The impacts of transfer learning, phylogenetic distance, and sample size on big-data

bioacoustics

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Running Title: Machine learning in bioacoustics

Abstract:

1. Vocalizations in animals, particularly birds, are critically important behaviors that influence their reproductive fitness, but automatically extracting vocalization data from existing large databases has only recently gained traction and has yet to be evaluated with respect to accuracy of different approaches.

2. Here, we use a recently-published machine learning framework to extract syllables from six bird species ranging in their phylogenetic relatedness from 1–85 million years, comparing how phylogenetic relatedness impacts accuracy as well as the utility of applying trained models to novel species.

3. Model performance is best on conspecifics, with accuracy progressively decreasing as phylogenetic distance increases between taxa; however, using models trained on multiple distantly related species can recover the lost accuracy.

4. When planning big-data bioacoustics studies, care must be taken in sample design to maximize sample size and minimize human labor without sacrificing accuracy.

Keywords: bioacoustics, birds, machine learning, phylogenetics, transfer learning

Introduction:

Avian bioacoustics

Vocalization is an important form of communication across many taxa. Birds (class Aves) make and respond to vocalizations profusely, for example in territory defense (de Kort et al., 2009), mate choice (Nowicki & Searcy, 2005; O'Loghlen & Rothstein, 1995; West et al., 1981), both conspecific and heterospecific communication (Magrath et al., 2015; Riebel et al., 2019), species discrimination (Gill & Lanyon, 1964; Derryberry, 2011), prey detection (Rice,

1982), and food solicitation via begging calls (Wright, & Leonard 2002; Catchpole & Slater,

1995). Bird vocalizations are generally classified into songs and calls. Significant attention has

been paid to song vocalizations, which are thought to mediate sexual selection (MacDougall-

Shackleton, 1997; Tinghitella et al., 2018; Burdfield-Steel & Shuker, 2018) and be important in

the generation of reproductive isolation in many bird taxa (Brambilla et al., 2008; Jenkins, 1978;

Uy et al., 2018; Dingle et al., 2010). Though less prominent, bird calls are also important as they

can mediate non-sexual behaviors (Kondo & Watanabe, 2009), for instance between parents and

offspring (Jiang et al., 2021) or even intra-specific communication (Keen et al., 2020). Despite

evidence that both the biotic and abiotic environment can impact the development and evolution

of vocalization, there are few studies that examine how the community soundscape evolves in

tandem (but see Mason et al., 2017; Freeman et al., 2022).

The sheer volume of recording data available to modern researchers represents a

tremendous asset to avian bioacoustic research. For example, the citizen-science initiative Xeno-

Canto (xeno-canto.org) having ~700,000 recordings and the Macaulay Library bioacoustics

repository (which includes eBird, Sullivan et al., 2009) having ~1.1 million recordings

(birds.cornell.edu/MacaulayLibrary). Even the smaller bioacoustic collections, which represent

important repositories of historical recordings, can have tens of thousands of songs to parse (e.g.,

the Borror Laboratory of Bioacoustics with ~36,000, blb.osu.edu). This volume of data

represents a methodological challenge because most bioacoustic workflows require the songs to

be segmented (Catchpole & Slater, 1995), either identifying them from background sounds or,

for more complex analyses, identifying individual syllables of song (e.g., Du & Troyer, 2009;

Alosnso et al., 2017).

Ideally, automated methods would be useful if they could reduce the amount of human

interaction without compromising data quality, as song segmentation is extremely time

consuming (e.g., Swiston et al., 2009 found manual annotation to take nearly five times as long,

even after accounting for needing to check automatic annotations), though less so than the gold-

standard methods of performing playback experiments to directly evaluate behavioral responses

to different vocalization treatments (e.g., Provost et al., 2018 performed ~4,600 minutes of

playback experiments to investigate a single species).

Machine Learning

Machine learning, which can be either unsupervised or supervised, is a useful tool for

data processing. Unsupervised machine learning is typically used to cluster data (Reynolds &

Rose, 1995, Greenspan et al., 2006, Edgington et al., 2006), while supervised learning is

frequently used to make predictions (Heileman & Myler, 1990). Many well-known algorithms in

ecology and evolution are derived from machine learning models in some capacity, for example

linear regression, k-means clustering (Lloyd, 1982), and MaxEnt niche modeling (Phillips et al.,

2006, Phillips et al., 2017). However, many of these techniques require inputs that are

specifically curated, such that the presence or absence of features and variables must be

determined beforehand, which can lead to a slew of problems.

More recently, deep learning via artificial neural networks (ANNs) has become popular.

ANNs are made from many layers of nodes which construct weighted mathematical functions to

perform tasks, typically from some kind of input data with diagnostic features (LeCun et al.,

2015). Deep learning methods are methods where the features used do not need to be pre-trained.

Because ANNs can iteratively learn their own features, use large amounts of data, and are less

constrained by assumptions about that data, they are extremely flexible and can handle many

kinds of tasks (e.g., Hopfield. 1982; Ripley. 1996, Sprengel et al., 2016; Zell, 1994; O'Mahony

et al., 2020; LeCun et al., 2015; Alom et al., 2018). These methods have been used on a variety

of topics including image processing, video segmentation, and speech recognition (Hemanth &

Estrela, 2017; Valipour & Wang, 2017; Kamath et al., 2019). Although challenges remain with

respect to scalability, computational efficiency, and how to handle depauperate data (Alom et al.,

2018), deep learning is one of the most powerful analytical tools in the modern researcher's

toolbox, particularly when human knowledge is lacking, or datasets are too large to be workable

by traditional means. In the context of ecology and evolutionary biology, there have been many

recent applications of both shallow and deep machine learning, including population genetics

and phylogeography (e.g., Schrider & Kern, 2018; Provost et al., 2021), bioacoustics (e.g.,

Nicholson, 2016; Zhong et al., 2020; Cohen et al., bioRxiv), species classification (e.g., Gupta et

al., 2021), phylogenetics (e.g., Halgaswaththa et al., 2012; Wang et al., 2016), sequencing and

genomics (e.g., Adrion et al., 2020; Boža et al., 2017), and phenotypic analyses and

morphometrics (e.g., Devine et al., 2020; Lürig et al., 2021). Neural networks and support vector

machines tend to be the most applied algorithms towards these analyses.

Though there are many ways to categorize machine learning algorithms, but for our

purposes dividing these into evaluative vs extractive algorithms is convenient. Evaluative

algorithms are typically used to make predictions from data, for example by training a model to

distinguish between simulated evolutionary scenarios (Pudlo et al., 2016; Sheehan & Song,

2016; Schrider & Kern, 2018; Gehara et al., bioRxiv; Provost et al., 2021). Extractive

algorithms, on the other hand, are designed to process data in some way. With respect to

bioacoustics, an extractive algorithm could, for example, segment out syllables within

vocalizations. A recently developed application, TweetyNet, was released to perform just this

task (Cohen et al., bioRxiv) using deep learning via ANNs. Specifically, TweetyNet uses

convolutional and recurrent ANNs. As such, it is one extractive algorithm that could be useful in

reducing the human workload in bioacoustics. However, though potentially much faster than

hand-processing data, ANNs like TweetyNet can still take prohibitively long times to train,

though they often only need to be trained once.

Minimizing training time can be done by co-opting existing trained ANNs to perform

similar tasks, known as transfer learning. This can be used to take a neural network trained on

one batch of data, and then applying it to new, related data (Alom et al., 2018; Weiss et al.,

2016). For example, a neural network that is trained to identify the leaves of oak trees from

images may be able to be co-opted to identify leaves of maple trees from images without having

to re-train the model to recognize a generic leaf shape. As an example, which is more applicable

for bioacoustics, a neural network that is trained to identify the song of one species against

background noise may be able to be applied to a different species.

Here we compare different treatments of bioacoustic data to optimize the overall

performance of machine learning models trained to segment syllables of avian song. We

investigate the utility of these methods across a subset of bird species, including birds from

different clades ranging in divergence times from ~1 to ~85 million years. We use those taxa to

evaluate how well models perform on closely vs distantly related taxa for use in maximizing

computational efficiency. Finally, we provide suggested practices for moving forward with

neural networks in avian bioacoustics.

Materials and Methods:

Unless otherwise noted, all analyses were performed using a MacBook Pro running

macOS Catalina (10.15.7) with a 2.8 GHz i7 core and 16 GB of RAM.

Acquiring and filtering song data

We downloaded bioacoustics data from two song databases: Xeno-Canto (xeno-

canto.org), and the Borror Laboratory of Bioacoustics (blb.osu.edu). We used the Xeno-Canto

database as it has a large amount of data of varying quality, due to it being a citizen science

initiative, and a convenient R package exists for accessing and filtering the files. The Borror Lab

of Bioacoustics database was used because it is full of historical data and based in North

America, the region we wished to focus on, in addition to being the lab where the authors are

based (Figure 1).

For the Xeno-Canto songs, we targeted songs from *Cardinalis sinuatus*, *Cardinalis*

cardinalis, *Zonotrichia leucophrys*, *Empidonax virescens*, and *Calypte anna*. We chose these

taxa because they were distributed across the phylogenetic tree and had relatively high numbers

of recordings available. We also targeted songs from *Melozone fusca* which we used to evaluate

the performance on a novel species. We restricted the data downloaded to all of those in North

America that were of Quality "A" according to the Xeno-Canto database using the package

warbleR version 1.1.26 (Araya-Salas & Smith-Vidaurre, 2017) in R version 4.0.3 (R Core Team

2020). Xeno-Canto recordings are available in the MP3 format, but since our downstream

pipeline requires WAV format, we converted all of them using the package tuneR version 1.3.3

(Ligges et al., 2018). For the Borror Laboratory of Bioacoustics songs, we downloaded all

available data from all species, which were already available as WAV format. Stereo recordings

were converted to mono using the 'mono' function in tuneR. We then retained recordings that

had a sample rate of 48,000 Hz, which was the most common sample rate in our dataset, because

downstream analyses require recordings of the same sample rate and will not work on stereo

recordings.

Figure 1: Flowchart describing the procedure from acquiring song recordings to having

segmented syllables for downstream analysis. Boxes show general steps, arrows show flow of

steps, and diamonds show decision point for data quality.

Identification of syllables via manual and automatic annotation of song data

We arbitrarily chose a subset of recordings to annotate from each species, starting with

Zonotrichia leucophrys which had the largest dataset, and moving down to *Calypte anna* which

had the smallest dataset. As few as three minutes of recording is sufficient for the TweetyNet

algorithm to perform accurately (Cohen et al., bioRxiv), so we ensured that all species had at

least 180 seconds worth of annotated syllables. We then added more recordings for species with

more data to investigate the influence of sample size. For *Melozone fusca*, we deliberately

annotated (and trained on) a disproportionately small number of songs to generate artificial

scarcity of data. In both the Borror Lab of Bioacoustics and the Xeno-Canto database, many

species have a small number of recordings. For the former, which primarily sampled North

American taxa, nearly 9,400 bird species lack any data, with an additional ~400 species with

fewer than 10 recordings. For the latter, ~600 species lack any data, with an additional ~2,500

with fewer than 10 recordings (Figure 2). As such, having methods which can handle species

with small numbers of recordings is critical.

Figure 2: Most species of bird have little bioacoustic data. X-axis shows, on a log scale, the

number of songs present per species. Y-axis shows the number of species per song bin. Top

panel: Xeno-Canto data. Bottom panel: Borror Laboratory of Bioacoustics (BLB) data; note the

x-axis has a gap to show an exceedingly high number of species with no data, as BLB is North

America focused.

Annotations were performed in Raven Pro version 1.6.1 using a spectral window size

(i.e., Fast Fourier Transform size) of 512 samples, which for a sample rate of 48,000 Hz was

~9.97 milliseconds. We drew boxes representing frequency and time boundaries for every

syllable in the recording of the focal species. These annotations were exported as text files using

Raven's built-in selection format. After annotating the songs, we then subsetted the recordings

such that individual recordings had approximately equal amounts of annotated song and silence.

Preliminary results showed that when TweetyNet was trained on full files, which had a high

proportion of non-annotated "silence", TweetyNet would fail to identify the songs and would

only predict silence, but with high accuracy. As such, we wrote a script using tuneR in R which

automatically sliced WAV files and their corresponding annotation files for training (see Data

Availability). Annotations that were separated by at least 1.0 second of non-annotated sound

were split. Splits were made such that at least 50% of the resulting file was annotated: in cases

where this was not possible, a larger percentage of the file may have been annotated. We then

converted the annotation files into XML files that TweetyNet could use following the

"Koumura" format (following Koumura & Okanoya, 2016). All annotations were given the label

"1".

Training of machine learning model using TweetyNet

When training TweetyNet, we set our spectrogram parameters as follows: Fast Fourier

Transform size of 512, step size of 32 samples, frequency cutoffs of 500 and 10,000 Hz, a log

transformed spectrogram, and a minimum power log threshold of 6.25. We also used a window

size of 88 spectral windows and normalized spectrograms while training. Unless otherwise

noted, we used an 80:10:10 split for training, validation, and test data, and tracked the total

training time for each model. These splits were generated manually for each species. Each model

used a batch size of three and was validated every 50 steps. Checkpoints were taken every 200

steps. We ran for 10 epochs (i.e., 10 passes the machine learning algorithm made over the full

training dataset) but stopped training prematurely if the model had gone 50 checkpoint steps

without an improvement in accuracy. We then used the maximum accuracy checkpoint as our

final trained model. Some models were restarted if they were killed before completing 50

checkpoints without improvement. Models that failed this process multiple times were restarted

and ran as needed. Models were run sequentially (i.e., one at a time).

We trained three sets of models: single-species, multi-species, and transfer learning. For

the single-species models, we trained a model in TweetyNet on a given species to 5-step and 50-

step checkpoints. We did this once for each of our five species with sufficient data, resulting in

ten total single-species models. For the multi-species models, we trained a model in TweetyNet

as above but using a combination of the five species with sufficient available data. We did this

for two models: an unbalanced and a balanced model. For the unbalanced, we included all the

training data from each of the five species irrespective of the sample sizes for each. For the

balanced model, we subset the training and validation data used such that each species had the

same amount of training and validation data (in seconds) as our species with the smallest sample

size, to mitigate any effects of biased sampling. Each was trained to 5-step and 50-step

checkpoints, resulting in four total multi-species models.

To incorporate transfer learning for use in the transfer models, we wrote a custom module

for TweetyNet ("train_checkpoint") which was able to continue training from an existing

checkpoint file from a previously completed trained model using new training data from a

different taxon. At the time of writing, this functionality has been planned but not executed in

TweetyNet's source code. After training both multi-species models as above, we then used that

finished checkpoint to train on data from *Melozone fusca*, which we manipulated to use a

relatively small amount of data (aka "few-shot learning"; Wang et al., 2019). Instead of training

for 50 checkpoints which would likely lead to overfitting, we trained for 5 checkpoints only.

Though we had a comparable amount of data for this species compared to the other five, we

generated artificial scarcity by only using ~30% of this as training and validation data, with the

remainder acting as test data. This resulted in four total transfer-learning models depending on

whether we trained from the balanced or unbalanced multi-species model, and whether we

trained from the 5-step or 50-step checkpoint of those models.

We tested the single-species and multi-species models on annotated data from each of the

five species we trained on with sufficient available data (*Zonotrichia*, *Cardinalis*, *Empidonax*,

and *Calypte* species; either zero-shot or normal training) as well as the species with artificially

insufficient available data (*Melozone fusca*; zero-shot training). For the transfer models, we only

tested on *Melozone fusca* (few-shot training). In all cases, we did this testing to assess the

accuracy of the model. We evaluated accuracy across species with respect to the predicted

genetic distance. In addition, we evaluated the performance with respect to different transfer

methods, the number of species present in the training dataset, the balancing approach for multi-

species models, and the impact of zero-shot vs few-shot vs normal training.

For some analyses we examined the phylogenetic distance between trained species and

test species. We mined these data from timetree.org (Kumar et al., 2017), taking the estimated

time listed as the time of divergence in millions of years (Mya). Intraspecific comparisons were

given an estimated time of 0.0 Mya; we also did this for the transfer learning done with

Melozone fusca. For the multiple-species models, we calculated the weighted average estimated

divergence time based on the sample sizes for each species, again taking conspecifics as 0.0

Mya.

Song diversity

We evaluated song diversity to determine whether this impacted our machine learning

model accuracy. Song diversity was evaluated in two ways: first, we consulted an analysis that

focused on Passeriformes which measured song complexity (Medina & Francis, 2012). Three of

the species had complexity analyzed in that study (*Zonotrichia leucophrys*, *Cardinalis*

cardinalis, *Cardinalis sinuatus*) which were calculated by performing a PCA on song properties

(e.g., number of notes, frequency bandwidth). We directly used those values as our complexity

measure. Our second metric of song diversity was done by estimating the hypervolume

complexity. To do this, we took the 5 longest WAV files for each species that had been manually

annotated. We then put them through the SoundShape pipeline (Rocha & Romano, 2021).

SoundShape aligns and normalizes syllables of sound to make them directly comparable with

respect to their size and shape. After aligned and normalized syllables were converted to TPS

format, we randomly selected 50 syllables per species and calculated a principal components

analysis on their values, retaining the first three principal components (PCs). We then used the

three PCs to calculate a hypervolume per species, as a percentage of the total hypervolume

occupied by all species. We repeated this 50 times and then calculated the mean percentage

hypervolume occupied per species as our complexity metric. Species with more diverse syllable

types, and therefore more song diversity, should have larger hypervolumes.

Impact of various metrics on accuracy and time of training

To investigate what aspects of our models determined accuracy and training time, we

used generalized linear regressions as well as ANOVA and Tukey's Honest Significant

Differences tests (Chambers et al., 1992; Miller, 1981; Yandell, 1997) using the stats package in

R. For all models, we took the log values of training time and sample size, as these varied by

orders of magnitude across taxa and models. Unless otherwise specified, we set our alpha level

to 0.05 to determine significance, and we applied linear models for 5-step and 50-step checkpoint

runs independently.

We tested the relationship between the size of the training dataset and the amount of

training time using a linear regression. We did this both with and without transfer learning

models. We also tested the relationship between accuracy and phylogenetic distance, accuracy

and song complexity, and/or accuracy and hypervolume diversity as linear regressions both for

single-species models only and for all models. The linear regressions involving song complexity

and hypervolume diversity were run calculating these metrics for the testing species as well as

the training species. Finally, we used an ANOVA to determine whether individual species

differed in accuracy as well.

Results:

Song data summary

Our dataset included 6,563 *Zonotrichia leucophrys* songs, 918 *Cardinalis cardinalis*

songs, 36 *Cardinalis sinuatus* songs, 128 *Empidonax virescens* songs, 102 *Calypte anna* songs,

and 72 *Melozone fusca* songs. We filtered out songs that did not have a sample rate of 48,000 Hz

and then annotated 197 *Zonotrichia leucophrys* songs, 61 *Cardinalis cardinalis* songs, 8

Cardinalis sinuatus songs, 22 *Empidonax virescens* songs, 15 *Calypte anna* songs, and 12

287 *Melozone fusca* songs for each species. After subsetting files to balance the relative amount of

288 annotated sound to silence, this resulted in total datasets of 4,843 seconds for *Zonotrichia*

289 *leucophrys*, 1,083 seconds for *Cardinalis cardinalis*, 401 seconds for *Cardinalis sinuatus*, 291

290 seconds for *Empidonax virescens*, 316 seconds for *Calypte anna*, and 272 seconds for *Melozone*

291 *fusca* (see Supplementary Table S1).

294 Table 1: Models trained in TweetyNet across multiple species vary in training time, dataset size,

295 and type of mode. "5 Species" models were trained on all data from the "Single species" type

296 models. "Balanced" models were trained on equal amounts of data from each of the "Single

297 species" type models.

Species Trained On	Training Time, 5 checkpoints (sec)	Training Time, 50 checkpoints (sec)	Training : Val : Test (sec)	Type
Calypte anna	310	4,506	256:29:31	Single species
Cardinalis cardinalis	377	12,464	867:108:108	Single species
Cardinalis sinuatus	468	7,484	320:40:41	Single species
Empidonax virescens	647	7,941	231:31:29	Single species
Zonotrichia leucophrys	1,999	44,694	3878:483:482	Single species
5 Species	4,214	25,863	5553:691:692	Multiple species
5 Species Balanced	557	10,348	1130:150:671	Multiple species
5 Species (5) + *Melozone fusca*	665	n/a	71:9:192	Transfer
5 Species Balanced (5) + *Melozone fusca*	553	n/a	71:9:192	Transfer
5 Species (50) + *Melozone fusca*	341	n/a	71:9:192	Transfer
5 Species Balanced (50) + *Melozone fusca*	246	n/a	71:9:192	Transfer

299   Table 2: Accuracy of models trained in TweetyNet across species, excluding *Melozone fusca*.

300   Accuracy values are given for models checkpointed at 5-steps and 50-steps. Estimated

301   divergence time for "5 Species" models is a weighted average, proportional to the sample sizes

302   used for each species trained on.

Species Trained On	Species Tested On	Accuracy 5-step checkpoints	Accuracy 50-step checkpoints	Estimated Divergence (Mya)
Calypte anna	*Calypte anna*	0.56	0.60	0.0
Calypte anna	*Cardinalis cardinalis*	0.71	0.67	85.0
Calypte anna	*Cardinalis sinuatus*	0.76	0.74	85.0
Calypte anna	*Empidonax virescens*	0.63	0.69	85.0
Calypte anna	*Zonotrichia leucophrys*	0.82	0.82	85.0
Cardinalis cardinalis	*Calypte anna*	0.50	0.52	85.0
Cardinalis cardinalis	*Cardinalis cardinalis*	0.67	0.91	0.0
Cardinalis cardinalis	*Cardinalis sinuatus*	0.87	0.92	6.6
Cardinalis cardinalis	*Empidonax virescens*	0.70	0.88	66.0
Cardinalis cardinalis	*Zonotrichia leucophrys*	0.57	0.87	38.0
Cardinalis sinuatus	*Calypte anna*	0.67	0.62	85.0
Cardinalis sinuatus	*Cardinalis cardinalis*	0.87	0.89	6.6
Cardinalis sinuatus	*Cardinalis sinuatus*	0.92	0.92	0.0
Cardinalis sinuatus	*Empidonax virescens*	0.80	0.79	66.0
Cardinalis sinuatus	*Zonotrichia leucophrys*	0.69	0.81	38.0
Empidonax virescens	*Calypte anna*	0.59	0.55	85.0
Empidonax virescens	*Cardinalis cardinalis*	0.65	0.71	66.0
Empidonax virescens	*Cardinalis sinuatus*	0.73	0.76	66.0
Empidonax virescens	*Empidonax virescens*	0.93	0.97	0.0
Empidonax virescens	*Zonotrichia leucophrys*	0.65	0.73	66.0
Zonotrichia leucophrys	*Calypte anna*	0.52	0.57	85.0
Zonotrichia leucophrys	*Cardinalis cardinalis*	0.82	0.76	38.0
Zonotrichia leucophrys	*Cardinalis sinuatus*	0.82	0.81	38.0
Zonotrichia leucophrys	*Empidonax virescens*	0.86	0.92	66.0
Zonotrichia leucophrys	*Zonotrichia leucophrys*	0.89	0.92	0 .0
5 Species	*Calypte anna*	0.68	0.77	81.1
5 Species	*Cardinalis cardinalis*	0.87	0.89	33.6
5 Species	*Cardinalis sinuatus*	0.87	0.91	34.2
5 Species	*Empidonax virescens*	0.90	0.94	64.1
5 Species	*Zonotrichia leucophrys*	0.90	0.91	14.8
5 Species Balanced	*Calypte anna*	0.69	0.76	68.9
5 Species Balanced	*Cardinalis cardinalis*	0.86	0.90	39.7
5 Species Balanced	*Cardinalis sinuatus*	0.87	0.92	39.9
5 Species Balanced	*Empidonax virescens*	0.86	0.95	57.6
5 Species Balanced	*Zonotrichia leucophrys*	0.81	0.88	41.3

Training time of the single species models to 5 checkpoints ranged from 310 seconds

(~5.2 minutes, *Calypte anna,* 256 seconds of song) to 1,999 seconds (~33.3 minutes, *Zonotrichia*

leucophrys, 3,878 seconds of song; Table 1). For 50 checkpoints, these times ranged from 4,506

seconds (~75.1 minutes or ~1.3 hours) to 44,694 seconds (~744.9 minutes or ~12.4 hours). The

multiple species models took 4,214 seconds (~70.2 minutes or ~1.2 hours) for the unbalanced

(5,553 seconds of song) and 557 seconds (~9.3 minutes) for the balanced (1,130 seconds of

song) to reach 5 checkpoints. For 50 checkpoints these numbers were 25,863 seconds (~431

minutes or ~7.2 hours) for unbalanced and 10,348 seconds (~172.4 minutes or ~2.9 hours) for

balanced. The four transfer learning models, which were all trained for 5 checkpoints each, took

between 246 seconds (~4.1 minutes) and 665 seconds (~11.1 minutes) to train on the *Melozone*

fusca data (71 seconds). Less additional training time was required when training using the 50-

step checkpoints than the 5-step checkpoints, though total training time was higher. The

relationship between the size of the training dataset and the amount of training time needed is

positive on a log scale for both the 5-step and 50-step checkpoint runs; this holds even after

excluding transfer learning models which were only run for 5-step checkpoints (adjusted R^2

range=0.51–0.77, p<0.021; Table 1).

Accuracy in the 50-step checkpoint runs was nearly always equal or higher than accuracy

in the 5-step checkpoint runs. Across all 42 pairs of single-species and multiple-species models,

32/42 had higher accuracy in the 50-step checkpoint (difference range=1–30%), 3/42 had the

same accuracy, and 7/42 had higher accuracy in the 5-step checkpoint (difference range=1–6%).

For models that were trained on single species, accuracy was generally highest when

models were tested on the same species that they were trained on (5-step range=89–93%; 50-step

range=91–97%), except for *Cardinalis cardinalis* in the 5-step checkpoints (in which accuracy

was highest for its congener *Cardinalis sinuatus*, 87% vs 67% for *Cardinalis cardinalis*) and

Calypte anna in both 5-step and 50-step checkpoints (where highest accuracy was also for

Cardinalis sinuatus, 62–67% vs 56–60% for *Calypte anna*).

Accuracy dropped off in a manner that scaled to the predicted phylogenetic distance in

the models that were not trained and tested on the same taxa (i.e., zero-shot training), with loss of

0–5% for congeneric comparisons (plus one gain of 20% in *Cardinalis cardinalis*), 4–32% for

taxa with the same oscine/suboscine status (plus one gain of 15% in *Cardinalis cardinalis*), 2–

24% for comparisons between birds in the same order, and 3–30% between passerines and non-

passerines (with gains of 2–11% in *Cardinalis cardinalis* and *Calypte anna*). This relationship is

significantly negative with respect to estimated phylogenetic distance for both 5-step and 50-step

models (adjusted R^2 range=0.27–0.35, p<0.0002; Figure 3).

Most of the multi-species models perform the same or worse at classifying a species than

the corresponding single-species model trained on that species (0–8% loss in accuracy across all

taxa), except for the *Cardinalis cardinalis* and *Calypte anna* models mentioned above (12–20%

gain in accuracy) and one *Zonotrichia leucophrys* 5-step checkpoint model (with a 1% gain).

However, multi-species models tend to perform better than zero-shot learning on other single-

species models (improvement of 0–33%) with a few exceptions that are worse (by 1–3%).

Multiple species models perform better than expected for their mean estimated divergence time:

of the 24 tests done across the models, 21 are above the line of best fit and three are below

(Figure 3).

Comparing the 12 unbalanced and balanced multi-species model pairs, the differences in

performance within species ranged from 4% better for balanced models to 9% better in

unbalanced models, with 8/12 of those pairs differing by less than 1%. *Zonotrichia leucophrys*,

the most over-represented species, has the unbalanced models performing much better (by 3–

9%).

There is no significant association between sample size of either training or test data and

average accuracy across single-species models (log sample sizes, p-value range=0.33–0.53),

although when multiple-species and transfer learning models are included, there is a slight

positive association with test data sample size and accuracy (p-value range=0.009–0.03, adjusted

R^2 range=0.09–0.12). However, *Calypte anna* vocalizations are significantly less likely to be

classified accurately than other species (p-value<0.03). Notably, this is the only non-passerine in

the dataset and has a very different type of song than the others.

Figure 3: Model performance by species drops with estimated divergence, but training on

multiple species performs better than expected given average divergence time. X-axis gives the

estimated divergence time. Y-axis gives the accuracy. Lines are drawn to connect points in

panels B through F. Points outlined with large squares were trained on the "5 Species" model.

Points outlined with large circles were trained on the "5 Species Balanced" model. Shades of red

indicate models run for 5-step checkpoints. Shades of blue indicate models run for 50-step

checkpoints. A) all data, excluding *Melozone fusca* (see Figure 4). Colored lines show the line of

best fit. B) *Calypte anna.* C) *Cardinalis cardinalis*. D) *Cardinalis sinuatus*. E) *Empidonax*

virescens. F) *Zonotrichia leucophrys*.

Table 3: Accuracy of models trained in TweetyNet for *Melozone fusca*. Accuracy values are

given for models checkpointed at 5-steps and 50-steps. Estimated divergence time for "5

Species" models is a weighted average, proportional to the sample sizes used for each species

trained on.

Trained On	Tested On	Accuracy 5-step checkpoints	Accuracy 50-step checkpoints	Estimated Divergence (Mya)
Calypte anna	*Melozone fusca*	0.77	0.80	85.0
Cardinalis cardinalis	*Melozone fusca*	0.81	0.91	38.0
Cardinalis sinuatus	*Melozone fusca*	0.91	0.91	38.0
Empidonax virescens	*Melozone fusca*	0.86	0.91	66.0
Zonotrichia leucophrys	*Melozone fusca*	0.88	0.91	8.6
5 Species	*Melozone fusca*	0.91	0.94	20.8
5 Species Balanced	*Melozone fusca*	0.91	0.93	43.8
5 Species (5) + *Melozone fusca*	*Melozone fusca*	0.95	n/a	0.0
5 Species Balanced (5) + *Melozone fusca*	*Melozone fusca*	0.94	n/a	0.0
5 Species (50) + *Melozone fusca*	*Melozone fusca*	0.95	n/a	0.0
5 Species Balanced (50) + *Melozone fusca*	*Melozone fusca*	0.93	n/a	0.0

Figure 4: Model performance on *Melozone fusca* drops with estimated divergence, but also

shows recovery when trained on multiple species. X-axis gives the estimated divergence time. Y-

axis gives the accuracy. Lines are drawn to connect points. Points outlined with large squares

were trained on a "5 Species" model. Points outlined with large circles were trained on a "5

Species Balanced" model. Large black squares and circles with cross-marks indicate transfer

models. Shades of red indicate models run for 5-step checkpoints. Shades of blue indicate

models run for 50-step checkpoints.

Zero-shot vs few-shot learning

        With respect to the small *Melozone fusca* dataset, classification of this species' songs is

overall highly accurate. For the single-species models, the least accurate was that of the distantly

related *Calypte anna* (72–80%). For the other species, the 50-step checkpoint models all perform

equivalently (91% accuracy) while in the 5-step checkpoint models, accuracy ranges (81–91%

accuracy). For the multi-species models, the unbalanced model (91–94%) performed slightly

better than the balanced model (88–93%), and slightly better than the closest related species

Zonotrichia leucophrys (88–91%), suggesting that having a model trained on a more diverse

subset of sounds improves performance of zero-shot learning. Lastly, the few-shot learning

transfer models perform the best of them all with little additional training time (93–95%),

suggesting that having a single multi-purpose model rich in data to "seed" the training of species

with few data may be beneficial.

Song diversity results

Song complexity values (from Medina & Francis, 2012) ranged such that *Cardinalis*

cardinalis had the lowest complexity, followed by *Cardinalis sinuatus,* and then *Zonotrichia*

leucophrys with the highest complexity. In contrast, our calculated hypervolume diversity metric

finds that instead *Cardinalis sinuatus* has the smallest hypervolume, followed by *Empidonax*

virescens, *Melozone fusca*, *Cardinalis cardinalis*, *Zonotrichia leucophrys*, and finally *Calypte*

anna with the largest hypervolume. The latter two species occupy much higher percentages of

hypervolumes compared to the remaining four species. With such a small number of points (N=3

for complexity from Medina & Francis, 2012; N=6 for hypervolume estimates) we do not have

the power to detect any correlations between the two values, but we expect that complexity and

diversity should be correlated.

Neither complexity (Medina & Francis, 2012) nor mean hypervolume of the species used

for training are significant predictors of accuracy (p>0.11). Likewise, song complexity (Medina

& Francis, 2012) of the species used for testing is not a significant predictor of accuracy

(p>0.45). However, the mean hypervolume of the species used for testing is negatively

associated with accuracy, such that the more diverse a species' song is, the less accurate the

model is at predicting it ($p < 0.0002$, adjusted $R^2 > 0.28$).

Table 4: Species vary with respect to song complexity and hypervolume diversity. Complexity

scores range from lower complexity (more negative) to higher complexity (more positive).

Hypervolume diversity is a percentage of total hypervolume across all six species, with means

and standard deviations from 50 independent runs of 50 random syllables per species.

Species	Complexity[1]	Hypervolume Mean ± SD
Calypte anna	n/a	0.5000 ± 0.1600
Zonotrichia leucophrys	-0.695	0.2400 ± 0.1100
Cardinalis cardinalis	-1.798	0.0140 ± 0.0100
Melozone fusca	n/a	0.0055 ± 0.0040
Empidonax virescens	n/a	0.0041 ± 0.0032
Cardinalis sinuatus	-1.086	0.0028 ± 0.0037

[1]from Medina and Francis 2012

Discussion:

Numerous initiatives have been undertaken over the past decades to accumulate and

archive biological information into searchable and accessible databases. Such efforts include

databases of morphological characteristics (MorphoNet; Leggio et al., 2019), localities for use in

modeling abundance and niche (GBIF; gbif.org), genetic, genomic, and protein sequences

(GenBank; Clark et al., 2016), and of course bioacoustics. Taking full advantage of all these

datasets requires increases in efficiency, including development of automated retrieval of data as

well as data extraction.

Bioacoustic data such as those we have used here have traditionally been parsed

manually, since spectrograms were developed in the 1950s (Borror & Reese, 1953). These

methods have required large amounts of time, both to extract the usable data but also to verify

and validate the homologous aspects of songs for comparisons within and between taxa. Songs

must be searched for the actual sounds, and then researchers must decide which parts of the

sound are considered unique syllables, before even beginning to classify them by which type of

syllable is being produced. Often this necessitates deep understanding of the species in question,

and while this approach is inherently valuable and has provided the foundation for avian

bioacoustics, it may preclude more broad understanding of vocalizations outside of those focal

taxa. Automated methods promise to improve the efficiency of bioacoustics by identifying data

more readily and by automatically calculating differences between sounds (e.g., Rocha &

Romano, 2021). Despite these improvements, the attributes and limitations of these methods are

not as well understood in comparison to more traditional manual analyses. One aspect that has

been investigated is the amount of time spent: processing songs in automated ways can be up to

~5 times faster compared to manual methods (Swiston et al., 2009).

In terms of performance, training a model on a species and applying it to that same

species appears to be the most accurate method of extracting syllables, provided that the sample

size is sufficiently high. This is irrespective of whether the training dataset includes only that

species or also includes other taxa (e.g., the 5-species models). However, the range in accuracy

varies highly across taxa. Species with more simple vocalizations (e.g., *Empidonax*) appear more

likely to be classified with high accuracy than species with more complex vocalizations (e.g.,

Zonotrichia). We speculate this is due to a combination of factors. Taxa with more complex

vocalizations must optimize the detection algorithm for multiple different kinds of syllables at

once, which is expected to be more difficult than categorizing fewer kinds of syllables. Our

models were run under the same parameters and for the same training times, though they were

allowed to exit early if they reached an accuracy plateau; given this, it is likely that the

convolutional neural network can solve a "simpler" problem in the same amount of time more

efficiently. Another consideration, however, is sample size given to the model. Although we did

not find an explicit correlation between sample size and accuracy, it is nevertheless possible that

our high accuracy on these species is due to over-fitting, rather than any biological aspect of the

song. However, we do not attribute these differences to learning vs. non-learning behavior;

although *Empidonax* are the only suboscine (non-learning) Passeriform bird in our dataset, there

is not a relationship between song complexity and oscine/suboscine status in Passeriformes

(Pearse et al., 2018).

We find that model accuracy drops with phylogenetic distance between the training data

and tested data. This suggests that there is commonality between closely related species in terms

of their song; indeed, evidence across passerines suggests that song is phylogenetically

constrained, in addition to being influenced by morphology and ecology (Pearse et al., 2018;

Mikula et al., 2021). From these results we would predict that any similar learning algorithm

should behave in this fashion with traits that are associated with phylogeny. This pattern may

also extend to species with convergent song features as well, though this remains to be tested.

Though the sample size does not appear to impact accuracy, it is an important

consideration when trying to decide how to best extract data from species. With sufficiently large

sample sizes (TweetyNet suggests 3 minutes of annotation, Cohen et al., bioRxiv), techniques

like convolutional neural networks will work effectively to learn and parse the diversity of

syllables and sounds present in these data. However, for some taxa the available data are not

sufficient for these types of learning. In this case, we have two recommendations depending on

the study system. If a study only concerns one single species without a large amount of data, it

would likely be best to manually annotate all the data available, rather than try to fit a

computationally expensive model to it. On the other hand, if a study is focused on multiple

species at once, especially if some of those species have a large sample size, we recommend

training a model on multiple different species, optionally then using transfer learning techniques

to fine-tune the model. The caveat with this latter approach is that the benefit of this learning

may decrease as phylogenetic distance (and song dissimilarity) increases with respect to your

focal taxa.

We find that transfer learning from multiple species models substantially increases

accuracy for taxa without a large amount of data. However, this might also be partially due to

overfitting of very small datasets. Transfer efficiency seems to be just as good when balanced vs

unbalanced. We recommend balancing, however, because it appears to perform just as well with

significantly less data and thus less training time. In terms of computational efficiency, we find

that training a model with multiple different species takes longer than training a model on each

species separately. We suspect this is because the algorithm needs to optimize segmentation of

many disparate kinds of syllables, much like with species that have more complex songs

compared to species with fewer songs. However, we also find that models trained on multiple

species perform better than models trained on single species when applied to a never-before-seen

taxon, even after considering average phylogenetic distance between training species and testing

species. Despite the increase in computation time, having a more general-purpose, multi-species

model trained can be worthwhile for researchers looking to maximize the number of taxa they

can segment syllables for, while minimizing the amount of manual annotation and training time

they need.

Downstream applications and implications

The approach we develop here is fast, flexible, and can be run on a standard laptop,

though performance will benefit from parallel processing. As such, our application can

supplement the deep avian bioacoustics literature with the breadth afforded by efficient data

generation. In addition, our overall framework will likely be applicable to other machine learning

algorithms: TweetyNet could be replaced by other convolutional/recurrent neural network

methods with these guidelines still being relevant. It remains to be seen whether other methods,

including random forests or evolving neural networks, will perform similarly (Knapp et al.,

2017; Juang & Chen, 2007; Ranjard & Ross, 2008; Neal et al., 2011).

Automatic methods can be used to validate and supplement manual segmentation

analyses, as are traditional in avian bioacoustics. From the most basic, this approach can be used

with sparser forms of data, for instance continuous recordings where the species of interest only

vocalizes for a short amount of time. Segmentation with this method and downstream scripts we

have developed here can reduce the amount of recording that needs to be searched by quickly

eliminating large portions of the files. This segmentation method can also be used to standardize

projects among individuals who are all working concurrently. Human biases in segmentation are

a known phenomenon (Tachibana et al., 2014; Ranjard et al., 2015; Milinski, 1997), but applying

a single model to begin from would potentially alleviate some of those issues. Further, when

working from automatically segmented data, the ability to recognize incorrect or otherwise

anomalous syllables where models and manual labeling do not match would make data cleaning

swifter and more reliable. Lastly, the use of these models can help assist in training younger

scientists without potentially compromising data quality; with a standard template to work from

and the ability for the machine learning algorithm to partially check classifications, students

could spend more time developing other research skills than simple song annotation.

Automatic segmentation can be useful not just to reduce the workload of people, but also

to set up for downstream applications. First, though not implemented here, TweetyNet can not

only identify the presence of syllables, but also individual types of syllables; indeed, this is

TweetyNet's original goal, and it does so with high accuracy. However, this requires knowledge

of what types of syllables exist in the dataset, and in taxa that have not been studied before, it is

hard to know *a priori* what those sounds are. However, an algorithm that can broadly detect

sounds can thus lead into automatic classification of syllables via clustering analyses, which exist

already in easily implemented fashions (e.g., K-means clustering, Lloyd, 1982). It is beyond the

scope of this manuscript to discuss clustering methods but being able to automatically

characterize the repertoire of a new species is within the realm of possibility. Further, any

methods that can segment the syllables as well as classify types of syllables allow for the fine

detection variation within those syllable classes, as well as changes in the syntax of bird song

(i.e., the arrangement of sounds into specific sequences). This contrasts with changes in other

features of song, like frequency, duration, or bandwidth, which are independent of the order of

specific syllables. Regardless, the ability to categorize these sounds will be particularly useful in

getting a more thorough assessment of song complexity within and between taxa.

Segmentation of sound is a critical part of analysis of song in birds, as well as other taxa

that communicate with sound including some insects, frogs, and bats. Given the sheer number of

species that this encompasses, having a standardized and automatic approach will allow for fast

performance of traditional bioacoustic analysis. Our method forms a starting point for this work.

Given enough time, we foresee that these and other models can be further refined and used to

segment all recordings available for all species, which would likely comprise multiple lifetimes

of work if done manually and be intractable with the current speed at which new recordings are

added. Further, it is not outlandish to think that as machine learning methods become more

sophisticated, software could be developed to auto-segment sounds as they are cataloged into

these databases and repositories, or even to be integrated into the recorders and microphones that

are commonly used today.

Conclusions

In this paper, we investigate the influence of phylogeny on machine learning methods to

extract syllables from recordings of bird song, finding that the accuracy of these methods

depends in part on the phylogenetic relatedness of the taxa being trained on and tested on.

However, the loss in accuracy associated with distantly related taxa can be ameliorated by

including multiple species. Further, species with smaller sample sizes can be classified with high

accuracy by tuning previously trained models. We suggest the following best practices for using

machine learning in bioacoustics, depending on the sample sizes of species of interest: for very

small amounts of total data, it is best to hand-annotate syllables to segment them irrespective of

the number of species. With larger amounts of data, hand-annotation of select species that span

the phylogenetic (and bioacoustic) variation in the dataset, then training a machine learning

model to segment syllables from it will reduce computational load without a major disruption to

accuracy. This framework will be broadly applicable to many regions of the world and many

taxa, allowing us to achieve a global perspective on vocalizations. Ultimately, this innovation

will change how biologists analyze songs by expanding the scope of what is possible from depth

to breadth.

Author Contributions:

KLP and BCC conceived the study; KLP and JY collected the data; KLP performed the analyses,

wrote custom code, and wrote the initial manuscript. All authors edited the manuscript.

Acknowledgements:

We thank all the individuals throughout the years who have contributed to the bioacoustics

repositories, including the Borror Laboratory of Bioacoustics and Xeno-Canto. For helpful

feedback on these analyses, we thank D Kelly, T Berger-Wolf, G Carter, R Adams, and S Gaunt.

Thoughtful reviews come from T Yuri, S Decker, F Mol Lanna, D Parsons, J Wieringa, D

Duckett, S Moshier, EM Da Fonesca, and MTC Thome. KLP and BCC were supported by NSF

DEB-2016189.

Conflict of Interest Statement:

The authors declare no conflicts of interest.

Data Availability:

Data used will be archived on Dryad. Bioacoustic data are publicly available from Xeno-Canto

and the Borror Laboratory of Bioacoustics. Code is available on GitHub at

github.com/kaiyaprovost/bioacoustics.

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