Differential effects of soil conservation practices on arthropods and crop yield

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Abstract

1. Various agricultural practices aim to promote biodiversity and ecosystem services by increasing habitat availability and diversity. However, responses to such practices are context dependent, differentially impacting functional groups of service-providing organisms and crop yields.

2. Low and no tillage are promoted as sustainable management practices that increase soil heterogeneity and habitat. Here we (a) investigated whether soil tillage practices in canola crop fields altered arthropod biodiversity or yield, and (b) compared effects of field-scale management to landscape-scale habitat context. We focused on effects of full (multiple tillage passes that leave soil surface bare), intermediate (tilled once and some stubble remains), or no (seed planted directly into last year's stubble) tillage on five functional groups with unique diets and reproductive strategies: (i) herbivores, (ii) kleptoparasites, (iii) parasitoids, (iv) pollinators, and (v) predators.

3. Effects of habitat context on arthropod abundance and diversity varied across functional groups. Pollinators responded to on-farm soil management, benefiting from intermediate tillage. Predators and herbivores responded most strongly to landscape-scale habitat composition and were more abundant in landscapes with more semi-natural habitat. Our results indicate that natural history differences among arthropod functional groups mediate how habitat availability affects biodiversity. However, variation in arthropod communities had little effect on canola crop yield, which was lowest in fields with no tillage.

4. *Policy implications:* Our research highlights that soil management impacts on aboveground arthropods are complex, and that practices thought to increase agricultural sustainability do not necessarily achieve this goal. While reducing tillage intensity may increase soil quality

and habitat availability, our results show that tillage can negatively impact arthropods and

crop yields. This highlights the increasingly-documented disconnect between biodiversity

conservation and enhancing crop yield. Truly sustainable agriculture requires finding and

implementing practices that benefit both biodiversity and human wellbeing.

Keywords: sustainable agriculture, biodiversity, functional groups, crop yield, canola, tillage,

pollinators, natural enemies, soil habitat

Introduction

Enhancing crop yield while supporting biodiversity and ecosystem services is a challenge

in agroecosystems. Management practices like intercropping and reduced tillage (here referred to

as sustainable agriculture) aim to support biodiversity and ecological services by increasing

habitat availability and diversity, although responses to such practices vary among unique

service-providing functional groups (Lefcheck et al., 2015; Mitchell et al., 2015). Indeed, there

has been a call to determine simultaneous impacts of habitat change on multiple ecosystem

services such as soil quality, pollination, and pest control (Bommarco et al., 2013; Tamburini et

al., 2020). These impacts cannot necessarily be extrapolated from studies of single services due

to interactions among organisms. It is thus key to determine how organisms in unique functional

groups, that provide separate ecosystem services, differentially respond to sustainable agriculture

practices.

Reducing tillage is a practice that can support biodiversity by increasing soil habitat

availability and heterogeneity (de Graaff et al., 2019; Tamburini et al., 2020). Reduced tillage

was originally implemented to limit soil erosion and conserve soil moisture. It can also improve

in-field habitat through reduced soil disturbance frequency, improved soil physical and chemical

profiles (Kennedy & Schillinger, 2006), and increased prevalence of flowering plants that feed

beneficial insects (e.g., He et al., 2021; Rowen et al., 2020). However, effects of soil

management on ecosystem service providers likely depend on how animal functional groups

interact with soil. For example, tillage harms ground-nesting bees (Ullmann et al., 2016) and

predators that shelter among weeds (Cranshaw, 2004), but often has little impact on herbivorous

pests (Tooker et al., 2020). Impacts of soil management on distinct functional groups may also

depend on landscape context. Semi-natural habitat near farms can facilitate dispersal via habitat

corridors or provide patches of suitable habitat amidst less hospitable matrixes (Kremen et al.,

2007; Tscharntke et al., 2012). However, prior studies find that organisms can benefit from field-

level sustainable agriculture practices mainly in relatively simple (Scheper et al., 2013;

Tscharntke et al., 2012) or complex (Kennedy et al., 2013; Lichtenberg et al., 2017) landscapes.

These complexities show that there is an urgent need to study effects of sustainable agriculture

practices, including soil management, on multiple functional groups at various scales.

Sustainable farming must also generate high crop yields. Because yield captures the total

contribution of biotic communities and farming practices, it can be difficult to ascribe yield

changes to single factors (Tamburini et al., 2019). Studies that use a single analytical framework

to measure habitat availability and diversification impacts on biodiversity enable assessing how

multiple elements of the biotic system, and farming practices, directly and indirectly alter crop

production (Birkhofer et al., 2015; Byrnes et al., 2014; Weekers et al., 2022).

Here, we assessed how soil management through reduced tillage, and landscape context,

affected arthropods in unique functional groups, and how habitat and arthropod communities

affected canola yield. Canola provides floral resources for pollinators and natural enemies, but

attracts pests like aphids and flea beetles (Delaplane & Mayer, 2000; Morandin & Winston,

2005; Reddy, 2017). Yield may be altered by interactions among such beneficial and harmful

arthropods (Lundin et al., 2020; Morandin & Winston, 2005; Ouvrard & Jacquemart, 2019;

Reddy, 2017). Specifically, we asked:

(1) Do effects of tillage on abundance and diversity of arthropods vary based on functional

group? While negative impacts of tillage on soil-dwelling animals are well documented (de

Graaff et al., 2019; Rowen et al., 2020), tillage impacts on insects that visit leaves, stems, and

flowers remain poorly studied. However, we can reasonably hypothesize that functional groups

with higher dependence on soil are more strongly impacted by high-intensity tillage (Rowen et

al., 2020). Thus, we predicted that pollinators (mainly ground-nesting bees in our study region)

and the kleptoparasites that depend on them would be harmed by tillage (as in Ullmann et al.,

2016). Conversely the directly of tillage impacts on herbivores, parasitoids, and predators varies

widely (e.g., Jasrotia et al., 2021; Rowen et al., 2020; Serée et al., 2021; Wichura et al., 2022).

(2) Which habitat scales do various functional groups respond most strongly to? Many

vegetation-visiting arthropods are highly mobile, and thus likely to be influenced by both field-

scale management and the surrounding landscape (Lichtenberg et al., 2017; Marja et al., 2022).

The relative importance of local and landscape habitat can vary by functional group (e.g.,

Lichtenberg et al., 2017; Sánchez et al., 2022), although multiple mechanisms likely drive these

differences (Sirami et al., 2019), and results published to date are not consistent enough to

predict differential effects for our study system.

(3) How do arthropods and habitat context affect crop yield? Reduced tillage often decreases

yield, including for oilseed crops like canola (e.g., Lundin, 2019; Tamburini et al., 2020). This

could potentially be counteracted if reduced tillage enhances pest control and pollination by

promoting abundance or biodiversity of predators and pollinators.

Materials and methods

Study sites

We sampled arthropod abundance and diversity in canola (*Brassica napus* and *B. rapa*)

fields in eastern Washington and northern Idaho during the summers of 2013 and 2014 (Fig. S1;

Table S1). This heavily agricultural region has only scattered patches of semi-natural habitat.

The primary crops grown are grains and legumes, but canola production has increased from 5K

to over 75K acres in the last decade because it promotes soil quality and grower profits (Painter

et al., 2006; USDA NASS, 2020). As a mass-flowering crop that blooms for up to a month,

canola attracts diverse flower-feeding arthropods such as pollinators and natural enemies like

predators and parasitoids (Delaplane & Mayer, 2000; Morandin & Winston, 2005). Canola fields

are thus an effective model system for studying multiple ecosystem service providers.

We selected 15 canola fields each year with sizes from 0.7 to 142 ha (mean $\pm$ SD = 44.4 $\pm$

41.0). Most fields were maintained by local farmers, but four were on research farms maintained

by local universities or seed companies. All seed was treated with a neonicotinoid (mainly

thiamethoxam) prior to planting, and some farmers applied a pyrethroid insecticide one time at

the end of or after bloom as needed to keep flea beetles and cabbage seed pod weevils below the

economic threshold. Most farmers also treated their fields with an herbicide (mainly glyphosate)

early in the year, well before bloom. Because farmers rotate canola with other crops, no field was

sampled both years. Fields were sufficiently far apart (nearest neighbor distance of 5.4 $\pm$ 2.9

(min 1.9) km in 2013 and 6.8 $\pm$ 6.0 (min 2.8) km in 2014), relative to insect flight distances and

the scale at which we measured the landscape (1 km radius), to be considered spatially

independent.

Arthropod sampling

To target diverse arthropods that associate with canola flowers, we used two collection

techniques: (i) traps typically used to sample bees and (ii) sweep netting. All sampling occurred

along one field edge on days where daytime temperatures were above 13 ˚C and the wind speed

below 4.5 m s-1. Sampling locations were haphazardly selected along accessible field edges,

where we could work without damaging crops and where traps on the ground would not be

covered in vegetation. At 10:00, we conducted 100 spatially continuous sweeps at the level of

the canola canopy, immediately emptied net contents into a plastic bag, and stored them on ice.

In the lab, we freeze-killed arthropods, then removed them and sorted to morphospecies (see

below for sorting details). Bees were pinned, while specimens in other taxa were stored in

ethanol.

Around noon at each site, we set out a line of bee traps that stayed in place for 24 h. This

line included two blue vane traps (SpringStar) and six pan traps painted in colors that attract bees

(Kearns & Inouye, 1993; Leong & Thorp, 1999). We used two each of white, fluorescent yellow,

and fluorescent blue pan traps, randomizing color order within the line. Each trap contained

unscented soapy water. We separated traps by 5 m, and located them ~0.5 m from the edge of the

field. Traps and sweep netting began at the same spot. The following day, we collected trapped

arthropods using a strainer, washed off excess soap, and stored the specimens in ethanol. In the

lab, we washed and dried bees, and all specimens were sorted to genus and morphospecies based

on morphological characteristics (Arnett et al., 2002; Arnett, 2000; Arnett & Thomas, 2000;

Boyle & Philogène, 1983; Derraik et al., 2002, 2010; Michener, 2000; Stehr, 1987a, 1987b).

Both adults and immature forms were considered.

To aid interpretation, specimens were assigned to functional groups based on literature and

information from local species (Arnett, 2000; Michener, 2000; Stehr, 1987a, 1987b). We found

five common functional groups: (i) pollinators, (ii) herbivores, (iii) predators, (iv) parasitoids,

and (v) kleptoparasites (Table S2). This classification considers both diet and reproductive

strategy. Herbivores included major regional canola pests (thrips, aphids, flea beetles, cabbage

seedpod weevils, and *Lygus* bugs; Reddy, 2017). Kleptoparasites are important regulators of bee

communities, and may be valuable indicators of pollinator biodiversity (Sheffield et al., 2013).

We also considered predators and parasitoids pooled together as "natural enemies".

Field and landscape variables

We assessed several agronomic and weather-related variables for each field: (i) field size,

(ii) tillage, (iii) growing degree days, and (iv) cumulative precipitation. We determined field size

in ArcGIS by creating a polygon that traced each field's boundary, and subtracting the areas of

all uncultivated patches. These patches were typically remnant prairie vegetation, and occurred

both within and adjacent to fields. We determined these uncultivated patches by walking through

fields and drawing patches on a map, ensuring that we covered sufficient area to see the entire

field. We later converted these maps to ArcGIS polygons. To categorize each site's tillage

regimen, we asked farmers about their practices. We categorized each field as having (i) full (4

to 7 passes that produce bare soil with no remaining stubble before seeding; 6 fields), (ii)

intermediate (field tilled once and then planted into remaining stubble; 13 fields), or (iii) no (seed

planted directly into the previous year's stubble; 11 fields) tillage. These represent the three main

tillage regimes used in the study region. Because canola is planted very shallow, spring tillage in

the region is recommended to be limited to 2 to 5 cm (Brown et al., 2009).

We incorporated temperature and precipitation variables into analyses because our sites

spanned a mild precipitation and temperature gradient (PRISM Climate Group, 2020), and

because weather can influence insect biodiversity (e.g., Forcella et al., 2021; Skellern et al.,

2017). To measure weather variables, we gathered daily temperature and precipitation estimates

from PRISM (PRISM Climate Group, 2020). We calculated growing degree days and cumulative

precipitation through the day before we sampled arthropods at a field. Growing degree days

indicate accumulated heat units that are likely to affect growth of a given taxonomic group.

Temperatures below 5 ˚C can hinder canola growth (Dickson, 2014). We thus calculated

growing degree days as the cumulative amount of heat above 5 ˚C since January 1.

We calculated landscape composition as the amount of semi-natural habitat within a 1 km

radius of each field. This radius is ecologically relevant for pollinators, herbivores, and natural

enemies (Greenleaf et al., 2007; Rusch et al., 2016). We determined land cover using CropScape

data (USDA SARS, 2014a, 2014b, 2015a, 2015b) and ArcGIS to calculate the areas of each land

cover type around each site. Semi-natural habitat included deciduous forest, evergreen forest,

grassland, shrubland, and wetlands, while crop habitats were classified as agricultural.

Landscape composition around our sites ranged from 0% to 36% semi-natural habitat, and was

independent of tillage regime (Kruskal-Wallis test: $X^2_2 = 2.47$, $P = 0.29$).

Data analysis

To assess how each functional group responded to local- and landscape-scale habitat, we

used a regression and model selection framework. We first calculated three common community

metrics – abundance, richness, and evenness (E_{var}, Smith & Wilson, 1996) – for each functional

group at each site as our response variables. Evenness describes how sampled individuals are

distributed across taxa, and can provide valuable insights into the degree that rare taxa are

contributing to taxonomic richness and to ecosystem service functioning (Crowder et al., 2010;

Winfree et al., 2015). We ran separate linear regressions for each functional group that included

tillage, the proportion of semi-natural habitat within 1 km, their interaction, year, degree days

through the day before sampling, cumulative precipitation through the day before sampling, and

field size as fixed effects. These variables were not collinear (Table S4). It was necessary to

control for year, weather, and field size because all are known to affect insect abundance and

biodiversity (e.g., Aldercotte et al., In press; Forcella et al., 2021; Fragoso et al., 2021; Skellern

et al., 2017; Smith et al., 2020). Because abundance data were overdispersed, we used negative

binomial regressions (MASS package, Venables & Ripley, 2002). We analyzed richness and

evenness data with linear models (identity link), although herbivore evenness had to be log-

transformed due to heteroscedasticity. We then used information-theoretic model selection to

determine the best fit models for each metric and functional group (MuMIn package, Barton,

2014), selecting models with AICc values within 2 of the smallest value (Burnham & Anderson,

1998). Low richness values, and thus lack of variation, prevented us from analyzing

kleptoparasite and parasitoid evenness, but all other metrics were assessed. This resulted in 13

models (one for each functional group and abundance/diversity metric), each with 30

observations (28 observations for predator evenness).

We next assessed how arthropod biodiversity, as well as local- and landscape-level habitat

factors, affected canola yields. We ran separate linear regressions for each community metric

(abundance, richness, evenness; three models total) that included tillage, the proportion of semi-

natural habitat within 1 km, their interaction, year, and abundance or diversity of each arthropod

functional group. All equations used linear regression (identity link), and met assumptions of this

modeling approach. As with assessment of how functional groups responded to habitat context,

we selected models with AICc values within 2 of the smallest value.

Ethics

This study did not require ethical approval.

Results

Overall, we collected 22,879 individuals across 131 taxa. This included 15,256 herbivores (20 taxa, 82% of individuals were pest species), 154 kleptoparasites (8 taxa), 1,080 parasitoids (6 taxa), 4,217 pollinators (64 taxa), and 739 predators (31 taxa). The most abundant taxonomic groups were thrips (7,184 individuals; herbivores, major canola pests), aphids (4,078; herbivores, major canola pests), sciaroid flies (2,041 individuals; herbivores), two halictid bee morphotaxa (813 and 581 individuals; pollinators), chalcidoid wasps (727 individuals; parasitoids), and chrysomelid beetles (576 individuals; herbivores, major canola pests). The most common predators were melyrid beetles (229 individuals).

Effects of tillage and landscape on arthropod communities

Effects of habitat context on arthropod abundance and diversity varied across functional groups. Broadly, pollinators and kleptoparasites were affected by tillage while herbivores and predators responded most strongly to landscape composition. Tillage and landscape composition never had an interactive effect. This pattern did not reflect a correlation between field- and landscape-scale habitat measures, as semi-natural habitat did not differ based on tillage regimen (Kruskal-Wallis test: $X^2_2 = 2.47$, $P = 0.29$). Overall, tillage and landscapes impacted arthropod abundance and richness more than evenness.

Pollinator richness was higher in fields with intermediate tillage than fields with no tillage (Fig. 1a; Tables S5, S6). Kleptoparasite abundance was higher in fields with either form of conservation tillage (intermediate or no tillage) than in heavily-tilled fields in one of four best models (and significant at $\alpha < 0.10$ in a second model; Fig. 1b; Tables S6, S7). Pollinator abundance and evenness, and kleptoparasite richness, were not affected by local or landscape

habitat context (Tables S5, S7, S8). There were too few kleptoparasite species (median 2 per site)

to assess evenness.

Landscapes with more semi-natural habitat promoted greater herbivore (Fig. 2a) and

predator (Fig. 2b) abundance regardless of tillage regime (Table S7). Herbivore diversity and

predator richness were unaffected by either tillage or landscape composition (Tables S5, S8).

These results did not qualitatively change if the site in the landscape with the most semi-natural

habitat was removed (Table S7). Parasitoids were unaffected by habitat context at either local or

landscape scale (Tables S5, S7, S8).

Environmental variables had mixed effects on arthropod abundance and diversity (Tables

S5, S7, S8). As expected, all functional groups except predators showed inter-annual variation in

abundance, richness, or evenness, typically with lower abundance and richness, but higher

evenness, in 2013 than 2014. Cumulative precipitation had stronger impacts than degree days at

time of insect sampling. Detritivore abundance, kleptoparasite abundance and richness,

pollinator abundance, and predator richness were higher at sites with less precipitation, while

parasitoid abundance and pollinator evenness showed the opposite pattern. Detritivore

abundance, kleptoparasite richness, parasitoid abundance, and predator richness were higher at

cooler sites. Field size affected only a few metrics. Kleptoparasites were more abundant and

speciose, and pollinator communities were less even, at larger fields.

Differential responses of arthropod functional groups could potentially indicate trade-offs

when aiming to manage for multiple functional groups. However, our results show stronger

evidence of synergies than trade-offs. Functional groups with known trophic relationships had

correlated metrics (Table S3). We found positive correlations in abundance and diversity of (i)

pollinators and kleptoparasites, and (ii) parasitoids, predators, and herbivores (and between

natural enemies and herbivores). Additionally, abundance and evenness of pollinators versus

predators and parasitoids (and of pollinators and natural enemies as a group) were positively

correlated (Table S3).

Effects of tillage, landscape and arthropod communities on crop yields

Field-scale habitat management directly impacted canola yield. In all analyses, yield was

lower in fields with no tillage than in fields with full or intermediate tillage (Fig. 3a, Tables S6,

S9). Landscape composition did not impact canola yield, however.

We found minimal evidence for effects of habitat context on crop yield, via arthropod

communities (Tables S6, S9; variance inflation factors in Table S10). Higher landscape-scale

habitat availability did promote more diverse herbivore communities, which lowered crop yields

(Fig. 3b; Tables S6, S9). However, arthropod abundance and evenness, and richness of other

functional groups, did not impact yield.

Discussion

Soil management can affect arthropod biodiversity, with impacts varying by functional

group or landscape context, due to differences in resource needs or dispersal among taxa

(Bommarco et al., 2013; Harmon-Threatt, 2020). Because many of these arthropods provide

ecosystem services or are pests, biodiversity impacts are predicted to affect crop yield. However,

impacts of soil-management on above-ground arthropods are poorly studied. We found that

agricultural landscapes can simultaneously support pollinators and predators, but that different

functional groups respond to habitat context at separate scales. Pollinators were affected by field-

scale availability and diversity via reduced tillage, while landscapes with more semi-natural

habitat promoted herbivores and predators. Although habitat context affected arthropod

biodiversity, arthropods did not strongly impact crop yield. Our results show that sustainable

agriculture practices can impact crop yield directly, but not always indirectly by promoting

higher arthropod biodiversity (as in Ricketts et al., 2016, but contrary to Dainese et al., 2019).

We found that pollinator and kleptoparasite, but not herbivore, predator, or parasitoid

guilds responded to tillage intensity. We expected reduced tillage would promote pollinators, and

kleptoparasites that depend on them, by destroying fewer ground-nesting bee nests and

improving soil quality (Kennedy & Schillinger, 2006; Ullmann et al., 2016). Intriguingly, we

found that intermediate tillage supported a more diverse pollinator community than no tillage.

One potential explanation is that untilled soil contains too thick a layer of decaying crop stems

for bees to dig through (Stinner & House, 1990). However, tillage impacts, and presence of

kleptoparasites that are typically found in the vicinity of bee nests, do suggest that bees are

nesting in canola crop fields or nearby grain or legume fields, which is often assumed to not

occur (Kleijn et al., 2011).

Effects of tillage on natural enemies and herbivores are more well studied than for bees,

although results have been mixed for each group (Furlan et al., 2021; Rowen et al., 2020; Tooker

et al., 2020). When reduced tillage promotes natural enemies, likely mechanisms are increased

availability of weeds that provide nectar, or surface mulch that provides shelter and prey (Clark

et al., 1993; Stinner & House, 1990). The first mechanism is likely not operating in our system

because canola provides abundant nectar and weeds were controlled with glyphosate. The second

mechanism may also not be relevant because crops rotated with canola do not contribute much

mulch (Hammel, 1996). When reduced tillage promotes herbivores, it mainly does so due to less

soil disturbance or by promoting larger or more diverse weed communities (Rowen et al., 2020).

Neither mechanism applies to our study, since we mainly sampled herbivores that reside near the

tops of plants rather than in soil, and weeds were controlled. Thus, it is not surprising that tillage

intensity did not affect natural enemies or herbivores. This highlights the importance of studying

mechanisms underlying habitat-biodiversity relationships, including how species' natural

histories relate to both food and shelter resources within a habitat (Carvalheiro et al., 2021).

We found that landscapes with more semi-natural habitat promoted predators and

herbivores. Heterogeneous landscapes offer opportunities for mobile consumers to exploit patchy

resources (Tscharntke et al., 2012). Predators and our main herbivores (aphids and thrips)

routinely travel long distances in search of suitable habitat patches (Loxdale & Lushai, 1999;

Schellhorn et al., 2014). In contrast, most of our pollinators (bees) are central place foragers that

repeatedly return to a single nest and visit food patches only within their flight range of this nest.

Indeed, bees visiting canola flowers in Alberta tended to travel only a few meters from their nest

when surrounded by canola flowers (Robinson, 2019). Thus, most of the bees we collected likely

were nesting in or near the fields we sampled, and were affected more by tillage than landscape

composition. This result mirrors meta-analyses that suggest that highly mobile organisms are

more likely to respond to landscape-scale habitat patterns than less mobile organisms

(Lichtenberg et al., 2017; Schneider et al., 2014).

While we show local and landscape habitat patterns affected arthropod communities, these

communities minimally impacted crop yields, similar to studies that have found no relationship

between multi-diversity and multifunctionality (Birkhofer et al., 2018). We did find that yield

was lower in fields with higher herbivore richness. Inspection of herbivore abundances at each

site (Table S11) suggests two potential drivers. First, sites with higher herbivore richness could

have higher pest abundance (Table S3). Second, sites with higher herbivore richness could be

more likely to contain a specific damaging pest. Our data shows high-herbivore-richness fields

contained large numbers of aphids, a key canola pest (Reddy, 2017). We also found more

chrysomelids, curculionids, meloids, scraptiids, pentatomids, and yponomeutids in sites with

higher herbivore richness. However, the only canola pests in these groups are seedpod weevils

(Curculionidae), which damage a later crop stage than we sampled (Reddy, 2017).

Abundance and diversity of "beneficial" pollinators and predators also did not affect canola

yield. Canola has high variability in pollinator dependence (Ouvrard & Jacquemart, 2019), and

the ≥ 15 varieties in our study may not be pollinator dependent (as in Perrot et al., 2018), or the

study region is windy enough to ensure high pollen dispersal. Variation in pollinator dependence

may explain the difference between our results and other studies that find increased yield of

oilseed crops with higher pollinator abundance (Catarino et al., 2019). Benefits from pollinators

and predators may have also been limited by insecticide use, as farmers grow insecticide-coated

seeds that provide plants with systemic insecticide coverage. This may effectively control

herbivore pests, or reduce pollinator abundance (Catarino et al., 2019). It is also possible that

pollinators or predators correlate with common measures of single ecosystem services, such as

pollen deposition or consumption of sentinel pests on a small subset of plants. If such patterns

were present, they did not scale up to the entire field (as found by Birkhofer et al., 2018; Sutter et

al., 2018; Tamburini et al., 2020), and thus are unlikely to affect farmer decisions.

Tillage intensity did affect crop yields. Reduced tillage has previously been found to reduce

yield for crops including oilseed rape (Lundin, 2019; Tamburini et al., 2020). Indeed, multiple

sustainability-oriented farming practices (e.g., organic farming) sometimes result in lower yield

than their conventional counterparts (Smith et al., 2020; Tamburini et al., 2020). Despite the

yield loss seen here, reduced tillage can provide other benefits such as improving soil infiltration,

reducing erosion, decreasing evaporative water loss from soil, and improving soil quality

(Hammel, 1996; Kennedy & Schillinger, 2006). These factors might ultimately increase yield of

other crops or reduce farmers' costs. This highlights the complex decisions that underlie farm

management decisions.

Overall, our study highlights the need to understand how biodiversity patterns and crop

yields are simultaneously affected by multiple mechanisms, including via soil management, at

various scales. We showed that field-scale soil habitat management impacted pollinators, while

landscape-scale habitat availability promoted predators and herbivores. These differences likely

reflect natural history differences among these functional groups. However, these habitat impacts

on biodiversity only minimally impacted yield. It is often assumed that enhancing biodiversity

promotes ecosystem services, although evidence from arthropod-mediated ecosystem services

such as pollination and pest control is mixed (e.g., Birkhofer et al., 2018; Dainese et al., 2019;

Ricketts et al., 2016). Without clear evidence that a conservation action such as reduced tillage is

likely to increase crop yield, adoption by farmers will likely remain low (Kleijn et al., 2019).

Thus, data-driven management of agricultural landscapes to simultaneously support natural

biodiversity and boost crop yield requires much more research to determine the contexts in

which given management practices, and soil diversification practices in particular, do or do not

meet this multi-faceted goal.

Author contributions

EML and DWC conceived the ideas and designed methodology; EML collected and analyzed the

data; IM identified arthropods; EML and AJC led the writing of the manuscript; All authors

contributed critically to the drafts and gave final approval for publication.

Statement on inclusion

Our study design and results interpretation incorporated local knowledge from stakeholders, and

results were communicated with stakeholders through outreach.

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References

Aldercotte, A. H., Simpson, D. T., & Winfree, R. (In press). Crop visitation by wild bees

declines over an 8-year time series: A dramatic trend, or just dramatic between-year

variation? *Insect Conservation and Diversity*. https://doi.org/10.1111/icad.12589

Arnett, Ross H., Jr., Thomas, M. C., Skelley, P. E., & Frank, J. H. (2002). *American Beetles,*

Volume II: Polyphaga: Scarabaeoidea through Curculionoidea. CRC Press.

Arnett, R. H., Jr. (2000). *American Insects: A Handbook of the Insects of America North of*

Mexico, Second Edition. CRC Press.

Arnett, R. H., Jr, & Thomas, M. C. (2000). *American Beetles: Archostemata, Myxophaga,*

Adephaga, Polyphaga: Staphyliniformia. CRC Press.

Barton, K. (2014). *MuMIn: Multi-model inference.* (1.10.5). http://CRAN.R-

project.org/package=MuMIn

Birkhofer, K., Andersson, G. K. S., Bengtsson, J., Bommarco, R., Dänhardt, J., Ekbom, B.,

Ekroos, J., Hahn, T., Hedlund, K., Jönsson, A. M., Lindborg, R., Olsson, O., Rader, R.,

Rusch, A., Stjernman, M., Williams, A., & Smith, H. G. (2018). Relationships between

multiple biodiversity components and ecosystem services along a landscape complexity

gradient. *Biological Conservation*, *218*, 247–253.

https://doi.org/10.1016/j.biocon.2017.12.027

Birkhofer, K., Diehl, E., Andersson, J., Ekroos, J., Früh-Müller, A., Machnikowski, F., Mader,

V. L., Nilsson, L., Sasaki, K., Rundlöf, M., Wolters, V., & Smith, H. G. (2015).

Ecosystem services—Current challenges and opportunities for ecological research.

Frontiers in Ecology and Evolution, *2*, 97. https://doi.org/10.3389/fevo.2014.00087

Bommarco, R., Kleijn, D., & Potts, S. G. (2013). Ecological intensification: Harnessing

ecosystem services for food security. *Trends in Ecology & Evolution*, *28*(4), 230–238.

https://doi.org/10.1016/j.tree.2012.10.012

Boyle, R. M. D., & Philogène, B. J. R. (1983). The native pollinators of an apple orchard:

Variations and significance. *The Journal of Horticultural Science*, *58*(3), 355–363.

https://doi.org/10.1080/00221589.1983.11515130

Brown, J., Davis, J. B., Lauver, M., & Wysocki, D. (2009). *United States Canola Association*

Canola Growers Manual. United States Canola Association.

https://www.webpages.uidaho.edu/jbrown/brassica/Canola%20Growers%20Manual%20

Final-3-14-09.pdf

Burnham, K. P., & Anderson, D. R. (1998). *Model Selection and Multimodel Inference: A*

Practical Information-Theoretic Approach (2nd ed.). Springer-Verlag.

Byrnes, J. E. K., Gamfeldt, L., Isbell, F., Lefcheck, J. S., Griffin, J. N., Hector, A., Cardinale, B.

J., Hooper, D. U., Dee, L. E., & Duffy, J. E. (2014). Investigating the relationship

between biodiversity and ecosystem multifunctionality: Challenges and solutions.

Methods in Ecology and Evolution, *5*(2), 111–124. https://doi.org/10.1111/2041-

210X.12143

Carvalheiro, L. G., Bartomeus, I., Rollin, O., Timóteo, S., & Tinoco, C. F. (2021). The role of

soils on pollination and seed dispersal. *Philosophical Transactions of the Royal Society*

B: Biological Sciences, *376*(1834), 20200171. https://doi.org/10.1098/rstb.2020.0171

Catarino, R., Bretagnolle, V., Perrot, T., Vialloux, F., & Gaba, S. (2019). Bee pollination

outperforms pesticides for oilseed crop production and profitability. *Proceedings of the*

Royal Society B: Biological Sciences, *286*(1912), 20191550.

https://doi.org/10.1098/rspb.2019.1550

Clark, M. S., Luna, J. M., Stone, N. D., & Youngman, R. R. (1993). Habitat preferences of

generalist predators in reduced-tillage corn. *Journal of Entomological Science*, *28*(4),

404–416. https://doi.org/10.18474/0749-8004-28.4.404

Cranshaw, W. (2004). *Garden Insects of North America*. Princeton University Press.

Crowder, D. W., Northfield, T. D., Strand, M. R., & Snyder, W. E. (2010). Organic agriculture

promotes evenness and natural pest control. *Nature*, *466*(7302), 109–112.

https://doi.org/10.1038/nature09183

Dainese, M., Martin, E. A., Aizen, M. A., Albrecht, M., Bartomeus, I., Bommarco, R.,

Carvalheiro, L. G., Chaplin-Kramer, R., Gagic, V., Garibaldi, L. A., Ghazoul, J., Grab,

H., Jonsson, M., Karp, D. S., Kennedy, C. M., Kleijn, D., Kremen, C., Landis, D. A.,

Letourneau, D. K., … Steffan-Dewenter, I. (2019). A global synthesis reveals

biodiversity-mediated benefits for crop production. *Science Advances*, *5*(10), eaax0121.

https://doi.org/10.1126/sciadv.aax0121

de Graaff, M.-A., Hornslein, N., Throop, H. L., Kardol, P., & van Diepen, L. T. A. (2019).

Effects of agricultural intensification on soil biodiversity and implications for ecosystem

functioning: A meta-analysis. In D. L. Sparks (Ed.), *Advances in Agronomy* (Vol. 155,

pp. 1–44). Academic Press. https://doi.org/10.1016/bs.agron.2019.01.001

Delaplane, K. S., & Mayer, D. F. (2000). *Crop Pollination by Bees*. CABI Publishing.

Derraik, J. G. B., Closs, G. P., Dickinson, K. J. M., Sirvid, P., Barratt, B. I. P., & Patrick, B. H.

(2002). Arthropod morphospecies versus taxonomic species: A case study with Araneae,

Coleoptera, and Lepidoptera. *Conservation Biology*, *16*(4), 1015–1023.

https://doi.org/10.1046/j.1523-1739.2002.00358.x

Derraik, J. G. B., Early, J. W., Closs, G. P., & Dickinson, K. J. M. (2010). Morphospecies and

taxonomic species comparison for Hymenoptera. *Journal of Insect Science*, *10*(108), 1–7.

https://doi.org/10.1673/031.010.10801

Dickson, T. J. (2014). *Growing Season Weather Impacts on Canola Phenological Development*

and Quality [Masters' thesis, University of Manitoba].

https://mspace.lib.umanitoba.ca/bitstream/handle/1993/23254/Dickson_Taryn.pdf

Forcella, F., Patel, S., Lenssen, A. W., Hoerning, C., Wells, M. S., Gesch, R. W., & Berti, M. T.

(2021). Weather and landscape influences on pollinator visitation of flowering winter

oilseeds (field pennycress and winter camelina). *Journal of Applied Entomology*, *145*(4),

286–294. https://doi.org/10.1111/jen.12854

Fragoso, F. P., Jiang, Q., Clayton, M. K., & Brunet, J. (2021). Patch selection by bumble bees

navigating discontinuous landscapes. *Scientific Reports*, *11*(1), 8986.

https://doi.org/10.1038/s41598-021-88394-2

Furlan, L., Milosavljević, I., Chiarini, F., & Benvegnù, I. (2021). Effects of conventional versus

no-tillage systems on the population dynamics of elaterid pests and the associated

damage at establishment of maize crops. *Crop Protection*, *149*, 105751.

https://doi.org/10.1016/j.cropro.2021.105751

Greenleaf, S. S., Williams, N. M., Winfree, R., & Kremen, C. (2007). Bee foraging ranges and

their relationship to body size. *Oecologia*, *153*, 589–596.

https://doi.org/10.1073/pnas.0600929103

Hammel, J. E. (1996). Water conservation practices for sustainable dryland farming systems in

the Pacific Northwest. *American Journal of Alternative Agriculture*, *11*(2/3), 58–63.

Harmon-Threatt, A. (2020). Influence of nesting characteristics on health of wild bee

communities. *Annual Review of Entomology*, *65*(1), 39–56.

https://doi.org/10.1146/annurev-ento-011019-024955

He, X., Kiær, L. P., Jensen, P. M., & Sigsgaard, L. (2021). The effect of floral resources on

predator longevity and fecundity: A systematic review and meta-analysis. *Biological*

Control, *153*, 104476. https://doi.org/10.1016/j.biocontrol.2020.104476

Jasrotia, P., Bhardwaj, A. K., Katare, S., Yadav, J., Kashyap, P. L., Kumar, S., & Singh, G. P.

(2021). Tillage intensity influences insect-pest and predator dynamics of wheat crop

grown under different conservation agriculture practices in rice-wheat cropping system of

Indo-Gangetic plain. *Agronomy*, *11*(6), 1087. https://doi.org/10.3390/agronomy11061087

Kearns, C. A., & Inouye, D. W. (1993). *Techniques for Pollination Biologists*. University Press

of Colorado.

Kennedy, A. C., & Schillinger, W. F. (2006). Soil quality and water intake in traditional-till vs.

No-till paired farms in Washington's Palouse region. *Soil Science Society of America*

Journal, *70*(3), 940–949. https://doi.org/10.2136/sssaj2005.0160

Kennedy, C. M., Lonsdorf, E., Neel, M. C., Williams, N. M., Ricketts, T. H., Winfree, R.,

Bommarco, R., Brittain, C., Burley, A. L., Cariveau, D., Carvalheiro, L. G., Chacoff, N.

P., Cunningham, S. A., Danforth, B. N., Dudenhöffer, J.-H., Elle, E., Gaines, H. R.,

Garibaldi, L. A., Gratton, C., … Kremen, C. (2013). A global quantitative synthesis of

local and landscape effects on wild bee pollinators in agroecosystems. *Ecology Letters*,

16(5), 584–599. https://doi.org/10.1111/ele.12082

Kleijn, D., Bommarco, R., Fijen, T. P. M., Garibaldi, L. A., Potts, S. G., & Putten, W. H. van

der. (2019). Ecological intensification: Bridging the gap between science and practice.

Trends in Ecology & Evolution, *34*(2), 154–166.

https://doi.org/10.1016/j.tree.2018.11.002

Kleijn, D., Rundlöf, M., Scheper, J., Smith, H. G., & Tscharntke, T. (2011). Does conservation

on farmland contribute to halting the biodiversity decline? *Trends in Ecology &*

Evolution, *26*(9), 474–481. https://doi.org/10.1016/j.tree.2011.05.009

Kremen, C., Williams, N. M., Aizen, M. A., Gemmill-Herren, B., LeBuhn, G., Packer, L., Potts,

S. G., Roulston, T., Steffan-Dewenter, I., Vásquez, D. P., Winfree, R., Adams, L., Crone,

E. E., Greenleaf, S. S., Keitt, T. H., Klein, A.-M., Regetz, J., & Ricketts, T. H. (2007).

Pollination and other ecosystem services produced by mobile organisms: A conceptual

framework for the effects of land-use change. *Ecology Letters*, *10*(4), 299–314.

https://doi.org/10.1111/j.1461-0248.2007.01018.x

Lefcheck, J. S., Byrnes, J. E. K., Isbell, F., Gamfeldt, L., Griffin, J. N., Eisenhauer, N., Hensel,

M. J. S., Hector, A., Cardinale, B. J., & Duffy, J. E. (2015). Biodiversity enhances

ecosystem multifunctionality across trophic levels and habitats. *Nature Communications*,

6(1), 1–7. https://doi.org/10.1038/ncomms7936

Leong, J. M., & Thorp, R. W. (1999). Colour-coded sampling: The pan trap colour preferences

of oligolectic and nonoligolectic bees associated with a vernal pool plant. *Ecological*

Entomology, *24*(3), 329–335. https://doi.org/10.1046/j.1365-2311.1999.00196.x

Lichtenberg, E. M., Kennedy, C. M., Kremen, C., Batáry, P., Berendse, F., Bommarco, R.,

Bosque-Pérez, N. A., Carvalheiro, L. G., Snyder, W. E., Williams, N. M., Winfree, R.,

Klatt, B. K., Åström, S., Benjamin, F., Brittain, C., Chaplin-Kramer, R., Clough, Y.,

Danforth, B., Diekötter, T., … Crowder, D. W. (2017). A global synthesis of the effects

of diversified farming systems on arthropod diversity within fields and across agricultural

landscapes. *Global Change Biology*, *23*(11), 4946–4957.

https://doi.org/10.1111/gcb.13714

Loxdale, H. D., & Lushai, G. (1999). Slaves of the environment: The movement of herbivorous

insects in relation to their ecology and genotype. *Philosophical Transactions of the Royal*

Society of London. Series B: Biological Sciences, *354*(1388), 1479–1495.

https://doi.org/10.1098/rstb.1999.0492

Lundin, O. (2019). No-till protects spring oilseed rape (*Brassica napus* L.) against crop damage

by flea beetles (*Phyllotreta* spp.). *Agriculture, Ecosystems & Environment*, *278*, 1–5.

https://doi.org/10.1016/j.agee.2019.03.014

Lundin, O., Malsher, G., Högfeldt, C., & Bommarco, R. (2020). Pest management and yield in

spring oilseed rape without neonicotinoid seed treatments. *Crop Protection*, *137*, 105261.

https://doi.org/10.1016/j.cropro.2020.105261

Marja, R., Tscharntke, T., & Batáry, P. (2022). Increasing landscape complexity enhances

species richness of farmland arthropods, agri-environment schemes also abundance – A

meta-analysis. *Agriculture, Ecosystems & Environment*, *326*, 107822.

https://doi.org/10.1016/j.agee.2021.107822

Michener, C. D. (2000). *The Bees of the World*. Johns Hopkins University Press.

Mitchell, M. G. E., Suarez-Castro, A. F., Martinez-Harms, M., Maron, M., McAlpine, C.,

Gaston, K. J., Johansen, K., & Rhodes, J. R. (2015). Reframing landscape

fragmentation's effects on ecosystem services. *Trends in Ecology & Evolution*, *30*(4),

190–198. https://doi.org/10.1016/j.tree.2015.01.011

Morandin, L. A., & Winston, M. L. (2005). Wild bee abundance and seed production in

conventional, organic, and genetically modified canola. *Ecological Applications*, *15*(3),

871–881. https://doi.org/10.1890/03-5271

Ouvrard, P., & Jacquemart, A.-L. (2019). Review of methods to investigate pollinator

dependency in oilseed rape (*Brassica napus*). *Field Crops Research*, *231*, 18–29.

https://doi.org/10.1016/j.fcr.2018.11.006

Painter, K., Hinman, H., & Roe, D. (2006). *Economics of spring canola production in dryland*

eastern Washington (No. EB2009E; p. 22). Washington State University.

http://wsldocs.sos.wa.gov/library/docs/wsu/extension/eb2009e_2008_004485.pdf

Perrot, T., Gaba, S., Roncoroni, M., Gautier, J.-L., & Bretagnolle, V. (2018). Bees increase

oilseed rape yield under real field conditions. *Agriculture, Ecosystems & Environment*,

266, 39–48. https://doi.org/10.1016/j.agee.2018.07.020

PRISM Climate Group. (2020). *Oregon State University*. http://prism.oregonstate.edu

Reddy, G. V. P. (2017). *Integrated Management of Insect Pests on Canola and Other Brassica*

Oilseed Crops. CABI.

Ricketts, T. H., Watson, K. B., Koh, I., Ellis, A. M., Nicholson, C. C., Posner, S., Richardson, L.

L., & Sonter, L. J. (2016). Disaggregating the evidence linking biodiversity and

ecosystem services. *Nature Communications*, *7*(1), 1–8.

https://doi.org/10.1038/ncomms13106

Robinson, S. V. J. (2019). *Central-place foraging, crop yield, and population change in bees: A*

study in canola agroecosystems [Doctoral thesis, University of Calgary]. PRISM.

http://hdl.handle.net/1880/111128

Rowen, E. K., Regan, K. H., Barbercheck, M. E., & Tooker, J. F. (2020). Is tillage beneficial or

detrimental for insect and slug management? A meta-analysis. *Agriculture, Ecosystems &*

Environment, *294*, 106849. https://doi.org/10.1016/j.agee.2020.106849

Rusch, A., Chaplin-Kramer, R., Gardiner, M. M., Hawro, V., Holland, J., Landis, D., Thies, C.,

Tscharntke, T., Weisser, W. W., Winqvist, C., Woltz, M., & Bommarco, R. (2016).

Agricultural landscape simplification reduces natural pest control: A quantitative

synthesis. *Agriculture, Ecosystems & Environment*, *221*, 198–204.

https://doi.org/10.1016/j.agee.2016.01.039

Sánchez, A. C., Jones, S. K., Purvis, A., Estrada-Carmona, N., & De Palma, A. (2022).

Landscape complexity and functional groups moderate the effect of diversified farming

on biodiversity: A global meta-analysis. *Agriculture, Ecosystems & Environment*, *332*,

107933. https://doi.org/10.1016/j.agee.2022.107933

Schellhorn, N. A., Bianchi, F. J. J. A., & Hsu, C. L. (2014). Movement of entomophagous

arthropods in agricultural landscapes: Links to pest suppression. *Annual Review of*

Entomology, *49*, 559–581. https://doi.org/10.1146/annurev-ento-011613-161952

Scheper, J., Holzschuh, A., Kuussaari, M., Potts, S. G., Rundlöf, M., Smith, H. G., & Kleijn, D.

(2013). Environmental factors driving the effectiveness of European agri-environmental

measures in mitigating pollinator loss – a meta-analysis. *Ecology Letters*, *16*(7), 912–920.

https://doi.org/10.1111/ele.12128

Schneider, M. K., Lüscher, G., Jeanneret, P., Arndorfer, M., Ammari, Y., Bailey, D., Balázs, K.,

Báldi, A., Choisis, J.-P., Dennis, P., Eiter, S., Fjellstad, W., Fraser, M. D., Frank, T.,

Friedel, J. K., Garchi, S., Geijzendorffer, I. R., Gomiero, T., Gonzalez-Bornay, G., …

Herzog, F. (2014). Gains to species diversity in organically farmed fields are not

propagated at the farm level. *Nature Communications*, *5*, 4151.

https://doi.org/10.1038/ncomms5151

Serée, L., Gardarin, A., Crouzet, O., Barbottin, A., Valantin-Morison, M., & Chiron, F. (2021).

Exploring multitrophic interactions in oilseed rape fields reveals the prevailing role of

Carabidae. *Ecology and Evolution*, *11*(21), 15377–15388.

https://doi.org/10.1002/ece3.8229

Sheffield, C. S., Pindar, A., Packer, L., & Kevan, P. G. (2013). The potential of cleptoparasitic

bees as indicator taxa for assessing bee communities. *Apidologie*, *44*(5), 501–510.

https://doi.org/10.1007/s13592-013-0200-2

Sirami, C., Gross, N., Baillod, A. B., Bertrand, C., Carrié, R., Hass, A., Henckel, L., Miguet, P.,

Vuillot, C., Alignier, A., Girard, J., Batáry, P., Clough, Y., Violle, C., Giralt, D., Bota,

G., Badenhausser, I., Lefebvre, G., Gauffre, B., … Fahrig, L. (2019). Increasing crop

heterogeneity enhances multitrophic diversity across agricultural regions. *Proceedings of*

the National Academy of Sciences, *116*(33), 16442–16447.

https://doi.org/10.1073/pnas.1906419116

Skellern, M. P., Welham, S. J., Watts, N. P., & Cook, S. M. (2017). Meteorological and

landscape influences on pollen beetle immigration into oilseed rape crops. *Agriculture,*

Ecosystems & Environment, *241*, 150–159. https://doi.org/10.1016/j.agee.2017.03.008

Smith, B., & Wilson, J. B. (1996). A consumer's guide to evenness indices. *Oikos*, *76*(1), 70–82.

https://doi.org/10.2307/3545749

Smith, O. M., Cohen, A. L., Reganold, J. P., Jones, M. S., Orpet, R. J., Taylor, J. M., Thurman, J.

H., Cornell, K. A., Olsson, R. L., Ge, Y., Kennedy, C. M., & Crowder, D. W. (2020).

Landscape context affects the sustainability of organic farming systems. *Proceedings of*

the National Academy of Sciences, *117*(6), 2870–2878.

https://doi.org/10.1073/pnas.1906909117

Stehr, F. W. (1987a). *Immature Insects* (Vol. 1). Kendall/Hunt Publishing Company.

Stehr, F. W. (1987b). *Immature Insects* (Vol. 2). Kendall/Hunt Publishing Company.

Stinner, B. R., & House, G. J. (1990). Arthropods and other invertebrates in conservation-tillage

agriculture. *Annual Review of Entomology*, *35*(1), 299–318.

https://doi.org/10.1146/annurev.en.35.010190.001503

Sutter, L., Albrecht, M., & Jeanneret, P. (2018). Landscape greening and local creation of

wildflower strips and hedgerows promote multiple ecosystem services. *Journal of*

Applied Ecology, *55*(2), 612–620. https://doi.org/10.1111/1365-2664.12977

Tamburini, G., Bommarco, R., Kleijn, D., van der Putten, W. H., & Marini, L. (2019).

Pollination contribution to crop yield is often context-dependent: A review of

experimental evidence. *Agriculture, Ecosystems & Environment*, *280*, 16–23.

https://doi.org/10.1016/j.agee.2019.04.022

Tamburini, G., Bommarco, R., Wanger, T. C., Kremen, C., Heijden, M. G. A. van der, Liebman,

M., & Hallin, S. (2020). Agricultural diversification promotes multiple ecosystem

services without compromising yield. *Science Advances*, *6*(45), eaba1715.

https://doi.org/10.1126/sciadv.aba1715

Tooker, J. F., O'Neal, M. E., & Rodriguez-Saona, C. (2020). Balancing disturbance and

conservation in agroecosystems to improve biological control. *Annual Review of*

Entomology, *65*(1), 81–100. https://doi.org/10.1146/annurev-ento-011019-025143

Tscharntke, T., Tylianakis, J. M., Rand, T. A., Didham, R. K., Fahrig, L., Batáry, P., Bengtsson,

J., Clough, Y., Crist, T. O., Dormann, C. F., Ewers, R. M., Fründ, J., Holt, R. D.,

Holzschuh, A., Klein, A. M., Kleijn, D., Kremen, C., Landis, D. A., Laurance, W., …

Westphal, C. (2012). Landscape moderation of biodiversity patterns and processes—

Eight hypotheses. *Biological Reviews*, *87*(3), 661–685. https://doi.org/10.1111/j.1469-

185X.2011.00216.x

Ullmann, K. S., Meisner, M. H., & Williams, N. M. (2016). Impact of tillage on the crop

pollinating, ground-nesting bee, *Peponapis pruinosa* in California. *Agriculture,*

Ecosystems & Environment, *232*, 240–246. https://doi.org/10.1016/j.agee.2016.08.002

USDA NASS. (2020). *Crop production historical track records* (p. 241). USDA, NASS.

https://www.nass.usda.gov/Publications/Todays_Reports/reports/croptr20.pdf

USDA SARS. (2014a). *2013 Idaho Cropland Data Layer*. USDA, NASS.

USDA SARS. (2014b). *2013 Washington Cropland Data Layer*. USDA, NASS.

USDA SARS. (2015a). *2014 Idaho Cropland Data Layer*. USDA, NASS.

USDA SARS. (2015b). *2014 Washington Cropland Data Layer*. USDA, NASS.

Venables, W. N., & Ripley, B. D. (2002). *Modern Applied Statistics with S* (4th ed.). Springer.

Weekers, T., Marshall, L., Leclercq, N., Wood, T. J., Cejas, D., Drepper, B., Garratt, M.,

Hutchinson, L., Roberts, S., Bosch, J., Roquer-Beni, L., Lhomme, P., Michez, D.,

Molenberg, J.-M., Smagghe, G., Vandamme, P., & Vereecken, N. J. (2022). Ecological,

environmental, and management data indicate apple production is driven by wild bee

diversity and management practices. *Ecological Indicators*, *139*, 108880.

https://doi.org/10.1016/j.ecolind.2022.108880

Wichura, A., Schorpp, Q., Kühlmann, V., & Hommes, M. (2022). Routine on-farm soil tillage

helps control asparagus fly (*Plioreocepta poeciloptera*). *Gesunde Pflanzen*, *74*(1), 1–8.

https://doi.org/10.1007/s10343-021-00609-8

Winfree, R., W. Fox, J., Williams, N. M., Reilly, J. R., & Cariveau, D. P. (2015). Abundance of

common species, not species richness, drives delivery of a real-world ecosystem service.

Ecology Letters, *18*(7), 626–635. https://doi.org/10.1111/ele.12424

Fig. 1: Box and whisker plots of (a) pollinator richness and (b) kleptoparasite abundance as a

function of tillage intensity. "Int." is intermediate tillage. Lines with asterisks indicate groups

that are statistically different from each other.

Fig. 2: (a) Herbivore and (b) predator abundance increase with landscape-scale habitat

availability (proportion of semi-natural habitat in a 1 km radius around a site). Curves show best-

fit lines from negative binomial regressions.

Fig. 3: Canola yield (a) was lowest with no tillage and (b) decreased as herbivore richness

increased. "Int." is intermediate tillage. Lines with asterisks (a) indicate groups that are

statistically different from each other. The curve (b) shows the best-fit line from linear

regression.