1 **Combining δ^{13}C and δ^{15}N from bone and dentine in marine mammal palaeoecological**
2 **research: insights from toothed whales**
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4 **Short title (up to 70 characters)**
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6 δ^{13}C and δ^{15}N from bone and dentine: insights from toothed whales
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26 **Abstract**
27
Rationale
Stable carbon (δ^{13}C) and nitrogen (δ^{15}N) isotope compositions of bone and dentine
collagen extracted from subfossil specimens of extinct and extant mammalian species have
been widely used to study the paleoecology of past populations. Due to possible systematic
differences in stable isotope values between bone and dentine, dentine values can be
transformed into bone-collagen equivalent using a correction factor. This approach has been
applied to terrestrial species, but correction factors specifically for marine mammals are
lacking. Here, we provide correction factors to transform dentine δ^{13}C and δ^{15}N values into
bone-collagen equivalent for two toothed whale sister species: narwhal and beluga.
37

Methods
We sampled bone and tooth dentine from the skulls of 11 narwhals and 26 belugas. In
narwhals, dentine was sampled from tusk and embedded tooth; in beluga, dentine was
sampled from tooth. δ^{13}C and δ^{15}N were measured using an elemental analyzer coupled to a
continuous flow isotope ratio mass spectrometer. Intraindividual bone and dentine isotopic
compositions were used to calculate correction factors for each species, and to translate
dentine isotopic values into bone-collagen equivalent.
45

Results
Our analysis revealed differences in δ^{13}C and δ^{15}N between bone and dentine. In
narwhals, we found (i) lower average δ^{13}C in bone compared with dentine from tusk and
embedded tooth; (ii) no difference in dentine δ^{13}C between tusk and embedded tooth; (iii)
lower average δ^{15}N in bone compared with dentine, with the highest values found in

embedded tooth. For belugas, we also detected lower δ^{13}C and δ^{15}N in bone compared with
tooth dentine.
53
54 **Conclusions**
55   Based on our analysis, we provide bone/dentine correction factors for narwhals (both
56 at species and population level), and for belugas. The correction factors, when applied to
57 dentine δ^{13}C and δ^{15}N values, enable the combined analysis of stable isotope data from bone
58 and dentine.
59
60 **Keywords**
61
62 Beluga whale, marine mammals, narwhal, paleoecology, stable isotopes
63
64 **Introduction**
65
Stable carbon (δ^{13}C) and nitrogen (δ^{15}N) isotope compositions of bone and dentine
collagen extracted from subfossil specimens have been used to study the paleoecology of past
populations and extinct species of mammals. Most of these studies have focused on terrestrial
taxa, including northern hemisphere species such as brown bear (*Ursus arctos*), hyena
(*Crocuta crocuta*), musk ox (*Ovibos moschatus*), saiga (*Saiga tatarica*), and woolly
rhinoceros (*Coelodonta antiquitatis*) [1-5], and South American species including camelids,
Equids, and Gomphotheres (extinct elephant-like proboscideans) [6,7].
73
74   Although to a lesser extent, stable isotope studies have also been applied to fossil
75 bone and dentine material of marine mammal species, including pinnipeds and cetaceans
76 [e.g., 8-12]. Stable isotope analysis of osseous materials has also been applied to
77 contemporary marine mammal populations to elucidate foraging ecology and dietary niche of
78 e.g., narwhals (*Monodon monoceros*) and belugas (*Delphinapterus leucas*) [13,14].
79
80   As a palaeoecological marker, carbon provides information on the photosynthetic
81 pathways and environmental parameters of the producers at the base of the food web. In
82 terrestrial environments, δ^{13}C represents plants [15], while in marine environments, δ^{13}C
83 primarily reflects phytoplankton and algae composition [12]. In both terrestrial and marine
84 ecosystems, δ^{15}N reflects the trophic position of an individual or species [16].
85
Combined, δ^{13}C and δ^{15}N from bone collagen provide information on the average diet
of an individual over multiple years [16,17]. In contrast with bone, which is continually
remodelled, dentine grows incrementally, and therefore the isotopic composition of dentine
corresponds to different years of an individual's life, depending on the specific tooth/tusk
development of the target species [18]. Thus, despite bone and dentine collagen being the
same substrate, they incorporate species' life histories differently. In narwhals, for example,
where males have one (and rarely two) long erupted tusks, stable isotope values from the
various dentine layers of the tusk represent an individual's diet in different years [19]. In
mammal species, where young nurse on their mother's milk until they are weaned, collagen
extracted from dentine of the first set of teeth in calves/juveniles, or from the roots of the
permanent teeth, usually have higher δ^{15}N values, reflecting the suckling period [2,20, 21].
97
If differences in stable isotope values between bone and dentine are systematic,
dentine isotopic values could be translated into the bone-collagen equivalent using a
correction factor. This would enable the direct comparison between δ^{13}C and δ^{15}N derived

from these distinct skeletal elements. The correction factor is an estimate of the average
difference between bone and dentine δ^{13}C and δ^{15}N from the same individual, and across
conspecific samples. This approach has been used to estimate the average difference in δ^{15}N
retrieved from bone and tooth in terrestrial mammals; e.g., wolves and hyenas, which across
samples resulted in a correction factor of 1.92 ± 0.19‰ and 1.08 ± 0.55‰, respectively [2].
However, it is important to keep in mind the difference in growth pattern between marine and
terrestrial mammals. Marine mammals' tooth growth pattern is characterized by the layered
deposition of dentine, which happens yearly in most marine mammal species [22].

Museum collections housing faunal remains are a crucial resource for studying
changes in populations and species across time and space. However, when analysing
specimens of the same species from collections or from the field, it is not always possible to
access the same skeletal element. Targeting only a specific skeletal element (e.g., cranial
bone or tooth) for analysis may drastically reduce the number of available specimens,
affecting the robustness and reliability of results. Here, we present a systematic approach to
circumvent this issue in two toothed whale species, narwhal and beluga (Monodontidae).

To calculate a correction factor for δ^{13}C and δ^{15}N in narwhal and beluga, we obtained
collagen δ^{13}C and δ^{15}N from bone and dentine collected from voucher specimens housed at
the Natural History Museum of Denmark, University of Copenhagen, and from the Greenland
Institute of Natural Resources. The various samples were used to estimate isotopic
differences between bone and dentine collagen within individuals, and average differences
among individuals within each species. Average differences were used to calculate a
correction factor, which can be applied to transform dentine collagen isotopic values into
bone-collagen equivalent.

Materials and Methods

1. Samples

The sampled material comprised skulls from 11 narwhals and 26 belugas. We
sampled skulls for both bone and dentine at the Natural History Museum of Denmark,
University of Copenhagen, and from the Greenland Institute of Natural Resources (see
detailed sample information in Supplementary Table 1). Five narwhal skulls were from West
Greenland (WG) and six from East Greenland (EG). All belugas were sampled in West
Greenland, as the species is not present in East Greenland. The skulls were collected in the
field between 1990 and 2018.

Narwhals have a unique dentition; they lack teeth in the lower jaw, and in the upper
jaw males have one long erupted tusk and one embedded tooth, while females have two
embedded teeth [23]. Hence for narwhals, we determined the isotopic compositions of three
skeletal elements: tusk, embedded tooth, and bone. For belugas, which have a row of
homodont teeth in both the upper and lower jaw, we sampled tooth and bone.

All individuals sampled for this study were adults, to avoid issues of high δ^{15}N values
in the bone and dentine of nursing calves/juveniles. Sex information for the samples was
based on the presence/absence of a tusk in narwhals, and/or the inspection of sexual organs in
both narwhals and belugas.

2. Stable isotope δ^{13}C and δ^{15}N analysis

We drilled 200-300 mg of each skeletal element. For narwhals, we drilled (i) bone, (ii)
tusk, and (iii) embedded tooth. For one of the narwhal samples (ID: 956), we did not have
access to the tusk, and therefore only sampled bone and embedded tooth. For belugas, we
drilled tooth dentine from each skull; δ^{13}C and δ^{15}N data from the same specimens have been
published previously in [13], and re-analyzed by [14]. See detailed sample information in
Supplementary Table 1.

Stable carbon and nitrogen isotope compositions were determined using a continuous
flow isotope ratio mass spectrometer at the Water Quality Centre, Trent University, Canada,
following the protocol described in. Full details are available in Supplementary Text.

Differences in δ^{13}C in samples collected at different time points may reflect changes
in δ^{13}C of atmospheric CO_2, reflecting the industrial revolution and the use of fossil fuels
[24]. Therefore, to avoid erroneous interpretations, δ^{13}C from samples collected across time
require correction. We corrected for this change, termed the Suess effect, following the
formula from [25].

To contextualise the data generated in our study, we combined our findings with
published δ^{13}C and δ^{15}N bone records (also from skulls) from 40 WG narwhals, 39 EG
narwhals, and one additional beluga [14].

3.   Skeletal element comparisons
169
To estimate a correction factor, we calculated pairwise average isotopic differences
between skeletal elements for each sampled individual (data for narwhal: Supplementary
Table 2; data for beluga: Supplementary Table 3). For narwhal, these estimates were also
calculated for each population, West Greenland and East Greenland; results are summarised
in Table 1. Correction factors were estimated as the average of the pairwise average
differences for each species, and for each population in narwhal.
176
177   **Results**
178
1.   Correction factor in narwhals

In narwhals, δ^{13}C values from bone collagen were lower than in dentine; this pattern
was consistent in both WG and EG narwhals (Supplementary Figure 1). In WG narwhals,
differences in δ^{13}C between tusk and bone (0.81 ± 0.13‰) were larger than differences
between embedded tooth and bone (0.37 ± 0.45‰). In EG narwhals, we observed the
opposite, and differences between tusk and bone (0.31 ± 0.25‰) were smaller than
differences between embedded tooth and bone (0.51 ± 0.35‰) (Table 1, Figure 1).

In both narwhal populations, δ^{15}N values from bone collagen were lower than in
dentine (Supplementary Figure 1). Both WG and EG narwhals showed smaller differences
between tusk and bone (0.49 ± 0.60‰ and 0.48 ± 0.37‰, respectively), than between
embedded tooth and bone (1.65 ± 0.69‰ and 1.45 ± 0.57‰, respectively) (Table 1, Figure
1).

2.   Correction factor in belugas

In belugas, $\delta^{13}C$ values from bone collagen were lower than in dentine
(Supplementary Figure 1). The average difference between tooth and bone was 0.37 ± 0.26‰
(Table 1, Figure 1). For $\delta^{15}N$, bone collagen values were lower than dentine (0.59 ± 0.69‰).

Average differences among skeletal elements for each species and population (for
narwhal) were used as a correction factor to translate dentine collagen isotopic values to
bone-collagen equivalent (Table 1). Differences in $\delta^{13}C$ and $\delta^{15}N$ before and after correcting
the data are visualised in Figure 1. Absolute comparisons can be found in Supplementary
Figure 1 and full data can be found in Supplementary Table 4.

We also summarized average $\delta^{13}C$ and $\delta^{15}N$ before and after correcting the data
derived from each dentine element (tusk, embedded tooth, tooth); we report these average
values for each species, and for the two geographic populations for narwhals in Table 2.
Overall, we detected a reduction in the difference between skeletal element average values
after application of the correction factor (Table 2, Figure 1). Dentine corrected values fall
within the bone average of a combined panel of newly generated and published data for these
species (Supplementary Figure 1, bone_reference).

207
208 **Discussion and Conclusions**
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We measured $\delta^{13}C$ and $\delta^{15}N$ from bone and dentine collagen derived from narwhal (n
= 11) and beluga (n = 26). The bone and dentine were sampled from whale skulls housed in
museum collections. We performed a systematic comparison of isotopic composition of
various skeletal elements from the same individuals, and across individuals within each
species and/or population (for narwhal). These values were successfully used as a correction
factor, allowing the direct comparison of data from different skeletal elements, and from
different populations.
217
For both narwhal and beluga, we detected lower values of $\delta^{13}C$ and $\delta^{15}N$ in bone
collagen than in dentine collagen. For narwhal, the highest $\delta^{15}N$ values were found in
embedded tooth. Due to the smaller size of the embedded tooth compared to the tusk, its
isotopic composition could still present higher $\delta^{15}N$ due to the suckling effect [18,20].
Differences in isotopic composition across tusk layers have been reported [19]; however, our
sampling strategy did not target a specific layer, but rather sampled a chunk of tusk
comprising several layers, and hence reflected average $\delta^{13}C$ and $\delta^{15}N$ across several years.
Because of this sampling strategy, the correction factors generated in this study are applicable
to samples collected in a similar manner.
227
Using $\delta^{13}C$ and $\delta^{15}N$ from bone collagen as dietary indicators, population differences
in the foraging ecology of narwhals in West Greenland and in East Greenland have been
reported [14]. Our narwhal bone and dentine data also showed systematic differences in $\delta^{13}C$
and $\delta^{15}N$ based on populations. Thus, for narwhal, we estimated a species-wide correction
factor, as well as a correction factor specifically for each population (Table 1, Figure 1).
When we applied the species-wide correction factor, the average differences between tusk
and bone, or embedded tooth and bone, were reduced to values approaching zero (Figure 1B,
Figure 1C). Thus, we show that in the absence of *a priori* information on population
differences due to life history in isotopic values within a species, it is feasible to apply a
species-wide correction factor to combine bone and dentine data. However, if possible, one

should consider life history differences among populations of a species when estimating
correction factors, as it might bias our results.
240
Our bone/dentine correction factor was estimated based on cranial bone and dentine
derived from three dentine elements: tusk and embedded tooth in narwhals, and tooth in
belugas. As only one bone element was included (skull), our correction factor may not be
fully representative of the intraskeletal bone isotopic variation in either species. δ^{13}C and
δ^{15}N values in bone tissue are dependent on bone remodeling and biochemical turnover rates
[17]. In humans, higher bone turnover rates are significantly and negatively correlated with
lower δ^{15}N [26]. Smith et al. 2020 [27] compiled published and new data on intraskeletal
bone variation in δ^{13}C and δ^{15}N across marine mammals. The data showed variation in
intraskeletal bone δ^{13}C ranging from 0.1‰ in walrus (*Odobenus rosmarus*) (between cranium
and mandible [28]) to 1.4‰ in dusky dolphin (*Lagenorhynchus obscurus*) (among atlas,
humerus and basioccipital [29]). Variation in intraskeletal bone δ^{15}N ranged from 0.3‰ in
walrus and dusky dolphin [28, 29] to 1.3‰ in South American sea lion (*Otaria flavescens*)
(among atlas, humerus and basioccipital [29]). However, other studies could not detect
intraskeletal variation when performing similar analysis; e.g., [30] did not detect differences
in isotopic composition between cortical and trabecular bone in harp seals (*Pagophilus*
groenlandicus).
257
Intraskeletal bone variation in δ^{13}C and δ^{15}N has also been investigated in terrestrial
mammals (e.g., in humans [26]; in cow (*Bos taurus*), pig (*Sus domesticus*), long-tailed weasel
(*Mustela frenata)*, and raccoon (*Procyon lotor*) [30]). Using an isotopic distance value
estimated from δ^{13}C and δ^{15}N bivariate plots, Hyland et al. 2021 [30] detected an average
isotopic distance value of bone sampled within individuals of 0.52‰. These data support the
existence of intraskeletal variation in isotopic composition; however, there are no clear
patterns, and variation appears to be species-specific. Our results showed intraskeletal
variation between bone and various dentine elements. In narwhal, we found a species-wide
variation in δ^{13}C of 0.51 ± 0.33‰ between tusk and bone, and 0.45 ± 0.38‰ between
embedded tooth and bone. For δ^{15}N, these values were 0.49 ± 0.44‰ and 1.54 ± 0.57‰,
respectively. In beluga, the δ^{13}C average difference between tooth and bone was 0.37 ±
0.26‰; for δ^{15}N, it was 0.59 ± 0.69‰.
270
Paleoecological inference from museum specimens is crucial for elucidating changes
in populations and species across time and space. However, the same skeletal element is often
not available across specimens. Bone and dentine collagen are known to show systematic
differences in δ^{13}C and δ^{15}N [31]. Thus, analyzing them together without applying a
correction factor may influence biological inferences. Here, we estimate and apply a
correction factor to our data from narwhal and beluga, and transform dentine values to bone-
collagen equivalent. Our approach assumes that one correction factor can be broadly applied
to transform dentine collagen to bone-collagen equivalent, and that data from different bone
skeletal elements can be analyzed in unison. Application of our correction factors allow the
direct comparison of δ^{13}C and δ^{15}N retrieved from these different skeletal elements (Figure 1
and Table 2). We acknowledge that our approach may not capture the full variation of
skeletal stable isotopes within an individual. However, using the correction factors when
analysing stable isotope data retrieved from bone and dentine reduces a large fraction of the
intraskeletal variation. This approach has been successfully applied to terrestrial mammals
(e.g., [2] reviewed the topic for several species). Here, we show its applicability in two
marine mammal species: narwhals and beluga. When dentine values were transformed to

bone-collagen equivalents using our approach, they fall within the range of known bone δ^{13}C and δ^{15}N values for these species demonstrating their applicability (Figure 1).

Data Accessibility:
Electronic Supplementary Table 1.

Author Contributions:

Research idea and study design: ARI, PS, EDL; data acquisition: ARI, MK, EG, MPHJ, EDL; data generation: TW, JR, PS; data analysis/interpretation: ARI, TW, JR; manuscript drafting: ARI, EDL with input from all the co-authors.

Competing interests:

We have no competing interests

Funding:

This research was supported by the Villum Fonden Young Investigator Programme, grant no 13151 and the Carlsberg Foundation Distinguished Associate Professor Fellowship, grant no CF16-0202 to EDL.

Acknowledgements:

We thank the Institutions that provided us with samples, and the fieldworkers and hunters collecting the samples. We thank Jeppe Møhl and Daniel K. Johansson for assisting with the sampling of the narwhal skulls at the Natural History Museum of Denmark, University of Copenhagen.

References:

1. Raghavan M, Themudo GE, Smith CI, Zazula G, Campos PF. Musk ox (*Ovibos moschatus*) of the mammoth steppe: tracing palaeodietary and palaeoenvironmental changes over the last 50,000 years using carbon and nitrogen isotopic analysis. *Quaternary Science Reviews*. 2014;102:192-201. doi:10.1016/j.quascirev.2014.08.001
2. Bocherens H. Isotopic tracking of large carnivore palaeoecology in the mammoth steppe. *Quat Sci Rev*. 2015;117:42-71. doi:10.1016/j.quascirev.2015.03.018
3. Jürgensen J, Drucker DG, Stuart AJ, Schneider M, Buuveibaatar B, Bocherens H. Diet and habitat of the saiga antelope during the late Quaternary using stable carbon and nitrogen isotope ratios. *Quat Sci Rev*. 2017;160:150-161. doi:10.1016/j.quascirev.2017.01.022
4. Rey-Iglesia A, García-Vázquez A, Treadaway EC, et al. Evolutionary history and palaeoecology of brown bear in North-East Siberia re-examined using ancient DNA and stable isotopes from skeletal remains. *Sci Rep*. 2019;9(1):4462.
5. Rey-Iglesia A, Lister AM, Stuart AJ, et al. Late Pleistocene paleoecology and phylogeography of woolly rhinoceroses. *Quat Sci Rev*. 2021;263:106993. doi:10.1016/j.quascirev.2021.106993
6. Domingo L, Prado JL, Alberdi MT. The effect of paleoecology and paleobiogeography on stable isotopes of Quaternary mammals from South America. *Quat Sci Rev*. 2012;55:103-113.

7. Díaz-Maroto P, Rey-Iglesia A, Cartajena I, et al. Ancient DNA Reveals the Lost Domestication History of South American Camelids in Northern Chile and Across the Andes. doi:10.1101/2020.10.16.337428

8. Burton RK, Snodgrass JJ, Gifford-Gonzalez D, Guilderson T, Brown T, Koch PL. Holocene changes in the ecology of northern fur seals: insights from stable isotopes and archaeofauna. *Oecologia*. 2001;128(1):107-115.

9. Alter SE, Newsome SD, Palumbi SR. Pre-whaling genetic diversity and population ecology in eastern Pacific gray whales: insights from ancient DNA and stable isotopes. *PLoS One*. 2012;7(5):e35039.

10. Vales DG, Cardona L, Zangrando AF, et al. Holocene changes in the trophic ecology of an apex marine predator in the South Atlantic Ocean. *Oecologia*. 2017;183(2):555-570.

11. Clark CT, Horstmann L, de Vernal A, Jensen AM, Misarti N. Pacific walrus diet across 4000 years of changing sea ice conditions. *Quat Res*. Published online March 14, 2019:1-17.

12. Szpak P, Savelle JM, Conolly J, Richards MP. Variation in late holocene marine environments in the Canadian Arctic Archipelago: Evidence from ringed seal bone collagen stable isotope compositions. *Quat Sci Rev*. 2019;211:136-155. doi:10.1016/j.quascirev.2019.03.016

13. Skovrind M, Castruita JAS, Haile J, et al. Hybridization between two high Arctic cetaceans confirmed by genomic analysis. *Sci Rep*. 2019;9(1):7729.

14. Louis M, Skovrind M, Garde E, Heide-Jørgensen MP, Szpak P, Lorenzen ED. Population-specific sex and size variation in long-term foraging ecology of belugas and narwhals. *R Soc Open Sci*. 2021;8(2):202226.

15. Bocherens H. Isotopic biogeochemistry and the palaeoecology of the mammoth steppe fauna. *Deinsea*. 2003;9(1):57-76.

16. Minagawa M, Wada E. Stepwise enrichment of ^{15}N along food chains: Further evidence and the relation between δ15N and animal age. *Geochim Cosmochim Acta*. 1984;48(5):1135-1140. doi:10.1016/0016-7037(84)90204-7

17. Hedges REM, Clement JG, Thomas CDL, O'connell TC. Collagen turnover in the adult femoral mid-shaft: modeled from anthropogenic radiocarbon tracer measurements. *Am J Phys Anthropol*. 2007;133(2):808-816.

18. Hobson KA, Sease JL. Stable isotope analyses of tooth annuli reveal temporal dietary records: an example using steller sea lions. *Mar Mamm Sci*. 1998;14(1):116-129. doi:10.1111/j.1748-7692.1998.tb00694.x

19. Dietz R, Desforges J-P, Rigét FF, et al. Analysis of narwhal tusks reveals lifelong feeding ecology and mercury exposure. *Curr Biol*. 2021;31(9):2012-2019.e2.

20. Fogel ML, Tuross N, Owsley DW. Nitrogen isotope tracers of human lactation in modern and archaeological populations. *Year B Carnegie Inst Wash*. 1989;88:111-117.

21. Fizet M, Mariotti A, Bocherens H, et al. Effect of diet, physiology and climate on carbon and nitrogen stable isotopes of collagen in a late pleistocene anthropic palaeoecosystem: Marillac, Charente, France. *J Archaeol Sci*. 1995;22(1):67-79. doi:10.1016/s0305-4403(95)80163-4

22. Scheffer VB, Myrick AC. *A Review of Studies to 1970 of Growth Layers in the Teeth of Marine Mammals*.1980.

23. Nweeia MT, Eichmiller FC, Hauschka PV, et al. Vestigial tooth anatomy and tusk nomenclature for monodon monoceros. *Anat Rec* . 2012;295(6):1006-1016.

24. Long ES, Sweitzer RA, Diefenbach DR, Ben-David M. Controlling for anthropogenically induced atmospheric variation in stable carbon isotope studies.

Oecologia. 2005;146(1):148-156. doi:10.1007/s00442-005-0181-6
25. Mellon SA. Investigating the ^{13}C Suess effect in the northwestern North Atlantic.
(Master's thesis, Dalhouse University, Canada). 2018. Retrieved from:
http://hdl.handle.net/10222/74718
26. Fahy GE, Deter C, Pitfield R, Miszkiewicz JJ, Mahoney P. Bone deep: Variation in
stable isotope ratios and histomorphometric measurements of bone remodelling
within adult humans. *J Archaeol Sci*. 2017;87:10-16. doi:10.1016/j.jas.2017.09.009
27. Smith KJ, Sparks JP, Timmons ZL, Peterson MJ. Cetacean Skeletons Demonstrate
Ecologically Relevant Variation in Intraskeletal Stable Isotopic Values. Front Mar
Sci. 2020;7. doi:10.3389/fmars.2020.00388
28. Clark CT, Horstmann L, Misarti N. Quantifying variability in stable carbon and
nitrogen isotope ratios within the skeletons of marine mammals of the suborder
Caniformia. *J Archaeol Sci Rep*. 2017;15:393-400. doi:10.1016/j.jasrep.2017.09.007
29. Bas M, García NA, Crespo EA, Cardona L. Intraskeletal variability in stable isotope
ratios of C and N among pinnipeds and cetaceans. *Mar Mamm Sci*. 2020;36(1):375-
385.
30. Hyland C, Scott MB, Routledge J, Szpak P. Stable carbon and nitrogen isotope
variability of bone collagen to determine the number of isotopically distinct
specimens. *J Archaeol Method Theory*. Published online September 1, 2021.
doi:10.1007/s10816-021-09533-7
31. Bocherens H, Fizet M, Mariotti A. Is collagen from teeth or bones equivalent for
isotopic (^{13}C, ^{15}N) diet investigations? *The Paleontological Society Special
Publications*. 1992;6:30-30. doi:10.1017/s2475262200005906

407
408   **Tables and Figures (with captions)**
409
Figure 1. Boxplots showing the average differences in δ^{13}C and δ^{15}N composition between
various skeletal elements analyzed from narwhal (tusk:bone, embedded tooth:bone) and
beluga (tooth:bone). (A) δ^{13}C and δ^{15}N raw composition, with δ^{13}C corrected for the Suess
effect; (B) δ^{13}C and δ^{15}N corrected using the correction factor estimated for each
species/skeletal element (data for narwhal: Supplementary Table 2; data for beluga:
Supplementary Table 3); and (C) δ^{13}C and δ^{15}N corrected using the correction factors
estimated for each narwhal population, considering known population differences [14]. All
data can be found in Supplementary Table 4. Colours represent species and populations.
Green palette: West Greenland (WG) narwhal, blue: East Greenland (EG) narwhal, and
purple: beluga.
420
421   **Table 1.** Average differences in δ^{13}C and δ^{15}N isotopic composition between different
422   skeletal elements in narwhal and beluga. These average differences were used as correction
423   factors to translate dentine values to bone-collagen equivalents.
424

Species	Population	Skeletal element	$\Delta\delta^{13}$C Average ± s.d. ‰	$\Delta\delta^{15}$N Average ± s.d. ‰
Narwhal	WG	Tusk/Bone	0.81 ± 0.13	0.49 ± 0.60
Narwhal	WG	Embedded /Bone	0.37 ± 0.45	1.65 ± 0.69
Narwhal	EG	Tusk/Bone	0.31 ± 0.25	0.48 ± 0.60

| Narwhal | EG | Embedded /Bone | 0.51 ± 0.35 | 1.45 ± 0.49 |
| Beluga | WG | Tooth/Bone | 0.37 ± 0.26 | 0.59 ± 0.60 |

Table 2. Average δ^{13}C and δ^{15}N values before and after correcting the data for systematic differences in skeletal element composition.

Species	Popula-tion	Skeletal element	Average δ^{13}C		Average δ^{15}N	
			Before correcting data (‰)	After correcting data (‰)	Before correcting data (‰)	After correcting data (‰)
Narwhal	WG	Tusk	-14.12 ± 0.42	-14.93 ± 0.42	17.44 ± 0.98	16.95 ± 0.98
		Embedded tooth	-14.35 ± 0.33	-14.72 ± 0.33	18.59 ± 0.94	16.94 ± 0.94
Narwhal	EG	Tusk	-15.72 ± 0.18	-16.03 ± 0.18	15.95 ± 0.34	15.47 ± 0.34
		Embedded tooth	-15.52 ± 0.56	-16.03 ± 0.56	16.92 ± 0.69	15.47 ± 0.69
Beluga	WG	Tooth	-13.65 ± 0.23	-14.02 ± 0.23	17.92 ± 0.63	17.33 ± 0.63

Supplementary Figures and Tables:

Supplementary Figure 1. Boxplots showing δ^{13}C and δ^{15}N composition of the various skeletal elements analyzed from narwhal and beluga. (A) δ^{13}C and δ^{15}N raw composition, with δ^{13}C corrected for the Suess effect; (B) δ^{13}C and δ^{15}N corrected using the correction factor estimated for each species/skeletal element (data for narwhal: Supplementary Table 2; data for beluga: Supplementary Table 3); and (C) δ^{13}C and δ^{15}N corrected using the correction factors estimated for each narwhal population, considering known population differences [14]. Colours represent species and /populations. Bone_reference represents the combined newly generated (for this study) and published bone data for beluga and narwhal [13,14]. Green palette: West Greenland (WG) narwhal, blue: East Greenland (EG) narwhal, and purple: beluga.

Supplementary Table 1. List of the narwhal and beluga samples used in this study to estimate correction factors.

Supplementary Table 2. Correction factor estimates for the narwhal samples.

Supplementary Table 3. Correction factor estimates for the beluga samples.

Supplementary Table 4. δ^{13}C and δ^{15}N data for the narwhal and beluga samples after applying the correction factor estimates. This table also includes the published data (bone_reference) for narwhal and beluga, which was used in Supplementary Figure 1.