Phenotypic and transcriptional response of *Daphnia pulicaria* to the combined effects of temperature and predation

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Abstract

Daphnia, an ecologically important zooplankton species in lakes, shows both genetic adaptation and phenotypic plasticity in response to temperature and fish predation, but little is known about the molecular basis of these responses and their potential interactions. We performed a factorial experiment exposing laboratory-propagated *Daphnia pulicaria* clones from two lakes in the Sierra Nevada mountains of California to normal or high temperature (15°C or 25°C) in the presence or absence of fish kairomones, then measured changes in life history and gene expression. Exposure to kairomones increased upper thermal tolerance limits for physiological activity in both clones. Cloned individuals matured at a younger age in response to higher temperature and kairomones, while size at maturity, fecundity and population intrinsic growth were only affected by temperature. At the molecular level, both clones expressed more genes differently in response to temperature than predation, but specific genes involved in metabolic, cellular, and genetic processes responded differently between the two clones. Although gene expression differed more between clones from different lakes than experimental treatments, similar phenotypic responses to predation risk and warming arose from these clone-specific patterns. Our results suggest that phenotypic plasticity responses to temperature and kairomones interact synergistically, with exposure to fish predators increasing the tolerance of *Daphnia pulicaria* to stressful temperatures, and that similar phenotypic responses to temperature and predator cues can be produced by divergent patterns of gene regulation.

Introduction

Many species are at risk of extinction as the environment changes at an unprecedented pace. Ecological and evolutionary research aims to predict species' responses to anthropogenic environmental change, such as global warming and introduction of predators [1,2]. To cope with such stressors, species can either migrate to more suitable habitats or adapt to new conditions. Understanding the potential and limits of genetic adaptation and phenotypic plasticity to maintain fitness is critical in order to predict persistence of populations and changes in biodiversity under environmental stresses [3]. The mechanisms by which organisms are adapted to present day conditions throughout their ranges therefore provide a basis for predicting extinction, persistence, and changes in biodiversity in a rapidly changing environment [4,5].

Plasticity and genetic adaptation impact organismal fitness in response to environmental change. Plasticity allows a genotype to express multiple phenotypes in different environments whereas genetic adaptation arises from environment-dependent variation in fitness of different genotypes [6]. Evidence for rapid evolution in natural systems, particularly in response to human-induced environmental change, indicates that most of the observed changes are not genetically based, but rather a consequence of plasticity [7]. Plasticity at level of gene expression is one of the most important mechanisms for coping with stress [8,9], yet the magnitude and genetic basis of such plasticity remains largely unknown.

Plasticity arises from differential gene expression patterns in response to environmental cues [10]. Variable levels of plasticity may evolve in a population if reaction norms differ across genotypes and the slope of the reaction norm is correlated with fitness

70 [11]. Either decreases or increases in phenotypic plasticity could contribute to adaptation
71 to variable environments depending on the rate and predictability of environmental change
72 [6,12].

73 Several factors could constrain the evolution of plasticity at the transcriptome level.
74 First, genetic variation potential for plasticity could be limited or absent. Alternatively,
75 even when genetic variation is present, its evolution could be constrained by costs of the
76 mechanisms underlying plastic responses [13]. A trade-off is expected where enhanced
77 plasticity would be beneficial in more spatially or temporally variable environments, but
78 detrimental in a stable environment [14,15]. In addition, if environmental variation is
79 unpredictable, then plasticity may fail to match phenotypes to the environment to produce
80 higher fitness.

81 *Daphnia* is an ecologically important genus of crustacean zooplankton in lakes that
82 transfers energy from phytoplankton to fish and invertebrate predators and exerts top-down
83 grazing control on primary production [16]. *Daphnia* shows both genetic adaptation and
84 plasticity at a phenotypic level in tolerance to two important determinants of fitness:
85 temperature and predation. Previous studies have documented that the response of *Daphnia*
86 populations to thermal stress is strongly correlated with environmental conditions and can
87 be either genetic, plastic or both [17–19]. Although these studies show plastic and
88 evolutionary changes, the co-occurrence of other environmental stressors, such as
89 predation, with temperature might interact in natural populations. Fish predation and
90 temperature impose selection on many of the same traits, and in the same direction [18,20].
91 For instance, *Daphnia* often mature at smaller sizes and younger ages in warmer water and

when fish are present, yet little is known about the molecular basis of this response among populations or their potential interactive effects.

We collected *Daphnia pulicaria* from two lakes in the Sierra Nevada Mountains, California, USA with different thermal conditions to conduct an experiment to evaluate the degree to which life history, thermal tolerance and gene expression are influenced by temperature and predation. We predicted that warm temperatures and exposure to predators should produce similar responses in life history traits, then used transcriptomics to ask whether the same genes underlie the plastic response to fish kairomones and warming, or if the similar phenotypic effects arise from pleiotropic effects where altered gene regulation affects multiple phenotypic traits. Our goal was to ask whether *D. pulicaria's* plastic response to one selective agent (fish or warming) magnified or dampened the response to the other in terms of both phenotype and gene expression.

Materials and methods

Lake sampling

Gardisky (37.955774, -119.251198) and Blue Lakes (38.051164, -119.270342), located in Inyo National Forest, California, USA, were sampled in late August and early September 2017, respectively. In both lakes we measured temperature throughout the water column at each meter using a field probe (YSI Incorporated, Yellow Springs, Ohio, USA). We also collected live zooplankton from the deepest point of the lake using a 30 cm diameter, 63 μm mesh conical net with a 1 m length through the water column, starting 1 m above the lake bottom. These samples were kept cold until returning to the laboratory, where we searched for *Daphnia pulicaria*. When present, 30 *D. pulicaria* females carrying

eggs in the brood pouch were separated in 50 ml centrifuge tubes filled with COMBO medium [21]. Each *Daphnia* individual was considered a maternal line. Each maternal line was cultured for at least twelve generations in separate 50 mL tubes filled with COMBO medium under ambient lab conditions. All *D. pulicaria* were fed non-viable cells of the green alga *Nannochloropsis* sp. (Brine Shrimp Direct, Ogden, Utah, USA) at a constant high rate of 24×10^6 cells every two days.

Experimental design

We randomly picked one clonal line from each lake to propagate. *Daphnia pulicaria* reproduces asexually under benign conditions. Thus, all individuals propagated asexually in the experiment. We started by randomly picking one mature female from each clone. Upon the release of the second clutch, we isolated three neonates that were separated and moved to separate 100 mL containers containing the same media and algae. All individuals were transferred to fresh media and algae three times a week and reared at 15 ± 1°C and photoperiod 12:12 h light/dark. Propagation continued until 100 neonates were collected for each of the clonal lines, with all individuals used for further study born within a 12-hour period. All neonates were placed into 100 ml jars containing COMBO medium at a density of 3 *Daphnia*/jar [21]. Each jar was randomly allocated to one of four treatments: (1) optimum temperature (15°C) without kairomones (fish cues), (2) high temperature (25°C) without fish cues, (3) optimum temperature with fish cues, and (4) high temperature with fish cues. An optimum temperature near 15°C is consistent with previous reports of thermal preference in *Daphnia pulicaria* [24], although there is variation in tolerable temperatures between *D. pulicaria* genotypes [25]. All *Daphnia* were transferred

138 to fresh medium, with algae (and kairomones in the fish cue treatment) daily. We
139 monitored jars daily for maturation (i.e., release of first clutch into the brood chamber).
140 Upon reaching maturity, individuals were either preserved in RNAlater (Qiagen, USA) and
141 kept in -20°C for subsequent RNAseq analyses or assigned to phenotypic assay (see below).
142 Since RNA was extracted from whole individuals, no food was added during the last 12
143 hours before sampling to minimize algal RNA contamination (most of which will be
144 digested and hence degraded after 12 hours). The period without food was kept short to
145 minimize starvation-dependent gene regulation.
146
147 *Kairomone collection*
148 COMBO medium conditioned by the presence of planktivorous fish was collected
149 daily from a tank containing 5 juvenile rainbow trout (*Oncorhynchus mykiss*; ~ 5 cm in
150 total length) in 72 L of COMBO. Each day, media containing fish chemical cues was
151 filtered through 0.7 µm mesh membrane filters and added at a concentration of 0.007 fish/L
152 to the predator treatments. This concentration of kairomone media is above the threshold
153 used by other studies to elicit a phenotypic response, such as the concentration of 5 µl of
154 kairomone media per 100 ml of solution [26].
155 *D. pulicaria* predation by *O. mykiss* in artificially stocked lakes is well established,
156 specifically predation by juvenile rainbow trout [27,28], *O. mykiss* is one of the most
157 abundant species for stocking previously fishless lakes in the Sierra Nevada mountains, the
158 source of our *Daphnia* samples [29]. *O. mykiss* were fed TetraMin Tropical Flakes (Tetra,
159 Blacksburg, Virginia, USA) prior to kairomone collection, rather than juvenile conspecific
160 daphniids as in some other studies [30,31]. We selected this method to minimize the

presence of conspecific alarm cues in the isolated kairomone media, so that we could measure the phenotypic and genetic responses to only predator-released, diet agnostic kairomone cues. Such research strategies are supported by Mitchell et al. [32], who call for further research to determine the relationship between predator odor and diet-based cues in aquatic systems. Following their nomenclature recommendations, we define the kairomones used in this study to be predator odor cues, as opposed to alarm cues released by injured conspecifics or diet cues released by predators after digesting conspecifics. All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of the University of California San Diego (IACUC protocol number: S14140).

Phenotype assays

Individuals from the experimental generation assigned to the phenotypic assay were scored for the following life-history variables: age and size at maturity, and age and number of offspring from the first three clutches. These data were used to calculate intrinsic population growth rate for each maternal line following the Lotka–Euler equation [33]. We also measured critical maximum temperature (CT_{max}) using a heat ramping assay. After a 30-minute resting period at ambient temperature, *Daphnia* from the four treatments were transferred to 0.5ml Eppendorf tubes and placed in a thermal heater (4x6 thermal heater, Corning Digital Dry Bath Heater Dual Block). The water temperature increased 0.1°C every 20 seconds. We continuously monitored the state of individuals and recorded the temperature when each individual *Daphnia* lost swimming ability and sank to the bottom of the tube. The temperature when *Daphnia* became immobilized was used as a proxy for

184 CT_{max} [17]. We measured life history traits of 11 to 13 individuals per treatment per lake
185 and CT_{max} of 10 to 12 individuals per treatment per lake. In total, we scored phenotypes of
186 186 individuals.
187
188 *Statistical analysis for phenotype*
189 We log-transformed all continuous variables, except intrinsic growth rate, after
190 visually assessing the probability distribution that best fit the data. We analyzed the effects
191 of the treatments using generalized linear models for each trait using the statistical software
192 R [34]. Temperature treatment, fish cue treatment, source lake and the interactions of these
193 variables were modeled as fixed effects for age and size at maturity, average number of
194 offspring, intrinsic growth rate and CT_{max}. Since we expected that CT_{max} would be
195 influenced by body size we included an additive effect of size.
196 Additionally, to test the effects of slopes of the reaction norms, we calculated the
197 pairwise differences for each dependent variable between each treatment combination
198 within a lake. We computed pairwise differences using the least square mean values based
199 on the generalized linear models for each trait using *lsmeans* function in the *lmerTest* R
200 package [35]. The pairwise difference test was corrected using Tukey's adjustment for
201 multiple comparisons [36].
202
203 *RNA isolation, library preparation and sequencing*
204 We extracted RNA using the TRI Reagent Protocol (Sigma-Aldrich, USA) from
205 samples preserved in RNAlater (Qiagen, USA). Any remaining genomic DNA was
206 removed using the TURBO DNA-free Kit (Invitrogen, USA). Each treatment included five

207 biological replicates, with each replicate comprised of 35 clonal *Daphnia* individuals.
208 Extracted RNA was stored in -80°C until sequencing. Quality of the isolated RNA was
209 assessed using RNA Nano 6000 Assay Kit of the Agilent Bioanalyzer 2100 system (Agilent
210 Technologies, CA, USA) and mRNA-seq libraries were constructed using NEBNext
211 UltraTM RNA Library Prep Kit for Ilumina (NEB, USA) following manufacturer's
212 recommendations. A total amount of 1 μg RNA per replicate was used as input material
213 for RNA-seq library preparations. Samples that did not reach 1 μg RNA were pooled with
214 a biological replicate grown under the same conditions, a process described by Takele
215 Assefa et al. [37], leaving 18 replicates from Blue Lake and 16 replicates from Gardisky
216 Lake. Index codes were added to attribute sequences to each sample and all 34 libraries
217 were clustered on a cBot System using the PE Cluster kit cBot-HS (Illumina). After
218 generating the clusters, libraries were sequenced using the Illumina HiSeq 2000 platform
219 and 150 bp paired-end reads were generated.
220
221 *Read cleaning and transcriptome comparison*
222 The raw reads were preprocessed using fastp v. 0.23.1 [38] with the command line
223 options "-x -f 18 -F 18". These options enabled polyX tail trimming and removed 18 low-
224 quality bases from the beginning of all forward and reverse reads, respectively. To check
225 for contamination in our read set, we used Kraken2 v. 2.0.9 [39] with a taxonomy database
226 constructed from the NCBI nr database [40] as of August 2021. A random subset of
227 1,000,000 read pairs was used for each Kraken2 run per sample. All downstream analyses
228 were based on the remaining cleaned, validated data.

229 Separate *de novo* assemblies for were performed on the Blue and Gardisky samples
230 using Trinity v. r20140413p1 [41] with parameter min_kmer_cov set to 2. Transcriptome
231 completeness was assessed using BUSCO v. 5.2.2 [42] with the arthopoda_odb10 lineage
232 dataset, and also by comparison to the following published, annotated genomes: *D.*
233 *pulicaria* LK16 [43], *D. pulex* TCO [44], *D. galeata* M5 [45], *D. magna* SK [46] and
234 *Eulimnadia texana* JT4(4)5-L [47].
235
236 *Differential expression analysis*
237 Relative expression levels of annotated genes were estimated by independently
238 mapping RNA-Seq reads from each lake sample back to the annotated *D. pulicaria* LK16
239 gene set using STAR v. 2.7.9a [48]. A read count matrix was generated from these
240 alignments using featureCounts v. 2.0.1 [49]. Genes with read counts below 10 across all
241 conditions were filtered from the remaining pipeline. The significance of difference in gene
242 expression of each treatment for each genotype was determined with the R package
243 DESeq2 v. 1.32.0 [50], using a model based on negative binomial distribution. Statistical
244 significance (p-value) was adjusted using the q-value obtained from the false discovery
245 rate [51], with a q-value < 0.05 and $|\log2(\text{foldchange})| > 1$ set as the threshold for
246 significant differential expression. Counts of differentially expressed genes across
247 treatments were visualized using the R package VennDiagram v. 1.7.1 [52]. A principal
248 component analysis (PCA) was visualized using read counts corrected through the variance
249 stabilizing transformation [53] function of DESeq2, The PCA was generated using
250 corrected read counts from every gene, rather than the DESeq2 default of using only the
251 500 most variable genes. These correct read counts were also used to perform a

252 PERMANOVA test [54] through the R package vegan v. 2.5-7 [55] with 999 permutations.
253 The groups tested with PERMANOVA were based on lake, temperature, fish cues, and
254 their interactions terms.

255 Gene Ontology (GO) enrichment analyses were performed to determine whether
256 significantly differentially expressed gene sets were enriched for any biologically relevant
257 functional categories [56]. GO term enrichment lists were generated using GOseq v. 1.44
258 [57] for the following one-factor comparisons: Blue, temperature rise with kairomones;
259 Blue, temperature rise without kairomones; Gardisky, temperature rise with kairomones;
260 Gardisky, temperature rise without kairomones. Further control of the False Discovery
261 Rate was performed using the Benjamini–Hochberg method [58], and the cutoff for
262 enriched GO terms was an adjusted p-value less than 0.05.

263

264 *Orthology of differentially expressed genes*

265 *Daphnia pulex* genes previously demonstrated to be differentially expressed in
266 response to temperature and/or predator kairomones were accessed from the *Daphnia*
267 Stressor Database [59] as of September 2021. Predicted protein sequences for these genes
268 were downloaded from wFleaBase [60]. These genes were compared to the annotated
269 *Daphnia pulicaria* LK16 genome using OrthoVenn2 [61] with an e-value of 1e-5.
270 OrthoVenn2 results were visualized using the R package VennDiagram v. 1.7.1 [52].

271

Results

Phenotype

Several traits in the two clones responded differently to temperature and fish cues (Table 1). The Gardisky Lake clone matured at a younger age than Blue Lake (Lake: $p < 0.001$, Table 1), but age at maturity was earlier for both clones in the presence of fish cues at high temperature (Temp x Fish: p-value = 0.006, Table 1, Fig 1). The post-hoc test (least square means) showed that fish had an effect only on the Blue Lake clone reared at 25°C. At 25°C, Blue Lake replicates reared with fish cues matured 0.55 ± 0.16 days earlier compared to replicates without fish cues (Fig 1).

Factor	**Df**	**Deviance**	**Deviance residuals**	**Pr (>Chi)**
Age at maturity				
Temperature	1, 184	16.0918	3.1609	**< 0.001**
Fish cues	1, 183	0.1340	3.0270	**0.002**
Lake	1, 182	0.3961	2.6309	**< 0.001**
Temp x Fish	1, 181	0.1036	2.5273	**0.006**
Temp x Lake	1, 180	0.0020	2.5253	0.7098
Fish x Lake	1, 179	0.0147	2.5106	0.3074
Temp x Fish x Lake	1, 178	0.0011	2.5096	0.7800
Size at maturity				
Temperature	1, 182	0.1078	0.9068	**< 0.001**
Fish cues	1, 181	0.00002	0.9068	0.9349
Lake	1, 180	0.2726	0.6341	**< 0.001**
Temp x Fish	1, 179	0.0054	0.6286	0.1989
Temp x Lake	1, 178	0.0382	0.5904	**< 0.001**
Fish x Lake	1, 177	0.0046	0.5858	0.2368
Temp x Fish x Lake	1, 176	0.0024	0.5833	0.3917
Average number of offspring				
Temperature	1, 87	2.7408	9.7072	**< 0.001**

Fish cues	1, 86	0.0403	9.6669	0.3436
Lake	1, 85	5.7215	3.9454	**< 0.001**
Temp x Fish	1, 84	0.0105	3.9349	0.6290
Temp x Lake	1, 83	0.0826	3.8524	0.1751
Fish x Lake	1, 82	0.0030	3.8494	0.7963
Temp x Fish x Lake	1, 81	0.2120	3.6374	**0.0297**
Intrinsic growth rate				
Temperature	1, 87	0.6194	0.0962	**< 0.001**
Fish cues	1, 86	0.0007	0.0954	0.3087
Lake	1, 85	0.0327	0.0627	**< 0.001**
Temp x Fish	1, 84	0.0000	0.0627	0.9713
Temp x Lake	1, 83	0.0003	0.0624	0.5032
Fish x Lake	1, 82	0.0003	0.0620	0.4853
Temp x Fish x Lake	1, 81	0.0014	0.0606	0.1665
Critical maximum temperature (CT_{max})				
Size at maturity	1, 91	0.0003	0.0256	0.1655
Temperature	1, 90	0.0082	0.0173	**< 0.001**
Fish cues	1, 89	0.0009	0.0163	**0.0125**
Lake	1, 88	0.0004	0.0159	0.1112
Temp x Fish	1, 87	0.0015	0.0144	**0.0019**
Temp x Lake	1, 86	0.0006	0.0138	0.0523
Fish x Lake	1, 85	0.0003	0.0134	0.1356
Temp x Fish x Lake	1, 84	0.0000	0.0134	0.5433

Table 1. Results of the generalized linear model analysis of the life-history traits of *Daphnia pulicaria*.

Clonal lines were collected from Blue and Gardisky Lakes and reared under different temperature (15°C or 25°C) and fish cue treatments (presence or absence).

Fig 1. Predation cues influence critical maximum temperature of Daphnia pulicaria.

Means ± 1 S.E.M. of age at maturity (days), size at maturity (mm), average number of

offspring of the first three clutches, intrinsic growth rate (r), and critical maximum temperature (CTmax, °C) of Daphnia pulicaria collected at Blue and Gardisky Lakes during summer 2017 in response to temperature (15°C and 25°C, x-axis) and predation cues treatments. Circles show response without fish cues and triangles response to fish cues.

Temperature affected size at maturity differently in the two populations (Temp x Lake: $p < 0.001$, Table 1) but fish cues had no main effect across clones (Fish: $p = 0.093$, Table 1, Fig 1). Post hoc tests revealed that the Blue Lake clone matured at 1.26 ± 0.006 mm regardless of temperature, while Gardisky matured at 15°C with 1.31 ± 0.01 mm compared to 1.41 ± 0.01 mm at 25°C.

The average number of offspring for the first three clutches showed a significant three-way interaction (Temp x Fish x Lake: $p = 0.029$, Table 1, Fig 1). For the Blue Lake clone, offspring number only varied with temperature; on average Blue had 1.1 ± 0.11 more offspring at 25°C compared to 15°C. In contrast, the effect of temperature on fecundity was greater for the Gardisky Lake clone when fish cues were present. Individuals had 1.67 ± 0.30 more offspring at 25°C than 15°C in the presence of fish cues, while without fish cues the difference between 25°C and 15°C response was 0.98 ± 0.23 offspring.

The intrinsic growth rate was significantly affected by the main effects of temperature and lake (Temp: $p < 0.001$ and Lake: $p < 0.001$, Table 1, Fig 1). At each temperature, the Gardisky Lake clone had higher intrinsic growth rates compared to the Blue Lake clone. For instance, the Gardisky clone had an $r = 0.15 \pm 0.001 * day^{-1}$ at 15°C, while Blue had $r = 0.11 \pm 0.004 * day^{-1}$. At 25°C, the intrinsic growth rate was 0.32 ±

313 0.006*day^{-1} for the Gardisky Lake clone, while the growth rate for the Blue Lake clone
314 was 0.28 ± 0.007*day^{-1}.
315 CT_{max} increased significantly when *Daphnia* were reared at 25°C compared to
316 15°C. CT_{max} was significantly affected by the interaction between temperature and fish
317 cues (Temp x Fish: p = 0.002, Table 1) and showed a marginally significant interaction
318 between temperature and lake (Temp x Lake: p = 0.052, Table 1), but no main effect of
319 lake. Post hoc tests showed that at 15°C, fish cues had no effect on CT_{max}. The thermal
320 tolerance of the Gardisky clone increased by 0.8 ± 0.12°C in the presence of fish cues,
321 while the Blue Lake clone increased by 0.3 ± 0.14°C at 25°C.
322
323 *Gene expression*
324 For the Blue Lake clone, sequencing produced on average 42,306,315 raw pair-ended reads
325 for each sample. Across all Blue Lake samples, 79.4% of the cleaned reads (after removing
326 adaptor sequences, low-quality and ambiguous sequences) were mapped to the D. pulicaria
327 LK16 primary gene set. The Gardisky clone produced on average 44,025,859 raw pair-
328 ended reads for each sample and 80.0% of the cleaned reads mapped to the D. pulicaria
329 LK16 primary gene set. A breakdown of read mapping rate and estimated read set
330 contamination by sample is visualized in S1 Fig. *De novo* assemblies of the Blue and
331 Gardisky clones show sequence similarity to previously annotated *Daphnia* genomes (S2
332 Fig) and high levels of completeness consistent with previously annotated arthropod
333 genomes (S3 Fig). A PCA plot generated using DESeq2-corrected read counts (Fig 2)
334 shows clear divergence in the gene expression between clones under all growth conditions.
335 A PERMANOVA test of gene expression data shows that gene expression was affected by

336 both source lake (Pseudo F = 9.6582, R^2 = 0.22284, p = 0.001) and temperature treatments
337 (Pseudo F = 4.367, R^2 = 0.10078, p = 0.001), but gene expression was not significantly
338 different between predation treatments (Pseudo F = 0.6706, R^2 = 0.01547, p = 0.746).
339
340 **Fig 2. Genotype, not treatment group, is the dominant source of differential gene**
341 **expression.**
342 Principal component analysis of gene expression patterns generated through DESeq2
343 prominently separates individuals by source lake along the first principal component.
344
345 Temperature treatments (25N vs. 15N and 25Y vs. 15Y) produced the greatest
346 number of differentially expressed genes (DEGs) for both the Blue and Gardisky clones
347 (Fig 3). A total of 320 genes were differentially expressed between one-factor comparisons
348 for the Blue Lake clone: 95 due to temperature differences with predation cues, 292 due to
349 temperature differences without predation cues, and 4 due to fish cues at 25°C (Fig 3a).
350 For the Gardisky Lake clone, we identified 575 DEGs. In total, 396 DEGs were specific to
351 25N vs. 15N and 52 DEGs were specific to 25Y vs. 15Y. These two treatment comparisons
352 shared 127 genes (Fig 3b). Predator cues alone had no measurable effect on gene
353 expression when comparing between Gardisky clones grown under the same temperature.
354 Tables listing all differentially expressed genes are presented in Supplementary Tables 1a
355 and 1b.
356

357 **Fig 3. Many genes are differentially expressed in response to temperature changes,**
358 **while kairomones have minimal observable effect at the level of transcript**
359 **expression.**
360 Venn diagram of differentially expressed genes for *Daphnia pulicaria* from **(a)** Blue and
361 **(b)** Gardisky Lakes in response to temperature and predation cues. Cyan and purple
362 sections illustrate differential expression from the introduction of predator cues (Y vs. N
363 for the presence and absence of fish cues), while the green and orange sections illustrate
364 differential expression due to temperature change. Treatment abbreviations: 15Y, 15°C
365 with kairomones; 15N, 15°C without kairomones; 25Y, 25°C with kairomones; 25N,
366 25°C without kairomones.
367
368 Comparisons of significant GO terms for differentially expressed genes were
369 difficult to interpret due to the limited specificity of the functional information they provide.
370 Genes related to proteolysis, chitin turnover, and vision appeared to be upregulated at
371 higher water temperatures, but no significantly over- or under-represented GO terms were
372 detected in response to kairomones. GO terms for phototransduction and visual perception
373 were enriched in all temperature comparisons, while protease and peptidase activity were
374 enriched in every comparison but 25Y vs. 15Y in the Blue clone. Meanwhile, terms
375 associated with chitin binding were enriched in every comparison but 25Y vs. 15Y in the
376 Gardisky clone. Complete lists of all enriched GO terms are given as Supplementary Tables
377 2a-2d.
378
379 *Gene orthology*

380 From the Daphnia Stressor Database, we retrieved 386 genes that responded to
381 kairomones in *D. pulex* across 7 studies, and 2660 genes that responded to temperature
382 changes in *D. pulex* across 4 studies. 315 of these previously documented kairomone-
383 responsive genes (81.6%) have orthologs in the *D. pulicaria* LK16 genome, while 1796 of
384 the temperature-responsive genes (67.5%) are orthologous to genes in *D. pulicaria* (Fig
385 4a). 35% of the temperature responsive *D. pulicaria* DEGs in this study are orthologous to
386 previously reported temperature-responsive genes from *D. pulex*, but only one of the four
387 genes in the Blue clone that responded to kairomones was orthologous to a previously
388 described kairomone-sensitive gene (Fig 4b). On the other hand, we observed that 18% of
389 described kairomone-responsive genes in *D. pulex* have orthologs that respond to
390 temperature in *D. pulicaria*, consistent with earlier phenotypic observations (Fig 1)
391 showing an interactive relationship between these variables. Lists of gene identifiers for
392 orthologous clusters and singleton genes are available in Supplementary Files 1 and 2.
393
394 **Fig 4. Gene orthology analysis reveals similarities between stress-responsive genes in**
395 ***D. pulicaria* and *D. pulex*.**
396 Venn diagrams of orthologous gene clusters between *D. pulex* DEGs from the *Daphnia*
397 Stressor Database and **(a)** the *D. pulicaria* genome or **(b)** DEGs from *Daphnia pulicaria.*
398 Numbers indicate protein families with at least two members, excluding "singletons" that
399 have no orthologous or paralogous matches. Gene identifiers for orthologous clusters and
400 singleton genes are available in **(a)** Supplementary File 1 and **(b)** Supplementary File 2.
401
402 **Discussion**

403 *Daphnia* clonal lines that were stressed by temperature and/or predator cues
404 exhibited similar phenotypic responses associated with distinctly different patterns of gene
405 regulation. Temperature was the most important factor driving changes in gene expression,
406 as most of the DEGs in both clones responded to this variable, versus only a handful to
407 kairomone cues. Yet fish cues magnified the effect of 10°C of warming on age of maturity
408 and increased the critical maximum temperature at which *D. pulicaria* lose motor control
409 when individuals were reared at high temperatures, but not at their optimum temperature.
410 Genetic pathways that underlie the plastic response to predators and temperature may be
411 subject to different regulatory pressures, but phenotypic responses suggest that exposure
412 to predator cues may increase thermal tolerance and affect life history response to warming.
413

414 *Phenotypic responses*

415 Both clones matured at a younger age and produced more offspring when reared
416 in high temperature, thus having a higher intrinsic growth rate compared to lower
417 temperature conditions (Fig 1). Indeed, high temperature increases metabolic rate, often
418 leading to a larger reproductive investment and faster development [62,63]. In addition, we
419 found an interactive effect of temperature and predation on age at maturity and critical
420 maximum temperature.

421 Our finding that individuals reared with predator cues had higher thermal
422 tolerance and matured younger at 25°C than those reared without predator cues agree
423 with other studies that also found synergistic effect of these two stressors in *Daphnia* and
424 other aquatic organisms [64–69]. In *D. pulex*, after seven generations, Tseng and
425 O'Connor observed increased thermal plasticity in individuals reared in higher

426 temperatures only when they were also reared with predators [64]. Zhang et al. tested
427 resurrected *D. magna* population from time periods without fish and from a high-density
428 fish period [65]. They found that *Daphnia* that coexisted with fish exhibited earlier
429 maturation in high temperatures compared to *Daphnia* from the pre-fish period. Earlier
430 maturation under predation risk is common in size-dependent predation of *Daphnia*
431 where predators, such as fish, prefer larger prey items, inducing a smaller size at maturity
432 and earlier age of first reproduction [63]. Our results agree with these studies showing
433 that fish cues magnify the life history response to temperature and show that exposure to
434 fish predation can increase the tolerance of stressfully high temperatures.
435
436 *Genetic responses to temperature*
437 We observed upregulation of serine peptidases in response to high temperature
438 treatment for both clones. These enzymes are the most important digestive proteases in *D.*
439 *magna* [70]. They are involved in proteolysis in the digestive system, the process by which
440 peptide bonds in proteins are broken generating free amino acids. This upregulation may
441 indicate a necessity to accommodate higher feeding rates caused by increased metabolism
442 in high temperatures [62,63]. Moreover, the *D. pulex* genome contains many peptidase
443 genes, which might indicate adaptation to high variation in food availability in aquatic
444 environment [71].
445 Genes related to visual perception were also upregulated in response to high
446 temperature treatment in both clones. Vision has previously been associated with vertical
447 migration, because visible light is a proxy for both *Daphnia's* visibility to predators and
448 the amount of incoming photodamage due to UV radiation at near-surface depths [72].

449 Vertical migration can serve as either a predator avoidance mechanism induced directly
450 from kairomones or a diel behavior triggered by temperature change [72]. Higher
451 temperatures can serve as a proxy for shallower depths, and therefore more incoming
452 radiation. This may explain why our dataset shows enrichment of phototransduction
453 genes as a response to temperature while previous studies have observed
454 phototransduction gene enrichment in response to kairomones [31].
455
456 *The missing kairomone response: behavioral changes and proximate cues*
457 In total, we only detected 4 DEGs across two *Daphnia pulicaria* clones in response
458 to kairomones from *Oncorhynchus mykiss.* Such a low number of DEGs related to
459 kairomone exposure is unexpected, but not unprecedented in previous *Daphnia* studies.
460 One study on *Daphnia galeata* M6 by Tams et al. also found 4 differentially expressed
461 transcripts in response to kairomones from the fish *Leuciscus idus* using a two-factor
462 analysis [73], and another study found no DEGs in *Daphnia magna* Iinb1 when exposed to
463 kairomones from stickleback fish [74]. Tams et al. hypothesize that life history changes
464 might be associated with only a few genes compared to morphological defenses, or that
465 regulation could be post-translational and therefore would not appear in an RNA
466 sequencing study [73]. Defense strategies can also include behavioral changes such as
467 vertical migration [75,76] and the transition into a state of alertness [77], which RNA
468 expression data may not fully capture. In contrast, morphological changes such as
469 neckteeth formation have been associated with far more genes, such as 230 DEGs found in
470 *D. pulex* in response to *Chaoborus* kairomones [78].

Miehls et al. find that the spiny water flea *Bythotrephes longimanus* displays no changes in life history or morphology when exposed to kairomones from the predator *Perca flavescens,* but increased temperature does induce morphologic defense against predation [79]. The authors hypothesize that temperature is a stronger predictor of predation risk than kairomones. This is because adult *P. flavescens* are present year-round but only juveniles are gape-limited in their consumption of *B. longimanus,* such that the morphologic response of increased tail length is only effective in discouraging predation from juveniles and not adult fish. If there is no difference between the kairomones released from juvenile and adult *P. flavescens,* then temperature could serve as a proximate cue for predation risk that could be damped through some phenotypic response.

The interactions between *B. longimanus* and *P. flavescens* closely mirror predation of *D. pulicaria* by *O. mykiss*, as *O. mykiss* are among the most abundant stocked fished in the Sierra Nevada lakes [29] and thus *D. pulicaria* are exposed to their kairomones year-round. Juvenile *O. mykiss* prey on *Daphnia* as a large percentage of their diet, while trout larger than 350 mm are primarily piscivorous [27]. A morphological defense response targeted solely at juvenile trout might require either a juvenile-specific odor, a dynamic response based on cue concentration, or some additional proxy for *O. mykiss* life stage that would suggest a high concentration of juveniles. Riessen and Gilbert propose that *D. pulex* may use water temperature to serve as a proximate cue for predator densities [80], so water temperature as a proxy for predator life stage is not implausible. Our dataset found some overlap between kairomone-responsive genes in *D. pulex* and temperature-responsive genes in *D. pulicaria,* so an intermediate defensive response may be inducible through heat alone as found by Suppa et al. [81].

494 However, behavior-based predator avoidance strategies may not require such
495 specificity. *O. mykiss* can produce the kairomone 5α-cyprinol sulfate in their bile without
496 consuming *Daphnia*, and this compound induces diel vertical migration in *D. pulex* [75].
497 Vertical migration still has an associated tradeoff in fitness [82], but this defense might be
498 preferred when food sources are abundant and temperature gradients are tolerable [83].
499 Another study found that *D. pulicaria* lineages that coexist with *O. mykiss* vertically
500 migrate without kairomone exposure, while *D. pulicaria* lineages without prior contact
501 with *O. mykiss* require kairomone exposure to induce diel vertical migration [84]. Thus,
502 some environments with high predation might select for *Daphnia* genotypes that respond
503 to predation primarily through behavioral responses, such as vertical migration.
504
505 *The missing kairomone response: predator- and diet-specific cues*
506 An alternative explanation for the lack of DEGs may lie in the subtle differences
507 between kairomone sources. Some kairomones are innately released by the predator
508 (predator odors), from damaged prey (alarm cues), and from digested prey excreted by the
509 predator (dietary cues). This study and Tams et al. explicitly avoided the addition of
510 conspecific alarm cues in our kairomone mixture as to measure only the effect of predator
511 odors [73]. Other studies, such as [30,31], tested responses to a mixture of predator-specific
512 odors and alarm cues released by consumed conspecifics. Mitchell et al. call for the
513 separation of nomenclature between cues released from the predator regardless of diet and
514 diet-specific cues [32], so that researchers can more accurately discern the independent
515 effects of these different kairomone compounds.

516 Previous research on the interaction between predator odors and alarm cues is
517 mixed. Some studies propose that defense systems rely on both types of compounds to
518 induce measurable defense responses [85,86], while others observe that alarm cues by
519 themselves can induce separate responses than predator odors alone [87,88]. Alarm cues
520 are thought to be nonspecific, as *Daphnia* respond differently to different predators [89]
521 and alarm cues alone might be unable to induce defense responses effective against a
522 specific predator [90]. The most effective defense responses may therefore require predator
523 odors to identify the type of predator combined with alarm cues to induce an immediate
524 response. A required mixture of predator odors and alarm cues for maximum observed
525 response in *D. pulicaria* would succinctly explain why we observed such a minor response
526 in our experiment. Results from this study support the conclusion that the detection of
527 predator odors alone may be insufficient to induce defense responses at the level of gene
528 expression, at least in the context of *D. pulicaria – O. mykiss* interactions.
529
530 **Conclusion**
531 Our study highlights that expression patterns of genes differed between *Daphnia*
532 *pulicaria* clones while phenotypic responses and interactions were qualitatively similar,
533 suggesting that different transcriptomic responses can result in similar phenotypes. Other
534 studies have found distinct gene expression patterns among individuals from different
535 zooplankton species [91,92]. These results suggest that diverse genetic pathways can give
536 rise to similar phenotypic plastic responses to environmental stress. We also showed
537 synergistic interactions between temperature and predation for some traits. Our results
538 show that *Daphnia* can have similar phenotypes through distinct molecular mechanisms

539 and the synergistic effects of temperature and predation may represent an important
540 mechanism for organisms to adapt to a rapidly changing environment. As one of the first
541 studies incorporating the *Daphnia pulicaria* genome, we hope to lay the groundwork for
542 future differential expression studies through incorporation into knowledge bases such as
543 the *Daphnia* Stressor Database. Overall, we find that the interactions between stressors are
544 heavily genotype dependent, and echo the sentiment that novel research strategies are
545 necessary [32] to further investigate the chemical signals of predation in *Daphnia*.
546

547 **Data Availability**

548 Read sets generated and/or analyzed during the current study are available on the
549 Sequence Read Archive (SRA) under BioProject PRJNA698341. The SRA Accession
550 numbers for the individual paired sets of reads associated with Blue Lake are
551 SRR13591461, SRR13591462, SRR13591463, SRR13591464, SRR13591465,
552 SRR13591466, SRR13591471, SRR13591482, SRR13591485, SRR13591486,
553 SRR13591487, SRR13591488, SRR13591489, SRR13591490, SRR13591491,
554 SRR13591492, SRR13591493, and SRR13591494, The SRA Accession numbers for the
555 individual paired sets of reads associated with Gardisky Lake are SRR13591467,
556 SRR13591468, SRR13591469, SRR13591470, SRR13591472, SRR13591473,
557 SRR13591474, SRR13591475, SRR13591476, SRR13591477, SRR13591478,
558 SRR13591479, SRR13591480, SRR13591481 ,SRR13591483, and SRR13591484.
559 Phenotype data, used to generated Fig 1, is included as S3 File.
560

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Supplementary Tables, Figures, and Files

S1 Tables 1a-1b. Data associated with Fig 3.

Differentially expressed genes in the **(a)** Blue Lake clone and **(b)** the Gardisky Lake clone. The comparison column denotes if genes were responsive to temperature and/or kairomone treatments.

S2 Tables 2a-2d. Gene Ontology of differentially expressed genes.

Upregulated Gene Ontology terms for comparisons between: **(a)** Blue Lake clone, 15Y and 25Y treatments, **(b)** Blue Lake clone, 15N and 25N treatments, **(c)** Gardisky Lake clone, 15Y and 25Y treatments, and **(d)** Gardisky Lake clone, 15N and 25N treatments.

S1 Fig. Read-based taxonomy assignment consistent with mapping rates to the *Daphnia pulicaria* genome.

Panel **(a)** shows Kraken2 read taxonomy on a random subset of one million cleaned reads from each sample. Panel **(b)** plots the alignment rate of the cleaned reads from each sample to the chromosomes of *Daphnia pulicaria* LK16 using the STAR aligner.

S2 Fig. Blue and Gardisky transcripts are highly similar to *Daphnia pulicaria* LK16.

Results of a two-way average nucleotide identity (ANI) matrix and associated dendrogram for *the de novo* assembled transcriptomes from Blue and Gardisky Lakes, predicted genes from *Daphnia* species with annotated genomes, and the annotated gene set of the related arthropod *Eulimnadia texana*. Our transcriptome sets are closest to those from *D. pulicaria*, within the well-described *Daphnia pulex-pulicaria* species complex.

S3 Fig. Blue and Gardisky transcriptomes are highly complete and highly duplicated.

Bar graph of gene set completeness as determined by BUSCO using the arthropod marker gene set. Our *de novo* assembled transcriptomes have completeness scores comparable to or better than genome-derived transcript sets from other *Daphnia* studies. However, Blue and Gardisky transcriptome sets have additional gene duplication in our transcriptome due to the inclusion of gene isoforms from *de novo* assembly.

S1 File. Orthologous clusters associated with Fig 4a.

Orthologous clusters and singletons for the gene orthology analysis between the *D. pulicaria* genome, temperature-responsive genes in *D. pulex*, and kairomone-responsive genes in *D. pulex*. *D. pulex* genes were collected from the *Daphnia* Stressor Database. Data is in Excel format.

S2 File. Orthologous clusters associated with Fig 4b.

Orthologous clusters and singletons for the gene orthology analysis between temperature-responsive *D. pulicaria* genes, kairomone-responsive *D. pulicaria* genes, temperature-responsive genes in *D. pulex*, and kairomone-responsive genes in *D. pulex*. *D. pulex* genes were collected from the *Daphnia* Stressor Database. Data is in Excel format.

S3 File. Phenotype and life history data associated with Fig 1.

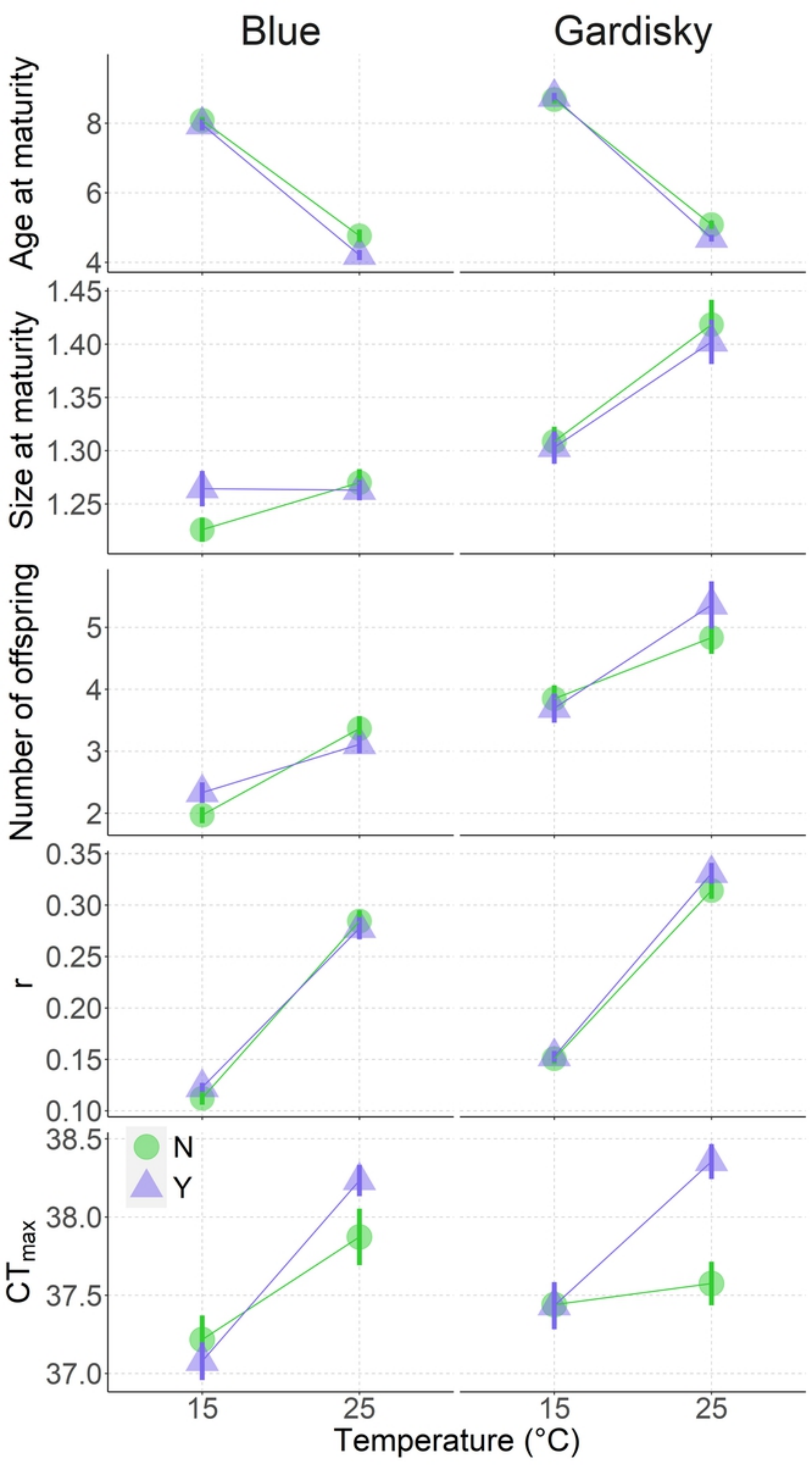
Blue
Gardisky
Age at maturity
8
6
4
Size at maturity
1.45
1.40
1.35
1.30
1.25
Number of offspring
5
4
3
2
r
0.35
0.30
0.25
0.20
0.15
0.10
CTmax
38.5
38.0
37.5
37.0
N
Y
15
25
15
25
Temperature (°C)

Figure1

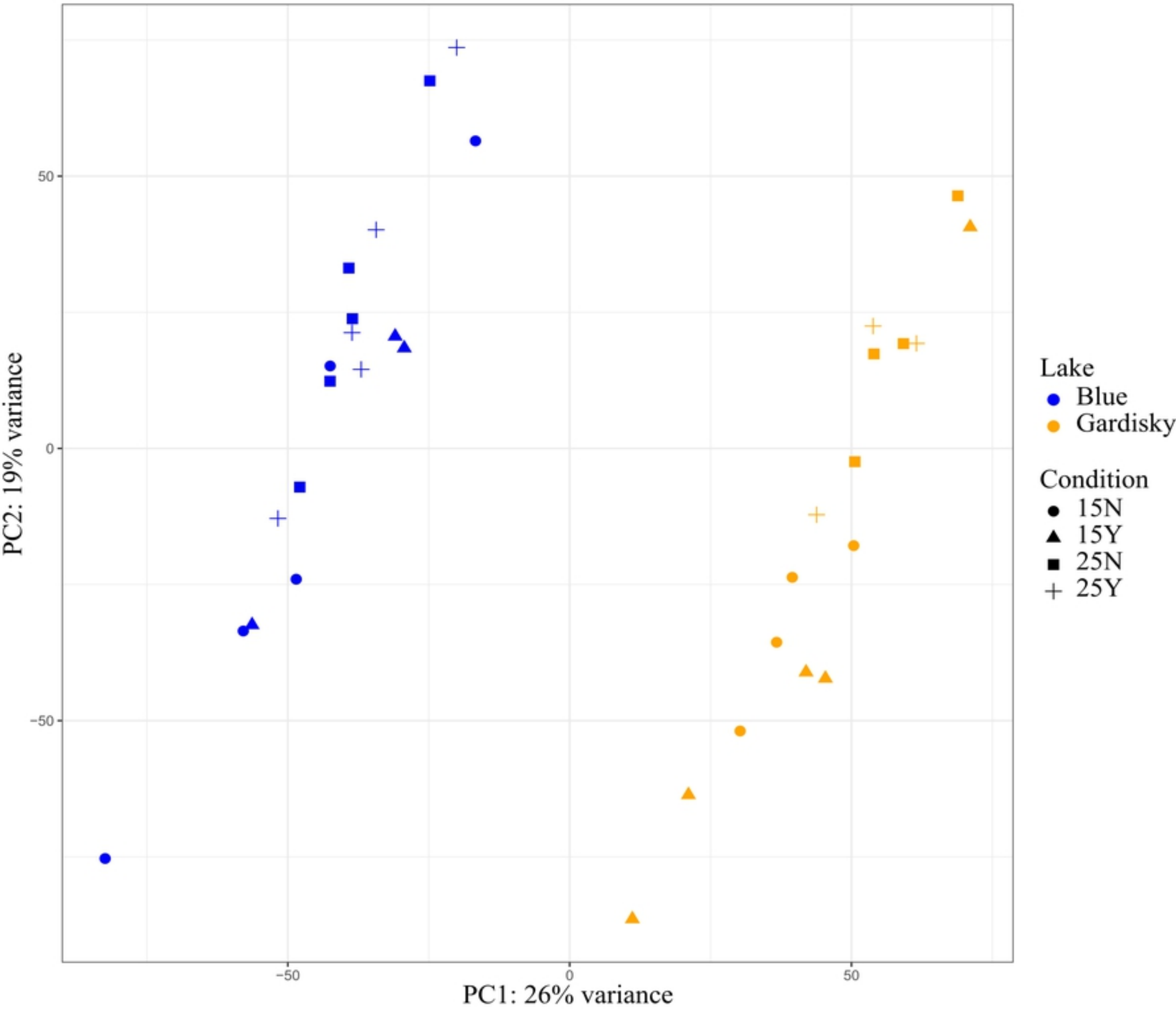
PC2: 19% variance
50
0
−50
−50
0
50
PC1: 26% variance
Lake
Blue
Gardisky
Condition
15N
15Y
25N
25Y

Figure2

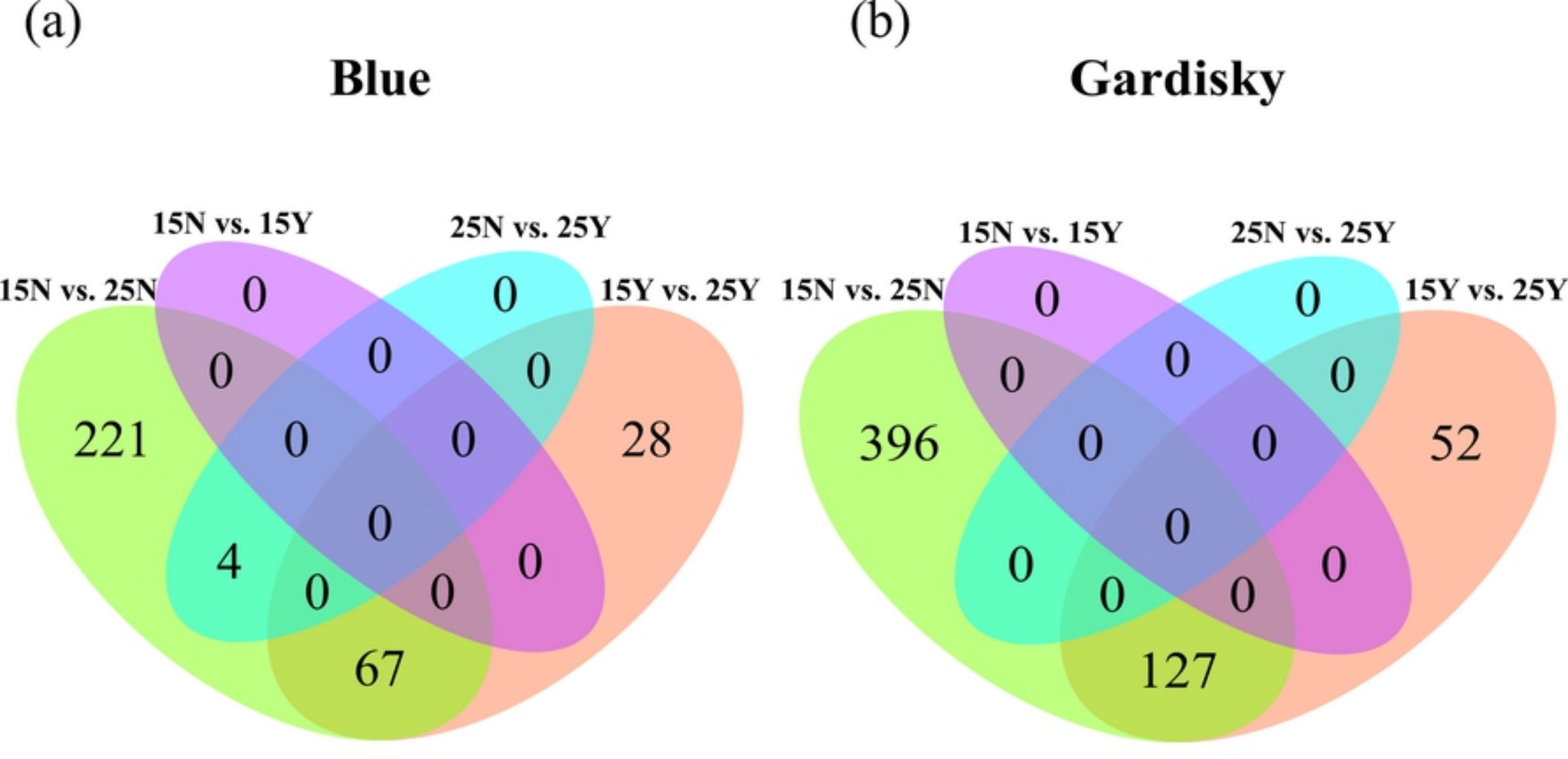
(a)
Blue
15N vs. 15Y
25N vs. 25Y
15N vs. 25N
15Y vs. 25Y
0
0
0
0
0
221
0
0
28
0
4
0
0
0
67
(b)
Gardisky
15N vs. 15Y
25N vs. 25Y
15N vs. 25N
15Y vs. 25Y
0
0
0
0
0
396
0
0
52
0
0
0
0
0
127

Figure3

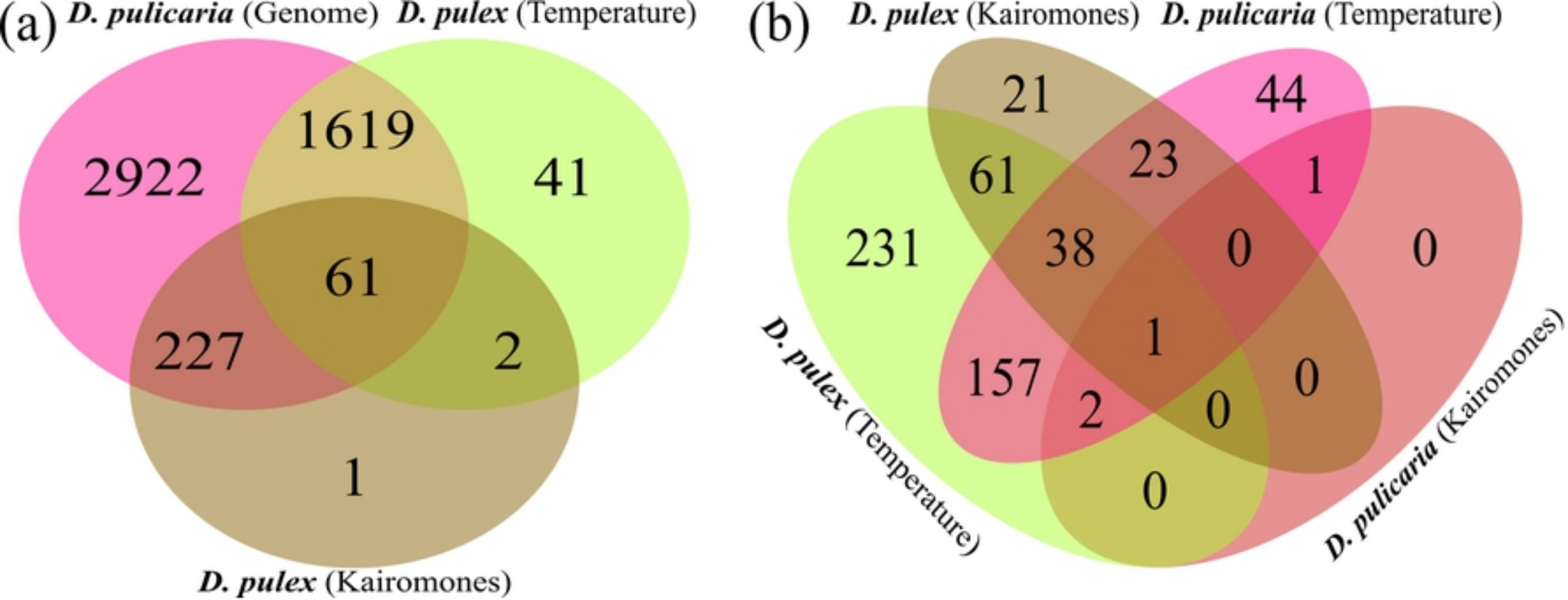
(a)
D. pulicaria (Genome)
D. pulex (Temperature)
D. pulex (Kairomones)
2922
1619
41
61
227
2
1
(b)
D. pulex (Kairomones)
D. pulicaria (Temperature)
D. pulex (Temperature)
D. pulicaria (Kairomones)
21
44
61
23
1
231
38
0
0
1
157
0
2
0
0

Figure4