Maternal investment evolves with larger body size and higher diversification

rate in sharks and rays

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Running head: Reproductive Mode Evolution of Sharks

Abstract

Across vertebrates, live-bearing has evolved at least 150 times from the ancestral

state of egg-laying into a diverse array of forms and degrees of prepartum maternal

investment. A key question is how this diversity of reproductive modes arose and

whether reproductive diversification underlies species diversification? To test these

questions, we evaluate the most basal jawed vertebrates, Chondrichthyans, which

have one of the greatest ranges of reproductive and ecological diversity among

vertebrates. We reconstructed the sequence of reproductive mode evolution across a

time-calibrated molecular phylogeny of 610 chondrichthyans. We find that egg-laying

is ancestral, and that live-bearing evolved at least seven times. Matrotrophy (i.e.

additional maternal contributions) evolved at least 15 times, with evidence of one

reversal. In sharks, transitions to live-bearing and matrotrophy are more prevalent in

larger-bodied species in the tropics. Further, the evolution of live-bearing is

associated with a near-doubling of the diversification rate, but, there is only a small

increase in diversification associated with the appearance of matrotrophy. The

chondrichthyan diversification and radiation, particularly throughout the shallow

tropical shelf seas and oceanic pelagic habitats, appears to be associated with the

evolution of live-bearing and the proliferation of a wide range of maternal investment

in their developing offspring.

Keywords: Live-bearing; Matrotrophy; Chondrichthyan; Shark; Trait Evolution;

Diversification

Introduction

A key transition in the evolution of vertebrate life is the appearance of live-

bearing and the evolution of maternal investment (Dulvy and Reynolds 1997;

Goodwin et al. 2002; Blackburn 2015b). Live-bearing is thought to have evolved to

increase offspring survival in the face of environmental and biological challenges

(Clutton-Brock and Godfrey 1991). The retention of eggs inside the maternal body

cavity provides a "safe harbor" for developing embryos (Shine 1978). For example, in

montane reptiles, the transition to live-bearing potentially protects eggs from freezing

(Shine 1995) or predation risk (Guillette 1993). However, mothers may suffer costs

from increased energetic investment, reduced fecundity, or greater predation risk and

reduced foraging ability due to pregnancy (Goodwin et al. 2002; Shine 2004). Body

size is important as internal body cavity space can limit female reproductive output.

Hence, live-bearing should be correlated with increasing maternal body size and the

space required to carry developing offspring to term in larger species (Qualls and

Shine 1995; Goodwin et al. 2002). In addition to body size, temperature may strongly

influence the evolution of reproductive modes in ecotherms (Shine 1995). In reptiles,

viviparity has evolved in cold climates because of the risk of freezing and lower

survival of eggs (Shine 2004).

Sister to all jawed vertebrates, the Class Chondrichthyes (hereafter 'shark and

rays'), exhibit live-bearing and varying modes of maternal investment (Wourms

1981; Dulvy and Reynolds 1997; Reynolds et al. 2002). Similar to reptiles,

reproductive transitions in chondrichthyans may be temperature-dependent but

potentially for different reasons (Compagno 1990). Sharks and rays that nourish

embryos solely via the yolk-sac (e.g. egg-laying or lecithotrophic live-bearing) are

predominantly found in colder habitats either in deep water or high latitude seas.

Whereas matrotrophic live-bearing, with maternal contribution beyond the yolk-sac,

is more prevalent in shallow tropical habitats, potentially reflecting temperature

effects on the rate and efficiency of development (Yampolsky and Scheiner 1996;

Dulvy 1998; Gillooly et al. 2002). Under this hypothesis, low maternal investment

strategies, such as lecithotrophic live-bearing, are more likely in colder environments

where metabolic rate (and hence maintenance costs) are lower, leaving a greater

fraction of energy available for embryonic growth. By contrast, the greater metabolic

maintenance costs in warmer environments may require additional maternal

investment through increased ovum size or post-fertilization trophic input to produce

a similar-sized offspring (Dulvy 1998). This temperature dependent – reproductive

investment hypothesis is consistent with thermal physiology (Gillooly et al. 2002),

but remains untested, simply because – until now – suitable molecular phylogenies

necessary for ancestral state reconstruction have been unavailable (Dulvy and

Reynolds 1997; Musick and Ellis 2005).

Reproductive mode evolution has the potential to influence speciation and

diversification dynamics through colonization of novel habitats (Yoder et al. 2010)

and / or parent-offspring conflicts (Zeh and Zeh 2000; Crespi and Semeniuk 2004).

Live-bearing sharks and rays tend to have larger geographic range sizes, potentially

spanning a greater range of habitats compared to smaller species (Goodwin et al.

2005). Speciation and diversification can also be driven by conflict between maternal

and offspring genomes in live-bearing species (Zeh and Zeh 2000), whereby conflict

over resource allocation during gestation drives antagonistic coevolution between

maternal and paternal genomes increasing the rate of species divergence. It is in the

father's best interest to have large, well-provisioned offspring, whereas it is in the

mother's best interest to weigh current offspring investment against her own survival

and future reproductive success (Zeh and Zeh 2000). Live-bearing provides an arena

in which this conflict can arise, and morphological adaptations for matrotrophic

nutrient transfer also provide an opportunity for embryos to influence maternal input

during gestation (Crespi and Semeniuk 2004). This resulting conflict over resources

could increase the rate of genetic divergence within populations, ultimately resulting

in speciation (Zeh and Zeh 2000). Hence, the potential for parent-offspring conflict in

live-bearing species may drive higher rates of diversification that may vary over the

range of maternal investment mechanisms. Increased rates of diversification

associated with live-bearing have been noted in squamate lizards (Pyron and Burbrink

2014) and teleost fishes (Helmstetter et al. 2016), although within live-bearers, the

degree to which matrotrophy versus lecithotrophy is related to species diversification

has yet to be tested.

Sharks and rays are an ideal group for studying the evolution of reproductive

mode and maternal investment. Sharks and rays exhibit egg-laying, live-bearing, and

multiple forms of matrotrophy along a continuum of maternal post-fertilization input

with yolk-sac live-bearing (lecithotrophy) at one end of the spectrum, with lipid

histotrophy, oophagy, and placentotrophy representing the extreme forms of

matrotrophy at the other. There remains considerable debate on whether the first

chondrichthyan laid eggs (Dulvy and Reynolds 1997) or gave birth to live young

(Musick and Ellis 2005).

The availability of a new molecular phylogeny of 610 chondrichthyan species

(Stein et al. 2018) with complete information on reproductive mode and habitat

provides a new opportunity to reconstruct the evolutionary history of reproductive

mode in chondrichthyans. We test the following predictions: (1) that the most recent

common ancestor of sharks and rays laid eggs based on the basal position of

Chimaeriformes; the evolution of live-bearing and varying forms of matrotrophy is

related to (2) larger body size, and (3) a radiation into shallow-water tropical habitats

(Compagno 1990). Finally, (4) that the evolution of live-bearing and matrotrophy is

associated with increasing rates of species diversification.

Methods

Trait Data and Phylogeny

Data on the reproductive mode and habitat type were collected for the 610
chondrichthyan species in our phylogeny, from primary literature and species
catalogues (Ebert et al. 2013; Dulvy et al. 2014; IUCN 2014). Chondrichthyans
exhibit eight distinct reproductive modes (Dulvy and Reynolds 1997), though we
focus the evolution of live bearing and maternal investment, therefore species were
categorized into three distinct modes: egg-laying, lecithotrophic live-bearing, and
matrotrophic live-bearing where embryos are nourished via the initial yolk-sac
investment and additional maternal contributions during gestation (oophagy,
histotrophy, and placentotrophy). We note that lecithotrophic live-bearing has also
been called yolk-sac viviparity, aplacental viviparity, or ovoviparity (Wourms 1981).
We collected data on maximum body size and depth ranges (minimum, mean,
median, and maximum) from species field guides and catalogues and primary
literature. Minimum and maximum latitudinal range was collected from species
Extent Of Occurrence geographic range maps from the International Union for the
Conservation of Nature (IUCN) Red List of Threatened Species database (IUCN
2014). Median latitude was calculated as the midpoint between minimum and
maximum latitude, and was expressed as an absolute value to represent distance north
or south from the equator. All continuous trait values were standardized, centered,
and divided by two standard deviations, using the `rescale` function in the `arm`
(version 1.9-3) (Gelman and Su 2016) package prior to analyses to facilitate
comparison of coefficients.

We conducted all analyses using a distribution of trees from a new 610 species

chondrichthyan molecular phylogeny (Stein et al. 2018). This phylogeny covers 51%

of all known species from every order, 98% of families, 88% of genera, and all

described character states are represented. Because the distribution of trees represents

a gradient of variation in root and node dating for the maximum likelihood tree, we

sequentially selected 21 trees, every 25th tree from one to 500 to account for the full

range of temporal calibrations. Results were pooled across all trees.

Ancestral State Reconstruction and Diversification

We reconstructed the evolutionary origins and sequence of reproductive mode and

habitat while estimating state dependent diversification rates using the multistate

speciation and extinction (MuSSE) method with maximum likelihood implemented

with `musse` (Pagel 1994) in the `diversitree` (version 0.9-9) package in R

(Fitzjohn 2012). Rabosky and Goldberg (2015) pointed out that state dependent

diversification models (including MuSSE) are susceptible to inflated false positives

when there is unmodeled heterogeneity in diversification rates – thereby associations

can be falsely detected between rates and states even when the diversification

dynamics are unrelated to the traits being considered. To minimize false positives, we

adopted the analytical approach of Uyeda et al. (2017) and integrated a hypothesis-

testing approach (i.e., MuSSE) with a more data-driven approach (what Uyeda et al.

refer to as "phylogenetic natural history"); in this way, we are able to tease out the

signal for our hypotheses that our focal traits are influencing diversification rates

while conditioning on the fact that there is likely substantial background variation in

these rates across the tree.

More specifically, we first used the medusa algorithm (as implemented in Geiger

v2.0.6; Pennell et al. 2014) to detect background variation in diversification rates

unrelated to our traits of interest. First, we infilled those species missing from the

molecular phylogeny (n = 582) using taxonomic constraints so that tips represented

unresolved clades. Importantly for earlier divergences, our sampling did not miss any

variability in reproductive mode (e.g. there are no live-bearing chimaeras) so missing

samples will not affect our conclusions. Under some circumstances, medusa may not

reliably correctly identify the placement of shifts (May and Moore 2016). However,

this issue does not pertain to our analysis because we are not making any inferences

about specific events or specific clades and are only interested in detecting broad-

scale differences in diversification dynamics. Our Medusa analysis only revealed

three clades with consistent increases in diversification rates across all trees: skates

(Rajiformes), South American freshwater stingrays (Potamotrygonidae), and ground

sharks (Carcharhiniformes). For the MuSSE analyses, we then partitioned the tree

into four diversification regimes (these three known radiations plus the background)

and fit a MuSSE model in which the diversification rate parameters were allowed to

vary among these four partitions. We also assigned each partition its own sampling

fraction based on the most up-to-date taxonomic treatment (Weigmann 2016; Stein et

al. 2018). Ideally, we would have fit an integrated model that included heterogeneity

in rates due to both the trait and to unmodeled "background" variation (Beaulieu and

O'Meara 2016; Uyeda et al. 2017) but unfortunately no such approach is available for

multi-state traits.

As we described above, speciation (λ), extinction (μ), and transition rates (q) were

estimated for each of the four partitions separately. Because the estimations of

extinction rates from molecular phylogenies can be difficult (Rabosky 2010), we ran

two models: with state dependent extinction rates (1) unconstrained ($\mu_{egg\text{-}laying} \neq \mu_{live\text{-}}$

$_{bearing} \neq \mu_{matrotrophic}$) and (2) constrained to be equal ($\mu_{egg\text{-}laying} = \mu_{live\text{-}bearing} = \mu_{matrotrophic}$).

We report findings from the unconstrained model as there was no significant

difference between models, and we are interested in how variation in this rate may

affect overall diversification. Additionally, the main difficulty with estimating

extinction rates arises from unaccounted for diversification rate heterogeneity, which

we have minimized by *a priori* identification with medusa and subsequent

partitioning. We report speciation (λ), extinction (μ), net diversification ($r = \lambda - \mu$),

and transition rates (q). We treated reproductive mode as an ordinal multistate

character and accordingly did not allow unlikely transitions such as directly between

egg-laying and matrotrophic live-bearing. Models were run for 10,000 generations

with the first 1,000 generations discarded as burn-in, using an exponential prior with

a rate of $1/(2r)$ where r is the character state independent diversification rate (Fitzjohn

2012). We checked that all parameter estimates had effective sample sizes (ESS: the

number of independent draws from an MCMC chain) greater than 200. To account

for autocorrelation in the estimates of state dependent diversification, we also

examined the posterior distribution of differences in state dependent rates across all

chains.

Evolutionary Covariation with Ecological Traits

We used a threshold model to test for the evolutionary covariation between

reproductive mode and continuous ecological traits. The threshold model assumes

that state changes in an ordinal variable (e.g. egg-laying to live-bearing to

matrotrophy) occur when a threshold value of an underlying continuous latent

variable, such as body size, is reached. Thus, it can be used to model the evolutionary

covariation between ordinal and continuous traits (Felsenstein 2012). Accurate

estimation of evolutionary covariation requires a suitable number of transitions and

distribution of traits across the phylogeny (Maddison and Fitzjohn 2015). We focus

on sharks (superorders Galeomorphii and Squalomorphii; n = 292) to evaluate

evolutionary covariance between reproductive transitions and three ecological traits

(body size, depth, and latitude), because there is only one transition in parity and few

appearances of matrotrophy within Chimaeriformes and rays (Batoideii). We

estimated the evolutionary covariance using Bayesian methods, sampling from the

posterior distribution using a special Reduced Animal Model implemented in a mixed

effects modeling framework while accounting for phylogeny, using the package

`MCMCglmmRAM` (version 2.24) in R (Hadfield 2015). This approach is equivalent to

estimating evolutionary covariance using the threshold model (Felsenstein 2012;

Revell 2014). These models are a special case of generalized linear mixed effects

models where heritability, akin to Pagel's λ, is set to a value of one corresponding to

Brownian motion with respect to the phylogenetic tree (Freckleton et al. 2002;

Housworth et al. 2004). Twenty chains were run for 2 million generations with the

first 200,000 iterations discarded as burn-in, using priors with an inverse-Wishart

distribution and the residual covariance matrix set to zero (Hadfield 2015). Samples

were drawn every 500 iterations to avoid temporal autocorrelation in parameter

estimates. Chains were visually inspected to ensure convergence using `coda` (version

0.19-4) (Plummer et al. 2006), and posterior samples were summarized to generate

mean and 95% highest posterior densities (HPD) with effective samples sizes greater

than 1000. Models were run using three different treatments of reproductive mode

with the threshold family: binary parity mode (egg-laying versus live-bearing), binary

embryo trophic mode (lecithotrophic versus matrotrophic), and ordinal multi-state

reproductive mode (egg-laying, lecithotrophic live-bearing, and matrotrophic live-

bearing).

Results

(1) Ancestral chondrichthyans laid eggs

The first chondrichthyans almost certainly laid eggs, as there is a high level of

support for egg-laying as the ancestral state of reproductive mode in chondrichthyans

(>99% probability; Figure 1). There are multiple independent origins of live-bearing

(seven) and matrotrophy (15), from the superordinal to subgeneric level, with few

instances of reversals (one reversal from matrotrophy to lecithotrophic live-bearing;

Figure 2). Specifically, live-bearing appears to have evolved from egg-laying at: (a)

base of Rhinopristiformes and Myliobatiformes, (b) base of Squalomorphii, (c) base

of clade encompassing Brachaeluridae, Orectolobidae, and Rhincodontidae, (d) base

of Ginglymostomatidae, (e) within the genus *Bythaelurus*, (f) in *Galeus polli*, and (g)

basal to clade encompassing Pseudotriakidae, Triakidae, Hemigaleidae, and

Carcharhinidae (Figure 1). We found no evidence of reversals from live-bearing to

egg-laying (Figure 2A).

Matrotrophy appears to have evolved independently from lecithotrophic live-

bearing at least 15 times with one reversal: (a-c) one to three origins within guitarfish

and wedgefish (Rhinopristiformes), (d) basal to stingrays (Myliobatiformes), (e-f) one

to three origins within sleeper sharks (Somniosidae), (g) great lanternsharks

(*Etmopterus princeps*), (h) tawny nurse shark (*Nebrius ferrugineus*), (i) mackerel

sharks (Lamniformes), (j) Pseudotriakidae, (k-m) one to three origins within

houndsharks (Triakidae), and (n) base of requiem sharks (Carcharhinidae). There was

evidence of a single instance of reversal from matrotrophy to lecithotrophic live-

bearing in the sharptooth houndshark (*Triakis megalopterus*). Overall transitions from

egg-laying to live-bearing and to matrotrophy occurred at higher rates then reversals

across all partitions that contained multiple reproductive modes (Figure 2B,C).

(2) Live-bearing and matrotrophy evolve with increasing maternal size

Reproductive mode was related to body size, such that larger bodied species had a

higher probability of live-bearing and matrotrophic investment (Figure 3A). We

found positive covariation with body size, using the threshold model to test for

evolutionary covariance between discrete values of reproductive mode and

continuous ecological traits, (median = 0.16, 95% CI = 0.09 to 0.22; Effective Sample

Size = 3351) indicating transitions in reproductive mode are more prevalent in

lineages with larger body size (Figure 3A). The relationship with larger body size was

slightly stronger for the transition from egg-laying to live-bearing (0.37 [0.17 to

0.55]; ESS = 3332) than for lecithotrophy to matrotrophy (0.3 [0.14 to 0.48]; ESS =

3600).

(3) Live-bearing and matrotrophy evolve in wide-ranging species at low latitudes

Live bearing species are more prevalent in the tropics, specifically transitions

from egg-laying to live-bearing are more prevalent in lineages at lower latitudes

(minimum latitude: -0.68 [-1.25 to -0.01], ESS = 2915, Figure 3C; median latitude: -

0.46 [-0.9 to -0.03]; ESS = 3189; Figure 3D). However, there was little evidence that

transitions in reproductive mode are related to either median latitude (-0.06 [-0.16 to

0.04], ESS = 3600) or median depth (0.06 [-0.04 to +0.15]; ESS = 3600; Figure 3

B,D). Transitions to live-bearing and in reproductive mode were more prevalent in

lineages with larger latitudinal ranges (0.058 [0.08 to 1.0], ESS = 2580; 0.11 [0.04 to

0.18], ESS = 3956; Figure 3E).

(4) Diversification rate is highest in matrotrophic lineages

Overall, the evolution of live-bearing and matrotrophy is associated with greater

diversification (Figure 4A-C), mainly due to a high relative extinction rate in egg-

laying species (Figure 4A,B). The evolution of matrotrophy is associated with greater

diversification than egg-laying (2.4 times), and there is weak evidence for higher

diversification than lecithotrophic live-bearing lineages (1.27 times; Figure 5A). Egg-

laying lineages have high turnover driven by high extinction rates because speciation

rate is highest in egg-laying lineages (mean 0.046 lineages/MY) compared with

lecithotrophic live-bearing (mean 0.03 lineages/MY) and matrotrophic lineages

(0.026 lineages/MY; Figure 4A-C).

Within the three radiations, the connection between reproductive mode and

diversification is more nuanced. Within the ground sharks (Carcharhiniformes), egg-

laying lineages have a higher diversification rate than lecithotrophic live-bearing

lineages (Figure 4D-F, Figure 5B), driven by high speciation in egg-laying cat sharks

that are found mainly in deepwater. Speciation is particularly high in both the skate

radiation (0.078 lineages/MY; Figure 4G) and the South American freshwater

stingray radiation (0.070 lineages/MY; Figure (4J), potentially reflecting their

colonization of novel deep-water and freshwater habitats, respectively.

Discussion

Here, we reveal the first chondrichthyan was an egg-layer and there have been

numerous transitions toward live-bearing and matrotrophy. The evolution of live-

bearing and matrotrophy covaries with increasing body size and is more prevalent in

shallow waters of tropical latitudes. Further, the evolution of live-bearing, and to a

lesser extent matrotrophy, appears to have resulted in greater species diversification.

Next, we consider three questions: What is the sequence of reproductive mode

evolution? What ecological factors have driven the evolution of live-bearing and

matrotrophy? Is chondrichthyan speciation and diversification explained by

viviparity-driven conflict or by novel ecological opportunity?

The sequence of reproductive mode evolution

We find support for egg-laying as the ancestral reproductive state, with numerous

independent origins of live-bearing and matrotrophy with few instances of reversals.

Previous analyses of reproductive evolution in chondrichthyans have been limited by

the available morphological phylogenetic hypotheses, particularly on the

phylogenetic position of rays (Shirai 1992; Naylor et al. 2012). Some morphological

analyses placed the rays as a highly derived group within the Squalimorph sharks, but

the emerging consensus from molecular data is that rays and sharks are sister taxa

(Naylor et al. 2012; Stein et al. 2018). With this new molecular tree, we find

transitions in reproductive mode are generally toward live-bearing or matrotrophy

though this not a strictly linear progression as reversals are infrequent though

plausible. We expect that improved biological sampling and phylogenetic

reconstructions may yield further examples, particularly in groups displaying

subgeneric transitions that are currently poorly phylogenetically resolved (e.g. within

catsharks, Scyliorhinidae). We find no support for live-bearing being ancestral nor

evidence of a high rate of reversals to egg-laying from live-bearing that this would

necessitate, as suggested previously (Musick and Ellis 2005). Interestingly, a similar

controversy has occurred in squamates, a group with highly labile reproductive

modes. The most likely model suggested an early origin of viviparity with a high rate

of reversals (Pyron and Burbrink 2014), though this result has been questioned based

on the choice of phylogenetic hypotheses and morphological features of live-bearing

lineages (Blackburn 2015a,b; Wright et al. 2015). Consequently, support remains for

the conventional hypothesis that the ancestral state was oviparity (Blackburn 2015b).

Live-bearing chondrichthyans, particularly lecithotrophic live-bearers, develop within

an egg envelope or candle which is similar to, but thinner than, that found in egg-

laying species (Castro 2009; Conrath and Musick 2012). This retention of the

morphological machinery for egg production could make reversals to egg-laying

feasible, and may allow for some of the subgeneric reproductive diversity seen within

catsharks (Scyliorhinidae). But these data, along with the emerging consensus in

reptiles, suggests that egg-laying is the ancestral vertebrate condition.

Transitions between matrotrophic modes can also occur, for example in hound

sharks (Family Triakidae) muccoid histotrophy can be used in lieu of, or in concert

with, placentotrophy (Hamlett et al. 2005) – resulting in intergeneric reproductive

diversity (López et al. 2006). Similarly, lecithotrophic live-bearing and some forms

of matrotrophy, particularly muccoid histotrophy found in Rhinopristiformes and

some sharks, may not represent discrete character states but rather a continuum that

will be revealed only with more detailed histological work in this lineage. Muccoid

histotrophy can be difficult to distinguish due to a paucity of uterine morphological

specializations and accurate measurements of ash-free dry weight of embryos and

ova. In several species of squaliform sharks the change in organic mass from ovum to

embryo has been used to identify muccoid histotrophy (Paiva et al. 2011; Cotton et al.

2014), though there is uncertainty given the exact threshold value of change in mass

that should be used to distinguish between modes (Frazer et al. 2012). Thus, large

groups predominantly composed of lecithotrophic live-bearing species (i.e.

Rhinopristiformes, Squalimorphii) may actually contain a greater diversity of

maternal investment than currently measurable or dealt with in this analysis. Despite

this uncertainty, chondrichthyans still exhibit remarkably labile reproductive modes

compared with other vertebrate groups, more similar to the transition rates seen in the

much larger clade Squamata (~10,000 species with >150 origins of live-bearing and 6

origins of matrotrophy) (Blackburn 2015a).

Evolutionary correlates of reproductive mode

Here we show that in sharks live-bearing and maternal investment has evolved in

association with larger body size, tropical latitudes, and increasing latitudinal range,

while there is little evidence of association with depth. We speculate that body size

may, in effect, be capturing allometric differences in predation pressure and access to

food resources, which have been hypothesized as drivers of reproductive evolution in

smaller more tractable freshwater livebearers (Trexler and Deangelis 2003; Marsh-

Matthews and Deaton 2006; Van Dyke et al. 2014). It seems that, in chondrichthyans

at least, the origin of live-bearing necessitates an increase in body size to

accommodate retained embryos throughout gestation in limited internal body space, if

larger offspring size is optimal (Blackburn 2015a). As a result, lecithotrophic live-

bearing sharks and rays typically have fewer, but larger, pups that are presumably

subject to less predation pressure and juvenile mortality as a result (Conrath and

Musick 2012; Kindsvater et al. 2016).

Despite detecting a relationship between reproductive mode and body size and

latitude in extant chondrichthyans (Rigby and Simpfendorfer 2013), our ability to test

for evolutionary covariation is hindered by the phylogenetic clustering of traits and

the use of depth and latitude as proxies for environmental temperature and resource

availability. As a result, it is currently difficult to disentangle evolutionary

hypotheses, as large lineages with similar character states may reflect a single origin

and subsequent coinheritance rather than a functional evolutionary covariation

(Maddison and Fitzjohn 2015). Rays exhibit an interesting transition – consistent with

the relationship we have revealed in sharks – from deep egg-laying skates to shallow

shelf and coastal live-bearing lineages, including: electric rays (Torpediniformes),

guitarfishes, wedgefishes, and sawfishes (Rhinopristiformes), and matrotrophic

stingrays (Myliobatiformes). However, our power to test for plausible correlations

between reproductive evolution and depth or latitude due to thermal physiology

(Dulvy 1998), predation (Harper and Peck 2016), and productivity are limited

because the batoids lineage contains a single origin of live-bearing and only one

certain origin of matrotrophy.

Viviparity-driven conflict versus novel ecological opportunity

At the large, coarse scale there appears to be evidence that radiations into novel

deepwater (Sorenson et al. 2014) or novel and highly dynamic fragmented freshwater

habitats (Lovejoy 1996; Lovejoy et al. 1998, 2006), rather than of reproductive mode

generally affecting diversification rates across all chondrichthyans. While

diversification rates are higher in live-bearing and matrotrophic lineages across the

main partition of the tree, resulting genomic conflicts may be a weak driver of

diversification compared to ecological forces in chondrichthyans.

The elevated diversification rates seen in skates (Rajiformes), South American

Freshwater stingrays (Potamotrygonidae), and ground sharks (Carcharhiniformes)

appear to be related to colonization of new ecological space. However, in each case,

the ecological space is different – skates radiated into deepwater habitats with the

opening up of the Atlantic Ocean (McEachran and Miyake 1990), subsequent

isolation of ocean basins (Long 1994), limited egg and adult dispersal ability

(McEachran and Miyake 1990), and a high degree of spatial niche differentiation

(Bizzarro et al. 2014; Humphries et al. 2016). By contrast the Neotropical

potamotrygonid stingrays (and other fish lineages) colonized freshwater upon the

closure of the isthmus of Panama (Lovejoy 1996, 1997; Lovejoy et al. 1998; de

Carvalho and Lovejoy 2011), radiating into novel and highly fragmented freshwater

habitat potentially facilitated by the emergence of unique morphological adaptations

(Kolmann et al. 2016). The increased diversification in ground sharks is driven by

both reinvasion of deepwater by catsharks (Scyliorhinidae) and the colonization of

shallow water coastal habitats (coral reefs and associated inshore and estuarine

habitats) by requiem sharks (Carcharhinidae) (Sorenson et al. 2014). At present, the

weight of evidence suggests that clade-specific increases in diversification rate are

associated with new ecological space, rather than being systematically driven by

reproductive evolution *per se* – nevertheless, we await stronger tests of this

hypothesis when more extensive phylogenies become available. Future investigations

of genomic conflict-driven speciation may require refined measurements of

ecological variation (Burin et al. 2016) or life history (Marki et al. 2015) as drivers of

diversification with tests restricted to smaller groups.

Conclusions

The evolution of chondrichthyan reproductive modes, ranging from egg-laying to

live-bearing and matrotrophy, appears strongly related to body size and temperature-

related biogeography. While patterns of species diversification in three major

radiations appear to be more strongly driven by colonization of novel habitats, the

evolution of the diversity of reproductive modes remains a fruitful area of research.

Parent-offspring conflict over resources during development and subsequent

antagonistic coevolution is an intriguing potential driver of reproductive mode

evolution worthy of further investigation. Chondrichthyans are an ideal group to test

for this given the diversity of reproductive modes and the frequency of polyandry,

though this requires a better understanding of maternal-fetal interactions across a

wider range of species. Future research could focus on improved measures of

maternal investment, particularly for identifying the continuum on which

lecithotrophic live-bearing and histotrophic matrotrophy may be expressed.

Combined with further refinement of phylogenetic hypotheses with more extensive

taxon sampling will help to clarify patterns of energetic investment, the degree of

income breeding in live-bearing species, and inter and intra specific plasticity. More

generally, we anticipate that this taxon will yield profound insights into the interplay

between reproductive life history evolution, ecology, and the biogeographic

patterning of species diversity across the Earth's oceans.

Ethics Statement

No live animals or human subjects were used in this study.

Data Availability

465   All phylogenetic trees are available at www.vertlife.org/sharktree.

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Author Contributions

476   CGM, KEY and NKD conceived the study; CGM and MWP designed the analytical approach; CGM performed the analyses; CGM, MWP, KEY and NKD drafted the manuscript. All authors gave final approval for publication.

Competing Interests

481   We have no competing interests.

Literature Cited

Beaulieu, J. M., and B. C. O'Meara. 2016. Detecting hidden diversification shifts in

models of trait-dependent speciation and extinction. Syst. Biol., doi:

10.1093/sysbio/syr025.

Bizzarro, J. J., K. M. Broms, M. G. Logsdon, D. A. Ebert, M. M. Yoklavich, L. A.

Kuhnz, and A. P. Summers. 2014. Spatial segregation in Eastern North pacific

skate assemblages. PLoS One 9.

Blackburn, D. G. 2015a. Evolution of vertebrate viviparity and specializations for

fetal nutrition: A quantitative and qualitative analysis. J. Morphol. 276:961–990.

Blackburn, D. G. 2015b. Evolution of viviparity in squamate reptiles: Reversibility

reconsidered. J. Exp. Zool. Part B Mol. Dev. Evol. 324:473–486.

Burin, G., W. D. Kissling, P. R. Guimarães, Ç. H. Sekercioglu, and T. B. Quental.

2016. Omnivory in birds is a macroevolutionary sink. Nat. Commun. 7:11250.

Castro, J. I. 2009. Observations on the reproductive cycles of some viviparous North

American sharks. Aqua, Int. J. Ichthyol. 15:205–222.

Clutton-Brock, T. H., and C. Godfrey. 1991. Parental Investment. Pp. 234–262 *in* J.

R. Krebs and N. B. Dabies, eds. Beahavioral Ecology: An Evolutionary

Approach. Cambridge University Press, Cambridge, UK.

Compagno, L. J. V. 1990. Alternative life-history styles of cartilaginous fishes in time

and space. Environ. Biol. Fishes 28:33–75.

Conrath, C. L., and J. A. Musick. 2012. Reproductive biology of elasmobranchs. Pp.

269–286 *in* J. C. Carrier, J. A. Musick, and M. R. Heithaus, eds. Biology of

sharks and their relatives. CRC Press, Boca Raton, FL.

Cotton, C. F., R. Dean Grubbs, J. E. Dyb, I. Fossen, and J. a. Musick. 2014.

Reproduction and embryonic development in two species of squaliform sharks,

Centrophorus granulosus and *Etmopterus princeps*: Evidence of matrotrophy?

Deep Sea Res. Part II Top. Stud. Oceanogr. 115:41–54. Elsevier.

Crespi, B., and C. Semeniuk. 2004. Parent-Offspring conflict in the evolution of

vertebrate reproductive mode. Am. Nat. 163:635–653.

de Carvalho, M. R., and N. R. Lovejoy. 2011. Morphological and phylogenetic

relationships of a remarkable new genus and two new species of Neotropical

freshwater stingrays from the Amazon basin (Chondrichthyes:

Potamotrygonidae). . Zootaxa 48:13–48.

Dulvy, N. K. 1998. Life histories and Conservation of Sharks and Rays. University of

East Anglia.

Dulvy, N. K., S. L. Fowler, J. A. Musick, R. D. Cavanagh, M. Kyne, L. R. Harrison,

J. K. Carlson, L. N. K. Davidson, V. Fordham, Sonja, M. P. Francis, C. M.

Pollock, C. A. Simpfendorfer, G. H. Burgess, K. E. Carpenter, L. J. V

Compagno, D. A. Ebert, C. Gibson, M. R. Heupel, S. R. Livingstone, J. C.

Sanciangco, J. D. Stevens, S. Valenti, and W. T. White. 2014. Extinction risk

and conservation of the world's sharks and rays. Elife 3:e00590.

Dulvy, N. K., and J. D. Reynolds. 1997. Evolutionary transitions among egg−laying,

live−bearing and maternal inputs in sharks and rays. Proc. R. Soc. B Biol. Sci.

264:1309–1315.

Ebert, D. A., S. L. Fowler, and L. J. V Compagno. 2013. Sharks of the world: a fully

illustrated guide. Wild Nature Press, Plymouth, UK.

536 Felsenstein, J. 2012. A Comparative Method for Both Discrete and Continuous

537   Characters Using the Threshold Model. Am. Nat. 179:145–156.

538 Fitzjohn, R. G. 2012. Diversitree: comparative phylogenetic analyses of

539   diversification in R. Methods Ecol. Evol. 1084–1092.

540 Frazer, H. a, M. Ellis, and C. Huveneers. 2012. Can a threshold value be used to

541   classify chondrichthyan reproductive modes: systematic review and validation

542   using an oviparous species. PLoS One 7:e50196.

543 Freckleton, R. P., P. H. Harvey, and M. Pagel. 2002. Phylogenetic analysis and

544   comparative data: A test and review of evidence. Am. Nat. 160:712–726.

545 Gelman, A., and Y.-S. Su. 2016. arm: data analysis using regression and

546   multilevel/hierarchical models.

547 Gillooly, J. F., E. L. Charnov, G. B. West, V. M. Savage, and J. H. Brown. 2002.

548   Effects of size and temperature on developmental time. Nature 417:70–73.

549 Goodwin, N. B., N. K. Dulvy, and J. D. Reynolds. 2002. Life-history correlates of the

550   evolution of live bearing in fishes. Philos. Trans. R. Soc. Lond. B. Biol. Sci.

551   357:259–67.

552 Goodwin, N. B., N. K. Dulvy, and J. D. Reynolds. 2005. Macroecology of live-

553   bearing in fishes: latitudinal and depth range comparisons with egg-laying

554   relatives. Oikos 110:209–218.

555 Guillette, L. J. 1993. The evolution of viviparity in lizards. Bioscience 43:742–751.

556 Hadfield, J. D. 2015. Increasing the efficiency of MCMC for hierarchical

557   phylogenetic models of categorical traits using reduced mixed models. Methods

558   Ecol. Evol. n/a-n/a.

Hamlett, W. C., G. Kormanik, M. Storrie, B. Stevens, and T. I. Walker. 2005.

Condrichthyan parity, lecithotrophy, and matrotrophy. Pp. 408–447 *in* W. C.

Hamlett, ed. Reproductive biology and phylogeny of chondrichthyans. Science

Publishers, Inc., Plymouth, UK.

Harper, E. M., and L. S. Peck. 2016. Latitudinal and depth gradients in marine

predation pressure. Glob. Ecol. Biogeogr. 25:670–678.

Helmstetter, A. J., A. S. T. Papadopulos, J. Igea, T. J. M. Van Dooren, A. M. Leroi,

and V. Savolainen. 2016. Viviparity stimulates diversification in an order of fish.

Nat. Commun. 7:11271. Nature Publishing Group.

Housworth, E. a, E. P. Martins, and M. Lynch. 2004. The phylogenetic mixed model.

Am. Nat. 163:84–96.

Humphries, N., S. Simpson, V. Wearmouth, and D. Sims. 2016. Two's company,

three's a crowd: fine-scale habitat partitioning by depth among sympatric species

of marine mesopredator. Mar. Ecol. Prog. Ser. 561:173–187.

IUCN. 2014. The IUCN Red List of Threatened Species. Version 2014.1.

Kindsvater, H. K., M. Mangel, J. D. Reynolds, and N. K. Dulvy. 2016. Ten principles

from evolutionary ecology essential for effective marine conservation. Ecol.

Evol. n/a-n/a.

Kolmann, M. A., K. C. Welch, A. P. Summers, and N. R. Lovejoy. 2016. Always

chew your food: freshwater stingrays use mastication to process tough insect

prey. Proc. R. Soc. B Biol. Sci. 283:20161392.

Long, D. J. 1994. Quaternary Colonization or Paleogene Persistence?: Historical

Biogeography of Skates (Chondrichtyes: Rajidae) in the Antarctic Ichthyofauna.

582   Paleobiology 20:215–228.

583  López, J. A., J. A. Ryburn, O. Fedrigo, and G. J. P. Naylor. 2006. Phylogeny of

584    sharks of the family Triakidae (Carcharhiniformes) and its implications for the

585    evolution of carcharhiniform placental viviparity. Mol. Phylogenet. Evol. 40:50–

586    60.

587  Lovejoy, N. R. 1997. Stingrays, parasites, and neotropical biogeography: a close look

588    at Borrks et al.'s hypotheses concerning the origins of neotropical freshwater

589    rays (Potamotrygonidae). Syst. Biol. 46:218–230.

590  Lovejoy, N. R. 1996. Systematics of myliobatoid elasmobranchs: with emphasis on

591    the phylogeny and historical biogeography of neotropical freshwater stingrays

592    (Potamotrygonidae: Zool. J. Linn. Soc. 117:207–257.

593  Lovejoy, N. R., J. S. Albert, and W. G. R. Crampton. 2006. Miocene marine

594    incursions and marine/freshwater transitions: Evidence from Neotropical fishes.

595    J. South Am. Earth Sci. 21:5–13.

596  Lovejoy, N. R., E. Bermingham, and A. P. Martin. 1998. Marine incursion into South

597    America. Nature 396:421–422.

598  Maddison, W. P., and R. G. Fitzjohn. 2015. The unsolved challenge to phylogenetic

599    correlation tests for categorical characters. Syst. Biol. 64:127–136.

600  Marki, P. Z., P. H. Fabre, K. A. Jønsson, C. Rahbek, J. Fjeldså, and J. D. Kennedy.

601    2015. Breeding system evolution influenced the geographic expansion and

602    diversification of the core Corvoidea (Aves: Passeriformes). Evolution (N. Y).

603    69:1874–1924.

604  Marsh-Matthews, E., and R. Deaton. 2006. Resources and offspring provisioning: A

test of the Trexler-DeAngelis model for matrotrophy evolution. Ecology

87:3014–3020.

May, M. R., and B. R. Moore. 2016. How Well Can We Detect Lineage-Specific

Diversification-Rate Shifts? A Simulation Study of Sequential AIC Methods.

Syst. Biol. 65:1076–1084.

McEachran, J. D., and T. Miyake. 1990. Zoogeography and bathymetry of skates

(Chondrichthyes, Rajoidei). Pp. 305–326 *in* H. L. Pratt, S. H. Gruber, and T.

Taniuchi, eds. Elasmobranchs as living resources: advances in the biology,

ecology, systematics and the status of the fisheries. National Oceanic and

Atmospheric Administration.

Musick, J. A., and J. K. Ellis. 2005. Reproductive evolution of chondrichthyans. Pp.

45–79 *in* W. C. Hamlett, ed. Reproductive biology and phylogeny of

chondrichthyans. Science Publishers, Inc., Plymouth, UK.

Naylor, G. J. P., J. N. Caira, K. Jensen, K. a M. Rosana, N. Straube, and C. Lakner.

2012. Elasmobranch Phylogeny: A mitochondrial estimate based on 595 species.

Pp. 31–56 *in* J. C. Carrier, J. A. Musick, and M. R. Heithaus, eds. The Biology

of Sharks and Their Relatives. CRC Press, Boca Raton, FL.

Pagel, M. 1994. Detecting correlated evolution on phylogenies: a general method for

the comparative analysis of discrete characters. Proc. R. Soc. B Biol. Sci.

255:37–45.

Paiva, R. B., A. Neves, V. Sequeira, M. L. Nunes, L. S. Gordo, and N. Bandarra.

2011. Reproductive strategy of the female deep-water shark birdbeak dogfish,

Deania calcea: lecithotrophy or matrotrophy? J. Mar. Biol. Assoc. United

Kingdom 92:387–394.

Pennell, M. W., J. M. Eastman, G. J. Slater, W. Brown, J. C. Uyeda, R. G. Fitzjohn,

M. E. Alfaro, and L. J. Harmon. 2014. geiger v2.0: an expanded suite of methods

for fitting macroevolutionary models to phylogenetic trees. Bioinformatics 1–3.

Plummer, M., N. Best, K. Cowles, and K. Vines. 2006. CODA: Convergence

Diagnosis and Output Analysis for MCMC. R News 6:7–11.

Pyron, R. A., and F. T. Burbrink. 2014. Early origin of viviparity and multiple

reversions to oviparity in squamate reptiles. Ecol. Lett. 17:13–21.

Qualls, C. P., and R. Shine. 1995. Maternal Body-Volume as a Constraint on

Reproductive Output in Lizards : Evidence from the Evolution of Viviparity.

Oecologia 103:73–78.

Rabosky, D. L. 2010. Extinction rates should not be estimated from molecular

phylogenies. Evolution (N. Y). 64:1816–1824.

Rabosky, D. L., and E. E. Goldberg. 2015. Model inadequacy and mistaken

inferences of trait-dependent speciation. Syst. Biol. 64:340–355.

Revell, L. J. 2014. Ancestral character estimation under the threshold model from

quantitative genetics. Evolution (N. Y). 68:743–759.

Reynolds, J. D., N. B. Goodwin, and R. P. Freckleton. 2002. Evolutionary transitions

in parental care and live bearing in vertebrates. Philos. Trans. R. Soc. Lond. B.

Biol. Sci. 357:269–81.

Rigby, C., and C. a. Simpfendorfer. 2013. Patterns in life history traits of deep-water

chondrichthyans. Deep Sea Res. Part II Top. Stud. Oceanogr. 1–11. Elsevier.

Shine, R. 1995. A New Hypothesis for the Evolution of Viviparity in Reptiles. Am.

Nat. 145:809–823.

Shine, R. 2004. Does viviparity evolve in cold climate reptiles because pregnant

females maintain stable (not high) body temperatures? Evolution 58:1809–1818.

Shine, R. 1978. Propagule size and parental care: The "safe harbor" hypothesis. J.

Theor. Biol. 75:417–424.

Shirai, S. 1992. Squalean Phylogeny: A new framework for "Squaloid" sharks and

their related taxa. Hokkaido University Press, Sapporo, Japan.

Sorenson, L., F. Santini, and M. E. Alfaro. 2014. The effect of habitat on modern

shark diversification. J. Evol. Biol. 27:1536–1548.

Stein, R., C. Mull, T. Kuhn, N. Aschliman, L. Davidson, J. Joy, G. Smith, N. Dulvy,

and A. Mooers. 2018. Global priorities for conserving the evolutionary history of

sharks, rays, and chimaeras. Nat. Ecol. Evol. 2. Springer US.

Trexler, J. C., and D. L. Deangelis. 2003. Resource Allocation in Offspring

Provisioning: An Evaluation of the Conditions Favoring the Evolution of

Matrotrophy. Am. Nat. 162:574–585.

Uyeda, J. C., R. Zenil-Ferguson, and M. W. Pennell. 2017. Rethinking phylogenetic

comparative methods. bioRxiv 222729.

Van Dyke, J. U., O. W. Griffith, and M. B. Thompson. 2014. High food abundance

permits the evolution of placentotrophy: evidence from a placental lizard,

Pseudemoia entrecasteauxii. Am. Nat. 184:198–210.

Weigmann, S. 2016. Annotated checklist of the living sharks, batoids and chimaeras

(Chondrichthyes) of the world, with a focus on biogeographical diversity. J. Fish

Biol. 88:837–1037.

Wourms, J. P. 1981. Viviparity : The Maternal-Fetal Relationship in Fishes. Am.

Zool. 21:473–515.

Wright, A. M., K. M. Lyons, M. C. Brandley, and D. M. Hillis. 2015. Which came

first: The lizard or the egg? Robustness in phylogenetic reconstruction of

ancestral states. J. Exp. Zool. Part B Mol. Dev. Evol. 324:504–516.

Yampolsky, L. Y., and S. M. Scheiner. 1996. Why Larger Offspring at Lower

Temperatures? A Demographic Approach. Am. Nat. 147:86–100.

Yoder, J. B., E. Clancey, S. Des Roches, J. M. Eastman, L. Gentry, W. Godsoe, T. J.

Hagey, D. Jochimsen, B. P. Oswald, J. Robertson, B. A. J. Sarver, J. J. Schenk,

S. F. Spear, and L. J. Harmon. 2010. Ecological opportunity and the origin of

adaptive radiations. J. Evol. Biol. 23:1581–1596.

Zeh, D. W., and J. A. Zeh. 2000. Reproductive mode and speciation: the viviparity-

drive conflict hypothesis. Bioessays 22:938–946.

Figure Legends
Figure 1. Ancestral state reconstruction reproductive mode on a representative tree of
610 species of chondrichthyans. Pie symbols represent the likelihood of the character
state for each node being egg-laying (black), live-bearing (blue), or matrotrophic
(red). Dark grey symbols denote the partitions encompassing diversification rate
shifts. Silhouettes depict representative species from the major orders.
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Figure 2. The number (A) evolutionary transitions in reproductive mode across
chondrichthyans and transition rates between modes in (B) main partition of the tree
and (C) within Carcharhiniformes. Origins of live-bearing from egg-laying are
depicted in blue with reversals in black, and origins of matrotrophy from
lecithotrophic live-bearing are depicted in red with reversals in green. Bars and
shaded regions in represent the 95% posterior density of transition rate estimates.
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Figure 3. Coefficient plots of evolutionary covariation between (A) body size, (b)
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circles denote live-bearing vs matrotrophy, and open circles denote reproductive
mode as an ordinal variable with all three character states. Horizontal bars represent
the 95% confidence intervals of the mean posterior estimate.
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state dependent speciation (λ), extinction (μ), and net diversification rate (r) from the
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order Rajiformes (G-I), and within the family Potamotrygonidae (J-L). Egg-laying is
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represent the mean and 95% confidence interval of the posterior mean estimate.
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diversification rates (r) from the main partition of the tree (A), and within the order
Carcharhiniformes (B). Live-bearing relative to egg-laying is depicted in black,
matrotrophy relative to live-bearing in blue, and matrotrophy relative to egg-laying in
red. Bars and circles represent the mean and 95% confidence interval of the posterior
mean estimate. The vertical lines denote no difference between the diversification
rates for the reproductive modes being compared.
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725   **Figure 1.**

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Figure 3.

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 Figure 5.