

1 **Determinants of trafficking, conduction, and disease within a K⁺ channel**
2 **revealed through multiparametric deep mutational scanning**

3
4 Short Title: Multiparametric Deep Mutational Scanning in the Inward Rectifier Kir2.1

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17
18 **Abstract:**

19 A longstanding goal in protein science and clinical genetics is to develop quantitative models of
20 sequence, structure, and function relationships and delineate the mechanisms by which mutations
21 cause disease. Deep Mutational Scanning (DMS) is a promising strategy to map how amino acids
22 contribute to protein structure and function and to advance clinical variant interpretation. Here, we
23 introduce 7,429 single residue missense mutation into the Inward Rectifier K⁺ channel Kir2.1 and
24 determine how this affects folding, assembly, and trafficking, as well as regulation by allosteric
25 ligands and ion conduction. Our data provides high-resolution information on a cotranslationally-
26 folded biogenic unit, trafficking and quality control signals, and segregated roles of different
27 structural elements in fold-stability and function. We show that Kir2.1 trafficking mutants are
28 underrepresented in variant effect databases, which has implications for clinical practice. By
29 comparing fitness scores with expert-reviewed variant effects, we can predict the pathogenicity
30 of ‘variants of unknown significance’ and disease mechanisms of know pathogenic mutations.
31 Our study in Kir2.1 provides a blueprint for how multiparametric DMS can help us understand the
32 mechanistic basis of genetic disorders and the structure-function relationships of proteins.

33

34 Introduction

35 Inward rectifier K⁺ (Kir) play a central role in physiology by setting, maintaining, and regulating a
36 cell's resting membrane potential ¹. Misregulated Kir cause neurological disorders (alcohol and
37 opiate addiction²⁻⁴, Down's syndrome ⁵, epilepsy ⁶, Parkinsons ⁷), cardiac disorders (Long QT
38 syndrome ⁸), hereditary renal diseases⁹, and diabetes ¹⁰.

39

40 While some mechanisms that underlie disease-causing Kir variants involve aberrant function
41 (gating, ion permeation, ligand binding ¹¹) or transcript processing ¹², disease-causing variants
42 are often linked to trafficking defects. For example, over half the missense mutations investigated
43 in Kir1.1 (ROMK1) affected surface expression, often caused by proteosomal degradation of
44 folding-deficient or mistrafficked channels ^{13,14}. A comparison of mutation hotspots in the Kir C-
45 terminal domain (CTD) revealed that ~60% of variants could be linked to impaired surface
46 expression ^{15,16}. Many mutations linked to neonatal diabetes reduced Kir6.2 surface expression
47 to varying degrees ¹⁷. Beyond known trafficking signals (e.g., ER and Golgi-export signal
48 sequences ¹⁸⁻²³), mutations along the entire Kir primary sequence can disrupt surface expression
49 ¹⁶. Several additional factors control surface expression, such as protein stability, interactions with
50 trafficking partners, and complexes that stabilize channels in the membrane ^{20,24,25}.

51

52 The apparent prevalence of Kir trafficking phenotypes has a significant caveat: phenotypes are
53 experimentally determined for a small fraction of possible or clinically observed variants. Of the
54 8,113 possible missense mutations for KCNJ2/Kir2.1 only 163 are represented in ClinVar— a
55 database of clinically observed variation and phenotypes ²⁶ (**Fig. 1A, Supplementary Fig. 1**).
56 Most studies focus on common natural variants observed in human genomes or exomes. Rare
57 genetic variants vastly outnumber common variants ²⁷ and are implicated in a substantial portion
58 of complex human disease ²⁸. Rarity means that clinical significance (benign or pathogenic) of
59 most variants cannot be established using genome-wide association studies due to low statistical
60 power for calling pathogenic variants. For KCNJ2/Kir2.1 ClinVar reports 144 missense mutations,
61 but most (72%) have “uncertain significance” or conflicting or no interpretation (**Fig. 1A**).
62 Computational algorithms (PolyPhen-2 ²⁹, SIFT ³⁰, EVE ³¹ etc.) are currently filling this gap by
63 predicting the consequence of mutations. However, computational approaches work best in
64 conserved regions of genes and cannot predict mechanism of pathogenicity.

65

66 Despite a central role for Kir trafficking and function in disease etiology, there are no large-scale
67 studies that comprehensively study sequence determinants of K_{IR} trafficking and function.
68 Previous studies focused on a small subset of common natural variants, while systematic variation
69 is needed for a global picture of Kir trafficking and function. Beyond explaining disease, large-
70 scale mutational studies can reveal intrinsic properties of proteins ³²⁻³⁴, determine protein
71 structures ³⁵, and inform mechanistic models of protein structure function relationships ³⁶.

72
73 Here, we combine programmable mutagenesis library generation with multi-parametric
74 sequencing-based assays for surface expression and function of Kir2.1. Our data reveal distinct
75 determinants for Kir trafficking, folding, tetramerization, and function. The phenotype of most
76 clinically observed variants can be explained by impacting function, which may be related to the
77 essential role of Kir2.1 in human physiology. We also find further support for the hypothesis that
78 a hierarchical organization of ion channels structure balances stability and flexibility for folding
79 and function ³⁴.

80
81 **A multi-parametric deep mutational scan of kir2.1**

82 For Kir2.1 to fulfill its roles in cellular physiology, it must be ER targeted, folded, tetramerized, and
83 surface trafficked (**Fig. 1B**). Once trafficked to the cell surface, Kir2.1 must be sensitive to
84 Phosphatidylinositol 4,5-bisphosphate (PIP_2) ^{1,37}, undergo gating associated conformational
85 transitions (pore opening and closing), and selectively conduct K^+ .

86
87 To probe how Kir2.1 mutations affect these processes, we use a programmable deep
88 mutagenesis approach (SPINE ^{38,39}) and mutated residues 2-392 of mouse Kir2.1 (Uniprot
89 P35561) to every other amino acid (**Fig. 1C**). We included synonymous mutations in 20 Kir2.1
90 positions as an internal standard to determine wildtype fitness. From this DMS library, we
91 generated a stable single copy library cell line using BxB1-mediated recombination ⁴⁰ for stringent
92 genotype-phenotype linkage. Endogenous channel currents in HEK293 are negligible when
93 compared to those of exogenous channel driven from a strong constitutive promoter (CAG) ⁴¹.

94
95 Since mutations can affect any of the abovementioned processes, multi-parametric assessment
96 of phenotypes is required to develop a comprehensive mechanistic understanding of Kir2.1
97 variation. We focused on separating a mutations effect on secretion, folding, and trafficking
98 ('surface expression') and allosteric regulation and ion conductance ('function'). Considering the

99 library size of variants (391x19 = 7,429 variants), assays that assess surface-expression and
100 function must be high-throughput. Cell-based assays coupled to fluorescently activated cell
101 sorting that can separate cell population (and the genotypes encoding in population) based on a
102 phenotype reported by fluorescent reporters are immensely flexible.

103

104 Mutations that interfere with secretion, folding, or trafficking will cause reduced Kir2.1 surface
105 expression ⁴²⁻⁴⁴. To measure Kir2.1 surface expression in cells we inserted an epitope tag into
106 the extracellular loop (T115 ⁴⁵). Cells with surface trafficked variants can be separated from poorly
107 expressed variants based on antibody-labeling and fluorescence-activated cell sorting (FACS)
108 (**Supplementary Fig. 2**). Phenotypes can be linked to genotype by NGS. We've extensively used
109 this method in FACS-based genotype-phenotype assays ^{34,38,46}.

110

111 Functional Kir2.1 hyperpolarizes HEK293 cells by driving the resting membrane potential (RMP)
112 towards the reversal potential of K⁺ (-80mV) while non-functional Kir2.1 variants do not affect the
113 RMP (-35mV). We can measure RMP with voltage sensitive dyes, such as the FLIPR Blue, a
114 concept that we and others demonstrated in optical Kir functional assays ^{46,47}. Changes in
115 membrane voltage alter dye membrane partitioning, and therefore extinction coefficient and
116 fluorescence. Hyperpolarized cells expressing functional Kir2.1 can be separated from more
117 depolarized cells expressing non-functional Kir2.1 by FACS using FLIPR dye fluorescence
118 (**Supplementary Fig. 3**). As with surface expression, phenotype can be linked to genotype by
119 NGS.

120

121 **A global view of trafficking determinants**

122 To learn how amino acids contribute to and mutations alter Kir2.1 trafficking and folding, we sorted
123 and sequenced the Kir2.1 DMS library based on surface expression by FACS-seq. Surface
124 trafficking fitness was determined for 6,898 Kir2.1 variants (93%). Biological replicates for
125 subpools were highly correlated (Pearson correlations 0.85-0.94, **Supplementary Fig. 4**) and
126 shows read depth is excellent (greater than 30-fold at most positions, **Supplementary Fig. 5**;
127 **Supplementary Table 1A**). Surface expression fitness scores and standard errors were
128 calculated using Enrich2 ⁴⁸. Enrich2 calculates log enrichment by fitting a weighted regression
129 based on changes in mutation frequency across samples within an experiment. Median fitness
130 error relative to measurement range was low (7.5%) in the surface expression assay based on
131 synonymous mutation standard deviations (**Fig. 2A, Supplementary Fig. 6**). Surface fitness

132 follows a bimodal distribution with one population of mutations centered at wildtype fitness and
133 the other population strongly decreasing surface expression (**Fig. 2A**).

134
135 As expected, mutations are highly deleterious within the Flag-tag used for surface labeling and
136 known trafficking signals (**Fig. 2B**). Substituting aromatic residues with charged residues strongly
137 decreases surface expression within the ⁸¹WRWMLLLL^{F88} motif that mediate interaction with
138 Caveolin-3 ⁴⁹. Glutamates within the di-acidic ER export signal, ³⁸²FCYENE³⁸⁷ ⁵⁰, are quite
139 sensitive to mutations with the second glutamate being far more sensitive. Interestingly,
140 negatively charged mutations, N386E or N386D, between the di-acidic motif increase surface
141 expression.

142
143 A Golgi export signal comprised of two residues patches at Kir2.1's N- and C-termini interacts
144 with the AP1 adaptin complex to target Kir2.1 to clathrin-coated vesicles for Golgi to plasma
145 membrane trafficking ²⁵. Others have used alanine mutagenesis screens to identify sequence
146 determinants for Golgi export. First, Ma et al. ²⁵ defined the minimal signal motif comprised of
147 hydrophobic residues (I321, W323, Y316) and a juxtaposed electrostatic interaction between R46
148 and E327. Later, Li et al. ¹⁸ included salt bridges between basic residues in the N-terminus (R44,
149 R46, K50) and C-terminus (E3201, E327), and hydrophobic residues (F203, L239, I240, F300).
150 This structured trafficking signal is formed when the CTD has folded correctly to include a
151 hydrophobic cleft, an adjacent cluster of basic residues, and a network of interactions between
152 termini and adjacent subunits. Ma et al. suggested this structured trafficking signal predisposes
153 Golgi exit on correct protein conformation; our comprehensive mutagenesis suggests this quality
154 control mechanism may extend beyond previously identified sequences. It appears to involve all
155 core beta sheet in the CTD (β D1, β H, β I, β J, β K, β B2, β C, β G) and helices α G and α F (**Fig.**
156 **2B,C**). Across CTD secondary structure elements, we find a distributed network of residues that,
157 when mutated, result in reduced surface expression (**Fig. 2D**). Changes to the packing of the
158 hydrophobic core of the CTD will likely impact the Golgi export motif orientation. We hypothesize
159 that the Golgi export signal is a reporter for folding of the entire CTD. Several lines of evidence
160 support this hypothesis: IgG-like domains are a beta sheet sandwich with conserved hydrophobic
161 residues at sheet interfaces that are crucial for fold stability ^{51,52}, computational mutagenesis in
162 Kir1.1(ROMK1) showed these residues are crucial for fold stability, and experimental
163 mutagenesis found mutations strongly reduces inward rectifier surface expression ^{15,53}. Residues
164 in this network belong to the same monomer; disrupting interactions between residues belonging

165 to neighboring monomers (e.g., carbon- δ and δ - δ interactions between F45, L246, Y338; **Fig.**
166 **2B,E**) do not uniformly reduce surface expression. Although Kir are presumably already
167 assembled into tetramers⁵⁴, our data suggests the conformational checkpoint for Golgi export
168 requires properly folded monomers not correctly assembled tetramers.

169
170 Sites enriched for beneficial or deleterious substitutions could point to additional mechanisms for
171 Kir2.1 trafficking control. Along those lines, we observe that substitution of N- and C terminal
172 residues (G2–G31, S377-S381) with polar mutations have neutral trafficking phenotypes, while
173 aromatic or hydrophobic mutations tend to decrease surface expression. While we do not know
174 the underlying mechanism, our data suggests disordered N- and C-termini are important in
175 targeting Kir2.1 to the cell surface.

176
177 Mutations along β B2(M211–K215), β DE(S242–P252), β K(R333, P336), β L(F339), β M(Y345),
178 β N(Y366), and α F helix(K372–L376) tend to improve surface-trafficking above WT especially
179 when aromatic residues are replaced with any non-aromatic amino acids (**Fig. 3A**, t test p-value
180 0.017). These sites form a patch on the outer face of the CTD domain where two subunits
181 interdigitate (β DE from one subunit, the remaining elements from another subunit; **Fig. 3B**). In
182 this region beneficial mutations have no clear physicochemical preferences beyond non-
183 aromatics. While no known trafficking signal matches, aromatic residues are often binding
184 interfaces^{49,55}. Perhaps mutations alter trafficking patterns by disrupting protein interactions (e.g.,
185 reduce ER retention, reduce forwarding to lysosomal recycling).

186
187 The agreement with earlier trafficking studies in Kir2.1 shows that our high-throughput surface
188 expression assay works. This allows us to establish a global view of sequence/trafficking
189 relationships in Kir2.1. Comprehensive mutagenesis allows us to validate and fill in the gaps on
190 existing models of channel trafficking^{18,25} while discovering new potential trafficking determinants.

191

192 **A cotranslationally-folded biogenic unit in Kir2.1**

193 By examining structured regions sensitive to mutations, we identify determinants of Kir2.1 folding.
194 In the pore domain, we find the lower halves of M1 and M2 are surprisingly tolerant to mutation,
195 in particular residues whose sidechains project towards the pore cavity. At the base of the pore
196 domain, mutations are highly deleterious within a cluster of hydrophobic residues (W81, L85, I178,
197 F182, I187, M191) that form inter-monomer interactions between M1 and M2 helices. The top of

198 the pore domain—a region comprised the upper halves of M1, M2, pore helix, and selectivity filter—
199 is extremely intolerant to substitutions. This includes the M2-pore helix loop (e.g., H110–E133),
200 whose amino acid sidechains contribute to tight packing centered around a hydrophobic pocket
201 formed by F103 (M1), F143 (pore helix), V169 (M2), and V131 M1-pore helix loop (**Fig. 3C**). The
202 same region contains two highly conserved cysteines (C130, C162) that form a disulfide bond
203 which is critical for folding, not function (K⁺ conductance)^{56,57,57,58}. Others found P156, at M2's
204 apex, is essential for efficient surface trafficking in Kir2.2⁵⁹. In agreement with folding critical roles,
205 all substitutions in these positions are deleterious to surface expression.

206
207 Glycines in the selectivity filter (¹⁴⁷IQTTIGYG¹⁵⁴ in Kir2.1; conserved residues are underlined),
208 which are absolutely conserved in all K⁺ channels, are immutable. Because glycine is achiral, it is
209 the only canonical amino acid that can satisfy the unusual conformation of the selectivity filter
210 main chain, which alternates between left-handed and right-handed α -helical dihedral angles⁶⁰.
211 Substitutions of either T150 (which occupies unfavorable backbone torsion angle conformation in
212 the Kir2.2. crystal structure^{37,56}) or Y153 (which forms π - π interaction to a neighboring subunit)
213 in the selectivity filter with glycine significantly increase surface expression. This suggests the
214 peculiar conformational requirements of the selectivity filter are a constraint in channel biogenesis.

215
216 Based on the extreme mutational sensitivity, high contact density of side chain packing in the
217 Kir2.2 crystal structure, and known disulfide-mediated fold stabilization, we propose that the top
218 halves of M1 & M2, pore helix, and the M2-pore helix loop promote tertiary structure formation
219 within nascent Kir2.1 monomers (perhaps in the ER translocon) to establish correct topology and
220 promote membrane insertion. This may be a general feature of K⁺ channel folding, as a similar
221 “cotranslationally folded biogenic unit” was proposed for Kv1.3, in which S5/pore loop/S6 interact
222 to establish a “reentrant pore architecture” and correct topology early in channel biogenesis
223 without requiring tetramerization^{61,62}. Interestingly, residues from neighboring subunits interact
224 with this putative biogenic unit in the fully assembled channel (e.g., selectivity filter residues Y153,
225 which forms a δ - δ interaction with F143 and F155, carbon- δ interaction with E133) are relatively
226 tolerant to mutations. This supports the idea that early trafficking-critical folding events involve
227 only intra-monomer interactions.

228

229 **Regions involved in gating transitions to do not contribute to fold stability**

230 From the perspective of surface-trafficking as the measured phenotype, several regions of Kir2.1
231 are tolerant to mutations. This includes the interface between pore and CTD (comprised of slide
232 helix $\alpha A/\alpha B$, inner helix gate (I183–M188), and tether helix αF), the G-loop ($\beta H-\beta I$ loop), and the
233 residues lining the CTD cavity (loops between $\beta C-\beta D$ and $\beta E-\beta G$) at the interface between
234 different subunits (**Fig. 3D**). Interestingly, these are involved in gating associated conformational
235 changes^{37,63} induced by the binding of PIP₂, Kir2.1 allosteric activator (two-sided Fisher's Exact
236 Test, p-value 8.091e-05, odds ratio 2.47). An explanation for high mutational tolerance in gating
237 structural elements is that they have higher conformational flexibility required for interconversion
238 between different gating states. They are therefore unlikely to contain motifs recognized by
239 folding-based quality control mechanisms.

240

241 Kir2.1 appears to have an internal organization into contiguous regions involved in fold-stability
242 (biogenic folding unit, IgG-like CTD) and gating transitions (TM/CTD interface, CTD subunit
243 interfaces) with distinct mutational tolerance. To understand the relationship between folding and
244 function, we assayed K⁺ conductance as a second phenotype for the DMS library.

245

246 **Functional phenotype assays map molecular determinants of conduction and PIP₂** 247 **sensitization**

248 To assay variant function, we sorted the Kir2.1 DMS library (in duplicate, on separate days) into
249 based on voltage-sensitive dye fluorescence, sequenced, and calculated fitness using the same
250 Enrich2 package. This assay is based on premise that Kir2.1 decreases RMP resulting in
251 decreased fluorescence of the FLIPR dye. Functional fitness was determined for 6,944 Kir2.1
252 variants (93.4%), with similar replicates (Pearson correlations 0.86-0.93, **Supplementary Fig. 7**)
253 and excellent read depth (greater than 30-fold at most positions, **Supplementary Fig. 8**;
254 **Supplementary Table 1B**). Based on synonymous mutation standard deviations (**Fig. 4A**),
255 median fitness error was low (15.3%). Unlike the bimodal distribution for surface-expression
256 fitness, functional fitness is unimodal, with most mutations neutral (**Fig. 4A**); there is no distinct
257 low function population likely due to lower dynamic range of the voltage sensitive dye compared
258 to fluorescent antibody labelling (**Supplementary Figs. 2-3**). Function fitness is the combination
259 of surface expression (number of channels on surface) and individual channel functional
260 properties (open probability, single channel conductance, K⁺ selectivity, etc.). Furthermore, as our
261 assay is measuring steady-state RMP and even partially functional Kir2.1 will eventually

262 hyperpolarize the cell. This means we must interpret functional fitness in context of surface
263 expression fitness.

264

265 Many variants in the pore domain have low functional fitness scores (e.g., pore helix, selectivity
266 filter, **Fig. 4B**). For example, Y153 mutations, which have relatively tolerant surface fitness,
267 strongly reduces function (**Fig. 4C**). Y153 interacts through π - π interactions with F143 and F155
268 on a neighboring subunit and mutations may preclude correct selectivity filter geometry to conduct
269 K^+ ions. The conservative substitution Y153F has a neutral phenotype, in agreement with earlier
270 electrophysiology studies ^{64,65}. Many positions along the pore helix that form a hydrogen bond
271 network to support a conductive and K^+ selective state ⁶⁶ are very sensitive to mutation. Mutations
272 of conserved hydrophobic residues in the CTD core beta sheet (e.g., W220) and the Golgi export
273 signal (e.g., hydrophobic cleft residues Y316, W323, and salt bridge R45, E327) are deleterious
274 as predicted by their effect on surface-expression.

275

276 Disordered N and C termini including the ER export signal FCYENE had moderately low surface-
277 expression fitness but their functional fitness is mostly near neutral (**Fig. 4D,F**). This suggests
278 mutations within the disordered regions only impact trafficking of otherwise properly folded and
279 functional channel. Mutations within the unstructured termini they are deleterious to surface-
280 expression and but neutral to individual channel function.

281

282 PIP_2 binding at the TM/CTD interface induces a disorder-to-order transition in the tether helix,
283 contracting it and thereby translating the CTD toward the pore domain. The engagement between
284 and CTD and pore domain allows the G-loop to wedge into the pore domain and the inner helix
285 activation gate to open ³⁷. PIP_2 's binding site includes pore domain residues (R80, W81, R82,
286 K190) and CTD residues (K186/193, K188/195, K189/196, R65, R219/226, R197). As expected,
287 surface-expression fitness was neutral and functional fitness was negative (**Fig. 4E,G**). By
288 focusing on mutation-sensitive positions, we can trace secondary structural elements that couple
289 the PIP_2 binding site to the Kir2.1's gate, the G-loop ⁶⁷. We find residues that directly interact with
290 PIP_2 in the pore domain (R80, W81, R82) and tether helix (K190, K193, K195, K196). We find
291 additional residues in the tether helix (R197), β B1 (L202), and interfacial helix (R65) interact with
292 the allosteric coupling β C- β D regions. Residues are highly sensitive in the entire β C- β D and its
293 interactions including a hydrophobic interface between β C and β I with each subunit, several

294 interactions couple to the β CD loop (e.g., H229) and β EG loop (e.g., F270) of one subunit to the
295 β H (I305), β I (R320), and the G-loop gate of another subunit (**Fig. 4H**).

296

297 Taken together, deep mutational scanning reinforces a central role of the β CD loop propagating
298 PIP₂ binding at the TM/CTD interface to the G-loop gate. Molecular dynamics simulations
299 uncovered an extended network of interactions between N-terminus, tether helix, CD loop keep
300 the G-loop gate in a closed state⁶³. Electrophysiology demonstrated that mutations in R219,
301 which is conserved in the K_{IR} family, attenuate PIP₂ affinity⁶⁸ and causes Anderson Tawill
302 Syndrome⁶⁹. Mutating the corresponding residues in GIRK2/Kir3.2 (R201A) constitutively
303 activates channels, presumably by forcing the β CD loop into conformation that mimics the G-
304 protein activated state⁷⁰.

305

306 In GIRK2, an aspartic acid in β CD (D228) is a key residue in mediating regulation by intracellular
307 sodium (an important physiological role of GIRK2). This Na⁺ binding site propagates
308 conformational changes in the CTD's β strands after G β γ binds to β DE and β LM loops, displacing
309 the β LM loop upward to interact with the N-terminus and β CD loop, ultimately stabilizing the open
310 conformational of the G loop gate^{71,72}. The corresponding wildtype amino acid in Kir2.1, which is
311 not regulated by Na⁺, is N224 (**Fig. 4B**). Interestingly, substitution with aspartate increases Kir2.1
312 function, perhaps by recapitulating the Na⁺ binding site, which also involves coordination from
313 main chain carbonyls (R226, K227, S228). The function increasing effect of N224K (and the
314 deleterious effect of N224R) could be explained through the Na⁺-independent stabilization of the
315 β CD loop through hydrogen bonding between the protonated epsilon-amino group of lysine
316 (mimicking Na⁺) and the abovementioned carbonyl oxygens.

317

318 Another striking feature revealed by functional assays are function-increasing mutations near the
319 helix bundle crossing gate of Kir2.1 (**Fig. 5A**). Previous studies suggested a kink occurring at
320 G176 in TM2 widens the pore at the helix bundle crossing (HBC) allowing for K⁺ to pass⁷³⁻⁷⁵. As
321 expected, mutating G176 decreased function fitness. Substitution in residues below the kink-
322 inducing G176 and above the actual bundle crossing (V187), increases functional fitness. This
323 effect is strongest in A173, I175, I176, G177 (corresponding to S174, M176, I177, G178 in Kir2.2;
324 **Fig. 5B**). Together, residues form a cuff above the HBC gate coupling TM2 from different subunits.
325 The cuff may stabilize the unkinked conformation of TM2 to decrease open probability. Consistent
326 with this idea, mutating cuff residues to large charged amino acids has the greatest function-

327 increasing effect (**Fig. 4C**). Perhaps mutating to large, charged residues disrupts nearby
328 hydrophobic interactions networks that stabilize the HBC close state. The HBC gate is also
329 stabilized by hydrophobic contacts to the TM1 of an adjacent subunit^{76,77}. Consistent with TM1
330 interactions forming a gating energy barrier, mutating interacting TM1 residues (F88; F86 in
331 Kir2.2) also increases function fitness.

332

333 To identify regions that are important for function and not surface expression, we z-scored and
334 subtracted surface expression from function fitness. We used t-tests with pooled variance
335 estimated from all mutations to test significance of difference in fitness and function scores for
336 each position (**Fig. 5C**). For many positions, surface scores were significantly lower than function
337 scores (**Fig. 5C, cyan circles**). This subset includes the FLAG tag that is required for surface
338 expression assay, but also Golgi export motifs, transmembrane helices, and CTD core beta
339 sheets. For another subset of positions, function scores were significantly lower than surface-
340 expression scores (**Fig. 5C, magenta circles**). This includes sites involved in gating (PIP₂ binding,
341 G-loop gate), and K⁺ conduction (selectivity filter). By making a binary assignment (based on p-
342 value) whether a position belongs to one subset or the other and mapping this assignment onto
343 the Kir2.2 structure, we again find structural organization around elements that are important for
344 fold stability (TM, CTD beta sheets) and function (PIP₂ sensitization, gating, K⁺ conduction) (**Fig.**
345 **5D**). This distinct organization is clearest in the pore domain, where all interactions within a
346 subunit are driving surface expression score (**Fig. 5E, cyan sticks**), while residues mediating
347 subunit interactions (which is required for stability of the selectivity filter) are driving function score
348 (**Fig. 5E, magenta sticks**). Other residues that drive function scores are distributed along an
349 interaction network that connects the PIP₂ binding site (M2, α F) with the slide helix (α A/B), β CD
350 loop, β EG loop, and the G-loop gate (β HI loop) (**Fig. 5F**). Intriguingly, this network involves
351 interfaces between adjacent subunits (N-terminus \leftrightarrow β LM loop interface, β I \leftrightarrow β CD loop,
352 β D \leftrightarrow β EG loop). Subunit interactions determining gating is consistent with recent crystal
353 structures of a forced-opened Kir2.2 in which inner helix gate opening requires CTD subunits to
354 move relative to each other⁷⁷.

355

356 Overall, the functional assay allows us to validate and test existing models of Kir2.1 gating while
357 also providing further evidence that inter-subunit interactions serve as an energy barrier to bias
358 Kir2.1's conformational ensemble towards closed states.

359

360 **Most pathogenic inward rectifier mutations are clustered within functionally important**
361 **residues**

362 A central goal in molecular genetics is identifying the mechanistic basis by which mutations cause
363 disease. Multiparametric DMS studies that interrogate multiple phenotypes associated with
364 variants are a promising strategy to answer this question. Based on the premise that most
365 mutation affect protein stability, approaches such as Vamp-Seq ⁷⁸ were developed as
366 generalizable measures of protein abundance. In cases such as Kir2.1, where proper trafficking,
367 localization, and gating are crucial for producing functioning proteins, measuring abundance likely
368 won't work. It is necessary to differentiate between mutations' effects on folding, trafficking, and
369 gating to learn the mechanism for how genotype affects Kir2.1 phenotype. To learn potential
370 pathogenicity and mechanism of action for reported disease-associated mutations, we compared
371 the surface expression and fitness data to clinically observed mutations.

372
373 Since the overall domain architecture and structure/function relationships are conserved within
374 the inward rectifier family, we reasoned that the mechanisms underlying variant effects are
375 conserved, as well. We therefore gathered missense variant effects reported in Clinvar and
376 dbSNP for inward rectifiers. By aligning all human Kir, we assigned the corresponding Kir2.1
377 position to each variant (total variant count: 613) and noted whether wildtype amino acid matches
378 between Kir2.1 and the aligned Kir (**Supplemental Table 2**). To test if variants in Clinvar or
379 dbSNP is related to surface-trafficking or function, we compared the empirical cumulative
380 distribution functions of trafficking and function scores variants in databases vs. those that are
381 not. They differed for trafficking scores but not for function scores (two-sided Kolmogorov-Smirnov
382 test p-values 4.28×10^{-8} and 0.51, respectively; **Fig. 6A**). This means being listed in genetic
383 variation databases is related to Kir2.1 function and less related to trafficking. Variants with low
384 surface scores in our DMS, are underrepresented in databases. A likely explanation is that Kir2.1
385 is essential for normal physiology and therefore under strong selection. Indeed, Kir2.1
386 homozygous knockout mice die 8 to 12 hours after birth ⁷⁹. Kir2.1's missense constraint score,
387 based on exome and whole-genomes sequencing deposited in gnomAD ⁸⁰, is 0.5 (i.e., 121
388 observed vs. 240 expected missense mutations based on background mutation rates). Variants
389 that misregulate surface trafficking likely are more deleterious (e.g., abolishing all K⁺
390 conductance) than variants affecting function (e.g., gating kinetics), which is why they are
391 extremely rare in the living human population. Consistent with this idea, exome and targeted
392 sequencing in products of conception showed a significant enrichment of pathogenic variants

393 associated with cardiac channelopathies in stillbirths ⁸¹. Furthermore, only one variant (W232C)
394 of the 5 most surface-trafficking impairing positions on Kir2.1's Golgi export motif (R44, S315,
395 E320, W323, G333) is reported in Clinvar.

396

397 The correspondence between variant listing and Inward Rectifier function is also apparent when
398 we annotate mean surface-trafficking and function scores with pathogenic variants (**Fig. 6C**).
399 Across the board, hotspots enriched for pathogenic disease-associated mutation have low
400 function scores, whereas their surface-expression scores are more varied. Variants of uncertain
401 significance followed this trend. In addition to regions involved in PIP₂-binding (α A/B slide helix,
402 lower part of M1, α F), and propagating conformational changes to the G-loop gate (β B1, β CD,
403 β HI loop), pore helix and selectivity filter are also hotspots for pathogenic variants. A notable
404 exception to down-trending function in mutational hotspots is a cluster of disease-associated
405 variants near the intracellular end of M2 (D180-A186). Interestingly, many Clinvar-reported
406 variants (e.g., I183L) are associated with Short QT syndrome, a disease that can be caused by
407 gain of function mutations in KNCJ2(Kir2.1) ⁸². Consistent with this clinical phenotype, many
408 mutations increase function (compared to wildtype Kir2.1). Based on known structure/function
409 relationship in inward rectifiers, we speculate that substitutions increase function by increasing
410 open probability of Kir2.1 by disrupting the inner helix gate (I183–M188).

411

412 We further divided fitness scores by assigned clinical significance to estimate the fitness score
413 bounds of expert-reviewed benign and pathogenic variants (**Fig. 6D,E**). For surface fitness, we
414 find that >97.5% of known benign variant have a fitness score > -0.43. With this lower bound
415 assumption for benign variants, we predict that any 'variant of unknown significance' (VUS)
416 scoring lower will be pathogenic with a loss of surface expression phenotype. Conversely, the
417 upper surface fitness score bound (10th percentile) for expert reviewed benign was 0.95, and VUS
418 with greater surface scores are predicted to have a gain of surface expression pathogenic
419 phenotype. We applied the same reasoning to function fitness scores. Using this estimate, we
420 can assign a predicted pathogenic phenotype (loss/gain of surface expression/function) to 69 of
421 370 (18%) Kir2.1 VUS (**Supplemental Table 2**). The primary sequence location of these
422 predicted pathogenic variants is shown in **Fig. 6C**. Many predicted loss of function variants
423 localize to the slide & tether helices, G-loop gate, and selectivity filter/pore helix. Gain of surface
424 expression variants map to the β BC and β K loop near the interface to β DE of a neighboring
425 subunit. The putative biogenic folding unit (top of M1 & M2, pore helix) contains several predicted

426 loss of surface expression variants. Interestingly, no expert-reviewed pathogenic variants map to
427 this region, which suggests that DMS-based prediction can identify new mutation hotspots.

428 We can also use estimated bounds of benign variants to predict the mechanism through which
429 27 out of 144 (18.8%) expert-reviewed pathogenic variants cause Kir2.1 disorders. Of those, eight
430 pathogenic variants have a predicted loss of surface expression while 19 are predicted loss of
431 function. In line with function-impairing variant being overrepresented in ClinVar, the odds ratio
432 between predicted loss of function phenotype of known pathogenic variants vs. all predicted
433 pathogenic variants is significantly different (two-sided Fisher's exact test, p-value = 0.003144).

434
435 Taken together, we find variants with deleterious surface-score are underreported in available
436 database, which we propose is due to the essential nature of Kir2.1 in human physiology. Using
437 surface and function scores for expert-assigned variant, we can estimate bounds for benign
438 variants, which then allows us to predict VUS that are above or below these bound to the be gain
439 or loss of surface expression/function, respectively. We can also predict the mechanism of action
440 through which known pathogenic mutations cause Kir2.1 disorders. This suggests that DMS-
441 derived surface-trafficking and function scores can be useful to predict pathogenicity and
442 underlying mechanism of inherited and de-novo mutations. Because of the conserved architecture
443 of the Inward Rectifier family, assignment of mutation hotspots may extend to the entire inward
444 rectifier family. Further corresponding DMS studies in other members of the inward rectifier family
445 are required to separate generalize themes from idiosyncrasies. For example, Kir6.2 requires
446 binding to the ABC-transporter SUR1 to be expressed to the cell surface and function. This means
447 that assay-reported phenotypes for a subset of variation at the Kir6.2/SUR1 interface (involving
448 α A/B slide helix, M1, β LM sheets) have greater bearing in Kir6.2-linked diseases (e.g., diabetes
449 mellitus). The methods described here are a blueprint for these studies.

450

451 **Discussion**

452 Deep Mutational Scanning (DMS) quantitatively links protein phenotypes to the genetic variation
453 within the protein's coding sequence. This style of systematic amino acid substitution can reveal
454 protein function^{83–85}, determine protein structure⁸⁶, and explain protein behavior in healthy and
455 diseased cellular contexts⁸⁷. Because protein don't exist in isolation –within the context of a cell
456 folding, trafficking, and function states can all be modified through protein interaction (e.g.,
457 chaperones) or ligand binding (e.g., allosteric modulators)– the phenotypic outcome of genetic
458 variation can be multi-faceted. To start to explore the numerous impacts of mutations, we

459 measure two phenotypes of Kir2.1 variation: surface-trafficking K⁺ conduction. Both assays are
460 based on cell sorting mediated by fluorescent signals (antibody binding to an extracellular loop
461 and voltage sensor dye). By measuring how variants are enriched or depleted, we assign
462 quantitative fitness scores. Many other assay types are compatible with this approach, providing
463 opportunities for even richer phenotypic description of ion channel variation. For example, a
464 recently described approach, HiLITR⁸⁸, could provide more granular resolution about trafficking
465 motifs and Kir2.1 localization in living cells. Spontaneous spiking HEK cells^{89,90}, which co-express
466 Kir2.1, a voltage-dependent Na⁺ channel (Nav1.7), the channelrhodopsin CheRiff, and genetically
467 encoded voltage indicator QuasAr2, could be adapted to evaluate how ion channel expression
468 levels and gating properties impact excitability⁹¹. When assays are conducted with chemical
469 screening, they may aid in the discovery of novel allosteric modulators, state-dependent blockers,
470 and molecular chaperones to precisely treat channelopathies based on genotype. Integration of
471 disparate assays into a common framework of protein variation and its role in structure and
472 function will be challenging but efforts to standardize reporting and unified statistical frameworks
473 for interpretation (e.g., Enrich2⁴⁸, the Atlas of Variant Effects⁹²) are on the horizon.

474

475 Many of our findings with respect to mutational sensitivity of trafficking motifs, core structural
476 elements, and regions of Kir2.1 involved in gating align with existing knowledge and biophysical
477 intuition. Unlike previous more limited screens and intuition, our data is quantitative and enables
478 data-driven approaches to construct global biophysical models of Kir2.1 structure and function.
479 For example, the Golgi export signal comprised of two patches at the N-terminus and CTD was
480 first described as a minimal set of hydrophobic residues along a CTD cleft and juxtaposed basic
481 residues²⁵ and later expanded to adjacent sites¹⁸. Our comprehensive screen shows this
482 trafficking motifs may indirectly probe the correct folding of the entire β -sheet core of the CTD. It
483 reveals scope of what this trafficking motif is reporting to the cellular quality control machinery;
484 not simply the proximity of a few basic and hydrophobic residues, but proper conformation of the
485 entire CTD and assembly into tetramers.

486

487 We observed contiguous regions with neutral expression fitness involved in gating transitions
488 while regions involved in fold-stability were highly sensitive to mutations. Looking at the same
489 regions in functional data, we find the inverse. This suggests two things. First, comprehensive
490 assessment of Kir2.1's phenotypes after perturbation (i.e., mutation) is a high-throughput method
491 to annotate protein sequences into classes linked to specific functions (e.g., putative trafficking

492 signals, folding units, etc.). Conceptually this is similar to other high-throughput biochemical
493 approaches that probe sequence-function relationships, such as circular permutation profiling⁹³
494 or high-throughput enzyme variant kinetics measurements⁹⁴. Second, combining multiple
495 assayed parameters is key to discover general organizational principles in Kir2.1, specifically two
496 distinct structural regions with distinct roles in providing fold stability or dynamics required for
497 gating transitions. Expanding to more measured phenotypes and integrating datasets may be the
498 blueprint for ‘sequencing-based’ biophysics that probes protein function, folding, and dynamics
499 through steady-state biochemical experiments.

500
501 Our DMS studies provide additional context for the mechanistic basis of structure/function relation
502 (e.g., reaffirming the β CD loop’s role in propagating PIP₂ binding at the TM/CTD interface to the
503 G-loop gate). We find a biogenic folding unit that represents an early quality control step of
504 reentrant pore architecture and correct topology likely while the monomer is in the translocon. A
505 similar biogenic unit was described in voltage-dependent K⁺ channels, where it is stabilized by an
506 extensive network of interactions⁶¹. Furthermore, many ion channels have structurally
507 homologous reentrant pore loop architectures and earlier studies suggested that the presence of
508 an ‘aromatic cuff’ is a general feature of their biogenesis^{61,95}. DMS studies may provide a path to
509 probe the energetics and biophysics of stabilizing interactions and to test the hypothesis that
510 hydrophobicity is a general stabilizing factor across reentrant pore loop architectures.

511
512 DMS may also shed light on how subunit interactions determine inward rectifier gating properties.
513 We find mutations near the interface of subunits strongly increase Kir2.1 function. This is
514 consistent with subunits roles in setting channel gating properties. Single channel patch
515 electrophysiology of Kir2.1/2.2 heterotetramers showed that addition of a Kir2.2 monomer
516 increases single channel conductance and decreases the open dwell time (τ_{open})⁹⁶. Perhaps this
517 feature contributes to the observed differences in gating between inward rectifier homo- and
518 heterotetramer, which differ in number and nature of subunit interactions.

519
520 Our finding that variants with deleterious surface-score are underreported in available database
521 supports the emerging theme that many disease-causing variants are linked to trafficking defects
522^{13–16,16–23}. Other large scale mutational analysis, as undertaken for the voltage-dependent K⁺
523 channel Kv11.1^{97,98}, has similarly shown that 88% of Long QT-linked variants have trafficking-

524 deficient mechanisms. They also demonstrate that data-driven approaches outperform smaller,
525 more limited studies that predicted normal trafficking for most mutants ⁹⁹.

526 Underrepresentation in variant databases has implications for clinical practice, since standards
527 for the interpretation of sequence variants are heavily focused on null variants (nonsense,
528 frameshift), population frequency (frequent variants are likely benign), predicted functional effect
529 (is the variant in an important domain), and case evidence ¹⁰⁰. In genes under strong purifying
530 selection, such as KCNJ2/Kir2.1, missense mutations cause severe developmental defects
531 (craniofacial structures, limb development ^{79,101}) likely contribute to or cause spontaneous
532 abortions and pregnancy loss. These variants therefore never enter population- or clinical-
533 associated variant databases. DMS studies are not limited by organismal viability to probe the
534 structural and functional consequences of genetic variation. DMS could be useful to identify
535 drivers of spontaneous abortions and the underlying mechanisms of recurrent pregnancy loss.

536 By comparing our experimentally determined fitness scores to expert-reviewed assignment of
537 variants effects, which are often based on smaller scale direct biochemical studies, we can
538 estimate the bounds of fitness scores for benign and pathogenic variants. This allows us to make
539 predictions about pathogenicity and mechanisms of action for variants of unknown significance
540 (VUS) and known pathogenic variants, closing the loop between high-throughput assays,
541 biophysical mechanisms underlying fitness scores, and clinical interpretation of human variation
542 in ion channel genes.

543 These examples demonstrate that deep mutational scanning of Kir2.1 combined with two simple
544 readouts, provides detailed insight into the mechanistic basis of ion channel folding, function
545 under steady state conditions, and how these processes become misregulated in disease.

546

547 **Materials and Methods**

548 *Kir2.1 DMS Library Generation:* Into mouse Kir2.1 (Uniprot P35561), we introduced a FLAG tag
549 into an extracellular loop (at position T115) and added a downstream expression marker
550 miRFP670 co-expressed via a P2A sequence. For Kir2.1 residues 2-391, each wildtype amino
551 acid was mutated to all other 19 amino acids weighted by their codon usage frequency in humans.
552 In addition to these missense mutations, mutations synonymous to wildtype were included for 20
553 positions as a benchmark. We included missense mutation into the FLAG as a negative control.
554 We generated this mouse Kir2.1 DMS library using the SPINE ³⁸, which we briefly summarize as
555 follows: SPINE based libraries employ synthesized pools of DNA oligos with mutations at each
556 position of a gene. Due to high error-rates in oligo synthesis, the current maximum length for

557 oligos is 230 base pairs. As most genes are longer than 230 base pairs, we break up our gene (8
558 section in the case of Kir2.1) and replace a subsection of the gene with a pool of mutated oligos.
559 The oligos are designed with unique barcodes for amplifying out a specific subpools library,
560 Golden Gate compatible BsmBI cut sites, and the mutation within a subsection of Kir2.1. OLS
561 oligos were designed using the SPINE scripts on Github (<https://github.com/schmidt-lab/spine>).
562 These scripts design oligo libraries, primers for amplifying oligo-sublibraries, and inverse PCR
563 primers for adding compatible cut sites to the Kir2.1 plasmid.

564 All backbone were amplified using a 25 cycle PCR with GXL polymerase and 1 ng of backbone
565 DNA as template. The PCR product was then gel purified. All oligo libraries were amplified using
566 25 cycles PCR with GXL polymerase and with 1 ul of the OLS library (resuspended in 1 ml TE)
567 as template. To assemble the backbone and library DNA, BsmBI Golden Gate reactions were set
568 up in 20 ul containing 100 ng of amplified backbone DNA, 20 ng of amplified oligo DNA, 0.2 µl
569 BsaI-HFv2 (New England Biolabs), 0.4 µl T4 DNA ligase (New England Biolabs), 2 µl T4 DNA
570 ligase buffer, and 2 µl 10 mg/ml BSA. These reactions were put in a thermocycler overnight using
571 the following program: (1) 5 min at 42°C, (2) 10 min at 16°C, (3) repeat 40 times, (3) 42°C for 20
572 min, (4) 80°C for 10 min. This reaction was cleaned using a Zymo Research Clean and
573 Concentrate 5 kit and eluted in 6 ul of elution buffer. The entirety of this reaction transformed in
574 E. Cloni 10G electrocompetent cells (Lucigen) according to manufacturer's instructions. Cells
575 were grown overnight with shaking at 30°C to avoid overgrowth in 30 ml of LB with 40ug/ml
576 kanamycin and library DNA was isolated using Zymo Zyppy miniprep kits. A small subset of
577 transformed cells was plated varying dilutions to assess transformation efficiency and validate
578 successful mutations. All libraries at this step yielded >300,000 colonies implying a 100x coverage
579 (assuming 1/3 of the variants were perfect based on our previous analysis of Agilent OLS based
580 libraries). Each sublibrary was combined at an equimolar ratio to make a complete library with all
581 intended mutations included.

582
583 *Stable Cell Line Generation:* To generate cell lines, we used a rapid single-copy mammalian cell
584 line generation pipeline⁴⁰. Briefly, mutational libraries are cloned into a staging plasmid with BxBI-
585 compatible *attB* recombination sites using BsmBI Golden Gate cloning. We amplify the staging
586 plasmid backbone using inverse PCR and the library of interest with primers that add
587 complementary BsmBI cut sites. Golden Gate cloning and subsequent transformation is
588 conducted with BsmBI (NEB), T4 Ligase (NEB) following manufacturer's using the same protocol
589 as previously described for library generation. Completed library landing pad constructs are co-

590 transfected (1:1) with a BxB1 expression construct (pCAG-NLS-Bxb1) into (TetBxB1BFP-iCasp-
591 Blast Clone 12 HEK293T cells) using Turbofect according to the manufacturer's instructions in 6
592 wells of a 6-well dish. This cell line has a genetically integrated tetracycline induction cassette,
593 followed by a BxB1 recombination site, and split rapalog inducible dimerizable Casp-9. Cells were
594 maintained in D10 (DMEM, 10% fetal bovine serum (FBS), 1% sodium pyruvate, and 1%
595 penicillin/streptomycin). Two days after transfection, doxycycline (2 ug/ml, Sigma-Aldrich) is
596 added to induce expression of our genes of interest (successful recombination) or the iCasp-9
597 selection system (no recombination). Successful recombination shifts the iCasp-9 out of frame,
598 thus only cells that have undergone recombination survive, while those that haven't will die from
599 iCasp-9-induced apoptosis. One day after doxycycline induction, AP1903 (10 nM,
600 MedChemExpress) is added to cause dimerization of Casp9 and selectively kill cells without
601 successful recombination. One day after AP1903-Casp9 selection, media is changed back to D10
602 + Doxycycline (2 ug/ml, Sigma-Aldrich) for recovery. Two days after cells have recovered, cells
603 are reseeded to enable normal cell growth. Once cells reach confluency, library cells are frozen
604 in 50% FBS and 10% DMSO stocks in aliquots for assays.

605

606 *Surface Expression cell sorting:* Thawed stocks of library cell lines were seeded into a 10 cm dish
607 and media was swapped the following day to D10. Cells were grown and split before confluency
608 to maintain cell health. Media was swapped to D10 + doxycycline (2 ug/ml, Sigma-Aldrich) two
609 days prior to the experiment. Cells were detached with 1 ml Accutase (Sigma-Aldrich), spun down
610 and washed three times with FACS buffer (2% FBS, 0.1% NaN₃, 1X PBS), incubated for 1-hour
611 rocking at 4°C with a BV421 anti-flag antibody (BD Bioscience), washed twice with FACS buffers,
612 filtered with cell strainer 5 ml tubes (Falcon), covered with aluminum foil, and kept on ice for
613 transfer to the flow cytometry core. Before sorting, 5% of cells were for processing and sequencing
614 as a baseline control.

615 Cells were sorted on a BD FACSAria II P69500132 cell sorter. miRFP670 fluorescence was
616 excited with a 640 nm laser and recorded with a 670/30 nm bandpass filter and 505 nm long-pass
617 filter. BV421 fluorescence was excited using a 405 nm laser. Cells were gated on forward
618 scattering area and side scattering area to find whole cells, forward scattering width, and height
619 to separate single cells, miRFP670 for cells that expressed variants without errors (our library
620 generation results in single base pair deletions that will not have miRFP670 expression because
621 deletions will shift the fluorescent protein out of frame ³⁸), and label for surface expressed cells.
622 The surface expression label gate boundaries were determined based on unlabeled cells from

623 the same population because controls tend to have non-representative distributions. Example of
624 the gating strategy for is depicted in **Supplemental Fig. 1**.

625 Cells were sorted based on surface expression into 4 populations (miRFP^{high}/BV421^{none},
626 miRFP^{high}/BV421^{low}, miRFP^{high}/BV421^{medium}, miRFP^{high}/BV421^{high}). We collected at least 2.1 million
627 cells in each population to ensure a greater than 100x coverage for each sample in each subpool.
628 The surface expression experiment was done in duplicate on separate days for two entirely
629 independent replicates.

630

631 *Resting membrane potential cell sorting:* Concurrently with preparing samples for surface labeling
632 another sample of the same cells were prepared for sorting based on resting membrane potential.
633 These cells were initially washed with FACS buffer and concentrated in this buffer for cell health
634 prior to sorting. 30 minutes prior to sorting cells were resuspended in Tyrode (125mM NaCl, 2mM
635 KCl, 3mM CaCl₂, 1mM MgCl₂, 10mM HEPES, 30mM glucose, pH 7.3) that contained FLIPR
636 membrane potential dye with a Blue quencher.

637 Cells were also sorted based on resting membrane potential on a BD FACSAria II P69500132
638 cell sorter. miRFP670 fluorescence was excited with a 640 nm laser and recorded with a 670/30
639 nm bandpass filter and 505 nm long-pass filter. FLIPR fluorescence was excited using a 488 nm
640 laser and recorded on a 525/50 nm bandpass filter. As before the same general sorting scheme
641 was used to identify whole single cells based on forward and side scatter and enriched for good
642 quality library members based on miRFP670 fluorescence. An example of the gating strategy is
643 in **Supplemental Fig. 2** Cells were sorted based on resting membrane potential into 3 populations
644 (miRFP^{high}/FLIPR^{Low}, miRFP^{high}/FLIPR^{medium}, miRFP^{high}/FLIPR^{high}). The resting membrane potential
645 experiment was done in duplicate on separate days for two entirely independent replicates.

646

647 *Sequencing:* For both biological replicates, DNA from pre-sort control and sorted cells was
648 extracted with Microprep DNA kits (Zymo Research) and triple-eluted with water. The elute was
649 diluted such that no more than 1.5ug of DNA was used per PCR reaction and amplified for 20
650 cycles of PCR using Primestar GXL (Takara Bio), run on a 1% agarose gel, and gel purified.
651 Primers that bind outside the recombination site ensure leftover plasmid DNA from the original
652 cell line construction step is not amplified. Purified DNA was quantified using Picogreen DNA
653 quantification. Equal amounts (by mass) of each domain insertion sample were pooled by cell
654 sorting category (“pre-sort control”, “surface expression”, “no surface expression”). Pooled
655 amplicons were prepared for sequencing using the Nextera XT sample preparation workflow and

656 sequenced using Illumina Novaseq in 2x150bp mode. Source sequencing data is available in the
657 NCBI Sequence Raw Archive (<https://www.ncbi.nlm.nih.gov/sra>) under accession code
658 PRJNA791691. Read count statistics are listed in **Supplemental Table 1A**.

659
660 *Alignment and Enrichment Calculation:* Variant frequency for single mutations were determined
661 by joining paired reads with bbmerge, aligning reads to the Kir2.1 reference gene sequence with
662 bbmap, and identifying mutations and filtering to find the OLS programmed mutations with a
663 custom python script. The 150 bp paired-end reads were trimmed to remove standard Illumina
664 adapters (bbmap/resources) using BBDuk with 'mink' at 8 bases. Overlapping reads were
665 corrected without merging using BBMerge with the 'ecco' and 'mix' setting. Reads were aligned
666 to Kir2.1 sequence using BBMap using 'maxindel' at 500 and 'local' alignment. The resulting sam
667 file was analyzed for mutations at each position and filtered to match only mutations that were
668 programmed on the OLS chip. To calculate enrichment of mutations we used Enrich2⁴⁸. Counts
669 for each mutation were input into the Enrich2 software with weighted least squares scoring and
670 wild type normalization. Scores were output with a standard error in a .csv file. Coverage of
671 mapped reads is listed in **Supplemental Table 1B-C**.

672
673 *Inward Rectifier Phylogenetic Alignment and Clinvar mutation assignment:* We downloaded all
674 human inward rectifiers (KCNJ1(Kir1.1/ROMK; Uniprot P48048), KCNJ10(Kir1.2/Kir4.1; Uniprot
675 P78508), KCNJ15(Kir1.3/Kir4.1; Uniprot Q99712), KCNJ2(Kir2.1; Uniprot P63252),
676 KCNJ12(Kir2.2; Uniprot Q14500), KCNJ4(Kir2.3; Uniprot P48050), KCNJ14(Kir2.4; Uniprot
677 Q9UNX9), KCNJ18(Kir2.6; Uniprot B7U540), KCNJ3(Kir3.1/GIRK1; Uniprot P48549),
678 KCNJ6(Kir3.2/GIRK2; Uniprot P48051), KCNJ9(Kir3.3/GIRK3; Uniprot Q92806),
679 KCNJ5(Kir3.4/GIRK4; Uniprot P48544), KCNJ16(Kir5.1; Uniprot Q9NPI9), KCNJ8(Kir6.1; Uniprot
680 Q15842), KCNJ11(Kir6.2; Uniprot Q14654), KCNJ13(Kir7.1; Uniprot O60928) and aligned these
681 together using MegaX¹⁰². Based on this alignment, we generated a master list of Inward Rectifier
682 numberings to translate residue numbering between Kir homologues. We assigned mutations
683 observed Clinvar and dbSNP and their pathogenic classification (ClinVar only) to this master
684 alignment (**Supplementary Table 2**).

685

686 **Figure Legends**

687 **Figure 1 Deep Mutational Scanning to improve clinical variant prediction. (A)** Clinically
688 observed variation in Kir2.1 and other Kir and their interpretation of pathogenicity. **(B)** Schematic

689 summarizing the various processes that must work correctly for Kir2.1 to fulfill its role in cellular
690 physiology. **(C)** Schematic summarizing DMS and multiparametric assessment of phenotype.
691 Mutations are introduced into Kir2.1 using SPINE. A stable single-copy insertion library is
692 generated by BxB1-mediated recombination in HEK293T. Cells are sorted based on channel
693 surface expression or function (K^+ conductance) as determined by antibody labeling of an
694 extracellular FLAG tag or voltage-sensitive dye, respectively. Genotypes of each sorted cell
695 population are recovered by NextGen Sequencing (NGS).

696

697 **Figure 2 Surface Expression Fitness. (A)** Distribution of surface expression fitness scores for
698 synonymous mutations (red line) and missense mutations (black line). **(B)** Heatmap of surface
699 expression fitness scores (gradient from blue to red). Wildtype residues are indicated by green
700 boxes. Missing data is indicated by yellow boxes. Synonymous mutations are indicated by red
701 box outline. **(C)** Cartoon of Kir2.2 (PDB 3SPI, 70% identity with Kir2.1). Monomer chains show in
702 different colors. Lipid bilayer boundaries are indicated by dashed lines. **(D)** Close-up of core beta
703 sheet of the CTD with surface expression fitness mapped onto each residue as a gradient from
704 blue to red. Residues that comprise the Golgi export signal are outlined green. **(E)** Interface close-
705 up of neighboring subunits (green and magenta cartoons). Sidechain are colored by surface
706 fitness score. Disrupting carbon- π and π - π interactions between F45, L246, and
707 Y338 does not uniformly decrease fitness.

708

709 **Figure 3 Determinant of improve surface expression and Kir2.1's biogenic folding unit. (A)**
710 Boxplots showing surface fitness in the β DE and β LM loop if the wildtype residues of a specific
711 type (e.g., glutamate, charged) is mutated to another residue of the same type (e.g., Aspartate)
712 or a different type (e.g., Tryptophan). Median is marked with a thick line, the vertical length of the
713 box represents the interquartile range (IQR), upper fence: 75th percentile +1.5 \times IQR, lower fence:
714 25th percentile -1.5 \times IQR, outlier points are shown as solid black circles. All data points are
715 indicated by transparent dots. Only replacing aromatic residues significantly increase surface
716 expression. Significance is tested using one-sided t-tests; p-values are indicated for each
717 comparison. **(B)** Close-up of surface expression fitness scores (gradient blue to red) in the β DE
718 and β LM loop. **(C)** Close-up of surface expression fitness scores in the pore domain. Residues
719 that are part of the putative biogenic unit of Kir2.1 are show as sticks. **(D)** Surface expression
720 fitness scores mapped onto a single Kir2.1 monomer highlighting the importance of the biogenic
721 folding unit in the pore domain and the core beta sheet of the CTD.

722

723 **Figure 4 Function Fitness. (A)** Distribution of function expression fitness scores for synonymous
724 mutations (red line) and missense mutations (black line). **(B)** Function fitness scores mapped onto
725 a single Kir2.1 monomer highlighting the importance of the pore helix and selectivity filter, and the
726 structural elements are the TM/CTD interface (e.g., interfacial helix, tether helix, G-loop gate) for
727 function. **(C)** Heatmap of function expression fitness scores (gradient from green to magenta).
728 Wildtype residues are indicated by blue boxes. Missing data is indicated by yellow boxes.
729 Synonymous mutations are indicated by red box outline. **(D-G)** Heatmap of fitness score in
730 regions that contain bona-fide trafficking signal **(D,E;** for surface and function scores, respectively)
731 or the PIP₂ binding site **(F,G;** for surface and function scores, respectively). **(H)** Closeup of
732 function fitness at the TM/CTD interface. Allosteric ligand PIP₂ is shown as sticks.

733

734 **Figure 5 HBC gate mutations and regions with distinct roles stability and function. (A)**
735 Function Fitness (gradient from green to magenta) in the pore domain. One subunit is removed
736 for clarity. K⁺ ions in the selectivity filter a show as purple spheres. Sidechain of residues that are
737 enriched for activating mutations are shown as sticks. **(B)** Detailed view of the HBC gate. **(C)**
738 Difference between z-score normalized mean Function and Surface Fitness scores mapped by
739 position. Filled circles (cyan or magenta) indicated significant differences (two-sided moderated t
740 test p-value < 0.05). Kir2.1 secondary structural element are shown as gray boxes; trafficking
741 signals are colored green. **(D-F)** Residues with significant function/surface fitness score
742 differences are mapped onto the Kir2.2 structure (PBD 3SPI). Backbone cartoons are colored by
743 chain. Residues with mutation that predominantly reduced function are shown in magenta.
744 Residues with mutation that predominantly reduced surface expression are shown in cyan. PIP₂
745 is shown in blue. Overview **(D)**, close-ups of selectivity filter **(E)** and TM/CTD **(F)**.

746

747 **Figure 6 Folding and trafficking variants in clinical databases.** Cumulative distributions of
748 surface **(A)** or function **(B)** fitness scores for variants that are represented in Clinvar and dbSNP
749 (red line) or that are not (black line). **(C)** Line plot by position of mean function or surface fitness
750 scores (gradient green to magenta and blue to red, respectively). Kir2.1 secondary structural
751 element are shown as gray boxes. Pathogenic variants are mapped as circles with fill color
752 indicating predicted pathogenic mechanism. Mutation hotspots are indicated by gray outlines.
753 Distribution of surface fitness **(D)** and function fitness **(E)** by ClinVar pathogenicity assignment.

754 Vertical dashed lines represent 2.5th and 97.5th percentiles bounds of expert reviewed pathogenic
755 variants (red) and benign (blue), respectively.

756

757 **Author contributions**

758 WCM, DN, and DS designed the work presented here. WCM and DN did the experiments with
759 support from YH. DN processed the sequencing data. WCM and DS did the analysis and wrote
760 the manuscript.

761

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769

770 **Competing Interests**

771 The authors declare no competing interests.

772

773 **References**

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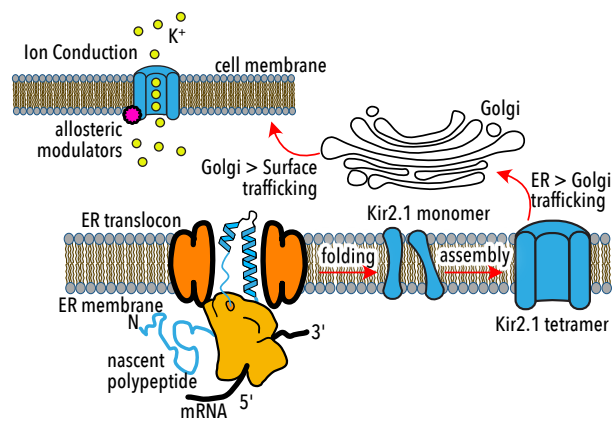
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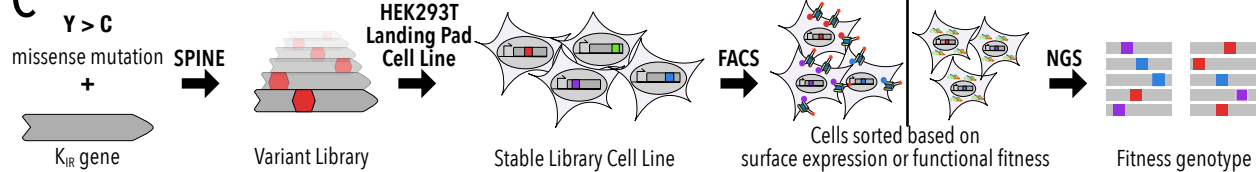
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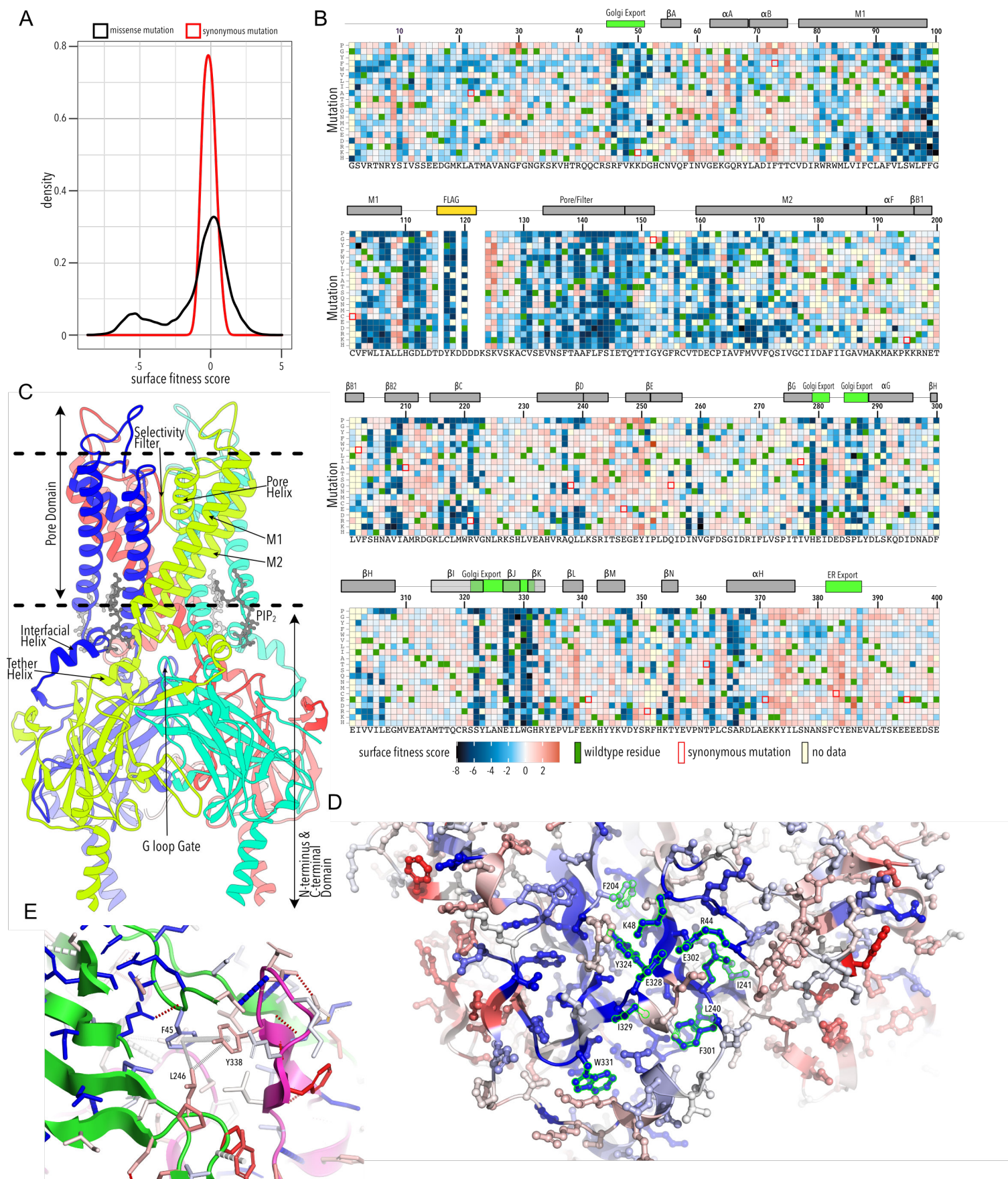
Clinical Interpretation	Kir2.1	other Kir
Benign	1	15
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Likely benign	1	10
Likely pathogenic	17	25
not provided	16	2
Pathogenic	27	79
Uncertain significance	89	299

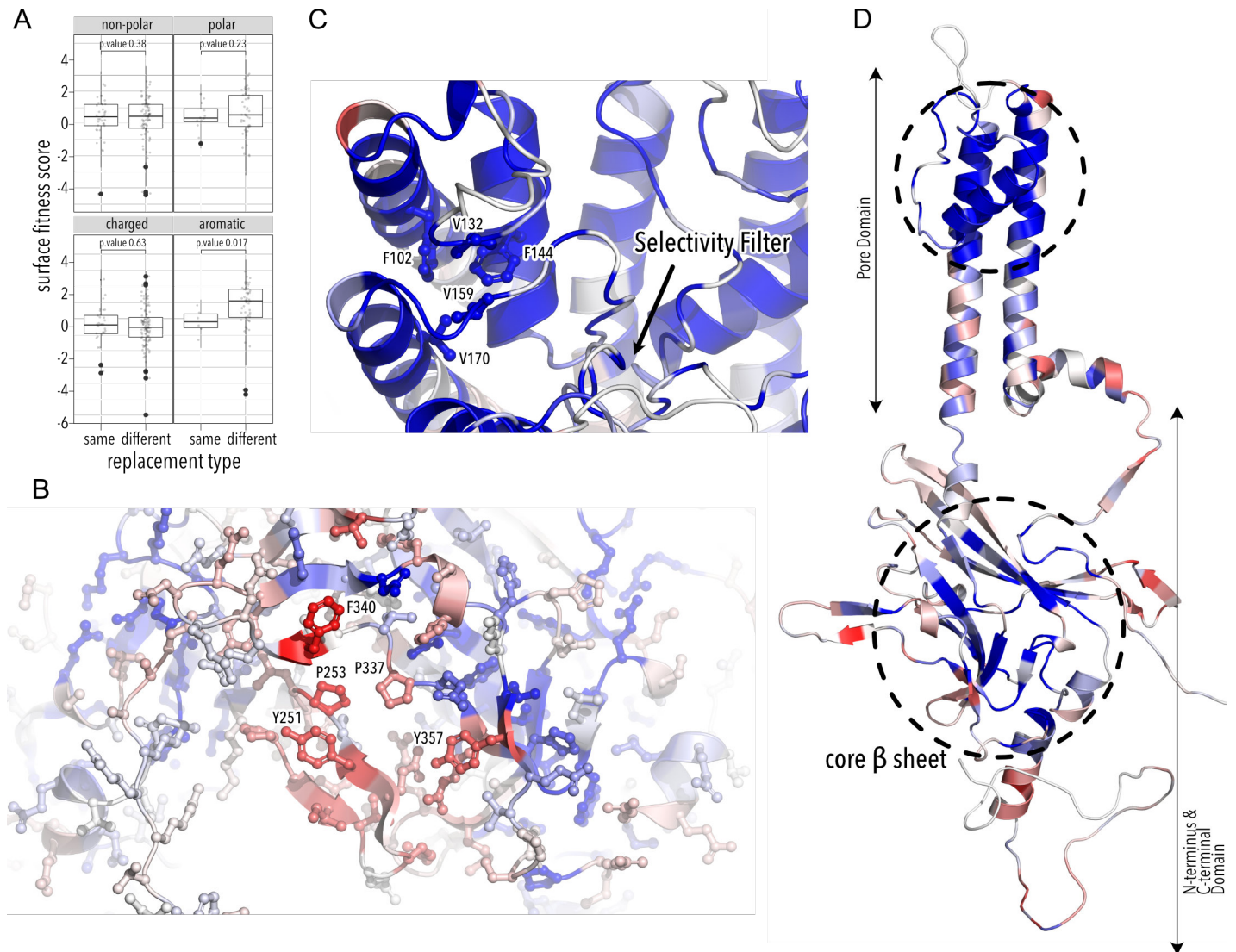
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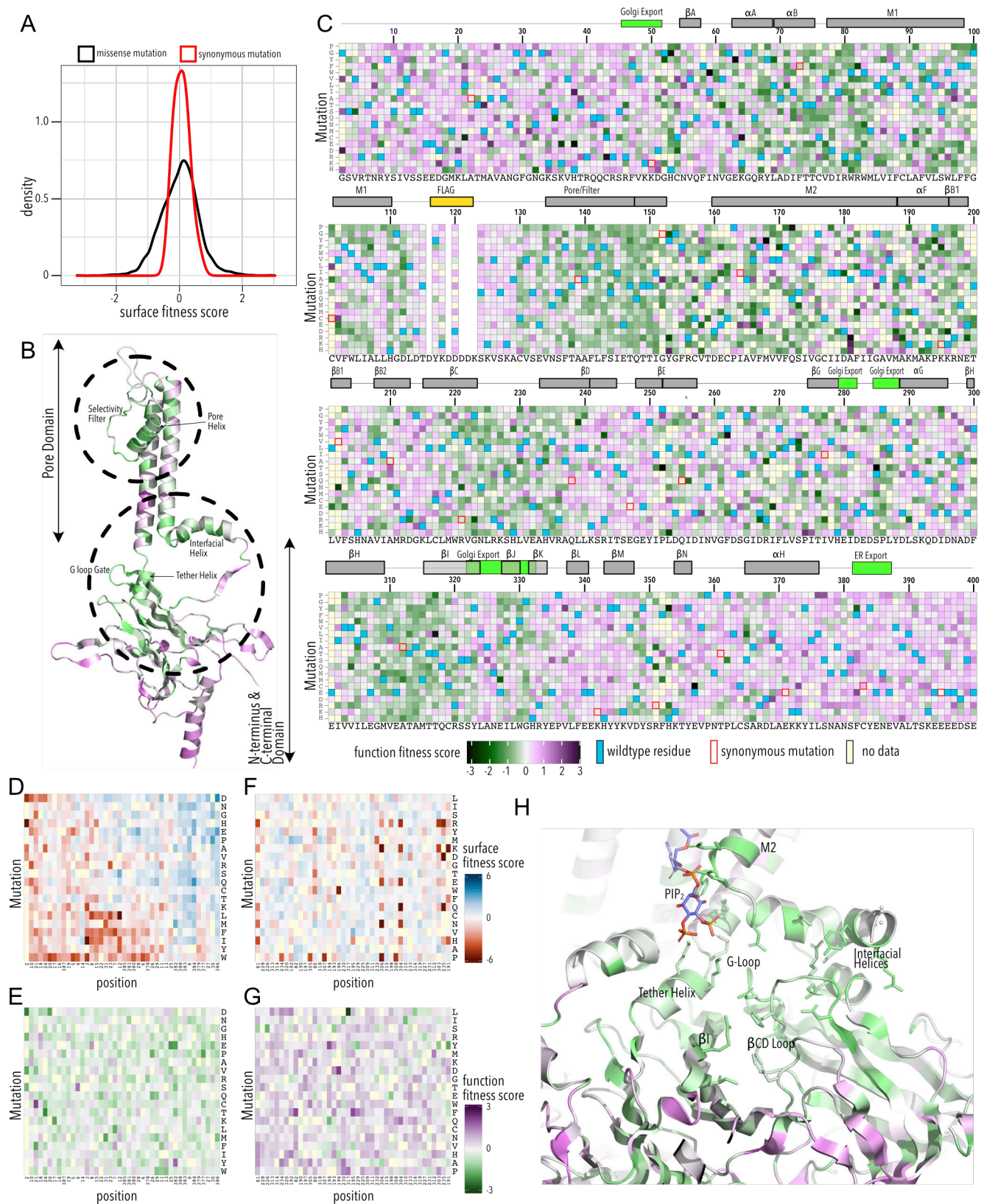


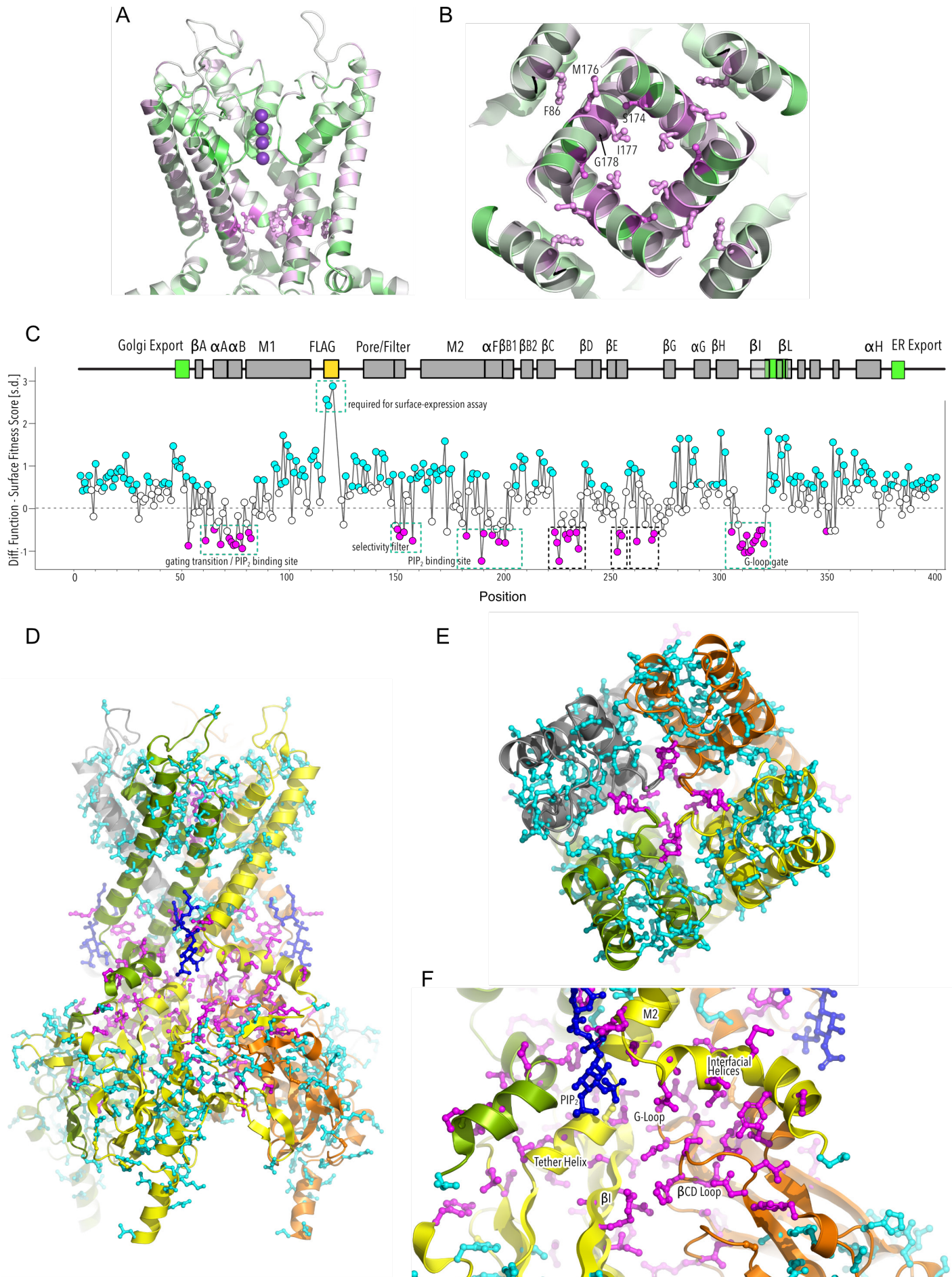
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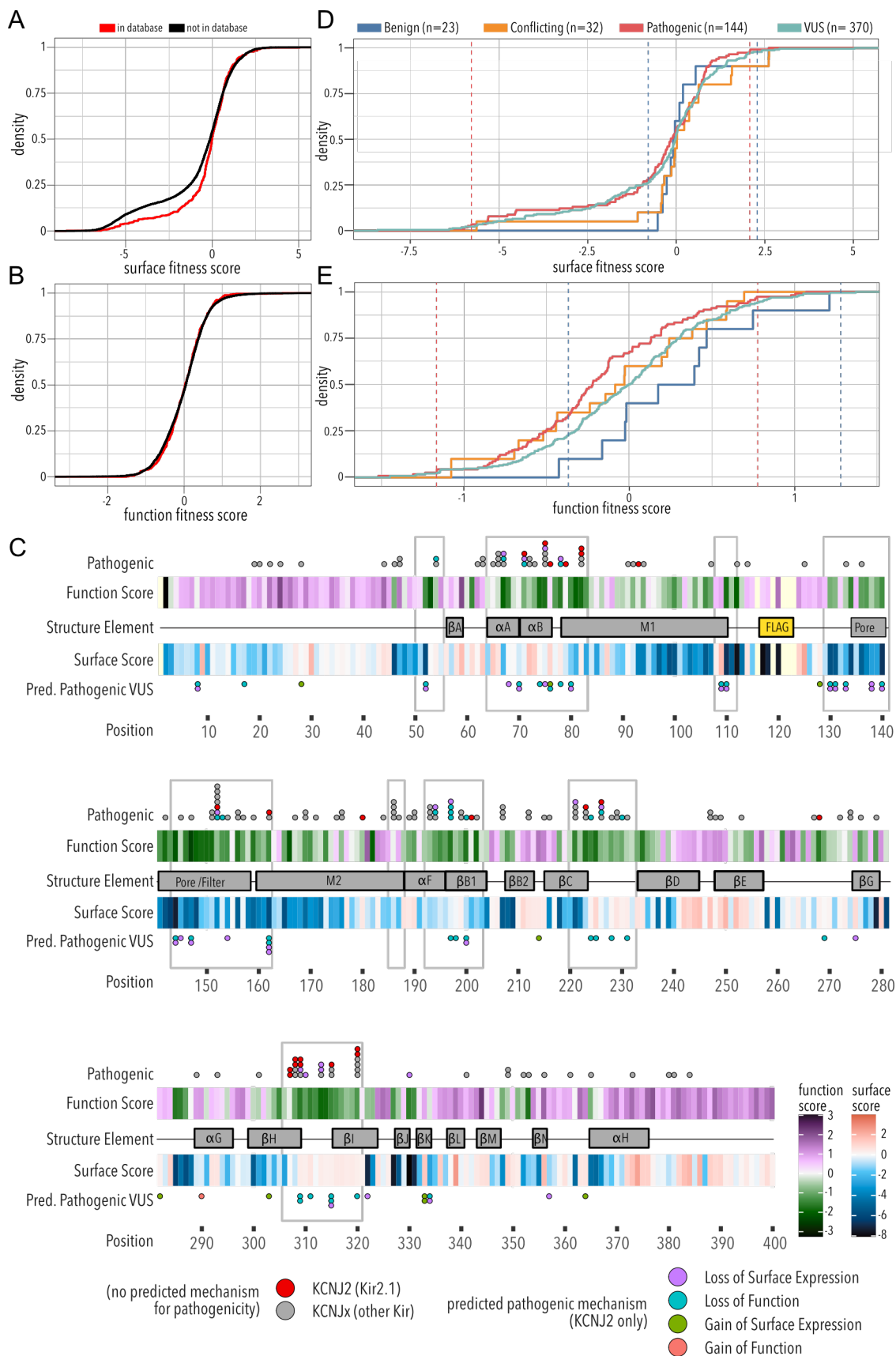












SUPPLEMENTARY MATERIALS FOR:

Determinants of trafficking, conduction, and disease within a K⁺ channel revealed through multiparametric deep mutational scanning

Short Title: Multiparametric Deep Mutational Scanning in the Inward Rectifier Kir2.1

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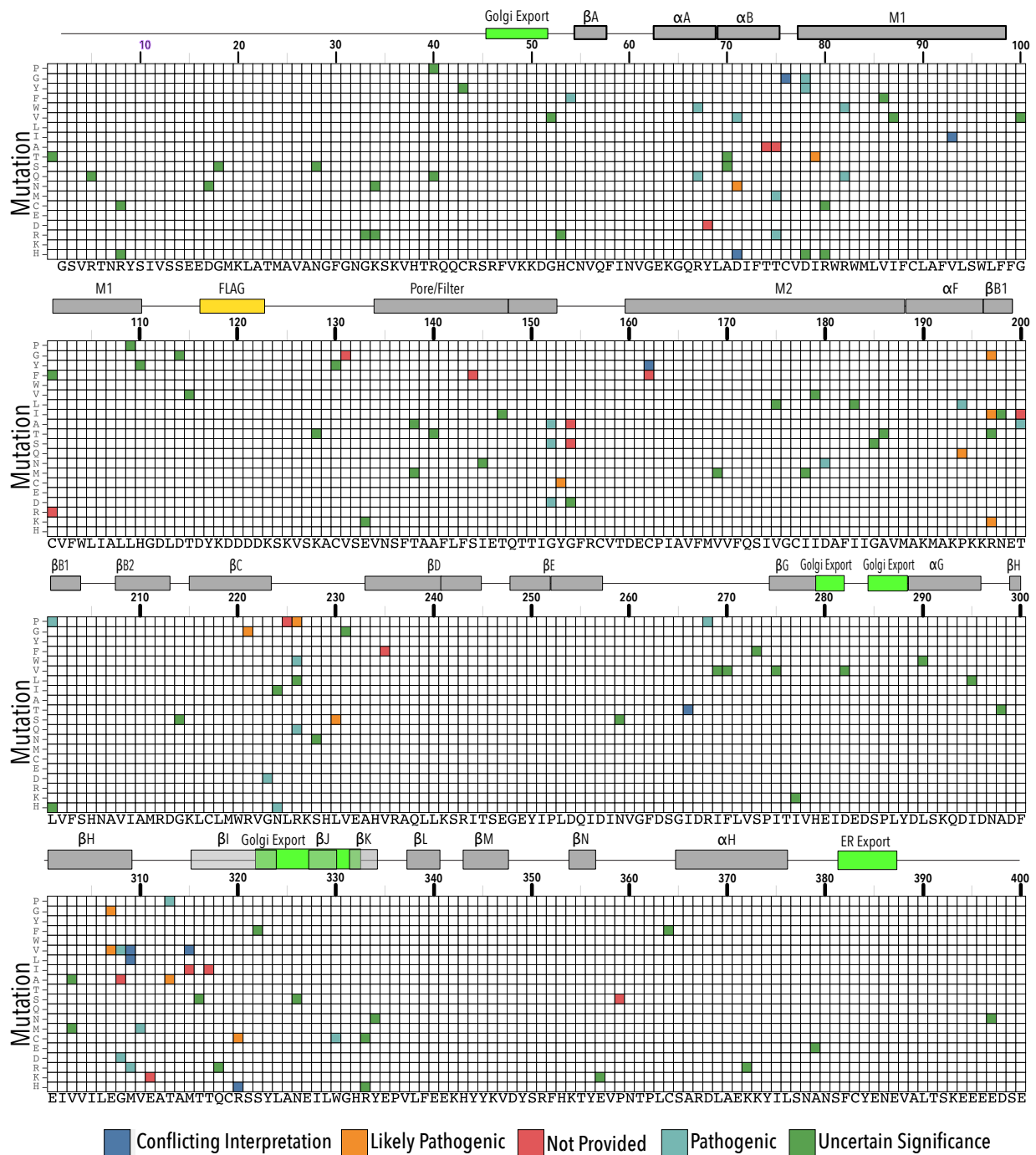
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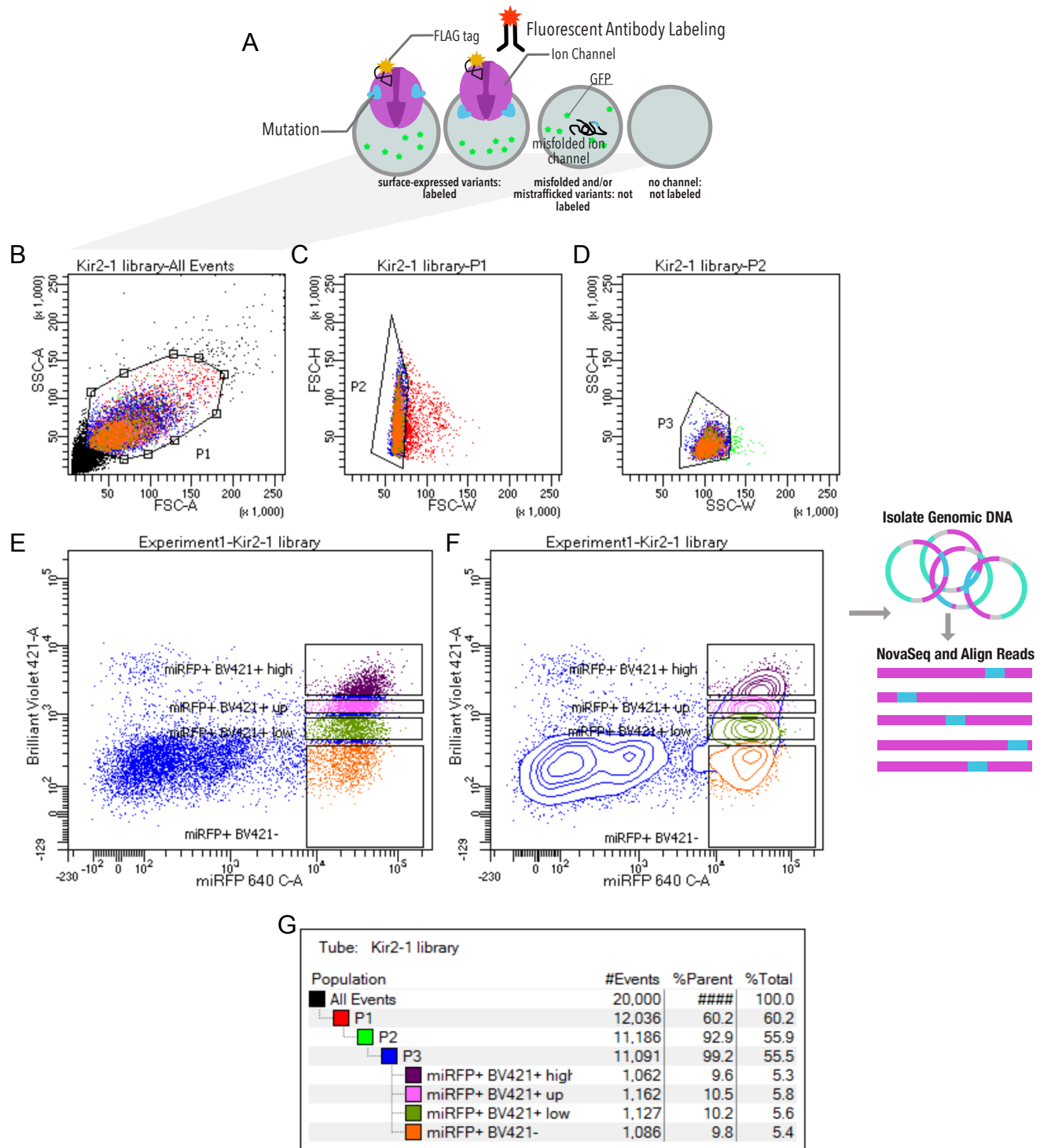
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Supplemental Figures 1 – 9

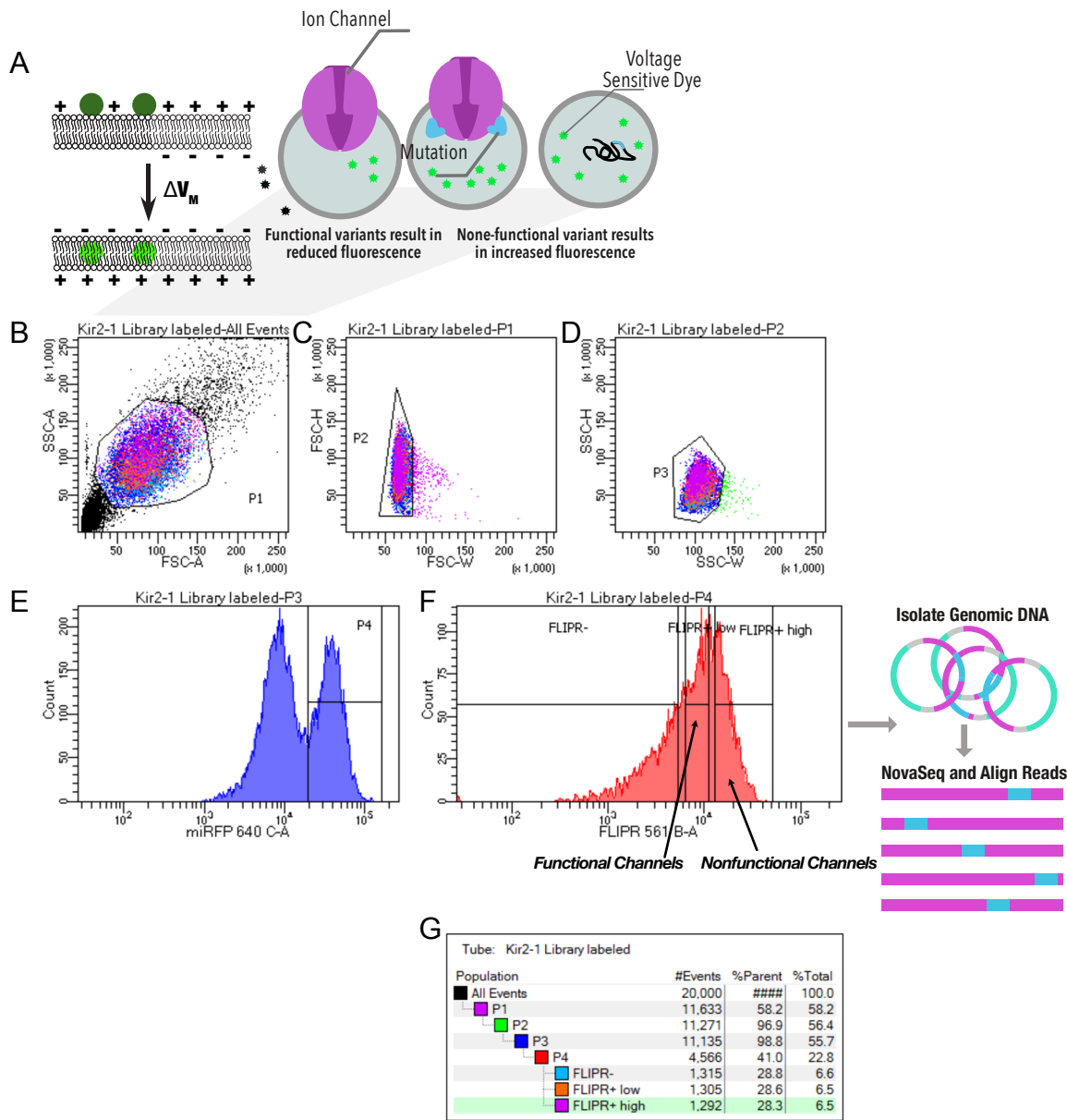
Supplemental Tables 1 – 3



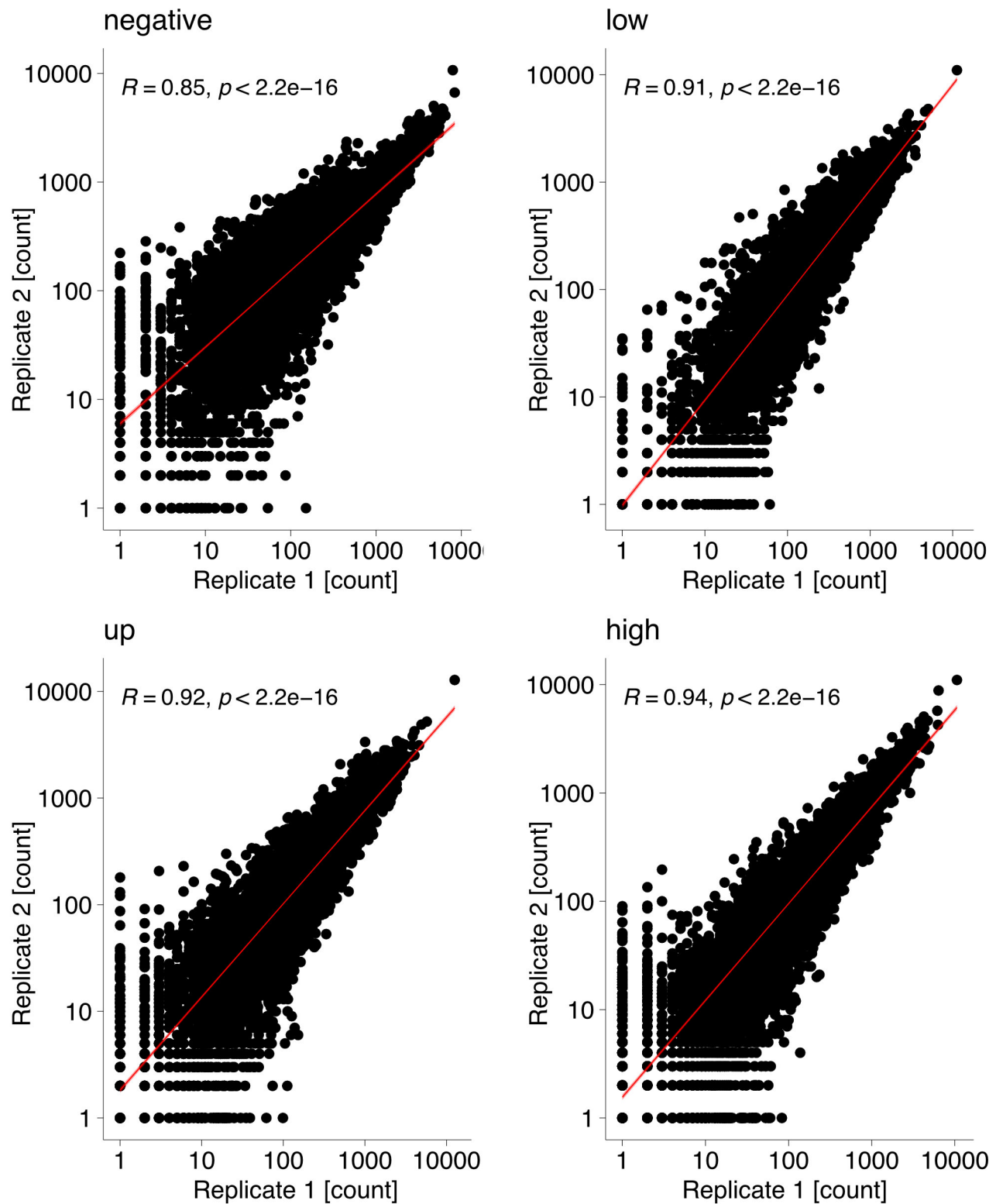
Supplementary Figure 1: Kir2.1 variant effect reported in ClinVar. Heatmap showing clinically observed Kir2.1 variants mapped by position, mutation, and pathogenicity interpretation. Kir2.1 secondary structural element are shown as gray boxes above each panel. Trafficking signals are colored green.



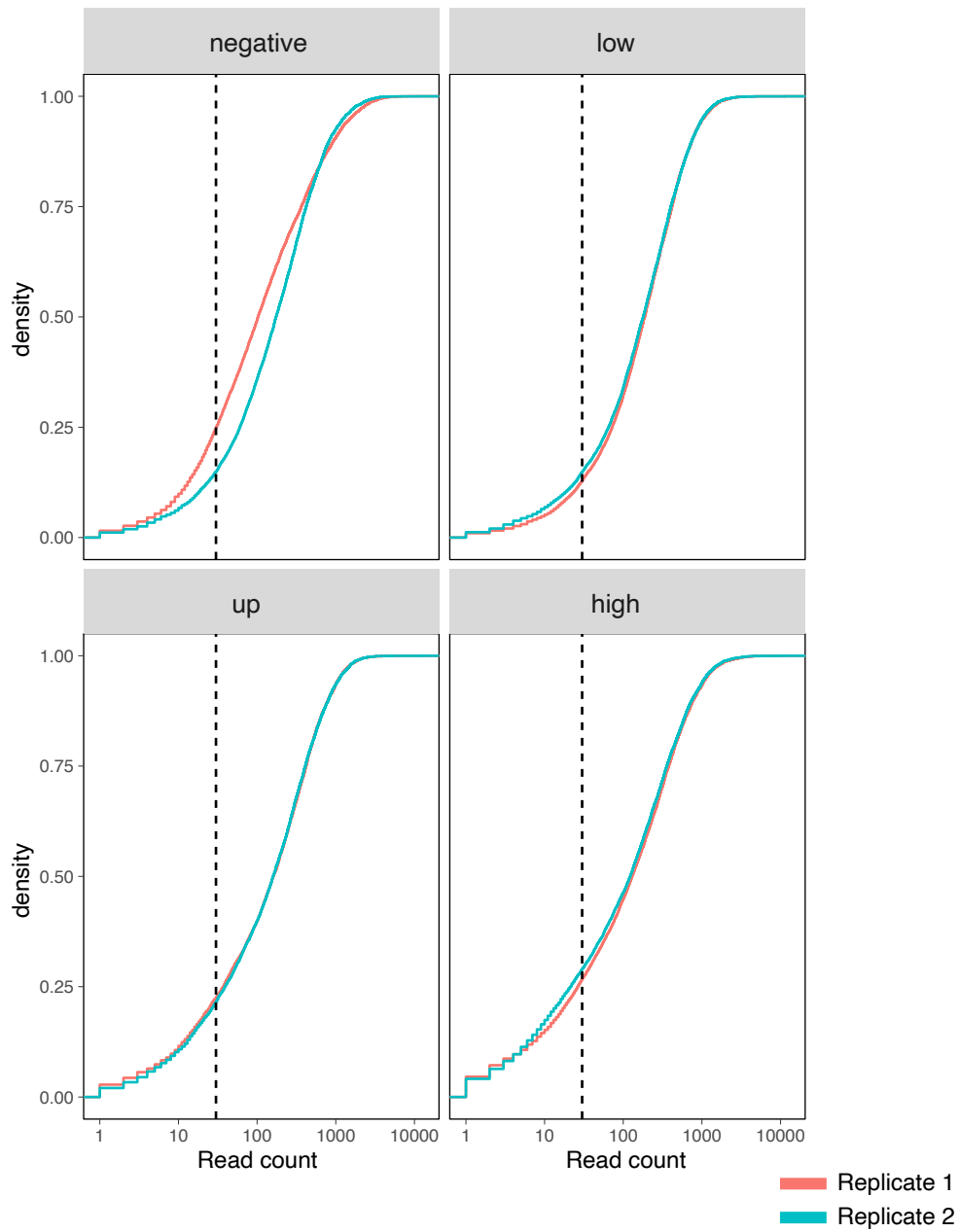
Supplementary Figure 2: Kir2.1 surface expression assay gating scheme. (A) HEK293 cells expressing a single variant are incubated with a fluorescent (Brilliant Violet) antibody that recognizes a FLAG tag inserted into an extracellular loop (Kir2.1 position T115). Cells expressing channel variants that can fold, assemble and traffic to the cell surface are labelled, while those that express variants with impaired folding, assembly, and/or trafficking are not labelled. **(B)** Using FACS, whole HEK293 cells are gated on side (SSC-A) and forward scattering (FSC-A). **(C-D)** Forward scattering height (FSC-H), forward scattering width (FSC-W), and Side scattering width (SSC-W) are used to gate single cells. Non-surface expressed variants (miRFP_{high}/Label_{low}) and surface expressed variants (miRFP_{high}/Label_{high}) populations are gated into 'negative', 'low', 'up', and 'high' populations based an expression marker (miRFP670) of Anti-Flag Brilliant Violet-421 fluorescence. Scatter plot **(E)** and contour plot **(F)** shown. Contour plots represent 95% confidence intervals with outliers shown as dots. **(G)** Sort statistic for each gated cell population.



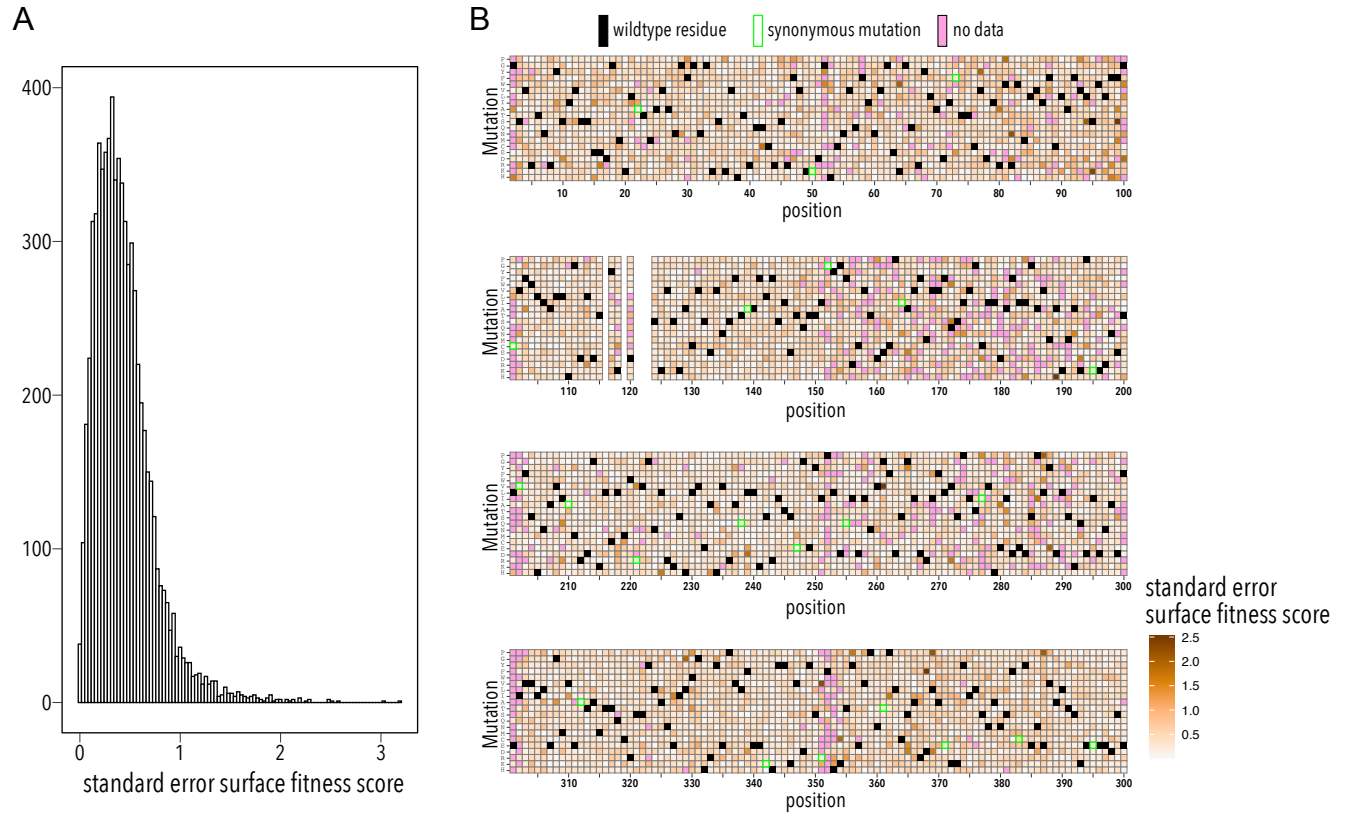
Supplementary Figure 3: Kir2.1 functional fitness assay gating scheme. (A) HEK293 cells expressing a single variant are incubated with a voltage-sensitive FLIPR dye. In strongly hyperpolarized cells, this dye partitions out the cell membrane into an aqueous environment, which decreases its extinction coefficient and thus decreases green fluorescence upon excitation with blue light. In depolarized cells, the FLIPR dye fluorescence is brighter because the dye partitions into the membrane, which increases its extinction coefficient. **(B)** Using FACS, whole HEK293 cells are gated on side (SSC-A) and forward scattering (FSC-A). **(C-D)** Forward scattering height (FSC-H), forward scattering width (FSC-W), and Side scattering width (SSC-W) are used to gate single cells. **(E)** Cells are further gated using miRFP670 as an expression marker. **(F)** Non-functional and functional channel population are separated by sorting based on FLIPR fluorescences. **(G)** Sort statistic for each gated cell population.



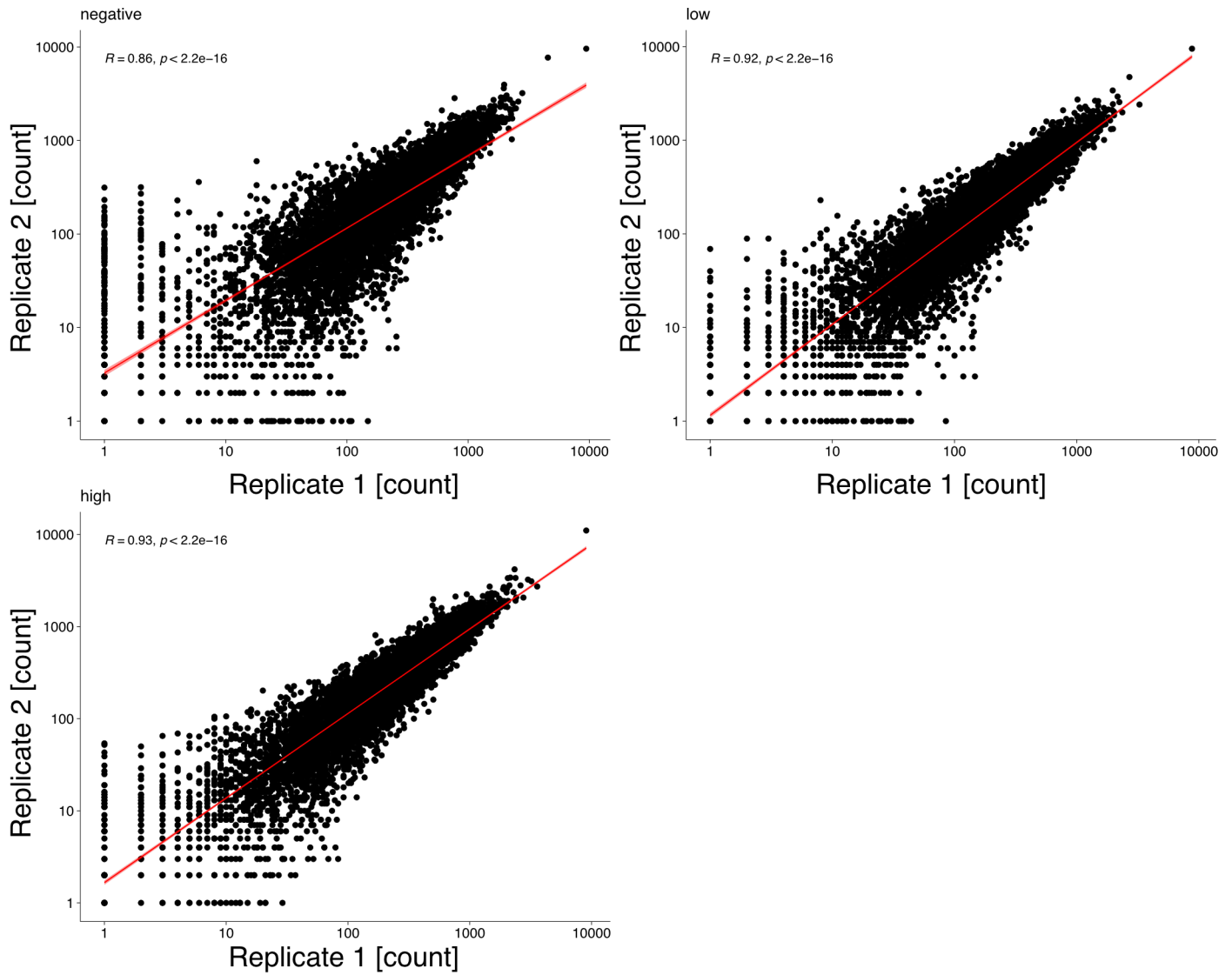
Supplementary Figure 4: Surface fitness assay biological replicates. Scatterplots show mapped read counts for each Kir2.1 variant in each replicate for different assay subpools. Spearman correlation coefficients are inset for each replicate pair. A linear model regression fit is shown as a red line.



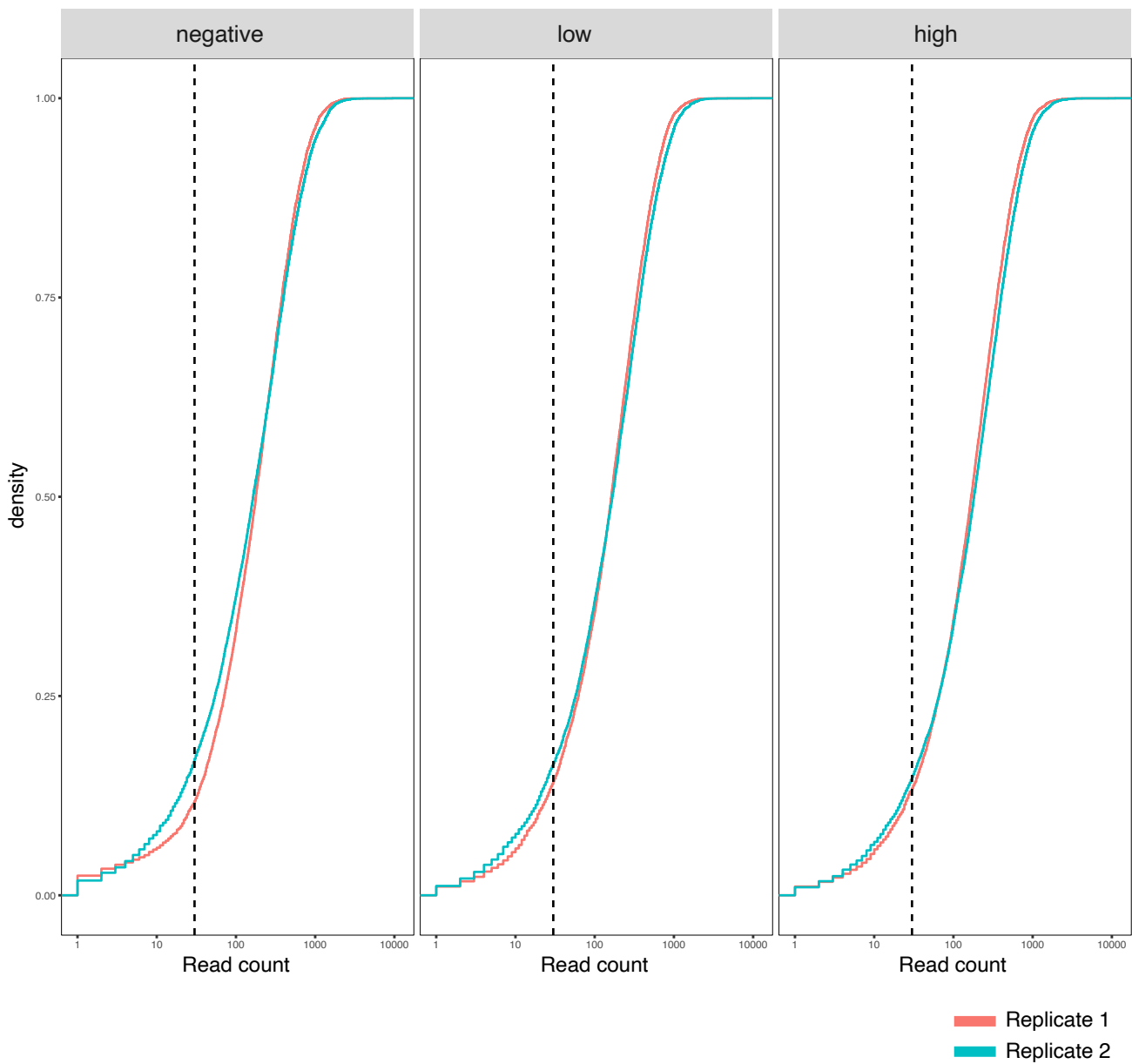
Supplementary Figure 5: Surface fitness assay read count depth. Cumulative density distribution of read counts for each replicate and assay condition subpool. Dashed vertical lines represent 30-fold coverage.



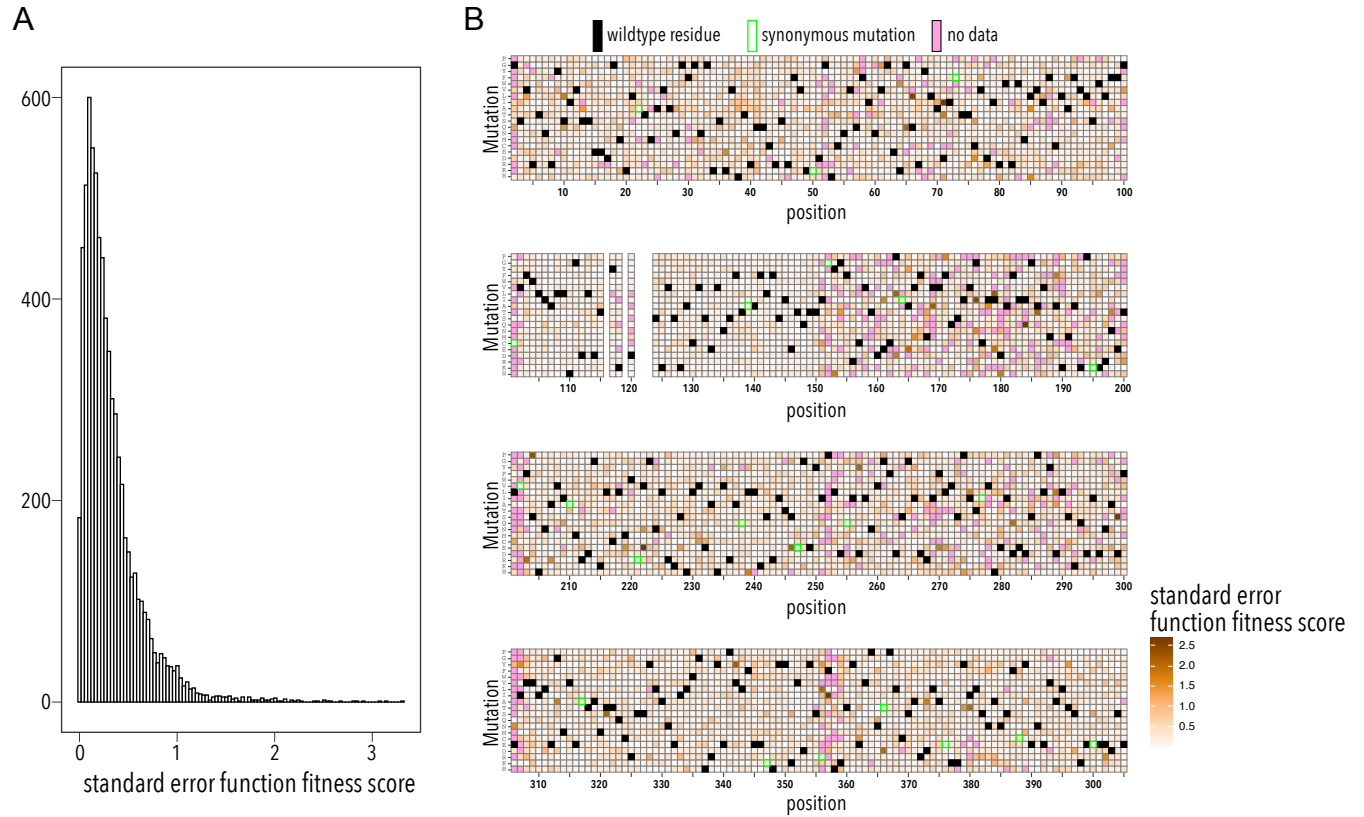
Supplementary Figure 6: Surface fitness assay standard error distribution. (A) Distribution of surface fitness score standard errors. **(B)** Heatmap showing surface fitness standard error (gradient white to brown) mapped to each position and mutation. Wildtype residues are shown as black boxes. Missing data is shown as magenta boxes. Synonymous mutations are indicated by a green outline.



Supplementary Figure 7: Surface fitness assay biological replicates. Scatterplots show mapped read counts for each Kir2.1 variant in each replicate for different assay subpools. Spearman correlation coefficients are inset for each replicate pair. A linear model regression fit is shown as a red line.



Supplementary Figure 8: Surface fitness assay biological replicates. Cumulative density distribution of read counts for each replicate and assay condition subpool. Dashed vertical lines represent 30-fold coverage.



Supplementary Figure 9: Function fitness assay standard error distribution. (A) Distribution of surface fitness score standard errors. **(B)** Heatmap showing surface fitness standard error (gradient white to brown) mapped to each position and mutation. Wildtype residues are shown as black boxes. Missing data is shown as magenta boxes. Synonymous mutations are indicated by a green outline.

A

Sample	Barcode sequence	PF Clusters	% of the lane	% Perfect barcode	% One mismatch barcode	Yield (Mbases)	% PF Clusters	% \geq Q30 bases	Mean Quality Score
dms_baseline	CGTACTAG+AGAGGATA	17,942,041	1.78	95.35	4.65	5,418	100	90.5	35.34
dms_function_high1	GTAGAGGA+ATAGAGAG	14,044,823	1.39	95.15	4.85	4,242	100	87.22	34.64
dms_function_high2	TAAGGCGA+AGAGGATA	18,810,378	1.86	96.5	3.5	5,681	100	89.08	35.01
dms_function_low1	CGAGGCTG+ATAGAGAG	14,199,546	1.41	97.46	2.54	4,288	100	88.63	34.99
dms_function_low2	GGAGCTAC+ATAGAGAG	14,903,201	1.48	95.74	4.26	4,501	100	89.28	35.1
dms_function_neg1	TAGGCATG+ATAGAGAG	15,073,953	1.49	97.14	2.86	4,552	100	91.1	35.45
dms_function_neg2	ACTCGCTA+ATAGAGAG	16,560,022	1.64	95.44	4.56	5,001	100	89.56	35.14
dms_surface_high1	TCCTGAGC+ATAGAGAG	15,200,690	1.51	96.22	3.78	4,591	100	90.8	35.37
dms_surface_high2	TAGGCATG+AGAGGATA	14,882,582	1.47	97.33	2.67	4,495	100	90.87	35.4
dms_surface_low1	CGTACTAG+ATAGAGAG	16,320,952	1.62	93.92	6.08	4,929	100	88.75	34.98
dms_surface_low2	TCCTGAGC+AGAGGATA	17,274,356	1.71	97.2	2.8	5,217	100	90.98	35.42
dms_surface_neg1	TAAGGCGA+ATAGAGAG	18,343,213	1.82	94.16	5.84	5,540	100	87.59	34.73
dms_surface_neg2	AGGCAGAA+AGAGGATA	20,844,383	2.07	96.24	3.76	6,295	100	90.49	35.31
dms_surface_up1	AGGCAGAA+ATAGAGAG	16,105,434	1.6	94.92	5.08	4,864	100	89.01	35.01
dms_surface_up2	GGACTCCT+AGAGGATA	16,752,187	1.66	97.54	2.46	5,059	100	90.65	35.36

B

Surface Fitness Assay – Mapped Reads

	Negative		Low		Up		High	
	Rep. 1	Rep. 2	Rep. 1	Rep. 2	Rep. 1	Rep. 2	Rep. 1	Rep. 2
N_{variants}	7,282	7,282	7,305	7,305	7,175	7,175	6,975	6,975
median	103	174	187	179	160.5	160	130	123
mean	342.3	344.6	315.5	307.4	310.4	309.5	301.4	285.5
>30-fold [%]	71.7	82.4	85.4	82.2	73.4	73.4	65.7	64.4

C

Function Fitness Assay – Mapped Reads

	Negative		Low		Up	
	Rep. 1	Rep. 2	Rep. 1	Rep. 2	Rep. 1	Rep. 2
N	7,168	7,168	7,284	7,284	7,320	7,320
median	178	165	162	167	168	182
mean	281.6	297.1	248.5	276.5	261.5	295.7
>30-fold [%]	79.4	79.5	82.7	80.9	83.3	83.5

Supplementary Table 1: NGS read statistics and assay repeatability. (A) Read statistics for each sequenced variant pool. Mapped reads for each replicate and surface fitness assay sub-pool (B) or function fitness assay (C). N_{variants} represented in each replicate (out of 7,429 total), median read count per variant, mean read count per variant, and percentage of variants that have greater than 30-fold reach coverage.

KCNJ2	309	301	TRUE	M	V	NP	NP	-0.6693282	0.19420483	2.02E-07	0.01573106	0.40628209	2.02E-98	Conflicting	lof_f
KCNJ2	315	307	TRUE	M	I	NP	NP	-0.7175479	0.09290881	1.99E-34	-0.481593	0.12160307	1.38E-33	VUS	lof_f
KCNJ8	57	46	TRUE	Q	V	P	NP	NA	NA	NA	NA	0.39448643	6.34E-68	VUS	NA
KCNJ2	71	71	TRUE	D	V	C	NP	1.00340855	0.85017164	2.22E-16	-1.0252169	0.2968519	4.06E-74	Pathog.	NA
KCNJ2	85	85	TRUE	L	V	NP	NP	0.27657576	0.36230995	6.49E-14	-0.6348007	0.30310343	0	NA	NA
KCNJ2	87	87	TRUE	I	V	NP	NP	0.14704835	0.09265711	3.92E-33	0.12908661	0.43399975	1.11E-16	VUS	NA
KCNJ2	91	91	TRUE	A	V	NP	NP	0.4535043	0.06087486	3.92E-36	0.24243356	0.57248274	1.32E-65	NA	NA
KCNJ2	98	98	TRUE	F	V	A	NP	0.48549801	0.42873784	0	-0.3763099	0.386465	0	NA	NA
KCNJ11	186	166	TRUE	A	W	NP	A	-0.8453095	0.62411791	1.11E-16	0.32397819	0.19439502	0	Pathog.	NA
KCNJ10	19	18	FALSE	M	W	NP	A	0.12648476	0.06042266	4.36E-05	-2.0238281	0.28166805	3.15E-83	VUS	NA
KCNJ10	193	171	TRUE	K	W	C	A	0.13171266	0.27994746	3.00E-10	-0.2004754	0.84091396	0	VUS	NA
KCNJ13	193	162	TRUE	K	W	C	A	0.13171266	0.27994746	3.00E-10	-0.2004754	0.84091396	0	Pathog.	NA
KCNJ5	212	211	TRUE	R	W	C	A	0.27986015	0.278617	0	-0.0793948	0.24729635	1.04E-09	VUS	NA
KCNJ5	221	220	TRUE	R	W	C	A	-0.4560462	0.10802933	3.39E-11	-1.94495	0.3794728	1.40E-67	VUS	NA
KCNJ2	226	218	TRUE	R	W	C	A	0.19756449	0.55813353	0	-0.1926924	0.26260942	9.19E-13	Pathog.	NA
KCNJ5	236	235	TRUE	R	W	C	A	-0.0787073	0.78303552	0.0008113	0.29973448	0.10473114	0	VUS	NA
KCNJ5	243	242	TRUE	R	W	C	A	NA	0.6317913	3.78E-117	NA	NA	NA	VUS	NA
KCNJ10	252	230	TRUE	P	W	NP	A	-0.1815275	0.29840475	9.92E-08	5.03654636	0.40913285	0	VUS	NA
KCNJ2	290	282	TRUE	L	W	NP	A	0.98491073	0.40905832	2.98E-20	0.61974861	0.39075808	1.68E-33	VUS	NA
KCNJ1	3	6	FALSE	S	W	P	A	-0.7700401	0.76472234	4.44E-16	-4.5934661	0.8304699	0	VUS	NA
KCNJ11	43	31	TRUE	C	W	P	A	-0.2069305	0.22551873	2.21E-40	-2.5670829	0.40805221	1.67E-16	VUS	NA
KCNJ10	44	27	TRUE	R	W	C	A	0.11121399	0.09370754	8.67E-18	0.44337584	0.05443669	9.52E-06	VUS	NA
KCNJ2	67	67	TRUE	R	W	C	A	-0.451578	0.11157025	5.46E-08	-0.5923685	0.07908518	8.54E-16	Pathog.	NA
KCNJ5	67	72	TRUE	R	W	C	A	-0.451578	0.11157025	5.46E-08	-0.5923685	0.07908518	8.54E-16	VUS	NA
KCNJ2	82	82	TRUE	R	W	C	A	0.05653513	0.10582424	1.63E-65	0.91338758	0.57536506	0	Pathog.	NA
KCNJ2	315	307	TRUE	M	V	NP	NP	-0.5209843	0.01909854	8.04E-06	-0.0590375	0.32528496	2.84E-144	Conflicting	lof_f
KCNJ2	320	312	TRUE	R	H	C	C	-1.0766496	0.23778485	0	0.36217952	0.28633596	2.79E-05	Conflicting	lof_f
KCNJ1	130	121	TRUE	C	Y	P	A	-0.5721161	0.30153904	0	-1.393564	0.30187403	1.92E-34	VUS	NA
KCNJ2	322	314	TRUE	S	F	P	A	0.00762798	0.52934667	0	-5.6902336	0.94147175	4.44E-16	VUS	lof_s
KCNJ2	334	326	TRUE	Y	N	A	P	-0.2505895	0.25987178	2.78E-17	-2.6558446	0.11581231	3.01E-43	VUS	lof_s
KCNJ2	357	349	TRUE	E	K	C	C	0.18102372	0.08803897	4.70E-08	-1.3287225	0.26255468	2.08E-109	VUS	lof_s
KCNJ10	182	160	TRUE	F	Y	A	A	0.28509142	0.33906686	0	-0.273535	0.15651211	2.08E-17	VUS	NA
KCNJ11	186	166	TRUE	A	Y	NP	A	NA	0.02221711	6.47E-64	NA	1.02236761	4.86E-89	Pathog.	NA
KCNJ2	254	246	TRUE	D	Y	C	A	-1.1908554	0.522666	0	0.39607077	0.2609136	0	NA	NA
KCNJ1	27	30	FALSE	A	Y	NP	A	0.37204555	0.88879096	0	-0.7394914	0.84563894	1.78E-116	VUS	NA
KCNJ13	270	236	TRUE	F	Y	A	A	0.25348327	0.43131981	1.11E-16	-0.7705225	0.15576727	2.18E-38	VUS	NA
KCNJ2	297	289	TRUE	N	Y	P	A	0.53498714	0.44835728	1.11E-16	-0.6996906	0.09562638	4.59E-10	NA	NA
KCNJ2	326	318	TRUE	N	Y	P	A	0.32799972	0.07318408	7.27E-46	0.01572495	0.30684738	5.43E-112	NA	NA
KCNJ11	346	327	TRUE	K	Y	C	A	0.05951968	0.11161787	2.79E-36	-1.5363827	0.68051062	8.82E-222	VUS	NA
KCNJ2	38	38	TRUE	H	Y	C	A	-0.4344452	0.99418305	0	-1.9377759	0.48055852	0	NA	NA
KCNJ2	43	43	TRUE	C	Y	P	A	-0.0557136	0.12115544	0	0.91054931	0.22182247	0	VUS	NA
KCNJ11	73	60	TRUE	F	Y	A	A	0.05992666	0.20735178	0	-0.3808293	0.09432038	1.50E-44	Pathog.	NA
KCNJ2	78	78	TRUE	D	Y	C	A	-0.3850291	0.10987519	2.34E-59	-0.5878599	0.33285326	3.69E-55	Pathog.	NA

Supplemental Table 3: Primers used in study.

>Landing_pad_genome_amplification_for
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Landing_pad_genome_amplification_rev
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>Library_swap_to_landing_bsmbi_for
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>backbone_swap_to_landing_bsmbi_rev
acagcCGTCTCcacggggtggcgtgtagttgcgg

>Kir_geneP_Mut-1_R Frag2-51 55.8C
ATACGTCTCCCATGGTGGCcggatc
>Kir_geneP_Mut-1_F Frag2-51 57.7C
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>Kir_geneP_Mut-2_R Frag52-101 61.5C
ATACGTCTCCATCTTTCTTGACAAAGCGTGACC
>Kir_geneP_Mut-2_F Frag52-101 59.7C
ATACGTCTCCGTTTTTTGGTTGATCGCTTTGTTG

>Kir_geneP_Mut-3_R Frag102-151 59.5C
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>Kir_geneP_Mut-3_F Frag102-151 59.3C
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>Kir_geneP_Mut-4_R Frag152-201 59.0C
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>Kir_geneP_Mut-5_R Frag202-251 59.5C
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>Kir_geneP_Mut-5_F Frag202-251 59.5C
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>Kir_geneP_Mut-6_R Frag252-301 62.0C
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>Kir_geneP_Mut-6_F Frag252-301 57.5C
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>Kir_geneP_Mut-8_R Frag352-401 59.5C
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>Kir_geneP_Mut-8_F Frag352-401 58.6C
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>Kir_oligoP_Mut-1_R Frag2-51_56.7C
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>Kir_oligoP_Mut-2_F Frag52-101_57.0C
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>Kir_oligoP_Mut-2_R Frag52-101_58.2C
TCAGGCCAAAATAATCTCCGTTC

>Kir_oligoP_Mut-3_F Frag102-151_57.6C
GTACATGAAACGATGGACGGTAA

>Kir_oligoP_Mut-3_R Frag102-151_58.8C
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>Kir_oligoP_Mut-4_F Frag152-201_56.5C
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>Kir_oligoP_Mut-5_F Frag202-251_56.9C
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