Tropical hummingbird pollination networks are resistant to short-term experimental

removal of a common flowering plant

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ABSTRACT

Theory predicts that the structure of plant-pollination networks should withstand disturbance, but experiments testing this prediction remain uncommon. In this study, we simulated the local extinction of a hummingbird-pollinated understory plant, *Heliconia tortuosa,* from tropical forest fragments using a replicated Before-After-Control-Impact design while quantifying hummingbird abundance and space use (383 hummingbird captures and 72 radio-tagged individuals), floral visitation rates (>19,000 observation hours), and pollination success (529 flowers). We expected that *H. tortuosa* removal would either result in (i) network collapse, in which hummingbirds vacate fragments and compromise the reproductive success of other flowering plants, or (ii) increased hummingbird reliance on alternative resources (rewiring), leading to sustained fragment use. In our experiment, hummingbird behavior and pollination were remarkably resistant to loss of *H. tortuosa*, a locally common plant species representing 30-40% of the available nectar resources on average. The exact mechanisms enabling short-term hummingbird persistence after resource removal remain unclear, as we did not discover evidence of rewiring. We hypothesize that physiological adaptations (e.g., torpor and insectivory) may have allowed hummingbird persistence, perhaps alongside high movement ability. With the important caution that short-term experiments may not emulate natural extinction processes, our study provides support for predictions that pollination networks may be robust to plant species loss.

Key words: mutualistic networks, ecosystem function, foraging ecology, Trochilidae, exclusion experiment

INTRODUCTION

Species loss can have cascading consequences for the broader community, including coextinctions of dependent species and declines in ecosystem function (Koh *et al.* 2004, Colwell *et al.* 2012). Linked extinctions may be especially likely for mutualistic interactions, such as those between plants and pollinators (Dunn *et al.* 2009, Aslan *et al.* 2013). For instance, plants that are not adequately pollinated produce fewer seeds, accruing extinction debt that becomes realized when the existing generation cannot be replaced by new recruits (Ashman *et al.* 2004, Vellend *et al.* 2006, Kuussaari *et al.* 2009). This repeated reproductive failure will eventually cause demographic collapse of plant populations (Anderson *et al.* 2011, Phillips *et al.* 2015), which could then hasten the decline of animals that consume pollen or nectar (Biesmeijer *et al.* 2006, Pauw and Hawkins 2011, Weiner *et al.* 2014, Scheper *et al.* 2014). Indeed, plants and pollinators are experiencing declines worldwide (Biesmeijer *et al.* 2006, Potts *et al.* 2010, Regan *et al.* 2015), which not only imperils the essential ecosystem services on which humans depend but also threatens the maintenance of global biodiversity (Allen-Wardell *et al.* 1998, Klein *et al.* 2007, Ollerton *et al.* 2011).

However, parallel declines of pollinators and plants are at odds with theoretical predictions suggesting that species loss should generally not result in coextinction of interaction partners (Memmott *et al.* 2004, Bascompte *et al.* 2006). Tolerance to subsequent extinctions (i.e., robustness) primarily arises from the expectation that interaction networks exhibit strongly nested topologies, such that specialists interact with subsets of partners linked to generalists (Bascompte *et al.* 2003). Under this interaction pattern, connections between specialists are uncommon, so the functional role of a specialist can be absorbed by its more abundant generalist partners (Bascompte *et al.* 2003, Bascompte and Jordano 2007). In addition to the redundancy

afforded by a nested structure, models suggest that flexibility in pollinator foraging behavior –

i.e., rewiring to use alternative resources – may buffer networks against disturbance (Kaiser-

Bunbury *et al.* 2010, Ramos-Jiliberto *et al.* 2012, Valdovinos *et al.* 2013, Timóteo *et al.* 2016).

Despite these overall predictions and mechanisms of robustness, simulation studies have

revealed that removal of highly connected, generalist species thwarts a network's capacity to

buffer perturbations, leading to rapid partner extinction (Memmott *et al.* 2004, Kaiser-Bunbury

et al. 2010, Traveset *et al.* 2017). Thus, if external stressors such as habitat fragmentation

threaten highly connected species ('topological keystones' sensu Jordán 2009), an extinction

cascade could occur – particularly without rewiring by pollinators. Uncertainty about the

robustness of real-world plant-pollinator networks still exists, however, because relatively few

studies have quantified pollinator responses to deletions of other species (Appendix S1: Table

S1).

In this study, we tested whether declines of a hummingbird-pollinated plant *(Heliconia*

tortuosa) could have cascading consequences for communities of hummingbirds and flowering

plants within a fragmented tropical forest landscape – or whether rewiring might forestall

network collapse. In Neotropical forests, the genus *Heliconia* (Heliconiaceae) comprises

understory herbs that have been suggested to function as "keystone mutualists" (Gilbert 1980).

Large, brightly colored *Heliconia* inflorescences not only provide nectar to large-bodied hermit

hummingbirds (Trochilidae: Phaethornithinae) throughout the year, but also support a variety of

other hummingbird species during periods of resource scarcity (Stiles 1975, 1985). Because most

hummingbird species visit multiple plant species (Rodríguez-Flores *et al.* 2019), loss of *H.*

tortuosa and its associated hummingbird partners could jeopardize the reproduction of co-

occurring plants, thus generating an extinction cascade (Gilbert 1980). Although *H. tortuosa* is

locally common in the mid-elevation, premontane tropical forests of Costa Rica, Hadley *et al.*

(2014) found that *H. tortuosa* in smaller forest fragments produced fewer seeds, possibly due to

short-distance pollen transfer associated with pollinator movement limitation.

To assess the potential consequences of *H. tortuosa* declines, we conducted one of most

highly replicated, large-scale removal experiments conducted within the context of plant-

pollinator interaction networks (Appendix S1: Table S1). We measured hummingbird space use,

floral visitation rates, and plant pollination success before and after temporary experimental

removal of *H. tortuosa* within forest fragments. We developed two alternative hypotheses about

how hummingbirds would respond to declines of *H. tortuosa* (hereafter *Heliconia*) and

potentially affect the pollination of co-occuring plant species (Table 1). Under the *robust*

network hypothesis (**H1**), hummingbirds do not permanently emigrate from forest fragments

following *Heliconia* decline, leading to uninterrupted visitation to remaining plants. We

hypothesized that hummingbirds compensate for *Heliconia* extinction by increasing their use of

non-*Heliconia* floral resources (rewiring). Alternatively, hummingbirds may be highly dependent

on *H. tortuosa* and inflexible in their foraging choices; therefore, local *Heliconia* declines should

reduce nectar availability such that hummingbirds cannot afford the energetic costs to remain in

(or travel to) focal forest fragments, leading to decreases in fragment use and pollination success

(**H2***: parallel extinction hypothesis*). Under both hypotheses, we expected hummingbirds to

change their behavior due to their exceptionally rapid metabolisms, high dependency on nectar,

and limited fat stores (Suarez 1992, Powers *et al.* 2003). Lastly, we expected that changes in

Heliconia density would affect green hermits (*Phaethornis guy*) and violet sabrewings

(*Campylopterus hemileucurus*) more strongly than other hummingbird species (**H3**). These two

species (hereafter '*Heliconia* specialists') have bills that are morphologically specialized for

long, curved flowers and are *Heliconia*'s primary pollinators (Stiles 1979, Wolf and Stiles 1989,

Taylor and White 2007, Betts *et al.* 2015). Both species inhabit the forest understory and behave

as high-reward trapliners (sensu Feinsinger and Colwell 1978), visiting spatially dispersed,

nectar-rich floral resources – such as *Heliconia* – in a routine sequence (Borgella *et al.* 2001).

MATERIAL AND METHODS

Study system

We conducted this study within a ∼122 km^2 area surrounding the Las Cruces Biological

Station in Costa Rica (Coto Brus Canton, Puntarenas Province, 8°47′7′′ N, 82°57′32′′ W, Fig.

1: A-B). This study area comprises cattle pastures, coffee plantations, and fragments of tropical

premontane wet forest created during a period of extensive land clearing concentrated between

1960 and 1980 (Zahawi *et al.* 2015). Remnant forest fragments (~30% of the study area) span a

wide range of sizes (<1 to >1000 ha), with otherwise isolated fragments often connected by

narrow riparian corridors and living fences (Hadley and Betts 2009, Zahawi *et al.* 2015). Mean

annual temperature at the Las Cruces Biological Station is ~21 ℃, mean annual rainfall is 3500-

4000 mm, and a dry season occurs from December to March (Borgella *et al.* 2001, Zahawi *et al.*

2015).

Based on visual examination of satellite imagery and previous work in this study area

(Hadley *et al.* 2014, 2018, Kormann *et al.* 2016), we selected 14 focal fragments ('sites', Fig.

1A). Sites were then included based on presence of flowering *Heliconia***,** landowner permission,

vehicle accessibility, and navigable terrain. To ensure site accessibility, we conducted our study

during the dry season and beginning of the rainy season (February – May), because localized

flooding can occur during periods of heavy rainfall. Elevation of sites ranged from ~900-1500 m

(Appendix S2: Table S1).

Experimental design

Site details and experimental timeline

This study used a replicated BACI (Before-After-Control-Impact) design. A BACI design

requires two paired sites: an Impact site (here, 'Treatment') that experiences a disturbance and a

Control site that does not. Additionally, each site is monitored concurrently throughout time

(here, 'Pre' and 'Post' disturbance) (Fig. 2). Therefore, the comparison of interest is the pre-to-

post change in the treatment site relative to the pre-to-post change in the control site. This design

separates treatment effects from natural processes occurring in the study system, either across

space (control *versus* treatment) or time (pre *versus* post).

Here, we paired control and treatment sites based on physical characteristics (elevation

and connectivity to additional forest: Appendix S2: Table S1) and ensured that paired sites were

at least 1000 m apart. This distance threshold corresponds to the estimated daily movement

distance of traplining hummingbirds in this study system (Volpe *et al.* 2014), making it unlikely

that hummingbirds from the treatment site would relocate to the control site. Actual distances

between sites ranged from 1917-5637 m (mean ± SD: 4256 ± 1528 m, median: 5086 m), and we

never detected hummingbird movement between paired sites — both in terms of hummingbird

captures and radio telemetry (see *'Response variables'*, below). Using the 14 sites, we repeated

the *Heliconia* removal experiment 16 times across three years (2016-2018), yielding a maximum

sample size of 16 control replicates and 16 treatment replicates (Appendix S2: Table S2). Thus,

most sites were used multiple times across the three-year study period, but only once per year

(Appendix S2; Table S3). Sites used multiple times received the opposite treatment of the

previous year, except for 3 of 16 instances (when covering *Heliconia* in that site again would

have posed substantial logistical challenges). Initial treatment assignments were random, which

reduces the likelihood that any treatment differences would be due to factors associated with

individual sites.

Each experimental replicate lasted for 9 days. Starting around noon on the fourth day, we

removed *Heliconia* from the treatment site and designated the following 36 hours as a behavioral

adjustment period during which no data were collected. Although this adjustment period was

necessarily relatively short due to the limited battery lifespan of hummingbird radio-transmitters

(see *'Radio telemetry'*, below), we expected hummingbirds to respond rapidly due to their

exceptionally high metabolisms and limited fat stores (Suarez 1992, Powers *et al.* 2003).

Heliconia removal (hummingbird exclusion)

Across all study sites and years, we removed between 8 and 526 *Heliconia* inflorescences

per treatment replicate (mean $\pm$ SD: 172 $\pm$ 138, median: 138, Appendix S2: Table 4) by covering

Heliconia inflorescences (mature and immature) with dark-colored fabric bags (Fig. 2A). This

approach prevented hummingbird access and obscured flowers from view while avoiding long-

term changes to floral resources associated with destructive sampling methods (e.g., cutting).

The covered inflorescences represented, on average, 97% ($\pm$ 5% SD) of the *Heliconia*

inflorescences encountered during removal (range: 80-99%); two focal *Heliconia* plants/site

remained uncovered for pre-to-post monitoring, and sometimes plant inaccessibility limited our

covering capacity. While covering *Heliconia*, we also collected data on the abundance and

location of resources available to hummingbirds (*Heliconia* and non-*Heliconia*). Resource data

were used to estimate the percentage of calories removed by our manipulation (see Appendix S3

for methods) and delineate 'focal areas' (see below). In 2017-2018, we also collected resource

data in the control sites, mimicked the trampling effect experienced by the treatment sites.

Focal areas

Within each site, we focused our observations within designated 'focal areas'. Focal areas

encompassed locations where we captured hummingbirds, installed trail cameras, and set up

plant arrays (Fig. 1C, see *'Response variables'*). We delineated focal areas as the 100%

minimum convex polygons of the plant GPS points. Treatment focal areas ranged from 0.27 to

7.49 ha (mean ± SD: 1.7 ± 1.9, median: 1.25, Appendix S2: Table S4), depending on the size of

the forest fragment and the navigability of the terrain. We acknowledge that our focal areas were

smaller, on average, than hummingbird home ranges in this study system (e.g., mean home range

size of 2.2 ha for green hermits: Volpe *et al.* 2016). However, here we were interested in any

behavioral changes occurring *within* the focal area, following similar removal studies (e.g., 20 *x*

20-m plots within the home range of a bumblebee: Brosi & Briggs 2013).

Response variables

Hummingbird captures

Within each focal area, we captured hummingbirds at the beginning and end of the

experimental period (Days 1 and 9). During each capture session, positioned 5-20 mist nets (12-

m and 6-m in length) near flowering plants visited by hummingbirds (e.g., the genera *Heliconia*,

Centropogon, Palicourea, Musa). Nets were opened 30 minutes after sunrise and remained open

for 2-5 hours per session. Capture effort (net-hours) and net locations were kept constant for both

capture sessions, except in one instance (Appendix S2: Fig. S1). Following the North American

Banding Council's hummingbird protocol (Russell and Russell 2001), we tagged hummingbirds

with an aluminum tarsus band, measured body mass to the nearest 0.01 g using an electronic

balance, and measured wing length to the nearest 0.5 mm using a wing rule. Tarsus bands

allowed detection of recaptured hummingbirds within and between capture sessions; individuals

recaptured during the same capture session were only counted once. In 2017-2018, we also

marked hummingbirds' heads with unique color combinations of nail polish following Hadley *et*

al. (2014); this additional mark allowed us to identify individual hummingbirds seen on camera

(see *'Flower visitation'*). All birds were handled in accordance with Oregon State University

Animal Care and Use Protocols (ACUP #4655, #5015).

Radio telemetry

We attached miniaturized radio transmitters (0.15-0.25 g, Blackburn Transmitters,

Nacogdoches, TX) to 72 hummingbirds according to the methods of Hadley & Betts (2009). We

then followed hummingbirds on foot using portable receivers (Wildlife Materials Inc TRX

1000S) and handheld Yagi antennae, recorded locations continuously when the bird was found

within receiver range (~250 m in the absence of topographical barriers). In total, we completed

450 hours of telemetry observation across Days 2-4 and Days 6-8 of the experiment, averaging

3.1 ± 0.9 hours per session (typically ~8:00-11:00) and 5.4 ± 0.9 sessions per site (mean $\pm$ SD).

From the recorded locations, we estimated the proportion of observation time that radio-tagged

birds spent in the focal area. Thirty-six hummingbirds spent time in the focal area during the

'pre' period and were included in analyses of the *Heliconia* removal experiment; for full methods

and a comprehensive summary of radio-tracking outcomes, see Appendix S2.

Flower visitation

To understand hummingbird resource use within each site and to monitor for the presence

of marked individuals, we positioned trail cameras (PlotWatcher Pro, Day 6 Outdoors) near

inflorescences of plant species known or suspected to be visited by hummingbirds ($N = 34$,

Appendix S2: Table S6). Effort within each site was concentrated at two locations ('stations',

Fig. 1C) located on roughly opposite ends of the focal area (mean $\pm$ SD distance between

stations: 60 ± 44 m). Stations were centered around a focal *Heliconia* plant expected to flower

throughout the experimental period, not in an extremely steep area, and >10 m from the fragment

edge. The two focal *Heliconia* plants remained uncovered for the entire experimental period,

which allowed us to observe whether visitation rates changed after declines in surrounding

Heliconia density. To partially standardize plant species composition between control and

treatment replicates, we added a floral array comprising ~3-5 potted plant species to each station

(see Appendix S2 and *'Pollination success'*).

Trail cameras took one photograph per second during daylight hours (5:30-17:30 on Days

2-4 and Days 6-8) and automatically combined these photographs into a time-lapse video. We

reviewed videos for hummingbird sightings using the motion detection program MotionMeerkat

(Weinstein 2015) or by manually watching the sped-up video. When hummingbirds were

detected on camera, we identified the species, recorded any color marks, and noted whether they

visited any flowers. From video, we also recorded the number of open flowers available per day.

Dates with no available flowers were excluded from analysis, yielding 20,735 total hours of

video. We used subsets of these data for a variety of purposes, including estimating the total

floral resources available for hummingbirds under 'normal' (non-experimental) circumstances.

The analysis for the experiment used 19,870 video hours (Appendix S2: Table S7).

Pollination success

To investigate whether *Heliconia* declines influenced pollination success, we examined

styles for the presence of pollen tubes, which indicate pollen grain germination and subsequent

movement toward the flower ovule(s) (Kearns and Inouye 1993). We focused our pollen tube

investigation on the two focal *Heliconia* plants per replicate, as well as several species from the

focal arrays (Appendix S2: Table S6). Throughout the experimental period, we collected flowers

from these focal plants every 1-2 days, which corresponds to the maximum floral longevity for

most of the selected species (*Heliconia tortuosa*, *Hamelia patens*: Stratton 1989, *Stachytarpheta*

frantzii: Thomas-Granger 2003). Styles were fixed in formalin-acetic acid-alcohol for at least 24

hours, stained with aniline blue, and examined for pollen tubes using epifluorescence microscopy

(Kearns and Inouye 1993, Betts *et al.* 2015). Due to low pollen tube presence and/or challenges

with pollen tube visualization, we were only able to analyze two plant species: *H. tortuosa* ($N =$

327 styles) and *H. patens* ($N = 202$ styles). Additional details about style collection and lab

procedures are available in Appendix S2.

Additional response variables

To supplement the primary response variables described above, we examined several

additional metrics: (1) recapture probability (i.e., proportion of hummingbirds recaptured in the

'post' period), (2) floral visitation rates from individual hummingbirds marked with nail polish,

and (3) body mass of hummingbirds captured during both experimental periods (pre and post).

The first two response variables assessed the possibility that hummingbirds caught during the

initial capture sessions were rapidly replaced by new individuals, potentially obscuring treatment

effects. Body mass measurements allowed us to explore whether birds that remained in (or

returned to) treatment focal areas paid an energetic cost. To compare mass changes across

hummingbirds of different structural sizes, we calculated a relative body mass measure: observed

mass divided by the mass predicted by species-specific allometric equations (i.e., Ln(body mass)

~ Ln(wing length), Appendix S2: Fig. S4).

Statistical analysis

We used linear mixed models (LMMs) and generalized linear mixed models (GLMMs) to

analyze how experimental *Heliconia* extinction influenced (i) hummingbird abundance, (ii)

proportion of time that radio-tagged hummingbirds spent in focal area, (iii) visitation rates to

flowering plants (*Heliconia* and non-*Heliconia*), (iv) plant pollination success, and (v) the

presence and condition of individual hummingbirds (see '*Additional response variables*'). For

visitation to non-*Heliconia*, our main analysis examined overall visitation pattern across all plant

species; however, we also analyzed individual species when sufficient data were available (nine

species had >10 replicates each). We modeled each response variable using appropriate

statistical distributions and controlled for variable effort when applicable (Appendix S2: Tables

S8-S10). All statistical analyses were conducted in R version 4.2.0 (R Core Team 2022) using

'glmmTMB' (Brooks *et al.* 2017). We checked statistical assumptions by visually inspecting

plots from the 'performance' package (Lüdecke *et al.* 2021) for LMMs and 'DHARMa' (Hartig

2020) for GLMMs.

In a BACI experimental design, an overall treatment effect is indicated by a statistically

significant interaction between the variables representing treatment and experimental period

(Underwood 1994, McDonald *et al.* 2000). Therefore, we tested the *parallel extinction*

hypothesis (**H1**) and *robust network hypothesis* (**H2**) using a two-way interaction between

Heliconia removal and period (pre *versus* post) for all response variables measured during both

experimental periods (recapture probability was measured in the post period only). All models

included a random intercept for 'site'. Except when modeling recapture probability, we also

included a nested random effect for 'replicate' (site-year), which paired pre-to-post observations

within each replicate. Additional nested random effects for plant species and individual birds

were included where applicable (Appendix S2: Tables S8-S9). Because we were interested in

pre-to-post changes in our response variables, we only included covariates that varied between

periods. Specifically, when analyzing flower visitation, we accounted for the attractiveness of a

given focal plant by including a covariate for the number of open flowers on camera per day,

averaged across the experimental period.

For most response variables, we implemented more than one statistical model. Since the

magnitude of *Heliconia* removal varied widely (Appendix S2: Table S3), we conducted analyses

with a categorical treatment variable (control/treatment) and a quantitative treatment variable,

Ln(calories removed/hectare + 1). Results were qualitatively similar, so only results from the

categorical approach are presented in the main text. Additionally, to determine whether declines

in *Heliconia* affected *Heliconia* specialists more strongly than other hummingbird species (**H3**),

we analyzed hummingbird-centric response variables using (i) the full dataset containing all

species, and (ii) a dataset including green hermits and violet sabrewings. The one exception was

when analyzing flower visitation to individual non-*Heliconia* plant species; here, we focused on

the full dataset because *Heliconia* specialists visited some species at very low rates (or not at all)

and only used the categorical treatment variable (to limit the total number of models run).

RESULTS

Across all sites and years, we detected 19 hummingbird species in the forest fragments

surrounding the Las Cruces Biological Station (Appendix S4: Table S1). We estimate that, on

average, the covered *Heliconia* plants supplied 30% ($\pm$ 25%, range: 0.4 – 87%) of the calories

available for this entire hummingbird community and 37% ($\pm$ 28%, range: 0.4 – 94%) of the

calories available for green hermits and violet sabrewings (Appendix S4: Table S2). Although

green hermits and violet sabrewings visited >20 plant species, highest visitation rates were

observed for *Pitcairnia imbricata*, *Erythrina costaricensis*, *Musa x paradisiaca*, *Heliconia*

tortuosa, and *Costus barbatus* (Appendix S4: Fig. S1). *Heliconia* inflorescences were visited by

hummingbirds 6.4 ± 5.7 times per day (range: 0-25, median: 5), mostly by green hermits (75% of

1,055 visits, Appendix S4: Table S3).

Mist net captures & radio telemetry

We found little evidence that *Heliconia* removal reduced captured hummingbird

abundance or decreased the time that radio-tagged hummingbirds spent in the focal area. In

treatment replicates, we observed a pre-to-post decrease in hummingbird captures that was, on

average, 19% greater relative to control replicates but confidence intervals suggest a wide range

of possible effects (95% CI: -47% to +20%, Fig. 3A, Appendix S4: Table S4). Radio-tagged

hummingbirds from treatment replicates showed a similar pre-to-post decrease in the percentage

of time spent in the focal area (-18% relative to control replicates, 95% CI: -53% to +40%, Fig.

3B, Appendix S4: Table S4). Results were consistent when examining all species together and

Heliconia specialists separately (Appendix S4: Fig. S2, Tables S5-S6).

Flower visitation & pollination success

We continuously monitored hummingbird visits to the two focal *Heliconia* inflorescences

that remained uncovered throughout the experimental period, as well as numerous other

flowering plant species (mean ± SD: 7.8 ± 2 species/replicate; range: 2-11). Under the *robust*

network hypothesis, we expected hummingbirds to rewire and increasingly use non-*Heliconia*

resources. However, pre-to-post changes in hummingbird visitation rate were non-

distinguishable between control and treatment replicates, both when examining the focal

Heliconia plants (95% CI: -19% to +80%) and non-*Heliconia* plant species analyzed together (-

29% to +10%) (Fig. 4A). Responses by *Heliconia* specialists mirrored the responses observed for

all species (Fig. 4A, Appendix S4: Tables S7-S8), although these species visited non-*Heliconia*

species relatively infrequently (<1 visit/day, on average: Appendix S4: Fig. S3). When analyzing

non-*Heliconia* plant species individually, treatment replicates showed greater pre-to-post

declines (on average) for 7 of 9 species, but confidence intervals encompassed a range of

possible effects (Fig. 4B, Appendix S3: Table S9). We also found no evidence that experimental

Heliconia removal influenced pollination success for the focal *Heliconia* plants ($z = 0.15$, $P =$

0.88) or another plant species from the floral arrays (*Hamelia patens: z = 1.03*, $P = 0.30$) (Fig.

4C, Appendix S3: Fig. S4, Table S10).

Additional response variables

When focusing further on the responses of marked individual hummingbirds, we also did

not observe an effect of *Heliconia* removal. On average, the estimated probability of recapturing

a hummingbird from the 'pre' capture session was 18% in control replicates (95% CI: 11% to

29%), compared to 14% in treatment replicates (95% CI: 8% to 23%). This difference was not

statistically significant, and recaptures of *Heliconia* specialists showed qualitatively similar

results (Fig. 5A, Appendix S4: Table S11). Results were consistent when examining pre-to-post

changes in flower visitation for individual color-marked hummingbirds (Fig. 5B, Appendix S4:

Fig. S5, Tables S12-13). Finally, we found no evidence that pre-to-post changes in hummingbird

body condition differed between control and treatment replicates (Fig. 5C), either for all

hummingbird species ($z = -0.49$, $P = 0.62$) or *Heliconia* specialists ($z = -0.22$, $P = 0.83$)

(Appendix S4: Table S14).

DISCUSSION

In this study, we experimentally induced the local extinction of a locally abundant

tropical plant, *Heliconia tortuosa*, to assess the robustness of plant-hummingbird interaction

networks in a fragmented tropical landscape. Based on suite of complementary response

variables we found that, even when high proportions of total available calories were

experimentally removed (~30-40% on average), hummingbird responses fell within the natural

variability of control replicates. As might be expected with such small changes in hummingbird

behavior, we did not observe any cascading effects of experimental manipulation on pollination

success of flowering plants, which supports the *robust network hypothesis* (**Hl**). However, the

mechanism underlying the robust network hypothesis — hummingbirds expanding their diet to

include to alternative floral resources — did not receive strong support, suggesting other

explanations for hummingbird persistence.

Our finding that that experimental removal of *Heliconia* did not elicit parallel extinctions

in pollinators aligns with theoretical predictions (Memmott *et al.* 2004, Kaiser-Bunbury *et al.*

2010) and several other experiments that removed plant species from mutualistic networks

(Ferrero *et al.* 2013, Goldstein and Zych 2016, Costa *et al.* 2018). If hummingbirds remained in

forest fragments despite *Heliconia* declines, we assumed that they would rewire to use

alternative floral resources; hummingbirds as a taxonomic group are highly opportunistic

foragers (Arizmendi and Ornelas 1990, Maruyama *et al.* 2013) and the most frequent *Heliconia*

visitor, the green hermit, has a relatively wide niche breadth (Borgella *et al.* 2001, Betts *et al.*

2015). Unexpectedly, we found no evidence that hummingbirds increasingly visited non-

Heliconia resources when *Heliconia* became less available, suggesting that the mechanisms

underlying network robustness in this study may not necessarily align with the rewiring observed

in previous empirical tests (e.g., Timóteo *et al.* 2016, Kaiser-Bunbury *et al.* 2017, Costa *et al.*

2018). This apparent lack of foraging flexibility may emerge from various constraints, such as

morphological trait-matching, search costs, and/or competition from other hummingbirds (see

also Weinstein and Graham 2017). For example, high levels of exploitative competition for non-

defendable resources may have discouraged hummingbirds from modifying their trapline and

risking overlap with another bird's foraging route (Young 1971, Gill 1988, Ohashi and Thomson

2009). Lastly, is possible that fine-scale rewiring did occur but was not detected using our

analytical approach, which aggregated visitation rates to the level of hummingbird species;

further analysis examining the diet of individual hummingbirds may shed light on the possibility

of rewiring at finer scales.

In the absence of rewiring, the resilience in hummingbird behavior, abundance, and

pollination services following experimental *Heliconia* removal is somewhat surprising. Brief

pauses in nectar intake can disrupt a hummingbird's daily energy balance, given their high

metabolic rate and limited energy reserves (Calder and Booser 1973, Tooze and Gass 1985,

Brice 1992). Moreover, previous nectar manipulations with hummingbirds – albeit within

defended territories – led to rapid changes in territory size, foraging patterns, and/or defense

behavior, generally within a day (e.g., Hixon *et al.* 1983, Gass and Sutherland 1985, Eberhard

and Ewald 1994, Temeles *et al.* 2004). One explanation for our result is that hummingbirds

permanently emigrated from fragments after *Heliconia* removal but, because of high intruder

pressure, were quickly replaced by new individuals (Stiles and Wolf 1970). However, we

monitored individual birds using several different approaches (radio telemetry, recaptures, flower

visitation) and found no evidence that *Heliconia* removal caused emigration and subsequent

replacement. An alternative possibility is that hummingbirds relied on physiological adaptations,

such as a torpor and/or insectivory, to maintain body condition despite large declines in nectar

availability – resulting in the stable body mass measurements we observed in recaptured

hummingbirds. Torpor is an energy-saving adaptation characterized by declines in body

temperature and metabolic rate (Hainsworth *et al.* 1977, Spence and Tingley 2021), and even

tropical hummingbirds inhabiting low-to-mid elevations appear to possess this capability,

including the green hermit (Krüger *et al.* 1982, Schuchmann and Prinzinger 1988, Bech *et al.*

1997, Shankar *et al.* 2020). Additionally, hummingbirds are highly insectivorous and, although

they cannot completely replace nectar with insects (Brice 1992, Stiles 1995), they may

supplement their diet with arthropods when nectar becomes scarce (Young 1971, Kuban and

Neill 1980, Montgomerie and Redsell 1980, Hazlehurst and Karubian 2018). Finding torpid

hummingbirds at night is extremely challenging (Carpenter and Hixon 1988), but future research

could explore changes in arthropod consumption using techniques such as DNA metabarcoding

(Moran *et al.* 2019).

Another explanation for hummingbird persistence following experimental *Heliconia*

removal involves their high movement ability. *H. tortuosa* is a patchily distributed species that

necessitates long-distance foraging movements, as a single clump does not make territoriality

energetically profitable (Stiles 1975). Densities can also vary substantially across short distances;

for instance, we observed a 150-fold difference in *Heliconia* availability across sites and years in

the present study (Appendix S4: Table S15). Similarly, Stiles (1979) notes that some Costa Rican

Heliconia species can be common in one watershed, yet nearly absent in an adjacent watershed <

1 km away (see also Bruna and Kress 2002). In accordance with this natural history, the most

frequent *Heliconia* visitor, the green hermit, can travel long distances (>500 m) while foraging

within home ranges of >150 ha (although home range sizes of ~2 ha are more typical: Volpe *et*

al. 2014, 2016). Thus, returning to our focal areas to visit the few remaining *Heliconia* resources

could be perceived as a relatively low-cost option for long-distance trapliners, at least in the short

term. Notably, although highly mobile pollinators might buffer against network collapse (Volpe

et al. 2016), they may be vulnerable to habitat loss and fragmentation once thresholds are

crossed, leading to parallel pollinator declines observed in nature (Andrén 1994, Hadley and

Betts 2012). This strategy is also unlikely to be profitable over longer time scales than measured

by our experiment, which only removed *Heliconia* for five days.

Although a short adjustment period (<1 day) is standard in similar species removal

experiments (e.g., Hixon *et al.* 1983, Brosi and Briggs 2013, Hazlehurst and Karubian 2018),

extending the experimental timeline may have more realistically simulated a future extinction

and provided more time for behavioral or demographic adjustment. If hummingbirds were not

permanent residents of our focal areas – as suggested by our relatively low recapture rates

(~15%) and proportion of time spent in the focal area (~20%) – then transient individuals might

have not immediately noticed changes in *Heliconia* density and changed their foraging patterns

accordingly. Likewise, hummingbirds might not have persisted had we removed *Heliconia* over

larger areas, because traveling to isolated plants would become even more energetically costly

(Heinrich and Raven 1972). Unfortunately, covering (and then uncovering) *Heliconia* plants in

steep terrain with dense understories limited the area we could manipulate.

We also acknowledge that our ability to detect subtle experimental effects may have been

limited by low statistical power, owing to high individual-level variation and limited sample size.

Nevertheless, we can rule out extreme declines in hummingbird space use, flower visitation, and

pollination success that would be indicative of complete network collapse; in the main analyses,

treatment replicates generally showed no more than a 50% pre-to-post decline compared to

control replicates (based on the lower confidence interval of the relevant contrast, e.g., Fig. 3A-

B, Fig. 4A, C). We also emphasize that the spatial scale of our experiment and amount of

replication surpasses all but two empirical studies simulating species extinctions within plant-

pollinator networks (Appendix S1: Table S1). Still, future removal experiments with highly

mobile pollinators should consider increasing the amount of replication, and, if possible,

manipulate larger areas.

CONCLUSIONS

Hummingbird pollinators seemed able to persist during our experiments that removed

floral resources for a short period of time (several days), but we caution that this persistence may

not be sustainable or reflect the outcome of a natural extinction process. Longer-term

experiments will be key for predicting consequences of species loss and understanding

mechanisms of network robustness – at least for pollinator species that forage at large spatial

scales. Our study also suggests that behavioral flexibility to use alternative floral resources might

be limited, even in taxonomic groups thought to be highly opportunistic foragers. Thus, even

rewiring rules with strong biological underpinnings (e.g., Vizentin-Bugoni *et al.* 2020) may not

necessarily predict real-world responses to species loss, perhaps due to additional constraints

such as competition. To further understand the mechanisms underlying network robustness or

collapse, we encourage future experiments that consider how temporal scale, species traits (e.g.,

movement ability), competition, and landscape context interact to determine the outcome of

species extinctions. These experiments could be especially valuable at geographic range limits,

where climate change will likely lead to species replacements (HilleRisLambers *et al.* 2013), and

in the tropics where landscape change and species loss are most rapid (Hansen *et al.* 2013).

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REFERENCES

Allen-Wardell, G., P. Bernhardt, R. Bitner, A. Burquez, S. Buchmann, J. Cane, P. A. Cox, V.

Dalton, P. Feinsinger, M. Ingram, D. Inouye, C. E. Jones, K. Kennedy, P. Kevan, H.

Koopowitz, R. Medellin, S. Medellin-Morales, G. P. Nabhan, B. Pavlik, V. Tepedino, P.

Torchio, and S. Walker. 1998. The potential consequences of pollinator declines on the

conservation of biodiversity and stability of food crop yields. Conservation Biology

12:8–17.

Anderson, S. H., D. Kelly, J. J. Ladley, S. Molloy, and J. Terry. 2011. Cascading effects of bird

functional extinction reduce pollination and plant density. Science 331:1068–1071.

Andrén, H. 1994. Effects of habitat fragmentation on birds and mammals in landscapes with

different proportions of suitable habitat: A review. Oikos 71:355–366.

Arizmendi, M. C., and J. F. Ornelas. 1990. Hummingbirds and their floral resources in a tropical

dry forest in Mexico. Biotropica 22:172–180.

Ashman, T. L., T. M. Knight, J. A. Steets, P. Amarasekare, M. Burd, D. R. Campbell, M. R.

Dudash, M. O. Johnston, S. J. Mazer, and R. J. Mitchell. 2004. Pollen limitation of plant

reproduction: ecological and evolutionary causes and consequences. Ecology 85:2408–

2421.

Aslan, C. E., E. S. Zavaleta, B. Tershy, and D. Croll. 2013. Mutualism disruption threatens

global plant biodiversity: A systematic review. PLOS ONE 8:e66993.

Bascompte, J., and P. Jordano. 2007. Plant-animal mutualistic networks: the architecture of

biodiversity. Annual Review of Ecology, Evolution, and Systematics 38:567–593.

Bascompte, J., P. Jordano, C. J. Melián, and J. M. Olesen. 2003. The nested assembly of plant–

animal mutualistic networks. Proceedings of the National Academy of Sciences

100:9383–9387.

Bascompte, J., P. Jordano, and J. M. Olesen. 2006. Asymmetric coevolutionary networks

facilitate biodiversity maintenance. Science 312:431–433.

Bech, C., A. S. Abe, J. F. Steffensen, M. Berger, and J. E. P. W. Bicudo. 1997. Torpor in three

species of Brazilian hummingbirds under semi-natural conditions. The Condor 99:780–

788.

Betts, M. G., A. S. Hadley, and W. J. Kress. 2015. Pollinator recognition by a keystone tropical

plant. Proceedings of the National Academy of Sciences 112:3433–3438.

Biesmeijer, J. C., S. P. M. Roberts, M. Reemer, R. Ohlemüller, M. Edwards, T. Peeters, A. P.

Schaffers, S. G. Potts, R. Kleukers, C. D. Thomas, J. Settele, and W. E. Kunin. 2006.

Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands.

Science 313:351–354.

Borgella, R., A. A. Snow, and T. A. Gavin. 2001. Species richness and pollen loads of

hummingbirds using forest fragments in southern Costa Rica. Biotropica 33:90–109.

Brice, A. T. 1992. The essentiality of nectar and arthropods in the diet of the Anna's

hummingbird (*Calypte anna*). Comparative Biochemistry and Physiology Part A:

Physiology 101:151–155.

Brooks, M. E., K. Kristensen, K. J. van Benthem, A. Magnusson, C. W. Berg, A. Nielsen, H. J.

Skaug, M. Maechler, and B. M. Bolker. 2017. glmmTMB Balances Speed and Flexibility

Among Packages for Zero-inflated Generalized Linear Mixed Modeling. The R Journal

9:378–400.

Brosi, B. J., and H. M. Briggs. 2013. Single pollinator species losses reduce floral fidelity and

plant reproductive function. Proceedings of the National Academy of Sciences

110:13044–13048.

Bruna, E. M., and W. J. Kress. 2002. Habitat fragmentation and the demographic structure of an

Amazonian understory herb (*Heliconia acuminata*). Conservation Biology 16:1256–

1266.

Calder, W. A., and J. Booser. 1973. Hypothermia of broad-tailed hummingbirds during

incubation in nature with ecological correlations. Science 180:751–753.

Carpenter, F. L., and M. A. Hixon. 1988. A new function for torpor: fat conservation in a wild

migrant hummingbird. The Condor 90:373–378.

Colwell, R. K., R. R. Dunn, and N. C. Harris. 2012. Coextinction and persistence of dependent

species in a changing world. Annual Review of Ecology, Evolution, and Systematics

43:183–203.

Costa, J. M., J. A. Ramos, L. P. da Silva, S. Timóteo, P. Andrade, P. M. Araújo, C. Carneiro, E.

Correia, P. Cortez, M. Felgueiras, C. Godinho, R. J. Lopes, C. Matos, A. C. Norte, P. F.

Pereira, A. Rosa, and R. H. Heleno. 2018. Rewiring of experimentally disturbed seed

dispersal networks might lead to unexpected network configurations. Basic and Applied

Ecology 30:11–22.

Dunn, R. R., N. C. Harris, R. K. Colwell, L. P. Koh, and N. S. Sodhi. 2009. The sixth mass

coextinction: are most endangered species parasites and mutualists? Proceedings of the

Royal Society B: Biological Sciences 276:3037–3045.

Eberhard, J. R., and P. W. Ewald. 1994. Food availability, intrusion pressure and territory size:

an experimental study of Anna's hummingbirds (*Calypte anna*). Behavioral Ecology and

Sociobiology 34:11–18.

Feinsinger, P., and R. K. Colwell. 1978. Community organization among Neotropical nectar-

feeding birds. American Zoologist 18:779–795.

Ferrero, V., S. Castro, J. Costa, P. Acuña, L. Navarro, and J. Loureiro. 2013. Effect of invader

removal: pollinators stay but some native plants miss their new friend. Biological

Invasions 15:2347–2358.

Gass, C. L., and G. D. Sutherland. 1985. Specialization by territorial hummingbirds on

experimentally enriched patches of flowers: energetic profitability and learning. Canadian

Journal of Zoology 63:2125–2133.

Gilbert, L. E. 1980. Food web organization and conservation of neotropical diversity. Pages 11–

33 *in* M. E. Soulé and B. A. Wilcox, editors. Conservation Biology: An Evolutionary-

Ecological Perspective. Sinauer, Sunderland.

Gill, F. B. 1988. Trapline foraging by hermit hummingbirds: competition for an undefended,

renewable resource. Ecology 69:1933–1942.

Goldstein, J., and M. Zych. 2016. What if we lose a hub? Experimental testing of pollination

network resilience to removal of keystone floral resources. Arthropod-Plant Interactions

10:263–271.

Hadley, A. S., and M. G. Betts. 2009. Tropical deforestation alters hummingbird movement

patterns. Biology Letters 5:207–210.

Hadley, A. S., and M. G. Betts. 2012. The effects of landscape fragmentation on pollination

dynamics: absence of evidence not evidence of absence. Biological Reviews 87:526–544.

Hadley, A. S., S. J. K. Frey, W. D. Robinson, and M. G. Betts. 2018. Forest fragmentation and

loss reduce richness, availability, and specialization in tropical hummingbird

communities. Biotropica 50:74–83.

Hadley, A. S., S. J. K. Frey, W. D. Robinson, W. J. Kress, and M. G. Betts. 2014. Tropical forest

fragmentation limits pollination of a keystone understory herb. Ecology 95:2202–2212.

Hainsworth, F. R., B. G. Collins, and L. L. Wolf. 1977. The function of torpor in hummingbirds.

Physiological Zoology 50:215–222.

Hansen, M. C., P. V. Potapov, R. Moore, M. Hancher, S. A. Turubanova, A. Tyukavina, D.

Thau, S. V. Stehman, S. J. Goetz, T. R. Loveland, A. Kommareddy, A. Egorov, L. Chini,

C. O. Justice, and J. R. G. Townshend. 2013. High-resolution global maps of 21st-

century forest cover change. Science 342:850–853.

Hartig, F. 2020. DHARMa: Residual Diagnostics for Hierarchical (Multi-Level / Mixed)

Regression Models.

Hazlehurst, J. A., and J. O. Karubian. 2018. Impacts of nectar robbing on the foraging ecology of

a territorial hummingbird. Behavioural Processes 149:27–34.

Heinrich, B., and P. H. Raven. 1972. Energetics and pollination ecology. Science 176:597–602.

HilleRisLambers, J., M. A. Harsch, A. K. Ettinger, K. R. Ford, and E. J. Theobald. 2013. How

will biotic interactions influence climate change–induced range shifts? Annals of the

New York Academy of Sciences 1297:112–125.

Hixon, M. A., F. L. Carpenter, and D. C. Paton. 1983. Territory area, flower density, and time

budgeting in hummingbirds: an experimental and theoretical analysis. The American

Naturalist 122:366–391.

Jordán, F. 2009. Keystone species and food webs. Philosophical Transactions of the Royal

Society B: Biological Sciences 364:1733–1741.

Kaiser-Bunbury, C. N., J. Mougal, A. E. Whittington, T. Valentin, R. Gabriel, J. M. Olesen, and

N. Blüthgen. 2017. Ecosystem restoration strengthens pollination network resilience and

function. Nature 542:223–227.

Kaiser-Bunbury, C. N., S. Muff, J. Memmott, C. B. Müller, and A. Caflisch. 2010. The

robustness of pollination networks to the loss of species and interactions: a quantitative

approach incorporating pollinator behaviour. Ecology Letters 13:442–452.

Kearns, C. A., and D. W. Inouye. 1993. Techniques for Pollination Biologists. University Press

of Colorado, Niwot.

Klein, A.-M., B. E. Vaissière, J. H. Cane, I. Steffan-Dewenter, S. A. Cunningham, C. Kremen,

and T. Tscharntke. 2007. Importance of pollinators in changing landscapes for world

crops. Proceedings of the Royal Society B: Biological Sciences 274:303–313.

Koh, L. P., R. R. Dunn, N. S. Sodhi, R. K. Colwell, H. C. Proctor, and V. S. Smith. 2004.

Species Coextinctions and the Biodiversity Crisis. Science 305:1632–1634.

Kormann, U., C. Scherber, T. Tscharntke, N. Klein, M. Larbig, J. J. Valente, A. S. Hadley, and

M. G. Betts. 2016. Corridors restore animal-mediated pollination in fragmented tropical

forest landscapes. Proceedings of the Royal Society B: Biological Sciences

283:20152347.

Krüger, K., R. Prinzinger, and K.-L. Schuchmann. 1982. Torpor and metabolism in

hummingbirds. Comparative Biochemistry and Physiology Part A: Physiology 73:679–

689.

Kuban, J. F., and R. L. Neill. 1980. Feeding ecology of hummingbirds in the highlands of the

Chisos Mountains, Texas. The Condor 82:180–185.

Kuussaari, M., R. Bommarco, R. K. Heikkinen, A. Helm, J. Krauss, R. Lindborg, E. Öckinger,

M. Pärtel, J. Pino, F. Roda, and others. 2009. Extinction debt: a challenge for biodiversity

conservation. Trends in ecology & evolution 24:564–571.

Lenth, R. 2020. emmeans: Estimated Marginal Means, aka Least-Squares Means.

Lüdecke, D., M. Ben-Shachar, I. Patil, P. Waggoner, and D. Makowski. 2021. performance: An

R Package for Assessment, Comparison and Testing of Statistical Models. Journal of

Open Source Software 6:3139.

Maruyama, P. K., G. M. Oliveira, C. Ferreira, B. Dalsgaard, and P. E. Oliveira. 2013. Pollination

syndromes ignored: importance of non-ornithophilous flowers to Neotropical savanna

hummingbirds. Naturwissenschaften 100:1061–1068.

McDonald, T. L., W. P. Erickson, and L. L. McDonald. 2000. Analysis of count data from

before-after control-impact studies. Journal of Agricultural, Biological, and

Environmental Statistics 5:262–279.

Memmott, J., N. M. Waser, and M. V. Price. 2004. Tolerance of pollination networks to species

extinctions. Proceedings of the Royal Society B: Biological Sciences 271:2605–2611.

Mendenhall, C. D., and A. M. Wrona. 2018. Improving tree cover estimates for fine-scale

landscape ecology. Landscape Ecology 33:1691–1696.

Montgomerie, R. D., and C. A. Redsell. 1980. A nesting hummingbird feeding solely on

arthropods. The Condor 82:463.

Moran, A. J., S. W. J. Prosser, and J. A. Moran. 2019. DNA metabarcoding allows non-invasive

identification of arthropod prey provisioned to nestling Rufous hummingbirds

(*Selasphorus rufus*). PeerJ 7:e6596.

Ohashi, K., and J. D. Thomson. 2009. Trapline foraging by pollinators: its ontogeny, economics

and possible consequences for plants. Annals of Botany 103:1365–1378.

Ollerton, J., R. Winfree, and S. Tarrant. 2011. How many flowering plants are pollinated by

animals? Oikos 120:321–326.

Pauw, A., and J. A. Hawkins. 2011. Reconstruction of historical pollination rates reveals linked

declines of pollinators and plants. Oikos 120:344–349.

Phillips, R. D., R. Peakall, B. A. Retter, K. Montgomery, M. H. M. Menz, B. J. Davis, C. Hayes,

G. R. Brown, N. D. Swarts, and K. W. Dixon. 2015. Pollinator rarity as a threat to a plant

with a specialized pollination system. Botanical Journal of the Linnean Society 179:511–

525.

Potts, S. G., J. C. Biesmeijer, C. Kremen, P. Neumann, O. Schweiger, and W. E. Kunin. 2010.

Global pollinator declines: trends, impacts and drivers. Trends in Ecology & Evolution

25:345–353.

Powers, D. R., A. R. Brown, and J. A. Van Hook. 2003. Influence of normal daytime fat

deposition on laboratory measurements of torpor use in territorial versus nonterritorial

hummingbirds. Physiological and Biochemical Zoology 76:389–397.

R Core Team. 2022. R: A Language and Environment for Statistical Computing. R Foundation

for Statistical Computing, Vienna, Austria.

Ramos-Jiliberto, R., F. S. Valdovinos, P. Moisset de Espanés, and J. D. Flores. 2012.

654   Topological plasticity increases robustness of mutualistic networks. Journal of Animal

655   Ecology 81:896–904.

656   Regan, E. C., L. Santini, L. Ingwall☐King, M. Hoffmann, C. Rondinini, A. Symes, J. Taylor,

657   and S. H. M. Butchart. 2015. Global trends in the status of bird and mammal pollinators.

658   Conservation Letters 8:397–403.

659   Rodríguez-Flores, C. I., J. F. Ornelas, S. Wethington, and M. C. Arizmendi. 2019. Are

660   hummingbirds generalists or specialists? Using network analysis to explore the

661   mechanisms influencing their interaction with nectar resources. PLOS ONE

662   14:e0211855.

663   Russell, S. M., and R. O. Russell. 2001. The North American Banders' Manual for Banding

664   Hummingbirds. North American Banding Council, Point Reyes Station, CA.

665   Scheper, J., M. Reemer, R. van Kats, W. A. Ozinga, G. T. J. van der Linden, J. H. J. Schaminée,

666   H. Siepel, and D. Kleijn. 2014. Museum specimens reveal loss of pollen host plants as

667   key factor driving wild bee decline in The Netherlands. Proceedings of the National

668   Academy of Sciences 111:17552–17557.

669   Schuchmann, K.-L., and R. Prinzinger. 1988. Energy metabolism, nocturnal torpor, and

670   respiration frequency in a Green Hermit (*Phaethornis guy*). Journal für Ornithologie

671   129:469–472.

672   Shankar, A., R. J. Schroeder, S. M. Wethington, C. H. Graham, and D. R. Powers. 2020.

673   Hummingbird torpor in context: duration, more than temperature, is the key to nighttime

674   energy savings. Journal of Avian Biology 51:e02305.

Spence, A. R., and M. W. Tingley. 2021. Body size and environment influence both intraspecific

and interspecific variation in daily torpor use across hummingbirds. Functional Ecology

35:870–883.

Stiles, F. G. 1975. Ecology, flowering phenology, and hummingbird pollination of some Costa

Rican Heliconia species. Ecology 56:285–301.

Stiles, F. G. 1979. Notes on the natural history of *Heliconia* (Musaceae) in Costa Rica. Notas

sobre la historia natural de *Heliconia* (Musaceae) en Costa Rica. Brenesia 15:151–180.

Stiles, F. G. 1985. Seasonal patterns and coevolution in the hummingbird-flower community of a

Costa Rican subtropical forest. Ornithological Monographs 36:757–787.

Stiles, F. G. 1995. Behavioral, ecological and morphological correlates of foraging for

arthropods by the hummingbirds of a tropical wet forest. The Condor 97:853–878.

Stiles, F. G., and L. L. Wolf. 1970. Hummingbird territoriality at a tropical flowering tree. The

Auk 87:467–491.

Stratton, D. A. 1989. Longevity of individual flowers in a Costa Rican cloud forest: ecological

correlates and phylogenetic constraints. Biotropica 21:308–318.

Suarez, R. K. 1992. Hummingbird flight: sustaining the highest mass-specific metabolic rates

among vertebrates. Experientia 48:565–570.

Taylor, J., and S. A. White. 2007. Observations of hummingbird feeding behavior at flowers of

Heliconia beckneri and *H. tortuosa* in southern Costa Rica. Ornitologia Neotropical

18:133–138.

Temeles, E. J., A. B. Muir, E. B. Slutsky, and M. N. Vitousek. 2004. Effect of food reductions on

territorial behavior of purple-throated caribs. The Condor 106:691–695.

Thomas-Granger, K. 2003. Light sum, nitrogen level and growth retardant effects on flower and

698   nectar production in *Stachytarpheta frantzii*. Master's thesis, University of Guelph.

699   Timóteo, S., J. A. Ramos, I. P. Vaughan, and J. Memmott. 2016. High resilience of seed

700   dispersal webs highlighted by the experimental removal of the dominant disperser.

701   Current Biology 26:910–915.

702   Tooze, Z. J., and C. L. Gass. 1985. Responses of rufous hummingbirds to midday fasts. Canadian

703   Journal of Zoology 63:2249–2253.

704   Traveset, A., C. Tur, and V. M. Eguíluz. 2017. Plant survival and keystone pollinator species in

705   stochastic coextinction models: role of intrinsic dependence on animal-pollination.

706   Scientific Reports 7:6915.

707   Underwood, A. J. 1994. On Beyond BACI: sampling designs that might reliably detect

708   environmental disturbances. Ecological Applications 4:3–15.

709   Valdovinos, F. S., P. Moisset de Espanés, J. D. Flores, and R. Ramos-Jiliberto. 2013. Adaptive

710   foraging allows the maintenance of biodiversity of pollination networks. Oikos 122:907–

711   917.

712   Vellend, M., K. Verheyen, H. Jacquemyn, A. Kolb, H. V. Calster, G. Peterken, and M. Hermy.

713   2006. Extinction debt of forest plants persists for more than a century following habitat

714   fragmentation. Ecology 87:542–548.

715   Vizentin-Bugoni, J., V. J. Debastiani, V. A. G. Bastazini, P. K. Maruyama, and J. H. Sperry.

716   2020. Including rewiring in the estimation of the robustness of mutualistic networks.

717   Methods in Ecology and Evolution 11:106–116.

Volpe, N. L., A. S. Hadley, W. D. Robinson, and M. G. Betts. 2014. Functional connectivity

experiments reflect routine movement behavior of a tropical hummingbird species.

Ecological Applications 24:2122–2131.

Volpe, N. L., W. D. Robinson, S. J. K. Frey, A. S. Hadley, and M. G. Betts. 2016. Tropical forest

fragmentation limits movements, but not occurrence of a generalist pollinator species.

PLOS ONE 11:e0167513.

Weiner, C. N., M. Werner, K. E. Linsenmair, and N. Blüthgen. 2014. Land-use impacts on

plant–pollinator networks: interaction strength and specialization predict pollinator

declines. Ecology 95:466–474.

Weinstein, B. G. 2015. MotionMeerkat: integrating motion video detection and ecological

monitoring. Methods in Ecology and Evolution 6:357–362.

Weinstein, B. G., and C. H. Graham. 2017. Persistent bill and corolla matching despite shifting

temporal resources in tropical hummingbird-plant interactions. Ecology Letters 20:326–

335.

Wolf, L. L., and F. G. Stiles. 1989. Adaptations for the "fail-safe" pollination of specialized

ornithophilous flowers. The American Midland Naturalist 121:1–10.

Young, A. M. 1971. Foraging for insects by a tropical hummingbird. The Condor 73:36–45.

Zahawi, R. A., G. Duran, and U. Kormann. 2015. Sixty-seven years of land-use change in

southern Costa Rica. PLOS ONE 10:e0143554.

TABLES

Table 1. Hypotheses testing whether declines of a locally common, tropical flowering plant (*Heliconia tortuosa*) affect hummingbird

space use, flower visitation rates, and plant pollination success. Predicted effects are indicated in the column below each hypothesis.

We experimentally removed *H. tortuosa* using a BACI (Before-After-Control-Impact) experimental design (Fig. 2), so predicted

effects can be conceptualized as pre-to-post changes in treatment replicates compared to pre-to-post changes in control replicates.

	Response variable	Methodological approach	How do hummingbirds respond to declines in *Heliconia,* and are there cascading effects on the remaining plants?		
			Robust network hypothesis (H1)	Parallel extinction hypothesis (H2)	Results from this study
Space use	Abundance	Mist net captures	No change	Decrease	**No change**
	Time spent in focal area	Radio telemetry			
Flower visitation rates	Visitation to focal *Heliconia* plants	Camera observations	No change or increase	Decrease	**No change**
	Visitation to non-*Heliconia* plants	Camera observations	Increase (rewiring)	Decrease	**No change**
Pollination services	Pollination success	Pollen tubes	No change or increase	Decrease	**No change**

  FIGURES

Figure 1. This study was conducted in the fragmented landscape surrounding the Las Cruces

Biological Station (A) in southern Costa Rica (B). Within the study system (rectangular extent),

we utilized fourteen sites. Forested areas are shown in grey, and non-forested areas (white)

primarily comprise pasture. The forest layer is a combination of areas digitized for this study

(generally within 1000 m of each site) and those digitized by Mendenhall & Wrona (2018). (C)

An example site, with red line representing a focal area [i.e., where we removed *Heliconia* (for

treatment replicates) and/or surveyed for floral resources (for treatment and control replicates)].

Within focal areas, we captured hummingbirds using mist nets (triangles) and monitored plant-

hummingbird interactions using trail cameras (camera icons).

Figure 2. We temporarily removed *Heliconia* by covering inflorescences with fabric bags (A) as

part of a Before-After-Control-Impact experimental design (B). Within a year (2016–2018), each

site received either *Heliconia* removal (Treatment) or was assigned to be a Control. Treatments

were reversed in alternate years for a total maximum sample size of 16 control replicates and 16

treatment replicates. Additionally, sites were sampled during two experimental periods: Before

('Pre') and After ('Post'). With this experimental design, the comparison of interest is the pre-to-

post change in treatment replicates compared to the pre-to-post change in control replicates.

Photos provided by M. Donald.

Bird group   • All species   ▲ *Heliconia* specialists

Figure 3. Effects of experimental *Heliconia* removal on hummingbird abundance (A) and space use (B). Hummingbird abundance was quantified as the number of hummingbirds captured in mist nets, controlling for capture effort (net-hours). Space use was quantified as the percentage of time that radio-tagged hummingbirds spent in the focal area. Following the Before-After-Control-Impact experimental design, the *y*-axis reflects the pre-to-post change in treatment replicates relative to pre-to-post change in control replicates. Estimates and 95% confidence intervals are contrasts calculated in 'emmeans' (Lenth 2020). The dashed line at 1 indicates no experimental effect; the grey area indicates strong support for the *network collapse hypothesis* (>50% decrease in treatment replicates relative to control replicates). *Heliconia* specialists refers to the two hummingbird species expected to rely most strongly on *H. tortuosa*: green hermits (*P. guy*) and violet sabrewings (*C. hemileucurus*).

A Flower visitation (visits/hr)

C Pollination success (% flowers pollinated)

B Flower visitation to individual non-*Heliconia* species (visits/hr)

Bird group ● All species ▲ *Heliconia* specialists

Figure 4. Effects of experimental *Heliconia* removal on flower visitation (A-B) and pollination

success (C). Flower visitation was quantified using trail cameras that recorded hummingbird

visits to focal *Heliconia* (2 inflorescences/site) and a 30 non-*Heliconia* plant species (Appendix

S2: Table S6). Pollination success was quantified as the percentage of flowers with pollen tubes.

We analyzed non-*Heliconia* visitation across all plant species (A) and, when sufficient data were

available, to individual plant species (B). Following the Before-After-Control-Impact

experimental design, the *y*-axis reflects the pre-to-post change in treatment replicates relative to

pre-to-post change in control replicates. Estimates and 95% confidence intervals are contrasts

calculated in 'emmeans' (Lenth 2020). The dashed line at 1 indicates no experimental effect; the

grey area indicates strong support for the *network collapse hypothesis* (>50% decrease in

treatment replicates relative to control replicates). *Heliconia* specialists refers to the two

hummingbird species expected to rely most strongly on *H. tortuosa*: green hermits (*P. guy*) and

violet sabrewings (*C. hemileucurus*).

A Recapture probability (% of birds recaptured)

B Flower visitation from marked birds (visits/hr)

focal *Heliconia*   non-*Heliconia* species (*N*=17)

C Bird condition (relative body mass)

Bird group ● All species ▲ *Heliconia* specialists

Figure 5. Effects of experimental *Heliconia* removal on additional response variables, which investigate whether hummingbirds present at the start of the experiment were replaced by new individuals (A-B, mist net recaptures and flower visitation of marked birds) or suffered an energetic cost by remaining in the focal area (C, bird condition). Following the Before-After-Control-Impact experimental design, the *y*-axis reflects treatment replicates relative to control replicates (A) or the pre-to-post change in treatment replicates relative to pre-to-post change in control replicates (B, C). Estimates and 95% confidence intervals are contrasts calculated with 'emmeans' (Lenth 2020). The dashed lines at 1 and 0 indicate no experimental effect; the grey area indicates strong support for the *network collapse hypothesis* (>50% decrease in treatment replicates relative to control replicates). *Heliconia* specialists refers to the two hummingbird species expected to rely most strongly on *H. tortuosa*: green hermits (*P. guy*) and violet sabrewings (*C. hemileucurus*).