

# Phenotypic Variation from Waterlogging in Multiple Perennial Ryegrass Varieties under Climate Change Conditions

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## Abstract

Identifying how various components of climate change will influence ecosystems and vegetation subsistence will be fundamental to mitigate negative effects. Climate change-induced waterlogging is understudied in comparison to temperature and CO<sub>2</sub>. Grasslands are especially vulnerable through the connection with global food security, with perennial ryegrass dominating many flood-prone pasturelands in North-western Europe. We investigated the effect of long-term waterlogging on phenotypic responses of perennial ryegrass using four varieties grown in atmospherically controlled growth chambers (ambient vs 2050, +2°C and eCO<sub>2</sub>) during two months of peak growth. Using image analysis and PCA methodologies, we assess how multiple predictors (phenotypic, environmental, genetic and temporal) influence overall plant performance and productivity. Long-term waterlogging was found to reduce leaf-colour intensity, with younger plants having purple hues indicative of anthocyanins. Plant performance and yield was lower in waterlogged plants, with tetraploid varieties coping better than diploid ones. The climate change treatment was found to reduce colour intensities further. Flooding was found to reduce plant productivity via reductions in colour pigments and root proliferation. These effects will have negative consequences for global food security from facing extreme weather events and flooding. Our approach can be adapted as plant health diagnostics tools via remote sensing and drone-technology.

## Keywords

Anthocyanins, Chamber Experiment, Climate Change, Colours, Harvest, Image Analysis, *Lolium perenne*, Waterlogging

## 1. Introduction

Predicting plant responses caused by climate change is a fundamental challenge that will increasingly impact coming generations. Identifying how the various components of climate change are likely to affect plant responses will inform options of optimal mitigation strategies to minimise any negative effects (Canadell and Raupach, 2008; Pareek et al., 2020). While this is a complex issue affecting all ecosystems, it is especially important for grassland ecosystems due to the unavoidable connection with global food security through agriculture, crop lands and pasture lands (Kipling et al., 2016a; Lecerf et al., 2019; Raza et al., 2019; Tester and Langridge, 2010). There is a growing concern for the biodiversity, health, productivity and ecological transformation of grasslands from the changing climate (Goliński et al., 2018; Jentsch et al., 2009; Kipling et al., 2016b; Qi et al., 2018; Wang et al., 2018).

Grasslands are defined as ecosystems dominated by the Poaceae (grass) taxonomic family, with the ecology varying widely depending on species composition, edaphic factors, topography, management and climate. Previous studies have suggested that elevated CO<sub>2</sub> levels are likely to increase photosynthetic capability in grasses, increasing Net Primary Production (NPP) and thereby total yield (Ergon et al., 2018; Yiotis et al., 2021). Meanwhile, higher temperatures are likely to extend the length of the growth season, providing longer time for sustained growth (Höglind et al., 2013; Pembleton et al., 2020). While increased ambient temperatures and elevated CO<sub>2</sub> act long-term and are the main components of many climate models, altered precipitation regimes are equally important, and predicted to act both short- and long-term (Brown et al., 2019; Cullen et al., 2009; Dore, 2005). The main focus of previous grass research from altered precipitation regimes has been the effects of prolonged droughts, reduced water-availability and increased desertification (Bothe et al., 2018; Buttler et al., 2019; Cullen et al., 2014; Farfan-Vignolo and Asard, 2012; Yates et al., 2019), however many areas might see the opposite effect, leading to increased flooding (Kiely, 1999; Rosenzweig et al., 2002).

Climate change-induced alterations to precipitation regimes are expected to contribute to increased return-intervals of extreme weather events, such as severe storms and extreme flooding (Easterling et al., 2000; Semmler and Jacob, 2004). This is partly due to warmer temperatures increasing evaporation, and warmer air being able to hold more moisture, increasing the total amount of water vapour in atmospheric circulation (Hu et al., 2000; O’Gorman and Muller, 2010). Additionally, predicted changes to the precipitation seasonality are likely to change the frequency and distribution of rain events, potentially enhancing the drought-flooding dichotomy (Feng et al., 2013; Kumar, 2013).

Perennial ryegrass (*Lolium perenne*) is a common cool season pasture grass grown extensively throughout its native Eurasian-range and cultivated worldwide due to its high nutritional quality and palatability for livestock (Hannaway et al., 1999; Hunt and Easton, 1989; Minneé et al., 2019; Smit et al., 2005; Tubritt et al., 2020). Many genetically different varieties of perennial ryegrass are bred and cultivated to match the climatological conditions of specific target regions, which in turn causes differences in plant health and yield depending on local environmental suitability (Grogan and Gilliland, 2011; Helgadóttir et al., 2018). Varieties suitable in current conditions under regular precipitation regimes might prove unsuitable under periods of increased flooding (Mustroph, 2018). Flooding can cause long-term waterlogging, affecting overall plant health and total yield (McFarlane et al., 2003; Striker, 2012) and grassland ecosystem function (Fay et al., 2008). It could also impact plant morphology due to phenotypic plasticity (Mizutani and Kanaoka, 2018; Münzbergová et al., 2017). Decrease in growth due to an altered phenotypic response can have devastating impacts on global food security due to the bottom-up reliance of agricultural system productivity from grassland areas (Baldos and Hertel, 2014; Hazell and Wood, 2008). In addition, economic consequences would be especially severe for countries like Ireland and the United Kingdom with large land areas consisting of ryegrass dominated pasture lands and many rivers currently prone and predicted in the future to flood (Blöschl et al., 2019).

Here, we quantify the effects from waterlogging on perennial ryegrass performance and plant health in the light of climate change. We hypothesised that waterlogging would decrease perennial ryegrass performance and lower plant yield. We further explored whether waterlogging impacted the phenotypic plasticity of the plants. We investigated this using atmospherically controlled growth chambers and multiple commercial high-producing perennial ryegrass varieties with varying genetic backgrounds in an image analysis framework.

## **2. Materials and Methods**

### **2.1. Experimental Setup**

This study used four atmospherically controlled CONVIRON BDW40 walk-in growth chambers located in the PÉAC (Programme for Experimental Atmospheres and Climate) facility in Rosemount Environmental Research Station belonging to the University College Dublin in Dublin, Ireland. This facility has been used in previous studies to investigate plant responses to elevated CO<sub>2</sub> (Batke et al., 2018; Yiotis et al., 2021), atmospheric paleoclimatic reconstruction (Evans-Fitz.Gerald et al., 2016; Porter et al., 2017; Yiotis et al., 2017). Two

chambers were chosen to represent typical North-western European climatological conditions (CO<sub>2</sub>-levels at 415 ppm) while two chambers were chosen to represent predicted 2050 climate change climatological conditions with elevated CO<sub>2</sub>-levels (550 ppm) and a 2°C increase in temperature (IPCC, 2014). The growth chamber climatological baselines were constructed from the last thirty years of meteorological data (1989 – 2018) collated from the meteorological station located at Cork Airport and publicly accessible via the Irish Meteorological Services (Met Éireann). The entire experiment simulated conditions from May to September but only climatological conditions replicating two months of optimal pasture growth (June and July) (McHugh et al., 2020) were used to investigate perennial ryegrass responses to waterlogging (**Table 1**). Four common internationally grown varieties of perennial ryegrass were used for the experiment: *Aberchoice*, *Abergain*, *Carraig* and *Dunluce* (European Commission, 2019). The varieties vary in heading date and ploidy (**Table 2**), which allowed for an intra-species comparison to identify whether genetic factors might contribute to the response to waterlogging. At the simulated start of May each chamber was populated with 80 PVC cores (50 cm x 16 cm (∅)) filled with John Innes No2 compost (320 cores in total for the four chambers). Each core was sealed inside a plastic bag to allow half of the cores to be waterlogged further on. The 50 cm tall cores allowed for the simulation of largely natural grassland root depth (Cougnon et al., 2017; Wedderburn et al., 2010). Each core was sown with ten seeds from one of the four varieties (DAS 0, Days after sowing) and allowed to germinate. At DAS 43, the most centrally germinated seedling was kept, and the other germinated seeds discarded. The seedlings were then cut to a height of 5 cm to simulate equal growth between all replicates. Waterlogging was initiated in half of all cores on DAS 48, with the additional watering being slowly initiated over three days to reach a stable water level 2 cm above the soil. Non-waterlogged cores continued to be watered normally. Waterlogging was actively enforced for one month and then allowed to dissipate naturally. Waterlogging experiments in grasses tend to last around 15 days (e.g. de la Cruz Jiménez et al., 2017; Liu and Jiang, 2015; Ploschuk et al., 2017), with few experiments lasting up to 30 days (McFarlane et al., 2003). The longer duration was implemented due to the increased chance (and consequences) of climate changed-induced extreme flooding events (Dore, 2005; Trenberth, 2011). Chamber treatment, waterlogging, and variety placement were stratified equally between the chambers and then randomised within chambers. The stratification allowed for equal numbers of each variety in each chamber, with half of the 80 cores per chamber being waterlogged for equal comparison for all factors.

**Table 1**

Experimental Growth Chamber setup for the Ambient chambers (#2 and #5) and Future climate chambers (#1 and #4) simulating the months June and July in Northwestern Europe. Dawn and dusk conditions are included in the light hours.

**Abbreviations:** SM - Simulated Months. DAS - Days after Sowing.

SM	DAS	Length Days	Treatment	Environmental Conditions					
				CO <sub>2</sub> (ppm)	Light / Dark Hours (h)	Max Light Intensity ( $\mu\text{mol-m}^{-2}\text{s}^{-1}$ )	Day Temp (°C)	Night Temp (°C)	Humidity (%)
June	44 - 72	28	Ambient	415	17 / 7	600	14	11	80
	44 - 72	28	eCO <sub>2</sub> + 2°C	550	17 / 7	600	16	13	80
July	73 - 101	28	Ambient	415	16 / 8	600	16	13	80
	73 - 101	28	eCO <sub>2</sub> + 2°C	550	16 / 8	600	18	15	80

**Table 2**

Perennial ryegrass (*Lolium perenne*) varieties grown and water status for all chambers and variety replicates. Each water status is equally divided between all four chambers.

Variety	Heading	Ploidity	Water Status		
			Logged	Normal	Total
Aberchoice	Late	Diploid	40	40	80
Abergain	Late	Tetraploid	40	40	80
Carraig	Intermediate	Tetraploid	40	40	80
Dunluce	Intermediate	Tetraploid	40	40	80

## 2.2. Phenotypic Data Collection

Multiple sets of phenotypic data were collected each month to identify the effects on plant performance from the waterlogging. SPAD (Soil and Plant Analyzer Development) readings were conducted using a SPAD-502 Plus Chlorophyll Meter (Konica Minolta) to sample leaf chlorophyll content (e.g. Bothe et al., 2018; Dong et al., 2019). Three representative leaves were sampled from each plant, with each leaf being sampled at three places and averaged for a reliable measurement. Soil moisture was measured at 10 cm depth using a HH2 Moisture Meter with a calibrated WET sensor type WET-2 attachment (Delta-T Devices). At the end of each month (DAS 72 and 101) the maximum height of the plants was measured and plant were then harvested by cutting at 5 cm above the soil-level. The harvested material from each plant was placed on a flat white background and photographed using a high-resolution LUMIX DC-G9 camera (Panasonic) with an accompanied ColorChecker Classic chart (X-rite). This enabled the harvested material to be processed using colour-corrective image analysis techniques to identify differences in leaf colour hues (e.g. de la Cruz Jiménez et al., 2017; Hu et al., 2013; Li et al., 2014; Zhang et al., 2014). The leaves were photographed immediately after harvest to prevent structural and colour degradation (Løkke et al., 2013; Yamauchi and Watada, 2019). After the photographs all harvest material was oven dried at 65°C for one week and then weighed to measure dried biomass.

### 2.3. Image Analysis

The first stage of image analysis was to convert the colour filter array (CFA) in each RAW image from the LUMIX DC-G9 camera into a colour-corrected red-green-blue (RGB) image. This was done using a standardised pipeline with the following nine steps: 1/ subtract a black value from all pixels in the CFA, 2/ set negative pixel values to zero, 3/ divide all pixels by the maximum pixel value, 4/ correct the white balance by scaling red and blue pixels relative to green pixels in the CFA, 5/ convert pixels to unsigned 16-bit integers, demosaiced the CFA into a true-colour image using a gradient-corrected linear interpolation (Malvar et al., 2004), 6/ transform the image from the camera's colour space to RGB, 7/ identify the 24 colours on the X-Rite ColorChecker Classic chart within the image, 8/ estimate an affine colour-transformation matrix that minimises the sum of squared deviations between the RGB colours in the image to the known colours of the 24 coloured squares, 9/ apply the colour-transformation matrix to produce a colour-correct RGB image. The camera's image metadata was used to obtain values for black level, white-balance correction and camera colour space to RGB conversion.

The second stage of image analysis extracted RGB values from pixels that corresponded to ryegrass leaves. Ryegrass leaves were placed upon a flat, white, rectangular background which enabled the image to be cropped to the white background. The cropped image was then converted into an HSV colour-space and an initial mask created with hue values in the range 0.0 - 0.4 and 0.875 - 1.0 (corresponding to yellow, green and red hues), saturation in the range 0.2 - 1.0 and value in the range 0.0 - 0.9. Regions of the mask with connected components containing fewer than 100 pixels were removed before the mask was refined using 50 iterations of an active contours region growing algorithm and saturation values retained if they were in the range 0.25 - 1.0. The final mask was used to extract the position of pixels in the mask and their RGB values. All image processing was performed using MatLAB (version R2021a.) and its image processing toolbox (Mathworks, 2021).

### 2.4. Statistical Analyses

The ryegrass leaf RGB values from the image analysis were further processed by calculating the median value for each hue from each image of the harvested ryegrass. Median values were used due to their robust statistical properties against outliers and skewed distributions (Chen, 1998; von Hippel, 2005). To dimensionally reduce the three hues into one variable the hues were analysed using principal component analysis (PCA) from the R package *vegan* (Oksanen

et al., 2020). Although variable standardisation is normally recommended (Jolliffe and Cadima, 2016), the hues were analysed without scaling and centring to preserve the relative values (Lever et al., 2017). We did not expect the removal of scaling to have a detrimental effect on model fitting due to the three bands having the same variances. The principal components (PC) were first tested for normality using the Shapiro-Wilks test (Shapiro and Wilk, 1965) and then analysed using Kendall's tau rank correlation (Kendall, 1938) and Wilcoxon's signed rank test (Wilcoxon, 1945) to test if there were any correlations and differences in mean values between the harvest groups (DAS 72 and 101) and between each water status. Kendall's tau was used due to the higher robustness and efficiency compared to the otherwise commonly used Spearman's rho (Croux and Dehon, 2010; Spearman, 1904). The first principal component was further used as a response variable to build a linear model to identify relevant covariates responsible for the combined hues. A similar approach has previously been used by Golzarian & Frick (2011) to investigate early growth stages of grasses. Multiple predictor variables were used to build the linear regression model: phenotypic (dried biomass, maximum height and SPAD measurements as a proxy for chlorophyll content), environmental (soil moisture as a proxy for waterlogging and chamber treatment as proxy for ambient and climate change climatological conditions), genetic (variety as a proxy for ploidy and heading date) and temporal (progression of the season and recovery from the waterlogging). The model terms were subsequently analysed using a Type II ANOVA (Langsrud, 2003; Smith and Cribbie, 2014). Model selection was then performed to analyse how removing variables in the full model would impact the AIC values and model understanding (Aho et al., 2014; Bozdogan, 1987). All statistical analyses were performed in the statistical software R (version 4.1.1.) (R Core Team, 2021).

### 3. Results

#### 3.1. Plant Appearance

To estimate the effects of long-term waterlogging on perennial ryegrass we grew 320 plants in fully atmospherically controlled growth chambers, simulating typical current climatic conditions in North-western Europe and conditions as predicted in 2050 (550 ppm CO<sub>2</sub> and a mean temperature increase of 2°C). Half of the plants were subjected to waterlogging 48 days after sowing (DAS) for one month. After waterlogging had begun, visible differences were observed between waterlogged and non-waterlogged plants at DAS 72 and continued to be visible at DAS 101 (**Figure 1**). The waterlogged plants by DAS 72 had stunted growth with dark brown leaf hues. The leaf morphology of the waterlogged plants also varied to the non-

waterlogged plants, because leaf unfolding was disrupted, causing a concave and folded appearance of many leaves in waterlogged plants (**Figure 2**). By DAS 101 many plants had started to change in leaf colour, with many leaves possessing light green shades. This contrasts with the non-waterlogged plants that had darker green shades and a lush appearance. Leaf morphology also differed at DAS 101 between waterlogged and non-waterlogged plants, where the leaves of the waterlogged plants remained concave and light green while the non-waterlogged plants had tall, wide and lush leaves. The median true colours of the harvests at DAS 72 (**Figure 3**) and DAS 101 (**Figure 4**) showed that there was substantial variation between plants of each water treatment and chamber condition, with darkening of leaves as the growth season progressed.

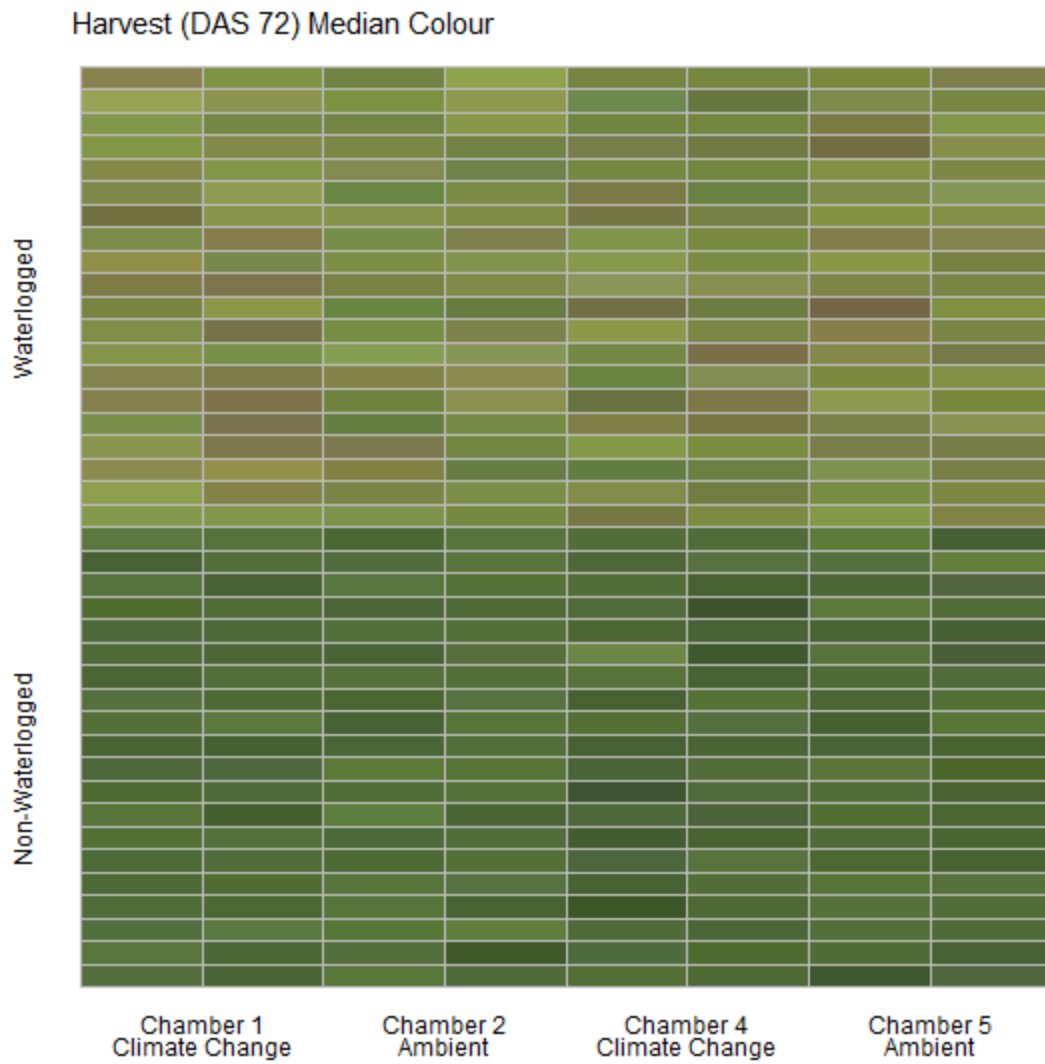




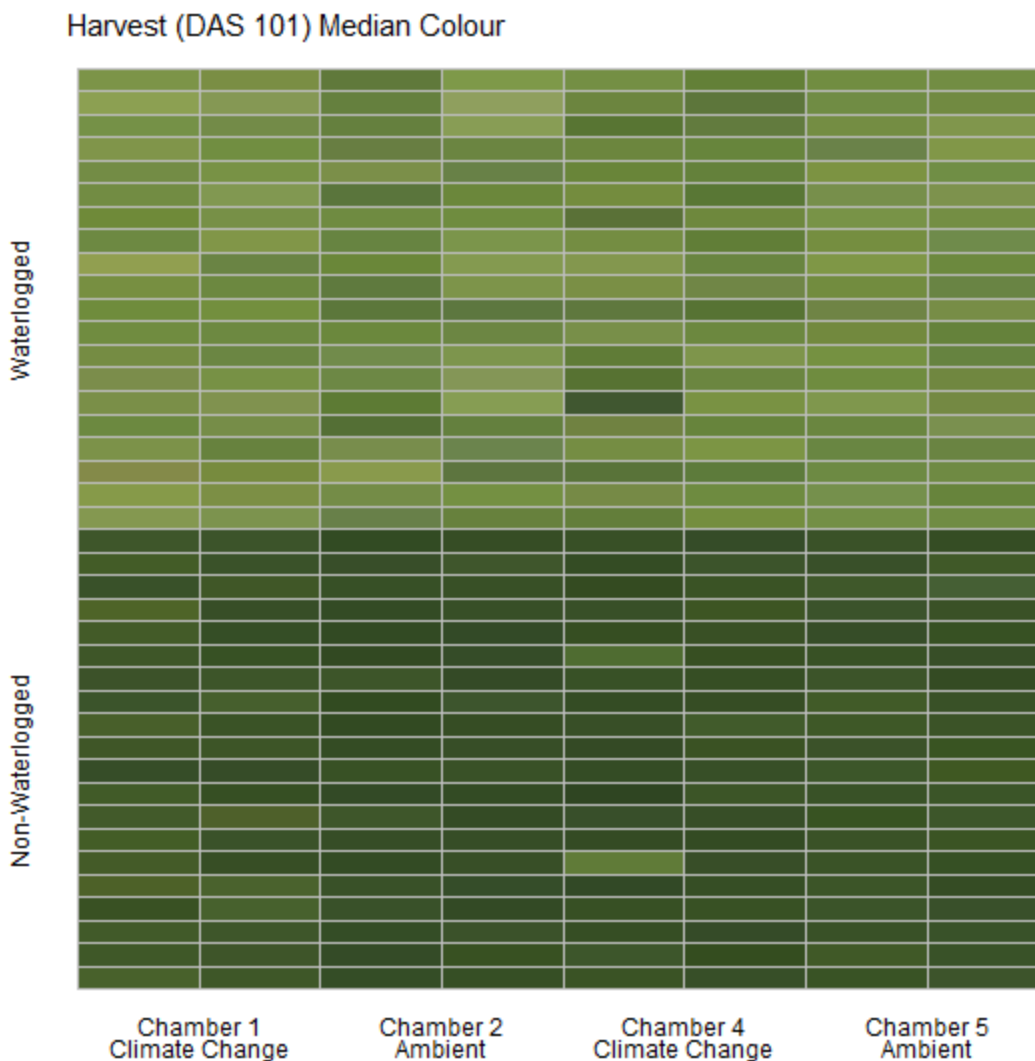
**Figure 1.** Perennial Ryegrass (*Lolium perenne*) whole plant appearance examples images for selected harvests and water status. A) DAS 72 non-waterlogged. B) DAS 72 waterlogged. C) DAS 101 non-waterlogged. D) DAS 101 waterlogged. Example images not colour-corrected.



**Figure 2.** Perennial Ryegrass (*Lolium perenne*) leaf morphology example images for selected harvests and water status. A) DAS 72 non-waterlogged. B) DAS 72 waterlogged C) DAS 72 waterlogged D) DAS 101 non-waterlogged. E) DAS 101 waterlogged. Example images not colour-corrected.



**Figure 3.** Median colour (RGB) of the harvested material for all cores on DAS 72 as identified by the image analysis, sorted on water status and chamber. Each cell represents the median true colour of the harvested material of a plant.

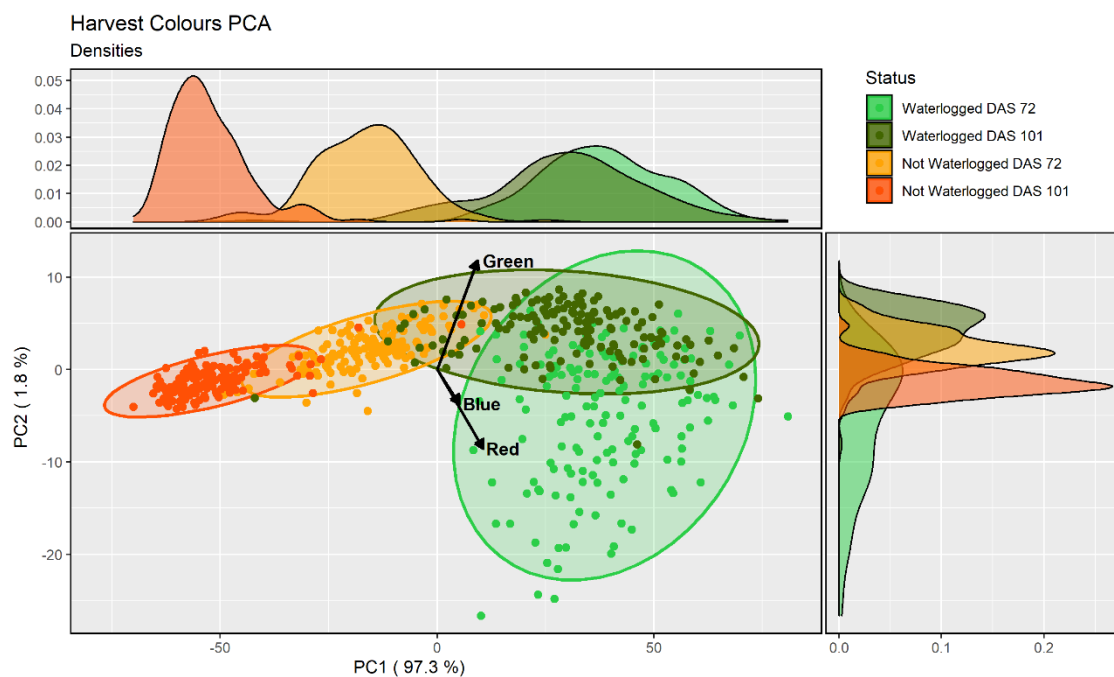


**Figure 4.** Median colour (RGB) of the harvested material for all cores on DAS 101 as identified by the image analysis, sorted on water status and chamber. Each cell represents the median true colour of the harvested material of a plant.

### 3.2. Colour Quantification Through RGB Fusion

To quantify the visible differences between the waterlogged and non-waterlogged plants, all plant leaves were harvested at DAS 72 and again at DAS 101, photographed and subjected to a colour analysis. The isolated RGB hues from the colour analysis were further modelled using a PCA-method. We hypothesised that there would be measurable colour differences within the water status, with the waterlogging contributing to lighter colours. We also expected colour differences between harvest dates, with darker colours as the growth season progressed. The PCA of the three RGB hues isolated from the harvested material from DAS 72 and 101 identified that the most variation (97.3 %) could be isolated in the first PC axis (PC1) (**Figure**

5). All three RGB hues showed a positive propensity for the first axis (**Supplementary Table 1**), illustrating that the axis describes a light beige to dark brown hue divergence (**Supplementary Figure 1**). To simplify this main leaf describing characteristics we classified the axis as overall colour intensity. The negative values of PC1 are therefore darker intensities, while the positive values are lighter intensities. The other two minor axes PC2 (1.8 %) and PC3 (0.9 %) describe pure colour hue gradients, a green-purple and an orange-blue hue divergence respectively. Kendall's tau and Wilcoxon's signed rank test showed that the harvested material from the waterlogged plants collected on DAS 72 were not correlated in colour to the non-waterlogged plants of the same harvest and were overall lighter ( $p < 0.001$ ) and more purple ( $p < 0.001$ ) (**Table 3**). The waterlogged plants on DAS 101 were positively correlated in colour intensity with the non-waterlogged plants of the same harvest while being darker ( $p < 0.001$ ) and greener ( $p < 0.001$ ). The harvested material from the waterlogged plants were positively correlated in colour intensity from DAS 72 to 101 with the plants becoming darker ( $p < 0.001$ ) and greener ( $p < 0.001$ ) as they started to recover from the waterlogging. The non-waterlogged plants were positively correlated in colour intensity between the harvests periods and became overall darker ( $p < 0.001$ ) as the growth season progressed. See supplementary material for the mean differences in PC axes values between the harvests and water status groupings (**Supplementary Table 2**).



**Figure 5.** PCA-analysis of the three RGB-hues of the harvested material for all cores, water status and harvest (DAS 72 and 101) as identified by the image analysis. Each ellipse represents a 95% confidence ellipse for each group.

**Table 3**

Model statistics and significance levels for the comparison of PC axes values for the groupings harvested material (DAS 72 and 101) and water status of the cores in contrast to the other grouping. The upper part of the table compares cores within the same water status between harvests. The lower part of the table compares between water status for the same harvest. **Abbreviation: DAS** - Days after Sowing.

PC Axis	Water Status		Harvest		Model Statistics				Wilcoxon Signed-Rank Test		
	Comparison		Comparison		z	tau (t)	P - value	Significance	V	P - value	Significance
PC1	Logged		DAS 72	DAS 101	9.979	0.532	< 1 x 10 <sup>-10</sup>	***	1,967	< 1 x 10 <sup>-10</sup>	***
	Normal		DAS 72	DAS 101	2.449	0.131	0.014	*	36	< 1 x 10 <sup>-10</sup>	***
PC2	Logged		DAS 72	DAS 101	1.757	0.094	0.079	NS	12,695	< 1 x 10 <sup>-10</sup>	***
	Normal		DAS 72	DAS 101	1.732	0.092	0.083	NS	445	< 1 x 10 <sup>-10</sup>	***
PC3	Logged		DAS 72	DAS 101	6.007	0.320	1.893 x 10 <sup>-9</sup>	***	4,232	1.694 x 10 <sup>-4</sup>	***
	Normal		DAS 72	DAS 101	5.495	0.293	3.907 x 10 <sup>-8</sup>	***	12,875	< 1 x 10 <sup>-10</sup>	***
PC1	Logged	Normal	DAS 72		0.282	0.015	0.778	NS	0	< 1 x 10 <sup>-10</sup>	***
	Logged	Normal	DAS 101		3.332	0.178	8.629 x 10 <sup>-4</sup>	***	2	< 1 x 10 <sup>-10</sup>	***
PC2	Logged	Normal	DAS 72		0.285	0.015	0.776	NS	12,053	< 1 x 10 <sup>-10</sup>	***
	Logged	Normal	DAS 101		-0.558	-0.030	0.577	NS	323	< 1 x 10 <sup>-10</sup>	***
PC3	Logged	Normal	DAS 72		1.374	0.073	0.170	NS	820	< 1 x 10 <sup>-10</sup>	***
	Logged	Normal	DAS 101		0.815	0.043	0.415	NS	9,506	1.765 x 10 <sup>-7</sup>	***

Significance: P < 0.001 - '\*\*\*', P < 0.01 - '\*\*', P < 0.05 - '\*', P > 0.05 - 'NS'.

### 3.3. Phenotypic Differentiation

To understand how phenotypic, environmental, genetic and temporal factors contribute to the quantified colours of the harvested leaf material a linear regression model was created and analysed. The first PC axis (PC1), identified as colour intensity, was modelled using three phenotypic variables, two environmental variables, one genetic variable and one temporal variable. The model was analysed using Type II ANOVA and AIC to see the model performance after subsequently removing each variable individually. We hypothesised that the effects from the predictor variables would be physiologically connected and cause differences in colour intensity. We expected that higher soil moisture would result in lighter colours and cause negative physiological effects on growth by lowering biomass and maximum height. We also expected that the climate change conditions would enhance growth through increased temperature and CO<sub>2</sub>, causing darker colours, along with darker colours as the growth season progressed, with variations between varieties caused by inherent genetic differences.

Phenotypically, harvested material from plants with lighter colour intensities had significantly lower dried biomass ( $F_{1,630} = 232.44$ ,  $p < 0.001$ ), significantly lower maximum height ( $F_{1,630} = 57.92$ ,  $p < 0.001$ ) and significantly lower SPAD values ( $F_{1,630} = 250.01$ ,  $p < 0.001$ ) (**Table 4**) (**Supplementary Figures 2 – 4**). For the water status, harvested material from cores with lighter colour intensities had significantly higher soil moisture ( $F_{1,630} = 174.67$ ,  $p < 0.001$ )

(**Supplementary Figure 5**). The harvested material from plants in chambers in predicted 2050 climate change conditions had significantly lighter intensities than harvested material from plants in ambient conditions ( $F_{1,630} = 19.20$ ,  $p < 0.001$ ) (**Supplementary Figure 6**).

Genetically, there were significant differences in colour intensity between the varieties ( $F_{3,630} = 33.41$ ,  $p < 0.001$ ). *Aberchoise*, the only diploid variety, had the lightest colour intensities, while the three tetraploid varieties, *Abergain*, *Dunluce* and *Carraig* had darker intensities in that order, with *Carraig* having the darkest leaf colour intensities (**Supplementary Figure 7**). The harvested material became darker overall as the season progressed ( $F_{1,630} = 4.27$ ,  $p = 0.039$ ), with the leaves in DAS 101 being significantly darker than DAS 72. The AIC revealed that the variables had a differential importance to the model performance. SPAD values ( $\Delta AIC = -211.9$ ), dried biomass ( $\Delta AIC = -199.0$ ) and soil moisture ( $\Delta AIC = -154.7$ ) were the most influential variables able to predict colour intensity, while progression of the season ( $\Delta AIC = -2.3$ ), climate treatment ( $\Delta AIC = -17.2$ ) and maximum height ( $\Delta AIC = -54.3$ ) were the least influential. Overall, the linear regression model had a very high accuracy in predicting leaf colour intensity, with an adjusted  $R^2$  of 91.8%. See supplementary material for the linear model estimates (**Supplementary Table 3**).

**Table 4**

Model statistics and significance levels for the linear model in regards to the first PCA axis (Colour Intensity) isolated from the colour analysis of the harvested perennial ryegrass material. **Abbreviations:** Df - Degrees of Freedom, RSS - Residual Sum of Squares.

**Model Statistics:** Anova (Type II) using the Drop1 function. **Model performance:** Adjusted  $R^2 = 91.8\%$ .

Variable	Model Statistics							
	Df	Sum Sq	RSS	AIC	$\Delta AIC$	F - Value	P - Value	Significance
<none>			79,595	3106.9				
Weight	1	29,367	108,962	3305.9	-199.0	232.44	$< 1 \times 10^{-10}$	***
Height	1	7,318	86,914	3161.2	-54.3	57.92	$< 1 \times 10^{-10}$	***
SPAD	1	31,587	111,182	3318.8	-211.9	250.01	$< 1 \times 10^{-10}$	***
Soil Moisture	1	22,068	101,663	3261.5	-154.6	174.67	$< 1 \times 10^{-10}$	***
Treatment	1	2,426	82,021	3124.1	-17.2	19.20	$1.379 \times 10^{-5}$	***
Variety	3	12,664	92,259	3195.4	-88.5	33.41	$< 1 \times 10^{-10}$	***
Month	1	539	80,134	3109.2	-2.3	4.27	0.039	*

Significance:  $P < 0.001$  - '\*\*\*',  $P < 0.01$  - '\*\*',  $P < 0.05$  - '\*',  $P > 0.05$  - 'NS'.

## 4. Discussion

Our aim was to investigate the phenotypic effect of long-term waterlogging on perennial ryegrass. We found that long-term waterlogging reduced perennial ryegrass productivity and changed the plant phenotype. Waterlogged plants were lighter in colour than controls, with younger waterlogged plants expressing purple hues, suggestive of anthocyanins. We also found that waterlogged plants had significantly lower dried biomass and maximum height,

demonstrating that waterlogging reduced overall plant performance and yield. Our study has experimentally determined the influence from waterlogging and climate change upon plant performance, with the effects from waterlogging largely overshadowing the effects from increased temperature and elevated CO<sub>2</sub>. The imaging methods could be developed as remote sensed diagnostics tools in combination with drone-technology to determine the influence of waterlogging on plant health in field environments.

#### **4.1. Physiology and Anthocyanins**

Our results identified substantial variation in leaf colouration in perennial ryegrass caused by long-term waterlogging. This variation was mainly in colour intensity, with waterlogging resulting in lighter colour intensities. Previous studies have shown that waterlogging can reduce concentrations of plant pigments, mainly chlorophyll, resulting in lighter colours and reduced photosynthetic capability (e.g. Barickman et al., 2019; Close and Davidson, 2003; Cotrozzi et al., 2021; Li et al., 2011; Ou et al., 2011; Pang et al., 2004; Simova-Stoilova et al., 2012; Smethurst and Shabala, 2003; Zhang et al., 2015). We also observed significant variation in hue, with waterlogging resulting in red/blue (purple) shades. This is especially pronounced for the waterlogged plants in DAS 72, with large variation being observed between the plants. These purple shades are likely caused by an accumulation of secondary anthocyanin metabolites, with multiple compounds having been identified in grasses (Clifford and Harborne, 1967; Fossen et al., 2002; Petrella et al., 2016). This suggests that individual plants respond differently to waterlogging by accumulating anthocyanins of varying degree. This is in agreement with previous research that has suggested that plants accumulate anthocyanins as a response to environmental stressors (Chalker-Scott, 1999), for example drought (e.g. Cirillo et al., 2021; Close and Beadle, 2003; Kovinich et al., 2015; Li et al., 2018). One of the main suggested benefits being protection against DNA damaging UV-B radiation that can reduce photosynthetic capability (Hoch et al., 2001; Rozema et al., 1997; Steyn et al., 2002; Teramura and Sullivan, 1994). Anthocyanin accumulation has also been found as a response to phosphorus deficiency (e.g. Sarker and Karmoker, 2011; Shaikh et al., 2008; Ulrychová and Sosnová, 1970). This is a likely cause of the observed purple shades in our waterlogged plants for two reasons. Firstly, phosphorus has been found to become soluble during excess water availability and migrate down the soil column (Sinaj et al., 2002) and secondly, altered plant nutrient uptake during periods of excess water abundance might make the soil nutrients (e.g., phosphorus) temporarily unavailable (Elzenga and van Veen, 2010). These effects are expected to be especially pronounced for younger plants, as we observed from our results, due to the



undeveloped root system occupying only the upper part of the soil column. Additional root growth and reduction in soil moisture would allow the plants to reach the migrated phosphorus and start to recover via normalised nutrient levels, as observed with the reduction in purple hues from the waterlogging in DAS 101. Our results strengthen the consensus of recent studies that phenotypic colour variations can effectively be quantified through image analysis and analysed to detect physiological effects from waterlogging (e.g. de la Cruz Jiménez et al., 2017; Ventura et al., 2020), although detailed soil nutrient analysis would be needed to confirm our findings in connection to phosphorus availability.

#### **4.2. Plant Performance**

Our results showed reductions in dried biomass and height in response to long-term waterlogging for all perennial ryegrass varieties. This likely stems from lower photosynthetic capability caused by a physiological reduction in plant pigments as identified from the SPAD values and colour analysis, but also from hindrance to root development. Complex interactions between soil moisture and root proliferation patterns are likely one of the main drivers governing growth performance via soil nutrient absorption (Brugge, 1985; Cougnon et al., 2017; Medlyn et al., 2016; Setter and Belford, 1990; Xu et al., 2013). Previous studies have linked waterlogging to reductions in grass performance caused by altered root development (Malik et al., 2002; Ploschuk et al., 2017). This agrees with our findings, as we observed a clear reduction in root proliferation in the waterlogged cores (visual inspection, results not shown), with long-term waterlogging having previously been shown to reduce root mass in ryegrass (McFarlane et al., 2003). Another potential consequence is reduced root respiration caused by a reduction in soil oxygen levels, which has been shown to have multiple negative feedbacks on grass biomass accumulation and nutrients absorption (Dunbabin et al., 1997; Fukao et al., 2019; Trought and Drew, 1980).

There is also variation in waterlogging tolerance between grass species (e.g. Rubio et al., 1995; Rubio and Lavado, 1999; Xiao and David, 2019) and wheat cultivars (e.g. Ghobadi et al., 2017; Cotrozzi et al., 2021) based on local adaptation and specific genotypes. Genotype specific tolerance to waterlogging has been found between perennial ryegrass varieties (Byrne et al., 2017; Liu and Jiang, 2015). This was expanded upon by Pearson et al. (2011) that found multiple quantitative trait loci (QTLs) that code for morphological traits influencing the tolerance to waterlogging in perennial ryegrass. This corroborates the differential performance to waterlogging amongst our varieties, with the tetraploid varieties generally tolerating

waterlogging better than the diploid variety. It is possible that the difference in performance is due to genotypic root development, which has been observed previously between perennial ryegrass varieties (Bonos et al., 2004; Deru et al., 2014; Wedderburn et al., 2010). Although, there is no clear connection between ploidy and stress tolerance to waterlogging or other environmental stressors (Kemesy et al., 2017; Lee et al., 2019; Tozer et al., 2017; Yu et al., 2012). We hypothesise that a wide range of varieties of varying genotypes grown under combinations of environmental stressors coupled with detailed genomics analyses would reveal the genetic basis of stress tolerance, and potential genetic trade-offs that occur between phenotypic traits.

### **4.3. Practical Implications**

We hypothesised that increases in temperature and CO<sub>2</sub> (predicted 2050-levels) would enhance plant performance to climate change-induced waterlogging, but our results showed that perennial ryegrass responded with lighter leaf shades, suggesting reduced photosynthetic capability and reduced yields. This is in contrast to the theory that climate change will generally increase photosynthesis and productivity (Chen et al., 1996; Dusenke et al., 2019; Yiotis et al., 2021). The developmental period of the grasses could be relevant here, with our study investigating the performance during the first few months containing younger plants. It is uncertain if these effects are specific to developing plants, or applicable to mature ones as well. Waterlogging has previously been shown to affect grain-crops differently depending on species and development, with wheat being able to sustain growth regardless of the period of waterlogging, while barley being disproportionately affected in later development (Ploschuk et al., 2021, 2020). Daepf *et al.* (2001) showed that the effects of elevated CO<sub>2</sub> will depend on the developmental stage of the ryegrass, suggesting that some stages are more sensitive to than others. For grasses in general, elevated CO<sub>2</sub> has been suggested to intensify the reproductive period from increases in NPP (Kurganskiy et al., 2021). Climate change will lead to multiple changes in the environment: increased temperature and CO<sub>2</sub> may have positive effects by extending the growing season for certain species while simultaneously flooding or drought may have negative effects on overall plant growth. To understand the effects of climate change we need to consider all environmental effects, their relative effect sizes and possible interactions to different developmental stages (Gray and Brady, 2016; Parmesan and Hanley, 2015; Tubiello et al., 2007; Zhou et al., 2020). In our case, the effect from waterlogging seems to largely overshadow the effects from increased temperature and elevated CO<sub>2</sub>.

To our knowledge this is the first study to investigate the combined effects of climate change (both temperature and CO<sub>2</sub>) and waterlogging experimentally in any plant. Previous studies have indicated that the interactive effects from elevated CO<sub>2</sub> in isolation with waterlogging are inconclusive (Pérez-Jiménez et al., 2018; Shimono et al., 2012), with one recent modelling-study suggesting that climate change will reduce waterlogging stress in barley (Liu et al., 2021). However, increases in flooding due to climate change would cause more severe impacts than from current precipitation regimes (Iglesias et al., 2012; Mirza, 2011). The decrease in overall dried biomass for all varieties as a consequence of waterlogging predicted by our study has implications for global food security. A reduction in overall plant yield due to poor plant growth from flooding would potentially cause increases in fodder prices with follow-up consequences to all agricultural sectors and industries relying on fodder from pasture lands (Hazell and Wood, 2008; Kipling et al., 2016b; Manik et al., 2019). Early detection of waterlogging will likely be important to employ mitigation strategies that could minimise reductions in plant health and production yield in pasture lands.

We demonstrate that image analysis approaches can be used as diagnostic tools to investigate plant performance reductions caused by waterlogging. While most field monitoring would be time consuming and most satellite-based remote sensing products would be low resolution for this type of analysis other more navigable high-resolution options are available, for example drones (Bansod et al., 2017; Cracknell, 2018; Simic Milas et al., 2018). Drones have been shown to be a useful and cost-effective tool for agricultural surveying (Kulbacki et al., 2018; Puri et al., 2017; Tripicchio et al., 2015) and plant ecological investigation (Cruzan et al., 2016; Sun et al., 2021; Tay et al., 2018; Zellweger et al., 2019). The main benefits comes from the use of high-resolution multi- and hyperspectral cameras which capture a wide-range of light wavelengths used to infer plant physiological parameters (e.g. Li et al., 2020; Tao et al., 2020; Papp et al., 2021). Our results showed that differences in colour intensity could be observed between perennial ryegrass varieties, suggesting that the imaging analysis method could be developed further to identify closely-related varieties or perhaps different grass species using remote-sensed colour distributions. Recent studies have also shown the usefulness of drones to monitor the effects of waterlogging on agricultural systems (e.g. Boiarskii et al., 2019; Den Besten et al., 2021; León-Rueda et al., 2021), illustrating that our imaging approaches could be adapted to work as diagnostic tools with drone-technology. Applications using integrated monitoring of plant health will become increasingly important as climate change-induced extreme weather events become more prevalent.

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## Author Contributions

**CAF** – Design of the research; Performance of the research; Data analysis, collection or interpretation; Writing the manuscript. **GX-S** – Performance of the research; Data analysis, collection or interpretation. **MO** – Performance of the research; Data analysis, collection or interpretation. **YB** – Performance of the research; Data analysis, collection or interpretation. **RM** - Design of the research; Data analysis, collection or interpretation; Writing the manuscript. **JMY** – Design of the research; Data analysis, collection or interpretation; Writing the manuscript.

## Data Availability

The data supporting the findings of this study (DOI: 10.5281/zenodo.6334191) will be publicly available upon journal acceptance on the general-purpose open-access repository Zenodo developed by the European OpenAIRE program and operated by CERN.

## References

- Aho, K., Derryberry, D., Peterson, T., 2014. Model selection for ecologists: the worldviews of AIC and BIC. *Ecology* 95, 631–636. <https://doi.org/10.1890/13-1452.1>
- Baldos, U.L.C., Hertel, T.W., 2014. Global food security in 2050: The role of agricultural productivity and climate change. *Aust. J. Agric. Resour. Econ.* 58, 554–570. <https://doi.org/10.1111/1467-8489.12048>
- Bansod, B., Singh, R., Thakur, R., Singhal, G., 2017. A comparison between satellite based and drone based remote sensing technology to achieve sustainable development: A review. *J. Agric. Environ. Int. Dev.* 111, 383–407. <https://doi.org/10.12895/jaeid.20172.690>
- Barickman, T.C., Simpson, C.R., Sams, C.E., 2019. Waterlogging Causes Early Modification in the Physiological Performance, Carotenoids, Chlorophylls, Proline, and Soluble Sugars of Cucumber Plants. *Plants* 160, 1–15. <https://doi.org/10.3390/plants8060160>

- Batke, S., Holohan, A., Hayden, R., Fricke, W., Porter, A.S., Christiana, C.M., 2018. The pressure is on – Epiphyte water-relations altered under elevated CO<sub>2</sub>. *Front. Plant Sci.* 871, 1–14. <https://doi.org/10.3389/fpls.2018.01758>
- Blöschl, G., Hall, J., Viglione, A., Perdigão, R.A.P., Parajka, J., Merz, B., Lun, D., Arheimer, B., Aronica, G.T., Bilibashi, A., Boháč, M., Bonacci, O., Borga, M., Čanjevac, I., Castellarin, A., Chirico, G.B., Claps, P., Frolova, N., Ganora, D., Gorbachova, L., Gül, A., Hannaford, J., Harrigan, S., Kireeva, M., Kiss, A., Kjeldsen, T.R., Kohnová, S., Koskela, J.J., Ledvinka, O., Macdonald, N., Mavrova-Guirguinova, M., Mediero, L., Merz, R., Molnar, P., Montanari, A., Murphy, C., Osuch, M., Ovcharuk, V., Radevski, I., Salinas, J.L., Sauquet, E., Šraj, M., Szolgay, J., Volpi, E., Wilson, D., Zaimi, K., Živković, N., 2019. Changing climate both increases and decreases European river floods. *Nature* 573, 108–111. <https://doi.org/10.1038/s41586-019-1495-6>
- Boiarskii, B., Hasegawa, H., Muratov, A., Sudeykin, V., 2019. Application of UAV-derived digital elevation model in agricultural field to determine waterlogged soil areas in Amur region, Russia. *Int. J. Eng. Adv. Technol.* 8, 520–523.
- Bonos, S.A., Rush, D., Hignight, K., Meyer, W.A., 2004. Selection for deep root production in tall fescue and perennial ryegrass. *Crop Sci.* 44, 1770–1775. <https://doi.org/10.2135/cropsci2004.1770>
- Bothe, A., Westermeier, P., Wosnitza, A., Willner, E., Schum, A., Dehmer, K.J., Hartmann, S., 2018. Drought tolerance in perennial ryegrass (*Lolium perenne* L.) as assessed by two contrasting phenotyping systems. *J. Agron. Crop Sci.* 204, 375–389. <https://doi.org/10.1111/jac.12269>
- Bozdogan, H., 1987. Model selection and Akaike's Information Criterion (AIC): The general theory and its analytical extensions. *Psychometrika* 52, 345–370. <https://doi.org/10.1007/BF02294361>
- Brown, J.N., Ash, A., MacLeod, N., McIntosh, P., 2019. Diagnosing the weather and climate features that influence pasture growth in Northern Australia. *Clim. Risk Manag.* 24, 1–12. <https://doi.org/10.1016/j.crm.2019.01.003>
- Brugge, R., 1985. A mechanistic model of grass root growth and development dependent upon photosynthesis and nitrogen uptake. *J. Theor. Biol.* 116, 443–467. [https://doi.org/10.1016/S0022-5193\(85\)80281-2](https://doi.org/10.1016/S0022-5193(85)80281-2)
- Buttler, A., Mariotte, P., Meisser, M., Guillaume, T., Signarbieux, C., Vitra, A., Preux, S., Mercier, G., Quezada, J., Bragazza, L., Gavazov, K., 2019. Drought-induced decline of productivity in the dominant grassland species *Lolium perenne* L. depends on soil type and prevailing climatic conditions. *Soil Biol. Biochem.* 132, 47–57. <https://doi.org/10.1016/j.soilbio.2019.01.026>
- Byrne, N., Gilliland, T.J., Mchugh, N., Delaby, L., Geoghegan, A., O'donovan, M., 2017. Establishing phenotypic performance of grass varieties on Irish grassland farms. *J. Agric. Sci.* 155, 1633–1645. <https://doi.org/10.1017/S0021859617000740>
- Canadell, J.G., Raupach, M.R., 2008. Managing forests for climate change mitigation. *SCIENCE* 320, 1456–1457. <https://doi.org/10.1126/science.1155458>

- Chalker-Scott, L., 1999. Environmental significance of anthocyanins in plant stress responses. *Photochem. Photobiol.* 70, 1–9. <https://doi.org/10.1111/j.1751-1097.1999.tb01944.x>
- Chen, D.X., Hunt, H.W., Morgan, J.A., 1996. Responses of a C3 and C4 perennial grass to CO2 enrichment and climate change: Comparison between model predictions and experimental data. *Ecol. Modell.* 87, 11–27. [https://doi.org/10.1016/0304-3800\(94\)00199-5](https://doi.org/10.1016/0304-3800(94)00199-5)
- Chen, Z., 1998. A note on bias robustness of the median. *Stat. Probab. Lett.* 38, 363–368. [https://doi.org/10.1016/S0167-7152\(98\)00049-2](https://doi.org/10.1016/S0167-7152(98)00049-2)
- Cirillo, V., D’Amelia, V., Esposito, M., Amitrano, C., Carillo, P., Carputo, D., Maggio, A., 2021. Anthocyanins are key regulators of drought stress tolerance in tobacco. *Biology (Basel)*. 10, 1–15. <https://doi.org/10.3390/biology10020139>
- Clifford, H.T., Harborne, J.B., 1967. Anthocyanin composition of and distribution in the Poaceae (Gramineae). *Proc. Linn. Soc. London* 178, 125–127. <https://doi.org/10.1111/j.1095-8312.1967.tb00968.x>
- Close, D.C., Beadle, C.L., 2003. The Ecophysiology of Foliar Anthocyanin. *Bot. Rev.* 69, 149–161. [https://doi.org/10.1663/0006-8101\(2003\)069\[0149:TEOFA\]2.0.CO;2](https://doi.org/10.1663/0006-8101(2003)069[0149:TEOFA]2.0.CO;2)
- Close, D.C., Davidson, N.J., 2003. Long-term waterlogging: Nutrient, gas exchange, photochemical and pigment characteristics of *Eucalyptus nitens* saplings. *Russ. J. Plant Physiol.* 50, 843–847. <https://doi.org/10.1023/B:RUPP.0000003284.25827.95>
- Commission, E., 2019. Common Catalogue of Varieties of Agricultural Plant Species (Document ID: C2019/013/01). *Off. J. Eur. Union* 37, 1-812.
- Cotrozzi, L., Lorenzini, G., Nali, C., Pisuttu, C., Pampana, S., Pellegrini, E., 2021. Transient waterlogging events impair shoot and root physiology and reduce grain yield of durum wheat cultivars. *Plants* 10, 1–19. <https://doi.org/10.3390/plants10112357>
- Cougnon, M., De Swaef, T., Lootens, P., Baert, J., De Frenne, P., Shahidi, R., Roldán-Ruiz, I., Reheul, D., 2017. In situ quantification of forage grass root biomass, distribution and diameter classes under two N fertilisation rates. *Plant Soil* 411, 409–422. <https://doi.org/10.1007/s11104-016-3034-7>
- Cracknell, A.P., 2018. The development of remote sensing in the last 40 years. *Int. J. Remote Sens.* 39, 8387–8427. <https://doi.org/10.1080/01431161.2018.1550919>
- Croux, C., Dehon, C., 2010. Influence functions of the Spearman and Kendall correlation measures. *Stat. Methods Appl.* 19, 497–515. <https://doi.org/10.1007/s10260-010-0142-z>
- Cruzan, M.B., Weinstein, B.G., Grasty, M.R., Kohn, B.F., Hendrickson, E.C., Arredondo, T.M., Thompson, P.G., 2016. Small Unmanned Aerial Vehicles (Micro-Uavs, Drones) in Plant Ecology. *Appl. Plant Sci.* 4, 1–11. <https://doi.org/10.3732/apps.1600041>
- Cullen, B.R., Johnson, I.R., Eckard, R.J., Lodge, G.M., Walker, R.G., Rawnsley, R.P., McCaskill, M.R., 2009. Climate change effects on pasture systems in south-eastern Australia. *Crop Pasture Sci.* 60, 933–942.

<https://doi.org/10.1071/CP09019>

Cullen, B.R., Rawnsley, R.P., Eckard, R.J., Christie, K.M., Bell, M.J., 2014. Use of modelling to identify perennial ryegrass plant traits for future warmer and drier climates. *Crop Pasture Sci.* 65, 758–766. <https://doi.org/10.1071/CP13408>

Daepf, M., Nösberger, J., Lüscher, A., 2001. Nitrogen fertilization and developmental stage alter the response of *Lolium perenne* to elevated CO<sub>2</sub>. *New Phytol.* 150, 347–358. <https://doi.org/10.1046/j.1469-8137.2001.00109.x>

de la Cruz Jiménez, J., Cardoso, J.A., Leiva, L.F., Gil, J., Forero, M.G., Worthington, M.L., Miles, J.W., Rao, I.M., 2017. Non-destructive phenotyping to identify *Brachiaria* hybrids tolerant to waterlogging stress under field conditions. *Front. Plant Sci.* 8, 1–10. <https://doi.org/10.3389/fpls.2017.00167>

Den Besten, N., Steele-Dunne, S., de Jeu, R., van der Zaag, P., 2021. Towards monitoring waterlogging with remote sensing for sustainable irrigated agriculture. *Remote Sens.* 13, 1–20. <https://doi.org/10.3390/rs13152929>

Deru, J., Schilder, H., van der Schoot, J.R., van Eekeren, N., 2014. Genetic differences in root mass of *Lolium perenne* varieties under field conditions. *Euphytica* 199, 223–232. <https://doi.org/10.1007/s10681-014-1129-x>

Dong, T., Shang, J., Chen, J.M., Liu, J., Quian, B., Ma, B., Morrison, M.J., Zhang, C., Liu, Y., Shi, Y., Pan, H., Zhou, G., 2019. Assessment of Portable Chlorophyll Meters for Measuring Crop Leaf Chlorophyll Concentrations. *Remote Sens.* 11, 2706. <https://doi.org/10.3390/rs11222706>

Dore, M.H.I., 2005. Climate change and changes in global precipitation patterns: What do we know? *Environ. Int.* 31, 1167–1181. <https://doi.org/10.1016/j.envint.2005.03.004>

Dunbabin, J.S., Hume, I.H., Ireson, M.E., 1997. Effects of irrigation frequency and transient waterlogging on the production of a perennial ryegrass-white clover pasture. *Aust. J. Exp. Agric.* 37, 165–171. <https://doi.org/10.1071/EA96057>

Dusenge, M.E., Duarte, A.G., Way, D.A., 2019. Plant carbon metabolism and climate change: elevated CO<sub>2</sub> and temperature impacts on photosynthesis, photorespiration and respiration. *New Phytol.* 221, 32–49. <https://doi.org/10.1111/nph.15283>

Easterling, D.R., Meehl, G.A., Parmesan, C., Changnon, S.A., Karl, T.R., Mearns, L.O., 2000. Climate Extremes: Observations, Modeling, and Impacts. *SCIENCE* 289, 2068–2074. <https://doi.org/10.1126/science.289.5487.2068>

Elzenga, J.T.M., van Veen, H., 2010. Waterlogging and Plant Nutrient Uptake, in: Mancuso, S., Shabala, S. (Eds.), *Waterlogging Signalling and Tolerance in Plants*. Springer Berlin Heidelberg, Berlin, pp. 23–35. [https://doi.org/10.1007/978-3-642-10305-6\\_2](https://doi.org/10.1007/978-3-642-10305-6_2)

Ergon, Seddaiu, G., Korhonen, P., Virkajärvi, P., Bellocchi, G., Jørgensen, M., Østrem, L., Reheul, D., Volaire, F., 2018. How can forage production in Nordic and Mediterranean Europe adapt to the challenges and

- opportunities arising from climate change? *Eur. J. Agron.* 92, 97–106.  
<https://doi.org/10.1016/j.eja.2017.09.016>
- Evans-FitzGerald, C., Porter, A.S., Yiotis, C., Elliott-Kingston, C., McElwain, J.C., 2016. Co-ordination in morphological leaf traits of early diverging angiosperms is maintained following exposure to experimental palaeo-atmospheric conditions of sub-ambient O<sub>2</sub> and elevated CO<sub>2</sub>. *Front. Plant Sci.* 7, 1–11.  
<https://doi.org/10.3389/fpls.2016.01368>
- Farfan-Vignolo, E.R., Asard, H., 2012. Effect of elevated CO<sub>2</sub> and temperature on the oxidative stress response to drought in *Lolium perenne* L. and *Medicago sativa* L. *Plant Physiol. Biochem.* 59, 55–62.  
<https://doi.org/10.1016/j.plaphy.2012.06.014>
- Fay, P.A., Kaufman, D.M., Nippert, J.B., Carlisle, J.D., Harper, C.W., 2008. Changes in grassland ecosystem function due to extreme rainfall events: Implications for responses to climate change. *Glob. Chang. Biol.* 14, 1600–1608. <https://doi.org/10.1111/j.1365-2486.2008.01605.x>
- Feng, X., Porporato, A., Rodriguez-Iturbe, I., 2013. Changes in rainfall seasonality in the tropics. *Nat. Clim. Chang.* 3, 811–815. <https://doi.org/10.1038/nclimate1907>
- Fossen, T., Slimestad, R., Øvstedal, D.O., Andersen, Ø.M., 2002. Anthocyanins of grasses. *Biochem. Syst. Ecol.* 30, 855–864. [https://doi.org/10.1016/S0305-1978\(02\)00028-5](https://doi.org/10.1016/S0305-1978(02)00028-5)
- Fukao, T., Barrera-Figueroa, B.E., Juntawong, P., Peña-Castro, J.M., 2019. Submergence and waterlogging stress in plants: A review highlighting research opportunities and understudied aspects. *Front. Plant Sci.* 10, 1–24.  
<https://doi.org/10.3389/fpls.2019.00340>
- Ghobadi, M.E., Ghobadi, M., Zebarjadi, A., 2017. Effect of waterlogging at different growth stages on some morphological traits of wheat varieties. *Int. J. Biometeorol.* 61, 635–645. <https://doi.org/10.1007/s00484-016-1240-x>
- Goliński, P., Czerwiński, M., Jørgensen, M., Mølmann, J.A.B., Golińska, B., Taff, G., 2018. Relationship between climate trends and grassland yield across contrasting European locations. *Open Life Sci.* 13, 589–598.  
<https://doi.org/10.1515/biol-2018-0070>
- Golzarian, M.R., Frick, R.A., 2011. Classification of images of wheat, ryegrass and brome grass species at early growth stages using principal component analysis. *Plant Methods* 7, 1–11. <https://doi.org/10.1186/1746-4811-7-28>
- Gray, S.B., Brady, S.M., 2016. Plant developmental responses to climate change. *Dev. Biol.* 419, 64–77.  
<https://doi.org/10.1016/j.ydbio.2016.07.023>
- Grogan, D., Gilliland, T.J., 2011. A review of perennial ryegrass variety evaluation in Ireland. *Irish J. Agric. Food Res.* 50, 65–81.
- Hannaway, D., Fransen, S., Cropper, J., Teel, M., Chaney, M., Griggs, T., Halse, R., Hart, J., Cheeke, P., Hansen, D., Klinger, R.G., Lane, W., 1999. Perennial Ryegrass (*Lolium perenne* L.). *A Pacific Northwest Ext. Publ.*



- Hazell, P., Wood, S., 2008. Drivers of change in global agriculture. *Philos. Trans. R. Soc. B Biol. Sci.* 363, 495–515. <https://doi.org/10.1098/rstb.2007.2166>
- Helgadóttir, Aavola, R., Isolahti, M., Marum, P., Persson, C., Aleliūnas, A., Brazauskas, G., Krisjánadóttir, T.A., Asp, T., Rognli, O.A., 2018. Adaptability and phenotypic stability of *Lolium perenne* L. cultivars of diverse origin grown at the margin of the species distribution. *J. Agron. Crop Sci.* 204, 493–504. <https://doi.org/10.1111/jac.12273>
- Hoch, W.A., Zeldin, E.L., McCown, B.H., 2001. Physiological significance of anthocyanins during autumnal leaf senescence. *Tree Physiol.* 21, 1–8. <https://doi.org/10.1093/treephys/21.1.1>
- Höglind, M., Thorsen, S.M., Semenov, M.A., 2013. Assessing uncertainties in impact of climate change on grass production in Northern Europe using ensembles of global climate models. *Agric. For. Meteorol.* 170, 103–113. <https://doi.org/10.1016/j.agrformet.2012.02.010>
- Hu, H., Oglesby, R.J., Saltzman, B., 2000. The relationship between atmospheric water vapor and temperature in simulations of climate change. *Geophys. Res. Lett.* 27, 3513–3516. <https://doi.org/10.1029/2000GL011680>
- Hu, H., Zhang, J., Sun, X., Zhang, X., 2013. Estimation of leaf chlorophyll content of rice using image color analysis. *Can. J. Remote Sens.* 39, 185–190. <https://doi.org/10.5589/m13-026>
- Hunt, W.F., Easton, H.S., 1989. Fifty Years of Ryegrass Research in New Zealand. *Proc. New Zeal. Grassl. Assoc.* 23, 1–23. <https://doi.org/10.33584/jnzc.1989.50.1876>
- Iglesias, A., Quiroga, S., Moneo, M., Garrote, L., 2012. From climate change impacts to the development of adaptation strategies: Challenges for agriculture in Europe. *Clim. Change* 112, 143–168. <https://doi.org/10.1007/s10584-011-0344-x>
- IPCC, 2014. *Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change.*
- Jentsch, A., Kreyling, J., Boettcher-Treschkow, J., Beierkuhnlein, C., 2009. Beyond gradual warming: Extreme weather events alter flower phenology of European grassland and heath species. *Glob. Chang. Biol.* 15, 837–849. <https://doi.org/10.1111/j.1365-2486.2008.01690.x>
- Jolliffe, I.T., Cadima, J., 2016. Principal component analysis: A review and recent developments. *Philos. Trans. R. Soc. A Math. Phys. Eng. Sci.* 374. <https://doi.org/10.1098/rsta.2015.0202>
- Kemesyte, V., Statkeviciute, G., Brazauskas, G., 2017. Perennial ryegrass yield performance under abiotic stress. *Crop Sci.* 57, 1935–1940. <https://doi.org/10.2135/cropsci2016.10.0864>
- Kendall, M.G., 1938. A New Measure of Rank Correlation. *Biometrika* 30, 81. <https://doi.org/10.2307/2332226>
- Kiely, G., 1999. Climate change in Ireland from precipitation and streamflow observations. *Adv. Water Resour.* 23, 141–151. [https://doi.org/10.1016/S0309-1708\(99\)00018-4](https://doi.org/10.1016/S0309-1708(99)00018-4)
- Kipling, R.P., Bannink, A., Bellocchi, G., Dalgaard, T., Fox, N.J., Hutchings, N.J., Kjeldsen, C., Lacetera, N., Sinabell, F., Topp, C.F.E., van Oijen, M., Virkajärvi, P., Scollan, N.D., 2016a. Modeling European ruminant

- production systems: Facing the challenges of climate change. *Agric. Syst.* 147, 24–37. <https://doi.org/10.1016/j.agsy.2016.05.007>
- Kipling, R.P., Virkajärvi, P., Breitsameter, L., Curnel, Y., De Swaef, T., Gustavsson, A.M., Hennart, S., Höglind, M., Järvenranta, K., Minet, J., Nendel, C., Persson, T., Picon-Cochard, C., Rolinski, S., Sandars, D.L., Scollan, N.D., Sebek, L., Seddaiu, G., Topp, C.F.E., Twardy, S., Van Middelkoop, J., Wu, L., Bellocchi, G., 2016b. Key challenges and priorities for modelling European grasslands under climate change. *Sci. Total Environ.* 566–567, 851–864. <https://doi.org/10.1016/j.scitotenv.2016.05.144>
- Kovinich, N., Kayanja, G., Chanoca, A., Otegui, M.S., Grotewold, E., 2015. Abiotic stresses induce different localizations of anthocyanins in *Arabidopsis*. *Plant Signal. Behav.* 10, 1–4. <https://doi.org/10.1080/15592324.2015.1027850>
- Kulbacki, M., Segen, J., Knieć, W., Klempous, R., Kluwak, K., Nikodem, J., Kulbacka, J., Serester, A., 2018. Survey of Drones for Agriculture Automation from Planting to Harvest, in: *INES 2018 - IEEE 22nd International Conference on Intelligent Engineering Systems, Proceedings*. pp. 000353–000358. <https://doi.org/10.1109/INES.2018.8523943>
- Kumar, P., 2013. Hydrology: Seasonal Rain Changes. *Nat. Clim. Chang.* 3, 783–784. <https://doi.org/10.1038/nclimate1996>
- Kurganskiy, A., Creer, S., Vere, N. De, Griffith, G.W., Osborne, N.J., Wheeler, B.W., McInnes, R.N., Clewlow, Y., Barber, A., Brennan, G.L., Hanlon, H.M., Hegarty, M., Potter, C., Rowney, F., Adams-groom, B., Petch, G.M., Pashley, C.H., Satchwell, J., Weger, L.A. De, Rasmussen, K., Oliver, G., Sindt, C., Bruffaerts, N., Allen, J., Bartle, J., Bevan, J., Frisk, C.A., Nielson, R., Potter, S., Selby, K., Tait, J., Zaragoza-Castells, J., Skjøth, C.A., 2021. Predicting the severity of the grass pollen season and the effect of climate change in Northwest Europe. *Sci. Adv.* 7, 1–12. <https://doi.org/10.1126/sciadv.abd7658>
- Langsrud, Ø., 2003. ANOVA for unbalanced data: Use Type II instead. *Stat. Comput.* 13, 163–167. <https://doi.org/10.1023/A:1023260610025>
- Lecerf, R., Ceglar, A., López-Lozano, R., Van Der Velde, M., Baruth, B., 2019. Assessing the information in crop model and meteorological indicators to forecast crop yield over Europe. *Agric. Syst.* 168, 191–202. <https://doi.org/10.1016/j.agsy.2018.03.002>
- Lee, M.A., Howard-Andrews, V., Chester, M., 2019. Resistance of multiple diploid and tetraploid perennial ryegrass (*Lolium perenne* L.) varieties to three projected drought scenarios for the UK in 2080. *Agronomy* 9, 1–7. <https://doi.org/10.3390/agronomy9030159>
- León-Rueda, W.A., León, C., Caro, S.G., Ramírez-Gil, J.G., 2021. Identification of diseases and physiological disorders in potato via multispectral drone imagery using machine learning tools. *Trop. Plant Pathol.* <https://doi.org/10.1007/s40858-021-00460-2>
- Lever, J., Krzywinski, M., Altman, N., 2017. Points of Significance: Principal component analysis. *Nat. Methods* 14, 641–642. <https://doi.org/10.1038/nmeth.4346>

- Li, B., Xu, X., Zhang, L., Han, J., Bian, C., Li, G., Liu, J., Jin, L., 2020. Above-ground biomass estimation and yield prediction in potato by using UAV-based RGB and hyperspectral imaging. *ISPRS J. Photogramm. Remote Sens.* 162, 161–172. <https://doi.org/10.1016/j.isprsjprs.2020.02.013>
- Li, C., Jiang, D., Wollenweber, B., Li, Y., Dai, T., Cao, W., 2011. Waterlogging pretreatment during vegetative growth improves tolerance to waterlogging after anthesis in wheat. *Plant Sci.* 180, 672–678. <https://doi.org/10.1016/j.plantsci.2011.01.009>
- Li, L., Zhang, Q., Huang, D., 2014. A review of imaging techniques for plant phenotyping. *Sensors (Switzerland)* 14, 20078–20111. <https://doi.org/10.3390/s141120078>
- Li, X., Lv, X., Wang, X., Wang, L., Zhang, M., Ren, M., 2018. Effects of abiotic stress on anthocyanin accumulation and grain weight in purple wheat. *Crop Pasture Sci.* 69, 1208–1214. <https://doi.org/10.1071/CP18341>
- Liu, K., Harrison, M.T., Archontoulis, S. V., Huth, N., Yang, R., Liu, D.L., Yan, H., Meinke, H., Huber, I., Feng, P., Ibrahim, A., Zhang, Y., Tian, X., Zhou, M., 2021. Climate change shifts forward flowering and reduces crop waterlogging stress. *Environ. Res. Lett.* 16. <https://doi.org/10.1088/1748-9326/ac1b5a>
- Liu, M., Jiang, Y., 2015. Genotypic variation in growth and metabolic responses of perennial ryegrass exposed to short-term waterlogging and submergence stress. *Plant Physiol. Biochem.* 95, 57–64. <https://doi.org/10.1016/j.plaphy.2015.07.008>
- Løkke, M.M., Seefeldt, H.F., Skov, T., Edelenbos, M., 2013. Color and textural quality of packaged wild rocket measured by multispectral imaging. *Postharvest Biol. Technol.* 75, 86–95. <https://doi.org/10.1016/j.postharvbio.2012.06.018>
- Malik, A.I., Colmer, T.D., Lambers, H., Setter, T.L., Schortemeyer, M., 2002. Short-term waterlogging has long-term effects on the growth and physiology of wheat. *New Phytol.* 153, 225–236. <https://doi.org/10.1046/j.0028-646X.2001.00318.x>
- Malvar, H.S., He, L.W., Cutler, R., 2004. High-quality linear interpolation for demosaicing of Bayer-patterned color images. *ICASSP, IEEE Int. Conf. Acoust. Speech Signal Process. - Proc.* 3, 2–5. <https://doi.org/10.1109/icassp.2004.1326587>
- Manik, S.M.N., Pengilley, G., Dean, G., Field, B., Shabala, S., Zhou, M., 2019. Soil and crop management practices to minimize the impact of waterlogging on crop productivity. *Front. Plant Sci.* 10, 1–23. <https://doi.org/10.3389/fpls.2019.00140>
- Mathworks, 2021. MATLAB R2021a. The Language for Technical Computing. Version 9.10.0.1710957. The MathWorks Inc., Natick, Massachusetts, United States.
- McFarlane, N.M., Ciavarella, T.A., Smith, K.F., 2003. The effects of waterlogging on growth, photosynthesis and biomass allocation in perennial ryegrass (*Lolium perenne* L.) genotypes with contrasting root development. *J. Agric. Sci.* 141, 241–248. <https://doi.org/10.1017/S0021859603003502>
- McHugh, O., Liu, J., Browne, F., Jordan, P., McConnell, D., 2020. Data-Driven Classifiers for Predicting Grass

- Growth in Northern Ireland: A Case Study, in: Information Processing and Management of Uncertainty in Knowledge-Based Systems, Communications in Computer and Information Science, Vol 1237. Springer International Publishing, pp. 301–312. [https://doi.org/10.1007/978-3-030-50146-4\\_23](https://doi.org/10.1007/978-3-030-50146-4_23)
- Medlyn, B.E., De Kauwe, M.G., Duursma, R.A., 2016. New developments in the effort to model ecosystems under water stress. *New Phytol.* 212, 5–7. <https://doi.org/10.1111/nph.14082>
- Minneé, E.M.K., Kuhn-Sherlock, B., Pinxterhuis, I.J.B., Chapman, D.F., 2019. Meta-analyses comparing the nutritional composition of perennial ryegrass (*Lolium perenne*) and plantain (*Plantago lanceolata*) pastures. *J. New Zeal. Grasslands* 81, 117–124. <https://doi.org/10.33584/jnzc.2019.81.402>
- Mirza, M.M.Q., 2011. Climate change, flooding in South Asia and implications. *Reg. Environ. Chang.* 11, 95–107. <https://doi.org/10.1007/s10113-010-0184-7>
- Mizutani, M., Kanaoka, M.M., 2018. Environmental sensing and morphological plasticity in plants. *Semin. Cell Dev. Biol.* 83, 69–77. <https://doi.org/10.1016/j.semcdb.2017.10.029>
- Münzbergová, Z., Hadincová, V., Skálová, H., Vandvik, V., 2017. Genetic differentiation and plasticity interact along temperature and precipitation gradients to determine plant performance under climate change. *J. Ecol.* 105, 1358–1373. <https://doi.org/10.1111/1365-2745.12762>
- Mustroph, A., 2018. Improving flooding tolerance of crop plants. *Agronomy* 8, 1–25. <https://doi.org/10.3390/agronomy8090160>
- O’Gorman, P.A., Muller, C.J., 2010. How closely do changes in surface and column water vapor follow Clausius-Clapeyron scaling in climate change simulations? *Environ. Res. Lett.* 5, 1–7. <https://doi.org/10.1088/1748-9326/5/2/025207>
- Oksanen, J., Guillaume Blanchet, F., Friendly, M., Kindt, R., Legendre, P., McGlenn, D., Minchin, P.R., O’Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., Wagner, H., 2020. *vegan: Community Ecology Package*.
- Ou, L.J., Dai, X.Z., Zhang, Z.Q., Zou, X.X., 2011. Responses of pepper to waterlogging stress. *Photosynthetica* 49, 339–345. <https://doi.org/10.1007/s11099-011-0043-x>
- Pang, J., Zhou, M., Mendham, N., Shabala, S., 2004. Growth and physiological responses of six barley genotypes to waterlogging and subsequent recovery. *Aust. J. Agric. Res.* 55, 895–906. <https://doi.org/10.1071/AR03097>
- Papp, L., van Leeuwen, B., Szilassi, P., Tobak, Z., Szatmári, J., Árvai, M., Mészáros, J., Pásztor, L., 2021. Monitoring invasive plant species using hyperspectral remote sensing data. *Land* 10, 1–18. <https://doi.org/10.3390/land10010029>
- Pareek, A., Dhankher, O.P., Foyer, C.H., 2020. Mitigating the impact of climate change on plant productivity and ecosystem sustainability. *J. Exp. Bot.* 71, 451–456. <https://doi.org/10.1093/jxb/erz518>
- Parmesan, C., Hanley, M.E., 2015. Plants and climate change: Complexities and surprises. *Ann. Bot.* 116, 849–

864. <https://doi.org/10.1093/aob/mcv169>

- Pearson, A., Cogan, N.O.I., Baillie, R.C., Hand, M.L., Bandaranayake, C.K., Erb, S., Wang, J., Kearney, G.A., Gendall, A.R., Smith, K.F., Forster, J.W., 2011. Identification of QTLs for morphological traits influencing waterlogging tolerance in perennial ryegrass (*Lolium perenne* L.). *Theor. Appl. Genet.* 122, 609–622. <https://doi.org/10.1007/s00122-010-1473-8>
- Pembleton, K.G., Cullen, B.R., Rawnsley, R.P., Ramilan, T., 2020. Climate change effects on pasture-based dairy systems in south-eastern Australia. *Crop Pasture Sci.* 72, 666–677. <https://doi.org/https://doi.org/10.1071/CP20108>
- Pérez-Jiménez, M., Hernández-Munuera, M., Piñero, M.C., López-Ortega, G., del Amor, F.M., 2018. Are commercial sweet cherry rootstocks adapted to climate change? Short-term waterlogging and CO<sub>2</sub> effects on sweet cherry cv. ‘Burlat.’ *Plant Cell Environ.* 41, 908–918. <https://doi.org/10.1111/pce.12920>
- Petrella, D.P., Metzger, J.D., Blakeslee, J.J., Nangle, E.J., Gardner, D.S., 2016. Anthocyanin production using rough bluegrass treated with high-intensity light. *HortScience* 51, 1111–1120. <https://doi.org/10.21273/HORTSCI10878-16>
- Ploschuk, R.A., Grimoldi, A.A., Ploschuk, E.L., Striker, G.G., 2017. Growth during recovery evidences the waterlogging tolerance of forage grasses. *Crop Pasture Sci.* 68, 574–582. <https://doi.org/10.1071/CP17137>
- Ploschuk, R.A., Miralles, D.J., Colmer, T.D., Striker, G.G., 2020. Waterlogging differentially affects yield and its components in wheat, barley, rapeseed and field pea depending on the timing of occurrence. *J. Agron. Crop Sci.* 206, 363–375. <https://doi.org/10.1111/jac.12396>
- Ploschuk, R.A., Miralles, D.J., Striker, G.G., 2021. Early- And late-waterlogging differentially affect the yield of wheat, barley, oilseed rape and field pea through changes in leaf area index, radiation interception and radiation use efficiency. *J. Agron. Crop Sci.* 207, 504–520. <https://doi.org/10.1111/jac.12486>
- Porter, A.S., Yiotis, C., Montañez, I.P., McElwain, J.C., 2017. Evolutionary differences in  $\Delta^{13}C$  detected between spore and seed bearing plants following exposure to a range of atmospheric O<sub>2</sub>:CO<sub>2</sub> ratios; implications for paleoatmosphere reconstruction. *Geochim. Cosmochim. Acta* 213, 517–533. <https://doi.org/10.1016/j.gca.2017.07.007>
- Puri, V., Nayyar, A., Raja, L., 2017. Agriculture drones: A modern breakthrough in precision agriculture. *J. Stat. Manag. Syst.* 20, 507–518. <https://doi.org/10.1080/09720510.2017.1395171>
- Qi, A., Holland, R.A., Taylor, G., Richter, G.M., 2018. Grassland futures in Great Britain – Productivity assessment and scenarios for land use change opportunities. *Sci. Total Environ.* 634, 1108–1118. <https://doi.org/10.1016/j.scitotenv.2018.03.395>
- Raza, A., Razzaq, A., Mehmood, S.S., Zou, X., Zhang, X., Lv, Y., Xu, J., 2019. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. *Plants* 8. <https://doi.org/10.3390/plants8020034>
- Rosenzweig, C., Tubiello, F.N., Goldberg, R., Mills, E., Bloomfield, J., 2002. Increased crop damage in the US from excess precipitation under climate change. *Glob. Environ. Chang.* 12, 197–202.

[https://doi.org/10.1016/S0959-3780\(02\)00008-0](https://doi.org/10.1016/S0959-3780(02)00008-0)

- Rozema, J., Van De Staaij, J., Björn, L.O., Caldwell, M., 1997. UV-B as an environmental factor in plant life: Stress and regulation. *Trends Ecol. Evol.* 12, 22–28. [https://doi.org/10.1016/S0169-5347\(96\)10062-8](https://doi.org/10.1016/S0169-5347(96)10062-8)
- Rubio, G., Casasola, G., Lavado, R.S., 1995. Adaptations and biomass production of two grasses in response to waterlogging and soil nutrient enrichment. *Oecologia* 102, 102–105. <https://doi.org/10.1007/BF00333316>
- Rubio, G., Lavado, R.S., 1999. Acquisition and allocation of resources in two waterlogging-tolerant grasses. *New Phytol.* 143, 539–546. <https://doi.org/10.1046/j.1469-8137.1999.00482.x>
- Sarker, B.C., Karmoker, J.L., 2011. Effects of phosphorus deficiency on accumulation of biochemical compounds in lentil (*Lens culinaris* Medik). *Bangladesh J. Bot.* 40, 23–27. <https://doi.org/10.3329/bjb.v40i1.7992>
- Semmler, T., Jacob, D., 2004. Modeling extreme precipitation events - A climate change simulation for Europe. *Glob. Planet. Change* 44, 119–127. <https://doi.org/10.1016/j.gloplacha.2004.06.008>
- Setter, T., Belford, B., 1990. Waterlogging : how it reduces plant growth and how plants can overcome its effects. *J. Dep. Agric. West. Aust. Ser.* 4 31, 51–55.
- Shaikh, N.P., Adjei, M.B., Scholberg, J.M., 2008. Interactive effect of phosphorus and nitrogen on leaf anthocyanins, tissue nutrient concentrations, and dry-matter yield of *Floralta* limpoggrass during short day length. *Commun. Soil Sci. Plant Anal.* 39, 1006–1015. <https://doi.org/10.1080/00103620801925414>
- Shapiro, S.S., Wilk, M.B., 1965. An Analysis of Variance Test for Normality (Complete Samples). *Biometrika* 52, 591–611. <https://doi.org/10.2307/2333709>
- Shimono, H., Konno, T., Sakai, H., Sameshima, R., 2012. Interactive effects of elevated atmospheric CO<sub>2</sub> and waterlogging on vegetative growth of soybean (*Glycine max* (L.) Merr.). *Plant Prod. Sci.* 15, 238–245. <https://doi.org/10.1626/pp.s.15.238>
- Simic Milas, A., Cracknell, A.P., Warner, T.A., 2018. Drones—the third generation source of remote sensing data. *Int. J. Remote Sens.* 39, 7125–7137. <https://doi.org/10.1080/01431161.2018.1523832>
- Simova-Stoilova, L., Demirevska, K., Kingston-Smith, A., Feller, U., 2012. Involvement of the leaf antioxidant system in the response to soil flooding in two *Trifolium* genotypes differing in their tolerance to waterlogging. *Plant Sci.* 183, 43–49. <https://doi.org/10.1016/j.plantsci.2011.11.006>
- Sinaj, S., Stamm, C., Toor, G.S., Condrón, L.M., Hendry, T., Di, H.J., Cameron, K.C., Frossard, E., 2002. Phosphorus Exchangeability and Leaching Losses from Two Grassland Soils. *J. Environ. Qual.* 31, 319–330. <https://doi.org/10.2134/jeq2002.0319>
- Smethurst, C.F., Shabala, S., 2003. Screening methods for waterlogging tolerance in lucerne: Comparative analysis of waterlogging effects on chlorophyll fluorescence, photosynthesis, biomass and chlorophyll content. *Funct. Plant Biol.* 30, 335–343. <https://doi.org/10.1071/FP02192>
- Smit, H.J., Tas, B.M., Taweel, H.Z., Tamminga, S., Elgersma, A., 2005. Effects of perennial ryegrass (*Lolium perenne* L.) cultivars on herbage production, nutritional quality and herbage intake of grazing dairy cows.

- Grass Forage Sci. 60, 297–309. <https://doi.org/10.1111/j.1365-2494.2005.00480.x>
- Smith, C.E., Cribbie, R., 2014. Factorial ANOVA with unbalanced data: A fresh look at the types of sums of squares. *J. Data Sci.* 12, 385–404. [https://doi.org/10.6339/jds.201407\\_12\(3\).0001](https://doi.org/10.6339/jds.201407_12(3).0001)
- Spearman, C., 1904. The Proof and Measurement of Association Between Two Things. *Am. J. Psychol.* 15, 72–101.
- Steyn, W.J., Wand, S.J.E., Holcroft, D.M., Jacobs, G., 2002. Anthocyanins in vegetative tissues: A proposed unified function in photoprotection. *New Phytol.* 155, 349–361. <https://doi.org/10.1046/j.1469-8137.2002.00482.x>
- Striker, G.G., 2012. Flooding Stress on Plants : Anatomical , Morphological and Physiological Responses, in: *Botany*. pp. 3–28.
- Sun, Z., Wang, X., Wang, Z., Yang, L., Xie, Y., Huang, Y., 2021. UAVs as remote sensing platforms in plant ecology: Review of applications and challenges. *J. Plant Ecol.* 14, 1003–1023. <https://doi.org/10.1093/jpe/rtab089>
- Tao, H., Feng, H., Xu, L., Miao, M., Yang, G., Yang, X., Fan, L., 2020. Estimation of the yield and plant height of winter wheat using UAV-based hyperspectral images. *Sensors (Switzerland)* 20, 1–19. <https://doi.org/10.3390/s20041231>
- Tay, J.Y.L., Erfmeier, A., Kalwij, J.M., 2018. Reaching new heights: can drones replace current methods to study plant population dynamics? *Plant Ecol.* 219, 1139–1150. <https://doi.org/10.1007/s11258-018-0865-8>
- Team, R.C., 2021. A Language and Environment for Statistical Computing. Version 4.1.1. R Foundation for Statistical Computing, Vienna, Austria.
- Teramura, A.H., Sullivan, J.H., 1994. Effects of UV-B radiation on photosynthesis and growth of terrestrial plants. *Photosynth. Res.* 39, 463–473. <https://doi.org/10.1007/BF00014599>
- Tester, M., Langridge, P., 2010. Breeding technologies to increase crop production in a changing world. *SCIENCE* 327, 818–822. <https://doi.org/10.1126/science.1183700>
- Tozer, K.N., Carswell, K., Griffiths, W.M., Crush, J.R., Cameron, C.A., Chapman, D.F., Popay, A., King, W., 2017. Growth responses of diploid and tetraploid perennial ryegrass (*Lolium perenne*) to soil-moisture deficit, defoliation and a root-feeding invertebrate. *Crop Pasture Sci.* 68, 632–642. <https://doi.org/10.1071/CP17154>
- Trenberth, K.E., 2011. Changes in precipitation with climate change. *Clim. Res.* 47, 123–138. <https://doi.org/10.3354/cr00953>
- Tripicchio, P., Satler, M., Dabisias, G., Ruffaldi, E., Avizzano, C.A., 2015. Towards Smart Farming and Sustainable Agriculture with Drones, in: *Proceedings - 2015 International Conference on Intelligent Environments, IE 2015*. pp. 140–143. <https://doi.org/10.1109/IE.2015.29>
- Trought, M.C.T., Drew, M.C., 1980. The development of waterlogging damage in young wheat plants in

- anaerobic solution cultures. *J. Exp. Bot.* 31, 1573–1585. <https://doi.org/10.1093/jxb/31.6.1573>
- Tubiello, F.N., Soussana, J.F., Howden, S.M., 2007. Crop and pasture response to climate change. *Proc. Natl. Acad. Sci. U. S. A.* 104, 19686–19690. <https://doi.org/10.1073/pnas.0701728104>
- Tubritt, T., Delaby, L., Gilliland, T.J., O'Donovan, M., 2020. The relationship between the grazing efficiency and the production, morphology and nutritional traits of perennial ryegrass varieties. *J. Agric. Sci.* 158, 583–593. <https://doi.org/10.1017/S0021859620000982>
- Ulrychová, M., Sosnová, V., 1970. Effect of phosphorus deficiency on anthocyanin content in tomato plants. *Biol. Plant.* 12, 231–235. <https://doi.org/10.1007/bf02920805>
- Ventura, I., Brunello, L., Iacopino, S., Valeri, M.C., Novi, G., Dornbusch, T., Perata, P., Loreti, E., 2020. Arabidopsis phenotyping reveals the importance of alcohol dehydrogenase and pyruvate decarboxylase for aerobic plant growth. *Sci. Rep.* 10, 1–14. <https://doi.org/10.1038/s41598-020-73704-x>
- von Hippel, P.T., 2005. Mean, median, and skew: Correcting a textbook rule. *J. Stat. Educ.* 13, 1–13. <https://doi.org/10.1080/10691898.2005.11910556>
- Wang, D., Wang, L., Liu, J., Zhu, H., Zhong, Z., 2018. Grassland ecology in China: Perspectives and challenges. *Front. Agric. Sci. Eng.* <https://doi.org/10.15302/J-FASE-2018205>
- Wedderburn, M.E., Crush, J.R., Pengelly, W.J., Walcroft, J.L., 2010. Root growth patterns of perennial ryegrasses under well-watered and drought conditions. *New Zeal. J. Agric. Res.* 53, 377–388. <https://doi.org/10.1080/00288233.2010.514927>
- Wilcoxon, F., 1945. Individual Comparisons by Ranking Methods. *Biometrics Bull.* 1, 80–83. <https://doi.org/10.2307/3001968>
- Xiao, B., David, D., 2019. Morphological and physiological responses of seashore paspalum and bermudagrass to waterlogging stress. *J. Am. Soc. Hortic. Sci.* 144, 305–313. <https://doi.org/10.21273/JASHS04737-19>
- Xu, C., McDowell, N.G., Sevanto, S., Fisher, R.A., 2013. Our limited ability to predict vegetation dynamics under water stress. *New Phytol.* 200, 298–300. <https://doi.org/10.1111/nph.12450>
- Yamauchi, N., Watada, A.E., 2019. Regulated Chlorophyll Degradation in Spinach Leaves during Storage. *J. Am. Soc. Hortic. Sci.* 116, 58–62. <https://doi.org/10.21273/jashs.116.1.58>
- Yates, S., Jaškūnė, K., Liebisch, F., Nagelmüller, S., Kirchgessner, N., Kölliker, R., Walter, A., Brazauskas, G., Studer, B., 2019. Phenotyping a dynamic trait: Leaf growth of perennial ryegrass under water limiting conditions. *Front. Plant Sci.* 10, 1–9. <https://doi.org/10.3389/fpls.2019.00344>
- Yiotis, C., Evans-Fitzgerald, C., McElwain, J.C., 2017. Differences in the photosynthetic plasticity of ferns and Ginkgo grown in experimentally controlled low [O<sub>2</sub>]:[CO<sub>2</sub>] atmospheres may explain their contrasting ecological fate across the Triassic-Jurassic mass extinction boundary. *Ann. Bot.* 119, 1385–1395. <https://doi.org/10.1093/aob/mcx018>
- Yiotis, C., McElwain, J.C., Osborne, B.A., 2021. Enhancing the productivity of ryegrass at elevated CO<sub>2</sub> is



- dependent on tillering and leaf area development rather than leaf-level photosynthesis. *J. Exp. Bot.* 72, 1962–1977. <https://doi.org/10.1093/jxb/eraa584>
- Yu, X., Luo, N., Yan, J., Tang, J., Liu, S., Jiang, Y., 2012. Differential growth response and carbohydrate metabolism of global collection of perennial ryegrass accessions to submergence and recovery following de-submergence. *J. Plant Physiol.* 169, 1040–1049. <https://doi.org/10.1016/j.jplph.2012.03.001>
- Zellweger, F., De Frenne, P., Lenoir, J., Rocchini, D., Coomes, D., 2019. Advances in Microclimate Ecology Arising from Remote Sensing. *Trends Ecol. Evol.* 34, 327–341. <https://doi.org/10.1016/j.tree.2018.12.012>
- Zhang, Y., Song, X., Yang, G., Li, Z., Lu, H., Kong, X., Eneji, A.E., Dong, H., 2015. Physiological and molecular adjustment of cotton to waterlogging at peak-flowering in relation to growth and yield. *F. Crop. Res.* 179, 164–172. <https://doi.org/10.1016/j.fcr.2015.05.001>
- Zhang, Y., Tang, L., Liu, X., Liu, L., Cao, W., Zhu, Y., 2014. Modeling dynamics of leaf color based on RGB value in rice. *J. Integr. Agric.* 13, 749–759. [https://doi.org/10.1016/S2095-3119\(13\)60391-3](https://doi.org/10.1016/S2095-3119(13)60391-3)
- Zhou, W., Chen, F., Meng, Y., Chandrasekaran, U., Luo, X., Yang, W., Shu, K., 2020. Plant waterlogging/flooding stress responses: From seed germination to maturation. *Plant Physiol. Biochem.* 148, 228–236. <https://doi.org/10.1016/j.plaphy.2020.01.020>