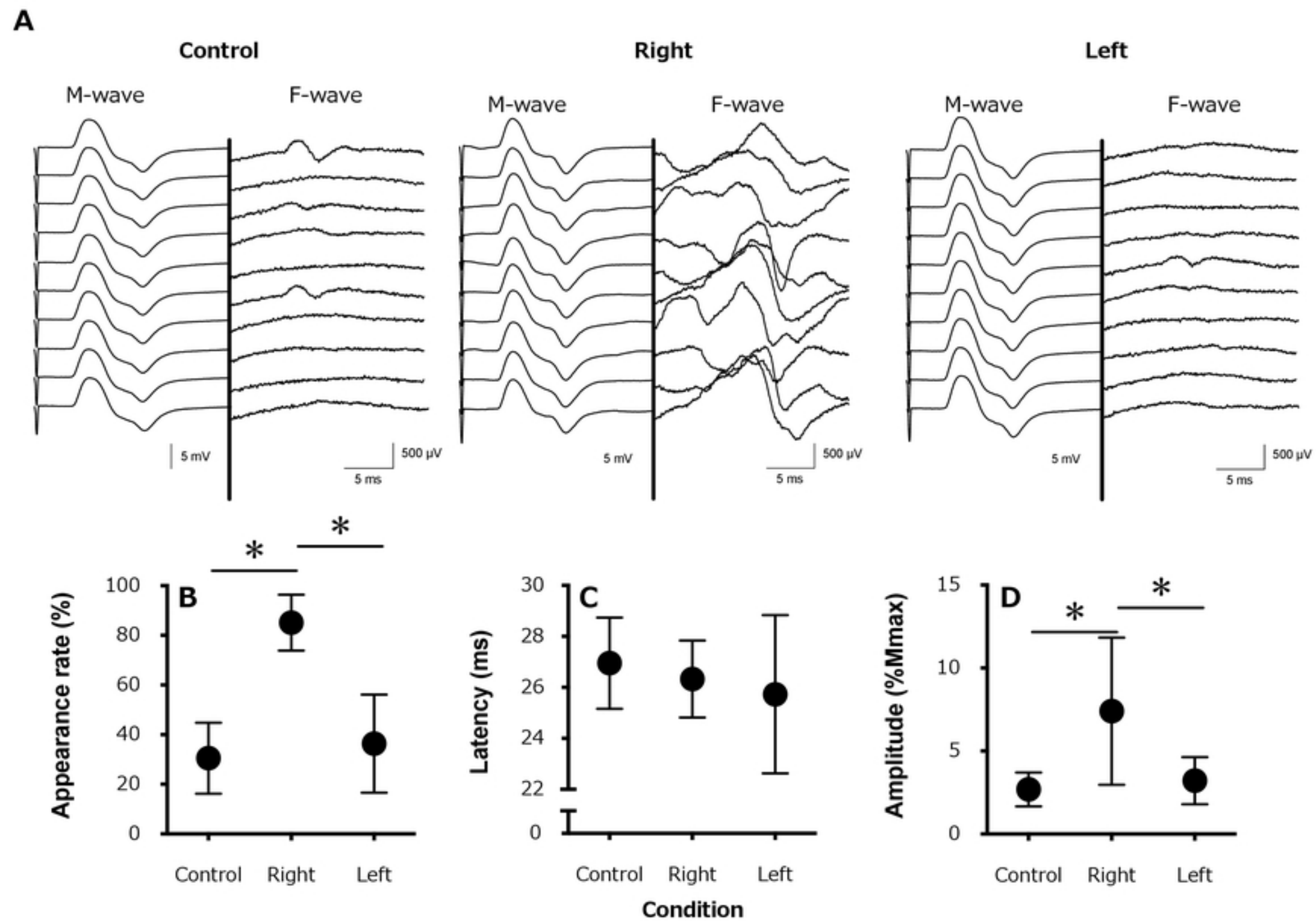
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268 **References**

- 269 1. Cheron G, Borenstein S. Gating of the early components of the frontal and parietal
270 somatosensory evoked potentials in different sensory-motor interference modalities.
271 Electroencephalogr Clin Neurophysiol. 1991;80: 522-530. doi: [10.1016/0168-5597\(91\)90134-](https://doi.org/10.1016/0168-5597(91)90134-j)
272 [j](https://doi.org/10.1016/0168-5597(91)90134-j), PubMed PMID: [1720728](https://pubmed.ncbi.nlm.nih.gov/1720728/).
- 273 2. Kakigi R, Koyama S, Hoshiyama M, Watanabe S, Shimojo M, Kitamura Y. Gating of
274 somatosensory evoked responses during active finger movements magnetoencephalographic
275 studies. J Neurol Sci. 1995;128: 195-204. doi: [10.1016/0022-510x\(94\)00230-l](https://doi.org/10.1016/0022-510x(94)00230-l), PubMed
276 PMID: [7738596](https://pubmed.ncbi.nlm.nih.gov/7738596/).
- 277 3. Nakata H, Inui K, Wasaka T, Nishihira Y, Kakigi R. Mechanisms of differences in gating
278 effects on short-and long-latency somatosensory evoked potentials relating to movement.
279 Brain Topogr. 2003;15: 211-222. doi: [10.1023/a:1023908707851](https://doi.org/10.1023/a:1023908707851), PubMed PMID: [12866825](https://pubmed.ncbi.nlm.nih.gov/12866825/).
- 280 4. Rushton DN, Rothwell JC, Craggs MD. Gating of somatosensory evoked potentials during
281 different kinds of movement in man. Brain. 1981;104: 465-491. doi: [10.1093/brain/104.3.465](https://doi.org/10.1093/brain/104.3.465),
282 PubMed PMID: [7272711](https://pubmed.ncbi.nlm.nih.gov/7272711/).
- 283 5. Kirimoto H, Tamaki H, Suzuki M, Matsumoto T, Sugawara K, Kojima S, et al. Sensorimotor
284 modulation differs with load type during constant finger force or position. PLoS ONE. 2014;9:
285 e108058. doi: [10.1371/journal.pone.0108058](https://doi.org/10.1371/journal.pone.0108058), PubMed PMID: [24169486](https://pubmed.ncbi.nlm.nih.gov/24169486/), PubMed Central PMCID:
286 [PMC4169486](https://pubmed.ncbi.nlm.nih.gov/24169486/).
- 287 6. Bocker KB, Forget R, Brunia CH. The modulation of somatosensory evoked potentials during
288 the foreperiod of a forewarned reaction time task. Electroencephalogr Clin Neurophysiol.
289 1993;88: 105-117. DOI: [10.1016/0168-5597\(93\)90061-s](https://doi.org/10.1016/0168-5597(93)90061-s), PubMed PMID: [7681751](https://pubmed.ncbi.nlm.nih.gov/7681751/).
- 290 7. Nishihira Y, Araki H, Ishihara A. Suppression of cerebral evoked potentials preceding rapid
291 reaction movement. J Sports Med Phys Fitness. 1990;30: 291-296. PubMed PMID: [2266761](https://pubmed.ncbi.nlm.nih.gov/2266761/).

- 292 8. Burke D, Adams RW, Skuse NF. The effects of voluntary contraction on the H reflex of human
293 limb muscles. *Brain*. 1989;112: 417-433. doi: [10.1093/brain/112.2.417](https://doi.org/10.1093/brain/112.2.417), PubMed PMID:
294 [2706438](https://pubmed.ncbi.nlm.nih.gov/2706438/).
- 295 9. Takahara T, Yamaguchi H, Seki K, Murata M, Onodera S. Effect of circulatory system
296 response to motor control in one-sided contractions. *Eur J Appl Physiol*. 2018;118: 1773-1780.
297 doi: [10.1007/s00421-018-3907-y](https://doi.org/10.1007/s00421-018-3907-y), PubMed PMID: [29869712](https://pubmed.ncbi.nlm.nih.gov/29869712/).
- 298 10. Takahara T, Yamaguchi H, Seki K, Onodera S. Sensory gating and suppression of subjective
299 peripheral sensations during voluntary muscle contraction. *BMC Neurosci*. 2020;21: 41. doi:
300 [10.1186/s12868-020-00592-2](https://doi.org/10.1186/s12868-020-00592-2), PubMed PMID: [PubMed PMID](https://pubmed.ncbi.nlm.nih.gov/34111111/). PubMed Central PMCID: [PMC7528260](https://pubmed.ncbi.nlm.nih.gov/PMC7528260/).
- 301 11. Zehr EP, Stein RB. Interaction of the Jendrassik maneuver with segmental presynaptic
302 inhibition. *Exp Brain Res*. 1999;124: 474-480. doi: [10.1007/s002210050643](https://doi.org/10.1007/s002210050643), PubMed PMID:
303 [10090659](https://pubmed.ncbi.nlm.nih.gov/10090659/).
- 304 12. Staines WR, Brooke JD, Cheng J, Misiaszek JE, MacKay WA. Movement-induced gain
305 modulation of somatosensory potentials and soleus H-reflexes evoked from the leg. I.
306 Kinaesthetic task demands. *Exp Brain Res*. 1997;115: 147-155. doi: [10.1007/pl00005674](https://doi.org/10.1007/pl00005674),
307 PubMed PMID: [9224842](https://pubmed.ncbi.nlm.nih.gov/9224842/).
- 308 13. Tinazzi M, Zanette G, La Porta F, Polo A, Volpato D, Fiaschi A, et al. Selective gating of
309 lower limb cortical somatosensory evoked potentials (SEPs) during passive and active foot
310 movements. *Electroencephalogr Clin Neurophysiol*. 1997;104: 312-321. doi: [10.1016/s0168-
311 5597\(97\)00023-3](https://doi.org/10.1016/s0168-5597(97)00023-3), PubMed PMID: [9246068](https://pubmed.ncbi.nlm.nih.gov/9246068/).
- 312 14. Rivner MH. The use of F-waves as a probe for motor cortex excitability. *Clin Neurophysiol*.
313 2008;119: 1215-1216. doi: [10.1016/j.clinph.2008.01.103](https://doi.org/10.1016/j.clinph.2008.01.103), PubMed PMID: [18406201](https://pubmed.ncbi.nlm.nih.gov/18406201/).
- 314 15. Belhaj-Saif A, Fourment A, Maton B. Adaptation of the precentral cortical command to elbow
315 muscle fatigue. *Exp Brain Res*. 1996;111: 405-416. doi: [10.1007/BF00228729](https://doi.org/10.1007/BF00228729), PubMed
316 PMID: [8911934](https://pubmed.ncbi.nlm.nih.gov/8911934/).

- 317 16. Cheney PD, Fetz EE. Functional classes of primate corticomotoneuronal cells and their relation
318 to active force. *J Neurophysiol.* 1980;44: 773-791. doi: [10.1152/jn.1980.44.4.773](https://doi.org/10.1152/jn.1980.44.4.773), PubMed
319 PMID: [6253605](https://pubmed.ncbi.nlm.nih.gov/6253605/).
- 320 17. Maier MA, Bennett KM, Hepp-Reymond MC, Lemon RN. Contribution of the monkey
321 corticomotoneuronal system to the control of force in precision grip. *J Neurophysiol.* 1993;69:
322 772-785. doi: [10.1152/jn.1993.69.3.772](https://doi.org/10.1152/jn.1993.69.3.772), PubMed PMID: [8463818](https://pubmed.ncbi.nlm.nih.gov/8463818/).
- 323 18. Muir RB, Lemon RN. Corticospinal neurons with a special role in precision grip. *Brain Res.*
324 1983;261: 312-316. doi: [10.1016/0006-8993\(83\)90635-2](https://doi.org/10.1016/0006-8993(83)90635-2), PubMed PMID: [6831213](https://pubmed.ncbi.nlm.nih.gov/6831213/).
- 325 19. Allison T, McCarthy G, Wood CC, Darcey TM, Spencer DD, Williamson PD. Human cortical
326 potentials evoked by stimulation of the median nerve. I. Cytoarchitectonic areas generating
327 short-latency activity. *J Neurophysiol.* 1989;62: 694-710. doi: [10.1152/jn.1989.62.3.694](https://doi.org/10.1152/jn.1989.62.3.694),
328 PubMed PMID: [2769354](https://pubmed.ncbi.nlm.nih.gov/2769354/).
- 329 20. Allison T, McCarthy G, Wood CC, Williamson PD, Spencer DD. Human cortical potentials
330 evoked by stimulation of the median nerve. II. Cytoarchitectonic areas generating long-latency
331 activity. *J Neurophysiol.* 1989;62: 711-722. doi: [10.1152/jn.1989.62.3.711](https://doi.org/10.1152/jn.1989.62.3.711), PubMed PMID:
332 [2769355](https://pubmed.ncbi.nlm.nih.gov/2769355/).
- 333 21. Gerrett N, Ouzzahra Y, Redortier B, Voelcker T, Havenith G. Female thermal sensitivity to
334 hot and cold during rest and exercise. *Physiol Behav.* 2015;152: 11-19. doi:
335 [10.1016/j.physbeh.2015.08.032](https://doi.org/10.1016/j.physbeh.2015.08.032), PubMed PMID: [26343771](https://pubmed.ncbi.nlm.nih.gov/26343771/).
- 336 22. Taylor JL, McCloskey DI. Detection of slow movements imposed at the elbow during active
337 flexion in man. *J Physiol.* 1992;457: 503-513. doi: [10.1113/jphysiol.1992.sp019390](https://doi.org/10.1113/jphysiol.1992.sp019390), [PubMed](#)
338 [PMID](#). PubMed Central PMCID: [PMC1175743](https://pubmed.ncbi.nlm.nih.gov/PMC1175743/).
- 339



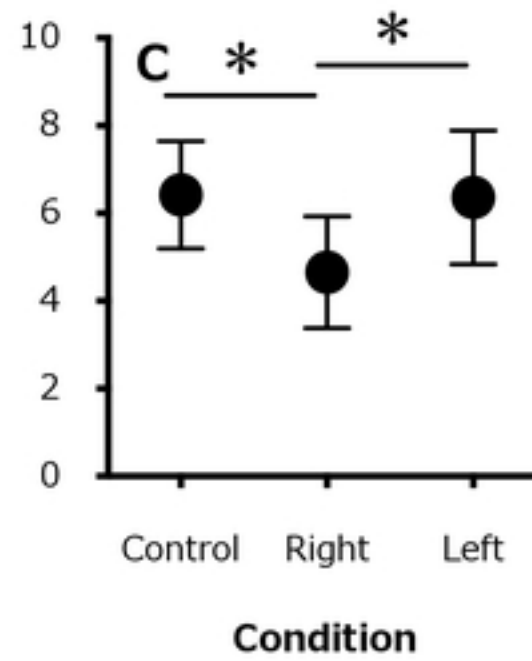
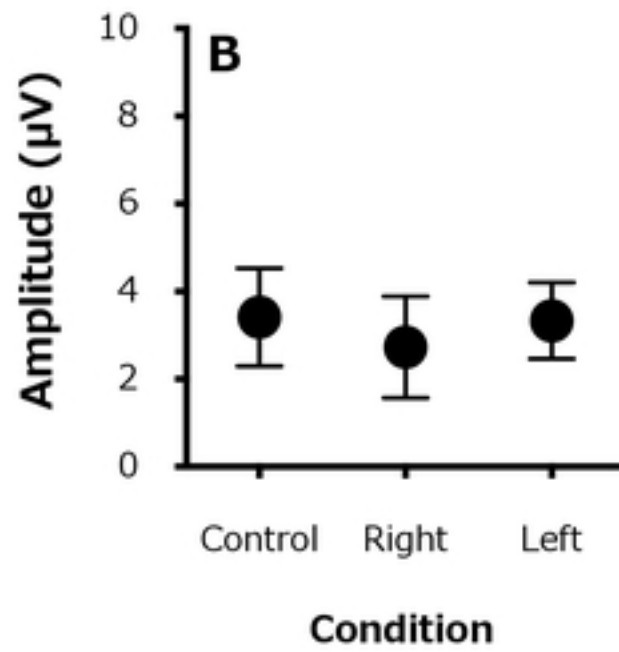
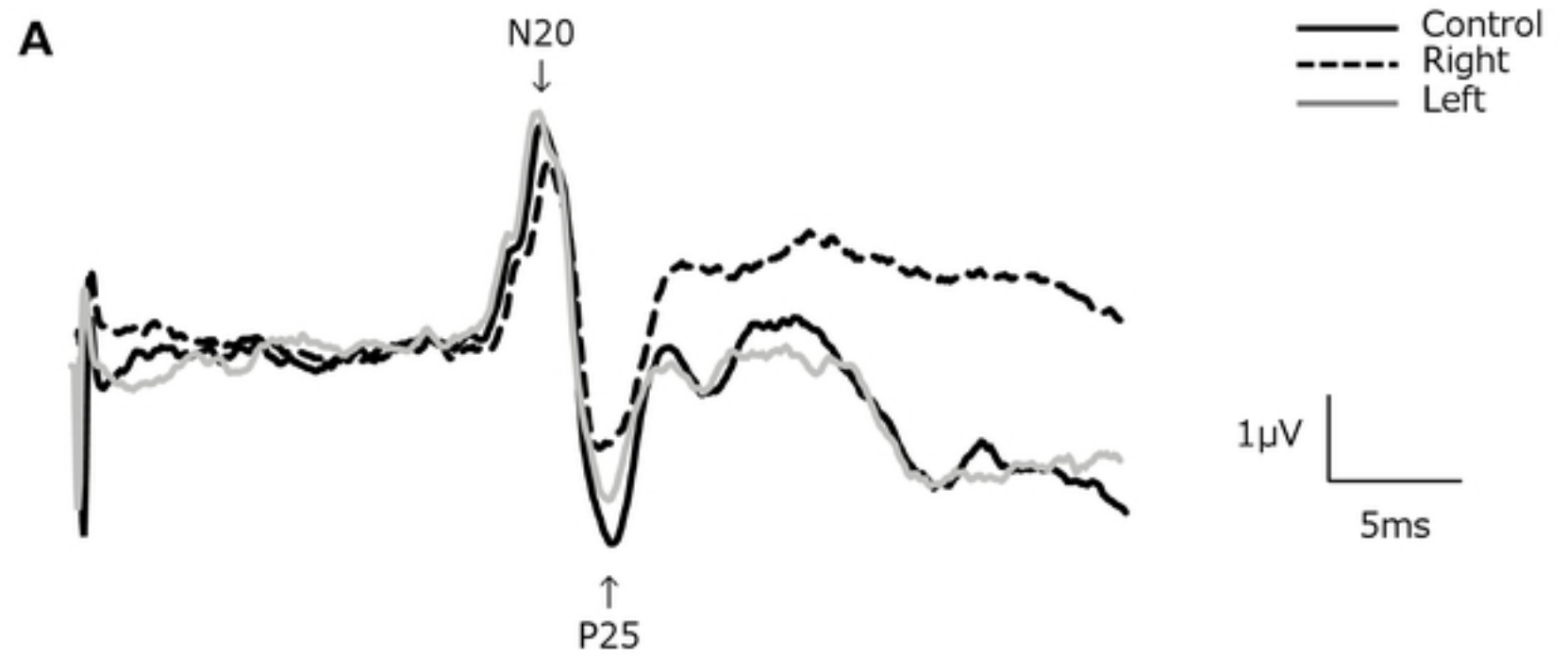


Figure2

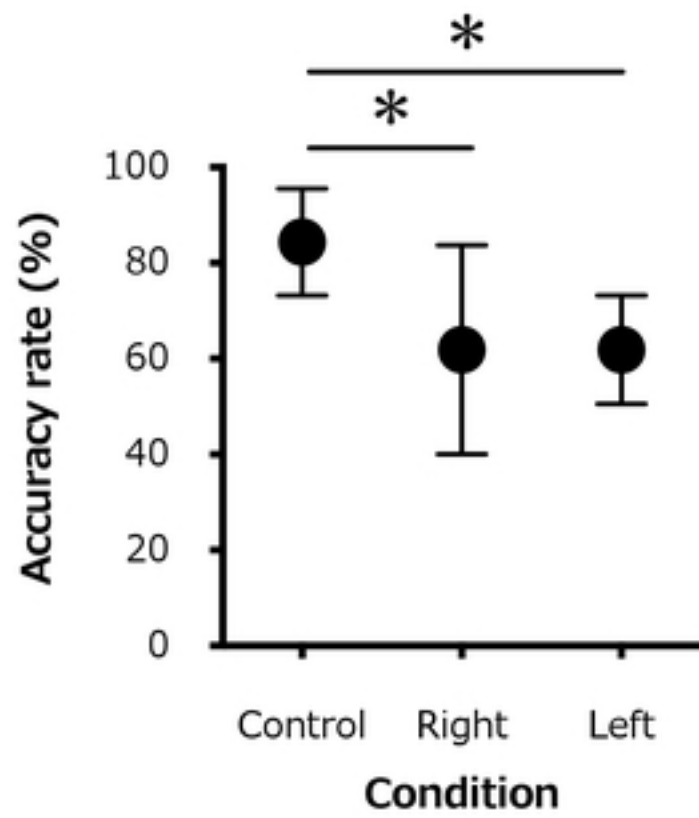


Figure3