Mitigating pseudoreplication and bias in resource selection functions with autocorrelation-informed weighting

Jesse M. Alston[1,2*†], Christen H. Fleming[3,4†], Roland Kays[5,6], Jarryd P. Streicher[7], Colleen T. Downs[7], Tharmalingam Ramesh[7,8], and Justin M. Calabrese[1,2,9]

[1]Center for Advanced Systems Understanding, Görlitz, Germany

[2]Helmholtz-Zentrum Dresden Rossendorf (HZDR), Dresden, Germany

[3]Smithsonian Conservation Biology Institute, National Zoological Park, Front Royal, VA USA

[4]Department of Biology, University of Maryland, College Park, MD USA

[5]Department of Forestry and Environmental Resources, North Carolina State University, Raleigh, NC USA

[6]North Carolina Museum of Natural Sciences, Raleigh, NC USA

[7]Centre for Functional Biodiversity, School of Life Sciences, University of KwaZulu-Natal, Pietermaritzburg, South Africa

[8]Sálim Ali Centre for Ornithology and Natural History (SACON), Coimbatore, Tamil Nadu, India

[9]Department of Ecological Modelling, Helmholtz Centre for Environmental Research (UFZ), Leipzig, Germany

[*]Corresponding author: j.alston@hzdr.de

[†]Equal contributions

₁ **Abstract**

₂ Resource selection functions are among the most commonly used statistical tools in both basic

₃ and applied animal ecology. They are typically parameterized using animal tracking data, and

₄ advances in animal tracking technology have led to increasing levels of autocorrelation between

₅ locations in such data sets. Because resource selection functions assume that data are indepen-

₆ dent and identically distributed, such autocorrelation can cause misleadingly narrow confidence

₇ intervals and biased parameter estimates. Data thinning, generalized estimating equations, and

₈ step selection functions have been suggested as techniques for mitigating the statistical problems

₉ posed by autocorrelation, but these approaches have notable limitations that include statistical in-

₁₀ efficiency, unclear or arbitrary targets for adequate levels of statistical independence, constraints

₁₁ in input data, and (in the case of step selection functions) scale-dependent inference. To remedy

₁₂ these problems, we introduce a method for likelihood weighting of animal locations to mitigate the

₁₃ negative consequences of autocorrelation on resource selection functions. This method weights

₁₄ each observed location in an animal's movement track according to its level of autocorrelation,

₁₅ expanding confidence intervals to match an objective target (i.e., the effective sample size for Au-

₁₆ tocorrelated Gaussian Density Estimation) and accounting for bias that can arise when there are

₁₇ gaps in the movement track. In this study, we describe the mathematical principles underlying our

₁₈ method, demonstrate its practical advantages versus conventional approaches using simulations

₁₉ and empirical data on a water mongoose (*Atilax paludinosus*), a caracal (*Caracal caracal*), and a

₂₀ serval (*Leptailurus serval*), and discuss pathways for further development of our method. We also

₂₁ provide a complete, annotated analytical workflow to help new users apply our method to their

₂₂ own animal tracking data using the `ctmm` R package.

₂₃ **Key-words**: continuous-time movement models, habitat selection, home range, Ornstein-Uhlenbeck

₂₄ process, space use, spatial point process, utilization distribution

Introduction

Resource selection functions (RSFs) have been a mainstay of basic and applied ecology for decades (Boyce & McDonald, 1999, Manly *et al.*, 2007), and are commonly used to answer questions such as "What landscape features do animals seek or avoid during their movements?" (e.g., Lamont *et al.*, 2019, Prokopenko *et al.*, 2017b, Valls-Fox *et al.*, 2018) and "In which areas of the landscape are animals at risk of predation or disease transmission?" (e.g., Ali *et al.*, 2017, Ng'weno *et al.*, 2019, Rayl *et al.*, 2019). They are typically fit in a use–availability framework, whereby environmental covariates at the locations where animals were known to be present (i.e. "used" locations) are compared with covariates at locations taken from an area assumed to be available for selection (i.e., "available" locations; Manly *et al.* 2007), and are usually formulated using logistic regression. This approach allows researchers to easily estimate selection or avoidance of covariates and to generate maps or data layers for use in downstream analyses (Northrup *et al.*, 2013, 2021).

The relative ease of fitting RSFs has contributed to their popularity in animal ecology. However, the results of RSFs are influenced by a number of methodological challenges (Aarts *et al.*, 2008, Northrup *et al.*, 2013, 2021). One major (and worsening) challenge is the degree of temporal autocorrelation in tracking data, which has increased considerably as advances in tracking devices facilitate the recording of animal locations at ever-smaller intervals (Fieberg *et al.*, 2010). Because RSFs assume independent and identically distributed (IID) data, autocorrelation can lead to incorrectly narrow confidence intervals, and therefore random variations being incorrectly identified as significant effects. This may (at least in part) explain why a growing number of studies have documented surprising levels of apparent variation in habitat selection within species and even populations (e.g., Leclerc *et al.*, 2016, Montgomery *et al.*, 2018, Newediuk *et al.*, 2022). While individual animals undoubtedly vary somewhat in their habitat preferences, this pattern of extreme variation in selection parameters can easily arise when pseudoreplication from autocorrelated data causes unimportant covariates to be (misleadingly) found to be significant. Accurate quantification of uncertainty is therefore paramount for accurate inference concerning habitat selection. In addition, RSFs that assume IID data are unable to account for varying levels of autocorrelation in

movement data. This might happen when environmental covariates inhibit successful estimation of animal locations (e.g., dense vegetation blocking Global Positioning System [GPS] satellite reception), leaving unsampled gaps in an animal movement track (Fleming *et al.*, 2018, Frair *et al.*, 2010, Lewis *et al.*, 2007). When the gaps in data are correlated with specific covariates of interest, the level of autocorrelation in animal locations is also correlated with those covariates. Despite widespread awareness of these problems, no generally accepted solutions have been described, and researchers often fit IID RSF models to autocorrelated tracking data.

Historically, data thinning has been the most common suggestion for reducing autocorrelation in tracking data used to inform RSFs (e.g., Hooten *et al.*, 2014, Northrup *et al.*, 2013, Swihart & Slade, 1985). While pragmatic and easy to implement, data thinning suffers from a number of limitations as a method of mitigating autocorrelation. First, thinning inherently requires discarding data, which results in the loss of useful information and can lead to imprecise parameter estimates. Second, thinning typically involves using statistical tests for the presence of statistically significant levels of autocorrelation—a target which is both arbitrary and not optimized for the task of estimating distributions (Aarts *et al.*, 2008). Under-thinning results in misleadingly narrow confidence intervals and possibly biased coefficient estimates, while over-thinning results in an unnecessary loss of information and statistical efficiency. Third, highly irregular time-series, such as those obtained from tracking aquatic animals (Breed *et al.*, 2011, Fleming *et al.*, 2018) or sampling designs targeted at identifying behaviors at multiple temporal scales (e.g., Scantlebury *et al.*, 2014, Ullmann *et al.*, 2020), may not be particularly amenable to thinning.

Increasingly, researchers use step-selection functions (hereafter SSFs; Avgar *et al.*, 2016, Fortin *et al.*, 2005, Thurfjell *et al.*, 2014) to account for autocorrelation in animal movements during studies of resource selection (e.g., Alston *et al.*, 2020, Dickie *et al.*, 2020, Kohl *et al.*, 2018, Merkle *et al.*, 2016, Prokopenko *et al.*, 2017a). This approach builds upon conventional RSFs by generating available points (or movement steps) using a biologically realistic, movement-informed sampling scheme. However, SSFs still have important limitations. First, because available points in SSFs depend on distributions of step lengths and turning angles on a fixed sampling interval, SSFs

cannot handle irregularly sampled data—SSFs can only be parameterized using animal location data that were sampled at a constant sampling interval. Second, parameters of SSFs are explicitly scale-dependent—parameters change as the sampling schedule of a movement track changes (Avgar *et al.*, 2016, Fieberg *et al.*, 2021, Signer *et al.*, 2017). In other words, if the same movement track were to be sampled at different intervals, the SSFs parameterized using the different samples would provide different estimates of selection. Third, SSFs do not invariably mitigate the pseudoreplication that arises from treating autocorrelated data as independent and identically distributed—step selection functions assume that steps are independent, so when the sampling interval is smaller than the length of time required for steps to be statistically independent of one another, SSFs will also produce misleadingly narrow confidence intervals. Additional measures such as data thinning (Babin *et al.*, 2011, Robb *et al.*, 2022), variance inflation (Nielsen *et al.*, 2002), and generalized estimating equations (Craiu *et al.*, 2008, Prima *et al.*, 2017) have been employed to widen confidence intervals of SSFs, but these methods lack an objective and easily implementable target for the appropriate width of confidence intervals.

Likelihood weighting offers another potential solution for mitigating the adverse effects of temporal autocorrelation on RSFs. In this framework, the contributions of individual animal locations are weighted according to a joint likelihood function, such that the sum of all weights matches an "effective sample size" derived from stochastic process models that describe the autocorrelation structure of an animal movement track. This framework avoids two problems caused by data thinning. First, instead of considering one thinned subsample of data and its set of resulting parameter estimates, or several thinned subsamples of the data and an average over their parameter estimates (which would reduce noise—and potentially bias—arising from thinning), all subsamples of independent data and an average of their log-likelihoods are evaluated during parameter estimation. Second, instead of thinning the data to an ad hoc threshold (e.g., less than 5% autocorrelation), data are weighted using an objective estimate of "effective sample size" that quantifies confidence in a home range estimate. As an additional benefit, likelihood weights can also account for sampling biases, such as highly irregular sampling. Such bias can occur, for example, when covariates

are over- or under-sampled, such as when environmental covariates are associated with differential probability of successful triangulation of animal locations (Fleming *et al.*, 2018, Frair *et al.*, 2010, Lewis *et al.*, 2007).

In this study, we introduce a method for autocorrelation-informed weighting of animal locations to mitigate pseudoreplication and bias in parameter estimates of RSFs. We provide a description of the mathematical principles underlying our method, demonstrate its practical advantages over conventional approaches using simulations and empirical animal tracking data for a water mongoose (*Atilax paludinosus*), a caracal (*Caracal caracal*), and a serval (*Leptailurus serval*), and discuss pathways for continuing to improve upon our method. We also provide an annotated analytical workflow for using the `ctmm` R package (Calabrese *et al.*, 2016) to apply our method to animal tracking data (Appendix 1).

Mathematical Concepts and Definitions

Conventional RSFs assume that data are sampled independently, which means that they do not allow for any autocorrelation in the data. Autocorrelated data are less informative than independent data when estimating coarse-scale parameters, because they share information with adjacent data points. Statistical estimates are therefore overconfident and often biased when autocorrelated data are modeled as independent. This is one form of pseudoreplication, which has long been acknowledged as a problem in ecology (Hurlbert, 1984). Animal tracking data are almost always autocorrelated (Noonan *et al.*, 2019), which poses a major problem for RSFs.

Here, we introduce an alternative method of correction—autocorrelation-informed likelihood weighting—that has several advantages over previous methods. First, all of the data are used; autocorrelated observations are down-weighted rather than discarded. Second, the amount of down-weighting applied is not ad hoc, but is chosen to obtain an "effective sample size" consistent with the estimate of the autocorrelated Gaussian area (the importance of which is explained in further detail below). This effective sample size is an objective estimate of the information content of a

set of autocorrelated data, expressed as the number of independent observations that would contain an equivalent amount of information. Third, and finally, we employ temporal weights that are optimized for non-parametric range distribution estimation (Fleming *et al.*, 2018), which give more weight to locations that are more independent on an animal movement track. For example, animal locations within rapid bursts of sampling in which the animal has not had time to move far are down-weighted more than samples with longer lags. This may occur when a landscape feature interferes with GPS triangulation, or when researchers employ duty cycling to capture some fine-scale movement behavior of interest while also attempting to preserve battery life. This means that irregular sampling of non-independent data (which is incompatible with the IID assumption underlying conventional RSFs) is explicitly accounted for in the model.

In a conventional RSF, the log-likelihood of n IID samples of an inhomogeneous Poisson point process model is given by

$$\ell(\text{data}|\lambda) = \sum_i^n \log \lambda(x(t_i), y(t_i)) - n \log \iint \lambda(X, Y) \, dX \, dY \, , \tag{1}$$

where $\lambda(x, y)$ is the intensity function, which contains all model parameters, and where, for simplicity, we have assumed that λ does not change in time. This likelihood function is typically approximated numerically via weighted logistic regression, and λ is usually constructed to be an exponential model (Fieberg *et al.*, 2021, Northrup *et al.*, 2021).

Within λ, we include area terms (x, y, and $x^2 + y^2$) that produce a bivariate Gaussian distribution of animal locations, so that when no resource selection occurs, the weighted likelihood reproduces a Gaussian home range estimate that approximates the maximum likelihood estimate of the autocorrelation model. This Gaussian distribution becomes the null RSF model (i.e., the domain of availability, and the area of movement when there is no habitat selection; Fieberg *et al.* 2021, Horne *et al.* 2008). The effective sample size is roughly the number of times an animal has crossed the linear extent of its home range, which dictates how well the area and shape of the Gaussian home range estimate can be resolved from the sampled movement path of an animal (Fleming

155 *et al.*, 2019). When selection parameters are supported, then we obtain a synoptic model of habitat

156 selection (*sensu* Horne *et al.*, 2008) within a Gaussian area. Further details of this configuration of

157 the RSF model are discussed in Fleming *et al* (in prep).

158 To the conventional IID log-likelihood, we incorporate our RSF log-likelihood weights, which

159 are given by

$$w(t_i) = w_{\mathrm{AKDE}}(t_i) \, N \,, \tag{2}$$

where $w_{\mathrm{AKDE}}(t_i)$ denotes the weights optimized for non-parametric autocorrelated kernel density

estimation at each sampled time t_i (Fleming *et al.*, 2018), which sum to 1, and where $N \leq n$ is

the effective sample size of the autocorrelated Gaussian area estimate, so that our weights, $w(t_i)$,

sum to N. The AKDE weights minimize error in non-parametric kernel density estimation, and

serve a similar purpose in the parametric Gaussian density estimation performed by the inclusion

of area terms in the RSF model. Weights (2) also provide identical estimates of the mean location

in both kernel and Gaussian density estimation, indicating that they provide a bias correction for

the Gaussian model. The weights $w(t_i)$ are used to weight each sampled location so that the final

log-likelihood is of the form

$$\ell(\mathrm{data}|\lambda) = \sum_i^n w(t_i) \log \lambda(x(t_i), y(t_i)) - N \log \iint \lambda(X, Y) \, dX \, dY \,. \tag{3}$$

Methods

Simulations

To validate our theoretical argument and demonstrate the value of our approach on animal location

data, we performed two sets of simulations. For both sets of simulations, we started by simulating

movement paths for an animal following an isotropic Ornstein-Uhlenbeck Foraging movement

model (Fleming *et al.*, 2014) with a location variance (σ) of 200,000 m^2, velocity persistence

timescale (τ_v) of 0.25 days, and home range crossing timescale (τ_p) of 1 day. This is equivalent to

an animal with a circular home range of 3.76 km^2, with four correlated bouts of movement each

day, and which crosses its home range roughly once per day. Because range crossings occurred

daily, the effective sample size in this movement track is equivalent to the number of days the

movement track is sampled, and sampling locations from the movement track more frequently than

once per day yields autocorrelated data. We overlaid each movement track on a raster consisting

of equal amounts of two habitat types (hereafter, Habitat 1 and Habitat 2) that each take up half of

the raster surface. All simulations were performed using the `ctmm` R package (v0.6.2; Calabrese

et al., 2016) in the R statistical software environment (v3.6.2; R Core Team, 2020). Habitat rasters

were created using the `raster` R package (v3.4-10; Hijmans, 2021).

Scenario 1: Triangulation Failure

We first simulated a scenario in which bias could arise when certain habitat covariates of in-

terest cause disproportionate amounts of triangulation failure in GPS devices. This is known to

happen, for example, in rugged terrain, thick vegetation, and underwater (e.g., Breed *et al.*, 2011,

O'Neill *et al.*, 2020, Streicher *et al.*, 2021). For this scenario, we simulated no habitat selection

occurring (i.e., the animal movement path was simulated independently of the habitat raster), but

we censored every other location in Habitat 2, which would be consistent with a habitat-specific

triangulation success rate of 50% (which simulates an extreme case of habitat-specific triangulation

failure). Because spatial autocorrelation in habitat affects estimates of resource selection (Northrup

et al., 2013), we configured our habitat rasters in two different ways to illustrate how weighting re-

sponds to extremely high and low habitat autocorrelation: (1) a high-habitat-autocorrelation case,

in which the two habitats were arrayed in two large contiguous blocks, and (2) a low-habitat-

autocorrelation case where the habitats were arrayed in alternating vertical stripes that were 10 m

in width. We sampled these tracks 1, 2, 4, 8, and 16 times per day, for 90 days, on 400 movement

tracks in each habitat configuration. We finally used weighted and IID RSFs to estimate strength of

selection for Habitat 1 compared with Habitat 2, and the confidence intervals around this parameter

estimate.

This scenario demonstrates several useful properties of our weighting scheme. First, by simulating habitat-specific location failure, we demonstrate that this weighting scheme can reduce sampling-induced bias in RSF parameter estimates compared with RSFs that assume data are IID. An IID RSF should detect a pattern of selection of Habitat 1 over Habitat 2 that is a statistical artifact of triangulation failure; a weighted RSF will shift the parameter estimate toward zero and expand the confidence intervals around the parameter estimate, correctly inferring that no habitat selection is occurring when effective sample sizes are large enough to reliably estimate parameters. Second, by varying the sampling rate, we can demonstrate that an RSF that treats data as IID becomes increasingly confident in its parameter estimates as data become more autocorrelated—even when they are biased—while weighted RSFs maintain stable confidence intervals as locations are sampled more frequently because the effective sample size of the data set is not increasing. In other words, increasing the rate of animal locations should cause the confidence intervals estimated using IID RSFs to become narrower (because increasing autocorrelation in the data causes pseudoreplication) but have no effect on weighted RSF confidence intervals (because autocorrelation is explicitly modeled and accounted for). Third, we can demonstrate that the amount of spatial autocorrelation in habitat covariates—which is known to influence RSF parameter estimates (Northrup *et al.*, 2013, Street *et al.*, 2021)—also influences differences between weighted RSF and IID RSF parameter estimates. In this case, as more consecutive locations occur within a single habitat type, the average weights of locations in the habitat type with frequent triangulation failure become larger, allowing the model to reduce the influence of the habitat-specific sampling intervals on parameter estimates.

Scenario 2: Absolute vs. Effective Sample Size

We then simulated a scenario that demonstrates that our weighting scheme does not reduce bias simply by inflating confidence intervals such that it becomes impossible to identify habitat selection when it occurs. For this scenario, we simulated habitat selection occurring by arraying habitat in 10 m vertical stripes (as in the low-habitat-autocorrelation scenario described above) and then simulated habitat-independent movement tracks over this layer. We sampled these tracks

while varying (1) the sampling rate (0.5, 2, 8, 32, 128, and 512 times per day for 100 days) or (2) the sampling duration (100 times per day for 16, 32, 64, 128, 256, and 512 days). We then shifted all locations that were in Habitat 1 in the right half of the movement track further right by 10 m, which moved those locations into Habitat 2. This alteration changed the fit of the underlying movement model very little, but led to the animals being located in Habitat 2 roughly three times as often as they were located in Habitat 1, emulating habitat selection. Finally, we used weighted RSFs to estimate strength of selection for Habitat 2 compared with Habitat 1, and the confidence intervals around this parameter estimate. In this scenario, parameter estimates should be constant around the true parameter in all scenarios, but confidence intervals should only meaningfully contract when the effective sample size (i.e., sampling duration) increases, and not just when the temporal resolution of the data increases.

Empirical Examples

To demonstrate the application of our method on real-world animal location data, we performed IID and weighted RSFs on tracking data from a water mongoose (144 locations over 58 days; Fig. S1), a caracal (504 locations over 85 days; Fig. S2), and a serval (3603 locations over 321 days; Fig. S3). All animals were tracked using GPS telemetry; further details on the specifics of animal capture and tracking can be found in Streicher *et al.* (2021), Ramesh *et al.* (2016a), and Ramesh *et al.* (2016b). All three animals are missing a meaningful number of locations because of GPS triangulation failure. For each animal, we estimated the effects of two land cover types (built-up [human settlements, roads, railways, and airfields] and plantation [exotic tree plantations]) on their habitat selection compared with a grouped reference category consisting of the remaining land cover types. For our land cover data, we used a land-use map with 20-m resolution from Ezemvelo KZN Wildlife (EKZN Wildlife & GeoTerraImage, 2018) reclassified into the three categories of interest (i.e., plantation, built-up, and reference).

For each animal, we used variogram analysis (Fleming *et al.*, 2014) to ensure animals were range-resident, fit and selected an autocorrelated movement model that best described the ani-

mal's movements using perturbative Hybrid Residual Maximum Likelihood (phREML; Fleming *et al.*, 2019) and Akaike's Information Criterion corrected for small sample sizes (AICc), estimated utilization distributions for each animal (wAKDE for weighted RSFs [Fleming *et al.* 2018]; conventional KDE for IID RSFs [Worton 1989]), and fit RSFs using the `ctmm` R package (v0.6.2; Calabrese *et al.*, 2016) in the R statistical software environment (v3.6.2; R Core Team, 2020).

Results

Simulations

Scenario 1: Triangulation Failure

In our simulations of triangulation failure, we found that weighted and IID RSFs performed as expected (Fig. 1). Across 400 individual movement paths, parameter estimates of weighted RSFs were closer to zero than parameter estimates of IID RSFs (Fig. 1). The mean confidence interval around parameter estimates of weighted RSFs contained zero at all sampling frequencies (Fig. 1A,C), while the mean confidence interval around parameter estimates of IID RSFs did not contain zero at any sampling rate above 1 (Figs. 1B,1D). Confidence intervals around parameter estimates of IID RSFs became narrower as the sampling rate (and thus autocorrelation) increased, while the width of confidence intervals around parameter estimates of weighted RSFs stayed consistent across all sampling frequencies (Fig. 1). Notably, weighted RSFs were better able to successfully identify a lack of habitat selection (i.e., weighted RSF parameter estimates became increasingly closer to zero and a higher proportion of confidence intervals contained zero) as the sampling rate increased, while IID RSFs performed worse as the sampling rate increased (i.e., parameter estimates were largely consistent, but confidence intervals around the [biased] estimates contracted sharply as the sampling rate increased).

Scenario 2: Absolute vs. Effective Sample Size

In our simulations of habitat selection, we found that weighted RSFs performed as expected,

Figure 1: *Mean parameter estimates and confidence intervals for weighted RSFs vs. IID RSFs for 400 simulations of habitat-specific GPS triangulation failure. Row 1 (Panels A and B) shows the performance of RSFs in a high-habitat-autocorrelation scenario, while Row 2 (Panels C and D) shows the performance of RSFs in a low-habitat-autocorrelation scenario. Black points in Column 1 (Panels A and C) indicate mean parameter estimates for weighted RSFs performed on 400 simulated movement tracks, while white points in Column 2 (Panels B and D) indicate mean parameter estimates for IID RSFs. Bars indicate average confidence intervals across all 400 simulations. In this case, the animal is not selecting habitat at all, yet 50% habitat-specific triangulation failure creates the appearance of strong avoidance of Habitat 2 in favor of Habitat 1. In both habitat scenarios, confidence intervals become narrower as sampling rate increases for IID RSFs, while wRSF confidence intervals largely remain stable. Parameter estimates improve (i.e., become closer to the truth at zero) for wRSFs as autocorrelation of the data increases, while parameter estimates of IID RSFs remain stable. The advantages of wRSFs are more clearly visible in the high-habitat-autocorrelation scenario, but weighting still improves both parameter estimates and confidence intervals in the low-habitat-autocorrelation scenario.*

with parameter estimates of weighted RSFs remaining consistently around the true parameter (Fig. 2). The width of confidence intervals remained stable as the sampling rate (i.e., autocorrelation) increased (Fig. 2A), but contracted as sampling duration (i.e., effective sample size) increased (Fig. 2B). Weighted RSFs could detect habitat selection when effective sample sizes were ≥ 32 (or 32 days) in this example.

Figure 2: *Mean parameter estimates and mean confidence intervals around those parameter estimates for weighted RSFs during simulations of habitat selection. In this case, the animal is selecting habitat such that its movement path contains three times as many locations in Habitat 2 as in Habitat 1. Panel A shows the performance of weighted RSFs as the sampling rate (i.e., autocorrelation) increases, while Panel B shows the performance of weighted RSFs as sampling duration (effective sample size) increases. As expected, point estimates of the selection parameter are constant across all scenarios, while confidence in those parameters increases as sampling duration (i.e., effective sample size), but not sampling rate (i.e., autocorrelation), increases.*

Empirical Examples

In the water mongoose we analyzed (Fig. 3A), we found that although point estimates for avoidance of plantation were similar between weighted and IID RSFs ($\hat{\beta}_{\mathrm{w}}$ = -2.22 vs. $\hat{\beta}_{\mathrm{IID}}$ = -2.33), avoidance of plantation areas was only statistically significant for the IID RSF (CI_w = -4.79 – 0.35 vs. $\mathrm{CI}_{\mathrm{IID}}$ = -3.66 – -1.01). This indicates that the IID RSF incorrectly identified avoidance of plantation areas. Both IID and weighted RSFs identified statistically significant avoidance of built-up areas. However, the point estimate of avoidance of built-up areas for the weighted RSF lies closer to zero and outside the 95% confidence interval generated by the IID RSF ($\hat{\beta}_{\mathrm{w}}$ = -1.23 vs. $\hat{\beta}_{\mathrm{IID}}$ = -1.90; CI_{w} = -2.04 – -0.42 vs. $\mathrm{CI}_{\mathrm{IID}}$ = -2.35 – -1.45), indicating that the IID RSF substantially overestimated avoidance of built-up areas.

In the caracal we analyzed (Fig. 3B), we found that although the IID RSF identified statistically significant avoidance of plantation areas ($\hat{\beta}_{\mathrm{IID}}$ = -2.02; $\mathrm{CI}_{\mathrm{IID}}$ = -3.96 – -0.08), the weighted RSF found no evidence for avoidance being statistically significant ($\hat{\beta}_{\mathrm{w}}$ = -1.17; CI_{w} = -8.25 – 5.91). Moreover, although point estimates for selection of built-up areas were similar ($\hat{\beta}_{\mathrm{w}}$ = 1.68 vs. $\hat{\beta}_{\mathrm{IID}}$

Figure 3: *Mean parameter estimates and confidence intervals for IID (white points) and weighted (black points) RSFs during empirical tests on a water mongoose (Panel A), a caracal (Panel B), and a serval (Panel C). Weighting meaningfully alters point estimates of selection for built-up areas for the mongoose, and appropriately expands confidence intervals around parameter estimates, changing inferred habitat selection for all three animals.*

= 1.58), confidence intervals were substantially wider for the weighted RSF (CI_w = 0.67 − 2.69 vs. CI_{IID} = 1.39 − 1.78), indicating that the IID RSF overstates confidence in selection for built-up areas.

In the serval we analyzed (Fig. 3C), we found that although point estimates for avoidance of plantation areas were similar between weighted and IID RSFs ($\hat{\beta}_w$ = -2.04 vs. $\hat{\beta}_{IID}$ = -1.88), avoidance of plantation areas was only statistically significant for the IID RSF (CI_w = -5.65 − 1.56 vs. CI_{IID} = -2.47 − -1.30). Point estimates for avoidance of built-up areas were also similar between weighted and IID RSFs ($\hat{\beta}_w$ = -0.57 vs. $\hat{\beta}_{IID}$ = -0.66), but this avoidance was again only statistically significant for the IID RSF (CI_w = -1.88 − 0.73 vs. CI_{IID} = -0.90 − -0.43). The IID RSF therefore incorrectly identified statistically significant avoidance of both plantation and built-up areas for this species.

Taken together, our analyses demonstrate that weighted RSFs conducted on real-world animal tracking data perform consistently with predictions and simulations. This indicates that coefficients from conventional RSFs that assume IID data can be both biased and overconfident in real-world data sets, even at absolute sample sizes that are relatively small for modern GPS data sets.

Discussion

Although the autocorrelated nature of modern animal tracking data is known to pose statistical challenges for conventional RSFs that assume IID data (Boyce, 2006, Fieberg *et al.*, 2010, Northrup *et al.*, 2013), no generally accepted solutions to these challenges have been described. Wildlife biologists therefore often fit IID RSF models to autocorrelated tracking data, which can lead to both bias and pseudoreplication in parameter estimates of RSFs. In this study, we introduced a method of weighting individual locations to account for autocorrelation, which reduces pseudoreplication and bias in RSFs. This method can improve estimates of confidence intervals provided by RSFs when tracking data are autocorrelated (as is the case for most modern animal tracking data sets), dramatically reducing the incidence of Type I error in studies of resource selection. It also reduces bias in parameter estimates when some covariates are over- or undersampled, such as when study animals inhabit areas where landscape features cause frequent triangulation failures in GPS devices, researchers employ duty cycling, or researchers use tracking technologies that do not produce regularly sampled location data (e.g., VHF telemetry or visual searches for marked individuals). In the future, our approach could be extended further to address other statistical problems associated with RSFs, which we discuss below.

Rigorous estimation of confidence intervals on RSF parameters is a challenge using conventional RSF methods. Because consecutive data points in modern animal tracking data are rarely statistically independent, using all the data available in an autocorrelated movement track leads to increasing (and increasingly misleading) confidence in parameter estimates with increasing sampling frequencies. Data thinning can be used to reduce autocorrelation between successive locations, which can widen confidence intervals that are known to be too narrow. In practice, however, data thinning inherently involves loss of useful information on an animal's movements, is generally applied in an ad-hoc and non-optimized way, and can pose other statistical problems (Noonan *et al.*, 2020). Generalized estimating equations can also be used to estimate more robust confidence intervals while using all the data in a movement track, but the "cluster" sizes (akin to effective sample sizes) are often also chosen arbitrarily (Prima *et al.*, 2017), and it is difficult

to implement generalized estimating equations jointly with other common techniques to mitigate statistical issues in RSFs (e.g., random effects). The method we propose in this study offers an objective way to estimate effective sample sizes for generating reliable confidence intervals.

In addition to estimating more robust confidence intervals, our method of weighting also offers potential to mitigate bias that can arise from inadequate sampling of landscape features that cause triangulation failure, irregular sampling designs intended to document animal behavior at multiple temporal scales, or technologies that do not allow regular sampling intervals. As shown by our simulations (Fig. 1), RSFs that assume IID data cannot handle frequent habitat-induced triangulation failure–estimates of selection of those habitats are negatively biased, and models become artificially more confident in this bias as the sampling rate increases. Our method of weighting reduces this bias, and the quality of estimated parameters improves rather than declines as the sampling rate increases. Duty cycling, by which researchers periodically suspend or reduce the sampling rate in an attempt to collect high-resolution data over longer total periods of time while conserving battery life, offers another promising use case for our method. For example, researchers may be interested in foraging behavior of a crepuscular animal and therefore program a tracking device to collect more locations near dawn and dusk, when an animal is actively foraging. In this case, an IID RSF would result in sampling foraging areas out of proportion to other areas where an animal spends its time, potentially underestimating use of landscape features that play important roles as rest sites, escape cover, and/or thermal refugia. A weighted RSF could account for this variation in sampling effort, providing a more objective parameter estimate for overall resource selection. Finally, many species are not amenable to wearing tracking devices that acquire fixes at regular intervals. Aquatic or semi-aquatic species may spend long periods of time underwater where they are inaccessible to satellites (Breed *et al.*, 2011, Fleming *et al.*, 2018), and many species remain too small for today's GPS technology (McMahon *et al.*, 2017, Weller *et al.*, 2016). Weighted RSFs provide a way for researchers to conduct RSFs that are less biased by such technological constraints.

Although applied here to tracking data of individual animals, the statistical principles underlying our method are also applicable at the species and population levels, which would be of great

use to researchers. Random effects are often used to account for non-independence of data for

individuals within a population. Gillies *et al.* (2006) recommended using random intercepts to

account for unequal sample sizes between individuals in studies of habitat selection, and random

slopes to account for differences in habitat selection between individuals. Similarly, Hebblewhite

& Merrill (2008) recommended using random intercepts to account for non-independence arising

from correlated locations in socially structured populations (e.g., repeated observations from in-

dividuals within the same herd or pack). Incorporating random effects into RSFs and SSFs has

since become standard practice (Fieberg *et al.*, 2010, Muff *et al.*, 2020), but remains an imperfect

solution for accounting for pseudoreplication. Random effects generate confidence intervals that

depend on variation in the point estimates of RSF parameters between individuals, which may or

may not reflect true parameter uncertainty. If autocorrelation in individual movement tracks is not

properly accounted for, individual sampling variance is underestimated, which inflates population

variance estimated using random effects. Checking for this is difficult and rarely (if ever) per-

formed. Population-level likelihood weighting, however, has potential to serve similar purposes.

Weighting based on effective sample sizes can account for individuals having been tracked for dif-

fering amounts of time, and autocorrelation-informed weighting can account for differing amounts

of spatial or temporal autocorrelation between individuals in a study area or study period. Iterated

over both individuals and populations, such weight optimization accounts for temporal sampling

bias within individual time-series, as it did for the single-individual weighting described in this

study, while also accounting for sampling biases among individuals, where certain individuals are

better sampled than others.

Because location weights are generated using weighted Autocorrelated Kernel Density Esti-

mate range distributions, which can only be validly calculated on animals that are range-resident,

this method cannot be used to conduct RSFs on individuals that are not range-resident. However,

it should be noted that because conventional IID RSFs assume stationary distributions of resource

use and availability, conventional RSFs also assume range residency (but this assumption is often

ignored). Step-selection functions are therefore more appropriate for studying resource selection

by migratory, dispersing, or nomadic animals because they do not assume stationary distributions of use and availability. The inferences gained from such studies should be treated with caution, however, because autocorrelation can still lead to pseudoreplication and bias in inferences gained from SSFs of these movement tracks.

Although ecologists and conservation biologists have access to better statistical and computational tools than ever before, new methods are still required to maximize the value of animal tracking data for informing our understanding of the natural world. In this study, we introduced a method of autocorrelation-informed weighting to reduce pseudoreplication and bias when parameterizing RSFs with autocorrelated and irregular animal tracking data, which are pervasive in studies of animal movement. Our method is easily implemented using the `ctmm` R package (Calabrese *et al.*, 2016), and we provide an annotated workflow in our supplementary materials (Appendix 1) so that other researchers can implement weighted RSFs on their own animal location data. We hope that the analytical technique we have provided here, which is grounded in statistical theory and validated using simulations and empirical data, allows continued progress toward more statistical rigor in studies of resource selection by animals.

Acknowledgements

We thank I. Silva for providing feedback on the weighted RSF tutorial in Appendix 1. JC and CF were supported by NSF IIBR 1915347. RK was supported by NSF ABI 1914928 and NASA Ecological Informatics 80NSSC21K1182. This work was partly funded by the Center for Advanced Systems Understanding (CASUS) which is financed by Germany's Federal Ministry of Education and Research (BMBF) and by the Saxon Ministry for Science, Culture and Tourism (SMWK) with tax funds on the basis of the budget approved by the Saxon State Parliament.

Data Accessibility

All data and code used in analyses for this manuscript has been included in the supplementary material for this manuscript and will be made publicly available on the *RODARE* data repository upon publication of this manuscript.

Authors' Contributions

JC and CF conceived the ideas; CF led the mathematical derivation and software implementation; JA conducted the simulations and empirical analyses; JS, CD, and TR contributed empirical data; and JA and CF led the writing of the manuscript. All authors contributed critically to drafts of the manuscript and gave final approval for publication.

References

Aarts, G., MacKenzie, M., McConnell, B., Fedak, M. & Matthiopoulos, J. (2008) Estimating space-use and habitat preference from wildlife telemetry data. *Ecography* **31**, 140–160.

Ali, A.H., Ford, A.T., Evans, J.S., Mallon, D.P., Hayes, M.M., King, J., Amin, R. & Goheen, J.R. (2017) Resource selection and landscape change reveal mechanisms suppressing population recovery for the world's most endangered antelope. *Journal of Applied Ecology* **54**, 1720–1729.

Alston, J.M., Joyce, M.J., Merkle, J.A. & Moen, R.A. (2020) Temperature shapes movement and habitat selection by a heat-sensitive ungulate. *Landscape Ecology* **35**, 1961–1973.

Avgar, T., Potts, J.R., Lewis, M.A. & Boyce, M.S. (2016) Integrated step selection analysis: bridging the gap between resource selection and animal movement. *Methods in Ecology and Evolution* **7**, 619–630.

Babin, J.S., Fortin, D., Wilmshurst, J.F. & Fortin, M.E. (2011) Energy gains predict the distribution of plains bison across populations and ecosystems. *Ecology* **92**, 240–252.

Boyce, M.S. (2006) Scale for resource selection functions. *Diversity and Distributions* **12**, 269–276.

Boyce, M.S. & McDonald, L.L. (1999) Relating populations to habitats using resource selection functions. *Trends in Ecology & Evolution* **14**, 268–272.

Breed, G.A., Costa, D.P., Goebel, M.E. & Robinson, P.W. (2011) Electronic tracking tag programming is critical to data collection for behavioral time-series analysis. *Ecosphere* **2**, art10.

Calabrese, J.M., Fleming, C.H. & Gurarie, E. (2016) ctmm: an r package for analyzing animal relocation data as a continuous-time stochastic process. *Methods in Ecology and Evolution* **7**, 1124–1132.

Craiu, R.V., Duchesne, T. & Fortin, D. (2008) Inference methods for the conditional logistic regression model with longitudinal data. *Biometrical Journal* **50**, 97–109.

Dickie, M., McNay, S.R., Sutherland, G.D., Cody, M. & Avgar, T. (2020) Corridors or risk? Movement along, and use of, linear features varies predictably among large mammal predator and prey species. *Journal of Animal Ecology* **89**, 623–634.

EKZN Wildlife, . & GeoTerraImage (2018) Updating the existing KZN provincial land-cover map 2011 to 2017: data users report and metadata (Version 001). Unpublished Report, Biodiversity Research and Assessment, Ezemvelo KZN Wildlife, PO Box 13053, Cascades, 3202, South Africa.

Fieberg, J., Matthiopoulos, J., Hebblewhite, M., Boyce, M.S. & Frair, J.L. (2010) Correlation and studies of habitat selection: problem, red herring or opportunity? *Philosophical Transactions of the Royal Society B: Biological Sciences* **365**, 2233–2244.

Fieberg, J., Signer, J., Smith, B. & Avgar, T. (2021) A 'how to' guide for interpreting parameters in habitat-selection analyses. *Journal of Animal Ecology* **90**, 1027–1043.

Fleming, C.H., Calabrese, J.M., Mueller, T., Olson, K.A., Leimgruber, P. & Fagan, W.F. (2014) From fine-scale foraging to home ranges: a semivariance approach to identifying movement modes across spatiotemporal scales. *American Naturalist* **183**, E154–E167.

Fleming, C.H., Noonan, M.J., Medici, E.P. & Calabrese, J.M. (2019) Overcoming the challenge of small effective sample sizes in home-range estimation. *Methods in Ecology and Evolution* **10**, 1679–1689.

Fleming, C.H., Sheldon, D., Fagan, W.F., Leimgruber, P., Mueller, T., Nandintsetseg, D., Noonan, M.J., Olson, K.A., Setyawan, E., Sianipar, A. & Calabrese, J.M. (2018) Correcting for missing and irregular data in home-range estimation. *Ecological Applications* **28**, 1003–1010.

Fortin, D., Beyer, H.L., Boyce, M.S., Smith, D.W., Duchesne, T. & Mao, J.S. (2005) Wolves influence elk movements: behavior shapes a trophic cascade in Yellowstone National Park. *Ecology* **86**, 1320–1330.

Frair, J.L., Fieberg, J., Hebblewhite, M., Cagnacci, F., DeCesare, N.J. & Pedrotti, L. (2010) Resolving issues of imprecise and habitat-biased locations in ecological analyses using GPS telemetry data. *Philosophical Transactions of the Royal Society B: Biological Sciences* **365**, 2187–2200.

Gillies, C.S., Hebblewhite, M., Nielsen, S.E., Krawchuk, M.A., Aldridge, C.L., Frair, J.L., Saher, D.J., Stevens, C.E. & Jerde, C.L. (2006) Application of random effects to the study of resource selection by animals. *Journal of Animal Ecology* **75**, 887–898.

Hebblewhite, M. & Merrill, E. (2008) Modelling wildlife–human relationships for social species with mixed-effects resource selection models. *Journal of Applied Ecology* **45**, 834–844.

Hijmans, R.J. (2021) raster: geographic data analysis and modeling.

Hooten, M.B., Hanks, E.M., Johnson, D.S. & Alldredge, M.W. (2014) Temporal variation and scale in movement-based resource selection functions. *Statistical Methodology* **17**, 82–98.

Horne, J.S., Garton, E.O. & Rachlow, J.L. (2008) A synoptic model of animal space use: simultaneous estimation of home range, habitat selection, and inter/intra-specific relationships. *Ecological Modelling* **214**, 338–348.

Hurlbert, S.H. (1984) Pseudoreplication and the design of ecological field experiments. *Ecological Monographs* **54**, 187–211.

Kohl, M.T., Stahler, D.R., Metz, M.C., Forester, J.D., Kauffman, M.J., Varley, N., White, P.J., Smith, D.W. & MacNulty, D.R. (2018) Diel predator activity drives a dynamic landscape of fear. *Ecological Monographs* **88**, 638–652.

Lamont, B.G., Monteith, K.L., Merkle, J.A., Mong, T.W., Albeke, S.E., Hayes, M.M. & Kauffman, M.J. (2019) Multi-scale habitat selection of elk in response to beetle-killed forest. *Journal of Wildlife Management* **83**, 679–693.

Leclerc, M., Vander Wal, E., Zedrosser, A., Swenson, J.E., Kindberg, J. & Pelletier, F. (2016) Quantifying consistent individual differences in habitat selection. *Oecologia* **180**, 697–705.

Lewis, J.S., Rachlow, J.L., Garton, E.O. & Vierling, L.A. (2007) Effects of habitat on GPS collar performance: using data screening to reduce location error. *Journal of Applied Ecology* **44**, 663–671.

Manly, B., McDonald, L., Thomas, D., McDonald, T. & Erickson, W. (2007) *Resource selection by animals: statistical design and analysis for field studies*. Springer Science & Business Media.

McMahon, L.A., Rachlow, J.L., Shipley, L.A., Forbey, J.S., Johnson, T.R. & Olsoy, P.J. (2017) Evaluation of micro-GPS receivers for tracking small-bodied mammals. *PLOS ONE* **12**, e0173185.

Merkle, J.A., Monteith, K.L., Aikens, E.O., Hayes, M.M., Hersey, K.R., Middleton, A.D., Oates, B.A., Sawyer, H., Scurlock, B.M. & Kauffman, M.J. (2016) Large herbivores surf waves of green-up during spring. *Proceedings of the Royal Society B: Biological Sciences* **283**, 20160456.

Montgomery, R.A., Redilla, K.M., Ortiz-Calo, W., Smith, T., Keller, B. & Millspaugh, J.J. (2018) Evaluating the individuality of animal-habitat relationships. *Ecology and Evolution* **8**, 10893–10901.

Muff, S., Signer, J. & Fieberg, J. (2020) Accounting for individual-specific variation in habitat-selection studies: Efficient estimation of mixed-effects models using Bayesian or frequentist computation. *Journal of Animal Ecology* **89**, 80–92.

Newediuk, L., Prokopenko, C.M. & Vander Wal, E. (2022) Individual differences in habitat selection mediate landscape level predictions of a functional response. *Oecologia* .

Ng'weno, C.C., Ford, A.T., Kibungei, A.K. & Goheen, J.R. (2019) Interspecific prey neighborhoods shape risk of predation in a savanna ecosystem. *Ecology* **100**, e02698.

Nielsen, S.E., Boyce, M.S., Stenhouse, G.B. & Munro, R.H.M. (2002) Modeling grizzly bear habitats in the Yellowhead Ecosystem of Alberta: taking autocorrelation seriously. *Ursus* **13**, 45–56.

Noonan, M.J., Fleming, C.H., Tucker, M.A., Kays, R., Harrison, A.L., Crofoot, M.C., Abrahms, B., Alberts, S.C., Ali, A.H., Altmann, J., Antunes, P.C., Attias, N., Belant, J.L., Beyer, D.E., Bidner, L.R., Blaum, N., Boone, R.B., Caillaud, D., Paula, R.C.d., Torre, J.A.d.l., Dekker, J., DePerno, C.S., Farhadinia, M., Fennessy, J., Fichtel, C., Fischer, C., Ford, A., Goheen, J.R., Havmøller, R.W., Hirsch, B.T., Hurtado, C., Isbell, L.A., Janssen, R., Jeltsch, F., Kaczensky, P., Kaneko, Y., Kappeler, P., Katna, A., Kauffman, M., Koch, F., Kulkarni, A., LaPoint, S., Leimgruber, P., Macdonald, D.W., Markham, A.C., McMahon, L., Mertes, K., Moorman, C.E., Morato, R.G., Moßbrucker, A.M., Mourão, G., O'Connor, D., Oliveira-Santos, L.G.R., Pastorini, J., Patterson, B.D., Rachlow, J., Ranglack, D.H., Reid, N., Scantlebury, D.M., Scott, D.M., Selva, N., Sergiel, A., Songer, M., Songsasen, N., Stabach, J.A., Stacy-Dawes, J., Swingen, M.B., Thompson, J.J., Ullmann, W., Vanak, A.T., Thaker, M., Wilson, J.W., Yamazaki, K., Yarnell, R.W., Zieba, F., Zwijacz-Kozica, T., Fagan, W.F., Mueller, T. & Calabrese, J.M. (2020)

Effects of body size on estimation of mammalian area requirements. *Conservation Biology* **34**, 1017–1028.

Noonan, M.J., Tucker, M.A., Fleming, C.H., Akre, T.S., Alberts, S.C., Ali, A.H., Altmann, J., Antunes, P.C., Belant, J.L., Beyer, D., Blaum, N., Böhning-Gaese, K., Cullen, L., Paula, R.C.d., Dekker, J., Drescher-Lehman, J., Farwig, N., Fichtel, C., Fischer, C., Ford, A.T., Goheen, J.R., Janssen, R., Jeltsch, F., Kauffman, M., Kappeler, P.M., Koch, F., LaPoint, S., Markham, A.C., Medici, E.P., Morato, R.G., Nathan, R., Oliveira-Santos, L.G.R., Olson, K.A., Patterson, B.D., Paviolo, A., Ramalho, E.E., Rösner, S., Schabo, D.G., Selva, N., Sergiel, A., Silva, M.X.d., Spiegel, O., Thompson, P., Ullmann, W., Zięba, F., Zwijacz-Kozica, T., Fagan, W.F., Mueller, T. & Calabrese, J.M. (2019) A comprehensive analysis of autocorrelation and bias in home range estimation. *Ecological Monographs* **89**, e01344.

Northrup, J.M., Hooten, M.B., Anderson Jr., C.R. & Wittemyer, G. (2013) Practical guidance on characterizing availability in resource selection functions under a use–availability design. *Ecology* **94**, 1456–1463.

Northrup, J.M., Wal, E.V., Bonar, M., Fieberg, J., Laforge, M.P., Leclerc, M., Prokopenko, C.M. & Gerber, B.D. (2021) Conceptual and methodological advances in habitat-selection modeling: guidelines for ecology and evolution. *Ecological Applications* p. e02470.

O'Neill, H.M.K., Durant, S.M. & Woodroffe, R. (2020) What wild dogs want: habitat selection differs across life stages and orders of selection in a wide-ranging carnivore. *BMC Zoology* **5**, 1.

Prima, M.C., Duchesne, T. & Fortin, D. (2017) Robust inference from conditional logistic regression applied to movement and habitat selection analysis. *PLOS ONE* **12**, e0169779.

Prokopenko, C.M., Boyce, M.S. & Avgar, T. (2017a) Characterizing wildlife behavioural responses to roads using integrated step selection analysis. *Journal of Applied Ecology* **54**, 470–479.

Prokopenko, C.M., Boyce, M.S. & Avgar, T. (2017b) Extent-dependent habitat selection in a migratory large herbivore: road avoidance across scales. *Landscape Ecology* **32**, 313–325.

R Core Team, . (2020) R: a language and environment for statistical computing.

Ramesh, T., Kalle, R. & Downs, C.T. (2016a) Space use in a South African agriculture landscape by the caracal (Caracal caracal). *European Journal of Wildlife Research* **63**, 11.

Ramesh, T., Kalle, R. & Downs, C.T. (2016b) Spatiotemporal variation in resource selection of servals: insights from a landscape under heavy land-use transformation. *Journal of Mammalogy* **97**, 554–567.

Rayl, N.D., Proffitt, K.M., Almberg, E.S., Jones, J.D., Merkle, J.A., Gude, J.A. & Cross, P.C. (2019) Modeling elk-to-livestock transmission risk to predict hotspots of brucellosis spillover. *The Journal of Wildlife Management* **83**, 817–829.

Robb, B.S., Merkle, J.A., Sawyer, H., Beck, J.L. & Kauffman, M.J. (2022) Nowhere to run: semi-permeable barriers affect pronghorn space use. *Journal of Wildlife Management* **early view**, e22212.

Scantlebury, D.M., Mills, M.G.L., Wilson, R.P., Wilson, J.W., Mills, M.E.J., Durant, S.M., Bennett, N.C., Bradford, P., Marks, N.J. & Speakman, J.R. (2014) Flexible energetics of cheetah hunting strategies provide resistance against kleptoparasitism. *Science* **346**, 79–81.

Signer, J., Fieberg, J. & Avgar, T. (2017) Estimating utilization distributions from fitted step-selection functions. *Ecosphere* **8**, e01771.

Street, G.M., Potts, J.R., Börger, L., Beasley, J.C., Demarais, S., Fryxell, J.M., McLoughlin, P.D., Monteith, K.L., Prokopenko, C.M., Ribeiro, M.C., Rodgers, A.R., Strickland, B.K., van Beest, F.M., Bernasconi, D.A., Beumer, L.T., Dharmarajan, G., Dwinnell, S.P., Keiter, D.A., Keuroghlian, A., Newediuk, L.J., Oshima, J.E.F., Rhodes Jr., O., Schlichting, P.E., Schmidt, N.M. & Vander Wal, E. (2021) Solving the sample size problem for resource selection functions. *Methods in Ecology and Evolution* **12**, 2421–2431.

Streicher, J.P., Ramesh, T. & Downs, C.T. (2021) An African urban mesocarnivore: Navigating the

urban matrix of Durban, South Africa. *Global Ecology and Conservation* **26**, e01482.

Swihart, R.K. & Slade, N.A. (1985) Testing for independence of observations in animal move-

ments. *Ecology* **66**, 1176–1184.

Thurfjell, H., Ciuti, S. & Boyce, M.S. (2014) Applications of step-selection functions in ecology

and conservation. *Movement Ecology* **2**, 4.

Ullmann, W., Fischer, C., Kramer-Schadt, S., Pirhofer-Walzl, K., Glemnitz, M. & Blaum, N.

(2020) How do agricultural practices affect the movement behaviour of European brown hares

(Lepus europaeus)? *Agriculture, Ecosystems & Environment* **292**, 106819.

Valls-Fox, H., De Garine-Wichatitsky, M., Fritz, H. & Chamaillé-Jammes, S. (2018) Resource de-

pletion versus landscape complementation: habitat selection by a multiple central place forager.

Landscape Ecology **33**, 127–140.

Weller, T.J., Castle, K.T., Liechti, F., Hein, C.D., Schirmacher, M.R. & Cryan, P.M. (2016) First di-

rect evidence of long-distance seasonal movements and hibernation in a migratory bat. *Scientific*

Reports **6**, 34585.

Worton, B.J. (1989) Kernel methods for estimating the utilization distribution in home-range stud-

ies. *Ecology* **70**, 164–168.

Appendix S2

Figure S1: *The movement track for the water mongoose used in our empirical example, super-imposed on the plantation (left) and built-up (right) cover types mapped by EKZN Wildlife & GeoTerraImage (2018). Green represents the cover type of interest.*

Figure S2: *The movement track for the caracal used in our empirical example, superimposed on the plantation (left) and built-up (right) cover types mapped by EKZN Wildlife & GeoTerraImage (2018). Green represents the cover type of interest.*

Figure S3: *The movement track for the serval used in our empirical example, superimposed on the plantation (left) and built-up (right) cover types mapped by EKZN Wildlife & GeoTerraImage (2018). Green represents the cover type of interest.*