Running Head: Can livestock be functional substitutes of native herbivores?

Functional substitutability of native herbivores by livestock for long-term soil carbon

sequestration depends on microbial decomposers.

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S.R. and S.B. designed research; S.R. performed research; S.R. and S.B. analysed data; and

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ABSTRACT

The influence of native mammalian herbivores on ecosystem functions is declining as

domestic livestock have displaced them to become the dominant grazers across the world.

Alongside rewilding, restoration, and conservation, it is now important to address whether

livestock can assume the functional roles of displaced native herbivores in grasslands,

savannas, and steppes. If yes, this has implications for the future of biodiversity, ecosystem

functions and services, food-security, and other social and environmental dimensions. We

test competing alternative hypotheses with a replicated, long-term, landscape-level

experiment to ask whether livestock in the Trans-Himalayan ecosystem can match decadal-

scale (2006-16) soil-C sequestration under native herbivores. Despite comparable settings,

livestock were imperfect substitutes and could only attain three-fourths of soil-C stocks under

native herbivores. We evaluate multiple lines of evidence surrounding this difference with a

grazing-exclusion experiment (nutrient-cycling, microbial metabolism, and e-DNA), and

asses their inter-relationships. Livestock and native herbivores differed in their effects on

several soil microbial variables. Structural equation models supported the hypotheses that

livestock alter soil microbial communities towards lower fungal abundance, which is

detrimental for microbial carbon use efficiency (CUE) and ultimately for soil-C storage. Lags

in soil-C storage can be addressed through microbial restoration and rewilding of soil.

Therefore, the idea of functional substitutability must be leveraged alongside microbial

rewilding to restore and conserve ecosystem functions. Together they can reconcile

conflicting demands from food-security and ecosystem services in ways that have eluded

conventional approaches to become an integral part of nature-based climate solutions.

INTRODUCTION

Large mammalian herbivores have undergone widespread declines in their abundance and distribution across the world's grasslands, steppes, and savannas. Pre-historic humans might have hunted many megafauna to extinction. More recently, humans have progressively replaced them with livestock across the world. Relatively intact assemblages of native herbivores are now increasingly confined to parks and reserves; elsewhere, livestock have not only become the most abundant herbivores, but they also represent the most geographically expansive land-use today (Asner et al. 2004, Bar-On et al. 2018). Further displacement of native herbivores seems inevitable due to steadily rising demands for livestock-products to accommodate evolving human-diets (Thornton 2010, Leclère et al. 2020). This severely restricts and alters the erstwhile influence of native herbivores on ecosystem structure and functions, their broad influence on global biogeochemical cycles, and their influence on soil-C in particular (Brodie et al. 2021, Roy and Bagchi 2021).

Increasingly, it is important to ask whether livestock can maintain the functional roles of the displaced native herbivores (Malhi et al. 2016, Cromsigt et al. 2018, Lundgren et al. 2020). If livestock can maintain ecological functions and services of their native counterparts, then such functional substitutability can have implications for the future of biodiversity, ecosystem functions and services, food-security, and other social and environmental dimensions (Sayer et al. 2013, Veblen et al. 2016, Willemen et al. 2020, Western et al. 2020, Delabre et al. 2021). The extent to which livestock can maintain ecosystem functions and services, particularly soil-C sequestration, becomes important to inform policies over sustainable use of grazing ecosystems and leverage their potential as natural climate solutions (Smith 2014, Cromsigt et al. 2018, Bossio et al. 2020).

Opportunities to restore and maintain ecosystem processes using livestock as surrogates of displaced native herbivores arise from the mutual similarity in their functional

traits (Malhi et al. 2016, Lundgren et al. 2020, Schowanek et al. 2021). Indeed, traits are

better predictors of ecosystem functions than taxonomic identity of species (Hooper et al.

2005). Analyses of key traits, such as body size, diet-choice, and fermentation type,

hypothesize that livestock can be approximate surrogates for native herbivores, i.e., nearest

neighbors in trait-space (Malhi et al. 2016, Lundgren et al. 2020, Schowanek et al. 2021).

However, empirical evidence on ecosystem functions and services is needed before these

ideas can be implemented toward ecosystem management, and such evidence is lacking

(Reinhart et al. 2021).

From this functional-substitutability hypothesis one can ask (Betts et al. 2021) – if

there is sufficient overlap in trait-space between livestock and native herbivores, then to what

extent can they act as surrogates? It is challenging to address this question because many

modern livestock-production systems no longer have native counterparts within comparable

ecological settings that can allow a fair comparison. Unlike intensive livestock production

systems that are often associated with various forms of environmental degradation, traditional

pastoral production systems where both livestock and native herbivores occur in multiple-use

landscapes use (Asner et al. 2004, du Toit et al. 2009, Veblen et al. 2016, Western et al.

2020) can be testbeds to assess whether livestock are good substitutes, or not. While this

functional premise does not expect livestock to be perfect replacements for the displaced

native herbivores (Bagchi and Ritchie 2010, Sitters et al. 2020), it is important to estimate the

degree to which their ecological roles are matched to identify gaps and shortfalls as well as

the scope for improved livestock production systems (Smith 2014, Western et al. 2020,

Reinhart et al. 2021, Delabre et al. 2021).

Here we address impacts of livestock and native herbivores on soil-C sequestration as

grazing by large mammals is an important component of the global carbon cycle (Schmitz et

al. 2014, Roy and Bagchi 2021). We compare their ecological impacts in Spiti region of

northern India, which is part of the greater Trans-Himalayan ecosystem spread across the

Tibetan plateau and surrounding mountains. Soil-C sequestration in the Trans-Himalaya is

particularly important under human land-use for livestock production and under ongoing

climate change (Murthy and Bagchi 2018, Naidu and Bagchi 2021). We investigate long-

term (decadal-scale) net soil-C sequestration under livestock and native herbivores, and their

influence on key microbial decomposer functions related to the C-cycle (Falkowski et al.

2008, Sinsabaugh et al. 2016, Roy and Bagchi 2021). We take advantage of several key

features in the Trans-Himalaya: (1) the livestock are a multi-species assemblage that show

considerable trait-overlap with the native herbivores (Fig. 1), (2) livestock and native

herbivores occupy distinct but juxtaposed watersheds with comparable environmental

settings (Fig. S1-S4), (3) their abundances are comparable so that grazing pressure on

vegetation is also similar (see Methods, Fig. S5). Absent major confounding influences, the

livestock and native assemblages in Trans-Himalaya can help evaluate their functional

substitutability (Malhi et al. 2016, Cromsigt et al. 2018, Lundgren et al. 2020, Schowanek et

al. 2021). Specifically, we investigate (1) whether, and by how much, does grazing by

livestock and native herbivores differ in its influence on long-term (decadal-scale) soil-C

sequestration, and (2) whether any differences can be attributed to how each assemblage

influences microbial decomposer functions in soil.

MATERIALS AND METHODS

STUDY AREA

Traditional pastoral livestock production is the major land-use in Spiti region of the Trans-

Himalayan grazing ecosystem. Watersheds near human settlements are used by a multi-

species livestock assemblage, and these are juxtaposed with watersheds that retain native

herbivores (Fig. 1, Fig. S1). Natural terrain and barriers (canyons, rivers, high ridges) that

restrict frequent animal movement, maintain replicate watersheds under two alternative types

of herbivore assemblages (Fig. S1, S2). Historical local extinctions of native herbivores and

their replacement by livestock over the past decades remains a major conservation challenge

in this region (Mishra et al. 2002, Namgail et al. 2013), and around the world (Veblen et al.

2016, Western et al. 2020). The extant native herbivores are ibex (*Capra sibirica*), bharal

(*Pseudois nayaur*), and yak (*Bos grunniens grunniens*) that are related to wild yak (Bos

grunniens mutus; also known as B. grunniens and B. mutus, respectively; Leslie and Schaller

2009). Currently, kiang (*Equus kiang*) are known from the fringes of the study area, and there

are sporadic reports of argali (*Ovis ammon*). But Tibetan antelope (*Pantholops hodgsonii*)

and Tibetan gazelle (*Procapra picticaudata*) are no longer found in the region. The livestock

consist of goat, sheep, donkey, horse, cattle and yak-cattle hybrids (Fig. 1). The livestock and

native herbivores (after including kiang and argali) show sufficiently high overlap in key

traits (Lundgren et al. 2020), such as body mass, dietary guild, fermentation type, sexual-

dimorphism, graminoid, limb morphology (Fig. 1). In recent decades, total animal biomass

has remained comparable (6.9×10^4 kg for native and 7.2×10^4 kg for livestock; Bagchi and

Ritchie 2010, Singh et al. 2015), such that grazing pressure on vegetation has also remained

similar for sufficiently long periods (Fig. S5).

The two types of watersheds are comparable in their abiotic conditions (Fig. S3–S4).

Climate in Spiti is cold and semi-arid with temperature ranging from 25 °C in summer to

below -30 °C in winters (Fig. S6). Precipitation occurs as winter-snow (100-200 cm yr^{-1}) and

summer-rain (150-300 mm yr^{-1}), and vegetation growth-season is short (May-August, Fig.

S6). Vegetation in this treeless landscape consists of grasses (*Poa*, *Elymus*, *Festuca*), sedges

(*Carex*, *Kobresia*), forbs and shrubs (*Artemisia*, *Lindelofia*, *Caragana*). Both types of

watersheds have an overlapping pool of plant species with spatial variation in vegetation

composition. Livestock watersheds have high abundance of sedges and grasses, have higher

ground cover as well as higher aboveground plant production than grass-and-forb vegetation

in the native watersheds (Fig. S5, Bagchi and Ritchie 2010).

SOIL CARBON

Between 2006 and 2016, we measured soil-C at regular intervals in four watersheds with

native herbivores, and another four watersheds with livestock (Fig. S1, S2). In each

watershed, we set up 3-4 replicated permanent plots (10×10 m^2), for a total of n=15 plots

under native herbivores, and another n=15 plots under livestock. We collected soils with a 5

cm diameter and 20 cm deep cores, and measured carbon content (TruSpec, Leco, USA, and

Elementar Analysensysteme, GmbH, Germany) to compare long-term soil-C stocks (mg C

cm^{-3}) under livestock and native herbivores.

GRAZING-EXCLUSION EXPERIMENT

We distinguish whether any observed difference in soil-C between the watersheds is merely

an artefact of pre-existing site-specific conditions, or it can be attributed to grazing by the

alternative herbivore assemblages. For this, starting in 2005, we set up experimental grazer-

exclusion with a paired-adjacent fence at each location (also 10×10 m^2, each). In 2017, after

up to 12 years of herbivore-exclusion, we estimated grazer-effects on key soil microbial

functions in the paired-adjacent fenced and control plots.

SOIL MICROBIAL FUNCTIONS

We estimated the response of key soil microbial functions to grazing, that are related to the

carbon cycle and net soil-C sequestration: (1) Basal respiration (BR), (2) Potential, or

substrate-induced respiration (SIR), (3) Soil microbial community composition, broadly as

fungal and bacterial fractions, (4) Microbial biomass C and N (MBC and MBN), (5) labile

Carbon (LC) and labile Nitrogen (LN) fractions of soil organic matter, (6) Metabolic quotient

(qCO_2), and (7) Microbial carbon use efficiency (CUE). Since these microbial processes have

seasonal dynamics (Bagchi et al. 2017), we measured them at five time-points during the

growing season, between May and September. We followed standard laboratory protocols to

measure these microbial variables (Robertson et al. 1999, Bagchi et al. 2017, Roy and Bagchi

2021). Briefly, for BR we used an alkali-trap where we incubated 4 g soil at 60% water

holding capacity and measured respired CO_2 (mg C g^{-1} soil day^{-1}). Similarly, we measured

potential microbial respiration (mg C g^{-1} soil day^{-1}) using substrate-induced respiration (SIR)

after adding 0.5% (w/v) of glucose to soil.

We estimated broad microbial community composition as fungal and bacterial

contributions to potential respiration under selective inhibition using Streptomycin (anti-

bacterial) and Cycloheximide (anti-fungal). We used chloroform fumigation-extraction to

estimate MBC (mg C g^{-1} soil), and MBN (mg N g^{-1} soil). We measured SOM from weight

loss on ignition at 450 °C after 4 h. We measured labile fractions (mg g^{-1} soil) of SOM using

two-step hydrolysis with H_2SO_4. We estimated metabolic quotient as the ratio of BR and

MBC (qCO_2, respiration per unit biomass, hr^{-1}). We estimated CUE from established

relationships between key microbial extracellular enzymes (carbon-acquiring Beta-

glucosidase, alongside nitrogen-acquiring Leucine aminopeptidase and *N*-acetyl

glucosaminidase), MBC and MBN, and labile fractions of C and N in SOM (Sinsabaugh et

al. 2013, 2016). We measured activity of the three enzymes using their specific fluorogenic

substrates (4-MUB beta-D-glucopyranoside for Beta-glucosidase, L-Leucine-7-amido-4-

methylcoumarin hydrochloride for Leucine aminopeptidase, and 4-MUB N-acetyl-beta-D-

glucosaminide for N-acetylglucosamindase) (German et al. 2011) in soils from growth season

of 2017. These parameters influence microbial metabolism to determine their investment in

growth (G) and respiration (R), so that $CUE = \frac{G}{G+R}$ (Sinsabaugh et al. 2013, 2016).

Theoretically CUE can range between 0 and 1, i.e., nil to perfect efficiency, with higher

values indicating greater potential for C-sequestration. But, CUE in nature generally does not

exceed 0.6 (Sinsabaugh et al. 2013, 2016).

SOIL MICROBIAL DNA

Based on patterns seen in fungal and bacterial contribution to soil respiration, we evaluated

community composition in more detail using their DNA markers in 2019 (Fierer et al. 2005,

Maestre et al. 2015). We collected soils from ten paired-plots at four time points during the

growing season and extracted microbial DNA using MOBIO Power Soil DNA Extraction Kit

following manufacturer's protocols. Next, we quantified the Internal Transcribed Spacer

(ITS-1 and ITS-2) regions of rRNA for fungal abundance, and region of 16s rRNA for

bacterial abundance with qPCR (Fierer et al. 2005, Maestre et al. 2015). We used these

genetic data to confirm whether DNA-based estimates align with respiration-based estimates

of fungal and bacterial abundance. Additional details of laboratory analyses are in Appendix.

DATA ANALYSIS

We evaluated competing hypotheses (Tredennick et al. 2021, Betts et al. 2021) to determine

whether the observed variation in the data merely represent site-specific and time-specific

heterogeneity between replicates, or it is necessary to invoke herbivore assemblage-type

(livestock or native) as an explanatory variable. We used a linear mixed-effects model to

evaluate long term trends in soil-C and grazer-exclusion effects on each microbial variable.

For soil-C, we used a null-model $\left(M_0^{soilC}\right)$ with time (years, 2006-16) and plot identity as

random-effects to account for background variation due to sampling locations and times. This

model was evaluated against the alternative competing model $\left(M_A^{soilC}\right)$ which included

assemblage-type as a fixed-effect, alongside both random-effects. We compared both models

based on parsimony, goodness-of-fit, and likelihood-ratio test (i.e., AIC, RMSE, and LR).

Next, we determined whether there was gain or loss of soil-C with a separate model where

time (years) was a fixed-effect and plot identity as random effect.

We estimated grazer-effect on each microbial variable as the log-ratio of values in the

fenced and paired-adjacent control plots i.e., $\Delta = ln\left(\frac{\text{Grazing}_i}{\text{Fenced}_i}\right)$ for the ith pair, and calculated

the mean and 95% CI of the log-ratio (Roy and Bagchi 2021). As earlier, we first evaluated a

null-model to assess whether the observed variation in grazer-effects can simply be attributed

to heterogeneity across sampling locations and times (months, May-September). As the null-

model for the jth microbial variable $\left(M_0^j\right)$, we used plot identity and month as random-effects.

In the competing alternative model for each variable $\left(M_A^j\right)$, we included herbivore

assemblage-type as a fixed-effect alongside the random-effects. As before, we used AIC,

RMSE and LR for model comparison to judge whether herbivore assemblage-type is a

necessary explanatory variable for the data (Tredennick et al. 2021). We assessed whether the

data met statistical assumptions from observed and theoretical quantiles of residuals ; the

variables required no further transformation (Δ is already in ln-scale). We used 'nlme' library

in R (Pinheiro et al. 2020) to perform these analyses.

We assessed the inter-relationships between grazer-effects (Δ) on different microbial

variables with structural equations models (SEM). SEM help evaluate hypothesized pathways

over how change in one variable can influence another (Grace 2006). We identified SEM

paths from *a-priori* examples in the literature (Fig. S9). To avoid multicollinearity in SEM

arising from redundancy among variables, we removed those variables which were either

highly correlated with each other ($|r| > 0.49$), or were derived from each other (Fig. S10). So,

ΔSIR, ΔB, ΔMBN, and ΔqCO_2 were not included in this step, and we built SEM using

ΔMBC, ΔLN, ΔLC and ΔF. We did not use genetic data from ITS and 16s rRNA in the SEM

because these were collected from a subset of plots (10 fenced and control pairs in each

watershed), were also not from the same year as the other microbial variables, and because

they broadly matched respiration-based estimates of community composition. As earlier, we

evaluated two competing SEMs – with and without herbivore assemblage-type as an

explanatory variable, i.e., SEM_0 and SEM_A. We incorporated SEM paths as mixed-effects

models for each variable as piece-wise structural equations (Lefcheck 2016). We assessed

goodness of fit with Fisher's C statistic (Shipley 2009, Roy and Bagchi 2021) and report

standardised path coefficients alongside their statistical significance ($\alpha=0.05$). When SEM

paths are supported by data, then they offer a plausible explanation for the underlying

process. When SEM paths are not supported, the hypothesized process may not give rise to

the observed data.

RESULTS

LONG-TERM TREND IN SOIL-CARBON

Model comparison showed that spatio-temporal heterogeneity was not sufficient to explain

long-term variation in soil-C, and it could instead be attributed to herbivore assemblage-type.

There was stronger support for M_A^{soilC} than for M_0^{soilC} (difference in AIC=-25.3, and

diffrence in RMSE=-0.61, and LR=27.3, P<0.001). Soils under native herbivores stored

7.7±1.5 SE more carbon (mg C cm^{-3}) than under livestock ($F_{1,176}=26.6$, P<0.001).

Effectively, areas under livestock could realize up to 71% of the soil-C found under native

herbivores (Fig. 2).

SOIL MICROBIAL RESPONSES TO GRAZING

Analysis of grazer-effects revealed that the observed difference in soil-C (Fig. 2) was related

to how soil microbes respond to grazing by livestock or native herbivores (Fig. 3). Livestock

and native herbivores had comparable effects on B, 16s rRNA abundance, MBC, MBN, LC,

LN, and qCO$_2$ (Fig. 3, Table S1). For these seven variables, the competing alternative models were not supported, and their variability could be attributed to spatio-temporal heterogeneity rather than to assemblage-type (Fig. 3, Table S1). But livestock and native herbivores differed in their influence on BR, SIR, F, ITS abundance, and CUE. For these five variables, there was stronger support for the competing models than the null models (Table S1), as assemblage-type was an important explanatory variable over and above background spatio-temporal heterogeneity (Fig. 3). Grazing by livestock increased both basal and potential microbial respiration (BR and SIR), compared to grazing by native herbivores (Fig. 3). But grazing by livestock reduced both the fungal fraction of microbial respiration and abundance of fungal genetic markers (F and ITS). Respiration-based estimates of fungal and bacterial fractions were consistent with DNA-based estimates (Fig. 3). Grazing by livestock reduced CUE compared to native herbivores (Fig. 3).

INTER-RELATIONSHIPS BETWEEN SOIL MICROBIAL RESPONSES

To assess how differences in grazer-effects on one microbial variable may influence other variables, we evaluated two competing SEM models. The null model was supported by the data (SEM$_0$: Fisher's C = 3.34, P=0.503, df = 4, AIC = 69.34). This showed, regardless of herbivore assemblage-type, grazer-effects on microbial community composition (fungal fraction) affects CUE, and grazers also influence the labile-C pool via their effects on microbial biomass and labile-N (Fig. 4). The alternative competing model was also supported by the data (SEM$_A$: Fisher's C = 2.108, P=0.716, df = 4, AIC = 88.11). It showed that change in assemblage-type can influence CUE via its effect on microbial community composition (Fig. 4). Specifically, change in herbivore-assemblage from native to livestock has negative influence on fungal abundance, which in turn is detrimental for CUE (Fig. 4). Both SEM models indicate that direction and magnitude of grazer-effects are consistent with the premise

that higher CUE under native herbivores improves net soil-C storage, compared to livestock

(Fig. 2, Fig. 4).

DISCUSSION

Previous studies from around the world have emphasized that grazing by large mammalian

herbivores leads to net soil-C storage (Conant and Paustian 2002, Witt et al. 2011, Abdalla et

al. 2018, Derner et al. 2019, Liu et al. 2020), and grazing ecosystems should be integral to

nature-based climate solutions (Smith 2014, Cromsigt et al. 2018, Willemen et al. 2020,

Bossio et al. 2020, Roy and Bagchi 2021). But it has remained unclear whether this service

can be maintained when natural ecosystems are repurposed for livestock production (Reinhart

et al. 2021). And if so, then to what extent?

Our results address several aspects of functional substitutability of native herbivores

by livestock. We find that a multi-species assemblage of livestock can have remarkable

functional similarities with the native assemblages they displace (Fig. 1). And, at comparable

abundances, both assemblages gain soil-C (Fig. 2) at approximately 1‰ to reiterate the role

for grazing ecosystems in nature-based climate solutions (Smith 2014, Cromsigt et al. 2018,

Willemen et al. 2020, Naidu and Bagchi 2021, Roy and Bagchi 2021). But livestock do not

emerge as perfect substitutes since they achieve only about three-fourths of the soil-C stock,

compared to the native herbivores (Fig. 2). Nevertheless, consistent with the functional

substitutability hypothesis, a key ecosystem service provided by native herbivores, soil-C

sequestration, can also be partly realized in livestock-production systems. This can have

implications for how we manage environmental impacts of food (Sayer et al. 2013, Smith

2014, Veblen et al. 2016, Willemen et al. 2020, Western et al. 2020, Delabre et al. 2021), and

meet the rising demand for livestock-products in the coming decades (Thornton 2010,

Leclère et al. 2020).

To determine the scope of improved management, it is important to identify the

potential drivers and mechanisms that explain why livestock lag native herbivores in soil-C

sequestration. Prevailing management practices have largely focused on two aspects—

overgrazing and vegetation community change—which result in altered C-input to soil from

plants (Bagchi and Ritchie 2010, Reinhart et al. 2021). However, this prevailing focus on C-

input alone can be simplistic and inadequate since it does not account for subsequent

metabolism of soil organic matter by microbes (Fontaine et al. 2004, Six et al. 2006,

Falkowski et al. 2008, Sinsabaugh et al. 2013, Roy and Bagchi 2021). Our results from the

grazer-exclusion experiment identify soil microbial responses as the proximate explanation

for why livestock are not perfect substitutes for native herbivores (Fig. 3). Tests of the

competing hypotheses address whether the observed patterns in soil-C reflects idiosyncratic

and/or pre-existing differences between watersheds, or they are linked to the identity of the

herbivores. We attribute this missing soil-C to how microbial processes respond to grazing

when native herbivores are replaced by livestock (Fig. 3-4). However, this still leaves the

possibility that livestock watersheds may have been historically overgrazed, and their soils

have not yet recovered. Even though vegetation may recover from historical overgrazing, it

may take much longer for soil microbial functions to recover (López Zieher et al. 2020). Seen

together, these highlight the importance of soil microbial restoration in tandem with

management of grazing intensity and stocking rates. CUE is a fundamental microbial trait

that controls residence time of carbon in the soil, and it determines the flow of carbon

through ecosystems (Six et al. 2006, Falkowski et al. 2008, Sinsabaugh et al. 2013). The

observed grazer-effects on fungi (Fig. 3) are also known from other ecosystems (Bardgett et

al. 1997, Zhang and Fu 2021). Direction and magnitude of difference in the microbial

community under livestock reduces CUE (Fig. 3-4), and this is consistent with lower soil-C

stocks (Fig. 2) because fungi are known to have higher CUE than bacteria (Fontaine et al.

2004, Six et al. 2006).

Several overlapping factors can alter microbial communities towards lower fungi, and

their individual roles may not be easily disentangled from each other. First, variation in

vegetation composition can result in differences in the quality of litter-input to soil, and this

in turn can influence the rate and efficiency with which microbes metabolize soil organic

matter (Fontaine et al. 2004, Six et al. 2006, Sinsabaugh et al. 2013). Second, as discussed

above, microbial communities may have not yet recovered from any historical overgrazing in

the past (López Zieher et al. 2020), even though vegetation biomass does not indicate

overgrazing is currently a concern (Fig. S5). Third, veterinary care offered to the livestock

leads to inadvertent release of antibiotics and other drugs to the soil from their dung and

urine, and this can alter microbial communities towards lower CUE (Wepking et al. 2019).

Such overlapping factors need to be addressed together to improve soil-C sequestration,

rather than in isolation.

Our results from alternative competing SEMs offer insights into how net soil-C

sequestration can be improved (Fig. 4). Since replacing native herbivores with livestock is

detrimental for CUE, promoting wildlife conservation over livestock-production would favor

soil-C (SEM$_A$, Fig. 4). This would compromise livelihoods and food-security, and it circles

back to the inconsistencies and conflicts between the goals of conservation and human land-

use. This conventional and exclusionary approach may no longer be tenable and is also

increasingly difficult to justify (Adams and Mulligan 2002); it is timely to assess alternative

ideas (Smith 2014). The null model (SEM$_0$, Fig. 4) can offer insights into ways to resolve

these conflicting demands on land-use. It suggests altered microbial communities

compromise CUE regardless of whether the community shift is a fallout of land-use, or not.

This opens opportunities for restoration of microbial communities to sustain ecosystem

services under human land-use (Paluch et al. 2013, Wubs et al. 2016, Mills et al. 2017).

Admittedly, there are many gaps in our current understanding of structure-function

relationships for hyper-diverse microbial communities (Bier et al. 2015). Yet, there are

encouraging signs that effective restoration can be achieved even with incomplete taxonomic

knowledge (Paluch et al. 2013, Wubs et al. 2016, Mills et al. 2017). Since livestock already

deliver 71% of the ecosystem service provided by native herbivores, further incorporating

microbial rewilding of soils into livestock-production may offset the 29% lag against native

herbivores—in the process, making livestock-production more environmentally responsible

(Smith 2014, Cromsigt et al. 2018, Willemen et al. 2020, Bossio et al. 2020).

In conclusion, our results highlight: (1) multi-species and functionally diverse

assemblages of livestock can partly approximate the ecological roles of native herbivores,

and (2) restoration and rewilding of soil microbial communities may enable livestock to

emulate the services provided by native herbivores. Alongside continued management of

stocking rates, functional substitutability and microbial restoration in-tandem may help

reconcile rising demands for livestock-products with conservation of ecosystem functions

and services.

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Figures

Fig. 1

Figure 1. Trait space of livestock and native herbivores in Spiti, India. We find there is

considerable overlap in multidimensional trait space between native herbivores (green) and

livestock (blue). Native herbivores in the study area are ibex, bharal, yak; asterix (kiang,

argali) indicates that these animals are found in nearby regions. The livestock consist of goat,

sheep, donkey, horse, cattle and yak-cattle hybrids. Six traits were used to construct the trait-

space: body mass, dietary guild, fermentation type, sexual-dimorphism, graminoid, and limb

morphology (Lundgren et al. 2020) with Principal Coordinates Analysis on the Gower's

distance matrix.

Fig. 2

Figure 2. Long-term change in soil-C (mean ± 95CI) under grazing by native herbivores and

livestock in Spiti, India. Watersheds with livestock had overall lower soil-C than those with

native herbivores, but both gained soil-C during the decade at approximately 1‰.

Fig. 3

Figure 3. Grazer-effect ($\Delta = ln\left(\frac{grazed}{fenced}\right)$), as mean ± 95CI) of livestock and native herbivores

on twelve soil microbial variables in Spiti, India. These variables are basal microbial

respiration (BR), potential microbial respiration (SIR), Fungal fraction (F), ITS abundance

(ITS), Bacterial fraction (B), 16s gene abundance (16s), Microbial Biomass Carbon (MBC),

Microbial Biomass Nitrogen (MBN), Labile-C (LC), Labile-N (LN), Metabolic quotient

(qCO_2), and microbial carbon use efficiency (CUE). Direction and magnitude of differences

in BR, SIR, F, ITS abundance, and CUE varied with herbivore-assemblage type. But the

remaining variables were not influenced by herbivore-assemblage type.

Fig. 4

SEM$_0$: Fisher's C = 3.34, P=0.503, df = 4, AIC = 69.34   SEM$_A$: Fisher's C = 2.108, P=0.716, df = 4, AIC = 88.11

Figure 4. Mixed-effects structural equation models to evaluate two competing hypotheses –

with and without herbivore assemblage-type as an explanatory variable, i.e., SEM$_A$ and

SEM$_0$. For each variable, grazer-effect ($\Delta = ln\left(\frac{grazed}{fenced}\right)$) was calculated from grazed and

ungrazed paired plots. The variables are ΔBR: basal respiration, ΔMBC: microbial biomass-

C, ΔLC: labile-C, ΔLN: labile-N, ΔF: fungal biomass, ΔCUE: carbon use efficiency. In

SEM$_A$, as herbivore-assemblage goes from native to livestock, a decrease in ΔF leads to a

decrease in ΔCUE. Alternatively, in SEM$_0$, increase in ΔF increases ΔCUE, independent of

herbivore-assemblage type. Green and red arrows represent significant positive and negative

standardized path coefficients respectively, while grey arrows represent non-significant paths

(α = 0.05). Marginal and conditional R^2 are reported, and asterix represent statistical

significance (*** P$\leq$0.001, ** P$\leq$0.01, and * P$\leq$0.05).