- 1 Title: In vitro effect of a non-immunosuppressive FKBP ligand, FK1706, on SARS-CoV-2
- 2 replication in combination with antivirals
- 3 Running Title: FK1706-remdesivir in vitro synergy against SARS-CoV-2
- 4 William E. Fitzsimmons^{a,b} #, Tracy L. Hartman^c, Michelle Mendenhall^d, Catherine Z. Chen^e
- 5 a. University of Illinois at Chicago, Chicago, IL
- 6 b. Tutela Pharmaceuticals Inc, Vernon Hills, IL
- 7 c. ImQuest Biosciences, Frederick, MD;
- 8 d. Institute for Antiviral Research, Utah State University, Logan, UT;
- 9 e. National Center for Advancing Translational Sciences, NIH, Rockville, MD;
- 10
- 11 #Address correspondence to: William E. Fitzsimmons
- 12 wfitzsim@uic.edu
- 13 *Present Address: University of Illinois at Chicago, College of Pharmacy
- 14 833 S. Wood Street
- 15 Chicago, IL 60612
- 16
- 17
- 18
- 19
- 20
- 21
- 22
- 23
- 23 24
- 25
- 25
- 27
- 28
- 29
- 30
- 31
 - 31
- 32

33 Abstract

- 34 FKBP, a naturally occurring ubiquitous intracellular protein, has been proposed as a potential
- 35 target for coronavirus replication. A non-immunosuppressive FKBP ligand, FK1706, was studied
- in vitro in a Vero cell model to assess potential activity alone and in combination with antivirals
- against SARS-CoV-2 replication. When combined with remdesivir, synergistic activity was seen
- 38 (summary synergy score 24.7<u>+</u>9.56). FK1706 warrants in vivo testing as a potential new
- 39 combination therapeutic for the treatment of COVID-19 infections.
- 40
- 41
- -
- 42
- 43
- 44
- 45
- 46
- 47
- 48
- 49
- 50
- 51
- 52
- 53
- 54
- ___
- 55

56 FKBP is one of the naturally occurring ubiquitous intracellular proteins called immunophilins that

57 has enzymatic activity as a peptidyl prolyl cis-trans isomerase and is also essential to the

58 pharmacologic activity of immunosuppressants. The binding of tacrolimus, everolimus, and

sirolimus, to FKBP is necessary but not sufficient to produce immunosuppression (1,2).

60 Replication of human coronaviruses is dependent on active immunophilin binding and inhibition

of cyclophilins, an intracellular immunophilin, by cyclosporine blocks the replication of CoVs of

all genera tested, including SARS-CoV, human CoV-229E and -NL-63, feline CoV, as well as

avian infectious bronchitis virus (3-6). More recently, the immunophilin FKBP has been

64 described as one of the potential targets for SARS-CoV-2 (7,8).

Two ligands to FKBP that are not immunosuppressive, FK1706 (9,10) and ElteN378 (11,12)

66 were studied. These compounds are structurally distinct; both bind to the core structure for

67 FKBP but do not have intact calcineurin or mTOR binding domains that produce

68 immunosuppression. Because these drugs target host cells and may work by a unique

69 mechanism to inhibit coronavirus replication, the additive or synergistic effect with known virus-

targeting antivirals with mechanisms of RNA polymerase inhibition (e.g., remdesivir), viral error

catastrophe or viral lethal mutagenesis (e.g., molnupiravir), or protease inhibition (e.g.,

72 M128533) were evaluated.

73 Vero E6 cells were infected with the live SARS-CoV-2 virus (USA-WA1/2020; World Reference

74 Center for Emerging Viruses and Arboviruses (WRCEVA)) at low MOI (multiplicity of infection)

and multiple rounds of viral replication occurred over the course of the assay. Percent CPE in

compound-treated virus-infected cells were normalized to infected untreated cells as 0% and

uninfected cells as 100% CPE protection. Based on these data, a concentration-response curve

78 was created. Toxicity was assessed and compared in untreated, uninfected cells compared to79 treated cells.

80 In vitro testing was conducted at two independent laboratories in sequence. The details of the 81 protocol followed by each laboratory are included in the appendix materials.

82 FK1706 (Shanghai SIMR Biotechnology Co. LQY20200910), ElteN378 (Glixx Laboratories Inc.

83 GLXC -20448), remdesivir, molnupiravir, and M128533 were solubilized in DMSO and were

84 diluted in culture test media to prepare compound concentrations.

85 Synergy was calculated using SynergyFinder 2.0 software (13). A summary synergy score

86 greater than 10 was considered synergistic.

87 The initial results of FK1706 alone and in combination with remdesivir, molnupiravir, and

- 88 M128533 are summarized in Table 1.
- 89 When combined, FK1706 (11-90 μ M) and remdesivir (3 μ M) were effective in inhibiting SARS
- 90 CoV-2 viral CPE (93-100%, see Appendix Fig A1). FK1706 (2.85-90 μM) and molnupiravir (0.3
- μ M) inhibited SARS CoV-2 CPE (up to 70% reduction in viral CPE at 90 μ M FK1706 with 0.3
- μM molnupiravir; see Appendix Fig A2). FK1706 (11-90 μM) and M128533 (1 $\mu g/mL$) reduced
- 93 SARS CoV-2 CPE (64-100%, see Appendix Fig A3).
- 94 Although FK1706 alone did not exhibit inhibitory activity against SARS-CoV-2, when combined
- 95 with suboptimal concentrations (less than the EC_{50}) of all three antivirals, increased inhibition
- 96 was observed. Additive effects of ElteN378 with either remdesivir or M128533 were also
- 97 demonstrated (see Appendix Table A1).
- 98 In follow-up confirmatory combination studies, FK1706 at multiple concentrations was tested in
- 99 combination with multiple concentrations of remdesivir. When combined, remdesivir and
- 100 FK1706 exhibited synergistic activity inhibiting SARS-CoV-2 and shifting the EC50 value of both
- 101 compounds when in combination with the other (Figures 1A,B). The summary synergy scores
- were 24.7 ± 9.56 by the ZIP (Supplementary Fig A4,A5), 24.8 ± 9.56 by the Bliss and 24.9 ± 9.56
- 103 by the HSA models. Scores >10 in all 3 models indicate synergy.
- Molnupiravir alone, nor in combination with FK1706, did not demonstrate activity in the follow-up confirmatory study. There was no evidence of cytotoxicity with FK1706 or remdesivir alone or in combination (Appendix Figs A6,A7).
- 107 The synergistic effects of FK1706 in combination remdesivir were demonstrated in a live SARS-
- 108 CoV-2 virus assay measuring the ability of compounds to inhibit viral-induced CPE in Vero E6
- 109 host cells in vitro. The CPE reduction assay is a popular and widely used assay format to
- screen for antiviral agents because of its ease of use in quantitative high-throughput screening.
- 111 The CPE reduction assay indirectly monitors the ability of compounds to inhibit viral replication
- and infection through various mechanisms, including direct inhibition of viral entry or enzymatic
- processes as well as acting on host pathways that modulate viral replication. This assay was
- 114 previously used to screen 8,810 approved and investigational drugs from the National Center for
- Advancing Translational Sciences (NCATS) small molecule collections (14). A cytotoxicity
- 116 counter-screen was conducted in parallel in host cells without addition of virus and
- demonstrated no substantial cytotoxicity of any of the test agents alone or in combination.

Since two chemically distinct FKBP ligands, FK1706 and ElteN378, both demonstrated activity,

- it is likely that FKBP is the key target. This target is in the host cells and complements the virus-
- targeted antivirals. The combination activity of these FKBP ligands was not limited to a single
- 121 virus-targeted mechanism as the three antivirals have distinctly different mechanisms.
- 122 Remdesivir (Veklury), currently the only FDA-approved antiviral for COVID-19 infections, is
- administered intravenously to patients (15). Molnupiravir has received Emergency Use
- Authorization as oral therapy for outpatient COVID-19 infections (16). Although both of these
- antivirals have demonstrated clinical efficacy, there is a need for higher response rates and
- 126 FK1706 may have utility in both settings. Additionally, these combinations should be active
- against variants with mutations in spike protein.
- Both live virus assays use Vero E6 as host cells. Vero E6 cells have been shown to have high
- drug efflux transporter P-glycoprotein (P-gp) activity, which can reduce cellular concentrations of
- test articles, and remdesivir is a known P-gp substrate (17). Therefore, synergy observed in
- 131 Vero E6 cells could be due to P-gp inhibition, which enhances the exposure of remdesivir *in*
- 132 *vitro*, and warrants repeating in other cell-based models.
- 133 FK1706 has completed all nonclinical safety pharmacology, ADME, and GLP toxicity studies to
- 134 support clinical development. Phase 1 healthy volunteer and Phase 2 studies in patients with
- neuropathy have been completed (9). This clinical experience would expedite the introduction of
- 136 FK1706 into clinical studies of patients infected with SARS-CoV-2.
- 137 In conclusion, these data demonstrate that FKBP is a valid target for coronavirus infections in
- 138 combination with virus-targeted antivirals such as remdesivir and molnupiravir. FK1706 warrants
- testing in an in vivo animal model of SARS-CoV-2 and if promising, rapid introduction into
- 140 COVID-19 infection clinical trials.
- 141
- 142
- 143
- 144
- 145
- 146
- 170
- 147

148 Acknowledgements:

149 The authors thank Diane M. Coniglio, Pharm.D., President, Opus Medical Communications for

- 150 editorial assistance with the manuscript.
- 151

152 **Funding:**

- 153 This research was funded by Tutela Pharmaceuticals Inc., a 501(c)(3) not-for-profit
- 154 pharmaceutical company. William E. Fitzsimmons is the Founder and Chair of Tutela
- 155 Pharmaceuticals Inc. This work was supported by the Intramural Research Program of National
- 156 Center for Advancing Translational Sciences, Sciences, National Institutes of Health.
- 157

158 **Figure Captions:**

- 159 Figure 1A. Concentration response of FK1706 when combined with remdesivir (RDM).
- 160 Figure 1B. Concentration response of remdesivir when combined with FK1706.
- 161
- 162
- 163
- 164
- 165
- 166
- 167
- 168
- 169
- 170
- ____
- 171
- 172
- 173
- _
- 174

175 **References:**

- 176 1. Fitzsimmons WE. 2012. Tacrolimus. In: Kaplan B., Burckart G.J., Lakkis F.G., eds.
- 177 Immunotherapy in Transplantation Principles and Practice. Wiley-Blackwell. Chichester, West
- 178 Sussex, UK, pp. 224-240.

179

Tamura K, Fujimura T, Iwasaki K, Sakuma S, Fujitsu T, Nakamura K, Shimomura K, Kuno T,
 Tanaka C, Kobayashi M. 1994. Interaction of tacrolimus (FK506) and its metabolites with FKBP
 and calcineurin. Biochem Biophys Res Commun. 202:437-43.

183

- 184 3. Carbajo-Lozoya J, Müller MA, Kallies S, Thiel V, Drosten C, von Brunn A. 2012. Replication
- of human coronaviruses SARS-CoV HCoV-NL63 and HCoV-229E is inhibited by the drug
- 186 FK506. Virus Res. 165: 112-117.

187

- 4. de Wilde AH, Falzarano D, Zevenhoven-Dobbe JC, Beugeling C, Fett C, Martellaro C,
- 189 Posthuma CC, Feldmann H, Perlman S, Snijder EJ. 2017. Alisporivir inhibits MERS-and SARS-
- 190 coronavirus replication in cell culture, but not SARS-coronavirus infection in a mouse model.
- 191 Virus Res. 228: 7-13.

192

- 5. Pfefferle S, Schöpf J, Kögl M, Friedel CC, Müller MA, Carbajo-Lozoya J, Stellberger T, von
 Dall'Armi E, Herzog P, Kallies S, Niemeyer D, Ditt V, Kuri T, Züst R, Pumpor K, Hilgenfeld R,
- 195 Schwarz F, Zimmer R, Steffen I, Weber F, Thiel V, Herrler G, Thiel HJ, Schwegmann-Wessels
- 196 C, Pöhlmann S, Haas J, Drosten C, von Brunn A. 2011. The SARS-coronavirus-host
- 197 interactome:identification of cyclophilins as target for pan-coronavirus inhibitors. PLoS Pathog.
- 198 7(10):e1002331.

199

6. Tanaka Y., Sato Y., Sasaki T. 2013. Suppression of coronavirus replication by cyclophilin
inhibitors. Viruses. 5:1250-60.

202

7. Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, White KM, O'Meara MJ, Rezelj VV,
Guo JZ, Swaney DL, Tummino TA, Hüttenhain R, Kaake RM, Richards AL, Tutuncuoglu B,

Foussard H, Batra J, Haas K, Modak M, Kim M, Haas P, Polacco BJ, Braberg H, Fabius JM, 205 Eckhardt M, Soucheray M, Bennett MJ, Cakir M, McGregor MJ, Li Q, Meyer B, Roesch F, Vallet 206 207 T, Mac Kain A, Miorin L, Moreno E, Naing ZZC, Zhou Y, Peng S, Shi Y, Zhang Z, Shen W, Kirby IT, Melnyk JE, Chorba JS, Lou K, Dai SA, Barrio-Hernandez I, Memon D, Hernandez-Armenta 208 209 C, Lyu J, Mathy CJP, Perica T, Pilla KB, Ganesan SJ, Saltzberg DJ, Rakesh R, Liu X, Rosenthal SB, Calviello L, Venkataramanan S, Liboy-Lugo J, Lin Y, Huang XP, Liu Y, 210 Wankowicz SA, Bohn M, Safari M, Ugur FS, Koh C, Savar NS, Tran QD, Shengjuler D, Fletcher 211 SJ. O'Neal MC. Cai Y. Chang JCJ. Broadhurst DJ. Klippsten S. Sharp PP. Wenzell NA. 212 213 Kuzuoglu-Ozturk D, Wang HY, Trenker R, Young JM, Cavero DA, Hiatt J, Roth TL, Rathore U, Subramanian A, Noack J, Hubert M, Stroud RM, Frankel AD, Rosenberg OS, Verba KA, Agard 214 215 DA, Ott M, Emerman M, Jura N, von Zastrow M, Verdin E, Ashworth A, Schwartz O, d'Enfert C, Mukherjee S, Jacobson M, Malik HS, Fujimori DG, Ideker T, Craik CS, Floor SN, Fraser JS, 216 Gross JD, Sali A, Roth BL, Ruggero D, Taunton J, Kortemme T, Beltrao P, Vignuzzi M, García-217 218 Sastre A, Shokat KM, Shoichet BK, Krogan NJ. 2020. A SARS-CoV-2 protein interaction map

reveals targets for drug repurposing. Nature. 583(7816):459-468.

220

8. Shigdel UK, Lee SJ, Sowa ME, Bowman BR, Robison K, Zhou M, Pua KH, Stiles DT,

Blodgett JAV, Udwary DW, Rajczewski AT, Mann AS, Mostafavi S, Hardy T, Arya S, Weng Z,

223 Stewart M, Kenyon K, Morgenstern JP, Pan E, Gray DC, Pollock RM, Fry AM, Klausner RD,

Townson SA, Verdine GL. 2020. Genomic discovery of an evolutionarily programmed modality

for small-molecule targeting of an intractable protein surface. Proc Natl Acad Sci USA.

226 117:17195-17203.

227

9. Minematsu T, Lee J, Zha J, Moy S, Kowalski D, Hori K, Ishibashi K, Usui T, Kamimura

229 H.2010. Time-dependent inhibitory effects of

230 (1R,9S,12S,13R,14S,17R,18E,21S,23S,24R,25S,27R)-1,14-dihydroxy-12-(E)-2-[(1R,3R,4R)-4-

hydroxy-3-methoxycyclohexyl]-1-methylvinyl-23,25-dimethoxy-13,19,21,27-tetramethyl-17-(2-

232 oxopropyl)-11,28-dioxa-4-azatricyclo[22.3.1.0(4.9)]octacos-18-ene-2,3,10,16-tetrone (FK1706),

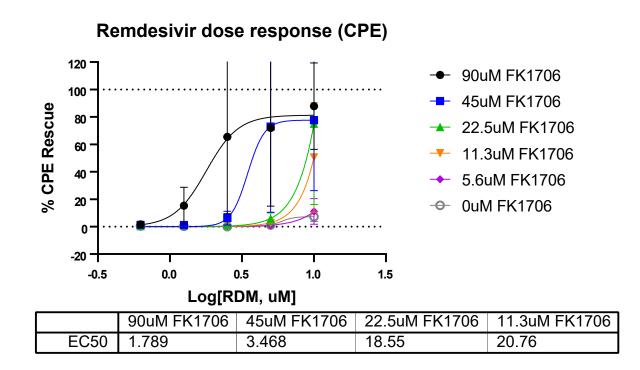
a novel nonimmunosuppressive immunophilin ligand, on CYP3A4/5 activity in humans in vivo

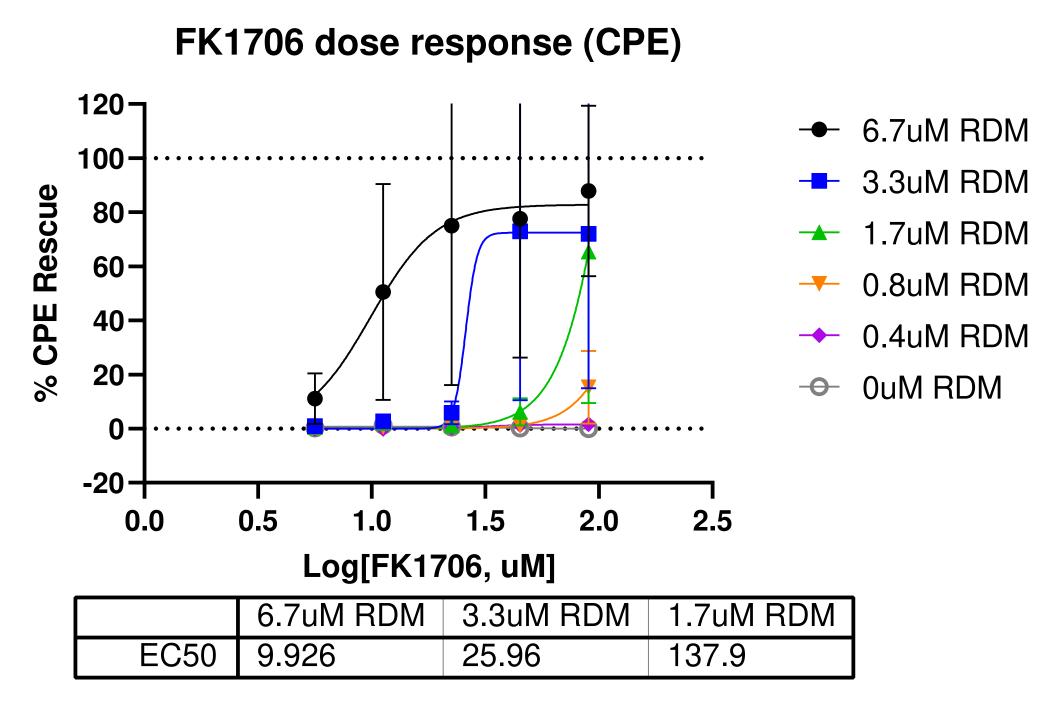
and in vitro. Drug Metab Dispos. 38:249-59.

235

| 236 237 238 | 10. Price RD, Yamaji T, Yamamoto H, Higashi Y, Hanaoka K, Yamazaki S, Ishiye M, Aramo Matsuoka N, Mutoh S, Yanagihara T, Gold BG. 2005. FK1706, a novel non-immunosuppres immunophilin: neurotrophic activity and mechanism of action. Eur J Pharmacol. 509:11-9. | | |
|---|---|--|--|
| 239 240 241 242 243 | 11. Caminati G, Martina MR, Menichetti S, Procacci P. 2019. Blocking the FKBP12 induced dendrimeric burst in aberrant aggregation of a-synuclein by using the ElteN378 synthetic inhibitor. J Enz Inhib Med Chem. 34: 1711-1715. | | |
| 244 245 246 247 | 12. Martina MR, Tenori E, Bizzarri M, Menichetti S, Caminati G, Procacci P. 2013. The precise chemical-physical nature of the pharmacore in FK506 binding protein inhibition: ElteX, a New class of nanomolar FKBP12 ligands. J Med Chem. 56:1041-51. | | |
| 248 249 250 | 13. Ianevski A., Giri A.K., Aittokallio T. 2020. SynergyFinder 2.0: visual analytics of multi-drug combination synergies.Nucleic Acids Research.48:W488–W493. | | |
| 251 252 253 254 255 | 14. Chen CZ, Shinn P, Itkin Z, Eastman RT, Bostwick R, Rasmussen L, Huang R, Shen M, Hu X, Wilson KM, Brooks BM, Guo H, Zhao T, Klump-Thomas C, Simeonov A, Michael SG, Lo DC, Hall MD, Zheng W. 2021. Drug repurposing screen for compounds inhibiting the cytopathic effect of SARS-CoV-2. Front Pharmacol. 11:592737. | | |
| 256 257 258 | 15. Rubin D, Chan-Tack K, Farley J, Sherwat A. 2020. FDA approval of remdesivir- a step in the right direction. N. Engl. J. Med. 383:2598-2600. | | |
| 259 260 261 262 263 264 | 16. Jayk Bernal A, Gomes da Silva MM, Musungaie DB, Kovalchuk E, Gonzalez A, Delos Reyes V, Martín-Quirós A, Caraco Y, Williams-Diaz A, Brown ML, Du J, Pedley A, Assaid C, Strizki J, Grobler JA, Shamsuddin HH, Tipping R, Wan H, Paschke A, Butterton JR, Johnson MG, De Anda C; MOVe-OUT Study Group. 2021. Molnupiravir for oral treatment of Covid-19 in nonhospitalized patients. N Engl J Med. 2021 Dec 16:NEJMoa2116044. | | |

- 265 17. Kumar D., Trivedi, N. 2021. Disease-drug and drug-drug interaction in COVID-19: Risk and
- assessment. Biomed Pharmacother. 2021.139: 111642.





| Table 1. Anti-SARS-CoV-2 Cytoprotection Assay Results for FK1706 and antivirals |
|---|
| against SARS-CoV-2 (USA-WA1/2020). |

| Compound or combination | EC ₅₀ (µM) | TC ₅₀ (μM) | TI |
|--------------------------------|-----------------------|-----------------------|-------|
| FK1706 | >90 | >90 | |
| FK1706 + Remdesivir (3 µM) | <11.3 | >90 | >7.96 |
| Remdesivir (3 µM) | >3 | >3 | |
| Remdesivir | 3.63 | >100 | >27.5 |
| FK1706 + M128533 (1 µg/mL) | <11.3 | >90 | >7.96 |
| M128533 (1 μM) | >1 | >1 | |
| M128533 (µg/mL) | 1.53 | 86.8 | 56.7 |
| FK1706 + Molnupiravir (0.3 µM) | 28.7 | >90 | >3.14 |
| Molnupiravir (single conc.) | >0.3 | >0.3 | |