

1 **Title:** Genomic underpinnings of population persistence in Isle Royale moose

2

3 **Authors:** Christopher C. Kyriazis^{1*}, Annabel C. Beichman², Kristin E. Brzeski³, Sarah R. Hoy³, Rolf
4 O. Peterson³, John A. Vucetich³, Leah M. Vucetich³, Kirk E. Lohmueller^{1,4,5†*}, Robert K. Wayne^{1†*}

5

6 ¹Department of Ecology and Evolutionary Biology, University of California, Los Angeles, CA
7 90095, USA.

8 ²Department of Genome Sciences, University of Washington, Seattle, WA, 98195 USA

9 ³College of Forest Resources and Environmental Science, Michigan Technological University,
10 Houghton, MI 49931, USA.

11 ⁴Interdepartmental Program in Bioinformatics, University of California, Los Angeles, CA 90095,
12 USA.

13 ⁵Department of Human Genetics, David Geffen School of Medicine, University of California, Los
14 Angeles, CA 90095, USA.

15

16 *correspondence: ckyriazis@g.ucla.edu, klohmueLLer@g.ucla.edu, rwayne@ucla.edu

17 †these authors contributed equally to this work

18

19 **Keywords:** *Alces alces*, bottlenecks, genetic load, inbreeding depression, purging

20 **Abstract:**

21 Island ecosystems provide models to assess the impacts of isolation on population persistence.
22 However, most studies of persistence have focused on a single species, without comparisons to
23 other organisms they interact with in the ecosystem. The simple predator-prey system of
24 moose and gray wolves on Isle Royale provides allows a direct contrast of genetic variation in a
25 prey species with their natural predator. Wolves on Isle Royale exhibited signs of severe
26 inbreeding depression, which nearly drove the population to extinction in 2019. In the relative
27 absence of wolves, the moose population has thrived and exhibits no obvious signs of
28 inbreeding depression despite being isolated for ~120 years and having low genetic diversity.
29 Here, we examine the genomic underpinnings of population persistence in the Isle Royale
30 moose population. We document high levels of inbreeding in the population, roughly as high as
31 the wolf population at the time of its decline. However, inbreeding in the moose population
32 manifests in the form of intermediate-length runs of homozygosity indicative of gradual
33 inbreeding, contrasting with the severe recent inbreeding observed in the wolf population.
34 Using simulations, we demonstrate that this more gradual inbreeding in the moose population
35 has resulted in an estimated 50% purging of the inbreeding load, helping to explain the
36 continued persistence of the population. However, we also document notable increases in
37 genetic load, which could eventually threaten population viability over the long term. Finally,
38 we document low diversity in mainland North American moose populations due to a severe
39 founder event occurring near the end of the Holocene. Overall, our results demonstrate a
40 complex relationship between inbreeding, genetic diversity, and population viability that
41 highlights the importance of maintaining isolated populations at moderate size to avert
42 extinction from genetic factors.

43

44 **Significance statement:**

45 Isolated wildlife populations face a high risk of extinction due in part to the deleterious
46 consequences of inbreeding. Whether purifying natural selection can overcome these negative
47 impacts by “purging” harmful recessive mutations is a topic of active debate. We characterized
48 the extent of purging in an isolated moose population. Our results demonstrate signatures of
49 gradual inbreeding in the population, ideal circumstances to facilitate purging. Using
50 simulations, we demonstrate substantial potential for purging in the population, though we
51 also show that fitness is reduced by small population size and inbreeding. Our findings provide
52 insight into the mechanisms enabling persistence in isolated populations, with implications for
53 conserving the growing number of isolated populations worldwide.

54

55 **Introduction**

56 Anthropogenic habitat fragmentation has dramatically increased the number of isolated and
57 inbred populations (1). To conserve these populations, a crucial question is whether they will be
58 able to persist in isolation, or if they will be driven to extinction by deleterious genetic factors,
59 such as inbreeding depression (2). Numerous examples exist of inbreeding depression driving
60 population decline in isolated populations (reviewed in (3)). However, in some populations,
61 harmful recessive mutations may potentially be ‘purged’ by purifying selection and such
62 purging may avert inbreeding depression (2, 4–8). Purging may be most effective in populations
63 where inbreeding is gradual due to a moderate population size (4–6, 9–11). However, the
64 extent to which purging is a relevant factor for the conservation of threatened populations, and
65 more broadly, the degree to which populations can persist with low genome-wide diversity, is
66 controversial (9, 12–18).

67
68 One of the best-studied examples of inbreeding depression driving population decline is the
69 gray wolf population on Isle Royale, an island in Lake Superior roughly 544 km² in area. After
70 ~70 years of isolation at a population size of ~25 individuals, the Isle Royale wolf population
71 declined nearly to extinction, with just two individuals remaining in the population in 2018 (19).
72 Recent research has demonstrated that this population collapse was a consequence of severe
73 inbreeding depression in the form of widespread congenital deformities (20, 21). The decline of
74 the Isle Royale wolf population allowed its main prey, moose, to thrive. The most recent moose
75 census count was ~2000 individuals, though the population generally numbers ~1000
76 individuals (19). Moreover, despite the moose population having low genetic diversity and
77 being isolated on the island for ~120 years (22–25), it exhibits no obvious signs of inbreeding
78 depression and has population growth rates similar to mainland populations (26). Thus, the
79 contrasting fates of the Isle Royale wolf and moose populations provides a compelling case
80 study for understanding the genetic underpinnings of population persistence in isolation and
81 effects on predator-prey dynamics.

82

83 Outside of the Isle Royale population, North American moose are also known to have low
84 genetic diversity relative to Eurasian moose, which is thought to be a consequence of a
85 relatively recent founder event following the Last Glacial Maximum (27–29). Evidence for this
86 recent founder also comes from a relative lack of population structure across North America as
87 well as the near absence of moose in the North American fossil record prior to 15,000 years ago
88 (27–30). Depending on how recent and severe this founding bottleneck was, the effects of
89 purging associated with the bottleneck may still be apparent in the North American moose
90 population. Thus, the ability of moose to persist in isolation on Isle Royale may be enhanced by
91 purging from historical bottlenecks.

92
93 Here, we use a dataset of high coverage whole genome sequences from 20 North American
94 moose and one Eurasian moose to characterize the impacts of bottlenecks, population
95 isolation, and purging in North American moose, focusing on the Isle Royale population. We
96 confirm previous findings of low genetic diversity in North American moose, especially Isle
97 Royale moose, where levels of inbreeding are comparable to that of the Isle Royale gray wolf
98 population at the time of its decline. Furthermore, we demonstrate that this low diversity is a
99 consequence of severe founder events in both the North American and Isle Royale populations.
100 Finally, we conduct extensive simulations exploring the impact of bottlenecks and population
101 isolation on genetic load and purging in North American moose. These results suggest
102 substantial purging associated with founding bottlenecks for the North American and Isle
103 Royale populations. However, this purging also has been accompanied by a notable increase in
104 genetic load. Overall, our analysis provides insight into how populations can persist despite
105 severe bottlenecks and high inbreeding and emphasizes the importance of maintaining
106 moderate population size to ensure viability in isolated populations. Moreover, our results
107 highlight the differential impacts of inbreeding depression in isolated predator and prey
108 populations, with implications for maintaining healthy ecosystems in the increasingly-
109 fragmented landscape of the Anthropocene.

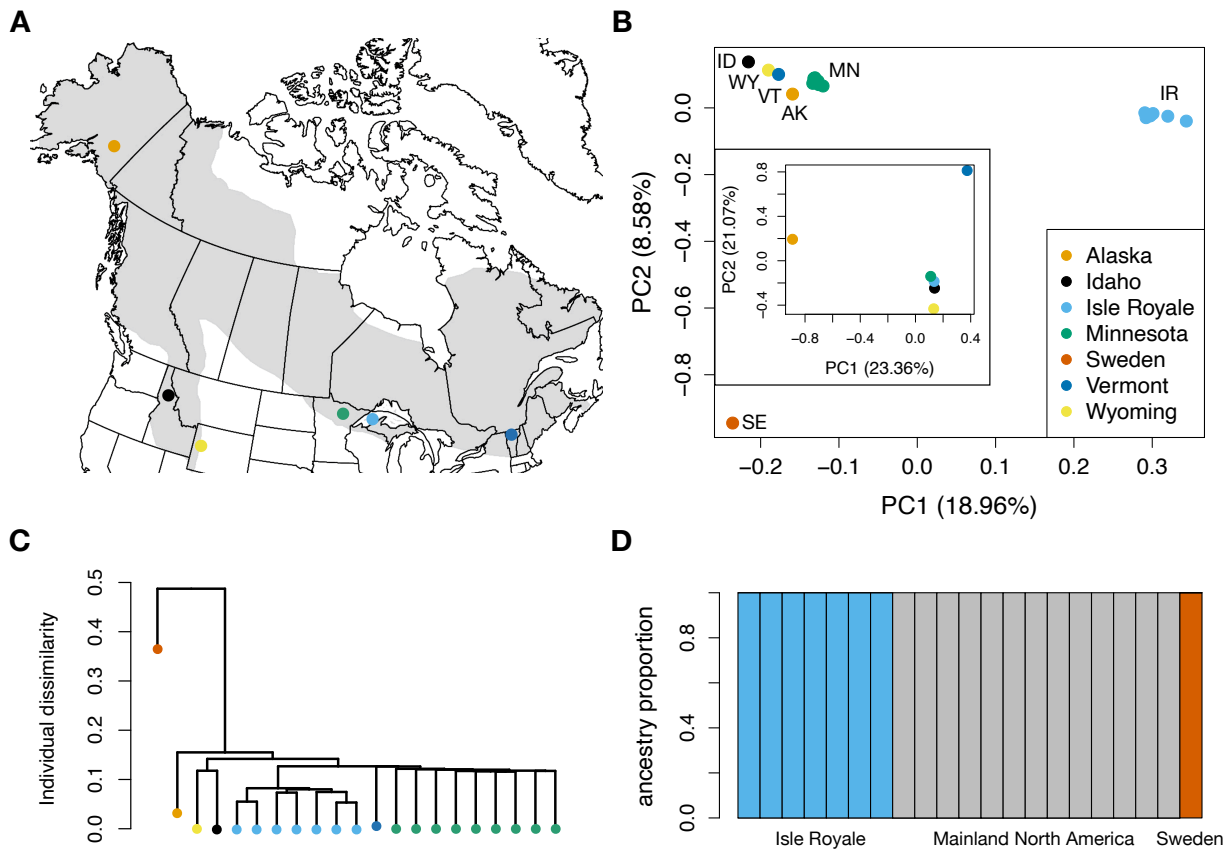


Figure 1: Moose sampling and population structure. (A) Map of North America including localities for individuals sampled for genomic data in our study. Note that Sweden is excluded. (B) PCA of 50,361 LD-pruned SNPs for all sequenced samples. Inset are results when down-sampling to one individual per population and excluding the Swedish sample. (C) Tree based on identity-by-state constructed using 50,361 LD-pruned SNPs. (D) fastSTRUCTURE results for K=3. See Fig. S1 for results with varying K values and Fig. S2 for results when down-sampling to four unrelated individuals each from Isle Royale and Minnesota.

110

111 Results

112 *Sampling and population structure*

113 To examine patterns of moose genetic diversity in North America, we generated a high-
114 coverage whole genome sequencing dataset for nine moose sampled from Minnesota and
115 seven moose sampled from Isle Royale between 2005 and 2014. We added existing moose
116 genomes to our dataset from Sweden, Alaska, Idaho, Wyoming, and Vermont. These genomes
117 were aligned, genotyped, and annotated relative to the cattle reference genome (ARS-UCD1.2).
118 Although a moose reference genome was recently published (30), we used the more distantly-

119 related cattle reference in order to leverage its fully assembled chromosomes and high-quality
120 annotations (see SI for further discussion). Average sequencing coverage after mapping was 21x
121 (range 11-27; Table S1).

122
123 We first used these data to characterize population structure among North American moose,
124 primarily aiming to assess evidence for isolation of the Isle Royale population. Principal
125 component analysis (PCA) revealed a tight clustering of Isle Royale samples relative to other
126 North American samples, which were distinctly clustered on the first PC (Fig. 1B). However,
127 when down-sampled to one individual per North American population, the Isle Royale and
128 Minnesota samples grouped more closely together, with overall patterns roughly reflecting
129 North American geography (Fig. 1B, inset). Nevertheless, we observe notable differentiation
130 between Isle Royale and Minnesota samples, with a mean $F_{ST} = 0.083$. These patterns were also
131 reflected in a tree based on identity-by-state, which found a tight clustering of Isle Royale
132 samples nested within other North American samples (Fig. 1C). Furthermore, using
133 fastSTRUCTURE analysis we found no evidence for admixture between Isle Royale and mainland
134 samples (Fig. 1D and S1-S2). Finally, we also estimated kinship for all North American samples,
135 and found that the mainland samples are not closely related to one another (Fig. S3). However,
136 two pairs of samples from Isle Royale exhibited kinship coefficients consistent with first-order
137 relationships (mean kinship = 0.234; Fig. S3). In summary, these findings suggest that the Isle
138 Royale population has been entirely isolated from nearby mainland moose populations as
139 suggested by previous work (24, 25), and provide a general characterization of moose
140 population structure in North America.

141

142 ***Genetic diversity and inbreeding***

143 Next, we examined levels of genetic diversity and inbreeding across sampled individuals.
144 Overall, we find that moose have relatively low diversity compared to other mammals (Fig. 2),
145 though these estimates may be slightly downward biased due to using a distant reference
146 genome (see SI for discussion). However, these biases do not impact estimates of relative
147 diversity across moose populations, where several notable patterns are apparent. First, we

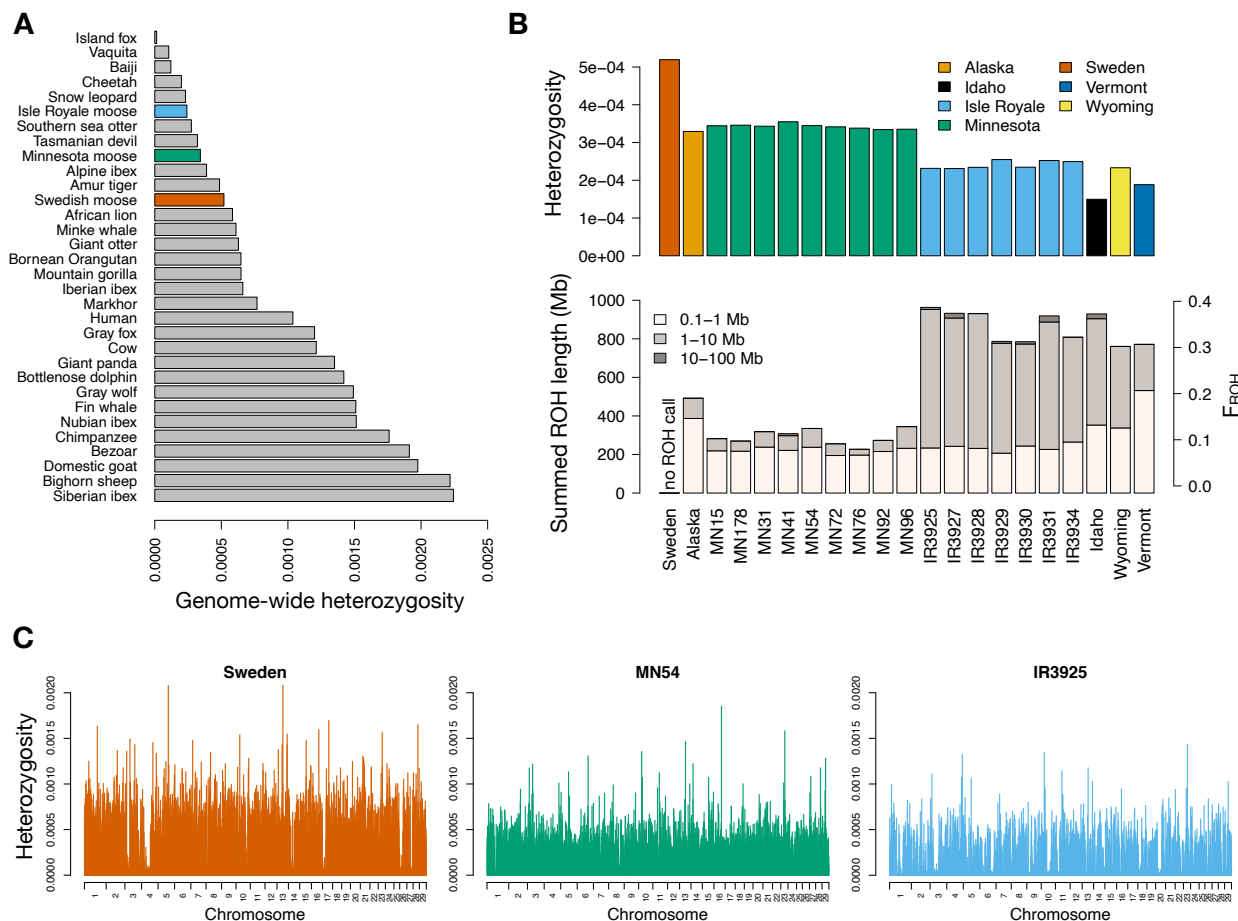


Figure 2: Moose genetic diversity and inbreeding. (A) Comparison of mean genome-wide diversity in three moose populations to published values for other mammals. (B). Plots of mean genome-wide diversity and summed ROH levels for North American moose genomes, with the corresponding F_{ROH} values on the right-hand axis. Note that we were not able to obtain ROH calls for the Sweden sample due to its differing population origin. (C) Per-site heterozygosity plotted in non-overlapping 1 Mb windows for representative individuals from Sweden, Minnesota, and Isle Royale. See Fig. S4 for plots of all individuals.

148 observe substantially lower diversity in North American samples relative to a sample from
 149 Sweden, with a decrease of at least $\sim 34\%$ (Fig. 2). This decrease in diversity is likely associated
 150 with a founder event for North American moose that is thought to have occurred during the
 151 last $\sim 15,000$ years (27–29). We observe further reductions in diversity in the Isle Royale
 152 population, with an estimated reduction of $\sim 30\%$ compared to samples from Minnesota (Fig. 2).
 153 Surprisingly, we find even lower diversity in mainland samples from Idaho, Wyoming, and
 154 Vermont, possibly due to these samples being near the southern range edge, where population
 155 densities are generally low and declining ((31); Fig. 2).

156
157 Mirroring these patterns of genetic diversity, the impact of inbreeding was prevalent across
158 North American samples in the form of abundant runs of homozygosity (ROH), chromosomal
159 segments that are inherited identical by descent from a recent common ancestor (32).
160 Specifically, we observed high levels of inbreeding in samples from Isle Royale, Vermont, Idaho,
161 and Wyoming, with ~35% of their autosomal genomes being covered by ROH >100 kb on
162 average (Fig. 2) and ~26% covered by ROH >1 Mb (Fig. S5). As this fraction represents an
163 estimate of the inbreeding coefficient (F_{ROH}), this result suggests that these populations are on
164 average more inbred than an offspring from a full-sib mating ($F=0.25$). Notably, these levels of
165 inbreeding are comparable to the Isle Royale gray wolf population, where ~20-50% of their
166 autosomal genomes contained ROH >100 kb (20). By contrast, much lower levels of inbreeding
167 were present in samples from Minnesota, Alaska, and Sweden, with ~12% of these genomes
168 covered by ROH >100 kb (Fig. 2), and ~3% covered in ROH >1 Mb (Fig. S5).

169 170 ***Demographic inference***

171 To understand the demographic processes accounting for these patterns of genetic diversity
172 and inbreeding, we fitted demographic models to the site frequency spectrum (SFS) using $\partial a \partial i$
173 (33). Briefly, this approach uses observed allele frequency information to estimate demographic
174 parameters for a model with an arbitrary number of population size changes (epochs). Our first
175 aim was to estimate the severity of the North American founding bottleneck, given the
176 apparent impact of this bottleneck on observed levels of genetic diversity between Eurasian
177 and North American moose (Fig. 2; (27)). We generated a folded SFS for our Minnesota sample,
178 and inferred various population size change models including one, two, three, and four epoch
179 models. Overall, the best-fitting model was a four-epoch model that included two ancestral
180 epochs followed by a severe bottleneck to an effective population size (N_e) of 49 for 29
181 generations and then expansion to $N_e=193,472$ for the last 1,179 generations (Fig. 3).
182 Bottlenecks that are mild with long duration can lead to similar patterns in the SFS as short and
183 severe bottlenecks (34). Consequently, we found a similar fit for a model with a slightly more
184 prolonged and less severe bottleneck of $N_e=218$ for 142 generations followed by expansion to

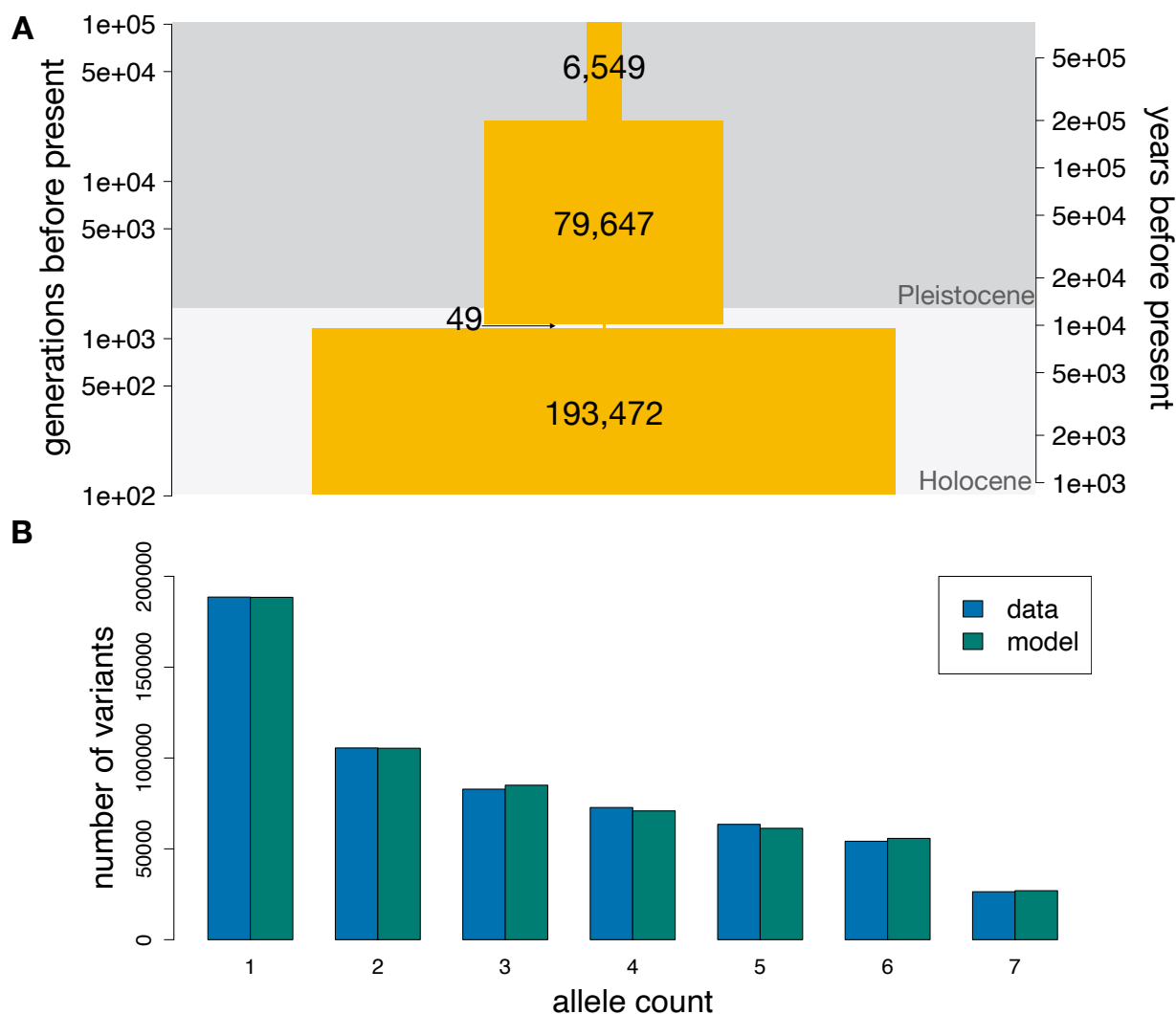


Figure 3: Demographic inference results. (A) Schematic of best-fit four epoch model based on the site frequency spectrum (SFS) for the Minnesota sample. Right-hand axis assumes a generation time of 8 years. Numbers denote maximum likelihood estimates of the effective population sizes at the various time points. Note the rapid and severe bottleneck occurring near the onset of the Holocene. See Table S2 for parameters of the second-best fitting run, which differs somewhat in bottleneck duration and magnitude and pre/post-bottleneck population sizes. (B) Comparison of the empirical projected folded SFS from the Minnesota sample with the SFS predicted by the model in shown in (A).

185 $N_e=105,531$ for the last 1,223 generations (Table S2). Overall, both of these models are
 186 consistent in detecting a strong bottleneck of $N_e = \sim 50-225$ for $\sim 30-150$ generations followed by
 187 dramatic population growth taking place $\sim 1,200$ years ago. The timing of expansion at $\sim 1,200$
 188 generations suggests a recent spread of moose across North America starting $\sim 9,600$ years ago,
 189 assuming a generation time of 8 years (35).

190

191 Our next aim for demographic inference was to obtain an estimate of the effective population
192 size of the Isle Royale moose population after its founding ~120 years ago using the SFS from
193 our Isle Royale sample. Given the shared evolutionary history of the Minnesota and Isle Royale
194 populations prior to their divergence, we fixed the demographic parameters of our four-epoch
195 model inferred from the Minnesota samples (Fig. 3), then added a fifth epoch to this model
196 representing the founding of Isle Royale. Furthermore, we fixed the timing of this fifth epoch to
197 15 generations ago, thus assuming that the population was founded in the early 1900s (120
198 years ago, assuming a generation time of 8 years; (35)), as suggested by available evidence (22,
199 23). We used this approach to retain power for estimating the Isle Royale effective population
200 size when fitting a complex five-epoch model to an SFS from a small sample size. When fixing
201 these parameters, we obtained an estimate of $N_e=187$ on Isle Royale, highlighting a dramatic
202 disparity in N_e between the North American and Isle Royale populations spanning three orders
203 of magnitude. Additionally, given that the Isle Royale moose population on average numbers
204 ~1000 individuals (19), these results suggest an $N_e:N$ ratio of ~0.19, consistent with those
205 observed in other species (36). Notably, we observe the same $N_e:N$ ratio of ~0.19 when
206 comparing our estimated North American $N_e=193,472$ (Fig. 3) to the current census estimate of
207 one million (31).

208

209 ***Quantifying putatively deleterious variation***

210 To understand how the vastly reduced effective population size on Isle Royale may have
211 impacted patterns of deleterious variation compared to mainland populations, we examined
212 variants in protein-coding regions that were predicted to be putatively damaging or benign on
213 the basis of evolutionary constraint (37). We observe a reduction in heterozygosity for both
214 damaging and benign variants on Isle Royale, mirrored by an increase in homozygosity for the
215 derived (i.e., mutant relative to the reference) allele (Fig. 4), as expected given the higher levels
216 of inbreeding in the Isle Royale population. Specifically, we find that homozygous derived
217 genotype counts are 9.7% higher for damaging variants and 6.8% higher for benign variants in
218 Isle Royale moose compared to mainland moose. However, we do not observe an excess of
219 derived alleles on Isle Royale (Fig. 4), as might be expected for a population that has

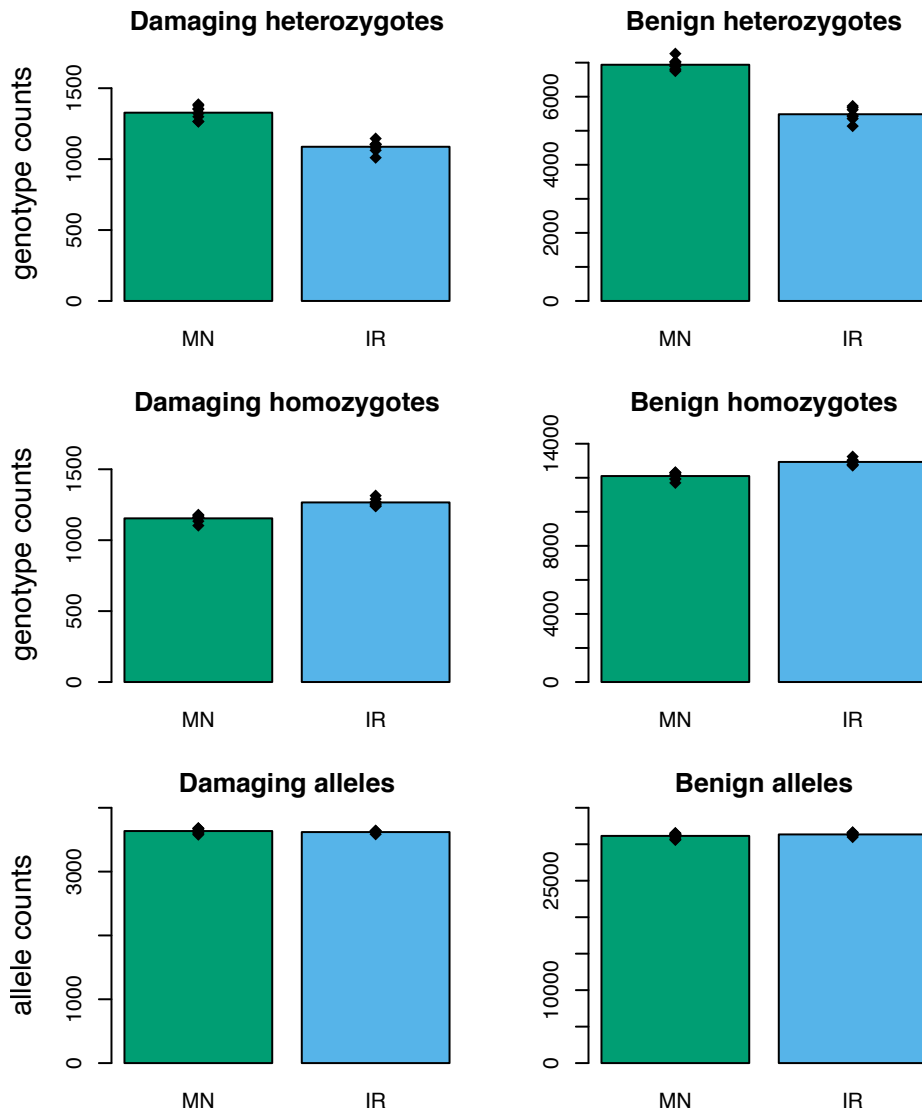


Figure 4: Empirical measures of deleterious variation in Isle Royale and Minnesota moose. Top row depicts counts of putatively damaging and benign heterozygotes, demonstrating that heterozygosity is reduced for both mutation types on Isle Royale. Middle row depicts counts of homozygotes for the derived allele at damaging and benign variants, similarly demonstrating increased homozygosity for both mutation types on Isle Royale. Bottom row depicts damaging and benign derived allele counts, demonstrating no differences between Isle Royale and Minnesota.

220 accumulated an excess of weakly deleterious mutations due to relaxed purifying selection (38,
221 39). Collectively, these results suggest that the genetic load attributable to an accumulation of
222 weakly deleterious mutations is negligible in Isle Royale moose.

223

224 *Simulations of deleterious variation and genetic load*

225 Empirical measures of deleterious variation are often challenging to interpret given that the
226 functional impact and dominance of mutations are uncertain (40, 41). Consequently, we also

227 conducted forward-in-time genetic simulations to assess the impact of bottlenecks on
228 deleterious genetic variation in North American moose using SLiM3 (42). These simulations
229 consisted of a 20 Mb chromosomal segment, which included a combination of introns, exons,
230 and intergenic regions. Neutral and deleterious mutations occurred at a rate of $7e-9$ per base
231 pair (30), with deleterious mutations only occurring within exons. Selection coefficients for
232 deleterious mutations were drawn from a distribution estimated from human genetic variation
233 data (43), and dominance coefficients were assumed to be inversely related to selection
234 coefficients, such that the most deleterious mutations were also the most recessive (see
235 Materials and Methods).

236
237 Our first aim was to examine the impact of the North American colonization bottleneck on
238 genetic diversity, genetic load, and purging. Here, we define “genetic load” as the realized
239 reduction in fitness due to segregating and fixed deleterious mutations (44), and quantify
240 purging as a reduction in the simulated “inbreeding load”, a measure of the quantity of
241 recessive deleterious variation concealed in heterozygosity (2). To examine the dynamics of
242 inbreeding, genetic diversity, and load in North American moose, we simulated under our best-
243 fit demographic model (Fig. 3), which includes a founding bottleneck of $N_e=49$ for 29
244 generations followed by expansion to $N_e=193,472$ for 1,179 generations. Over the duration of
245 this bottleneck, we observe a decrease in genetic diversity of 21%, along with a decrease in the
246 inbreeding load of 24%, an increase in genetic load of 282% and an increase in F_{ROH} to 0.22 (Fig.
247 5). However, these increases in genetic load and F_{ROH} are largely absent after 1,179 generations
248 of recovery, though levels of inbreeding notably remain above zero, in agreement with our
249 empirical data (Fig. 2B). By contrast, genetic diversity and inbreeding load do not greatly
250 increase after recovery, with the inbreeding load continuing to decline after the bottleneck and
251 remaining 34% below its pre-bottleneck value even after 1,179 generations of recovery (Fig. 5).
252 Thus, this result suggests that the North American moose population may still be experiencing
253 the lingering purging effects of this founding bottleneck, despite occurring $\sim 9,600$ years ago.
254 Importantly, we observe qualitatively similar patterns when simulating under a model with a

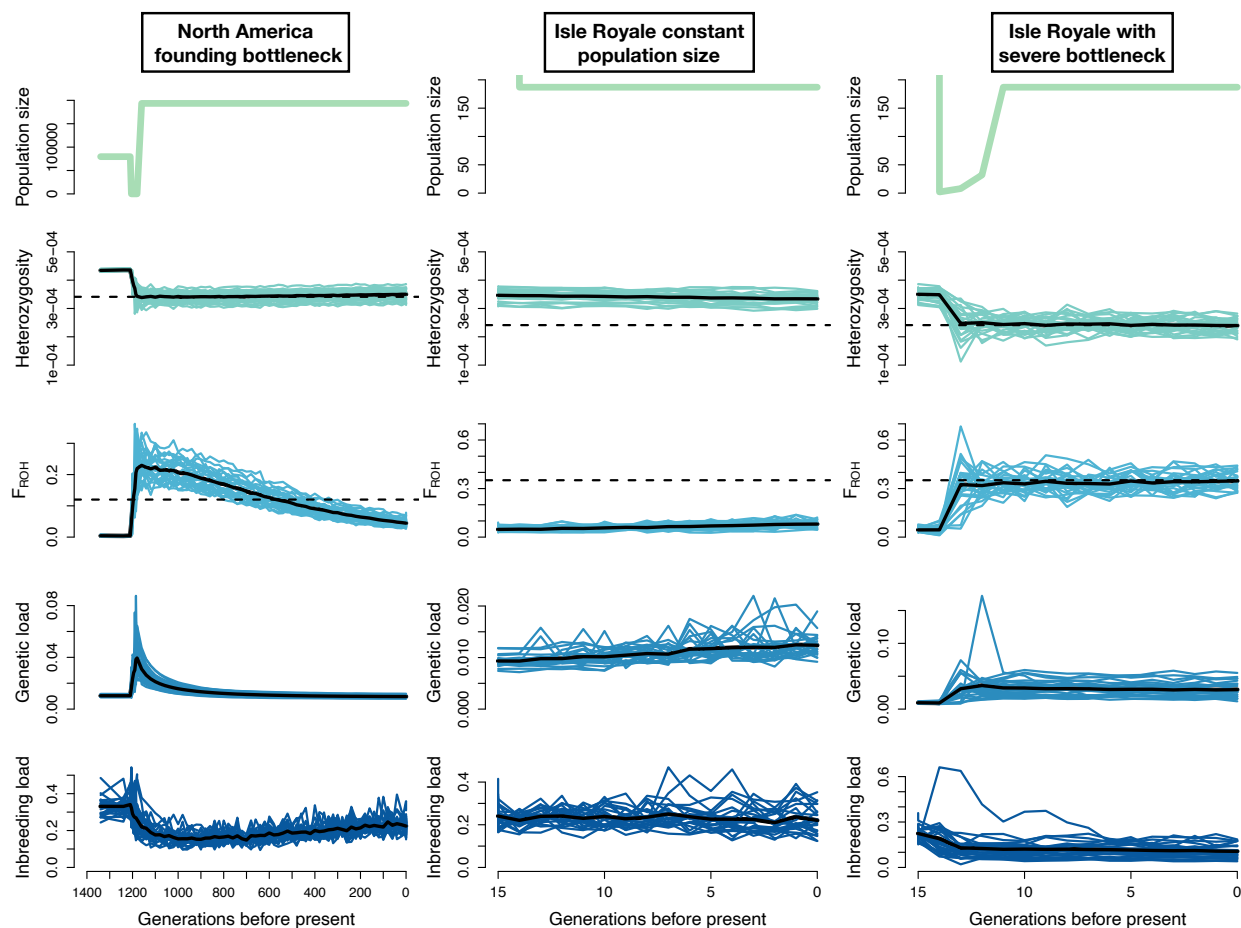


Figure 5: Simulation results under three demographic scenarios. Left column depicts simulation dynamics during the North America founding bottleneck; middle column depicts results when simulating the Isle Royale population at constant population size; right column depicts results when simulating the Isle Royale population including a severe founder event ($N_e=\{2,8,32\}$ for the first three generations). Each column includes plots of the simulated effective population size, mean heterozygosity, mean levels of inbreeding ($F_{ROH>100kb}$), mean genetic load, and mean inbreeding load from 25 simulation replicates. The black line represents the average from all replicates. The dashed lines represent the empirical estimates for heterozygosity and F_{ROH} from the Minnesota and Isle Royale populations, respectively. Note that the simulation trajectories do not reach these empirical estimates when assuming constant population size (middle column) but do when a founder event is included (right column). See Fig. S7 for results under additional bottleneck parameters.

255 slightly longer and less severe bottleneck (Fig. S7), suggesting that these simulation results are
 256 robust to uncertainty in our estimated demographic parameters.

257

258 Next, we examined the impact of isolation and small population size on Isle Royale on patterns
 259 of genetic variation and genetic load. We again simulated under our North America

260 demographic model, though added a final epoch with the estimated Isle Royale demographic

261 parameters of $N_e=187$ for 15 generations. When simulating under this demography, however,
262 we do not recapitulate the differences in genetic diversity and inbreeding observed in our
263 empirical data between Isle Royale and mainland samples (Fig. 5). Specifically, heterozygosity
264 decreased by only 3.6% compared to a ~30% difference between Minnesota and Isle Royale
265 samples in our empirical data, and levels of inbreeding increase only to $F_{ROH}=0.08$ compared to
266 $F_{ROH}=0.35$ from our empirical data (Table S3).

267
268 We hypothesized that this discrepancy may be due to the absence of a severe founder event at
269 the origination of the Isle Royale population in our model, given that the population is believed
270 to be founded by a small number of individuals (22, 23). To test this hypothesis, we ran
271 simulations where we included a bottleneck during the first three generations following the
272 founding of Isle Royale. We tested three bottleneck severities with effective population sizes
273 during the first three generations of $N_e=\{6,24,96\}$, $N_e=\{4,16,64\}$, and $N_e=\{2,8,32\}$, each followed
274 by expansion to $N_e=187$ for the final 12 generations. These bottleneck parameters were
275 selected because available evidence suggests that population density was low soon after
276 founding, particularly from 1900-1920, though it is unclear exactly how low or how many
277 founders there were (22, 23). When varying these bottleneck parameters, we find that only the
278 most severe bottleneck of $N_e=\{2,8,32\}$ recapitulated the observed differences in genetic
279 diversity and inbreeding, yielding a decrease in heterozygosity of 32% and increase in
280 inbreeding to $F_{ROH}=0.35$, in agreement with our empirical results (Figs. 5 and S7-S8; Table S3).
281 Under this model, we also observe a relative increase in genetic load on Isle Royale of 206% as
282 well as a 53% reduction in the inbreeding load (Fig. 5; Table S3). Thus, these results suggest that
283 the Isle Royale moose population may have been founded by just two individuals, and that this
284 severe founder event has been an essential factor in shaping patterns of genetic diversity,
285 inbreeding, genetic load, and purging on the island. Finally, we do not observe any differences
286 in allele counts between simulated island and mainland populations for mutations with
287 selection coefficient (s) > -0.01 (Fig. S9), in agreement with our empirical result suggesting
288 negligible impacts on load due to weakly deleterious mutations (Fig. 4). However, we do
289 observe a sharp reduction in the number of strongly deleterious ($s < -0.1$) alleles per individual

290 in the simulated Isle Royale population, suggesting that purging has largely been driven by a
291 reduction in the number of strongly deleterious recessive alleles (Fig. S9).

292
293 Although our results suggest a substantial decrease in genetic diversity and increase in
294 inbreeding in Isle Royale moose, field observations of the population have not detected obvious
295 signs of inbreeding depression or reduced population growth rates (26). We hypothesized that
296 this may be in part due to the purging that occurred during the North America founding event,
297 which could enhance the ability of North American moose to persist at small population size. To
298 test this hypothesis, we ran simulations under the above parameters including a severe Isle
299 Royale founding bottleneck, but excluding the North America founding bottleneck. Here, we
300 observe a much greater increase in genetic load on Isle Royale of 350%, compared to 206%
301 when including the North America founding event (Table S3). Thus, these results suggest that
302 the lingering effects of purging due to the North American founder event may have aided the
303 ability of moose to persist at small population size on Isle Royale. In other words, the negative
304 genetic consequences of small population size on Isle Royale may have been greater if the
305 North American moose population had not experienced a strong bottleneck during
306 colonization.

307
308 Next, we explored the potential impact of a low rate of historical migration on genetic variation
309 in the Isle Royale population. Specifically, we explored the effect of a low rate of migration on
310 genetic diversity, genetic load, levels of inbreeding, and inbreeding load. We ran simulations
311 with migration fractions of 0.5% and 5%, roughly corresponding to 1 and 10 effective migrants
312 per generation, respectively, chosen to model two relatively low but plausible rates of
313 migration. Under the low migration scenario of 0.5%, results are nearly identical to the no
314 migration scenario (Fig. S10; Table S3), implying that a very low level of historical migration (~1
315 migrant per generation) would not have had much impact on the genetic state of the
316 population. These results imply that we cannot fully rule out the possibility of a low rate of
317 migration to Isle Royale, as suggested by direct observations of moose swimming between Isle
318 Royale and the mainland (45). By contrast, when the migration fraction is increased to 5%,

319 heterozygosity is higher and inbreeding lower relative to empirical values (Fig. S10; Table S3). In
320 sum, these results further confirm that historical migration to Isle Royale was either absent or
321 very low. Moreover, these results also suggest that any future attempts to restore genetic
322 diversity and reduce genetic load in the Isle Royale moose population would require a relatively
323 high rate of migration (>10 effective migrants per generation).

324
325 Finally, we explored the sensitivity of our results to selection and dominance parameters.
326 Specifically, we simulated under parameters proposed by Kardos et al. (12), which assume that
327 inbreeding depression is primarily due to recessive lethals and that deleterious mutations with
328 $s > -0.1$ have largely additive effects on fitness. When simulating the North America founder
329 event with these parameters, we observe a much smaller 22% increase in genetic load and a
330 more substantial 60% decrease in the inbreeding load (Fig. S11). Additionally, the inbreeding
331 load recovers much more rapidly following the bottleneck, due to the faster increase towards
332 equilibrium of recessive lethal mutations (Fig. S11). When simulating a severe founder event for
333 Isle Royale, we observe a much greater initial increase in genetic load; however, genetic load
334 quickly decreases as recessive lethals are purged from the population, with a net increase of
335 66% (Fig. S12). Additionally, we observe substantial purging on Isle Royale, with a 75%
336 reduction in the inbreeding load (Fig. S12). Thus, simulations under these parameters predict a
337 much smaller increase in genetic load and much larger impacts of purging. This greater impact
338 of purging is likely a consequence of the increased emphasis on recessive lethals in this model,
339 which are most easily purged (5, 46).

340

341 **Discussion**

342 Highly inbred populations are often thought to be doomed to extinction. However, some can
343 persist, and understanding the factors enabling persistence can aid in conservation efforts. Our
344 results document high inbreeding in the Isle Royale moose population ($F_{ROH}=0.35$ on average;
345 Fig. 2), roughly as high as the gray wolf population at the time of its decline. Yet, despite these
346 high levels of inbreeding, the Isle Royale moose population does not exhibit obvious signs of
347 inbreeding depression, and maintains population growth rates that do not noticeably differ

348 from mainland moose (26). A key factor that likely underlies these different outcomes is the
349 pace of inbreeding in these two populations: whereas the wolf population became quickly
350 inbred while isolated at a population size of ~ 25 for ~ 70 years, inbreeding in the moose
351 population was more gradual due to its more moderate population size of ~ 1000 for a longer
352 duration of ~ 120 years. These differing demographic histories are reflected in the distribution
353 of ROH lengths in the wolf and moose populations. In the wolf population, ROH were
354 predominantly long (>10 Mb), reflecting recent and severe inbreeding (20), whereas the moose
355 population exhibits an abundance of intermediate-length ROH (1-10 Mb; Fig. 2). Several recent
356 studies have highlighted the severe fitness consequences of long ROH, which tend to be
357 enriched for highly deleterious recessive alleles, whereas more intermediate-length ROH may
358 be largely purged of such variation (20, 47–49). Although our results imply an elevated genetic
359 load in the Isle Royale moose population (Fig. 5), this load has apparently not impacted
360 population growth rates substantially, perhaps due to reduced interspecific competition on Isle
361 Royale and soft selection (50). Overall, our results emphasize the importance of maintaining
362 moderate size ($N_e > 100$) in isolated populations to enable purging and avert extinction in the
363 short to intermediate term, in agreement with other studies (4–6, 9–11). Over the longer term,
364 maintaining even larger population sizes ($N_e > 1000$) is preferable whenever possible to avoid
365 the impacts of increasing drift load and loss of adaptive potential (12, 18).

366
367 Our results suggest that roughly half of the inbreeding load in Isle Royale moose may have been
368 purged in the ~ 15 generations or ~ 120 years since founding (Fig. 5). The relatively rapid pace of
369 this purging is notable, given that most existing examples of purging in wild populations
370 occurred after thousands of years of isolation (4, 8, 51, 52). In Isle Royale moose, purging
371 appears to have been accelerated by a severe founding bottleneck of perhaps just two
372 individuals (Fig. 5). The impacts of severe bottlenecks on purging are well known (44), and have
373 also been recently documented in an analysis of Alpine ibex genomes (7). For both Isle Royale
374 moose and Alpine ibex, a severe bottleneck followed by relatively prompt recovery appears to
375 have driven rapid purging on a timescale of ~ 100 years. Thus, rapid purging on the timescale of
376 anthropogenic fragmentation may only be possible in the presence of severe bottlenecks,

377 perhaps precluding intentional purging [as](#) a viable conservation strategy. Nevertheless, many
378 populations of at-risk species may have experienced historical purging due to severe
379 bottlenecks or long-term moderate population size and identifying these populations could
380 prove useful for future management actions.

381
382 Our findings also have important implications for understanding the evolutionary history and
383 conservation status of mainland North American moose populations. Across all North American
384 moose samples, we observe a reduction in genome-wide diversity of at least 34% relative to a
385 sample from Sweden (Fig. 2), consistent with previous work (27, 30). Our demographic
386 modeling indicates this reduction in diversity is due to a severe bottleneck in the ancestral
387 North American moose population occurring ~9,600 years ago (Fig. 3). This timing closely aligns
388 with glacial recession at the onset of the Holocene 11,000 years ago as well as the North
389 American fossil record (29). Furthermore, our simulation results suggest a substantial 34%
390 purging of the inbreeding load associated with this founding bottleneck, the effects of which
391 may persist until present day (Fig. 5). This phenomenon could further explain the success of the
392 isolated Isle Royale moose population, implying that the founding individuals may have been
393 'pre-purged' of inbreeding depression. Moreover, the possibility of 'pre-purging' in North
394 American moose could also help explain the success of other introduced moose populations in
395 North American, such as the Newfoundland population, which was founded by just six
396 individuals and now numbers >100,000 individuals (53). Nevertheless, many fragmented North
397 American moose populations near the southern range edge have experienced recent declines
398 (31). Though these declines have generally been linked to synergistic impacts of climate change
399 and increasing disease and pathogen load (31, 54), the potential role of genetic factors has
400 been largely overlooked. For example, we observed low genetic diversity in samples from Idaho
401 and Wyoming (Fig. 2), perhaps due to the recent founding of these populations in the mid 19th
402 century and low population density (55). Notably, moose in this region exhibit low adult
403 pregnancy rates (56), which could potentially be a consequence of inbreeding depression.
404 Moreover, it is possible that low genetic diversity in these populations has increased their
405 susceptibility to parasites (57). Overall, the causes of moose population declines near the

406 southern range edge appear to be complex, and additional genomic sampling of these
407 populations will be necessary to more fully investigate the potential role of genetic factors.

408

409 In conclusion, our results depict a complex relationship between genetic diversity, inbreeding,
410 and population viability in isolated and fragmented populations. The contrasting fates of the
411 Isle Royale wolf and moose populations serve as a dramatic example of the importance of
412 maintaining isolated populations at moderate size to facilitate purging and avert extinction over
413 the short to intermediate term. Moreover, this case study of predator and prey hints at a more
414 far-reaching phenomenon, in which isolated predator populations may be doomed to
415 extinction by inbreeding depression due to their naturally lower density, whereas the higher
416 abundance of prey populations may enable them to purge the most severe impacts of
417 inbreeding depression. In light of the well-documented connections among gray wolf, moose
418 and plant abundance on Isle Royale (58), we suggest the possibility of an eco-evolutionary link
419 between purging and the dynamics of the Isle Royale ecosystem. In general, purging may have
420 system-wide effects in other isolated and fragmented ecosystems, where predator populations
421 are declining in part due to inbreeding depression, and prey populations are thriving in their
422 absence, often to the detriment of the broader ecosystem (59, 60). Thus, our results highlight a
423 unique connection between deleterious genetic variation and ecosystem health, with
424 implications for best management practices of small and fragmented populations.

425

426 **Materials and Methods**

427 **Sampling and sequencing**

428 Tissue samples were obtained opportunistically from moose carcasses on Isle Royale and
429 Minnesota samples were collected during regular management activities by the Minnesota
430 Department of Natural Resources (MN DNR). Isle Royale tissue samples were frozen and
431 archived at Michigan Technological University and Minnesota tissue samples were provided by
432 the MN DNR. DNA was extracted from samples using Qiagen kits and quantified using a Qubit
433 fluorometer. Whole-genome sequencing was performed on an Illumina NovaSeq at the Vincent
434 J. Coates Genomics Sequencing Laboratory at University of California, Berkeley and
435 MedGenome. Existing genomes from (61) and (30) were downloaded from the National Center
436 for Biotechnology Information (NCBI) Sequence Read Archive (see Table S1).

437

438 **Read processing and alignment**

439 We processed raw reads using a pipeline adapted from the Genome Analysis Toolkit (GATK)
440 (62) Best Practices Guide. We aligned paired-end 150bp raw sequence reads to the cattle
441 genome (ARS-UCD1.2) using BWA-MEM (63), followed by removal of low-quality reads and PCR
442 duplicates. Given that we do not have a database of known variants, we did not carry out Base
443 Quality Score Recalibration, but instead carried out hard filtering of genotypes (see below).
444 Although the cattle genome is highly divergent from moose, we opted to use it due to its much
445 higher quality and contiguity compared to existing moose genomes (scaffold N50 of 103 Mb for
446 ARS-UCD1.2 vs 1.7 Mb for NRM_Aalces_1_0) as well as its high-quality annotations and existing
447 resources on the Ensembl Variant Effect Predictor database (64). To explore the potential
448 impact of this on our downstream analyses, we also mapped a subset of nine genomes to the
449 more closely related hog deer reference genome (ASM379854v1), which has high contiguity
450 with a scaffold N50 of 20.7 Mb. Importantly, we found that the choice of reference genome
451 here does not appear to qualitatively impact our genetic diversity and runs of homozygosity
452 results. Thus, we use the cattle reference genome for all downstream analyses (see SI text for
453 further discussion).

454

455 **Genotype calling and filtering**

456 We performed joint genotype calling at all sites (including invariant sites) using GATK
457 HaplotypeCaller. Genotypes were filtered to include only high-quality biallelic SNPs and
458 monomorphic sites, removing sites with Phred score below 30 and depth exceeding the 99th
459 percentile of total depth across samples. In addition, we removed sites that failed slightly
460 modified GATK hard filtering recommendations (QD < 4.0 || FS > 12.0 || MQ < 40.0 ||
461 MQRankSum < -12.5 || ReadPosRankSum < -8.0 || SOR > 3.0), as well as those with >25% of
462 genotypes missing or >35% of genotypes heterozygous. We masked repetitive regions using a
463 mask file downloaded from ftp://ftp.ncbi.nlm.nih.gov/genomes/Bos_taurus/. Finally, we
464 applied a per-individual excess depth filter, removing genotypes exceeding the 99th percentile
465 of depth for each individual, as well as a minimum depth filter of six reads.

466

467 **Population structure and relatedness**

468 We used SNPrelate v1.14 (65) to run principal component analysis (PCA), construct a tree based
469 on identity-by-state (IBS), and estimate kinship among sampled genomes. For all analyses, we
470 pruned SNPs for linkage (ld.threshold=0.2) and filtered out sites with minor allele frequency
471 below 0.05, resulting in 50,361 SNPs for analysis. PCA was run both for all sampled individuals
472 as well as for North American individuals down-sampled to one individual per population. We
473 used the KING method of moments approach (66) to estimate kinship among North American
474 moose samples. Finally, we estimated IBS among all samples, then performed hierarchical
475 clustering on the resulting matrix to construct a dendrogram.

476

477 As another means of characterizing population structure, we used fastSTRUCTURE v1.0 (67) to
478 test for admixture among sampled individuals. We converted our vcf to PLINK bed format with
479 a minor allele frequency of 0.05 and maintained the order of alleles from the original vcf file.
480 We ran fastSTRUCTURE on all sampled individuals as well as only Minnesota and Isle Royale
481 individuals, each down-sampled to five unrelated individuals. For both analyses, we ran
482 fastSTRUCTURE using values of k from 1-4. Finally, we used vcftools (68) to estimate Weir and
483 Cockerham's (69) F_{ST} between all Minnesota and Isle Royale samples using default settings.

484

485 **Genetic diversity and runs of homozygosity**

486 We calculated heterozygosity for each individual in non-overlapping 1 Mb windows across the
487 autosomal genome. We removed windows with fewer than 80% of sites called, as well as
488 windows below the 5th percentile of the total number of calls, as these windows have high
489 variance in heterozygosity. We estimated mean genome-wide heterozygosity by averaging
490 heterozygosity across windows for each individual.

491

492 Runs of homozygosity were called using BCFtools/RoH (70). We used the -G30 flag and allowed
493 BCFtools to estimate allele frequencies. Due to the Swedish sample coming from a highly
494 divergent population with differing allele frequencies, we excluded it from this analysis. We
495 used a custom R script (71) to partition the resulting ROH calls into length categories 0.1-1 Mb,
496 1-10 Mb, and 10-100 Mb. We calculated F_{ROH} by summing the total length of all ROH calls >100
497 kb (or >1 Mb) and dividing by 2489.4 Mb, the autosomal genome length for the cattle reference
498 genome. When conducting this analysis for the subset of samples mapped to the hog deer
499 reference genome, we only used scaffolds >1 Mb in length, which together sum to 2479 Mb
500 (~93% of the total reference length).

501

502 **Identifying putatively deleterious variation**

503 Variant sites were annotated using the Ensembl Variant Effect Predictor (VEP) v.97 (64). We
504 used SIFT (37) to determine whether a nonsynonymous mutation is likely to be damaging or
505 benign based on phylogenetic constraint. We classified protein-coding variants as “damaging” if
506 they were determined to be “deleterious” nonsynonymous variants (SIFT score of <0.05) or
507 variants that disrupted splice sites, start codons, or stop codons. Variants were classified as
508 “benign” if they were determined to be “tolerated” nonsynonymous variants (SIFT score of
509 ≥ 0.05) or synonymous mutations. Using these annotations, we tallied the number of derived
510 alleles of each category relative to the cattle reference genome, as well as the number of
511 heterozygous and homozygous derived genotypes, comparing these tallies for genomes

512 sampled from Isle Royale and Minnesota. Variants that were that were fixed derived across the
513 entire sample were ignored.

514

515 **Demographic inference**

516 We estimated historical demographic parameters for North American moose based on the
517 neutral site frequency spectrum (SFS) using $\partial a\partial i$ (33). In brief, we first focused on estimating
518 parameters for the mainland North American population based on the neutral SFS for our nine
519 Minnesota genomes, then used these results to guide inference of the effective population size
520 on Isle Royale based on a neutral SFS from five genomes of unrelated Isle Royale individuals.

521

522 To generate a neutral SFS, we began by identifying regions that were >10kb from coding
523 regions and did not overlap with repetitive regions (downloaded from
524 ftp://ftp.ncbi.nlm.nih.gov/genomes/Bos_taurus/). We also excluded un-annotated highly
525 conserved regions that are under strong evolutionary constraint, identified by aligning the
526 remaining regions against the zebra fish genome using BLASTv2.7.1 (72) and removing any
527 region which had a hit above a 1e-10 threshold.

528

529 We then generated a folded neutral SFS for these regions using a modified version of EasySFS
530 (<https://github.com/isaacovercast/easySFS>), which implements $\partial a\partial i$'s hypergeometric
531 projection to account for missing genotypes. We found that the number of SNPs was
532 maximized by using a projection value of seven diploids for the Minnesota sample and four
533 diploids for the Isle Royale sample. In addition, we counted the number of monomorphic sites
534 passing the projection threshold in neutral regions and added these to the 0 bin of the SFS.

535

536 We then used these SFSs to conduct demographic inference using the diffusion approximation
537 approach implemented in $\partial a\partial i$ (33). Using the Minnesota SFS, we fit 1-epoch, 2-epoch, 3-
538 epoch, and 4-epoch models. These models included the following parameters: N_{anc} (the
539 ancestral effective population size), N_{1-3} (the effective size of the subsequent 1-3 epochs), and
540 T_{1-3} (the duration of the subsequent 1-3 epochs; Table S2). In other words, a 3-epoch model

541 includes the parameters N_{anc} , N_1 , N_2 , T_1 , and T_2 . Overall, we found the best fit for a 4-epoch
542 model including expansion in the second epoch followed by a strong bottleneck and a final
543 epoch of expansion, though with poor convergence of estimated parameters. Based on initial
544 results, we constrained parameter space for the 4-epoch model by setting a limit on N_1 to be in
545 the range $[10, 30] * N_{anc}$, N_2 to be in the range $[1e-2, 5] * N_{anc}$, and N_3 to be in the range $[10,$
546 $40] * N_{anc}$.

547
548 We next sought to obtain an estimate of the effective population size on Isle Royale using a
549 folded neutral SFS from five unrelated individuals, projected to four diploids. Given this limited
550 sample size and the shared evolutionary history of Isle Royale and Minnesota moose, we fixed
551 the parameters estimated from our 4-epoch model inferred above based on the Minnesota SFS.
552 We then added a fifth epoch to the model, fixing the duration of this epoch to 15 generations,
553 based on an estimated date of colonization of 1900 and 8 year generation time (35). Thus, the
554 only estimated parameter in this approach is N_5 , the effective population size on Isle Royale.

555
556 We carried out inference by permuting the starting parameter values and conducting 50 runs
557 for each model. We calculated the log-likelihood using *∂a∂i*'s optimized parameter values
558 comparing the expected and observed SFSs. For each model, we selected the maximum
559 likelihood estimate from the 50 runs and used AIC to compare across models. We then used a
560 mutation rate of $7e-9$ mutations/site/generation and the total sequence length (L) to calculate
561 the diploid ancestral effective population size as $N_{anc} = \Theta / (4 * \mu * L)$. We scaled other inferred
562 population size parameters by N_{anc} and time parameters by $2 * N_{anc}$, in order to obtain values in
563 units of diploids and numbers of generations.

564

565 **Simulations of deleterious genetic variation**

566 We performed forward-in-time genetic simulations using SLiM v3.6 (42). We simulated a 20 Mb
567 chromosomal segment with randomly generated introns, exons, and intergenic regions
568 following the approach from (73). Thus, our aim with these simulations is not to quantify
569 genome-wide effects of deleterious mutations, but rather to examine relative changes in

570 deleterious mutations within a 20 Mb chromosomal segment. Deleterious (nonsynonymous)
571 mutations occurred in exonic regions at a ratio of 2.31:1 to neutral (synonymous) mutations
572 (74), and only neutral mutations occurred in intronic and intergenic regions. Following (30), we
573 assumed a mutation rate of $7e-9$ mutations per site per generation. Selection coefficients (s) for
574 deleterious mutations were drawn from a distribution estimated using human genetic variation
575 data by (43), consisting of a gamma distribution with mean s of -0.01314833 and shape = 0.186 .
576 Additionally, we augmented this distribution such that 0.5% of deleterious mutations were
577 recessive lethal, given that this distribution may underestimate the fraction of lethal mutations
578 (12). The dominance coefficients (h) of our simulations were set to model an inverse
579 relationship between h and s , given that highly deleterious mutations also tend to be highly
580 recessive (75, 76). Specifically, we assumed $h=0.0$ for very strongly deleterious mutations ($s < -$
581 0.1), $h=0.01$ for strongly deleterious mutations ($-0.1 \leq s < -0.01$), $h=0.1$ for moderately
582 deleterious mutations ($-0.01 \leq s < -0.001$), and $h=0.4$ for weakly deleterious mutations ($s > -$
583 0.001). To test the sensitivity of our analysis to our assumed selection and dominance
584 parameters, we also ran simulations under the selection and dominance parameters proposed
585 by (12). Specifically, this model assumes that deleterious mutations come from a gamma
586 distribution with mean s of -0.05 and shape = 0.5 , augmented with an additional 5% of
587 deleterious mutations being lethal. Dominance coefficients follow the relationship $h = 0.5 * \exp(-$
588 $13*s)$; however, we simplified this to five dominance partitions for computational efficiency:
589 $h=0.48$ for $s \geq -0.01$, $h=0.31$ for $-0.1 \leq s < -0.01$, $h=0.07$ for $-0.4 \leq s < -0.1$, $h=0.001$ for $-1.0 \leq s$
590 < -0.4 , and $h=0.0$ for $s=-1.0$. For all simulations, we retained fixed mutations, such that their
591 impact on fitness was allowed to accumulate.

592
593 We set the population sizes of our simulations according to our best-fit 4-epoch demographic
594 model based on the SFS from our Minnesota moose genomes (Fig. 3; Table S2). Specifically, this
595 model estimated an ancestral effective population size of $N_{anc}=6,548$ diploids, followed by
596 expansion to $N_1=79,647$ for $T_1=22,628$ generations, then contraction to $N_2=49$ for $T_2=29$
597 generations, and finally expansion to $N_3=193,472$ for $T_3=1,179$ generations. We also ran
598 simulations under a second 4-epoch model that had similar log-likelihood and somewhat

599 differing parameters of $N_{anc}=7,017$, $N_1=145,662$, $T_1=20,883$, $N_2=218$, $T_2=142$, $N_3=105,531$ and
600 $T_3=1,223$. In both cases, we allowed the ancestral population to get to mutation-selection-drift
601 equilibrium by running a burn-in at N_{anc} for 70,000 generations.

602
603 Following the fourth epoch of both models, we added a fifth and final epoch representing the
604 founding of the Isle Royale population, consisting of $N_e=187$ for 15 generations. However, when
605 simulating under this demography, we observed that the simulated levels of inbreeding and
606 genetic diversity for the Isle Royale population did not recapitulate those observed in our
607 empirical data (Fig. 5). Specifically, we observed only a 3.6% reduction in heterozygosity
608 (compared to ~30% in our empirical data) and an increase in F_{ROH} to just 0.08 (compared to 0.35
609 in our empirical data). We hypothesized that this was due to the lack of a founder event at the
610 origination of the Isle Royale population in our model. To explore the impact of a founder
611 event, we modified the effective population sizes during the first three generations of the Isle
612 Royale population, using three plausible bottleneck parameters of $N_e=\{6,24,96\}$, $N_e=\{4,16,64\}$,
613 and $N_e=\{2,8,32\}$. We focused on the three initial generations after founding, reflecting the
614 period from ~1900-1924 when census estimates are crude and/or unavailable (22, 23).
615 Specifically, little is known about the number of founding individuals, though it is likely this
616 number was small, particularly if the population was naturally founded. Additionally, available
617 records indicate a population size of ~300 by 1920 and perhaps several thousand by 1930,
618 suggesting that population growth was rapid following founding (22, 23). Following this three-
619 generation bottleneck, we simulated the final 12 generations at our estimated $N_e=187$,
620 representing an average effective population size for the period ~1924-2020 when census
621 estimates ranged from ~500-2000 (average of ~1000; (19)).

622
623 During simulations, we recorded mean heterozygosity, mean F_{ROH} for ROH >100 kb and >1 Mb,
624 mean genetic load (calculated multiplicatively across sites), mean inbreeding load (measured as
625 the number of diploid lethal equivalents), and the mean number of strongly deleterious ($s < -$
626 0.01), moderately deleterious ($-0.01 \leq s < -0.001$), and weakly deleterious ($s > -0.001$) alleles per
627 individual. These quantities were estimated from a sample of 40 diploids every 1,000

628 generations during the burn-in, every 100 generations during the second epoch, every 5
629 generations during the North America founding bottleneck, every 20 generations during the
630 fourth epoch, and every generation during the Isle Royale bottleneck. For all simulated
631 scenarios, we ran 25 replicates.

632 **Acknowledgements**

633 We are grateful to members of the Wayne and Lohmueller labs for helpful input on this work.

634 We thank Michelle Carstensen and the Minnesota Department of Natural Resources for

635 providing tissue samples used in this study. C.C.K. and K.E.L. were supported by National

636 Institutes of Health grant R35GM119856 (to K.E.L.). A.C.B was supported by the Biological

637 Mechanisms of Healthy Aging Training Program NIH T32AG066574. This work was supported by

638 the National Science Foundation (DEB Small Grant #1556705).

639

640 **Data Availability**

641 All scripts are available at https://github.com/ckyrizis/moose_WGS_project and raw data will

642 be available on SRA upon publication.

643

644 **Author Contributions**

645 C.C.K., R.K.W., and K.E.L. conceived the study. K.E.B., J.A.V., L.M.V., S.R.H. and R.O.P. acquired

646 samples. C.C.K. conducted all analyses with input from A.C.B. and K.E.L. and wrote the

647 manuscript with input from all authors. R.K.W. and K.E.L. jointly supervised this work.

648

649 References

- 650 1. N. M. Haddad, *et al.*, Habitat fragmentation and its lasting impact on Earth's ecosystems.
651 *Sci. Adv.* **1**, 1–10 (2015).
- 652 2. P. W. Hedrick, A. Garcia-Dorado, Understanding Inbreeding Depression, Purging, and
653 Genetic Rescue. *Trends Ecol. Evol.* **31**, 940–952 (2016).
- 654 3. L. Keller, D. M. Waller, Inbreeding effects in wild populations. *Trends Ecol. Evol.* **17**, 19–23
655 (2002).
- 656 4. J. A. Robinson, C. Brown, B. Y. Kim, K. E. Lohmueller, R. K. Wayne, Purging of Strongly
657 Deleterious Mutations Explains Long-Term Persistence and Absence of Inbreeding
658 Depression in Island Foxes. *Curr. Biol.* **28**, 3487–3494.e4 (2018).
- 659 5. N. Pérez-Pereira, *et al.*, Long-term exhaustion of the inbreeding load in *Drosophila*
660 *melanogaster*. *Heredity (Edinb.)*, 1–11 (2021).
- 661 6. S. Glémin, How Are Deleterious Mutations Purged? Drift versus Nonrandom Mating.
662 *Evolution (N. Y.)*. **57**, 2678–2687 (2003).
- 663 7. C. Grossen, F. Guillaume, L. F. Keller, D. Croll, Purging of highly deleterious mutations
664 through severe bottlenecks in Alpine ibex. *Nat. Commun.* **11**, 1001 (2020).
- 665 8. Y. Xue, *et al.*, Mountain gorilla genomes reveal the impact of long-term population
666 decline and inbreeding. *Science (80-)*. **348**, 242–245 (2015).
- 667 9. C. C. Kyriazis, R. K. Wayne, K. E. Lohmueller, Strongly deleterious mutations are a primary
668 determinant of extinction risk due to inbreeding depression. *Evol. Lett.* **5**, 33–47 (2021).
- 669 10. S. B. Day, E. H. Bryant, L. M. Meffert, The influence of variable rates of inbreeding on
670 fitness, environmental responsiveness, and evolutionary potential. *Evolution (N. Y.)*. **57**,
671 1314–1324 (2003).
- 672 11. N. Pekkala, K. E. Knott, J. S. Kotiaho, M. Puurtinen, Inbreeding rate modifies the dynamics
673 of genetic load in small populations. *Ecol. Evol.* **2**, 1791–1804 (2012).
- 674 12. M. Kardos, *et al.*, The crucial role of genome-wide genetic variation in conservation. *Proc.*
675 *Natl. Acad. Sci. U. S. A.* **118**, 1–10 (2021).
- 676 13. J. C. Teixeira, C. D. Huber, The inflated significance of neutral genetic diversity in
677 conservation genetics. *Proc. Natl. Acad. Sci.* **118**, 1–10 (2021).
- 678 14. K. Ralls, P. Sunnucks, R. C. Lacy, R. Frankham, Genetic rescue: A critique of the evidence
679 supports maximizing genetic diversity rather than minimizing the introduction of
680 putatively harmful genetic variation. *Biol. Conserv.* **251**, 108784 (2020).
- 681 15. A. Khan, *et al.*, Genomic evidence for inbreeding depression and purging of deleterious
682 genetic variation in Indian tigers. *Proc. Natl. Acad. Sci. U. S. A.* **118** (2021).
- 683 16. D. Kleinman-Ruiz, *et al.*, Purging of deleterious burden in the endangered Iberian lynx.
684 *Proc. Natl. Acad. Sci.* **119** (2022).
- 685 17. N. Pérez-Pereira, A. Caballero, A. García-Dorado, Reviewing the consequences of genetic
686 purging on the success of rescue programs. *Conserv. Genet.* **23**, 1–17 (2022).
- 687 18. Y. Willi, *et al.*, Conservation genetics as a management tool: The five best-supported
688 paradigms to assist the management of threatened species. *Proc. Natl. Acad. Sci. U. S. A.*
689 **119**, 1–10 (2022).
- 690 19. S. R. Hoy, R. O. Peterson, J. A. Vucetich, “Ecological Studies of Wolves on Isle Royale:
691 Annual Report 2019-2020” (2020).

- 692 20. J. A. Robinson, *et al.*, Genomic signatures of extensive inbreeding in Isle Royale wolves, a
693 population on the threshold of extinction. *Sci. Adv.* **5**, 1–13 (2019).
- 694 21. P. W. Hedrick, J. A. Robinson, R. O. Peterson, J. A. Vucetich, Genetics and extinction and
695 the example of Isle Royale wolves. *Anim. Conserv.* **22**, 302–309 (2019).
- 696 22. L. D. Mech, “The Wolves of Isle Royale” (1966).
- 697 23. A. Murie, Moose of Isle Royale. *Univ. Michigan Museum Zool. Misc. Publ. No. 25* (1934).
- 698 24. R. L. Sattler, J. R. Willoughby, B. J. Swanson, Decline of heterozygosity in a large but
699 isolated population: a 45-year examination of moose genetic diversity on Isle Royale.
700 *PeerJ* **5**, 1–18 (2017).
- 701 25. P. J. Wilson, *et al.*, Genetic variation and population structure of moose (*Alces alces*) at
702 neutral and functional DNA loci. *Can. J. Zool.* **683**, 670–683 (2003).
- 703 26. S. R. Hoy, *et al.*, Fluctuations in age structure and their variable influence on population
704 growth. *Funct. Ecol.* **34**, 203–216 (2020).
- 705 27. K. J. Hundertmark, *et al.*, Mitochondrial Phylogeography of Moose (*Alces alces*): Late
706 Pleistocene Divergence and Population Expansion. **22**, 375–387 (2002).
- 707 28. K. J. Hundertmark, R. T. Bowyer, G. F. Shields, C. C. Schwartz, Mitochondrial
708 Phylogeography of Moose (*Alces alces*) in North America. *J. Mammal.* **84**, 718–728
709 (2003).
- 710 29. N. J. Decesare, *et al.*, Phylogeography of moose in western North America. *J. Mammal.*
711 **101**, 10–23 (2020).
- 712 30. N. Dussex, Moose genomes reveal past glacial demography and the origin of modern
713 lineages. *BMC Genomics* **21**, 1–13 (2020).
- 714 31. H. R. Timmermann, A. R. Rodgers, The Status and Management of Moose in North
715 America - Circa 2015. *Alces A J. Devoted to Biol. Manag. Moose* **53**, 1–22 (2017).
- 716 32. M. Kirin, *et al.*, Genomic runs of homozygosity record population history and
717 consanguinity. *PLoS One* **5**, 1–7 (2010).
- 718 33. R. N. Gutenkunst, R. D. Hernandez, S. H. Williamson, C. D. Bustamante, Inferring the joint
719 demographic history of multiple populations from multidimensional SNP frequency data.
720 *PLoS Genet.* **5**, 1–11 (2009).
- 721 34. A. C. Beichman, *et al.*, Genomic analyses reveal range-wide devastation of sea otter
722 populations. *Mol. Ecol.*, 1–18 (2022).
- 723 35. J.-M. Gaillard, Are Moose Only a Large Deer?: Some Life History Considerations. *Alces* **43**,
724 1–12 (2007).
- 725 36. R. Frankham, Effective population size/adult population size ratios in wildlife: A review.
726 *Genet. Res. Cambridge* **66**, 95–107 (1995).
- 727 37. R. Vaser, S. Adusumalli, S. N. Leng, M. Sikic, P. C. Ng, SIFT missense predictions for
728 genomes. *Nat. Protoc.* **11**, 1–9 (2016).
- 729 38. K. E. Lohmueller, *et al.*, Proportionally more deleterious genetic variation in European
730 than in African populations. *Nature* **451**, 994–997 (2008).
- 731 39. R. Do, *et al.*, No evidence that selection has been less effective at removing deleterious
732 mutations in Europeans than in Africans. *Nat. Genet.* **47**, 126–131 (2015).
- 733 40. G. M. Cooper, J. Shendure, Needles in stacks of needles: Finding disease-causal variants
734 in a wealth of genomic data. *Nat. Rev. Genet.* **12**, 628–640 (2011).
- 735 41. C. E. T. Pedersen, *et al.*, The effect of an extreme and prolonged population bottleneck

- 736 on patterns of deleterious variation: Insights from the Greenlandic Inuit. *Genetics* **205**,
737 787–801 (2017).
- 738 42. B. C. Haller, P. W. Messer, SLiM 3: Forward Genetic Simulations Beyond the Wright-Fisher
739 Model. *Mol. Biol. Evol.* **36**, 632–637 (2019).
- 740 43. B. Y. Kim, C. D. Huber, K. E. Lohmueller, Inference of the Distribution of Selection
741 Coefficients for New Nonsynonymous Mutations Using Large Samples. *Genetics* **206**,
742 345–361 (2017).
- 743 44. M. Kirkpatrick, P. Jarne, The Effects of a Bottleneck on Inbreeding Depression and the
744 Genetic Load. *Am. Nat.* **155**, 154–167 (2000).
- 745 45. J. A. Vucetich, *Restoring the Balance: What Wolves Tell Us about Our Relationship with*
746 *Nature* (Johns Hopkins Press, 2021).
- 747 46. P. W. Hedrick, Purging inbreeding depression and the probability of extinction: full-sib
748 mating. *Heredity (Edinb.)* **73**, 363–372 (1994).
- 749 47. M. A. Stoffel, S. E. Johnston, J. G. Pilkington, J. M. Pemberton, Mutation load decreases
750 with haplotype age in wild Soay sheep. *Evol. Lett.* **5**, 187–195 (2021).
- 751 48. Z. A. Szpiech, *et al.*, Long runs of homozygosity are enriched for deleterious variation.
752 *Am. J. Hum. Genet.* **93**, 90–102 (2013).
- 753 49. Z. A. Szpiech, *et al.*, Ancestry-Dependent Enrichment of Deleterious Homozygotes in Runs
754 of Homozygosity. *Am. J. Hum. Genet.* **105**, 747–762 (2019).
- 755 50. A. F. Agrawal, M. C. Whitlock, Mutation Load: The Fitness of Individuals in Populations
756 Where Deleterious Alleles Are Abundant. *Annu. Rev. Ecol. Evol. Syst.* **43**, 115–135 (2012).
- 757 51. Y. Yang, *et al.*, Genomic effects of population collapse in a critically endangered ironwood
758 tree *Ostrya rehderiana*. *Nat. Commun.* **9**, 5449 (2018).
- 759 52. M. A. Stoffel, S. E. Johnston, J. G. Pilkington, J. M. Pemberton, Genetic architecture and
760 lifetime dynamics of inbreeding depression in a wild mammal. *Nat. Commun.* **12**, 1–10
761 (2021).
- 762 53. H. G. Broders, S. P. Mahoney, W. A. Montevicchi, W. S. Davidson, Population genetic
763 structure and the effect of founder events on the genetic variability of moose , *Alces*
764 *alces* , in Canada. *Mol. Ecol.* **8**, 1309–1315 (1999).
- 765 54. D. L. Murray, *et al.*, Pathogens, Nutritional Deficiency, and Climate Influences on a
766 Declining Moose Population. *Wildl. Monogr.* **166**, 1–30 (2006).
- 767 55. M. L. Wolfe, K. R. Hersey, D. C. Stoner, A History of Moose Management in Utah. *Alces A*
768 *J. Devoted to Biol. Manag. Moose* **46**, 37-52–52 (2010).
- 769 56. J. S. Ruprecht, *et al.*, Reproduction in moose at their southern range limit. *J. Mammal.*
770 **97**, 1355–1365 (2016).
- 771 57. A. K. Gibson, A. E. Nguyen, Does genetic diversity protect host populations from
772 parasites? A meta-analysis across natural and agricultural systems. *Evol. Lett.* **5**, 16–32
773 (2021).
- 774 58. B. E. McLaren, R. O. Peterson, Wolves, Moose, and Tree Rings on Isle Royale. *Science (80-*
775 *.)* **266**, 1555–1558 (1994).
- 776 59. W. J. Ripple, *et al.*, Status and ecological effects of the world’s largest carnivores. *Science*
777 *(80-.)* **343** (2014).
- 778 60. J. A. Estes, *et al.*, Trophic downgrading of planet earth. *Science (80-.)* **333**, 301–306
779 (2011).

- 780 61. T. S. Kalbfleisch, *et al.*, A SNP resource for studying North American moose.
781 *F1000Research* **7**, 1–17 (2018).
- 782 62. G. A. Van der Auwera, *et al.*, *From fastQ data to high-confidence variant calls: The*
783 *genome analysis toolkit best practices pipeline* (2013)
784 <https://doi.org/10.1002/0471250953.bi1110s43>.
- 785 63. H. Li, Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM.
786 *arXiv:1303.3997v2* **00**, 1–3 (2013).
- 787 64. W. McLaren, *et al.*, The Ensembl Variant Effect Predictor. *Genome Biol.* **17**, 1–14 (2016).
- 788 65. X. Zheng, *et al.*, A High-performance Computing Toolset for Relatedness and Principal
789 Component Analysis of SNP Data. *Bioinformatics* **28**, 3326–3328 (2012).
- 790 66. A. Manichaikul, *et al.*, Robust relationship inference in genome-wide association studies.
791 *Bioinformatics* **26**, 2867–2873 (2010).
- 792 67. A. Raj, M. Stephens, J. K. Pritchard, FastSTRUCTURE: Variational inference of population
793 structure in large SNP data sets. *Genetics* **197**, 573–589 (2014).
- 794 68. P. Danecek, *et al.*, The variant call format and VCFtools. *Bioinformatics* **27**, 2156–2158
795 (2011).
- 796 69. B. S. Weir, C. C. Cockerham, Estimating F-statistics for the analysis of population
797 structure. *Evolution (N. Y.)*, 1358–1370 (1984).
- 798 70. V. Narasimhan, *et al.*, BCFtools/RoH: A hidden Markov model approach for detecting
799 autozygosity from next-generation sequencing data. *Bioinformatics* **32**, 1749–1751
800 (2016).
- 801 71. R Core Team, R: A language and environment for statistical computing. (2021).
- 802 72. C. Camacho, *et al.*, BLAST+: architecture and applications. [BMC Bioinformatics. 2009] -
803 PubMed - NCBI. *BMC Bioinformatics* **10**, 421 (2009).
- 804 73. J. A. Mooney, *et al.*, Understanding the Hidden Complexity of Latin American Population
805 Isolates. *Am. J. Hum. Genet.*, 1–53 (2018).
- 806 74. C. D. Huber, B. Y. Kim, C. D. Marsden, K. E. Lohmueller, Determining the factors driving
807 selective effects of new nonsynonymous mutations. *Proc. Natl. Acad. Sci.* **114**, 4465–
808 4470 (2017).
- 809 75. C. D. Huber, A. Durvasula, A. M. Hancock, Gene expression drives the evolution of
810 dominance. *Nat. Commun.* **9**, 1–11 (2018).
- 811 76. A. F. Agrawal, M. C. Whitlock, Inferences about the distribution of dominance drawn
812 from yeast gene knockout data. *Genetics* **187**, 553–566 (2011).
- 813