Factors determining distributions of rainforest *Drosophila* shift from

interspecific competition to high temperature with decreasing elevation

Running title: Biotic and abiotic effects on distributions

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Author Contributions:

JC and OTL both contributed to the development of ideas. JC designed and

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manuscript. OTL contributed to the writing.

Funding:

This work was supported by NERC grant NE/N010221/1 to OTL and a tuition

grant from the China Scholarship Council to JC.

Conflict of Interest:

There is no conflict of interest to declare.

Data Accessibility:

All original data are achieved on Zenedo (10.5281/zenodo.6400611) and the code of

analysis is available on GitHub (https://github.com/Jinlinc/DistributionRegulators.git).

Acknowledgements:

We thank the laboratories of Jan Hrček (Czech Academy of Sciences) and

Megan Higgie (James Cook University, Townsville) for their assistance in

establishing *Drosophila* laboratory cultures. Jan Hrček, Chia-Hua Lue, Nick Pardikes,

Mélanie Thierry and the Oxford Fly group provided valuable advice and shared

facilities. We thank Chris Terry, Mélanie Thierry, Benjamin Van Doren, Eleanor

O'Brien and Mukhlish Jamal Musa Holle for advice on data analysis and comments

on the manuscript.

Abstract:

Species turnover with elevation is a widespread phenomenon and provides valuable information on why and how ecological communities might reorganize as the climate warms. Tropical mountains typically have pronounced thermal gradients and intense species interactions, providing a testing ground for investigating the relationship between thermal tolerances and biotic interactions as the proximate factors determining species' distributions. We investigated thermal tolerances and interspecific competition as causes of species turnover in the nine most abundant species of *Drosophila* along elevational gradients in the Australian Wet Tropics. Upper thermal limits varied less among species than lower thermal limits; nonetheless, at low elevation sites, these small differences determined the composition of communities through environmental sorting. In contrast, community composition at cool, high elevations was driven by interspecific competition rather than tolerance to low temperatures. These results run counter to common assumptions about the role of abiotic and biotic factors in structuring communities along thermal gradients, and indicate that tropical insects may be vulnerable to future warming wherever their distributions lie along elevational and other temperature gradients.

Keywords:

Species distribution, climate change, *Drosophila*, tropics, thermal tolerance, thermal performance curve, biotic competition, coexistence.

Introduction

Temperature has a fundamental impact on the reproduction, survival, growth, and behaviour of organisms (Huey and Kingsolver 1989; Huey and Stevenson 1979), strongly influencing species' ranges and abundances (Hoffmann and Blows 1994; Wilson et al. 2005). As a result, estimates of thermal tolerances based on laboratory assays or species' distributions have been used widely to evaluate species' sensitivity to climate change (Deutsch et al. 2008; Kearney and Porter 2009). Tropical ecosystems comprise many species that live close to their upper thermal limits (Deutsch et al. 2008; Diamond et al. 2012; Huey et al. 2009), and which may not tolerate or adapt to warmer temperatures (Bonebrake and Deutsch 2012; Kellermann et al. 2012). The narrow thermal ranges of tropical insects (Khaliq et al. 2014) also mean that they will need to undertake relatively large latitudinal or elevational range-shifts to track their climate envelopes as the climate warms, increasing the risk of extinction and community disassembly (Colwell et al. 2008; Sheldon, Yang, and Tewksbury 2011).

Despite high relevance for understanding species' responses to climate change in tropical ecosystems (Corlett 2012), it remains questioned whether temperature is the major proximate factor setting the position of the 'warm' (low-latitude or low-elevation) limits of species' distributions. Global analyses found smaller distribution shifts in response to warming at warm range limits than at cool range limits (Chen, Hill, Shiu, et al. 2011; Sunday, Bates, and Dulvy 2012), and some laboratory assays have found similar upper thermal limits for tropical and temperate species (MacLean et al. 2019; Nowrouzi et al. 2018; Overgaard, Kearney, and Hoffmann 2014). Instead, other abiotic factors (e.g. precipitation) or biotic

interactions may be critical in determining range limits in the tropics (Engelbrecht et

al. 2007; Louthan, Doak, and Angert 2015).

A common expectation, first expressed by Charles Darwin, is that tolerance to

low temperatures will set cool range limits, while biotic interactions will more often

set warm range limits (O'Brien et al. 2017; Paquette and Hargreaves 2021).

However, a recent analysis of data for 654 taxa found that biotic factors became less

important in determining warm limits towards the tropics, while abiotic factors

remained consistently important across latitudes (Paquette and Hargreaves 2021),

highlighting the regional differences in the relative roles of abiotic and biotic factors

on local thermal gradients.

There remains limited empirical evidence on the proximate factors setting

range limits in species-rich tropical communities (Feeley, Stroud, and Perez 2017;

Jankowski et al. 2013), and most such studies examine only the correlation between

thermal traits (such as critical temperatures and optimal temperatures) and species'

distributions (Cahill et al. 2014). A significant correlation between tolerance and

distribution does not rule out a role for biotic interactions, which are themselves

sensitive to temperature (Gilman et al. 2010). Additionally, the abundance and

distribution of a species may still be sensitive to temperature change, even when

biotic interaction is the proximate factor determining species composition. To

understand the relationship between temperature and distribution, studies

quantifying both thermal traits and temperature-dependent biotic interactions are

required (Cahill et al. 2014). Such studies can help unify the long-separate concepts

of environmental and biotic filters (HilleRisLambers et al. 2012), as well as providing

practical information to inform conservation efforts in the face of climate change.

To investigate the roles of thermal tolerances and biotic interactions in

determining species' range limits and structuring communities, we focused on the

community of *Drosophila* flies occupying mountain rainforest habitats in north-

eastern Australia. These and other tropical mountains provide natural environmental

gradients to test the sensitivity of species to temperature (Corlett 2011), with

pronounced changes in species composition with elevation for many taxa (Williams,

Bolitho, and Fox 2003). We hypothesized that species turnover along the elevational

gradient would result from thermal constraints at cool, high elevation sites, but from

competitive exclusion at warmer, lowland sites. Thus, we predicted that cold-tolerant

species would become more abundant at higher elevations. We also predicted that

species with reduced abundance at low elevations would be those limited by

competition with species with lowland-biased distributions, and not necessarily those

with lower heat tolerance. To test these predictions, we first examined correlations

between species' distributions and multiple thermal tolerance traits. We then used

laboratory experiments mimicking upland and lowland thermal conditions to examine

short-term and long-term competitive outcomes for pairs of species with similar or

different distribution types.

Methods

1. Species distributions along elevational gradients

Field survey. Field data on *Drosophila* communities were collected from rainforest

sites at Paluma Range (S18° 59.031' E146° 14.096') and Kirrama Range (S18°

12.134' E145° 53.102'), Queensland, Australia. These sites lie within the Wet Tropics

bioregion which has high levels of endemism associated with cool and moist upland

refugia (Williams et al. 2003). Abundances of *Drosophila* at these sites peak during

the period March to June. *Drosophila* pupae were sampled using bottle traps baited

with fermented banana from 11th March – 12th April 2016 for three sites at elevation

of 70m, 350-390m, and 730-880m (subsequently referred to as low-, mid- and high-

elevation, respectively) on the two mountains. 182 pupae were sampled at each site

and 716 pupae in total were successfully identified to species by DNA

metabarcoding (Jeffs et al. 2021), with 86 – 134 pupae at each site. Two infrequent

species, *Drosophila serrata* (1 sample) and *D. immigrans* (4 samples), were

excluded from analyses.

Distribution analysis. Two methods were used to assess trends in relative

abundance with elevation for individual species. First, the probability of detecting a

particular *Drosophila* species among other co-occurring *Drosophila* species was

modeled as a function of *elevation*, *transect* and their interaction in a generalized

linear model (family = "binomial") using the *stats* package in R. For each species, the

response variable was 1 if the pupa was identified as the focal species and 0 if it was

identified as any other species. Second, since none of the species had a bimodal

distribution with elevation, abundance-weighted mean elevation (*hIndex*) was used

as a simple alternative method to quantify distributions. The location of each sample

was assigned a value of 0, 0.5, or 1 if it was collected at low-, mid- and high-

elevation sites, respectively. *hIndex* was calculated for each species by averaging

these values for samples from both transects. Correspondence between the two

measures was tested by Spearman's rank correlation test between the coefficient

value for *elevation* in the regression and *hIndex* across species.

2. Maintenance of laboratory culture

Drosophila isofemale cultures were established in 2017 and 2018 from adults

collected from high- and low-elevation sites. Cultures were maintained initially at

24°C at the Biology Centre, Czech Academy of Sciences, and then transferred to the

Department of Zoology, University of Oxford, UK, in December 2018, where they

were maintained at 25°C. Cultures and all experiments mentioned below were

maintained under a 12h/12h light/dark cycle. There were approximately 15 to 30

non-overlapping generations in the Czech Republic and four to seven non-

overlapping generations in Oxford before cultures were used to establish mass bred

lines (MBLs).

To revive genetic variation, we made MBLs of each *Drosophila* species by

combining four isofemale lines (except for *D. pandora*, where only three isofemale

lines were available). The four lines were selected from different mountains and

different elevations if possible (detailed information in Supplementary Table 1). Each

mass bred population was initiated using two independently-reared MBLs of the

same species. Large populations were maintained at 25°C for more than four

generations before measuring the thermal traits. The MBLs were maintained at 25°C

until 2020 and at 23°C thereafter. In this way, measurements should not have been

influenced by maternal effects, acclimation, or isofemale line effects.

Nine tropical *Drosophila* species (listed in Supplementary Table 1) were included

in laboratory thermal trait measurements. Of the available species, only *D. rubida*

was not included because it was difficult to raise in large numbers and to

synchronize with the other species. Isofemale lines of *D. pseudotakihashii* were

contaminated by another species before measuring the thermal traits. Therefore, its

thermal traits were not measured. A new MBL originated from the two available *D.*

pseudotakihashii isofemale lines was used in the competition experiment. *Drosophila*

melanogaster does not occur naturally at the study sites, but a laboratory strain (wild

type, *Dah* strain) was measured for thermal performance together with the focal

species, as a benchmark for future comparisons.

3. Reproductive thermal performance

Experimental measurements: We exposed flies to temperatures ranging from

14°C to 32°C and measured how their reproductive success changed with

temperature (procedures are detailed in Supplementary Figure 1A). To generate the

adults flies to initiate the experiment, fly eggs collected from the population cage

were reared at low density (<100 eggs per vial) at 25°C. Emerging adults were

separated by sex within 12 hours to guarantee that they were unmated. Rearing was

started on different days for different species to synchronize the first day of egg-

laying. Two additional vials with five pairs of flies were monitored daily for sexual

maturation, indicated by egg laying. Two days after the first observation of egg-laying

in both vials, two virgin females and two virgin males were paired in a new vial

containing 4ml *Drosophila* medium (weight/volume concentration: 8% corn flour, 4%

yeast, 5% sugar, 1% agar, and 1.67% methyl-4-hydroxybenzoate). Vials were

randomly assigned to water baths set at one of seven constant temperatures (14°C,

17°C, 20°C, 23°C, 26°C, 29°C, 32°C). For each species and each temperature

treatment, eight replicates were evenly split between two experimental blocks,

making up a total of 1512 vials. Vials were submerged with the water level kept

above the zone within which flies could freely move. The temperature and relative

humidity of vials in each water bath were monitored within two empty tubes placed at

the centre and the corner. Observed temperatures varied ±0.5°C around the set

temperature. Temperatures in the centre of the water bath were on average 0.5°C

higher than at the corners; the average of the former during the experimental period

was used as the corrected temperature in analyses. Relative humidity levels were

similar to field conditions, ranging between 80% - 95%.

As temperature influenced how fecundity changed through time (Supplementary

Figure 1B), offspring numbers were measured for the 1st – 2nd day and the 7th – 8th

day and then averaged to reflect relative fecundity in early adult life. After eight days

of exposure to temperature treatments, all flies were returned to 25°C for another

four days to examine their recovery of reproduction. Surviving flies were recorded at

the beginning and end of each period. Eggs produced during the test periods

developed at the same temperature as their parents. Vials were left for 5 – 7 days

after the first emergence was observed and then the offspring was counted. The

experiment was conducted from May to August 2019.

Thermal performance curve: A multi-level, non-linear piecewise model was fitted

to describe how reproductive success changed with temperature for each *Drosophila*

species. The average daily fecundity per female was square root transformed, then

modelled with the Briere2 function (Briere et al. 1999):

$$P_i(T) = \begin{cases} a_i \times T \times (T - RTmin_i) \times (RTmax_i - T)^{1/b_i} & (for\ RTmax_i > T > RTmin_i) \\ 0 & (for\ T \leq RTmin_i\ or\ T \geq RTmax_i) \end{cases}$$

where $P_i(T)$ is the reproductive performance of species i in temperature T, *RTmin*

and *RTmax* are the minimum and maximum temperatures for the species to

reproduce, *a* is a scaling factor and *b* is a shape factor of the curve. Values of *RTmin*

of the nine species were modelled by a Gaussian distribution whose mean, μ_{RTmin},

and variance, σ_{RTmin}^2, were the hyper parameters in the multi-level model. The

same applied to $RTmax_i$, a_i, and b_i. $P(T)$ was modelled assuming a Gaussian

distribution of errors. Although a Gaussian distribution is not ideal to model the

transformed count data, which are all positive, models using untransformed count

numbers with Poisson, zero-inflated Poisson, negative binomial, and lognormal

distributions did not adequately converge, potentially due to the piecewise nature of

the thermal performance functions. The standard deviations of the Gaussian

distribution of *P(T)* were assumed different in different temperature treatments and

followed a Gaussian distribution, $\sigma_{P(T)} \sim Gaussian(\mu_{\sigma_P}, \ \sigma_{\sigma_P}^2)$. This generated better

fits than assuming the same standard deviation of *P(T)* across temperature

treatments, as judged by leave-one-out cross-validation. This was because when the

temperature was equal or close to *RTmin* and *RTmax*, *P(T)* was close to zero and its

standard deviation was also small.

The prior distributions of the hyper parameters μ_{RTmin}, μ_{RTmax}, μ_a, μ_b, and μ_{σ_P}

are Gaussian distributions with reasonable means (15°C, 30°C, 0, 0, 0) and

relatively large standard deviations (10, 10, 1, 10, 10). The prior distribution of the

hyper parameters σ_{RTmin}^2, σ_{RTmax}^2, σ_a^2, σ_b^2, and $\sigma_{\sigma_P}^2$ is inverse-gamma (0.001,

0.001), which is a commonly used non-informative distribution for priors of variance.

The values of a_i and b_i were bounded to be positive. The values of $RTmin_i$ were

bounded between 0°C to 17°C and the values of $RTmax_i$ were bounded between

26°C to 35°C based on prior knowledge of the range of temperatures under which

Australian *Drosophila* can reproduce. The multi-level model was fitted under a

Bayesian framework using MCMC sampling within the *rstan* package (Stan

Development Team, 2021) in R. Models converged and performance was

acceptable in diagnostic plots (Supplementary Figure 2). Medians of the posterior

distributions were used as the parameter values to construct the thermal

performance curves. Optimal temperatures for peak reproduction were calculated

directly from known parameters:

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$$RTopt = \frac{2bRTmax+(b+1)RTmin+\sqrt{4b^2RTmax^2+(b+1)^2RTmin^2-4b^2RTminRTmax}}{4b+2}$$

Regression analysis: The medians of the posterior distributions of *RTmin*,

RTmax, and *RTopt* were modelled by *hIndex* in a linear model with phylogenetic

correction. Phylogenetic tree were modified from Finet et al. 2021 (Supplementary

Figure 3). Fecundity at 29°C and 17°C, and recovered fecundity after 29°C and

14°C, were used as direct measurements of performance in high or low

temperatures. These fecundities were modelled as a function of *hIndex* and

experimental block as fixed effects, and *species* as a random effect in a generalized

linear mixed-effect model (family = "negative binomial") with phylogenetic correction.

These regression analyses were conducted using the *brms* package (Bürkner 2021)

in R. *Drosophila melanogaster* and *D. simulans* were not included in the regression

because their distribution patterns were unavailable from the field survey.

4. Thermal knockdown

Experimental measurements: Resistance to extreme cold temperature was

measured as knockdown time for each individual at 5°C and the time for recovery of

mobility after a 30-minute exposure to 5°C. The constant temperature chosen for

cold stress studies is often around 0°C (Gibert et al. 2001). As tropical species often

have significantly weaker cold resistance (Gibert et al. 2001), 5°C was used instead

to increase the variation among the tested species after pilot trials. Heat stress was

chosen to be 40°C, which follows common practice for *Drosophila* species

(Hoffmann, Sørensen, and Loeschcke 2003) and is expected to capture the

between-species variance in heat tolerance over a time scale which is convenient to

measure (Jørgensen, Malte, and Overgaard 2019). After being knocked down by

heat (40°C), most flies did not survive. In this case, only knockdown time was used

to evaluate resistance to heat. The knockdown experiments were conducted from

May to Jun 2019.

Virgin adult flies (siblings of those used for reproduction measurements) were

kept in same-sex groups at 25°C for 9-10 days before knockdown assays. Assays

were conducted for male and female flies separately. An observation rack was

divided into nine (3x3) cells. Each cell was randomly assigned one of the nine

Drosophila species and held seven flat-bottomed 3ml glass vials, each with a

randomly-selected individual of the allocated species. One set of observations on

such a set-up represents a single block. We repeated measurements for three

experimental blocks, and the allocation of species to cells was redrawn for each

block. In total, we measured 21 individuals per species per sex . During

measurement, the observation rack was moved immediately into the incubator pre-

set at 5°C or 40°C. Every tube was examined once every minute and flies that lost

their ability to stand in that minute were recorded. After exposure to 5°C for 30

minutes, all flies were in a chill coma. The observation rack was moved to a 25°C

room. Flies were left undisturbed and the time taken until each fly regained its ability

to stand was recorded.

Regression analysis: The knockdown time by heat, knockdown time by cold and

recovery time from cold of both sexes were first compared among species using

ANOVA. When interspecific variation was observed, they were modelled by *hIndex*,

block, and *cell position* as fixed effects, and *species* as a random effect in linear

mixed-effect models with phylogenetic correction using the *brms* package in R.

Species was included as a random effect to account for repeated measures.

5. Short-term competition

Experimental design: Pairs of species were reared in the same vials for one

generation to evaluate how the densities of intraspecific and interspecific competitors

influenced reproductive success under temperature regimes typical of lowland and

upland sites. We used incubators set at alternating temperatures mimicking day-time

and night-time temperatures in February at high-elevation (23°C / 21°C) and low-

elevation sites (28.5°C / 24°C) (Supplementary Figure 4). Adults to establish the

competition experiment were reared at moderate density (300 – 500 per bottle) at

their testing temperature regimes. After eclosion, individuals that emerged within the

same 48 hours were kept together in mixed-sex containers. Two days after the first

observation of egg-laying, adults of different sexes were separated, and from the

following day were allowed to compete and lay eggs in the 5 ml experimental vials

for two days before being discarded.

Five species were chosen as representative species for upland-biased, elevation-

generalist and lowland-biased distribution types (see Results for definition of

distribution types). The two-species combinations were: lowland species vs. upland

species: *D. bipectinata* vs. *D. pallidifrons*, *D. bipectinata* vs. *D. pseudotakahashii*,

D.pandora vs. *D. pallidifrons*; lowland species vs. lowland species: *D. bipectinata* vs.

D.pandora; lowland species vs. elevation-generalist: *D. bipectinata* vs. *D.*

sulfurigaster; upland species vs. elevation-generalist: *D. pallidifrons* vs. *D.*

sulfurigaster. Each combination was tested at different founding densities in a

factorial design: (4 pairs of species A, 2 pairs of species B), (4A, 4B), (4A, 8B), (2A,

4B) and (8A, 4B). We also included monocultures of each species with 2, 4, and 8

pairs. Each density and species combination was replicated ten times across two or

three blocks staggered by two days (two blocks for the *D. pandora* vs. *D. pallidifrons*

combination; three blocks for the other five pairs). Offspring that successfully

developed to adulthood were identified to species and counted. Experiment of *D.*

pandora vs. *D. pallidifrons* combination was conducted from September to

December, 2020 and the other five pairs were experimented from January to March,

2021.

Competition models: We used the Beverton-Holt model to describe the

population growth of a single generation of flies on discrete and temporary

resources:

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$$N_{focal,t+1} = \frac{R_0 N_{focal,t}}{1 + a(N_{focal,t} + \beta N_{competitor,t})}$$

where R_0 is the generational reproduction rate and a is a constant defining the

form of the density-dependence relationship. β represents the interspecific

competition coefficient of the competitor species to the focal species, which defines

the equivalence between the two competing species. Offspring numbers of the focal

species were modelled assuming a negative binomial error distribution, under a

Bayesian framework using MCMC sampling within the *rstan* (Stan Development

Team 2021) package in R, as described by Terry et al. (2021). Model diagnostics are

shown in Supplementary Figure 5A-B. The medians of the posterior distribution were

used as the parameter values to infer the equilibrium state of each pair following

Hassell and Comins (1976).

6. Long-term competition

Experimental design: To evaluate the long-term impact of temperature and

competition on population size, a lowland species, *D. pandora*, and an upland

species, *D. pallidifrons*, were reared in monoculture and mixed-culture environments

for multiple generations under low-elevation and high-elevation temperature regimes

(Supplementary Figure 4).

Four monocultures of each species and eight mixed-species cultures were

maintained at each temperature regime for 13 weeks, totalling 24 cultures.

Monocultures were started with ten pairs of individuals. Mixed-species cultures were

started with ten pairs of individuals of each species. The starting density was very

low compared to the equilibrium density. The cultures were evenly divided into two

blocks starting on different dates. Each was maintained in a series of five bottles

following Ayala, Gilpin, and Ehrenfeld (1973). At the start of each week, adults

surviving in the most recent bottle and adults which were freshly emerged in the

older four bottles were separately collected, photographed and transferred together

into a new bottle with fresh food. In this way, adult survival and reproduction were

recorded separately. The total population size of each species was counted at the

end of the experiment. To avoid pseudo-replication introduced by 'incubator' effects,

the two incubators were switched between temperature regimes every week, with

their contents moved accordingly. Trays were shuffled inside the incubator every two

days. Temperature and humidity were recorded and the temperature regimes were

confirmed during and at the end of experiments. The experiment was conducted

from September to December, 2020.

Data analysis: Population sizes were modelled as a function of *temperature*,

species, *presence/absence of competitors*, and their interactions, with *culture ID* as a

random effect with a generalized linear mixed-effect model (family = "zero-inflated

negative binomial") using the *brms* package in R. Model diagnostics are shown in

Supplementary Figure 5C. To visualize the three-way interactive effects, the

posterior estimates of the high-temperature effect were plot against zero for the two

species with or without the presence of competitors; the posterior estimates of the

effect of the competition were plotted against zero for the two species in each

temperature regime.

All statistics mentioned in the *Methods* section were performed with R statistical

software, version 4.0.3 (R Core Team, 2020). Original data are achieved on Zenedo

(10.5281/zenodo.6400611) and code of analysis are available on GitHub

(https://github.com/Jinlinc/DistributionRegulators.git).

Results

1.Field distributions

The numbers of samples found at low-, medium- or high-elevation sites for

each of the nine major *Drosophila* species (accounting for 99% of all samples) are

shown in Figure 1. Distributions quantified using regression and by weighted

elevation (*hIndex*) were consistent (Spearman's rank correlation rho = 0.93, p value

= 0.0007). *Drosophila bipectinate* and *D. pandora* had significantly decreased

detection probability with increasing elevation and were thus categorized as lowland-

biased species with high confidence. *Drosophila pseudoananassae* showed a

lowland bias on one of the two mountains, and a mid-elevation peak on the other.

Drosophila rubida, *D. sulfurigaster* and *D. birchii* showed no significant change with

elevation and were thus defined as elevation generalists. *Drosophila pallidifrons* and

D. pseudotakahashii were significantly more likely to be found at high elevations and

were thus defined as upland-biased species. Coefficients and p values of the

regressions and *hIndex* are shown in Supplementary Table 2. For *D. bunnanda,* the

six records were insufficient for model fitting but all occurred at lowland sites, and

data from a larger-scale study suggest it is a lowland-biased species (Schiffer and

McEvey 2006).

2.Thermal performance curves

Thermal performance curves of daily fecundity per female vary among

species in terms of the range, optimal temperature, peak fecundity, and shape

factors (Figure 2; Table 1; see Supplementary Figure 2C for original data and fitted

curves for each species). The temperature for optimal reproductive performance,

RTopt, did not correlate with distribution patterns (Coefficient = 0.09, 95% credible

interval = -2.83 – 3.01; Supplementary Figure 6A). There was no general trade-off

between cold tolerance (estimated *RTmin*) versus heat tolerance (estimated *RTmax*)

corresponding to species' distribution types (Spearman's rank correlation rho = -0.6,

p value = 0.10; Supplementary Figure 6B). For example, the lowland-biased species

D. bunnanda has higher heat tolerance and lower cold tolerance than its elevation-

generalist relative, *D. birchii*. In contrast, *D. sulfurigaster* outperforms its upland-

biased relative, *D. pallidifrons*, across the temperature range.

3.Cold tolerance

Values of *RTmin* were not correlated with species distribution patterns (Figure

3a. Coefficient = -0.41, 95% credible interval = -4.15 – 3.43). Similarly, upland-

biased species did not show higher fecundity at the low temperature, 17°C (Figure

3b. Coefficient = -0.27, 95% credible interval -3.80 – 3.05). When exposed to acute

sublethal low temperature (5°C), all seven tropical *Drosophila* species showed

similarly poor performance compared to *D. simulans* and *D. melanogaster*

(Supplementary Table 3 for Tukey multiple pairwise comparisons). All species

recovered their fecundity after eight days at 14°C. This recovered fecundity was

slightly but non-significantly higher among upland species (Figure 3c. Coefficient =

0.33, 95% credible interval = -0.55 – 1.14). It took longer for upland species to regain

mobility after chill coma (Figure 3d. Male: coefficient = 13.04 (-9.31 – 35.05); female:

coefficient = 8.49 (-2.18 – 20.02).)

4.Heat tolerance

Regardless of the small variation of *RTmax* (standard deviation = 1.01)

compared with *RTmin* (standard deviation = 2.82), *RTmax* was significantly higher

among species with more lowland-biased distributions (Figure 3e. Coefficient = -

3.06, 95% credible interval = -5.30 – -0.88). Reproductive performance at 29°C also

decreased with *hIndex* (Figure 3f. Coefficient = -5.80, 95% credible interval = -9.37 –

-2.50). After exposure to 29°C for eight days, neither upland species could

reproduce when transferred back to 25°C, while four out of the five elevation-

generalist and lowland-biased species resumed reproduction (Figure 3g).

Knockdown time at lethally high temperature (40°C) was shorter among upland

species (Figure 3h. Male: coefficient = -6.71 (-13.70 – 0.50); female: coefficient = -

1.99 (-9.48 – 5.71)), indicating these species lose their mobility faster at high

temperatures.

5.Interspecific competition

When raised in a laboratory condition mimicking the warmer, lowland sites,

reproductive success was highest for the two lowland-biased species, followed by

the elevation-generalist species, *D. sulfurigaster*. The two upland species could

barely reproduce regardless of the presence of competitors (proliferation rates (R0)

of *D. pallidifrons* and *D. pseudotakahashii* were lower than 1; Figure 4; Table 2).

Their competitive effect on lowland species was minimal, indicated by low β values

(Table 2). The two lowland species, *Drosophila pandora* and *D. bipectinata*, are

expected to coexist stably, based on their reproductive and competitive parameters.

When raised a under cooler, upland condition, all species could reproduce

and sustain their populations (all proliferation rates (R0) were higher than 1; Table

2). Lowland species were strongly affected by the density of *D. pallidifrons*, an

upland species, while upland species were significantly less affected by lowland

species. Competition with *D.pallidirons* under upland conditions was predicted to

drive *D. pandora* and *D. bipectinata* to exclusion (Table 2).

In the long-term competition experiment, high temperature drove the upland-

species *D. pallidifrons* to extinction regardless of the starting species composition

(monoculture versus mixed-species culture) (Figure 5a). In contrast, the monoculture

of the lowland-species *D. pandora* remained abundant at both temperatures (Figure

5b: 90% credible interval of temperature effect in monoculture overlaps with zero).

The interspecific competition effect was only significant at the upland regime (Figure

5c), which significantly reduced the population size of *D. pandora* in mixed-species

cultures compared to the monocultures (Figure 5a and Figure 5b).

Discussion

Counter to expectation, our results do not support the common assumption

that cool boundaries to species' ranges are constrained abiotically (reflecting thermal

niches), while biotic interactions (such as interspecific competition) define warm

boundaries. Instead, we found that maximum temperatures experienced on a daily

basis at lowland sites were sufficiently high to serve as an environmental filter. In

cooler, high-elevation conditions, lowland species were outcompeted by certain

upland species whose distribution were confined to high elevations as a result of

their intolerance to heat.

1.Contributions of abiotic and biotic factors in determining distributions

The idea that the contributions of abiotic versus biotic factors might differ for

species' warm and cool range boundaries can be traced back to Charles Darwin,

and remains an area of active debate (Cahill et al. 2014; Hargreaves, Samis, and

Eckert 2014; Schemske et al. 2009). There is especially mixed evidence on the

importance of upper thermal limits in deciding warm boundaries (positive

relationships: Batista et al. 2018; Duarte et al. 2012; García-Robledo et al. 2016;

Kellermann et al. 2012; Merrill et al. 2008; null relationships: Gaston and Chown

1999; Huang and Tu 2008; Kimura 2004; Nowrouzi et al. 2018). This inconsistency

could arise for at least three reasons. First, data available for synthetic studies are

over-represented by systems from temperate latitudes in the northern hemisphere

(Feeley et al. 2017) and relate largely to cool limits (Cahill et al. 2014); a recent

synthesis finds that the difference in the relative importance of biotic factors to warm

versus cool boundaries tends to disappear toward the equator (Paquette and

Hargreaves 2021). Second, mechanisms governing latitudinal or elevational

distribution patterns could differ (Chen, Hill, Ohlemüller, et al. 2011), and may only

become apparent as more datasets accumulate from less-studied locations (Duarte

et al. 2012; Freeman et al. 2021). Third, some of the uncertainty might reflect

methodological differences when choosing, defining and measuring thermal traits.

Studies like the current one that examine a range of thermal traits and biotic

interactions may be needed to understand how and why the governing rules might

change under different thermal and biotic regimes (Amundrud and Srivastava 2020;

Srinivasan et al. 2018).

Indeed, different choices of indicators of thermal performance, and different

experimental methods may explain why some of our results contrast with some other

studies of Australian rainforest *Drosophila*. Overgaard et al. (2014) found no

evidence that thermal traits of tropical rainforest *Drosophila* differed among species.

Unlike Overgaard et al. (2014), we measured parental fecundity by allowing their

eggs to develop to adults under the relevant temperature; we observed that eggs laid

at high temperatures never hatched, perhaps as a result of sperm sterilization

(Parratt et al. 2021). We also observed different speeds of senescence after

exposure to different temperatures (Supplementary Figure 1B), which the three-day

period of fecundity measurement in Overgaard's study would not have captured.

Nevertheless, both studies highlight the conserved nature of heat tolerance and

suggest small thermal safety margins to future warming.

Field experiments on *D. birchii* (O'Brien et al. 2017, 2020), found that its

reproduction when raised alone was highest at low elevations, where both

intraspecific and interspecific competition with its sister species *D. bunnanda* were

also high. In our experiments, *D. birchii* was one of the most heat-sensitive species,

and its populations were unable to persist at high temperatures typical of low

elevations (pilot experiment, unpublished data). This discrepancy can be explained

by different temperature regimes used in our study (corresponding to February

temperatures in the field, with an average of 26°C and average daily maximum of

30.6°C at lowland) and by O'Brien et al. (April temperatures, which are around 2°C

lower on average temperature and on average daily maximum). A further possibility

is that single-generation experiments initiated with mated adult flies did not capture

the impact of high temperatures prior to sexual maturation. These observations

further highlight the sensitivity of measures of thermal performance to experimental

conditions. They also suggest that the relative importance of abiotic and biotic

factors in determining species distributions may change seasonally.

2.Low variation in upper thermal limits

Upper thermal limits vary little among species (Hoffmann 2010) and have

limited adaptive potential (van Heerwaarden and Sgrò 2021; Kellermann et al. 2012).

Consistent with other studies (Goulet, Thompson, and Chapple 2017; Hangartner

and Hoffmann 2016), we found that heat tolerance is a systematic trait manifested in

critical temperatures, fecundities at sub-sterile temperatures, recovered fecundities,

and locomotive responses. Modest changes in critical temperature can therefore

lead to pronounced differences in overall performance under real and variable

temperature regimes.

Such small thermal safety margins suggest a severe threat of biotic attrition in

tropical lowlands (Colwell et al. 2008; Deutsch et al. 2008; Duarte et al. 2012; van

Heerwaarden and Sgrò 2021). Laboratory-measured critical temperatures can be

sensitive to experimental conditions, making it difficult to relate them to climatological

means or maxima, and hence the threat of rising temperatures (Sinclair et al. 2016).

In this context, our study benefits from a comparative approach, which reveals that

upland species are already constrained by high temperatures, and that a very small

difference (about 1°C) distinguishes the upper thermal limits (*RTmax*s) of lowland

and upland species. Given the low evolutionary potential of heat tolerance

(Hoffmann, Chown, and Clusella-Trullas 2013), both lowland species and upland

species are likely to be vulnerable to modest temperature increases across the

elevation gradient. Thus, lowland biotic attrition and upland range contraction are

likely to occur with future warming, leading to cascading effects in lowland

communities and threatening endemic upland species on tropical mountains such as

those in the Australian Wet Tropics (Freeman et al. 2018).

3.Daily peak and extreme high temperature as the main abiotic filters

Daily maxima can be more important than mean temperatures in structuring

distributions (Lynch et al. 2014). In our study system, the mean temperature during

the study season at our lowland sites is around 26°C. At this temperature, all studied

species are close to their peak reproductive performance. However, upland and

lowland sites differ greatly in the average daily maximum temperature and the daily

duration that the temperature reaches or exceeds stressful levels for *Drosophila*

reproduction (Supplementary Figure 7). Brief exposure to stressful thermal

environments can have similar fitness costs to continuously stressful conditions

(Saxon, O'Brien, and Bridle 2018). This highlights the importance of considering

daily temperature variations and extreme temperature events when studying species

distributions and projecting responses to climate change (Kingsolver, Diamond, and

Buckley 2013; Ma, Hoffmann, and Ma 2015).

4. Context-dependence of thermal performance

Inevitably, thermal traits measured under laboratory conditions cannot

account for the tendency of thermal tolerances to vary with a range of abiotic factors

including, but not limited to precipitation (Bozinovic and Pörtner 2015; Kellermann et

al. 2012), diet, and larval conditions (Bubli, Imasheva, and Loeschcke 1998). Such

variables are to a large extent standardized in our study, allowing dependence on

abiotic conditions to be resolved. Our long-term experiment showed that the thermal

response of population size to temperature depended on competitive conditions,

highlighting the difference between thermal performance curves of organisms and

their populations when biotic interactions are present (Davis et al. 1998). There is a

growing realisation that sensitivity to temperature depends on the biotic environment

(Gilman et al. 2010): it is not only the demographics of the interacting species that

are affected by temperature (Huey and Kingsolver 1989); interactions themselves

are altered (Burnside et al. 2014).

Finally, between-population variation and local adaptation (Hoffmann,

Anderson, and Hallas 2002), especially adaptation to abiotic environments at

distribution boundaries (Peterson, Doak, and Morris 2019), should not be neglected

if studying distributions at a large spatial scales (e.g., across latitudes). The relatively

restricted spatial scale of our study (within a mountain range), the use of mass bred

lines sourced from different elevations, and the limited plasticity and local adaptation

documented for our study species (MacLean et al. 2019; O'Brien et al. 2017) mean

that plastic and evolutionary response to thermal conditions are unlikely to

complicate interpretation of our results.

5.Conclusions

Tropical ecosystems host an exceptional diversity of endemic species

(Laurance et al. 2011). Predicting their sensitivity to climate change is a high priority

and requires an understanding of the proximate causes of current distributions as

well as the interactions between thermal tolerance and other environmental factors,

both biotic and abiotic. Our study contributes to the growing literature demonstrating

that species can be sensitive to warming at the warm boundaries of their

distributions. In particular, we highlight the important role of daily maximum

temperatures in structuring community at tropical lowlands, the essential role of

interspecific competition at uplands, and the vulnerability of species there to

increasing mean temperatures and temperature extremes.

References:

Amundrud, Sarah L. and Diane S. Srivastava. 2020. "Thermal Tolerances and
    Species Interactions Determine the Elevational Distributions of Insects." *Global
    Ecology and Biogeography* 29(8):1315–27.

Ayala, Francisco J., Michael E. Gilpin, and Joan G. Ehrenfeld. 1973. "Competition
    between Species: Theoretical Models and Experimental Tests." *Theoretical
    Population Biology* 4(3):331–56.

Batista, Marcos Roberto Dias, Felipe Bastos Rocha, and Louis Bernard Klaczko.
    2018. "Altitudinal Distribution of Two Sibling Species of the Drosophila
    Tripunctata Group in a Preserved Tropical Forest and Their Male Sterility
    Thermal Thresholds." *Journal of Thermal Biology* 71:69–73.

Bonebrake, Timothy C. and Curtis A. Deutsch. 2012. "Climate Heterogeneity
    Modulates Impact of Warming on Tropical Insects." *Ecology* 93(3):449–55.

Bozinovic, Francisco and Hans Otto Pörtner. 2015. "Physiological Ecology Meets
    Climate Change." *Ecology and Evolution* 5(5):1025–30.

Briere, Jean Francois, Pascale Pracros, Alain Yves Le Roux, and Jean Sebastien
    Pierre. 1999. "A Novel Rate Model of Temperature-Dependent Development for
    Arthropods." *Environmental Entomology* 28(1):22–29.

636 Bubli, Oleg A., Alexandra G. Imasheva, and Volker Loeschcke. 1998. "Selection for Knockdown Resistance to Heat in Drosophila Melanogaster at High and Low Larval Densities." *Evolution* 52(2):619–25.

639 Bürkner, Paul-Christian. 2021. "Bayesian Item Response Modeling in R with Brms and Stan." *Journal of Statistical Software* 100(5):1–54.

641 Burnside, William R., Erik B. Erhardt, Sean T. Hammond, and James H. Brown. 2014. "Rates of Biotic Interactions Scale Predictably with Temperature despite Variation." *Oikos* 123(12):1449–56.

644 Cahill, Abigail E., Matthew E. Aiello-Lammens, M. Caitlin Fisher-Reid, Xia Hua, Caitlin J. Karanewsky, Hae Yeong Ryu, Gena C. Sbeglia, Fabrizio Spagnolo, John B. Waldron, and John J. Wiens. 2014. "Causes of Warm-Edge Range Limits: Systematic Review, Proximate Factors and Implications for Climate Change" edited by W. Daniel Kissling. *Journal of Biogeography* 41(3):429–42.

Chen, I. Ching, Jane K. Hill, Ralf Ohlemüller, David B. Roy, and Chris D. Thomas. 2011. "Rapid Range Shifts of Species Associated with High Levels of Climate Warming." *Science* 333(6045):1024–26.

Chen, I. Ching, Jane K. Hill, Hau Jie Shiu, Jeremy D. Holloway, Suzan Benedick, Vun Khen Chey, Henry S. Barlow, and Chris D. Thomas. 2011. "Asymmetric Boundary Shifts of Tropical Montane Lepidoptera over Four Decades of Climate Warming." *Global Ecology and Biogeography* 20(1):34–45.

Colwell, Robert K., Gunnar Brehm, Catherine L. Cardelús, Alex C. Gilman, and John T. Longino. 2008. "Global Warming, Elevational Range Shifts, and Lowland Biotic Attrition in the Wet Tropics." *Science* 322(5899):258–61.

Corlett, Richard T. 2011. "Impacts of Warming on Tropical Lowland Rainforests." *Trends in Ecology and Evolution* 26(11):606–13.

Corlett, Richard T. 2012. "Climate Change in the Tropics: The End of the World as

We Know It?" *Biological Conservation* 151(1):22–25.

Davis, A. J., L. S. Jenkinson, J. H. Lawton, B. Shorrocks, and S. Wood. 1998.

"Making Mistakes When Predicting Shifts in Species Range in Response to

Global Warming." *Nature* 391(6669):783–86.

Deutsch, Curtis A., Joshua J. Tewksbury, Raymond B. Huey, Kimberly S. Sheldon,

Cameron K. Ghalambor, David C. Haak, and Paul R. Martin. 2008. "Impacts of

Climate Warming on Terrestrial Ectotherms across Latitude." *Proceedings of the*

National Academy of Sciences of the United States of America 105(18):6668–

72.

Diamond, Sarah E., D. Magdalena Sorger, Jiri Hulcr, Shannon L. Pelini, Israel Del

Toro, Christopher Hirsch, Erik Oberg, and Robert R. Dunn. 2012. "Who Likes It

Hot? A Global Analysis of the Climatic, Ecological, and Evolutionary

Determinants of Warming Tolerance in Ants." *Global Change Biology*

18(2):448–56.

Duarte, Helder, Miguel Tejedo, Marco Katzenberger, Federico Marangoni, Diego

Baldo, Juan Francisco Beltrán, Dardo Andrea Martí, Alex Richter-Boix, and

Alejandro Gonzalez-Voyer. 2012. "Can Amphibians Take the Heat? Vulnerability

to Climate Warming in Subtropical and Temperate Larval Amphibian

Communities." *Global Change Biology* 18(2):412–21.

Engelbrecht, Bettina M. J., Liza S. Comita, Richard Condit, Thomas A. Kursar,

Melvin T. Tyree, Benjamin L. Turner, and Stephen P. Hubbell. 2007. "Drought

Sensitivity Shapes Species Distribution Patterns in Tropical Forests." *Nature*

447(7140):80–82.

Feeley, Kenneth J., James T. Stroud, and Timothy M. Perez. 2017. "Most 'Global'

Reviews of Species' Responses to Climate Change Are Not Truly Global" edited

by I. Kühn. *Diversity and Distributions* 23(3):231–34.

Finet, Cédric, Victoria A. Kassner, Antonio B. Carvalho, Henry Chung, Jonathan P.

Day, Stephanie Day, Emily K. Delaney, Francine C. De Ré, Héloïse D. Dufour,

Eduardo Dupim, Hiroyuki F. Izumitani, Thaísa B. Gautério, Jessa Justen, Toru

Katoh, Artyom Kopp, Shigeyuki Koshikawa, Ben Longdon, Elgion L. Loreto,

Maria D. S. Nunes, Komal K. B. Raja, Mark Rebeiz, Michael G. Ritchie, Gayane

Saakyan, Tanya Sneddon, Machiko Teramoto, Venera Tyukmaeva, Thyago

Vanderlinde, Emily E. Wey, Thomas Werner, Thomas M. Williams, Lizandra J.

Robe, Masanori J. Toda, and Ferdinand Marlétaz. 2021. "DrosoPhyla:

Resources for Drosophilid Phylogeny and Systematics" edited by J. Gonzalez.

Genome Biology and Evolution 13(8).

Freeman, Benjamin G., Micah N. Scholer, Viviana Ruiz-Gutierrez, and John W.

Fitzpatrick. 2018. "Climate Change Causes Upslope Shifts and Mountaintop

Extirpations in a Tropical Bird Community." *Proceedings of the National*

Academy of Sciences of the United States of America 115(47):11982–87.

Freeman, Benjamin G., Yiluan Song, Kenneth J. Feeley, and Kai Zhu. 2021.

"Montane Species Track Rising Temperatures Better in the Tropics than in the

Temperate Zone." *Ecology Letters* ele.13762.

García-Robledo, Carlos, Erin K. Kuprewicz, Charles L. Staines, Terry L. Erwin, and

W. John Kress. 2016. "Limited Tolerance by Insects to High Temperatures

across Tropical Elevational Gradients and the Implications of Global Warming

for Extinction." *Proceedings of the National Academy of Sciences of the United*

States of America 113(3):680–85.

Gaston, Kevin J. and Steven L. Chown. 1999. "Elevation and Climatic Tolerance: A

Test Using Dung Beetles." *Oikos* 86(3):584.

Gibert, Patricia, Brigitte Moreteau, Georges Pétavy, Dev Karan, and Jean R. David.

2001. "Chill-Coma Tolerance, a Major Climatic Adaptation among Drosophila

Species." *Evolution* 55(5):1063–68.

Gilman, Sarah E., Mark C. Urban, Joshua Tewksbury, George W. Gilchrist, and

Robert D. Holt. 2010. "A Framework for Community Interactions under Climate

Change." *Trends in Ecology and Evolution* 25(6):325–31.

Goulet, Celine T., Michael B. Thompson, and David G. Chapple. 2017. "Repeatability

and Correlation of Physiological Traits: Do Ectotherms Have a 'Thermal Type'?"

Ecology and Evolution 7(2):710–19.

Hangartner, Sandra and Ary A. Hoffmann. 2016. "Evolutionary Potential of Multiple

Measures of Upper Thermal Tolerance in Drosophila Melanogaster." *Functional*

Ecology 30(3):442–52.

Hargreaves, Anna L., Karen E. Samis, and Christopher G. Eckert. 2014. "Are

Species' Range Limits Simply Niche Limits Writ Large? A Review of Transplant

Experiments beyond the Range." *American Naturalist* 183(2):157–73.

Hassell, M. P. and H. N. Comins. 1976. "Discrete Time Models for Two-Species

Competition." *Theoretical Population Biology* 9(2):202–21.

van Heerwaarden, Belinda and Carla M. Sgrò. 2021. "Male Fertility Thermal Limits

Predict Vulnerability to Climate Warming." *Nature Communications* 12(1):1–11.

HilleRisLambers, J., P. B. Adler, W. S. Harpole, J. M. Levine, and M. M. Mayfield.

2012. "Rethinking Community Assembly through the Lens of Coexistence

Theory." *Annual Review of Ecology, Evolution, and Systematics* 43(1):227–48.

Hoffmann, A. A. 2010. "Physiological Climatic Limits in Drosophila: Patterns and

Implications." *Journal of Experimental Biology* 213(6):870–80.

Hoffmann, Ary A., Alisha Anderson, and Rebecca Hallas. 2002. "Opposing Clines for High and Low Temperature Resistance in Drosophila Melanogaster." *Ecology Letters* 5(5):614–18.

Hoffmann, Ary A. and Mark W. Blows. 1994. "Species Borders: Ecological and Evolutionary Perspectives." *Trends in Ecology and Evolution* 9(6):223–27.

Hoffmann, Ary A., Steven L. Chown, and Susana Clusella-Trullas. 2013. "Upper Thermal Limits in Terrestrial Ectotherms: How Constrained Are They?" edited by C. Fox. *Functional Ecology* 27(4):934–49.

Hoffmann, Ary A., Jesper G. Sørensen, and Volker Loeschcke. 2003. "Adaptation of Drosophila to Temperature Extremes: Bringing Together Quantitative and Molecular Approaches." *Journal of Thermal Biology* 28(3):175–216.

Huang, Shu Ping and Ming Chung Tu. 2008. "Heat Tolerance and Altitudinal Distribution of a Mountainous Lizard, Takydromus Hsuehshanensis, in Taiwan." *Journal of Thermal Biology* 33(1):48–56.

Huey, Raymond B., Curtis A. Deutsch, Joshua J. Tewksbury, Laurie J. Vitt, Paul E. Hertz, Héctor J. Álvare. Pérez, and Theodore Garland. 2009. "Why Tropical Forest Lizards Are Vulnerable to Climate Warming." *Proceedings of the Royal Society B: Biological Sciences* 276(1664):1939–48.

Huey, Raymond B. and Joel G. Kingsolver. 1989. "Evolution of Thermal Sensitivity of Ectotherm Performance." *Trends in Ecology and Evolution* 4(5):131–35.

Huey, Raymond B. and R. D. Stevenson. 1979. "Integrating Thermal Physiology and Ecology of Ectotherms: A Discussion of Approaches." *Integrative and Comparative Biology* 19(1):357–66.

Jankowski, Jill E., Gustavo A. Londoño, Scott K. Robinson, and Mark A. Chappell. 2013. "Exploring the Role of Physiology and Biotic Interactions in Determining

Elevational Ranges of Tropical Animals." *Ecography* 36(1):1–12.

Jeffs, Christopher T., J. Christopher D. Terry, Megan Higgie, Anna Jandová, Hana

Konvičková, Joel J. Brown, Chia Hua Lue, Michele Schiffer, Eleanor K. O'Brien,

Jon Bridle, Jan Hrček, and Owen T. Lewis. 2021. "Molecular Analyses Reveal

Consistent Food Web Structure with Elevation in Rainforest Drosophila –

Parasitoid Communities." *Ecography* 44(3):403–13.

Jørgensen, Lisa Bjerregaard, Hans Malte, and Johannes Overgaard. 2019. "How to

Assess Drosophila Heat Tolerance: Unifying Static and Dynamic Tolerance

Assays to Predict Heat Distribution Limits" edited by C. White. *Functional*

Ecology 33(4):629–42.

Kearney, Michael and Warren Porter. 2009. "Mechanistic Niche Modelling:

Combining Physiological and Spatial Data to Predict Species' Ranges." *Ecology*

Letters 12(4):334–50.

Kellermann, Vanessa, Johannes Overgaard, Ary A. Hoffmann, Camilla Fljøgaard,

Jens Christian Svenning, and Volker Loeschcke. 2012. "Upper Thermal Limits of

Drosophila Are Linked to Species Distributions and Strongly Constrained

Phylogenetically." *Proceedings of the National Academy of Sciences of the*

United States of America 109(40):16228–33.

Khaliq, Imran, Christian Hof, Roland Prinzinger, Katrin Böhning-Gaese, and Markus

Pfenninger. 2014. "Global Variation in Thermal Tolerances and Vulnerability of

Endotherms to Climate Change." *Proceedings of the Royal Society B: Biological*

Sciences 281(1789).

Kimura, Masahito T. 2004. "Cold and Heat Tolerance of Drosophilid Flies with

Reference to Their Latitudinal Distributions." *Oecologia* 140(3):442–49.

Kingsolver, Joel G., Sarah E. Diamond, and Lauren B. Buckley. 2013. "Heat Stress

786  and the Fitness Consequences of Climate Change for Terrestrial Ectotherms"

787  edited by J. Grindstaff. *Functional Ecology* 27(6):1415–23.

788 Laurance, William F., D. Carolina Useche, Luke P. Shoo, Sebastian K. Herzog,

789  Michael Kessler, Federico Escobar, Gunnar Brehm, Jan C. Axmacher, I. Ching

790  Chen, Lucrecia Arellano Gámez, Peter Hietz, Konrad Fiedler, Tomasz Pyrcz,

791  Jan Wolf, Christopher L. Merkord, Catherine Cardelus, Andrew R. Marshall,

792  Claudine Ah-Peng, Gregory H. Aplet, M. del Coro Arizmendi, William J. Baker,

793  John Barone, Carsten A. Brühl, Rainer W. Bussmann, Daniele Cicuzza, Gerald

794  Eilu, Mario E. Favila, Andreas Hemp, Claudia Hemp, Jürgen Homeier, Johanna

795  Hurtado, Jill Jankowski, Gustavo Kattán, Jürgen Kluge, Thorsten Krömer, David

796  C. Lees, Marcus Lehnert, John T. Longino, Jon Lovett, Patrick H. Martin, Bruce

797  D. Patterson, Richard G. Pearson, Kelvin S. H. Peh, Barbara Richardson,

798  Michael Richardson, Michael J. Samways, Feyera Senbeta, Thomas B. Smith,

799  Timothy M. A. Utteridge, James E. Watkins, Rohan Wilson, Stephen E.

800  Williams, and Chris D. Thomas. 2011. "Global Warming, Elevational Ranges

801  and the Vulnerability of Tropical Biota." *Biological Conservation* 144(1):548–57.

802 Louthan, Allison M., Daniel F. Doak, and Amy L. Angert. 2015. "Where and When Do

803  Species Interactions Set Range Limits?" *Trends in Ecology and Evolution*

804  30(12):780–92.

805 Lynch, Heather J., Marc Rhainds, Justin M. Calabrese, Stephen Cantrell, Chris

806  Cosner, and William F. Fagan. 2014. "How Climate Extremes—Not Means—

807  Define a Species' Geographic Range Boundary via a Demographic Tipping

808  Point." *Ecological Monographs* 84(1):131–49.

809 Ma, Gang, Ary A. Hoffmann, and Chun Sen Ma. 2015. "Daily Temperature Extremes

810  Play an Important Role in Predicting Thermal Effects." *Journal of Experimental*

Biology 218(14):2289–96.

MacLean, Heidi J., Jesper G. Sørensen, Torsten N. Kristensen, Volker Loeschcke,

Kristian Beedholm, Vanessa Kellermann, and Johannes Overgaard. 2019.

"Evolution and Plasticity of Thermal Performance: An Analysis of Variation in

Thermal Tolerance and Fitness in 22 *Drosophila* Species." *Philosophical*

Transactions of the Royal Society B: Biological Sciences 374(1778):20180548.

Merrill, Richard M., David Gutiérrez, Owen T. Lewis, Javier Gutiérrez, Sonia B. Díez,

and Robert J. Wilson. 2008. "Combined Effects of Climate and Biotic

Interactions on the Elevational Range of a Phytophagous Insect." *Journal of*

Animal Ecology 77(1):145–55.

Nowrouzi, Somayeh, Alan N. Andersen, Tom R. Bishop, and Simon K. A. Robson.

2018. "Is Thermal Limitation the Primary Driver of Elevational Distributions? Not

for Montane Rainforest Ants in the Australian Wet Tropics." *Oecologia*

188(2):333–42.

O'Brien, Eleanor K., Megan Higgie, Christopher T. Jeffs, Ary A. Hoffmann, Jan

Hrček, Owen T. Lewis, and Jon R. Bridle. 2020. "Fitness Effects of Competition

within and between Species Change across Species' Ranges, and Reveal

Limited Local Adaptation in Rainforest <em>Drosophila</Em>." *BioRxiv* 395624.

O'Brien, Eleanor K., Megan Higgie, Alan Reynolds, Ary A. Hoffmann, and Jon R.

Bridle. 2017. "Testing for Local Adaptation and Evolutionary Potential along

Altitudinal Gradients in Rainforest Drosophila: Beyond Laboratory Estimates."

Global Change Biology 23(5):1847–60.

Overgaard, Johannes, Michael R. Kearney, and Ary A. Hoffmann. 2014. "Sensitivity

to Thermal Extremes in Australian Drosophila Implies Similar Impacts of Climate

Change on the Distribution of Widespread and Tropical Species." *Global*

Change Biology 20(6):1738–50.

Paquette, Alexandra and Anna L. Hargreaves. 2021. "Biotic Interactions Are More

Often Important at Species' Warm versus Cool Range Edges" edited by C.

Ghalambor. *Ecology Letters* 24(11):2427–38.

Parratt, Steven R., Benjamin S. Walsh, Soeren Metelmann, Nicola White, Andri

Manser, Amanda J. Bretman, Ary A. Hoffmann, Rhonda R. Snook, and Tom A.

R. Price. 2021. "Temperatures That Sterilize Males Better Match Global Species

Distributions than Lethal Temperatures." *Nature Climate Change* 11(6):481–84.

Peterson, Megan L., Daniel F. Doak, and William F. Morris. 2019. "Incorporating

Local Adaptation into Forecasts of Species' Distribution and Abundance under

Climate Change." *Global Change Biology* 25(3):775–93.

R Core Team (2020). R: A language and environment for statistical computing. R

Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-

project.org/.

Saxon, A. D., E. K. O'Brien, and J. R. Bridle. 2018. "Temperature Fluctuations during

Development Reduce Male Fitness and May Limit Adaptive Potential in Tropical

Rainforest Drosophila." *Journal of Evolutionary Biology* 31(3):405–15.

Schemske, Douglas W., Gary G. Mittelbach, Howard V. Cornell, James M. Sobel,

and Kaustuv Roy. 2009. "Is There a Latitudinal Gradient in the Importance of

Biotic Interactions?" *Annual Review of Ecology, Evolution, and Systematics*

40(1):245–69.

Schiffer, Michele and Shane F. McEvey. 2006. "Drosophila Bunnanda - A New

Species from Northern Australia with Notes on Other Australian Members of the

Montium Subgroup (Diptera: Drosophilidae)." *Zootaxa* (1333):1–23.

Sheldon, Kimberly S., Sylvia Yang, and Joshua J. Tewksbury. 2011. "Climate

Change and Community Disassembly: Impacts of Warming on Tropical and

Temperate Montane Community Structure." *Ecology Letters* 14(12):1191–1200.

Sinclair, Brent J., Katie E. Marshall, Mary A. Sewell, Danielle L. Levesque,

Christopher S. Willett, Stine Slotsbo, Yunwei Dong, Christopher D. G. Harley,

David J. Marshall, Brian S. Helmuth, and Raymond B. Huey. 2016. "Can We

Predict Ectotherm Responses to Climate Change Using Thermal Performance

Curves and Body Temperatures?" edited by D. Vasseur. *Ecology Letters*

19(11):1372–85.

Srinivasan, Umesh, Paul R. Elsen, Morgan W. Tingley, and David S. Wilcove. 2018.

"Temperature and Competition Interact to Structure Himalayan Bird

Communities." *Proceedings of the Royal Society B: Biological Sciences*

285(1874):20172593.

Stan Development Team. 2021. "RStan: The R Interface to Stan." R package version

2.21.3.

Sunday, Jennifer M., Amanda E. Bates, and Nicholas K. Dulvy. 2012. "Thermal

Tolerance and the Global Redistribution of Animals." *Nature Climate Change*

2(9):686–90.

Terry, J. Christopher D., Jinlin Chen, and Owen T. Lewis. 2021. "Natural Enemies

Have Inconsistent Impacts on the Coexistence of Competing Species." *Journal*

of Animal Ecology 1365-2656.13534.

Williams, Stephen E., Elizabeth E. Bolitho, and Samantha Fox. 2003. "Climate

Change in Australian Tropical Rainforests: An Impending Environmental

Catastrophe." *Proceedings of the Royal Society B: Biological Sciences*

270(1527):1887–92.

Wilson, Robert J., David Gutiérrez, Javier Gutiérrez, David Martínez, Rosa Agudo,

and Victor J. Monserrat. 2005. "Changes to the Elevational Limits and Extent of

Species Ranges Associated with Climate Change." *Ecology Letters* 8(11):1138–

46.

Figure 1. Distribution patterns of nine rainforest *Drosophila* species at two study

sites, Mount Kirrama and Mount Paluma. The proportion of samples found at sites at

low (yellow), medium (green) and high (blue) elevations is shown for each species.

Numbers on the bars show the counts of samples for each combination of species

and site.

Figure 2. Thermal performance curves for reproduction of nine *Drosophila*

species. The fitted numbers of adult offspring produced per female parent per day

are plotted against temperature. Line colours indicate species' distribution types, with

upland-biased species represented by cold colours and lowland-biased species

represented by warm colours.

Figure 3. Reproductive and physiological thermal tolerance of seven tropical

Drosophila species. Species are ordered by their distribution patterns, from lowland-

biased species (left) to upland-biased species (right). Cold tolerance is represented

by *RTmin* (a), fecundity at 17°C (b), recovered fecundity after 14°C (c) and recovery

time after chill coma (d). Heat tolerance is represented by *RTmax* (e), fecundity at

29°C (f), recovered fecundity after 29°C (g) and knockdown time at high temperature

(h). Graph (a) and (e) show the medians and 90% credible intervals of the posterior

distribution of the estimated parameters. Boxplots in b-d and f-h show the minimum,

25th percentile, median, 75th percentile, maximum and potential outliers. In (d) and

(h), data for females (shaded) and males (unshaded) are plotted separately.

Figure 4. Interspecific competition for species pairs under upland (left) and lowland

(right) temperature regimes. Each line shows the fecundity of the focal species when

its founder number is kept at four pairs while changing the number of individuals of

the competing species. The identity of the focal species in each pair is indicated by

the colour of the line (see legend) and by the label (e.g. for BIP_PST, the first code

BIP is the focal species and the competitor species is PST). The shaded area

indicates the 90% credible interval of the fitted values under the Beverton-Holt model

for pairwise species competition. PST = *D. pseudotakahashii*, PAL = *D. pallidifrons*,

SUL = *D. sulfurigaster*, BIP = *D. bipectinata*, PAN = *D. pandora*.

Figure 5. Effects of temperature and interspecific competition on population sizes of

D. pallidifrons and *D. pandora*. a) The ending population sizes of *D. pallidifrons* and

D. pandora which were initiated in monoculture (single) or mixed-species culture

(mix) under upland (cold) and lowland (hot) temperature regimes. b) the posterior

distributions of the effect of high temperature (coefficient of temperature) when the

indicated species were maintained alone (single) or with the other species (mix). c)

the posterior distributions of the effect of competition (coefficient of competition)

when the indicated species were maintained at upland (cold) and lowland (hot)

temperature regimes.

Table 1. Estimated parameters of thermal performance functions and their 90% credible intervals (ci90) of the nine species. *RTmin*

and *RTmax* are the minimum and maximum temperatures for the species to reproduce. *a* is the scaling factor and *b* is the shape

factor of the Briere2 function.

species	a	a.ci90	b	b.ci90	RTmin	RTmin.ci90	RTmax	RTmax.ci90
D. bipectinata	0.0046	0.0031 - 0.0060	1.27	1.02 - 1.55	15.28	14.54 - 15.87	30.44	30.08 - 31.05
D. birchii	0.0034	0.0022 - 0.0056	1.17	0.95 - 1.56	13.45	13.07 - 13.77	29.27	28.12 - 29.81
D. bunnanda	0.0016	0.0012 - 0.0025	0.88	0.81 - 1.06	14.57	14.10 - 15.18	31.21	30.60 - 31.78
D. melanogaster	0.0037	0.0032 - 0.0042	1.72	1.48 - 2.03	8.29	6.89 - 9.37	32.13	32.03 - 32.28
D. pallidifrons	0.0073	0.0056 - 0.0098	1.74	1.37 - 2.38	16.23	15.54 - 16.75	29.06	28.19 - 29.38
D. pandora	0.0052	0.0037 - 0.0065	1.26	1.03 - 1.51	15.25	14.58 - 15.80	30.13	29.87 - 30.58
D. pseudoananassae	0.0053	0.0035 - 0.0071	1.68	1.24 - 2.32	15.06	14.17 - 15.89	29.22	28.38 - 29.77
D. simulans	0.0035	0.0027 - 0.0046	1.69	1.36 - 2.21	8.51	6.94 - 9.64	31.09	30.39 - 31.78
D. sulfurigaster	0.0040	0.0028 - 0.0051	1.27	1.03 - 1.52	14.37	13.91 - 14.96	30.11	29.84 - 30.63

942 **Table 2.** Fitted values of the parameters of the competition model and predicted equilibrium states for pairwise interspecific

943 competition. R_0 is the reproductive rate, a is a constant defining the form of the density-dependence relationship, β is the

944 interspecific competition coefficient, and ci90 represents the 90% credible intervals of each parameter. The equilibrium states of the

945 focal species are inferred based on Hassell and Comins (1976).

Temperature	Focal species	R_0	R_0.ci90	a	a.ci90	Competitor	β	β.ci90	Equilibrium state of the focal species
Cool, upland	BIP	11.36	8.06-15.21	0.05	0.02-0.09	PAL	2.26	1.4-4.08	excluded
						PAN	0.99	0.47-1.95	excluded
						PST	0.47	0.12-1.09	stable coexistence
						SUL	2.95	1.91-5.25	excluded
	PAL	27.94	19.52-38.66	0.42	0.27-0.64	BIP	0.3	0.15-0.49	dominant
						PAN	0.32	0.14-0.52	dominant
						SUL	1.22	0.9-1.62	unstable coexistence
	PAN	13.68	10.4-17.85	0.07	0.04-0.12	BIP	0.74	0.33-1.36	dominant
						PAL	3.41	2.26-5.59	excluded

	PST	6.27	3.4-10.66	0.08	0.03-0.19	BIP	0.79	0.35-1.76	stable coexistence
	SUL	20.96	14.27-31.13	0.25	0.14-0.44	BIP	0.41	0.19-0.67	dominant
						PAL	1.05	0.71-1.53	unstable coexistence
Warm, lowland	BIP	15.35	12.51-19.05	0.07	0.05-0.11	PAL	0.29	0.07-0.63	dominant
						PAN	0.87	0.54-1.35	stable coexistence
						PST	0.31	0.07-0.63	dominant
						SUL	1.35	0.93-2	excluded
	PAL	0.99	0.19-2.37	0.12	0.02-0.46	BIP	6.81	2.98-22.27	unable to establish
						PAN	2.99	1.52-8.93	unable to establish
						SUL	3.98	1.77-12.83	unable to establish
	PAN	17.18	14.2-21.24	0.09	0.06-0.14	BIP	0.27	0.08-0.51	stable coexistence
						PAL	0.11	0.01-0.29	dominant
	PST	0	\	\	\	BIP	\	\	unable to establish
	SUL	13.65	10.26-19.03	0.15	0.09-0.24	BIP	0.37	0.18-0.62	dominant
						PAL	0.04	0-0.14	dominant

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