# Increased male investment in sperm competition results in offspring of lower quality

Mareike Koppik<sup>1, 2, \*</sup>, Julian Baur<sup>1</sup> & David Berger<sup>1</sup>

Affiliation: <sup>1</sup>Department of Ecology and Genetics, Animal Ecology, Uppsala University, 75236

Uppsala, Sweden

Present address: <sup>2</sup>Department of Zoology, Animal Ecology, Martin-Luther University Halle-

Wittenberg, 06120 Halle (Saale), Germany

\*Correspondence: mareike.koppik@zoologie.uni-halle.de

## Abstract

Male animals often show higher mutation rates than their female conspecifics. A hypothesis for this male-bias is that competition over fertilization of female gametes leads to sexual selection for increased male germline replication at the expense of maintenance and repair, resulting in a trade-off between male success in sperm competition and offspring quality. Here we test this hypothesis using experimental evolution lines of the seed beetle Callosobruchus maculatus, maintained for >50 generations under three alternative mating regimes: natural and sexual selection (N+S-lines), natural selection only (N-lines) or sexual selection only (S-lines). Previous findings suggest that S-males reduce germline maintenance when engaging in reproduction compared to N- and N+S-males. Here, we first show that S-males are superior in sperm competition compared to both N- and N+S-males, suggesting that the removal of trade-offs between naturally and sexually selected male fitness components has resulted in the evolution of increased post-copulatory reproductive success. We then show that S-males produce progeny of lower quality if engaging in socio-sexual interactions with conspecifics prior to being challenged with a dose of irradiation introducing DNA-damage in their germline. We identify 18 candidate genes that showed differential expression in response to the induced germline damage. These genes also showed significant expression changes across socio-sexual treatments of fathers and predicted the reduction in quality of their offspring. Moreover, sex differences in expression of the same 18 genes indicate a substantially higher female investment in germline maintenance. Our findings provide evidence for a trade-off between male success in sperm

competition and germline maintenance, suggesting that sex-differences in the relative strengths

of sexual and natural selection are causally linked to male-mutation bias.

Key words: sexual selection, sperm competition, experimental evolution, mutation rate, post

copulatory, DNA repair, oxidative damage, genetic variation, offspring quality, genetic load

## Introduction

The germline mutation rate impacts on a range of evolutionary processes such as the rate of adaption<sup>1,2</sup> and risk of extinction <sup>3,4</sup>, sequence evolution <sup>5</sup> and the preservation of genetic variation <sup>6</sup>. Contrary to the typical assumption in population genetic models, recent studies have shown that mutation rate can be a state-dependent property and vary both within and between individuals within a given species <sup>7–17</sup>. Such variability can affect genetic load at mutation-selection balance <sup>18,19</sup>, rates of adaptation in stressful environments <sup>9,20,21</sup>, the prevalence of genetic disease <sup>22–25</sup>, and may cause errors in evolutionary inferences based on molecular clocks <sup>26</sup>. Despite such important implications, experimental evidence providing ultimate causation for the observed intraspecific variability in mutation rates remains scarce <sup>27–29</sup>.

One type of intraspecific variation is that between males and females of a given species. Males often show higher germline mutation rates in animal taxa <sup>30–34</sup>, including humans <sup>35,36</sup> and other primates <sup>28,34</sup>. This male mutation bias has been ascribed to the greater number of cell divisions occurring in the male germline prior to fertilization, and the higher number of divisions in males is itself thought to be a result of anisogamy and sexual selection promoting increased gamete production in the sex competing most intensively for fertilization success <sup>37,38</sup>. Indeed, a need for fast-dividing male germline cells would inevitably lead to an elevated risk of unrepaired replication errors in male gametes, all else equal, as the DNA-repair system must constantly attend single and double strand breaks that occur during meiosis and mitosis <sup>39–42</sup> and in postmeiotic chromatin remodelling during spermiogenesis <sup>35,42,43</sup>. This should result in a trade-off between increased male germline replication rates, granting greater success in sperm

4

competition, and increased germline mutation rate, reducing offspring quality <sup>8,39,44–47</sup>. This potential feedback loop between the strength of sexual selection and male mutation rate has implications for mate choice processes <sup>8,31,46,48–50</sup>.

However, sex differences in the number of germline cell division do not perfectly predict male mutation bias across species <sup>28,31,33,51,52</sup> and in humans differences in mutation rate between males of the same age can be many times greater than that between the sexes <sup>13,14,36</sup>. This suggests that a major determinant of the mutation rate is the energetically costly maintenance of the germline <sup>42,43,53</sup>, comprising interrelated processes such as antioxidant defence <sup>54–56</sup>, repair of DNA damage <sup>21,40</sup> and programmed cell death of damaged sperm <sup>57</sup>. Indeed, the male gonad is a highly oxidative environment that, without antioxidant defence devoted to dealing with reactive oxygen species <sup>47,54–57</sup>, produces DNA damage that when left unchecked results in germline mutations <sup>41–43,58</sup>. Accordingly, recent studies show that ejaculate composition <sup>59–64</sup> as well as sperm phenotype <sup>65</sup> can strongly depend on the allocation decisions of the male. However, direct experimental evidence showing that increased male investment in sperm competition results in reduced genetic quality of offspring remains very scarce indeed.

Here we test this prediction using experimental evolution lines of the seed beetle *Callosobruchus maculatus*, a model organism for sexual selection where sperm competition is rife <sup>8,66–70</sup>. These lines have been maintained for >50 generations under three alternative mating regimes manipulating the relative strength of natural and sexual selection: natural polygamy applying both natural and sexual selection (N+S regime), enforced monogamy applying natural selection only (N regime), or a sex-limited middle class neighborhood breeding design <sup>71,72</sup> applying sexual selection only (S regime). Previous findings suggest that S-males pass on a greater genetic load

5

to their progeny if having engaged in socio-sexual interactions prior to being challenged with a dose of irradiation introducing DNA-damage in their germline, an evolved response to socio-sexual interactions not seen in N- or N+S-males<sup>8</sup>. As the S-regime does not show any strong signs of decline in other fitness traits when assayed at standard conditions <sup>73,74</sup>, a plausible explanation for the result is that S-males have evolved reduced germline maintenance as a response to increased post-copulatory sexual selection coupled with weakened constraints on the evolution of male sperm and ejaculate traits.

## Results

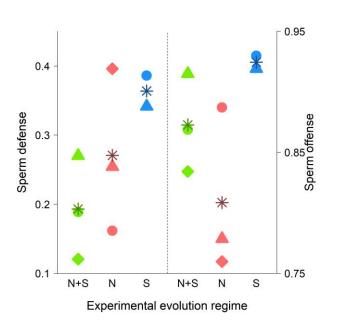
#### Sperm competition

To test whether the removal of natural selection in S-males has led to reduced constraints and evolution of traits ensuring higher post-copulatory competition success, we assayed male sperm competition success in defense (P1: focal male is first to mate) and offense (P2: focal male is second to mate) for all three evolution regimes. Females from the ancestral population, from which the experimental evolution lines were derived, were mated twice (once to a focal male and once to a competitor) with 24 h in between matings, during which time they were provided with beans for egg laying. The competitor males were from a black mutant strain <sup>75</sup> such that paternity in offspring could be determined. Focal males were either held singly or in groups of five males from the same line prior to mating, and for sperm offense males were also tested in 5 consecutive matings to determine sperm and seminal fluid depletion patterns.

Sperm competition success was highest in S-males for both, sperm offense and defense (Fig. 1). S-males had a significantly higher overall sperm competition success compared to N-males ( $P_{MCMC}$  = 0.018) and marginally higher success compared to N+S males ( $P_{MCMC}$  = 0.078). We found no indication that the advantage of S-males depended on being previously exposed to male competitors or having been mated repeatedly prior to the trial. Similarly, the sperm competition advantage of S-males was not significantly different for offense and defense (for detailed statistics see Supplement). Thus, rather than leading to evolution of specific post-copulatory

7

traits, removal of natural selection led to an overall increase in post-copulatory reproductive success (Fig. 1, Supplementary Information S2).

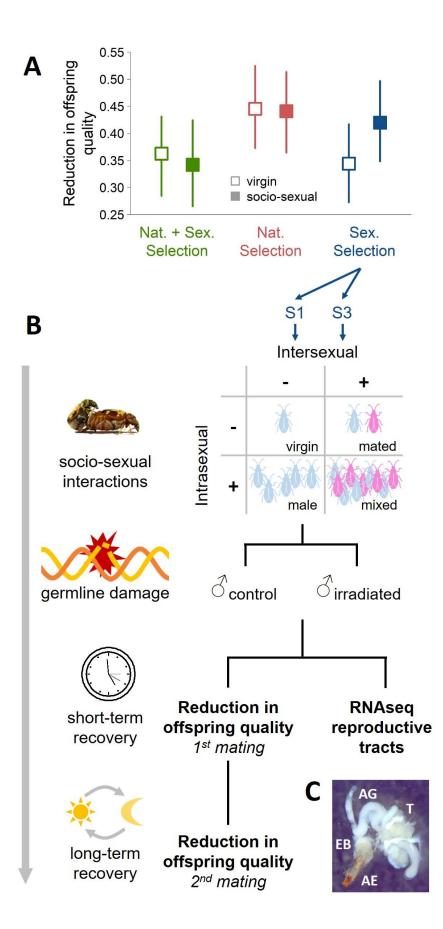


**Figure 1:** Sperm competition success in males from the experimental evolution lines. Line specific sperm defense (paternity share when male is first to mate, P1) and offense (paternity share when male is second to mate, P2) success in double mated females. Asterisks depict means of experimental evolution regimes.

## Germline maintenance

## **Offspring quality**

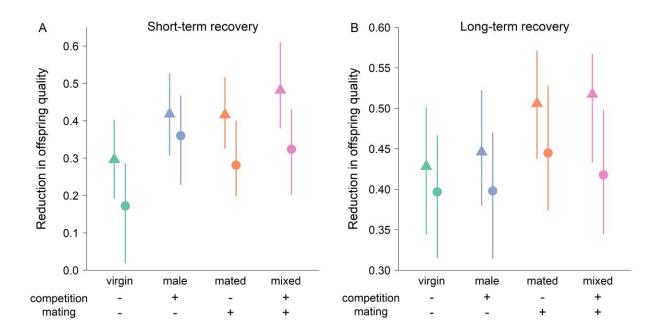
Since S-males evolved enhanced post-copulatory competitiveness, we hypothesized that they evolved to invest more into mating and competition than N- and N+S-males at a potential cost of reduced germline maintenance. Indeed, S-males have been previously shown to reduce germline maintenance when engaging in inter-and intrasexual interactions with conspecifics (Fig. 2A and Baur and Berger<sup>8</sup>). To dissect the effects of inter-and intrasexual interactions on germline maintenance, we manipulated the social environment of S-males in a full-factorial design (with or without male competitors and with or without female mating partners, Fig. 2B). We then measured the reduction in offspring quality for those males after a short (~ 3h) and long (~24h) recovery period after induction of germline damage through gamma radiation. To this end, we mated males to a single virgin female at each time point and established a second generation from the resulting offspring to estimate the quality of offspring from F0 irradiated fathers by counting the number of F2 progeny produced in those lineages relative to F2 progeny production in lineages deriving from unirradiated F0 control males. Hence, the reduction in offspring quality could be calculated as: 1-[F2<sub>IRRADIATED</sub>/F2<sub>CONTROL</sub>].



**Figure 2:** Overview of main experiment (germline maintenance). (A) Reduction in offspring quality after induction of germline DNA damage through irradiation of male beetles. Virgin males (open symbols) with an evolutionary history of sexual selection (N+S- and S-males) suffer less reduction in offspring quality than males from lines with only natural selection acting (N-males). However, socio-sexual interactions prior to the challenge to the germline decreases offspring quality further in S-males but not in N+S- and N-males (closed symbols). Data represent posterior means and 95% confidence intervals from Baur and Berger<sup>8</sup>. (B) Schematic overview of the main experiment. Males from two S-lines were exposed to one of four socio-sexual environments, manipulating the presence of conspecific males and females. Afterwards, we induced germline damage via gamma radiation and determined reduction in quality of offspring produced by those males after a short (~3h) and long (~24h) recovery period. Additionally, we examined gene expression in male reproductive tracts at the end of the short recovery period. (C) Picture of a male reproductive tract. In *C. maculatus* the male reproductive tract consists of the aedagus (AE), ejaculatory bulb (EB), five accessory gland (AG) pairs (two large and three small AG pairs) and a pair of bilobed testes (T). For the gene expression the two large AG pairs were not included.

In agreement with the previously reported negative effect of socio-sexual interactions on offspring quality in S-males<sup>8</sup>, males in the mixed treatment, including both, male-male and male-female (mating) interactions, consistently fathered offspring of lower quality in both matings and for both lines (Fig. 3). Importantly, we find a decrease in offspring quality due to independent effects of both, male-male interactions (Irradiation × Intrasexual interactions:  $P_{MCMC} = 0.010$ ) and mating (Irradiation × Intersexual interactions:  $P_{MCMC} = 0.088$ ), after a short-term recovery period (Fig. 3A). After a long-term recovery period (Fig. 3B), with males being held in isolation during the 24 h post irradiation resting period, the effect of male-male interactions is no longer detectable (Irradiation × Intrasexual interactions:  $P_{MCMC} = 0.030$ ). For both recovery periods, there was no significant interaction effect of inter- and intrasexual interactions on offspring quality (Irradiation × Intrasexual:  $1^{st}$  mating:  $P_{MCMC} = 0.454$ ,  $2^{nd}$  mating:  $P_{MCMC} = 0.796$ ). While the two lines differed overall in the reduction of offspring quality, the response to the socio-sexual

treatments was similar in the two lines (Fig. 3, for detailed statistics see: Supplementary Information S3 and S4).



**Figure 3:** Reduction in offspring quality (1-[F2<sub>IRRADIATED</sub>/F2<sub>CONTROL</sub>]) for first (A) and second (B) mating of males from two experimental evolution lines (S1: triangles, S3: circles) with an evolutionary history of intense sexual selection. Males were held in one of four different social environments before irradiation: 1. Solitary, without any competitors or mating partners (virgin, green symbols), 2. without mating partners but with 4 male competitors (male, blue symbols), 3. without competitors but with one female mating partner (mated, orange symbols), or 4. with 4 male competitors and 5 female mating partners (mixed, pink symbols). Values are posterior means and 95% highest posterior density intervals.

## **Differential gene expression**

To explore the molecular underpinnings of the reduction in germline maintenance, we took a subset of males from all treatment groups to analyze gene expression in the male reproductive tracts (Fig. 2C) after short term recovery. Analyzing all 12161 genes expressed in our data set, we found strong differences between the two experimental evolution lines (5910 DEGs, 49 %, Table

1) reflecting that these lines have been evolving separately for more than 50 generations. When looking at the effect of inter- and intrasexual interactions, most differences occurred in the environment that combined both types of interactions (mixed, 3418 DEG, 28 %, Table 1), with the effect of mating (2747 DEGs, 23 %, Table 1) contributing more than the effect of male-male competition (2 DEGs, < 1 %, Table 1). Irradiation resulted in only very few gene expression changes (18 DEGs, < 1 %, Table 1) and only one of those showed a larger than two-fold change (Table S1, Fig. 4B), which may in part be due to the timing of the measurements. Nevertheless, among the up-regulated genes, three (CALMAC LOCUS18783, CALMAC LOCUS9511 and CALMAC LOCUS2860) code for proteins containing a MADF domain <sup>76</sup>, which can also be found in the putative transcription factor stonewall in Drosophila melanogaster, that is involved in female germline stem cell maintenance 77. Also upregulated is the gene CALMAC\_LOCUS8201 coding for a protein containing a ULP protease domain <sup>76</sup>; Ulp1 in yeast is involved in the sumoylation dynamics that play a critical role in DNA damage response, specifically in the repair of doublestrand breaks <sup>78</sup>. Among the down-regulated genes, CALMAC\_LOCUS9612 codes for a protein containing a BIR domain <sup>76</sup>, which can for example be found in the *D. melanogaster* apoptosis inhibitors lap1 and lap2 79.

<b>Table 1:</b> Summary of differential gene expression analysis for the contrasts of interest, significance at 5
% False Discovery Rate

	Line	Irradiation	Male	Mated	Mixed
	(MA3 – MA1)	(irrad control)	(male - virgin)	(mated - virgin)	(mixed - virgin)
down	2867	8	2	1440	1720
not sign.	6251	12143	12159	9414	8743
up	3043	10	0	1307	1698

Moreover, there was a considerable overlap between genes responding to irradiation and to the social environments (specifically those including intersexual interactions, Fig. S2). To explore effects of the social environment on irradiation responsive genes, we tested the 18 irradiation response candidate genes (Fig. 4A) in a MANOVA. Here, we took advantage of our full-factorial design and tested the interaction between intersexual interactions, intrasexual interactions and irradiation (Table 2). Both, inter- and intrasexual interactions significantly influenced overall expression of irradiation responsive genes independently (Table 2). Furthermore, intersexual interactions even significantly affected the irradiation response itself (Table 2).

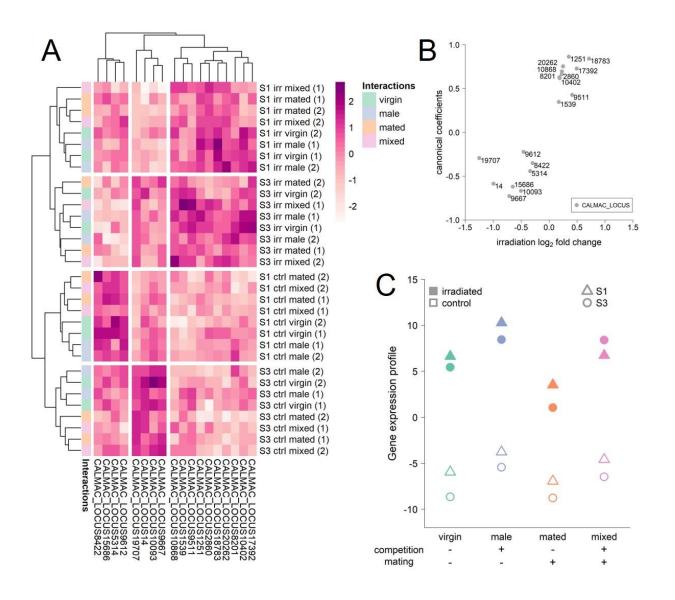
**Table 2:** Test statistics of a multivariate analysis of variance with all 18 irradiation response genes as

 dependent variables

	df	Pillai's trace	Р
Line	1	0.98861	< 0.001
Irradiation	1	0.99632	< 0.001
Inter(sexual interactions)	1	0.98812	< 0.001
Intra(sexual interactions)	1	0.93388	0.032
Irradiation × Inter	1	0.96718	0.005
Irradiation × Intra	1	0.62005	0.852
Inter × Intra	1	0.70972	0.662
Irradiation × Inter × Intra	1	0.77902	0.452

To further explore this link, we first conducted a canonical discriminant analysis to find a linear combination of expression values of irradiation responsive genes that best separates the irradiation and control samples. To get the best representation of the irradiation effect while avoiding overfitting the data, we controlled for variation due to line, social environment and day and limited our interpretation to the first canonical axis. As expected, canonical coefficients for the 18 candidate genes roughly followed the log<sub>2</sub> fold change induced by irradiation (Fig. 4B). The resulting canonical scores of samples not only separated irradiated from control samples, but

also showed differences between the social environments (Fig. 4C). For example, males with intrasexual interactions already have slightly more "irradiation-like" canonical scores in controls, thus without any germline damage induction. This may indicate that they already have an increased need for germline maintenance due to increased investment in reproduction or other stressors linked to male-male competition.



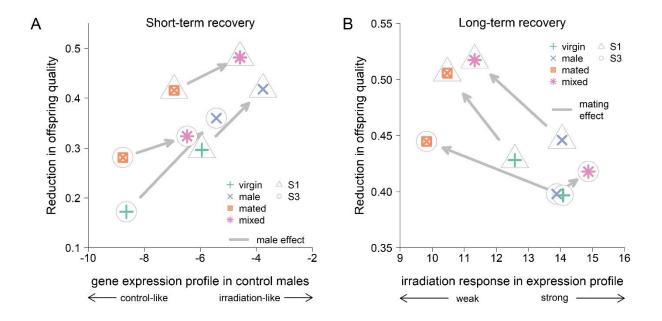
**Figure 4:** Gene expression in the reproductive tracts of S-males. (A) Heatmap of scaled normalized log<sub>2</sub> expression of the 18 genes that responded to the irradiation treatment, expression is clearly separated between irradiation treatments (ctrl: control, irr: irradiated) and experimental evolution lines (S1, S3), within these blocks a separation between mated (orange and pink) and non-mated (green and blue) males can be observed. (B) Canonical coefficients of the 18 irradiation responsive genes that make up the canonical scores of each sample against their log<sub>2</sub> fold change in response to irradiation. (C) Canonical scores separating control (open) and irradiated (closed) samples based on expression of the 18 irradiation responsive genes (triangles: S1; circles: S3).

To determine whether the gene expression profiles of fathers from our treatment groups predicted the quality of their offspring, we applied a canonical correlation analysis. Here, gene expression canonical scores from FO fathers of the two lines and four socio-sexual treatments were entered as x variables and the corresponding measures of reduction in offspring quality after the short and long recovery period as y variables resulting in 8 independent samples with 2 explanatory (gene expression) and 2 response (offspring quality) variables. Using Fapproximations of Pillai-Bartlett's trace both canonical dimensions taken together are significant (F = 7.10, df1 = 4, df2 = 10, P = 0.006) with the second dimension significant on its own (F = 6.37, P = 0.006)df1 =1, df2 = 14, P = 0.024). From the canonical coefficients for dimension 1 (Table 3) it can be concluded that more irradiation-like gene expression profiles (those having a higher canonical score, see Fig. 4C) of control samples were associated with an increased reduction in the quality of offspring resulting from the first mating, taking place shortly after induction of DNA damage (Fig. 5A). This dynamic was thus strongly influenced by male-male interactions, which drove changes in offspring quality after short-term recovery. For dimension 2, canonical coefficients (Table 3) describe a correlation between reduction in quality of offspring resulting from the second mating, following long-term recovery from induction of DNA damage, and the magnitude of the gene expression response to irradiation found in fathers. This thus provides direct evidence

that offspring quality is connected to the capacity of the fathers to change their gene expression in order to deal with the induced damage with stronger responses mitigating the consequences of germline damage (Fig. 5B). Here, mating rather than male-male interactions were the main drivers of differences between socio-sexual treatments.

Table 3: Canonica	l correlation	coefficients
-------------------	---------------	--------------

	Raw coe	fficients	Standardized coefficients			
	Dimension 1	Dimension 2	Dimension 1	Dimension 2		
	(corr.: 0.92)	(corr.: 0.79)	(corr.: 0.92)	(corr.: 0.79)		
Gene expression						
Control samples	-0.83	-0.39	-1.48	-0.70		
Irradiated samples	0.25	0.49	0.74	1.46		
Offspring quality						
Short-term recovery	-11.71	10.38	-1.14	1.01		
Long-term recovery	4.27	-33.16	0.19	-1.51		



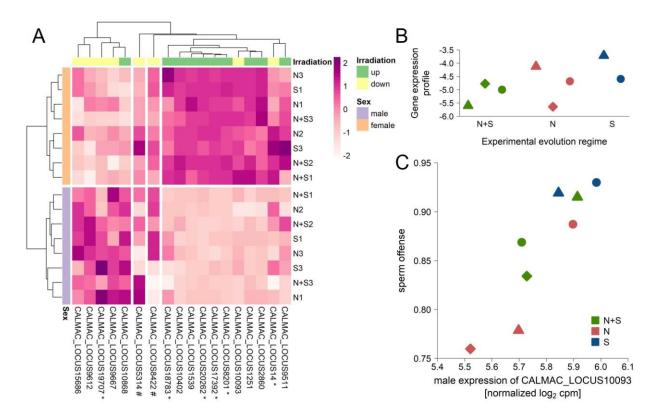
**Figure 5:** Relationship between gene expression profile (as canonical scores, see Fig. 4) and reduction in offspring quality fathered by irradiated males. (A) As indicated by the first dimension of the canonical correlation analysis, stronger reduction in offspring quality after a short-term recovery can be connected to a more irradiation-like gene expression profile in control males (corresponding to baseline expression before irradiation), with male-male interactions leading to more irradiation-like gene expression profiles and stronger reduction of offspring quality. Arrows indicate the effect of adding males to the socio-sexual environment. (B) According to the second dimension of the canonical correlation analysis, larger gene expression response to irradiation (seen in unmated males), led to a smaller reduction in offspring quality. Arrows indicate the effect of a smaller reduction in offspring quality. Arrows indicate the effect of action analysis, larger gene expression response to irradiation (seen in unmated males), led to a smaller reduction in offspring quality. Arrows indicate the effect of adding females (and thus mating opportunities) to the socio-sexual environment.

## Expression of irradiation responsive genes in experimental evolution lines

It has been previously established that males from the different selection regimes differ in their capacity to maintain their germline after induced damage (Fig. 2A and Baur and Berger<sup>8</sup>). We therefore compared the expression of the 18 irradiation responsive genes across all 8 experimental evolution lines. To this end, we analyzed available data that focused on the reproductive tissue. These data were RNA sequences from male and female abdomens from the experimental evolution lines, taken 24 h after a single mating with females having access to beans and males being held in groups of five males during the 24 h period.

We first calculated canonical scores for males from all experimental evolution lines using the canonical coefficients from our previous canonical discriminant analysis (Fig. 4B). While there was a tendency for S males to have a more irradiation-like gene expression profile than N and N+S males (Fig. 6B), we found no significant difference between the three experimental evolution regimes (ANOVA:  $F_{2,5} = 1.02$ , P = 0.425), which in part may reflect the low statistical power for the test, comparing regimes represented by only 2 or 3 replicate lines. In contrast, when analyzing sex differences across all 8 replicate lines, all but two genes showed a significant differential expression between males and females (Table S2), though only 6 of them (see gene names with asterisks in Fig. 6A) pass the two-fold threshold for sex-biased expression. Genes upregulated due to irradiation tend to be female-biased and genes that are downregulated due to irradiation tend to be female-biased and genes that are downregulated females generally invest more heavily in germline maintenance than males do.

Our data are consistent with a trade-off between germline maintenance and investment in postcopulatory traits conferring advantages in sperm competition; thus, we were interested in whether any of the irradiation responsive genes might be involved in sperm offense or defense. Therefore, we tested whether male expression of any of the 18 genes correlated with the estimated sperm competition ability (Fig. 1) in the 8 experimental evolution lines (Table S3 and S4). After p-value correction for multiple testing, one gene (*CALMAC\_LOCUS10093*) retained a significant positive correlation with sperm offense (Fig. 6C). A higher expression of this gene is strongly statistically associated with a higher sperm offense success in *C. maculatus* males, while being downregulated in response to germline-damage. Since the predicted protein for this gene contains a *TFIIS N-terminal* domain <sup>76</sup>, it is likely to be a transcription factor and thus may have a regulatory role in mediating the trade-off between investment into sperm competition and germline maintenance.



**Figure 6:** Expression of the 18 irradiation responsive genes in the 8 experimental evolution lines. (A) Heatmap of scaled normalized log<sub>2</sub> expression values. Samples are separated by sex (females: orange; male: purple) and genes are separated by sex-bias, which roughly coincides with the direction of irradiation response (upregulated: green; downregulated: yellow). All but two genes (#, middle) show a significant differential expression between males and females, 6 genes show a significant sex-bias of at least twofold difference (\*). Genes being upregulated in response to irradiation tend to be female-biased (right block), while genes being downregulated in response to irradiation tend to be male-biased (left block). (B) Scores (based on canonical coefficients used previously to separate control and irradiated samples) of male samples from the experimental evolution lines (24 h after a single mating). Higher scores indicate a more irradiation-like gene expression profile. (C) Positive correlation between normalized log<sub>2</sub> expression of the irradiation responsive gene *CALMAC\_LOCUS10093* (down-regulated in response to germline damage) and sperm offense (P2) ability of males from the experimental evolution lines. Experimental evolution regimes: N+S – polygamy - natural and sexual selection acting; N – monogamy - only natural selection acting; S - male-limited - mainly sexual selection acting.

## Discussion

We hypothesized that male mutation rate and resulting offspring quality can be governed by male strategies balancing the competing needs for post-copulatory reproductive success and germline maintenance. We therefore predicted that the removal of constraints imposed by natural selection would lead to the evolution of increased male reproductive competitiveness at the cost of germline maintenance under intense sexual selection.

Here, we first confirmed a key expectation under this prediction by showing that the relaxation of natural selection while maintaining strong sexual selection (in S-males) indeed resulted in the evolution of increased post-copulatory reproductive success (compared to N- and N+S-males). We then showed that previous findings, demonstrating that S-males pass on more DNA damage to their progeny if having engaged in socio-sexual interactions<sup>8</sup>, are repeatable and that malemale interactions (without mating) can be enough to elicit this response. We sequenced male reproductive tissue and identified 18 candidate genes that show differential expression in response to induced damage in the germline of S-males. Many of those genes' predicted protein products contain domains implicating their involvement in cellular maintenance and DNA repair. The 18 candidate genes also show significant expression changes across socio-sexual treatments and mating generally limited their damage-response, implicating their involvement in a trade-off between germline replication and maintenance. We found that the expression of these candidate genes in the reproductive tissue of fathers predicted the reduction in quality of their progeny brought about by the induced germline damage. Furthermore, we identified one gene whose expression was strongly positively correlated to sperm offense success but down-regulated in

response to germline damage, suggesting that the gene could play a central role in mediating the trade-off between post-copulatory reproductive success and offspring quality in *C. maculatus*. Our findings thus demonstrate how changes in the relative strength of sexual selection can lead to the evolution of phenotypic plasticity in the male reproductive tract with likely consequences for germline mutation rates and offspring quality.

In line with a growing body of studies, we found considerable within-species plasticity in germline maintenance <sup>7,9,11,80</sup>. Notably, in our study different socio-sexual interactions, that should signal the need to modify reproductive effort to males, caused a shift in males' germline maintenance capacity. Male reproductive investment is known to be responsive to female characteristics such as mating status <sup>81</sup>, and the presence of conspecific males <sup>61,82</sup>, most likely because they serve as cues for predicting the level of competition a male's sperm may encounter<sup>83</sup>. Here, we show that similarly to reproductive effort, male capacity to maintain their germline is plastic and may tradeoff with investment into reproduction. Indeed, there is evidence for germline maintenance to be costly reviewed in 53. For example, in Drosophila melanogaster low-condition females have a lower capacity to repair DNA damage of received sperm <sup>7</sup> and *D. melanogaster* lines constructed to be of lower genetic quality accumulate more mutations than control lines in a mutation accumulation study <sup>15</sup>. In zebrafish, *Danio rerio*, germline-free males have a higher capacity for somatic recovery than germline-carrying fish after being exposed to DNA damage via irradiation <sup>10</sup>. Furthermore, under intense male-male competition, a zebrafish male's ejaculate contains more sperm showing signs of DNA damage compared to ejaculates from males held under low competition conditions <sup>65</sup>. Thus, resource allocation decisions could explain the observed decrease in male germline maintenance after inter- and intrasexual interactions in C. maculatus.

Our gene expression data offer potential mechanistic insights into the allocation trade-off between sperm competition success and germline maintenance and the plasticity of the reproductive tract in response to the socio-sexual environment. The observed reductions in quality of offspring fathered by males engaging in socio-sexual interactions could theoretically result from an increase in sperm production while keeping maintenance constant, rendering more replication errors per gamete. The gene expression data, however, indicate that these males also have a decreased capacity to respond to DNA damage. Indeed, this result is also in line with previous finding from these lines showing that reductions in offspring quality under inflated levels of germline damage is not a simple relationship of either sperm age or sperm maturation rate, but might reflect allocation decisions affecting the maintenance of the germline <sup>8</sup>. While the need for investment into reproduction after mating is obvious as males need to replenish sperm and seminal fluid proteins, encounters with rival males do not result in an expenditure of those components, and thus may not necessarily stimulate further production of sperm and seminal fluid. However, encounters with conspecific males can serve as a signal for increased sexual competition that might warrant an increased investment into sperm and seminal fluid proteins that enhance post-copulatory fertilization success. Indeed, in D. melanogaster, male-male interactions change a male's gene expression in the somatic (head/thorax) and reproductive (abdomen) tissue <sup>84</sup>. Though these changes were highly variable between replicates, upregulation of several ejaculate component genes was found <sup>84</sup>, indicating that males increase the production of seminal fluid proteins in response to perceived risk of sperm competition.

A trade-off between germline maintenance and competitive reproductive success offers an explanation for male mutation bias beyond differences in germline replication rate <sup>8,44,46</sup>, since sexual selection acts more strongly in males in most species <sup>85,86</sup>, including *C. maculatus* <sup>87,88</sup>. In accord with this hypothesis, our gene expression data indicate that C. maculatus females have an overall higher investment into germline maintenance. Data from other species are scarce, but there is some correlative comparative evidence to support a trade-off between germline mutation rate and post-copulatory reproductive investment. For example, testes mass - a possible adaptation to sperm competition - and substitution rates have been shown to covary in primates <sup>89</sup>, and across bird taxa estimates of mutation rate have been shown to correlate with extra-pair paternity (as a measure of sperm competition intensity), but notably not with relative testes mass <sup>45</sup>. Additionally, reproductive demands in males can be highly variable, preventing males from evolving an optimal resource allocation strategy <sup>90</sup>. In conclusion, the evolution of phenotypic plasticity in germline maintenance in response to investment in sexual competition, as demonstrated here by experimental manipulation, might help to explain both, systematic differences in mutation rate between the sexes as well as among male variation within species.

## Acknowledgements

We thank P. E. Eady for sharing the *C. maculatus* black line with us and J. Liljestrand-Rönn for help in the lab. Sequencing was performed by the SNP&SEQ Technology Platform in Uppsala. The facility is part of the National Genomics Infrastructure (NGI) Sweden and Science for Life Laboratory. The SNP&SEQ Platform is also supported by the *Swedish Research Council* and the *Knut and Alice Wallenberg Foundation*. This work was supported by the *Carl Tryggers Stiftelse för Vetenskaplig Forskning* (grant no. CTS18:32), by the *Vetenskapsrådet* (grant no. 2019-05024), and by the *Fysiografiska sällskapet i Lund*.

## **Author Contributions**

MK, JB and DB designed the study, MK and JB conducted the research, MK and DB analysed the data and drafted the manuscript. All authors contributed to the final version of the manuscript.

## **Declaration of interests**

The authors declare no competing interests.

## **Materials and Methods**

#### <u>Beetles</u>

The seed beetle Callosobruchus maculatus develops on the seeds of legumes. All beetles for the experiments were reared and kept on black-eyed beans (Vigna unquiculata) in constant climate chambers at 29 °C, 50 % relative humidity and a 12:12 L:D cycle. Where applicable, we used beetles from the ancestral Lome population see: 91,92 as standardized mating partners and males from a black *C. maculatus* line as competitors <sup>75</sup>. Focal individuals came from 8 experimental evolution lines that originated from the *Lome* population and are described in detail in <sup>8,74</sup>. In short, those beetles evolved for > 50 generations under one of three experimental evolution regimes: N+S beetles evolved under polygamy with opportunities for natural (N) and sexual (S) selection to act, N beetles evolved under enforced monogamy with sexual competition between males removed and thus mainly natural (N) selection acting, S beetles evolved under polygamy but with a middle-class neighborhood breeding design applied to females weakening natural selection and leaving mainly sexual (S) selection to act. For the germline maintenance experiment, we focused on males from the two S lines, which were both mated to females from a third (polygamous) line to exclude any female derived and/or co-evolution effects. For the sperm competition experiment and the second gene expression dataset we included beetles from all 8 experimental evolution lines.

#### Sperm competition

Males from all 8 experimental evolution lines were tested for the sperm competitiveness when being first (sperm defense, P1) or second (sperm offense, P2) to mate with a double mated female. To avoid potential confounding effects of females, we used females from the ancestral *Lome* population that were mated to two males (observed single matings) 24 hours apart. Focal males were held in one of two social environments for approximately 24 h before their mating: solitary (single males in 30 mm dishes) or competition (in groups of five males in 90 mm dishes). The experiment was conducted twice. Within each of the two blocks, lines were separated into one of three sub-blocks, two sub-blocks contained one line from each of the three experimental evolution regimes and the last block contained the third replicate line of the N+S and N regime.

For sperm defense, females were mated to a focal male from one of the experimental evolution lines in a single observed mating and afterwards kept on beans for 24 h before their second mating. Beans were incubated for approximately 30 days and then frozen at -20 °C to determine 24 h offspring production elicited by the first mating. 24 h after the first mating all females were given the opportunity to mate with a black competitor male within 40 minutes. Successful mating pairs were separated and females were moved onto fresh beans for 48 h. Beans were incubated for approximately 30 days and frozen at -20 °C, afterwards offspring was counted separated by black and wildtype fathers based on their coloration.

For sperm offense, males were consecutively mated five times to already mated females in one of the two blocks to determine the rate of decline in sperm competitiveness. Females were first

28

mated to a black male in observed single matings and kept on beans for egg laying in groups of max. 25 for 24 h. Afterwards females were given the opportunity to mate with one focal male from the experimental evolution lines for 40 min. Successful pairs were separated, females from matings 1,3 and 5 were put on fresh beans for 48h to determine focal male sperm offense success. Beans were incubated for approximately 30 days and frozen at -20 °C. Afterwards, offspring was counted separated by black and wildtype fathers based on their morphology.

Statistical analyses and preparation of graphs were done in R 4.1.1 <sup>93</sup> using Bayesian Generalized Linear Models implemented within the package *MCMCgImm* <sup>94</sup>. Proportion of the focal (wildtype) father's offspring was modelled with Binomial error distribution corrected for overdispersion. We included experimental evolution regime as well as its interaction with competition mode (sperm offense or defense), number of mating (three-level factor), and social environment (twolevel factor) as fixed effects. To ease interpretation, non-significant interaction terms were removed stepwise. We included experimental evolution line crossed with competition mode, number of mating, and social environment as random terms. As additional random terms, we included experimental block, experimental evolution line, and male ID. For graphical presentation, line specific defense and offense success for a focal male's first mating were calculated in separate models using package *MCMCgImm* <sup>94</sup> including block and social environment as random.

## Germline maintenance

## **Experimental assay**

In order to measure germline maintenance capacity, we induced DNA-damage in adult males by exposing them to gamma radiation (for 35 min at a dose rate of 0.72 Gray/min from a cesium-137 source). Gamma radiation causes double and single strand DNA breaks as well as increases the amount of reactive oxygen species in cells <sup>40</sup>, which in turn can induce further DNA damage <sup>40,54</sup>. While our treatment drastically increased germline damage, DNA breaks occur naturally during both recombination and chromatin remodeling during sperm development, and errors in the repair of those breaks give rise to point mutations <sup>35,40,95</sup>. The number of DNA lesions induced by a given dose of gamma radiation is surprisingly constant per DNA base pair <sup>95</sup>, and thus differences in mutation rate are mainly caused by to the amount and type of repair molecules <sup>40,95,96</sup>, which makes this assay ideal for measuring germline maintenance. Because most mutations are neutral or deleterious <sup>22,97</sup>, the amount of mutations transferred from parents to offspring can be approximated by the decline in offspring quality of parents that were challenged to deal with elevated levels of reactive oxygen species and DNA damage <sup>7–9,11,16</sup>.

Assays were replicated on two consecutive days. Males and females (from the two S-lines) and females (from a third, polygamous line) were picked as virgins within 24 h after emergence. Females (from the third, polygamous line) were held in groups of ten in petri dishes (90 mm) until mating assays and males were immediately transferred to their respective socio-sexual environment using females from their own experimental evolution line where applicable. Socio-

30

sexual treatments were set up by manipulating the presence of conspecific males and females in a full factorial design. Males were held in a 35 mm petri dish without any conspecifics or with a single virgin female, or in a 90 mm dish together with four conspecific males or with 4 conspecific males and an additional 5 virgin females. Males were held in their respective socio-sexual environment for approximately 24 h until shortly (< 1 h) before the irradiation treatment. Then, males were separated into individual 0.5 ml reaction tubes with a hole punched into the lid. Roughly half of the males then underwent a radiation treatment (a dose of 25 Gray of gamma radiation from a cesium-137 source over 35 min) while the other half served as controls.

#### **Offspring quality**

Germline maintenance was assessed by measuring fitness effects of the induced germline damage in subsequent generations. Shortly after irradiation (1.5-3 h day 1, 2.5-4 h day 2) males were mated once to a single virgin female (0-48 h old) in a 60 mm petri dish on a heating plate set to 29 °C. Females were put on beans to lay eggs for 72 h and males remained in their individual petri dishes to renew their ejaculate, thus making sure that all males were challenged to deal with the competing tasks of both replicating and maintaining their germline. One day after irradiation (22-24 h day 1, 22-23 h day 2) males were again mated to a single virgin female (24-48 h old) in 60 mm dishes on a heating plate. Females were put on beans for 72 h to lay eggs and males were discarded. All beans were incubated at 29°C, 50% r. h. and 12:12 L:D cycle in a climate chamber for 30 days to ensure that all viable offspring had emerged. Before offspring eclosion, beans were transferred to virgin chambers. Some of the offspring were used in the assay below.

Remaining offspring were frozen at -20 °C and then counted to determine male offspring production from their first and second ejaculate.

To determine the reduction in quality of offspring fathered by irradiated males, we crossed the  $F_1$  offspring of each male with the  $F_1$  offspring of other males within the same treatment, experimental evolution line, and experimental day using a Middle-Class Neighbourhood breeding design (relaxing selection on the induced mutations). For the first ejaculate, we aimed at crossing one  $F_1$  male and one  $F_1$  female per male. For the second ejaculate, we aimed at crossing 3  $F_1$  males and 3  $F_1$  females, as we wished to focus on the recovery of the male germline. Pairs were kept on beans for their entire life and we incubated dishes for 33 days before freezing them at - 20°C to count  $F_2$  production. Counts of  $F_2$  adult offspring emerging from these irradiated lineages (n = 224) were compared to counts from corresponding control lineages (n = 163) to calculate reduction in offspring quality as:  $1-[F2_{IRRADIATED}/F2_{CONTROL}]$ . Thus, we could explore phenotypic plasticity in germline maintenance in response to the socio-sexual treatments by comparing reduction in quality of offspring from grandfathers kept under the four treatments (Fig. 2B).

Again, we used Bayesian Generalized Linear Models implemented within the package *MCMCgImm*<sup>94</sup> in R 4.1.1<sup>93</sup> for statistical analyses. Number of F<sub>1</sub> and F<sub>2</sub> offspring were modelled with Poisson error distribution corrected for overdispersion, for analysis of F<sub>2</sub> offspring, dam and sire (IDs of the two grandfathers) were entered as a multiple-membership random term. Socio-sexual interactions were modelled as two two-level fixed effects (Inter- and Intrasexual interactions) testing for a significant interaction with irradiation treatment. We also added experimental evolution line and day as fixed effects to test for any differences between the two lines and the two assay days. To ease interpretation, non-significant interaction terms were

removed. For graphical presentation, line specific means (and their 95% HPD interval) of the reduction in offspring quality were calculated per socio-sexual environment with Bayesian Generalized Linear Models and a Gaussian distribution. Similarly, those values were calculated for each of the two assay days separately for use in further analyses. Packages *Hmisc* <sup>98</sup> and *RColorBrewer* <sup>99</sup> were used to generate graphs.

## **Gene expression**

For RNA extraction, beetles were snap frozen 2 h after irradiation treatment and stored at -80°C until dissections. During dissections, beetles and dissected tissues were kept on dry ice and afterwards stored at -80°C until RNA extraction. Males were dissected on ice in a droplet of PBS, the entire reproductive tract (Fig. 2C) was removed and the two large accessory gland pairs cut off. We decided to remove the two large accessory gland pairs in order to keep dissections consistent, as the large accessory glands easily detach and/rupture during dissections. Afterwards the remaining tissue (aedeagus, ejaculatory bulb, 2 bilobed testes and 3 pairs of smaller accessory glands [ectadenial glands] <sup>100</sup>), was quickly rinsed in a fresh droplet of PBS and then transferred to a reaction tube on dry ice. We aimed to pool tissue from 10 males per sample, for two samples (1 mated irradiated line S3 and 1 mated control line S3) we only obtained tissue from nine males. Each sample consisted only of males from the same treatment, line and experimental day. This resulted in a total of 32 samples with 4 replicates per treatment (1 per day and line).

RNA was extracted with Qiagen RNeasy Mini Kit and on-column DNA digestion was performed with Qiagen RNase free DNase Kit. We followed the manufacturer's instructions, beta-mercapto-Ethanol was added to the lysis buffer, tissue lysis was done with one stainless steel bead in a bead mill at 28 Hz for 90 s. Two samples underwent an additional clean-up using the Qiagen RNeasy Mini Kit. RNA concentration and purity were assessed with NanoDrop and additional quality controls were performed at the sequencing facility. Samples were sequenced at the SNP&SEQ Technology Platform in Uppsala. Libraries were multiplexed and sequenced as paired-end 50 bp reads in 2 lanes of a NovaSeq SP flow cell resulting in roughly 11M-26M reads per sample.

Raw reads were inspected with *FastQC*<sup>101</sup> and quality information summarized with *MultiQC*<sup>102</sup>. We then mapped all reads to the *C. maculatus* transcriptome and genome (GCA\_900659725.1; ASM90065972v1)<sup>103</sup> with *TopHat 2.1.1*<sup>104</sup> allowing for up to two mismatches per read. We only kept reads where both mates successfully mapped to the *C. maculatus* genome. Those reads were then counted per gene using *HTSeq*<sup>105</sup> with default settings. Statistical analyses were done in R 4.1.1<sup>93</sup>, with packages *edgeR*<sup>106</sup> and *limma*<sup>107</sup>. Libraries from the two lanes were merged into one sample. Genes that were not at least expressed as 1 count per million (cpm) in at least 2 samples were excluded from the analysis resulting in a total of 12161 genes being analyzed. Counts were normalized with the 'Trimmed Mean of M-values' method and normalized log<sub>2</sub> cpm values were analyzed in linear models within *limma*<sup>107</sup>. Experimental evolution line and experimental day were added as additive terms to control for variance between lines and days. Socio-sexual environment was entered as a 4-level factor and irradiation treatment, we lacked statistical power to analyze the interaction between social environment and irradiation with the

full set of genes. Therefore, the interaction was removed from the model and we analyzed the interaction in a separate model considering only genes that responded to the irradiation treatment. We used information available on *UniProt* <sup>76</sup> accessed on 10.03.2022 to gain insight on the potential function of some of the genes found to be differentially expressed.

For further analyses we always used normalized log<sub>2</sub> cpm values. Using the 18 irradiation responsive genes, we ran a multivariate Anova. Additionally, we ran a linear discriminant analysis on gene expression in the 18 irradiation responsive genes to find a linear combination of the expression of these genes that best separates irradiated from control samples. To this end we controlled for variation arising through differences in irradiation response between lines, sociosexual environments or experimental days using package candisc <sup>108</sup>. To avoid overfitting the data, we calculated canonical scores of the 32 samples with the first canonical axis only. For further analyses and graphical representation, we used mean canonical scores across the two experimental days. To estimate how well differences in gene expression correspond to differences in reduction in offspring quality due to germline damage, we conducted a Canonical Correlation Analysis. We added canonical scores of irradiated and control samples (averaged across the two experimental days) as two x variables and reduction in offspring quality after a short- and long-term recovery period as two y variables to the Canonical Correlation Analysis implemented in the package CCA <sup>109</sup>. For graphical presentation, heatmaps were constructed on scaled normalized log<sub>2</sub> cpm values using hierarchical clustering and Manhattan distance metrics in *pheatmap*<sup>110</sup>, and additional packages *VennDiagram*<sup>111</sup> and RColorBrewer<sup>99</sup> were used.

## Expression of irradiation responsive genes in experimental evolution lines

## **Experimental assay**

To analyze the expression of irradiation responsive genes in the 8 lines from all 3 experimental evolution regimes, we made use of an existing data set designed to study the evolution of sexbiased gene expression under these selection regimes. Before collecting individuals for sequencing, all experimental evolution lines underwent 3 generations of common garden rearing (i.e., a polygamous mating setting). Males and females from the experimental evolution lines were mated once (observed) to a standardized mating partner from the *Lome* population on heating plates set to 29 °C. Matings were separated into 4 blocks and in each block, we set up 6 mating pairs per line and sex. Beetles from the first 5 successful matings per line and sex were separated after the end of the mating, focal females were kept singly on beans for 24 h and focal males were held together in a 90 mm dish (in groups of 5 individuals) for 24 h.

#### Gene expression

After 24 h, males and females were flash frozen in liquid nitrogen and kept at -80 °C until sample preparation. Since we were interested in the reproductive tissues, we only sampled the abdomen of males and females. To that end, we separated the abdomen from the rest of the body on ice, while storing samples on dry ice during preparation. We pooled 12 abdomen per group balanced over the four blocks (except for the male sample from line N+S2, which only contained 10 abdomen, block information on the two lost abdomen is not available). After dissection, samples were stored again at -80 °C until RNA extraction.

RNA was extracted with Qiagen RNeasy Mini Kit and on-column DNA digestion was performed with Qiagen RNase free DNase Kit. We followed the manufacturer's instructions, beta-mercapto-Ethanol was added to the lysis buffer and tissue lysis was done with two stainless steel beads in a bead mill at 28 Hz for 90 s in 700 µl lysis buffer. After centrifugation the entire supernatant was transferred to a fresh tube, mixed quickly and 350 µl went onto the extraction column while the rest was discarded to avoid overloading the column. RNA was eluted 2 times in 50 µl of RNase free water each. RNA concentration and purity were assessed with NanoDrop, gel electrophoresis and Qbit, additional quality controls were performed at the sequencing facility. Samples were sequenced at the SNP&SEQ Technology Platform in Uppsala. Libraries were multiplexed and sequenced as paired-end 150 bp reads in 1 lane of a NovaSeq S4 flow cell resulting in roughly 24M-56M reads per sample.

Gene counts were obtained and analyzed as in the main experiment with the exception of an additional quality and adapter trimming step with *Trimmomatic*<sup>112</sup> and allowing for up to eight mismatches per read during mapping due to the higher read length. Normalized log<sub>2</sub> cpm counts of all 12874 retained genes were analyzed in a linear model within *limma*<sup>107</sup>. We constructed an additive model with sex (2-level factor) and experimental evolution regime (3-level factor) as explanatory variables. Sex bias was estimated across all experimental evolution regimes (male - female) and p-values were corrected with Benjamini-Hochberg method using a 5% FDR cut-off. We then extracted normalized log<sub>2</sub> cpm values of the 18 irradiation responsive genes for all

samples for further analysis. Using these values, we predicted canonical scores for males from all

experimental evolution lines based on the linear coefficients from the previous analysis.

## References

- 1. Cobben, M.M.P., Mitesser, O., and Kubisch, A. (2017). Evolving mutation rate advances the invasion speed of a sexual species. BMC Evol. Biol. *17*, 150.
- 2. Sprouffske, K., Aguilar-Rodríguez, J., Sniegowski, P., and Wagner, A. (2018). High mutation rates limit evolutionary adaptation in *Escherichia coli*. PLOS Genet. *14*, e1007324.
- 3. Bürger, R., and Lynch, M. (1995). Evolution and extinction in a changing environment: a quantitative-genetic analysis. Evolution *49*, 151–163.
- 4. Lande, R., and Shannon, S. (1996). The Role of genetic variation in adaptation and population persistence in a changing environment. Evolution *50*, 434–437.
- 5. Kimura, M. (1983). The neutral theory of molecular evolution (Cambridge University Press).
- 6. Xu, S., Stapley, J., Gablenz, S., Boyer, J., Appenroth, K.J., Sree, K.S., Gershenzon, J., Widmer, A., and Huber, M. (2019). Low genetic variation is associated with low mutation rate in the giant duckweed. Nat. Commun. *10*, 1243.
- 7. Agrawal, A.F., and Wang, A.D. (2008). Increased transmission of mutations by low-condition females: evidence for condition-dependent DNA repair. PLoS Biol. *6*, e30.
- 8. Baur, J., and Berger, D. (2020). Experimental evidence for effects of sexual selection on conditiondependent mutation rates. Nat. Ecol. Evol. *4*, 737–744.
- 9. Berger, D., Stångberg, J., Grieshop, K., Martinossi-Allibert, I., and Arnqvist, G. (2017). Temperature effects on life-history trade-offs, germline maintenance and mutation rate under simulated climate warming. Proc. R. Soc. B Biol. Sci. *284*, 20171721.
- Chen, H., Jolly, C., Bublys, K., Marcu, D., and Immler, S. (2020). Trade-off between somatic and germline repair in a vertebrate supports the expensive germ line hypothesis. Proc. Natl. Acad. Sci. U. S. A. *117*, 8973–8979.
- 11. Maklakov, A.A., Immler, S., Løvlie, H., Flis, I., and Friberg, U. (2013). The effect of sexual harassment on lethal mutation rate in female *Drosophila melanogaster*. Proc. R. Soc. B Biol. Sci. *280*, 20121874.
- 12. Martincorena, I., Seshasayee, A.S.N., and Luscombe, N.M. (2012). Evidence of non-random mutation rates suggests an evolutionary risk management strategy. Nature *485*, 95–98.
- 13. the 1000 Genome Project, Conrad, D.F., Keebler, J.E.M., DePristo, M.A., Lindsay, S.J., Zhang, Y., Casals, F., Idaghdour, Y., Hartl, C.L., Torroja, C., et al. (2011). Variation in genome-wide mutation rates within and between human families. Nat. Genet. *43*, 712–714.

- 14. Rahbari, R., Wuster, A., Lindsay, S.J., Hardwick, R.J., Alexandrov, L.B., Al Turki, S., Dominiczak, A., Morris, A., Porteous, D., Smith, B., et al. (2016). Timing, rates and spectra of human germline mutation. Nat. Genet. *48*, 126–133.
- 15. Sharp, N.P., and Agrawal, A.F. (2012). Evidence for elevated mutation rates in low-quality genotypes. Proc. Natl. Acad. Sci. U. S. A. *109*, 6142–6146.
- 16. Svetec, N., Cridland, J.M., Zhao, L., and Begun, D.J. (2016). The adaptive significance of natural genetic variation in the DNA damage response of *Drosophila melanogaster*. PLoS Genet. *12*, e1005869.
- Yang, S., Wang, L., Huang, J., Zhang, X., Yuan, Y., Chen, J.-Q., Hurst, L.D., and Tian, D. (2015). Parent–progeny sequencing indicates higher mutation rates in heterozygotes. Nature 523, 463–467.
- 18. Agrawal, A.F. (2002). Genetic loads under fitness-dependent mutation rates. J. Evol. Biol. 15, 1004– 1010.
- 19. Shaw, F.H., and Baer, C.F. (2011). Fitness-dependent mutation rates in finite populations. J. Evol. Biol. 24, 1677–1684.
- 20. McGaughran, A., Laver, R., and Fraser, C. (2021). Evolutionary responses to warming. Trends Ecol. Evol. *36*, 591–600.
- 21. Sniegowski, P.D., Gerrish, P.J., Johnson, T., and Shaver, A. (2000). The evolution of mutation rates: separating causes from consequences. BioEssays *22*, 1057–1066.
- 22. Agrawal, A.F., and Whitlock, M.C. (2012). Mutation load: the fitness of individuals in populations where deleterious alleles are abundant. Annu. Rev. Ecol. Evol. Syst. *43*, 115–135.
- 23. Crow, J.F. (1997). The high spontaneous mutation rate: Is it a health risk? Proc. Natl. Acad. Sci. *94*, 8380–8386.
- 24. Keightley, P.D. (2012). Rates and fitness consequences of new mutations in humans. Genetics *190*, 295–304.
- 25. Lynch, M. (2016). Mutation and human exceptionalism: our future genetic load. Genetics *202*, 869–875.
- 26. Moorjani, P., Gao, Z., and Przeworski, M. (2016). Human germline mutation and the erratic evolutionary clock. PLoS Biol. *14*, e2000744.
- 27. Baer, C.F., Miyamoto, M.M., and Denver, D.R. (2007). Mutation rate variation in multicellular eukaryotes: causes and consequences. Nat. Rev. Genet. *8*, 619–631.
- 28. Chintalapati, M., and Moorjani, P. (2020). Evolution of the mutation rate across primates. Curr. Opin. Genet. Dev. *62*, 58–64.

- 29. MacLean, R.C., Torres-Barceló, C., and Moxon, R. (2013). Evaluating evolutionary models of stressinduced mutagenesis in bacteria. Nat. Rev. Genet. 14, 221–227.
- 30. Bartosch -Harlid, A., Berlin, S., Smith, N.G.C., Mosller, A.P., and Ellegren, H. (2003). Life history and the male mutation bias. Evolution *57*, 2398–2406.
- 31. Ellegren, H. (2007). Characteristics, causes and evolutionary consequences of male-biased mutation. Proc. R. Soc. B Biol. Sci. 274, 1–10.
- 32. Kirkpatrick, M., and Hall, D.W. (2004). Male-biased mutation, sex linkage, and the rate of adaptive evolution. Evolution *58*, 437–440.
- 33. Wilson Sayres, M.A., and Makova, K.D. (2011). Genome analyses substantiate male mutation bias in many species. BioEssays *33*, 938–945.
- 34. Venn, O., Turner, I., Mathieson, I., de Groot, N., Bontrop, R., and McVean, G. (2014). Strong male bias drives germline mutation in chimpanzees. Science *344*, 1272–1275.
- 35. Grégoire, M.-C., Massonneau, J., Simard, O., Gouraud, A., Brazeau, M.-A., Arguin, M., Leduc, F., and Boissonneault, G. (2013). Male-driven *de novo* mutations in haploid germ cells. Mol. Hum. Reprod. *19*, 495–499.
- 36. Ségurel, L., Wyman, M.J., and Przeworski, M. (2014). Determinants of mutation rate variation in the human germline. Annu. Rev. Genomics Hum. Genet. *15*, 47–70.
- 37. Clutton-Brock, T.H., and Parker, G.A. (1992). Potential reproductive rates and the operation of sexual selection. Q. Rev. Biol. *67*, 437–456.
- 38. Schärer, L., Rowe, L., and Arnqvist, G. (2012). Anisogamy, chance and the evolution of sex roles. Trends Ecol. Evol. *27*, 260–264.
- Bergero, R., Ellis, P., Haerty, W., Larcombe, L., Macaulay, I., Mehta, T., Mogensen, M., Murray, D., Nash, W., Neale, M.J., et al. (2021). Meiosis and beyond – understanding the mechanistic and evolutionary processes shaping the germline genome. Biol. Rev. *96*, 822–841.
- 40. Friedberg, E.C., Walker, G.C., Siede, W., Wood, R.D., Schultz, R.A., and Ellenberger, T. (2005). DNA Repair and Mutagenesis 2nd ed. (American Society for Microbiology Press).
- 41. González-Marín, C., Gosálvez, J., and Roy, R. (2012). Types, causes, detection and repair of DNA fragmentation in animal and human sperm cells. Int. J. Mol. Sci. *13*, 14026–14052.
- 42. Lemaître, J.-F., Gaillard, J.-M., and Ramm, S.A. (2020). The hidden ageing costs of sperm competition. Ecol. Lett. *23*, 1573–1588.
- 43. Aitken, R.J., and De Iuliis, G.N. (2010). On the possible origins of DNA damage in human spermatozoa. Mol. Hum. Reprod. *16*, 3–13.
- 44. Blumenstiel, J.P. (2007). Sperm competition can drive a male-biased mutation rate. J. Theor. Biol. 249, 624–632.

- 45. Møller, A., and Cuervo, J. (2003). Sexual selection, germline mutation rate and sperm competition. BMC Evol. Biol. *3*, 6.
- 46. Petrie, M., and Roberts, G. (2007). Sexual selection and the evolution of evolvability. Heredity *98*, 198–205.
- 47. Reinhardt, K. (2007). Evolutionary consequences of sperm cell aging. Q. Rev. Biol. 82, 375–393.
- 48. Segami, J.C., Lind, M.I., and Qvarnström, A. (2021). Should females prefer old males? Evol. Lett. 5, 507–520.
- 49. Kokko, H., and Lindström, J. (1996). Evolution of female preference for old mates. Proc. R. Soc. Lond. B Biol. Sci. *263*, 1533–1538.
- 50. Beck, C.W., and Promislow, D.E.L. (2007). Evolution of female preference for younger males. PLoS ONE *2*, e939.
- Gao, Z., Moorjani, P., Sasani, T.A., Pedersen, B.S., Quinlan, A.R., Jorde, L.B., Amster, G., and Przeworski, M. (2019). Overlooked roles of DNA damage and maternal age in generating human germline mutations. Proc. Natl. Acad. Sci. U. S. A. *116*, 9491–9500.
- 52. Wu, F.L., Strand, A.I., Cox, L.A., Ober, C., Wall, J.D., Moorjani, P., and Przeworski, M. (2020). A comparison of humans and baboons suggests germline mutation rates do not track cell divisions. PLoS Biol. *18*, e3000838.
- 53. Maklakov, A.A., and Immler, S. (2016). The expensive germline and the evolution of ageing. Curr. Biol. *26*, R577–R586.
- 54. Dowling, D.K., and Simmons, L.W. (2009). Reactive oxygen species as universal constraints in lifehistory evolution. Proc. R. Soc. B Biol. Sci. 276, 1737–1745.
- 55. Friesen, C.R., Noble, D.W.A., and Olsson, M. (2020). The role of oxidative stress in postcopulatory selection. Philos. Trans. R. Soc. B Biol. Sci. *375*, 20200065.
- 56. von Schantz, T., Bensch, S., Grahn, M., Hasselquist, D., and Wittzell, H. (1999). Good genes, oxidative stress and condition-dependent sexual signals. Proc. R. Soc. B Biol. Sci. *266*, 1–12.
- 57. Ball, B.A. (2008). Oxidative stress, osmotic stress and apoptosis: impacts on sperm function and preservation in the horse. Anim. Reprod. Sci. *107*, 257–267.
- 58. Ramm, S.A., Schärer, L., Ehmcke, J., and Wistuba, J. (2014). Sperm competition and the evolution of spermatogenesis. Mol. Hum. Reprod. *20*, 1169–1179.
- 59. Evans, J.P., Wilson, A.J., Pilastro, A., and Garcia-Gonzalez, F. (2019). Ejaculate-mediated paternal effects: evidence, mechanisms and evolutionary implications. Reproduction *157*, R109–R126.
- 60. Fitzpatrick, J.L., and Lüpold, S. (2014). Sexual selection and the evolution of sperm quality. Mol. Hum. Reprod. *20*, 1180–1189.

- Hopkins, B.R., Sepil, I., Thézénas, M.-L., Craig, J.F., Miller, T., Charles, P.D., Fischer, R., Kessler, B.M., Bretman, A., Pizzari, T., et al. (2019). Divergent allocation of sperm and the seminal proteome along a competition gradient in *Drosophila melanogaster*. Proc. Natl. Acad. Sci. U. S. A. *116*, 17925–17933.
- 62. Ramm, S.A., Edward, D.A., Claydon, A.J., Hammond, D.E., Brownridge, P., Hurst, J.L., Beynon, R.J., and Stockley, P. (2015). Sperm competition risk drives plasticity in seminal fluid composition. BMC Biol. *13*, 87.
- Sepil, I., Hopkins, B.R., Dean, R., Bath, E., Friedman, S., Swanson, B., Ostridge, H.J., Harper, L., Buehner, N.A., Wolfner, M.F., et al. (2020). Male reproductive aging arises via multifaceted matingdependent sperm and seminal proteome declines, but is postponable in *Drosophila*. Proc. Natl. Acad. Sci. U. S. A. *117*, 17094–17103.
- 64. Simmons, L.W., and Lovegrove, M. (2017). Socially cued seminal fluid gene expression mediates responses in ejaculate quality to sperm competition risk. Proc. R. Soc. B Biol. Sci. 284, 20171486.
- 65. Silva, W.T.A.F., Sáez-Espinosa, P., Torijo-Boix, S., Romero, A., Devaux, C., Durieux, M., Gómez-Torres, M.J., and Immler, S. (2019). The effects of male social environment on sperm phenotype and genome integrity. J. Evol. Biol. *32*, 535–544.
- 66. Eady, P. (1994). Sperm transfer and storage in relation to sperm competition in *Callosobruchus maculatus*. Behav. Ecol. Sociobiol. *35*, 123–129.
- 67. Eady, P.E. (1995). Why do male *Callosobruchus maculatus* beetles inseminate so many sperm? Behav. Ecol. Sociobiol. *36*, 25–32.
- Gay, L., Hosken, D.J., Vasudev, R., Tregenza, T., and Eady, P.E. (2009). Sperm competition and maternal effects differentially influence testis and sperm size in *Callosobruchus maculatus*. J. Evol. Biol. 22, 1143–1150.
- 69. Hotzy, C., and Arnqvist, G. (2009). Sperm competition favors harmful males in seed beetles. Curr. Biol. *19*, 404–407.
- 70. Rönn, J., Katvala, M., and Arnqvist, G. (2007). Coevolution between harmful male genitalia and female resistance in seed beetles. Proc. Natl. Acad. Sci. U. S. A. *104*, 10921–10925.
- 71. Chenoweth, S.F., Appleton, N.C., Allen, S.L., and Rundle, H.D. (2015). Genomic evidence that sexual selection impedes adaptation to a novel environment. Curr. Biol. *25*, 1860–1866.
- 72. Morrow, E.H., Stewart, A.D., and Rice, W.R. (2008). Assessing the extent of genome-wide intralocus sexual conflict via experimentally enforced gender-limited selection. J. Evol. Biol. *21*, 1046–1054.
- 73. Bagchi, B., Corbel, Q., Khan, I., Payne, E., Banerji, D., Liljestrand-Rönn, J., Martinossi-Allibert, I., Baur, J., Sayadi, A., Immonen, E., et al. (2021). Sexual conflict drives micro- and macroevolution of sexual dimorphism in immunity. BMC Biol. *19*, 114.

- 74. Martinossi-Allibert, I., Thilliez, E., Arnqvist, G., and Berger, D. (2019). Sexual selection, environmental robustness, and evolutionary demography of maladapted populations: a test using experimental evolution in seed beetles. Evol. Appl. *12*, 1371–1384.
- 75. Eady, P.E. (1991). Sperm competition in *Callosobruchus maculatus* (Coleoptera: Bruchidae): a comparison of two methods used to estimate paternity. Ecol. Entomol. *16*, 45–53.
- 76. The UniProt Consortium, Bateman, A., Martin, M.-J., Orchard, S., Magrane, M., Agivetova, R., Ahmad, S., Alpi, E., Bowler-Barnett, E.H., Britto, R., et al. (2021). UniProt: the universal protein knowledgebase in 2021. Nucleic Acids Res. *49*, D480–D489.
- Shukla, V., Dhiman, N., Nayak, P., Dahanukar, N., Deshpande, G., and Ratnaparkhi, G.S. (2018). Stonewall and Brickwall: Two Partially Redundant Determinants Required for the Maintenance of Female Germline in *Drosophila*. G3 GenesGenomesGenetics *8*, 2027–2041.
- 78. Cremona, C.A., Sarangi, P., and Zhao, X. (2012). Sumoylation and the DNA Damage Response. Biomolecules *2*, 376–388.
- 79. Hay, B.A. (2000). Understanding IAP function and regulation: a view from *Drosophila*. Cell Death Differ. *7*, 1045–1056.
- 80. Waldvogel, A.-M., and Pfenninger, M. (2021). Temperature dependence of spontaneous mutation rates. Genome Res. *31*, 1582–1589.
- Lüpold, S., Manier, M.K., Ala-Honkola, O., Belote, J.M., and Pitnick, S. (2011). Male *Drosophila* melanogaster adjust ejaculate size based on female mating status, fecundity, and age. Behav. Ecol. 22, 184–191.
- Bretman, A., Fricke, C., and Chapman, T. (2009). Plastic responses of male *Drosophila melanogaster* to the level of sperm competition increase male reproductive fitness. Proc. R. Soc. B Biol. Sci. 276, 1705–1711.
- 83. Parker, G.A., and Pizzari, T. (2010). Sperm competition and ejaculate economics. Biol. Rev., 897– 934.
- 84. Mohorianu, I., Bretman, A., Smith, D.T., Fowler, E.K., Dalmay, T., and Chapman, T. (2017). Genomic responses to the socio-sexual environment in male *Drosophila melanogaster* exposed to conspecific rivals. RNA *23*, 1048–1059.
- 85. Whitlock, M.C., and Agrawal, A.F. (2009). Purging the genome with sexual selection: reducing mutation load through selection on males. Evolution *63*, 569–582.
- 86. Winkler, L., Moiron, M., Morrow, E.H., and Janicke, T. (2021). Stronger net selection on males across animals. eLife *10*, e68316.
- 87. Grieshop, K., Maurizio, P.L., Arnqvist, G., and Berger, D. (2021). Selection in males purges the mutation load on female fitness. Evol. Lett. *5*, 328–343.

- 88. Grieshop, K., Stångberg, J., Martinossi-Allibert, I., Arnqvist, G., and Berger, D. (2016). Strong sexual selection in males against a mutation load that reduces offspring production in seed beetles. J. Evol. Biol. *29*, 1201–1210.
- 89. Wong, A. (2014). Covariance between testes size and substitution rates in primates. Mol. Biol. Evol. *31*, 1432–1436.
- 90. Ramm, S.A., and Schärer, L. (2014). The evolutionary ecology of testicular function: size isn't everything: The evolutionary ecology of testicular function. Biol. Rev. *89*, 874–888.
- 91. Berger, D., Martinossi-Allibert, I., Grieshop, K., Lind, M.I., Maklakov, A.A., and Arnqvist, G. (2016). Intralocus sexual conflict and the tragedy of the commons in seed beetles. Am. Nat. *188*, E98– E112.
- 92. Berger, D., Grieshop, K., Lind, M.I., Goenaga, J., Maklakov, A.A., and Arnqvist, G. (2014). Intralocus sexual conflict and environmental stress: sex, genes, and conflict in stressful environments. Evolution *68*, 2184–2196.
- 93. R Core Team (2021). R: A language and environment for statistical computing (R Foundation for Statistical Computing).
- 94. Hadfield, J.D. (2010). MCMC Methods for Multi-Response Generalized Linear Mixed Models: The MCMCglmm *R* Package. J. Stat. Softw. *33*.
- 95. Daly, M.J. (2012). Death by protein damage in irradiated cells. DNA Repair 11, 12–21.
- 96. Supek, F., and Lehner, B. (2015). Differential DNA mismatch repair underlies mutation rate variation across the human genome. Nature *521*, 81–84.
- 97. Lynch, M., Blanchard, J., Houle, D., Kibota, T., Schultz, S., Vassilieva, L., and Willis, J. (1999). Perspective: spontaneous deleterious mutation. Evolution *53*, 645–663.
- 98. Harrell Jr, F.E. (2021). Hmisc: Harrell Miscellaneous.
- 99. Neuwirth, E. (2014). RColorBrewer: ColorBrewer Palettes.
- Bayram, H., Sayadi, A., Immonen, E., and Arnqvist, G. (2019). Identification of novel ejaculate proteins in a seed beetle and division of labour across male accessory reproductive glands. Insect Biochem. Mol. Biol. *104*, 50–57.
- 101. Andrews, S. (2010). FastQC: A Quality Control Tool for High Throughput Sequence Data.
- 102. Ewels, P., Magnusson, M., Lundin, S., and Käller, M. (2016). MultiQC: summarize analysis results for multiple tools and samples in a single report. Bioinformatics *32*, 3047–3048.
- 103. Sayadi, A., Martinez Barrio, A., Immonen, E., Dainat, J., Berger, D., Tellgren-Roth, C., Nystedt, B., and Arnqvist, G. (2019). The genomic footprint of sexual conflict. Nat. Ecol. Evol. *3*, 1725–1730.

- 104. Kim, D., Pertea, G., Trapnell, C., Pimentel, H., Kelley, R., and Salzberg, S.L. (2013). TopHat2: accurate alignment of transcriptomes in the presence of insertions, deletions and gene fusions. Genome Biol. *14*, R36.
- 105. Anders, S., Pyl, P.T., and Huber, W. (2015). HTSeq--a Python framework to work with high-throughput sequencing data. Bioinformatics *31*, 166–169.
- 106. McCarthy, D.J., Chen, Y., and Smyth, G.K. (2012). Differential expression analysis of multifactor RNA-Seq experiments with respect to biological variation. Nucleic Acids Res. *40*, 4288–4297.
- 107. Ritchie, M.E., Phipson, B., Wu, D., Hu, Y., Law, C.W., Shi, W., and Smyth, G.K. (2015). limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res. *43*, e47–e47.
- 108. Friendly, M., and Fox, J. (2021). candisc: Visualizing Generalized Canonical Discriminant and Canonical Correlation Analysis.
- 109. González, I., and Déjean, S. (2021). CCA: Canonical Correlation Analysis.
- 110. Kolde, R. (2019). pheatmap: Pretty Heatmaps.
- 111. Chen, H. (2021). VennDiagram: Generate High-Resolution Venn and Euler Plots.
- 112. Bolger, A.M., Lohse, M., and Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics *30*, 2114–2120.