Role of posterodorsal medial amygdala kisspeptin and urocortin-3 in pubertal timing in

female mice

Deyana Ivanova[1], Xiao-Feng Li[1], Yali Liu[2], Caitlin McIntyre[1], Cathy Fernandes[3,4], Geffen

Lass[1], Lingsi Kong[1] and Kevin T O'Byrne[1]

[1]Department of Women and Children's Health, Faculty of Life Science and Medicine, King's

College London, UK

[2]Department of Assisted Reproduction, Shanghai Ninth People's Hospital, Shanghai Jiaotong

University School of Medicine, Shanghai, People's Republic of China.

[3]Social, Genetic & Developmental Psychiatry Centre, Institute of Psychiatry, Psychology &

Neuroscience, King's College London, UK

[4] MRC Centre for Neurodevelopmental Disorders, King's College London, UK

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Corresponding author's contact details: Deyana Ivanova (deyana.ivanova@kcl.ac.uk) and

Kevin T O'Byrne (kevin.obyrne@kcl.ac.uk), Department of Women and Children's Health,

School of Life Course and Population Sciences, Faculty of Life Science and Medicine, King's

College London, 2.92W Hodgkin Building, Guy's Campus, London SE1 1UL, UK.

Reprint requests: Deyana Ivanova, Department of Women and Children's Health, School of

Life Course Sciences, Faculty of Life Science and Medicine, King's College London, 2.92W

Hodgkin Building, Guy's Campus, London SE1 1UL, UK.

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Abstract

Post-traumatic stress disorder impedes pubertal development and disrupts pulsatile LH secretion. Kisspeptin (kiss1) neurons in the posterodorsal sub-nucleus of the medial amygdala (MePD) are an upstream modulator of the hypothalamic gonadotropin-releasing hormone (GnRH) pulse generator, pubertal timing, as well as emotional processing and anxiety. Psychosocial stress exposure alters neuronal activity within the MePD increasing the expression of Urocortin3 (Ucn3) and its receptor corticotropin-releasing factor type-2 receptor (CRFR2) whilst enhancing the inhibitory output from the MePD to key hypothalamic reproductive centres. We test the hypothesis that psychosocial stress, processed by the MePD, is relayed to the hypothalamic GnRH pulse generator to delay puberty. Firstly, we virally infected Kiss-cre-tdTomato mice with stimulatory DREADDs targeting MePD kiss1 neurons and monitored the effect on pubertal timing. Next, we exposed C57Bl6/J mice to the predator odor, 2,4,5-Trimethylthiazole (TMT), during pubertal transition and examined the effect on pubertal timing, pre-pubertal LH pulses and anxiety-like behaviour. Subsequently, we virally infected Ucn3-cre-tdTomato mice with stimulatory DREADDs targeting MePD Ucn3 neurons and determined the effect on pubertal timing. Activation of MePD kiss1 neurons during pubertal development advanced puberty. Additionally, exposure to TMT during pubertal development delayed puberty, suppressed pre-pubertal LH pulsatility and enhanced anxiety-like behaviour, while activation of MePD Ucn3 neurons delayed puberty. Kiss1 activation in the MePD modulates the maturation of the reproductive axis advancing puberty in female mice. Moreover, early psychosocial stress exposure decreases GnRH pulse generator frequency delaying puberty whilst inducing anxiety-behaviour in female mice, an effect potentially mediated by Ucn3 neurons in the MePD.

Introduction

In humans post-traumatic stress disorder (PTSD) is associated with altered pubertal timing and the development of anxiety disorders (1,2). Predator odor exposure is a classic model for PTSD in rodents (3) and has been shown to delay puberty (4), suppress luteinizing hormone (LH) pulse frequency (5) and inhibit the pre-ovulatory LH surge (6). The gonadotropin-releasing hormone (GnRH) pulse generator that controls the hypothalamus-pituitary-gonadal (HPG) axis and pubertal development is restrained in the juvenile, but reactivated at a critical time point with increased frequency leading to the initiation of puberty (7). However, the neural mechanisms controlling pubertal timing are not fully established.

Kisspeptin (kiss1) is known to be a major gatekeeper of pubertal onset. An absence of the *kiss1* gene and kiss1 receptor (kiss1r) in mice and humans leads to a loss of gonadal maturation and a lack of pubertal onset (8,9), and treatment with kisspeptin can reactivate the HPG axis during the juvenile hiatus in monkeys and rats (10,11). Kiss1 neurons are located in the anteroventral periventricular nucleus (AVPV) and arcuate nucleus (ARC) of the hypothalamus in mice (12). The ARC kiss1 neurons, known as KNDy because they co-express neurokinin B (NKB) and dynorphin A (Dyn), are a critical component of the GnRH pulse generator (12–14). The activation of KNDy neurons induces pulsatile release of kiss1 acting on GnRH dendrons in the median eminence to stimulate pulsatile GnRH secretion (15). The AVPV kiss1 neurons project to GnRH cell bodies and proximal dendrites and are known to control the preovulatory LH surge and ovulation in rodents (16). In the juvenile, a neurobiological break exerted on the ARC GnRH pulse generator suppresses pulsatile kiss1 release in the median eminence, however the nature of this upstream inhibition of the GnRH pulse generator, which is released at the end of the juvenile phase to trigger puberty is unknown (7).

The amygdala, a stress-sensitive part of the limbic brain, is involved in processing anxiety and fear as well as modulating pubertal timing and the HPG axis. Lesioning of the amygdala

advances menarche in female rhesus macaques (17), whereas electrical stimulation of this

region delays puberty in rats (18). Specific lesioning of the posterodorsal sub-nucleus of the

medial amygdala (MePD) advances puberty in rats (19), suggesting this region may play a key

role in exerting an inhibitory break on pubertal timing. The MePD sends GABAergic

projections to various hypothalamic reproductive centres (20,21) and has been shown to project

directly to ARC KNDy neurons, although their neurochemical phenotype is undetermined

(22,23). Interestingly, the MePD contains a kiss1 neuronal population (24) and antagonism of

MePD kiss1 delays puberty, disrupts estrous cyclicity and the LH surge in rats (25). Selective

optogenetic stimulation of MePD kiss1 neurons increases LH pulse frequency in adult mice

(26) however, it is unknown whether pre-pubertal activation of MePD kiss1 neurons may be

involved in driving puberty onset. Moreover, the MePD is highly responsive to psychosocial

stress, exhibiting a distinct firing pattern in response to predator odor (27) and recently cat odor

was found to delay puberty, disrupt estrous cyclicity and induce a fear response in rats (4). The

MePD may be a central hub involved in the integration of external olfactory and anxiogenic

signals with the GnRH pulse generator.

Central administration of the stress neuropeptide corticotrophin releasing factor (CRF) dose-

dependently delays puberty, while CRF-receptor antagonism advances puberty in rats (28).

Urocortin-3 (Ucn3), a member of the CRF family and an endogenous ligand for CRF type 2

receptors (CRFR2) is found in abundance in the MePD (29). Psychosocial stressors, including

social defeat and restraint, increase cfos expression in CRFR2 positive neurons and increase

Ucn3 mRNA expression in the MePD of rodents (30). Moreover, we have recently shown that

MePD Ucn3 mediates predator odor and restraint stress-induced suppression of pulsatile LH

secretion in mice (5).

In this study, we aimed to determine whether MePD kiss1 neurons are involved in modulating

pubertal timing. We also tested the hypothesis that predator odor stress suppresses pre-pubertal

LH pulse frequency and delays pubertal timing. Additionally, we assessed social, anxiety and

fear-like behaviour as measures of psychosocial stress in predator odor-exposed pre-pubertal

female mice. Finally, we aimed to determine whether MePD Ucn3 signalling is involved in

modulating pubertal timing.

Methods

Mice

Breeding pairs of C57Bl6/J mice were purchased from Charles River Laboratories

International, Inc.   Breeding pairs of heterozygous transgenic Kiss-cre mice (strain

129S6Sv/Ev) were obtained from the Department of Physiology, Development and

Neuroscience, University of Cambridge, UK. Genotyping of Kiss-cre mice was performed

using PCR to detect heterozygosity. Heterozygous Kiss-cre mice were bred with cre-activated

tdTomato reporter mice to obtain Kiss-cre-tdTomato mice, as previously described (31). For

Ucn3-cre mice, cryopreserved sperm of strain (Tg(Ucn3-cre)KF43Gsat/Mmucd; congenic on

C57BL/6 background) was acquired from MMRRC GENSAT. Breeding pairs of heterozygous

transgenic Ucn3-cre mice were recovered by insemination of female C57Bl6/J mice at King's

College London.   Genotyping of Ucn3-cre mice was performed using PCR to detect

heterozygosity. Heterozygous Ucn3-cre mice were bred with cre-activated tdTomato reporter

mice (strain B6.Cg-Gt(ROSA)26Sortm9(CAG-tdTomato)Hze/J; congenic on C57BL/6

background obtained from The Jackson Laboratory, Bar Harbor, ME, USA) to obtain Ucn3-

cre-tdTomato mice, as previously described (5,29). Litter size was reduced to 6-8 pups 2-3

days after birth (on pnd 2-3) to standardise body weight, which can alter pubertal development.

Mice aged between 21 to 45 days were group housed in individually ventilated cages equipped

with wood-chip bedding and nesting material with food and water ad libitum and sealed with

a HEPA-filter at 25 ± 1 °C in a 12:12 h light/dark cycle, lights on at 07:00 h. All procedures

were carried out following the United Kingdom Home Office Regulations and approved by the

Animal Welfare and Ethical Review Body Committee at King's College London.

Stereotaxic adeno-associated-virus injection

All surgical procedures were carried out with aseptic conditions and under general anaesthesia

using ketamine (Vetalar, 100 mg/kg, i.p.; Pfizer, Sandwich, UK) and xylazine (Rompun, 10

mg/kg, i.p.; Bayer, Leverkusen, Germany). The mouse brain atlas of Paxinos and Franklin

(32) was used to obtain target coordinates for the MePD (2.15 mm lateral, -1.25 mm from

bregma, at a depth of -5.30 mm below the skull surface) of pre-pubertal mice. Mice at pnd 14

were temporarily taken from their mother and secured in a David Kopf stereotaxic frame (Kopf

Instruments), a small skin incision was made to reveal the skull and two small holes were

drilled above the location of the MePD. Bilateral stereotaxic viral injections of the stimulatory

adeno-associated-virus (AAV) carrying the DIO-hM3D-mCitrine, DREADD, construct

(AAV-hSyn-DIO-HA-hM3D(Gq)-IRES-mCitrine, 3×10^{11} GC/ml, Serotype:5; Addgene) was

administered intra-MePD using the robot stereotaxic system (Neurostar, Tubingen, Germany),

performed for the targeted expression of DIO-hM3D-mCitrine in MePD kiss1 neurons in Kiss-

cre-tdTomato mice and MePD Ucn3 neurons in Ucn3-cre-tdTomato mice. AAV-hSyn-DIO-

HA-hM3D(Gq)-IRES-mCitrine (150 nl) was bilaterally injected into the MePD using a 2-μl

Hamilton micro syringe (Esslab, Essex, UK) over 10 min and the needle was left in position

for a further 5 min then slowly lifted over 2 min. Mice that were cre-positive received the

AAV-hM3D injection (test mice) or a control virus AAV-YFP (Addgene) (control mice) where

the control virus does not contain the DIO-hM3D-mCitrine construct. The mice were placed

back with the mother for a further 7 days until wean day (pnd 21). Ten Kiss-cre-tdTomato

mice received the AAV-hM3D injection and 5 Kiss-cre-tdTomato mice received the control

AAV-YFP. Nine Ucn3-cre-tdTomato mice received the AAV-hM3D injection and 6 Ucn3-

cre-tdTomato mice received the control AAV-YFP.

Chronic pre-pubertal predator-odor exposure and puberty evaluation

Female pups were weaned on pnd 21 and separated into control and test groups. To investigate the effects of chronic pre-pubertal psychosocial stress mice were removed from their home cages, singly housed in new cages and left to habituate for 10 min. After the habituation period, the mice were exposed to 12 µl of 2,4,5-Trimethylthiazole (TMT; synthetic extract of fox urine; ≥98% purity; Sigma-Aldrich, UK) pipetted on a small circular piece of filter paper in a petri dish placed in the centre of the cage for 20 min. The mice were exposed to TMT daily, at random time points, for 14 days (from pnd 21 to 35). Control mice were exposed to filter paper soaked with 12 µl of ddH$_2$O water. To rule out the possibility that physiological and behavioural responses in TMT-exposed mice resulted from novelty of scent we repeated the experiment with 85 mM of ethyl vanillin (≥98% purity; Sigma-Aldrich UK) as a control scent in a separate cohort of mice. Mice were monitored from pnd 24 for vaginal opening (VO) and first estrous (FE) indicated by epithelial cell cornification. Once the occurrence of VO was detected, vaginal smears were taken to determine the exact day of FE.

Pre-pubertal blood sampling

The effect of TMT exposure on pre-pubertal LH pulses was measured, on pnd 26 and 29. Mice were handled twice daily for 10 min to habituate to tail-tip blood collection for at least 7 days prior to blood sampling, as described previously (5). For LH measurement, 3 µl of blood was collected every 5 min for 80 min. Blood sampling was performed on pnd 26 and 29 during the period of daily TMT exposure (pnd 21-35). Blood samples were collected at least 2 h before TMT-exposure on these days. Blood collection was performed between 09:00-12:00 h.

Behavioural tests

Light Dark Box

Mice were transported to a new room (lights off) and left to habituate for 30 min. Mice were placed in the centre of the light compartment facing towards the dark compartment. The time spent in each compartment (entry with all 4 limbs) was recorded manually over a period of 5 min by two independent observers. The LDB test was carried out on pnd 19, 27 and 40. Behaviour testing performed during the period of daily TMT exposure (pnd 21-35) and was measured before the exposure to TMT on that day. This applies to all behaviour tests performed during the TMT exposure period. All behaviour tests were performed between 11:00-13:00 h.

Social interaction with familiar conspecific

Mice were transported to a new dimly lit room and left to habituate for 30 min. Experimental mice and a same sex familiar conspecific (similar age, weight and strain) were placed simultaneously into the test arena (a new cage with clean wood-chip bedding). The time spent following, sniffing, grooming and mounting the conspecific was monitored and recorded manually over a period of 5 min by two independent observers. The social interaction test was carried out on pnd 19, 27 and 40.

Elevated Plus maze

Mice were transported to a new room (lights on) and left to habituate for 30 min. Mice were placed in the centre of the maze facing towards the closed arms. The time spent in each part of the maze (open, centre and closed; entry with all 4 limbs) was recorded manually over a period of 5 min by two independent observers. The EPM test was carried out on pnd 20, 28 and 41.

Chronic DREADD activation of kiss1 or Ucn3 neurons in the MePD

For DREADDs experiments, stock solution of Clozapine-N-oxide (CNO) was made by dissolving 5 mg of CNO in 1 ml of 0.9% sterile saline solution and stored at 4 °C. CNO was made fresh daily and administered via drinking water at a final concentration of 0.5 mg CNO/ kg, as described previously (33) for 14 days from pnd 21-35. Mice were monitored from pnd 22 for vaginal opening (VO) and first estrous (FE), as described above.

Validation of AAV injection

On pnd 50, Kiss-cre-tdTomato and Ucn3-cre-tdTomato mice were anaesthetised with a lethal dose of ketamine. Transcardial perfusion was performed with heparinised saline for 5 min followed by ice-cold 4% paraformaldehyde (PFA) in phosphate buffer (pH 7.4) for 15 min with a pump (Minipuls, Gilson, Villiers Le Bel, France). Brains were immediately collected and fixed in 15% sucrose in 4% PFA at 4 °C and left to sink. Brains were then transferred to 30% sucrose in phosphate-buffered saline (PBS) and left to sink. Brains were snap-frozen in isopropanol on dry ice and stored in -80°C. Every third coronal brain section, 30-μm/section, was collected using a cryostat (Bright Instrument Co., Luton, UK) through-out the MePD region corresponding to -1.34 mm to -2.70 mm from bregma. Brain sections were mounted on microscope slides, air dried and covered with ProLong Antifade mounting medium (Molecular Probes, Inc. OR, USA). The number of tdTomato labelled and AAV-hM3D-mCitrine infected kiss1 or Ucn3 neurons per side per slice in the MePD was quantified from 4 slices. For AAV-hM3D-mCitrine injected Kiss-cre-tdTomato or Ucn3-cre-tdTomato mice we determined whether kiss1 or Ucn3 neurons were infected in the MePD region by merging td-Tomato fluorescence of kiss1 or Ucn3 neurons with mCitrine fluorescence in the MePD. Images were taken using Axioskop 2 Plus microscope (Carl Zeiss) equipped with axiovision, version 4.7 (Carl Zeiss). Only data from animals with correct AAV injection were analysed.

LH pulse detection and analysis

Blood samples were processed with a LH ELISA, as previously reported (34). Capture antibody (monoclonal antibody, anti-bovine LHβ subunit, AB_2665514) was purchased from Department of Animal Science at the University of California, Davis. Mouse LH standard (AFP-5306A) and primary antibody (polyclonal antibody, rabbit LH antiserum, AB_2665533) were obtained from Harbour-UCLA (California, USA). Secondary antibody (Horseradish-Peroxidase (HRP)-linked donkey anti-rabbit IgG polyclonal antibody, AB_772206) was purchased from VWR International (Leicestershire, UK). Inter-assay and intra-assay variations were 4.6% and 10.2%, respectively. ODs of the standards were plotted against the log of the standard concentrations, non-linear regression to fit the points and parameters were extracted to calculate the concentration of LH (ng/ml) in blood samples, as previously described (34). The LH concentration at every time point of blood collection was plotted as a line and scatter graph using Igor Pro 7, Wavemetrics, Lake Oswego, OR, USA. DynPeak algorithm was used for the detection of LH pulses (35).

Statistics

For TMT-exposed and DREADD injected female pre-pubertal mice, data obtained for VO and FE was analysed using RM one-way ANOVA. Data obtained for performance on LDB, SI and EPM, LH pulse frequency and body weight measure were analysed using RM two-way ANOVA. Statistics were performed using Igor Pro 7, Wavemetrics, Lake Oswego, OR, USA. Data was represented as mean ± SEM and +$p < 0.05$, ++$p < 0.001$ and +++$p < 0.0001$ were considered to be significant.

Results

Selective DREADD activation of kiss1 neurons in the MePD advances puberty onset

Bilateral DREADD activation of MePD kiss1 neurons during the pubertal transition period, pnd 21 to 35, of Kiss-cre-tdTomato mice advanced FE without altering VO compared to non-surgery and AAV-YFP controls (Fig. 1, A and B; Control vs DREADD, +p<0.05; DREADD, n=8, control AAV, n=5, no surgery, n=5). Data for non-surgery and control AAV-YFP injected mice were combined as control since there was no significant difference between the two control groups. Body weight was unaffected between the experimental groups (Fig. 1, C; Control vs DREADD). The data from this study shows that activation of MePD kiss1 during the pubertal transition advances pubertal timing. Mice with misplaced injections (n=2) were excluded from the analysis.

Chronic TMT exposure delays puberty onset

TMT exposure during the pubertal transition period, pnd 21 to 35, delayed FE without altering VO compared water and ethyl vanillin controls (Fig. 2, A and B; Control vs TMT, +++p<0.0001; TMT, n=14, water control, n=6, ethyl vanillin control, n=4). Data for water and ethyl vanillin exposed control mice were combined as control since there was no significant difference between the two control groups. Body weight was unaffected between the experimental groups (Fig. 2, C; Control vs TMT). The data from this study shows that predator-odor stress exposure during the pubertal transition delays pubertal onset.

Chronic TMT exposure decreases pre-pubertal LH pulse frequency

TMT exposure during the pubertal transition period, pnd 21 to 35, suppressed pre-pubertal LH pulse frequency on pnd 26 and 29 compared to water and ethyl vanillin controls (Fig. 3, A-E; Control vs TMT, ++p<0.001; TMT, n=8, water control, n=6, ethyl vanillin control, n=3). On

pnd 29, the average LH pulse frequency for the control group tended to increase compared to

the control group on pnd 26 potentially marking an acceleration of GnRH pulse generator

activity approaching the onset of puberty (Fig. 3, A, C and E). The results of this experiment

are summarised in the figure 3E. Data for water and ethyl vanillin exposed control mice were

combined as control since there was no significant difference between the two control groups.

The data from this study shows that predator-odor stress exposure during the pubertal transition

inhibits pre-pubertal LH pulsatility.

Chronic TMT exposure induced long-lasting anxiety and fear-like behaviour

We investigated anxiety and fear-like behaviour induced by TMT-exposure during the pubertal

transition period, pnd 21 to 35. Anxiety and fear-behaviour were assessed using standard

anxiety and fear tests, the LDB, SI and EPM, on pnd 19 or 20 (prior to day of weaning and

TMT-exposure), pnd 27 or 28 (during the TMT exposure period) and pnd 40 or 41 (a week

after termination of TMT-exposure). The first LDB, SI and EPM trial on pnd 19 or 20, prior

to weaning, confirms no group difference in anxiety before the beginning of TMT-exposure

(Fig. 4, A-C). TMT-exposed mice spent significantly less time in the light compartment of the

LDB as well as socialising with a familiar conspecific compared to the control group on pnd

27, but not on pnd 40 (Fig. 4, A, B; Control vs TMT, $+p<0.05$; TMT, n=14, water control, n=6,

ethyl vanillin control, n=4). TMT-exposed mice spent less time in the open arms of the EPM

on pnd 28 and pnd 41 compared to controls (Fig. 4, C; Control vs TMT, $++p<0.001$; TMT,

n=14, water control, n=6, ethyl vanillin control, n=4). It is acknowledged that repeat testing

on the EPM assesses phobia/fear (36). On pnd 20, pnd 28 and 41, within-group analysis

showed no significant differences in the total time spent in the open arms of the EPM across

the different test days in the control group. However, within-group analysis of the TMT-

exposed group showed a significant difference in time-spent in the open arm of the EPM on

pnd 41 compared to pnd 28 (Fig. 4, C; TMT on pnd 28 vs TMT on pnd 41, ###p<0.0001).

TMT-exposure during pubertal transition had a long-lasting effect on anxiety and fear/phobia.

Selective DREADD activation of Ucn3 neurons in the MePD delays puberty onset

Bilateral DREADD activation of MePD Ucn3 neurons during the pubertal transition period, pnd 21 to 35, of Ucn3-cre-tdTomato mice delayed FE without altering VO compared to non-surgery and AAV-YFP controls (Fig. 5, A and B; Control vs DREADD, +$p<0.05$; DREADD, n=8, control AAV, n=6, no surgery, n=4). Data for non-surgery and control AAV-YFP injected mice were combined as control since there was no significant difference between the two control groups. Body weight was unaffected between the experimental groups (Fig. 5, C; Control vs DREADD). These data show that activation of MePD Ucn3 during the pubertal transition delays pubertal timing. Mice with misplaced injections (n=1) were excluded from the analysis

Selective expression of DREAD(Gq) in MePD kiss1 and Ucn3 neurons

Evaluation of m-Citrine, hM3D, expression in tdTomato labelled neurons from AAV-injected Kiss-cre-tdTomato mice revealed that $78 \pm 3\%$ of MePD Kiss neurons expressed hM3D and the number of tdTomato labelled kiss1 neurons per side per slice in the MePD was counted at 7.38 ± 1.66 (mean $\pm$ SEM) with the number of AAV-hM3D-mCitrine infected neurons being 5.78 ± 1.42 (mean $\pm$ SEM) (n=8). A representative example is shown in Fig 6, A-F. Evaluation of m-Citrine, hM3D, expression in tdTomato labelled neurons from AAV-injected Ucn3-cre-tdTomato mice revealed that $86 \pm 5\%$ of MePD Ucn3 neurons expressed hM3D and the number of tdTomato labelled Ucn3 neurons per side per slice in the MePD was counted at 72.90 ± 5.48 (mean $\pm$ SEM) with the number of AAV-hM3D-mCitrine infected neurons being 63.00 ± 6.83 (mean $\pm$ SEM) (n=8). A representative example is shown in Fig 7, A-F.

Discussion

The present study shows for the first time that DREADD activation of MePD kiss1 neurons from weaning on pnd 21 for 14 days advances puberty onset, revealing a crucial stimulatory role of MePD kiss1 signalling in regulating the HPG axis during puberty. Additionally, chronic exposure to the psychogenic stressor, predator odor, from pnd 21 for 14 days suppresses the pubertal acceleration of LH pulse frequency, delaying puberty onset, whilst concurrently inducing long-lasting anxiety and fear-like behaviour. Moreover, DREADD activation of MePD Ucn3 neurons during this same developmental period delays puberty onset, demonstrating that MePD Ucn3 signalling may play an important role in stress-related changes in pubertal timing.

Hypothalamic kiss1 signalling is pivotal in regulating puberty onset in various species, including rodents and humans (7,11). Pubertal onset is thought to be timed by an increase in GnRH pulse generator frequency, however the mechanisms underlying the timing of ARC kiss1 neuronal activation to trigger pubertal onset are not well established. The amygdala is known to influence pubertal timing in primates (17), but more specifically, the MePD has been shown to regulate pubertal timing and reproductive function in rodents (4,19), vis-à-vis MePD kiss1 receptor antagonism delays puberty, reduces the occurrence of preovulatory LH surges in rats (25) and MePD kiss1 neuronal activation heightens sexual partner preference in male mice (37). Recently, we have shown optogenetic stimulation of MePD kiss1 neurons increases GnRH pulse generator frequency in adult female mice (26), and in the present study we show that DREADD activation of MePD kiss1 neurons during the critical pubertal development period advances FE without affecting VO or body weight in female mice. In mice, VO and FE do not coincide, wherein VO is not tightly coupled to the first ovulation, but rather a variable time gap exists between VO and FE; the latter associated with first ovulation and considered the unequivocal marker of puberty (38). Although the present study clearly shows an

advancement of puberty onset following activation of MePD kiss1 neurons, the underlying

neural networks engaged remain to be established.

In mice, the maturation of the kiss1 system begins with an increase in the number of

hypothalamic kiss1 neurons in peripubertal mice (39) occurring concomitantly with an increase

in connectivity between kiss1 and GnRH neurons which is considered critical for normal

puberty onset (40). Kiss1 expression in the rostral periventricular area of the third ventricle

increases across the pubertal transition in mice (41) and knockdown of AVPV kiss1 delays

puberty and reduces the occurrence of LH surges (42). In ovine species, exposure to a sexually

active ram can advance puberty (43) and stimulate ovulation in anoestrous ewes via the

resurgence of pulsatile and surge release of LH coinciding with elevated cfos expression in

kiss1 neurons of the ARC, pre-optic area (POA) and medial amygdala (44,45). MePD kiss1

neurons have been shown to project directly to GnRH neurons in the medial POA (46), hence

revealing a possible route via which MePD kiss1 neurons may control puberty. However,

puberty onset has been associated with increased ARC Tac2 (encoding NKB) and kiss1 mRNA

expression in mice (47,48), which are critical components of the GnRH pulse generator.

Moreover, there is an increase in ARC kiss1 promoter activity during pubertal development

(49). We know direct neural projections, of unknown phenotype, extend from the MePD to

ARC kiss1 neurons (22,23) and MePD kiss1 signalling functions as an upstream regulator of

the ARC KNDy network, accelerating GnRH pulse generator frequency (26,50). Therefore,

an alternative route may involve upstream MePD kiss1 signalling accelerating ARC KNDy

neuronal activity to trigger pubertal onset.

Stress exposure during the juvenile period is known to alter pubertal timing and induce anxiety

behaviour in mammals, including rodents, primates and humans (1,2,4,51). Predator odor

exposure in rodents is a model for PTSD; increasing fear and anxiety behaviour as well as

reducing social interaction (3). Previously, we have shown that acute TMT exposure

suppresses LH pulsatility (5) and restraint stress, another psychological stressor, inhibits LH

pulse frequency while reducing ARC kiss1 cfos expression in adult mice (52). Moreover, we

have shown that cat odor delays puberty and induces a fear response in rats (4). In the present

study we show that the predator odor, TMT, reduces LH pulsatility and delays puberty in mice.

Pre-pubertal LH pulse frequency in the control group tended to increase potentially marking

the acceleration of the GnRH pulse generator as the time of puberty onset approaches.

Contrastingly, pre-pubertal LH pulse frequency in the TMT-exposed group was significantly

lower compared to controls and we did not observe an increase in GnRH pulse generator

activity between pnd 26 and 29.

Puberty is a sensitive period where enhanced neuroplasticity provides a context for the impact

of stress on the development of long-term anxiety disorders in humans (53). Early-life stress

increases anxiety-like behaviour in juvenile and adult mice linked to hyper-connectivity

between fronto-limbic circuits, involving the amygdala (54). The medial amygdala is activated

in response to external stressors and electrophysiological recordings reveal that predator urine

robustly activates the MePD, which is associated with delaying pubertal onset (4) and

supressing LH pulsatility in rodents (5,55). We found that predator odor-induced pubertal

delay was accompanied by increased anxiety-like behaviour, with mice spending significantly

less time in the light compartment of the LDB and in the open arms of the EPM during stress

exposure. Moreover, we observed that mice exposed to TMT exhibited long lasting fear-like

behaviour where they spent significantly less time in the open arm of the EPM compared to

controls on pnd 41; 6 days after termination of stress exposure. This is consistent with our

previous studies showing rats exposed to cat odor during pubertal development display

increased long-lasting fear-like behaviour on the EPM even after the termination of stress

exposure (4). Moreover, exposure to fox odor induces long-lasting anxiety on the EPM in rats

(56), while mice exposed to foot-shock stress during adolescence exhibit increased startle

reflexes into adulthood indicative of lasting anxiety-like behaviour (57). Additionally, TMT-

exposed mice showed significantly reduced social interaction with familiar con-specifics

during stress exposure whereas sociability tended to increase along with reproductive

maturation in the control group, which is consistent with our previous observations where

exposure to cat odor during puberty decreased sociability in rats (4).

Central administration of CRF inhibits kiss1 expression in the ARC and POA in rats (58). In

women with functional hypothalamic amenorrhea there is a correlation between decreased

serum kisspeptin and increased CRF concentrations (59). Moreover, an endogenous CRF tone

regulates pubertal timing where central administration of CRF in pnd 28 rats delays puberty

onset and administration of astressin-B, a CRFR1 antagonist, advances puberty (28). Central

antagonism of CRFR2 blocks the suppressive effect of stress on LH pulsatility in rodents (60).

These data indicate stress neuropeptides may provide an inhibitory tone on the timing of

puberty. Predator odor activates CRFR2 positive neurons in the medial amygdala of rats (61)

and we have recently shown that TMT-induced suppression of LH pulsatility is mediated by

MePD Ucn3 and CRFR2 signalling in mice (5). In the present study, selective activation of

MePD Ucn3 neurons during pubertal development delayed puberty, demonstrating that MePD

Ucn3 signalling plays a major role in regulating pubertal timing. The MePD has a major

GABAergic output to key hypothalamic reproductive nuclei (21,62) and we know the

amygdala exerts an inhibitory brake on pubertal timing (19,63). Ucn3 neurons in the medial

amygdala have an interneuron-like appearance (64) where Ucn3 fibres overlap with CRFR2

expression (65,66) and we have shown that antagonism of MePD CRFR2 blocks the

suppressive effect of TMT on LH pulses in adult mice (5), thus Ucn3 neurons possibly connect

to and signal via CRFR2 within the MePD. Moreover, the majority of MePD CRFR2 neurons

co-express GAD65 and 67 (29) indicating they are GABAergic and may be involved in

mediating stress-induced suppression of the GnRH pulse generator (5). MePD Ucn3 activity

modulates puberty possibly via enhancing GABAergic output whilst suppressing kiss1

signalling within the MePD, as antagonism of kisspeptin receptor signalling in the MePD

decreases GnRH pulse generator frequency (50) while delaying puberty (25).

Our findings show for the first time that MePD kiss1 signalling plays a fundamental role in

controlling pubertal timing. Moreover, early life exposure to a psychosocial stressor disrupts

GnRH pulse generator frequency to delay puberty, and Ucn3 signalling in the MePD may be

involved in mediating this response. These findings provide novel insight into the key

interactions between the emotional stress centres and reproductive centres in the brain that

control pubertal onset in response to the external environment. Alterations in the timed onset

of puberty has major implications later in life with increased risk of diverse adverse outcomes,

including cancer, gynaecologic/obstetric, cardio-metabolic and neuro-cognitive categories in

humans (67). Understanding the mechanisms involved in controlling pubertal timing and the

impact of stress is crucial to ultimately aid future development of more effective treatments for

stress-related disorders of puberty.

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Figure 1. DREADD activation of kisspeptin (kiss1) neurons in the posterodorsal sub-nucleus of the medial amygdala (MePD) from post-natal day (pnd) 21 for 14 days advances first estrous (FE) without affecting vaginal opening (VO) and body weight (BW) in Kiss-cre-tdTomato female mice. Effect of MePD kiss1 activation on day of VO (A), FE (B) and (C) BW weight gain between pnd 21 and 45. +$p < 0.05$ Control group (combined control AAV: n=5; no surgery: n=5) vs DREADD group (n=8).

Figure 2. Chronic 2,4,5-Trimethylthiazole (TMT) exposure from post-natal day (pnd) 21 for 14 days delays first estrous (FE) without affecting vaginal opening (VO) and body weight (BW) in C57Bl6/J female mice. Effect of chronic TMT exposure on day of VO (A), FE (B) and (C) BW weight gain between pnd 21 and 45. +++$p < 0.0001$ Control group (combined water control: n=6; ethyl vanillin control: n=4) vs TMT group (n=14).

Figure 3. Chronic 2,4,5-Trimethylthiazole (TMT) exposure from post-natal day (pnd) 21 for 14 days suppresses pre-pubertal pulsatile luteinising hormone (LH) secretion on pnd 26 and 29 in C57Bl6/J female mice. Representative LH pulse profile with (A) control mouse on pnd 26, (B) TMT-exposed mouse on pnd 26, (C) control mouse on pnd 29 and (D) TMT-exposed mouse on pnd 29. (E), Summary of LH pulse frequency on pnd 26 and 29 of control and TMT-exposed group during the 80 min blood sampling period. LH pulses detected by the DynePeak algorithm are indicated with an asterisk located above each pulse on the representative LH pulse profiles. ++$p < 0.001$ control group (combined water control: n=6; ethyl vanillin control: n=3) vs. TMT-exposed group (n=8) on pnd 26 and 29.

Figure 4. Chronic 2,4,5-Trimethylthiazole (TMT) exposure increased anxiety-like behavior on the light-dark box (LDB) and elevated plus maze (EPM) and decreased social interaction with familiar conspecifics on the social interaction (SI) test. (A) Summary of time spent in the

light compartment of the LDB on post-natal day (pnd) 19 (before beginning of TMT-exposure),

27 (during TMT exposure) and 40 (after termination of TMT-exposure), (B) time spent socially

interacting with familiar conspecifics on pnd 19, 27 and 40, and (C) time spent in the open arm

of the EPM on pnd 20, 28 and 41. $+p<0.05$ time spent on LDB, SI and EPM of control group

(combined water control: n=6; ethyl vanillin control: n=4) vs TMT-exposed group (n=14) on

pnd 27 or 28 (during TMT-exposure); $\#\#\#p<0.0001$ time spent in the open arm of the EPM in

the TMT-exposed group on pnd 28 vs pnd 41.

Figure 5. DREADD activation of urocortin 3 (Ucn3) neurons in the posterodorsal sub-nucleus

of the medial amygdala (MePD) from post-natal day (pnd) 21 for 14 days delays first estrous

(FE) without affecting vaginal opening (VO) and body weight (BW) in urocortin3-cre-

tdTomato female mice. Effect of MePD Ucn3 activation on day of VO (A), FE (B) and (C)

BW weight gain between pnd 21 and 45. $+p<0.05$ Control group (combined control AAV: n=6;

no surgery: n=4) vs DREADD group (n=8).

Figure 6. Expression of AAV5-hSyn-DIO-HA-hM3D(Gq)-IRES-mCitrine in posterodorsal

medial amygdala (MePD) Kisspeptin (kiss1) neurons. (A-F) Representative dual fluorescence

photomicrographs of the MePD from a Kiss-cre-tdTomato female mouse injected with AAV5-

hSyn-DIO-HA-hM3D(Gq)-IRES-mCitrine. kiss1 neurons labelled with m-Citrine (A) and

tdTomato (C) appear yellow/orange (E). (B), (D) and (F) are a higher power view of (A), (C)

and (E) respectively. Scale bars represent A, C, E 100 µm and B, D, F 25 µm; OT, Optic tract

(blue line).

Figure 7. Expression of AAV5-hSyn-DIO-HA-hM3D(Gq)-IRES-mCitrine in posterodorsal

medial amygdala (MePD) Urocortin 3 (Ucn3) neurons. (A-F) Representative dual fluorescence

photomicrographs of the MePD from a Ucn3-cre-tdTomato female mouse injected with

AAV5-hSyn-DIO-HA-hM3D(Gq)-IRES-mCitrine. Ucn3 neurons labelled with m-Citrine (A)

and tdTomato (C) appear yellow/orange (E). (B), (D) and (F) are a higher power view of (A),

(C) and (E) respectively. Scale bars represent A, C, E 100 μm and B, D, F 25 μm; OT, Optic

tract (blue line).

Figure 1

Figure 2

Figure 3

Figure 4

Figure 5

A Vaginal Opening

B First Estrous

C

Figure 6

Figure 7