Retrotransposon instability dominates the acquired mutation landscape of mouse induced pluripotent stem cells

Patricia Gerdes[1,10], Sue Mei Lim[2,3,4,10], Adam D. Ewing[1,10], Michael R. Larcombe[2,3,4], Dorothy Chan[1], Francisco J. Sanchez-Luque[1,5], Lucinda Walker[1], Alexander L. Carleton[1], Cini James[1], Anja S. Knaupp[2,3,4], Patricia E. Carreira[1], Christian M. Nefzger[2,3,4], Ryan Lister[6,7,], Sandra R. Richardson[1,*], Jose M. Polo[2,3,4,9,*], and Geoffrey J. Faulkner[1,8,*]

[1]Mater Research Institute - University of Queensland, TRI Building, Woolloongabba QLD 4102, Australia.

[2]Department of Anatomy & Developmental Biology, Monash University, Melbourne VIC 3800, Australia.

[3]Development and Stem Cells Program, Monash Biomedicine Discovery Institute, Melbourne VIC 3800, Australia.

[4]Australian Regenerative Medicine Institute, Monash University, Melbourne VIC 3800, Australia.

[5]GENYO. Pfizer-University of Granada-Andalusian Government Centre for Genomics and Oncological Research, PTS Granada 18016, Spain.

[6]Australian Research Council Centre of Excellence in Plant Energy Biology, School of Molecular Sciences, The University of Western Australia, Perth WA 6009, Australia.

[7]Harry Perkins Institute of Medical Research, Perth WA 6009, Australia.

[8]Queensland Brain Institute, University of Queensland, Brisbane QLD 4072, Australia.

[9]Adelaide Centre for Epigenetics and The South Australian Immunogenomics Cancer Institute, The University of Adelaide, Adelaide SA 5005, Australia.

[10]These authors contributed equally.

[*]Corresponding authors: sandra.richardson@mater.uq.edu.au (S.R.R.); jose.polo@monash.edu (J.M.P.); faulknergj@gmail.com (G.J.F.)

Abstract

Induced pluripotent stem cells (iPSCs) may differentiate into any cell of the body and as such have revolutionized biomedical research and regenerative medicine. Unlike their human counterparts, mouse iPSCs (miPSCs) are reported to silence transposable elements (TEs) and prevent TE-mediated mutagenesis. Here we applied short- or long-read genome sequencing to 38 bulk miPSC lines reprogrammed from 10 parental cell types, as well as 18 single-cell miPSC clones. While single nucleotide variants and structural variants restricted to miPSCs were rare, we found 83 *de novo* TE insertions, including examples intronic to *Brca1* and *Dmd*. LINE-1 (L1) retrotransposon families were profoundly hypomethylated in miPSCs, beyond other TEs and the genome overall, and harbored alternative promoters for protein-coding genes. Treatment with the L1 reverse transcriptase inhibitor lamivudine did not hinder reprogramming, pointing to a viable strategy to block retrotransposition. These experiments reveal the complete spectrum and potential significance of mutations acquired by miPSCs.

Introduction

Induced pluripotent stem cells (iPSCs) resemble embryonic stem cells (ESCs) in their near unlimited capacity for self-renewal and differentiation potential[1]. These properties have driven widespread uptake of iPSCs in clinical and research applications[2–4]. Despite their immense therapeutic promise, the reprogramming process required to generate iPSCs can produce genomic and epigenomic aberrations[4–8]. These abnormalities could undermine the functional equivalence of iPSCs and ESCs, or alter the phenotype of iPSC-derived differentiated cells, and hence necessitate genetic and functional screening of iPSCs prior to their use in the clinic[9]. Fortunately, whole genome sequencing (WGS) analyses of single nucleotide variants (SNVs), copy number variants, and structural variants (SVs) restricted to human and mouse iPSC lines have found relatively few conclusive reprogramming-associated mutations[10–12]. Instead, most mutations acquired by iPSCs appear to occur before and after reprogramming[10,11,13], implying they are not caused by molecular processes intrinsic to iPSC generation. Transposable elements (TEs) may present an important exception to this rule, where the attainment of a pluripotent state via reprogramming leaves iPSCs vulnerable to TE-mediated mutagenesis.

The retrotransposon long interspersed element 1 (LINE-1, or L1) is active in nearly all mammals[14]. L1 autonomously mobilizes via a copy-and-paste process called target-primed reverse

transcription (TPRT), which involves reverse transcription of L1 mRNA *in cis*, and is characterized by the generation of target site duplications (TSDs) upon L1 integration[15–20]. The C57BL/6 mouse reference genome contains ~3,000 potentially mobile L1 copies belonging to three subfamilies (T_F, G_F and A) defined by their monomeric 5′ promoter sequences, in addition to several active endogenous retrovirus (ERV) and short interspersed element (SINE) families[21–23]. By contrast, only ~100 mobile L1s from the transcribed subset Ta (-Ta)[24] subfamily are present in each individual human genome, with the vast majority of retrotransposition potential concentrated in fewer than 10 of these elements[25,26]. Perhaps owing to the disparate count of mobile TEs in each species, the rate of L1 mobilization in the mouse germline is estimated to be at least an order of magnitude higher than that of humans[27–30].

TE mobility is regulated by DNA methylation and histone modifications, as well as various post-transcriptional and post-translational mechanisms[31–41]. Reprogramming somatic cells to generate human iPSCs (hiPSCs) and mouse iPSCs (miPSCs) leads to epigenome-wide remodeling, including broad de-repression of L1 promoters[7,42–47]. L1 mRNA abundance increases strongly during reprogramming, and remains approximately 10-fold higher in cultured miPSCs than in parental mouse embryonic fibroblasts (MEFs)[46]. As a corollary, the early mouse embryo is a major niche for new heritable L1 retrotransposition events[28]. Mouse ESCs cultured in standard media containing serum permit genome-wide L1 hypomethylation, express endogenous L1 proteins and support engineered L1 mobilization[41,48]. Naïve ESCs grown in media containing two small-molecule kinase inhibitors (2i) in place of serum also exhibit L1 promoter hypomethylation[37,48,49]. Engineered and endogenous L1 retrotransposition are supported by hiPSCs and ESCs[45,50–52]. These observations collectively suggest L1 hypomethylation may be an inherent aspect of pluripotency accentuated by the molecular roadmap to an induced pluripotent state. Consequently, miPSCs are likely to harbor *de novo* retrotransposition events. However, a prior WGS analysis of 3 miPSC lines, employing paired-end 42mer reads and ~11× genome-wide sequencing depth, found no *de novo* TE insertions, and concluded that endogenous retrotransposition did not occur during miPSC production[12]. The apparent lack of TE mobility in this context remains an unresolved and yet potentially important source of miPSC mutagenesis[4]. Here, we analyze a diverse panel of miPSC genomes with short- or long-read sequencing and, as reported for hiPSCs and other pluripotent human cells[45,50–52], we detect numerous *de novo* TE insertions acquired by miPSCs.

Results

Mutational spectra of bulk miPSC populations generated from diverse cell lineages

To survey genomic variation among miPSC lines generated from a broad range of parental cell types, we bred triple transgenic C57BL/6×129S4Sv/Jae animals carrying a GFP reporter knocked into the *Oct4* locus (*Oct4*-GFP), a transcriptional activator (m2rtTA) under the control of the ubiquitously expressed Rosa26 locus (R26-m2rtTA), and a doxycycline-inducible polycistronic reprogramming cassette (Col1a1-tetO-OKSM)[53]. From each of three animals (labeled A67, A82 and A172), we used fluorescence activated cell sorting (FACS) and a range of surface markers to isolate nine isogenic primary cell populations, including three representing each germ layer (**Fig. 1a**). Bulk cultures were then treated with doxycycline to induce reprogramming, followed by FACS to purify *Oct4*-GFP^{+} miPSCs. Twenty-six miPSC lines were successfully expanded and cultured in standard media containing serum (**Supplementary Table 1** and **Extended Data Fig. 1**). Illumina paired-end 150mer read WGS (~41× average genome-wide depth) was then applied to each miPSC line at passage 4 (p4), as well as to 3 MEF genotypic controls (**Supplementary Table 1**).

Concordant SNVs detected by GATK HaplotypeCaller and freebayes[54,55] were filtered to remove known mouse strain germline variants[56], yielding 3,603 SNVs private to a single miPSC line (average ~140 per line) and absent from the corresponding MEF samples (**Supplementary Table 2**). Of these, 27 in total were non-synonymous exonic mutations. We then called concordant SVs using Delly and GRIDSS[57,58], finding 34 private SVs (~1 per line). These included a 210kbp deletion of the *de novo* methyltransferase *Dnmt3a* in miPSCs derived from the hematopoietic stem cells of animal A172 (**Supplementary Table 2**). Considering private SNVs and SVs together, we observed no significant ($p<0.05$, one-way ANOVA with Tukey's multiple comparison test) difference in miPSC variant counts associated with parental cell type or germ layer, and SNV and SV rates resembled those found previously for fibroblast-derived miPSCs[10,12]. Choice of primary cell type, at least among the diverse panel assembled here, may therefore not significantly impact the frequency of SNVs and SVs later found in miPSC lines.

Bulk miPSC populations harbor *de novo* L1 insertions

As *de novo* TE insertions can be overlooked by generalized SV calling algorithms[59], we used TEBreak[60] to identify non-reference TE insertions. Known non-reference genome TE insertions[56],

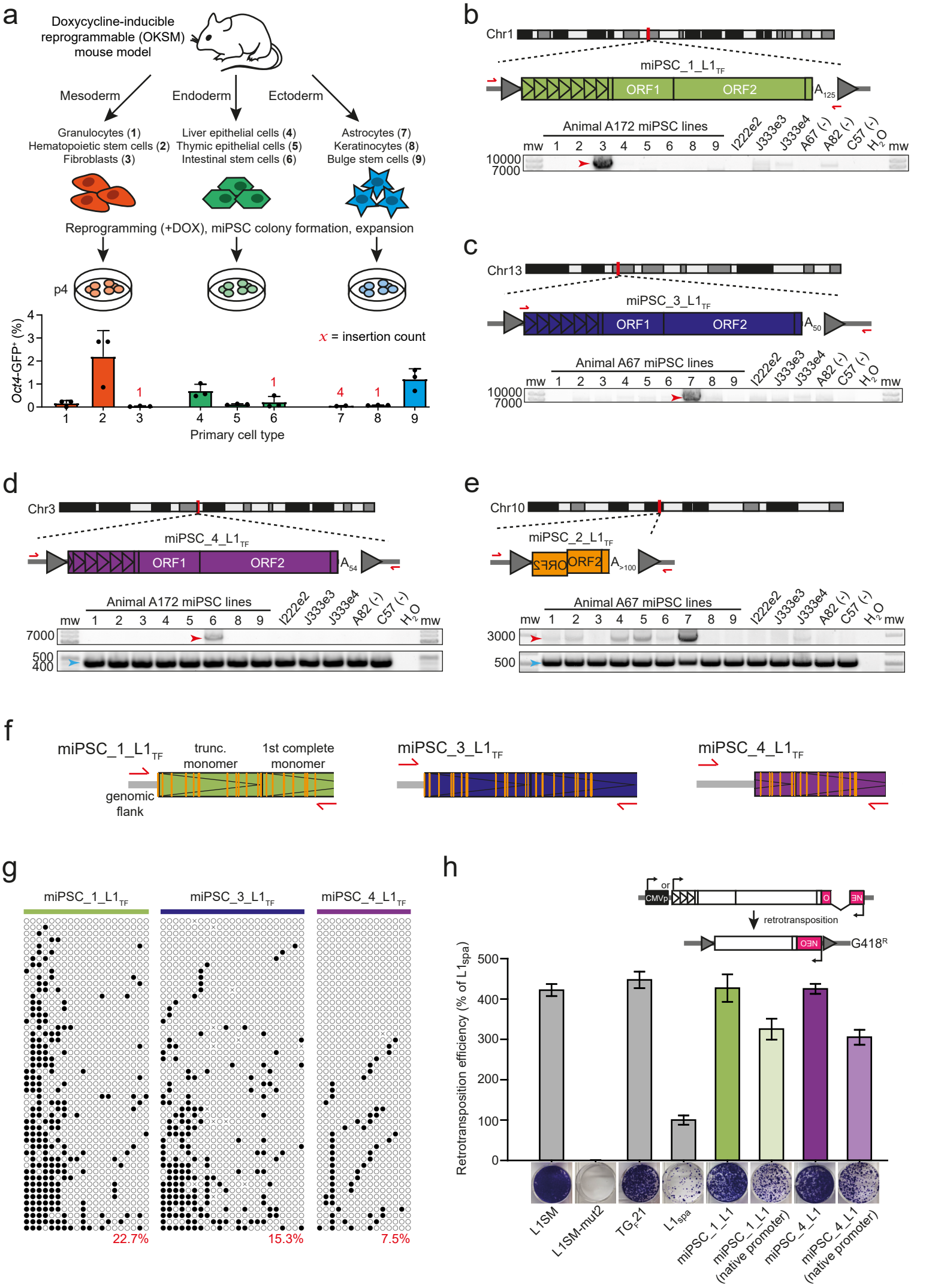

a
Doxycycline-inducible reprogrammable (OKSM) mouse model
Mesoderm
Endoderm
Ectoderm
Granulocytes (1)
Hematopoietic stem cells (2)
Fibroblasts (3)
Liver epithelial cells (4)
Thymic epithelial cells (5)
Intestinal stem cells (6)
Astrocytes (7)
Keratinocytes (8)
Bulge stem cells (9)
Reprogramming (+DOX), miPSC colony formation, expansion
p4
Oct4-GFP+ (%)
x = insertion count
Primary cell type
b
Chr1
miPSC_1_L1TF
ORF1
ORF2
A125
Animal A172 miPSC lines
mw 1 2 3 4 5 6 8 9 I222e2 J333e3 J333e4 A67 (-) A82 (-) C57 (-) H2O mw
10000
7000
c
Chr13
miPSC_3_L1TF
ORF1
ORF2
A50
Animal A67 miPSC lines
mw 1 2 3 4 5 6 7 8 9 I222e2 J333e3 J333e4 A82 (-) C57 (-) H2O mw
10000
7000
d
Chr3
miPSC_4_L1TF
ORF1
ORF2
A54
Animal A172 miPSC lines
mw 1 2 3 4 5 6 8 9 I222e2 J333e3 J333e4 A82 (-) C57 (-) H2O mw
7000
500
400
e
Chr10
miPSC_2_L1TF
ORF2
A>100
Animal A67 miPSC lines
mw 1 2 3 4 5 6 7 8 9 I222e2 J333e3 J333e4 A82 (-) C57 (-) H2O mw
3000
500
f
miPSC_1_L1TF
trunc. monomer
1st complete monomer
genomic flank
miPSC_3_L1TF
miPSC_4_L1TF
g
miPSC_1_L1TF
miPSC_3_L1TF
miPSC_4_L1TF
22.7%
15.3%
7.5%
h
or
CMVp
O
NE
retrotransposition
NEO
G418R
Retrotransposition efficiency (% of L1spa)
L1SM
L1SM-mut2
TGF21
L1spa
miPSC_1_L1
miPSC_1_L1 (native promoter)
miPSC_4_L1
miPSC_4_L1 (native promoter)

Fig. 1: *De novo* **L1 insertions in germ layer specific bulk expanded miPSC lines. a,** Experimental design of bulk miPSC generation using a Col1a1-tetO-OKSM mouse model containing a doxycycline-inducible reprogramming cassette. Tissues were isolated and sorted by FACS to obtain 9 primary cell types (named and numbered 1-9) from each of 3 mice (A67, A82, A172). After reprogramming with doxycycline (+DOX), *Oct4*-GFP positive miPSCs were sorted and expanded in cell culture. DNA was extracted from miPSCs, sequenced via WGS and mRC-seq, and analyzed for *de novo* TE insertions with TEBreak. The histogram displayed underneath indicates the mean reprogramming efficiency (*Oct4*-GFP+) ± SD for each primary cell type, with individual replicate values marked by black dots. Red numerals indicate the count of reprogramming-associated L1 insertions found in the miPSCs obtained from each primary cell type. Note: astrocyte-derived miPSCs were not produced for animal A172. **b,** A full-length (6.8 monomers) intergenic *de novo* L1 T_F insertion. Promoter monomers are shown as triangles within the L1 5′UTR. PolyA (A_n) tract length is indicated immediately 3′ of the L1. Target site duplications (TSDs) are depicted as gray arrows flanking the L1. PCR validation primers are shown as red arrows. A PCR validation agarose gel containing the full-length PCR product (red arrow) only in the fibroblast-derived miPSC line where the L1 was detected by genomic analysis is shown. miPSC line numbers are provided in panel (a). Note: given the variable, and in some cases very low, reprogramming efficiencies shown in panel (a), and the objective to obtain a full set of 9 miPSC lines from each animal, we entirely used each sorted primary cell population for reprogramming, relying on PCR amplification in a single miPSC line to validate reprogramming-associated retrotransposition events. DNA from other animals included in the study are however shown at right as additional controls. **c,** As for panel (b), except for an L1 T_F with 5.8 promoter monomers. Note: the very faint, smaller-sized gel band observed in most of the samples was an off-target product. **d,** As for panel (b), except for an L1 T_F with 5.3 promoter monomers, and using an empty/filled PCR design where both primers are outside of the L1 insertion, generating "filled" L1 (red arrow) and "empty" wild-type (blue arrow) products. **e,** As for panel (b), except showing a 5′ truncated and inverted/deleted L1 T_F insertion and using an empty/filled PCR validation design, as per panel (d). **f,** Locus-specific methylation analysis schematic representation for 3 full-length *de novo* L1 insertions (panels b-d). After bisulfite conversion, the 5′ monomeric sequences of each L1 were PCR amplified using primer pairs (red arrows) specific to that locus. Amplicons were then pooled and sequenced as 2×300mer Illumina reads. Orange strokes indicate CpG dinucleotides covered by the assay. **g,** Methylation of the 3 L1 promoter sequences shown in panel (f), in the miPSC line where each *de novo* L1 insertion was identified. Each cartoon panel corresponds to an amplicon and displays 50 non-identical sequences (black circle, methylated CpG; white circle, unmethylated CpG; ×, mutated CpG) extracted at random from a much larger pool of available Illumina reads. The percentage of methylated CpG is indicated in the lower right corner of each cartoon in red. **h,** *top*: Rationale of a cultured cell retrotransposition assay[19,67]. A mouse L1 driven by its native promoter alone, or with the addition of a CMV promoter (CMVp), is tagged with an antisense orientated neomycin (NEO) reporter cassette interrupted by an intron. Cells harboring this construct become neomycin (G418) resistant upon retrotransposition. *bottom*: Retrotransposition assays conducted in HeLa cells. Constructs included: L1SM[68], a highly mobile synthetic L1 (positive control); L1SMmut2, L1SM with endonuclease and reverse transcriptase active site mutations (negative control); TG_F21, a mobile L1 G_F element[21]; $L1_{spa}$, a mobile L1 T_F element[22]; miPSC_1_L1 (panel b); miPSC_4_L1 (panel d). Data were normalized to $L1_{spa}$ and are shown as mean ± SD of three independent biological replicates, each of which comprised three technical replicates. Representative well pictures are shown below each construct. Note: L1SM retrotransposed very efficiently, leading to cell colony crowding in wells, and a likely underestimate of retrotransposition. Unless otherwise stated, L1 constructs incorporated a CMVp element.

and those found in MEF genotypic controls or multiple miPSC lines, were filtered, leaving 4 putative *de novo* L1 T_F insertions (**Fig. 1b-d**, **Table 1**, **Extended Data Fig. 2**, **Supplementary Table 3**). To achieve even greater coverage of potentially active TEs, we performed mouse retrotransposon capture sequencing (mRC-seq), which uses sequence capture probes to enrich Illumina libraries for the 5′ and 3′ genomic junctions of mobile TEs, including T_F, G_F and A subfamily L1s, B1 and B2 SINEs, and IAP and ETn ERVs (**Supplementary Table 1**)[28,61]. The combination of WGS and mRC-seq identified an additional 4 putative *de novo* L1 G_F and T_F insertions (**Extended Data Fig. 3**, **Table 1** and **Supplementary Table 3**).

We PCR amplified and fully characterized each putative L1 insertion sequence. Six events were full-length, retaining 2-7 monomers at their 5′ end, and could only be PCR amplified in the miPSC line where they were detected by genomic analysis (**Fig. 1b-d**, **Extended Data Fig. 2** and **Extended Data Fig. 3**). An additional L1 (labeled miPSC_6_L1) was very heavily 5′ truncated and confirmed by PCR to be private to one miPSC line (**Extended Data Fig. 3**). The final example (miPSC_2_L1) was heavily 5′ truncated and inverted[62] and could be PCR amplified in 7/9 miPSC lines representing all 3 germ layers of animal A67 (**Fig. 1e** and **Extended Data Fig. 2**). miPSC_2_L1 most likely represented a mosaic insertion that arose early in the embryonic development of animal A67, as found previously[28,30,63,64]. Each insertion carried TSDs of 13-19nt, a long and pure 3′ polyA tract, and integrated at a degenerate L1 endonuclease recognition motif (5′-TTTT/AA-3′) (**Table 1**). These hallmarks were consistent with *bona fide* TPRT-mediated L1 retrotransposition events[15,16,19,65,66]. In sum, 10/26 miPSC lines harbored at least one PCR validated *de novo* L1 insertion. Not counting the mosaic miPSC_2_L1 insertion, miPSCs from all 3 animals and 4/9 primary cell types, representing each germ layer, presented at least one *de novo* L1 insertion (**Fig. 1a** and **Supplementary Table 3**). Of these insertions, 4/7 were detected in astrocyte-derived miPSCs and 6/7 in miPSCs obtained from primary cells in the bottom 50% of reprogramming efficiencies (**Fig. 1a**), though neither of these proportions were statistically significant (binomial test). Notably, down-sampling to 11× depth WGS, as per[12], indicated an expected 95% probability of finding none of the validated *de novo* insertions (**Extended Data Fig. 4a**).

Comprehensive capillary sequencing of the 3 full-length insertions (miPSC_1_L1, miPSC_3_L1 and miPSC_4_L1) revealed that each had intact ORFs (**Fig. 1b-d**). To assess the potential for further mobilization of these newly retrotransposed elements, we first used

multiplexed L1 locus-specific bisulfite sequencing[34,61] to measure CpG methylation of their most 5′ promoter monomers (**Fig. 1f**). All 3 full-length insertions were fully unmethylated in a subset of miPSCs, and their methylation decreased with distance from the L1 5′ end (**Fig. 1g**). Next, we cloned and tested miPSC_1_L1 and miPSC_4_L1 in a cultured cell retrotransposition assay[19,67], using the natural elements $L1_{spa}$ (T_F subfamily)[22] and TG_F21 (G_F subfamily)[21] as positive controls, as well as the highly mobile synthetic L1 T_F element L1SM[68]. As evidence of their *in vitro* mobility, miPSC_1_L1 and miPSC_4_L1 retrotransposed efficiently in HeLa cells, when expressed from their native promoter alone or with the addition of a cytomegalovirus promoter (**Fig. 1h**). Thus, endogenous L1 retrotransposition in miPSCs is driven by highly mobile donor L1s that can produce incompletely methylated, retrotransposition-competent offspring L1s.

Single-cell miPSC clones reveal extensive L1-mediated endogenous retrotransposition

Despite *de novo* L1 insertions being present in 10/26 miPSC lines, we were concerned that additional mutations could be obscured by the heterogeneous mixture of cellular clones contained in bulk reprogrammed miPSCs. We therefore reprogrammed MEFs from one of our C57BL/6×129S4Sv/Jae animals (labeled I222e2), isolated individual miPSCs via FACS, and expanded 18 clones cultivated in serum until p3 (**Fig. 2a**). To assess the impact of standard and naïve culture conditions, respectively, upon L1 activity, each clone was divided and then further expanded in serum or 2i until p6 (**Fig. 2a**). We then applied ~41× average genome-wide depth Illumina WGS and mRC-seq to miPSC single-cell clones 1-9, and mRC-seq only to clones 10-18, with each clone analyzed after culture in serum or 2i media (**Fig. 2a**, **Extended Data Fig. 1** and **Supplementary Table 1**). Deep WGS was performed on the parental I222e2 MEF population, attaining cumulative 117× genome-wide depth, in addition to mRC-seq (**Supplementary Table 1**). Using the WGS data, we again called concordant SNVs and SVs private to one miPSC clone, while excluding known germline variants and those found in the parental MEFs. We found, on average, ~100 and ~1 private SNVs and SVs per miPSC clone, respectively, almost all of which were detected in both the serum and 2i conditions for each clone (**Supplementary Table 2**). These frequencies resembled those found by genomic analysis of bulk miPSCs, underlining that heterogeneous and homogeneous fibroblast-derived miPSC populations are relatively free of genomic abnormalities[10,12]. This experiment also indicated choice of serum or 2i media did not impact the frequency of SNVs or SVs present in miPSCs.

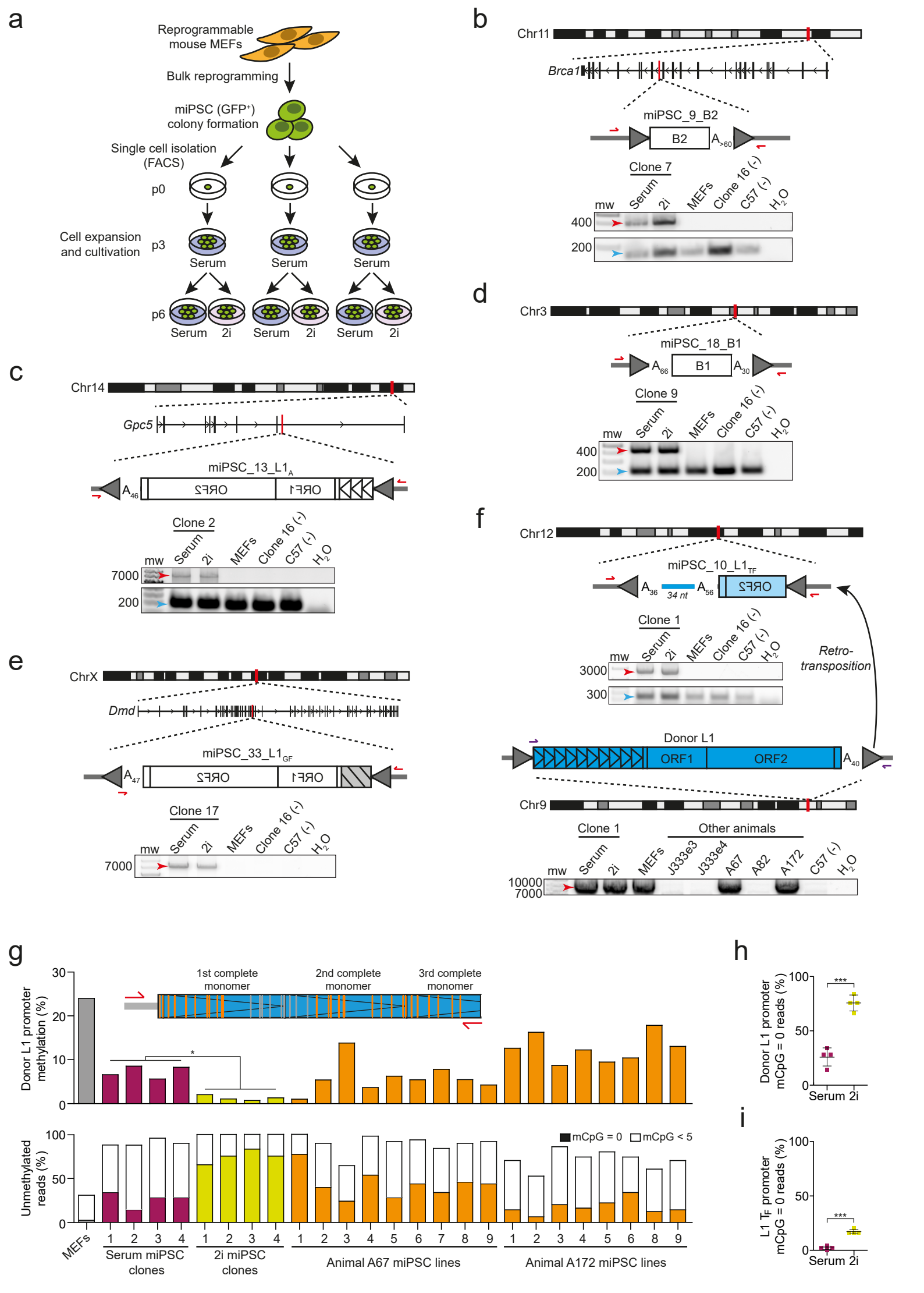

a
Reprogrammable mouse MEFs
Bulk reprogramming
miPSC (GFP+) colony formation
Single cell isolation (FACS)
p0
Cell expansion and cultivation
p3
Serum
p6
Serum
2i
b
Chr11
Brca1
miPSC_9_B2
B2
Clone 7
mw
Serum
2i
MEFs
Clone 16 (-)
C57 (-)
H2O
400
200
c
Chr14
Gpc5
miPSC_13_L1A
ORF2
ORF1
Clone 2
7000
200
d
Chr3
miPSC_18_B1
B1
Clone 9
e
ChrX
Dmd
miPSC_33_L1GF
Clone 17
f
Chr12
miPSC_10_L1TF
34 nt
Clone 1
3000
300
Retro-transposition
Donor L1
ORF1
ORF2
Chr9
Other animals
J333e3
J333e4
A67
A82
A172
10000
7000
g
1st complete monomer
2nd complete monomer
3rd complete monomer
Donor L1 promoter methylation (%)
Unmethylated reads (%)
mCpG = 0
mCpG < 5
MEFs
Serum miPSC clones
2i miPSC clones
Animal A67 miPSC lines
Animal A172 miPSC lines
h
Donor L1 promoter mCpG = 0 reads (%)
Serum
2i
i
L1 TF promoter mCpG = 0 reads (%)

Fig. 2: Frequent *de novo* TE insertions in MEF-derived clonally expanded miPSC lines. a, Experimental design to generate single-cell miPSC clonal lines. Bulk MEFs from a Col1a1-tetO-OKSM mouse (animal I222e2) were purified and reprogrammed by addition of doxycycline. Individual *Oct4*-GFP positive miPSCs were then isolated via FACS, expanded in serum for 3 passages, and then cultured in serum- or 2i-containing miPSC media for 3 additional passages. DNA was then extracted and analyzed by WGS and mRC-seq for 9 single-cell clones (for both serum and 2i conditions), with 9 further clones analyzed only with mRC-seq. **b,** A full-length *de novo* B2 inserted and orientated in antisense to intron 15 of *Brca1*. PolyA tract length is indicated immediately 3′ of the B2. TSDs are depicted as gray arrows flanking the B2. PCR validation (gel pictures shown) involved an empty/filled PCR design where both primers (red arrows) are outside of the B2, generating "filled" B2 (red arrow) and "empty" wild-type (blue arrow) products. The B2 was amplified only in either the serum or 2i conditions for the single-cell clone (number 7) where the B2 was detected by genomic analysis, and not in the matched parental MEFs, the C57BL/6 strain, or a single-cell clone (number 16) selected at random. **c,** A full-length (3 monomers) L1 A subfamily element inserted *de novo* antisense to intron 7 of *Gpc5*. Sequence characteristics and PCR validation results are shown as in panel (b). Promoter monomers are shown as triangles within the L1 5′UTR. **d,** As in panel (b), except showing an unusual intergenic B1 insertion flanked by both 5′ and 3′ polyA tracts. **e,** A full-length L1 G_F inserted *de novo* antisense to intron 60 of *Dmd*. PCR validation involved a 5′ genomic primer and a 3′ junction primer (red arrows). As indicated by a gray box with black stripes, the number of monomers is unknown but was >3. **f,** A heavily 5′ truncated, intergenic *de novo* L1 T_F insertion validated by empty/filled PCR, as per panel (b). Sequence features are annotated as per panel (b), with the addition of a 34nt 3′ transduction matching a donor L1 T_F located on Chromosome 9. PCR using primers (purple arrows) designed to amplify the entire donor L1 indicated it was polymorphic in our colony. Capillary sequencing indicated the donor L1 retained a promoter of 10 monomers and had intact ORFs. **g,** Locus-specific bisulfite sequencing analysis of the donor L1 promoter identified in panel (f), in MEFs, single-cell miPSC clones, and miPSC lines derived from primary cells. *top*: Assay design and primer locations. CpGs located in the first 3 monomers of the donor L1 were assessed. Orange and gray strokes indicate CpGs covered and not covered, respectively, by sequencing the amplicon with 2×300mer Illumina reads. *middle*: Mean percentages of donor L1 CpG methylation for 50 non-identical sequences selected at random from each sample. A two-tailed t test (*$p<0.05$) was used to compare serum and 2i culture conditions for single-cell miPSC clones 1-4. *bottom*: Percentages of fully unmethylated (mCpG=0, filled bars) and heavily unmethylated (0<mCpG<5, white bars) reads using the same sequencing data as displayed in the above histogram. **h,** Percentages of fully unmethylated (mCpG=0) reads corresponding to the donor L1 promoter identified in panel (f), for miPSCs cultured in serum or 2i conditions. Data represent mean methylation ± SD observed for single-cell miPSC clones 1-4. Significance testing was via two-tailed t test (***$p<0.0001$). **i,** As for panel (h), except using an assay targeting the L1 T_F subfamily monomer.

By contrast, TEBreak revealed 35 putative *de novo* TE insertions absent from the parental MEFs, all of which were found in both serum and 2i culture conditions for at least one miPSC clone. Of these, 27 were detected by both WGS and mRC-seq, 6 by mRC-seq only, and 2 by WGS only (**Supplementary Table 3**). Note that the 6 events found only by mRC-seq were detected in the 9 miPSC clones (numbered 10-18) not analyzed with WGS. The 2 insertions found by WGS alone were both moderately 5′ truncated and carried a 3′ transduction, two features that reduced their probability of detection by mRC-seq, where enrichment probes target the 5′ and 3′ ends of L1 consensus sequences[28,61]. We were able to PCR amplify 32 insertions in full and capillary sequence at least their 5′ and 3′ junctions (**Fig. 2b-f**, **Extended Data Fig. 3** and **Supplementary Table 3**). Two other putative TE insertions could only be amplified at their 5′ genome junction; one of these (miPSC_29_L1) however also had strong 3′ WGS and mRC-seq support. We therefore considered 33 TE insertions as validated *de novo* events (**Table 1**). Thirty-one of these were PCR validated as private to only one miPSC clone, whereas the remaining two events were found in either 2 clones (miPSC_23_B2) or 4 clones (miPSC_43_L1) (**Extended Data Fig. 3**). These last two insertions were therefore present in subclones of the parental MEF population.

The 33 *de novo* insertions included 20, 3 and 1 T_F, G_F and A L1 subfamily members, respectively, as well as 2 B1 and 7 B2 elements (**Supplementary Table 3**). All insertions generated TSDs and a 3′ polyA tract, and integrated at a degenerate L1 endonuclease motif (**Table 1**). 14/24 L1 insertions retained at least one promoter monomer and were therefore considered full-length (**Table 1**). Of the remaining 10 L1s, 3 were 5′ inverted. One unusual B1 insertion, miPSC_18_B1, was flanked by 5′ and 3′ polyA tracts as well as TSDs (**Fig. 2d**), likely arising via a variant of TPRT[69]. While no TE insertions were found in protein-coding exons, 14 were intronic, including a B2 antisense to the tumor suppressor gene *Brca1* (**Fig. 2b**) and an L1 G_F antisense to the dystrophin gene *Dmd* (**Fig. 2e**). 16/18 miPSC clones (88.9%) harbored at least one *de novo* TE insertion, including all clones analyzed with both WGS and mRC-seq (**Supplementary Table 1**). Clone 2 contained the most (6) insertions. No *de novo* ERV insertions were found.

Among 277 high confidence heterozygous non-reference TE insertions (**Supplementary Table 4**) found in the parental MEF population, 97.0% were detected on average in each miPSC clone surveyed with WGS and mRC-seq. Down-sampling followed by seeking at least one WGS or mRC-seq read in support of these non-reference insertions suggested our approach would distinguish approximately 50%, 95% and 99% of *de novo* TE insertions from pre-existing

subclonal TE insertions present in 1%, 5% and 10% of cells, respectively (**Extended Data Fig. 4b**), whereas mRC-seq alone would achieve respective sensitivities of approximately 22%, 69% and 89%. Consistently, only 2/33 PCR validated TE insertions in the miPSC clones were subclonal in the parental MEFs (**Table 1**, **Extended Data Fig. 3**). An additional down-sampling analysis indicated *de novo* TE insertions were likely to be detected at a lower average WGS depth in the single-cell miPSC clones than insertions found in the bulk miPSC experiments (**Extended Data Fig. 4a**), in agreement with the greater homogeneity of the clonal miPSC cultivars. Deep sequencing of miPSCs and parental MEFs therefore enabled reliable detection and distinction of TE insertions arising before and during reprogramming.

A polymorphic retrotransposition-competent L1 eludes methylation

Six *de novo* L1 insertions found in the miPSC clones carried 3′ transductions, flanking sequences generated when PolII bypasses the native L1 polyA signal in favor of a downstream alternative[70–74] (**Table 1**). Of these transductions, 5 were either too short to reliably map to the genome, or mapped to multiple locations (**Supplementary Table 3**). The remaining 34bp transduction accompanied a 5′ truncated L1 T_F insertion on Chromosome 12 (miPSC_10_L1) (**Fig. 2f**). While the transduction aligned uniquely to Chromosome 9, a donor L1 was not present adjacent to this reference genome location. However, PCR amplification revealed an L1 T_F immediately upstream of the transduced sequence (**Fig. 2f**). This donor L1 was polymorphic in our C57BL/6×129S4Sv/Jae animals and retained a 5′ promoter comprising an unusually high number of monomers (10). Capillary sequencing confirmed the donor L1 possessed intact ORFs. L1 locus-specific bisulfite sequencing revealed that few (24.1%) of the CpG dinucleotides in the first two monomers of the donor L1 promoter were methylated in MEFs (**Fig. 2g** and **Extended Data Fig. 5**), as opposed to 7.3% in a subset of single-cell miPSC clones cultured in serum, and 1.3% for the same miPSC clones when cultured in 2i conditions (**Fig. 2g**). This difference in CpG methylation between culture conditions was significant ($p<0.05$, two-tailed t test). The donor L1 promoter was fully unmethylated in nearly all miPSCs cultured in 2i (**Fig. 2g** and **Extended Data Fig. 5**). Indeed, significantly more ($p<0.0001$, two-tailed t test) fully unmethylated sequences were found for the donor L1 promoter in 2i conditions than in serum, possibly as a consequence of global naïve state hypomethylation (**Fig. 2h**). Among the bulk reprogrammed miPSCs obtained from animals A67 and A172, which carried the donor L1 (**Fig. 2f**), only 9.1% of CpG dinucleotides were methylated

in the donor L1 promoter, and fully unmethylated sequences were identified in all miPSC lines (**Fig. 2g** and **Extended Data Fig. 5**). By contrast, in MEFs, 83.6% of CpG dinucleotides in L1 T_F promoter monomers genome-wide were methylated, compared to 45.2% among the A67 and A172 miPSC lines (**Extended Data Fig. 6**), and resembled L1 T_F promoter methylation reported elsewhere for differentiated primary cells[48,61]. L1 T_F subfamily monomers were also significantly ($p<0.001$, two-tailed t test) less methylated in 2i (34.3%) miPSC conditions than serum (53.5%), leading to an increase in fully unmethylated monomers (**Fig. 2i** and **Extended Data Fig. 6**). These bisulfite sequencing analyses highlighted genome-wide and persistent relaxation of L1 T_F methylation in miPSCs, leaving mobile L1 promoters completely unmethylated.

Reprogramming is unaffected by L1 reverse transcriptase inhibition

Lamivudine (3TC) is a potent nucleoside reverse transcriptase inhibitor known to limit engineered L1 retrotransposition without impacting telomerase or ERV mobility[75,76]. In previous retrotransposition assays conducted in cultured HeLa cells, 3TC was tested at a maximum concentration of 25µM against the codon-optimized L1SM element, reducing its mobility by ~50%[75]. By performing titration experiments to optimize the use of 3TC during miPSC generation, we determined that 3TC concentrations of up to 100µM did not reduce viability of cultured MEFs or miPSCs (**Extended Data Fig. 7a**) or MEF reprogramming efficiency (**Fig. 3a** and **Extended Data Fig. 7b-e**). Using a wild-type L1 T_F carrying an mCherry retrotransposition indicator cassette (L1-mCherry), we found 100µM 3TC reduced mouse L1 retrotransposition by ~95% in cultured HeLa cells (**Fig. 3b**). Thus, 3TC could be used, without apparent drawbacks, to limit L1-mediated mutagenesis arising during reprogramming and miPSC cultivation.

Nanopore genomic analysis of TE insertions in bulk miPSCs

A single DNA sequencing read, if of sufficient length and quality, can completely resolve a *de novo* TE insertion present in a heterogeneous cell population, as well its genomic flanks and accompanying TPRT hallmarks[77]. Long-read sequencing, as for example developed by Oxford Nanopore Technologies (ONT), is well suited to this application, and can locate TE insertions within repetitive genomic regions refractory to short-read methods[35,77–79]. As a proof-of-principle before analyzing miPSCs, we applied PCR-free ONT sequencing to 5 HeLa cell lines expanded from single colonies (~5× genome-wide depth per colony) harboring mouse L1-mCherry

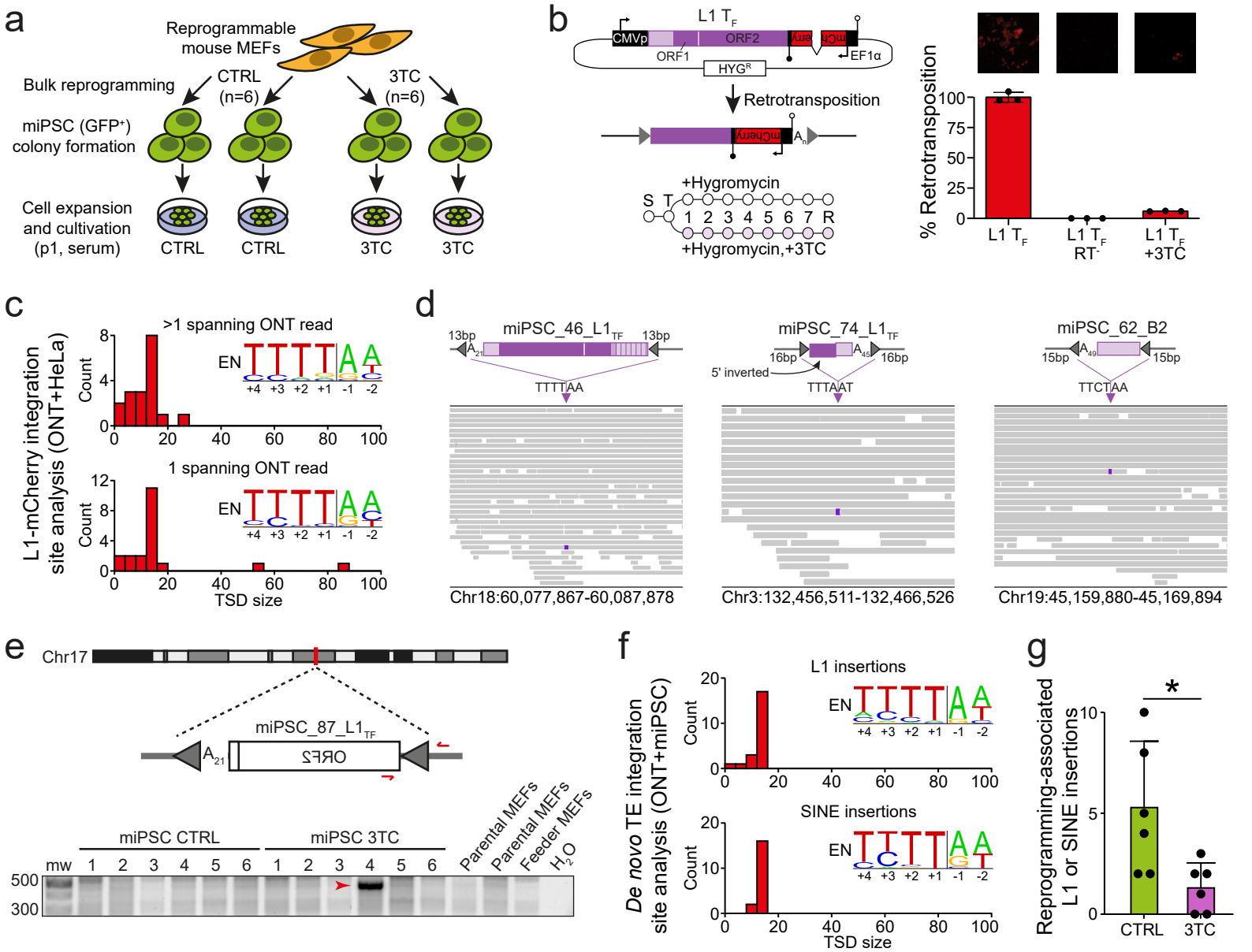


Fig. 3: Long-read detection of retrotransposition in HeLa cells and MEF-derived bulk miPSC lines. a, Bulk MEFs were reprogrammed by the addition of doxycycline. *Oct4*-GFP positive miPSCs were then sorted and expanded in serum. Six miPSC lines were reprogrammed and cultured in media containing 100µM lamivudine (3TC), and six miPSC lines generated without lamivudine (CTRL). DNA was extracted from MEFs and miPSCs and then ONT sequenced. **b,** *top left*: retrotransposition indicator plasmid L1-mCherry consists of the pCEP4 backbone (CMV promoter, black; SV40 polyadenylation signal, open lollipop; hygromycin resistance gene, white) containing a wild-type L1 T_F element (5′UTR, light purple; ORFs, dark purple). An mCherry reporter gene equipped with an EF1α promoter and HSVtk polyadenylation signal (black lollipop) is inserted into the L1 3′UTR antisense to the L1. The mCherry sequence is interrupted by an intron in sense orientation relative to the L1, ensuring mCherry expression only upon retrotransposition. *bottom left*: retrotransposition assay timeline. Cells were split (S), transfected (T), and cultured in hygromycin-containing medium with and without 100µM 3TC. Retrotransposition efficiency was assessed by flow cytometry 8 days post-transfection (R). *top right*: fluorescence microscopy images showing representative wells at 8 days post-transfection with L1 T_F (left), reverse transcriptase mutant (RT^-) L1 T_F (middle), and L1 T_F treated with 100µM 3TC (right). *bottom right*: Retrotransposition efficiency assessed by flow cytometry, relative to L1 T_F. Histogram depicts the mean and standard deviation of three independent biological replicates (black dots) consisting of three technical replicates each. **c,** Target site duplication (TSD) size distributions for L1-mCherry retrotransposition events recovered from HeLa cells via TLDR analysis of ONT sequencing, divided into integrants detected by >1 (top) or 1 (bottom) spanning read. Inset sequence logos[120] display the observed integration site motif, as preferred by the L1 endonuclease (EN). **d,** Integrative genomics viewer (IGV)[121] inspection of 3 example *de novo* TE insertions. Cartoons (top) show a full-length (>6 monomers) L1 T_F insertion, a heavily 5′ truncated L1 T_F insertion, and a SINE B2 insertion, each flanked by TSDs (gray triangles) and a 3′ polyA tract, and integrated at the indicated L1 EN motif. IGV snapshots (bottom) show ONT read alignments from the miPSC line carrying each TE insertion (purple box). **e,** A 5′ truncated intergenic *de novo* L1 T_F insertion. PolyA (A_n) tract length is indicated immediately 3′ of the L1. PCR validation primers are shown as red arrows. Underneath is shown a PCR validation agarose gel containing the 5′ junction PCR product (red arrow) only in the miPSC line where the L1 was detected by ONT sequencing. Note: DNA was obtained from two parental MEF aliquots, one corresponding to miPSC CTRL/3TC lines 1-3 and one to miPSC CTRL/3TC lines 4-6. **f,** TSD size distributions for *de novo* L1 (top) and SINE (bottom) insertions detected in miPSC lines via ONT sequencing, with inset integration site sequence logo, as per panel (c). **g,** Putative reprogramming-associated L1-mediated insertion counts detected by ONT sequencing in CTRL and 3TC-treated miPSCs. Replicate (n=6 per group) data points are marked by black dots and represented as the mean ± SD, *p<0.02, two-tailed t test.

retrotransposition events (**Supplementary Table 1**). Using the TLDR long-read TE insertion detection pipeline[35], we identified 41 L1-mCherry insertions spanned by at least one ONT read (**Supplementary Table 3**). L1-mCherry insertions overwhelmingly bore TPRT hallmarks regardless of whether they were detected by one, or more than one, ONT read (**Fig. 3c**), showing that single spanning ONT reads can reliably recover *bona fide* retrotransposition events.

Next, to survey endogenous retrotransposition in miPSCs, we ONT sequenced (~20× average genome-wide depth) 6 bulk miPSC lines reprogrammed in the presence of 100µM 3TC, 6 control miPSC lines not treated with 3TC, and matched parental MEFs (**Fig. 3a** and **Supplementary Table 1**). Using these data, TLDR identified 3,975 non-reference TE insertions carried by the parental MEFs (**Supplementary Table 4**). Of these, 3,429 (86.3%) corresponded to previously known insertions[56] and 99.6% were found in each miPSC line, on average (**Supplementary Table 4**). To gauge the general tractability of PCR validation applied to this dataset, we used a panel of 4 heterozygous non-reference TE insertions (**Supplementary Table 3**) and confirmed these all PCR amplified in the MEFs and miPSCs (**Extended Data Fig. 8a**).

An additional 43 TE insertions were each detected in only one miPSC line and not the parental MEFs, or in the earlier Illumina sequencing, and were supported by at least one spanning ONT read (**Fig. 3d** and **Supplementary Table 3**). Performing PCR validation of these insertions, a step mainly intended to exclude them being polymorphisms carried by the parental or feeder MEF populations, we could amplify one (miPSC_50_B1) in the parental MEFs (**Extended Data Fig. 8b**). The remaining 42 putative *de novo* events comprised 16, 4 and 2 L1 T_F, G_F, and A insertions, respectively, as well as 5 and 13 SINE B1 and B2 insertions, respectively, and 2 ERV insertions. One L1 T_F insertion (miPSC_69_L1) PCR amplified in multiple miPSC lines (**Extended Data Fig. 8c**) and one L1 T_F insertion (miPSC_87_L1) amplified only in the miPSC line where it was detected by ONT sequencing (**Fig. 3e**). While the remaining insertions could not be PCR amplified in any sample, all 40 of the putative *de novo* L1 and SINE insertions carried clear TPRT hallmarks (**Fig. 3f** and **Supplementary Table 3**) and on this basis we considered them *bona fide* retrotransposition events. As well, both ERVs corresponded to the mobile intracisternal A-particle (IAP) family, presented a typical proviral structure of two long terminal repeats flanking an internal coding sequence, and generated TSDs of the expected size (6bp)[80,81] (**Supplementary Table 3**). It was not clear whether the difference in (two) *de novo* ERV insertions being found here by ONT sequencing, and none by the earlier Illumina sequencing of different miPSC lines, was

due to chance or unknown technical reasons. Significantly fewer ($p<0.02$, two-tailed t test) putative reprogramming-associated L1-mediated insertions were found on average in the 3TC-treated miPSCs (~1.3 per line) than in the control miPSCs (~5.2 per line) (**Fig. 3g**), consistent with L1 inhibition by 3TC (**Fig. 3b**). Overall, detection of endogenous retrotransposition events in bulk miPSCs by ONT sequencing yielded results orthogonal and complementary to our short-read genomic analyses.

Genome-wide DNA demethylation during reprogramming focused on young L1 loci

A major feature of reprogramming mouse fibroblasts to a pluripotent state is globally reduced DNA methylation[43,44,47,48]. Although bisulfite sequencing can estimate the overall methylation of TE families, at specific genomic loci it can typically only resolve CpGs close to the termini of full-length L1s not located in highly repetitive regions. To generate a comprehensive genome-wide view of DNA methylation changes during reprogramming, and complement our bisulfite sequencing data, we analyzed the ONT data obtained from MEFs and miPSCs (control and 3TC-treated) using Methylartist[35,82]. While methylation in control miPSCs was reduced genome-wide when compared to MEFs, on protein-coding gene promoters and amongst all of the TE families considered, the very youngest L1 subfamilies (T_{FI} and T_{FII}) displayed by far the greatest median methylation change (-54.0%) (**Fig. 4a**, **Extended Data Fig. 9a,b** and **Supplementary Table 5**). 90.3% of full-length L1 T_{FI} and T_{FII} copies were significantly ($p<0.01$, Fisher's exact test with Bonferroni correction, and ΔmC >25%) less methylated in control miPSCs than MEFs (**Supplementary Table 5**), with this demethylation most pronounced in the monomeric L1 5′UTR (**Fig. 4b**). By contrast, we observed no significant local or global differences in L1 T_F or protein-coding gene promoter methylation between control and 3TC-treated miPSCs (**Fig. 4** and **Supplementary Table 5**). Thirty-six L1s initiated transcription of a spliced mRNA from their 5′UTR, as defined by GenBank expressed sequence tags, including alternative promoters for protein-coding genes expressed in pluripotent cells, such as *Fsd1l* (**Fig. 4c** and **Supplementary Table 5**). We also identified full-length L1s demethylated in both MEFs and miPSCs (**Extended Data Fig. 9c**), in line with prior human data suggesting certain L1 loci evade DNA methylation in differentiated cells[34,83]. In sum, ONT analysis showed global reprogramming-associated demethylation was most accentuated for the youngest L1s, where retrotransposition potential is concentrated, creating opportunities for L1-driven mobilization and protein-coding gene

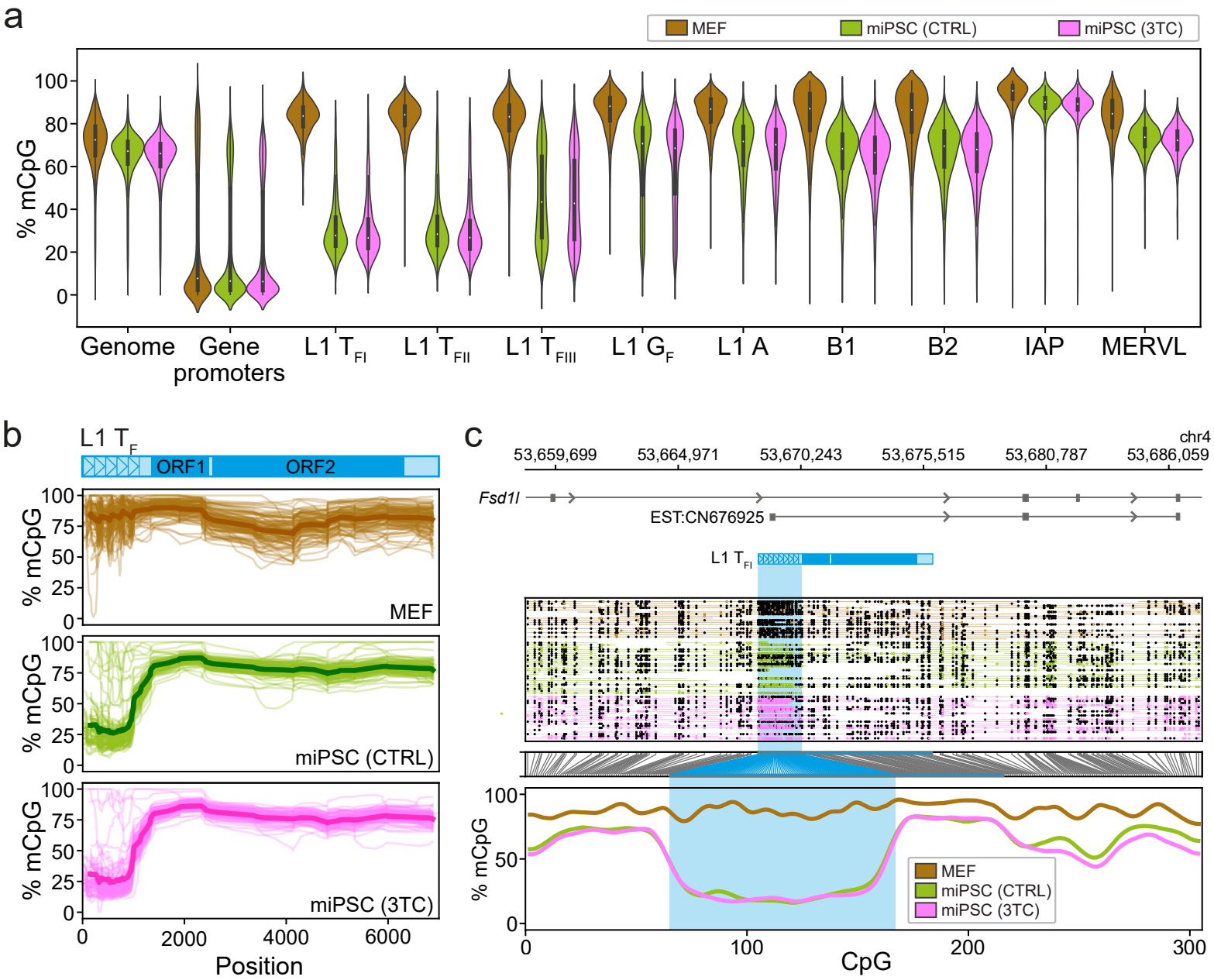


Fig. 4: Genome-wide methylation analyses via ONT sequencing. a, CpG methylation ascertained by ONT sequencing of MEFs (brown), control miPSCs (CTRL, green), and miPSCs derived in the presence of lamivudine (3TC, pink). Results are shown for the whole genome (10kbp windows), the proximal promoters (-1000,+500) of protein-coding genes[119], the 5′UTR of T_F, G_F, and A-type L1s >6kbp, B1 and B2 SINEs, and MERVL MT2 and IAP long terminal repeats. Note: replicates from control and 3TC-treated miPSCs are displayed in aggregate. **b,** Composite L1 T_F methylation profiles. Each graph displays 100 profiles. A schematic of the T_F consensus is provided at top. Average values are indicated by more thickly colored lines. **c,** Methylation profile of the *Fsd1l* locus obtained by ONT sequencing. The first panel shows an L1 T_F orientated in sense to intron 6 of *Fsd1l*, as well as an expressed sequence tag (EST) obtained from a mouse ESC sample and supporting a transcript initiated in the T_F 5′UTR and spliced into a downstream *Fsd1l* exon. The second panel displays ONT read alignments, with unmethylated CpGs colored in brown (MEFs), green (control miPSCs) and pink (3TC-treated miPSCs), methylated CpGs colored black, and CpGs not confidently called, i.e. abs(log-likelihood ratio) > 2.5, omitted. The displayed miPSC data were downsampled to the approximate depth of the MEF data with Picard DownsampleSam. The third panel indicates the relationship between CpG positions in genome space and CpG space, including those corresponding to the T_F 5′UTR (shaded light blue). The fourth panel indicates the fraction of methylated CpGs. Note: this L1 T_F is polymorphic amongst inbred mouse strains[56].

alternative promoters.

Discussion

This study demonstrates miPSCs incompletely silence mobile TE families and routinely harbor *de novo* TE insertions. While some TE insertions occur in parental cells and are inherited by miPSCs, our data suggest the majority arise during reprogramming or very early upon reaching pluripotency. In support of this view, firstly, we observed profound hypomethylation of young L1 promoters in miPSCs and not parental cells. As shown elsewhere, L1 mRNA abundance is low in fibroblasts and increases greatly upon reprogramming[45,46,52], while engineered L1 reporter genes retrotranspose >10-fold more frequently in hiPSCs and hESCs than in fibroblasts[51,52]. Secondly, 38/41 *de novo* TE insertions detected by Illumina sequencing PCR validated in only one miPSC line each. These and the 42 putative *de novo* TE insertions identified by ONT sequencing were absent from all other samples in the study, as assayed by PCR and deep WGS. Finally, given the similar mutation calling thresholds of Illumina sequencing applied to heterogeneous (bulk) and homogenous (single-cell clone) miPSC populations, we note that private SNVs (~100 per line) and SVs (~1 per line) were detected at similar frequencies in either experiment, whereas far more *de novo* retrotransposition events were found in the latter dataset. One explanation for this result is that a relatively small number of clones dominate bulk reprogramming experiments[84] and most SNVs and SVs predate reprogramming[10,11,13], while retrotransposons mainly mobilize during reprogramming. This position is consistent with a prior WGS analysis that, alongside thousands of SNVs, identified no somatic L1 insertions among 10 human fibroblast clones generated from single cells[85]. It is nonetheless possible that additional somatic variants would have been annotated if primary single-cell clones, as analyzed elsewhere[86], were reprogrammed. However, the introduction of multiple bottlenecks followed by clonal expansion prolongs cell culture and could thereby exaggerate mutation frequencies. The parental MEFs used here cannot in any case be clonally expanded from single cells, and reprogram extremely inefficiently after more than three passages in culture[87,88], and for these reasons we did not prepare single-cell MEF clones prior to reprogramming.

Previous experiments employing hiPSCs and mouse and human ESCs showed L1 de-repression and mobilization were likely to take place in pluripotent cells[34,41,42,45,46,48,50–52,89]. Notably, 31/56 (55.4%) *de novo* L1 insertions found here in miPSCs were full-length, a similar

percentage to that observed previously in hiPSCs (57.1%)[45]. New full-length L1 insertions have potential for further retrotransposition and were largely unmethylated in miPSCs. Their CpG dinucleotides presented a "sloping shore" of methylation, as found elsewhere for newly retrotransposed CpG islands[34,35,48,90], where methylation decreases from the L1 5′ genome junction and forms a trough before sharply increasing over the L1 ORFs. Only 3/56 (5.4%) L1 insertions corresponded to the A subfamily, while the remainder were T_F and G_F elements, consistent with relative activity levels revealed by sequencing extended mouse pedigrees and mouse tumors[28,61]. *De novo* SINE B1 and B2 insertions, mediated *in trans* by the L1 protein machinery[91], were also detected in miPSCs, in line with L1-mediated *Alu* SINE insertions arising in hiPSCs and hESCs[45,89,92]. Discovery of *de novo* TE insertions in low-passage miPSCs derived from multiple parental cell types suggests endogenous retrotransposition may be an intrinsic risk of the epigenome remodeling required for the acquisition of pluripotency[7,43–45,47].

Exonic retrotransposon insertions are clear in their potential to cause disease[93]. However, the L1 endonuclease does not favor exons[65,66], which are depleted for its AT-rich recognition motif and make up only ~2% of the genome[94], and none of the 81 *de novo* L1-mediated insertions reported here in miPSCs were exonic. By comparison, L1 integration within introns occurs much more frequently. We observed 30/81 (37.0%) L1-mediated insertions in introns, in line with the proportion of the genome occupied by these regions[65,66]. Whilst intronic events are less likely to be pathogenic than exonic insertions, they can perturb gene expression, for example through reduced transcriptional elongation[95] or, as shown here, the provision of new promoter elements[96]. Retrotransposition into the introns of protein-coding genes, as observed here for *Brca1* and *Dmd*, could therefore undermine miPSC models of human disease. Such mutations necessitate screening of miPSC lines[4]. Fortunately, strategies to minimize TE-mediated mutagenesis, including via the use of 3TC or another L1 reverse transcriptase inhibitor[75], appear achievable without affecting DNA methylation or reprogramming efficiency. While the frequency of *de novo* endogenous L1-mediated insertions found in miPSCs was significantly reduced by 3TC, a handful of events, likely arising prior to reprogramming, were nonetheless observed in 3TC-treated miPSCs. Reverse transcriptase inhibitors thus may be incorporated into future miPSC derivation protocols to attenuate *de novo* retrotransposition, in addition to genomic screening of miPSC lines.

Methods

Ethics statement

All animal experimentation was performed under the auspices and approval of the Monash University Animal Research Platform Animal Ethics Committee (Approval Numbers MARP-2011-172-Polo, MARP-2011-171-BC-Polo, MARP-2017-151-BC-Polo, and ERM# 21634).

Adult *Oct4*GFP-OKSM-M2rtTA mouse tissue somatic cell isolation and reprogramming

Induced pluripotent stem cells were generated from adult and embryonic *Oct4*GFP-OKSM-M2rtTA doxycycline inducible reprogrammable mice[53]. These animals are heterozygous for an *Oct4*-GFP reporter and an OKSM cassette targeted to the *Collagen1α1* locus, and homozygous for the ROSA26-M2rtTA allele from the ubiquitous *ROSA26* locus. The polycistronic cassette is under the control of a tetracycline-dependent promoter *(tetOP)*. Hence, upon the addition of doxycycline, M2rtTA binds to the *tetOP*, thereby inducing OKSM expression. *Oct4*GFP-OKSM-M2rtTA mice were housed at the Monash University Animal Research Platform animal facility.

Bone marrow extraction and FACS purification of granulocytes and hematopoietic stem (LSK) cells were performed as previously described[97]. In brief, harvested bone marrow cells were labeled using a two-step sequential antibody labeling procedure using the following primary conjugated antibodies: 1:200 dilution of Anti-Mouse CD5 FITC antibody (BD Biosciences, Cat#: 553020), 1:100 dilution of Anti-Mouse B220 FITC antibody (BD Biosciences, Cat#: 557669), 1:200 dilution of Anti-Mouse TER-119 FITC antibody (BD Biosciences, Cat#: 557915), 1:400 dilution of Anti-Mouse Sca-1 PB antibody (Biolegend, Cat#: E13-161.7), 1:200 dilution of Anti-Mouse cKit APC antibody (BD Biosciences, Cat#: 553356), 1:200 dilution of Anti-Mouse SSEA1 Biotinylated antibody (Thermo Fisher Scientific, Cat#: 13-8813-80), 1:200 dilution of Anti-Mouse Gr-1 APC-Cy7 antibody (Biolegend, Cat#: 108423) and 1:1000 dilution of Anti-Mouse Mac1 PE antibody (Biolegend, Cat#: 101207). This was followed by the secondary labeling step with 1:200 dilution of Streptavidin PE-Cy7 antibody (BD Biosciences, Cat#: 557598). Cells were isolated and sorted using an Influx Cell Sorter Instrument (BD Biosciences) with a 100µm nozzle. Samples were resuspended in phosphate buffered saline (PBS) supplemented with 2% fetal bovine serum (Thermo Fisher Scientific, Cat#: SH30071.03FBS, Hyclone). FACS sorting for these and the cell types below were performed with 2µg/mL Propidium Iodide (PI) (Sigma Aldrich, Cat#: P4864) in order to exclude non-viable cells. Granulocytes were isolated using the following cell surface

marker profile: $CD5^-/B220^-/Ter119^-/Sca1^-/cKit^-/SSEA1^-/Gr1^+/Mac1^+$, whilst LSK cells were isolated from bone marrow using the following cell surface marker profile: $CD5^-/B220^-/Ter119^-/Sca1^+/cKit^+/SSEA1^-/Gr1^-/Mac1^-$.

Fibroblasts were isolated from both ear lobes from each mouse. Tissue pieces were resuspended in 0.25% Trypsin-EDTA (Thermo Fisher Scientific, Cat#: 25200-072) solution, and after 5min incubation at room temperature, were mechanically minced using two surgical blades for a further 2min. iPSC medium was used to inactivate trypsin, and dissociated pieces were transferred to a 15mL centrifuge tube (Corning). Tissue pieces were then transferred to a gelatin coated T-75 flask (Corning) and cells were left to grow for a further 7 days. $CD45^-/CD31^-/Thy1.2^{hi+}$ fibroblasts were fractionated by FACs using the following antibodies: a 1:100 dilution of Anti-Mouse CD31 antibody conjugated to FITC (Thermo Fisher Scientific, Cat#: 11-0311-81), a 1:100 dilution of Anti-Mouse CD45 antibody conjugated to FITC (Thermo Fisher Scientific, Cat#: 11-0451-810) and a 1:400 dilution of Anti-Mouse Thy-1.2 antibody conjugated to APC (Thermo Fisher Scientific, Cat#: 17-0902-81).

Liver epithelial cells were isolated according to an adaptation of a previously described method[98]. Briefly, 3mg/mL Collagenase Type 1 (Sigma-Aldrich, Cat#: C1639) solution was prepared in sterile PBS. Whole liver was transferred into a sterile 6cm petri dish and finely minced using fine dissecting scissors. Minced liver pieces were transferred to 15mL tube with preheated Collagenase Type 1 (Sigma, Cat#: C1639). Tubes were left to agitate on a Thermomix (Eppendorf) at 750rpm, 37°C for 15min. Following digestion, the tube was removed and the cellular suspension was triturated with an 18G needle, until tissue chunks were mostly dissociated. Sample tubes were then left to agitate for an additional 15min, until liver fragments were completely digested. The sample suspension was again triturated, with a 21G needle, to generate a single cell suspension, and then processed through a 40µm cell strainer into a clean 50mL centrifuge tube (Corning). After rinsing in 2% FCS/PBS (wash buffer) and centrifuging for 5min at 1380rpm for 4°C, the supernatant was removed and cells were resuspended in wash buffer and centrifuged once again. Cells were counted and 5×10^6 cells were resuspended for sorting. Cells were labeled with primary antibodies using a 1:100 dilution of Anti-mouse CD31 antibody conjugated to FITC (Thermo Fisher Scientific, Cat#: 11-0311-81), followed by a 1:100 dilution of Anti-mouse CD45 antibody conjugated to FITC (Thermo Fisher Scientific, Cat#: 11-0451-81) and 1:100 dilution of Anti-mouse EpCAM antibody conjugated to eFluor450 (Thermo Fisher Scientific, Cat#: 48-5791-82).

Liver epithelial cells were isolated using the following cell surface marker profile: $CD45^-/CD31^-/EpCAM^{+hi}$.

Thymus tissue was processed for thymic epithelial cell isolation as previously described[99]. Cells were labeled with the following antibodies: 1:400 dilution of Anti-mouse CD45 antibody conjugated to APC-Cy7 (BD Biosciences, Cat#: 557659), 1:200 dilution of Anti-mouse TER-119 antibody conjugated to APC-Cy7 (BD Biosciences, Cat#: 560509), 1:6000 dilution of Anti-mouse MHC Class II antibody conjugated to PB (Biolegend, Cat#: 107620) and 1:1000 dilution of Anti-mouse EpCAM antibody conjugated to APC (Biolegend, Cat#: 118214). Thymic epithelial cells were sorted according to the following cell surface marker profile: $CD45^-/Ter119^-$/MHC Class $II^+/EpCAM^+$.

Intestinal stem cells were purified as previously described[100]. Cells were labeled with a 1:200 dilution of Anti-mouse CD45 antibody conjugated to BV510 (BD Biosciences, Cat#: 563891), 1:200 dilution of Anti-mouse CD31 antibody conjugated to BV510 (BD Biosciences, Cat#: 563089), a 1:100 dilution of Anti-mouse CD24 antibody conjugated to Pe-Cy7 (Thermo Fisher Scientific, Cat#: 25-0242-82), a 1:100 dilution of Anti-mouse EpCAM antibody conjugated to eFluor450 (Thermo Fisher Scientific, Cat#: 48-5791-82), and 1:100 Anti-EphrinB2 unconjugated antibody (BD Biosciences, Cat#: 743763). In the secondary labeling step, a 1:200 dilution of Anti-mouse Alexa Fluor 555 polyclonal antibody (Thermo Fisher Scientific, Cat#: A-31570) was used to detect the EphrinB2 antibody. Intestinal stem cells were fractionated according to the following cell surface marker profile: $CD45^-/CD31^-/CD24^+/EpCAM^+/Ephrin^+$.

To obtain astrocytes, brain tissue was processed using a MACS Neural Tissue Dissociation Kit (T) (Miltenyi Biotec, Cat#: 130-093-231) and manually dissected according to manufacturer's instructions. Cells were then collected and incubated with antibodies directed against Glast1 (Allophycocyanin-conjugated, ACSA-1, 1:10 dilution) (Miltenyi Biotec, Cat#: 130-098-803), 1:100 dilution of Anti-mouse CD133 antibody conjugated to PE (Thermo Fisher Scientific, Cat#: 12-1331-80), 1:200 dilution of Anti-mouse CD45 antibody conjugated to PE-Cy7 (BD Biosciences, Cat#: 552848) and 1:200 dilution of Anti-mouse CD31 antibody conjugated to PE-Cy7 (Thermo Fisher Scientific, Cat#: 25-0311-82). Astrocytes were sorted and purified according to the following cell surface marker profile: $CD45^-/CD31^-/CD133^-/GLAST1^+$.

Keratinocytes and bulge stem cells were isolated from epidermis as previously described[101]. Cells were collected and incubated with antibodies against Anti-Mouse Integrin alpha

6 antibody (GoH3) conjugated to PE (1:600) (Abcam, Cat#: ab95703), a 1:200 CD104 antibody conjugated to FITC (Biolegend, Cat#: 346-11A) and a 1:100 dilution of Anti-mouse CD34 biotinylated antibody (Thermo Fisher Scientific, Cat#: 13-0341-85) for 20min at 4°C. For secondary antibody labeling, cells were incubated with 1:200 APC-Streptavidin antibody (Biolegend, Cat#: 405207) to detect CD34 biotinylated antibody for 20min at 4°C. They were then washed and resuspended in PI (2µg/mL) 1% BSA/PBS (Sigma-Aldrich, Cat#: A8412) and passed through a 40µm cell strainer (BD Falcon) to produce single cell suspensions. Cells with the surface marker profile of $CD104^+/CD34^+/\alpha6\text{-integrin}^+$ were defined as bulge stem cells, and those marked as $\alpha6\text{-integrin}^-/CD34^+$ were defined as keratinocytes.

Reprogramming of the above 9 primary cell types was performed as follows: cells were seeded into gelatinized tissue culture treated 6-well plates (Corning Costar, Cat#: CLS3506) and cultured at 37°C and 5% CO_2 in iPSC media containing KnockOut DMEM (Thermo Fisher Scientific, Cat#: 10829-018), 15% Fetal Bovine Serum (FBS) (Thermo Fisher Scientific, Cat#: SH30071.03), GlutaMAX Supplement (Thermo Fisher Scientific, Cat#: 35050061), Penicillin-Streptomycin (Thermo Fisher Scientific, Cat#: 15070063), MEM Non-Essential Amino Acids Solution (Thermo Fisher Scientific, Cat#: 11140050), 2-Mercaptoethanol (Thermo Fisher Scientific, Cat#: 21985023) and 1000U/mL Leukemia Inhibitory Factor (LIF) (Merck Millipore, Cat#: ESG1107), supplemented with 2µg/mL of doxycycline (dox) (Sigma-Aldrich, Cat#: 33429-100MG-R). iPSC medium supplemented with dox was replaced every alternate day after the first 3 days of reprogramming and withdrawn 4 days after the presence of iPSC-like colonies had formed, with typical dome-shaped iPSC morphology. Cells were then cultured to confluency on a layer of irradiated MEFs prior to further FACs purification and enrichment for Oct-GFP^+ cells. Purified *Oct4*-GFP iPSCs were then bulk expanded in $175cm^2$ cell culture flasks (Corning, Cat#: CLS430825) and then frozen at a density of 1×10^6 cells/vial.

Mouse embryonic fibroblast isolation and reprogramming

Reprogrammable mouse embryonic fibroblast (MEF) cultures were derived as described previously[102] from a E13.5dpc *Oct4*GFP-OKSM-M2rtTA embryo (animal I222e2) and cultivated at 37°C, 5% O_2, 5% CO_2 in MEF medium containing DMEM High Glucose (Thermo Fisher Scientific, Cat# 11960-044) with 10% FBS (Thermo Fisher Scientific, Cat#: SH30071.03), 1mM Sodium Pyruvate (Thermo Fisher Scientific, Cat#: 11360-070), GlutaMAX Supplement (Thermo

Fisher Scientific, Cat#: 35050061), Penicillin-Streptomycin (Thermo Fisher Scientific, Cat#: 15070063), MEM Non-Essential Amino Acids Solution (Thermo Fisher Scientific, Cat#: 11140050) and 2-Mercaptoethanol (Thermo Fisher Scientific, Cat#: 21985023). MEFs were reprogrammed by being placed in iPSC medium supplemented with 2µg/mL dox (Sigma-Aldrich, Cat#: 33429-100MG-R) and cultured on irradiated MEFs at 37°C, 5% CO_2. iPSC colonies were discerned according to GFP expression in the absence of dox. In addition to bulk iPSC cultures (see below), single *Oct4*-GFP$^+$ cells were deposited via FACS individually into 96-well pre-gelatinized tissue culture plates (Falcon, Cat#: 353072). Eighteen single-cell clones were bulk expanded on 6-well pre-gelatinized tissue culture plates (Falcon, Cat#: 353046) and maintained in serum or 2i conditions (see below).

Oct4-GFP$^+$ iPSC flow cytometry

For flow cytometry, cells were harvested by dissociating in 0.25% Trypsin EDTA (Life Technologies) to yield a single cell suspension, and then resuspended in FACS wash (Phosphate Buffered Saline with 2% Fetal Calf Serum) containing PI. Live cells were gated on the basis of forward scatter, side scatter and PI exclusion. Flow cytometric gates were set using control iPSCs that did not have endogenous GFP expression. Tubes were sorted according to GFP expression using an Influx Cell Sorter Instrument (Becton Dickinson). Data collected were analyzed and presented using FlowJo software. Sorted GFP$^+$ cells were then plated down on T-25 flasks (Corning) and expanded onto T-150 flasks (Corning), before being frozen down at a density of 1×10^6 cells/vial.

Serum and serum-free iPSC culture

Mouse iPSCs were maintained on irradiated primary MEFs, as previously described[8,103]. Briefly, iPSCs were cultured on 0.2% Porcine Gelatin (Sigma-Aldrich, Cat#: G1890-500G) coated tissue culture plates and flasks (Corning) on a feeder layer of irradiated MEFs (2×10^4 cells/cm^2). iPSC medium was changed daily and cells were cultured at 37°C and 5% CO_2. Passaging was performed when iPSCs reached 70% confluency. Alternatively, iPSCs were cultured on irradiated MEFs in serum-free media containing knockout serum replacement (KOSR) and 2i/LIF[104]. Here, cells were cultured in DMEM (Thermo Fisher Scientific, Cat#: 11960-044), 1000U/mL LIF (Merck Millipore, Cat#: ESG1107), 0.1mM 2-Mercaptoethanol (Thermo Fisher Scientific, Cat#:

21985023), 1mM GlutaMAX Supplement (Thermo Fisher Scientific, Cat#: 35050061), 1% Sodium Pyruvate (Thermo Fisher Scientific, Cat#: 11360-070), 0.1mM MEM Non-Essential Amino Acids Solution (Thermo Fisher Scientific, Cat#: 11140050), 1% Penicillin-Streptomycin (Thermo Fisher Scientific, Cat#: 15070063), with medium supplemented with 15% KOSR (Thermo Fisher Scientific, Cat#: 10828-028), 1µm Mek1/2 Inhibitor (PD0325901) (Tocris, Cat#: 4192) and 3µm GSK3a/b inhibitor (CHIR99021) (Tocris, Cat#: 4423). Prior to genomic DNA extraction, iPSCs depleted from irradiated feeders were dissociated with 0.5% Trypsin EDTA (Thermo Fisher Scientific, Cat#: 25200-072). The irradiated MEFs were feeder depleted with 10mL of iPSC media for 45min in non-gelatinized T-25 flasks (Corning, Cat#: CLS3056). The resultant iPSCs were collected as a supernatant in suspension medium.

Lamivudine titration experiments

iPSCs were cultured with primary irradiated MEFs, as above, for 9 days in concentrations of lamivudine (3TC, Sigma-Aldrich, Cat#: L1295-10MG) ranging from 0 to 200µM and cell survival calculated as a % of the starting population. Reprogrammable OKSM, rtTA3 MEFs were isolated from embryonic day 13.5 embryos from *Oct4*-GFP;ROSA-rtTA-out;OKSM-72 mice as previously described[44,105]. Doxycycline inducible reprogrammable MEFs were grown in media containing 2µg/mL dox (Sigma Aldrich Cat#: 33429-100MG-R) and 0-200µM 3TC for 15 days, with the percentage cell survival calculated at days 3, 7, 10 and 15. Once 100µM was identified as the optimal concentration of 3TC to assess its impact on L1 retrotransposition, 30,000 reprogrammable MEFs at passage 2 were seeded onto gelatinized 6-well plates and reprogrammed in dox for 12 days, then cultured for an additional 4 days without dox. *Oct4*-GFP^{+} iPSCs were then purified via flow cytometric sorting and expanded on irradiated MEFs for an additional 11 days, then feeder depleted prior to DNA extraction for ONT sequencing. Reprogramming and iPSC media contained serum, and either 100µM 3TC or no 3TC.

Illumina sequencing and genomic analysis

Genomic DNA was harvested from MEFs and iPSCs using a DNeasy Blood and Tissue Kit (Qiagen, Cat#: 60594). DNA was quantified by a Qubit dsDNA HS Assay Kit (Life Technologies, Cat#: Q32851) on a Qubit Fluorometer 3.0 (Life Technologies). For WGS, libraries were

generated using an Illumina TruSeq DNA PCR-free kit (Illumina, Cat#: 20015962) and sequenced separately on an Illumina HiSeq X Ten platform (Macrogen, Korea).

For mRC-seq, libraries were prepared as follows: 1µg genomic DNA was sheared using a Covaris M220 Focused Ultrasonicator in a 130µL microTUBE AFA fiber snap-cap vial (Covaris, Cat#: 520045). The following parameters were used to gain 500bp insert libraries: 50W, duty factor 20%, 200 cycles per burst, duration 55s. Size selection to remove fragments <300 bp was performed using Agencourt AMPure XP beads (Beckman Coulter, Cat#: A63881) with a 1:0.6 DNA:beads ratio. Libraries were then generated by TruSeq Nano DNA LT kit (Illumina, Cat#: 20015964) using TruSeq DNA Single Indexes (Illumina, Cat#: 20015960 and 20015961) and run on a 2% agarose gel (Bioline, Cat#: BIO-41025) pre-stained with SYBR Safe Nucleic Acid Gel Stain (Invitrogen, Cat#: S33102). For ~500bp insert size libraries the target gel fragment size was 600-650bp, which was excised under a Safe Imager 2.0 Blue-Light Transilluminator (Invitrogen). DNA was purified using a MinElute Gel Extraction Kit (Qiagen, Cat#: 28606) according to the manufacturer's instructions. DNA was eluted in 25µL molecular grade water. Enrichment of DNA fragments was performed as described for Illumina TruSeq Nano DNA LT Kit (Illumina, Cat#: 20015964). Sample clean up was performed with Agencourt AMPure XP beads (Beckman Coulter, Cat#: A63881) using a 1:1.1 ratio of DNA to beads. Amplified libraries were eluted in 30µL molecular grade water and quantified using a Bioanalyzer DNA 1000 chip (Agilent Technologies, Cat#: 5067-1504).

mRC-seq hybridization was performed as previously described[28]. Hybridization reactions were washed using SeqCap Hybridization and Wash Kit (Roche, Cat#: 05634261001) and DNA eluted in 50µL molecular grade water. Two post-hybridization LM-PCR reactions per sample were performed using 20µL Enhanced PCR Mix, 5µL PCR Primer Cocktail from the Illumina TruSeq Nano DNA LT Kit (Illumina, Cat#: 20015964) and 25µL sample. PCR was performed with the following cycling conditions: 95˚C for 3min, 8 cycles of 98˚C for 20s, 60˚C for 15s, and 72˚C for 30s, followed by 72˚C for 5min. The two PCR reactions for each sample were pooled and cleaned up using the QIAquick PCR Purification Kit (Qiagen) and samples eluted in 15µL Elution Buffer (Qiagen, Cat#: 28706). Quantity and fragment size were determined using a Bioanalyzer DNA 1000 chip (Agilent Technologies, Cat#: 5067-1504). Libraries were pooled and sequenced on an Illumina HiSeq X Ten platform (Macrogen, Korea).

Reads were aligned to the mm10 reference genome using bwa-mem[106] version 0.7.12 with

parameters -M -Y. Duplicate reads were marked via Picard MarkDuplicates version 1.128. Indel Realignment was carried out via GATK IndelRealigner (3.7). SNVs were called by GATK HaplotypeCaller 3.7[54] to generate GVCFs and GenotypeGVCFs to obtain cohort-level calls. SNVs were also called using freebayes[55] filtered to remove known mouse strain germline variants[56]. SVs were called using Delly2 and GRIDSS 2.0.0[57,58], using calls with concordant non-filtered precise breakends. SNVs and SVs, including potential private variants, were genotyped using the full catalog of variants obtained from all samples, including MEFs. Variant impact prediction and annotation was carried out using SnpEff version 4.3T[107]. WGS and mRC-seq aligned BAMs were processed to identify non-reference TE insertions using TEBreak[60] (https://github.com/adamewing/tebreak) as previously described[61].

TE insertion PCR validation experiments

Reads supporting putative *de novo* TE insertions were manually examined using Serial Cloner (http://serialbasics.free.fr/Serial_Cloner.html), the UCSC Genome Browser BLAT tool[108] and the Repbase CENSOR tool[109]. PCR primers were designed with Primer3[110] against TE insertion sequences and their 5′ and 3′ genomic flanks (**Supplementary Table 3**). Empty/filled PCRs (combining 5′ and 3′ flanking primers) and full-length PCRs (using junction-spanning primers) were performed using an Expand Long Range dNTPack (Roche, Cat#: 4829034001). Reaction mixes contained 5µL 5× Expand Long Range Buffer with 12.5mM $MgCl_2$, 1.25µL dNTP Mix (dATP, dCTP, dGTP, dTTP at 10mM each), 1.25µL DMSO (100%), 1µL primer mix (25µM of each primer), 0.35µL Expand Long Range Enzyme Mix (5U/µL), 4-10ng genomic DNA template, and molecular grade water up to a total volume of 25µL. PCR was performed with the following cycling conditions: 92˚C for 3min, 10 cycles of 92˚C for 30s, 56-60˚C for 30s, and 68˚C for 7min 30s 25 cycles of 92˚C for 30s, 56-60˚C for 30s, and 68˚C for 7min + 20s cycle elongation for each successive cycle, followed by 68˚C for 10min. TE-genome junction validation PCRs were performed using MyTaq HS DNA Polymerase (Bioline, Cat#: BIO-2111). Reaction mixes contained 5µL 5× MyTaq Reaction Buffer, 0.5µL primer mix (25µM of each primer), 0.2µL MyTaq HS DNA Polymerase, 2-4ng genomic DNA template, and molecular grade water up to a total volume of 25µL. PCRs were performed using the following conditions: 95˚C for 2min, 35 cycles of 95˚C for 15s, 55/57˚C for 15s, and 72˚C for 10s, followed by 72˚C for 10min. PCR products were run on 0.8-2% agarose gels (Bioline, Cat#: BIO-41025), depending on fragment

size, pre-stained with SYBR Safe Nucleic Acid Gel Stain (Invitrogen, Cat#: S33102). A Typhoon FLA 9000 (GE Healthcare Life Sciences) was used for gel imaging. Gel fragments were excised under a Safe Imager 2.0 Blue-Light Transilluminator (Invitrogen). DNA purification was performed using the QIAquick Gel Extraction Kit (Qiagen, Cat#: 28706) or MinElute Gel Extraction Kit (Qiagen, Cat#: 28606) according to the manufacturer's instructions. PCR fragments were either sequenced directly or cloned using the pGEM-T Easy Vector System (Promega, Cat#: A1360) and Sanger sequenced to resolve insertion characteristics, as shown in **Supplementary Table 3**.

L1-mCherry retrotransposition assays

The L1-mCherry construct was derived from the construct pTN201, a pCEP4-based vector containing the native mouse element $L1_{spa}$[22]. The $L1_{spa}$ coding sequence was modified by site-directed mutagenesis to include two nonsynonymous nucleotide substitutions, rendering the ORF1p amino acid sequence identical to that of the L1 T_F subfamily consensus sequence[111,112]. The 3′UTR was interrupted by a reporter cassette based on previously described L1 retrotransposition indicator plasmids[19,113]. This reporter cassette consisted of the mCherry coding sequence in antisense orientation to the L1 and was equipped with an EF1α promoter and HSVtk polyadenylation signal. The mCherry ORF was interrupted by a β-globin intron oriented in sense to the L1. The mCherry cassette was cloned using G-block double-stranded DNA fragments synthesized by Integrated DNA Technologies (IDT) and PCR products generated using Q5 DNA polymerase (New England Biolabs, Cat#: M0492). The mCherry coding sequence was synthesized with silent mutations ablating potential splice donor and splice acceptor sites that could interfere with intended splicing of the intron. In the L1-mCherry construct, the final 157bp of the $L1_{spa}$ 3′UTR, which included a conserved poly-purine tract, was situated downstream of the mCherry cassette and immediately upstream of the pCEP4 SV40 polyadenylation signal[112]. The L1-mCherry_RT- mutant contained a missense mutation in the reverse transcriptase domain of ORF2 (D709Y)[22]. Plasmids were prepared using a Qiagen Plasmid Plus Midi Kit and a QIAvac vacuum manifold (Qiagen, Cat#: 12145).

HeLa-JVM cells[19] were cultured at 37°C and 5% CO_2 in HeLa complete medium (DMEM, Life Technologies, Cat#: 11960044) supplemented with 10% FBS (Life Technologies, Cat#: 10099141), 1% Glutamax (Life Technologies, Cat#: 35050061) and 1% penicillin-streptomycin

(Life Technologies, Cat#: 15140122). Cells were passaged at 70-80% confluency using 0.25% Trypsin-EDTA (Life Technologies, Cat#: 25200072). Cultured cell retrotransposition assays were then performed as described previously[67,113], except retrotransposition was detected by mCherry fluorescence instead of EGFP fluorescence. Briefly, 1×10^5 HeLa-JVM cells were seeded per well of a 6-well plate. Eighteen hours later, cells were transfected with 1µg L1-mCherry or L1-mCherry_RT- plasmid per well using 3µL FuGENE HD transfection reagent (Promega, Cat#: E2311) and 97µL Opti-MEM (Life Technologies, Cat#: 31985047) per well according to the manufacturer's protocol. Twenty-four hours post-transfection, medium was replaced with either HeLa complete medium with 200µg/mL Hygromycin (Life Technologies, Cat#: 10687010), or HeLa complete medium with 200µg/mL Hygromycin and 100µM Lamivudine (Sigma-Aldrich, Cat#: L1295-10MG). Medium was replaced every other day, and at 8 days post-transfection cells were collected by trypsinization, resuspended in sterile PBS, and analyzed on a CytoFLEX flow cytometer (Beckman Coulter) to determine the percentage of mCherry positive cells. Three biological replicate assays were performed, each consisting of 3 assayed wells per condition (technical replicates).

L1-mneoI retrotransposition assays

To prepare reporter constructs, miPSC_1_L1 and miPSC_4_L1 were amplified from genomic DNA using an Expand Long Range dNTPack (Roche, Cat#: 4829034001). Reaction mixes contained 5µL 5× Expand Long Range Buffer with 12.5mM $MgCl_2$, 1.25µL dNTP Mix (dATP, dCTP, dGTP, dTTP at 10mM each), 1.25µL DMSO (100%), 1µL primer mix (50µM of each primer), 0.35µL Expand Long Range Enzyme Mix (5U/µL), 10ng genomic DNA template and molecular grade water, up to a total volume of 25µL. PCRs were performed with the following cycling conditions: 92˚C for 3min, 10 cycles of 92˚C for 30s, 58˚C for 30s, and 68˚C for 7min 30s; 25 cycles of 92˚C for 30sec, 58˚C for 30s, and 68˚C for 7min plus 20s elongation for each successive cycle, followed by 68˚C for 10min. Primers introduced a *NotI* restriction site at the L1 5′ end (miPSC_1_L1_F, 5′-tttgcggccgcagaaagggaataatcgaggtg-3′; miPSC_1_L1_R, 5′-gctaagcttgagaataagtgaagga-3′; miPSC_4_L1_F, 5′-agggcggccgcaggattaagaacccaatcaccag-3′; miPSC_4_L1_R, 5′-aaaatgcctgttgtgccaat-3′). Reactions were purified using agarose gel electrophoresis. Target fragments were excised and purified using either traditional phenol-chloroform extraction or QIAquick and MinElute Gel Extraction Kits (Qiagen, Cat#: 28706 and

28604). Each L1 was then cloned into pGEMT Easy Vector (Promega, Cat#: A1360). Ligations were incubated overnight at 4˚C. Ligation reactions were transformed using One Shot TOP10 chemically competent *E. coli* (Invitrogen, Cat#: C404010). Blue/white screening was performed using LB/ampicillin/IPTG/X-Gal plates. At least 3 positive colonies per L1 were chosen for Miniprep culture and plasmid DNA was isolated using a QIAprep Spin Miniprep Kit (Qiagen, Cat#: 27106). At least three clones per element were capillary sequenced and compared to identify PCR-induced mutations. Full-length L1s were then reconstructed by combining PCR-mutation free fragments from different clones using restriction enzymes (New England Biolabs) recognizing the L1 sequence. Reactions were purified using agarose gel electrophoresis and target fragments were excised and purified using QIAquick and MinElute Gel Extraction Kits (Qiagen, Cat#: 28706 and 28604).

pTN201 was used to generate L1 reporter constructs. pTN201 is composed of a pCEP4 backbone (Life Technologies) containing $L1_{spa}$, a retrotransposition-competent L1 T_F[22] and a downstream mneoI retrotransposition reporter cassette[114]. The mneoI cassette is driven by an SV40 promoter and holds the neomycin resistance gene, which is interrupted by an intron and is positioned antisense to L1spa. In this assay, neomycin (or its analog, Geneticin/G418) resistance only occurs via transcription, splicing and integration of the L1 and mneoI cassette into genomic DNA[19,67]. To measure miPSC_1_L1 and miPSC_4_L1 retrotransposition efficiency, $L1_{spa}$ was removed from the pCEP4 backbone by digesting with *NotI* and *PacI*. The pCEP4 backbone was dephosphorylated using Calf Intestinal Alkaline Phosphatase (CIP) (New England Biolabs, Cat#: M0290). The backbone and fragments of either miPSC_1_L1 or miPSC_4_L1 were combined in a single ligation reaction using T4 DNA Ligase (New England Biolabs, Cat#: M0202) and incubated overnight at 16˚C. Ligations were transformed using One Shot TOP10 chemically competent *E. coli* (Invitrogen, Cat#: C404010) and plasmid DNA of positive clones was obtained using QIAprep Spin Miniprep Kit (Qiagen, Cat#: 27106). Clones were verified as mutation-free by capillary sequencing. Plasmid DNA for retrotransposition assays was obtained using a Plasmid Maxi Kit (Qiagen, Cat#: 12163). Each construct was built with and without a cytomegalovirus promoter (CMVp) preceding the L1. In addition, the following controls, each based on a pCEP4 backbone containing the mneoI cassette, were employed: TGF21, a retrotransposition-competent L1 G_F[21]; L1SM, a synthetic codon optimized mouse L1[68]; L1SMmut2, L1SM immobilized by reverse transcriptase and endonuclease domains mutations[68].

Retrotransposition assays were performed as previously described[67], with minor modifications. HeLa-JVM cells were grown in HeLa complete medium (DMEM, Life Technologies, Cat#: 11960044) supplemented with 10% FBS (Life Technologies, Cat#: 10099141), 1% Glutamax (Life Technologies, Cat#: 35050061) and 1% penicillin-streptomycin (Life Technologies, Cat#: 15140122), and then seeded at a density of 4×10^4 cells/well in 6-well tissue culture plates. 14-16h after plating, cells were transfected with L1 reporter constructs using 4µL FuGENE HD transfection reagent (Promega, Cat#: E2311) 96µL Opti-MEM (Life Technologies, Cat#: 31985047) and 1µg plasmid DNA per well. Transfection efficiencies were determined in parallel by preparing transfection mixes containing 4µL FuGENE HD transfection reagent (Promega, Cat#: E2311), 96µL Opti-MEM (Life Technologies, Cat#: 31985047), 0.5µg L1 expression plasmid and 0.5µg pCEP4-eGFP. The transfection mixture was added to each well containing 2mL DMEM-complete medium. Plates were incubated at 37˚C and 5% CO_2, medium replaced 24h post-transfection, and transfection efficiency determined 72h post-transfection. pCEP4-eGFP transfected wells were trypsinized and cells were collected from each well and centrifuged at 2000g for 5min. Cell pellets were resuspended in 300-500µL 1× PBS. The number of eGFP-positive cells was determined using a CytoFLEX flow cytometer (Beckman Coulter). The percentage of eGFP-positive cells was used to normalize the G418-resistant colony counts for each L1 reporter construct[67]. G418 (400µg/mL) (Thermo Fisher Scientific, Cat#: 10131035) selection was started 3 days post-transfection and performed for 12 days. G418 foci were washed with 1× PBS and fixed using 2% Formaldehyde/0.2% Glutaraldehyde in 1× PBS (Sigma-Aldrich) fixing solution at room temperature for 30min. Staining was done using 0.1% Crystal Violet solution (Sigma-Aldrich) at room temperature for 10min. Foci were counted in each well to quantify retrotransposition.

L1 bisulfite sequencing experiments

Bisulfite conversion was performed with 200ng input genomic DNA from miPSC lines and MEFs using a EZ DNA Methylation-Lightning Kit (Zymo Research, Cat#: D5030), following the manufacturer's instructions. DNA was eluted in 10µL Elution Buffer. The internal sequences of L1 T_F monomers were amplified genome-wide with the following primers: BS_TfIII_mono_F, 5′-GGAAATTAGTTTGAATAGGTTAGAGGGTG; BS_TfIII_mono_R, 5′-TCCTAAATTCCAAAAAATCCTAAAACCAAA. The following locus-specific primers were

used to target the 5′ promoter region of the following elements of interest: BS_miPSC_1_L1_F, 5′-TGATTTATTTTTGATTGAATTTATTTTTAT; BS_miPSC_1_L1_R/donor_L1_R, 5′-CTATTCAAACTAATTTCCTAAATTCTACTA; BS_miPSC_3_L1_F, 5′-TAGTTGGGGTTGTATGATGTAAGTT; BS_miPSC_3_L1_R, 5′-TCCCAAAAACTATCTAATTCTCTAAC; BS_miPSC_4_L1_F, 5′-TTTATATTGAAGGTTTGGATGATTTTATAT; BS_miPSC_4_L1_R, 5′-TCCAATTCTCTAATACACCCTCTAAC; BS_donor_L1_F, 5′-TTAAAGAAGTTAGTGATTTTTTAGAATTTT.

PCRs were performed using MyTaq HS DNA Polymerase (Bioline, Cat#: BIO-21111). Reaction mixes contained 5µL 5× MyTaq Reaction Buffer, 0.5µL primer mix (25µM of each primer), 0.2µL MyTaq HS DNA Polymerase, DMSO at a final concentration of 0.1%, 2µL bisulfite converted DNA template, and molecular grade water up to a total volume of 25µL. PCR cycling parameters were as follows: 95˚C for 2min, 40 cycles of 95˚C for 30s, 54˚C for 30s, and 72˚C for 30s, followed by 72˚C for 5min. PCR products were run on a 2% agarose gel, excised and purified using a MinElute Gel Extraction Kit (Qiagen, Cat#: 28604) according to the manufacturer's instructions. Illumina libraries were constructed using a NEBNext Ultra™ II DNA Library Prep Kit (New England Biolabs, Cat#: E7645). Libraries were quantified using a Bioanalyzer DNA 1000 chip (Agilent Technologies, Cat#: 5067-1504). Barcoded libraries were pooled in equimolar amounts and sequenced as 2x300mer reads on an Illumina MiSeq platform using a MiSeq Reagent Kit v3 (Illumina, Cat#: MS-102-3003). 50% PhiX Control v3 (Illumina, Cat#: FC-110-3001) was used as a spike-in. Sequencing data were analyzed as described previously[34]. To summarize, for the L1 T_F genome-wide analysis, paired-end reads were considered separately and those with the L1 T_F bisulfite PCR primers at their termini were retained and aligned to the mock converted T_F monomer target amplicon sequence with blastn. Reads where non-CpG cytosine bisulfite conversion was <95%, or ≥5% of CpG dinucleotides were mutated, or ≥5% of adenine and guanine nucleotides were mutated, were removed. 50 reads per sample, excluding identical bisulfite sequences, were randomly selected and analyzed using QUMA[115] with default parameters, with strict CpG recognition. Specific L1 loci were analyzed in a similar fashion, except paired-end reads were assembled into contigs, as described elsewhere[34], prior to blastn alignment to the mock converted L1 locus target amplicon.

Nanopore sequencing analyses

Genomic DNA was extracted from 6 miPSC lines reprogrammed in the presence of 100µM 3TC, 6 miPSC lines generated without 3TC, and the parental MEFs, with a Nanobind CBB Big DNA Kit (Circulomics, Cat#: NB-900-001-01) according to the manufacturer's instructions. DNA libraries were prepared at the Kinghorn Centre for Clinical Genomics (KCCG, Australia) using 3µg input DNA, without shearing, and an SQK-LSK110 ligation sequencing kit. Libraries were each sequenced separately on a PromethION (Oxford Nanopore Technologies) flow cell (FLO-PRO002, R9.4.1 chemistry) (**Supplementary Table 1**). Bases were called with guppy 5.0.13 (Oxford Nanopore Technologies).

Non-reference TE insertions were detected with TLDR[35]. Briefly, this involved aligning ONT reads to the mm10 reference genome using minimap2[116] version 2.20 (index parameter: -x map-ont; alignment parameters: -ax map-ont -L -t 32) and SAMtools[117] version 1.12. BAM files were then processed as a group with TLDR[35] version 1.2.2 (parameters -e teref.mouse.fa -p 128 -m 1 -r mm10.fa -n nonref.collection.mm10.chr.bed.gz --keep_pickles). The files teref.mouse.fa, composed of TE family consensus sequences, and nonref.collection.mm10.chr.bed.gz, a collection of known non-reference retrotransposon insertions, are available from github.com/adamewing/tldr/. The TLDR output table was further processed to remove calls not passing relevant TLDR filters, where family = "NA" or remappable = "FALSE" or UnmapCover < 0.5 or LengthIns < 100 or EndTE-StartTE < 100 or strand = "None" or SpanReads < 1. 3′ truncated TE insertions, and B1 or B2 insertions 5′ truncated by more than 2bp, were removed. Events detected in only one miPSC line and not matching a known non-reference insertion were designated as putative *de novo* insertions (**Supplementary Table 3**).

Reference TE methylation was assessed for parental MEFs and miPSC lines aggregated by condition (reprogrammed with or without 3TC), using Methylartist version 1.2.4[82]. Briefly, CpG methylation calls were generated from ONT reads using nanopolish version 0.13.2[118]. Using Methylartist commands db-nanopolish, segmeth and segplot with default parameters, methylation statistics were generated for the genome divided into 10kbp bins, protein-coding gene promoters defined the Eukaryotic Promoter Database (-1000bp,+500bp)[119], and reference TEs defined by RepeatMasker coordinates (http://www.repeatmasker.org/). TE families displayed in **Fig. 4a** included T_F, G_F, and A-type L1s >6kbp, B1 and B2 SINEs, and MERVL MT2 and IAP elements represented by their long terminal repeats. Methylation values were calculated for L1 5′UTRs only,

excluding the L1 body. Methylation profiles for individual loci were generated using the Methylartist command locus. L1 T_F methylation profiles shown in **Fig. 4b** were generated for elements >7kbp with the Methylartist command composite. To identify individual differentially methylated TEs (**Supplementary Table 5**), we required elements to have at least 4 reads and 20 methylation calls in each of the MEF, aggregated control miPSC and aggregated 3TC-treated miPSC datasets. Statistical comparisons were performed based on methylated and unmethylated CpG call counts, using Fisher's exact test with Bonferroni correction for multiple testing.

L1-mCherry retrotransposition events were recovered via ONT sequencing in a similar fashion to the analysis of mouse samples, except genomic DNA obtained from 5 HeLa cell lines expanded over 3-5 passages from single L1-mCherry insertion-harboring colonies was barcoded using a SQK-MLK111.96-XL kit, pooled in equimolar amounts, and sequenced on a single PromethION flow cell. Reads were aligned to the hg38 human reference genome build. L1-mCherry insertions were then identified using TLDR, as for the endogenous mouse TE analysis described above.

Data availability

All Oxford Nanopore Technologies and Illumina sequencing data generated by this study were deposited in the European Nucleotide Archive (ENA) under project PRJEB20569.

Code availability

TEBreak, TLDR and Methylartist, and instructions for their use, are available at https://github.com/adamewing/tebreak, https://github.com/adamewing/TLDR and https://github.com/adamewing/methylartist, respectively.

References

1. Takahashi, K. & Yamanaka, S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* **126**, 663–676 (2006).
2. Mandai, M. *et al.* Autologous Induced Stem-Cell–Derived Retinal Cells for Macular Degeneration. *N. Engl. J. Med.* **376**, 1038–1046 (2017).
3. Takahashi, K. & Yamanaka, S. A decade of transcription factor-mediated reprogramming to pluripotency. *Nat. Rev. Mol. Cell Biol.* **17**, 183–193 (2016).

4. Tapia, N. & Schöler, H. R. Molecular Obstacles to Clinical Translation of iPSCs. *Cell Stem Cell* **19**, 298–309 (2016).
5. Gore, A. *et al.* Somatic coding mutations in human induced pluripotent stem cells. *Nature* **471**, 63–67 (2011).
6. Hussein, S. M. *et al.* Copy number variation and selection during reprogramming to pluripotency. *Nature* **471**, 58–62 (2011).
7. Lister, R. *et al.* Hotspots of aberrant epigenomic reprogramming in human induced pluripotent stem cells. *Nature* **471**, 68–73 (2011).
8. Polo, J. M. *et al.* Cell type of origin influences the molecular and functional properties of mouse induced pluripotent stem cells. *Nat. Biotechnol.* **28**, 848–855 (2010).
9. Laurent, L. C. *et al.* Dynamic changes in the copy number of pluripotency and cell proliferation genes in human ESCs and iPSCs during reprogramming and time in culture. *Cell Stem Cell* **8**, 106–118 (2011).
10. Young, M. A. *et al.* Background mutations in parental cells account for most of the genetic heterogeneity of induced pluripotent stem cells. *Cell Stem Cell* **10**, 570–582 (2012).
11. Cheng, L. *et al.* Low incidence of DNA sequence variation in human induced pluripotent stem cells generated by nonintegrating plasmid expression. *Cell Stem Cell* **10**, 337–344 (2012).
12. Quinlan, A. R. *et al.* Genome sequencing of mouse induced pluripotent stem cells reveals retroelement stability and infrequent DNA rearrangement during reprogramming. *Cell Stem Cell* **9**, 366–373 (2011).
13. Abyzov, A. *et al.* Somatic copy number mosaicism in human skin revealed by induced pluripotent stem cells. *Nature* **492**, 438–442 (2012).
14. Kazazian, H. H., Jr & Moran, J. V. Mobile DNA in Health and Disease. *N. Engl. J. Med.* **377**, 361–370 (2017).
15. Doucet, A. J., Wilusz, J. E., Miyoshi, T., Liu, Y. & Moran, J. V. A 3' Poly(A) Tract Is Required for LINE-1 Retrotransposition. *Mol. Cell* **60**, 728–741 (2015).
16. Jurka, J. Sequence patterns indicate an enzymatic involvement in integration of mammalian retroposons. *Proc. Natl. Acad. Sci. U. S. A.* **94**, 1872–1877 (1997).
17. Luan, D. D., Korman, M. H., Jakubczak, J. L. & Eickbush, T. H. Reverse transcription of R2Bm RNA is primed by a nick at the chromosomal target site: a mechanism for non-LTR

retrotransposition. *Cell* **72**, 595–605 (1993).

18. Monot, C. *et al.* The specificity and flexibility of l1 reverse transcription priming at imperfect T-tracts. *PLoS Genet.* **9**, e1003499 (2013).
19. Moran, J. V. *et al.* High frequency retrotransposition in cultured mammalian cells. *Cell* **87**, 917–927 (1996).
20. Wei, W. *et al.* Human L1 retrotransposition: cis preference versus trans complementation. *Mol. Cell. Biol.* **21**, 1429–1439 (2001).
21. Goodier, J. L., Ostertag, E. M., Du, K. & Kazazian, H. H., Jr. A novel active L1 retrotransposon subfamily in the mouse. *Genome Res.* **11**, 1677–1685 (2001).
22. Naas, T. P. *et al.* An actively retrotransposing, novel subfamily of mouse L1 elements. *EMBO J.* **17**, 590–597 (1998).
23. Nellåker, C. *et al.* The genomic landscape shaped by selection on transposable elements across 18 mouse strains. *Genome Biol.* **13**, R45 (2012).
24. Skowronski, J., Fanning, T. G. & Singer, M. F. Unit-length line-1 transcripts in human teratocarcinoma cells. *Mol. Cell. Biol.* **8**, 1385–1397 (1988).
25. Beck, C. R. *et al.* LINE-1 retrotransposition activity in human genomes. *Cell* **141**, 1159–1170 (2010).
26. Brouha, B. *et al.* Hot L1s account for the bulk of retrotransposition in the human population. *Proc. Natl. Acad. Sci. U. S. A.* **100**, 5280–5285 (2003).
27. Ewing, A. D. & Kazazian, H. H., Jr. High-throughput sequencing reveals extensive variation in human-specific L1 content in individual human genomes. *Genome Res.* **20**, 1262–1270 (2010).
28. Richardson, S. R. *et al.* Heritable L1 retrotransposition in the mouse primordial germline and early embryo. *Genome Res.* **27**, 1395–1405 (2017).
29. Stewart, C. *et al.* A comprehensive map of mobile element insertion polymorphisms in humans. *PLoS Genet.* **7**, e1002236 (2011).
30. Feusier, J. *et al.* Pedigree-based estimation of human mobile element retrotransposition rates. *Genome Res.* **29**, 1567–1577 (2019).
31. de la Rica, L. *et al.* TET-dependent regulation of retrotransposable elements in mouse embryonic stem cells. *Genome Biol.* **17**, 234 (2016).
32. Warkocki, Z. *et al.* Uridylation by TUT4/7 Restricts Retrotransposition of Human LINE-1s.

Cell **174**, 1537–1548.e29 (2018).

33. Deniz, Ö., Frost, J. M. & Branco, M. R. Regulation of transposable elements by DNA modifications. *Nat. Rev. Genet.* **20**, 417–431 (2019).
34. Sanchez-Luque, F. J. *et al.* LINE-1 Evasion of Epigenetic Repression in Humans. *Mol. Cell* **75**, 590–604 (2019).
35. Ewing, A. D. *et al.* Nanopore Sequencing Enables Comprehensive Transposable Element Epigenomic Profiling. *Mol. Cell* **80**, 915–928.e5 (2020).
36. Bourc'his, D. & Bestor, T. H. Meiotic catastrophe and retrotransposon reactivation in male germ cells lacking Dnmt3L. *Nature* **431**, 96–99 (2004).
37. Walter, M., Teissandier, A., Pérez-Palacios, R. & Bourc'his, D. An epigenetic switch ensures transposon repression upon dynamic loss of DNA methylation in embryonic stem cells. *Elife* **5**, e11418 (2016).
38. Greenberg, M. V. C. & Bourc'his, D. The diverse roles of DNA methylation in mammalian development and disease. *Nat. Rev. Mol. Cell Biol.* **20**, 590–607 (2019).
39. Goodier, J. L. Restricting retrotransposons: a review. *Mob. DNA* **7**, 16 (2016).
40. Chelmicki, T. *et al.* m6A RNA methylation regulates the fate of endogenous retroviruses. *Nature* **591**, 312–316 (2021).
41. MacLennan, M. *et al.* Mobilization of LINE-1 retrotransposons is restricted by Tex19.1 in mouse embryonic stem cells. *Elife* **6**, e26152 (2017).
42. Salvador-Palomeque, C. *et al.* Dynamic Methylation of an L1 Transduction Family during Reprogramming and Neurodifferentiation. *Mol. Cell. Biol.* **39**, e00499–18 (2019).
43. Lee, D.-S. *et al.* An epigenomic roadmap to induced pluripotency reveals DNA methylation as a reprogramming modulator. *Nat. Commun.* **5**, 5619 (2014).
44. Knaupp, A. S. *et al.* Transient and Permanent Reconfiguration of Chromatin and Transcription Factor Occupancy Drive Reprogramming. *Cell Stem Cell* **21**, 834–845.e6 (2017).
45. Klawitter, S. *et al.* Reprogramming triggers endogenous L1 and Alu retrotransposition in human induced pluripotent stem cells. *Nat. Commun.* **7**, e10286 (2016).
46. Friedli, M. *et al.* Loss of transcriptional control over endogenous retroelements during reprogramming to pluripotency. *Genome Res.* **24**, 1251–1259 (2014).
47. Maherali, N. *et al.* Directly reprogrammed fibroblasts show global epigenetic remodeling

and widespread tissue contribution. *Cell Stem Cell* **1**, 55–70 (2007).

48. Gerdes, P. *et al.* Mouse L1s fade with age: a methylation-enforced mechanism for attenuation of L1 retrotransposition potential. *bioRxiv* (2022) doi:10.1101/2022.08.06.500894.
49. Habibi, E. *et al.* Whole-genome bisulfite sequencing of two distinct interconvertible DNA methylomes of mouse embryonic stem cells. *Cell Stem Cell* **13**, 360–369 (2013).
50. Garcia-Perez, J. L. *et al.* LINE-1 retrotransposition in human embryonic stem cells. *Hum. Mol. Genet.* **16**, 1569–1577 (2007).
51. Macia, A. *et al.* Engineered LINE-1 retrotransposition in nondividing human neurons. *Genome Res.* **27**, 335–348 (2017).
52. Wissing, S. *et al.* Reprogramming somatic cells into iPS cells activates LINE-1 retroelement mobility. *Hum. Mol. Genet.* **21**, 208–218 (2012).
53. Stadtfeld, M., Maherali, N., Borkent, M. & Hochedlinger, K. A reprogrammable mouse strain from gene-targeted embryonic stem cells. *Nat. Methods* **7**, 53–55 (2010).
54. McKenna, A. *et al.* The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* **20**, 1297–1303 (2010).
55. Garrison, E. & Marth, G. Haplotype-based variant detection from short-read sequencing. *arXiv [q-bio.GN]* arXiv:1207.3907 (2012).
56. Keane, T. M. *et al.* Mouse genomic variation and its effect on phenotypes and gene regulation. *Nature* **477**, 289–294 (2011).
57. Rausch, T. *et al.* DELLY: structural variant discovery by integrated paired-end and split-read analysis. *Bioinformatics* **28**, i333–i339 (2012).
58. Cameron, D. L. *et al.* GRIDSS: sensitive and specific genomic rearrangement detection using positional de Bruijn graph assembly. *Genome Res.* **27**, 2050–2060 (2017).
59. Ewing, A. D. Transposable element detection from whole genome sequence data. *Mob. DNA* **6**, 24 (2015).
60. Carreira, P. E. *et al.* Evidence for L1-associated DNA rearrangements and negligible L1 retrotransposition in glioblastoma multiforme. *Mob. DNA* **7**, 21 (2016).
61. Schauer, S. N. *et al.* L1 retrotransposition is a common feature of mammalian hepatocarcinogenesis. *Genome Res.* **28**, 639–653 (2018).
62. Ostertag, E. M. & Kazazian, H. H., Jr. Twin priming: a proposed mechanism for the

creation of inversions in L1 retrotransposition. *Genome Res.* **11**, 2059–2065 (2001).

63. Kano, H. *et al.* L1 retrotransposition occurs mainly in embryogenesis and creates somatic mosaicism. *Genes Dev.* **23**, 1303–1312 (2009).
64. van den Hurk, J. A. J. M. *et al.* L1 retrotransposition can occur early in human embryonic development. *Hum. Mol. Genet.* **16**, 1587–1592 (2007).
65. Sultana, T. *et al.* The Landscape of L1 Retrotransposons in the Human Genome Is Shaped by Pre-insertion Sequence Biases and Post-insertion Selection. *Mol. Cell* **74**, 555–570 (2019).
66. Flasch, D. A. *et al.* Genome-wide de novo L1 Retrotransposition Connects Endonuclease Activity with Replication. *Cell* **177**, 837–851 (2019).
67. Kopera, H. C. *et al.* LINE-1 cultured cell retrotransposition assay. *Methods Mol. Biol.* **1400**, 139–156 (2016).
68. Han, J. S. & Boeke, J. D. A highly active synthetic mammalian retrotransposon. *Nature* **429**, 314–318 (2004).
69. Meyer, T. J., Srikanta, D., Conlin, E. M. & Batzer, M. A. Heads or tails: L1 insertion-associated 5' homopolymeric sequences. *Mob. DNA* **1**, 7 (2010).
70. Miki, Y. *et al.* Disruption of the APC gene by a retrotransposal insertion of L1 sequence in a colon cancer. *Cancer Res.* **52**, 643–645 (1992).
71. Goodier, J. L., Ostertag, E. M. & Kazazian, H. H., Jr. Transduction of 3'-flanking sequences is common in L1 retrotransposition. *Hum. Mol. Genet.* **9**, 653–657 (2000).
72. Pickeral, O. K., Makałowski, W., Boguski, M. S. & Boeke, J. D. Frequent human genomic DNA transduction driven by LINE-1 retrotransposition. *Genome Res.* **10**, 411–415 (2000).
73. Moran, J. V., DeBerardinis, R. J. & Kazazian, H. H., Jr. Exon shuffling by L1 retrotransposition. *Science* **283**, 1530–1534 (1999).
74. Holmes, S. E., Dombroski, B. A., Krebs, C. M., Boehm, C. D. & Kazazian, H. H., Jr. A new retrotransposable human L1 element from the LRE2 locus on chromosome 1q produces a chimaeric insertion. *Nat. Genet.* **7**, 143–148 (1994).
75. Banuelos-Sanchez, G. *et al.* Synthesis and Characterization of Specific Reverse Transcriptase Inhibitors for Mammalian LINE-1 Retrotransposons. *Cell Chem Biol* **26**, 1095–1109.e14 (2019).
76. Dai, L., Huang, Q. & Boeke, J. D. Effect of reverse transcriptase inhibitors on LINE-1 and

Ty1 reverse transcriptase activities and on LINE-1 retrotransposition. *BMC Biochem.* **12**, 18 (2011).

77. Smits, N. *et al.* No evidence of human genome integration of SARS-CoV-2 found by long-read DNA sequencing. *Cell Rep.* **36**, 109530 (2021).
78. Lanciano, S. & Cristofari, G. Measuring and interpreting transposable element expression. *Nat. Rev. Genet.* **21**, 721–736 (2020).
79. Zhou, W. *et al.* Identification and characterization of occult human-specific LINE-1 insertions using long-read sequencing technology. *Nucleic Acids Res.* **48**, 1146–1163 (2020).
80. Wolf, G. *et al.* KRAB-zinc finger protein gene expansion in response to active retrotransposons in the murine lineage. *Elife* **9**, e56337 (2020).
81. Heidmann, O. & Heidmann, T. Retrotransposition of a mouse IAP sequence tagged with an indicator gene. *Cell* **64**, 159–170 (1991).
82. Cheetham, S. W., Kindlova, M. & Ewing, A. D. Methylartist: Tools for Visualising Modified Bases from Nanopore Sequence Data. *Bioinformatics* **38**, 3109–3112 (2022).
83. Scott, E. C. *et al.* A hot L1 retrotransposon evades somatic repression and initiates human colorectal cancer. *Genome Res.* **26**, 745–755 (2016).
84. Shakiba, N. *et al.* Cell competition during reprogramming gives rise to dominant clones. *Science* **364**, eaan0925 (2019).
85. Saini, N. *et al.* The Impact of Environmental and Endogenous Damage on Somatic Mutation Load in Human Skin Fibroblasts. *PLoS Genet.* **12**, e1006385 (2016).
86. Kuijk, E. *et al.* The mutational impact of culturing human pluripotent and adult stem cells. *Nat. Commun.* **11**, 2493 (2020).
87. Nefzger, C. M., Alaei, S., Knaupp, A. S., Holmes, M. L. & Polo, J. M. Cell surface marker mediated purification of iPS cell intermediates from a reprogrammable mouse model. *J. Vis. Exp.* e51728 (2014).
88. Utikal, J. *et al.* Immortalization eliminates a roadblock during cellular reprogramming into iPS cells. *Nature* **460**, 1145–1148 (2009).
89. Macia, A. *et al.* Epigenetic control of retrotransposon expression in human embryonic stem cells. *Mol. Cell. Biol.* **31**, 300–316 (2011).
90. Grandi, F. C. *et al.* Retrotransposition creates sloping shores: a graded influence of

hypomethylated CpG islands on flanking CpG sites. *Genome Res.* **25**, 1135–1146 (2015).

91. Dewannieux, M. & Heidmann, T. L1-mediated retrotransposition of murine B1 and B2 SINEs recapitulated in cultured cells. *J. Mol. Biol.* **349**, 241–247 (2005).

92. Dewannieux, M., Esnault, C. & Heidmann, T. LINE-mediated retrotransposition of marked Alu sequences. *Nat. Genet.* **35**, 41–48 (2003).

93. Kazazian, H. H., Jr *et al.* Haemophilia A resulting from de novo insertion of L1 sequences represents a novel mechanism for mutation in man. *Nature* **332**, 164–166 (1988).

94. International Human Genome Sequencing Consortium. Initial sequencing and analysis of the human genome. *Nature* **409**, 860–921 (2001).

95. Han, J. S., Szak, S. T. & Boeke, J. D. Transcriptional disruption by the L1 retrotransposon and implications for mammalian transcriptomes. *Nature* **429**, 268–274 (2004).

96. Faulkner, G. J. *et al.* The regulated retrotransposon transcriptome of mammalian cells. *Nat. Genet.* **41**, 563–571 (2009).

97. Firas, J., Liu, X., Nefzger, C. M. & Polo, J. M. GM-CSF and MEF-conditioned media support feeder-free reprogramming of mouse granulocytes to iPS cells. *Differentiation* **87**, 193–199 (2014).

98. Bertoncello, I. & McQualter, J. Isolation and clonal assay of adult lung epithelial stem/progenitor cells. *Current Protocols in Stem Cell Biology* **16**, (2011).

99. Seach, N., Wong, K., Hammett, M., Boyd, R. L. & Chidgey, A. P. Purified enzymes improve isolation and characterization of the adult thymic epithelium. *J. Immunol. Methods* **385**, 23–34 (2012).

100. Merlos-Suárez, A. *et al.* The intestinal stem cell signature identifies colorectal cancer stem cells and predicts disease relapse. *Cell Stem Cell* **8**, 511–524 (2011).

101. Jensen, K. B., Driskell, R. R. & Watt, F. M. Assaying proliferation and differentiation capacity of stem cells using disaggregated adult mouse epidermis. *Nat. Protoc.* **5**, 898–911 (2010).

102. Alaei, S. *et al.* An improved reprogrammable mouse model harbouring the reverse tetracycline-controlled transcriptional transactivator 3. *Stem Cell Res.* **17**, 49–53 (2016).

103. Lim, S. M. *et al.* Temporal restriction of pancreatic branching competence during embryogenesis is mirrored in differentiating embryonic stem cells. *Stem Cells Dev.* **21**, 1662–1674 (2012).

104. Liu, K. *et al.* KSR-based medium improves the generation of high-quality mouse iPS cells. *PLoS One* **9**, e105309 (2014).
105. Knaupp, A. S. *et al.* TINC—A Method to Dissect Regulatory Complexes at Single-Locus Resolution—Reveals an Extensive Protein Complex at the Nanog Promoter. *Stem cell reports* **15**, 1246–1259 (2020).
106. Li, H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv [q-bio.GN]* arXiv:1303.3997 (2013).
107. Cingolani, P. *et al.* A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of Drosophila melanogaster strain w1118; iso-2; iso-3. *Fly* **6**, 80–92 (2012).
108. Kent, W. J. *et al.* The human genome browser at UCSC. *Genome Res.* **12**, 996–1006 (2002).
109. Bao, W., Kojima, K. K. & Kohany, O. Repbase Update, a database of repetitive elements in eukaryotic genomes. *Mob. DNA* **6**, 11 (2015).
110. Untergasser, A. *et al.* Primer3--new capabilities and interfaces. *Nucleic Acids Res.* **40**, e115 (2012).
111. Martin, S. L. *et al.* A single amino acid substitution in ORF1 dramatically decreases L1 retrotransposition and provides insight into nucleic acid chaperone activity. *Nucleic Acids Res.* **36**, 5845–5854 (2008).
112. Richardson, S. R. *et al.* Revisiting the impact of synthetic ORF sequences on engineered LINE-1 retrotransposition. *bioRxiv* (2022) doi:10.1101/2022.08.29.505632.
113. Ostertag, E. M., Prak, E. T., DeBerardinis, R. J., Moran, J. V. & Kazazian, H. H., Jr. Determination of L1 retrotransposition kinetics in cultured cells. *Nucleic Acids Res.* **28**, 1418–1423 (2000).
114. Freeman, J. D., Goodchild, N. L. & Mager, D. L. A modified indicator gene for selection of retrotransposition events in mammalian cells. *Biotechniques* **17**, 46, 48–9, 52 (1994).
115. Kumaki, Y., Oda, M. & Okano, M. QUMA: quantification tool for methylation analysis. *Nucleic Acids Res.* **36**, W170–5 (2008).
116. Li, H. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* **34**, 3094–3100 (2018).
117. Li, H. *et al.* The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**, 2078–2079 (2009).

118. Simpson, J. T. *et al.* Detecting DNA cytosine methylation using nanopore sequencing. *Nat. Methods* **14**, 407–410 (2017).
119. Dreos, R., Ambrosini, G., Groux, R., Cavin Périer, R. & Bucher, P. The eukaryotic promoter database in its 30th year: focus on non-vertebrate organisms. *Nucleic Acids Res.* **45**, D51–D55 (2017).
120. Crooks, G. E., Hon, G., Chandonia, J.-M. & Brenner, S. E. WebLogo: a sequence logo generator. *Genome Res.* **14**, 1188–1190 (2004).
121. Robinson, J. T. *et al.* Integrative genomics viewer. *Nat. Biotechnol.* **29**, 24–26 (2011).

Acknowledgements

The authors thank Jef D. Boeke and John V. Moran for sharing L1SM plasmids and the HeLa-JVM cell line, respectively. This study was supported by an Australian Government Research Training Program (RTP) Scholarship and a Mater Research Frank Clair Scholarship awarded to P.G., ARC Discovery Early Career Researcher Award (DE150101117), Discovery Project (DP170101198) and Australian Department of Health Medical Frontiers Future Fund (MRFF) (MRF1175457) grants awarded to A.D.E., NHMRC Project Grant (GNT1051117), ARC Future Fellowship (FT180100674) and Sylvia and Charles Viertel Senior Medical Research Fellowship funds awarded to J.M.P., NHMRC Investigator Grants (GNT1178460 to R.L., GNT1173476 to S.R.R., GNT1173711 to G.J.F.), and CSL Centenary Fellowship and NHMRC Project Grant (GNT1106206, GNT1125645, GNT1126393, GNT1138795) funding awarded to G.J.F.. A.D.E., S.R.R. and G.J.F. additionally acknowledge support from the Mater Foundation.

Author contributions

P.G., S.M.L., M.R.L., D.C., F.J.S-L., L.W., A.L.C., C.J., A.S.K., P.E.C., C.M.N. and S.R.R. performed experiments. R.L., J.M.P. and G.J.F. provided resources. A.D.E. and G.J.F. performed bioinformatic analyses. P.G., S.R.R., J.M.P. and G.J.F. conceived the study and designed experiments. P.G., S.R.R. and G.J.F. generated figures. G.J.F. wrote the manuscript and directed the study. All authors commented on the manuscript.

Competing interests

The authors declare no competing interests.

Table 1: *De novo* TE insertions detected and PCR validated in miPSC lines.

Insertion #	Subfamily	Location	Monomers	EN motif	TSD (bp)	PolyA (bp)	Origin
miPSC_1_L1	T_F	1q	6.8	TCTT/AG	16	~125	Reprogramming
miPSC_2_L1	T_F	10q	0	TTCT/GT	14	>100	Mosaic
miPSC_3_L1	T_F	13q	5.8	ATTC/AA	15	~50	Reprogramming
miPSC_4_L1	T_F	3q	5.3	TCTT/AA	13	~54	Reprogramming
miPSC_5_L1	G_F	19q	2	TTAT/AT	14	~50	Reprogramming
miPSC_6_L1	T_F	7q	0	TTTA/AA	17	~51	Reprogramming
miPSC_7_L1	G_F	Xq	5	TCTT/AT	16	>80	Reprogramming
miPSC_8_L1	T_F	19q	3.7	TTTC/AA	19	~24	Reprogramming
miPSC_9_B2	B2	11q	NA	TCTT/AC	16	>60	Reprogramming
miPSC_10_L1	T_F	12q	0	TTTT/GT	6	~36*	Reprogramming
miPSC_11_B2	B2	13q	NA	TTTT/GA	14	>73	Reprogramming
miPSC_12_L1	T_F	13q	0	TCTT/AG	17	~97	Reprogramming
miPSC_13_L1	A	14q	3	TTTC/AT	13	~46	Reprogramming
miPSC_14_B2	B2	15q	NA	TTTT/AC	16	>66	Reprogramming
miPSC_15_L1	G_F	2q	0	TTTC/AA	17	~28*	Reprogramming
miPSC_16_L1	T_F	2q	>3	TTTT/AA	16	>100	Reprogramming
miPSC_17_L1	T_F	3q	>3	ACTT/AA	14	~45	Reprogramming
miPSC_18_B1	B1	3q	NA	TTTT/AA	15	~30	Reprogramming
miPSC_19_L1	T_F	3q	>3	GTTT/AT	15	>80	Reprogramming
miPSC_21_L1	T_F	4q	0	TTTT/CA	17	>150	Reprogramming
miPSC_22_B2	B2	6q	NA	TCTT/GA	15	~52	Reprogramming
miPSC_23_B2	B2	9q	NA	TTTT/AT	16	~50	Mosaic
miPSC_24_B2	B2	Xq	NA	TTTT/AA	15	>100	Reprogramming
miPSC_26_L1	T_F	1q	>3	TCTT/AT	22	~58	Reprogramming
miPSC_27_B2	B2	11q	NA	TTTC/AA	14	>60	Reprogramming
miPSC_28_L1	T_F	13q	3.6	TCCT/AA	15	~93*	Reprogramming
miPSC_29_L1	T_F	15q	0	TCTT/AA	16	>80	Reprogramming
miPSC_30_L1	T_F	6q	>3	TCTT/AT	16	~72	Reprogramming
miPSC_31_L1	T_F	7q	>3	TTTG/AC	15	~43	Reprogramming
miPSC_32_L1	T_F	Xq	2	TCTT/AT	13	~37	Reprogramming
miPSC_33_L1	G_F	Xq	>3	TTTT/AA	15	~47	Reprogramming
miPSC_34_L1	T_F	8q	0	TCTT/AA	6	~36*	Reprogramming
miPSC_35_L1	T_F	1q	0	TTTA/AA	15	~38	Reprogramming
miPSC_36_L1	G_F	8q	0	ATGT/GA	6	~42	Reprogramming
miPSC_37_L1	T_F	1q	1.2	TTTT/GT	14	~20	Reprogramming
miPSC_38_L1	T_F	10q	0	TTCT/AA	15	~55	Reprogramming
miPSC_39_L1	T_F	10q	0	TTTT/AA	8	>140*	Reprogramming
miPSC_40_L1	T_F	11q	>3	TTTT/GA	14	>120	Reprogramming
miPSC_41_L1	T_F	12q	2.6	TCTT/GC	16	~49	Reprogramming
miPSC_42_B1	B1	14q	NA	TTCT/AA	15	>50	Reprogramming
miPSC_43_L1	T_F	16q	>3	ATTT/AA	14	~42*	Mosaic
miPSC_69_L1	T_F	13q	2	CTTT/AT	16	~61	Mosaic
miPSC_87_L1	T_F	17q	0	TTTT/GT	16	~21	Reprogramming

Insertions marked with an asterisk carry a 3′ transduction. miPSC_1_L1 - miPSC_8_L1 were detected in adult primary cell-derived bulk miPSCs by Illumina sequencing (WGS or mRC-seq), miPSC_9_B2 - miPSC_43_L1 we found in MEF-derived single-cell miPSC clones by Illumina sequencing, and miPSC_69_L1 and miPSC_87_L1 were identified in MEF-derived bulk miPSCs by ONT sequencing.

Supplementary Table Legends

Supplementary Table 1: WGS and mRC-seq library statistics.

Supplementary Table 2: Genomic variants private to individual bulk or single-cell miPSC lines, and their predicated impacts. Note: Variants (SNVs, INDELs, SVs) were called using Illumina WGS data only.

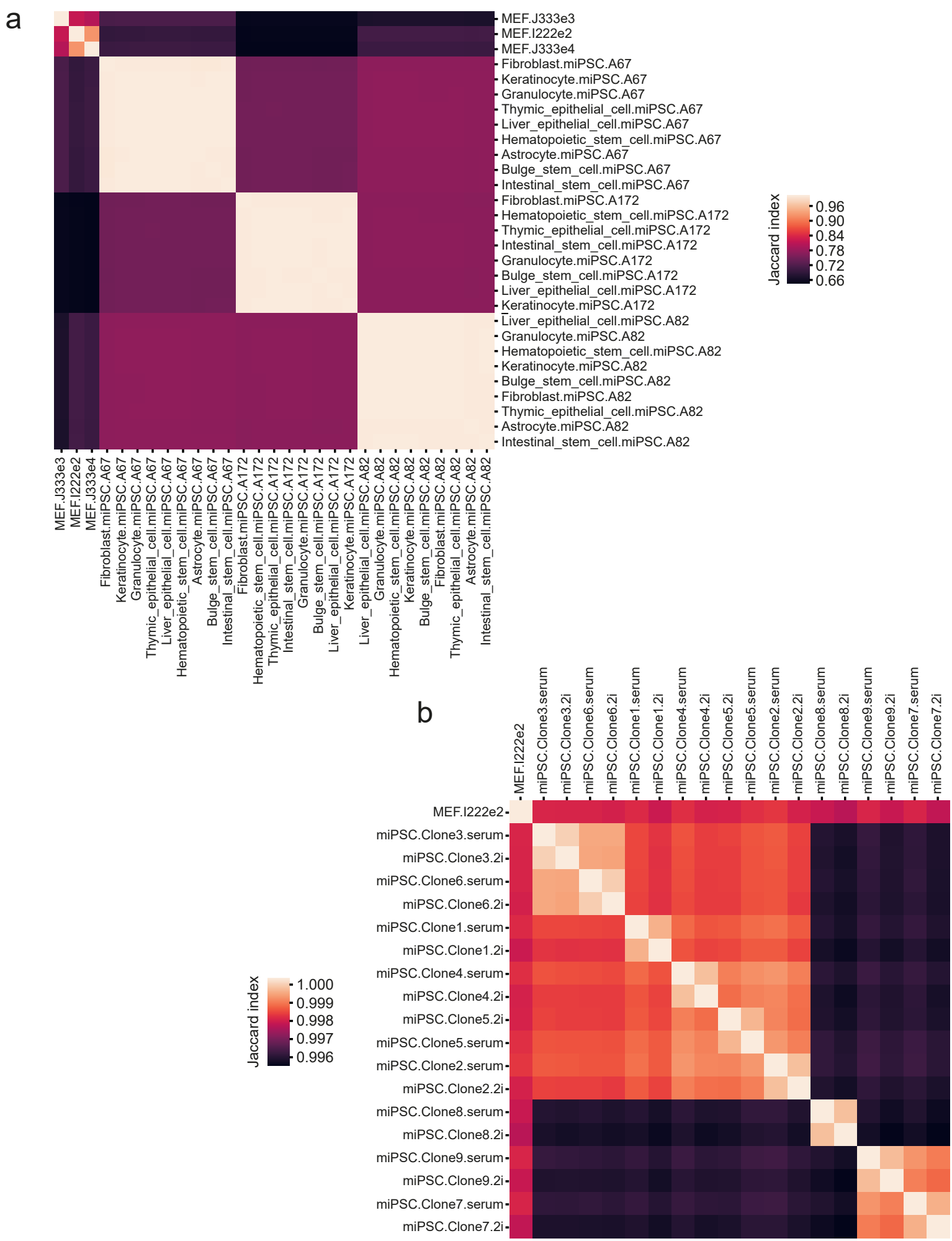
Supplementary Table 3: TE insertion validation details. Four tabs provide information for *de novo* insertions detected via (i) Illumina sequencing of bulk miPSCs reprogrammed from various tissues, (ii) Illumina sequencing of single-cell miPSC clones derived from MEFs, (iii) ONT sequencing of bulk miPSCs derived from MEFs and (iv) ONT sequencing to detect L1-mCherry retrotransposition events in cultured HeLa cells. Illumina data were analyzed with TEBreak, while ONT data were analyzed with TLDR. In tabs (i), (ii) and (iii), Column 1 lists a universally unique identifier (UUID) for each insertion. Columns 2-6 show the insertion name used in other figures and tables, the chromosomal location including genomic 5′ and 3′ end positions and whether the insertion is intergenic or intragenic. For tabs (i) and (ii), columns 7-14 provide the 5′ and 3′ end orientations and whether these provisionally indicate a 5′ inversion, the TE family, the 5′ and 3′ end nucleotide positions of each insertion relative to the TE consensus sequence, and the WGS/mRC-seq read counts for the 5′ and 3′ ends. Column 15 displays the TSD sequence and columns 16 and 17 respectively contain the 5′ and 3′ junction consensus sequences constructed from WGS/mRC-seq reads. Columns 18 and 19 contain the name of the samples where the TE insertion was detected at its 5′ and 3′ end, respectively, and the number of reads present for each sample. For tab (iii), columns 7-17 provide the orientation of the insertion, the TE family and subfamily, the 5′ and 3′ end nucleotide positions of each insertion relative to the TE consensus sequence, the length of the insertion, whether a 5′ inversion is present, the total number of ONT reads supporting the insertion, the total number of spanning reads supporting the insertion, the name of the samples where the TE insertion was detected and the number of reads present for each sample, and whether the TE insertion has previously been observed in genomic analysis of mouse strains. Columns 18-20 display the TSD sequence, the consensus sequencing constructed from ONT reads, and any associated TLDR flags. Columns 21-23 indicate TSD sequences and other

TPRT hallmarks associated with TE insertions obtained by manual inspection of consensus sequences. For tab (iv), Column 1 lists a universally unique identifier (UUID) for each insertion. Columns 2-4 show the chromosomal location including genomic 5′ and 3′ end positions. Columns 5-13 provide the orientation of the insertion, the TE family (all L1-mCherry), the 5′ and 3′ end nucleotide positions of each insertion relative to the spliced L1-mCherry sequence, the length of the insertion, whether a 5′ inversion is present, the total number of ONT reads supporting the insertion, the total number of spanning reads supporting the insertion, and the name of the HeLa colony carrying the TE insertion along with the corresponding number of reads. Columns 14-16 display the TSD sequence, the consensus sequencing constructed from ONT reads, and any associated TLDR flags. Columns 17-19 indicate TSD sequences and other TPRT hallmarks obtained by manual inspection of consensus sequences. Finally, in tabs (i), (ii) and (iii), the last 10 columns comprise the validation PCR type, validation PCR primers and capillary sequencing reads for each insertion, and any relevant notes.

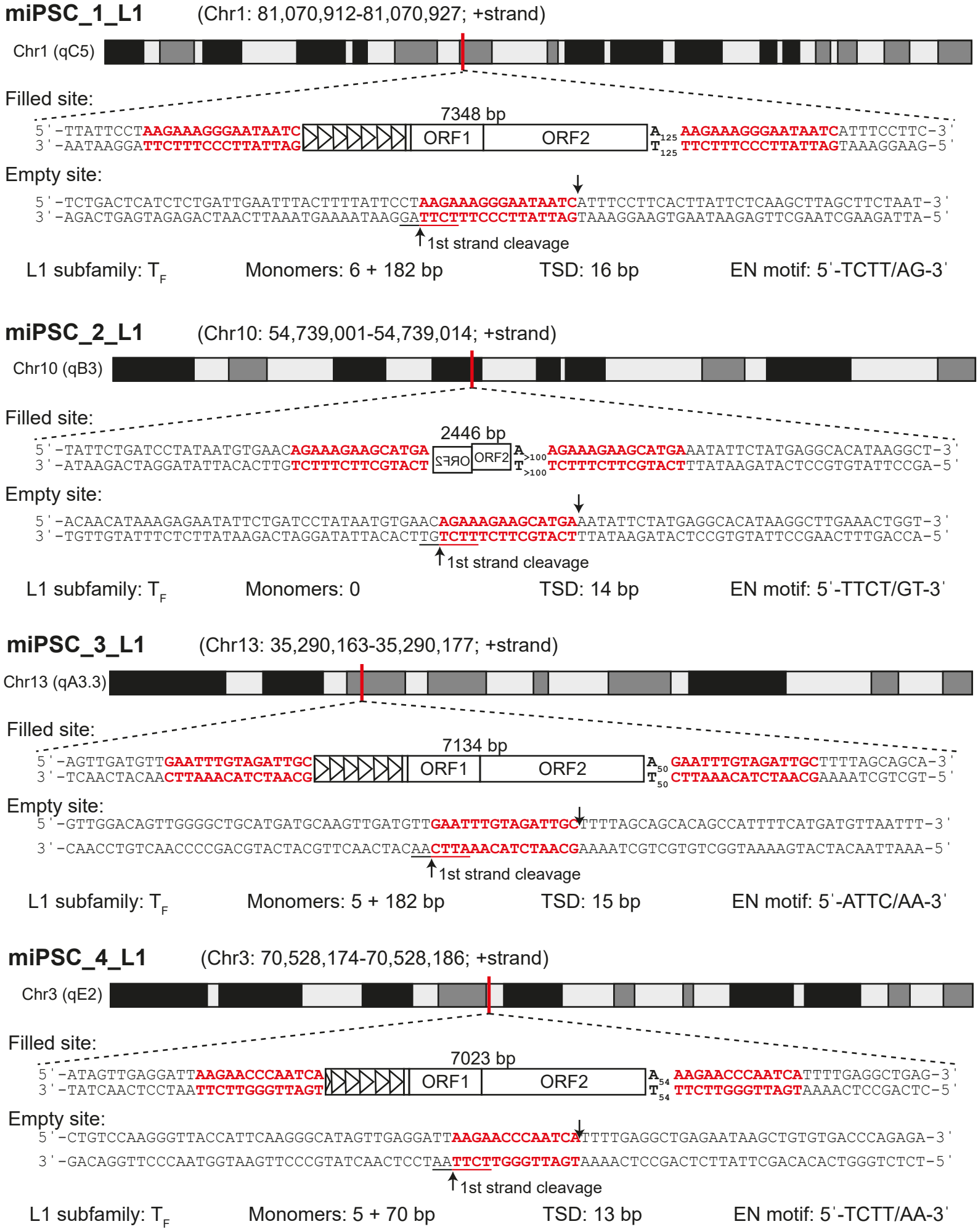
Supplementary Table 4: Non-reference TE insertions detected in MEFs. Two tabs provide information for (i) 277 gold standard heterozygous insertions detected via Illumina sequencing and (ii) insertions detected via ONT sequencing. Each tab is arranged approximately as per **Supplementary Table 3**, except without PCR validation information. All listed insertions were detected in the parental MEFs.

Supplementary Table 5: Reference genome TE methylation values. Tab (i) contains information for TEs and tab (ii) contains information for gene promoters, as defined by the Eukaryotic Promoter Database[119]. Columns are as follows. seg_chrom, seg_start, seg_end: position of the reference TE or gene promoter. seg_name: TE subfamily or gene promoter name. seg_strand: + or - orientation relative to the genome assembly. *_meth_calls: number of methylated CpG calls for each sample. *_unmeth_calls: number of unmethylated CpG calls for each sample. *_no_calls: number of ambiguous CpG calls for each sample (log likelihood ratio between -2.5 and 2.5). *_methfrac: fraction of non-ambiguous calls indicating methylation. *_readcount: number of reads aligned to the element. methfrac_difference: difference between the indicated sample methylation percentages. uncorrected_pvalue: Fisher's exact test p-value from comparison of methylation/demethylation counts between samples. fdr_corrected_pvalue: the

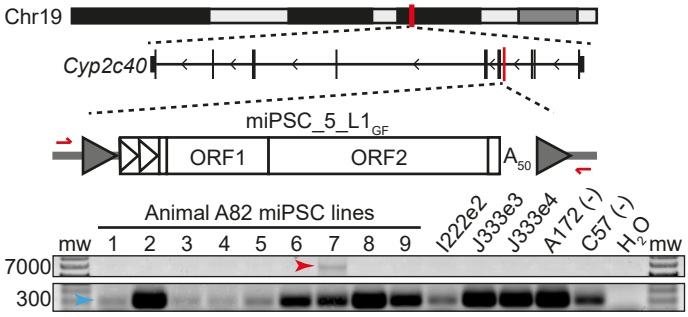
1182 value in “uncorrected_pvalue” Bonferroni corrected for multiple testing. significant: whether the
1183 corrected p-value is less than 0.01 and the absolute methylation difference (ΔmC) is >25%. In tab
1184 (i), L1_TSS_support indicates whether the 5′ end of a spliced EST or mRNA obtained from the
1185 UCSC Genome Browser coincides with the listed L1. Note: full-length L1 methylation values were
1186 calculated for the given 5′UTR coordinates.

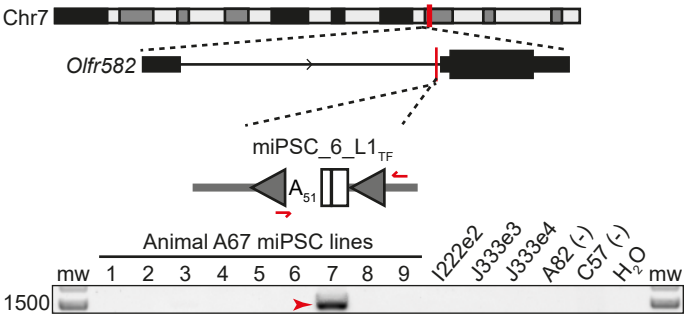


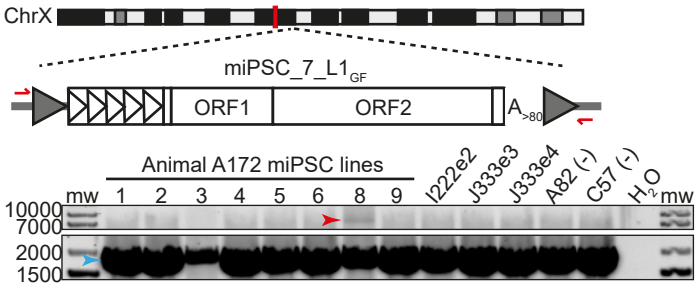
Extended Data Fig. 1: miPSC line genotypic relationships. a, Clustering of miPSC lines derived from 9 primary cell types isolated from 3 animals (A67, A82, A172), and 3 MEF genotypic controls. For each pairwise comparison, the Jaccard index (J) was calculated as the ratio of the union and intersection of SNP/INDEL variants called from WGS data and shared by the sample pair. Known SNPs/INDELs were removed and filtered as described in the Methods. J=1 (light color on key) indicates an identical variant profile between a sample pair, whereas J=0 (dark color on key) indicates no variants in common. Hierarchical clustering was performed using average linkage and a Euclidean distance metric via the seaborn clustermap function. **b,** As for panel (a), except for 9 single-cell clones derived from animal I222e2 MEFs and cultured in serum or 2i conditions.

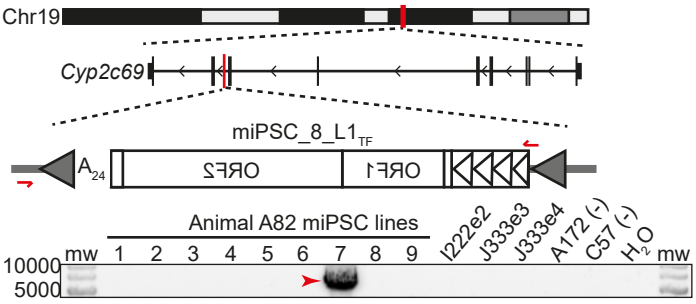


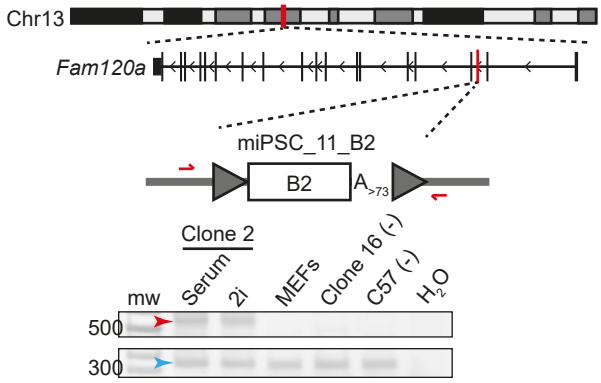
Extended Data Fig. 2: Sequence characteristics of *de novo* L1 insertions detected in bulk tissue-derived miPSCs. For each of four insertions, the following information is provided: the chromosomal location; a filled site illustration indicating target site duplication (TSD) sequences in red, the number of promoter monomers (black triangles) if applicable, and 3′ polyA tract length (A_n/T_n); an empty site illustration depicting TSD sequence and first strand endonuclease (EN) cleavage motif (underlined); summary characteristics (L1 subfamily, number of monomers, TSD length and EN motif).

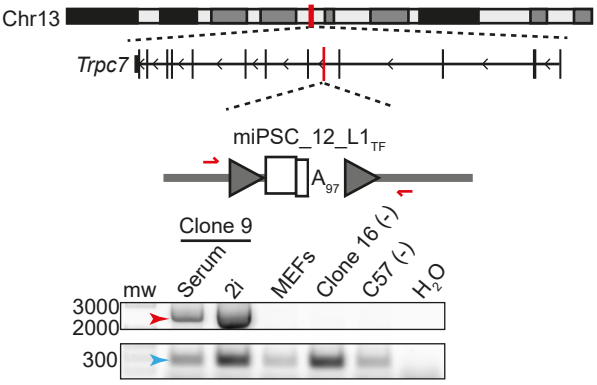

Chr19
Cyp2c40
miPSC_5_L1GF
ORF1
ORF2
A50
Animal A82 miPSC lines
mw 1 2 3 4 5 6 7 8 9
I222e2 J333e3 J333e4 A172 (-) C57 (-) H2O mw
7000
300

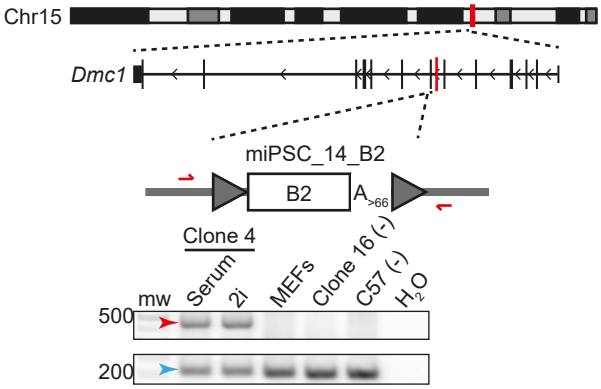

Chr7
Olfr582
miPSC_6_L1TF
A51
Animal A67 miPSC lines
mw 1 2 3 4 5 6 7 8 9
I222e2 J333e3 J333e4 A82 (-) C57 (-) H2O mw
1500

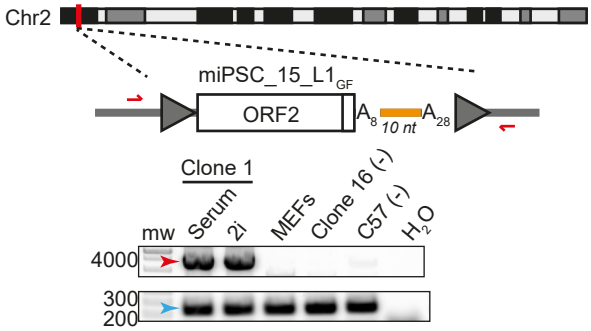

ChrX
miPSC_7_L1GF
ORF1
ORF2
A>80
Animal A172 miPSC lines
mw 1 2 3 4 5 6 8 9
I222e2 J333e3 J333e4 A82 (-) C57 (-) H2O mw
10000
7000
2000
1500

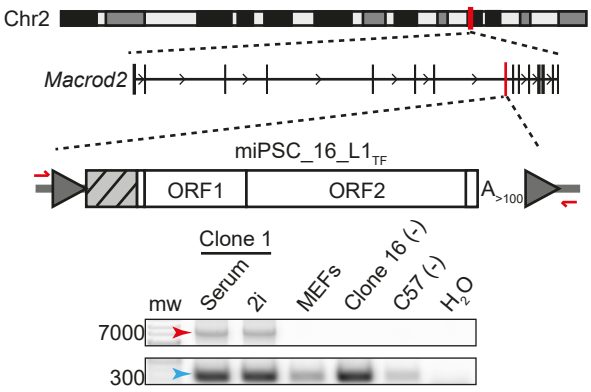

Chr19
Cyp2c69
miPSC_8_L1TF
A24
ORF2
ORF1
Animal A82 miPSC lines
mw 1 2 3 4 5 6 7 8 9
I222e2 J333e3 J333e4 A172 (-) C57 (-) H2O mw
10000
5000

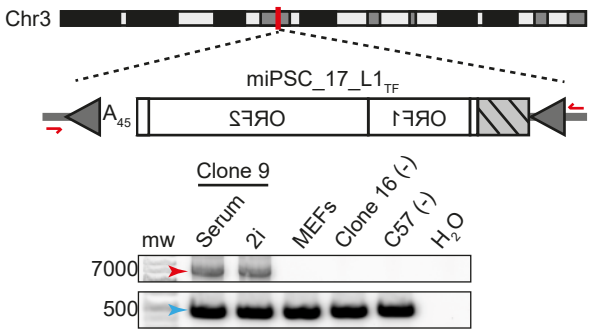

Chr13
Fam120a
miPSC_11_B2
B2
A>73
Clone 2
mw Serum 2i MEFs Clone 16 (-) C57 (-) H2O
500
300

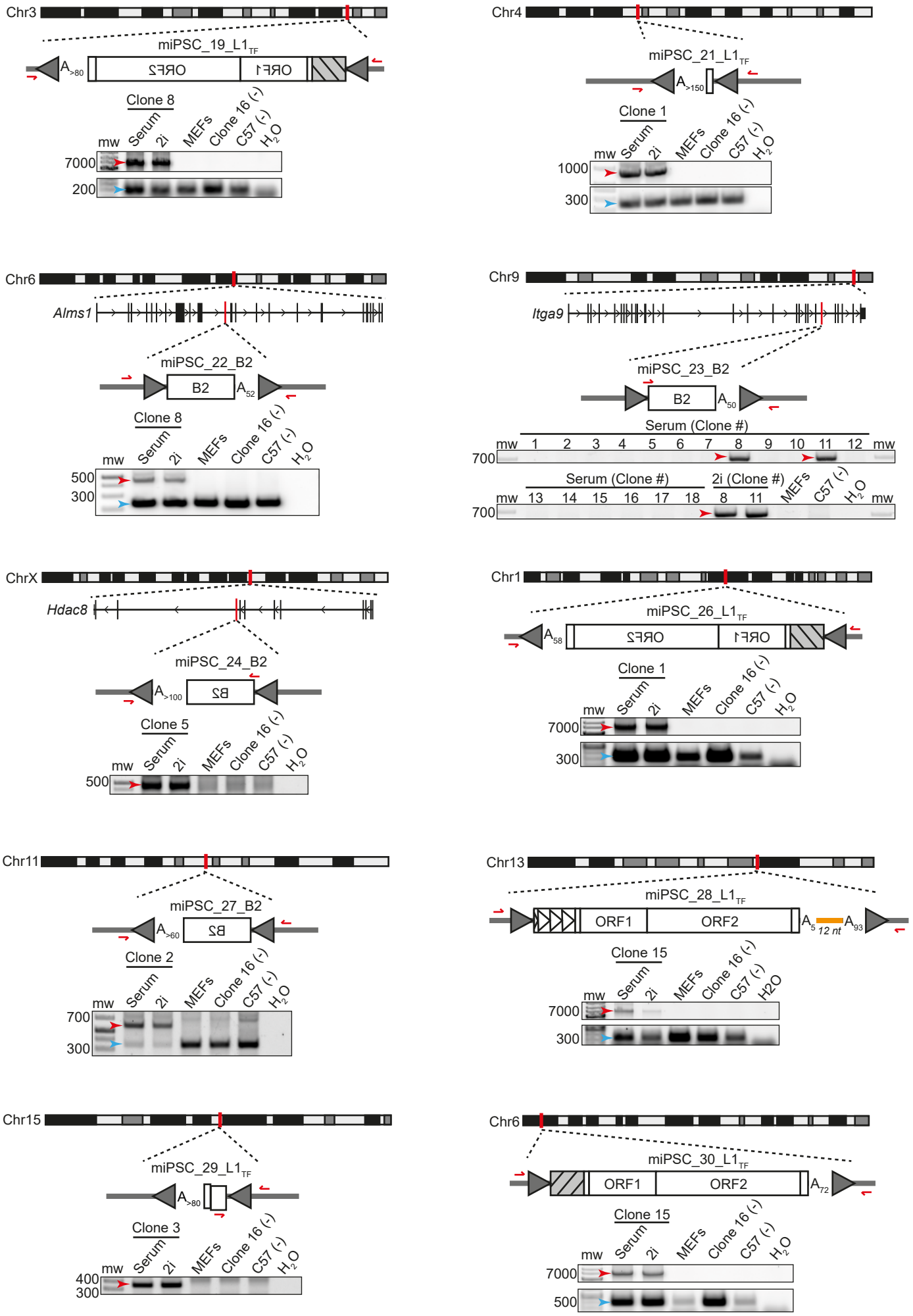

Chr13
Trpc7
miPSC_12_L1TF
A97
Clone 9
mw Serum 2i MEFs Clone 16 (-) C57 (-) H2O
3000
2000
300

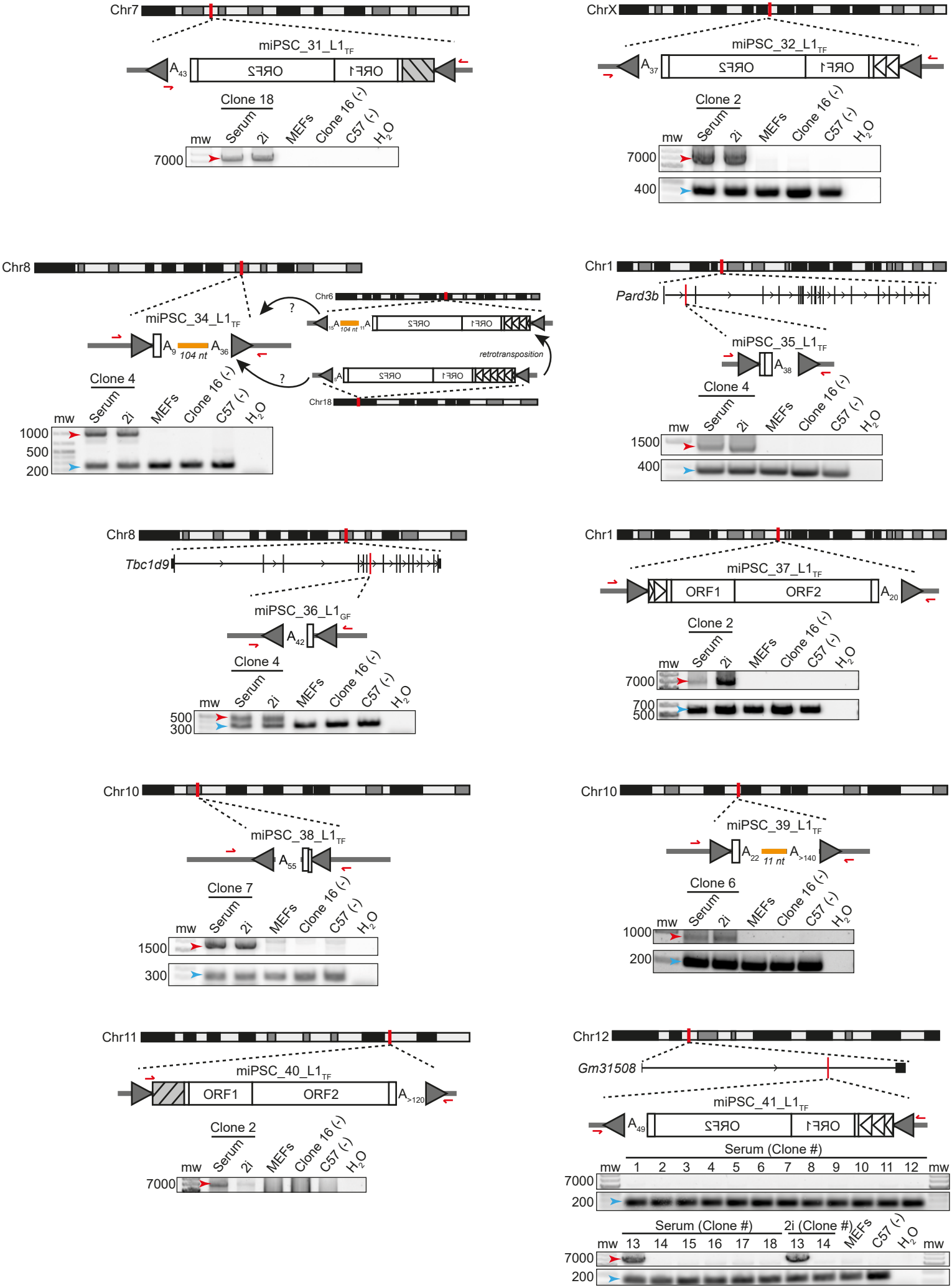

Chr15
Dmc1
miPSC_14_B2
B2
A>66
Clone 4
mw Serum 2i MEFs Clone 16 (-) C57 (-) H2O
500
200

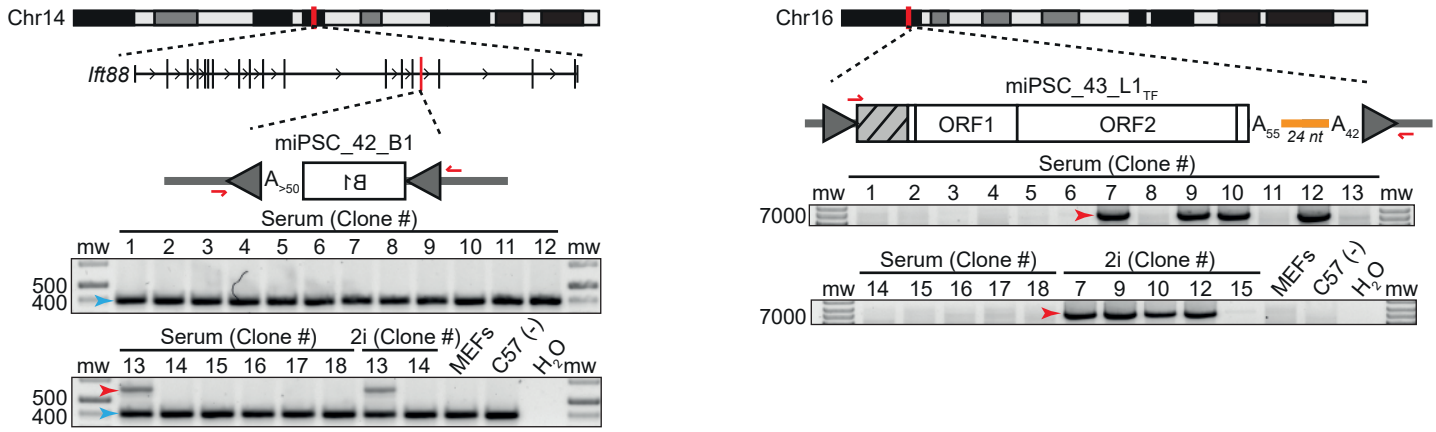

Chr2
miPSC_15_L1GF
ORF2
A8
10 nt
A28
Clone 1
mw Serum 2i MEFs Clone 16 (-) C57 (-) H2O
4000
300
200


Chr2
Macrod2
miPSC_16_L1TF
ORF1
ORF2
A>100
Clone 1
mw Serum 2i MEFs Clone 16 (-) C57 (-) H2O
7000
300

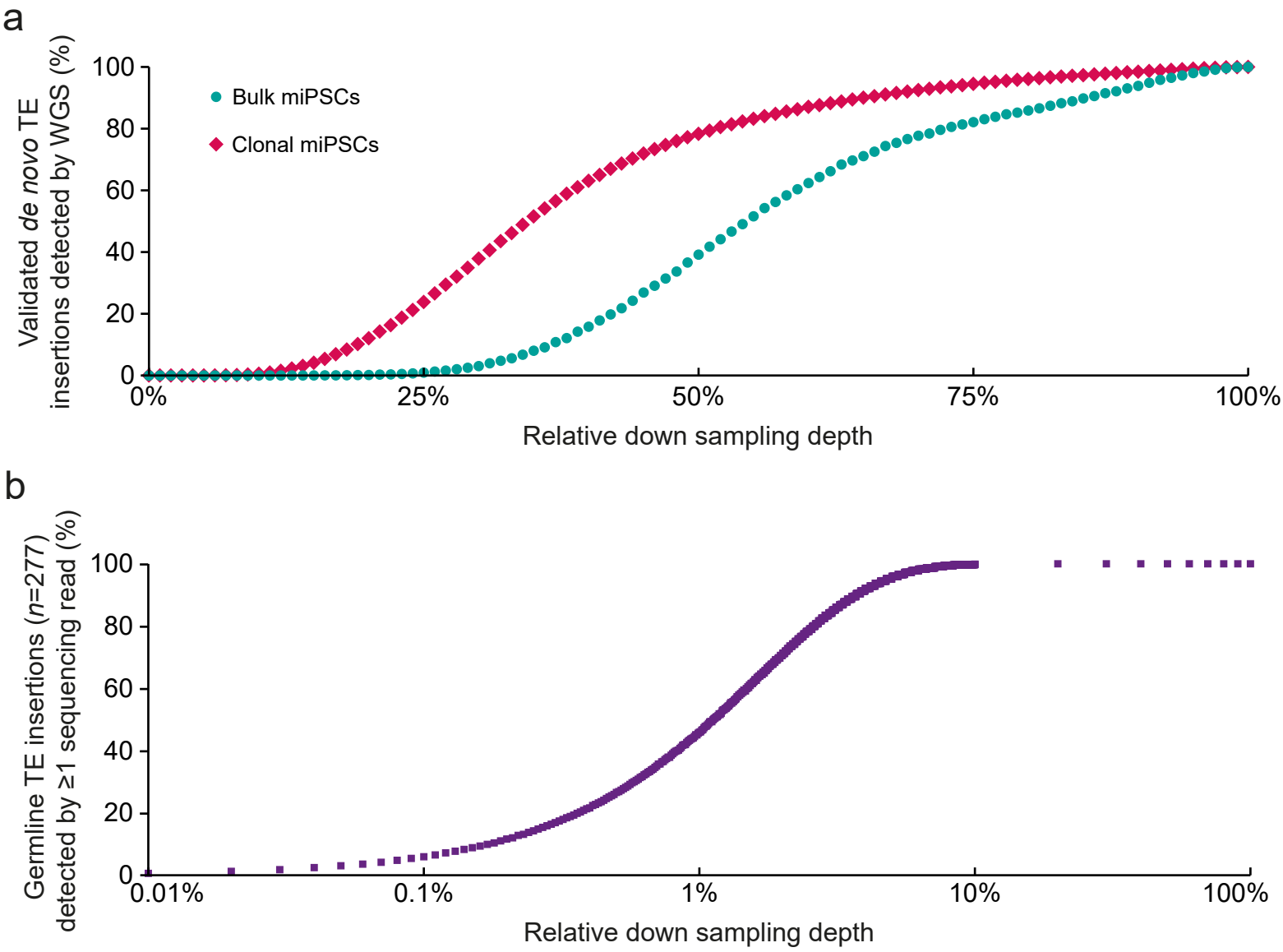

Chr3
miPSC_17_L1TF
A45
ORF2
ORF1
Clone 9
mw Serum 2i MEFs Clone 16 (-) C57 (-) H2O
7000
500


Chr3
miPSC_19_L1TF
A>80
ORF2
ORF1
Clone 8
mw
Serum
2i
MEFs
Clone 16 (-)
C57 (-)
H2O
7000
200
Chr4
miPSC_21_L1TF
A>150
Clone 1
1000
300
Chr6
Alms1
miPSC_22_B2
B2
A52
Clone 8
500
300
Chr9
Itga9
miPSC_23_B2
A50
Serum (Clone #)
1 2 3 4 5 6 7 8 9 10 11 12
700
2i (Clone #)
13 14 15 16 17 18 8 11
ChrX
Hdac8
miPSC_24_B2
A>100
Clone 5
Chr1
miPSC_26_L1TF
A58
Clone 1
Chr11
miPSC_27_B2
A>60
Clone 2
Chr13
miPSC_28_L1TF
ORF1
ORF2
A5
12 nt
A93
Clone 15
H2O
Chr15
miPSC_29_L1TF
A>80
Clone 3
400
300
Chr6
miPSC_30_L1TF
A72
Clone 15
500

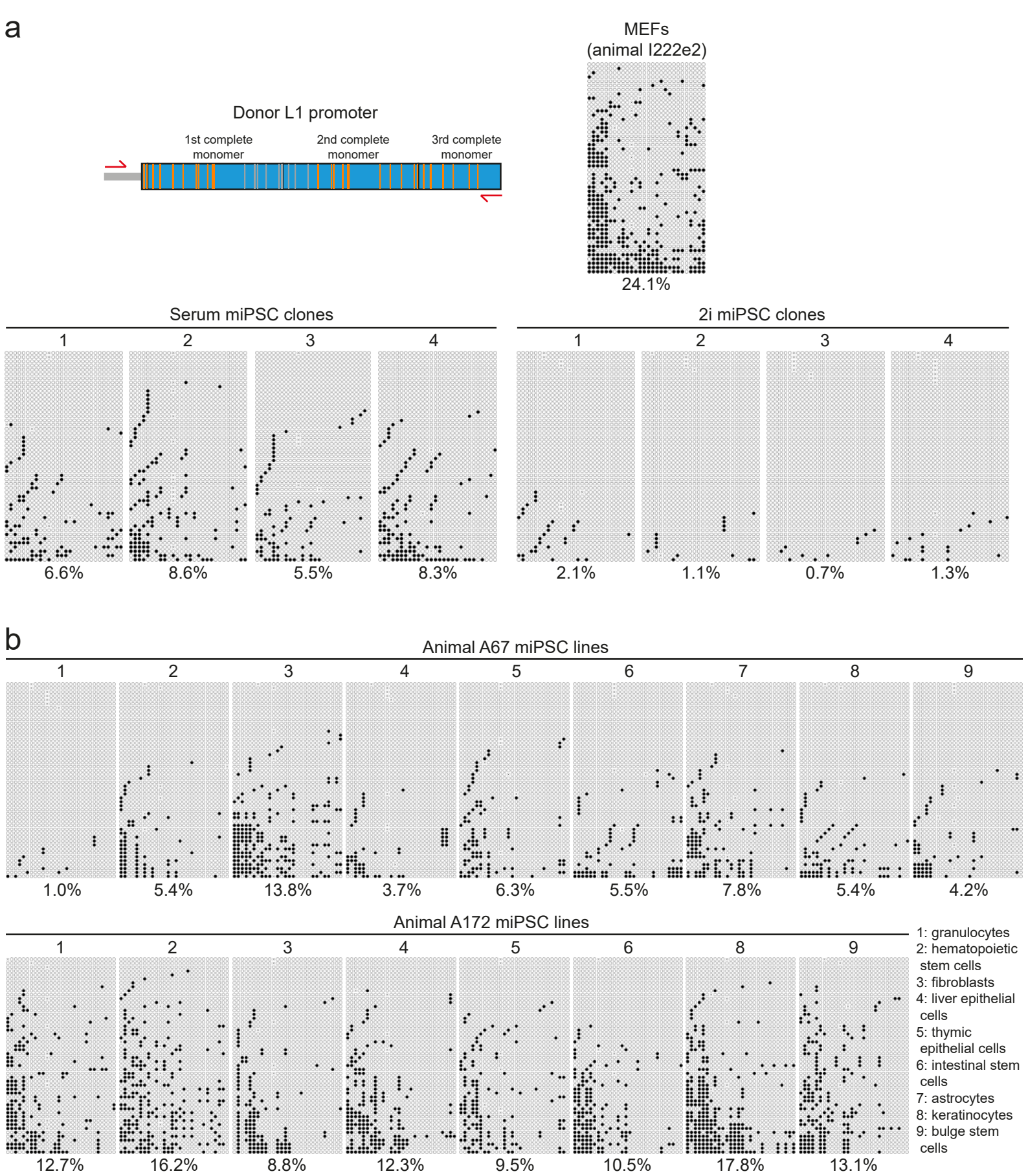

Chr7
miPSC_31_L1TF
A43
ORF2
ORF1
Clone 18
mw
Serum
2i
MEFs
Clone 16 (-)
C57 (-)
H2O
7000
ChrX
miPSC_32_L1TF
A37
Clone 2
7000
400
Chr8
miPSC_34_L1TF
A9
104 nt
A36
Clone 4
1000
500
200
Chr6
Chr18
retrotransposition
Chr1
Pard3b
miPSC_35_L1TF
A38
1500
400
Chr8
Tbc1d9
miPSC_36_L1GF
A42
500
300
Chr1
miPSC_37_L1TF
ORF1
ORF2
A20
700
500
Chr10
miPSC_38_L1TF
A55
Clone 7
1500
300
Chr10
miPSC_39_L1TF
A22
11 nt
A>140
Clone 6
1000
200
Chr11
miPSC_40_L1TF
A>120
Chr12
Gm31508
miPSC_41_L1TF
A49
Serum (Clone #)
1 2 3 4 5 6 7 8 9 10 11 12
2i (Clone #)
13 14 15 16 17 18



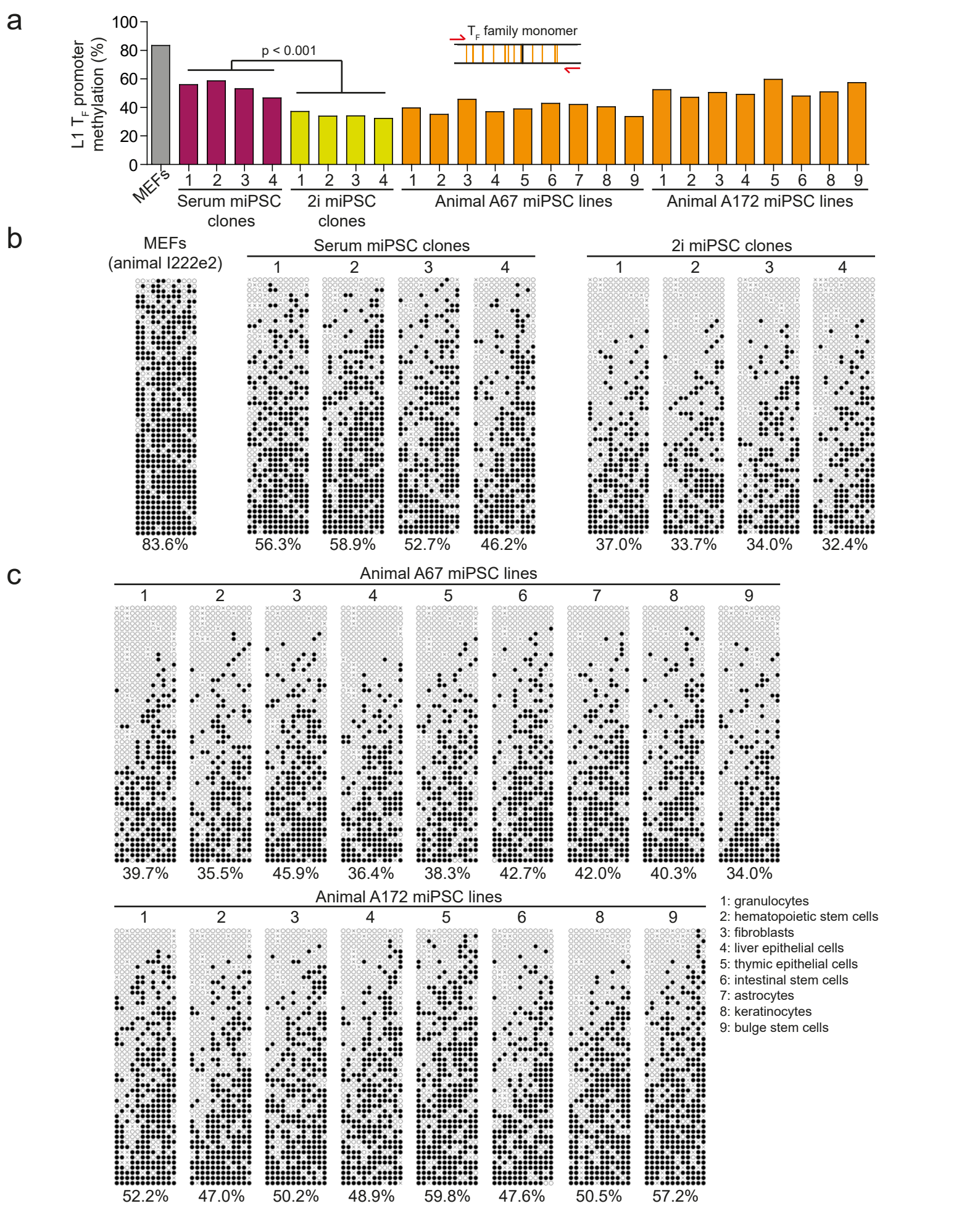
Extended Data Fig. 3: Additional *de novo* TE insertion validation and characterization. *De novo* TE insertions found in 26 bulk miPSC lines generated from primary cells or 18 single-cell miPSC clones derived from MEFs, each subjected to Illumina sequencing. For each insertion, the chromosomal location and orientation are shown. L1 and SINE B1 and B2 insertions are represented by white rectangles. L1 5′UTR promoter monomers, if present, are indicated by triangles or, if the number of monomers is unknown, a gray box with black stripes. Poly(A) tracts and their length are indicated (A_n), and target site duplications (TSDs) are depicted as gray arrows. 3′ transductions are shown as orange lines. PCR validation primers are shown as red arrows. PCR products in agarose gels used to confirm TE insertions are indicated by red arrows. Empty site (wild-type) amplicons are indicated by blue arrows, where applicable.



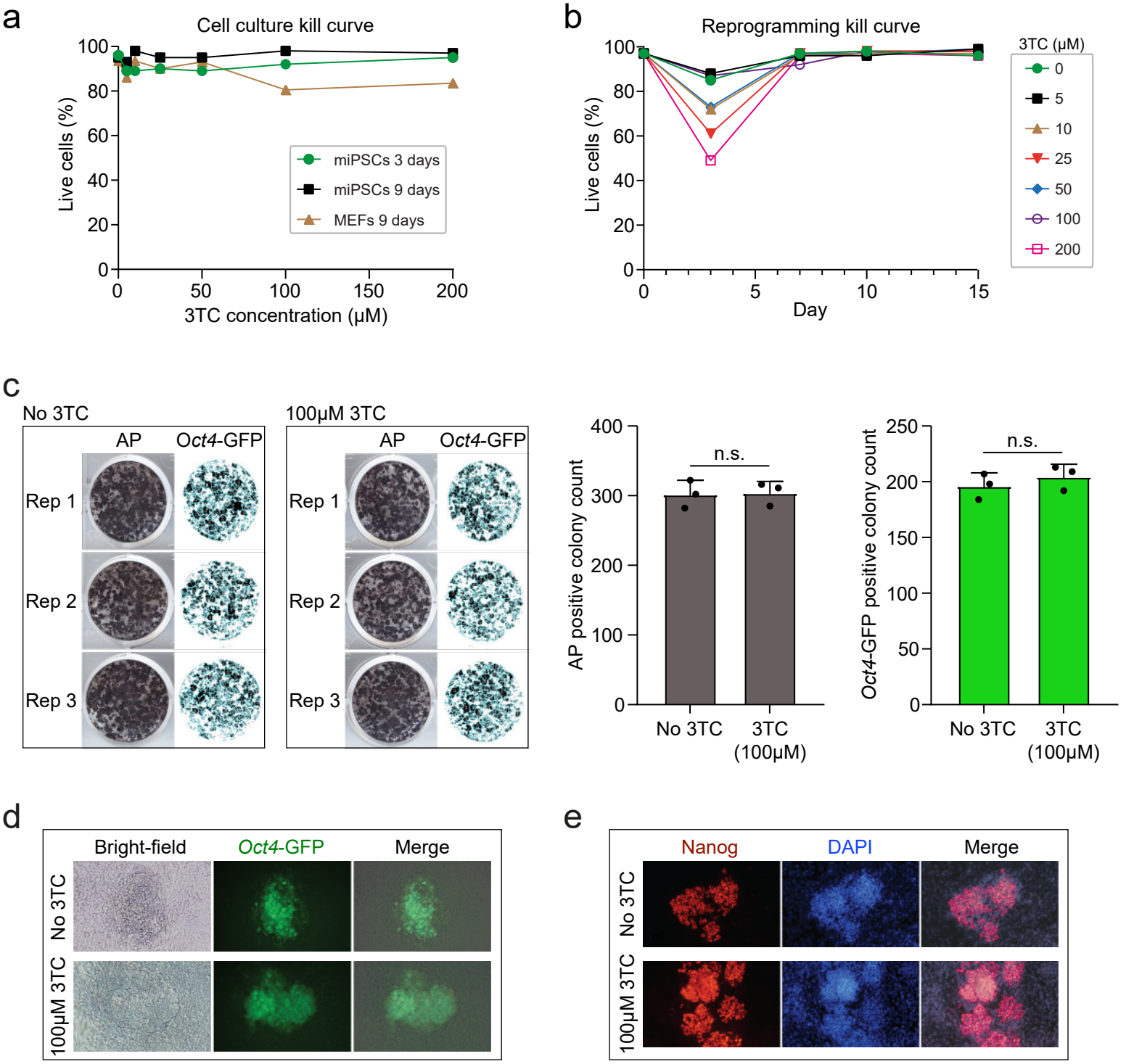
Extended Data Fig. 4: TE detection sensitivities at simulated sequencing depths. a, To assess whether PCR validated *de novo* TE insertions would have been initially overlooked by lower coverage WGS, we down sampled our ~41× average depth WGS in percentile increments. In order to be called as present, *de novo* insertions found in the bulk (*top*) and single-cell (*bottom*) miPSC experiments required ≥1 WGS read at each of their 5′ and 3′ junctions, and ≥10 WGS reads in total. **b,** To estimate the likelihood of a mosaic TE insertion being overlooked in the parental animal I222e2 MEF population, and called as *de novo* in one of the associated clonal miPSC lines, we defined a set of 277 heterozygous germline TE insertions found in I222e2 and that were detected by ≥25 WGS or mRC-seq reads at each of their 5′ and 3′ junctions. We then simulated the probability of at least one read being found for an insertion when the reads assigned to that insertion were assigned probabilities to achieve random sampling depths ranging from 0.01% to 100% of the parental MEF bulk sequencing data. Note: at each depth in panels (a) and (b), simulations were repeated 10,000 times.



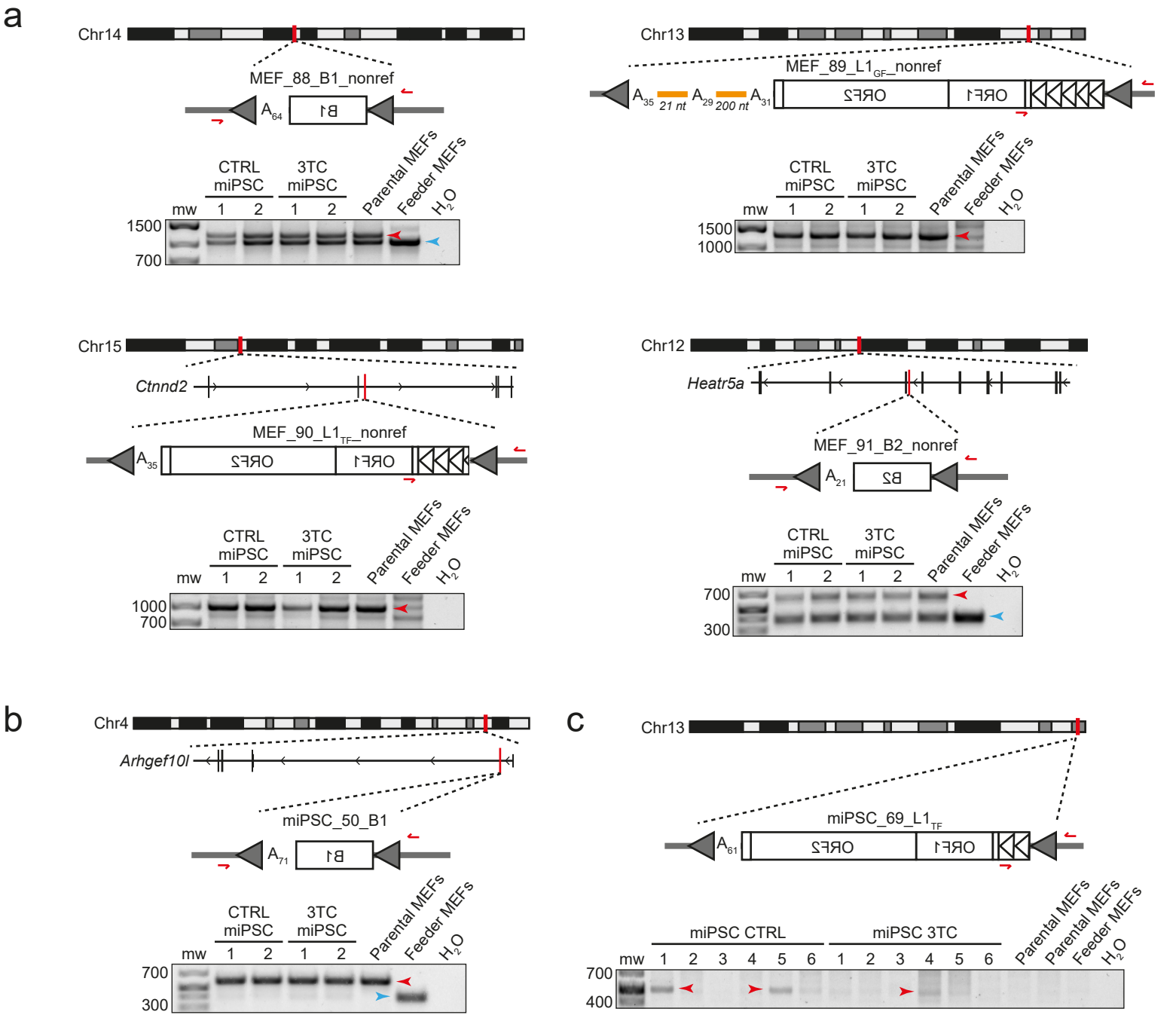
Extended Data Fig. 5. Donor L1 hypomethylation in MEFs and miPSCs. a, *top left*: Locus-specific methylation analysis design for a donor L1 found to generate insertion miPSC_10_L1 in a MEF-derived single-cell miPSC clone (Clone 1). CpGs located in the first 3 monomers of the donor L1 were assessed. Orange and gray strokes indicate CpGs covered and not covered, respectively, by sequencing the amplicon with 2×300mer Illumina reads. *bottom right*: Methylation of the donor L1 promoter sequence in four single-cell miPSC clones, including Clone 1, cultured in either serum or 2i conditions, and the parental MEF population. Each cartoon panel corresponds to an amplicon and displays 50 non-identical randomly selected sequences (black circle, methylated CpG; white circle, unmethylated CpG; ×, mutated CpG). The percentage of methylated CpG is indicated underneath each cartoon. **b,** Donor L1 methylation data as per (a) except for bulk miPSC lines derived from two animals (A67 and A172) carrying the polymorphic donor L1.



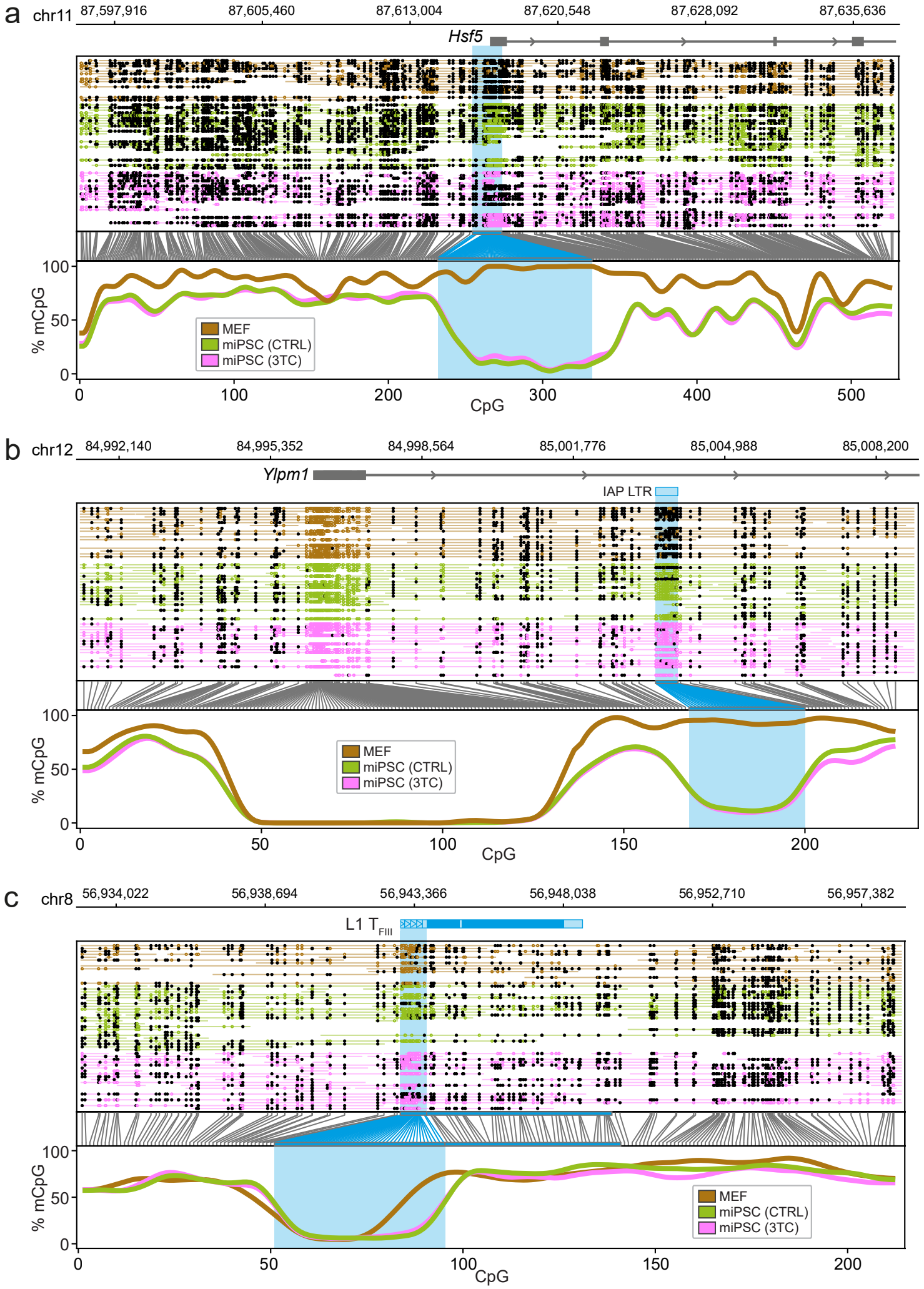
Extended Data Fig. 6. L1 T_F subfamily promoter monomer methylation. a, L1 T_F monomer CpG methylation in MEFs, single-cell miPSC clones, and bulk miPSCs derived from primary cells. *top*: Assay design and primer locations with respect to L1 T_F monomer structure. Orange strokes indicate CpGs covered by the assay. *bottom*: Histogram data represent the mean percentage methylation of 50 non-identical bisulfite converted sequences selected at random from each sample. A two-tailed t test ($p<0.001$) was used to compare serum and 2i culture conditions for single-cell miPSC clones 1-4. **b,** L1 T_F methylation in four single-cell miPSC clones and parental MEFs. Each cartoon panel corresponds to an amplicon and displays 50 non-identical randomly selected sequences (black circle, methylated CpG; white circle, unmethylated CpG; ×, mutated CpG). Methylated CpG percentage is indicated underneath each cartoon. **c,** As per (b) except for bulk miPSCs derived from animals A67 and A172. Note: this assay surveys CpG methylation for T_F monomers genome-wide without retaining their position within individual L1 loci.



Extended Data Fig. 7: Lamivudine does not impact reprogramming efficiency or miPSC survival. a, Cultured MEF and miPSC viability as a function of lamivudine (3TC) concentration. miPSCs were tested for 3 and 9 days in culture with 3TC, and MEFs tested for 9 days. **b,** Cell viability during MEF reprogramming to miPSCs in the presence of varying concentrations of 3TC, as a function of days since reprogramming was induced by the addition of doxycycline. **c,** Reprogrammed MEFs stained for alkaline phosphatase (AP) and *Oct4*-GFP pluripotency markers 15 days after the addition of doxycycline, in media containing 100μM 3TC or no 3TC. Results are shown as microscopy images of stained cultured cells (left) and as histograms (right) representing the mean observed AP positive or *Oct4*-GFP positive colony counts ± SD. Differences were not significant (two-tailed t test). **d,** Representative microscopy images of day 15 miPSC colonies obtained when MEFs were reprogrammed in the presence of 100μM 3TC or no 3TC. **e,** Representative immunostaining of Nanog (red), an additional pluripotency marker, in day 15 miPSCs.



Extended Data Fig. 8: Supporting data for ONT sequencing analysis of miPSCs. a, Non-reference polymorphic TE insertions found by ONT sequencing, used as positive controls for PCR validation experimental designs. **b,** A putative *de novo* B1 insertion detected in one miPSC line by ONT sequencing and annotated as a false positive based on PCR amplification in the parental MEF template DNA. **c,** A putative *de novo* L1 T_F insertion detected in one miPSC line by ONT sequencing and amplified in multiple miPSC lines by PCR. Here, DNA was obtained from two parental MEF aliquots, one corresponding to miPSC CTRL/3TC lines 1-3 and one to miPSC CTRL/3TC lines 4-6. Note: in each panel, the insertion chromosomal location and orientation are shown. L1 and SINE B1 and B2 insertions are represented by white rectangles. L1 5′UTR promoter monomers are indicated by triangles. PolyA tracts and their length are indicated (A_n), and target site duplications (TSDs) are depicted as gray arrows. 3′ transductions are shown as orange lines. PCR validation primers are shown as red arrows. PCR products in agarose gels used to confirm TE insertions are indicated by red arrows. Empty site (wild-type) amplicons are indicated by blue arrows, where applicable.



Extended Data Fig. 9: Examples of protein-coding gene and TE methylation, as surveyed by ONT sequencing. Methylation profiles are shown for **a,** the *Hsf5* gene promoter **b,** an IAP LTR intronic to *Ylpm1*, and **c,** an intergenic L1 T_F. For each example, the panels are arranged as per **Fig. 4c.** ONT data are shown for MEFs (brown) and for control (green) and 3TC-treated (pink) miPSC datasets. miPSC replicates were aggregated for each condition and down-sampled to approximate MEF dataset read depth.