

1 **Reproductive tradeoffs govern sexually dimorphic tubular lysosome induction in *C. elegans***

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7

8 **Abstract**

9 Animals of different sexes often exhibit unique behaviors that benefit their specific reproductive
10 interests. In the nematode *Caenorhabditis elegans*, self-fertilizing hermaphrodites can reproduce
11 without a mate and thus prioritize feeding to satisfy the high energetic costs of reproduction.
12 However, males, which rely on finding potential mates for reproduction, sacrifice feeding and
13 instead prioritize exploratory behavior. Here, we demonstrate that these differences in behavior
14 are linked to sexual dimorphism at the cellular level; young males raised on a rich food source
15 show constitutive induction of gut tubular lysosomes, a non-canonical lysosome morphology that
16 typically forms in the gut of young hermaphrodites only when food is limited. We find that male-
17 specific induction of gut tubular lysosomes on abundant food is due to self-imposed dietary
18 restriction through *daf-7/TGF β* signaling, which promotes mate-searching at the cost of feeding.
19 While gut tubular lysosomes are largely absent from well-fed hermaphrodites at the start of
20 adulthood, their induction accelerates in hermaphrodites in early aging, dependent on the
21 presence of sperm and, partly, on embryo production. These findings identify tubular lysosome
22 induction as a sexually dimorphic cellular event that may integrate animal physiology with sex-
23 specific behavioral differences important for reproductive success.

1 **Introduction**

2
3 Many animal traits and behaviors, especially those linked to reproduction, display sexual
4 dimorphism (Portman, 2007; Yamamoto, 2007; Zilkha et al., 2021). Such phenotypes vary from
5 one sex to another within a single species, and maintenance of these differences is often vital for
6 efficient reproduction and, ultimately, species survival. In the nematode *C. elegans*, the choice
7 between feeding and mate searching presents an interesting example of a sexually dimorphic
8 behavior; young male worms prioritize mate searching over feeding, whereas hermaphrodites,
9 which reproduce on their own using self-sperm and oocytes, constantly prioritize feeding (Lipton
10 et al., 2004; Ryan et al., 2014). The male-specific preference for mating over feeding is
11 controlled by the *daf-7*/TGF β neuroendocrine signaling axis. In well-fed young males, elevated
12 DAF-7 inhibits expression of the odorant receptor *odr-10*, thereby reducing the preference to
13 feed (Hilbert and Kim, 2017; Wexler et al., 2020). In contrast, inhibiting *daf-7* in young males is
14 sufficient to prevent mate-searching behavior and to promote feeding behavior instead (Wexler
15 et al., 2020). While sacrificing feeding for exploratory behavior is a critical element of male
16 reproductive behavior and success, its effects on other aspects of animal physiology is less clear.
17 In principle, metabolic parameters linked to nutritional status could be impacted in males. To
18 what degree this occurs, and how it compares to changes in hermaphrodites, which assume high
19 metabolic costs in producing embryos, is unknown.

20 In previous work, we demonstrated that nutritional cues govern the induction of
21 autophagic tubular lysosomes (TLs) in the digestive tissues of worms and flies (Dolese et al.,
22 2021; Villalobos et al., 2021). Upon starvation or dietary restriction (DR), gut lysosomes
23 transform from vesicles into expansive tubular networks that show high degradative activity
24 (Dolese et al., 2021; Villalobos et al., 2021). Importantly, this morphological transformation in

1 lysosome structure supports lifespan extension in nutrient deprived conditions, and can even be
2 artificially mimicked in well-fed animals for health benefits (Villalobos et al., 2021). Given that
3 nutritional cues are intimately linked to sexually dimorphic feeding/mating behaviors in *C.*
4 *elegans*, it is conceivable that TL induction might naturally vary between the biological sexes in
5 this species. If so, this could contribute to sex-specific differences in animal health and
6 physiology.

7 Here, we demonstrate that young male *C. elegans* induce TLs in their gut even in the
8 presence of abundant nutrient sources. We further demonstrate that this is linked to the self-
9 imposed dietary avoidance that permits male worms to spend more time searching for a mate. In
10 contrast, hermaphrodites show lower TL-related signaling in young adulthood, but this increases
11 dramatically, surpassing even the male, during aging. Using sperm-defective mutants, we find
12 that elevated TL induction with age in hermaphrodites relates to both the presence of sperm and
13 embryo production, potentially as a mechanism to supply nutrients to the developing progeny
14 and/or the mother. Collectively, our results suggest that reproductive tradeoffs dictate TL
15 induction in *C. elegans* and may provide physiological support to animals as they prioritize
16 distinct modes of reproductive success.

17

18 **Materials and Methods**

19 **Strains**

20 Table S1 provides a complete list of strains used in this study. Endogenously-tagged *spin-*
21 *1::mCherry* was generated by *In Vivo* Biosystems using CRISPR technology. For genetic
22 crosses, the endogenous *spin-1::mCherry* transgene was tracked by stereomicroscopy, and
23 genetic mutations were verified by phenotypic characterization and/or sequencing.

1

2 **Animal maintenance**

3 Unless otherwise noted, worms were raised at 20°C on NGM agar (51.3 mM NaCl,
4 0.25% peptone, 1.7% agar, 1 mM CaCl₂, 1 mM MgSO₄, 25 mM KPO₄, 12.9 μM cholesterol, pH
5 6.0). For standard experiments, fed worms were maintained on NGM agar plates that had been
6 seeded with *E. coli* OP50 bacteria. To obtain starved adult worms, worms were washed 5x in 5
7 mL M9 buffer and transferred onto NGM agar that lacked OP50 bacteria. Synchronous
8 populations of worms were obtained by bleaching young-adult hermaphrodites. Briefly, adult
9 hermaphrodites were vortexed in 1 mL bleaching solution (0.5 M NaOH, 20% bleach) for 5
10 minutes to isolate eggs, and eggs were then washed three times in M9 buffer (22 mM KH₂PO₄,
11 42 mM Na₂HPO₄, 85.5 mM NaCl, 1 mM MgSO₄) before plating.

12 For RNAi experiments, synchronous populations of animals were grown on OP50-seeded
13 NGM plates until late L4 or day 1 of adulthood, at which time they were transferred to RNAi
14 plates (NGM plus 100 ng/μl carbenicillin and 1 mM IPTG) that had been seeded with bacteria
15 expressing the relevant RNAi clone. An empty L4440 vector was used as a negative control.

16 In experiments involving the *fog-2* strain, virgin females were isolated by transferring
17 feminized individuals onto NGM plates seeded with OP50 bacteria without males. Populations
18 of mated feminized worms were kept on plates with young males throughout the experiments.

19 Sperm-defective *fer-1* and *spe-9* strains are fertile at 15°C but, when raised at 25°C,
20 produce sperm that signal appropriately but are defective in fertilizing oocytes. In experiments
21 involving these mutants, strains were routinely maintained at 15°C until the experiment was
22 conducted. To obtain experimental, synchronous populations of *fer-1* and *spe-9* mutants, strains
23 were bleached, and NGM agar plates with eggs and OP50 bacteria were shifted to 25°C to render

1 animals self-sterile. Control strains for these experiments were treated identically at the same
2 time.

3 For aging experiments, synchronous populations of worms were obtained by bleaching
4 young adult hermaphrodites and plating their eggs onto NGM plates seeded with OP50 bacteria.
5 For progeny-producing strains, the sample populations were transferred to fresh NGM plates
6 every two days to isolate them from their progeny and to ensure a continuous food supply.

7

8 **Male generation and propagation**

9 Males were generated by heat shocking hermaphrodites. Briefly, hermaphrodite animals
10 were subjected to a persistent heat shock at 25°C, or, alternatively, L4 hermaphrodites were
11 subjected to a briefer 4-6 hour heat shock at 30°C. Male progeny isolated in the next generation
12 were propagated by mating. For mating, 8-10 hermaphrodites were placed on a 35 mm NGM
13 plate with roughly 20 males and maintained at 20°C overnight. This plate was seeded with a
14 small scoop of OP50 bacteria at the center of the plate to increase the likelihood of mating
15 encounters. On the following day, hermaphrodites were transferred to 60 mm NGM plates and
16 allowed to lay eggs. The mating process could be repeated iteratively in subsequent generations
17 to maintain a consistent population of males.

18

19 **Male-conditioned plates**

20 30 male worms were transferred onto NGM plates seeded with OP50 bacteria to allow
21 males to secrete pheromones onto the plates (Maures et al., 2014). After two days, males were
22 transferred off the plates, and *fog-2* feminized virgins were transferred onto the plates at day one

1 of adulthood to expose them to the male-conditioned environment. *fog-2* feminized virgins were
2 imaged two or four days after exposure.

3

4 **Microscopy**

5 4% agarose (Fisher Bioreagents) pads were dried on a Kimwipe (Kimtech) and then
6 placed on top of a Gold Seal™ glass microscope slide (ThermoFisher Scientific). A small
7 volume of 20 mM levamisole (Acros Organics) was spotted on the agarose pad. Worms were
8 transferred to the levamisole spot, and a glass cover slip (Fisher Scientific) was placed on top to
9 complete the mounting. Live-animal fluorescence microscopy was performed using a Leica
10 DMI8 THUNDER imager, equipped with 10X (NA 0.32), 40X (NA 1.30), and 100X (NA 1.40)
11 objectives and GFP and Texas Red filter sets.

12

13 **Image analysis**

14 Images were processed using LAS X software (Leica) and FIJI/ImageJ (NIH). Lysosome
15 networks were analyzed using “Skeleton” analysis plugins in FIJI. Briefly, images were
16 converted to binary 8-bit images and then to skeleton images using the “Skeletonize” plugin.
17 Skeleton images were then quantified using the “Analyze Skeleton” plugin. Number of objects,
18 number of junctions, and object lengths were scored. An “object” is defined by the Analyze
19 Skeleton plugin as a branch connecting two endpoints, an endpoint and a junction, or two
20 junctions. Junctions/object was used as a parameter to quantify network integrity. For SPIN-
21 1:mCherry fluorescence quantification, the gut tissue was outlined using the free-draw tool in
22 FIJI/ImageJ, and average fluorescence intensity of the outlined area was measured. For all

1 fluorescence intensity experiments, the same laser intensity (50%), exposure time (300 ms), and
2 FIM (100%) were used.

3

4 **Statistical analyses**

5 Data were statistically analyzed using GraphPad Prism. For two sample comparisons, an
6 unpaired t-test was used to determine significance ($\alpha=0.05$). For three or more samples, a one-
7 way ANOVA with Dunnett's multiple comparisons was used to determine significance ($\alpha=0.05$).

8

9 **Results and Discussion**

10 ***Young male worms show constitutive TL induction in the gut, even under nutrient rich*** 11 ***conditions***

12 Previously, we demonstrated that well-fed *C. elegans* hermaphrodites show gut
13 lysosomes that are morphologically static and predominantly vesicular in structure; however,
14 upon starvation, these lysosomes transform into dynamic, autophagic, tubular networks (Dolese
15 et al., 2021; Villalobos et al., 2021). Thus, starvation acts as a natural trigger for TL induction in
16 the gut of *C. elegans* hermaphrodites. To extend these studies, we explored whether male
17 animals, which normally sacrifice feeding for mating, show differences in TL induction, perhaps
18 even in the presence of food. We tracked lysosomes in males on and off of food using
19 endogenous *spin-1::mCherry*, which encodes a Spinster ortholog that robustly labels TLs
20 (Villalobos et al., 2021). We found that young male worms, unlike young hermaphrodites
21 (Villalobos et al., 2021), in fact exhibited TLs in the gut when food was abundant (Figure 1A-B).
22 As in starved hermaphrodites (Villalobos et al., 2021), TL induction in young males on food was
23 accompanied by a relative increase in endogenous SPIN-1 protein intensity (Figure 1C-D),

1 suggesting SPIN-1 protein expression serves as a proxy for TL induction. Additionally, young
2 male worms that were raised without food exhibited no further increase in SPIN-1::mCherry
3 protein intensity compared to males raised on food (Figure 1E-F). Thus, in young males, TLs
4 appear to be constitutively induced in the gut, regardless of food status.

5 Like starvation, aging also induces gut TLs in hermaphrodite worms (Dolese et al., 2021;
6 Villalobos et al., 2021). Given that young male worms exhibited TL induction and higher SPIN-
7 1::mCherry fluorescence intensity compared to young hermaphrodites, we surmised that male
8 worms might have a higher basal level of SPIN-1, which would continue to increase relative to
9 hermaphrodite levels during aging. However, this was not the case; by day five of adulthood,
10 SPIN-1::mCherry intensity in hermaphrodites superseded that in males, and this trend continued
11 into late life (Figure 1D). Thus, the stronger TL induction in nutrient rich conditions was specific
12 to young male worms. Moreover, some biological activities specific to hermaphrodites in young
13 adulthood may contribute to their relatively fast increase in SPIN-1 protein levels with age.

14

15 ***Elevated TL induction in young males results from daf-7-dependent prioritization of mating***
16 ***over feeding***

17 Our observation that TLs were induced in male worms even on a rich food source could
18 suggest that the same starvation-based mechanisms of TL induction seen in hermaphrodites do
19 not apply to the male sex. Yet, given the consideration that young male worms trade off feeding
20 in order to spend more time searching for a mate (Lipton et al., 2004; Ryan et al., 2014), we
21 reasoned that a self-imposed DR due to prioritization of exploratory behavior may explain the
22 constitutive TL induction in males raised on food. To test this hypothesis, we manipulated the
23 *daf-7/TGF β* signaling axis that differentially regulates feeding/mating decision-making in *C.*

1 *C. elegans* hermaphrodites and males (Figure 2A) (Hilbert and Kim, 2017; Milward et al., 2011;
2 Wexler et al., 2020; You et al., 2008). Strikingly, inhibition of *daf-7* by RNAi prevented the
3 male-specific increase in SPIN-1::mCherry fluorescence intensities and TL induction during
4 young age (Figure 2B-C). These data support the notion that TL induction in young male worms
5 is a consequence of a self-imposed DR caused by prioritized mate-searching behaviors.

6

7 ***Sperm signaling and embryo production contribute to increased SPIN-1 intensities in mothers***
8 ***during early aging***

9 We next considered mechanisms that possibly contribute to elevated SPIN-1 protein
10 expression and TL induction in hermaphrodites with age. Although the two natural sexes of *C.*
11 *elegans* are hermaphrodite (XX) and male (XO), “feminized” worms can be obtained using XX
12 animals incapable of producing sperm (Barton and Kimble, 1990). For example, in *fog-2* mutant
13 animals, germ cells that would normally differentiate into sperm instead differentiate into
14 oocytes (Schedl and Kimble, 1988). Using SPIN-1::mCherry intensity levels as a proxy for TL
15 induction, we compared SPIN-1::mCherry intensities in hermaphrodites, virgin feminized
16 animals, and mated feminized animals throughout adulthood. At day one, no significant
17 differences were observed between the three groups (Figure 3A-B). However, by days five and
18 ten, SPIN-1::mCherry intensities were significantly lower in virgin feminized animals compared
19 to both hermaphrodite and mated feminized animals (Figure 3A-B). These data suggest that the
20 presence of sperm might drive the steep increase in hermaphrodite SPIN-1 expression during
21 adulthood.

22 Intriguingly, the mere presence of mating-competent male worms has been demonstrated
23 to depreciate physiological health and lifespan of hermaphrodite worms cultured in the same

1 environment (Maures et al., 2014). Moreover, pre-conditioning plates with male pheromones
2 alone is sufficient to cause reduced lifespan in hermaphrodites, indicating that exposure to male
3 pheromones rather than mating triggers accelerated aging phenotypes in hermaphrodite worms
4 (Maures et al., 2014). These studies prompted us to test whether exposure to male pheromones
5 was also sufficient to induce age-related changes to *spin-1* expression levels in feminized
6 animals. We found that virgin feminized worms exposed to the male-conditioned plates failed to
7 exhibit increased SPIN-1::mCherry intensities compared to control virgin feminized worms after
8 either two or four days of exposure to male-conditioned plates (Figure S1A-B). Thus, exposure
9 to male-specific pheromones is insufficient to induce SPIN-1 levels, consistent with sperm
10 instead playing a causal role.

11 The above results suggested three possibilities: (i) signals emanating from sperm trigger
12 an age-related increase in SPIN-1 and TLs in the mother, independent of fertilization; (ii)
13 production of embryos upon fertilization of oocytes by sperm instead triggers TL induction; or
14 (iii) signals from both sperm and embryo production contribute to increased SPIN-1 and TL
15 induction. To distinguish between these possibilities, we examined SPIN-1::mCherry intensities
16 in *spe-9* and *fer-1* mutants, which can produce both gametes (sperm and oocytes) but have
17 mutations that render the sperm incapable of fertilization (L'Hernault et al., 1988; Singson et al.,
18 1998; Ward and Miwa, 1978; Ward et al., 1981). These sperm-defective mutants allowed us to
19 examine a biological scenario in which sperm signals are present, but embryo production is
20 disabled. At day one of adulthood, no significant increase in SPIN-1::mCherry intensity was
21 detected in either *spe-9* and *fer-1* mutants compared to virgin feminized worms (Figure 3C-D).
22 However, by days three and five, SPIN-1::mCherry intensity in *spe-9* and *fer-1* mutants
23 increased significantly compared to virgin feminized worms, albeit not to the level of

1 hermaphrodite worms (Figure 3C-D). These results suggest that signals from both sperm and
2 embryo production contribute to increasing SPIN-1::mCherry levels and TL induction in the
3 mother.

4 In conclusion, we have uncovered two sexually dimorphic properties of TL induction in
5 *C. elegans*: (1) young males show constitutive TL induction due to a self-imposed DR that
6 permits enhanced mate-searching behavior; and (2) TL induction in hermaphrodites commences
7 later during aging, dependent on previous reproductive signaling and activity. We propose that
8 TLs are induced by different mechanisms in each sex to meet the nutritional demands imposed
9 by their distinct reproductive activities. In young males, the induction of TLs could provide
10 health benefits during this self-imposed DR period to boost their reproductive fitness. DR has
11 been long known to confer health benefits and extend lifespan in many species; however, in *C.*
12 *elegans*, lifespan is extended by DR in hermaphrodites, but not in males (Honjoh et al., 2017).
13 This supports the notion that male worms, which are calorically restricted by choice, already
14 exhibit the health benefits of DR as a natural consequence of this behavior and, thus, do not
15 exhibit any further lifespan extension when put under experimental dietary constraints.
16 Moreover, we have shown previously that artificial induction of TLs allows worms to sustain
17 their mobility longer in life (Villalobos et al., 2021). Thus, it is interesting to speculate whether
18 induction of TLs in young males improves their physical fitness and ability to find a mate. In the
19 case of hermaphrodites, developing embryos inside the uterus require significant nutritional
20 support, which must come from the mother. Thus, TL induction might allow mothers to recycle
21 nutrients, such that they can provide additional nutritional sustenance to the developing embryos
22 and/or themselves during reproduction. Collectively, these findings add to growing evidence
23 indicating that different sexes have distinct nutritional requirements during their reproductive

1 lifespan, and they also suggest that TL induction may contribute to sustaining reproductive
2 fitness by different mechanisms in each sex.

3

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7

8 **Competing interests**

9 The authors declare that they have no competing interests.

10

11 **Author contributions**

12 Conceptualization: KAB, AEJ; Methodology: CDR; Investigation: CDR, KAB, AEJ;
13 Visualization: CDR, KAB, AEJ; Funding acquisition: KAB, AEJ; Project administration: KAB,
14 AEJ; Supervision: KAB, AEJ; Writing – original draft: KAB, AEJ; Writing – review & editing:
15 CDR, KAB, AEJ

16

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1 **References**

- 2 Akagi, K., Wilson, K.A., Katewa, S.D., Ortega, M., Simons, J., Hilsabeck, T.A., Kapuria, S.,
3 Sharma, A., Jasper, H., and Kapahi, P. (2018). Dietary restriction improves intestinal cellular
4 fitness to enhance gut barrier function and lifespan in *D. melanogaster*. *PLOS Genet.* *14*,
5 e1007777.
- 6 Barton, M.K., and Kimble, J. (1990). *fog-1*, a regulatory gene required for specification of
7 spermatogenesis in the germ line of *Caenorhabditis elegans*. *Genetics* *125*, 29–39.
- 8 Dolese, D.A., Junot, M.P., Ghosh, B., Butsch, T.J., Johnson, A.E., and Bohnert, K.A. (2021).
9 Degradative tubular lysosomes link pexophagy to starvation and early aging in *C. elegans*.
10 Autophagy.
- 11 Hilbert, Z.A., and Kim, D.H. (2017). Sexually dimorphic control of gene expression in sensory
12 neurons regulates decision-making behavior in *C. elegans*. *Elife* *6*.
- 13 Honjoh, S., Ihara, A., Kajiwara, Y., Yamamoto, T., and Nishida, E. (2017). The Sexual
14 Dimorphism of Dietary Restriction Responsiveness in *Caenorhabditis elegans*. *Cell Rep.* *21*,
15 3646–3652.
- 16 Kapahi, P., Kaeberlein, M., and Hansen, M. (2017). Dietary restriction and lifespan: Lessons
17 from invertebrate models. *Ageing Res. Rev.* *39*, 3–14.
- 18 L’Hernault, S.W., Shakes, D.C., and Ward, S. (1988). Developmental genetics of chromosome I
19 spermatogenesis-defective mutants in the nematode *Caenorhabditis elegans*. *Genetics* *120*, 435–
20 452.
- 21 Lakowski, B., and Hekimi, S. (1998). The genetics of caloric restriction in *Caenorhabditis*
22 *elegans*. *Proc. Natl. Acad. Sci. U. S. A.* *95*, 13091.
- 23 Lipton, J., Kleemann, G., Ghosh, R., Lints, R., and Emmons, S.W. (2004). Mate searching in

1 *Caenorhabditis elegans*: a genetic model for sex drive in a simple invertebrate. *J. Neurosci.* *24*,
2 7427–7434.

3 Magwere, T., Chapman, T., and Partridge, L. (2004). Sex differences in the effect of dietary
4 restriction on life span and mortality rates in female and male *Drosophila melanogaster*. *J.*
5 *Gerontol. A. Biol. Sci. Med. Sci.* *59*, 3–9.

6 Maures, T.J., Booth, L.N., Benayoun, B.A., Izrayelit, Y., Schroeder, F.C., and Brunet, A. (2014).
7 Males shorten the life span of *C. elegans* hermaphrodites via secreted compounds. *Science* *343*,
8 541–544.

9 Milward, K., Busch, K.E., Murphy, R.J., De Bono, M., and Olofsson, B. (2011). Neuronal and
10 molecular substrates for optimal foraging in *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. U. S.*
11 *A.* *108*, 20672–20677.

12 Portman, D.S. (2007). Genetic Control of Sex Differences in *C. elegans* *Neurobiology and*
13 *Behavior. Adv. Genet.* *59*, 1–37.

14 Ryan, D.A., Miller, R.M., Lee, K., Neal, S.J., Fagan, K.A., Sengupta, P., and Portman, D.S.
15 (2014). Sex, age, and hunger regulate behavioral prioritization through dynamic modulation of
16 chemoreceptor expression. *Curr. Biol.* *24*, 2509–2517.

17 Schedl, T., and Kimble, J. (1988). *fog-2*, a germ-line-specific sex determination gene required
18 for hermaphrodite spermatogenesis in *Caenorhabditis elegans*. *Genetics* *119*, 43–61.

19 Singson, A., Mercer, K.B., and L’Hernault, S.W. (1998). The *C. elegans spe-9* Gene Encodes a
20 Sperm Transmembrane Protein that Contains EGF-like Repeats and Is Required for Fertilization.
21 *Cell* *93*, 71–79.

22 Villalobos, T. V., Ghosh, B., Alam, S., Butsch, T.J., Mercola, B.M., Ramos, C.D., Das, S.,
23 Eymard, E.D., Bohnert, K.A., and Johnson, A.E. (2021). Tubular lysosome induction couples

- 1 animal starvation to healthy aging. *BioRxiv* 2021.10.28.466256.
- 2 Ward, S., and Miwa, J. (1978). Characterization of temperature-sensitive, fertilization-defective
3 mutants of the nematode *Caenorhabditis elegans*. *Genetics* 88, 285–303.
- 4 Ward, S., Argon, Y., and Nelson, G.A. (1981). Sperm morphogenesis in wild-type and
5 fertilization-defective mutants of *Caenorhabditis elegans*. *J. Cell Biol.* 91, 26–44.
- 6 Weindruch, R., Walford, R.L., Fligiel, S., and Guthrie, D. (1986). The retardation of aging in
7 mice by dietary restriction: longevity, cancer, immunity and lifetime energy intake. *J. Nutr.* 116,
8 641–654.
- 9 Wexler, L.R., Miller, R.M., and Portman, D.S. (2020). *C. elegans* Males Integrate Food Signals
10 and Biological Sex to Modulate State-Dependent Chemosensation and Behavioral Prioritization.
11 *Curr. Biol.* 30, 2695-2706.e4.
- 12 Yamamoto, D. (2007). The Neural and Genetic Substrates of Sexual Behavior in *Drosophila*.
13 *Adv. Genet.* 59, 39–66.
- 14 You, Y. jai, Kim, J., Raizen, D.M., and Avery, L. (2008). Insulin, cGMP, and TGF- β Signals
15 Regulate Food Intake and Quiescence in *C. elegans*: A Model for Satiety. *Cell Metab.* 7, 249–
16 257.
- 17 Zilkha, N., Sofer, Y., Kashash, Y., and Kimchi, T. (2021). The social network: Neural control of
18 sex differences in reproductive behaviors, motivation, and response to social isolation. *Curr.*
19 *Opin. Neurobiol.* 68, 137–151.
- 20

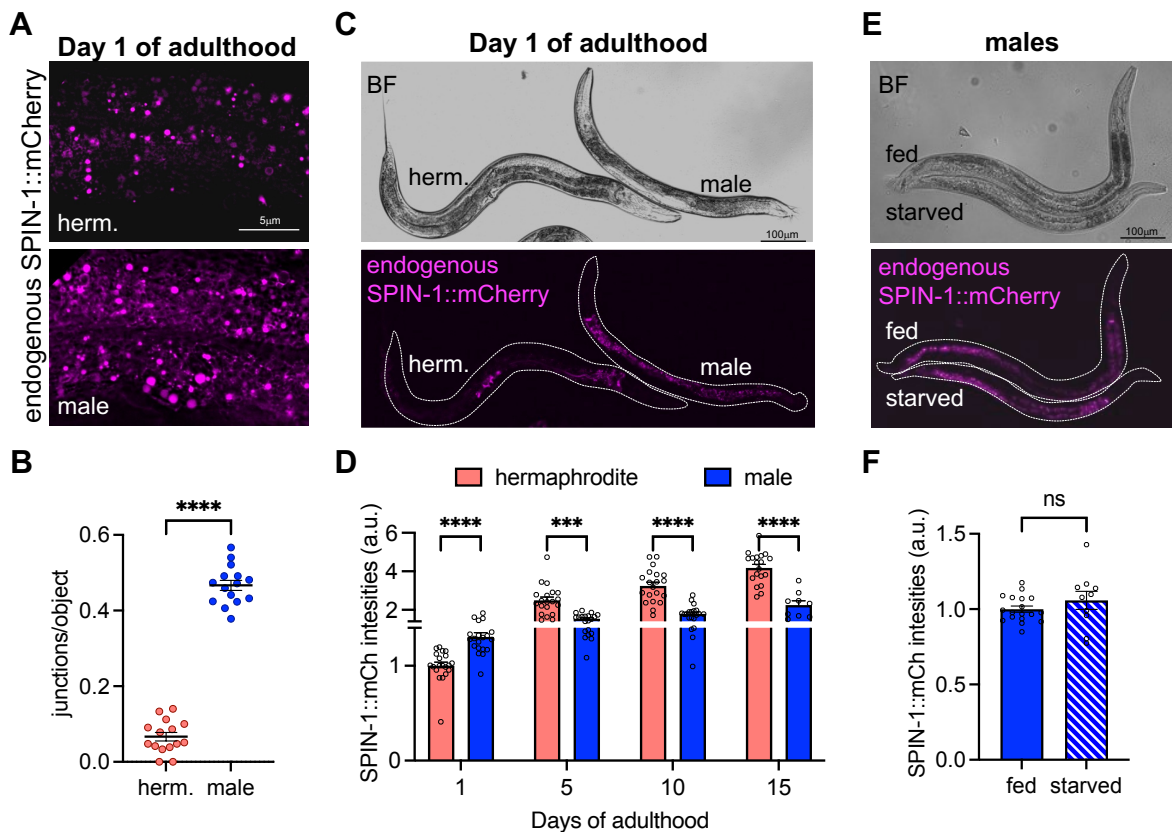


Figure 1: Young male worms show constitutive TL induction in the gut, even under nutrient rich conditions.

A. Representative images of endogenously tagged SPIN-1::mCherry in hermaphrodite and male worms. **B.** Quantification of lysosome junctions/object in hermaphrodite and male worms. **C.** Representative images of *spin-1* expression in hermaphrodite and male worms. **D.** Quantification of SPIN-1::mCherry intensities in hermaphrodite and male worms throughout adulthood. **E.** Representative images of *spin-1* expression in fed and starved male worms. **F.** Quantification of SPIN-1::mCherry intensities in fed and starved male worms.

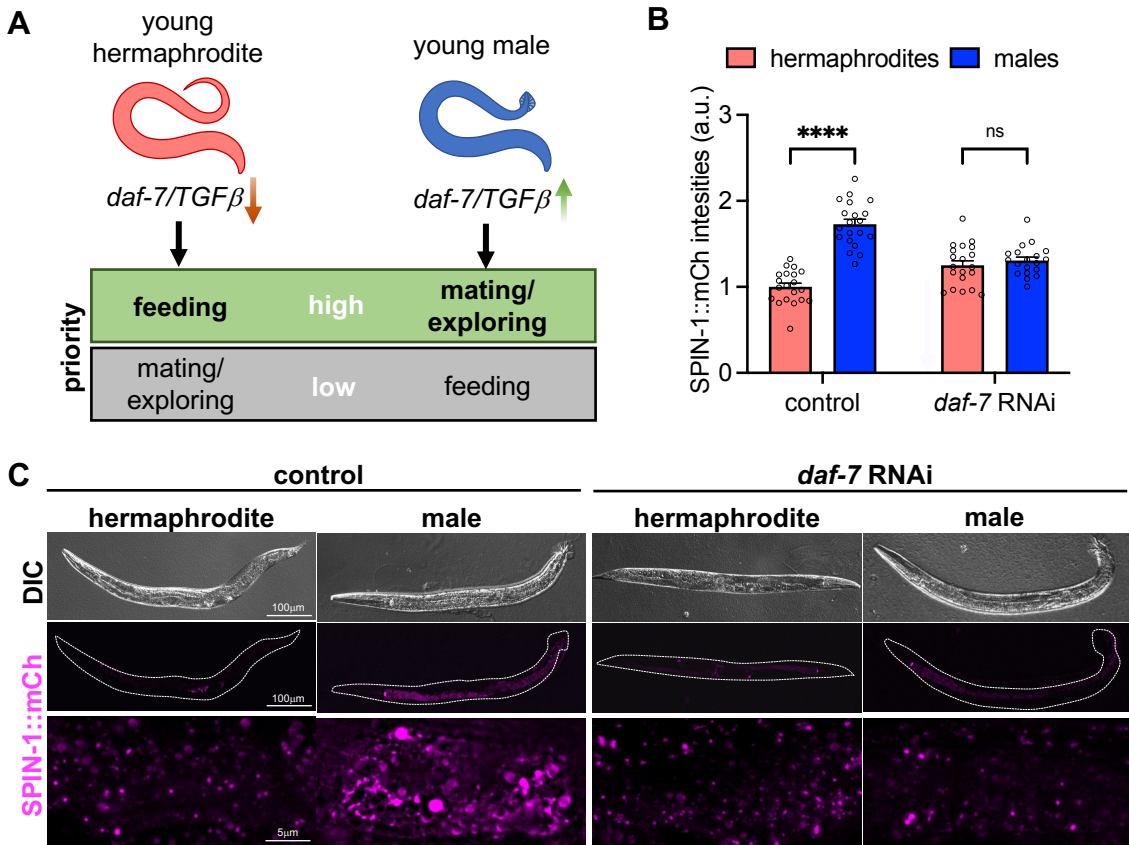


Figure 2: Elevated TL induction in young males results from *daf-7*-dependent prioritization of mating over feeding.

A. Schematic diagram illustrating the *daf-7*-dependent sexually dimorphic feeding/exploring behavior in *C. elegans*. In young hermaphrodites, *daf-7* signaling is downregulated, which promotes feeding behaviors. In contrast, *daf-7* signaling is upregulated in young male worms to promote exploratory behaviors. **B.** Quantification of SPIN-1::mCherry intensities in control and *daf-7* RNAi treated worms. **C.** Representative images of *spin-1* expression and TL induction in control and *daf-7* RNAi treated worms.

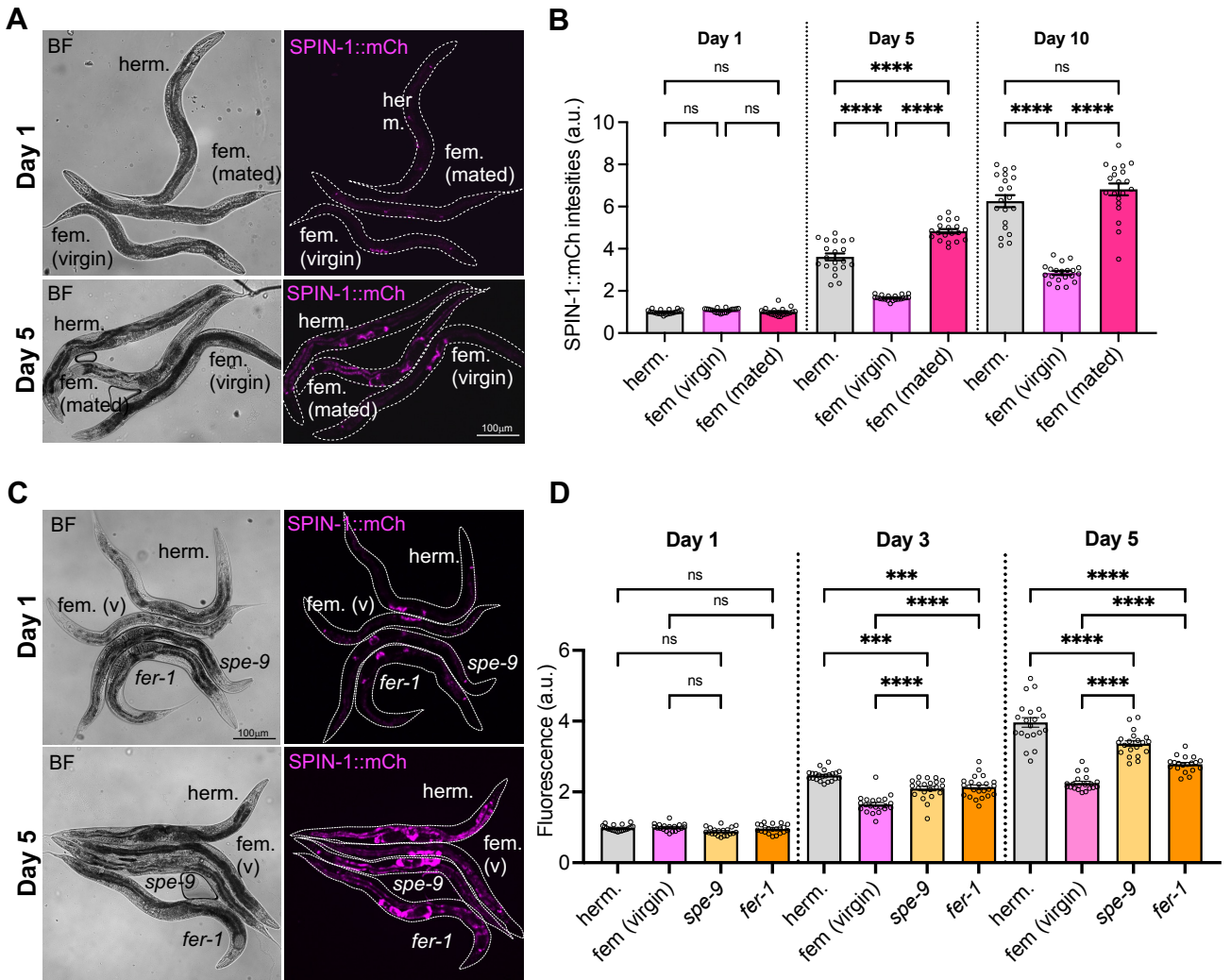


Figure 3: Sperm signaling and embryo production contribute to increased SPIN-1 intensities in mothers during early aging.

A-B. Representative images of *spin-1* expression (A) and quantification of SPIN-1::mCherry intensities (B) in hermaphrodite and feminized worms (virgin and mated) throughout adulthood.

C-D. Representative images of *spin-1* expression (C) and quantification of SPIN-1::mCherry intensities (D) in hermaphrodites, virgin feminized worms, and mutants with fertilization-incompetent sperm (*spe-9* and *fer-1*) during early adulthood.