Title: A novel method for reliably measuring miniature and spontaneous postsynaptic

potentials/currents in whole-cell patch clamp recordings in the central nervous system

Abbreviated Title: Novel method for measuring mPSPs/mPSCs in the CNS

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Abstract

Measurements of miniature postsynaptic currents (mPSCs) or potentials (mPSPs) in the soma of

neurons of the central nervous system (CNS) provide a way of quantifying the synaptic function at

the network level and, therefore, are routine in the neurophysiology literature. These miniature

responses (or 'minis') are thought to be elicited by the spontaneous release of a single

neurotransmitter vesicle, also called quantum. As such their measurement at the soma can

potentially offer a technically straightforward way of estimating 'quantal sizes' of central synapses.

However, popular methods for detecting minis in whole-cell recordings fall short of being able to

reliably distinguish them from background physiological noise. This issue has received very limited

attention in the literature and its scope as well as the relative performance of existing algorithms

have not been quantified. As a result, solutions for reliably measuring the quantal size in somatic

recordings also do not exist. As the first step in proposing and testing a potential solution, we

developed and described a novel mPSP/mPSC detection algorithm as part of our quantal analysis

software called 'minis'. We tested its performance in detecting real and simulated minis in whole-

cell recordings from pyramidal neurons in rat neocortical slices and compared it to two of the most-

used mini detection algorithms. This benchmarking revealed superior detection by our algorithm.

The release version of the algorithm also offers great flexibility via graphical and programming

interfaces.

Abbreviations

aCSF – artificial cerebrospinal fluid

AMPA – α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid

AP – action potential

API - application programming interface

AUC – area under the curve

CNS – central nervous system

CPP – (*RS*)-3-(2-carboxypiperazin-4-yl)-propyl-1-phosphonic acid

d' – sensitivity index 'd-prime' (defined in Methods)

EPSC – excitatory postsynaptic current

EPSP – excitatory postsynaptic potential

FNR – false negative rate ('miss' rate)

FPR – false positive rate ('false alarm' rate)

GABA – gamma-aminobutyric acid

IPSC – inhibitory post-synaptic current

IPSP – inhibitory post-synaptic potential

mEPSC– miniature excitatory post-synaptic current

mEPSP– miniature excitatory post-synaptic potential

mIPSC– miniature inhibitory post-synaptic current

mIPSP– miniature inhibitory post-synaptic potential

mPSC – miniature post-synaptic current

mPSP – miniature post-synaptic potential

NBQX – 2,3-dioxo-6-nitro-1,2,3,4-tetrahydrobenzo[*f*]quinoxaline-7-sulfonamide

NMDA – N-methyl-D-aspartate

ROC – receiver operator characteristic

sEPSC– spontaneous excitatory post-synaptic current

sEPSP– spontaneous excitatory post-synaptic potential

sIPSC– spontaneous inhibitory post-synaptic current

sIPSP– spontaneous inhibitory post-synaptic potential

smEPSC – simulated mEPSC

smEPSP – simulated mEPSP

smPSC – simulated mPSC

smPSP – simulated mPSP

sPSC – spontaneous post-synaptic current

sPSP – spontaneous post-synaptic potential

TNR – true negative rate

TPR – true positive rate ('hit' rate, sensitivity)

TTX – tetrodotoxin

V_m – membrane potential

v-ROC – virtual ROC (ROC-like plot)

Introduction

Miniature postsynaptic potentials (mPSPs; under voltage recording) or currents (mPSCs; under voltage clamp), often simply called minis, are thought to correspond to the postsynaptic response to a spontaneous release of a single neurotransmitter vesicle (Brown et al., 1979; del Castillo and Katz, 1954; Fatt and Katz, 1952; Isaacson and Walmsley, 1995; Wall and Usowicz, 1998). Somatic measurements of their properties in the CNS provide a window onto the synaptic function at the network level and, therefore, are routine in the literature. Their kinetics (i.e., rise and decay times) are thought to reflect the underlying kinetics of individual ion channels (α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors, AMPARs (Sara et al., 2011), or γ-Aminobutyric acid type A receptors, GABA$_A$Rs (El Khoueiry et al., 2022); i.e., their probabilities of opening and staying open) (De Koninck and Mody, 1994), as well as the locations of the source synapses across the neuronal dendritic tree (proximal vs. distal) and the passive electric properties of neuronal membranes (Agmon-Snir and Segev, 1993; Major et al., 1994; Rinzel and Rall, 1974). Their incidence rates (often called frequencies, although they do not occur rhythmically) are typically thought to reflect the overall number of active synapses on the dendritic tree (Isaac et al., 1995; Liao et al., 1995; Segal, 2010) but can also be indicative of the rate at which presynaptic vesicles are fusing spontaneously with the neuronal membrane (Glasgow et al., 2019). Finally, the size of the mean amplitude of minis is often thought to reflect the overall strength of synapses across the dendritic tree (Segal, 2010). Assuming that minis are postsynaptic responses to the release of a single neurotransmitter vesicle, their amplitude measurements at the soma can also be used as a straightforward method of estimating the quantal size of synapses onto a recorded neuron. These estimates should, however, be viewed with caution as minis amplitudes at the soma tend to be small and difficult to distinguish from noise in the CNS (Larkum et al., 2009; Major et al., 2013; Nevian et al., 2007; Stuart and Spruston, 1998; Williams and Mitchell, 2008; Williams and Stuart, 2002).

'Template matching' and 'thresholded amplitude detection' have emerged as the two main

approaches to detecting minis (Shi et al., 2010). The first approach relies on the observation that

minis often have a fast rise phase and a slower decay phase while noise fluctuation typically do not

show such regularities (Liao et al., 1992; Rall, 1967). The somatic current or voltage recording trace is

scanned with a sliding window and an average minis template is scaled and fitted to the traces to

detect minis-like events of varying amplitudes (Clements and Bekkers, 1997). This approach has

been implemented in a commercially available software suite pClamp (Clampfit application). The

second approach detects peaks in the recording traces and applies an amplitude threshold combined

with shape criteria like the fast rise time and often a limit on the area under the trace (total charge)

(Clements and Bekkers, 1997; Shi et al., 2010). This approach has been implemented in a popular

commercially available software application MiniAnalysis.

'Thresholded amplitude detection' algorithms are thought to suffer from a lack in selectivity

(Clements and Bekkers, 1997). As a result, their performance can either inflate false positive

detection rates and confuse noise fluctuations with the small amplitude minis if a low amplitude

threshold is used or inflate false negative rejection rates and bias the detection towards large

amplitude events if a large amplitude detection threshold is used instead. Similarly, change in the

signal to noise ratio due to changes in the noise level or the amplitude of minis would artifactually

affect estimates of both amplitudes and incidence rates of minis. 'Template matching' algorithms

were introduced as a remedy to the selectivity problem (Clements and Bekkers, 1997), yet in

practice they were found to bias detection towards unambiguous clearcut PSPs that often were also

large, and, therefore, they suffer from the opposite problem of the lack in sensitivity (Hwang and

Copenhagen, 1999). The lower popularity of 'template matching' algorithms compared to their

counterpart suggests this problem to be more severe than the one intended to be remedied.

Given these considerations and our goal of developing a reliable method for estimating quantal sizes

of central synapses based on minis' amplitude somatic measurements (presented in (Dervinis and

Major, 2024)), we developed a novel algorithm, called 'minis', to detect miniature (and action

potential (AP)-triggered) postsynaptic events. A novel algorithm was needed for several reasons.

Firstly, existing algorithms have not been described in sufficient detail in the peer-reviewed

literature. Popular algorithms like MiniAnalysis and Clampfit are proprietary. Secondly, because

existing algorithms are not open, they are not modifiable. Thus, they cannot be automated and

integrated with other software. Lastly, existing algorithms have not yet, to our knowledge, been

benchmarked in peer-reviewed articles. To fill these gaps, we developed an algorithm that is

transparent, flexible, open-source, and objectively evaluated using standardised criteria.

Materials and Methods

Published values for amplitudes and incidence rates of minis.

We searched the neuroscience literature between January 1, 2020 – January 31, 2022 for a

representative sample of recently reported amplitudes and incidence rate of miniature excitatory

postsynaptic currents (mEPSCs) and potentials (mEPSPs) in cortical pyramidal cells, and common

data analysis practices in the CNS. We also compared reported mEPSC rates per second to a

predicted range of values. We limited our search to publications in English only that reported

research results on synaptic function in the CNS. We examined the title and abstract fields of studies

in MEDLINE database using the PubMed search engine with the following search keywords:

("spontaneous neurotransmission" OR "spontaneous neurotransmitter release" OR "spontaneous

transmitter release" OR "spontaneous postsynaptic" OR "miniature postsynaptic" OR "mEPSP" OR

"mEPSC" OR "mPSP" OR "mPSC" OR "mIPSP" OR "mIPSC" OR OR "sEPSP"[Title/Abstract] OR

"sEPSC"[Title/Abstract] OR "sPSP"[Title/Abstract] OR "sPSC"[Title/Abstract] OR

"sIPSP"[Title/Abstract] OR "sIPSC"[Title/Abstract]) AND ("cortex"[Title/Abstract] OR

"cortical"[Title/Abstract] OR "neocortex"[Title/Abstract] OR "neocortical"[Title/Abstract] OR

"hippocampus"[Title/Abstract] OR "hippocampal"[Title/Abstract] OR "thalamus"[Title/Abstract] OR

"thalamic"[Title/Abstract] OR "brain"[Title/Abstract] OR "cerebral"[Title/Abstract] OR "central

nervous system"[Title/Abstract] OR "pyramidal"[Title/Abstract] OR "CNS"[Title/Abstract]).

Abbreviations ending in PSP and PP stand for postsynaptic potentials, while PSC and PC stand for

postsynaptic currents and should not be confused with end-plate potentials and currents.

Animals and electrophysiology

All experimental procedures were carried out in accordance with the UK Animals (Scientific

Procedures) Act 1986 at Cardiff University under Home Office personal and project licenses.

19 to 27-day old Wistar rats (RRID:RGD_13508588; n = 11) of either sex were anaesthetised with

isoflurane and decapitated, and the brain removed under cold (0-2 °C) artificial cerebrospinal fluid

(aCSF), composed of (in mM) 125 NaCl, 26 $NaHCO_3$, 2.3 KCl, 1.26 KH_2PO_4, 1 $MgSO_4$, 10 glucose, and 2

$CaCl_2$ bubbled with 95% O_2 5%CO_2. Coronal somatosensory cortical slices 350 μm thick were cut and

held in an interface chamber with their surfaces kept moist for 1 to 3 hours by aCSF at room

temperature (21-23°C) under a 95% O_2 5% CO_2 atmosphere. Individual slices were then placed in the

recording chamber and held at 35-37°C with aCSF flowing over both surfaces. For recording

purposes at 35-37°C the aCSF solution contained (in mM) 125 NaCl, 24 $NaHCO_3$, 2.3 KCl, 1.26 KH_2PO_4,

1 $MgSO_4$, 10 glucose, and 2 $CaCl_2$.

Whole-cell recordings (n = 14) were obtained under visual control using infrared scanning gradient

contrast imaging with the imaging laser beam and a Dodt tube. Recordings were made with 4.5-9

MΩ borosilicate glass pipettes (Harvard Apparatus, 1.5 mm outside diameter and 0.86 mm, internal

diameter, omega-dot filament; Sutter P2000 puller), filled with (mM): 140 KGluconate, 10 HEPES, 2

$MgCl_2$, 3 ATP-Na_2, 0.3 GTP-Na, 0.1 Magnesium Green, 0.1 Alexa 594; 7.35 pH, 273 mOsm/L

osmolarity. Signals were amplified, low-pass filtered (5 KHz) and digitised (Axopatch-200B, custom

amplifier/filters, Digidata 1440, Clampex software; Molecular Devices). Pipettes and electrodes were

positioned with Sutter MP265 and 285 manipulators with a diagonal (axial) mode, using

parallelogram trajectories.

The electrophysiological recordings consisted of two phases. During the first phase we recorded

mEPSPs against the background of physiological noise ('noise + minis' condition) by applying aCSF

with AP and inhibitory postsynaptic potential blockers added: 0.5 or 1 μM tetrodotoxin (TTX;

selective, potent Na^+ channel blocker), and 12.5 μM gabazine (selective, potent $GABA_A$ receptor

antagonist to block (fast components) of inhibitory synaptic inputs – IPSCs and resulting IPSPs). AP

blockade by TTX was explicitly tested for with long current pulses pushing membrane potential (V_m)

well above normal AP threshold levels, to around -20 mV. The average duration of this phase of the

recordings was 21.8 (± s.d. 0.9) minutes. During a second phase, we recorded noise with mEPSPs

blocked ('noise-alone' condition). This recording was carried out after further adding 40 µM 2,3-

dioxo-6-nitro-7-sulfamoyl-benzo[f]quinoxaline (NBQX) (selective, potent α-amino-3-hydroxy-5-

methyl-4-isoxazolepropionic acid (AMPA) receptor antagonist) and 50 µM CPP (competitive, non-use

dependent selective N-methyl-D-aspartate (NMDA) receptor antagonist) to the solution used in the

'noise + minis' condition, to block all excitatory glutamatergic synaptic inputs (and thus EPSPs),

verified by inspecting and analysing the traces across a number of recordings with the same flow

rates. The average duration of this phase of the recordings was 44.8 (± s.d. 2.9) minutes.

Experimental design

The goal of this benchmarking study was to compare mEPSP detection performance by three

different detection algorithms, mainly in terms of standard signal detection theory measures. Two of

them were the most popular software products used in the field of synaptic function research in the

CNS, while the third one was our novel algorithm called 'minis'.

Current clamp whole-cell patch recordings obtained during the initial recording phase ('noise +

minis' condition) underwent mEPSP detection analysis using 'minis', MiniAnalysis (Bluecell;

RRID:SCR_002184), and Clampfit (part of the pClamp software suite, Molecular Devices;

RRID:SCR_011323). Detected mEPSPs were analysed in terms of their amplitude, 10-90% rise and

decay times, and incidence rate.

The current clamp recordings obtained during the second recording phase ('noise-alone' condition)

served as the background data in the mEPSP ('simulated minis') computer simulations (n=14 cells).

Computationally simulated mEPSPs (smEPSPs) were added onto segments of these V_m noise

recordings (1 of them 200 seconds, 1 more of them 40 seconds, while the remaining 12 were 100

seconds long) and the resultant hybrid waveforms (noise + simulated minis) were then subjected to

mEPSP detection analyses using 'minis', MiniAnalysis, and Clampfit software. Detection performance

was quantified using signal detection theory measures and compared between the three algorithms.

Simulations

Simulating distributions of miniature excitatory postsynaptic potentials with a range of amplitudes

and rise times

All simulations were carried out within the 'minis' software environment which is based on Matlab

(Mathworks; RRID:SCR_001622). Simulation of minis-like events was based on an analytical solution

to the passive cable equation for a transient current (Rall, 1977) using 100 lumped terms with a

double exponential synaptic current with τ_1 = 0.1 ms and τ_2 = 2 ms (Major et al., 1993).

For a given membrane time constant, an initial cable segment was constructed with an electrotonic

length l/λ = 0.6, where l is the physical length of a dendrite (μm) and λ is the length (μm) constant

(Rall, 1977). Charge (Q) was injected at a location (x μm) along this cable and the resulting simulated

postsynaptic V_m was measured at one end of this cable segment. The precise location of the injected

charge was varied systematically, depending on the desired shape of the simulated event. Increasing

the distance of the injected charge along the cable away from the measurement site gradually

increased the rise time of the simulated event. To further increase the rise time, electrotonic length

was gradually increased until the required rise time was obtained. Increasing the amount of injected

charge increased the amplitude of the simulated event. Therefore, one was able to construct a

simulated postsynaptic potential of any rise time or amplitude, for the particular membrane time

constant used. A pool of simulated events could be created having any kind of distribution of

amplitudes and rise times, for a given decay time constant, with individual events' onsets

timed/placed pseudo-randomly along the time axis of the V_m recording.

Simulations used to construct full receiver operating characteristic (ROC)-like curves, using relatively

fixed but realistic ('moderately-sized') mini amplitudes

All simulated mEPSPs were drawn from a two-dimensional normal distribution (independent

amplitude and rise time dimensions) with the following amplitude parameters: mean μ_1 = 0.3 mV

and standard deviation σ_1 = 0.05 mV (i.e., amplitudes relatively constant, with a relatively small

amount of variation). The following (independent) 10-90% rise time parameters were used: μ_2 = 0.05

ms and σ_2 = 2.5 ms (with a minimum rise time of 0.05 ms; i.e., only upper half of normal distribution

used). Simulated events were drawn pseudo-randomly from a distribution at one of the following

incidence rates (minis/second): 640, 320, 160, 80, 40, 20, 10, 5, 2.5, 2.5, 2.5, 2.5, and 2.5.

Corresponding noise amplitude scale factors for these 14 incidence rates conditions were as follows:

1, 1, 1, 1, 1, 1, 1, 1, 1, 1.2, 1.4, 1.8, 2.6, and 4.2. The simulated events were then positioned at

pseudo-randomly determined times along the V_m noise recording epochs obtained during the

second recording phase ('noise-alone' condition). Two of the noise-alone epochs were 200 and 40

seconds long, while the remaining twelve were 100 seconds long. Four different simulation traces

per recording, each of them having different pseudo-random event timings, were generated for

every incidence rate/noise scale condition, resulting in a total number of 14×4×14 = 784 traces.

Simulations of miniature excitatory postsynaptic potentials using a range of biologically realistic

amplitudes at realistic incidence rates

The simulation parameters used to construct full ROC-like curves cover only a part of the likely

biological range. In reality, in cortex, minis vary far more in their amplitudes, with large numbers of

smaller minis, but a minority of bigger minis too - some much bigger. Therefore, we set out to

compare all three algorithms using a range of plausible simulated mini amplitudes, within the

restricted range of what we deemed to be realistic incidence rates based on our estimation (from

recent peer-reviewed publications) in the first Results subsection.

All simulated events were drawn from a two-dimensional distribution with the following

parameters: amplitudes were chosen from ten discrete values (0.05, 0.1, 0.15, 0.2, 0.25, 0.3, 0.35,

0.4, 0.45, 0.5 mV) with equal probabilities, rise times were independently chosen from the upper

half of a normal distribution with mean $\mu_2 = 0.05$ ms, and standard deviation $\sigma_2 = 2.5$ ms (i.e., no rise

time shorter than 0.05 ms). Three 'realistic' minis' incidence rates were used: either 61, 38, or 27

minis/s. The simulated events were positioned at pseudo-random times along the same noise

recording epochs as used in the previous subsection. Four different simulation traces per recording,

each of them with different random event timings, were generated for every incidence rate

condition, resulting in a total number of 3×4×14 = 168 traces.

Detection of spontaneous and miniature postsynaptic potentials

'minis' detection algorithm

We developed a new algorithm to detect postsynaptic potentials and currents, that we have

incorporated into a data analysis software package called 'minis', coded in Matlab (Mathworks). It is

distributed as an application programming interface (API) in the form of a packaged Matlab app or a

Python (Python Software Foundation; RRID:SCR_008394) package or as a compiled standalone

desktop application with a graphical user interface. In brief, the detection algorithm takes a filtered

V_m or clamp current trace in the Axon Binary File (ABF) format and detects peaks and rising phases

and estimates their amplitudes and rise and decay times after applying certain processing steps.

These steps are outlined below (with parameter values tailored for detecting smEPSPs in cortical

pyramidal neurons). A schematic illustration of the key steps is shown in Figure 1:

A. Steps to find minis:

1. The data can be band-stop filtered to remove mains noise (e.g., small integer multiples of

299   50 Hz frequency components: optional). For this, a Butterworth filter (Matlab's 'butter'

300   function) is used with a stopband attenuation of 10 dB and a passband 'ripple' of 0.05 dB.

301   The stopband size is 1 Hz, and the passband is the entire remaining frequency range except

for the ±3 Hz window either side of the stopband frequency. Filtering effects are illustrated

in Figure 2.

2. The recording trace can be smoothed using a Gaussian window (optional). We use a 1.5-

millisecond window (with standard deviation of the Gaussian of 0.53 ms). This step removes

high frequency noise. Smoothing effects are illustrated in Figure 2.

3. Peaks (local maxima) are identified in the filtered, smoothed recording trace. Each peak is

then examined consecutively, starting with the initial peak.

4. Going through all the peaks one by one, any peak within the "Peak Integration Period"

('duration') following the current ('working') peak of interest is discarded if it is *smaller*

(lower) than the peak of interest. For the analysis in this paper, we use a Peak Integration

Period of 2.5 ms; this can be matched to the different 'sharpnesses' or 'breadths' of peaks

in different data.

5. If there is a larger peak within the peak integration period, the 'working' peak of interest is

moved to this new peak, and the old peak and all intervening peaks between it and the new

'working' peak of interest are discarded.

6. The baseline of the peak is positioned asymmetrically around the lowest value (local

minimum) before the peak. Because of the faster upstroke of minis, compared with their

decays, 80% of the baseline period falls before the lowest voltage (local minimum) and the

remaining 20% after it (see Figure 1, inset). The interval from the baseline to the peak is

limited by the *Maximum Time-to-Peak* parameter (10 ms used here). The last 20% of the

baseline cannot start earlier than this.

7. The length in time of the baseline is controlled by the *baseline duration* parameter. We

used 2-ms. (N.B. a baseline, or indeed an entire mini, can occur during the summated decay

'tails' of previous minis, particularly when the minis incidence rate is high relative to their

decay time, as often seen with mPSPs under voltage recording at body temperature – but

also under voltage clamp).

8. If the baseline overlaps with or falls before the previous peak, its start is delayed till

immediately after the previous peak (to stop the baseline being pulled down by the

previous rising phase), but the end time is kept the same.

9. To reject any large artefactual 'glitches', or small events that are putative noise, if the

amplitude of the peak is outside of the range of acceptable amplitudes, between a

detection threshold and a maximum, the peak is discarded. We allowed a range of

amplitudes of 0.1-10 mV in detection conditions used to construct full 'virtual' receiver

operating characteristic (ROC) curves. In other conditions we applied a range of 0.05-10 mV.

To detect real minis, we used a range of 0.01-10 mV. (N.B. 0.02 or 0.03 mV detection

thresholds can also be used for real whole-cell patch recording data from neocortical

pyramidal neurons, under our experimental conditions, guided by the range of amplitudes

of dendritically detected, somatically recorded mEPSPs in (Nevian et al., 2007) from layer 5

pyramidal neurons from rat somatosensory cortex; in general, detection thresholds will

need to be adjusted to the cell type and experimental conditions).

B. Steps against multiple small minis being erroneously lumped together (merged):

10. Rising phase asymmetries: if the first half of the rise time (10-50% or 20-50%, depending on

the rise time duration of choice) is longer than the second half (50-90% or 50-80%) by a

factor of 5 or more (an asymmetry incompatible with typical single, non-summating EPSP or

EPSC rising phases), the end of the baseline is extended slightly into the 'take-off' of the

rising phase. (This correction can be repeated one more time).

11. The amplitude range test is applied again (Step 9).

12. If the second half of the rise time is longer than the first half by a factor of 5 or more, the

peak is shifted to a previously discarded smaller peak, if exists. (Two minis could be

summating on their rising phases, which is treated as a single mini, or the second mini could

occur just after the peak of the first, which is treated as two minis, if two peaks are

distinguishable).

13. The amplitude range test is applied again (Step 9).

14. If the 10% or 20% rise time mark (depending on whether 10-90% or 20-80% rise time is

measured) is later than the end of the baseline by more than a half of the baseline duration

(1 ms in our case), a new baseline is established closer to the peak (by moving the baseline

to the next local minimum).

15. Finally, the amplitude range test is applied again (Step 9).

16. A mini (mPSP/mPSC) is established.

Detection using MiniAnalysis software

MiniAnalysis software version 6.0.7 was used to detect mEPSPs for a benchmarking purpose. We

used the following detection parameters: As with the 'minis' program, an amplitude threshold of 0.1

mV (when constructing the full ROC-like curve) or 0.05 mV (all other simulated conditions), in both

cases somewhat matched to the simulated data, or 0.01 mV when detecting real minis, a "Period to

search for a Local maximum" of 12.5 ms, "Time before a peak for baseline" of 10 ms, a "Period to

search a decay time" of 30 ms, a "Fraction of peak to find a decay time" of 0.37 (one effective time

constant for exponential decay), a "Period to average a baseline" of 2 ms, an "Area threshold" of 0

mV×ms (i.e., not used), a "Number of points to average for peak" of 1, and using the "Detect

complex peak" option (to avoid missing close-together events). The chosen detection parameters

closely resembled similarly named parameters in the 'minis' detection algorithm with minor

deviations to improve performance. Arrived by trial and error, these parameters were also found to

give the most optimal detection performance of the MiniAnalysis software in comparison to other

tested parameter sets. Event detection was carried out on the recording data that was filtered and

smoothed in the same way as described in steps 1 and 2 of the previous sub-section (Figure 2).

Detection using Clampfit

Clampfit software, part of the pClamp 11.0.3.03 software suite, was used to detect real and

simulated mEPSPs and mEPSCs for a benchmarking purpose. Recordings were band-stop filtered as

described in step 1 of the 'minis' detection algorithm. Data was not smoothed. A template search

algorithm was used with 9 templates. The mEPSP template set was constructed based on the

simulated events. Events having 0.5 ms, 1.5 ms, 3.5 ms, and 6.5 ms 10-90% rise times were selected

to produce 4 distinct mEPSP templates (with the correct decay time constants). An additional 5

single-peaked templates were constructed in a way that made them potentially a mix of multiple

simulated waveforms. They were constructed to have extra-fast, fast, intermediate, slow, and extra-

slow rise times corresponding to 2 ms, 2.75 ms, 6 ms, 7 ms, and 8 ms 0-100% rise times and 8 ms,

9.25 ms, 17 ms, 23 ms, and 37 ms duration full decays, respectively. Combining single and mixed

waveforms gave better detection performance than having waveforms based only on any one of

these types alone. Template match threshold was set to the default of 4 noise standard deviations

with the rest of the detection parameters being set to their default values as well. The same set of

templates was used to analyse all simulated current clamp data, as well as real data. Using the same

templates with real data was justified, because they resulted in a superior detection performance

compared to templates created directly from the real data.

Detection algorithm response classification

At the outset we identified prominent noise events in the V_m noise recordings (the 2nd recording

phase: 'noise-alone' condition) that could potentially be confused for true minis (mEPSPs) by any of

the algorithms (noise traces in Figures 2A and B). All noise recordings were filtered and smoothed as

in Steps 1 and 2 of the 'minis' detection algorithm (Figure 2). The data was further smoothed by a

rectangular 'box-car' moving average window of 20 ms. Then all peaks larger than 0.01 mV and

having a half-width of at least 0.5 ms were identified. These peaks were classed as (notional) noise

'events' (for the purpose of generating 'sensible' true negative rates, TNRs, which do not 'swamp'

false positive rates across the entire virtual ROC; see below). Subsequently we simulated mEPSPs

and added them at pseudo-random times to a zero-trace having the same length as the noise

recording of interest (Simulation traces in Figures 2A and B). The peaks of these events were classed

as signal events. The simulated trace was then added to the noise recording ("Simulation + noise"

traces in Figures 2A and B). Locations of signal and designated noise events formed the 'ground

truth' information.

Following the detection process, all detected events were associated with one of the signal and

noise events depending on their proximity to these predefined 'ground truth' events. A detected

event was classed as a true positive (a 'hit') if it was the closest detection event within 5ms of a

signal (simulated mEPSP/mini) event. If no detection event occurred within the 10 ms symmetrical

window surrounding a signal (simulated mini) event (-5 to 5 ms), the signal event was classed as a

false negative (a 'miss'). If no detection event (except correct detections) occurred within the same

duration window surrounding a noise event, the noise event was classed as a true negative ('correct

rejection'). Although in theory there could potentially be a much higher number of possible time

points which could be labelled as correct rejections, or true negatives, in this situation, because of

their inherently somewhat arbitrary nature, we wanted to keep the maximum number comparable

to the maximum number of true positives (simulated minis), so the ROC plot had a reasonable

chance of being relatively 'square' (i.e. having a useful dynamic range). The remaining detection

events were classed as false positives ('false alarms'). Examples of classified detection events are

shown in Figure 2 separately for all three detection algorithms.

Detection response time to the nearest neighbour

The time to the peak of the nearest smEPSP was available for every classified response. Detection

performance was then evaluated as a function of the time to the nearest neighbour.

Membrane potential 'phase' (trend) classification

We attempted to address common detection errors, in particular missed minis on the decaying

phase of a previous mini (causing second peak to drop relative to its baseline), or on the rising phase

of another mini (so the rises of both minis get lumped together). Algorithm detection performance

was assessed not only for full recordings but also separately for periods when the 'underlying'

simulated V_m (before adding the noise) was rising, decaying, or remained roughly stable (Figure 3).

The 'decaying phases' (downward trend) of the purely simulated V_m were set to be a) the period

from 0.0625 ms after the peak until the purely simulated V_m trace dropped to the ratio of 0.3/e mV

(0.11 mV) (amplitudes of simulated minis were ~0.3 mV in the full virtual-ROC curve conditions, so

this would be about one membrane time constant after an 'average' simulated mini) or b) until the

V_m started rising again to reach a new peak that was above the initial peak (at the start of this decay

phase). If the new peak was lower than the initial one, the entire new peak was classified as part of

the ongoing decay phase. Peaks occurring on the decay phase of the V_m were assigned the V_m decay

rate of change that was present 0.25 ms (5 sample points) prior to the appearance of the V_m

inflection points associated with the start of the rise of a new mini (indicated by the blue arrow in

Figure 3A). The rising 'phase' was treated essentially somewhat as a decay phase in reverse. The only

difference was that the ratio of 0.3/e mV was replaced by the ratio of 0.3/10 mV (0.03 mV). Peaks

occurring on a V_m rise phase (upward trend) were assigned a V_m rate of rise value that was present

0.125 ms *after* the appearance of an upward V_m inflection point following the initial decay of the

previous peak (indicated by green arrows in Figure 3A). Periods that were already classified as decay

phases could not be reclassified as rising phases. Periods of relatively slowly-changing simulated V_m,

outside of the rise and decay phases were classed as "stable" (or a flattish phase). The classification

of purely simulated membrane potentials into three different trend phases was used to assign

phases to the combined noise recordings + simulated minis (Figure 3B). The detection performance

during rise and decay trend phases was evaluated as a function of the V_m rate of change.

Data Analyses

Performance of the three algorithms was compared using measures from signal detection theory. All

detection responses were classified as described in the *Detection algorithm response classification*

subsection. True positive ('hit') rate (TPR) was calculated as follows:

$$TPR = \frac{True\ positive\ count}{True\ positive\ count + False\ negative\ count}.$$

Throughout this text we use TPR, detection probability, and sensitivity interchangeably. False

positive ('false alarm') rate (FPR) was calculated as follows:

$$FPR = \frac{False\ positive\ count}{False\ positive\ count + True\ negative\ count}.$$

The true negative count we have used is perforce notional, as explained above, but still useful for

standardised, non-subjective comparisons and benchmarking. Throughout we use FPR and 'false

alarm rate' interchangeably.

Calculation of TPR and (notional) FPR allowed us to produce ROC-like curves and to calculate area

under the curve (AUC; area under the ROC curve, for each complete ROC curve, with 0.5 indicating

chance performance (the area of the triangle under the diagonal from (0,0) to (1,1) and 1 indicating

perfect performance, i.e., the area of a right angle starting at (0,0), vertical to (0,1) then going

horizontal to (1,1)) as detection performance indicators. These are the most commonly used

performance indicators for classifiers, in addition to sensitivity (TPR), and d' (discriminability or

sensitivity index) which we often use in parallel throughout this text. The d' statistic was calculated

as follows:

$$d' = z(TPR) - z(FPR),$$

where z is the z-score defined as the inverse of the normal cumulative distribution function, i.e.

z(arg) is the number of sd's from the mean giving the area equal to its argument arg under the

standard unit normal distribution (mean 0, sd 1), starting from $-\infty$. TPR and FPR values are essentially

treated as cumulative probabilities as explained in (Macmillan and Creelman, 2005). So, d' tells us

how many standard deviations 'apart' the TPR and FPR are, assuming they are cumulative

probabilities (areas under curve) of a standard normal distribution. N.B. If FPR is 0, z(FPR) is -∞ so d'

is undefined, but arbitrarily large and positive (as occurs in some of our figures below). These plots

are truncated for the detection method in question, but the performance is still superior to any plots

with finite values.

The above detection performance indicators were calculated for full recordings, as well as separately

for rising and decaying periods of V_m.

In addition, we also looked at cumulative (left-to-right over the x-axis) TPR and cumulative FPR. The

cumulative TPR was defined as follows, for each detection algorithm and simulated waveform:

$$Cumulative\ TPR = \frac{Cumulative\ true\ positive\ count}{Total\ unbanded\ true\ positive\ count} * Overall\ unbanded\ TPR.$$

Likewise, the cumulative FPR was defined as:

$$Cumulative\ FPR = \frac{Cumulative\ false\ positive\ count}{Total\ unbanded\ false\ positive\ count} * Overall\ unbanded\ FPR.$$

The latter two measures were not only useful for measuring cumulative hits and false alarms over a

range of time intervals and V_m rates of change, but also provided overall estimates of TPR and FPR

(the right side of the x-axis).

Statistical analyses

All inferential statistics were based on the assumption that distributions of all measures were

normal. Only repeated samples or single-sample (often when comparing with baseline true positive

values in tables) t-tests were used across the Results section. All mean distribution values are

stated/depicted with 95% confidence limits of the mean (if exist).

Data accessibility

All data analysed in this study is publicly available (GIN DOI). The available data include 'noise +

minis' and 'noise-alone' whole-cell patch membrane potential recordings that were used in minis

detection procedures and computer simulations. The hybrid V_m traces containing smEPSPs added to

noise recordings are also available. All electrophysiological recordings were stored in Axon Binary

File (ABF) format.

mEPSP templates used for detecting real and simulated events via Clampfit are also in the same

dataset in the form of Axon Template Files (ATFs).

Code accessibility

All analyses were carried out in Matlab and the analysis code is publicly available on GitHub

(available once published). The code is complete with instructions on how to reproduce all figures

reported in this study.

Software accessibility

The present study reported the use of a novel mEPSP/mEPSC detection algorithm that is part of

'minis' software available on GitHub (Dervinis, 2022).

Results

Research literature measuring minis in the central nervous system

The full results of the literature survey are reported in Supplementary Results. Here we only briefly

report key findings. First, we established that the two most popular software applications used to

detect minis are MiniAnalysis followed by Clampfit with 48% and 26% of surveyed studies reporting

to use these algorithms, respectively. Moreover, these studies reported a mean minis' incidence rate

of 3.6 ± s.d. 0.34 per second (range of 1.0 to 8.0 per second). Meanwhile, we suspect that true minis

incidence rates range somewhere between ca. 10 minis/s and 100 minis/s based on research on

synapse counts and release probabilities in the CNS (an order of magnitude higher than reported

direct measurements using voltage clamp).

Algorithm performance comparison for detecting moderately-sized (~0.3 mV) simulated miniature

excitatory postsynaptic potentials

Detection under a wide range of incidence rate conditions: overall performance

In the Results subsection 'Research literature measuring minis in the central nervous system', we

reported that detection of spontaneous postsynaptic events is typically carried out using an

amplitude threshold. This approach implies that the range of amplitudes of spontaneous

postsynaptic events is somewhat known and that, on the smaller end, the amplitudes are

moderately-sized and sufficiently larger than pure noise fluctuations. Leaving aside the question of

the empirical validity of this (somewhat dubious?) assumption, we evaluated automated detection

of moderately-sized (0.3 ± s.d. 0.05 mV) smEPSPs added to whole-cell patch clamp noise recordings

(the 2nd recording phase: 'noise-alone' condition).

We compared our novel 'minis' algorithm with MiniAnalysis and Clampfit. We created 14 simulation

incidence rate conditions with different effective signal-to-noise ratios that loosely mimicked the

effects of varying the signal detection threshold. On the one hand, increasing the incidence rate of

smEPSPs tended to decrease TPR as simulated events started to merge. On the other hand,

increasing the noise amplitude (scaling up the noise that was added to the simulated minis) tended

primarily to increase the FPR.

We plotted the detection performance in terms of TPRs against FPRs, as 'virtual' (notional) Receiver

Operator Characteristic (v-ROC) curves, for all 14 conditions and all three algorithms in Figure 4A.

We found that in the first six conditions (high minis incidence rates), the v-ROC curve for 'minis' had

minimal overlap with any other curve and was positioned furthest away from the diagonal (starting

from left to right; 640, 320, 160, 80, 40, and 20 minis/s with x1 noise scaling), indicative of a superior

performance in this (relatively high) incidence rate range (the highest incidence rates were used to

create the full v-ROC; higher incidence rates increase overlaps, i.e. temporal summation, between

minis, which are particularly challenging to detect). The same observation was supported by another

measure, the sensitivity index d', shown in Figure 4B. Detection performance in the remaining 8

conditions (10, 5, 2.5, 2.5, 2.5, 2.5, 2.5, and 2.5 minis/s with noise scaling factors of x 1, 1, 1, 1.2, 1.4,

1.8, 2.6, and 4.2, respectively) was very similar for both 'minis' and MiniAnalysis: neither of the two

showed a clear superiority on both performance measures simultaneously: v-ROC and d' curves

overlapped. In contrast, Clampfit consistently performed worst across all 14 conditions, particularly

with high amplitude noise. Clampfit becomes progressively 'confused' as noise increases, placing

templates on noise events, but ignoring 'swamped' smEPSPs.

When overall performance was assessed averaging across individual incidence rate and signal-to-

noise ratio conditions, we found that 'minis' showed the best performance with the mean Area

Under the (v-ROC) Curve (AUC) value of 0.960 ± 0.002 (95% confidence interval here onwards; Figure

4C). A paired samples t-test comparing to MiniAnalysis which had a mean AUC value of 0.937 ±

0.0022 gave t(13) = 12.4 and p = 1.37×10^{-8}. Clampfit showed the poorest overall performance with

the mean AUC value of 0.706 ± 0.0043, t(13) = 33.1, and p = $6.14×10^{-14}$ when compared to the

MiniAnalysis mean AUC value in a paired samples t-test. These differences in detection performance

were further corroborated by the d' measure averaged across all signal-to-noise ratio conditions

(Figure 4D). A paired samples t-test comparing mean d' values from 'minis' (2.45 ± 0.045) versus

MiniAnalysis (2.11 ± 0.04) gave t(13) = 14.0 and p = $3.24×10^{-9}$. The same test comparing the mean d'

from MiniAnalysis and Clampfit (1.23 ± 0.024) gave t(13) = 13.3 and p = $6.07×10^{-9}$. In summary, these

findings clearly demonstrated that our 'minis' detection algorithm outperformed the other two

algorithms, for ~0.3 mV simulated minis added to real recording noise, at incidence rates above 10

minis/s, while Clampfit consistently showed the poorest detection performance across the entire

incidence rate and signal-to-noise range examined, for these mPSP shapes and templates.

Detection under a wide range of incidence rate conditions: common errors

Both 'minis' and MiniAnalysis were fairly good at detecting moderately-sized (0.3 ± s.d. 0.05 mV)

smEPSPs added to our noise-alone recordings. Yet all three algorithms, including 'minis', missed a

sizeable proportion. Figure 5 shows examples of common mistakes. One type of mistake was missing

simulated events that occurred very close to other events (Figures 5A, B, C, D, E, F, and G).

Unsurprisingly, none of the three algorithms could deal with events that overlapped nearly perfectly,

always lumping them together. Detecting peaks in the data using thresholds or fitting templates to

the data cannot offer a way of identifying overlaps. Another common error type was missing events

with peaks on the rising phase of another mEPSP (Figures 5A, B, C, D, E, and F; green arrowheads).

Unlike perfect or near-perfect overlaps, these events had a clear but brief decay period that was

quickly upended by the rising phase of the next mini. Because these decays were brief, they were

often lumped with a subsequent mEPSP or they could not be adequately fit with a template by

Clampfit and, therefore, were discarded. The 'minis' program (but not MiniAnalysis) does offer a way

to minimise these errors by reducing the peak integration period, but this comes at the expense of

noise fluctuations on the rising phase increasingly passing as postsynaptic potentials (a type of false

positive; Figure 5H, red arrowheads). An effective solution to this problem would, therefore, require

isolating and measuring the noise component in whole-cell patch clamp recordings - which currently

is not implemented in either MiniAnalysis or Clampfit (but is in 'minis').

An even more common error type was to miss (smaller) mEPSPs that occurred on the decay phase of

another, often larger, earlier mEPSP (Figures 5D and G; blue arrowheads). The amplitude of mEPSPs

occurring on decays is effectively reduced (as the peak drops relative to the baseline, due to the

underlying decay of V_m) and the steeper the decay, the more the amplitude is reduced. The

amplitude of such a reduced mini may fall below the detection threshold and/or shape distortions

may prevent fitting a template. As a result, such mEPSPs were occasionally discarded (this turns out

to be a significant problem with real minis recorded from cortical pyramidal neurons, which exhibit

significant temporal summation, because of their relatively high incidence rates and slow decays).

There is no easy way of correcting these errors, as it would (for example) require estimating the

underlying decaying potential and subtracting it from the overall V_m or estimating the fall in the peak

and adding it back to the amplitude before deciding whether to accept that candidate putative mini.

mEPSPs that neither occurred during the rise nor decay phases were also occasionally missed

(Figures 5A, D, E, and F; black arrowheads). This could have happened due to a failure to fit a

template (Clampfit) or due to the amplitude being reduced below threshold by *downward*

background noise fluctuations. In recordings of real mEPSPs, these errors would mainly affect small

amplitude mEPSPs. Again, an effective solution to this problem would require isolating and

measuring the noise component in whole-cell patch clamp recordings similarly to previously

discussed errors occurring on the V_m rise phase.

A whole other class of detection errors appeared in the form of false alarms (false positives)

occurring on the rising (Figure 5E; red arrowhead) or decaying phases (Figures 5A, B, and H; red

arrowheads) of underlying mEPSPs or in the absence of any other minis in the vicinity (Figures 5B

and H; red arrowheads). These mistakes were often caused by *upward* background noise

fluctuations passing as real minis and might have been 'enabled' by low amplitude detection

thresholds or an inability to discern shapes of real minis from mini-like noise. Just like the previously

discussed error type, a solution would require isolating and measuring properties of the noise

component. In addition, there were errors that uniquely affected only one of the algorithms, like the

misplacement of detected events (peaks) by MiniAnalysis (Figure 5C; green arrowheads).

We saw that distinct detection errors could be made depending on how close they were to

neighbouring mEPSPs and whether they occurred on the rise or decay phases of other nearby

mEPSPs or in the absence of any other mEPSPs in the background. All three algorithms occasionally

committed these errors, but their incidence was uneven.

Detection under a wide range of incidence rate conditions: the membrane potential 'slope trend'

effect

In order to quantify the detection performance in different circumstances, for the same ~0.3 mV

smEPSPs, we evaluated the probability of making errors in relation to the *proximity* to neighbouring

minis and the phase (*trend*) of the background V_m: rising or decaying. We looked at TPR as a function

of time interval to the nearest-neighbour mini (Figure 6A) taken together for all minis incidence rate

conditions used above (but excluding conditions where noise was scaled): i.e., 640, 320, 160, 80, 40,

20, 10, 5, and 2.5 minis/s conditions (9 in total). We found that detection was superior for our 'minis'

program across the entire inter-mini interval range. MiniAnalysis came second, with Clampfit

consistently worst. These differences in detection performance were also reflected in the *cumulative*

TPR (Figure 6D). As for the FPR (Figure 6B), 'minis' was superior at detecting ~0.3 mV smEPSPs that

were within 10 ms of the nearest neighbour (peak-to-peak). This performance edge over

MiniAnalysis was not maintained for larger inter-mini peak intervals, while Clampfit again showed

consistently the worst performance across the entire interval range. These performance differences

were also reflected in the *cumulative* FPR (Figure 6E). Taken together, true and false positive rates

allowed as to calculate a combined performance indicator, sensitivity index d', as a function of time

to the nearest neighbour (Figure 6C). The combined measure indicated that 'minis' had the superior

detection performance within 10 ms of the nearest neighbour while Clampfit showed the worst

performance across the entire inter-mini interval range.

During the V_m rise phase performance differences were unequivocal. There were very few events

and, therefore, positive detections when the background V_m was rising at 200 µV/s or faster (Figure

7D). Most of the detected events coinciding with the rise phase actually occurred when V_m was

changing at the rate of 10 to 100 µV/s. In this range 'minis' showed the best detection performance

with d' between 1 and 2 (Figure 7C). MiniAnalysis was the second best, while Clampfit showed the

worst performance. This order of performance was maintained for rates below 10 µV/s with overall

performance improving for all algorithms. The performance edge of 'minis' over the other

algorithms was primarily due to low FPR (Figure 7B). The TPR of 'minis' was also superior to the

other two algorithms but, in terms of sensitivity, MiniAnalysis was not far behind (Figure 7A). In the

range between 15 and 35 µV/s 'minis' actually fell slightly behind MiniAnalysis. The cumulative rates

clearly supported the observation that 'minis' was superior to the other two algorithms with

MiniAnalysis coming second and Clampfit having the worst performance (Figures 7D and E).

The same behaviour of the three algorithms was also observed for detecting ~0.3 mV smEPSPs

occurring on the decay phase of the background V_m (caused by summation of other smEPSPs). Most

of the detected simulated events were associated with rates of V_m change in the range of -100 and -

10 µV/s (Figure 8D). In this range and above, 'minis' demonstrated the best performance relative to

the other two algorithms with d' ranging between 1.5 and 2.5 (Figure 8C). MiniAnalysis again came

second, while Clampfit showed very poor performance. Clampfit often had a d' that was close to 0,

meaning that its performance was often not different from chance level when background V_m was

decaying. This was mainly due to the large FPR for this algorithm (Figure 8B); 'minis' had

unequivocally the highest TPR and the lowest FPR (Figures 8A and B) with cumulative rates very

clearly reflecting this (Figures 8D and E). The overall performance of the runner-up, MiniAnalysis,

was substantially poorer.

There are only 3 conditions with realistic incidence rates (20, 40, and 80 minis/s). Although, higher or

lower incidence rates were useful in constructing the full v-ROC curve, it can be argued that they are

artificial and experimental electrophysiologists may not be concerned with them. Therefore, we also

examined the membrane potential 'slope trend' effect separately for the three realistic incidence

rate conditions in the Supplementary Results section demonstrating similar findings to the overall

results.

Algorithm performance comparison for detecting simulated excitatory postsynaptic potentials

with a wide set of discrete amplitudes, at realistic incidence rates

So far, we have only tested performance of the three algorithms when detecting moderately-sized

(0.3 mV ± s.d. 0.05 mV) mEPSPs with a relatively narrow range of amplitudes. The existing evidence

does not indicate that either mPSPs or mPSCs form a distinct amplitude distribution that can be

separated from the amplitude distribution of noise fluctuations. Therefore, here we aimed to test

the performance of detecting smEPSPs of a wide range of amplitudes, to evaluate the three

algorithms in more realistic settings.

We used three incidence rate conditions: 27, 38, and 61 minis/s. Within each of these conditions

smEPSPs were drawn pseudo-randomly from a uniform distribution of the following ten discrete

amplitudes: 0.05, 0.1, 0.15, 0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5 mV (4 different simulations per noise

trace per incidence rate conditions; see Methods for shapes). Just like in previous simulations, we

positioned these smEPSPs pseudo-randomly over noise recordings and detected them using the

three algorithms. The overall performance of these algorithms is shown in terms of true and false

positive rates in Figure 12A and d' in Figure 12B. A few things are prominent in these graphs. First of

all, the detection performance of all algorithms was considerably worse than when detecting

moderately-sized (~0.3 mV) mEPSPs. Secondly, 'minis' again had the edge over two other algorithms,

especially at higher minis' incidence rates. Third, Clampfit's performance was consistently the

poorest, in our hands.

Performance was also assessed pooling the incidence rate conditions. We found 'minis' to have the

best performance with the mean AUC value of 0.827 ± 0.003 (95% confidence interval from here on;

Figure 12C). A paired samples t-test comparing to MiniAnalysis with the mean AUC value of 0.809 ±

0.003 gave $p = 1.0\times10^{-6}$. Clampfit showed the poorest performance, with the mean AUC value of

0.665 ± 0.004, $p = 2.2\times10^{-10}$ when compared to the MiniAnalysis mean AUC value in a paired samples

t-test. These differences in detection performance were further corroborated by the d' measure

averaged across all incidence rate conditions (Figure 12D). The d' discriminability index scores of

'minis' (1.5 ± 0.05) and MiniAnalysis (1.33 ± 0.04) were significantly different ($p < 10^{-5}$), as were the

d' from MiniAnalysis and Clampfit (0.61 ± 0.02) ($p < 4\times10^{-6}$) The conclusion based on combined

measures was in line with the assessment based on Individual incidence rate conditions.

Other measures like true positive and false positive rates as functions of the interval to the nearest

neighbour and V_m rate of change during the rise or decay phases using more realistic settings

pointed to a very similar conclusion to that made with the full virtual ROC curve. First, we found that

'minis' performed better than the other algorithms at detecting mEPSPs that were within 10 ms of

other mEPSPs, and that at larger intervals, MiniAnalysis performed just as well (Figure 13).

Meanwhile, Clampfit performed consistently the worst across the range of inter-mini intervals

whether in terms of d' (Figure 13C) or true and false positive rates (Figures 13A, B, D, and E). When it

came to V_m rate of change, 'minis' performed better than MiniAnalysis, which in turn performed

better than Clampfit when detecting smEPSPs occurring during rise and decay phases of background

simulated V_m (Figures 14 and 15). One difference compared to the full virtual ROC curve analysis was

that the detection performance of 'minis' and MiniAnalysis overlapped in the range of 15-25 µV/s

when detecting smEPSPs occurring on the rise phase of the background V_m (Figure 14C). This is

largely caused by an increased FPR of 'minis' in this rate range (Figure 14B). The overall performance

of all algorithms was better (Figures 14C and 15C) compared to the full virtual ROC curve analysis

(Figures 7C and 8C), but still worse when compared to detecting moderately-sized (~0.3 mV)

smEPSPs with realistic incidence rates (Figures 10C and 11C), as described in the previous

subsection. In summary, 'minis' had a substantial advantage when detecting minis in close proximity

to neighbouring minis, as well as during rising or falling background V_m, with Clampfit severely

underperforming.

Comparison of amplitudes, time course, and incidence rates of detected real and simulated

excitatory postsynaptic potentials by different algorithms

In the final section we looked at how the three tested algorithms fare across all simulation

conditions in terms of their minis' amplitude, time course, and incidence rate estimates. We also

carried out detection of real minis in the same recorded cells prior to blocking minis ('noise + minis'

condition) and compared the same estimates for the three algorithms. A large number of estimates

and inferential statistics was compiled into several tables providing a thorough performance

overview.

Table 1 below shows the compiled mean amplitude estimates for simulated and real minis detected

by the three algorithms while Supplementary Table 1 shows repeated samples t-test p-values for

comparisons of mean amplitude values for the three different algorithms with respect to ground

truth values. Mean amplitude values of simulated and real mPSPs detected by 'minis' were

consistently lower than mean amplitude values of events detected by both MiniAnalysis and

Clampfit and were closest to ground truth values. Detection of simulated events by the 'minis'

algorithm in conditions with realistic smPSP incidence rates (i.e., Type 1 RI and Type 2 rows in the

tables) were extremely close to the ground truth values (0.3 mV vs. 0.31 mV and 0.275 mV vs. 0.29

mV, respectively; Table 1). These observations further confirm that 'minis' detection algorithm is

exceptionally good at picking up small PSPs. The tendency to overestimate amplitudes of detected

smPSPs in high incidence conditions (i.e., Type 1 HI row in the tables) by all three algorithms reflects

the increased probability of major overlap between simulated events in these conditions.

Table 2 below shows the compiled mean 10-90% rise time estimates for simulated and real minis

detected by the three algorithms while Supplementary Table 2 shows repeated samples t-test p-

values for comparisons of mean 10-90% rise time values for the three different algorithms with

respect to ground truth values. All three algorithms tended to significantly overestimate the rise

times of detected minis in all simulated conditions. This can be explained by the fact that noise

fluctuations affect the baseline positioning by shifting it to the lowest trough some time prior to the

real start of the event. The rise times of detected events are also more likely to be overlaps of more

than a single event and, therefore, would tend to be longer if lumped together. This was especially

true for events detected by Clampfit which had a pronounced tendency to lump smaller events into

fewer bigger ones. Both MiniAnalysis and Clampfit had large range values. For example, MiniAnalysis

had a range of 0.02-18 ms and Clampfit had a range of 0.09-37 ms when detecting real mPSPs, while

'minis' had a range of 0.35-9.4 ms when detecting the same type of events (Table 2). This

observation is indicative of the fact that MiniAnalysis and Clampfit occasionally failed to accurately

position the baseline of a detected event and failed to reject 'wildly' inaccurate estimates.

Table 3 below shows the compiled mean 1/e decay time estimates for simulated and real minis

detected by the three algorithms while Supplementary Table 3 shows repeated samples t-test p-

values for comparisons of mean decay time values for the three different algorithms with respect to

ground truth values. Decay times of smPSPs detected by 'minis' were the closest to the ground truth

values when compared to the other two algorithms. For simulated realistic incidence rate conditions

(i.e., Type 1 RI and Type 2 rows in the tables) p-values were not significant for the 'minis' vs. the

ground truth comparison (Table 3). Unfortunately, the same cannot be said of MiniAnalysis and

pClamp which consistently over- and underestimated mean decay times, respectively (Table 3 and

Supplementary Table 3). These differences between the algorithms may stem from differences in

how decays are being estimated: Whether effective decays are measured or detected events are

fitted with exponential decay functions. 'Minis' reduce these discrepancies by averaging the two

methods to obtain a single estimate.

Table 4 below shows the compiled mean incidence rates for simulated and real minis detected by

the three algorithms while Supplementary Table 4 shows repeated samples t-test p-values for

comparisons of mean incidence rates for the three different algorithms with respect to ground truth.

Values produced by 'minis' and MiniAnalysis were close to each other as well as ground truth values

(albeit less so for MiniAnalysis) as indicated by corresponding p-values of the t-test statistic (Table 4

and Supplementary Table 4). Incidence rate values produced by Clampfit were considerably lower

than the other two algorithms or the ground truth values indicating that Clampfit missed many

events, as well as tended to lump events together. A distinct ability by 'minis' algorithm to pick up

smaller events was further supported by a considerably larger incidence estimate for real data

(comparable to the range of incidence rates reported in our companion article (Dervinis and Major,

2024)) when compared with the other two algorithms. Clampfit values for real data were smaller by

almost an order of magnitude revealing a really poor performance by this algorithm.

Discussion

Our analysis suggests that mini (mEPSC/P) incidence rates typically reported in neocortical slices in

the CNS synaptic function literature may be underestimated, by up to an order of magnitude.

Discrepancy of this size indicates that signal-to-noise ratio is a severe issue preventing accurate

detection of minis irrespective of the detection method used: That is, both 'template matching' and

'thresholded amplitude detection' approaches are significantly flawed in distinguishing real minis

from the background physiological noise. Having stated that, however, we also demonstrated that

the two approaches are not equally flawed and that 'thresholded amplitude detection' performs

reasonably well and is further improved by our novel mEPSC/P detection algorithm called 'minis'.

We quantified the performance of 'minis' using standard measures from signal detection theory –

namely, ROC (receiver operating characteristic) curves and sensitivity index d', alongside measures

of sensitivity – true positive rate (TPR) and false positive rate (FPR = 1 – specificity) – and

demonstrated superior performance of our algorithm relative to the other two most popular

algorithms in the field of minis research: MiniAnalysis and Clampfit. Our algorithm had a

performance edge over the other two algorithms not only for detecting moderately-sized (~0.3 mV)

mEPSPs, but also across a wide range of realistic mEPSP amplitudes, shapes and incidence rates.

Crucially, the latter included the higher minis' incidence rates observed in neocortical slice

experiments at body temperature, 13-80 minis/s, where temporal summation becomes an

increasing challenge for detection. As a necessary step in building a reliable quantal size estimation

method, we succeeded in developing an algorithm for detecting postsynaptic events that is

transparent, systematically evaluated, and flexible.

Notwithstanding its good performance, the 'minis' algorithm has important limitations. First, the

'minis' algorithm, just like the other two detection algorithms, cannot by itself alone be used to

estimate quantal sizes. Due to amplitude attenuation of spontaneous postsynaptic events and the

need to use amplitude detection thresholds, the estimated amplitude will typically only be an upper

limit on the actual population mean amplitude (unless a very low threshold is used in which case FPR

would increase). We have also demonstrated that mini detection is impaired on rising and decaying

trends of V_m. The severity of this problem worsens with the rate of change of V_m. The problem is

exacerbated as the time between neighbouring minis gets shorter. Therefore, the mean amplitude

estimate is not only affected by the presence of noise and by amplitude attenuation, but also by the

presence of other miniature postsynaptic events. All the above limitations also apply to estimating

the incidence rate of postsynaptic events. All the mentioned factors reduce event detectability and,

therefore, result in an underestimated incidence rate. However, 'detecting' noise events as false

minis can cause errors in the other direction if too low a detection threshold is set. Despite these

limitations, our program 'minis' fares better on this front compared to the other two algorithms:

'minis' performance is also superior on rising and decaying trends in V_m, and for closely-spaced

events. To our knowledge, this is the first time these problems have been addressed, and their

impact quantified in a systematic manner, and their implications discussed.

Other caveats concern the method for assessing the performance of detection algorithms. We did

not use the full plausible range of realistic distributions for simulating mEPSPs in terms of their

incidence rates, amplitudes, and their rise and decay times. However, the shapes of these

distributions are not entirely known, and this is an empirical question that remains unanswered.

Because of this reason, we refrained for trying to address this question here and avoided making

empirically dubious assumptions. Instead, we explored a range of scenarios where simulation

incidence rates ranged from more to less realistic. We explored the effects of varying signal/noise

ratios. We believe that the large size of the parameter space we have explored does allow us to

conclude that the three algorithms differ systematically in their performance, with our algorithm –

'minis' – being superior to the other two (in our hands at least).

We have not constructed ROC curves in the usual fashion by varying a detection threshold of each

algorithm. Unfortunately, we were limited by the nature of the algorithms, specifically Clampfit

which does not use a pure amplitude detection threshold but uses convolution of the data trace with

each of the templates. However, our manipulations of varying the incidence rate of simulated

events, as well as varying the amplitude of noise, allowed us to mimic effects of varying the

detection threshold. We are also aware of one particular issue regarding the classification of correct

rejections (true negatives). One could argue that a proper definition of correct rejections would

require treating every data sample point as a potential noise event that could either be falsely

detected as a signal event or correctly rejected as a noise event. Unfortunately, such a definition

would result in large numbers of correct rejections (true negatives) swamping the number of other

detection measures (true positives, false alarms (false positives), and misses (false negatives)) and,

therefore, would make the detection performance analysis essentially meaningless. Alternatively,

treating every 10 ms noise window as a basis for correct rejection would still produce a relatively

large number of correct rejections, as would similar, more complicated schemes. There is also no

reason why a single noise event should be limited to a single 10 ms window. Therefore, we thought

that a reasonably practical definition of correct rejections as rejected prominent noise fluctuations

fared better than the alternatives. Having explicit, transparent, and reproducible, methods to

benchmark the performance is better than relying on subjective examination of recording traces.

Finally, we would like to address the question of whether we used optimal detection settings for

each algorithm. MiniAnalysis settings were similar to those of 'minis'. The two algorithms are

somewhat similar and chosen settings were observed to give the best performance for both

algorithms when detecting simulated mPSPs, after considerable exploration. In case there is any

doubt about our method, we have made our recorded and simulated data and Matlab analysis code

publicly available. With regards to using Clampfit to analyse simulated data, we used 9 templates

(the maximum allowed) constructed in a typical fashion by averaging a number of waveforms with

similar shapes but separating qualitatively different shapes. These templates have also been made

available publicly. We tested different detection regimes with templates based on both smoothed or

unsmoothed traces and comparing detection of smoothed and unsmoothed recording traces and

chose the best regime. However, we did not construct new templates for every new recording or

every new simulation condition, because this is extremely time-consuming and is not an efficient

way to conduct this type of analysis, although Clampfit detection performance might have improved

marginally. Clampfit might perform better if the number of templates was not limited to 9, and if a

wide range of inbuilt templates was available. These issues are, however, outside the scope of this

study. Given these considerations, we are confident in the validity of our approach.

We defer the discussion of possible reasons why there is such a big discrepancy between reported

and expected minis' incidence rates in the synaptic physiology literature and how they can be

effectively addressed to our companion article (Dervinis and Major, 2024). It is worth noting,

however, that 'minis' or another similar algorithm can be used to objectively measure the properties

of pure noise fluctuations (with minis blocked). This would allow one, in principle, to separate

'signal' (minis) from noise components in traces of minis recorded in the presence of noise. Noise

events could potentially be 'removed' from the combined signal and noise event distribution by

direct histogram subtraction, although in practice it turns out one needs to compensate for the

decay of the underlying waveform, following minis, and noise peaks coinciding with minis peaks:

noise and pure noise-free minis histograms don't simply summate (Dervinis and Major, 2024).

Alternatively, noise traces can be used to more accurately 'reverse-engineer' the signal distribution

via simulations, by iteratively adjusting the shapes, amplitudes, and incidence rates of simulated

minis, and adding these to real noise, under the control of an optimisation algorithm to match up

the distributions of *simulated* minis plus real noise to *real* minis plus real noise. The companion

article (Dervinis and Major, 2024) explores this and presents a quantal analysis method based on the

'minis' spontaneous postsynaptic event detection algorithm.

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Figures

Figure 1: A Schematic representation of key steps in 'minis' detection algorithm.

An illustration of a step-by-step application of the mPSP/C detection algorithm using simulated data.

Numbered steps dealing with mini peaks/amplitudes are shown in red, steps used to determine the

baseline are shown in blue. Briefly, the steps are:

1. Bandstop filter (against mains).

2. Lowpass filter to remove high frequency noise.

3. Find peaks.

4. Discard smaller peaks with the peak integration period represented by the length of the

   bracket and the union symbol (∪).

5. Move to a higher peak (if exists) within the peak integration period discarding the earlier

   peak.

6. Position the baseline at a local minimum located within the time-to-peak period prior to the

   peak.

7. Position the baseline asymmetrically (80:20) around the minimum (colour change at the

inset).

8. Correct the start of the baseline if it overlaps with the previous peak.

9. Discard the peak if it falls outside the acceptable amplitude range (repeated three more

times).

10. Compare rise time halves to correct the baseline (can be repeated once more).

11. Repeat Step 9.

12. Compare rise time halves to correct the peak.

13. Repeat Step 9.

14. Correct the baseline if it significantly deviates from the 10%/20% rise time mark.

15. Repeat Step 9.

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Figure 2: Filtering and smoothing effects.

(A) A brief segment of V_m trace recording containing real minis and noise. The black trace is raw

(unfiltered and unsmoothed), whereas the red trace is the same data after filtering and smoothing

as described in Materials and Methods subsection called *'minis' detection algorithm*.

(B) Same as (A) but containing varying amplitude smEPSPs with a 20 minis/s incidence rate instead of

real minis.

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Figure 2: Simulation and detection performance.

(A) An example of simulated membrane potential recording with average simulated mEPSP

incidence rate of 20 minis/s. The top voltage trace shows a brief segment with randomly timed

smEPSPs with random, normally-distributed amplitudes (μ_1= 0.3 mV, σ_1 = 0.05 mV) and 10-90% rise

times (drawn from the upper half of the distribution with μ_2 = 0.05 ms, σ_2 = 2.5 ms); see Methods.

Dotted green vertical lines mark signal events corresponding to smEPSP peaks. The paler shaded

colour demarcates a 10-ms (± 5 ms) window for accepting the detection of a signal event. The

second traces from the top are corresponding recordings of V_m noise fluctuations (the 2nd recording

phase: 'noise-alone' condition, in TTX and transmitter blockers), after initial filtering and smoothing

(steps 1 and 2 of the 'minis' detection algorithm; Figure 2). Dotted vertical cyan lines indicate noise

events; the shaded colour demarcates a window of exclusion for correctly rejecting these events

(see Methods). The V_m third from the top trace is a hybrid one with smEPSPs added to the noise

recording and with signal events and noise 'events' indicated. The coloured V_m traces below show

detection performance for the three different algorithms.

(B) An example of simulated V_m recording with mEPSP incidence rate of 40 minis/s.

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Figure 3: Membrane potential rising, steady-ish and falling 'trend phases'.

(A) A short segment of a V_m trace showing five smEPSPs. Different V_m 'trend phases', based on the

overall upward or downward trend in *simulated* voltage, are marked in colour. Green arrows

indicate V_m points where the V_m rate of increase is assigned to the preceding mEPSP peak. The blue

arrow indicates a V_m point where the V_m rate of decay is assigned to the following mEPSP peak.

(B) Simulated V_m trace in (A) added to a noise recording. V_m 'trend phases' were determined in (A)

and are used to classify detected events in (B).

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Figure 4: Overall performance for detecting 'moderately'-sized smEPSPs (~0.3 mV).

(A) 'virtual' ROC (Receiver Operator Characteristic)-like curve showing performance for detecting

'moderately'-sized smEPSPs in terms of True Positive Rate TPR (sensitivity) and False Positive Rate

FPR (1 – specificity) for all three algorithms, averaged across all 14 cells. In each case, dataset varies

from point to point, as opposed to detection threshold itself (which is varied systematically in a

traditional ROC, using one dataset). Ideal performance would be a right-angle plot going vertically up

TPR-axis at FPR=0, then horizontally along the top of the plot at TPR=1; minis is close to ideal, and

superior to MiniAnalysis, which is not too far from ideal (despite its 'missed minis' problems).

Clampfit is far from ideal. Different points represent 14 different incidence rate/signal-to-noise ratio

conditions, averaged across all 14 cells. Moving (generally) from bottom to top/left to right,

simulated minis rates and signal-to-noise ratio are decreasing. Vertical and horizontal bars indicate

95% confidence interval around each mean. The dotted diagonal line indicates chance performance.

The hollow circles mark three realistic smEPSP incidence rates (80, 40, and 20 minis/s).

(B) Sensitivity (discriminability) index (d') in the same 14 conditions as in (A) for all three algorithms,

averaged across all 14 cells.

(C) 'Violin' plots for Area under the virtual-ROC curve in (A) for all three algorithms. Individual data

points represent the 14 individual recordings (cells), horizontally 'swarmed' to avoid overlaps. AUC

(Area Under Curve) is averaged across all 14 incidence rate/signal-to-noise conditions for each cell

for each detection algorithm. The mean is marked by a solid line over the 'violin' centre. The dashed

line indicates the 95% confidence limits of the mean, and coloured shapes indicate the approximate

(smoothed) probability densities (wider = higher probability of those values).

**** indicates (highly) significantly different at $p \leq 10^{-5}$ level, paired t-test.

(D) Similar plot for sensitivity index d' averaged across all 14 incidence rate/signal-to-noise

conditions for each cell and each detection algorithm.

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Figure 5: Moderately-sized (~0.3 mV) smEPSPs (40 minis/s): detection errors made by the three

algorithms.

(A-H) Examples of smEPSP detection errors commonly committed by the three algorithms. Traces in

blue, red, and magenta represent detection performance for 'minis', MiniAnalysis, and Clampfit,

respectively. The top black trace shows actual smEPSPs. Vertical green dotted lines mark actual

individual (i.e., non-summated) smEPSP peak locations with the fainter shaded colour demarcating a

10-ms window for accepting the detection of an mEPSP. Arrowheads indicate common detection

errors made by the three algorithms. (Details in text).

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Figure 6: Performance when detecting moderately-sized (~0.3 mV) smEPSPs as a function of time to

the nearest neighbour (mean inter-mini peak-to-peak interval). Pale shaded colours indicate 95%

confidence intervals.

(A) True positive rate (TPR).

(B) False positive rate (FPR).

(C) Sensitivity index d' (undefined but arbitrarily high for TPR > 0 and FPR = 0, e.g. for inter-mini

intervals below 1 ms, for minis algorithm).

(D) Cumulative true positive rate (TPR) (cumulative true positives normalised by the total unbanded

true positive count and scaled by the overall unbanded TPR; see text for details).

(E) Cumulative FPR (cumulative false positives normalised by the total unbanded false positive count

and scaled by the overall unbanded FPR).

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Figure 7: Performance when detecting moderately-sized (~0.3 mV) smEPSPs on a rising trend V_m as a

function of the V_m rate of change. Paler shaded colours indicate 95% confidence intervals.

(A) True positive rate (TPR).

(B) False positive rate (FPR).

(C) Sensitivity index d'.

(D) Cumulative TPR.

(E) Cumulative FPR.

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Figure 8: Performance when detecting moderately-sized (~0.3 mV) smEPSPs on the membrane

potential decaying trend phase as a function of the membrane potential rate of change. Paler

shaded colours indicate 95% confidence intervals.

(A) True positive rate (TPR)

(B) False positive rate (FPR).

(C) Sensitivity Index d'.

(D) Cumulative TPR.

(E) Cumulative FPR.

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Figure 12: Overall performance for detecting smEPSPs of various (realistic) discrete amplitudes at

'realistic' minis' rates (details in text).

(A) Partial virtual "ROC" curve showing performance for detecting small-to-large-sized smEPSPs (0.05

to 0.5 mV in increments of 0.05 mV selected and added at random to a noise trace from each of the

14 cells) in terms of TPR (sensitivity) and FPR (1 – specificity) for all three algorithms. Moving from

left to right different data points represent 3 different biologically realistic incidence rate conditions:

61, 38, and 27 minis/s. Vertical and horizontal bars indicate 95% confidence interval around the

mean. The dotted diagonal line indicates chance performance.

(B) Sensitivity index (d') when detecting smEPSPs in the same 3 conditions as in (A) for all three

algorithms.

(C) Area under the virtual-ROC curve in (A) for all three algorithms. Individual data points represent

individual recordings (n=14 cells), averaged over the three simulated minis incidence rates. The

mean is marked by a solid line over the violin centre. The dashed line indicates the 95% confidence

limits. **** indicates (highly) significantly different at $p \leq 10^{-5}$ level, paired t-test.

(D) Sensitivity index d' averaged across all three simulated minis incidence rates, for each cell (n=14),

separately for each detection algorithm.

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Figure 13: Performance for detecting smEPSPs of various (realistic) discrete amplitudes as a function

of the time to the nearest neighbour.

(A) True positive rate (TPR) when detecting smEPSPs of various discrete amplitudes (0.05 to 0.5 mV

in increments of 0.05 mV) added at random to noise waveforms, as in previous figure, as a function

of the time to the nearest neighbour. Pale shaded colours indicate 95% confidence intervals.

(B) False positive rate (FPR).

(C) d' (undefined but arbitrarily high for TPR > 0 and FPR = 0, e.g. for inter-mini intervals below 1 ms,

for minis algorithm).

(D) Cumulative TPR.

(E) Cumulative FPR.

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Figure 14: Performance for detecting smEPSPs of (realistic) discrete amplitudes on the membrane

potential rise phase.

(A) True positive rate (TPR) when detecting smEPSPs of various amplitudes (0.05 to 0.5 mV in

increments of 0.05 mV, added at random to noise waveforms, as in the previous figure), on the V_m

rise phase as a function of the V_m rate of change. Pale shaded colours indicate 95% confidence

intervals.

(B) False positive rate (FPR).

(C) d'.

(D) Cumulative TPR.

(E) Cumulative FPR.

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Figure 15: Performance for detecting smEPSPs of (realistic) discrete amplitudes on the membrane

potential decay phase under realistic incidence rate conditions.

(A) True positive rate (TPR) when detecting smEPSPs of various amplitudes (0.05 to 0.5 mV in

increments of 0.05 mV, as in previous figure) on the V_m decay phase, as a function of the V_m rate of

change. Pale shaded colours indicate 95% confidence intervals.

(B) false positive rate (FPR).

(C) d'.

(D) Cumulative TPR.

(E) Cumulative FPR.

Tables

Condition	Truth	Minis ± 95% CI	Range	MiniAnalysis ± 95% CI	Range	Clampfit ± 95% CI	Range
Type 1 All	0.3	0.36±0.003	0.1-4.49	0.43±0.003	0.1-5.78	0.45±0.005	0.005-8.32
Type 1 HI	0.3	0.44±0.003	0.1-4.49	0.53±0.002	0.1-4.32	0.49±0.004	0.005-8.32
Type 1 RI	0.3	**0.31±0.004**	0.1-2.46	0.36±0.004	0.1-2.48	0.35±0.006	0.005-2.73
Type 2	0.275	0.29±0.002	0.06-2.48	0.32±0.002	0.06-2.35	0.34±0.004	0.05-1.2
Real data	-	0.05±0.001	0.02-3.97	0.09±0.002	0.02-4.87	0.21±0.006	0.07-2.67

Table 1: Mean amplitudes of detected simulated and real minis by the three different algorithms. All values are given in mV. Abbreviations: Confidence interval (CI), all conditions with 0.3 mV amplitude simulated minis used to construct the virtual ROC curve (Type 1 All), high incidence rate conditions (640, 320, and 160 minis/s) with 0.3 mV amplitude simulated minis used to construct the virtual ROC curve (Type 1 HI), realistic incidence rate conditions (80, 40, and 20 minis/s) with 0.3 mV amplitude simulated minis used to construct the virtual ROC curve (Type 1 RI), all realistic incidence rate conditions with varying amplitude simulated minis (Type 2). Statistically not significant comparisons to ground truth values are emphasised in bold (p-values are provided in Supplementary Table 1).

Condition	Truth	Minis ± 95% CI	Range	MiniAnalysis ± 95% CI	Range	Clampfit ± 95% CI	Range
Type 1 All	2.6	3.31±0.018	0.55-9.15	3.7±0.015	0.01-22.3	5.05±0.022	0.05-39.4
Type 1 HI	2.6	3.05±0.005	0.55-8.8	3.37±0.004	0.01-20.8	5.12±0.018	0.05-39.4
Type 1 RI	2.6	3.97±0.037	0.75-9.1	3.9±0.041	0.01-22.3	4.8±0.032	0.05-39.4
Type 2	2.6	3.69±0.014	0.65-9	3.29±0.022	0.02-57.5	4.77±0.013	0.06-35.9
Real data	-	3.37±0.044	0.35-9.4	3.42±0.051	0.02-18	4.72±0.052	0.09-37

Table 2: Mean 10-90% rise times of detected simulated and real minis by the three different algorithms**.** All values are given in ms. Abbreviations are the same as in Table 1. All comparisons to ground truth values were significant (p-values are provided in Supplementary Table 2).

Condition	Truth ± 95% CI	Minis ± 95% CI	Range	MiniAnalysis ± 95% CI	Range	Clampfit ± 95% CI	Range
Type 1 All	10.9±0.33	**11.2±0.22**	0.65-30.1	13.2±0.1	0.05-30.1	7.75±0.03	0.18-30.1
Type 1 HI	10.9±0.33	9.8±0.16	0.65-30.1	12.7±0.078	0.05-30.1	7.73±0.02	0.18-30.1
Type 1 RI	10.9±0.33	**10.9±0.19**	0.8-30.1	14.1±0.14	0.05-30.1	7.63±0.05	0.2-29.7
Type 2	10.9±0.33	**10.9±0.19**	0.75-30.1	13.9±0.13	0.05-30.1	7.37±0.04	0.16-28.2
Real data	-	11.2±0.44	0.45-30.1	12.3±0.24	0.05-30.1	7.02±0.06	0.28-28.9

1185   **Table 3:** Mean decay times of detected simulated and real minis by the three different algorithms.

1186   All values are given in ms. Abbreviations are the same as in Table 1. Statistically not significant

1187   comparisons to ground truth values are emphasised in bold (p-values are provided in Supplementary

1188   Table 3).

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Condition	Truth	Minis ± 95% CI	Range	MiniAnalysis ± 95% CI	Range	Clampfit ± 95% CI	Range
Type 1 All	93.6	40.8±4.54	29.3-146.5	34.7±0.43	30.4-41.8	12.3±0.13	10.5-13.9
Type 1 HI	379.2	64.1±6.04	41-205.1	53.9±0.3	48.5-55.5	21.5±0.1	20.4-22.4
Type 1 RI	47.4	**50±6.85**	32.9-209.4	39±0.81	33.2-52.3	20.2±0.39	15.9-24.4
Type 2	39	**40.1±5.66**	28.5-172.9	32.7±0.41	29.9-37.5	17.9±0.08	17-18.8
Real data	-	94.7±2.67	63.1-124.5	74.3±1.38	55.7-89.7	8.19±0.34	4.34-12.4

1190 **Table 4:** Mean incidence rates of detected simulated and real minis by the three different

1191   algorithms. All values are given in minis/s. Abbreviations are the same as in Table 1. Statistically not

1192   significant comparisons to ground truth values are emphasised in bold (p-values are provided in

1193   Supplementary Table 4).

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Supplementary Results

Research literature measuring minis in the central nervous system: Extended report

There were 107 studies in total that matched our criteria. All of these studies used voltage clamp as the electrophysiological method for recording spontaneous and miniature postsynaptic events. We surveyed the software they used to detect these events. The biggest group, ~48%, reported using MiniAnalysis. 26% reported using Clampfit, while 13% used a custom algorithm. The remaining 13% did not specify. We compared the performance of the two most popular detection algorithms, MiniAnalysis and Clampfit, both commercially available, with our new algorithm 'minis'.

35 studies (ca. 32%) reported using a current amplitude detection threshold of which 19 studies specified actual numeric value used (-6.8 ± s.d. 0.44 pA, range of -3 to -20 pA). It is likely that a higher fraction of studies made use of amplitude thresholds as only two explicitly reported not using any detection thresholds. Most of the studies did not give enough details (65% of all studies) to judge one way or the other. However, studies that used MiniAnalysis for detection must have used a detection threshold, as this software requires it.

Out of all the studies, 18 carried out research in neocortical pyramidal cells looking at mPSCs only (spontaneous PSCs excluded). 12 studies reported a mean mEPSC amplitude value of -11.74 ± s.d. 0.77 pA (range of -6.0 to -20.9 pA). 13 studies reported a mean incidence rate value of 3.6 ± s.d. 0.34 per second (range of 1.0 to 8.0 per second).

The total putative excitatory synapse count on a single cortical pyramidal cell can number in the tens of thousands (Eyal et al., 2018b). Layer 2/3 pyramidal cells in rats are thought to have 5,000 to 30,000 putative excitatory synapses (Larkman, 1991; Eyal et al., 2018b). Large-thick-trunk/tufted layer 5 pyramidal cells would have an even larger number of contacts (Larkman, 1991). Therefore, we can roughly assume that for most cortical pyramidal cells the number of putative excitatory

synapses ranges between 5,000 and 50,000. Conservatively, we could assume that only a minority,

roughly 40%, of them are functional ('non-silent') synapses with active zones and post-synaptic

densities containing AMPA receptors (Holler et al., 2021; Motta et al., 2019). Given that a

spontaneous vesicle release at a single release site has been estimated to occur with an approximate

incidence rate of 0.0021 per second (Murthy and Stevens, 1999) and given that there may be at least

2.7 release sites per cortical synapse (Holler et al., 2021), the lower bound incidence rate estimate of

such events within a single cortical pyramidal neuron might be expected to range somewhere

between ca. 10 minis/s and 100 minis/s (5,000*0.4*0.0021*2.7– 50,000*0.4*0.0021*2.7). Incidence

rates values reported in the recent literature are below this predicted range by an order of

magnitude, suggesting a significant fraction of smaller minis may commonly be 'missed' under some

recording conditions. This problem has been observed directly (Nevian et al., 2007), using paired

dendritic and somatic whole-cell patch recordings, albeit using voltage recording rather than voltage

clamp, recording from large layer 5 neocortical pyramidal neurons.

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Detection under realistic incidence rate conditions (20, 40, 80 minis/s)

Data points used to construct the full v-ROC curve include incidence rate and noise conditions that

were not necessarily realistic. Based on our calculations, the range of mEPSP incidence rates one

might commonly expect during real recordings from neocortical pyramidal neurons is likely to be

between 10 and 100 minis/s (see above; also G.M., unpublished experimental observations); In

Figures 4A and B, 20, 40 and 80 minis/s (from right to left) are highlighted by open circles. Just as

across the entire ROC curve, the performance of 'minis' was superior in this incidence rate range

compared to the other two algorithms with MiniAnalysis coming second, and Clampfit being worst,

by a substantial margin.

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Other measures like true positive and false positive rates as functions of the time to the nearest

neighbour and V_m rate of change during the rise or decay phases pointed to a very similar conclusion

made regarding the full ROC curve. First, we found that 'minis' performed better than other

algorithms at detecting mEPSPs that were within 10 ms of other mEPSPs and that at longer delays

MiniAnalysis performed just as well (Supplementary Figure 1). Meanwhile, Clampfit performed

consistently worst across the range of inter-mini intervals, whether measured by d' (Supplementary

Figure 1C) or true positive and false positive rates (Supplementary Figures 1A, B, D, and E). When it

came to V_m rate of change, 'minis' consistently showed the best performance at detecting smEPSPs

occurring with the background V_m rate of change ranging between -40 and 100 µV/s (Supplementary

Figures 2 and 3). This is the range where the vast majority of simulated events occurred

(Supplementary Figures 2D and 3D). In terms of individual TPR and FPR (Supplementary Figures 2A

and B and 3A and B) and cumulative rates (Supplementary Figures 2D and E and 3D and E), the

conclusions are much the same as those made regarding the full virtual ROC curve. Namely, that

'minis' had the largest TPR and the smallest FPR with an exception being the TPR in relation to rising

background (simulated) membrane potential trends (Supplementary Figures 2A and D), with there

being a range (45-100 µV/s) where MiniAnalysis had a slightly higher TPR, but not overall

performance measured as d' (Supplementary Figure 2C). Similar to conditions used to construct the

full ROC curve, Clampfit showed consistently the worst performance by all measures. The

performance of all three algorithms was better with realistic smEPSP incidence rates (Supplementary

Figures 2C and 3C) than over the full 'virtual' ROC curve (Figures 7C and 8C). In summary, evaluation

of detection of mEPSPs simulated at realistic incidence rates supported conclusions made regarding

detection performance under a much broader range of simulation incidence rates.

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Supplementary References

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 Supplementary Figures

Supplementary Figure 1: Performance when detecting moderately-sized (~0.3 mV) smEPSPs as a

function of time to the nearest neighbour with realistic mini rates (20, 40, 80 minis/s). Paler shaded

colours indicate 95% confidence intervals.

(A) True positive rate (TPR).

(B) False positive rate (FPR).

(C) Sensitivity index d' (undefined but arbitrarily high for TPR > 0 and FPR = 0, e.g. for inter-mini

intervals below 1 ms, for minis algorithm).

(D) Cumulative TPR.

(E) Cumulative FPR.

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Supplementary Figure 2: Performance when detecting moderately-sized (~0.3 mV) smEPSPs on the

rising trend V_m phases with realistic minis' rates (20, 40, 80 minis/s, as in Fig. 9), as a function of the

rate of change of V_m. Pale shaded colours indicate 95% confidence intervals.

(A) True positive rate (TPR)

(B) False positive rate (FPR).

(C) Sensitivity index d'.

(D) Cumulative TPR.

(E) Cumulative FPR.

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Supplementary Figure 3: Performance when detecting moderately-sized (~0.3 mV) smEPSPs on the

membrane potential decay phase with realistic minis rates (20, 40, 80 minis/s, as in Figs. 9 and 10).

Pale shaded colours indicate 95% confidence intervals.

(A) True positive rate (TPR) when detecting moderately-sized smEPSPs on the V_m decay phase as a

function of the V_m rate of change.

(B) False positive rate (FPR).

(C) Sensitivity index d'.

(D) Cumulative TPR.

(E) Cumulative FPR.

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 # Supplementary Tables

Condition	Minis vs. truth	MiniAnalysis vs. truth	Clampfit vs. truth
Type 1 All	1.01×10^{-7}	3.91×10^{-12}	7.49×10^{-10}
Type 1 HI	3.49×10^{-13}	5.84×10^{-17}	1.39×10^{-12}
Type 1 RI	**0.17**	8.89×10^{-6}	0.00021
Type 2	1.18×10^{-5}	7.48×10^{-9}	1.26×10^{-7}
Real data	-	-	-

1318 **Supplementary Table 1:** T-test p-values for mean amplitude values of detected minis using different

1319 algorithms. Repeated measures t-tests were used when comparing performance of two algorithms

1320 and single sample t-tests were used when comparing mean values with ground truth (subtracting

1321 ground truth values). Abbreviations are the same as in Table 1.

1322

Condition	Minis vs. truth	MiniAnalysis vs. truth	Clampfit vs. truth
Type 1 All	1.33×10^{-11}	5.03×10^{-15}	1.54×10^{-17}
Type 1 HI	4.68×10^{-16}	4.33×10^{-21}	9.95×10^{-19}
Type 1 RI	4.14×10^{-10}	2.16×10^{-10}	5.47×10^{-15}
Type 2	1.31×10^{-13}	2.13×10^{-10}	5.98×10^{-20}
Real data	-	-	-

1323 **Supplementary Table 2:** T-test p-values for mean 10-90% rise time values of detected minis using

1324 different algorithms**.** Repeated measures t-tests were used when comparing performance of two

1325 algorithms and single sample t-tests were used when comparing mean values with ground truth.

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Condition	Minis vs. truth	MiniAnalysis vs. truth	Clampfit vs. truth
Type 1 All	**0.3**	0.0006	0.0002
Type 1 HI	0.006	0.002	0.0002
Type 1 RI	**0.91**	1.07×10^{-5}	0.0002
Type 2	**0.97**	3.02×10^{-6}	6.61×10^{-5}
Real data	-	-	-

1327 **Supplementary Table 3:** T-test p-values for mean decay time values of detected minis using different

1328 algorithms. Repeated measures t-tests were used when comparing mean values. Abbreviations are

1329 the same as in Table 1.

1330

Condition	Minis vs. truth	MiniAnalysis vs. truth	Clampfit vs. truth
Type 1 All	2.09×10^{-5}	1.4×10^{-18}	3.93×10^{-27}
Type 1 HI	3.31×10^{-13}	2.64×10^{-30}	3.6×10^{-37}
Type 1 RI	**0.835**	6.6×10^{-5}	6.7×10^{-15}

| | Type 2 | **0.852** | 1.12×10^{-6} | 4.96×10^{-22} |
| | Real data | - | - | - |

1331 **Supplementary Table 4.** T-test p-values for mean incidence rates values of detected minis using

1332 different algorithms. Repeated measures t-tests were used when comparing performance of two

1333 algorithms and single sample t-tests were used when comparing mean values with ground truth.

1334 Abbreviations are the same as in Table 1.