Lethal and sub-lethal concentrations of pesticides detected in milkweeds purchased from

nurseries across the United States

Christopher A. Halsch[1], Sarah M. Hoyle[2], Aimee Code[2], James A. Fordyce[3], and Matthew

L. Forister[1]

[1]Department of Biology, Program in Ecology, Evolution and Conservation Biology, University

of Nevada, Reno, NV, U.S.A.

[2]Xerces Society for Invertebrate Conservation, Portland, OR, U.S.A.

[3]Department of Ecology & Evolutionary Biology, University of Tennessee, Knoxville, TN,

U.S.A.

* **Correspondence:**

Christopher Halsch

cahalsch@nevada.unr.edu

Keywords: Monarch, milkweed, pesticides, non-target insects, nurseries, butterflies

Abstract

The decline of monarch butterflies in both the eastern and western United States has garnered

widespread public interest. Planting milkweed has been promoted as one action that individuals

can take, but very little is known with respect to potential pesticide contamination of store-

bought milkweeds during the process of production and transport to market. In this study, we

collected 235 milkweed leaf samples from 33 retail nurseries across the US to screen for

pesticides. Across all samples, we detected 61 different pesticides with an average of 12.2

compounds per leaf. While most of the compounds have unknown effects on caterpillars, 48

samples contained a pesticide at a concentration that exceeds a published lepidopteran LD_{50} and

89 contained a pesticide above a concentration shown to have a sub-lethal effect for a monarch.

We detected only modest effects of store size and milkweed species; however, plants with labels

advertising their value for wildlife were more likely to contain pesticides at concentrations

known to have negative effects on monarchs. These results demonstrate the extensiveness of

pesticide exposure within nursery milkweeds and the potential impacts on monarchs and other

insects consuming store-bought plants.

Introduction

Insect populations are facing an expansive and interacting set of stressors [1–3]. Among native insects declining in the US, the monarch (*Danaus plexippus*) is a widely recognized butterfly whose declines have been substantial in both the eastern and western populations [4–6]. The causes of these declines are complex and region-specific, but proposed hypotheses implicate climate change, the loss of overwintering habitat, natural enemies, and pesticides, especially herbicides affecting the abundance of native milkweeds (*Asclepias sp.*). [7–10]. Many monarch conservation strategies have been suggested, from changes in agricultural practices to the planting of milkweeds by individuals in gardens and yards [11]. For people wanting to help imperiled insects, planting beneficial plants is a seemingly simple action, however the practices used to bring nursery plants to shelf often involve pesticide treatments [12,13]. A recent study found pesticides in milkweeds growing across diverse landscapes in the Central Valley of California, including up to 31 compounds in individual retail plants [14]. Milkweeds in that study were purchased from only two nurseries in one metropolitan area, thus one of the conclusions was that retail locations required further investigation.

In this study, we address the need to better understand contamination of retail plants by quantifying the concentrations of pesticides (insecticides, fungicides, and herbicides) found in the leaves of milkweed plants sold in nurseries across the United States. Stores ranged from local nurseries to large national chains and were all in areas where monarchs breed. First, we present an exploratory analysis of the detected compounds, including a search of published toxicological data for lethal effects of those compounds on lepidopterans. Second, we examine associations between observed pesticide concentrations and factors relevant to a milkweed buyer. Finally, we offer suggestions for buyers interested in planting milkweeds for monarch conservation.

Methods

Sample collection

Five species of milkweed plants were purchased from 33 retail stores across 15 states from May 15 to June 29, 2021 (Table S1). Whenever possible, five plants of each type (species x supplier) were purchased (fewer plants were available in two places). These collections were intended to represent a snapshot of milkweed plants available to the public across the monarch's migratory range. After purchase, collectors clipped at least 5 grams of leaves from an individual plant, and wrapped samples in tin foil stored in sealed plastic bags. Clippers were cleaned with soap and water or rubbing alcohol between samples. Samples were shipped with ice packs to a central location for temporary storage in a freezer, and ultimately shipped on dry ice to the lab for chemical analysis. In one instance, 10 leaf samples were collected immediately after purchase, and another 10 were collected after growing the plants outdoors in pots with daily watering for two weeks (these additional samples were excluded from the primary analysis but discussed below).

Chemical analysis

Using methods similar to previously mentioned milkweed study [14], frozen milkweed leaves were extracted by a modified version of the EN 15662 QuEChERS procedure [15] and screened for 92 pesticides by liquid chromatography mass spectrometry (LC-MS/MS). Sample analysis was carried out with a UHPLC system coupled with a TSQ Quantis mass spectrometer. MS/MS detection was carried out using the selected reaction monitoring (SRM) mode. Additional details including a full table of retention times and optimized SRM acquisition parameters for pesticides and internal standards can be found in the supplement (Table S2).

Literature search

To gain insight into potential biological impacts of observed concentrations, we collected

published oral and contact LD_{50} values for honeybees from EPA records in the ECOTOX and

PubChem databases as well as from the University of Hertfordshire's Pesticide Properties

Database (Table S3). Honeybee LD_{50} values are widely used to estimate risk to terrestrial

invertebrates, however, adult bee and caterpillar physiology are of course different, and we only

use these values in a heuristic sense. To find the most directly relevant data, we also conducted a

literature search for bioassay studies that quantified LD_{50} values on caterpillars. For studies that

specifically looked at monarchs, we also made note of any reported sub-lethal effects. This was

done in January of 2022 for all compounds identified in our samples using ISI Web of Science

and the terms (lepidopt* OR butterfl* OR moth*) (Table S4).

Statistical analysis

To examine relationships between pesticides and explanatory variables, we modeled pesticide

richness, diversity, and the number of exceedances of a published lethal concentration in

lepidoptera (including monarchs), or sub-lethal concentration in monarchs. Diversity was

represented as the effective number of compounds by taking the exponential of the Shannon

diversity index [16]. Exceedance was treated as a binary category: either a sample did or did not

contain a pesticide at or above a concentration known to exhibit lethal or sub-lethal effects. The

predictor variables we explored were store size, milkweed species, region (eastern or western

US), and whether the plant contained a label at the point of purchase with information or a claim

about its value for wildlife (e.g. "attracts butterflies"). Pesticide richness, diversity, and the

number of exceedances were each modeled using a generalized mixed effects model with a

Poisson error structure (log link), a Gaussian error structure, and a binomial error structure (logit

link) respectively. The individual store the plants were purchased from was included as a random

intercept to account for store-specific effects not encompassed by other explanatory variables.

We could not separately evaluate the identity of suppliers because in most cases wholesalers

provided plants to a single store.

Random effects models were built in R using the lme4 package [17] and variance partitioning

was performed using the partR2 and modEvA packages [18,19]. We also explored compositional

differences using principal coordinates analysis, but the results added little to the overall picture

of contamination and are not discussed further but are included in supplementary material (Fig

S1). Finally, to investigate the importance of wildlife labels, we used a random forest classifier

with the presence of a label as the response and the presence of each compound as predictor

variables (using the randomForest package [20]).

Results

We detected 61 unique compounds out of a possible 92 in the test panel: 24 insecticides, 26

fungicides, 10 herbicides, and 1 synergist (Fig. 1). The fungicides azoxystrobin, metalaxyl, and

difenoconazole were the most common, found in over 75 percent of samples. We obtained

honeybee contact and oral LD_{50} values for 59 of the 61 compounds, and lepidoptera LD_{50} values

for 15 of the 61 compounds (from 69 studies). We found that 72 of the 235 samples contained a

pesticide at a concentration that exceeded a honeybee contact or oral LD_{50}. These were

associated with ten compounds and with 13 of the 25 locations (towns or cities) (Table S5). We

found that 48 of the 235 samples contained a pesticide at a concentration that exceeded a

published lepidopteran LD_{50}, coming from 7 locations and involving five compounds (Fig. 1,

Table S6). We also detected exceedances of published monarch sub-lethal effects in 89 samples,

driven by azoxystrobin, clothianidin, and trifloxystrobin, occurring in 18 of 25 locations (Table

S7).

Milkweed species and store size explained the greatest variation in pesticide richness, and

store size and the presence of wildlife-friendly labels explained the largest amount of variation in

pesticide diversity (Table 1, Fig. S2). Specifically, larger stores were associated with higher

richness ($z = 2.644$, $p = 0.01$, Table 2, Table S8, Fig. S3) and higher diversity ($t_{23.1} = 2.764$, $p =$

0.01, Table 2, Table S9, Fig. S4). The presence of a wildlife friendly label was associated with

lower diversity, likely due to lower evenness across compounds ($t_{30.8} = -2.707$, $p = 0.01$, Table 2,

Table S9, Fig. S4). While pesticide richness and diversity are one aspect of contamination, these

metrics are not sufficient for understanding the realized effects of these pesticides on caterpillars.

When considering the number of exceedances of a lethal or sub-lethal concentration, we found

that milkweed species and the presence of wildlife labels explained the most variation (Table 1,

Fig. S2). Most notably, plants with a label claiming the plant's value for wildlife had an

increased chance of exceeding a lethal or sub-lethal concentration of at least one pesticide ($z =$

2.262, $p = 0.02$, Fig. 2, Table S10). Random forest analysis identified azoxystrobin and

trifloxystrobin among the most predictive of whether a sample was labeled, both of which were

often detected above the sub-lethal threshold in labeled plants (Fig. 2).

Discussion

Monarch butterflies are in decline in both the eastern and western United States, leading to an

upswell in conservation efforts, including planting milkweeds in gardens and yards. We detected

pesticides in milkweeds bought from nurseries in regions of the US where monarchs breed. The

number of compounds and their concentrations were highly variable, however many plants

contained pesticides at levels known to have deleterious effects on insects. Plants from larger

stores contained higher numbers of pesticides (but not necessarily more lethal or sub-lethal

concentrations), and most staggeringly, plants with a label referring to its value for wildlife were

more than twice as likely to contain at least one pesticide at a level potentially dangerous to

monarchs.

Pesticides are also known to have sub-lethal effects that can affect survival and reproduction

[21]. In monarchs, sub-lethal effects are under-studied, but a few have been identified [22,23].

Pecenka and Lundgren found that clothianidin as low as 1ppb, a concentration we observed in 5

samples from 2 states [22], can reduce larval size. Olaya☐Arenas et al. investigated the impacts

of six compounds, all of which we detected, and found that azoxystrobin and trifloxystrobin both

reduce adult monarch wing size [23]. The concentrations that caused these effects in their study

were exceeded 73 times by azoxystrobin and 22 times by trifloxystrobin in our samples.

Azoxystrobin and trifloxystrobin were more common in samples labeled as valuable for wildlife

and were the primary drivers of higher exceedances in those samples compared to those that

were unlabeled. This pattern of use might be explained by the fact that these compounds are

fungicides and are not used to target caterpillars, but fungicides do have direct impacts on

monarchs and other insects [23–27]. Critically, most of the compounds we detected (including

some of the most prevalent) have not been directly tested on monarchs, so harmful levels of

contamination are likely underestimated and may be widespread in nursery plants.

In addition to exceedances of lethal and sub-lethal concentrations, there are other findings in

these results worth discussing. The herbicide atrazine was found in 62% of samples. While one

study did not find it to exhibit effects on monarchs [23], its detection frequency poses the

question of how these plants became exposed in the first place. In one case, plants that were

certified organic still contained atrazine, raising the possibility of environmental conditions in

addition to nursery practices being a source of contamination. Another noteworthy result is the

potential for additive and synergistic effects from mixtures of compounds. On average, plants

contained 12.2 different pesticides and some of these pesticides have been shown to interact to

increase lethality [28–30]. Interestingly, Olaya☐Arenas et al. also examined mixtures and did not

identify lethal or sub-lethal effects [23], but it is unclear how these findings, pertaining to a

mixture treatment containing 6 compounds (with 1 insecticide), might apply to our samples

which contained, on average over 12 compounds including multiple insecticides. Finally, in one

instance leaves from the same plants were screened at two different time points, immediately

after purchase and two weeks later, to gain some insight into pesticide degradation over time.

Although sample sizes were too small for statistical analyses, we observed reductions in the

concentrations of some pesticides over this time. Compounds like spinosad and acephate which

have short half-lives saw the greatest reduction, while other compounds like azoxystrobin (which

has a longer half-life) did not change (Table S11, Fig. S5).

What should the average person do if they want to support monarch conservation by planting

milkweed? We recommend that plants be purchased from nurseries that implement a robust

approach to minimizing their reliance on pesticides. This extends beyond insecticides, as

fungicides can also have negative effects on monarch caterpillars and were ubiquitous in our

samples. Consumers should ask retailers to source plants grown using ecologically sound pest

management strategies [31,32]. On the regulatory front, the U.S. Environmental Protection

Agency could take steps to address risk, including reducing permissible nursery application rates

to prevent residues toxic to pollinators. Finally, further work could investigate sources of

contamination in nursery plants, which may become contaminated at multiple points from

production to sale. The monarch is the most famous declining butterfly in the US, but it is one of

many, and may not even be the most dire case [33]. The planting of beneficial plants can help

imperiled insects, yet, for small scale insect conservation efforts like native plant gardens to be

effective, it is critical that nurseries provide plants free from harmful pesticide residues.

Acknowledgements

We thank all the volunteers and Xerces staff who purchased milkweeds; Wayne Anderson
(Cornell Chemical Ecology Core Facility) for performing the chemical analysis; and Sharon
Selvaggio for discussion of recommendations.
199

Contribution of authors

C.H.: data curation, formal analysis, methodology, visualization, writing – original draft, writing
– review and editing; S.H.: data curation, project administration, funding acquisition, writing –
review and editing; A.C.: funding acquisition, writing – review and editing; J.F.: methodology,
writing – review and editing; M.F.: methodology, supervision, visualization, writing – review
and editing.
206

207   **Funding information**
208   This work was funded through the generous support of Linda S. Raynolds, who donated to the
209   Xerces Society. CH was supported by a National Institute of Food and Agriculture fellowship
210   (NEVW-2021-09427), and MF acknowledges National Science Foundation support (DEB-
211   2114793).
212

Competing interests

The authors declare no competing interests.

Data availability

Data will be made available on dryad when the paper is accepted for publication.

Supporting Information

Full description of chemical analysis.
221
222   Table S1. Summary of purchased milkweeds.
223
224   Table S2. Retention times and optimized SRM acquisition parameters for pesticides and internal
225   standards (RT: Retention time, CE: Collision Energy)
226
227   Table S3. Contact and oral LD_{50} data for honey bees.
228
229   Table S4. Studies from Lepidoptera literature review.
230

References:

1. Wagner DL, Grames EM, Forister ML, Berenbaum MR, Stopak D. 2021 Insect decline in the Anthropocene: Death by a thousand cuts. *Proc. Natl. Acad. Sci.* **118**. (doi:10.1073/PNAS.2023989118)

2. Habel JC, Samways MJ, Schmitt T. 2019 Mitigating the precipitous decline of terrestrial European insects: Requirements for a new strategy. *Biodivers. Conserv.* **28**, 1343–1360. (doi:10.1007/s10531-019-01741-8)

3. Sánchez-Bayo F, Wyckhuys KAG. 2019 Worldwide decline of the entomofauna: A review of its drivers. *Biol. Conserv.* **232**, 8–27. (doi:10.1016/j.biocon.2019.01.020)

4. Espeset AE, Harrison JG, Shapiro AM, Nice CC, Thorne JH, Waetjen DP, Fordyce JA, Forister ML. 2016 Understanding a migratory species in a changing world: climatic effects and demographic declines in the western monarch revealed by four decades of intensive monitoring. *Oecologia* **181**, 819–830. (doi:10.1007/s00442-016-3600-y)

5. Pelton EM, Schultz CB, Jepsen SJ, Black SH, Crone EE. 2019 Western Monarch Population Plummets: Status, Probable Causes, and Recommended Conservation Actions. *Front. Ecol. Evol.* **7**. (doi:10.3389/fevo.2019.00258)

6. Agrawal AA, Inamine H. 2018 Mechanisms behind the monarch's decline. *Science (80-.).* **360**. (doi:10.1126/science.aat5066)

7. Zylstra ER, Ries L, Neupane N, Saunders SP, Ramírez MI, Rendón-Salinas E, Oberhauser KS, Farr MT, Zipkin EF. 2021 Changes in climate drive recent monarch butterfly dynamics. *Nat. Ecol. Evol. 2021 510* **5**, 1441–1452. (doi:10.1038/s41559-021-01504-1)

8. Crone EE, Pelton EM, Brown LM, Thomas CC, Schultz CB. 2019 Why are monarch butterflies declining in the West? Understanding the importance of multiple correlated

drivers. *Ecol. Appl.* **29**. (doi:10.1002/eap.1975)

9.   Thogmartin WE *et al.* 2017 Monarch butterfly population decline in North America:

identifying the threatening processes. *R. Soc. Open Sci.* **4**. (doi:10.1098/RSOS.170760)

10.   Stenoien C, Nail KR, Zalucki JM, Parry H, Oberhauser KS, Zalucki MP. 2018 Monarchs

in decline: a collateral landscape-level effect of modern agriculture. *Insect Sci.* **25**, 528–

541. (doi:10.1111/1744-7917.12404)

11.   Thogmartin WE *et al.* 2017 Restoring monarch butterfly habitat in the Midwestern US:

'all hands on deck'. *Environ. Res. Lett.* **12**, 074005. (doi:10.1088/1748-9326/AA7637)

12.   Lentola A, David A, Abdul-Sada A, Tapparo A, Goulson D, Hill EM. 2017 Ornamental

plants on sale to the public are a significant source of pesticide residues with implications

for the health of pollinating insects. *Environ. Pollut.* **228**, 297–304.

(doi:10.1016/J.ENVPOL.2017.03.084)

13.   Krischik V, Rogers M, Gupta G, Varshney A. 2015 Soil-applied imidacloprid translocates

to ornamental flowers and reduces survival of adult coleomegilla maculata, harmonia

axyridis, and hippodamia convergens lady beetles, and larval danaus plexippus and

vanessa cardui butterflies. *PLoS One* **10**. (doi:10.1371/journal.pone.0119133)

14.   Halsch CA, Code A, Hoyle SM, Fordyce JA, Baert N, Forister ML. 2020 Pesticide

Contamination of Milkweeds Across the Agricultural, Urban, and Open Spaces of Low-

Elevation Northern California. *Front. Ecol. Evol.* **8**. (doi:10.3389/fevo.2020.00162)

15.   Standardization EC for. 2008 Foods of plant origin-Determination of pesticide residues

using GC-MS and/or LC-MS/MS following acetonitrile extraction/partitioning and clean-

up by dispersive SPE. *QuEChERS-method. EN* **15662**, 2008.

16.   Jost L. 2006 Entropy and diversity. *Oikos* **113**, 363–375. (doi:10.1111/j.2006.0030-

306   1299.14714.x)

307 17. Bates D, Maechler M, Bolker B, Walker S. 2020 lme4: Linear Mixed-Effects Models

308   using Eigen and S4.

309 18. Stoffel MA, Nakagawa S, Schielzeth H. 2021 partR2: Partitioning R2 in generalized linear

310   mixed models. *PeerJ* **9**, e11414. (doi:10.7717/PEERJ.11414/SUPP-1)

311 19. A.M. B, J.A. B, A. J-V, R. R. 2021 modEvA: Model Evaluation and Analysis.

312 20. Breiman L, Cutler A, Liaw A, Wiener M. 2018 randomForest: Breiman and Cutler's

313   Random Forests for Classification and Regression.

314 21. Desneux N, Decourtye A, Delpuech J-M. 2006 The Sublethal Effects of Pesticides on

315   Beneficial Arthropods. *Annu. Rev. Entomol.* **52**, 81–106.

316   (doi:10.1146/annurev.ento.52.110405.091440)

317 22. Pecenka JR, Lundgren JG. 2015 Non-target effects of clothianidin on monarch butterflies.

318   *Sci. Nat.* **102**. (doi:10.1007/s00114-015-1270-y)

319 23. Olaya-Arenas P, Hauri K, Scharf ME, Kaplan I. 2020 Larval pesticide exposure impacts

320   monarch butterfly performance. *Sci. Reports 2020 101* **10**, 1–12. (doi:10.1038/s41598-

321   020-71211-7)

322 24. Su J, Wang YC, Zhang SK, Ren XB. 2014 Antifungal agents against aspergillus niger for

323   rearing rice leaffolder larvae (Lepidoptera: Pyralidae) on artificial diet. *J. Econ. Entomol.*

324   **107**. (doi:10.1603/EC13296)

325 25. Bernauer OM, Gaines-Day HR, Steffan SA. 2015 Colonies of Bumble Bees (Bombus

326   impatiens) Produce Fewer Workers, Less Bee Biomass, and Have Smaller Mother Queens

327   Following Fungicide Exposure. *Insects 2015, Vol. 6, Pages 478-488* **6**, 478–488.

328   (doi:10.3390/INSECTS6020478)

329   26.   Zhu W, Schmehl DR, Mullin CA, Frazier JL. 2014 Four common pesticides, their

330   mixtures and a formulation solvent in the hive environment have high oral toxicity to

331   honey bee larvae. *PLoS One* **9**. (doi:10.1371/journal.pone.0077547)

332   27.   Tsvetkov N, Samson-Robert O, Sood K, Patel HS, Malena DA, Gajiwala PH,

333   Maciukiewicz P, Fournier V, Zayed A. 2017 Chronic exposure to neonicotinoids reduces

334   honey bee health near corn crops. *Science (80-.).* **356**, 1395–1397.

335   (doi:10.1126/SCIENCE.AAM7470/SUPPL_FILE/AAM7470_TABLES5.XLSX)

336   28.   Jones MM, Robertson JL, Weinzierl RA. 2012 Toxicity of Thiamethoxam and Mixtures

337   of Chlorantraniliprole Plus Acetamiprid, Esfenvalerate, or Thiamethoxam to Neonates of

338   Oriental Fruit Moth (Lepidoptera: Tortricidae). *J. Econ. Entomol.* **105**, 1426–1431.

339   (doi:10.1603/ec11349)

340   29.   Liu Y, Zhang H, He F, Li X, Tan H, Zeng D. 2018 Combined toxicity of

341   chlorantraniliprole, lambda-cyhalothrin, and imidacloprid to the silkworm Bombyx mori

342   (Lepidoptera: Bombycidae). *Environ. Sci. Pollut. Res.* **25**, 22598–22605.

343   (doi:10.1007/s11356-018-2374-7)

344   30.   Chen J, Jiang W, Hu H, Ma X, Li Q, Song X, Ren X, Ma Y. 2019 Joint toxicity of

345   methoxyfenozide and lufenuron on larvae of Spodoptera exigua Hübner (Lepidoptera:

346   Noctuidae). *J. Asia. Pac. Entomol.* **22**, 795–801. (doi:10.1016/j.aspen.2019.06.004)

347   31.   Selvaggio S, Code A. 2020 Buying Bee-Safe Plants.

348   32.   Selvaggio S, Code A. 2020 Offering Bee-Safe Plants: A Guide for Nurseries.

349   33.   Forister ML *et al.* 2021 Fewer butterflies seen by community scientists across the

350   warming and drying landscapes of the American West. *Science (80-.).* **371**, 1042–1045.

351   (doi:10.1126/science.abe5585)

Table 1. Variance explained by covariates predicting pesticide richness, pesticide diversity, and the presence of a lethal or sub-lethal exceedance. The total explained variance is the explained variance of the model when including all terms and the covariate specific rows are the semi-partial variance explained by each term.

	Pesticide richness	Pesticide diversity	Exceedance
Total explained variance	20.5%	30.0 %	33.5 %
Store size	9.8%	3.9%	1.0 %
Milkweed species	9.5 %	7.9%	13.2%
Region	0.5%	0%	0.0%
Label	0%	17.2%	25.4%

Table 2. Means and standard errors of pesticide richness and pesticide diversity for each covariate level.

Covariate		Pesticide richness	Pesticide diversity
Store size	Single store	11.25 (± 0.39)	2.98 (± 0.13)
	2-100 stores	11.85 (± 0.58)	2.94 (± 0.18)
	> 100 stores	16.43 (± 0.83)	4.02 (± 0.32)
Milkweed species	*Asc. curassavica*	12.18 (± 0.59)	2.69 (± 0.17)
	Asc. fascicularis	13.50 (± 0.67)	4.53 (± 0.73)
	Asc. incarnata	12.03 (± 0.59)	3.58 (± 0.22)
	Asc. speciosa	9.12 (± 0.68)	2.70 (± 0.37)
	Asc. tuberosa	12.92 (± 0.65)	3.13 (± 0.17)
Region	East	12.32 (± 0.40)	3.04 (± 0.12)
	West	12.04 (± 0.52)	3.23 (± 0.18)
Label	Yes	13.33 (± 0.49)	2.77 (± 0.14)
	No	10.64 (± 0.84)	3.79 (± 0.20)

Figure 1. Pesticides found in milkweed samples. A) Locations where milkweeds were purchased.
Orange arrows denote major monarch migration routes. B) Adult monarch butterfly and
caterpillar on *Asclepias tuberosa*. C) Mean concentrations of compounds by location. Values are
shown in parts per billion on a log scale. White circles indicate compounds found above a
lepidopteran LD_{50}. Black circles indicate compounds above a monarch sub-lethal concentration.

Figure 2. Summary of exceedance associations. Panels A-D show the percentage of plants from each store with at least one exceedance by store size, species, region, and wildlife label. Gray points show the mean percentage (bars are standard errors). E) Compounds that are the most predictive of whether a sample was labeled safe for wildlife based on random forest analysis. F)

371   Differences in the total number of detections between labeled and un-labeled samples for the
372   three most predictive compounds.
373