Title: Climate and competition combine to set elevational distributions of tropical

rainforest *Drosophila*

Running head: biotic and abiotic effects on distributions

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Abstract

Species turnover with elevation is a widespread phenomenon and provides valuable

information on why and how ecological communities might reorganize as the climate warms.

Tropical mountains typically have pronounced thermal gradients and intense species interactions,

providing a testing ground for investigating the relationship between thermal tolerances and

biotic interactions as the proximate factors influencing species' distributions. We investigated

temperature and interspecific competition as causes of species turnover and abundance changes

of the nine most abundant species of *Drosophila* along elevational gradients in the Australian

Wet Tropics. Thermal performance curves revealed that species' distributions were better

explained by their performance at extreme temperatures, rather than their thermal optima. Upper

thermal limits varied less among species than lower thermal limits. Nonetheless, these small

differences were associated with differences in centred elevation of distribution, consistent with

environmental sorting as a driver of community composition at low-elevation sites. In contrast,

community composition at cool, high elevations was driven by temperature-dependent

interspecific competition rather than tolerance to low temperatures. These results run counter to

common assumptions about the role of abiotic and biotic factors in structuring communities

along thermal gradients, and indicate that tropical insects may be highly vulnerable to future

warming. Our study illustrates the importance of experimental, quantitative tests across

biological levels (i.e., individuals to populations) and temporal scales (i.e., within-generation to

multi-generation) for characterizing effects of climate on a guild of closely-interacting species.

Keywords

Species distribution, climate change, *Drosophila*, tropics, thermal tolerance, thermal

performance curve, biotic competition, coexistence.

INTRODUCTION

Temperature has a fundamental impact on the reproduction, survival, growth, and behaviour of organisms (Huey and Kingsolver 1989; Huey and Stevenson 1979), strongly influencing species' ranges and abundances (Hoffmann and Blows 1994; Wilson et al. 2005). As a result, estimates of thermal tolerances based on laboratory assays or species' distributions have been used widely to evaluate species' sensitivity to climate change (Deutsch et al. 2008; Kearney and Porter 2009). Tropical ecosystems comprise many species that live close to their upper thermal limits (Deutsch et al. 2008; Diamond et al. 2012; Huey et al. 2009), and which may not tolerate or adapt to warmer temperatures (Bonebrake and Deutsch 2012; Kellermann et al. 2012). The narrow thermal ranges of tropical insects (Khaliq et al. 2014) also mean that they will need to undertake relatively large latitudinal or elevational range shifts to track their climate envelopes as the climate warms, increasing the risk of extinction and community disassembly (Colwell et al. 2008; Sheldon, Yang, and Tewksbury 2011).

Despite high relevance for understanding species' responses to climate change in tropical ecosystems (Corlett 2012), it remains questioned whether temperature is the major proximate factor setting the position of the 'warm' (low-latitude or low-elevation) limits of species' distributions. Global analyses found smaller distribution shifts in response to warming at warm range limits than at cool range limits (Chen et al. 2011; Sunday, Bates, and Dulvy 2012), and some laboratory assays have found similar upper thermal limits for tropical and temperate species (MacLean et al. 2019; Nowrouzi et al. 2018; Overgaard, Kearney, and Hoffmann 2014). Instead, other abiotic factors (e.g. precipitation) or biotic interactions may be critical in setting range limits in the tropics (Engelbrecht et al. 2007; Louthan, Doak, and Angert 2015).

A common expectation, first expressed by Charles Darwin, is that tolerance to low

temperatures will set cool range limits, while biotic interactions will more often set warm range

limits (O'Brien et al. 2017; Paquette and Hargreaves 2021). However, a recent analysis of data

for 654 taxa found that biotic factors became less important in determining warm limits towards

the tropics, while abiotic factors remained consistently important across latitudes (Paquette and

Hargreaves 2021), highlighting the regional differences in the relative roles of abiotic and biotic

factors on local thermal gradients.

There remains limited empirical evidence on the proximate factors setting range limits in

species-rich tropical communities (Feeley, Stroud, and Perez 2017; Jankowski et al. 2013), and

most such studies examine only the correlation between thermal traits (such as critical

temperatures and optimal temperatures) and species' distributions (Amundrud and Srivastava

2020; Cahill et al. 2014; García-Robledo et al. 2016; von May et al. 2017; Nowrouzi et al. 2018;

Pintanel et al. 2021). A significant correlation between tolerance and distribution does not rule

out a role for biotic interactions: differing sensitivities of reproductive performance to

temperature can further increase the competitive advantage of tolerant species over less tolerant

species, sometimes driving qualitative changes in persistence (Lyu and Alexander 2022).

Additionally, the abundance and distribution of a species may be sensitive to temperature change

because of the thermal sensitivities of competitors, hosts, consumers, and other interacting

species (Gifford and Kozak 2012; Merrill et al. 2008; Van Der Putten, Macel, and Visser 2010).

In studies of species' distributions along thermal gradients, niche underfilling has been often

used to indicate the influence of biotic interactions (Chick et al. 2020; O'Brien et al. 2017), but

biotic interactions such as competition, have rarely been measured directly. A coupled

characterization of fundamental thermal performance and temperature-dependent biotic

interactions within a complete guild along an environmental gradient is lacking. Such studies

can help unify the long-separate concepts of environmental and biotic filters (HilleRisLambers et

al. 2012), as well as provide realistic information on thermal sensitivities to inform conservation

efforts in the face of climate change.

To investigate the roles of thermal tolerances and biotic interactions in influencing

species' range limits and structuring communities, we focused on the community of *Drosophila*

flies occupying mountain rainforest habitats in northeastern Australia. These and other tropical

mountains provide natural environmental gradients to test the sensitivity of species to

temperature (Corlett 2011), with pronounced changes in species composition with elevation for

many taxa (Williams, Bolitho, and Fox 2003). We hypothesized that species turnover along the

elevational gradient would result from thermal constraints at cool, high-elevation sites, but from

competitive exclusion at warmer, lowland sites. We first quantified species' elevational

distributions in the field, and then measured thermal performances and competitive outcomes to

test the following predictions. (1) Species with relatively low abundance at upland sites

("lowland species") would be the least cold-tolerant, but (2) species with relatively low

abundance at lowland sites ("upland species") would not be the least heat-tolerant. We further

predicted that (3) species are competitively excluded from local assemblages by locally-abundant

species under the thermal conditions typical at low elevations, but not under high elevation

conditions.

METHODS

We collected data on species' elevational distributions in the field; their reproductive

performance on thermal gradients and adults' resistance to acute thermal stress (by knockdown

assay); and the short-term and long-term outcomes of interspecific competition under low and

high temperatures. All statistics mentioned in this section were performed with R version 4.0.3

(R Core Team, 2020). Original data are archived on Zenodo (10.5281/zenodo.6400611) and code

for analyses is available on GitHub (https://github.com/Jinlinc/DistributionRegulators.git).

Species distributions along elevational gradients

We used a standardized method to survey the relative abundance of different *Drosophila*

species, allowing us to calculate indices of elevational distributions. These were used in later

sections to test whether thermal traits correlate with distribution patterns.

Field survey

Field data on *Drosophila* communities were collected from rainforest sites at Paluma

Range (S18° 59.031' E146° 14.096') and Kirrama Range (S18° 12.134' E145° 53.102'),

Queensland, Australia. These sites lie within the Wet Tropics bioregion which has high levels of

endemism associated with cool and moist upland refugia (Williams et al. 2003). Abundances of

Drosophila at these sites peak from March to June. *Drosophila* pupae were sampled using bottle

traps baited with fermented banana from 11th March – 12th April 2016 for three sites at elevations

of 70m, 350/390m, and 730/880m (subsequently referred to as low-, mid- and high-elevation,

respectively) on each of the two mountains. At each site, we randomly sorted 182 *Drosophila*

pupae from 15 traps (five traps of 12, 15 or 24-day exposure time) hanging at the understory

level. From the 1092 pupae sent for sequencing, 716 pupae in total were successfully identified

to species by DNA metabarcoding, with 86 – 134 pupae at each site (Jeffs et al. 2021). Two

infrequent species, *Drosophila serrata* (1 pupa) and *D. immigrans* (4 pupae), were excluded

from analyses.

Distribution analysis

Two methods were used to assess elevational distribution for individual species. First, the

probability of detecting a particular *Drosophila* species among other co-occurring *Drosophila*

species was modelled as a function of *elevation*, *mountain* and their interaction in a generalized

linear model (family = "binomial"). For each species, the response variable was 1 if the pupa was

identified as the focal species and 0 if it was identified as any other species. Second, since none

of the species had a bimodal distribution with elevation, abundance-weighted mean elevation

(*hIndex*) was used as a simple alternative method to quantify distributions. The location of each

sample was assigned a value of 0, 0.5, or 1 if it was collected at low-, mid- or high-elevation

sites, respectively. *hIndex* was calculated for each species by averaging these values for samples

from both mountains. Correspondence between the two measures was tested by Spearman's rank

correlation test between the coefficient value for *elevation* in the regression and *hIndex*.

Maintenance of laboratory cultures

To minimize adaptation to laboratory conditions, we maintained the *Drosophila* species

by isofemale lines and then crossed them into mass-bred lines (MBLs) shortly (4-6 generations)

before measuring thermal traits. The isofemale cultures were established in 2017 and 2018 from

adults collected from high- and low-elevation sites. Cultures were maintained initially at 24°C at

the Biology Centre, Czech Academy of Sciences, and then transferred to the Department of

Zoology, University of Oxford, UK, in December 2018, where they were maintained at 25°C.

Cultures and all experiments mentioned below were maintained under a 12h/12h light/dark cycle.

There were approximately 15 to 30 non-overlapping generations in the Czech Republic and four

to seven non-overlapping generations in Oxford before establishing mass-bred lines.

Each MBL is made of four isofemale lines (except for *D. pandora*, where only three

isofemale lines were available). Previous studies have found no local adaptation to elevations

and high gene flow between mountains (O'Brien et al. 2017; Schiffer et al. 2007). To reflect the

average value of traits, the four isofemale lines were from different mountains and elevations if

possible (Supplementary Table 1). A large population of each species was initiated using two

independently-reared MBLs. These mass-bred populations were maintained at 25°C until 2020,

and at 23°C thereafter. Measurements taken from these mass-bred populations should not have

been influenced by maternal effects, acclimation, or isofemale line effects.

Eight *Drosophila* species (*D. bipectinata*, *D. birchii*, *D. bunnanda*, *D. pallidifrons*, *D.*

pandora, *D. pseudoananassae*, *D. simulans*, *D. sulfurigaster*) sourced from the Queensland

tropical rainforest, plus a laboratory population of *D. melanogaster* (wild type, *Dahomey* strain)

were measured for cold resistance, heat resistance and thermal performance curves (collectively

called thermal traits). *Drosophila melanogaster* does not occur naturally at the study sites, but

was measured as a benchmark for future comparisons. Competitive outcomes under different

temperature regimes were assessed for five species (*D. bipectinata*, *D. pallidifrons*, *D. pandora*,

D. pseudotakahashii, *D. sulfurigaster*). *Drosophila rubida* was not measured because it was

difficult to raise to a large number and to synchronize with the other species. The thermal traits

of *D. pseudotakahashii* were not measured because its population was contaminated by another

species at the time. A new mass-bred population crossed by two uncontaminated isofemale lines

was constructed later for the competition experiment.

Reproductive thermal performance

We exposed adult *Drosophila* to seven temperatures ranging from 14°C to 32°C and

measured how their reproductive success changed with temperature (procedures are detailed in

Supplementary Figure 1A). We extracted reproductive thermal traits from the thermal

performance curves and examined their correlations with centred elevations of distribution.

Experimental measurements

To generate the adult flies for measurements, fly eggs collected from the population cage

were reared at low density (<100 eggs per vial) at 25°C. Emerging adults were separated by sex

within 12 hours to guarantee that they were unmated. Rearing was started on different days for

different species to synchronize the first day of egg-laying. Two additional vials with five pairs

of flies were monitored daily for sexual maturation, indicated by egg laying. Two days after the

first observation of egg-laying in both vials, two virgin females and two virgin males were paired

in a new vial containing 4ml *Drosophila* medium (weight/volume concentration: 8% corn flour,

4% yeast, 5% sugar, 1% agar, and 1.67% methyl-4-hydroxybenzoate). Vials were randomly

assigned to water baths set at one of seven constant temperatures (14°C, 17°C, 20°C, 23°C, 26°C,

29°C, 32°C). This range of temperature spans the temperatures our *Drosophila* usually

experience in the field. For each species and each temperature treatment, eight replicates were

evenly split between two experimental blocks, making up a total of 1512 vials. To measure

productivity at different temperatures, vials were submerged with the water level kept above the

zone within which flies could freely move. The temperature and relative humidity of vials in

each water bath was monitored within two empty tubes placed at the centre and the corner.

Observed temperatures varied ±0.5°C around the set temperature. Temperatures in the centre of

the water bath were on average 0.5°C higher than at the corners; the average of the former during

the experimental period was used as the corrected temperature in analyses. Relative humidity

levels were similar to field conditions, ranging between 80% - 95%.

Flies in the experimental vials were left to lay eggs on fresh food for 48 hours (vials from

the 1st and 2nd days). They were then transferred to new food and kept in their corresponding

water baths for a further four days before being put onto new food for another 48 hours (vials

from the 7th and 8th days). Vials containing eggs from the two 48-hour periods were kept at the

corresponding temperature until adults emerged. Productivities for the two periods are shown

separately in Supplementary Figure 1B. We averaged these values to reflect the relative

reproductive success in early adult life (within 8 days of sexual maturity).

After eight days in water baths, all flies were returned to 25°C, a favourable temperature,

in new vials with fresh food for a further four days before being sacrificed. Their offspring

developed at 25°C until eclosion. These adult offspring numbers reflect the recovered

reproduction following eight-day exposure to different temperatures. The numbers of surviving

parents were recorded when flies were transferred onto new food on the 3rd, 7th, 9th, and 13th day.

Offspring numbers were counted 5 – 7 days after the first emergence was observed. The

experiment was conducted from May to August 2019.

Thermal performance curves

A multi-level, non-linear piecewise model was fitted to describe how reproductive

success changed with temperature for all *Drosophila* species. We used the Briere2 function

(Briere et al. 1999) to describe how reproductive performance (square-rooted daily productivity

per female) changed with temperature:

$$\widehat{P(\iota,T)} = \begin{cases} a_i \times T \times (T - RTmin_i) \times (RTmax_i - T)^{1/b_i} & (if\ RTmax_i > T > RTmin_i) \\ 0 & (if\ T \leq RTmin_i\ or\ T \geq RTmax_i) \end{cases} \quad (1)$$

where $\widehat{P(\iota,T)}$ is the theoretical reproductive performance of species *i* at temperature *T*, $RTmin_i$

and $RTmax_i$ are the minimum and maximum temperatures for species *i* to reproduce, a_i is a

scaling factor, and b_i is a shape factor of the curve for species *i*.

The observed reproductive performance, $P(i,T)$, was modelled assuming a Gaussian

distribution of errors as shown in Equation (2). A Gaussian distribution is not ideal to model

transformed count data, which are all positive. However, models using untransformed counts

with Poisson, zero-inflated Poisson, negative binomial, or lognormal distributions did not

adequately converge.

$$P(i, T) \sim Gaussian(\widehat{P(i, T)}, \sigma_{P(T)}{}^2) \ (2)$$

227   As standard deviations, $\sigma_{\sigma_P(T)}{}^2$, tend to get smaller as temperature approaches the critical

228   points (beyond which performance is zero), we assumed seven different $\sigma_{P(T)}$ for our seven test

229   temperatures. They follow:

$$\sigma_{P(T)} \sim Gaussian \ (\mu_{\sigma_P}, \sigma_{\sigma_P}{}^2) \ (3)$$

where mean, μ_{σ_P}, and variance, $\sigma_{\sigma_P}{}^2$, are hyperparameters that describe the mean and variance

of $\sigma_{P(T)}$.

$RTmin_i$, $RTmax_i$, a_i and b_i of the Briere2 function are features of *Drosophila* species. We

assumed that each value showed some similarities as they are all *Drosophila* species. To reflect

this structure, $RTmin_i$, $RTmax_i$, a_i and b_i are modelled by equations (4) – (7). Their means and

variances are also hyperparameters defined by priors. Additionally, the values of a_i and b_i were

bounded to be positive. The values of $RTmin_i$ were bounded to be lower than 17°C and the values

of $RTmax_i$ were bounded between 26°C to 35°C based on prior knowledge of the range of

temperatures under which our species can reproduce.

$$RTmin_i \sim Gaussian(\mu_{RTmin}, \sigma_{RTmin}{}^2) \ (4)$$

$$RTmax_i \sim Gaussian(\mu_{RTmax}, \sigma_{RTmax}{}^2) \ (5)$$

$$a_i \sim Gaussian(\mu_a, \sigma_a{}^2) \ (6)$$

$$b_i \sim Gaussian(\mu_b, \sigma_b{}^2) \ (7)$$

Prior distributions of hyperparameters μ_{RTmin}, μ_{RTmax}, μ_a, μ_b, and μ_{σ_P} are Gaussian

distributions with reasonable means (15°C, 30°C, 0, 0, 0) and large standard deviations (10, 10, 1,

10, 10) compared with the realistic range of these parameters in our species. The prior

distribution of the hyper parameters $\sigma_{RTmin}{}^2$, $\sigma_{RTmax}{}^2$, $\sigma_a{}^2$, $\sigma_b{}^2$, and $\sigma_{\sigma_P}{}^2$ is inverse-gamma

(0.001, 0.001), which is a commonly used non-informative distribution for prior of variance.

The multi-level model was fitted under a Bayesian framework using MCMC sampling by

the *rstan* package (Stan Development Team 2021). Models converged and performance was

acceptable in diagnostic plots (Supplementary Figure 2). Medians of the posterior distributions

for $RTmin_i$, $RTmax_i$, a_i and b_i were used to construct the thermal performance curve of species *i*.

Optimal temperatures for peak reproduction (*RTopt*) were deduced from known parameters:

$$RTopt = \frac{2bRTmax + (b+1)RTmin + \sqrt{4b^2 RTmax^2 + (b+1)^2 RTmin^2 - 4b^2 RTminRTmax}}{4b+2} \quad (8)$$

Testing for associations between reproductive thermal traits and species distributions

Among the tropical *Drosophila* species, the medians of the posterior distributions of

RTmin, *RTmax*, and *RTopt* were modelled as a function of *hIndex* in a linear model with

phylogenetic correction. The phylogenetic tree was derived with modification from Finet et al.

2021 (Supplementary Figure 3). Productivity at 29°C and 17°C, and recovered productivity after

29°C and 14°C, were used as direct measurements of performance at high or low temperatures.

These temperatures were chosen because they are known to cause stress to the reproduction of

the focal *Drosophila* species, and correspond to realistic understory air temperatures during the

hottest and coolest months. These productivities were modelled as a function of *hIndex* and

experimental block as fixed effects, and *species* as a random effect in generalized linear mixed-

effect models (family = "negative binomial") with phylogenetic correction, using the *brms*

package (Bürkner 2021). *Drosophila simulans* has not been found in the standardized assay,

therefore, it was not included in the regression. It is potentially a lowland-biased species because

we only found their isofemale lines from lowland sites when we tried to collect as many

isofemale lines as possible from different elevations. Adding *D. simulans* as a lowland-only

species into the regression will not change the conclusions.

Thermal knockdown

We measured the physiological tolerance of each *Drosophila* species to extremely cold

and hot environments, and then examine their correlations with distribution patterns.

Experimental measurements

Resistance to extreme cold temperature was measured as knockdown time for each

individual at 5°C and the time for recovering muscle coordination after a 30-minute exposure to

5°C. The constant temperature chosen for cold stress studies is often around 0°C (Gibert et al.

2001). As tropical species often have significantly weaker cold resistance (Gibert et al. 2001),

5°C was used instead to increase the variation among the tested species after pilot trials. Heat

stress was chosen to be 40°C, which follows common practice for *Drosophila* species

(Hoffmann, Sørensen, and Loeschcke 2003) and is expected to capture the between-species

variance in heat tolerance over a timescale which is convenient to measure (Jørgensen, Malte,

and Overgaard 2019). After being knocked down by heat (40°C), most flies did not survive. In

this case, only knockdown time was used to evaluate resistance to heat. The knockdown

experiments were conducted from May to Jun 2019.

Virgin adult flies (siblings of those used for reproduction measurements) were kept in

same-sex groups at 25°C for 9-10 days before knockdown assays. Assays were conducted for

male and female flies separately. An observation rack was divided into nine (3x3) cells. Each

cell was randomly assigned one of the nine *Drosophila* species and held seven flat-bottomed 3ml

glass vials, each with a randomly-selected individual of the allocated species. One set of

observations on such a set-up represents a single block. We repeated measurements for three

experimental blocks, and the allocation of species to cells was redrawn for each block. In total,

we measured 21 individuals per sex per species. During measurement, the observation rack was

moved immediately into the incubator pre-set at 5°C or 40°C. Every tube was examined once

every minute and flies that lost their ability to stand in that minute were recorded. After exposure

to 5°C for 30 minutes, all flies were in a chill coma. The observation rack was moved to a 25°C

room. Flies were left undisturbed and the time taken until each fly regained its ability to stand

was recorded.

Testing for associations between knockdown resistance and species distributions

The knockdown time by the heat, knockdown time by the cold and recovery time from

the cold of each sex were first compared among species using ANOVA. When interspecific

variation was observed, they were modelled as a function of *hIndex*, *block*, and *cell position* as

fixed effects, and *species* as a random effect in linear mixed-effect models with phylogenetic

correction using the *brms* package. *Species* was included as a random effect to account for

repeated measures.

Short-term competition

Pairs of *Drosophila* species were reared together in the same vials for one generation to

evaluate how interspecific competition affects the observed reproductive success and predicted

coexistence patterns under temperature regimes typical of low-elevation and high-elevation sites.

Experimental design

We used incubators set at alternating temperatures mimicking average day-time and

night-time temperatures in February at high-elevation (23°C / 21°C) and low-elevation sites

(28.5°C / 24°C) (Supplementary Figure 4). Adults to establish the competition experiment were

reared in bottles (30ml fresh medium) at moderate density (300 – 500 per bottle) at their testing

temperatures. After eclosion, individuals that emerged within 48 hours were kept together in

mixed-sex containers. Two days after the first observation of egg-laying, adults of different sexes

were separated, and from the following day were allowed to pair and lay eggs in new

experimental vials (5ml medium) for two days, without or with another competitor species.

Five species were chosen as representative species for upland-biased, elevation-generalist

and lowland-biased distribution types (see Results for the definition of distribution types). The

two-species combinations were lowland species vs. upland species: *D.bipectinata* vs. *D.*

pallidifrons, *D.bipectinata* vs. *D. pseudotakahashii*, *D.pandora* vs. *D. pallidifrons*; lowland

species vs. lowland species: *D. bipectinata* vs. *D.pandora*; lowland species vs. elevation-

generalist: *D. bipectinata* vs. *D. sulfurigaster*; upland species vs. elevation-generalist: *D.*

pallidifrons vs. *D. sulfurigaster*. Each combination was tested at different founding densities in a

factorial design: (4 pairs of species A, 2 pairs of species B), (4A, 4B), (4A, 8B), (2A, 4B) and

(8A, 4B). We also included monocultures of each species with 2, 4, and 8 pairs. Each density and

species combination was replicated ten times across two or three blocks staggered by two days

(two blocks for the *D. pandora* vs. *D. pallidifrons* combination; three blocks for the other five

pairs). Offspring that successfully developed to adulthood were identified to species and counted.

The experiment of *D. pandora* vs. *D. pallidifrons* was conducted from September to December

2020 and the other five pairs were experimented with from January to March 2021.

Short-term competitive outcomes

We used the Beverton-Holt model to describe the population growth of a single

generation of flies on discrete and temporary resources:

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$$N_{focal,1} = \frac{R_0 N_{focal,0}}{1 + a(N_{focal,0} + \beta N_{competitor,0})} \quad (9)$$

Where $N_{focal,0}$ is the number of parents and $N_{focal,1}$ is the number of offspring of the focal

species. $N_{competitor,0}$ is the number of parents of the competing species. R_0 is the generational

reproduction rate and a is a constant defining the form of the density-dependence relationship. β

represents the interspecific competition coefficient of the competitor species to the focal species,

and defines the equivalence between the two competing species. Offspring numbers of the focal

species were modelled assuming a negative binomial error distribution, under a Bayesian

framework using MCMC sampling by the *rstan* package, as described by Terry et al. (2021).

Model diagnostics are shown in Supplementary Figure 5A-B. Raw data and fitted curves of each

pair are shown in Supplementary Figure 6. Median values of posterior distributions were used as

parameters to infer the equilibrium states of each pair following Hassell and Comins (1976).

Long-term competition

To validate the long-term population-level outcomes of asymmetrical competition shown

in the short-term competition experiment, we raised one lowland species and one upland species

in monoculture and mixed-species culture for multiple overlapping generations under

temperature regimes typical of low-elevation and high-elevation sites. We then evaluated the

long-term effects of temperature and competition on population sizes.

Experimental design

Four monocultures of each species and eight mixed-species cultures were maintained at

each temperature regime for 13 weeks, totalling 32 cultures. The cultures were evenly divided

into two blocks starting on different dates. Monocultures were started with ten pairs of

individuals. Mixed-species cultures were started with ten pairs of individuals of each species.

The starting density was very low compared to the equilibrium density. Each culture was

maintained in a series of five bottles (30ml fly medium) following Ayala, Gilpin, and Ehrenfeld

(1973). At the start of each week, adults surviving in the most recent bottle and adults which

were freshly emerged in the older four bottles were separately collected, photographed and

transferred together into a new bottle with fresh food. In this way, adult survival and

reproduction were recorded separately. As the total population sizes were relatively stable, they

were only counted when the populations were terminated after 13 weeks of maintenance. To

avoid pseudo-replication introduced by 'incubator' effects, the two incubators were switched

between temperature regimes every week, with their contents moved accordingly. Trays were

shuffled inside the incubator every two days. Temperature and humidity were recorded and the

temperature regimes were confirmed during and at the end of experiments. The experiment was

conducted from September to December 2020.

Testing temperature and competition effects on long-term population persistence

Population sizes were modelled as a function of *temperature*, *species*, *presence/absence*

of competitors, and their interactions, with *culture ID* as a random effect in a generalized linear

mixed-effect model (family = "zero-inflated negative binomial") using the *brms* package. Model

diagnostic is shown in Supplementary Figure 5C.

RESULTS

Field distributions

The numbers of samples found at low-, medium- or high-elevation sites for each of the

nine major *Drosophila* species (accounting for 99% of all samples) are shown in Figure 1.

Distributions quantified using regression and by centred elevation (*hIndex*) were consistent

(Spearman's rank correlation rho = 0.93, p value = 0.0007). Among the three elevations sampled,

the detection probability of *D. bipectinata* and *D. pandora* increased monotonically towards

lower-elevation sites and were thus categorized as lowland-biased species with high confidence.

Drosophila pseudoananassae showed a lowland bias on one of the two mountains, and a mid-

elevation peak on the other. *Drosophila rubida*, *D. sulfurigaster* and *D. birchii* showed no

significant change with elevation and were thus defined as elevation generalists. *Drosophila*

pallidifrons and *D. pseudotakahashii* were significantly more likely to be found at high

elevations and were thus defined as upland-biased species. Coefficients and p values of the

regressions and *hIndex* values are shown in Supplementary Table 2. For *D. bunnanda,* the six

records were insufficient for model fitting but all occurred at lowland sites, and data from a

large-scale study suggest that it is a lowland-biased species (Schiffer and McEvey 2006).

Thermal performance curves

Thermal performance curves of daily productivity per female parent vary among species

in terms of the range, optimal temperature, peak productivity, and shape factors (Figure 2; Table

1; see Supplementary Figure 2C for original data and fitted curves for each species). The

temperature for optimal reproductive performance, *RTopt*, did not correlate with *hIndex*

(Coefficient = 0.09, 95% CI (credible interval) = -2.83 – 3.01; Figure 2b). Cold tolerances,

*RTmin*s, varied more among species than heat tolerances, *RTmax*s (standard deviation: 2.82

versus 1.01, respectively). There was no general trade-off between *RTmin* and *RTmax*

(Spearman's rank correlation test: p value = 0.10; Figure 2c). For example, *D. sulfurigaster*

outperforms its upland-biased relative, *D. pallidifrons*, across the temperature range.

Testing correlations between cold tolerance and distribution

*RTmin*s were not correlated with species distribution patterns (Figure 3a. Coefficient = -

0.41, 95% CI = -4.15 – 3.43). Similarly, upland-biased species did not show higher productivity

at the low temperature, 17°C (Figure 3b; Coefficient = -0.27, 95% CI = -3.80 – 3.05). When

exposed to acute sublethal low temperature (5°C), all seven tropical *Drosophila* species showed

similarly poor performance compared to *D. simulans* and *D. melanogaster* (Supplementary Table

3). Most species did not reproduce during their eight-day exposure to 14°C (Supplementary 1B),

however, all of them were able to resume productivity once transferred back to 25°C. Species

with a higher centred elevation of distribution had slightly but non-significantly higher recovered

productivity (Figure 3c; Coefficient = 0.33, 95% CI = -0.55 – 1.14). It took longer, but not

significantly so, for upland species to regain muscle coordination after chill coma (Figure 3d;

Male: coefficient = 13.04, 95% CI = -9.31 – 35.05; Female: coefficient = 8.49, 95% CI = -2.18 –

20.02)

Testing correlations between heat tolerance and distribution

Species whose distribution were more biased towards lowland have higher critical

temperatures for reproduction, *RTmax*s (Figure 3e; Coefficient = -3.06, 95% CI = -5.30 – -0.88).

Reproductive performance at 29°C also decreased with *hIndex* (Figure 3f; Coefficient = -5.80, 95%

CI = -9.37 – -2.50). After exposure to 29°C for eight days, the two species with the highest

hIndex could not reproduce when transferred back to 25°C, while four out of the other five

elevation-generalist and lowland-biased species resumed reproduction (Figure 3g). Knockdown

time at lethally high temperature (40°C) was shorter among species of higher *hIndex* (Figure 3h;

Male: coefficient = -6.71, 95% CI = -13.70 – 0.50; Female: coefficient = -1.99, 95% CI = -9.48 –

5.71), indicating these species lose their muscle coordination faster at high temperatures.

Effects of interspecific competition at upland and lowland temperatures

When raised in a laboratory condition mimicking the warmer, lowland sites, reproductive

success was highest for the two lowland-biased species, followed by the elevation-generalist

species, *D. sulfurigaster*. The two upland species could barely reproduce regardless of the

presence of competitors (Figure 4; Table 2: proliferation rates $R_0 < 1$). *Drosophila*

pseudotakahashii, whose distribution was most constrained to upland sites, had an even lower

proliferation rate at high temperatures than *D. pallidifrons*. Their competitive effect on lowland

species was minimal, indicated by low *β* values (Table 2). The two lowland species, *Drosophila*

pandora and *D. bipectinata*, are expected to coexist stably in lowland, based on their

reproductive and competitive parameters.

When raised under a cooler, upland condition, all species could reproduce and sustain

their populations (Table 2: all R_0s were higher than 1). Lowland species were strongly affected

by the density of *D. pallidifrons*, an upland species (shown by large *β* values), while upland

species were significantly less affected by lowland species. Competition with *D. pallidirons*

under upland conditions was predicted to drive *D. pandora* and *D. bipectinata* to exclusion

(Table 2).

In the long-term competition experiment (Figure 5), high temperature drove the upland-

species *D. pallidifrons* to extinction both in monocultures (except in one replicate, where two

adults were found), and when reared together with the lowland competitors. In contrast, the

monoculture of the lowland species *D. pandora* remained abundant at both low and high

temperatures (temperature's effect: 95% CI = -0.26 – 0.12, overlapping with zero). The presence

of *D. pandora* only slightly decreased the population size of *D. pallidifrons* (median coefficient

of competition = -0.32, 95% CI = -0.14 – -0.49). The presence of *D. pallidifrons* had no

significant effect on the population size of *D. pandora* in warm, lowland temperatures (95% CI =

-0.28 – 0.06), but had a strong negative effect (median coefficient of competition = -1.56, 95%

CI = -1.73 – -1.40) in cool, upland temperatures.

DISCUSSION

Our results do not support the common assumption that cool boundaries to species'
ranges are constrained abiotically (reflecting thermal niches), while biotic interactions (such as
interspecific competition) define warm boundaries. Our study adds to a growing literature for
tropical mountain systems showing significant associations between upper thermal limits and
patterns of species turnover with elevation (Amundrud and Srivastava 2020; García-Robledo et
al. 2016; von May et al. 2017; Pintanel et al. 2021). Noteworthy, this result does not rule out the
potential contribution of competition with warm-adapted species to raising the warm boundaries
to higher elevations. However, the observation that upland species were excluded from the
lowest elevations solely by high temperatures experienced on a daily basis implies that further
warming will likely drive contraction of their ranges directly. In contrast, lowland species were
not worse than upland species at surviving or reproducing at low temperatures. However,
lowland species were outcompeted by one upland species whose distribution was confined to
high elevations as a result of intolerance to heat. This important role of interspecific competition
at cold boundaries on elevational gradients (also shown by Gifford and Kozak 2012; Lyu and
Alexander 2022; Rodrıguez-Castañeda et al. 2017) helps to explain the maintenance of species
richness and endemism in mountain regions.

Abiotic versus biotic limits of distribution

The idea that contributions of abiotic versus biotic factors might differ for species' warm
and cool range boundaries can be traced back to Charles Darwin, and remains an area of active
debate (Cahill et al. 2014; Hargreaves, Samis, and Eckert 2014; Schemske et al. 2009). There is
especially mixed evidence on the importance of upper thermal limits in setting warm boundaries
(positive relationships: Batista et al. 2018; Duarte et al. 2012; García-Robledo et al. 2016;

Kellermann et al. 2012; Merrill et al. 2008; null relationships: Brandt et al. 2020; Gaston and

Chown 1999; Huang and Tu 2008; Kimura 2004; Nowrouzi et al. 2018).

This inconsistency could arise for at least three reasons. First, data available for synthetic

studies over-represent systems from temperate latitudes in the northern hemisphere (Feeley et al.

2017; Parker 2022) and relate largely to cool limits (Cahill et al. 2014). Comparative studies

have shown in lower latitudes that local variations in thermal tolerance have a tighter association

with environmental variations (Duarte et al. 2012), and that species distribution appears more

sensitive to warming (Freeman et al. 2021). Second, mechanisms governing latitudinal or

elevational distribution patterns could differ. Studies of *CTmin* (the critical temperature at which

organisms lose their muscle coordination) have found a tight relationship between this lower

thermal limit and species' latitudinal distributions (Kellermann et al. 2012; Overgaard et al.

2014). In contrast, we found that various cold tolerances of our species, including *CTmin* sourced

from the literature, were not indicative of elevational distribution patterns. Third, some of the

uncertainty might reflect methodological differences when choosing, defining and measuring

thermal traits.

Different choices of indicators of thermal performance, and different experimental

methods may explain why some of our results contrast with some other studies of Australian

rainforest *Drosophila*. Overgaard et al. (2014) found no evidence that the upper thermal limits of

tropical rainforest *Drosophila* differed among species. Unlike Overgaard et al. (2014), we

measured parental productivity by allowing their eggs to develop into adults under the relevant

temperature. We observed that many eggs laid at high temperatures never hatched, perhaps as a

result of sperm sterilization (Parratt et al. 2021). We also observed different speeds of senescence

after exposure to different temperatures (Supplementary Figure 1B), which the three-day period

of fecundity measurement in Overgaard's study would not have captured. Nevertheless, both

studies highlight the conserved nature of heat tolerance and suggest small thermal safety margins

for future warming.

Field experiments on *D. birchii* (O'Brien et al. 2017) found that its single-generation

reproduction in monoculture was highest at low elevations. In our experiments, *D. birchii* was

one of the most heat-sensitive species, and its populations were unable to persist within a few

generations at high temperatures typical of low elevations (pilot experiment, unpublished data).

The single-generation set-up, with lab-prepared mated adults, could not capture the impact of

high temperatures prior to sexual maturation. A further possibility is seasonal climatic changes:

our experimental conditions correspond to February temperatures in the field, when the average

daily maximum temperature reaches 30.6°C at lowland. This is when the rainy season starts,

driving an increase in *Drosophila* abundance and may serve as an annual reset of species'

distributions. The field study of O'Brien et al. occurred in April, which is 2°C lower in average

temperature and average daily maximum. As the weather cools, mid- and low-elevation areas

will become more suitable for the upland species to colonize. The relative importance of abiotic

and biotic factors in affecting species distributions may change seasonally. Spatial and temporal

data on species distributions are not yet available for our system but would provide a further test

of our conclusions.

Low variations in upper thermal limits

Upper thermal limits vary little among species (Hoffmann 2010) and have limited

adaptive potential (van Heerwaarden and Sgrò 2021; Kellermann et al. 2012). Nevertheless,

ecologically meaningful variations do exist, especially in tropical regions (Amundrud and

Srivastava 2020; García-Robledo et al. 2016; von May et al. 2017; Pintanel et al. 2021),

indicating ecological sorting or adaptation. Consistent with other studies (Goulet, Thompson, and

Chapple 2017; Hangartner and Hoffmann 2016), we found that heat tolerance is a systematic

trait manifested in critical temperatures, productivities at sub-sterile temperatures, recovered

productivities, and locomotive responses. Modest variations in critical temperature may signify

ecologically meaningful differences in overall performance under real and variable thermal

conditions.

Such small thermal safety margins suggest a severe threat of biotic attrition in tropical

lowlands (Colwell et al. 2008; Deutsch et al. 2008; Duarte et al. 2012; van Heerwaarden and

Sgrò 2021). Laboratory-measured critical temperatures are sensitive to experimental conditions,

making it difficult to relate the exact values to climatological means or maxima, and hence the

threat of rising temperatures (Sinclair et al. 2016). In this context, our study benefits from a

comparative approach, which reveals that upland species are already constrained by high

temperatures at low elevations, and that a very small difference (about 1°C) distinguishes the

upper thermal limits (*RTmax*s) of lowland and upland species. Given the low evolutionary

potential of heat tolerance (Hoffmann, Chown, and Clusella-Trullas 2013), both lowland species

and upland species are likely to be vulnerable to modest temperature increases across the

elevation gradient. Thus, lowland biotic attrition and upland range contraction are likely to occur

with future warming. This could lead to cascading effects in lowland communities and threaten

endemic upland species on tropical mountains such as those in the Australian Wet Tropics

(Freeman et al. 2018).

Daily peak temperature as the main abiotic filter

Daily maxima can be more important than mean temperatures in structuring distributions

(Lynch et al. 2014). In our study system, the mean temperature during the study season at our

lowland sites is around 26°C. At this temperature, all studied species are close to their peak

reproductive performance. However, upland and lowland sites differ greatly in the average daily

maximum temperature and the daily duration that the temperature reaches or exceeds stressful

levels for *Drosophila* reproduction (Supplementary Figure 7). In a preliminary experiment where

populations were maintained for multiple generations in mixed-species cultures at constant 26°C,

D. pallidifrons always out-numbered *D. pandora* (Jinlin Chen, data not shown), contrasting poor

performance of *D. pallidifrons* at 24°C /28.5°C. Brief exposure to stressful thermal environments

can have similar fitness costs to continuously stressful conditions (Saxon, O'Brien, and Bridle

2018). These results highlight the importance of considering daily temperature variations and

extreme temperature events when studying species distributions and projecting responses to

climate change (Kingsolver, Diamond, and Buckley 2013; Ma, Hoffmann, and Ma 2015).

Limitations

Our laboratory environment has low variations in temperature, spatial structure, and diet,

limiting the mechanisms organisms use to withstand thermal stress and competitive pressure.

Behavioural thermoregulation allows species with distinctive fundamental thermal tolerance to

exist in one area but using different microhabitats (Vives-Ingla et al. 2022). This spatial or

temporal partitioning can also influence the competition between resource-sharing species

(Bestelmeyer 2008; Porras et al. 2020). Furthermore, competition in reproduction and larval

survival may be overrepresented in restricted space inside containers, while differences in

abilities to forage, escape, and use alternative resources were not captured. Nevertheless, our

thermal performance curves and monoculture experiments showed that lowland species could

persist in abiotic conditions outside their observed range. It implies that biotic mechanisms,

likely competitive exclusion, play a role in upland community composition. The match between

reproduction limits of upland species and recorded air temperature in lowlands implies their lack

of ability to mitigate the adverse impact of high temperature on natural populations, such as

direct heat stress on larvae on rotten fruits or deficiency in foraging under heat.

Finally, between-population variation and local adaptation (Hoffmann, Anderson, and

Hallas 2002), especially adaptation to abiotic environments at distribution boundaries (Peterson,

Doak, and Morris 2019), should not be neglected if studying distributions at large spatial scales

(e.g., across latitudes). The relatively restricted spatial scale of our study (within a mountain

range), the use of mass-bred lines sourced from different elevations, and the limited plasticity

and local adaptation documented for our study species (MacLean et al. 2019; O'Brien et al. 2017)

mean that plastic and evolutionary response to thermal conditions is unlikely to complicate

interpretation of our results.

Conclusions

Tropical ecosystems host an exceptional diversity of endemic species (Laurance et al.

2011). Predicting their sensitivity to climate change is a high priority and requires an

understanding of the proximate causes of current distributions as well as the interactions between

thermal tolerance and other environmental factors, both biotic and abiotic. Our study contributes

to the growing literature demonstrating that species are sensitive to high temperatures at the

warm boundaries of their distributions in the tropics. In particular, we highlight the important

role of daily maximum temperatures in structuring communities in tropical lowland sites, and the

essential role of interspecific competition at upland sites. Our results imply the vulnerability of

both upland and lowland communities to increasing temperatures. These results demonstrate that

reliable predictions of the sensitivity of species to rising temperatures may not always be

achieved using simple correlational approaches; instead, a detailed understanding of multiple

thermal traits and interactions with other species may be needed.

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JC and OTL both contributed to the development of ideas. JC designed and conducted the

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Conflict of Interest:

There is no conflict of interest to declare.

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Table 1. Parameters of thermal performance functions and their 90% credible intervals (ci90) of the nine species. *RTmin* and *RTmax*

are the minimum and maximum temperatures for the species to reproduce. *a* is the scaling factor and *b* is the shape factor of the

Briere2 function.

species	a	a.ci90	b	b.ci90	RTmin	RTmin.ci90	RTmax	RTmax.ci90
D. bipectinata	0.0046	0.0031 - 0.0060	1.27	1.02 - 1.55	15.28	14.54 - 15.87	30.44	30.08 - 31.05
D. birchii	0.0034	0.0022 - 0.0056	1.17	0.95 - 1.56	13.45	13.07 - 13.77	29.27	28.12 - 29.81
D. bunnanda	0.0016	0.0012 - 0.0025	0.88	0.81 - 1.06	14.57	14.10 - 15.18	31.21	30.60 - 31.78
D. melanogaster	0.0037	0.0032 - 0.0042	1.72	1.48 - 2.03	8.29	6.89 - 9.37	32.13	32.03 - 32.28
D. pallidifrons	0.0073	0.0056 - 0.0098	1.74	1.37 - 2.38	16.23	15.54 - 16.75	29.06	28.19 - 29.38
D. pandora	0.0052	0.0037 - 0.0065	1.26	1.03 - 1.51	15.25	14.58 - 15.80	30.13	29.87 - 30.58
D. pseudoananassae	0.0053	0.0035 - 0.0071	1.68	1.24 - 2.32	15.06	14.17 - 15.89	29.22	28.38 - 29.77
D. simulans	0.0035	0.0027 - 0.0046	1.69	1.36 - 2.21	8.51	6.94 - 9.64	31.09	30.39 - 31.78
D. sulfurigaster	0.0040	0.0028 - 0.0051	1.27	1.03 - 1.52	14.37	13.91 - 14.96	30.11	29.84 - 30.63

884   **Table 2.** Fitted values of the parameters of the competition model and predicted equilibrium states for pairwise interspecific

885   competition. R_0 is the reproductive rate, a is a constant defining the form of the density-dependence relationship, β is the interspecific

886   competition coefficient, and ci90 represents the 90% credible intervals of each parameter.

Temperature	Focal species	R_0	R_0.ci90	a	a.ci90	Competitor	β	β.ci90	Equilibrium state of the focal species
Cool, upland	BIP	11.36	8.06-15.21	0.05	0.02-0.09	PAL	2.26	1.4-4.08	excluded
						PAN	0.99	0.47-1.95	excluded
						PST	0.47	0.12-1.09	stable coexistence
						SUL	2.95	1.91-5.25	excluded
	PAL	27.94	19.52-38.66	0.42	0.27-0.64	BIP	0.3	0.15-0.49	dominant
						PAN	0.32	0.14-0.52	dominant
						SUL	1.22	0.9-1.62	unstable coexistence
	PAN	13.68	10.4-17.85	0.07	0.04-0.12	BIP	0.74	0.33-1.36	dominant
						PAL	3.41	2.26-5.59	excluded
	PST	6.27	3.4-10.66	0.08	0.03-0.19	BIP	0.79	0.35-1.76	stable coexistence
	SUL	20.96	14.27-31.13	0.25	0.14-0.44	BIP	0.41	0.19-0.67	dominant
						PAL	1.05	0.71-1.53	unstable coexistence
Warm, lowland	BIP	15.35	12.51-19.05	0.07	0.05-0.11	PAL	0.29	0.07-0.63	dominant
						PAN	0.87	0.54-1.35	stable coexistence
						PST	0.31	0.07-0.63	dominant
						SUL	1.35	0.93-2	excluded
	PAL	0.99	0.19-2.37	0.12	0.02-0.46	BIP	6.81	2.98-22.27	unable to establish
						PAN	2.99	1.52-8.93	unable to establish
						SUL	3.98	1.77-12.83	unable to establish
	PAN	17.18	14.2-21.24	0.09	0.06-0.14	BIP	0.27	0.08-0.51	stable coexistence
						PAL	0.11	0.01-0.29	dominant
	PST	0	\	\	\	BIP	\	\	unable to establish
	SUL	13.65	10.26-19.03	0.15	0.09-0.24	BIP	0.37	0.18-0.62	dominant
						PAL	0.04	0-0.14	dominant

 Figure legends

Figure 1. Distribution patterns of nine rainforest *Drosophila* species in Kirrama Range or

Paluma Range. The proportion of samples found at sites at low (yellow), medium (green) and

high (blue) elevations is shown for each species. Numbers on the bars show the counts of

samples for each combination of species and site.

Figure 2. Reproductive thermal traits. (a) Thermal performance curves for reproduction of nine

Drosophila species. The fitted adult offspring number (per female parent per day) are plotted

against temperature. (b) Optimal temperatures for reproduction of the seven tropical *Drosophila*

species, ordered by their *hIndex*. (c) Posterior distribution of *RTmax*s and *RTmin*s of each

Drosophila species. Colours indicate distribution types: cold colours indicate upland-biased

species represented and warm colours indicate lowland-biased species; the grey colour represents

D. melanogaster.

Figure 3. Reproductive and physiological thermal tolerance of seven tropical *Drosophila* species.

Species are ordered by their *hIndex*, from lowland-biased species (left) to upland-biased species

(right). Colours represent distribution types: orange for lowland species, green for elevation

generalist and blue for upland species. Cold tolerance is represented by *RTmin* (a), productivity

at 17°C (b), recovered productivity after 14°C (c) and recovery time after chill coma (d). Heat

tolerance is represented by *RTmax* (e), productivity at 29°C (f), recovered productivity after

29°C (g) and knockdown time at high temperature (h). Graph (a) and (e) show the medians and

90% credible intervals of the posterior distribution of the estimated parameters. Boxplots in b-d

and f-h show the minimum, 25th percentile, median, 75th percentile, maximum and potential

outliers. In (d) and (h), data for females (shaded) and males (unshaded) are plotted separately.

Figure 4. Interspecific competition for species pairs under upland (left) and lowland (right)

temperature regimes. Each line shows the two-day productivity per pair of the focal species (four

pairs in total) while changing the number of individuals of the competing species. The identity of

the focal species in each pair is indicated by the colour of the line (see legend) and by the label

(e.g., for BIP_PST, the first code BIP is the focal species and the competitor species is PST). The

shaded area indicates the 90% credible interval of the fitted values under the Beverton-Holt

model for pairwise species competition. PST = *D. pseudotakahashii*, PAL = *D. pallidifrons*, SUL

= *D. sulfurigaster*, BIP = *D. bipectinata*, PAN = *D. pandora*.

Figure 5. Effects of temperature and interspecific competition on population sizes of *D.*

pallidifrons (upland species, blue) and *D. pandora* (lowland species, red). Each point indicates

the ending population size of each culture (n(monoculture) = 4, n(mixed-species) = 8). Dashed

(monoculture) and solid (mixed-species culture) lines connect the means of population sizes in

the two temperature regimes.

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Paluma Range. The proportion of samples found at sites at low (yellow), medium (green) and
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Figure 3. Reproductive and physiological thermal tolerance of seven tropical *Drosophila* species. Species are ordered by their *hIndex*, from lowland-biased species (left) to upland-biased species (right). Colours represent distribution types: orange for lowland species, green for elevation generalist and blue for upland species. Cold tolerance is represented by *RTmin* (a), productivity at 17°C (b), recovered productivity after 14°C (c) and recovery time after chill coma (d). Heat tolerance is represented by *RTmax* (e), productivity at 29°C (f), recovered productivity after 29°C (g) and knockdown time at high temperature (h). Graph (a) and (e) show the medians and 90% credible intervals of the posterior distribution of the estimated parameters. Boxplots in b-d and f-h show the minimum, 25th percentile, median, 75th percentile, maximum and potential outliers. In (d) and (h), data for females (shaded) and males (unshaded) are plotted separately.

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