The GR2D2 Estimator for the Precision Matrices

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Abstract

Biological networks are important for the analysis of human diseases, which summarize the regulatory interactions and other relationships between different molecules. Understanding and constructing networks for molecules, such as DNA, RNA and proteins, can help elucidate the mechanisms of complex biological systems. The Gaussian Graphical Models (GGMs) are popular tools for the estimation of biological networks. Nonetheless, reconstructing GGMs from high-dimensional datasets is still challenging. Current methods cannot handle the sparsity and high-dimensionality issues arising from datasets very well. Here we developed a new GGM, called the GR2D2 (Graphical R^2-induced Dirichlet Decomposition) model, based on the R2D2 priors for linear models. Besides, we provided a data-augmented block Gibbs sampler algorithm. The R code is available at https://github.com/RavenGan/GR2D2. The GR2D2 estimator shows superior performance in estimating the precision matrices compared to existing techniques in various simulation settings. When the true precision matrix is sparse and of high dimension, the GR2D2 provides the estimates with smallest information divergence from the underlying truth. We also compare the GR2D2 estimator to the graphical horseshoe estimator in five cancer RNA-seq gene expression datasets grouped by three cancer types. Our results show that GR2D2 successfully identifies common cancer pathways and cancer-specific pathways for each dataset.

1 Introduction

Biological networks, which capture the regulatory interactions or other relationships between molecules, such as DNA, RNA and protiens, are important for us to understand the mechanisms of complicated biological systems [3, 13]. The genetic regulatory networks, for instance, have played important roles of analysing gene expression data, for they can capture the complex gene-gene interactions and make the inferences of the dependency structures of genes possible [39, 40]. The Gaussian Graphical Models (GGMs) have been a popular tool in genomics to describe the functional dependencies between genes of interests in graphs,

since these models can circumvent indirect associations between genes by evaluating conditional dependencies in multivariate Gaussian distributions [17, 18, 30, 39]. GGMs have also been successfully applied to different scenarios to infer the genetic regulatory networks [7, 19, 24, 29, 31, 41]. In GGMs, we assume that the multivariate vector of genes follows a multivariate normal distribution with a particular structure of the inverse of the covariance matrix, called the precision matrix. Under this assumption, the conditional independence of two genes is equivalent to a zero value of the corresponding element in the precision matrix. The graph generated by GGMs can provide lucid interpretations of the relationship between genes in which nodes represent genes and edges between them represent interactions. The set of edges, which decides the network topology, can also be used to generate hypotheses about genetic interactions [18].

However, reconstructing GGMs from high-dimensional datasets remain a difficult task. The standard estimation of the partial correlation of genes either estimates the precision matrix or the parameters in p least squares regression problems, where p is the number of genes. If the sample size n is much smaller than the number of genes p, both methods are inappropriate [16]. Possible alternatives are either using penalization or the Bayesian framework to estimate the precision matrix. Consider n samples from a p-dimensional multivariate normal distribution with zero mean and a $p \times p$ covariance matrix Ω^{-1}, which is

$$\boldsymbol{y}_k \sim \text{Normal}(\boldsymbol{0}, \Omega^{-1}),$$

where $k = 1, 2, \ldots, n$ and Ω is the precision matrix. The i, j-th off-diagonal element of Ω is the negative of the partial covariance between features i and j, and the i-th feature is regressed on all the other features [26]. Besides, under the multivariate normal assumption, zero off-diagonal elements in Ω are conditionally independent given the remaining features.

One major challenge in estimating precision matrices is that the number of parameters increases quadratically with the number of features, leading to a sparse matrix. One approach to deal with sparsity is to penalized the likelihood functions. In graphical lasso [9], they maximize the penalized likelihood

$$\log(|\Omega|) - \text{tr}(S\Omega/n) - \sum_{i<j} \phi_\lambda(|\omega_{ij}|), \tag{1}$$

where $S = \sum_{i=1}^{n} \boldsymbol{y}_i^T \boldsymbol{y}_i$, $\Omega = (\omega_{ij})$, $\phi_\lambda(|\omega_{ij}|)$ is the l_1 penalty, and λ is a tuning parameter. A Bayesian version of the graphical lasso [36] is proposed afterwards, and the goal is to maximize a posterior estimate of Ω under the prior

$$p(\Omega|\lambda) \propto \prod_{i<j} (\text{DE}(\omega_{ij}|\lambda)) \prod_{i=1}^{p} (\exp(\omega_{ii}|\lambda/2)) \boldsymbol{1}_{\Omega \in \mathcal{S}_p},$$

where $\text{DE}(x|\lambda)$ denotes the double exponential distribution with rate λ, $\exp(x|\lambda)$ denotes the exponential distribution with rate λ, and $\mathcal{S}_p$ is the space of $p \times p$ positive definite matrices.

The smoothly clipped absolute deviation (SCAD) penalty [8] was also introduced to estimate the precision matrix, in which they maximized the penalized likelihood in equation 1 where the penalty term has the first-order derivative

$$\phi_\lambda^{'}(|x|) = \lambda(\boldsymbol{1}_{|x|\leq\lambda} + \frac{(a\lambda - |x|)}{(a-1)\lambda}\boldsymbol{1}_{|x|>\lambda}),$$

with $a > 2$ and $\lambda > 0$. However, the performance of this method relies on the choice of tuning parameters, which cannot be easily implemented in reality.

The graphical horseshoe estimator employs a Bayesian framework and puts horseshoe priors on the off-diagonal elements of precision matrices, and puts uninformative prior on the diagonal elements [20]. The model is summarized in the following

$$
\begin{aligned}
\omega_{ii} &\propto 1, \\
\omega_{ij;i<j} &\sim \text{Normal}(0, \lambda_{ij}^2 \tau^2), \\
\lambda_{ij;i<j} &\sim C^+(0, 1), \\
\tau &\sim C^+(0, 1)
\end{aligned}
$$

where $C^+(0, 1)$ denotes a half-cauchy random variable with density $p(x) \propto (1 + x^2)^{-1}; x > 0$. The distinctive scale parameter λ_{ij} is treated as a local shrinkage parameter, and the scale parameter τ is shared by all dimensions, treated as a global shrinkage parameter.

The graphical lasso, Bayesian graphical lasso, and the graphical SCAD face the problem of accumulated estimation errors due to increasing number of parameters [20], while the graphical horseshoe estimator shows weak performance in high-dimensional settings when the number of features p is much larger than the sample size n. Here, we propose an alternative Bayesian estimator called the GR2D2 (Graphical R^2-induced Dirichlet Decomposition) estimator that can handle sparse and high-dimensional issues faced by other methods. The R2D2 shrinkage priors have shown promising performance in high-dimensional linear regression models, having greater concentration near the origin and heavier tails than current shrinkage priors [42]. We extended the R2D2 priors from linear models to graphical models and our simulation results show that the GR2D2 estimators achieved the state-of-art performance in sparse and high-dimensional settings, compared with the graphical lasso and the graphical horseshoe estimator. We further compare the GR2D2 estimator to the graphical horseshoe estimator in five cancer RNA-seq gene expression datasets grouped by three cancer types. Our results show that the graphical horseshoe estimator always gives highly densely connected networks and it is very hard to identify relevant gene sets for a specific cancer pathway. GR2D2 outputs sparse networks with clear structures and it successfully identifies common cancer pathways and cancer-specific pathways.

The remaining sections are organized as follows. We outlines the GR2D2 estimator with a full Gibbs sampler for efficient sampling. Next we present our simulation studies to show the efficiency of the GR2D2 followed by real data analysis.

2   Methods

For Gaussian graphical models, it is usually assumed that the patterns of variation in expression of a gene can be predicted by those of a small subset of other genes, which leads to a sparse structure in the precision matrix [40]. For such precision matrix, we need a shrinkage method that is able to give zero or very small estimate for the zero elements. Besides, this method should also be able to sustain the nonzero values and shrink them as little as possible. We propose the GR2D2 estimator to achieve this goal.

2.1 The GR2D2 Hierarchical Model

The GR2D2 model puts R2D2 priors on the off-diagonal elements of the precision matrix. The element-wise priors are specified for $i, j = 1, \ldots, p$ as follows

$$\omega_{ii} \propto 1, \quad \omega_{ij} \sim \mathrm{N}(0, \phi_{ij}\psi_{ij}\omega/2),$$
$$\Phi \sim \mathrm{Dir}(\alpha_\pi, \alpha_\pi, \ldots, \alpha_\pi), \quad \psi_{ij} \sim \exp(1/2),$$
$$\omega|\xi \sim \mathrm{Gamma}(a, \xi), \quad \xi \sim \mathrm{Gamma}(b, 1),$$

where $\mathrm{N}(0, 1)$ denotes a normal random variable with mean 0 and variance 1; $\mathrm{Dir}(\alpha_\pi, \alpha_\pi, \ldots, \alpha_\pi)$ denotes a vector of Dirichlet distributed random variables with a parameterized vector $\boldsymbol{\alpha} = [\alpha_\pi, \alpha_\pi, \ldots, \alpha_\pi]^T$; and the vectors $\boldsymbol{\alpha}$ and Φ are of length p^2. However, in the following notations, we use Φ as a symmetric matrix of dimension $p \times p$ with entries ϕ_{ij} and use $\boldsymbol{\phi}$ as a row or column vector of Φ. Further, let $\exp(1/2)$ denote an exponential random variable with rate $1/2$; let $\mathrm{Gamma}(1, 1)$ denote a Gamma random variable with shape 1 and rate 1. The distinctive scale parameters ϕ_{ij} and ψ_{ij} on each dimension are referred to as the local shrinkage parameters, and the scale parameter ω shared by all dimensions is referred to as the global shrinkage parameter. Note that ξ is used as an auxiliary parameter to help sample ω. Since conditional independence of each entry is guaranteed in precision matrices given the remaining entries under the multivariate normal assumption, the prior on Ω under the GR2D2 model can be written as the product of off-diagonal entries

$$p(\Omega|\boldsymbol{\phi}, \psi_{ij}, \omega) \propto \prod_{i<j} f(\omega_{ij}|\phi_{ij}, \psi_{ij}, \omega)\mathbf{1}_{\Omega \in \mathcal{S}_p},$$

where $f(\omega_{ij}|\phi_{ij}, \psi_{ij}, \omega)$ stands for the normal density function of ω_{ij} given parameters $\phi_{ij}, \psi_{ij}, \omega$, and $\mathcal{S}_p$ is the space of $p \times p$ positive definite matrices.

In high-dimensional precision matrix estimation using GR2D2, the global shrinkage parameter ω adapts to the sparsity of the entire matrix Ω and shrinks the estimates of the off-diagonal elements toward zero. On the flip side, the local shrinkage parameters, ϕ_{ij} and ψ_{ij} for $i < j$, preserve the magnitude of nonzero off-diagonal elements, and ensure that the element-wise biases are not very large.

2.2 A Data-Augmented Block Gibbs Sampler

Posterior samples under the GR2D2 hierarchical model are drawn by an augmented block Gibbs sampler, utilizing the scheme proposed by [22] for linear regression. In each iteration, each column and row of Ω and $\Lambda = (\phi_{ij}\psi_{ij})$ are partitioned from a $p \times p$ matrix of parameters and updated in a block. Then the global shrinkage parameter ω and its auxiliary variable ξ are updated.

Given data $Y_{n \times p}$ and shrinkage parameters, we provide necessary derivation of the posterior distribution of the precision matrix as follows. Define the matrices

$$\Omega = \begin{bmatrix} \Omega_{(-p)(-p)} & \boldsymbol{\omega}_{(-p)p} \\ \boldsymbol{\omega}_{(-p)p}^T & \omega_{pp} \end{bmatrix}, \quad S = \begin{bmatrix} S_{(-p)(-p)} & \boldsymbol{s}_{(-p)p} \\ \boldsymbol{s}_{(-p)p}^T & s_{pp} \end{bmatrix},$$

$$\Lambda = \begin{bmatrix} \Lambda_{(-p)(-p)} & \boldsymbol{\lambda}_{(-p)p} \\ \boldsymbol{\lambda}_{(-p)p}^T & \lambda_{pp} \end{bmatrix},$$

where the subindex $_{(-p)}$ denotes the set of all indices except for p, and the matrix Λ has entries $\lambda_{ij} = \phi_{ij}\psi_{ij}$. Diagonal elements of $\Lambda_{(-p)(-p)}$ are set to be 1. The posterior distribution of Ω is

$$p(\Omega|\Lambda, Y, \omega) \propto p(S|\Omega, \Lambda, \omega)p(\Omega|\Lambda, \omega)$$

$$\propto |\Omega|^{n/2} \exp(-\frac{1}{2}tr(\Omega S)) \prod_{i<j} \exp(-\frac{\omega_{ij}^2}{\psi_{ij}\phi_{ij}\omega}),$$

where $S = Y^T Y \sim W_p(\Omega^{-1}, n)$ and W stands for a Wishart distribution. After some calculation, the full conditional of the last column of Ω is

$$p(\boldsymbol{\omega}_{(-p)p}, \omega_{pp}|Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$

$$\propto (\omega_{pp} - \boldsymbol{\omega}_{(-p)p}^T \Omega_{(-p)(-p)}^{-1} \boldsymbol{\omega}_{(-p)p})^{n/2}$$

$$\times \exp(-\frac{1}{2}(2\boldsymbol{s}_{(-p)p}^T \boldsymbol{\omega}_{(-p)p} + s_{pp}\omega_{pp}))$$

$$\times \exp(-\boldsymbol{\omega}_{(-p)p}^T (\Lambda^* \omega)^{-1} \boldsymbol{\omega}_{(-p)p}),$$

where $\Lambda^* = \mathrm{diag}(\boldsymbol{\lambda}_{(-p)p})$. Let $\boldsymbol{\beta} = \boldsymbol{\omega}_{(-p)p}$, $\gamma = \omega_{pp} - \boldsymbol{\beta}^T \Omega_{(-p)(-p)}^{-1} \boldsymbol{\beta}$, and $\omega_{pp} = \gamma + \boldsymbol{\beta}^T \Omega_{(-p)(-p)}^{-1} \boldsymbol{\beta}$. The Jacobian of this transformation is a constant, and the full conditional of β and γ is

$$p(\boldsymbol{\beta}, \gamma|Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$

$$\propto \gamma^{n/2} \exp(-\boldsymbol{\beta}^T (\Lambda^* \omega)^{-1} \boldsymbol{\beta})$$

$$\times \exp(-\frac{1}{2}(s_{pp}\gamma + s_{pp}\boldsymbol{\beta}^T \Omega_{(-p)(-p)}^{-1} \boldsymbol{\beta} + 2\boldsymbol{S}_{(-p)p}^T \boldsymbol{\beta}))$$

$$\propto \gamma^{n/2} \exp(-\frac{1}{2}s_{pp}\gamma)$$

$$\times \exp(-\frac{1}{2}(\boldsymbol{\beta} + D\boldsymbol{s}_{(-p)p})^T D^{-1}(\boldsymbol{\beta} + D\boldsymbol{s}_{(-p)p})),$$

where $D = (2(\Lambda^* \omega)^{-1} + s_{pp}\Omega_{(-p)(-p)})^{-1}$. The above formula can be summarized as

$$p(\boldsymbol{\beta}, \gamma|Y, \Lambda, \omega, \Omega_{(-p)(-p)}) \propto \mathrm{Gamma}(\frac{n}{2} + 1, \frac{s_{pp}}{2})$$

$$\times \mathrm{Normal}(-D\boldsymbol{s}_{(-p)p}, D).$$

Hence, the posterior distribution of the last row and column of Ω is obtained. Since the derivation of the posterior distributions of ψ_{ij}, ω, ξ and $\boldsymbol{\phi}$ is standard, we summarize them in the sampling procedure section with a proposed algorithm structure.

2.3 Gibbs sampling procedures with initial settings

Based on the results derived in the previous section, we provide the initial settings used in our sampling procedures. We use the same initial settings as [20] and [42], and set p to be the

number of rows (or columns) in $S = Y^T Y$, $\Omega = I_{p\times p}$, $\Sigma = I_{p\times p}$, $\Phi = \mathbf{1}_{p\times p}$, $\Psi = \mathbf{1}_{p\times p}$, $\omega = 1$, $\xi = 1$, $b = 0.5$, $r = 1$, $C = 1$, $p_n = \frac{p(p-1)}{2}$, $\alpha_\pi = \frac{C}{p_n^{b/2} n^{rb/2} \log(n)}$, $a = \frac{p(p-1)}{2}\alpha_{pi}$, $\Lambda = \Phi \odot \Psi$, where Φ has entries of ϕ_{ij}, Ψ has entries of ψ_{ij}. The Gibbs sampling procedures are as follows:

(a) Set initial values as above.

(b) Choose the p-th column and do the following until the matrix Ω is updated.

    (b.1) Sample $\gamma | Y, \Lambda, \omega, \Omega_{(-p)(-p)} \sim \text{Gamma}(\text{shape} = \frac{n}{2} + 1, \text{rate} = \frac{s_{pp}}{2})$.

    (b.2) Sample $\boldsymbol{\beta} | Y, \Lambda, \omega, \Omega_{(-p)(-p)} \sim \text{Normal}(-D\boldsymbol{s}_{(-i)i}, D)$, where $D = (2(\Lambda^*\omega)^{-1} + s_{pp}\Omega_{(-p)(-p)})^{-1}$, $\Lambda^* = \text{diag}(\boldsymbol{\lambda}_{(-p)p})$, and $\lambda_{ij} = \phi_{ip}\psi_{ip}$.

    (b.2) Sample $\psi_{ij}^{-1} | \omega_{ij}, \phi_{ij}, \omega \sim \text{IG}(\mu = \sqrt{\phi_{ij}\omega/2}/|\omega_{ij}|, \lambda = 1)$, and then take the reciprocal to get ψ_{ij}. IG stands for an inverse Gamma distribution.

(c) Sample $\omega | \Omega, \Lambda, \xi \sim \text{giG}(\chi = 2\sum_{i<j} \frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}}, \rho = 2\xi, \lambda_0 = a - \frac{p(p-1)}{4})$, where giG stands for a generalized inverse Gaussian distribution.

(d) Sample $\xi | \omega \sim \text{Gamma}(\text{shape} = a + b, \text{rate} = \omega + 1)$

(e) Sample $\boldsymbol{\phi} | \Omega, \psi, \omega, \xi$. Based on our derivation, if $a = \frac{p(p-1)}{2}\alpha_\pi$, one can draw $T_1, \ldots, T_{\frac{p(p-1)}{2}}$ independently with $T_j \sim \text{giG}(\chi = 2\omega_j^2/\psi_j, \rho = 2\xi, \lambda_0 = \alpha_\pi - \frac{1}{2})$. Then set $\phi_j = T_j/T$ with $T = \sum_{j=1}^{\frac{p(p-1)}{2}} T_j$.

(f) Repeat steps (b)-(g) until convergence.

The algorithm 1 of the GR2D2, along with simulation examples, is available at this github website.

3 Simulation Studies

In the graphical horseshoe estimator [20], they compared the performance of the graphical horseshoe with the graphical SCAD[8], frequentist graphical lasso [9] and the Bayesian graphical lasso [36] using diverse assessment criteria and different structures of the precision matrices. Their results showed that the graphical horseshoe estimator achieved the best performance. Therefore, we only compare the GR2D2 estimator with the graphical horseshoe estimator and the graphical lasso, where the graphical lasso works as the baseline.

3.1 Simulation Setting and Assessment Criterion

We consider six precision matrix structures with different pairs of features and observations. These pairs are $(p, n) = (100, 50)$, $(150, 50)$, $(200, 50)$, and $(250, 50)$. The reason of this setting is that we intend to compare the GR2D2 and other estimators when the ratio p/n increases, which corresponds to high-dimensional settings. The diagonal entries of the precision matrix ω_0 is 1 for all six structures, while the off-diagonal entries depend on one of the following structures. We take the pair $(p, n) = (100, 50)$ as an example.

Algorithm 1 The GR2D2 Sampler

1: **function** $\text{GR2D2}(S, n, burnin, nmc)$ ▷ where $S = Y^T Y, n = $ sample size
2:     Set p to be the number of rows (or columns) in S
3:     Set initial values $\Omega = I_{p\times p}$, $\Sigma = I_{p\times p}$, $\Phi = \mathbf{1}_{p\times p}$, $\Psi = \mathbf{1}_{p\times p}$
4:     Set $\omega = 1$, $\xi = 1$, $b = 0.5$, $r = 1$, $C = 1$
5:     Set $p_n = \frac{p(p-1)}{2}$, $\alpha_\pi = \frac{C}{p_n^{b/2} n^{rb/2} \log(n)}$, $a = \frac{p(p-1)}{2}\alpha_{pi}$, $\Lambda = \Phi \odot \Psi$,
6:     where Φ has entris of ϕ_{ij}, Ψ has entris of ψ_{ij}
7:     **for** $iter = 1$ to $(\text{burnin} + \text{nmc})$ **do**
8:        **for** $i = 1$ to p **do**
9:           $\gamma \sim \text{Gamma}(\text{shape} = \frac{n}{2} + 1, \text{rate} = \frac{s_{ii}}{2})$
10:         $\Omega^{-1}_{(-i)(-i)} = \Sigma_{(-i)(-i)} - \boldsymbol{\sigma}_{(-i)i}\boldsymbol{\sigma}^T_{(-i)i}/\sigma_{ii}$
11:         $D = (s_{ii}\Omega^{-1}_{(-i)(-i)} + 2\text{diag}(\boldsymbol{\lambda}_{(-i)i}\omega)^{-1})^{-1}$
12:         $\boldsymbol{\beta} \sim \text{Normal}(-D\boldsymbol{s}_{(-i)i}, D)$ ▷ sample $\boldsymbol{\beta}$
13:         $\boldsymbol{\omega}_{(-i)i} = \boldsymbol{\beta}$, $\omega_{ii} = \gamma + \boldsymbol{\beta}^T \Omega^{-1}_{(-i)(-i)}\boldsymbol{\beta}$ ▷ variable transformation
14:         $\psi^{-1}_{ij} \sim \text{InvGaussian}(\mu_{ij} = \frac{\sqrt{\phi_{ij}\omega/2}}{|\omega_{ij}|}, \lambda = 1)$
15:         Take the reciprocal of ψ^{-1}_{ij} to get ψ_{ij} ▷ sample Ψ
16:         Save updated Ω
17:         $\Sigma_{(-i)(-i)} = \Omega^{-1}_{(-i)(-i)} + (\Omega^{-1}_{(-i)(-i)}\boldsymbol{\beta})(\Omega^{-1}_{(-i)(-i)}\boldsymbol{\beta})^T/\gamma$
18:         $\boldsymbol{\sigma}_{(-i)i} = -(\Omega^{-1}_{(-i)(-i)}\boldsymbol{\beta})/\gamma$, $\sigma_{ii} = 1/\gamma$
19:         Save updated Σ
20:        **end for**
21:        Motivated by [4], since $a = \frac{p(p-1)}{2}\alpha_\pi$
22:        $T_{ij} \sim \text{giG}(\chi = 2\omega^2_{ij}/\phi_{ij}, \rho = 2\xi, \lambda = \alpha_\pi - 1/2)$
23:        $\phi_{ij} = T_{ij}/T$, where $T = \sum_{i<j} T_{ij}$
24:        $\lambda_{ij} = \phi_{ij}\psi_{ij}$
25:        Save updated Λ
26:        $\omega \sim \text{giG}(\chi = \sum^{p-1}_{i=1}\sum^{p}_{j=i+1}\frac{2\omega^2_{ij}}{\phi_{ij}\psi_{ij}}, \rho = 2\xi, \lambda = a - \frac{p(p-1)}{4})$ ▷ sample ω
27:        $\xi \sim \text{Gamma}(\text{shape} = a + b, \text{rate} = \omega + 1)$ ▷ sample ξ
28:     **end for**
29:     Return MC samples Ω
30: **end function**

Hubs. The rows or columns are partitioned into disjoint groups $\{G_k\}_1^K$. Each group has a row k where off-diagonal elements are taken to be $\omega_{ik} = 0.25$ corresponding to negative partial correlations for $i \in G_k$ and $\omega_{ij} = 0$ otherwise, and we allow four members within each group. We consider 10 groups and 10 members within each group, giving 40 nonzero off-diagonal elements in Ω_0. The topological structure of this pattern is like a star with four members connected with one member at the center.

Cliques positive. The rows or columns are partitioned into disjoint groups and $\{\omega_{ij} : i, j \in G_k, i \neq j\}$ are set to -0.45 corresponding to positive partial correlation. We also use 10 groups in this setting but only also three members within each group resulting in 30 nonzero off-diagonal elements in Ω_0. The topological structure of this pattern is like a small circle with only three points connected with each other.

Cliques negative. The rows or columns are partitioned into disjoint groups and $\{\omega_{ij} : i, j \in G_k, i \neq j\}$ are set to 0.75 corresponding to positive partial correlation. Other settings are the same as those in cliques positive.

Scale-free positive We use the function barabasi.game from the igraph R package to generate a scale-free graph according to the Barabasi-Albert model (BA model) [2]. 100 (n = 100) vertices are generated with the power of the preferential attachment equal to 1 (power = 1). We set m = 1 to add one edge in each time step, resulting in a sparse network structure. The arguments n, power and m are used in the function barabasi.game. We then turn the graph into an adjacency matrix, which is treated as the precision matrix in our estimation. The off-diagonal elements are set to be -0.15, corresponding to the positive partial correlations.

Scale-free negative The settings in this structure are identical to those in scale-free positive except that the off-diagonal elements are set to be 0.15, corresponding to the negative partial correlations.

Scale-free half The settings in this structure are identical to those in scale-free positive except that we randomly sample half of the off-diagonal elements to be 0.15 and the other half to be -0.15.

We choose the hubs, cliques positive and negative structures since they are commonly used structures to verify the performance of graphical models [10]. One more structure, random, mentioned in [10], is not used here since the generated samples from the random structure may not always be positive definite and hence not invertible. We use three scale-free graph structures based on the BA model. This model generates graphs whose vertex connectivities follow a scale-free power-law distribution. This power-law distribution has been found in many natural and man-made networks [6]. Hence, we apply this scale-free structure to generate graphs that may represent realistic network structures.

The results of four pairs of observations and features and six structures are summarized in Figures 1 and 2, and in Supplementary Information Tables S1-S8. We also include the results

of three scale-free structures using $(p, n) = (250, 30)$ in Table S9 to show the performance of graphical models when the sample size is small. Considering the variation in the network topology [11], in Tables S10 and S11, we generated 30 different networks for Hubs, Cliques positive and Cliques negative structures with 30 different non-zero ω's and $(p, n) = (100, 50)$ and $(150, 50)$. (For Hubs, ω is ranging from 0.11 to 0.4 with step size 0.01; for Cliques positive, ω is ranging from -0.40 to -0.1 with step size 0.01; for Cliques negative, ω is ranging from 0.11 to 0.4 with step size 0.01) In these figures, we report the results of Frobenius norm, Stein's loss, precision, and recall, while in tables we also report false positive rate (FPR), accuracy, and F scores with corresponding standard deviation (s.d.).

For each choice of Ω_0, 30 datasets are generated and Ω is estimated using the graphical lasso, the graphical horseshoe and GR2D2 estimators. The graphical horseshoe estimator is implemented in MATLAB (2022a), while the graphical lasso is implemented using the package "glasso" in Rstudio (version 4.1.2). For the graphical horseshoe and the GR2D2 estimators, we drop the initial 500 burnin samples and keep the following 500 MCMC samples with convergence. We also use the median of the remaining 500 samples as our estimate for the graphical horseshoe and GR2D2.

The Stein's loss of the estimated precision matrix $\hat{\Omega}$ is equal to 2 times the Kullback-Leibler divergence of $\hat{\Omega}$ from Ω_0, with the formula

$$\mathcal{L}_S(\hat{\Omega}, \Omega_0) = \text{tr}(\hat{\Omega}\Omega_0^{-1}) - \log|\hat{\Omega}\Omega_0^{-1}| - p.$$

The Frobenius norm between $\hat{\Omega}$ and Ω_0 is

$$\mathcal{L}_F(\hat{\Omega}, \Omega_0) = ||\hat{\Omega} - \Omega_0||_F = \sqrt{\sum_{i=1}^{p}\sum_{j=1}^{p}|\hat{\omega}_{ij} - \omega_{ij}|^2}.$$

Since the graphical horseshoe and the GR2D2 are shrinkage methods that do not estimate elements to be exactly equal to zero, we use the 95% quantile intervals in the MCMC samples for variable selection. This is to say, if the 95% quantile intervals of an off-diagonal entry of Ω does not contain zero, this entry is considered as a discovery and the median values across the MCMC samples are used as the estimate. If the discoveries identified in $\hat{\Omega}$ have the same signs as those in Ω_0, we call them true positive discoveries and use them to calculate the precision, recall, positive rate, accuracy and F scores. For each measurement, we report the mean and s.d. computed over 30 datasets.

3.2 Estimation

Based on Figures 1 and 2, the GR2D2 estimates always have the smallest Steins' loss and most of the time have the smallest Frobenius norm (F norm). When the ratio p/n is 2, the graphical horseshoe estimators show better performance than GR2D2 in F norm in the structures cliques positive and cliques negative; and when the ratio between p and n is 3, the graphical horseshoe estimators show better performance than GR2D2 in F norm in the pattern cliques negative only. When p/n is 4 and 5, the GR2D2 shows consistent superior performance to the graphical horseshoe in all six structures. The GR2D2 is expected to

Figure 1: Frobenius norm (F-norm), Mean Stein's loss, precision, and recall of precision matrix estimates for hubs, cliques positive and negative network structures over 30 datasets generated by multivariate normal distribution with the sample size $n = 50$ and number of features $p = (100, 150, 200, 250)$. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the GR2D2. We use 2log(GL), 10log(GL) and 10log(GL) for the GL estimator in hubs, cliques positive and negative respectively when comparing F-norm and Stein's loss. For F-norm, GR2D2 performs better than or comparably well to GHS while GL performs poorly. For Stein's loss, GR2D2 consistently shows the best performance. For precision scores, GR2D2 and GHS show comparable good performance while GL provides the worst precision scores. For recall scores, GR2D2 and GHS show comparable performance in hubs structure while GHS performs better in cliques structure. GL performs the best regarding to the recall scores. Overall, GR2D2 shows the best estimation results according to F-norm and Stein's loss and comparable results to GHS in variable selection under hubs, cliques positive and negative network structures.

Figure 2: Frobenius norm (F-norm), Mean Stein's loss, precision, and recall of precision matrix estimates for scale-free positive, negative and half network structures over 30 datasets generated by multivariate normal distribution with the sample size $n = 50$ and number of features $p = (100, 150, 200, 250)$. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the GR2D2. We use 5log(GL) for the GL estimator in scale-free positive, negative and half when comparing F-norm and Stein's loss. For F-norm and Stein's loss, GR2D2 consistently shows the best performance. For precision scores, GR2D2 and GHS show comparable good performance while GL provides the worst precision scores. For recall scores, GR2D2 and GHS show comparable performance while GL performs the best. Overall, GR2D2 shows the best estimation results according to F-norm and Stein's loss and comparable results to GHS in variable selection under scale-free positive, negative and half network structures.

perform well when the precision matrix is sparse and the absolute values of scaled nonzero elements are large as mentioned in the work of [43]. Besides, the GR2D2 is able to deal with high dimensional datasets and always provides the best results as shown in $p/n = 4$ and $p/n = 5$. These results show that in both commonly used network structures and scale-free network structures, GR2D2 performs overall the best.

Besides, it could be noted that as the ratio between p and n increases, the Stein's loss and F norm increase accordingly, which is expected since when the datasets become sparser and the dimension becomes higher, it is harder to estimate the precision matrices. Considering the standard deviation across 30 datasets, the GR2D2 always gives 0 values, indicating that the estimation of GR2D2 could provide the most concentrated estimation results with least variation, as shown in Tables S1-S11. For the graphical horseshoe estimator, it shows relatively small variation while the graphical lasso provides the biggest variations.

3.3   Variable Selection

As shown in Figures 1 and 2, for the precision, GR2D2 always gives 1, indicating the all the positive observations are true observations. The graphical horseshoe provides results close to 1, showing comparably good results, while the graphical lasso provides quite small precision scores. For the scores of recall, the graphical lasso always gives the best recall scores while the results of graphical horseshoe and the GR2D2 improve as the ratio between p and n increases. Besides, the graphical horseshoe estimator most of the time gives better scores than the GR2D2. For FPR, as shown in Tables S1-S11, the GR2D2 always gives 0 values, indicating that there are 0 false positive result across all 30 datasets. The graphical horseshoe estimators provide results quite close to zero across 30 datasets, showing comparably good results to the GR2D2. However, the graphical lasso always provides the worst FPR scores, indicating that this method always provides false positive estimations. For the accuracy scores, both the graphical horseshoe and the GR2D2 show comparably good results with values more than 0.99, while the graphical lasso provides poorer accuracy results. One point that is worth mentioning is that since the precision matrices are quite sparse, there are always much less observations than zero values, resulting in unbalanced datasets. Therefore, the accuracy scores may not be a good measurement to assess the performance of three estimation methods. Next we turn to F scores, which can cooperate the results of precision and recall, giving measurement free from the effects of unbalanced datasets. According to results of F scores, the graphical horseshoe method and GR2D2 show comparable results in the six structures, while the graphical lasso performs poorly. We also put the formulae of precision, recall, FPR, accuracy and F scores in the Supplementary Information Section 2. Overall, both GR2D2 and the graphical horseshoe show promising abilities to correctly identify the true observations in variable selection under our simulation settings, while the graphical lasso shows weak performance.

4 Real data analysis

4.1 Datasets and pre-processing steps

Five RNA-seq gene expression datasets of cancerous tissues are obtained from the TCGA database via TCGAbiolinks (v. 2.25.2) [37]. These datasets are breast invasive carcinoma (BRCA) with 1094 cancer patients, kidney renal clear cell carcinoma (KIRC) with 533 cancer patients, kidney renal papillary cell carcinoma (KIRP) with 290 cancer patients, lung adenocarcinoma (LUAD) with 515 cancer patients, and lung squamous cell carcinoma (LUSC) with 502 cancer patients. We use the Kyoto Encyclopedia of Genes and Genomes database to download three cancer pathways [15], which are breast cancer (hsa05224), renal cell carcinoma (hsa05211), and non-small-cell lung cancer (hsa05223). In the following analysis, the cancer gene expression datasets from a specific organ are pooled to form one dataset for that organ, resulting in three datasets, BRCA, KIRC_KIRP, and LUAD_LUSC.

For the pre-processing steps, since the downloaded datasets have been normalized by fragments per kilobase of transcript per million mapped reads, we replace 0 by 1 and add 1 to all the non-zero values before the log-transformation. Genes specific to each pathway are extracted and pooled to form the subset of genes for all three datasets. Finally, we have three datasets with 212 genes each before applying graphical models.

We assume the gene expressions of individuals are identically distributed with a multivariate normal distribution. Simular to the steps we use for simulation studies, we drop the initial 500 burnin samples and keep the following 500 MCMC samples. We use 99% quantile intervals in the MCMC samples for variable selection to get sparse networks. If 0 is not included in the quantile interval, we treated it as a discovery. Based on the results in our simulation studies, since both GR2D2 and the graphical horseshoe show comparably good performance in variable selection, we compare both methods in our real data analysis.

Results

The inferred graphs by GR2D2 of datasets BRCA, KIRC_KIRP and LUAD_LUSC are shown in Figures 3, 4, 5. We also provide the inferred graph by the graphical horseshoe of the LUAD_LUSC dataset in Supplementary Information Figures S1. With 99% quantile intervals, the graphical horseshoe method identifies 1704 edges for the LUAD_LUSC dataset, resulting in a densely connected graph. This densely connected graph indicate that the graphical horseshoe estimator fails to select the most relevant edges for the dataset, and it becomes hard for the users to identify useful information from the network structure. Similar results on datasets BRCA and KIRC_KIRP can be expected. On the other hand, the GR2D2 method identifies 94 edges for the BRCA dataset, 76 edges for the KIRC_KIRP dataset, and 108 edges for the LUAD_LUSC dataset, resulting in sparse graphs for all three datasets.

We further scrutinize the graphs inferred by GR2D2 and check whether important cancer pathways have been identified. At the center to the right of Figure 3, the upper part of Figure 4, and the bottom of Figure 5, we notice a community of genes formed by RAF1, MAPK1, MAP2K1, MAPK2K2, and MAPK3, which correspond to the genes in the MAPK signaling pathway. This pathway comprises several necessary signaling components that play

Figure 3: The inferred graph for the BRCA dataset by the GR2D2 estimator. The size of node is proportional to the degrees of edges. At the center to the right part and the top right part of this figure, four pathways can be identified, which are MAPK, Wnt, Notch and ESR1 signaling pathways. MAPK and Wnt pathways are commonly found in many cancers while Notch and ESR1 have special relevance to the breast cancer.

a role in tumorigenesis, and the alternation of it has been reported in human cancer as a result of abnormal activation of receptor tyrosine kinases [28, 35]. Besides, this pathway is considered as potential therapeutic targets for cancer treatment and several inhibitors targeting this pathway have been developed [28]. At the top right part of Figure 3 and the left part of Figure 5, there are two communities of genes, most of which come from the Wnt signaling pathway. This pathway regulates many cellular processes, like cell proliferation, differentiation and cell fate, and aberrant Wnt signaling pathway is a hallmark of many cancers, including breast cancer [25, 38] and non-small cell lung cancer [33].

We then check pathways and genes that seem only appear in specific cancer types. At the top right of Figure 3, besides genes from the Wnt siganling pathway, genes (NOTCH1/3/4, DLL4, FLT4) from the Notch signaling pathway and genes (ESR1, CCND1) from the ESR1 signaling pathway are identified. Activated Notch signaling pathway and upregulation of tumor-promoting Notch target genes have been reported in human breast cancer and this pathway has been implicated in both the development and progression of breast cancer [1, 27, 34]. The ESR1 ESR1 signaling pathway has been found to be responsible for the luminal subtype of breast cancer and the mutation of the gene ESR1 has been reported in approximately 70% of breast cancer patients [14]. The ESR1 gene is also treated as a biomarker in breast cancer [5]. At the bottom of Figure 4, genes (VEGFA, TGFA, SLC2A1, TGCB1, and TGFB3) from the HIF-1 signaling pathway are identified. The activation of this pathway has special relevance to the kidney disease [12, 23] and we cannot find it on

Figure 4: The inferred graph for the KIRC_KIRP dataset by the GR2D2 estimator. The size of node is proportional to the degrees of edges. At the upper part and the lower part of the figure, genes from MAPK and HIF-1 signaling pathways can be found. HIF-1 pathway has special relevance to kidney disease while the MAPK pathway is related to many human cancers.

the other two graphs. At the left bottom of Figure 5, genes (JAK3, STAT5A and STAT5B) from the JAK-STAT signaling pathway are identified. This pathway has been found to be related to the progression of non-small cell lung cancer [44]. We further notice that the gene KIT at the center to the left part of Figure 5 is also identified. The mutation of KIT has special relevance to the lung cancer [21, 32]. Based on above findings, we can conclude that the GR2D2 estimator identifies important cancer pathways and also cancer-specific pathways for each cancer type.

5 Discussion

In high-dimensional data analysis, how to estimate the precision matrix is always an attractive challenge under multivariate Gaussian assumption. In this work, we proposed the GR2D2 estimator with easy implementation by a full Gibbs sampler. By using a prior with high density near the origin, a combination of an exponential distributed parameter and a Dirichlet distributed parameter on each dimension, the GR2D2 hierarchical model provides estimates close to the true distribution in Kullback-Leibler divergence and with the smallest bias for nonzero elements among current published methods. Simulation studies also confirm that the GR2D2 outperforms alternative methods like the graphical horseshoe and the graphical lasso in different settings. Furthermore, real data analysis shows that the GR2D2 estimator can capture important cancer-specific genetic interactions and have the ability to provide

Figure 5: The inferred graph for the LUAD_LUSC dataset by the GR2D2 estimator. The size of node is proportional to the degrees of edges. At the center to the left part, lower left part and lower right part of the figure, Wnt, JAK-STAT and MAPK signaling pathways can be identified. MAPK and Wnt pathways are commonly found in many cancers while JAK-STAT has been found to be related to the progression of non-small cell lung cancer. The KIT gene at the left part of the figure is also identified, which shows special relevance to lung cancer.

insights for future experiment design. Therefore, we may conclude that the GR2D2 could provide the best precision matrix estimation in sparse and high-dimensional settings, and infer meaningful genetic relationships.

6   Data availability

Data used in this study were downloaded from public databases that have been described in the main text.

7   Supplementary information and code

Supplementary information are available online with detailed derivation of GR2D2. The source code is available at https://github.com/RavenGan/GR2D2.

References

[1] Hamed Al-Hussaini, Deepa Subramanyam, Michael Reedijk, and Srikala S Sridhar. Notch signaling pathway as a therapeutic target in breast cancernotch in breast cancer. *Molecular cancer therapeutics*, 10(1):9–15, 2011.

[2] Albert-László Barabási and Réka Albert. Emergence of scaling in random networks. *science*, 286(5439):509–512, 1999.

[3] Albert-Laszlo Barabasi and Zoltan N Oltvai. Network biology: understanding the cell's functional organization. *Nature reviews genetics*, 5(2):101–113, 2004.

[4] Anirban Bhattacharya, Debdeep Pati, Natesh S Pillai, and David B Dunson. Dirichlet–laplace priors for optimal shrinkage. *Journal of the American Statistical Association*, 110(512):1479–1490, 2015.

[5] Marcela Carausu, François-Clément Bidard, Celine Callens, Samia Melaabi, Emmanuelle Jeannot, Jean-Yves Pierga, and Luc Cabel. Esr1 mutations: a new biomarker in breast cancer. *Expert Review of Molecular Diagnostics*, 19(7):599–611, 2019.

[6] Aaron Clauset, Cosma Rohilla Shalizi, and Mark EJ Newman. Power-law distributions in empirical data. *SIAM review*, 51(4):661–703, 2009.

[7] Adrian Dobra, Chris Hans, Beatrix Jones, Joseph R Nevins, Guang Yao, and Mike West. Sparse graphical models for exploring gene expression data. *Journal of Multivariate Analysis*, 90(1):196–212, 2004.

[8] Jianqing Fan and Runze Li. Variable selection via nonconcave penalized likelihood and its oracle properties. *Journal of the American statistical Association*, 96(456):1348–1360, 2001.

[9] Jerome Friedman, Trevor Hastie, and Robert Tibshirani. Sparse inverse covariance estimation with the graphical lasso. *Biostatistics*, 9(3):432–441, 2008.

[10] Jerome Friedman, Trevor Hastie, and Robert Tibshirani. Applications of the lasso and grouped lasso to the estimation of sparse graphical models. Technical report, Technical report, Stanford University, 2010.

[11] Tyler Grimes and Somnath Datta. Seqnet: An r package for generating gene-gene networks and simulating rna-seq data. *Journal of statistical software*, 98(12), 2021.

[12] VH Haase. The vhl/hif oxygen-sensing pathway and its relevance to kidney disease. *Kidney international*, 69(8):1302–1307, 2006.

[13] Trey Ideker and Nevan J Krogan. Differential network biology. *Molecular systems biology*, 8(1):565, 2012.

[14] Rinath Jeselsohn, Gilles Buchwalter, Carmine De Angelis, Myles Brown, and Rachel Schiff. Esr1 mutations—a mechanism for acquired endocrine resistance in breast cancer. *Nature reviews Clinical oncology*, 12(10):573–583, 2015.

[15] Minoru Kanehisa and Susumu Goto. Kegg: kyoto encyclopedia of genes and genomes. *Nucleic acids research*, 28(1):27–30, 2000.

[16] Nicole Krämer, Juliane Schäfer, and Anne-Laure Boulesteix. Regularized estimation of large-scale gene association networks using graphical gaussian models. *BMC bioinformatics*, 10(1):1–24, 2009.

[17] Jan Krumsiek, Karsten Suhre, Thomas Illig, Jerzy Adamski, and Fabian J Theis. Gaussian graphical modeling reconstructs pathway reactions from high-throughput metabolomics data. *BMC systems biology*, 5(1):1–16, 2011.

[18] Gwenaël GR Leday, Mathisca CM De Gunst, Gino B Kpogbezan, Aad W Van der Vaart, Wessel N Van Wieringen, and Mark A Van De Wiel. Gene network reconstruction using global-local shrinkage priors. *The annals of applied statistics*, 11(1):41, 2017.

[19] Hongzhe Li and Jiang Gui. Gradient directed regularization for sparse gaussian concentration graphs, with applications to inference of genetic networks. *Biostatistics*, 7(2):302–317, 2006.

[20] Yunfan Li, Bruce A Craig, and Anindya Bhadra. The graphical horseshoe estimator for inverse covariance matrices. *Journal of Computational and Graphical Statistics*, 28(3):747–757, 2019.

[21] Hong-Yang Lu, Gu Zhang, Qiao-Yuan Cheng, Bo Chen, Ju-Fen Cai, Xiao-Jia Wang, Yi-Ping Zhang, Zeng Wang, Zhen-Yi Lu, Fa-Jun Xie, et al. Expression and mutation of the c-kit gene and correlation with prognosis of small cell lung cancer. *Oncology Letters*, 4(1):89–93, 2012.

[22] Enes Makalic and Daniel F Schmidt. A simple sampler for the horseshoe estimator. *IEEE Signal Processing Letters*, 23(1):179–182, 2015.

[23] Patrick Maxwell. Hif-1: an oxygen response system with special relevance to the kidney. *Journal of the American Society of Nephrology*, 14(11):2712–2722, 2003.

[24] Vasyl Pihur, Somnath Datta, and Susmita Datta. Reconstruction of genetic association networks from microarray data: a partial least squares approach. *Bioinformatics*, 24(4):561–568, 2008.

[25] SÖ-G Pohl, N Brook, Mark Agostino, Frank Arfuso, Alan Prem Kumar, and Arunasalam Dharmarajan. Wnt signaling in triple-negative breast cancer. *Oncogenesis*, 6(4):e310–e310, 2017.

[26] Mohsen Pourahmadi. Covariance estimation: The glm and regularization perspectives. *Statistical Science*, pages 369–387, 2011.

[27] Michael Reedijk. Notch signaling and breast cancer. *Notch Signaling in Embryology and Cancer*, pages 241–257, 2012.

[28] Libero Santarpia, Scott M Lippman, and Adel K El-Naggar. Targeting the mapk–ras–raf signaling pathway in cancer therapy. *Expert opinion on therapeutic targets*, 16(1):103–119, 2012.

[29] Juliane Schäfer and Korbinian Strimmer. An empirical bayes approach to inferring large-scale gene association networks. *Bioinformatics*, 21(6):754–764, 2005.

[30] Juliane Schäfer and Korbinian Strimmer. Learning large-scale graphical gaussian models from genomic data. In *AIP Conference Proceedings*, volume 776, pages 263–276. American Institute of Physics, 2005.

[31] Juliane Schäfer and Korbinian Strimmer. A shrinkage approach to large-scale covariance matrix estimation and implications for functional genomics. *Statistical applications in genetics and molecular biology*, 4(1), 2005.

[32] Alice T Shaw and Jeffrey A Engelman. Alk in lung cancer: past, present, and future. *Journal of clinical oncology*, 31(8):1105, 2013.

[33] David J Stewart. Wnt signaling pathway in non–small cell lung cancer. *JNCI: Journal of the National Cancer Institute*, 106(1), 2014.

[34] Spyros Stylianou, Rob B Clarke, and Keith Brennan. Aberrant activation of notch signaling in human breast cancer. *Cancer research*, 66(3):1517–1525, 2006.

[35] Yu Sun, Wen-Zhou Liu, Tao Liu, Xu Feng, Nuo Yang, and Hua-Fu Zhou. Signaling pathway of mapk/erk in cell proliferation, differentiation, migration, senescence and apoptosis. *Journal of Receptors and Signal Transduction*, 35(6):600–604, 2015.

[36] Hao Wang et al. Bayesian graphical lasso models and efficient posterior computation. *Bayesian Analysis*, 7(4):867–886, 2012.

[37] Lin Wei, Zhilin Jin, Shengjie Yang, Yanxun Xu, Yitan Zhu, and Yuan Ji. Tcga-assembler 2: software pipeline for retrieval and processing of tcga/cptac data. *Bioinformatics*, 34(9):1615–1617, 2018.

[38] Xiufang Xu, Miaofeng Zhang, Faying Xu, and Shaojie Jiang. Wnt signaling in breast cancer: biological mechanisms, challenges and opportunities. *Molecular cancer*, 19(1):1–35, 2020.

[39] Huangdi Yi, Qingzhao Zhang, Cunjie Lin, and Shuangge Ma. Information-incorporated gaussian graphical model for gene expression data. *Biometrics*, 2021.

[40] Jianxin Yin and Hongzhe Li. A sparse conditional gaussian graphical model for analysis of genetical genomics data. *The annals of applied statistics*, 5(4):2630, 2011.

[41] Ming Yuan and Yi Lin. Model selection and estimation in the gaussian graphical model. *Biometrika*, 94(1):19–35, 2007.

[42] Yan Dora Zhang, Brian P Naughton, Howard D Bondell, and Brian J Reich. Bayesian regression using a prior on the model fit: The r2-d2 shrinkage prior. *Journal of the American Statistical Association*, pages 1–13, 2020.

[43] Zhen Zhang, Jiajun Bu, Martin Ester, Jianfeng Zhang, Chengwei Yao, Zhi Yu, and Can Wang. Hierarchical graph pooling with structure learning. *arXiv preprint arXiv:1911.05954*, 2019.

[44] Dengyan Zhu, Yang Yu, Wei Wang, Kai Wu, Donglei Liu, Yang Yang, Chunyang Zhang, Yu Qi, and Song Zhao. Long noncoding rna part1 promotes progression of non-small cell lung cancer cells via jak-stat signaling pathway. *Cancer medicine*, 8(13):6064–6081, 2019.

Supplementary Information for
The GR2D2 Estimator for the Precision Matrices

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1. A Data-Augmented Block Gibbs Sampler

Posterior samples under the graphical R2D2 hierarchical model are drawn by an augmented block Gibbs sampler, utilizing the scheme proposed by [3] for linear regression. In each iteration, each column and row of Ω and $\Lambda = (\phi_{ij}\psi_{ij})$ are partitioned from a $p \times p$ matrix of parameters and updated in a block. Then the global shrinkage parameter ω and its auxiliary variable ξ are updated.

The following part derives the posterior distribution of the precision matrix. Given data $Y_{n \times p}$ and the shrinkage parameters, we provide the step-by-step derivation in the following. Define the following matrices

$$\Omega = \begin{bmatrix} \Omega_{(-p)(-p)} & \boldsymbol{\omega}_{(-p)p} \\ \boldsymbol{\omega}_{(-p)p}^T & \omega_{pp} \end{bmatrix}, \quad S = \begin{bmatrix} S_{(-p)(-p)} & \boldsymbol{s}_{(-p)p} \\ \boldsymbol{s}_{(-p)p}^T & s_{pp} \end{bmatrix},$$

$$\Lambda = \begin{bmatrix} \Lambda_{(-p)(-p)} & \boldsymbol{\lambda}_{(-p)p} \\ \boldsymbol{\lambda}_{(-p)p}^T & \lambda_{pp} \end{bmatrix},$$

where $(-p)$ denotes the set of all indices except for p, and $\Lambda_{(-p)(-p)}$ and $\boldsymbol{\lambda}_{(-p)p}$ have entries $\phi_{ij}\psi_{ij}$. Diagonal elements of $\Lambda_{(-p)(-p)}$ are set to be 1. The posterior distribution of Ω is

$$p(\Omega|\Lambda, Y, \omega) \propto p(S|\Omega, \Lambda, \omega)p(\Omega|\Lambda, \omega),$$

where $S = Y^TY \sim W_p(\Omega^{-1}, n)$ has the Wishart density function

$$f(S) = \frac{1}{2^{np/2}|\Omega^{-1}|^{n/2}\Gamma_p(n/2)}|S|^{(n-p-1)/2}exp(-\frac{1}{2}tr(\Omega S)).$$

Then, the posterior distribution of Ω can be written as

$$p(\Omega|\Lambda, Y, \omega) \propto |\Omega|^{n/2}exp(-\frac{1}{2}tr(\Omega S))\prod_{i<j}Normal(\omega_{ij}|\phi_{ij}, \psi_{ij}, \omega)$$

$$\propto |\Omega|^{n/2}exp(-\frac{1}{2}tr(\Omega S))\prod_{i<j}exp(-\frac{\omega_{ij}^2}{\psi_{ij}\phi_{ij}\omega}).$$

By block determinant, we have

$$|\Omega| = |\Omega_{(-p)(-p)}|(\omega_{pp} - \boldsymbol{\omega}_{(-p)p}^T \Omega_{(-p)(-p)}^{-1} \boldsymbol{\omega}_{(-p)p}).$$

Therefore, we have

$$p(\Omega|\Lambda, Y, \omega) \propto (|\Omega_{(-p)(-p)}|(\omega_{pp} - \boldsymbol{\omega}_{(-p)p}^T \Omega_{(-p)(-p)}^{-1} \boldsymbol{\omega}_{(-p)p}))^{n/2}$$

$$exp(-\frac{1}{2}tr(\Omega S)) \prod_{i<j} exp(-\frac{\omega_{ij}^2}{\psi_{ij}\phi_{ij}\omega}).$$

Then we consider $tr(S\Omega)$,

$$S\Omega = \begin{bmatrix} M_{1,1} & M_{1,2} \\ M_{2,1} & M_{2,2} . \end{bmatrix}$$

where

- $M_{1,1} = S_{(-p)(-p)}\Omega_{(-p)(-p)} + \boldsymbol{s_{(-p)p}}\boldsymbol{\omega_{(-p)p}^T}$

- $M_{1,2} = S_{(-p)(-p)}\boldsymbol{\omega_{(-p)p}} + \omega_{pp}\boldsymbol{s_{(-p)p}}$

- $M_{2,1} = s_{pp}\boldsymbol{\omega_{(-p)p}^T} + \boldsymbol{s_{(-p)p}^T}\Omega_{(-p)(-p)}$

- $M_{2,2} = \boldsymbol{s_{(-p)p}^T}\boldsymbol{\omega_{(-p)p}} + s_{pp}\omega_{pp}$

Note that

$$tr(\boldsymbol{s_{(-p)p}}\boldsymbol{\omega_{(-p)p}^T}) = \sum_{i=1}^{p} s_{(-p)i}\omega_{(-p)i} = \boldsymbol{s_{(-p)i}^T}\boldsymbol{\omega_{(-p)p}}.$$

Hence, we obtain

$$tr(S\Omega) = 2\boldsymbol{s_{(-p)p}^T}\boldsymbol{\omega_{(-p)p}} + s_{pp}\omega_{pp}$$

If we only consider the last column

$$p(\boldsymbol{\omega_{(-p)p}}, \omega_{pp}|Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$

$$\propto (\omega_{pp} - \boldsymbol{\omega_{(-p)p}^T}\Omega_{(-p)(-p)}^{-1}\boldsymbol{\omega_{(-p)p}})^{n/2}$$

$$exp(-\frac{1}{2}(2\boldsymbol{s_{(-p)p}^T}\boldsymbol{\omega_{(-p)p}} + s_{pp}\omega_{pp}))$$

$$\prod_{i=1}^{p-1} exp(-\frac{\omega_{ip}^2}{\phi_{ip}\psi_{ip}\omega}).$$

Note that

$$\prod_{i=1}^{p-1} exp(-\frac{\omega_{ip}^2}{\phi_{ip}\psi_{ip}\omega}) = exp(-\sum_{i=1}^{p-1} \omega_{ip}(\phi_{ij}\psi_{ij}\omega)^{-1}\omega_{ip})$$

$$= exp(-\boldsymbol{\omega_{(-p)p}^T}(\Lambda^*\omega)^{-1}\boldsymbol{\omega_{(-p)p}}),$$

where $\Lambda^* = diag(\boldsymbol{\lambda_{(-p)p}})$.
Therefore, we have

$$p(\boldsymbol{\omega}_{(-p)p}, \omega_{pp}|Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$
$$\propto (\omega_{pp} - \boldsymbol{\omega}^T_{(-p)p}\Omega^{-1}_{(-p)(-p)}\boldsymbol{\omega}_{(-p)p})^{n/2}$$
$$exp(-\frac{1}{2}(2\boldsymbol{s}^T_{(-p)p}\boldsymbol{\omega}_{(-p)p} + s_{pp}\omega_{pp}))$$
$$exp(-\boldsymbol{\omega}^T_{(-p)p}(\Lambda^*\omega)^{-1}\boldsymbol{\omega}_{(-p)p}).$$

Let

$$\boldsymbol{\beta} = \boldsymbol{\omega}_{(-p)p},$$
$$\gamma = \omega_{pp} - \boldsymbol{\omega}^T_{(-p)p}\Omega^{-1}_{(-p)(-p)}\boldsymbol{\omega}_{(-p)p} = \omega_{pp} - \boldsymbol{\beta}^T\Omega^{-1}_{(-p)(-p)}\boldsymbol{\beta},$$
$$\omega_{pp} = \gamma + \boldsymbol{\beta}^T\Omega^{-1}_{(-p)(-p)}\boldsymbol{\beta}$$

The Jacobian of this transformation is a constant, and the full conditional of β and γ is

$$p(\boldsymbol{\beta}, \gamma|Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$
$$\propto \gamma^{n/2}$$
$$exp(-\frac{1}{2}(s_{pp}\gamma + s_{pp}\boldsymbol{\beta}^T\Omega^{-1}_{(-p)(-p)}\boldsymbol{\beta} + 2\boldsymbol{S}^T_{(-p)p}\boldsymbol{\beta}))$$
$$exp(-\boldsymbol{\beta}^T(\Lambda^*\omega)^{-1}\boldsymbol{\beta}).$$

Then, we obtain

$$p(\boldsymbol{\beta}, \gamma|Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$
$$\propto \gamma^{n/2}exp(-\frac{1}{2}s_{pp}\gamma)$$
$$exp(-\frac{1}{2}(\boldsymbol{\beta}^T(2(\Lambda^*\omega)^{-1} + s_{pp}\Omega_{(-p)(-p)})\boldsymbol{\beta} + 2\boldsymbol{s}^T_{(-p)p}\boldsymbol{\beta})).$$

Let $D = (2(\Lambda^*\omega)^{-1} + s_{pp}\Omega_{(-p)(-p)})^{-1}$. Since Ω is positive definite by assumption, we have $\Omega_{(-p)(-p)}$ is symmetric. Hence D is symmetric and $D^TD_{-1} = \boldsymbol{I}$. Note that $\boldsymbol{s}^T_{(-p)p}\boldsymbol{\beta} = \boldsymbol{\beta}^T\boldsymbol{s}_{(-p)p} = \boldsymbol{\beta}^TD^{-1}D\boldsymbol{s}_{(-p)p}$. Then, we consider

$$exp(-\frac{1}{2}(\boldsymbol{\beta}^T(2(\Lambda^*\omega)^{-1} + s_{pp}\Omega_{(-p)(-p)})\boldsymbol{\beta} + 2\boldsymbol{s}^T_{(-p)p}\boldsymbol{\beta}))$$
$$= exp(-\frac{1}{2}(\boldsymbol{\beta}^TD\boldsymbol{\beta} + 2\boldsymbol{s}^T_{(-p)p}\boldsymbol{\beta}))$$
$$= exp(-\frac{1}{2}(\boldsymbol{\beta}^TD\boldsymbol{\beta} + \boldsymbol{s}^T_{(-p)p}\boldsymbol{\beta} + \boldsymbol{s}^T_{(-p)p}D^TD^{-1}\boldsymbol{\beta}))$$
$$\propto exp(-\frac{1}{2}(\boldsymbol{\beta} + D\boldsymbol{s}_{(-p)p})^TD^{-1}(\boldsymbol{\beta} + D\boldsymbol{s}_{(-p)p}))$$

Therefore, we have

$$p(\boldsymbol{\beta}, \gamma | Y, \Lambda, \omega, \Omega_{(-p)(-p)})$$
$$\propto \gamma^{n/2} exp(-\frac{1}{2} s_{pp} \gamma)$$
$$exp(-\frac{1}{2}(\boldsymbol{\beta} + D\boldsymbol{s}_{(-p)p})^T D^{-1}(\boldsymbol{\beta} + D\boldsymbol{s}_{(-p)p})),$$

which can be summarized as

$$p(\boldsymbol{\beta}, \gamma | Y, \Lambda, \omega, \Omega_{(-p)(-p)}) \propto Gamma(\frac{n}{2} + 1, \frac{s_{pp}}{2})$$
$$Normal(-D\boldsymbol{s}_{(-p)p}, D),$$

where $D = (2(\Lambda^* \omega)^{-1} + s_{pp}\Omega_{(-p)(-p)})^{-1}$, $\Lambda^* = diag(\boldsymbol{\lambda}_{(-p)p})$, and $\lambda_{ij} = \phi_{ij}\psi_{ij}$.

Above part is about how to sample Ω one column each time. In the following part, we provide standard derivation of the posterior distribution of ψ_{ij}, ω, ξ and $\boldsymbol{\phi}$.

We first consider $\psi_{ij} \sim Exp(\frac{1}{2})$ with density function $p(\psi_{ij}) = \frac{1}{2}exp(-\frac{1}{2}\psi_{ij})$. The posterior distribution is

$$p(\psi_{ij}|\omega_{ij}, \phi_{ij}, \omega) \propto p(\omega_{ij}|\psi_{ij}, \phi_{ij}, \omega)p(\psi_{ij})$$
$$\propto \frac{1}{\sqrt{\psi_{ij}}}exp(-\frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}\omega})exp(-\frac{1}{2}\psi_{ij})$$

Let $\psi = \psi_{ij}^{-1}$, and the Jacobian term is $|J| = |\frac{\partial \psi}{\partial \psi_{ij}}| = |\psi_{ij}^{-2}|$. With this transformation, we have

$$p(\psi|\omega_{ij}, \phi_{ij}, \omega) \propto \frac{1}{\psi_{ij}^{3/2}}exp(-\frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}\omega})exp(-\frac{1}{2}\psi_{ij})$$
$$= \psi^{3/2}exp(-\frac{\omega_{ij}^2}{\phi_{ij}\omega}\psi)exp(-\frac{1}{2\psi})$$
$$= \psi^{3/2}exp(-\frac{2\omega_{ij}^2\psi^2 + \phi_{ij}\omega}{2\psi\phi_{ij}\omega})$$
$$= \psi^{3/2}exp(-\frac{\psi^2 + \frac{\phi_{ij}\omega}{2\omega_{ij}^2}}{2\psi\frac{\phi_{ij}\omega}{2\omega_{ij}^2}})$$
$$\propto \psi^{3/2}exp(-\frac{(\psi - \mu)^2}{2\mu^2\psi})$$

where $\mu = \sqrt{\frac{\phi_{ij}\omega}{2\omega_{ij}^2}} = \sqrt{\phi_{ij}\omega/2}/|\omega_{ij}|$. Hence, we can conclude that

$$p(\psi|\omega_{ij}, \phi_{ij}, \omega) \sim IG(\mu = \sqrt{\phi_{ij}\omega/2}/|\omega_{ij}|, \lambda = 1).$$

When we sample ϕ_{ij}, we first sample ϕ_{ij}^{-1} based on above formula and take the reciprocal.

Next we consider $\omega|\xi \sim Gamma(shape = a, rate = \xi)$ with density function $p(\omega|\xi) = \frac{\xi^a}{\Gamma(a)}\omega^{a-1}exp(-\xi\omega)$. Denote $Z \sim giG(\chi, \rho, \lambda_0)$, the generalized inverse Gaussian distribution, if $p(z) \propto z^{\lambda_0-1}exp(-(\rho z + \chi/z)/2)$. The posterior distribution of ω is

$$p(\omega|\Omega, \Lambda, \xi) \propto p(\Omega|\Lambda, \xi, \omega)p(\omega|\xi)$$

$$\propto (\prod_{i<i} \frac{1}{\sqrt{\omega}}exp(-\frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}\omega}))\omega^{a-1}exp(-\xi\omega).$$

Since we only consider the upper diagonal entries of the precision matrix, there are $p(p-1)/2$ entries in total. We have

$$p(\omega|\Omega, \Lambda, \xi) \propto \omega^{-\frac{p(p-1)}{4}}exp(-\sum_{i<j} \frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}\omega})\omega^{a-1}exp(-\xi\omega)$$

$$\propto \omega^{a-\frac{p(p-1)}{4}-1}exp(-\frac{1}{2}(2\xi\omega + \frac{1}{\omega}2\sum_{i<j} \frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}})).$$

In summary, we have

$$p(\omega|\Omega, \Lambda, \xi) \sim giG(\chi = 2\sum_{i<j} \frac{\omega_{ij}^2}{\phi_{ij}\psi_{ij}}, \rho = 2\xi, \lambda_0 = a - \frac{p(p-1)}{4}).$$

Next, we derive the posterior distribution of $\xi \sim Gamma(b, 1)$ with probability density function $p(\xi) = \frac{1^b}{\Gamma(b)}\xi^{b-1}exp(-\xi)$.

$$p(\xi|\Omega, \omega, \Lambda) \propto p(\omega|\xi)p(\xi)$$

$$\propto \frac{\xi^a}{\Gamma(a)}\omega^{a-1}exp(-\xi\omega)\xi^{b-1}exp(-\xi)$$

$$\propto \xi^{a+b-1}exp(-(\omega + 1)\xi).$$

In summary, we have

$$p(\xi|\omega) \sim Gamma(shape = a + b, rate = \omega + 1).$$

In order to sample $\phi \sim Dir(\alpha_{pi}, \alpha_{pi}, \ldots, \alpha_{pi})$, where there are $p(p-1)/2\alpha'_{pi}s$ in total, we resort to the work of [1] and the sampling procedures for ϕ are in the following. We first

integrate out ω. The joint distribution of $\phi_i | \boldsymbol{\theta}, i = 1, 2, \ldots, \frac{p(p-1)}{2}$ has the form

$$p(\phi_1, \ldots, \phi_{\frac{p(p-1)}{2}} | \boldsymbol{\theta})$$

$$\propto \prod_{j=1}^{\frac{p(p-1)}{2}} \phi_j^{\alpha_\pi - 1} \int_0^\infty \omega^{a-1} exp(-\xi\omega)$$

$$\prod_{i<j} \frac{1}{\sqrt{\phi_{ij}\omega}} exp(-\frac{\omega_{ij}^2}{\psi_{ij}\phi_{ij}\omega}) dw \tag{1}$$

$$\propto (\prod_{j=1}^{\frac{p(p-1)}{2}} \phi_j^{\alpha_\pi - 3/2})$$

$$\int_0^\infty \omega^{a-1} \omega^{-\frac{p(p-1)}{4}} exp(-\xi\omega) exp(-\sum_{j=1}^{\frac{p(p-1)}{2}} \frac{\omega_j^2}{\phi_j\psi_j\omega}) dw$$

We next use the results from the theory of normalized random measures [2]. Suppose $T_1, \ldots, T_{\frac{p(p-1)}{2}}$ are independent random variables with T_j having a density f_j on $(0, +\infty)$. Let $\phi_j = T_j/T$ with $T = \sum_{j=1}^{p(p-1)/2} T_j$. Then, the joint density f of $(\phi_1, \ldots, \phi_{\frac{p(p-1)}{2}-1})$ supported on the simplex S^{n-1} has the form

$$f(\phi_1, \ldots, \phi_{\frac{p(p-1)}{2}-1}) = \int_{t=1}^\infty t^{\frac{p(p-1)}{2}-1} \prod_{j=1}^{\frac{p(p-1)}{2}} f_j(\phi_j t) dt,$$

where $\phi_{\frac{p(p-1)}{2}} = 1 - \sum_{j=1}^{\frac{p(p-1)}{2}-1} \phi_j$. Set

$$f_j(x) \propto \frac{1}{x^\sigma} exp(-\frac{\omega_j^2}{\psi_j}\frac{1}{x} - \frac{1}{2}2\xi x)$$

$$= x^{-\sigma} exp(-\frac{1}{2}(2\xi x + \frac{2\omega_j^2}{\psi_j}\frac{1}{x})).$$

We get

$$f(\boldsymbol{\phi}) \propto \int_0^\infty t^{\frac{p(p-1)}{2}-1} \prod_{j=1}^{\frac{p(p-1)}{2}} (\frac{1}{\phi_j t})^\sigma$$

$$exp(-\frac{\omega_j^2}{\psi_j}\frac{1}{\phi_j t} - \frac{1}{2}2\xi\phi_j t)dt$$

$$\propto \prod_{j=1}^{\frac{p(p-1)}{2}} \frac{1}{\phi_j^\sigma} \int_{t=0}^\infty t^{\frac{p(p-1)}{2}-1}$$

$$\prod_{j=1}^{\frac{p(p-1)}{2}} (\frac{1}{t^\sigma}) exp(- \sum_{j=1}^{\frac{p(p-1)}{2}} \frac{\omega_j^2}{\psi_j\phi_j t} - \frac{1}{2}2\xi t)dt \tag{2}$$

$$\propto \prod_{j=1}^{\frac{p(p-1)}{2}} \frac{1}{\phi_j^\sigma} \int_{t=0}^\infty t^{\frac{p(p-1)}{2}-1-\frac{p(p-1)}{2}\sigma}$$

$$exp(-\xi t)exp(- \sum_{j=1}^{\frac{p(p-1)}{2}} \frac{\omega_j^2}{\psi_j\phi_j t})dt$$

Then, we equate the results from functions 1 and 2 and obtain

$$\alpha_\pi - \frac{3}{2} = -\sigma$$

$$\frac{p(p-1)}{2} - 1 - \frac{p(p-1)}{2}\sigma = \frac{p(p-1)}{2}\alpha_\pi - 1 - \frac{1}{2}\frac{p(p-1)}{2}.$$

Hence, $\sigma = \frac{3}{2} - \alpha_\pi$ and we have

$$f_j(x) \propto x^{\alpha_\pi - \frac{3}{2}} exp(-\frac{1}{2}(2\xi x + \frac{2\omega_j^2}{\psi_j}\frac{1}{x})).$$

In summary, we have

$$T_j \sim giG(\chi = 2\omega_j^2/\psi_j, \rho = 2\xi, \lambda_0 = \alpha_\pi - \frac{1}{2}).$$

2. Formulae of precision, recall, FPR, accuracy and F score

Denote True Positive, False Positive, False Negative and True Negative as TP, FP, FN, and TN respectively. The formulae are

$$\text{Precision} = \frac{\text{TP}}{\text{TP} + \text{FP}}, \quad \text{Recall} = \frac{\text{TP}}{\text{TP} + \text{FN}}, \quad \text{FPR} = \frac{\text{FP}}{\text{FP} + \text{TN}}$$

$$\text{Accuracy} = \frac{\text{TP} + \text{TN}}{\text{TP} + \text{FP} + \text{TN} + \text{FN}}$$

$$\text{F-score} = 2 \times \frac{\text{Precision} \times \text{Recall}}{\text{Precision} + \text{Recall}}.$$

3. Supplementary figures

Figure S1: The inferred graph for the LUAD_LUSC dataset by the graphical horseshoe estimator. The size of node is proportional to the degrees of edges.

4. Supplementary tables

(P100, n50)	Hubs			Cliques positive			Cliques negative		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.2188**	3.4443	82.4434	5.0073	**4.869**	86.3335	6.9366	**6.7698**	86.7184
(s.d.)	0.1112	0.1385	0.5057	0.1201	0.2189	0.667	0.0632	0.2046	0.5267
Stein's loss	**5.8591**	6.7082	484.8487	**53.1149**	58.6612	793.8653	**23.9461**	31.6169	679.9838
(s.d.)	0.3417	0.4307	4.4029	2.6451	4.7103	16.9524	1.1757	1.9982	10.2098
Precision	1	0.9981	0.0218	1	0.9969	0.0197	1	0.9989	0.0205
(s.d.)	0	0.0059	0.0008	0	0.0074	0.0004	0	0.0044	0.0003
Recall	0.5556	0.56	**0.9563**	0.625	0.7967	**0.9702**	0.625	0.7142	**0.9962**
(s.d.)	0	0.0075	0.0267	0	0.0222	0.0157	0	0.0191	0.0068
FPR	**0**	2.04E-05	0.7655	**0**	4.07E-05	0.7650	**0**	1.36E-05	0.7667
(s.d.)	0	6.21E-05	0.0056	0	9.84E-05	0.0053	0	5.16E-05	0.0056
Accuracy	0.992	**0.9921**	0.2471	0.994	**0.9967**	0.2465	0.994	**0.9954**	0.2454
(s.d.)	0	1.30E-04	0.0055	0	3.59E-04	0.0052	0	3.10E-04	0.0055
F score	0.7143	**7.17E-01**	0.0427	0.7692	**0.8854**	0.0387	0.7692	**0.8327**	0.0402
(s.d.)	0	0.0058	0.0015	0	0.0137	0.0007	0	0.0132	0.0005

Table S1. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for network structures Hubs, Cliques positive and Cliques negative over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 100 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P150, n50)	Hubs			Cliques positive			Cliques negative		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.6462**	4.1748	140.6986	**5.2716**	5.4002	146.4574	7.1302	**7.0766**	146.1478
(s.d.)	0.1699	0.2507	0.3868	0.1144	0.1894	0.6187	0.0792	0.1833	0.628
Stein's loss	**6.9167**	8.7052	1154.8461	**54.249**	61.8153	1631.2601	**24.4415**	33.3348	1468.3316
(s.d.)	0.4852	0.7139	4.7164	1.8619	3.4329	19.6544	1.2492	1.9639	11.2402
Precision	**1**	0.9965	0.0150	**1**	0.997	0.0141	**1**	0.9976	0.0145
(s.d.)	0	0.0068	0.0004	0	0.0058	0.0002	0	0.0048	0.0002
Recall	0.6522	0.6545	**0.9490**	0.7143	0.8435	**0.9640**	0.7143	0.7911	**0.9911**
(s.d.)	0	0.0051	0.0202	0	0.0192	0.0118	0	0.0214	0.009
FPR	**0**	2.39E-05	0.6356	**0**	2.39E-05	0.6315	**0**	1.79E-05	0.6314
(s.d.)	0	4.68E-05	0.0022	0	4.67E-05	0.0029	0	3.65E-05	0.0041
Accuracy	**0.9964**	**0.9964**	0.3703	0.9973	**0.9985**	0.3740	0.9973	**0.998**	0.3743
(s.d.)	0	7.38E-05	0.0023	0	1.94E-04	0.0029	0	1.96E-04	0.0041
F score	0.7895	**0.7901**	0.0295	0.8333	**0.9137**	0.0277	0.8333	**0.8823**	0.0285
(s.d.)	0	0.0045	0.0008	0	0.012	0.0004	0	0.0131	0.0003

Table S2. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for network structures Hubs, Cliques positive and Cliques negative over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where $p = 150$ and $n = 50$. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P200, n50)	Hubs			Cliques positive			Cliques negative		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.9499**	4.9696	191.1832	**5.5071**	5.9812	197.9015	**7.3117**	7.6966	197.0806
(s.d.)	0.1986	0.297	0.6503	0.1106	0.2666	0.7279	0.0932	0.311	0.6971
Stein's loss	**8.0758**	11.6529	1922.5238	**55.1247**	64.9922	2528.4258	**26.321**	38.1823	2327.1573
(s.d.)	0.6396	1.055	7.8913	3.4851	4.4964	29.9884	1.2098	2.7868	13.0926
Precision	1	0.9948	0.0123	1	0.9968	0.0117	1	0.9945	0.0120
(s.d.)	0	0.0084	0.0003	0	0.0043	0.0002	0	0.0062	0.0002
Recall	0.7143	0.7169	**0.9521**	0.7692	0.8751	**0.9648**	0.7692	0.8277	**0.992**
(s.d.)	0	0.004	0.0156	0	0.0124	0.0106	0	0.0141	0.0066
FPR	**0**	2.69E-05	0.5357	**0**	1.85E-05	0.5311	**0**	3.02E-05	0.5315
(s.d.)	0	4.33E-05	0.0024	0	2.47E-05	0.0026	0	3.40E-05	0.0021
Accuracy	**0.998**	**0.998**	0.4677	0.9985	**0.9992**	0.4722	0.9985	**0.9989**	0.4719
(s.d.)	0	5.10E-05	0.0024	0	8.67E-05	0.0026	0	9.19E-05	0.0021
F score	**0.8333**	**0.8333**	0.0243	0.8696	**0.932**	0.0231	0.8696	**0.9034**	0.0237
(s.d.)	0	0.004	0.0005	0	0.0075	0.0003	0	0.0084	0.0003

Table S3. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for network structures Hubs, Cliques positive and Cliques negative over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 200 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P250, n50)	Hubs			Cliques positive			Cliques negative		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**4.2889**	5.973	236.344	**5.7492**	6.816	243.6115	**7.5132**	8.2696	242.1377
(s.d.)	0.1823	0.2517	0.657	0.1092	0.2221	0.6882	0.1297	0.3703	0.9657
Stein's loss	**9.1185**	15.6072	2745.0562	**54.7514**	69.4763	3444.198	**27.779**	41.837	3220.5177
(s.d.)	0.5655	1.038	8.2134	1.9759	3.7656	14.67	1.9426	3.6747	15.7963
Precision	**1**	0.9921	0.0108	**1**	0.9948	0.0104	**1**	0.9938	0.0106
(s.d.)	0	0.0082	0.0002	0	0.0049	0.0001	0	0.0052	0.0001
Recall	0.7576	0.759	**0.9518**	0.8065	0.8946	**0.9700**	0.8065	0.8576	**0.9889**
(s.d.)	0	0.0031	0.0168	0	0.0137	0.0097	0	0.0106	0.0084
FPR	**0**	3.22E-05	0.4617	**0**	2.36E-05	0.4575	**0**	2.68E-05	0.4583
(s.d.)	0	3.38E-05	0.0018	0	2.22E-05	0.0013	0	2.25E-05	0.0014
Accuracy	**0.9987**	**0.9987**	0.5405	0.999	**0.9995**	0.5446	0.999	**0.9993**	0.5439
(s.d.)	0	3.43E-05	0.0018	0	7.27E-05	0.0013	0	6.02E-05	0.0014
F score	**0.8621**	0.86	0.0213	0.8929	**0.942**	0.0207	0.8929	**0.9207**	0.0211
(s.d.)	0	0.0033	0.0004	0	0.0081	0.0002	0	0.0069	0.0002

Table S4. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for network structures Hubs, Cliques positive and Cliques negative over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where $p = 250$ and $n = 50$. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P100, n50)	Scale-free positive			Scale-free negative			Scale-free half		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.0680**	3.2880	82.4385	**3.0928**	3.3613	82.5090	**3.1336**	3.3995	82.6982
(s.d.)	0.1308	0.2082	0.5793	0.1814	0.2342	0.5100	0.1245	0.1798	0.4436
Stein's loss	**4.8533**	5.5368	480.9690	**4.9342**	5.7451	482.3797	**5.0948**	5.8694	483.5125
(s.d.)	0.3414	0.5583	4.8474	0.4550	0.5823	4.3023	0.3007	0.4279	3.7743
Precision	**1**	0.9981	0.0312	**1**	0.9961	0.0313	**1**	0.9967	0.0310
(s.d.)	0	0.0078	0.0012	0	0.0080	0.0012	0	0.0074	0.0010
Recall	0.3356	0.3358	**0.8826**	0.3356	0.3362	**0.8864**	0.3356	0.3360	**0.8856**
(s.d.)	0	0.0012	0.0258	0	0.0020	0.0244	0	0.0017	0.0230
FPR	**0**	2.0614E-05	0.7646	**0**	4.1229E-05	0.7641	**0**	3.4357E-05	0.7629
(s.d.)	0	8.2989E-05	0.0056	0	8.3867E-05	0.0051	0	7.8138E-05	0.0047
Accuracy	**0.9802**	**0.9802**	0.2530	**0.9802**	**0.9802**	0.2535	**0.9802**	**0.9802**	0.2544
(s.d.)	0	8.9955E-05	0.0055	0	9.6132E-05	0.0052	0	9.6132E-05	0.0046
F score	**0.5025**	**0.5025**	0.0602	0.5025	**0.5028**	0.0605	0.5025	**0.5026**	0.0599
(s.d.)	0	0.0017	0.0024	0	0.0024	0.0022	0	0.0022	0.0018

Table S5. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for scale-free network structures Positive, Negative and Half over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 100 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P150, n50)	Scale-free positive			Scale-free negative			Scale-free half		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.7772**	4.2308	140.6210	**3.8144**	4.3399	140.7319	**3.8134**	4.2667	140.8609
(s.d.)	0.1417	0.1987	0.4297	0.1485	0.2350	0.5925	0.1359	0.1944	0.6803
Stein's loss	**7.3703**	9.0517	1163.8198	**7.5436**	9.5356	1166.5813	**7.6187**	9.2930	1167.6915
(s.d.)	0.4423	0.6678	5.0521	0.4635	0.7542	6.3172	0.4388	0.7099	7.4508
Precision	**1**	0.9961	0.0241	**1**	0.9965	0.0240	**1**	0.9969	0.0241
(s.d.)	0	0.0070	0.0010	0	0.0068	0.0010	0	0.0066	0.0010
Recall	0.3348	0.3351	**0.8232**	0.3348	0.3350	**0.8287**	0.3348	0.3353	**0.8216**
(s.d.)	0	0.0011	0.0269	0	8.1505E-04	0.0288	0	0.0018	0.0322
FPR	**0**	2.7208E-05	0.6350	**0**	2.4185E-05	0.6345	**0**	2.1162E-05	0.6340
(s.d.)	0	4.8520E-05	0.0027	0	4.7237E-05	0.0036	0	4.5711E-05	0.0042
Accuracy	**0.9868**	0.9867	0.3736	**0.9868**	0.9867	0.3740	**0.9868**	0.9867	0.3745
(s.d.)	0	3.8238E-05	0.0026	0	5.0517E-05	0.0036	0	6.0573E-05	0.0042
F score	**0.5017**	0.5015	0.0469	**0.5017**	0.5014	0.0467	0.5017	**0.5018**	0.0469
(s.d.)	0	0.0010	0.0019	0	0.0013	0.0019	0	0.0023	0.0020

Table S6. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for scale-free network structures Positive, Negative and Half over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 150 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P200, n50)	Scale-free positive			Scale-free negative			Scale-free half		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**4.3981**	5.1910	191.2565	**4.4003**	5.2303	191.2011	**4.4253**	5.2939	191.5233
(s.d.)	0.1650	0.2692	0.6659	0.1272	0.2377	0.6863	0.1461	0.2736	0.5432
Stein's loss	**9.8990**	13.3903	1958.7836	**10.0976**	13.7089	1961.7145	**10.1667**	13.9406	1964.9255
(s.d.)	0.6240	1.0591	6.8868	0.4597	0.8692	7.5419	0.5174	0.9857	7.6765
Precision	**1**	0.9938	0.0206	**1**	0.9957	0.0206	**1**	0.9944	0.0205
(s.d.)	0	0.0079	0.0006	0	0.0050	0.0008	0	0.0067	0.0005
Recall	0.3344	0.3348	**0.7768**	0.3344	0.3349	**0.7789**	0.3344	0.3349	**0.7776**
(s.d.)	0	0.0010	0.0205	0	0.0012	0.0276	0	0.0012	0.0141
FPR	**0**	3.2147E-05	0.5348	**0**	2.1995E-05	0.5357	**0**	2.8763E-05	0.5343
(s.d.)	0	4.1050E-05	0.0025	0	2.5583E-05	0.0025	0	3.4461E-05	0.0028
Accuracy	**0.9901**	0.9900	0.4697	**0.9901**	0.9900	0.4688	**0.9901**	0.9900	0.4701
(s.d.)	0	4.3018E-05	0.0025	0	2.6749E-05	0.0025	0	3.8693E-05	0.0028
F score	**0.5013**	0.5008	0.0402	**0.5013**	0.5012	0.0401	**0.5013**	0.5010	0.0400
(s.d.)	0	0.0015	0.0011	0	0.0013	0.0016	0	0.0016	0.0009

Table S7. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for scale-free network structures Positive, Negative and Half over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 200 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P250, n50)	Scale-free positive			Scale-free negative			Scale-free half		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**5.0374**	6.2186	236.6587	**5.0073**	6.2395	236.1609	**5.0266**	6.3137	236.6091
(s.d.)	0.1091	0.3048	0.6145	0.0174	0.2449	0.8115	0.2099	0.3806	0.6069
Stein's loss	**12.9199**	18.8915	2819.6789	**12.9766**	19.2947	2817.7321	**13.0276**	19.4635	2822.2823
(s.d.)	0.3989	1.2837	6.0606	0.4700	1.2020	11.9869	0.8417	1.7436	7.2261
Precision	**1**	0.9927	0.0187	**1**	0.9937	0.0187	**1**	0.9911	0.0185
(s.d.)	0	0.0086	0.0006	0	0.0063	0.0004	0	0.0070	0.0003
Recall	0.3342	0.3348	**0.7505**	0.3342	0.3350	**0.7564**	0.3342	0.3346	**0.7487**
(s.d.)	0	0.0012	0.0217	0	0.0016	0.0169	0	9.2445E-04	0.0157
FPR	**0**	3.0228E-05	0.4597	**0**	2.5910E-05	0.4621	**0**	3.6706E-05	0.4615
(s.d.)	0	3.6018E-05	0.0023	0	2.6077E-05	0.0018	0	2.9134E-05	0.0022
Accuracy	**0.9920**	0.9920	0.5427	**0.9920**	**0.9920**	0.5404	**0.9920**	**0.9920**	0.5410
(s.d.)	0	2.8061E-05	0.0024	0	3.1161E-05	0.0017	0	2.9111E-05	0.0021
F score	**0.5010**	0.5008	0.0364	0.5010	**0.5011**	0.0365	**0.5010**	0.5003	0.0361
(s.d.)	0	9.7054E-04	0.0011	0	0.0019	0.0008	0	0.0013	0.0006

Table S8. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for scale-free network structures Positive, Negative and Half over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 250 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P250, n30)	Scale-free positive			Scale-free negative			Scale-free half		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**5.9317**	11.8837	347.7816	**6.2410**	12.1086	348.2220	**6.1445**	11.9801	347.5982
(s.d.)	0.2609	0.7882	1.0779	0.3908	0.7533	0.7812	0.2882	0.7372	1.0481
Stein's loss	**16.8981**	52.2173	4506.4513	**18.2984**	54.1731	4528.8644	**17.7533**	53.0650	4515.2779
(s.d.)	1.0161	5.1706	14.3722	1.3418	4.7993	10.3968	1.4126	5.0977	20.0845
Precision	**1**	0.9862	0.0224	**1**	0.9916	0.0221	**1**	0.9882	0.0221
(s.d.)	0	0.0107	0.0008	0	0.0076	0.0003	0	0.0079	0.0009
Recall	0.3342	0.3344	**0.6267**	0.3342	0.3346	**0.6192**	0.3342	0.3348	**0.6186**
(s.d.)	0	9.7633E-04	0.0236	0	9.2445E-04	0.0067	0	0.0011	0.0279
FPR	**0**	5.7218E-05	0.3213	**0**	3.4547E-05	0.3199	**0**	4.8581E-05	0.3209
(s.d.)	0	4.4751E-05	0.0026	0	3.1748E-05	0.0020	0	3.2666E-05	0.0017
Accuracy	**0.9920**	**0.9920**	0.6781	**0.9920**	**0.9920**	0.6794	**0.9920**	**0.9920**	0.6784
(s.d.)	0	4.3788E-05	0.0025	0	3.3544E-05	0.0019	0	3.6775E-05	0.0015
F score	**0.5010**	0.4994	0.0432	**0.5010**	0.5003	0.0426	**0.5010**	0.5001	0.0426
(s.d.)	0	0.0016	0.0016	0	0.0014	0.0006	0	0.0017	0.0018

Table S9. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for scale-free network structures Positive, Negative and Half over 30 datasets generated by multivariate normal distributions with precision matrix Ω, where p = 250 and n = 30. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P100, n50)	Different.Hubs			Different.Cliques positive			Different.Cliques negative		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.4594**	3.6141	82.9108	**3.1724**	3.4886	82.8688	**3.0308**	3.3812	82.5949
(s.d.)	0.8119	0.6405	**0.6288**	0.6422	**0.4880**	0.7030	0.4422	0.4894	**0.3415**
Stein's loss	**8.6314**	9.3594	501.4684	**7.6612**	8.8059	494.3325	**4.0098**	4.9211	478.5753
(s.d.)	**6.0021**	6.3452	33.4307	5.3531	**5.2763**	28.2741	**1.0906**	1.3944	8.7506
Precision	**1**	0.9934	0.02201	**1**	0.9969	0.0196	**1**	0.9968	0.0191
(s.d.)	**0**	0.0152	0.0010	**0**	0.0070	0.0007	**0**	0.0089	0.0013
Recall	0.5556	0.5726	**0.9585**	0.625	0.6550	**0.9597**	0.625	0.6267	**0.9476**
(s.d.)	**0**	0.0265	0.0263	**0**	0.0397	0.0196	**0**	0.0043	0.0355
FPR	**0**	7.47E-05	0.7628	**0**	3.39E-05	0.7649	**0**	3.39E-05	0.7653
(s.d.)	**0**	1.73E-04	0.0057	**0**	7.70E-05	0.0037	**0**	9.37E-05	0.0054
Accuracy	0.992	**0.9922**	0.2498	0.994	**0.9944**	0.2465	**0.994**	0.9940	0.2457
(s.d.)	**0**	4.58E-04	0.0056	**0**	6.25E-04	0.0038	**0**	1.23E-04	0.0056
F score	0.7143	**7.26E-01**	0.0430	0.7692	**0.7899**	0.0384	0.7692	**0.7695**	0.0373
(s.d.)	**0**	0.0200	0.0020	**0**	0.0277	0.0014	**0**	0.0045	0.0025

Table S10. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for network structures Hubs, Cliques positive and Cliques negative over 30 different datasets generated by multivariate normal distributions with 30 different precision matrices Ω, where p = 100 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

(P150, n50)	Different.Hubs			Different.Cliques positive			Different.Cliques negative		
	GR2D2	GHS	GL	GR2D2	GHS	GL	GR2D2	GHS	GL
F-norm	**3.8987**	4.3683	141.2842	**3.7038**	4.2616	141.2350	**3.6989**	4.1376	140.8548
(s.d.)	0.7092	0.6009	**0.5567**	0.6311	0.4984	**0.4553**	0.4910	0.4638	**0.4384**
Stein's loss	**10.1162**	11.5502	1184.3517	**9.7600**	11.4109	1175.2010	**5.8451**	7.1299	1148.7890
(s.d.)	6.5300	**6.4673**	58.1842	**5.9547**	6.2515	53.1417	**1.4292**	1.6567	17.7214
Precision	1	0.9904	0.0151	1	0.9949	0.0138	1	0.9969	0.0136
(s.d.)	0	0.0103	0.0006	0	0.0071	0.0006	0	0.0056	0.0008
Recall	0.6522	0.6672	**0.9528**	0.7143	0.7368	**0.9554**	0.7143	0.7149	**0.9482**
(s.d.)	0	0.0245	0.0278	0	0.0314	0.0279	0	0.0024	0.0400
FPR	0	6.89E-05	0.6342	0	3.59E-05	0.6339	0	2.09E-05	0.6339
(s.d.)	0	7.71E-05	0.0033	0	5.05E-05	0.0038	0	3.86E-05	0.0021
Accuracy	0.9964	**0.9965**	0.3717	0.9973	**0.9975**	0.3715	0.9973	**0.9973**	0.3714
(s.d.)	0	2.08E-04	0.0033	0	2.95E-04	0.0036	0	4.10E-05	0.0023
F score	0.7895	**0.7970**	0.0297	0.8333	**0.8463**	0.0273	**0.8333**	0.8327	0.0269
(s.d.)	0	0.0152	0.0011	0	0.0204	0.0012	0	0.0023	0.0016

Table S11. Frobenius norm (F-norm), Mean Stein's loss, precision, recall, false positive rates, accuracy and F scores of precision matrix estimates for network structures Hubs, Cliques positive and Cliques negative over 30 different datasets generated by multivariate normal distributions with 30 different precision matrices Ω, where p = 150 and n = 50. The precision matrix is estimated by frequentist graphical lasso (GL), the graphical horseshoe (GHS), and the graphical R2D2 (GR2D2). The best performance in each row is shown in bold.

References

[1] Anirban Bhattacharya, Debdeep Pati, Natesh S Pillai, and David B Dunson. Dirichlet–laplace priors for optimal shrinkage. *Journal of the American Statistical Association*, 110(512):1479–1490, 2015.

[2] Willem Kruijer, Judith Rousseau, Aad Van Der Vaart, et al. Adaptive bayesian density estimation with location-scale mixtures. *Electronic journal of statistics*, 4:1225–1257, 2010.

[3] Enes Makalic and Daniel F Schmidt. A simple sampler for the horseshoe estimator. *IEEE Signal Processing Letters*, 23(1):179–182, 2015.