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3 **Multimodal deep neural decoding of visual object representation in humans**
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Abstract

Perception and categorization of objects in a visual scene are essential to grasp the

surrounding situation. However, it is unclear how neural activities in spatially-

distributed brain regions, especially in the aspect of temporal dynamics, represent visual

objects. To address this issue, we explored spatial and temporal organization of visual

object representations using concurrent functional magnetic resonance imaging (fMRI)

and electroencephalography (EEG), combined with neural decoding by deep neural

networks (DNNs). Visualization of the fMRI DNN revealed that visual categorization

(faces or non-face objects) occurred in brain-wide cortical regions, including the ventral

temporal cortex. Interestingly, the EEG DNN valued the earlier phase of neural

responses for categorization and the later phase of neural responses for sub-

categorization. Combination of both DNNs improved classification performance for

both categorization and sub-categorization. These deep learning-based results

demonstrate a categorization principle in which visual objects are represented in a

spatially organized and coarse-to-fine manner.

Introduction

Visual perception is a rapid cognitive process that allows immediate judgement of safety/risk, enabling to quick planning and acting. The recognition and categorization of the visual scene requires a series of processing stages in the ventral visual processing stream (Grill-Spector and Weiner, 2014; Grill-Spector et al., 2017; Hung et al., 2005; Logothetis and Sheinberg, 1996; Peelen and Downing, 2017). The last stage of the ventral stream, where both perceptual and mnemonic signals meet, plays a crucial role in this cognitive process by referring the perceived object to the neural representation of memorized objects stored in the long-term memory (Landi et al., 2021; Suzuki and Naya, 2014; Takeda et al., 2018).

The functional role of the ventral stream on visual categorization has been investigated by neuropsychological studies. Lesions to the ventral stream can cause various forms of agnosia depending on the location of the core lesion (Barton, 2011; Cavina-Pratesi et al., 2010; Gainotti and Marra, 2011; Konen et al., 2011; Schiltz et al., 2006), suggesting module-like spatial organization for visual categories, such as faces and objects. These observations are further supported by accumulating evidence from functional magnetic resonance imaging (fMRI) in humans, such as the identification of category-selective regions for faces (Haxby et al., 2001; Ishai et al., 1999; Kanwisher et al., 1997), objects (Haxby *et al.*, 2001; Ishai *et al.*, 1999; Kourtzi and Kanwisher, 2000; Malach et al., 1995), and places (Epstein and Kanwisher, 1998) in not only the ventral stream, but also other cortical regions. Spatial organization in the categorization was also demonstrated in non-human primates (Kriegeskorte et al., 2008; Tsao et al., 2003; Tsao et al., 2006; Turesson et al., 2012).

In contrast, neural representations for categorical subdivision (sub-categorical representation), especially in the temporal aspects, remain unclear. At the finest spatial scale, single neurons show selective responses to identities within a single category in non-human primates (Miyashita and Chang, 1988; Tanaka, 1996). At the larger spatial scale, high-frequency local field potential carried spike-coupled information, such as the species, view, and identity of the face (Miyakawa et al., 2018). In human studies, extensive evidence has been obtained from magneto-/electroencephalography (MEG/EEG) (Carlson et al., 2013; Cichy et al., 2014; Dobs et al., 2019; Liu et al., 2002; Zheng et al., 2012) (see also fMRI studies for sub-categorical representation (Kriegeskorte et al., 2007; Nestor et al., 2011)). However, the temporal trajectory of categorical and sub-categorical representation, that is, whether our brains process visual object categorization in a fine-to-coarse or coarse-to-fine manner, is still controversial

(Carlson *et al.*, 2013; Cichy *et al.*, 2014; Dobs *et al.*, 2019; Liu *et al.*, 2002; Mack and
Palmeri, 2011).
Unraveling the whole picture of the spatiotemporal profile of categorical and
sub-categorical representations of visual objects is indispensable for understanding the
computational model of visual object processing in the brain. To address this issue, we
introduce two technical implementations here. First, we employed concurrent fMRI and
EEG recordings in humans, to obtain neural responses to visual stimuli in high
spatiotemporal resolution. Second, we utilized the deep neural network (DNN) to
resolve spatiotemporal patterns of the neural activity responsible for the categorical/sub-
categorical classification of visual objects. We hypothesized that (1) fMRI data involve
spatially resolved patterns of neural activity that contribute to visual object
classification between categories (faces or objects), and (2) EEG data involve
temporally resolved patterns of neural activity that contribute to visual object
classification not only between categories but also within a category (sub-
categorization; male faces or female faces, and natural objects or artificial objects), both
of which would be processed at different periods (Figure 1A). These hypotheses were
tested using newly-developed DNN classifiers for multimodal neural data recorded by
concurrent fMRI and EEG (Figure 2) as well as the univariate analysis for fMRI and
EEG data.

Figure 1. Experimental paradigm and behavioral performance. (A) A hypothesis for the neural representation of visual object categorization and sub-categorization. Categorical information, such as identification of faces or objects, is mainly represented in the spatial domain, whereas sub-categorical information such as male faces or female faces is mainly represented in the temporal domain. **(B)** Sequence of a task run. At the start of a task run, the association between visual objects and buttons is instructed. Each visual stimulus from four sub-categories (male face, female face, natural object, artificial object stimuli) is presented for 500 ms, with a 10-s interval. **(C)** Exemplar visual stimuli. Face and object images are selected from ATR Facial Expression Database DB99 and ALOI (Amsterdam Library of Object Images), respectively. See also Figure 1 – figure supplement 1 for their low-level visual features. **(D–E)** Correct performance (D) and response latency (E) for each category and sub-category. **(D)** Differences in the correct performance in both face sub-category and object sub-category are not statistically significant (right, Wilcoxon signed rank test, $z < 0.4$, $P > 0.68$), though performance in the face trial is higher than that in the object trial (left, face and object; $z = 4.1$, $P = 4.19 \times 10^{-5}$). **(E)** Left, reaction times to face stimuli are significantly faster than those to object stimuli (Wilcoxon signed rank test, $z = 5.56$, $P = 2.63 \times 10^{-8}$). Right, within face sub-category, reaction times to male face are faster than those to female face ($z = 3.50$, $P = 4.63 \times 10^{-4}$), while difference in reaction times within object sub-category is not significant ($z = 0.71$, $P = 0.48$). $N = 50$ participants. Boxplots show median (thick line), 25th/75th percentile (box), and minimum/maximum (whisker).

Figure 2

Figure 2. DNN models for multimodal neural data. (A) Temporal relationship across stimulus presentation and fMRI/EEG data windows used for DNN. fMRI data were taken within 4–6 s from stimulus onset to consider the delayed hemodynamic response of the BOLD signal. EEG data were taken during the period from stimulus onset to 0.5 s after stimulus offset. **(B)** Schematics of the train/validation/test architecture. In the training/validation dataset (N = 45 participants, 1-fold data include data from N = 5 participants), we applied 9-fold cross-validation to tune hyperparameters and constructed learned DNN models. Then, the generalization performances of these models for the independent test dataset were assessed (N = 5 participants). **(C)** A DNN model for fMRI data. The model includes three convolution layers and two fully connected layers. **(D)** A DNN model for EEG data. The model includes four temporal blocks and one fully connected layer. **(E)** A temporal block of EEG DNN. This block includes two dilated convolution layers with a residual connection.

Results

Behavioral results

Participants performed a visual object classification task, in which they judged whether the presented visual stimuli belonged to either the male face, female face, natural object, or artificial object image category (Figure 1B–C). To minimize the effect of low-level visual features of stimuli on differential brain activations and resultant differences in decoding performances, we controlled for contrast and luminance of images (Stigliani et al., 2015) (Figure 1 – figure supplement 1; see Materials and Methods for details).

Participants almost perfectly categorized the presented visual stimuli (Figure 1D; $N = 50$). Performance was not significantly different between male face and female face trials and between natural object and artificial object trials (Wilcoxon signed rank test, $z < 0.4$, $P > 0.68$). Thought reaction times to male face were faster than those to female face ($z = 3.50$, $P = 4.63 \times 10^{-4}$), there was no significant difference in reaction times within the object sub-category ($z = 0.71$, $P = 0.48$) (Figure 1E).

Spatial organization of category-specific information revealed by fMRI activations

We explored brain regions associated with categorical processing based on standard univariate general linear model (GLM) analysis of fMRI data (Figure 3, Figure 3 – figure supplement 1). Robust activations for faces compared with those for objects, were observed in various regions, including the temporal occipital fusiform cortex (TOF), which includes the region known as the fusiform face area (FFA), and lateral occipital complex (LOC) in the ventral temporal cortex (VTC), as well as the occipital pole, angular gyrus, frontal medial cortex, amygdala, frontal orbital cortex, precuneous (PcC), and middle temporal gyrus (Figure 3A). In contrast, the brain regions activated by objects included the TOF, LOC, middle frontal gyrus, insula, superior frontal gyrus, thalamus, and paracingulate gyrus. These results were consistent with previous studies (Haxby *et al.*, 2001; Ishai *et al.*, 1999; Kanwisher *et al.*, 1997; Kourtzi and Kanwisher, 2000; Malach *et al.*, 1995).

Subsequently, we explored brain regions associated with sub-categorical processing (Figure 3 – figure supplement 2, 3). The face gender contrast (male face vs female face) revealed weak activations; there were no male face dominant clusters and three female face dominant clusters (Figure 3B). The contrast between natural and artificial objects also showed weak activations; there were no natural object dominant clusters and two artificial object dominant clusters (Figure 3C). These results suggest that the difference in sub-categorical processing of visual objects is hardly detected at the spatiotemporal resolution of fMRI used in this study.

Figure 3

Figure 3. Spatial representation of categorization/sub-categorization for visual objects revealed by fMRI. (A–C) Activation map for the contrast between face and object (A), between male face and female face trials (B) and between natural object and artificial object trials (C). Color bars denote t-value. Significant brain regions are denoted using a threshold of $P < 0.05$ corrected with cluster-wise family-wise error rate based on a non-parametric permutation test. Identified clusters in the subcortical structures and the insula are shown in the horizontal sections. See also Figure 3 – figure supplement 1–3. **(D)** Generalization decoding performance for the test dataset. Ensemble accuracy (purple), as well as average accuracy ± S.D (thick line) across models, are shown as a function of the number of averaged trials. Thin lines represent accuracy in each model. *, binomial test for ensemble accuracy against chance level (50%), FDR corrected $q < 0.05$. †, t-test for accuracy distribution against chance level, FDR corrected $q < 0.05$. The number of significant models (binomial test, $q < 0.05$) is shown above the abscissa. **(E)** Spatial organization of the guided Grad-CAM values for the contrast of category (between face and object trials). The threshold of significance is set at $P < 0.05$ corrected with cluster-wise FWE based on a non-parametric permutation test. **(F)** ROI analysis. Colored and grayed bars denote the guided Grad-CAM values in learned and non-learned models, respectively. For display purpose, the results are plotted in separate panels according to the magnitude of values. *, t-test against values obtained from the non-learned model, FDR corrected $q < 0.001$. Values in non-learned models are nearly zeros. Boxplots show median (black line), $25^{th}/75^{th}$ percentile (box), nonoutlier minimum/maximum (whisker), and outlier (dot, data points outside 1.5× interquartile range). Cluster information is listed in Figure 3 – figure supplement 1. L, left; R, right; LH, left hemisphere; RH, right hemisphere; AG, angular gyrus; FMC, frontal medial cortex; FOC, frontal orbital cortex; Ins, insular cortex; LOC, lateral occipital cortex; MFG, middle frontal gyrus; MTG, middle temporal gyrus; OcP, occipital pole; PcC, precuneous cortex; PcG, paracingulate gyrus; SFG, superior frontal gyrus; TOF, temporal occipital fusiform cortex; i, inferior; s, superior; p, posterior.

Deep neural decoding of visual object categorization by fMRI DNN

To decode information about the visual object category and identify the spatial
organization of responsible brain regions, we used a DNN classifier for fMRI data,
which was based on three-dimensional convolution layers (Figure 2C). We trained the
fMRI DNN using an fMRI volume acquired 4 s after the presentation of visual stimuli
in each trial (Figure 2A). Training and validation of the DNN were conducted by 9-fold
cross-validation, and the generalization performance of the resultant nine models was
evaluated using an independent test dataset (Figure 2B; see Materials and Methods for
details).

The ensemble learning method using a majority voting scheme (Rokach, 2009)
revealed that classification performance in categorization reached statistical significance
when trials were not averaged (72.90%; $q = 2.61\times10^{-72}$, binomial test against chance
level 50%, corrected for multiple comparisons by false discovery rate (FDR) of $q <$
0.05) (Figure 3D). The performance improved as a function of the number of averaged
trials, reaching 85.83% with an average of nine trials ($q = 3.89\times10^{-185}$). In contrast, both
face sub-categorical and object sub-categorical classification were not significant in the
averaged trial condition ($q > 0.21$), except for object sub-categorization when nine trials
were averaged.

Two additional analyses were performed to examine the robustness of these
results. In the first analysis, a t-test showed that for categorical classification, the
performance distribution statistically reached above chance when trials were not
averaged [68.80 ± 0.39% (mean ± s.e.m.); $t = 48.80$, $P = 3.44\times10^{-11}$, corrected by FDR
$q < 0.05$]. The performance improved as a function of the number of averaged trials
(linear regression, $R^2 = 0.82$, slope coefficient, $t = 13.99$, $P = 1.32\times10^{-17}$), reaching
84.96 ± 0.71% with an average of nine trials ($t = 49.04$, $P = 3.31\times10^{-11}$). The highest
accuracy reached was 87.60% in the best performing DNN model. In contrast, both face
sub-categorical and object sub-categorical classification did not reach statistical
significance in either averaged trial conditions ($q > 0.26$).

In the second analysis, we counted the number of models showing significant
performance against chance level using a binomial test (threshold at $P = 0.05$, FDR
corrected). Although all nine models were significant in categorical classification across
all averaged trial conditions, there were less than two models with significant sub-
categorical classification. The results in the two additional analyses support the result of
the ensemble learning method.

167   With respect to important spatial neural activity for visual object classification, guided Gradient-weighted Class Activation Mapping (Selvaraju et al., 2019) (guided Grad-CAM) showed that the fMRI DNN model valued brain-wide regions, including the frontal, parietal, temporal, and occipital cortices as well as subcortical structures in both face and object trials (Figure 3E; Figure 3 – figure supplement 4). Whole-brain activations identified in the univariate analysis and guided Grad-CAM were significantly correlated at the voxel-by-voxel level, although majority of voxels showed guided Grad-CAM values around zero, leading to a relatively low effect size (Figure 3 – figure supplement 5A; Spearman's rank correlation, $\rho = 0.048$, $t = 19.73$, $P < 0.001$ for face trial; $\rho = -0.21$, $t = -90.13$, $P < 0.001$ for object trial). The correlation coefficient increased when target voxels were masked by the maps of meta-analysis obtained from Neurosynth (Yarkoni et al., 2011) using the terms "face" and "object" (Figure 3 – figure supplement 5B; $\rho = 0.27$, $t = 46.27$, $P < 0.001$ for face trial; $\rho = -0.30$, $t = -56.21$, $P < 0.001$ for object trial). This suggests that voxels with nearly-zero values in the guided Grad-CAM tended to be located outside significant clusters in the univariate analysis. Indeed, the region-of-interest (ROI) analysis revealed that most of the identified clusters in the univariate analysis showed significant guided Grad-CAM values in both face and object trials, when comparing with those obtained in non-learned DNN models where weight values were assigned randomly (Figure 3F; Wilcoxon signed rank test against guided Grad-CAM values in non-learned models, FDR corrected $q < 0.05$). Similar results were obtained when ROIs were defined based on the anatomical structure or functional meta-analysis by Neurosynth (Figure 3 – figure supplement 6); among the three ROI-based analyses, consistent results were obtained in the VTC (LOC, TOF), FMC, and PcC for the face trial, and the VTC (LOC), and thalamus for the object trial.

191   These results indicate that, for the categorization between faces and objects, our DNN model paid attention to the neural signals in the distributed brain regions identified in the univariate analysis.

Temporal trajectory of categorical/sub-categorical information by EEG responses

To examine temporal characteristics of categorical and sub-categorical processing of visual objects, we analyzed the event-related potential (ERP) and power spectrogram of the EEG signals (Figure 4; Figure 4 – figure supplement 1). Regarding ERPs for categorization, face stimuli induced an early negative component (< 200 ms from stimulus onset) with a prolonged component (around 200–700 ms), wherein the polarity of the potential and its dynamics differed across channels. Meanwhile, object stimuli did not evoke the clear early negative component (Figure 4 – figure supplement 1A).

Figure 4. Temporal representation of categorization/sub-categorization for visual objects revealed by EEG. (A) Differential ERP and power spectrogram in the contrast of face minus object trials. Line colors correspond to the channel location depicted on the left side. A, anterior, P, posterior, R, right, L, left. **(B)** Differential ERP and power spectrogram in the contrast of male face minus female face trials. **(C)** Differential ERP and power spectrogram in the contrast of natural object minus artificial object trials. **(D)** Generalization decoding performance for the test dataset. Configurations are the same as in Figure 3d. **(E)** Temporal trajectories of the guided Grad-CAM values for each trial type. Data from all channels are averaged, and values are normalized to the maximum guided Grad-CAM value (=1). Dashed lines represent a threshold (20% of maximum value). Arrows depict maximum latency that first exceeded the threshold. **(F)** Temporal distribution of maximum latency for each trial (dot). Trials for face, object, male face, female face, natural object, and artificial object are separately colored. *, Tukey–Kramer test, $P < 0.05$ after Kruskal–Wallis test, $\chi^2 = 9.70$, $P = 0.0078$. **(G)** Comparison of the guided Grad-CAM values at the maximum latency between learned and non-learned models. *, t-test against values obtained from the non-learned model, FDR corrected $q < 0.001$. Note values in non-learned models are nearly zeros. Boxplots show median (black line), 25th/75th percentile (box), nonoutlier minimum/maximum (whisker), and outlier (dot, data points outside 1.5× interquartile range).

This led to differential ERPs between face and object trials (Figure 4A). The ERPs
between faces and objects were significantly different at around 200–600 ms from
stimulus onset at brain-wide (50/63) channels (Figure 4 – figure supplement 1B; cluster-
defining threshold of $P < 0.05$, corrected significance level of $P < 0.05$). In contrast, the
power spectrogram did not show clear differences (Figure 4A; Figure 4 – figure
supplement 1B).

209   In contrast to categorization, spectral power was more informative in sub-
categorization. Specifically, in face sub-category processing, male face stimuli elicited
larger beta power than did female face stimuli at a prolonged time window (–1,000 ms).
Meanwhile, these two stimuli did not evoke distinctive ERP responses (Figure 4B and
Figure 4 – figure supplement 1C). In object sub-category processing, artificial stimuli
evoked larger beta power than did natural stimuli at a prolonged time window (–1,000
ms) in several channels at the posterior part of the brain. Distinctive ERP responses
were also observed at 200–300 ms (21/63 channels; Figure 4D and Figure 4 – figure
supplement 1D).

Deep neural decoding of visual object categorization/sub-categorization by EEG
DNN
To decode information about the visual object category and sub-category as well as to
identify its temporal dynamics, we used a DNN classifier for EEG data, which was
based on a temporal convolutional network (TCN) (Bai et al., 2018) (Figure 2D–E). We
trained the EEG DNN using 63-ch EEG time series during 1 s after stimulus onset
(Figure 2A). The ensemble learning method revealed that classification performance in
categorization statistically reached above chance when trials were not averaged
(59.33%; binomial test against chance level 50%, $q = 5.75 \times 10^{-13}$, corrected for multiple
comparisons by FDR $q < 0.05$) (Figure 4D). The performance improved as a function of
the number of averaged trials, reaching 75.41% with an average of nine trials ($q =$
4.95×10^{-89}). Regarding face and object sub-categorization, classification performance
improved from 53.50% (no trial average; $q = 0.056$) to 60.75% (average of 9 trials; $q =$
1.25×10^{-8}) and from 53.48% (no trial average; $q = 0.067$) to 60.72% (average of 9 trials;
$q = 4.99 \times 10^{-8}$), respectively. These results suggest that the EEG DNN succeeded to
decode sub-categorization of visual objects while the fMRI DNN did not.

235   Performance distribution and the number of significant models were further
analyzed to confirm the robustness of the results. First, performance distribution of nine
models for categorization statistically reached above chance when trials were not
averaged (57.07 ± 0.15%; t-test against chance level 50%, $t = 47.33$, $P = 4.39 \times 10^{-11}$,

FDR corrected $q < 0.05$) (Figure 4D). The performance improved as a function of the

number of averaged trials (linear regression, $R^2 = 0.87$, slope coefficient, $t = 16.90$, $P =$

1.34×10^{-20}), resulting in $73.41 \pm 0.74\%$ with an average of nine trials ($t = 31.73$, $P =$

1.06×10^{-9}). Decoding performance for both face and object sub-categorization also

improved as the number of averaged trials increased (face sub-categorization: $R^2 = 0.44$,

slope coefficient, $t = 5.78$, $P = 7.51 \times 10^{-7}$; object sub-categorization: $R^2 = 0.59$, slope

coefficient, $t = 8.00$, $P = 4.78 \times 10^{-10}$). As for male face/female face classification, the

mean accuracy in the condition where trials were not averaged was significantly above

chance ($51.90 \pm 0.60\%$; $t = 3.19$, $P = 0.0129$, FDR corrected). When nine trials were

averaged, the accuracy reached $57.51 \pm 0.64\%$ ($t = 11.76$, $P = 2.49 \times 10^{-6}$). Notably, the

highest accuracy was 59.46% in the best performing model. As for natural

object/artificial object classification, the accuracy improved from $52.91 \pm 0.60\%$ (no

trial average; $t = 4.86$, $P = 0.0013$, FDR corrected) to $58.71 \pm 0.62\%$ (average of 9

trials, $t = 14.08$, $P = 6.30 \times 10^{-7}$). The highest accuracy was 61.56% in the best

performing model.

Second, while all nine models were significant in categorical classification

across all averaged trial conditions, the number of models with significant sub-

categorical classification increased as a function of the number of averaged trials

(binomial test, threshold at $P = 0.05$, FDR corrected). In face sub-categorization, the

number of significant models increased from one to nine, whereas in object sub-

categorization, the number increased from two to nine.

Visualization by guided Grad-CAM revealed that our DNN model valued

differential periods between categorization and sub-categorization (Figure 4E). The

model started to value the timepoint at 120 ms and 336 ms in the face and object trials,

respectively ("emergence latency"; see Materials and Methods for details). Regarding

sub-categorization, the four emergence latencies were 788 ms for male face trials, 216

ms for female face trials, 288 ms for natural object trials, and 328 ms for artificial object

trials, all of which were slower than that for face trials. Meanwhile, emergence latencies

for female face and natural object trials were longer than those for object trials. This

"categorization-then-subcategorization" tendency in the latency of first valued

timepoints was also observed in the latency with the maximum guided Grad-CAM

value ("maximum latency") (Figure 4F). The maximum latency varied among trials, and

their ranges differed between categorization and sub-categorization: the maximum

latency in categorization varied between 104 ms and 912 ms (median = 498 ms), while

that in face sub-categorization and object sub-categorization varied between 236 ms and

960 ms (median = 556 ms) and between 228 ms and 932 ms (median = 541 ms),

respectively. The maximum latency in categorization was significantly shorter than that
in the face and object sub-categorization (Kruskal–Wallis test, $\chi^2 = 9.70$, $P = 0.0078$;
post-hoc Tukey–Kramer test, $P < 0.05$). Notably, the maximum guided Grad-CAM
values in each trial type were significantly larger than those in non-learned models
(Figure 4G; Wilcoxon signed rank test against guided Grad-CAM values in non-learned
models, FDR corrected $q < 0.05$). When grouping guided Grad-CAM values into six
brain regions by locations of electrode channels, all regions showed significant guided
Grad-CAM values (Figure 4 – figure supplement 2). Of note, the highest value of
guided Grad-CAM in categorization was in the occipital cortex, while the frontal and
parietal cortices were more valued in sub-categorization (Friedman's test with post-hoc
Tukey-Kramer method, $P < 0.05$).

Collectively, these results indicate that our EEG DNN model successfully
captured not only the categorical representation, but also the sub-categorical
representation of visual objects in the temporal domain of neural responses. Our results
experimentally support that the visual object categorization is processed in a coarse-to-
fine manner in the temporal axis.

Concurrent neural decoding by combined DNN model
Simultaneous acquisition of fMRI and EEG data enables concurrent neural decoding
using an integrated DNN. Thus, we performed neural decoding for concurrent fMRI and
EEG data with the combined fMRI–EEG DNN (Figure 5A). In the DNN, feature
extraction layers, 3D convolution layers for fMRI data and temporal blocks for EEG
data, were finally concatenated and fully-connected.

When the fMRI and EEG activities of nine trials were averaged, ensemble
performance of the fMRI–EEG DNN was 90.05%, 60.70%, and 63.64% for
categorization, face sub-categorization, and object sub-categorization, respectively, all
of which were significantly above the chance level (Figure 5B; binomial test, $P =$
3.03×10^{-240}, $P = 2.51 \times 10^{-9}$, and $P = 2.44 \times 10^{-13}$). Distributions of performance in the
nine models were also significant against chance level (categorical, $87.49 \pm 1.10\%$, $t =$
78.97, $P = 7.37 \times 10^{-13}$; face sub-categorical, $58.40 \pm 1.43\%$, $t = 40.84$, $P = 1.42 \times 10^{-10}$;
object sub-categorical, $59.97 \pm 2.36\%$, $t = 25.44$, $P = 6.11 \times 10^{-9}$; all $q < 0.05$ in FDR
correction). The best performing models showed performances of 90.46%, 63.81%, and
70.04%. A binomial test revealed that nine, seven, and eight models had significant
decoding performance for categorization, face sub-categorization, and object sub-
categorization, respectively (threshold at $P = 0.05$, FDR corrected).

A

B

C

Figure 5. Combined fMRI–EEG DNN model. (A) Architecture of the model. First, fMRI volume data and EEG time series data are separately convolved by conv3Ds and temporal blocks shown in Figure 2. Next, resultant outputs are concatenated, which are then processed using four fully connected layers. **(B)** Generalization decoding performance for the test dataset (trial average of nine). Bar denotes mean accuracy across models. Circles, accuracy in each model. *, binomial test for ensemble accuracy against chance level (50%), FDR corrected $q < 0.05$. †, t-test for accuracy distribution against chance level, FDR corrected $q < 0.05$. The number of significant models (binomial test, $q < 0.05$) is shown above the abscissa. **(C)** Improvement in decoding accuracy of fMRI–EEG DNN against accuracy in the fMRI DNN model and the EEG DNN model. Differences in accuracy are color-coded. Note that none of the fMRI DNN or the EEG DNN shows higher accuracy than the fMRI–EEG DNN. *, t-test, FDR corrected $q < 0.05$. •, $P < 0.05$, uncorrected.

In the comparison of the classification accuracy of the combined fMRI–EEG
DNN with accuracy for fMRI data alone (Figure 3) or EEG data alone (Figure 4),
accuracies in the fMRI–EEG DNN were higher than those in the fMRI DNN and the
EEG DNN in all averaged trial conditions (Figure 5C). Categorical classification
accuracy of the fMRI–EEG DNN was significantly increased in all averaged trial
conditions (t-test, FDR corrected $q < 0.05$). Regarding sub-categorical classification, the
fMRI–EEG DNN showed significantly higher performance than did the fMRI DNN in
all averaged trial conditions ($q < 0.05$). However, the difference in performance
between the fMRI–EEG DNN and the EEG DNN did not reach statistical significance
except for when three trials were averaged. These results suggest that concurrent usage
of spatial and temporal neural data contributes to improve neural decoding of visual
objects.

Discussion

This study found that the categorical representation of visual objects is supported by spatially-organized brain-wide activations in a coarse-to-fine manner. To our knowledge, the present study is the first to examine neural representation of visual objects using multimodal DNNs. This approach enabled highly resolved brain-wide spatiotemporal neural decoding. The present findings would contribute to construct a novel paradigm for trial-based neural decoding of internal processes in patients with impaired consciousness in clinical practice.

DNNs, which achieved state-of-the-art results for the large-scale image recognition (Simonyan and Zisserman, 2015), is spreading to apply to neural data, known as neural decoding (Hong et al., 2016; Horikawa and Kamitani, 2017; Yamins and DiCarlo, 2016). In fMRI DNN, we managed to use 3D voxel data as an input, while some previous studies used 2D slice data (Sarraf et al., 2017) or 2D flattened cortical data as an input (Tsumura et al., 2021b) (see also other studies where 3D voxel data were used as input data (Frey et al., 2021; Vu et al., 2020; Wang et al., 2020; Zhao et al., 2018)). Such image-like input could be learned by DNNs that have already been trained with a large-scale image set (e.g., ImageNet), such as VGG. The re-training approaches such as fine-tuning or transfer learning help improve decoding performance. Given that trained models for spatio-temporal data, which are suitable for applying to concurrent fMRI and EEG data, are currently unavailable, we could not employ such a strategy in this study. However, there are several clear advantages of our approach, such as (1) inclusion of neural activities in the subcortical structures, which are ignored in the 2D flattened map, and (2) coverage of whole brain structures, which are impossible in 2D slice data. Indeed, we could identify the involvement of the amygdala and thalamus in categorical classification of visual objects. Accumulating open 3D-voxel data available for deep learning is promising for further improving in 3D fMRI decoding using DNNs.

In EEG DNN, we employed a TCN (Bai *et al.*, 2018) in which time-series data are convoluted in a dilated manner. The TCN has excellent performance for non-neural data such as character-level and word-level language modeling. Successful application of the TCN to EEG data in the present study would expand the possibility for the application of the TCN to other biological data such as electromyography.

We obtained high decoding performance for both categorical and sub-categorical classification (*c.f.,* in the fMRI–EEG DNN, ensemble performance reached 90.05%, 60.70%, and 63.64% for categorical and two sub-categorical classifications, and the best performing model showed 90.46%, 63.81%, and 70.04%). It is difficult to

directly compare the performance in this study with that in other studies because
experimental paradigms (e.g., decoding technique, stimulus, and sample size) are quite
different. Nevertheless, the present finding that the fMRI–EEG DNN outperformed the
fMRI DNN and the EEG DNN indicates that a larger number of modal data would
improve the precise identification of the neural representation of visual objects.
In machine learning including deep learning, learning performance is usually
evaluated once for the test dataset (Raschka, 2020). This study employed the ensemble
learning approach (Rokach, 2009) in which obtained classification labels from nine
models were voted and the majority label was treated as the predicted label for the trial.
We confirmed the robustness of this ensemble approach using two distinct indices,
performance distribution and the number of significant models. Considering the
inherent properties of neural data as noisy and variable between trials, this
combinational approach is suitable for the application of deep learning to neural data.
For example, when comparing an fMRI volume or an EEG time series with pictures in
ImageNet, neural data have some background noise, c.f., we can hardly detect the
internal process of the subject by glimpsing an fMRI volume or an EEG time series. In
addition, it is difficult to collect adequate amount of neural data that are comparable to
the number of pictures in ImageNet (>10,000,000). The noisiness and smaller sample
size would result in higher variance in the decoding performance. The ensemble
learning approach would reduce the variance of classification errors, and in addition to a
point estimate of the classification performance, our approach can also evaluate the
variance of performance across models.
Brain regions where the fMRI DNN valued for categorization closely matched
with regions identified in the fMRI univariate analysis, including the ventral temporal
cortex (FFA for face stimuli and LOC for object stimuli), consistent with previous
findings (Haxby *et al.*, 2001; Kanwisher *et al.*, 1997; Kourtzi and Kanwisher, 2000;
Malach *et al.*, 1995). Notably, we revealed the involvement of the prefrontal cortex and
amygdala, which have been also demonstrated in monkey (Freedman et al., 2001) and
human (Kreiman et al., 2000) electrophysiology. Considering the wider distribution of
significant voxels in the guided Grad-CAM than that in the univariate analysis, deep
learning techniques would have greater sensitivity to detect responsible brain regions
for visual object categorization.
Fine and coarse information processing in visual object categorization is still
controversial (Carlson *et al.*, 2013; Cichy *et al.*, 2014; Dobs *et al.*, 2019; Liu *et al.*,
2002; Mack and Palmeri, 2011). Our deep learning-based approach provided robust
evidence to this issue: temporal trajectory of neural decoding by the EEG DNN

demonstrated that the neural activities first conveyed categorical information, followed
by sub-categorical information. These results are in line with the theoretical model of
"coarse-to-fine" categorization (Bar, 2004; Rousselet et al., 2002; Serre et al., 2007).
One notable observation in this study is that peak time in the guided Grad-CAM value
(maximum latency) varied among trials, which could be possibly caused by temporal
variability of behavioral states (e.g., attention) among trials. EEG DNN could detect
such variability at any time point owing to shift invariance in the temporal domain by
dilated convolutions. Trial-dependent latencies of category/sub-category information in
the neural responses would be related to the persistent time windows with significant
object decoding reported in other studies (Cichy *et al.*, 2014; Dobs *et al.*, 2019).

Compared with the situation in which fMRI and EEG recordings are separately
conducted, the concurrent fMRI/EEG recording has several advantages. First, the
variance of behavioral outcomes such as the correct performance and reaction time
could be lower than those obtained in separate sessions or days. Second, concurrent
usage of fMRI and EEG data can result in better decoding performance than the usage
of fMRI alone or EEG alone, as shown in this study. It is promising to add other bio-
signals for detecting other aspects of information of visual objects such as valence or
arousal.

An advanced M/EEG–fMRI fusion approach has recently attracted attention as
it links multivariate neural response patterns based on representational similarity (Cichy
and Oliva, 2020; Cichy *et al.*, 2014) while M/EEG and fMRI data are acquired on
separate sessions. The combination of concurrent multimodal recording and
representational similarity-based fusion approach would be a useful method to more
accurately elucidate neural representation in the sensory system.

Materials and Methods

Participants

A total of 53 healthy right-handed subjects (34 males and 19 females; age, 18–25 years) with normal or corrected-to-normal vision participated in the study. Of them, data from 3 participants were excluded due to low behavioral performance: 2 participants had low correct performance (< 80% at least for one sub-category) and 1 participant had slow responses (mean response latency > 1.5 s). Participants received 1,000 yen per hour for each session. The target number of participants was determined based on the number of trials required in previous deep learning research in neuroscience (e.g., Supratak et al (Supratak et al., 2017)) before starting the collection of neuroimaging data. This study was approved by the institutional review board of Kochi University of Technology, Kami, Japan and complied with the Declaration of Helsinki. Written informed consent was obtained from all participants.

Experimental design

Stimuli. Stimuli consisted of color images (size, 7.4° × 5.8°) and a white fixation cross (size, 0.3° × 0.3°). Forty color images from face and inanimate object categories were used as experimental stimuli. The 20 face and 20 object images were selected from ATR Facial Expression Database DB99 (http://www.atr-p.com/products/face-db.html) and ALOI (Amsterdam Library of Object Images; https://aloi.science.uva.nl/) (Geusebroek et al., 2005), respectively. Facial images were obtained from 4 identities: two male individuals (male face sub-category) and two female individuals (female face sub-category), with neutral facial expressions and 5 different angles. The object images were obtained from 4 identities: two natural objects (paprika and broccoli; natural object sub-category) and two artificial objects (hat and whistle; artificial object sub-category), with 5 different angles.

All images were contrast- and luminance-matched using in-house MATLAB (MathWorks, Natick, MA) code (Keerativittayayut et al., 2018) adapted from SHINE toolbox (*lumMatch* function) (Willenbockel et al., 2010) for each RGB channel, after figure-ground separation. Based on a prior study (Stigliani *et al.*, 2015), we evaluated (1) contrast, (2) luminance, (3) similarity to other stimuli of the same category, (4) visual field coverage, and (5) spatial frequency power distributions, after converting RGB color images to grayscale images by *rgb2gray* function in MATLAB (Figure 1 – figure supplement 1). In addition, hue histograms of color images were also evaluated for each sub-category.

Task procedure. During the recording of brain activities by concurrent fMRI and EEG,
participants performed a visual object classification task, in which sequences of
experimental stimuli were presented. Participants judged the corresponding visual sub-
categories (male face, female face, natural object, and artificial object) and pressed a
corresponding button (Figure 1B–C). At the beginning of each scanning run, a task
instruction was presented that indicated the correspondence between 4 positions of
buttons and 4 sub-categories. To exclude the possibility that the learning of stimulus-
response mapping would affect neural activities, the correspondence was
pseudorandomized across runs. In the scanning run, a fixation cross was first presented
for 30 s, followed by a sequence of 50 trials. In 40 trials, face or object images were
presented, and in 10 trials, blank images were presented to improve dissociation of
hemodynamic responses between trials. In each trial, a stimulus was presented for 500
ms, followed by a fixation cross for 9,500 ms. Thus, one scanning run took a total of
530 s. All stimulus images were presented once in each scanning run, and the order of
stimulus presentation was pseudorandomized. No visual feedback was presented.
Participants practiced the task before starting concurrent fMRI and EEG.
Participants conducted 9.96 ± 2.88 runs (mean ± SD; total 496 runs) while
simultaneous fMRI and EEG recordings were performed. Neural data were accordingly
obtained from a total of 24,800 trials (9,920 face trials, 9,920 object trials, and 4,960
blank trials). Trials wherein participants did not respond within 2 s (1,005 trials, 5% of
total face/object trials) were excluded. Correct performance and reaction time were
calculated for each category and sub-category (Figure 1D–E). Only data from correct
trials were used for the succeeding analyses (18,007 face and object trials). The task was
programmed and administered using Psychtoolbox version 3 (http://psychtoolbox.org/).
The visual stimuli were projected on a screen located behind the scanner using
PROPixx DLP LED projector (VPixx Technologies, Canada). Participants viewed the
projected visual stimuli through a mirror attached to a head coil.
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fMRI procedure and preprocessing
MRI scanning was performed using a 3T MRI scanner (Siemens Prisma, Germany) with
a 64-channel head coil. Functional images were acquired using an echo-planar imaging
sequence [repetition time (TR): 2.0 s; echo time (TE): 27 ms; flip angle (FA): 90 deg;
36 slices; slice thickness: 3 mm; in-plane resolution: 2 × 2 mm]. Multiband accelerated
imaging was unavailable because the guideline for the simultaneous fMRI–EEG
recoding by the developer of the MRI-compatible EEG system did not permit it (Brain

Products, Germany). Each functional run involved 265 volume acquisitions. The first 5

volumes were discarded for analyses to take into account the equilibrium of longitudinal

magnetization. High-resolution anatomical images were acquired using an MP-RAGE

T1-weighted sequence (TR: 2,400 ms; TE = 2.32 ms; FA: 8 deg; 192 slices; slice

thickness: 0.9 mm; in-plane resolution: 0.93×0.93 mm).

All preprocessing was performed using SPM12 software

(http://fil.ion.ac.uk/spm/). Functional images were first temporally realigned across

volumes and runs, and the anatomical image was co-registered to a mean image of the

functional images. The functional images were then spatially normalized to a standard

Montreal Neurological Institute (MNI) template, with normalization parameters

estimated for the anatomical scans. The images were resampled into 2-mm isotropic

voxels. For the univariate fMRI analysis, functional volumes were spatially smoothed

with a 6-mm full-width at half-maximum Gaussian kernel. For the deep learning

analysis, we did not apply spatial smoothing to fMRI data.

EEG procedure and preprocessing

High-density EEG data were acquired from MRI-compatible 64 sintered Ag/AgCl

electrodes with extended scalp coverage, including 63 scalp recording electrodes and an

electrode on the upper back for electrocardiogram (ECG) (BrainCap-MR, Brain

Products, Germany). The electrodes AFz and FCz served as ground and online

reference, respectively. All signals were recorded with a bandpass of 0.016–250 Hz and

digitized at 5,000 Hz (BrainAmp MR Plus, Brain Products). Electrode impedances were

lowered to <10 kΩ before recording and monitored throughout the experiment.

Offline preprocessing was firstly performed in Brain Vision Analyzer 2 (Brain

Products). First, we removed the MR gradient artifact using an average template

subtraction method (Allen et al., 2000) and down-sampled the resultant EEG data to 250

Hz. Second, the EEG data were bandpass filtered between 1 Hz and 100 Hz. Third, the

cardiovascular artifact was removed from the EEG data. Heartbeat detection using the

ECG channel was then performed using a semiautomatic template-matching procedure.

Fourth, eye blink correction was performed using independent component analysis

(ICA). Since specific sensors for vertical/horizontal electrooculography (EOG) were not

used, we used the EEG data obtained from electrode FP1 as a substitute for the vertical

activity of the EOG channel. Finally, we applied ICA-based denoising techniques

implemented in EEGLAB (*runica* function). Then, each independent component was

labeled using ICLabel plugin (https://github.com/sccn/ICLabel). Components with

<75% probability of being in the brain signal were flagged as artifacts and rejected.

The resultant artifact-removed EEG data were further processed in EEGLAB as

follows. If the signal was continuously noisy at a certain channel in a scanning run, the

EEG data were interpolated at the corresponding channel by replacing it with averaged

EEG data from the neighboring channels. Accordingly, 16 out of 3,150 channels (50

participants × 63 scalp channels) were interpolated. Thereafter, the EEG data were

segmented into 1.5-s epochs (0.5 s pre-stimulus to 1.0 s post-stimulus) time locked to

the stimulus onset and were baseline-normalized. The data were then inspected

manually to detect any trials containing any excessive noises. After excluding 742 trials,

17,265 trials were included in the analyses. Data were then digitally re-referenced to the

average of all scalp channels.

For the ERPs, we first trial-averaged EEG time series for each EEG channel

separately. Then, the resultant ERPs were grand-averaged across participants to obtain

population ERPs for each EEG channel. Spectral power was also calculated using

mtspecgramc function in Chronux toolbox for MATLAB (http://chronux.org/). The

parameters for multi-taper time-frequency spectrum were as follows: window = 200 ms,

step size = 80 ms, time-bandwidth product = 3, and number of tapers = 5.

Respiration and eye-tracking recording

Respiratory data were recorded by using the BIOPAC MP160 system with

AcqKnowledge software (BIOPAC Systems, Inc., Goleta, CA). Raw physiological time

series of respiration were sampled at 2,000 Hz and then were preprocessed in PhysIO

toolbox (Kasper et al., 2017) for MATLAB as follows. First, the respiration data were

down-sampled to 250 Hz and bandpass-filtered between 0.1 Hz and 5.0 Hz. Then, both

the respiratory phase and respiratory volume per time (RVT) for the respiratory data

were obtained. From these physiological determinants with noise modeling, (1) 8

RETROICOR (RETROspective Image CORrection) regressors were computed via a 4th

order Fourier expansion of respiratory phase (Harvey et al., 2008), and (2) respiratory

response was calculated by convolution of RVT with the respiration response function

(Birn et al., 2006). Accordingly, we obtained a total of 9 regressors and treated them as

nuisance regressors in the fMRI univariate analysis (see "*fMRI univariate analysis*"

section below).

To check whether participants fixated the eye position, the eye position during

task runs was monitored online at a sampling rate of 1,000 Hz using an MRI-compatible

infrared camera-based eye tracker (EyeLink 1000, SR Research, Ontario, Canada).

Deep neural network classifier

To explore spatiotemporal neural activity representing visual object category and sub-

category, in-house deep neural network (DNN) classifiers (Simonyan and Zisserman,

2015) were constructed (Figure 2). The architecture of the models is described below.

The model training, validation, and testing framework for binary classification

(face/object, male face/female face, or natural object/artificial object images) were

implemented using PyTorch (https://pytorch.org/). The scripts used for network

modeling and analyses are available in GitHub (https://github.com/masaki-takeda/dld).

The whole dataset included 17,265 trial-based concurrent fMRI–EEG data

from 50 participants. Data from 45 participants were used as a training/validation

dataset for optimizing hyperparameters, and independent data from 5 participants were

used as a hold-out test dataset. In the training/validation dataset, data from 5 participants

were treated as a unit of the fold, resulting in 9-fold datasets (Figure 2B). After

preliminarily determining network architectures, we conducted grid searches for

hyperparameter tuning by using the train/validation dataset (see "*Grid search*" section).

After completing hyperparameter tuning, we next evaluated the generalization of neural

decoding performance for the independent test dataset: 9-fold cross-validation was

performed for the training/validation dataset, and then the resultant 9 learned models

were applied to the hold-out test dataset. For each binary classification for the test

dataset, we equalized the number of trials for face/object, male face/female face, and

natural object/artificial object by excluding a surplus of trials in the test dataset. This

resulted in the chance level of 50% (categorization: 734 face trials vs 734 object trials;

face sub-categorization: 386 male face trials vs 386 female face trials; object sub-

categorization: 359 natural object trials vs 359 artificial object trials). The decoding

performance for visual object categorization and sub-categorization was then

investigated using the following network models based on fMRI data alone (Figure 2C),

EEG data alone (Figure 2D–E), and combined fMRI–EEG data (Figure 5A).

fMRI DNN. The current DNN model for fMRI data (fMRI DNN) consisted of a three-

dimensional (3D) convolutional neural network (Frey *et al.*, 2021) (Figure 2C). The 3D

CNN included three convolution layers (*conv3d* function) for extracting spatial features

and two fully connected layers. We also examined other networks with a greater number

of convolution and fully connected layers, but decoding performance did not change

drastically. Based on the preliminary test for the training/validation dataset, the

following hyperparameters and settings were selected: batch size of 100, early stopping,

and Adam optimizer (Kingma and Ba, 2015). We then grid-searched for the number of
averaged trials, learning rate, and weight decay.

The input datasets were trial-based non-smoothed fMRI volumes and were
prepared as follows. To consider the delayed hemodynamic response of the BOLD
signal, an fMRI volume that was acquired at 4 s after the onset of the visual stimulus
presentation was collected (Figure 2A). The input fMRI volume was spatially
normalized to standard MNI space, and signal intensities outside of the brain were
masked to zero. Then, the signal intensity in each voxel was z-scored by the mean and
SD values of the signal intensities in the volume.

EEG DNN. Seven DNN models for EEG data (EEG DNN) were tested in the
preliminary inquiry for training/validation dataset: (1) a custom DNN model including
four *conv1d* layers, (2) a custom DNN model including four *conv2d* layers for band-
passed EEG time series (delta, theta, alpha, beta, and gamma frequency band × time),
(3) a recurrent neural network (RNN) model using long short term memory (LSTM), (4)
a convolutional RNN (ConvRNN) model using LSTM after two *conv1d* layers, (5) a
spatial-temporal neural network originally designed for P300 detection (Zhang et al.,
2021), (6) a generic temporal convolutional network (TCN) (Bai *et al.*, 2018), and (7) a
modified version of the TCN. Among them, we adopted the TCN model (6) because of
its superior decoding performance and its simplicity (Figure 2D–E). The TCN model
has several distinguishing characteristics such as the dilated convolutions (dilated
conv1d) and the causal architecture (no information leakage from future to past). Based
on the preliminary test, a batch size of 100 and early stopping were selected. We then
grid-searched for the number of averaged trials, learning rate, and kernel size. The
dilation factors for each kernel size were determined to cover all the input time series.

The input datasets were trial-based EEG time series data and were prepared as
follows. Preprocessed EEG time series data were collected from 63 scalp channels. The
segmented data obtained at 1 s from the stimulus onset (250 samples at a sampling rate
of 250 Hz) was used (Figure 2A). The resultant segmented data were further z-scored
by the mean and SD values of the signal amplitude for each channel.

DNN for combined data. We also tested whether concurrent usage of both fMRI and
EEG data improves the decoding performance relative to the usage of fMRI data alone
or EEG data alone. The DNN architecture for combined data (fMRI–EEG DNN) is
shown in Figure 5A. In the model, the feature extraction layers of the two networks for
fMRI and EEG data were concatenated and then fully connected. Both fMRI and EEG

networks were pre-trained separately; thereafter the initial parameters of convolution

layers in the fMRI–EEG DNN were set to the pre-trained parameters.

Grid search. Before conducting the grid search, we preliminarily searched several

hyperparameters such as the number of layers and kernel sizes, and found that several

hyperparameters affected the decoding performance: the number of averaged trials,

weight decay, and learning rate for fMRI data, and the number of averaged trials, kernel

size for delated convolutional layers, and learning rate for EEG data. Thus, the grid

searches were conducted to obtain the optimal combination of these hyperparameters.

For the fMRI DNN, the following hyperparameters were tuned: the number of averaged

trials (1, 3, 5, 7, 9), learning rate (0.0001–0.01 in 5 divisions), and weight decay (0–0.01

in 5 divisions). For the EEG DNN, the following hyperparameters were tuned: the

number of averaged trials (1, 3, 5, 7, 9), learning rate (0.0001–0.01 in 5 divisions), and

kernel size (2–9 in 5 divisions). For the combined fMRI–EEG DNN, the following

hyperparameters were tuned: the number of averaged trials (1, 3, 5, 7, 9), learning rate

of feature extraction layers for fMRI data (0.001 or 0.0001), weight decay for fMRI data

(0 or 0.001), learning rate of feature extraction layers for EEG data (0.0001 or 0.0005),

and kernel size for EEG data (2, 5, 7, 9). For the combined fMRI–EEG DNN, we fixed

the learning rate and weight decay for fully connected layers as 0.001 and 0,

respectively. All grid search analyses were conducted using the training/validation

dataset, and the hold-out test dataset was completely free from the hyperparameter

settings.

Data averaging. A single-trial neural signal contains noises derived from the brain and

measurement hardware, and one of the solutions to increase the signal to noise (S/N)

ratio is to average the data. To estimate how the S/N ratio of brain signals affects the

decoding performance, we tested the decoding using both data from a single trial data

and averaged data (number of averaged trials ≤ 9). We averaged fMRI or EEG data

within each subject and then augmented the data to match the total number of averaged

data with the number of all trials (e.g., averaged data from 9 trials were augmented by a

factor of 9).

Visualization of relevant neural activity in DNNs. To highlight important spatiotemporal

neural activities for visual object classification, visualization approaches in deep

learning were applied. For both the fMRI DNN and EEG DNN, guided Gradient-

weighted Class Activation Mapping (guided Grad-CAM) (Selvaraju *et al.*, 2019) was

used to create a high-resolution class-discriminative visualization. Guided Grad-CAM
was created by multiplying guided backpropagation at the input of the first convolution
layer with Grad-CAM at the last convolution layer. The original Grad-CAM uses global
average pooling of the gradients (Selvaraju *et al.*, 2019), but we did not use it like
Layer-CAM (Jiang et al., 2021), because the smoothing effect of pooling was extremely
strong, especially for the 3D convolution. In the end, we obtained multiple guided Grad-
CAM maps, the number of which corresponded to the number of trials. The obtained
guided Grad-CAM values were not normalized. As for EEG guided Grad-CAM, the
model without residual connections was used because those connections can cause
gridding artifacts with the dilated convolution layers and obfuscate the visualization
results (Yu et al., 2017). The resultant guided Grad-CAM values in the EEG DNN were
then smoothed using the Gaussian-weighted moving average with a 40-ms window.
680
681 **Statistics**
682 *General.* All statistical analyses were performed in MATLAB and Python. All data are
683 shown as mean ± standard error of the mean unless otherwise stated. Sample sizes were
684 not based on a priori power calculations but are comparable to other studies in the field
685 using similar techniques (Supratak *et al.*, 2017).
686
fMRI univariate analysis. The fMRI univariate analysis was conducted as in previous
studies (Matsui et al., 2022; Tanaka et al., 2020; Tsumura et al., 2021a; Tsumura *et al.*,
2021b) (Figure 3A–C). In the first-level analysis, a GLM (Worsley and Friston, 1995)
approach was used to estimate parameter values for task events. The events of interest
were the presentation of male face, female face, natural object, artificial object stimuli
(4 regressors). Only correct trials were coded as regressors. Those task events were time
locked to the onset of stimulus presentation and then convolved with a canonical
hemodynamic response function implemented in SPM. Additionally, the onset of blank
stimuli, six-axis head movement parameters, white matter signal, CSF signal, and effect
of respiration (9 regressors) were included in GLM as nuisance effects, totaling to 18
nuisance regressors. Then, parameters were estimated for each voxel across the whole
brain.
In the group level analysis, maps of parameter estimates were first contrasted
within individual participants. The contrasted maps were then collected from
participants and subjected to group-mean tests. Voxel clusters were first identified using
a voxel-wise uncorrected threshold of $P < 0.001$. The voxel clusters were then tested for
a significance across the whole brain with a threshold of $P < 0.05$ corrected by family-

wise error (FWE) rate based on permutation methods (5,000 permutations) implemented

in the *randomise* function in FSL suite (http://fmrib.ox.ac.uk/fsl/), which was

empirically validated to appropriately control the false positive rate in a previous study

(Eklund et al., 2016). Peaks of significant clusters were then identified and listed in

tables. If multiple peaks were identified within 12 mm, the most significant peak was

retained.

EEG univariate analysis. For each participant, a single ERP and power spectrogram

were constructed by averaging data across all trials separately for each visual category

or sub-category in each channel. We then averaged ERPs/spectrograms from all

participants, which resulted in a grand-averaged of 63-ch ERPs/spectrograms for face,

object, male face, female face, natural object, and artificial object trials (Figure 4A–C;

Figure 4 – figure supplement 1A). The subsequently obtained spectrograms were further

bandpass-filtered to divide them into alpha (8–15 Hz), beta (15–30 Hz), and gamma

(30–70 Hz) frequency bands.

For statistical inference on differential EEG time series of ERP and power

spectrogram (e.g., face minus object trials) against zero, permutation-based cluster-size

inference (Dobs *et al.*, 2019; Maris and Oostenveld, 2007) (i.e., a cluster refers to a set

of contiguous time points) was performed separately for each channel. Time points that

exceeded the 97.5th percentile of the permutation distribution ($N = 5,000$) served as

cluster-inducing time points (i.e., equivalent to $P < 0.05$, two-sided). The significance of

clusters was set at the threshold of $P < 0.05$ (Figure 4 – figure supplement 1B–D).

Neural decoding performance. We trained our DNN models using 9-fold cross-

validation with the training/validation dataset and then evaluated their generalization

performance with the test dataset. Thus, a total of 9 generalization performances were

obtained for the fMRI DNN model, EEG DNN model, and combined fMRI–EEG DNN

model. Three different statistical tests were performed to examine the significance of

decoding accuracies and their robustness (Figure 3D, 4D, and 5B). First, an ensemble

learning method with majority voting (Rokach, 2009) was conducted. In this scheme,

each base model ($N = 9$) made a prediction and voted for each sample (a stimulus in a

trial). Only the sample class with the highest votes was included in the final predictive

class. Statistical significance was tested using the binomial test against chance level

(50%). Second, the t-test was used for distribution of accuracies obtained by the nine

models against chance level, with FDR corrections ($q < 0.05$). Third, the binomial test

was used for each model against chance level, with FDR corrections ($q < 0.05$), and the

number of significant models was counted.

Visualization of neural decoding. The model that showed maximum accuracy in the

average of nine trials was selected for the succeeding visualization analysis.

Statistical significance of guided Grad-CAM for fMRI data was evaluated by

permutation-based cluster-size inference (Dobs *et al.*, 2019; Maris and Oostenveld,

2007) used in the present univariate analyses (Figure 3E; see "*fMRI univariate analysis*"

section). The spatial distribution of the significant guided Grad-CAM was also

evaluated based on the Harvard-Oxford cortical and subcortical structural atlases, which

covered 48 cortical and 21 subcortical structural areas, implemented in FSL (Figure 3 –

figure supplement 4; *HarvardOxford-cort-maxprob-thr50-2mm* and *HarvardOxford-*

sub-maxprob-thr50-2mm). Regarding cortical areas, we divided them into areas in the

right and left hemispheres, and obtained 95 cortical regions (note no voxels for left

supracalcarine cortex). Regarding subcortical areas, we bilaterally removed areas

labeled as the cerebral white matter, lateral ventricle and cerebral cortex, and obtained

15 subcortical regions. We examined whether guided Grad-CAM values in each brain

regions ($N = 110$) were significantly higher than values obtained when DNNs were not

learned. In the non-learned model, weight values were initialized and fixed as random

values. Statistical significance was evaluated by Wilcoxon signed rank test with FDR

corrections ($q < 0.05$).

We also conducted two types of correlation analysis (Spearman's rank

correlation; Figure 3 – figure supplement 5) and three types of ROI analysis to examine

the spatial overlap between activations in the univariate analysis and guided Grad-CAM

(Figure 3F, Figure 3 – figure supplement 6). The correlation was calculated either for

the voxels masked by FSL's gray matter prior volume (*avg152T1_gray* template,

threshold = 0.5) or for the voxels masked by the maps of meta-analysis obtained from

Neurosynth (Yarkoni *et al.*, 2011) (https://neurosynth.org) using the terms "face" and

"object" (uniformity test). In the ROI analysis, we examined whether guided Grad-

CAM values in each ROI were significantly higher than the values obtained using the

non-learned DNNs. ROIs were selected based on (1) the univariate analysis (Figure 3 –

figure supplement 1; Figure 3F), on (2) the anatomical structures from the Harvard–

Oxford atlas (Figure 3 – figure supplement 6A), or on (3) the maps of meta-analysis

obtained from Neurosynth (Figure 3 – figure supplement 6B). In the Harvard-Oxford

atlas (2), we created ROIs, in which the peak voxel of clusters identified by the

univariate analysis was included, by thresholding the probabilistic map of the region by

50%. Note that we combined MFGi and MFGs for object trials identified in the

univariate analysis into one ROI as MFG. In the Neurosynth (3), ROIs were selected

with the uniformity test using the terms "face" and "object." ROIs with more than 50

voxels were used in the analysis. In all ROI analyses, statistical significance was

evaluated using Wilcoxon signed rank test with FDR corrections ($q < 0.05$).

Guided Grad-CAM for EEG data were quantified according to two indices,

emergence latency and maximum latency. First, guided Grad-CAM values for each trial

were averaged across 63 channels. Then, for each trial type, we defined the emergence

latency as the time point where the value first exceeded the level at 20% of the

maximum of channel-averaged guided Grad-CAM values (Figure 4E). We obtained

similar results when the threshold was changed to 10%, 15%, 25%, and 30%. The

maximum latency for each trial was also defined as the time point where the guided

Grad-CAM showed maximum value in each trial (Figure 4F), and statistical

significance in the maximum latency across categorization/sub-categorization was

tested using Kruskal–Wallis test with post-hoc Tukey-Kramer test ($P < 0.05$). We also

calculated the maximal latencies and corresponding guided Grad-CAM values when the

DNN was not learned. Then, we examined whether guided Grad-CAM values at the

maximum latency were significantly higher in the learned model than those in the non-

learned model using Wilcoxon signed rank test with FDR corrections ($q < 0.05$; Figure

4G). Finally, we tested whether the guided Grad-CAM values at the maximum latency

differed across brain regions. EEG channels were grouped into 6 brain regions (frontal

cortex, central cortex, parietal cortex, occipital cortex, left temporal cortex, and right

temporal cortex; Figure 4 – figure supplement 2). The electrodes for the 6 regions are as

follows: Fp1, Fp2, Fz, F1, F2, AF3, AF4, AF7, AF8, and Fpz for the frontal region; C3,

C4, Cz, FC1, FC2, C1, C2, FC3, and FC4 for the central region; P3, P4, Pz, CP1, CP2,

P1, P2, CP3, CP4, P5, P6, and CPz for the parietal region; F3, F7, T7, P7, FC5, CP5,

TP9, F5, C5, FT7, TP7, and FT9 for the left temporal region; and F4, F8, T8, P8, FC6,

CP6, TP10, F6, C6, FT8, TP8, and FT10 for the right temporal region. The values

obtained by each channel in a region were averaged, and regional differences were

tested using Friedman's test with the post-hoc Tukey–Kramer method ($P < 0.05$).

Data and code availability

All data supporting the findings of this study are provided within the paper. All custom

Python codes used for deep learning and learned model data in this study are available

at https://github.com/masaki-takeda/dld. All additional information will be made

available upon reasonable request to the authors.

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Author contributions

N.W., K.J., K.N., and M.T. designed the experiment and study. N.W., R.K., and M.T.

collected the data. N.W., K.M., and M.T. analyzed the data. N.W., K.M., K.J., D.S.,

K.N., and M.T. wrote the manuscript.

Competing interests

The authors declare no competing interests.

Figure 1 – figure supplement 1

Figure 1 – figure supplement 1. Low-level visual features of stimulus images. (A) Michelson contrast of images as the ratio between the difference and the sum of the maximum and minimum pixel intensities. **(B)** Luminance as the mean pixel intensity of images. **(C)** Pixel-based pairwise similarity. To estimate visual similarity among images of a category, the mean Euclidean distance between normalized grayscale values (scaled to be between 0 and 1) of each stimulus and every other stimulus of the same sub-category is measured. Pairwise similarity values range from 0 (images with inverted intensity difference at each pixel) to 1 (identical images). Note that the order of the size of the effect [0.0062, standard deviation (S.D.) of category means in normalized grayscale values] is comparable to the resolution of the display (0.0039, minimum pixel-wise distance resolved by the display in normalized values, corresponding to the change in one gray-level value of a possible 255 increments). Mean ± S.D. are shown in (A–C). **(D)** Average spatial frequency across images. **(E)** Average pixel intensity for each pixel. **(F)** Average hue histogram across images.

Figure 3 – figure supplement 1. Brain regions showing significant signal increase and decrease in the contrast of face vs. object. Coordinates are listed in MNI space.

Increase in face trial

Cluster index	Cluster name	t value	x (mm)	y (mm)	z (mm)	Harvard-Oxford Atlas
1	FMC	8.37	4	48	-10	Frontal Medial Cortex
		7.76	8	58	12	Frontal Pole
		6.87	-8	60	20	Frontal Pole
		5.69	-12	48	10	Paracingulate Gyrus
		4.89	4	32	6	Cingulate Gyrus, anterior division
		4.43	-24	42	-4	Frontal Pole
		4.17	-10	48	32	Frontal Pole
		4.04	-8	32	-6	Cingulate Gyrus, anterior division
		3.77	8	48	36	Superior Frontal Gyrus
2	R. AG	7.25	52	-56	18	Angular Gyrus
		6.76	40	-56	24	Angular Gyrus
		5.34	60	-38	48	Supramarginal Gyrus, posterior division
		5.34	54	-60	36	Lateral Occipital Cortex, superior division
		4.64	46	-42	10	Supramarginal Gyrus, posterior division
		4.48	64	-44	2	Middle Temporal Gyrus, temporooccipital part
		4.23	64	-46	34	Supramarginal Gyrus, posterior division
		3.67	46	-62	52	Lateral Occipital Cortex, superior division
3	PcC	6.49	6	-56	20	Precuneous Cortex
		6.28	-14	-52	32	Cingulate Gyrus, posterior division
		5.51	6	-56	34	Precuneous Cortex
		4.51	-12	-48	46	Precuneous Cortex
4	L. AG	5.30	-52	-52	20	Angular Gyrus
		5.27	-54	-64	38	Lateral Occipital Cortex, superior division
		4.77	-62	-52	34	Supramarginal Gyrus, posterior division
		4.59	-38	-60	18	Angular Gyrus
5	R. TOF	9.05	44	-48	-22	Temporal Occipital Fusiform Cortex
6	R. FOC	5.61	40	30	-12	Frontal Orbital Cortex
		4.44	26	14	-20	Frontal Orbital Cortex
7	L. MTGp	4.79	-66	-10	-24	Middle Temporal Gyrus, posterior division
8	R. LOCi	5.71	44	-80	-10	Lateral Occipital Cortex, inferior division
9	R. MTGp	5.37	60	-8	-16	Middle Temporal Gyrus, posterior division
		4.47	60	0	-26	Middle Temporal Gyrus, anterior division
10	R. Amygdala	8.46	20	-8	-16	Right Amygdala
11	L. LOCi	5.24	-44	-84	-6	Lateral Occipital Cortex, inferior division
		3.47	-50	-80	6	Lateral Occipital Cortex, inferior division
12	R. OcP	5.96	12	-98	12	Occipital Pole

Increase in object trial

Cluster index	Cluster name	t value	x (mm)	y (mm)	z (mm)	Harvard-Oxford Atlas
1	L. TOF/LOC	-15.40	-28	-52	-16	Temporal Occipital Fusiform Cortex
		-11.40	-50	-62	-8	Lateral Occipital Cortex, inferior division
		-10.60	-26	-70	-12	Occipital Fusiform Gyrus
		-9.75	-24	-64	46	Lateral Occipital Cortex, superior division
		-9.66	-30	-84	20	Lateral Occipital Cortex, superior division
		-9.64	-28	-86	4	Lateral Occipital Cortex, inferior division
		-9.44	-12	-94	-4	Occipital Pole
		-8.24	-26	-66	34	Lateral Occipital Cortex, superior division
		-7.55	-32	-30	-20	Temporal Fusiform Cortex, posterior division
		-7.31	-34	-50	44	Superior Parietal Lobule
		-6.69	-46	-30	40	Supramarginal Gyrus, anterior division
		-5.41	-12	-76	40	Precuneous Cortex
		-5.39	-24	-62	-28	Cerebellum
		-4.06	-50	-38	56	Postcentral Gyrus

		-3.86	-40	-64	-30	Cerebellum
2	R. TOF/LOC	-15.40	28	-46	-16	Temporal Occipital Fusiform Cortex
		-14.00	38	-86	14	Lateral Occipital Cortex, superior division
		-12.60	28	-62	-12	Occipital Fusiform Gyrus
		-10.10	50	-60	-8	Lateral Occipital Cortex, inferior division
		-9.81	24	-78	-10	Occipital Fusiform Gyrus
		-9.42	16	-88	0	Occipital Pole
		-7.47	28	-34	-22	Temporal Fusiform Cortex, posterior division
		-7.18	28	-68	50	Lateral Occipital Cortex, superior division
		-6.52	8	-76	-18	Lingual Gyrus
		-6.44	32	-82	32	Lateral Occipital Cortex, superior division
		-6.22	34	-72	-44	Cerebellum
		-5.98	30	-64	36	Lateral Occipital Cortex, superior division
		-5.81	14	-74	-44	Cerebellum
		-5.78	32	-68	24	Lateral Occipital Cortex, superior division
		-5.48	28	-62	-28	Cerebellum
		-5.44	22	-84	40	Lateral Occipital Cortex, superior division
		-5.10	10	-70	50	Precuneous Cortex
		-5.07	10	-84	-28	Occipital Fusiform Gyrus
		-4.88	32	-56	62	Superior Parietal Lobule
		-4.76	-8	-70	-42	Cerebellum
		-4.45	18	-64	60	Lateral Occipital Cortex, superior division
		-4.44	-26	-64	-48	Cerebellum
		-4.35	20	-60	-48	Cerebellum
		-4.14	0	-74	-32	Cerebellum
		-4.09	50	-44	-10	Inferior Temporal Gyrus, temporooccipital part
		-3.92	14	-56	-34	Cerebellum
		-3.84	-38	-56	-48	Cerebellum
		-3.45	-30	-60	-36	Cerebellum
		-3.44	26	-50	-34	Cerebellum
3	L. MFGi	-9.85	-48	10	34	Middle Frontal Gyrus
		-8.34	-46	32	14	Inferior Frontal Gyrus, pars triangularis
		-7.15	-42	24	24	Middle Frontal Gyrus
		-7.04	-30	24	4	Insular Cortex
		-4.85	-52	14	-6	Inferior Frontal Gyrus, pars opercularis
		-4.84	-30	32	-10	Frontal Orbital Cortex
		-3.98	-42	8	4	Central Opercular Cortex
		-3.92	-38	2	42	Precentral Gyrus
4	PcG	-6.81	6	10	50	Paracingulate Gyrus
		-6.63	-6	18	42	Paracingulate Gyrus
		-6.11	10	20	32	Cingulate Gyrus, anterior division
		-5.32	-8	4	54	Supplementary Motor Cortex
		-4.79	-10	-4	70	Superior Frontal Gyrus
		-4.66	-2	-10	58	Supplementary Motor Cortex
		-3.84	10	4	70	Superior Frontal Gyrus
5	L. MFGs	-5.60	-38	-2	60	Middle Frontal Gyrus
		-4.76	-22	2	58	Superior Frontal Gyrus
6	R. Ins	-5.60	32	24	4	Insular Cortex
		-4.57	54	16	-2	Inferior Frontal Gyrus, pars opercularis
		-3.79	52	14	-16	Temporal Pole
7	L. Thalamus	-6.39	-10	-18	10	Left Thalamus
		-4.05	-14	-4	16	Left Caudate
		-3.60	-10	-20	-8	Left Thalamus
8	R. SFG	-5.55	26	4	62	Superior Frontal Gyrus
		-3.96	36	-2	58	Middle Frontal Gyrus

Abbreviations. R, right; L, left; AG, angular gyrus; FG, fusiform gyrus; FMC, frontal medial cortex; FOC, frontal orbital cortex; Ins, insular cortex; LOC, lateral occipital cortex; MFG, middle frontal gyrus; MTG, middle temporal gyrus; OcP, occipital pole; OFC, orbitofrontal cortex; OP, occipital pole; PcC, precuneous cortex; PcG, paracingulate gyrus; SFG, superior frontal gyrus; TOF, temporal occipital fusiform cortex; i, inferior; s, superior; p, posterior

Figure 3 – figure supplement 2. Brain regions showing significant signal increase and decrease in the contrast of male-face vs. female-face. Coordinates are listed in MNI space.

Increase in male-face trial

Cluster index	t value	x (mm)	y (mm)	z (mm)	Harvard-Oxford Atlas

Increase in female-face trial

Cluster index	t value	x (mm)	y (mm)	z (mm)	Harvard-Oxford Atlas
1	-5.34	-30	-68	-12	Occipital Fusiform Gyrus
	-4.91	-16	-90	-6	Occipital Pole
	-4.87	-36	-78	-16	Occipital Fusiform Gyrus
	-4.84	-28	-72	22	Lateral Occipital Cortex, superior division
	-4.67	-20	-80	-14	Occipital Fusiform Gyrus
	-4.42	-42	-80	12	Lateral Occipital Cortex, inferior division
	-4.40	-54	-68	-8	Lateral Occipital Cortex, inferior division
	-4.31	-30	-54	-18	Temporal Occipital Fusiform Cortex
	-4.13	-34	-76	0	Lateral Occipital Cortex, inferior division
	-4.07	-16	-96	8	Occipital Pole
	-4.05	-48	-56	-16	Inferior Temporal Gyrus, temporooccipital part
	-4.02	-38	-88	0	Lateral Occipital Cortex, inferior division
	-3.61	-46	-78	-4	Lateral Occipital Cortex, inferior division
2	-4.84	28	-66	-14	Occipital Fusiform Gyrus
	-4.49	36	-46	-16	Temporal Occipital Fusiform Cortex
	-3.94	36	-88	-12	Lateral Occipital Cortex, inferior division
	-3.91	50	-70	-14	Lateral Occipital Cortex, inferior division
	-3.89	24	-82	-6	Occipital Fusiform Gyrus
	-3.67	42	-74	-2	Lateral Occipital Cortex, inferior division
3	-4.62	36	-76	4	Lateral Occipital Cortex, inferior division

Figure 3 – figure supplement 3. Brain regions showing significant signal increase and decrease in the contrast of natural-object vs. artificial-object. Coordinates are listed in MNI space.

Increase in natural-object trial

Cluster index	t value	x (mm)	y (mm)	z (mm)	Harvard-Oxford Atlas

Increase in artificial-object trial

Cluster index	t value	x (mm)	y (mm)	z (mm)	Harvard-Oxford Atlas
1	-5.61	38	-74	-6	Lateral Occipital Cortex, inferior division
	-5.45	38	-80	6	Lateral Occipital Cortex, inferior division
	-5.28	28	-78	-12	Occipital Fusiform Gyrus
	-4.75	34	-82	18	Lateral Occipital Cortex, superior division
	-3.95	42	-68	-16	Occipital Fusiform Gyrus
	-3.53	26	-78	0	Occipital Fusiform Gyrus
2	-4.18	-32	-92	12	Occipital Pole
	-3.66	-18	-94	22	Occipital Pole

Figure 3 – figure supplement 4. Guided Grad-CAM values for face and object trials in anatomically-defined brain regions. Values are sorted by their magnitudes. An asterisk denotes FDR q < 0.05.

Face trial

#	Label	Region	Value	FDR q
1	L. LOCi	Occipital	0.0067	1.10E-113 *
2	R. LOCi	Occipital	0.0061	1.16E-114 *
3	R. TOF	Temporal	0.0023	3.85E-93 *
4	L. OcP	Occipital	0.0013	3.43E-113 *
5	R. OcP	Occipital	0.00049	2.02E-112 *
6	R. MTGtp	Temporal	0.00041	3.33E-89 *
7	L. TOF	Temporal	0.00031	9.49E-58 *
8	L. IcC	Occipital	0.00012	1.22E-40 *
9	R. AG	Parietal	0.00010	4.93E-33 *
10	R. SmGp	Parietal	0.00010	2.74E-68 *
11	L. COpc	Frontal	8.85E-05	8.35E-99 *
12	L. PrG	Frontal	7.39E-05	1.11E-108 *
13	L. LG	Occipital	7.27E-05	7.71E-34 *
14	R. LG	Occipital	7.26E-05	2.95E-31 *
15	L. SMC	Frontal	5.43E-05	6.79E-98 *
16	L. PoG	Parietal	5.40E-05	2.75E-103 *
17	L. H1/H2	Parietal	4.98E-05	3.52E-39 *
18	L. LOCs	Parietal	4.92E-05	6.98E-51 *
19	R. LOCs	Parietal	4.40E-05	7.36E-47 *
20	L. POpc	Parietal	3.88E-05	1.12E-37 *
21	L. PcC	Parietal	3.84E-05	2.00E-67 *
22	L. SmGa	Parietal	3.82E-05	1.66E-36 *
23	R. CGa	Frontal	3.65E-05	2.41E-60 *
24	L. SPL	Parietal	2.94E-05	5.83E-85 *
25	L. CGa	Frontal	2.90E-05	7.12E-57 *
26	L. Ins	Insula	2.84E-05	1.07E-28 *
27	L. SmGp	Parietal	2.27E-05	5.32E-13 *
28	R. ScC	Frontal	2.24E-05	1.56E-53 *
29	L. PP	Temporal	2.21E-05	3.47E-19 *
30	L Putamen	Subcortex	2.21E-05	1.64E-41 *
31	R. SccC	Occipital	2.00E-05	0.00018 *
32	R. FMC	Frontal	1.95E-05	2.15E-37 *
33	L. OFG	Occipital	1.85E-05	1
34	R Amygdala	Subcortex	1.78E-05	1.78E-28 *
35	R. SMC	Frontal	1.76E-05	1.82E-47 *
36	R. CGp	Parietal	1.75E-05	9.07E-36 *
37	L. TFCp	Temporal	1.71E-05	6.56E-11 *
38	R. TFCa	Temporal	1.66E-05	1.10E-40 *
39	L. FOpc	Frontal	1.63E-05	8.63E-29 *
40	R. IcC	Occipital	1.60E-05	0.0034 *
41	L. TP	Temporal	1.60E-05	2.96E-46 *
42	R Putamen	Subcortex	1.53E-05	3.82E-27 *
43	R. FOC	Frontal	1.49E-05	9.81E-49 *
44	L. PcG	Frontal	1.46E-05	7.25E-21 *
45	R. PcG	Frontal	1.46E-05	8.66E-14 *
46	R. COpC	Frontal	1.39E-05	3.19E-13 *
47	L. AG	Parietal	1.16E-05	0.0043 *
48	R. TFCp	Temporal	1.11E-05	0.042 *
49	R. PoG	Parietal	1.01E-05	1.73E-41 *
50	R. PrG	Frontal	9.93E-06	2.87E-39 *
51	R. IFGpt	Frontal	9.56E-06	4.09E-20 *
52	L. TFCa	Temporal	6.93E-06	9.98E-07 *
53	L. FOC	Frontal	5.89E-06	2.01E-19 *
54	R. FOpC	Frontal	5.63E-06	9.07E-09 *
55	L Amygdala	Subcortex	5.61E-06	1.46E-06 *
56	Brain-Stem	Subcortex	5.33E-06	1.06E-30 *
57	R. IFGpo	Frontal	4.42E-06	1.72E-05 *
58	R. PhGa	Temporal	4.30E-06	0.0018 *
59	R. SPL	Parietal	4.03E-06	2.99E-13 *
60	R Accumbens	Subcortex	3.77E-06	1.53E-05 *
61	L. PT	Temporal	3.65E-06	0.016 *
62	L. ITGa	Temporal	3.45E-06	0.00014 *
63	L. IFGpo	Frontal	3.01E-06	0.099
64	L Thalamus	Subcortex	2.43E-06	5.62E-07 *
65	R Pallidum	Subcortex	2.23E-06	0.079
66	R. MFG	Frontal	2.03E-06	0.0056 *
67	L. MTGa	Temporal	2.03E-06	0.23
68	R Caudate	Subcortex	1.93E-06	0.0025 *
69	L Caudate	Subcortex	1.91E-06	4.15E-05 *
70	R. ITGa	Temporal	1.62E-06	0.017 *
71	L. STGa	Temporal	1.22E-06	0.022 *
72	R. SFG	Frontal	1.21E-06	0.0041 *
73	R. PcC	Parietal	6.89E-07	0.0017 *
74	R. TP	Temporal	6.14E-07	0.54
75	R. STGp	Temporal	5.48E-07	0.54
76	L. FMC	Frontal	1.49E-07	0.28
77	R Thalamus	Subcortex	1.25E-07	0.31
78	R. SmGa	Parietal	-8.87E-07	0.59
79	L. PhGa	Temporal	-1.01E-06	0.54
80	L Accumbens	Subcortex	-1.27E-06	1
81	R. Ins	Insula	-1.96E-06	0.91
82	R. MTGa	Temporal	-2.18E-06	1
83	L. MFG	Frontal	-2.39E-06	1
84	L. ScC	Frontal	-2.39E-06	1
85	R. STGa	Temporal	-2.79E-06	1
86	L. SFG	Frontal	-3.02E-06	1
87	R. PP	Temporal	-4.09E-06	1
88	L. STGp	Temporal	-6.18E-06	1
89	L Pallidum	Subcortex	-7.76E-06	1
90	R. FP	Frontal	-9.19E-06	1
91	R. MTGp	Temporal	-9.29E-06	1
92	L. CGp	Parietal	-9.78E-06	1
93	L. FP	Frontal	-1.02E-05	1
94	L. IFGpt	Frontal	-1.02E-05	1
95	L. PhGp	Temporal	-1.16E-05	1
96	L. MTGp	Temporal	-1.30E-05	1
97	L. ITGp	Temporal	-1.54E-05	1
98	R. H1/H2	Parietal	-1.65E-05	1
99	L Hippocampus	Subcortex	-2.84E-05	1
100	R. POpC	Parietal	-6.59E-05	1
101	R. ITGp	Temporal	-7.27E-05	1
102	R Hippocampus	Subcortex	-7.60E-05	1
103	L. CC	Occipital	-8.47E-05	1
104	R. CC	Occipital	-9.99E-05	1
105	R. PhGp	Temporal	-0.00012	1
106	L. MTGtp	Temporal	-0.00014	1
107	R. PT	Temporal	-0.00016	1
108	R. ITGtp	Temporal	-0.00033	1
109	L. ITGtp	Temporal	-0.00039	1
110	R. OFG	Occipital	-0.00040	1

Object trial

#	Label	Region	Value	FDR q
1	L. TOF	Temporal	0.0046	7.44E-92 *
2	L. ITGtp	Temporal	0.0035	9.32E-93 *
3	R. TOF	Temporal	0.0014	5.46E-84 *
4	L. LOCi	Occipital	0.0014	6.64E-86 *
5	L. OcP	Occipital	0.00088	8.90E-95 *
6	R. ITGtp	Temporal	0.00084	2.63E-76 *
7	R. OFG	Occipital	0.00066	6.97E-87 *
8	R. OcP	Occipital	0.00060	8.90E-95 *
9	L. OFG	Occipital	0.00049	2.21E-71 *
10	R. LOCi	Occipital	0.00045	1.47E-68 *
11	R. LOCs	Parietal	0.00020	9.14E-84 *
12	L. ITGp	Temporal	0.00010	1.70E-16 *
13	L. LOCs	Parietal	9.80E-05	3.04E-60 *
14	L. TFCp	Temporal	7.93E-05	6.54E-18 *
15	R. FOpC	Frontal	5.52E-05	4.75E-55 *
16	R. SPL	Parietal	3.93E-05	1.22E-67 *
17	L. IFGpt	Frontal	3.74E-05	1.15E-18 *
18	R. IcC	Occipital	3.49E-05	7.43E-12 *
19	L. SPL	Parietal	2.82E-05	7.56E-47 *
20	L. PrG	Frontal	2.71E-05	9.67E-62 *
21	L. LG	Occipital	2.66E-05	5.88E-06 *
22	R. TFCp	Temporal	2.62E-05	9.81E-11 *
23	R Caudate	Subcortex	1.83E-05	2.79E-24 *
24	L. SMC	Frontal	1.76E-05	3.32E-09 *
25	R. PcC	Parietal	1.73E-05	1.89E-26 *
26	L. POpc	Parietal	1.50E-05	1.16E-29 *
27	R. CC	Occipital	1.30E-05	2.72E-05 *
28	L. H1/H2	Parietal	1.30E-05	3.78E-14 *
29	R. ITGa	Temporal	1.05E-05	2.85E-33 *
30	R Thalamus	Subcortex	9.74E-06	1.80E-34 *
31	R. CGa	Frontal	8.03E-06	1.18E-17 *
32	L. ITGa	Temporal	7.80E-06	1.20E-29 *
33	R. CGp	Parietal	6.71E-06	0.00098 *
34	R. PrG	Frontal	6.39E-06	2.71E-21 *
35	R. PoG	Parietal	5.97E-06	1.22E-38 *
36	R. SMC	Frontal	5.66E-06	2.33E-10 *
37	R. PcG	Frontal	5.60E-06	3.42E-09 *
38	R. LG	Occipital	5.42E-06	1
39	L. FOC	Frontal	4.93E-06	1.92E-17 *
40	L. TFCa	Temporal	4.65E-06	5.25E-11 *
41	L Caudate	Subcortex	4.58E-06	1.58E-13 *
42	R. IFGpo	Frontal	4.52E-06	4.07E-10 *
43	Brain-Stem	Subcortex	3.49E-06	2.52E-15 *
44	R. STGa	Temporal	3.12E-06	3.21E-08 *
45	L. CGp	Parietal	3.01E-06	0.0040 *
46	L Thalamus	Subcortex	2.81E-06	5.80E-08 *
47	L. CGa	Frontal	2.70E-06	0.0051 *
48	R. POpC	Parietal	2.68E-06	3.36E-05 *
49	L. MTGa	Temporal	2.63E-06	0.0031 *
50	R. MTGa	Temporal	2.63E-06	3.37E-07 *
51	R. FOC	Frontal	2.07E-06	1.96E-06 *
52	R. PhGa	Temporal	1.69E-06	0.0082 *
53	L. MFG	Frontal	1.62E-06	0.0067 *
54	R Putamen	Subcortex	1.30E-06	0.042 *
55	R. PT	Temporal	9.14E-07	0.17
56	R. SmGa	Parietal	8.67E-07	0.27
57	L. FP	Frontal	6.47E-07	0.0019 *
58	L. ScC	Frontal	3.93E-07	0.12
59	L. PoG	Parietal	3.44E-07	0.00012 *
60	L Accumbens	Subcortex	2.39E-07	0.012 *
61	L. PhGa	Temporal	2.26E-07	0.68
62	R. Ins	Insula	3.67E-08	1
63	L Putamen	Subcortex	-1.04E-08	1
64	R Accumbens	Subcortex	-2.52E-08	0.73
65	R. TP	Temporal	-3.08E-07	0.52
66	R Pallidum	Subcortex	-4.39E-07	1
67	L Pallidum	Subcortex	-4.97E-07	1
68	L. TP	Temporal	-1.10E-06	1
69	R. COpC	Frontal	-1.24E-06	1
70	L. IFGpo	Frontal	-1.28E-06	0.30
71	R. MFG	Frontal	-1.43E-06	1
72	L. PcG	Frontal	-1.70E-06	1
73	L. FOpc	Frontal	-2.24E-06	0.62
74	R. SFG	Frontal	-2.71E-06	1
75	R. H1/H2	Parietal	-2.91E-06	1
76	L. STGa	Temporal	-2.97E-06	1
77	R. PP	Temporal	-3.36E-06	1
78	R Amygdala	Subcortex	-3.41E-06	1
79	L Amygdala	Subcortex	-3.51E-06	1
80	R. ScC	Frontal	-3.55E-06	1
81	L. Ins	Insula	-3.58E-06	1
82	L. FMC	Frontal	-3.67E-06	1
83	L. SmGa	Parietal	-3.78E-06	1
84	L. SFG	Frontal	-3.92E-06	1
85	L. COpc	Frontal	-4.58E-06	1
86	R. FP	Frontal	-5.28E-06	1
87	R. FMC	Frontal	-5.34E-06	1
88	R. SccC	Occipital	-5.57E-06	1
89	R. TFCa	Temporal	-7.03E-06	1
90	R. AG	Parietal	-8.02E-06	1
91	R. STGp	Temporal	-8.02E-06	1
92	L. SmGp	Parietal	-1.04E-05	1
93	L. CC	Occipital	-1.05E-05	1
94	R. SmGp	Parietal	-1.19E-05	1
95	R. ITGp	Temporal	-1.26E-05	1
96	L. STGp	Temporal	-1.37E-05	1
97	R. MTGtp	Temporal	-1.55E-05	1
98	L. PP	Temporal	-1.59E-05	1
99	L. IcC	Occipital	-1.80E-05	1
100	R. IFGpt	Frontal	-1.88E-05	1
101	R. MTGp	Temporal	-1.91E-05	1
102	L Hippocampus	Subcortex	-1.91E-05	1
103	L. PcC	Parietal	-2.22E-05	1
104	L. PT	Temporal	-3.06E-05	1
105	R Hippocampus	Subcortex	-4.91E-05	1
106	L. AG	Parietal	-5.36E-05	1
107	R. PhGp	Temporal	-5.48E-05	1
108	L. MTGp	Temporal	-6.69E-05	1
109	L. PhGp	Temporal	-0.00011	1
110	L. MTGtp	Temporal	-0.00030	1

A All voxels (masked by avg152T1_gray.img)

B Selected voxels (masked by Neurosynth-defined ROIs)

Figure 3 – figure supplement 5

Figure 3 – figure supplement 5. Voxel-based relationship between t-values in the univariate analysis and the guided Grad-CAM values in the face and object trial. (A) Relationship between the two values when voxels are masked by FSL's gray matter prior volume. **(B)** Relationship between the two values when voxels are masked by functional ROIs obtained from Neurosynth using the terms "face" and "object." *, $P < 0.001$.

Figure 3 – figure supplement 6

Figure 3 — figure supplement 6. ROI analysis. Guided Grad-CAM values for each ROI when ROIs are defined **(A)** anatomically (Harvard–Oxford atlas) and **(B)** functionally (Neurosynth). Figure configurations are the same as in Figure 3f. L, left; R, right; AG, angular gyrus; FMC, frontal medial cortex; FOC, frontal orbital cortex; FP, frontal pole; IFG, inferior frontal gyrus; Ins, insular cortex; LOC, lateral occipital cortex; MFG, middle frontal gyrus; MTG, middle temporal gyrus; OcP, occipital pole; OFG, occipital fusiform gyrus; PcC, precuneous cortex; PcG, paracingulate gyrus; SFG, superior frontal gyrus; SPL, superior parietal lobule; TOF, temporal occipital fusiform cortex; i, inferior; s, superior; p, posterior.

Figure 4 – figure supplement 1

Figure 4 — figure supplement 1. EEG responses. (A) ERPs for face trials, object trials, male face trials, female face trials, natural object trials, and artificial object trials. Line colors correspond to the channel location depicted on the left side. A, anterior, P, posterior, R, right, L, left. **(B–D)** Significance of temporal trajectories in ERPs and power spectrograms. Colored lines depict time points showing significant differential ERP amplitude or power spectrogram against zero in respective channels. The color of each line depicts the location of the corresponding channels (left). Time points that exceeded the 97.5th percentile of the permutation distribution ($N = 5,000$) served as cluster-inducing time points, and the significance of clusters is set at the threshold of $P < 0.05$.

Figure 4 – figure supplement 2

Figure 4 — figure supplement 2. Regional analysis of guided Grad-CAM.
(A–C) Comparison of the guided Grad-CAM values at the maximum latency in
face and object trials (A), in male face and female face trials (B), and in natural
object and artificial object trials (C) across six regions. All 63 channels are
categorized as either frontal, central, parietal, occipital, left temporal, and right
temporal regions based on their locations. Shown are trial-based median value
distributions across channels in a region. Regional differences in the maximum
guided Grad-CAM values are statistically significant in all trial types (Friedman's
test, $P < 0.001$). **(D–F)** Results of multiple comparison test (Tukey–Kramer
method, $P < 0.05$). Significant differences in the guided Grad-CAM values
between regions are color-coded.

References

Allen, P.J., Josephs, O., and Turner, R. (2000). A method for removing imaging artifact from continuous EEG recorded during functional MRI. Neuroimage *12*, 230-239. 10.1006/nimg.2000.0599.

Bai, S., Kolter, J.Z., and Koltun, V. (2018). An Empirical Evaluation of Generic Convolutional and Recurrent Networksfor Sequence Modeling. arXiv:1803.01271v2. 10.48550/arXiv.1803.01271.

Bar, M. (2004). Visual objects in context. Nat Rev Neurosci *5*, 617-629. 10.1038/nrn1476.

Barton, J.J. (2011). Disorder of higher visual function. Curr Opin Neurol *24*, 1-5. 10.1097/WCO.0b013e328341a5c2.

Birn, R.M., Diamond, J.B., Smith, M.A., and Bandettini, P.A. (2006). Separating respiratory-variation-related fluctuations from neuronal-activity-related fluctuations in fMRI. Neuroimage *31*, 1536-1548. 10.1016/j.neuroimage.2006.02.048.

Carlson, T., Tovar, D.A., Alink, A., and Kriegeskorte, N. (2013). Representational dynamics of object vision: the first 1000 ms. J Vis *13*, 1. 10.1167/13.10.1.

Cavina-Pratesi, C., Kentridge, R.W., Heywood, C.A., and Milner, A.D. (2010). Separate processing of texture and form in the ventral stream: evidence from FMRI and visual agnosia. Cereb Cortex *20*, 433-446. 10.1093/cercor/bhp111.

Cichy, R.M., and Oliva, A. (2020). A M/EEG-fMRI Fusion Primer: Resolving Human Brain Responses in Space and Time. Neuron *107*, 772-781. 10.1016/j.neuron.2020.07.001.

Cichy, R.M., Pantazis, D., and Oliva, A. (2014). Resolving human object recognition in space and time. Nat Neurosci *17*, 455-462. 10.1038/nn.3635.

Dobs, K., Isik, L., Pantazis, D., and Kanwisher, N. (2019). How face perception unfolds over time. Nat Commun *10*, 1258. 10.1038/s41467-019-09239-1.

Eklund, A., Nichols, T.E., and Knutsson, H. (2016). Cluster failure: Why fMRI inferences for spatial extent have inflated false-positive rates. Proc Natl Acad Sci U S A *113*, 7900-7905. 10.1073/pnas.1602413113.

Epstein, R., and Kanwisher, N. (1998). A cortical representation of the local visual environment. Nature *392*, 598-601. 10.1038/33402.

Freedman, D.J., Riesenhuber, M., Poggio, T., and Miller, E.K. (2001). Categorical representation of visual stimuli in the primate prefrontal cortex. Science *291*, 312-316. 10.1126/science.291.5502.312.

Frey, M., Nau, M., and Doeller, C.F. (2021). Magnetic resonance-based eye tracking
using deep neural networks. Nat Neurosci *24*, 1772-1779. 10.1038/s41593-021-
00947-w.
Gainotti, G., and Marra, C. (2011). Differential contribution of right and left temporo-
occipital and anterior temporal lesions to face recognition disorders. Front Hum
Neurosci *5*, 55. 10.3389/fnhum.2011.00055.
Geusebroek, J., Burghouts, G.J., and Smeulders, A.W.M. (2005). The Amsterdam
Library of Object Images. Int. J. Comput. Vis. *61*, 103-112.
10.1023/B:VISI.0000042993.50813.60.
Grill-Spector, K., and Weiner, K.S. (2014). The functional architecture of the ventral
temporal cortex and its role in categorization. Nat Rev Neurosci *15*, 536-548.
10.1038/nrn3747.
Grill-Spector, K., Weiner, K.S., Kay, K., and Gomez, J. (2017). The Functional
Neuroanatomy of Human Face Perception. Annu Rev Vis Sci *3*, 167-196.
10.1146/annurev-vision-102016-061214.
Harvey, A.K., Pattinson, K.T., Brooks, J.C., Mayhew, S.D., Jenkinson, M., and Wise,
R.G. (2008). Brainstem functional magnetic resonance imaging: disentangling signal
from physiological noise. J Magn Reson Imaging *28*, 1337-1344.
10.1002/jmri.21623.
Haxby, J.V., Gobbini, M.I., Furey, M.L., Ishai, A., Schouten, J.L., and Pietrini, P.
(2001). Distributed and overlapping representations of faces and objects in ventral
temporal cortex. Science *293*, 2425-2430. 10.1126/science.1063736.
Hong, H., Yamins, D.L., Majaj, N.J., and DiCarlo, J.J. (2016). Explicit information for
category-orthogonal object properties increases along the ventral stream. Nat
Neurosci *19*, 613-622. 10.1038/nn.4247.
Horikawa, T., and Kamitani, Y. (2017). Generic decoding of seen and imagined objects
using hierarchical visual features. Nat Commun *8*, 15037. 10.1038/ncomms15037.
Hung, C.P., Kreiman, G., Poggio, T., and DiCarlo, J.J. (2005). Fast readout of object
identity from macaque inferior temporal cortex. Science *310*, 863-866.
10.1126/science.1117593.
Ishai, A., Ungerleider, L.G., Martin, A., Schouten, J.L., and Haxby, J.V. (1999).
Distributed representation of objects in the human ventral visual pathway. Proc Natl
Acad Sci U S A *96*, 9379-9384. 10.1073/pnas.96.16.9379.
Jiang, P.T., Zhang, C.B., Hou, Q., Cheng, M.M., and Wei, Y. (2021). LayerCAM:
Exploring Hierarchical Class Activation Maps for Localization. IEEE Trans Image
Process *30*, 5875-5888. 10.1109/TIP.2021.3089943.

Kanwisher, N., McDermott, J., and Chun, M.M. (1997). The fusiform face area: a module in human extrastriate cortex specialized for face perception. J Neurosci *17*, 4302-4311. 10.1523/JNEUROSCI.17-11-04302.1997.

Kasper, L., Bollmann, S., Diaconescu, A.O., Hutton, C., Heinzle, J., Iglesias, S., Hauser, T.U., Sebold, M., Manjaly, Z.M., Pruessmann, K.P., and Stephan, K.E. (2017). The PhysIO Toolbox for Modeling Physiological Noise in fMRI Data. J Neurosci Methods *276*, 56-72. 10.1016/j.jneumeth.2016.10.019.

Keerativittayayut, R., Aoki, R., Sarabi, M.T., Jimura, K., and Nakahara, K. (2018). Large-scale network integration in the human brain tracks temporal fluctuations in memory encoding performance. Elife *7*, e32696. 10.7554/eLife.32696.

Kingma, D., and Ba, J. (2015). Adam: A Method for Stochastic Optimization. ICLR 2015. 10.48550/arXiv.1412.6980.

Konen, C.S., Behrmann, M., Nishimura, M., and Kastner, S. (2011). The functional neuroanatomy of object agnosia: a case study. Neuron *71*, 49-60. 10.1016/j.neuron.2011.05.030.

Kourtzi, Z., and Kanwisher, N. (2000). Cortical regions involved in perceiving object shape. J Neurosci *20*, 3310-3318. 10.1523/JNEUROSCI.20-09-03310.2000.

Kreiman, G., Koch, C., and Fried, I. (2000). Category-specific visual responses of single neurons in the human medial temporal lobe. Nat Neurosci *3*, 946-953. 10.1038/78868.

Kriegeskorte, N., Formisano, E., Sorger, B., and Goebel, R. (2007). Individual faces elicit distinct response patterns in human anterior temporal cortex. Proc Natl Acad Sci U S A *104*, 20600-20605. 10.1073/pnas.0705654104.

Kriegeskorte, N., Mur, M., Ruff, D.A., Kiani, R., Bodurka, J., Esteky, H., Tanaka, K., and Bandettini, P.A. (2008). Matching categorical object representations in inferior temporal cortex of man and monkey. Neuron *60*, 1126-1141. 10.1016/j.neuron.2008.10.043.

Landi, S.M., Viswanathan, P., Serene, S., and Freiwald, W.A. (2021). A fast link between face perception and memory in the temporal pole. Science *373*, 581-585. 10.1126/science.abi6671.

Liu, J., Harris, A., and Kanwisher, N. (2002). Stages of processing in face perception: an MEG study. Nat Neurosci *5*, 910-916. 10.1038/nn909.

Logothetis, N.K., and Sheinberg, D.L. (1996). Visual object recognition. Annu Rev Neurosci *19*, 577-621. 10.1146/annurev.ne.19.030196.003045.

Mack, M.L., and Palmeri, T.J. (2011). The timing of visual object categorization. Front Psychol *2*, 165. 10.3389/fpsyg.2011.00165.

Malach, R., Reppas, J.B., Benson, R.R., Kwong, K.K., Jiang, H., Kennedy, W.A.,
Ledden, P.J., Brady, T.J., Rosen, B.R., and Tootell, R.B. (1995). Object-related
activity revealed by functional magnetic resonance imaging in human occipital
cortex. Proc Natl Acad Sci U S A *92*, 8135-8139. 10.1073/pnas.92.18.8135.
Maris, E., and Oostenveld, R. (2007). Nonparametric statistical testing of EEG- and
MEG-data. J Neurosci Methods *164*, 177-190. 10.1016/j.jneumeth.2007.03.024.
Matsui, T., Hattori, Y., Tsumura, K., Aoki, R., Takeda, M., Nakahara, K., and Jimura,
K. (2022). Executive control by fronto-parietal activity explains counterintuitive
decision behavior in complex value-based decision-making. Neuroimage *249*,
118892. 10.1016/j.neuroimage.2022.118892.
Miyakawa, N., Majima, K., Sawahata, H., Kawasaki, K., Matsuo, T., Kotake, N.,
Suzuki, T., Kamitani, Y., and Hasegawa, I. (2018). Heterogeneous Redistribution of
Facial Subcategory Information Within and Outside the Face-Selective Domain in
Primate Inferior Temporal Cortex. Cereb Cortex *28*, 1416-1431.
10.1093/cercor/bhx342.
Miyashita, Y., and Chang, H.S. (1988). Neuronal correlate of pictorial short-term
memory in the primate temporal cortex. Nature *331*, 68-70. 10.1038/331068a0.
Nestor, A., Plaut, D.C., and Behrmann, M. (2011). Unraveling the distributed neural
code of facial identity through spatiotemporal pattern analysis. Proc Natl Acad Sci U
S A *108*, 9998-10003. 10.1073/pnas.1102433108.
Peelen, M.V., and Downing, P.E. (2017). Category selectivity in human visual cortex:
Beyond visual object recognition. Neuropsychologia *105*, 177-183.
10.1016/j.neuropsychologia.2017.03.033.
Raschka, S. (2020). Model Evaluation, Model Selection, and Algorithm Selection in
Machine Learning. arXiv:1811.12808v3. 10.48550/arXiv.1811.12808.
Rokach, L. (2009). Ensemble-based classifiers. Artificial Intelligence Review *33*, 1-39.
10.1007/s10462-009-9124-7.
Rousselet, G.A., Fabre-Thorpe, M., and Thorpe, S.J. (2002). Parallel processing in high-
level categorization of natural images. Nat Neurosci *5*, 629-630. 10.1038/nn866.
Sarraf, S., DeSouza, D.D., Anderson, J., and Tofighi, G. (2017). DeepAD: Alzheimer's
Disease Classification via Deep Convolutional Neural Networks using MRI and
fMRI. bioRxiv. 10.1101/070441.
Schiltz, C., Sorger, B., Caldara, R., Ahmed, F., Mayer, E., Goebel, R., and Rossion, B.
(2006). Impaired face discrimination in acquired prosopagnosia is associated with
abnormal response to individual faces in the right middle fusiform gyrus. Cereb
Cortex *16*, 574-586. 10.1093/cercor/bhj005.

Selvaraju, R.R., Cogswell, M., Das, A., Vedantam, R., Parikh, D., and Batra, D. (2019).

Grad-CAM: Visual Explanations from Deep Networks via Gradient-Based

Localization. International Journal of Computer Vision *128*, 336-359.

10.1007/s11263-019-01228-7.

Serre, T., Oliva, A., and Poggio, T. (2007). A feedforward architecture accounts for

rapid categorization. Proc Natl Acad Sci U S A *104*, 6424-6429.

10.1073/pnas.0700622104.

Simonyan, K., and Zisserman, A. (2015). Very Deep Convolutional Networks for

Large-Scale Image Recognition. ICLR2015. 10.48550/arXiv.1409.1556.

Stigliani, A., Weiner, K.S., and Grill-Spector, K. (2015). Temporal Processing Capacity

in High-Level Visual Cortex Is Domain Specific. J Neurosci *35*, 12412-12424.

10.1523/JNEUROSCI.4822-14.2015.

Supratak, A., Dong, H., Wu, C., and Guo, Y. (2017). DeepSleepNet: A Model for

Automatic Sleep Stage Scoring Based on Raw Single-Channel EEG. IEEE Trans

Neural Syst Rehabil Eng *25*, 1998-2008. 10.1109/TNSRE.2017.2721116.

Suzuki, W.A., and Naya, Y. (2014). The perirhinal cortex. Annu Rev Neurosci *37*, 39-

53. 10.1146/annurev-neuro-071013-014207.

Takeda, M., Hirabayashi, T., Adachi, Y., and Miyashita, Y. (2018). Dynamic laminar

rerouting of inter-areal mnemonic signal by cognitive operations in primate temporal

cortex. Nat Commun *9*, 4629. 10.1038/s41467-018-07007-1.

Tanaka, D., Aoki, R., Suzuki, S., Takeda, M., Nakahara, K., and Jimura, K. (2020).

Self-Controlled Choice Arises from Dynamic Prefrontal Signals That Enable Future

Anticipation. J Neurosci *40*, 9736-9750. 10.1523/JNEUROSCI.1702-20.2020.

Tanaka, K. (1996). Inferotemporal cortex and object vision. Annu Rev Neurosci *19*,

109-139. 10.1146/annurev.ne.19.030196.000545.

Tsao, D.Y., Freiwald, W.A., Knutsen, T.A., Mandeville, J.B., and Tootell, R.B. (2003).

Faces and objects in macaque cerebral cortex. Nat Neurosci *6*, 989-995.

10.1038/nn1111.

Tsao, D.Y., Freiwald, W.A., Tootell, R.B., and Livingstone, M.S. (2006). A cortical

region consisting entirely of face-selective cells. Science *311*, 670-674.

10.1126/science.1119983.

Tsumura, K., Aoki, R., Takeda, M., Nakahara, K., and Jimura, K. (2021a). Cross-

Hemispheric Complementary Prefrontal Mechanisms during Task Switching under

Perceptual Uncertainty. J Neurosci *41*, 2197-2213. 10.1523/JNEUROSCI.2096-

20.2021.

Tsumura, K., Kosugi, K., Hattori, Y., Aoki, R., Takeda, M., Chikazoe, J., Nakahara, K.,
and Jimura, K. (2021b). Reversible Fronto-occipitotemporal Signaling Complements
Task Encoding and Switching under Ambiguous Cues. Cereb Cortex, bhab324.
10.1093/cercor/bhab324.

Turesson, H.K., Logothetis, N.K., and Hoffman, K.L. (2012). Category-selective phase
coding in the superior temporal sulcus. Proc Natl Acad Sci U S A *109*, 19438-19443.
10.1073/pnas.1217012109.

Vu, H., Kim, H.C., Jung, M., and Lee, J.H. (2020). fMRI volume classification using a
3D convolutional neural network robust to shifted and scaled neuronal activations.
Neuroimage *223*, 117328. 10.1016/j.neuroimage.2020.117328.

Wang, X., Liang, X., Jiang, Z., Nguchu, B.A., Zhou, Y., Wang, Y., Wang, H., Li, Y.,
Zhu, Y., Wu, F., et al. (2020). Decoding and mapping task states of the human brain
via deep learning. Hum Brain Mapp *41*, 1505-1519. 10.1002/hbm.24891.

Willenbockel, V., Sadr, J., Fiset, D., Horne, G.O., Gosselin, F., and Tanaka, J.W.
(2010). Controlling low-level image properties: the SHINE toolbox. Behav Res
Methods *42*, 671-684. 10.3758/BRM.42.3.671.

Worsley, K.J., and Friston, K.J. (1995). Analysis of fMRI time-series revisited--again.
Neuroimage *2*, 173-181. 10.1006/nimg.1995.1023.

Yamins, D.L., and DiCarlo, J.J. (2016). Using goal-driven deep learning models to
understand sensory cortex. Nat Neurosci *19*, 356-365. 10.1038/nn.4244.

Yarkoni, T., Poldrack, R.A., Nichols, T.E., Van Essen, D.C., and Wager, T.D. (2011).
Large-scale automated synthesis of human functional neuroimaging data. Nat
Methods *8*, 665-670. 10.1038/nmeth.1635.

Yu, F., Koltun, V., and Funkhouser, T. (2017). Dilated Residual Networks. CVPR 2017,
636-644. 10.1109/cvpr.2017.75.

Zhang, Z., Yu, X., Rong, X., and Iwata, M. (2021). Spatial-Temporal Neural Network
for P300 Detection. IEEE Access *9*, 163441-163455. 10.1109/access.2021.3132024.

Zhao, Y., Dong, Q., Zhang, S., Zhang, W., Chen, H., Jiang, X., Guo, L., Hu, X., Han, J.,
and Liu, T. (2018). Automatic Recognition of fMRI-Derived Functional Networks
Using 3-D Convolutional Neural Networks. IEEE Trans Biomed Eng *65*, 1975-1984.
10.1109/TBME.2017.2715281.

Zheng, X., Mondloch, C.J., and Segalowitz, S.J. (2012). The timing of individual face
recognition in the brain. Neuropsychologia *50*, 1451-1461.
10.1016/j.neuropsychologia.2012.02.030.