The interplay between habitat fragmentation and life traits affects

specialization in lichen symbioses

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Abstract

1. Interactions between organisms are determined by species traits and differ in specialization,

from generalist to highly specialized relationships. Although we expect that the effect of habitat

loss and fragmentation on the abundance and survival of species depends on their degree of

specialization, few studies have deepened into the interplay between interaction specialization

and habitat loss.

2. Here, we investigate the combined effect of habitat fragmentation and life traits (growth type

and reproductive mode) on the specialization of lichen-forming fungi (mycobionts) towards its

photosynthetic partners (photobionts) in lichen symbioses.

3. We studied mycobiont specialization in epiphytic lichen communities present in ten

fragments of *Quercus rotundifolia* trees embedded in an agricultural matrix. Both mycobionts

and photobionts were identified through DNA sequencing and mycobiont specialization was

measured through interaction parameters calculating the relative number of interactions

(normalized degree; ND) and the specialization of each species based on its discrimination from

a random selection of partners (d'). Phylogenetic generalized linear mixed models were used to

analyze the effect of patch size as well as the life traits growth type (crustose, foliose, fruticose)

and reproduction mode (sexual vs asexual) on mycobiont specialization.

4. Both mycobiont and photobiont richness along the patch size gradient followed a hump-back

pattern, which was more pronounced in photobionts. Mycobionts forming crustose biotypes

established the largest number of interactions. Mycobiont specialization (d') was larger for

fruticose and foliose biotypes and for species with vegetative reproduction. Along the gradient

of fragment size, the relative number of interactions decreased and the specialization of

mycobionts with vegetative reproduction increased.

5. *Synthesis*: The analysis of mycobiont specialization towards their photobionts in communities

of epiphytic lichens in a fragmented Mediterranean forest revealed that the interaction between

species life traits and habitat loss on specialization cannot be neglected. The results also pointed

to the ability of some species to modulate their interactions to certain extent, suggesting that

species might have a greater resilience to abiotic changes than expected, presumably creating

extinction debt or even avoiding extinction processes to some extent.

Keywords: Epiphytic, lichen-forming fungi, Mediterranean forest, photobionts, selectivity.

Resumen

1. Las interacciones entre organismos están determinadas por los rasgos funcionales de las

especies y difieren en su grado de especialización, desde interacciones generalistas hasta las

altamente especializadas. Aunque se espera que el efecto de la pérdida y fragmentación del

hábitat sobre la abundancia y la supervivencia de las especies dependa de su grado de

especialización, pocos estudios han profundizado en la interacción entre la especialización de

las interacciones y la pérdida de hábitat.

2. En este artículo investigamos el efecto combinado de la fragmentación del hábitat y los rasgos

funcionales de las especies (tipo de crecimiento y modo reproductivo) en la especialización de

los hongos liquenizados (micobiontes) hacia sus socios fotosintéticos (fotobiontes) en las simbiosis de líquenes.

3. Se estudió la especialización de micobiontes en comunidades de líquenes epífitos presentes en diez fragmentos de árboles de *Quercus rotundifolia* incrustados en una matriz agrícola. Tanto los micobiontes como los fotobiontes fueron identificados mediante la secuenciación del ADN y la especialización de los micobiontes se midió a través de parámetros de interacción calculando el número relativo de interacciones (grado normalizado; ND) y la especialización de cada especie en función de su discriminación de una selección aleatoria de simbiontes (d'). Se utilizaron modelos lineales mixtos generalizados filogenéticos para analizar el efecto del tamaño del parche, así como los rasgos de tipo de crecimiento (crustáceo, folioso, frutal) y el modo de reproducción (sexual frente a asexual) sobre la especialización de los micobiontes.

4. Tanto la riqueza de micobiontes como de fotobiontes a lo largo del gradiente de tamaño de los parches siguió un patrón de U inversa, que fue más pronunciado en los fotobiontes. Los micobiontes que forman biotipos crustáceos establecieron el mayor número de interacciones. La especialización de los micobiontes (d') fue mayor para los biotipos fruticulosos y foliosos y para las especies con reproducción vegetativa. A lo largo del gradiente del tamaño del fragmento, el número relativo de interacciones disminuyó y la especialización de los micobiontes con reproducción vegetativa aumentó.

5. Síntesis: El análisis de la especialización de los micobiontes hacia sus fotobiontes en comunidades de líquenes epífitos en un bosque mediterráneo fragmentado reveló que no se puede descartar la interacción entre los rasgos de vida de las especies y la pérdida de hábitat sobre la especialización. Los resultados también apuntaron a la capacidad de algunas especies para modular sus interacciones hasta cierto punto, sugiriendo que las especies podrían tener una resiliencia a los cambios abióticos mayor de lo esperado, presumiblemente creando una deuda de extinción o incluso evitando los procesos de extinción hasta cierto punto.

Introduction

Habitat loss due to anthropogenic land use represents one of the main drivers of biodiversity

loss under the current scenario of global change (Haddad et al., 2015). A common process

following habitat loss is habitat fragmentation in which natural homogeneous habitat is

transformed into smaller fragments embedded in a matrix different from the original one

(Wilcove et al., 1986). Consequently, fragmentation leads to a reduction in the amount of

available habitat for species as well as its connectivity, and to changes of abiotic and biotic

conditions in the remaining habitat (Fahrig, 2003). This process usually entails the decrease of

population sizes, increasing the risk of species extinctions, and subsequent reduction of

ecosystem structure, functioning and services (Isbell et al., 2011; Millennium ecosystem

assessment 2005).

Changes derived from habitat fragmentation affect species unevenly depending on life

strategies, population sizes and reproductive and physiological traits (Bovo et al., 2018; Farneda

et al., 2015; Henle et al., 2004). Ecological specialization towards abiotic and biotic conditions is

a highly relevant trait that may determine species fate during habitat fragmentation, being

specialists more prone to suffer fragmentation effects (Devictor et al., 2008; Nordén et al.,

2013).

As species interact with other species in natural habitats, fragmentation and habitat loss

jeopardize species interactions (Tylianakis et al., 2008). Whereas the loss of taxonomic diversity

has received large attention during the last decades, knowledge on the effect of habitat loss on

interactions is still scarce (Gonzalez et al., 2011). The loss of interactions often precedes the loss

of species diversity in ecosystems and it is seen as an early signal of ecosystem decay (Aizen et

al., 2012; Valiente-Banuet et al., 2015). Not all kinds of interactions are equally affected by

fragmentation, which has been shown to produce more negative effects on mutualisms than on

antagonisms (Magrach et al., 2014). Most of the knowledge we have on the effect of

fragmentation on mutualisms comes from non-intimate interactions (e.g., seed dispersal, plant

pollination) whereas intimate mutualisms have been largely ignored.

Independently of the interaction type, species show variability regarding the degree of

specialization towards their partners, which ranges from generalists, interacting with many

partners, to specialists, which interact with a few or even a single partner (Solé and Montoya

2001; Thompson, 1988). These strategies involve different evolutionary and ecological

advantages and constraints (Dennis et al., 2011; Ollerton, 2006). Furthermore, they have been

highly relevant to predicting the loss of interactions and species in gradients of habitat

availability (Aizen et al., 2012). In addition, specialization is correlated with morphological and

reproductive traits (Maglianesi et al., 2014; Otálora et al., 2013; Reif et al., 2016; Santamaría and

Rodríguez-Gironés, 2007) as well as with species´ evolutionary history (Webb et al., 2010).

However, it is largely ignored whether species might modulate their specialization under

changing environmental conditions in order to survive. Overall specialization tends to decrease

together with habitat availability (Hadley et al., 2018; Jauker et al., 2019). This decrease may be

due to the loss of specialists (Aizen et al., 2012; Bagchi et al., 2018) or due to the variation of

specialization within species, which may show certain plasticity to cope with new habitat

conditions.

Lichens, the symbiotic phenotype of lichen-forming fungi (hereafter, the mycobiont) associated

with, at least, one photosynthetic partner (the photobiont) a cyanobacteria and/or a green alga

(Grube and Hawksworth 2007) are paradigmatic examples of intimate mutualisms. Due to their

high sensibility to subtle changes in abiotic conditions in the environment, they are renowned

bioindicators of habitat conditions (Marmor et al., 2011; McCune 2000; Nascimbene and Marini

2015; Rivas Plata et al., 2008), including changes in land management and habitat fragmentation

(Aragón et al., 2010; Brunialti et al., 2013; Matos et al., 2017; Svoboda et al., 2010).

Some functional traits of lichens, mostly related to growth types and reproductive modes, are

commonly used in ecological and indicator studies as a response to many environmental or

anthropogenic changes (Ellis et al., 2021; Giordani et al., 2012; Hurtado et al., 2020; Koch et al.,

2019; Nelson et al., 2015). Another functional trait commonly used is the photobiont type, often

categorized as highly broad taxonomic categories (e.g. Giordani et al., 2012). The last two

decades have seen great advances in the understanding of photobiont diversity and the range

of fungal-algal interactions in lichen symbioses (Muggia et al., 2020), especially to know the

breadth of compatible photobionts at different systematic levels (e.g. (Dal Grande et al., 2012;

Fernández-Mendoza et al., 2011; Leavitt et al., 2015; Thüs et al., 2011)). During this time, it has

become clear that mycobiont specialization towards the photobiont varies among species and

it is likely a species-specific trait (Magain et al., 2016; Pérez-Ortega et al., 2012). However, the

knowledge about the influence of certain functional traits on photobiont specialization is scarce.

Lichens show a wide range of reproductive strategies (Tripp and Lendemer, 2018), but they can

be simply divided between asexual versus sexual reproduction. Most common asexual

reproductive strategies involve vegetative clonal propagules (soredia, isidia…) in which both

bionts are dispersed together ensuring the availability of compatible photobionts for the fungi

at the expense of suppressing genetic variability in both bionts (Buschbom and Mueller 2005;

Yahr et al., 2006). On the contrary, the dispersal of the mycobiont via ascospores implies the

propagule to find a compatible photobiont after its establishment on the new substrate

(Fernández-Mendoza et al., 2011; Yahr et al., 2006). The scarce studies in which specialization

towards the photobiont has been explored within a reproductive framework offer contradictory

results (Otálora et al., 2013; Wornik and Grube, 2010), likely due to the possibility of photobiont

switches after the vegetative propagule is settled (Nelsen and Gargas, 2008)

Lichen thalli also show a number of growth forms named biotypes (Honegger, 2001), which have

numerous implications for associated photobionts since different architectures and topologies

imply the existence of different microniches (Hawksworth and Grube, 2020). The most common

biotypes, which are the focus of this study, are: 1) crustose, in which the thallus lacks a lower

cortex and is attached to the substrate through the medulla, 2) foliose, in which the thallus has

a laminar form, partially adhered to the substrate by certain fixation structures, 3) fruticose, in

which thallus shows a shrub-like form attached to the substrate usually by a single point. The

role of thallus morphologies on photobiont specialization has not been formally tested, although

it is known that, for instance, crustose species usually display a high phylogenetic breadth of

photobiont partners (Blaha et al., 2006; Guzow-Krzeminska, 2006; Muggia et al., 2014).

In this study, we explore the effect of habitat fragmentation on mycobiont specialization toward

their photobionts as well as the effect of growth and reproduction traits on the specialization

along the fragmented habitat. We study the richness and specialization of epiphytic lichen-

forming fungi and their associated photobionts in habitat fragments of Mediterranean forest of

increasing size. We hypothesize that habitat loss reduces both biont richness and specialization.

Finally, we explore whether species specialization may be affected by species-specific functional

traits such as reproductive mode and growth form.

Material and Methods

Study area

The area of study was located in Central Spain (42° 5-8' N; 3º 36-44' W; 923-1093 m a.s.l.). The

area harbors remnants of forest embedded in an agricultural matrix (cereal crops) (Figure 1).

Fragments are either dominated by one of the main dominating tree species in the area (*Quercus*

rotundifolia, Q. pyrenaica and *Q. faginea*) or show mixed formations in which *J. oxycedrus* also

occasionally occur.

Figure 1. Map detailing the area where the study was carried out. Images obtained from Google

Earth (14/12/2015) Europe. (Landsat/Copernicus Data SIO, NOAA, U.S. Navy, NIGA, GEBCO,

GeoBasis-DE/BKG (©2009) Map GISrael Instituto Geográfico Nacional; 45°09'57"N 11°06'33"E.

https://earth.google.com/web/@45.16589093,11.10934345,1020.81870028a,3304630.75835

109d,35y,0h,0t,0r) The second map (smaller one on the right) is also obtained from Google Earth

(20/08/2018). Madrigal del Monte. Instituto Geográfico Nacional. 42º07'17"N 3º38'41"W.

https://earth.google.com/web/@42.12151496,3.64472552,955.37218271a,9380.0627212d,35

y,0h,0t,0r

Sampling

The selection of the habitat fragments was made as follows: Orthorectified photographs were

downloaded from the National Centre for Geographic Information

(http://centrodedescargas.cnig.es/CentroDescargas/-index.jsp) corresponding to the year 2017,

and by means of the Google Earth Pro software (available at

https://www.google.com/earth/download/gep/agree.html) all forest fragments present in the

area were identified. Perimeter and area were measured for each fragment. Fragments with

highly irregular geometries (low area/perimeter ratio) were discarded to avoid disproportionate

edge effects. The remaining fragments were inspected during the fall of 2017 in order to check

the tree composition homogeneity. Considering the available diversity of fragment sizes, we

chose fragments in which *Quercus rotundifolia* was the dominant species (> 75 %). Finally, a total

of 10 fragments were selected in a size gradient ranging from 0.002 ha (a single isolated tree) to

250ha (Table 1).

Table 1. Mycobiont and photobiont richness and mean mycobiont specialization in each studied
forest patch. A= Area of each forest patch, in hectares; M_d= Number of mycobiont species; M_e=
Number of exclusive mycobiont species in a forest patch; P_d= Number of photobiont ASVs; P_e=
Number of exclusive ASV species in a forest patch; d= mean degree; ND: mean normalized
degree.

A	M_d/M_e	P_d/P_e	d	ND
0.002	9/1	25/3	9	0.33
0.1	17/1	34/8	7.64	0.22
0.3	22/1	42/6	9.13	0.217
1.02	23/1	63/12	10.83	0.17
4.23	27/1	51/13	8.37	0.17
12.21	28/1	75/20	9.54	0.13
16.13	23/0	57/8	11.87	0.21
35.85	23/1	52/9	8.39	0.16
48.01	27/0	61/12	8.89	0.15
250	20/2	46/7	2.9	0.06

In each fragment, epiphytic lichen communities were surveyed at the geographic center of the

fragments. Sampling was carried out according to Asta et al., (2002). In summary, we sampled

the lichen community in a total of ten tree trunks (diameter ≥ 12 cm) in both north and south

orientation, using a 10×50 cm grid divided into five 10×10 cm squares and annotated all species

present and their abundance which was measured as the number of squares where the species

occurs. Once the inventory was completed, we attempted to collect a total of 10 individuals of

all the species identified in each fragment. This search lasted up to a maximum of 4 hours per

plot (two people). Samples were dried at room temperature and stored until further processing.

Molecular methods

Mycobiont barcoding

Field identifications of mycobionts were subsequently checked using currently used methods in

lichen taxonomy and available literature. Further, we sequenced the nrITS region, the universal

fungal barcode for fungi (Schoch et al., 2012) to corroborate identifications in each of the

identified morphospecies. Briefly, DNA extractions were performed using cationic exchange

resin Chelex 100 (BioRad, Madrid) following Ferencova et al., (2017). PCRs were performed using

the primers ITS1F and ITS4-Kyo2 (Gardes and Bruns 1993; Toju et al., 2012). Reactions included

6.5 µl of MyTaq Red Mix (BioLine, UK), 2 µl of diluted (1:10) DNA template, 0.5 µl (10uM) of each

primer and 5.5 µl of sterile distilled H_2O. PCR conditions were set as follows: a denaturation step

of 94ºC for 5 min followed by 35 cycles of 95ºC for 30 s, 57ºC for 30 s, and 72 ºC for 1 min ending

with an elongation of 72ºC for 7 min. Amplicons were sequenced in Macrogen (Madrid, Spain)

using the same primers as in the amplification reaction.

Mycobiont phylogeny

We constructed the phylogenetic tree depicting the evolutionary relationships of mycobionts to

account for statistical non-independence derived from common ancestry in the models. DNA

extractions were either using extractions obtained in the previous section or using E.Z.N.A.®

Forensic Kit (Omega Bio-Tek, Norcross, Georgia, USA) according to the protocol defined by the

company. To determine phylogenetic affinities, we used DNA three nuclear genomic regions, *i.e.*

the ribosomal internal transcribed spacer (nrITS) obtained in the previous section, the nuclear

large subunit region (nrLSU), and the largest subunit of RNA polymerase II (RPB1), as well as the

mitochondrial small subunit locus (mtSSU). The primers used for the amplification of these

regions are provided in Supplementary Material Table 1.

PCR reactions were performed in a total volume of 15 µL, containing 2 µL of template DNA for

amplification of the nuLSU, mtSSU (or 4 µL for the amplification of the RPB1 region), 0.5 µL of

each primer (10 mM) and 6.5 µL of MyTaq Mix wich contains MyTaq DNA Polymerase (Bioline,

UK)) and dNTPs; distilled water was added to reach the final volume.

The PCR conditions used for amplification for RPB1, nuLSU and mtSSU were the same as

described in Perez-Ortega et al., (2016). PCR products were sequenced by Macrogen Inc.

(Madrid, Spain) using the same primers as in the amplification reaction.

The four regions were aligned through MAFT tool v 1.4.0 (Katoh and Standley 2013) using the

algorithm "auto" implemented in Geneious Prime v 2019.0.3

(https://www.geneious.com/prime/). To eliminate the ambiguously aligned regions of those

alignments, Gblocks v 0.91b (http://molevol.cmima.csic.es/castresana/Gblocks_server.html)

has been used (Castresana, 2000) allowing the least stringent parameters smaller final blocks,

gap position within the blocks and less strict flanking positions. In order to select the best-fit

partitioning schemes and nucleotide evolution models, we used PartitionFinder v 2.1.1 (Lanfear

et al., 2017). Phylogenetic relationships among taxa were calculated using Bayesian inference in

BEAST2 v 2.6 (Bouckaert et al., 2014) as implemented in CIPRES Science Gateway v 3

(https://www.phylo.org/) (Miller et al., 2010). We used an uncorrelated relaxed model with a

log-normal prior for modelling clock and a Yule process to model tree prior (Drummond et al.,

2006). The MCMC chain length was run during 5×10^9 generations and results were logged every

5000 generations. Trace plots and effective sample sizes (ESS) were examined by TRACER v 1.7

(Rambaut et al., 2018). Finally, after discarding the first 25% sampled trees (burning), the results

were summarized and annotated in a maximum clade credibility tree (MCC) through

TreeAnnotator v 1.8.4 (https://beast.community/treeannotator) (Drummond et al., 2007). For

the visualization of the resulting consensus tree and comparison of retrieved phylogenetic

relationships with previous literature, we used Figtree v 1.4.4 (Rambaut, 2012). Accession

numbers for the sequences generated and used during this study are available in Supplementary

Material Table 2.

Photobiont

A small fragment (1-2 mm^2) of the thallus was taken from each specimen. Special care was taken

to avoid thallus areas with clear signs of epiphytic fungi or algae as well as necrotic areas. Up to

ten fragments per species and forest fragment were pooled in a single microcentrifuge tube.

DNA extractions were performed using E.Z.N.A. Forensic DNA Kit (Omega Bio-Tek, Norcross,

Georgia, USA) following the manufacturer´s instructions. The second part of the internal

transcribed spacers (ITS2) of the photobiont was amplified using the primers FDGITS2-f y

FDGITS2-r (Dal Grande et al., 2018) with Illumina adapters CS1 y CS2 attached (Available in:

https://rtsf.natsci.msu.edu/-genomics/sequencing-services/sample-requirements-for-illumina-

sequencing/). PCR reactions were performed using a mix of 25 µl which contained 0.625 U

PrimerSTAR GXL DNA Polymerase (Takara BioInc, Japan), 5 µl of Buffer, 2 µl of dNTP Mixture (2.5

mM), 3 µl of DNA template, 0.5 µl of each primer at 10 mM and 13.5 µl of sterile distilled H$_2$O.

PCR conditions were set as follows: a 94ºC denaturation step for 1 min followed by 30 cycles of

95ºC for 15 s, 52ºC for 15 s and 72 ºC for 30 s ending with an elongation of 72ºC for 1 min. PCR

products were visualized in agarose gels and quantified by using a Qubit® Fluorometer (Life

Technologies, Darmstadt, Germany) and normalized. Samples were indexed and pooled in a

single Illumina MiSeq run (2 x 250bp paired-end sequencing, v2 Standard 500 cycle) at the

Research Technology Support Facility Genomics Core at Michigan State University (USA).

Bioinformatic analysis

MiSeq reads were merged, demultiplexed and filtered using the package DADA2 v 1.8.0

(https://benjjneb.github.io/dada2/index.html)   for   R   studio   (v   3.4.3   https://cran.r-

project.org/bin/windows/base/old/3.4.3/). The first-left 22 bp corresponding to primer

sequences were trimmed using the *filterAndTrim* function. Based on quality plots forward and

reverse reads were trimmed to 200 and 170 bp respectively. Sequence filtering parameters was

set to maxN = 0, maxEE=c (1,4), trunQ =2. The remaining parameters were operated as default.

Finally, amplicon sequence variants (ASVs) were generated and a data matrix was built in which

the rows represented the species of mycobiont, columns the ASVs of the photobiont and each

cell contained the number of reads of each ASV inside the lichen, being the last a proxy for the

interaction strength. The table containing the data was rarefied using the function "*rrarefy*"

from the package "*vegan*".

We could not rule out the presence of contaminations in spite of the special care taken during

sample tissue collection, as thallus surface may dwell single algae cells not visible under the

stereoscope. Further, some algal species may occur inside the thallus but not taking part in the

symbiosis (Moya et al., 2017). In order to overcome this problem, we performed a double

filtering of the obtained results. First of all, we collected information available in the literature

about the photobionts associated with the species and/or genera found in our study (Blaha et

al., 2006; Muggia et al., 2014; Tschermak-Woess, 1988; Wornik and Grube, 2010). Secondly, we

established a limit of reads (100) below which we considered that the signal corresponded to

contaminations of transients or algae not primarily associated with the mycobiont. Thus,

sequences of taxa which based on the available literature do not represent the actual

photobiont of the species and/or those with less than 100 reads were eliminated from

subsequent analyses. All sequences obtained in this study will be available in the SRA (NCBI).

Species specialization

An interaction matrix between mycobionts and photobionts was built to account for interactions

in each forest patch. Mycobiont specialization towards their photobionts was explored by means

of two metrics, normalized degree and the parameter *d'*. Normalized degree is the number of

links (partners) per species normalized by the total number of possible partners. The parameter

d' measures the specialization of each species based on its discrimination from a random

selection of partners, ranging from 0 to 1 (a species interacts with only one other species, and

that species only interacts with the first one) (Blüthgen et al., 2006). Both metrics were

calculated for every species at all studied forest fragments using the function *specieslevel* from

the *bipartite* v. 2.16 R package (Dormann et al., 2008).

Statistical analyses

We first explored changes in ND and *d'* at the species level using Pearson correlation tests

between each of the two specialization parameters and the fragment size gradient in those

species present in at least five fragments. Analyses were carried out using the function *cor.test*

of the *stats* v. 3.6.2 R package. Then, we explored the independent and cross effect of fragment

size and two life traits, growth form and reproduction mode, on species specialization. All

species recorded during the inventories fitted well within the main three growth forms

recognized in lichens (Nash, 1996), i.e. crustose, whose thalli are strongly attached to the

substrate and cannot be separated from it; foliose, whose thalli have a more or less flat

appearance and are attached to the substratum by specialized structures in the lower cortex;

and fruticose, whose thalli have a three-dimensional architecture, similar to tiny bushes and are

attached to the substratum from a single point. Secondly, we categorized all species based on

their reproduction modes. Although several reproductive strategies may be found in lichens

(Tripp and Lendemer, 2018), species may be easily divided between those producing meiotic

propagules (ascospores) which once settled in a new substrate they must find a compatible

photobiont to reestablish the symbiosis, and those producing asexual propagules in which both

partners, fungus and alga, are dispersed together, and the new thalli correspond to clones of

the source thallus. We first explored the effect of area on the number of algae ASV and

mycobiont species using the function *lm* of the package *stats*.

We investigated the effect of both functional traits, growth form and reproduction type in the

mycobiont specialization towards their photosynthetic partners independently and in the

context of a gradient of habitat availability. We used generalized linear mixed models in a

Bayesian framework (Hadfield and Nakagawa, 2010) implemented in the package MCMCglmm

v. 2.29 (Hadfield, 2010) in order to control the phylogenetic effects on these relationships. The

uncertainty in the phylogenetic reconstructions was accounted for by running three

MCMCglmm´s, each one using a phylogenetic tree randomly chosen from the distribution of tree

topologies obtained in BEAST after discarding 25% of samples as burnin, and integrated over the

posterior samples by drawing 1000 random samples across models and using *HPDinterval*

function from the *coda* R package Plummer *et al.,* 2013). Models priors follow de Villemereuil

and Nakagawa (2014) using an inverse-Gamma distribution with shape and scale parameters

equal to 0.01 as priors for the random effects and for the residual variance.

All graphs for the analyses performed were created with the R packages *ggplot2* (Wickham

2016), *gridExtra* (https://CRAN.R-project.org/package=gridExtraand) and *wesanderson*

(https://CRAN.R-project.org/package=wesanderson).

Interaction turnover

In order to shed light on the origin of changes in mycobiont specialization along the fragment

size gradient, we analyzed the interaction beta diversity using the framework proposed by Poisot

et al. (2012). This approach decomposes the total beta diversity of interactions (β_{WN}) between

two sites into the interaction dissimilarity due to species turnover species (β_{ST}) and interaction

dissimilarity when species occur in both sites but interaction not (β_{OS}) (Poisot et al. 2012).

RESULTS

Mycobionts

We detected a total of 44 species of lichen-forming fungi in the study area (Sup. Mat. Table 2).

17 species were crustose, 19 foliose and 8 fruticose. Twenty species reproduced mainly sexually

whereas 24 showed vegetative reproduction ((Sup. Mat. Table 2). Number of mycobiont species

per forest fragment ranged from 9 at the 0.002 ha fragment to 28 at the 12.21 ha, with an

average of 22 species per fragment (Table 1), with a distribution along the area that was adjusted

to a quadratic model (R^2= 0.82; p<0.05; Figure 2). The number of exclusive taxa was low in all

fragments, occurring the highest, 2, at the largest forest fragment (Table 1).

The richness of crustose and foliose species did not vary along the area gradient while that of

fruticose species showed a quadratic relation (R^2= 0.571; F= 6.998, 2; d.f.=7; p-value<0.05), being

maximum in intermediate plots. Regarding reproduction mode, the richness of sexual species

showed a quadratic relationship with the log of area (R^2= 0.617; F= 8.237, 2; d.f.=7; p-

value<0.05), while the relationship for asexual species followed a positive linear trend (R^2=

0.346; F= 5.756, 1; d.f.=8; p-value<0.05).

Figure 2. Distribution of number (N) of mycobiont species and ASVs along the patch size gradient
(log(hectares)). Quadratic adjusted models (bold line) and confidence intervals (paler colored

area) are represented for the number of mycobiont species in red (R^2= 0.82; p<0.05) and green

for ASVs (R^2= 0.52; p <0.05).

Photobionts

A total of 2272 thalli were collected, corresponding to the 229 unique pools used for Illumina

MiSeq analysis. The average number of thalli per species and fragment was 9.92. A total of

8,478,480 paired-end reads were obtained from the photobiont ITS2 amplicons. After quality

filtering, a total of 4,956,878 reads remained. The processing of these reads resulted in 985

ASVs. After this processing we applied a manual filter removing the ASVs with less than 100

reads, obtaining a final number of 175 ASVs. Furthermore, the total number of reads remaining

after this manual filter was 4,903,976 with an average of 21,415 reads per sample. BLAST

searches of these remaining ASVs resulted in 158 corresponding to the genus *Trebouxia*, 13 to

Dictyochloropsis and 4 to *Asterochloris*. *Asterochloris* ASVs were restricted to *Cladonia*, while

the genus *Dictyochloropsis* was mostly found in the species *Phlyctis argena*.

The number of ASVs per forest fragment ranged from 25 at 0.002 ha to 75 at 12.21 ha (Table 1).

ASV diversity along the patch size gradient followed a quadratic relationship (R^2= 0.52; p <0.05;

Figure 2). The highest number of exclusive photobiont ASVs (20) occurred at the 12.21 ha forest

fragment (Table 1).

Specialization

The mean number of ASVs per mycobiont species, species degree, ranged from 2.9 at the 250

ha fragment to 11.87 at the 16.13 ha fragment (Table 1). The normalized degree ranged from

0.01 in *Evernia prunastri* at the 12.21 ha forest fragment to 0.54 in *Blastenia xerothermica* at the

0.3 ha forest fragment (Supplementary Material Table 3). The specialization parameter *d'* ranged

from 1 in *Cladonia fimbriata* at the 563.9 ha forest fragment to 0.16 in *Lecanora subcarpinea* at

the 1.02 ha forest fragment (Supplementary Material Table 3).

The correlation analyses between ND and area in species with occurrences in ≥ 5 forest patches

showed negative Pearson's correlation coefficients for all species (Supplementary Material

Table 4)., although they were only statistically significant in six of them, i.e. *Candelariella*

xanthostigma, *Melanelixia subaurifera*, *Parmelina tiliacea*, *Physconia perisidiosa* and *Ramalina*

fastigiata, all but the last one are vegetatively reproducing species. Regarding correlations

between d' and area, they were only significant in *Physconia enteroxantha* and *Physconia*

perisidiosa, for which d' increased along the forest fragment size gradient (Supplementary

Material Table 4).

Phylogenetic generalized linear models showed differences in average normalized degree

between crustose and fruticose species (Figure 3A), but not between reproduction modes

(Figure 3B, Supplementary Material Table 4). Regarding the specialization parameter *d'* we

found significant differences between crustose species and foliose and fruticose species (Figure

3C) as well as between reproduction modes, with vegetative species showing higher average d'

(Figure 3D, Supplementary Material Table 4).

Figure 3. Boxplots show differences in specialization among species with different functional traits. A) Differences among biotypes in the specialization parameter *d'*; B) Differences between reproduction modes in the specialization parameter *d'*; C) Differences among biotypes in normalized degree (ND); D) Differences between reproduction modes in normalized degree (ND). The letters above boxplot depict significant differences among functional groups. Significance is based on phylogenetic controlled MCMCglm models (see. Mat. Supp. Table 3).

Phylogenetic generalized linear models also showed that area had a significant and negative effect on normalized degree (Figures 4A, B, Table 2). There were significant differences between crustose taxa and foliose and fruticose species and these differences were independent of the area (Table 2). No significant differences were found between reproduction modes regarding normalized degree along the area gradient (Table 2). Concerning the specialization parameter

d', the area had a significant effect, the larger the patch size the larger d' when considering

species biotypes (Figures 4C, D, Table 2). In addition, fruticose species had larger d' than crustose

species. Interestingly, there is a significant interaction between area and fruticose species, in

which the increase of the parameter d' along the area gradient is lower than in crustose species

(Figure 4C, Table 2). When considering the effect of area and reproductive modes on d' we

found differences between sexual and asexual species, with significant effects of interactions

(Figure 4D, Table 2). Vegetatively reproduced species increased their d' along the area gradient,

but the pattern was the contrary in sexual species.

Table 2. Results of Phylogenetic Generalized Linear Models explaining Normalized degree and
specialization (d´) as a function of biotype, reproductive mode and area. Variables with
significant effect, those in which the highest posterior density interval does not include zero, are
shown in bold. Area was implemented in all analyses as log(area+1).

	Highest Posterior Density intervals
*Normalized degree ~ biotype*area*	
Intercept	**[0.161 — 0.301]**
Area	**[-0.035 — -0.019]**
Biotype crustose	**[0.000 — 0.146]**
Biotype fruticose	[-0.134 — 0.033]
Area: Biotype crustose	[-0.015 — 0.012]
Area: Biotype fruticose	[-0.007 — 0.025]
*Normalized degree ~ reproductive mode*area*	
Intercept	**[0.203 — 0.353]**
Area	**[-0.037 — -0.019]**
Vegetative reproduction	[-0.092 — 0.016]
Area: Vegetative reproduction	[-0.002 — 0.023]
*d' ~ biotype*area*	
Intercept	**[0.434 — 0.815]**
Area	**[0.008 — 0.038]**
Biotype crustose	[-0.285 — 0.004]
Biotype fruticose	[-0.003 — 0.337]
Area: Biotype crustose	[-0.043 — 0.003]
Area: Biotype fruticose	**[-0.068 — -0.013]**
*d' ~ reproductive mode*area*	
Intercept	**[0.384 — 0.656]**
Area	[-0.024 — 0.009]
Vegetative reproduction	**[0.038 — 0.223]**
Area: Vegetative reproduction	**[0.001 — 0.044]**

436 **Figure 4**. Representation of the interaction effect between area and reproduction type on ND
437 (A) and d'(B), as well as the interaction effect between area and biotype on ND (C) and d'(D).

439 Interaction turnover analyses revealed that most of the differences between forest patches

440 were due to the establishment of new interactions even though partners were present rather

441 than dissimilarity due to partner turnover (β_{ST}) (Figure. 5; t = 5.2741, df = 74.8, p-value<0.001).

Figure 5. Percentage of beta diversity of interactions explained by the dissimilarity of interactions established between species common to both communities (BOS) and due to species turnover (BST) (t-test: t = 5.2741, df = 74.8, p-value<0.001).

Discussion

Our study represents the first attempt to disentangle the effect of habitat loss on interaction

specialization in symbiotic organisms. The obtained results shed light on how habitat loss

interplays with life traits to affect fungal-algal interactions in epiphytic lichen symbioses.

Diversity

Mycobionts and photobiont richness showed a humped-shaped distribution along the patch size

gradient. This pattern is more pronounced for algae suggesting a higher sensitivity of algae to

abiotic conditions. Previous studies dealing with lichen diversity along gradients of habitat loss

in different forest ecosystems have reported a gradual loss of diversity as the available habitat

diminished (Svoboda et al., 2010), with changes in abiotic conditions associated to an increasing

fragment edge effect proposed as the main reason for such reduction (Asplund et al., 2014;

Belinchón et al., 2007; Boudreault et al., 2008; Esseen and Renhorn, 1998). Hump-shaped

diversity patterns have been reported for lichen-forming fungi along stand age (Asplund et al.,

2014; Miller et al., 2020) and elevational gradients (Nanda et al., 2021) so-far this is the first

report of a hump-shaped pattern of diversity along a fragmentation gradient. The intermediate

disturbance hypothesis (IDH) (Connell, 1978) predicts highest levels of biodiversity at

intermediate levels of frequency or intensity of disturbance (but see Mackey and Currie (2001)

and Fox (2013) for criticism). Although it has been proposed that the main driver for this peak

of diversity is the decrease of species densities, which entails reduced species competition (Fox

2013), it is not clear that this assumption may be applied to sessile organisms such as lichen-

forming fungi with high habitat specialization (Fritz et al., 2009). It could be that fragments of

intermediate size have intermediate abiotic conditions that allow the presence of habitat

specialist populations as well as species associated with the abiotic conditions existing in smaller

fragments. It is noteworthy that intermediate patch sizes dwell the largest number of exclusive

ASVs, a pattern not shared by mycobionts. Therefore, the hump-shaped pattern found in

mycobionts and photobionts may not respond to the same factors, or they may affect both

organisms differently. Intermediate abiotic conditions could be translated into a higher range of

available microhabitats, and higher spatial heterogeneity, promoting a larger amount of habitat

specialists and exclusive ASVs in photobionts (Rosenzweig, 1995). If mycobionts had broader

ecological requirements than photobionts, this would explain the difference in the observed

pattern between the two organisms.

Our results showed that a gradient of habitat availability influences the distribution of functional

traits in the studied lichen communities. Thus, except for crustose and foliose biotypes, the

number of fruticose, vegetative and sexually reproduced species changed along the gradient.

Changes in the distribution of functional traits in epiphytic lichen communities are directly

related to abiotic conditions such as solar radiation, water availability or air pollution (Giordani

et al., 2014; Koch et al., 2019; Matos et al. 2015; Paoli et al., 2017), and they change along

environmental gradients such as elevation (Bässler et al., 2016) or land use (Pinho et al., 2012).

Functional traits and specialization

Levels of specialization are affected by life traits in all known organisms (e.g. Armbruster, 2017;

Dehling et al., 2016). Our results showed differences in specialization among biotypes and

between reproductive modes, both in terms of ND and d'. Crustose species interacted on

average with a higher number of ASVs than fruticose and foliose species, although only the first

difference was significant. Regarding the parameter d', crustose species showed significantly

lower specialization than foliose and fruticose species. So far, no study had compared

differences in specialization among biotypes although most studies on photobionts associated

with crustose species have shown that they interact with a large number of species, in a wide

phylogenetic spectrum (Blaha et al., 2006; Guzow-Krzeminska, 2006; Muggia et al., 2014)

although examples of high specialization also occur (Hauck et al., 2007; Singh et al., 2015).

The interaction with a wider range of photobionts facilitates the settlement in a wider range of

substrates and ecological conditions (Muggia et al., 2014) since changes in photobiont allow

species to increase their ecological niche (Peksa and Škaloud, 2011; Fernández-Mendoza and

Printzen, 2013; Rolshausen et al., 2018). However, why most crustose lichens studied so far

show a higher number of associated photobionts still remains unclear. The constant contact with

the substrate during lichen growth mediated by prothallus ensures the constant contact with

new suitable free-living photobionts (Asmtrong and Bradwell, 2010; Sanders, 2005) that may be

incorporated into the lichen thallus in the course of development (Sanders and Lücking, 2002).

New photobionts might coexist with previous algal strains or replace them (Ott 1987). On the

contrary, chances to incorporate new algal strains in foliose and fruticose species are much more

reduced due to the scarce contact (single fixation points in fruticose species, rhizines in foliose

not adnate species) with substrate and global isolation due to cortical tissues. Molins et al.,

(2021) has recently shown that early developmental stages of *R. farinacea* show higher algal

diversity than older stages. This fact might indicate that, likewise it occurs in corals (Abrego et

al., 2009); it is in early stages of foliose or fruticose species when there is more intimate contact

with the substrate, therefore the mycobiont being able to establish contact with a wider array

of algal strains, although only one or a few of these will remain in the later stages.

The reproduction mode also showed a clear effect on mycobiont specialization. Sexual and

vegetatively reproduced species had on average a similar number of partners but the latter

showed considerably higher values of d'. Contrary to our findings, previous studies comparing

closely related species pairs had shown that species of lichen-forming fungi with clonal

reproduction via joint dispersal propagules tend to have lower associated algal diversity and are

more selective than sexual species (Cao et al., 2015; Otálora et al., 2013). The lower diversity

associated with species that reproduce by vegetative propagules has been attributed to the joint

dispersal of both bionts and the lack of need to search for new partners if the fitness of the

combination is optimal or suboptimal in the new environment. However, this interaction may

be broken and the mycobiont may interact with new photobionts in the new substrate, using

the original photobiont only for the settlement stage, thus avoiding the restriction to colonize

new environments. So far, no study had analyzed a high number of species to have a broad

overview. Thus, we showed that, on average, sexual and asexual species did not differ in terms

of the number of associated photosynthetic partners. Wornik and Grube (2010) showed that the

ease to switch photobiont in the vegetatively reproduced *Physconia grisea*, a Mediterranean

species dispersed via soredia and also found in our study area, was the only suitable explanation

for comparable photobiont diversity between a sexual and an asexual species. However,

regarding d' differences between both reproductive modes were significant and outstanding,

showing asexual species much higher specialization. Cao et al. (2015) suggested that the

decrease of selectivity in sexual species may be due to the need of finding a new photobiont

before the ascospores perish. In this case, the similar number of partners in both groups does

not point in that direction. The greater specialization found in vegetatively dispersing species

may reflect the higher fitness of a particular myco-photobiont combination at the local scale

which is favored by the clonal reproduction.

Specialization along the gradient of area size

Our results showed that average specialization at the species level changed along the gradient

of fragment size in the same way as occurs with other parameters that describe the structure of

interaction networks (Tylianakis and Morris, 2017). The variation of species specialization in such

gradients has only been scarcely studied (Aizen et al., 2012). Specialization is often considered

a species-level trait, conserved across the phylogeny (Gómez et al., 2010) and with relatively

little variation, although it is known to vary at geographical and phylogenetic scales (Poisot et

al., 2015; Trøjelsgaard et al., 2015). Carstensen et al. (2018) found that local and regional

specialization were strongly correlated in 14 plant-pollinator networks from two different

continents, considering species specialization consistent level across scales. On the contrary, we

found that average specialization may change due to habitat abiotic conditions, here derived

from habitat fragmentation. We observed a strong decrease in the average number of

photobionts associated with each fungal species as the fragment size decreased and an overall

higher specialization (*d'*) along the area gradient. The higher values of the normalized degree

found in smaller patch sizes regarding larger ones could be an artifact derived from the lower

number of photobionts available in small fragments (Fig. 2). However, the specialization

parameter *d'* did clearly point towards higher mycobiont specialization in larger fragments.

Although the number of available partners increased along most of the gradient, fungal species

become more selective. In addition, the analysis of the interaction beta diversity showed that

most of the turnover among forest patches was not due to changes in biont composition, that

is, the observed pattern of increasing specialization is not due to the absence of certain bionts,

but to an increase in the level of selectivity among the those available. Assuming a local

adaptation of the photobiont, the smaller fragments, with more homogeneous abiotic

conditions due to the greater edge effect (greater insolation, wind, nitrogen inputs, etc.) may

host only photobiont species that we could call weeds, adapted to the greater intensity of the

disturbance. New propagules arriving in the patch will have at their disposal only a pool of

equally suboptimal photobionts, perhaps interchangeable in terms of holobiont fitness, so the

degree of selectivity is low. On the contrary, in larger fragments, where a larger pool of

photobionts is available but also abiotic conditions are more heterogenous propagules of

different species will be able to establish more specialized interactions, thus increasing

holobiont fitness.

Interplay between specialization, traits and fragmentation

Changes in specialization along the area gradient were not even among morphological groups

and between the reproduction modes, showing that changes in specialization along the gradient

are also influenced by their life traits. For example, although the number of interacting

photobionts decreases along the area gradient in all biotypes, there was a significant difference

between how this number decreases in foliose and fruticose species and how it does in crustose

species. In addition, we found a significant interplay between the fruticose biotype and area,

regarding the specialization parameter d', showing that changes in habitat conditions and

availability do not affect similarly all biotypes in terms of discriminating among their

photosynthetic partners. It is known that different growth forms represent different strategies

of exploitation of water sources in lichens (Gauslaa, 2014). Thus, it is likely that the changing

abiotic conditions along the patch size gradient do not affect similarly photobionts occurring in

different growth forms since thallus biotypes may have different buffering potential of

environmental conditions.

Regarding reproductive modes, it is certainly striking that although the ND of both functional

groups similarly decreased along the area gradient, they showed contrasting trends regarding

the specialization parameter d' with sexual species decreasing in their specialization as the patch

area increases. The mode of reproduction has relevant implications with respect to constraints

on dispersal and colonization of new habitats (Ellis et al., 2021; Walser, 2004), thallus physiology

(Tretiach et al., 2005), and response to disturbance gradients such as air pollution (Koch et al.,

2019). The contrasting trends along the area gradient between reproductive modes in the

average d' may have their origin in differences in species composition and the distribution of

reproductive modes in the communities and also, in the different modulation of specialization

along the gradient at the individual level. Models indicated a significant change of reproduction

modes along the fragment size area. In addition, linear models for individual changes in

specialization along the gradient revealed five species in which species significantly modulated

their specialization in terms of ND (four of them asexually reproduced via soredia) and two cases

in which d' significantly increased along the gradient, both asexually reproduced via soredia.

Thus, the explanation of this contrasting pattern may be derived both from changes in species

composition and modulation of specialization in certain species. Modulation of specialization at

the species level is certainly a phenomenon that should be further explored in the future.

Implications for conservation

Our results have clear implications in conservation biology. Interactions among species are

certainly influenced by local abiotic conditions, so a certain biont combination would not have

the same fitness in two patches with different sizes. Small changes in forest conditions can have

drastic consequences and cause existing biont combinations to lose their ability to compete. In

addition, if some species can modulate their specialization to a certain degree, then those

species would display higher resilience to environmental changes such as those derived from

habitat fragmentation than other species. Their ability to adapt to changes in local conditions

through changes in specialization towards their photobionts may buffer the effect of more

stressful abiotic conditions due to the increase of edge effect in small fragments, at least until

phylogenetic constraints prevent interactions in some species. This buffering effect produced by

the modulation of specialization may postpone the extinction debt (Berglund and Jonsson,

2005), or, depending on the intensity and frequency of perturbation, even avoid extinction, with

remarkable implications for biodiversity conservation (Kuussaari et al., 2009). Further studies

will show whether the analyses of interaction dynamics (partner fidelity, link conservatism and

rewiring) in myco-photobiont symbioses in the context of interaction bipartite networks may be

used as suitable tools to understand and predict long-term changes in epiphyte lichen

communities.

Author contributions

SPO, MV, MVdP and ABdG; SPO and ABdG designed the study; SPO and ABdG carried out the

field sampling; ABdG and MVdP collected laboratory data; MV, SPO, ABdG, MVdP and MV

analyzed the data; SPO and ABdG led the writing of the manuscript; All authors contributed

critically to the drafts and gave final approval for publication.

Data availability statement

All NGS sequences obtained in this study will be available in the SRA (NCBI). Sequences obtained

through Sanger sequencing are available in Gen Bank (Accession numbers provided in

Supplementary Material Table 2).

Conflict of interest statement

The authors have no conflict of interests to declare.

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