Research Articles

Unique genomic sequences in novel *Mycobacterium avium* subsp. *hominissuis* lineage

enabled fine scale tracing the transmission route during pig movement

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The order of authors: We determined the order of co-authors according to the

magnitude of their contribution to the study. Of these, the two authors who made

particular contributions to the design of this study, analyses, interpretation and

discussion of the results, and preparation of the paper were designated as the first

authors according to the magnitude of their contributions.

Running title: NOVEL MAH LINEAGE CAN TRACE THE TRANSMISSION ROUTE

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Abstract

Mycobacterium avium subsp. *hominissuis* (MAH) is one of the most prevalent

mycobacteria causing non-tuberculous mycobacterial (NTM) disease in human and

animals. Of note, MAH is a major cause of mycobacterial granulomatous mesenteric

lymphadenitis outbreaks in pig populations. To determine the precise source of infection

of MAH in a pig farm and to clarify the epidemiological relationship among pig and

human MAH lineages, we collected 50 MAH isolates from pigs reared in Japan and

determined draft genome sequences of 30 isolates. We conducted a variable number of

tandem repeat (VNTR), genome phylogenetic tree analysis, historical recombination

analysis and genome content analysis. Consistent with the results of previous studies,

VNTR revealed that most pig MAH isolates in Japan were closely related to North

American, European and Russian human isolates but not to those from East Asian

human. Recombination analysis indicated that most pig isolates could be classified into

SC2/4 and SC3. To our surprise, four isolates belonged to a new lineage in the global

MAH population. Members of this group had few footprints of inter-lineage

recombination in the genome, and carried 80 unique genes, most of which were located

on lineage specific-genomic islands. As the members of this new lineage were rare, we

were able to trace the putative transmission route via their host pigs. Together, these

results reveal the diversity in the pig MAH population and clarify the difference of source

of infection and the vector of transmission between humans and pigs.

Importance

   Mycobacterium avium subsp. *hominissuis* (MAH) is a dominant cause of non-

tuberculous mycobacterial (NTM) disease in human and animals. In this study, in order

to clarify the epidemiological relatedness between MAH isolates from human and pigs,

and to determine the MAH transmission route among pigs, we conducted molecular and

genomic-based epidemiological research. The pig isolates were genetically distant from

those of humans in Japan, however, they were closely related to human isolates from

North America, Europe and Russia, suggesting a proportion of pig MAH isolates are

imported from abroad. Importantly, we identified a novel lineage of MAH that was

distributed in a specific area in Japan. Epidemiological survey and its unique genomic

features allowed us to trace putative transmission routes via pig movement. Our study

provides critical information for understanding the diversity of MAH and will be useful for

countermeasures of pig and human NTM disease.

Introduction

Mycobacterium avium complex (MAC) is the most common causative agent of

non-tuberculous mycobacterial disease (NTM) in humans in Asia and Europe (1, 2). The

frequency of pulmonary MAC infection is increasing worldwide (3). *Mycobacterium*

avium subsp. *hominissuis* (MAH) is the most prevalent subspecies of MAC clinical

isolates in Asia and Europe (1, 2). MAH causes pulmonary disease (4) and sometimes

causes systemic infection in immunocompromised people, such as those infected with

human immunodeficiency virus (HIV) (5).

MAH also causes lymphadenitis most often involving mesenteric lymph nodes in

pigs (6). MAH sometimes causes systemic infection in pigs (7). Pigs infected with MAH

usually have no clinical signs and the lesions are incidentally found during slaughter (8),

bringing great financial losses in pig farm management in the event of an outbreak (1).

It is thought that environmental materials in pig farms such as sawdust (9) and

peat (10) are sources of MAH infection in pigs, suggesting that the environmental

materials are the most important factor in the spread of MAH (11). In a previous study,

use of IS1245 restriction fragment length polymorphism (RFLP) analysis indicated that

the pig flow and the mother sow are important sources of MAH infection (12). However,

the MAH infection by pig flow and sow to piglet transmission in that study were

anecdotal events within a single farm and it is possible that employees and mobile farm

equipment could also serve as sources of MAH exposure. In addition, RFLP analysis is

inferior to VNTR analysis (13) for MAH typing and for more precise determination of

source of MAH infection, molecular and genomic epidemiological study using the

isolates from several farms and environments is essential.

Pigs are thought to be a potential source of exposure for human MAH infection

(14, 15). Recently, several molecular epidemiological studies of MAH involving variable

number of tandem repeat (VNTR) analysis have been described from around the world

(6, 16, 17, 18, 19, 20). Some reports showed epidemiological relationships of MAH

strains between humans and pigs (6, 20, 21, 22). From these studies, it was suggested

that MAH would have region-specific genetic diversity and distribution. In East Asian

countries, human MAH strains isolated from patients with lung disease were not

identical to MAH isolates of pigs (6, 20). On the contrary, MAH strains of pigs were

identical to those of humans from Europe (18, 22). However, MAH strains of pigs used

in these studies were isolated from limited regions (Kinki - Western region of Honshu

Island, Hokkaido, Okinawa), that harbor about 9.8% of farmed pigs in Japan. VNTR

analysis of MAH isolates from pigs in Kyushu area in Japan, where 31.2% of pigs in

Japan are reared, was also reported (23). However, this study was limited due to use of

only 8 loci for VNTR, which was thought to be less useful than 16 loci used in other

analyses (13). To circumvent these shortcomings, MAH isolates from additional regions

are needed to provide a more robust view of MAH distribution.

Phylogenetic and population structure analysis based on MAH genomes has

recently revealed that MAH can be divided into six major clades, including EastAsia1

(EA1), EastAsia2 (EA2) and SC1 - 4, suggesting that MAH adaptation is geographically

restricted around the world (24, 25). For example, EA1 and EA2 lineages were mainly

distributed in humans in Japan and Korea although SC1 - 4 lineages were distributed in

Europe and USA (24, 25). Although these studies utilized many MAH genomes, only a

few pig isolates were included. Therefore, it is still unknown what lineages are

distributed in pigs in Japan, and highlights a major knowledge gap in understanding

likely zoonotic sources of MAH infection in humans.

To understand the diversity of MAH and its potential risk for humans, it is

essential to characterize the distribution of MAH lineages in farmed pigs in Japan and

their genetic relationship with human MAH isolates. In addition, it is also critical to

conduct detailed epidemiological study at a genomic level for the determination of

transmission route of MAH for pigs. In this study, we collected 50 MAH of pig isolates

from Tokai and Hokuriku area, which produce about 10% of pigs in Japan. Initially, we

determined the genetic relatedness of human and pig isolates via VNTR and the

genomic features of major and minor MAH lineages distributed in pig population in

Japan. Next, we characterized genomic and epidemiological features of a novel MAH

lineage distributed in a limited geographic area. The unique genes of these isolates may

be useful to trace MAH transmission routes via pig movement.

Results

Insertion Sequence (IS) and SNP analysis of *hsp65*.

To investigate molecular features of pig MAH, we compared *hsp65* gene

sequences and IS patterns with those of some reference strains (Table S1). Among

MAH isolates, IS*1245* was absent in 14/50 isolates (28.0%). No isolates had IS*900*,

IS*901* and IS*902*. Four isolates (GM10, GM12, GM21 and GM32), which formed a

distinct new lineage as described in the following section, had a conserved SNP pattern

in the *hsp65* region that was unique to this lineage (Figure S1). These isolates had two

characteristic SNPs, G486A and G828T. GM16 also had a characteristic SNP, G441A.

Mycobacterial Interspersed Repetitive Unit (MIRU) and *Mycobacterium*

***avium* Tandem Repeat (MATR) VNTR.**

Next, to investigate molecular epidemiological relationship between human and

pig MAH strains isolated from the world and Japan, VNTR analysis was performed and

two types of MST trees were generated using 7 loci for comparison with domestic MAH

isolates and 13 loci for those of global isolates.

VNTR based on 7 loci. MST based on 7 loci is shown in Figure 1. From VNTR

analysis, MAH isolates were classified into two groups. Group 1 contained mainly the

human isolates, although almost all isolates derived from pigs in Japan were classified

in Group 2. Some isolates in this study were classified in Group 1, however, no isolate

was clustered with human isolates. The pig isolates also formed a cluster with

environmental isolates in Group 2. The Isolates of the new lineage described above

formed a distinct subgroup with Group 2.

VNTR based on 13 loci. MST based on 13 loci is shown in Figure 2. Isolates

were divided into six clonal complexes. Most isolates of human origin from Japan and

Korea were classified into complex 1. Complex 2 was composed of MAH isolates from

USA, Europe, Russia and some human derived isolates from Korea and pig isolates

from Japan. Complex 4 and 5 were mainly composed of pig isolates from Japan,

although, complex 3 contained human and pig isolates from Japan. The isolates of the

newly described lineage were classified into complex 6 and were genetically far from

any adjacent node.

Phylogenetic tree analysis.

To clarify the phylogenetic features of pig MAH at the genome level, phylogenetic

analysis was performed using draft genome sequences of pig isolates, reference strains

described by Yano et al (25) and *Mycobacterium avium* subsp. *paratuberculosis* (MAP)

strain K-10 as an outgroup. Phylogenetic trees generated by the neighbor joining (NJ)

tree method and maximum likelihood (ML) tree method via CSIphylogeny (26) and IQ-

TREE (27) are shown in Figure 3, Figure S2a and Figure S2b, respectively. As a result

of topological evaluation of both phylogenetic trees via IQ-TREE, phylogenetic tree of

CSI phylogeny considered to be more appropriate than that of IQ-TREE. From the

phylogenetic tree based on draft genome sequences of 30 MAH isolates, 24 isolates

were classified into 3 main lineages (SC2: 13, SC3: 1, SC4: 10), and 4 isolates were

classified into a new lineage (Figure 3). This new lineage was the earliest branch from

the MAP (Figure 3, Figure S2a, Figure S2b). The isolates GM16 and OCU484 were

unclassified and placed between EA2 and SC3 (Figure 3, Figure S2a), however, they

were classified in EA2 by IQ-TREE.

SNP analysis for *cinA* and *sugA* genes.

SNP analysis for *cinA* and *sugA* genes, which are considered as epidemiological

markers to distinguish the six major lineages of MAH (25), are shown in Figure S4 and

Figure S5. The new lineage shared a unique SNP pattern in both genes compared with

the other lineage (Figure S3a and Figure S3b). The isolate GM5 had a different SNP

pattern from SC4 in *cinA* (Figure 3a). GM16 had the same SNP patterns as EA1 in *cinA*

(Figure S3b). GM16 and OCU484 had unique SNP patterns in *sugA*, respectively

(Figure S3b). Interestingly, SNP in the *sugA* gene of the isolates of lineage SC2 were

divided into two patterns, SC2a and SC2b. SNP patterns of SC2a were of a new type

and had a characteristic SNP, C25T. The patterns of SC2b were the same as

Mycobacterium avium subsp *avium* (MAA) (Figure S3b).

Bayesian analysis of population structure (BAPS) analysis and

recombination analysis for lineage classification.

In order to predict precise MAH lineage, we conducted BAPS analysis and

recombination analysis. From population structure analysis using fastBAPs (28), MAH

isolates in this study were divided into 4 lineages, new lineage, SC1, SC2 and large SC

(Figure S4). The occurrence of inter-lineage recombination events assessed by

fastGEAR (29) is shown in Figure 5. The strains and isolates were divided into 5

lineages (new lineage, EastAsia, SC2/4, and SC3). Each of the pig isolates was

classified into three lineages (new lineage: 4, SC2/4: 25, SC3: 1). The novel lineage had

few recombination events with any other lineages. Similarly, some isolates of lineage

SC2/4, which were classified into SC2a by SNP analysis, had few occurrences of

recombination among lineages.

Pan and core genome analysis.

To search the unique genes of new lineage MAH, pan and core genome analysis

were conducted using Roary software (30). From results of pan and core genome

analysis, core and accessory gene content were 2,314 and 12,115, respectively. Six

core genes were absent only in the new lineage and 97 genes were characteristic in this

lineage (Table S2). From Basic Local Alignment Search Tool (BLAST) analysis, 80

genes of the new lineage had less than 95% coverage and similarity with the other MAH

isolates. Most of the genes unique to the new lineage were clustered and were located

on some genomic islands (GIs), which were not previously reported (31) (Table S2).

Clustering analysis based on accessory genes showed that the new lineage made one

cluster (Figure S5).

Detection of mobile elements (Plasmid, phage and GI) in MAH genome.

In order to search for mobile elements of pig isolates, plasmid, phage region and

GI were identified from draft genome sequences via online programs (32-36). No

plasmids were detected in any isolates via Plasmidfinder (32), however,

*avium*MD30_plasmid_pMD2-like sequence was detected in 9/30 isolates of draft

genome sequences via JSpecies (33) (Table S3a). Interestingly, all isolates of the new

lineage had these plasmid-like sequences. Intact and questionable phage regions were

also not detected, although incomplete sequences of phage were detected in all isolates

(34-35) (Table S3b). GIs were detected in all isolates (36), however, there was diversity

in the number of GIs (Table 3c).

Farm relationship where the novel lineage MAH was isolated.

GM10, GM12, GM21 and GM32 were isolated from pigs reared in three farms

located in Gifu, Shiga and Aichi Prefecture (Table S1). From interviewing farm owners,

these 3 farms were related epidemiologically (Figure 6). Aichi B and Gifu A farms had

introduced boars and sows from Aichi Y farm. Shiga A farm had introduced fattening

pigs from Aichi X farm, whose sows were introduced from Aichi Y farm.

Discussion

Discussion

In the present study, we revealed two critical points, first was the genetic

relationship among MAH isolates of pigs, humans and the environment in Japan, and

the second was the identification of a new MAH lineage and its genomic and

epidemiological features.

Genetic relationship of pig isolates in Japan

From VNTR analysis using both 7 and13 loci, most isolates in this study were

grouped with those from pigs and the environment in Japan and humans in Europe,

Russia and USA (Figure: Group 2-5), supporting previous studies (6, 16, 18, 20). In

Japan, sow farms in various regions supply sows to each region. Nevertheless, pig

MAH isolates from various regional origins are closely genetically related, except for the

new lineage. This observation indicates that genetically close MAHs are distributed

among Japanese pigs.

In addition, by phylogenetic tree analysis based on draft genome sequences, 30

isolates were classified into SC2 (13 isolates), SC4 (10), SC3 (1), unclassified (2) and a

new lineage (4) (24, 25). These results are consistent with previous reports (24, 25), but

it is worth noting that SC3 was isolated from pigs in Japan for the first time.

Recombination analysis clarified two previously unclassified isolates GM16 and

OCU484 that were classified into SC2/4. GM16 and OCU484 had many recombination

events with EastAsia. In addition, these isolates were classified into EA1 and SC4 by

SNP analysis of *cinA* gene, respectively. Frequent recombination events and multiple

SNP in recombination-cold regions may contribute to the result of these isolates being

unclassified in genome-based phylogenetic tree analysis.

On the other hand, SNP analysis reveals there is diversity in pig isolates in Japan

at the level of a single gene, such as *sugA*. Our SC2/4 isolates were divided into 5

patterns by phylogenetic analysis of *sugA*, SC4 (10 isolates), SC2a (7), SC2b (6) and

two unclassified (2). These isolates had been isolated from multiple farms across the

prefectures. Focusing on SC2a, these isolates had a unique SNP pattern in their core

gene and had few recombination events with the other lineages. This indicates that

SC2a has evolved in a species- or region-specific manner in Japan.

In lineage SC4, most isolates had the same SNP pattern as the reference strain

and had relatively frequent recombination events among lineages, consistent with a

past study (25). SC4 was isolated from human and the environment in Europe, USA

and Russia (25) and Japanese pig isolates are genetically close to those of

environmental isolates in these areas (16). According to a recent annual report from the

Animal Quarantine Service in Japan, most live pigs that are imported to Japan originate

from Canada (43.7%), America (33.4%) and Denmark (12.9%) in 2019

(https://www.maff.go.jp/aqs/tokei/toukeinen.html). These observations suggest that SC4

may be transmitted from overseas countries by a pig movement. Our study also reveals

that phylogenetic analysis at the level of genome or SNP analysis of genes such as

cinA and *sugA* are useful to correctly understand the ecology of human, pig and

environmental MAH.

Genomic and epidemiological features of the novel lineage

Four isolates (GM10, GM12, GM21, GM32) had the same characteristic SNP

patterns in the *hsp65*, *cinA* and *sugA* genes and possessed related IS patterns. In

addition, these isolates also had the same VNTR profiles and were present in a very

unique place in MST trees. These results including genome based phylogenetic tree,

BAPS and recombination analysis indicated that this cluster was the novel lineage. This

new lineage of MAH had 80 unique genes in their genomes, which were not observed in

the other lineages (Table S3). These genes were clustered and most of them were on

what is predicted to be a GI. In addition, these genes contained mobile element

associated genes, such as IS*481* family transposase IS*3514* and putative prophage

phiRv2 integrase, although any plasmids and prophage regions were not detected via

finder programs. These results suggest that this new lineage can be partly

characterized by mobile elements in addition to recombination events (24, 25). These

unique genes can serve as candidate markers for molecular epidemiological study of

this new lineage.

It was unknown why this new lineage was isolated to the Tokai area in Japan. In

general, the *Mycobacterium* genus is mainly present in the environment, such as in

water sources, soil and dust (37, 38), however, our results of VNTR revealed that this

new lineage is genetically distinct from the other MAH isolates including environmental,

human and pig isolates from Japan, East Asia, Europe, Russia and USA. Based on this

observation, it is unlikely that new MAH lineage was introduced from these regions. It is

reported that MAH is isolated from wild boars in Europe (39), however, MAH is not

isolated from wild animals in the same area of this study (40). Epidemiological

information from farms harboring infected pigs revealed that the source of the new MAH

lineage was likely introduced by pig movement. Specifically, Aichi B and Gifu A farms

had directly introduced pigs from the same farm (Aichi Y), and Shiga A farm had contact

through Aichi X farm, which obtained pigs from Aichi Y farm. There was nothing in

common about using animal housing materials, such as sawdust, among farms. Further,

the other farms including in the Hokuriku area in this study had not introduced pigs from

the Tokai area. These facts suggest that this new lineage is spreading via pig

movement. Based on our study, the transfer of bacteria with the movement of infected

pigs seems to be the most plausible model. This hypothesis is also supported by the

observation that there were few inter-lineage recombination events in the new lineage,

because it is thought that acquisition of genetic variation through recombination (24) or

plasmid transfer (41) occurs more frequently in the environment than in living hosts. To

determine whether this new lineage of MAH is distributed only in pigs in the limited area,

it is necessary to investigate more MAH isolates from different regions, animals, and

environments.

In conclusion, we have clarified the relationship among MAH isolates of pigs

and humans in Japan and the world using draft genome sequences and VNTR analysis.

We have also identified a new lineage of MAH distributed only in the Tokai area in

Japan and based on its genomic features we were able to identify the route of infection

for this MAH as a pig movement. This one-health approach revealed some novel

aspects of dynamics of MAH, which can be useful for implementing countermeasures of

pig and human NTM disease.

Materials and methods

Bacterial isolation, Insertion Sequence and *hsp65* profile and SNP analysis.

Bacterial isolation and identification of MAH isolates of 30 isolates (GM5, GM10, GM12,

GM16, GM17, GM21, GM32, GM44, OCU467 – OCU486 and Toy194 – Toy195) were

referred in our past study (42). 20 isolates used for VNTR analysis (gifu-1 – gifu-92,

GM6, GM11, GM23 – GM25) were isolates from mesenteric lymph nodes, feces,

sawdust and farm soil (Table S1). Isolation and identification of these isolates including

determination of IS profiles and *hsp* code were conducted by our study (42). All of them

were isolated from pigs or environment in Tokai area (Gifu, Aichi and Shiga Prefecture)

and Hokuriku area (Toyama and Ishikawa Prefecture), whose geographical relationship

was showed in Figure 6. The sequences of *hsp65* gene were referred from repository

related to our past study (43) with genes retrieved from genomes of representative

lineages (OCU404: MahEastAsia1, TH135: MahEastAsia2, A5: SC1, 104: SC2, 100:

SC3, OCU491: SC4) and were aligned with Molecular Evolutionary Genetics Analysis

(MEGA) and were used for SNP analysis.

MIRU and MATR VNTR. Primer sets for 15 loci of MATR (MATR1-9, 11-16) and 7

loci selected from MIRU loci (32, 292, X3, 25, 7, 10, 47) were used to get VNTR profiles, as

previously reported (20). Three loci were overlapped between MATR-VNTR (MATR-2,

MATR-3 MATR-9) and MIRU-VNTR (MIRU-292, MIRU-X3, MIRU-10). From all data, we

selected 13 MATR (MATR1-8, 11-13, 15-16) and 7 MIRU profiles for VNTR analysis to

compare with variable data of past reports: Japanese patients and Japanese bathroom of

healthy volunteers (6, 16, 18, 20), Korea patients (18, 44), European patients (18, 45),

Japanese pig (6, 16, 18, 20, 23) and the Japanese natural environment (16, 20). The data

profiles 50 MAH isolates in this study were available in Table S6. Detail number of VNTR

profiles using in this study were showed in Table S7. MSTs based on 7 loci and 13 loci

using above data set and data in this study were generated using Bionumerics version 7.0

(Applied Maths, Sint-Martens-Latem, Belgium) for clustering analysis. Clonal complex in

MST of 13 loci was defined as a different group when there were two or more differences in

the VNTR profile of adjacent nodes according to the past study with one modify, that a

clonal complex was composed when it had over five genotypes (15).

Draft genome sequence. Draft genome sequences of 30 isolates were obtained

according to our past study (42).

Genome phylogenetic tree and SNP analysis of *cinA* and *sugA* gene.

Genome phylogenetic tree analysis based on 30 MAH isolates in this study and 20

reference strains were performed via online tool CSI Phylogeny version 1.4

(https://cge.cbs.dtu.dk/services/CSIPhylogeny/) with default parameters (26) and IQ-

TREE (27). Obtained newick file was visualized by MEGA according to our past study

(42). Alignment file generated by CSI Phylogeny was used for phylogenetic analysis by

NJ method with p-distance method via MEGA. Topological evaluation of obtained

newick file was conducted by IQ-TREE (27). All of the reference genome sequences

were retrieved from National Center for Biotechnology Information (NCBI) genome

database. The sequence of two marker genes, *cinA* (product name: cytochrome P-450)

and *sugA* (product name: trehalose ABC transporter) of 30 MAH located in

recombination cold region, which are offered to be able to distinguish major six MAH

lineages (25), were retrieved from draft genome sequences. These sequences in this

study and the reference strains shown in the past report (25) were aligned and used for

the phylogenetic tree analysis by ML method via MEGA according to our past study (42).

BAPS and recombination analysis for determination of MAH lineage. MAH

lineage in this study was predicted by using fastBAPS software (28) and fastGEAR (29),

which was able to infer the MAH lineages, considering genetic linkage (25, 28). Used

genome sequences were the same as using for followed phylogenetic tree analysis except

for MAP. Filtered polymorphic sites in vcf file generated by CSI Phylogeny were used for

fastBAPS (28). In addition, these sites were combined with intervening reference genome

sequences TH135, and were organized in an alignment file (25) and then used as an input

for fastGEAR (29). The lineage is generated by a result of hierarchical clustering (29). We

adopted the result of fastGEAR as the lineage classification in this study because it was the

most consistent with previous reports (24, 25).

Pan and core genome analysis. The pan and core genome analysis was

conducted using Roary software (30). Used genomes sequences were the same as BAPS

and recombination analysis. Draft genome sequences in this study and references were re-

annotated to get gff files by Prokka v1.14.5 software (46). Genes which were present or

absent only in new lineage (GM10, GM12, GM21, GM32) were searched by Roary

software. BLAST analysis was performed with the genes present only in GM10, GM12,

GM21 and GM32 and marked the genes which had high coverage and similarity (over

95%) with genes of MAH in NCBI gene database. We also researched whether these

genes were located on GI and/or phage regions or not in next paragraph. Hierarchical

clustering analysis based on the present or absent accessory genes in MAH were also

performed by Cluster 3.0 and visualized by TreeView 3.0

(https://zenodo.org/record/1303402#.X4cME-ZxeUk).

Detection of mobile element (Plasmid, phage and GI) in MAH genome. Plasmid

was identified from draft genome sequences by PlasmidFinder version 2.1

(https://cge.cbs.dtu.dk/services/PlasmidFinder/) with following setting (threshold for

minimum % identity: 50%, minimum % coverage: 20%) (32). In addition, we retrieved MAH

plasmid sequences which were more than 2 kbp and used them for pairwise genome

comparisons with draft genome sequences in this study by MUMmer program via JSpecies

(33). Prophage sequences were identified by PHASTER (PHAge Search Tool Enhanced

Release) (https://phaster.ca) (34, 35). GIs were predicted by IslandViewer version 4 (36).

Epidemiological information of farms. Epidemiological information of each farm

condition and pig movement was obtained via interviewing farm owners.

Data availability.

The draft genome sequence data used in this study have been submitted to the NCBI and

all of accession number was available in our past study (42).

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Supporting Information Legends

Table S1. Information of MAH isolates in this study.

Table S2. Uniquely present or absent genes in new lineage.

Table S3. Detection of mobile elements in the draft genome sequences of 30 MAH

isolates in this study.

Table S4. VNTR profiles of 50 MAH isolates in this study.

Table S5. Information of MAH strains or isolates used for VNTR analysis in this

study.

Figure S1. SNP position of *hsp65* gene. Characteristic SNPs were detected and

shared in the new lineage (indicated by gray background). *1: Turenne et al., 2006 J

Clin Microbiol. *2: Iwamoto et al., 2012 Infect Genet Evol (15).

Figure S2. Phylogenetic tree based on draft genome sequences generated by

CSIphylogeny and IQ-TREE. a) Maximum likelihood tree was generated by CSI

phylogeny version 1.4 and was visualized via MEGA 7.0. The area and Prefecture of

isolates in this study were indicated by the color dots. Lineage classification except for

new lineage was based on Yano et al., 2019. BMC Genomics. **b) Phylogenetic tree**

based on draft genome sequences generated by IQ-TREE. Maximum likelihood tree

was generated by IQ-TREE and visualized via MEGA 7.0. The Area and Prefecture of

isolates in this study were indicated by the color dots. Lineage classification except for

new lineage was based on Yano et al., 2019. BMC Genomics.

Figure S3. Phylogenetic tree analysis and SNP positions in *cinA* and *sugA* genes

of MAH. a) Phylogenetic tree analysis and SNP positions in *cinA* gene. A)

Phylogenetic tree of *cinA* gene was generated by maximum likelihood method using

MEGA 7.0. Bootstrap values were calculated by 1,000 replications. The scale bar

indicates genetic distance of each strain. Lineage classification except for new lineage

was based on Yano et al., 2019. BMC Genomics. B) New lineage shared the same SNP

patterns, which were highlighted by gray. Reference SNP patterns were referred from

Yano et al., 2019 BMC Genomics. **b) Phylogenetic tree analysis and SNP positions**

in *sugA* gene. A) Phylogenetic tree of *sugA* gene was generated by maximum

likelihood method using MEGA 7.0. Bootstrap values were calculated by 1,000

replications. The scale bar indicates genetic distance of each strain. Lineage

classification except for new lineage was based on Yano et al., 2019. BMC Genomics.

B) New lineage shared the same SNP patterns, which were highlighted by gray.

Reference SNP patterns were referred from Yano et al., 2019 BMC Genomics.

Figure S4. Lineage classification by fastBAPS. Lineage classification was predicted

by fastBAPS (Tonkin-Hil et al., 2019 Nucleic Acids Res). Bootstrap values were

calculated by 1,000 replications. The scale bar indicates genetic distance of each strain.

Lineage classification in the right side except for new lineage was based on Yano et al.,

2019. BMC Genomics. Neighbor joining tree was generated by via MEGA 7.0 using

alignment file obtained from CSI phylogeny version 1.4.

Figure S5. Clustering analysis based on the presence or absence of accessory

genes in MAH. Phylogenetic tree based on the presence or absence of accessory

genes in MAH was constructed by Cluster 3.0 and visualized by TreeView 3.0.

<center>**Figure Legends**</center>

Figure 1. Minimum spanning tree (MST) based on 7 loci MIRU-VNTR genotyping

of MAH isolates in Japan. Circles indicate different VNTR profiles. The size of each

circle depended on the number of isolates sharing the same profiles. New lineages

indicated by an arrow were clustered and consisted of isolates GM10, GM11, GM12,

GM21, GM23, GM24, GM25, GM32.

Fig 2. Minimum spanning tree (MST) based on 13 loci MATR-VNTR genotyping of

MAH isolates from various geographic regions and sources. Circles indicate

different VNTR profiles. The size of each circle depended on the number of isolates

sharing the same profiles. Clonal complexes were highlighted in gray. Each clonal

complex consisted of over five isolates.

Figure 3. Phylogenetic tree based on draft genomes. Neighbor joining tree was

generated by MEGA 6.0 using alignment file obtained from CSI phylogeny version 1.4.

Bootstrap values were calculated by 1,000 replications. The scale bar indicates genetic

distance of each strain. Lineage classification except for new lineage was based on

Yano et al (21).

Figure 4. Determination of MAH lineages. Left: Visualization of recent horizontal

acquisition events generated by fastGEAR. Color of chromosomal regions expresses

<center>29</center>

donor lineage of the regions. Right: Neighbor joining tree was generated by via MEGA

7.0 using alignment file obtained from CSI phylogeny version 1.4.

Figure 5. Relationship among farms where new lineage MAH was isolated. Aichi Y

farm bred and supplied pigs to Aichi X, Aichi B and Gifu A farm, where these farms had

the novel lineage (GM21, GM10 and GM32). Aichi X farm whose sows were introduced

from Aichi Y farm supplied pigs to Shiga A farm where GM12 was isolated. Finally, pigs

of Gifu A, Aichi B and Shiga A farm were slaughtered in the same slaughtered plant

(Gifu meat inspection center).

Figure 6. Location of each regions and prefectures in Japan. Pigs from Tokai area

(southern Gifu, Shiga and Aichi Prefecture) are slaughtered in Gifu meat inspection

center. Pigs from Hokuriku area (northern Gifu, Toyama and Ishikawa Prefecture) are

slaughtered in Toyama meat inspection center.

Group 1

Group 2

Area, origin
and No. of isolates

- 🟦 Tokai pigs (22)
- 🟪 Toyama pigs (13)
- 🟧 Ishikawa pigs (8)
- 🟪 Kinki pigs (70)
- 🟫 Kyushu pigs (36)
- ⬛ Hokkaido pigs (5)
- 🟥 Kinki human (93)
- 🟨 Hokkaido human (31)
- 🟦 Tokai farm (7)
- 🟩 Hokkaido bathroom (3)
- 🟦 Chubu bathroom human (4)
- 🟫 Kinki bathroom human (60)
- 🟩 Kinki river (17)
- ⬜ Kinki environment (5)
- 🟦 Chugoku & Shikoku bathroom (8)
- ⬜ Kanto bathroom (15)
- 🟫 Tohoku bathroom (3)

Fig 1. Minimum spanning tree (MST) based on 7 loci MIRU-VNTR genotyping of MAH isolates in Japan. Circles indicate different VNTR profiles. The size of each circle depended on the number of isolates sharing the same profiles. New lineages indicated by an arrow were clustered and consisted of isolates GM10, GM11, GM12, GM21, GM23, GM24, GM25, GM32.

Fig 2. Minimum spanning tree (MST) based on 13 loci MATR-VNTR genotyping of MAH isolates from various geographic regions and sources.

Circles indicate different VNTR profiles. The size of each circle depended on the number of isolates sharing the same profiles. Clonal complexes were highlighted in gray. Each clonal complex was consisted of over five isolates.

Fig 3. Phylogenetic tree based on draft genomes.
Neighbor joining tree was generated by MEGA 6.0 using alignment file obtained from CSI phylogeny version 1.4. Bootstrap values were calculated by 1,000 replications. The scale bar indicates genetic distance of each strain. Lineage classification except for new lineage was based on Yano et al (21).

Fig 4. Determination of MAH lineages.

Left: Visualization of recent horizontal acquisition events generated by fastGEAR. Color of chromosomal regions expresses donor lineage of the regions. The isolates with dots were classified into SC2a by SNP analysis. Right: Neighbor joining tree was generated by via MEGA 7.0 using alignment file obtained from CSI phylogeny version 1.4.

Fig 5. Relationship among farms where new lineage MAH was isolated.
Aichi Y farm bred and supplied pigs to Aichi X, Aichi B and Gifu A farm, where these farms had the novel lineage (GM21, GM10 and GM32). Aichi X farm whose sows were introduced from Aichi Y farm supplied pigs to Shiga A farm where GM12 was isolated. Finally, pigs of Gifu A, Aichi B and Shiga A farm were slaughtered in the same slaughtered plant (Gifu meat inspection center).

Fig 6. Location of each regions and prefectures in Japan.
Pigs from Tokai area (southern Gifu, Shiga and Aichi Prefecture) are slaughtered in Gifu meat inspection center. Pigs from Hokuriku area (northern Gifu, Toyama and Ishikawa Prefecture) are slaughtered in Toyama meat inspection center.