Why the architecture of environmental fluctuation matters for fitness

(Manuscript type: Perspective)

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Statement of authorship: JSP developed the theoretical ideas with input from AF. JSP conducted the simulations and wrote the initial draft. AF contributed substantially to the writing.

Short running title: Architecture of fluctuation and fitness

Keywords: fluctuating environments, cyclical environments, temporal autocorrelation, life history, evolutionary demography, stochastic fitness, nonlinearity, Jensen's Inequality

Data accessibility statement: No new data were used in this study.

Number of words in abstract: 197

Number of words in main text: 4902

Number of figures: 5

Number of tables: 0

Number of text boxes: 0

Number of references: 79

Abstract

The physical environment represents the very stage upon which the eco-evolutionary play unfolds.

How fluctuations in the environment affect fitness is thus central to demographic projections,

selection predictions, life history analyses, and conservation of populations. Here we

systematically compare and explain why different 'architectures' of fluctuations—both in how

they are generated in nature and constructed theoretically—alter fitness. Modelling efforts have

mostly characterized variable environments with stochasticity. However, stochasticity refers to a

random process. Many fluctuating environments in nature are non-random, driven by geophysical

forces that create feedbacks or cyclicality: for example, seasons, tides, day/night, and disturbances

such as floods, fires, and hurricanes. We show that evolutionary fitness $log\lambda_s$ of a simple

structured population is sensitive to the nature of fluctuations, even if different fluctuations appear

indistinguishable in long-run statistical distributions. Importantly, we uncover two mathematical

explanations of why fitness depends on the architecture of environmental fluctuations—

consecutiveness of deviations from the environmental mean, and Jensen's Inequality acting on

nonlinear biological parameters—both arguably relevant features in virtually all populations

inhabiting variable environments. The fitness divergence we demonstrate and explain is a useful

step towards developing evolutionary demographic theory for the prevalent categories of non-

random environments.

Introduction

Predicting population growth—or fitness—is a centerpiece of life history theory, evolutionary demography, and conservation biology. Cole (1954) first conceived of the fitness of life history strategies as directly linked to the strategies' influence on population growth rate. Variations in the age distribution of survival and reproduction shape the fitness landscapes of life histories (Gadgil & Bossert 1970; Stearns 1976), upon which natural selection can act. Classical demography (Keyfitz 1968; Caswell 2001) leverages these concepts to analyze how the trajectories and sensitivities of population growth are influenced by population structures (*e.g.* in age, stage, or size) and their variations.

Early theories began with assumptions of constant environments for simplicity, and because it was necessary to lay the groundwork for comparing species in a systematic manner (Fisher 1958; MacArthur 1962). Developments since (Levins 1968; Schaffer 1974) have largely focused on extending analyses to include stochastic environment assumptions to reflect the complexities of nature (Lande *et al.* 2003; Tuljapurkar *et al.* 2003), and to turn our attention to increasing environmental variability under climate change (Easterling *et al.* 2000; Drake 2005; Boyce *et al.* 2006; Morris *et al.* 2008; Lawson *et al.* 2015). Seminal work by Tuljapurkar *et al.* (2009) demonstrated how and why the expected optimal life histories differ between constant and stochastic environments in a general sense.

In ecology and evolution, stochastic environmental formulations are now so commonplace that the term 'stochastic' is often used interchangeably with 'fluctuating' or 'variable' environments. Stochasticity indeed offers a powerful suite of tools to model an extensive spectrum of environments, principally afforded by mature statistical theories of probability distributions. In

short, a variable environment can typically be statistically summarized with moments of

distributions, e.g. the mean ($\bar{x}$) and standard deviation (σ) of a normal or lognormal distribution,

derivations of which lead to tractable analytical solutions for biological predictions in that

environment (Lawson *et al.* 2015), *e.g.* bet-hedging strategies (Cohen 1966; Childs *et al.* 2010).

Higher order moments of stochastic environments can be decomposed with Taylor expansion

methods to analyze the effects of, for example, intra- and inter-annual variability and temporal

autocorrelation on fitness (Tuljapurkar *et al.* 2009). Indeed, there is substantial theoretical and

empirical evidence that 'color' of noise in stochastic environments (negative to zero to positive

autocorrelation) influences fitness and extinction risk (Cohen 1966; Lande & Orzack 1988;

Vasseur & Yodzis 2004; Halley 2005; Tuljapurkar & Haridas 2006; Reuman *et al.* 2008; Engen *et*

al. 2013; Chevin *et al.* 2017). In summary, modern 'stochastic demography' is now a rich

discipline that places stochastic environmental formulation at its core to ask ecological and

evolutionary questions (Fieberg & Ellner 2001; Engen *et al.* 2009; Tuljapurkar 2013; Chevin *et*

al. 2017; Lande *et al.* 2017; Crewe *et al.* 2018; Schreiber & Moore 2018; Davison *et al.* 2019).

Mathematically, however, stochasticity is specific: it describes a random process, usually

implemented in biological models with stationary probability distributions.

Vast categories of environments are non-randomly driven by external forces in nature. For

example, fundamental geophysical fixtures such as the rotation of Earth (governing diel cycles),

its axial tilt and revolution around the Sun (seasonal cycles), and the revolution of the Moon around

Earth (tidal cycles) set sinusoidal oscillations in most terrestrial and aquatic ecosystems around the

planet. Physical processes such as the large mass and heat capacity of deep ocean waters

moderating the short-term fluctuations in ocean surface temperatures buffer chance events over

time and cause temporal autocorrelation (Steele 1985). Distinct from stochastic variability,

geophysically-driven fluctuations are 1) not emergent products of ecological and evolutionary

dynamics, 2) nor are they causally driven by past environmental states as in Markovian processes

or in phenomenologically autocorrelated environments. Analogous to driven harmonic oscillators

in physics, many environments are governed by permanent (at least on time scales relevant to

ecology and evolution) external drivers of fluctuations.

Here we envision generating functions that produce the timing patterns of events or state-changes

in an environment—from purely random to cyclical—to compare four broadly representative

architectures of fluctuating environments. Events here represent ecological disturbances (Lytle

2001) such as fires, floods, hurricanes, avalanches, droughts, and heatwaves, or, in seasonal

systems, onsets of winter that can largely impede biological activity (Inouye 2008). Stearns (1976,

Fig. 7) outlined the possible outcomes of environmental fluctuation frequency and predictability

for the age distribution of reproduction. Tuljapurkar (1985) analyzed how the relative scaling

between the period of environmental cycles and generation time influences population growth rate.

Recent theoretical work demonstrated that environmental periodicity is a distinct selective force

that determines optimal (Park 2019) as well as individual variation in life histories (Park &

Wootton 2021). Further, perturbations to seasonality and disturbance regimes constitute some of

the most widespread signatures of climate change, with far-reaching consequences such as

phenological shifts (Walther *et al.* 2002; Westerling *et al.* 2006; Marlon *et al.* 2009). Importantly,

frequency and predictability can vary independently in disturbance regimes (Lytle 2001; Lytle *et*

al. 2008; Denny *et al.* 2009). Thus, teasing apart the different generating functions of

environmental variability—beyond collapsing all environmental variability to stochasticity—is

crucial not only for more accurate predictions of population growth (Tuljapurkar & Haridas 2006),

but for understanding different causal drivers of population dynamics and life history selection.

Here we demonstrate why long-term fitness differs between stochastic *vs*. non-randomly

fluctuating environments. We construct a simple demographic fitness model to be systematically

compared across four broadly representative categories of fluctuating environments. We identify

two general mathematical mechanisms that cause divergences in fitness estimates across the four

categories. The first mechanism involves the pattern of consecutiveness of positive or negative

deviations from the mean environment; this pattern depends on the generating functions of

fluctuations. The second involves Jensen's Inequality acting on nonlinear life history traits, or vital

rates; the prevalence of such nonlinearity in nature is likely universal. Both are very general

properties of populations in variable environments, and we reveal how the two operate in concert

to drive fitness estimates to diverge from purely stochastic models. Finally, we discuss the general

relevance of our findings and identify key theoretical extensions and empirical tests that will be

important moving forward.

The generating functions of fluctuating environments

We start by setting up four generating functions of fluctuation in environments (Fig. 1). We mean

generating functions in the mathematical sense of rules underlying discrete encodings of infinite

series of events on a timeline. For all four generating functions, our focal parameter is the *interval*

between events. All four generating functions will have an error term σ, and it will be some

proportion of mean interval, both of which will be held constant for systematic comparison. Again,

these events represent ecological disturbances, or the onset of winter in seasonal systems with the

simplifying assumption that winters are harsh enough to perturb or halt biological activity. For the

seasonal case, note that celestial seasons are perfectly periodic, but the meteorological (*e.g.*

temperature, frost) onset of winter and arrival of spring are variable, making the intervening season

(the interval) also variable in length ('climatological growing seasons', *sensu* Linderholm 2006).

Figure 1. Four generating functions of fluctuating environments. Each fluctuation architecture is determined by the process of generating a sequence of discrete intervals between events on a continuous timeline.

112   *Generating function I: Stochastic (random)*

113   We generate a stochastic environment by drawing event intervals X_t from a normal distribution:

$$X_t \sim N(\bar{X}, \sigma) \tag{1}$$

where $\bar{X}$ is the mean interval. We do this instead of generating a series of events via a Poisson

process, which is a more common way of generating random events on a timeline, to make this

case comparable with the other three cases with respect to their error distribution $N(0, \sigma)$, and

because in subsequent sections we will manipulate the amount of noise (σ) while holding the mean

interval constant (which is not possible with Poisson distributions). Note that this manner of

generating a fluctuating environment does not consider the source or nature of the noise, nor the

length of preceding intervals (*i.e.,* there is no memory of past fluctuations), but only assumes the

long-run statistical distribution of variability. While measured environmental noise may always

have a physical source in reality, and all systems may have some memory process to varying

degrees, some systems are so complex and noisy that they are well approximated by pure

randomness. Stochastic environments are experienced, for example, by mice living in dryland

ecosystems with variable resource availability (Noble *et al.* 2019), by weevil species using strongly

fluctuating oak acorns as egg-laying sites (Pélisson *et al.* 2012), and by annual herbs growing in

or near vernal pools created by stochastic winter precipitation events (Emery & La Rosa 2019).

Generating function II: Positively autoregressive

We generate a positively autocorrelated environment with a simple AR(1) process:

$$X_t = \bar{X} + \varphi X_{t-1} + \varepsilon_t \sim N(0, \sigma) \tag{2}$$

where $\bar{X}$ is the mean interval, and each interval X_t is positively correlated ($+\varphi$) with the interval

immediately preceding it (X_{t-1}). The i.i.d. (independent identically distributed) noise ε_t is drawn

from $N(0, \sigma)$. Positively autocorrelated environments are characterized by a 'compounding'

behavior where a longer or shorter interval than the mean is likely to beget another longer or shorter

interval in the immediate future, respectively. Examples of positively autocorrelated fluctuations

in nature include some fire regimes (Caswell & Kaye 2001), oceanic temperatures (Steele 1985;

Halley 2005), and flood regimes (Sabo & Post 2008).

Generating function III: Negatively autoregressive

The negative counterpart to the positively autoregressive environment is one with a negative

autoregressive term $(-\varphi)$ and all other terms identical to Eq. 2:

$$X_t = \bar{X} - \varphi X_{t-1} + \varepsilon_t \sim N(0, \sigma) \tag{3}$$

Negatively autocorrelated environments are characterized by a 'compensating' or self-correcting

behavior where a longer or shorter interval than the mean is likely to be counterbalanced with a

shorter or longer interval in the immediate future, respectively. Real examples of negatively

autocorrelated environments in nature, though much more rarely considered than positively

autocorrelated ones (Metcalf & Koons 2007), include terrestrial plant systems with delayed

density-dependence driven by nutrient cycling (Gonzalez-Andujar *et al.* 2006), and tree masting

(Hacket-Pain & Bogdziewicz 2021).

Generating function IV: 'Noisy Clock'

The last generating function describes a fluctuating environment with a fundamental external

driver of cyclicality, whose deterministic periodicity is not affected by the realized noise. We

henceforth refer to this case as the 'Noisy Clock' to describe the clocked (deterministic) nature of

fluctuations that underlies the noise. Here, we first punctuate the timeline with a series of

deterministic 'due' times of events much like the celestial periodicity of Earth's revolution that

determines the boundaries of the annual cycle. Intervals are not generated by counting time from

the preceding event, but imperfectly timed events occur around each 'due' time with i.i.d. noise

$N(0, \sigma)$. Then the intervals are computed by measuring gaps between subsequent realized events:

$X_t = Realized\ event_{t+1} - Realized\ event_t$

$$= [Due_{t+1} + \varepsilon_{t+1} \sim N(0, \sigma)] - [Due_t + \varepsilon_t \sim N(0, \sigma)] \tag{4}$$

Counting time from preceding events (which is the case for generating functions I, II, and III) as

opposed to in reference to an external clock would be an unrealistic way to model fundamentally

cyclical systems such as seasonal ones. For instance, if one were to model a winter event (*e.g.* first

snowfall), but stochastically modelled the length of intervening summer seasons with no regard to

due times, then after some additive sequence of stochastic lengths it would be possible to have

winters starting in July when lined up against a deterministic calendar. This would be unrealistic

under any reasonable geophysical stationarity assumptions. Adding error to due times has the

effect of gravitating noisy events back to realistic due times set by the underlying forces of

harmonic oscillations in the environment. Real fluctuations with clocked due times driven by

geophysical forces—but with error in the measured proxy variables—are exemplified by a myriad

of diel (*e.g.* hunting behavior of African wild dogs; Nouvellet *et al.* 2012), tidal (*e.g.* foraging of

intertidal snails; Hayford *et al.* 2018), and seasonal environments (*e.g.* advancement of egg-laying

with warming springs in numerous bird species; Dunn & Møller 2014). Despite widespread

attention to seasonal environments, particularly with respect to global phenological shifts (Walther

et al. 2002), the theoretical foundation of demographic dynamics and life history evolution in

cyclical environments is not yet well understood (Lande *et al.* 2017).

Comparing apples to apples: indistinguishable sample distributions

Despite the fact that the four simulated architectures of fluctuation (Fig. 2A) are known *a priori*

and fundamentally different, when one samples event intervals from each environmental time

series, one recovers normal distributions that are indistinguishable from each other (Fig. 2B&C).

The main purpose of demonstrating this sample similarity is to argue that long-run statistical

summaries of environments might give, incorrectly, the assurance that one can model any of the

four environments stochastically by drawing from the sampled normal distribution. However, we

will show shortly that the underlying generating functions, though invisible with sample

distributions, change fitness estimates. To our knowledge there is no simple derivation to force the

fundamentally different generating functions (Eq. 1-4) to be analytically equivalent with respect

to the expected sample distribution. The stochastic case, if generated with the same σ as the other

three, produces a slightly narrower sample distribution. We thus applied a multiplier of 1.3 to the

σ of the stochastic generating function, which heuristically makes its sample distribution

statistically indistinguishable from the other three (Fig. 2C). Our goal was to make the sample

distributions as similar as possible to demonstrate in the following steps that, despite their

similarity, the unequal architectures of environmental fluctuations produce fitness differences

through mechanisms that are invisible from time series summary statistics.

Figure 2. Visualizing the four generated environments. A) Time series of intervals over 1000 events are simulated with $\bar{X} = 200$ and $\sigma = 0.2 \times \bar{X}$. A multiplier of 1.3 was applied to the σ of the stochastic generating function to make the width of all four sample distributions more similar. Dotted lines represent the mean interval. B) Sampling 600 intervals from each environment with replacement recovers normal distributions. Dotted lines represent the mean interval. C) Shapiro-Wilk tests of normality on each independent sample distribution show that all are statistically normal ($p > 0.05$). Further, all six pairwise comparisons between the four sample distributions with Kolmogorov-Smirnov tests show that one cannot reject the null hypothesis that they came from the same distribution (p-value range: 0.15~0.64). AR: autoregressive.

A simple demographic model in fluctuating environments

Upon the environmental fluctuation backgrounds (Fig. 3A), we model a simple population that proceeds through time according to defined life history (or vital) rates. The population, described in a system of discrete difference equations, has a general two-stage structure consisting of juveniles and reproducing adults:

$$\begin{cases} J_{t+1} = A_t f \\ A_{t+1} = A_t S_A + J_t S_J \end{cases} \tag{5}$$

where J_t and A_t are numbers of juveniles and adults at time t respectively, f is adult fecundity,

and S_J and S_A are juvenile and adult survival rates respectively. The model is deliberately kept

simple in terms of structure for generality, and to focus on the effect that the different generating

functions have on fitness. Structural complexity (*e.g.* the number of developmental stages reflected

by the number of equations in the system, or the number of relevant vital rate terms) would be an

obvious area of customization for system-specific inquiry. As we will show, the important

mechanisms should propagate to overall fitness regardless of demographic complexity.

Next, suppose that a vital rate, S_J, changes nonlinearly between events. Verbally, this means that

if two disturbance events happen in quick succession, then S_J will be low, and if a long time passes

between events, then S_J will be high; and this change in S_J as a function of interval length is

nonlinear. For illustrative purposes, we assume the nonlinearity of S_J has a logistic shape as a

function of time since last event (Fig. 3B). Biologically, this function signifies the time needed for

juveniles to sufficiently develop, grow, or learn to endure the next impending perturbation in the

environment. Though not always logistic, nonlinearity in vital rate response functions is likely

common (Caswell 2008; Childs *et al.* 2010; Sletvold *et al.* 2013; Taylor *et al.* 2014; Ehrlén *et al.*

2016; Shriver 2016). For example, Morris et al. (2006) showed that, across five species of

perennial plants in the USA, vital rates including seedling survival (equivalent to S_J here), survival

at other stages, growth, germination, and seed production (equivalent to f here) showed nonlinear

change as a function of time since last hurricane or fire disturbance (disturbance phase). Similarly,

Bürkli & Jokela (2017) showed that embryonic development success (early-stage juvenile survival)

of a freshwater snail changed nonlinearly across the species' reproductive season. The exact shape

of vital rate response functions is another key point to be tailored for system-specific inquiry,

whether with empirical measurements or as realistic assumptions built into models.

Time-averaged fitness varies across the fluctuating environments

For each of the four architectures of environmental fluctuations, we ask what the long-run fitness

of a life history would be. The modelled life history is defined as the set of vital rate parameters

$\{f, S_A, S_J\}$ in the demographic model, scaled for per time-step rates. We hold $f = 0.2$ and $S_A =$

0.95 constant to analyze the effect of fluctuations in S_J along its logistic function about its mean

= 0.5 (Fig. 3B). Note that these values are arbitrary but biologically realistic. For fitness, we use

the 'small-noise approximation' (Tuljapurkar 1982) of time-averaged fitness in variable

environments:

$$log\lambda_s = \lim_{t \to \infty} \left(\frac{1}{t}\right) log[N(t)/N(0)] \qquad (6)$$

which gives the long-run growth rate in fluctuating environments (also known as 'a') where $N(t)$

is the total population size consisting of the sum of both juvenile and adult stages at time t. This

metric can be computed from numerical simulations over a long time course of the environment

(Caswell 2001), and is a robust approximation of fitness even under considerably large fluctuations

(Lande *et al.* 2003; Tuljapurkar *et al.* 2003, 2009). We simulate demographic dynamics (Eq. 5) in

each fluctuating environment (Fig. 1) for 1000 event intervals, calculate $log\lambda_s$, and repeat these

calculations over 500 iterations. Lastly, we compute all of the above across a spectrum of the

generating σ [0,0.3], *i.e.,* the error proportional to mean interval $\bar{X}$, to assess how the amount of

fluctuation noise affects the divergence of fitness estimates among the four architectures.

Figure 3. The interaction of environmental fluctuation architecture and nonlinearity in a vital rate creates divergence in fitness estimates. A) Diagrammatic representation of the four architectures of fluctuation. B) Nonlinear response function of a vital rate relative to time since last event. Here, juvenile survival rate (S_J) increases nonlinearly as the length of the interval between subsequent disturbance events increases. Dashed lines indicate the mean*s* of each axis, held constant in all modelled environments. C) Time-averaged fitness estimate*s* ($log\lambda_S$) are driven apart by the generating functions of environmental fluctuations as the amount of noise, or predictability, in the environment increases (modelled with standard deviation of event intervals). Each colored point depicts a single computation of $log\lambda_S$ after 1000 event intervals, corresponding to a given fluctuating environment and a given value of σ [0, 0.3]. The thicker lines correspond to the means of 500 iterations for each environment. The divergence in fitness estimates is largely consistent over the iterations.

Our results show that fitness estimates of the same life history model differ substantially among

the four architectures of fluctuating environments (Fig. 3C), despite the close similarity of the

environments' long-term time series statistics (Fig. 2). The long-run growth rate $log\lambda_S$ was overall

highest for the negatively autoregressive environment, slightly lower for the cyclically driven but

noisy environment (Noisy Clock), still lower for the purely stochastic, and by far the lowest for

the positively autoregressive environment. Mean fitness (thicker lines, Fig. 3C) was roughly

inversely correlated with amount of variance (spread of dots, Fig. 3C): the lower the mean, the

greater the spread. Furthermore, the four environments exhibited unequal responses of fitness to

an increase in σ: while increasing σ had a small positive effect on fitness in the negatively

autoregressive and Noisy Clock environments, it caused considerable negative trends of fitness in

stochastic and positively autoregressive environments. Importantly, all above differences in fitness

among the four environments are revealed as amount of noise (σ of event intervals) is increased.

This gradual divergence in fitness occurs up to $\sigma \approx 0.1$, after which it qualitatively stabilizes (*i.e.,*

further increase in σ seems to have relatively little effect on fitness across all four environments).

Mechanisms of fitness divergence: Consecutiveness of deviations and Jensen's Inequality

The central question that arises from Fig. 3C is why fitness estimates diverge between the four

architectures of fluctuating environments, given that the long-term statistical summaries of the

environments were hardly different (Fig. 2). There are two mechanisms that work in concert to

drive the divergence: (1) *Consecutiveness in deviations from the environmental mean,* and (2)

Jensen's Inequality in vital rate fluctuations.

(1) Consecutiveness in deviations from the environmental mean

In fluctuating environments with a mean interval, deviations to one side of the mean interval (*i.e.,*

below or above) can have short consecutive runs (*e.g.* flipping equally and quickly between below

and above the mean) or long consecutive runs (*e.g.* many deviations below or above the mean in a

row) even if the long-term overall distribution of deviations is normally distributed. The four

generating functions have different levels of compounding or compensatory behavior of

fluctuations, creating different distributions of consecutiveness (Fig. 4A). For instance, negatively

autoregressive and Noisy Clock environments both have a tendency to self-correct deviations back

to the mean, driven by the $-\varphi$ in the former, and the deterministic 'due times' in the latter. Thus,

the two environments have short tails in their distributions of consecutive deviation length, and

higher concentrations of short consecutive runs. In contrast, the positively autoregressive

environment by definition tends to repeat deviations to one side of the mean in a compounding

manner, resulting in a long tail in its consecutive deviation length distribution. The stochastic

environment exhibits an intermediate exponential decay distribution of consecutive deviations.

Figure 4. Mechanisms of fitness divergence between the architectures of fluctuating environments. A) Despite very similar long-term distributions of event intervals (Fig. 2), environments have very different distributions of consecutive runs to one side of (below or above) the mean interval depending on the nature of the generating function. Negatively autoregressive and Noisy Clock environments inherently exhibit compensatory or self-correcting behavior, resulting in shorter tails in the consecutive run distribution. The opposite is true of the positively autoregressive case. The stochastic environment exhibits an intermediate exponential distribution. B) Jensen's Inequality exists in any convex/cave fluctuating variable, and introduces asymmetry between environmental and demographic fluctuations. Nonlinear functions in vital rates with respect to intervals between ecological events such as disturbances are likely common. AR: autoregressive.

(2) Jensen's Inequality in vital rate fluctuations

Deviations in the environmental fluctuations below and above the mean interval have

asymmetrical consequences for the corresponding nonlinear demographic parameter (*i.e.,* vital rate,

here the juvenile survival rate S_J). This asymmetry is referred to as Jensen's Inequality (Jensen

1906; Ruel & Ayres 1999), which states that for a convex or concave function $f(x)$, the average

of the function, $\overline{f(x)}$, does not equal the function of the average, $f(\bar{x})$. Verbally, this means that

symmetrical fluctuation about the mean of a variable x along the x-axis does not translate

equivalently to symmetrical fluctuation about the mean along the y-axis. If we take fluctuation on

the x-axis as that of the environment, and fluctuation on the y-axis as that of demography, Jensen's

Inequality introduces a break in the environment-demography link in fluctuating environments

(Fig. 4B). The direction of that asymmetry (*i.e.,* $f(\bar{x}) \leq or \geq \overline{f(x)}$) as well as its magnitude

depend on the convex/cavity of the region of the nonlinear function where the mean lies. Here that

nonlinearly fluctuating variable is juvenile survival S_J, which has a direct effect on fitness $log\lambda_s$.

In summary, the mechanisms of fitness divergence between the architectures operate as follows:

i)   generating functions cause inherent differences in patterns of consecutive environmental

deviations towards one side of the mean, and

ii) since S_J is asymmetrically affected by environmental deviations to one or the other side of

the mean due to Jensen's Inequality, time-averaged fitness diverges between environments.

One last element that amplifies the fitness differences further is that $log\lambda_s$ is a multiplicative, or

geometric, fitness measure. In other words, short-term fitness differences caused by asymmetric

deviations per event are multiplied over the long-run, thus magnifying the divergences.

A) Convex/cavity of vital rate around the fluctuation mean

(y-axis: S_J (Juvenile survival); x-axis: Time since last event)

B) Corresponding time-averaged fitness in fluctuating environments

(y-axis: Fitness (log λ_s); x-axis: Amount of noise (σ of event intervals))

Figure 5. Demonstration of the effect of Jensen's Inequality on fitness divergence across architectures of environmental fluctuation. A) We morph the response function of S_J to manipulate its nonlinear shape around the mean event interval (dashed blue line), which is held constant. Going from left to right, the response function is more convex to more concave around the mean interval, highlighted in the yellow rectangles. This flips the sign of Jensen's Inequality, so that the overall effect of environmental fluctuations on fitness would generally become more positive to negative. B) The corresponding fitness plots demonstrate the sign flip in the effect of Jensen's Inequality as a result of the convex/cavity around the mean. Thick lines are means over all iterations, and bands are ± 1 sd. Colors correspond to the four architectures in previous figures. The middle panel is equivalent to a composite of Fig. 3C.

By manipulating the convex/cavity of the nonlinear demographic parameter S_J around the mean

(Fig. 5A), we explore further the biological consequences of the two interacting mathematical

mechanisms. When mean juvenile survival is low and its response function convex around the

mean (Fig. 5A, left panel), fitness increases with an increase in environmental variability (σ) due

to Jensen's Inequality (corresponding panel in Fig. 5B). Conversely, when mean juvenile survival

is high and its response function concave around the mean (Fig. 5A, right panel), fitness on average

is pulled down with an increase in environmental variability for the same reason (corresponding

panel in Fig. 5B). Across the convex/cavity spectrum of the logistic response function of juvenile

survival, as σ of the environmental fluctuation is increased from $0 \rightarrow 0.3$ ($0 \rightarrow 60$ time-steps,

since σ is proportional to mean interval $= 200$), the range of fitness estimates begins to converge

for all four environments ($log\lambda_s = 0.02{\sim}0.05$; Fig. 5B, note different y-axis ranges); this occurs

because with higher variance, fluctuations begin to sample from wider breadths of the logistic

response function which has a combination of convex and concave regions, rendering the role of

Jensen's Inequality weaker. Nevertheless, the relative ordering of fitness across environments (Fig.

5B, all panels) remains stable, demonstrating the interactive role that the consecutive deviation

patterns of environmental fluctuation architectures (mechanism 1) and the geometric nature of

$log\lambda_s$ play in differentiating fitness.

Summary and Future Outlook

We show in this Perspective how relying on simple stochastic modelling to describe fluctuating

environments, while powerful, can give erroneous and unreliable estimates of life history fitness

in non-randomly fluctuating environments. *A priori* knowledge or quantitative characterization of

the nature of fluctuations can thus help us explain a greater breadth of population dynamics and

evolutionary trajectories of life histories across environments. Non-random fluctuations abound in

nature, from temporally autocorrelated to externally driven cycles (Sabo & Post 2008; Dunn &

Møller 2014; Hacket-Pain & Bogdziewicz 2021). Incorporating non-randomness in environmental

models is not new to many researchers working on life history theory or demography.

Developments in periodic matrix models (Caswell & Trevisan 1994), the seasonal forcing of

disease dynamics (Altizer *et al.* 2006), bet-hedging strategies (Venable 2007), and the effect of

temporal autocorrelation on life history evolution and population persistence (Pike *et al.* 2004;

Metcalf & Koons 2007) are just a few of the examples. The novelty of our results is two-fold: we

show for the first time that a systematic comparison of random and non-random fluctuations

reveals consistent differences in fitness estimates of a life history; then, we resolve two simple

mathematical mechanisms that drive this divergence in fitness. These features are prevalent, and

their treatment should open new doors for investigating the implications for demography and

selection dynamics in the diverse—and changing—fluctuation architectures in nature.

In reality, environmental fluctuations will seldom neatly fall into only one of the architectural

categories simulated here. For example, a coastal system that is strongly perturbed by periodic

tidal disturbances will also be controlled by a lower resonance of seasonal fluctuations (Denny *et*

al. 2009). How resonances of non-random fluctuations constructively or destructively interfere to

produce a variety of fluctuation superpositions in different ecosystems might explain a wider

breadth of observed life history diversity and population trajectories. Even if an environment is

predominantly driven by one type of fluctuation architecture, the similarity of the sample

distributions (Fig. 2) can make it difficult to unequivocally place a given environment into one

fluctuation category. Empiricists could consider plotting the distribution of consecutive deviations

relative to a long-term mean interval of events—or periodicity of fluctuation—as we did in Fig.

4A. The shape of this distribution, particularly compared against the null hypothesis of a stochastic

(purely random) distribution of consecutive deviations simulated with the measured long-term

mean and standard deviation, might give an indication of whether an environment is fluctuating

positively autoregressively (long tail), or negatively autoregressively / in a Noisy Clock fashion

(short tail). Climate scientists and population ecologists regularly use spectral analyses to detect

dominant periodicities and lag times in time series of disturbance events or continuous

environmental variables (Priestley 1981; Chatfield 2003; Sabo & Post 2008), though often to

remove temporal autocorrelation so long-term trends can be detected (Pyper & Peterman 1998;

Brown *et al.* 2011). We argue that consideration of such methods should become more common

for life history and evolutionary demography analyses in nature. Our demonstrations suggest that

interactions between the architecture of environmental fluctuation and nonlinear biological vital

rates (*e.g.* survival, growth, reproduction) will alter fitness estimates in a broad range of situations.

For generality, we strived to simplify our assumptions and structure of the modelling framework,

particularly with respect to life history complexity. We emphasize that these are likely fruitful

areas for expansion and customization for specific empirical tests. For instance, many species are

naturally structured into more than two stages, for example amphibians (*e.g.* egg, larva, adult) and

insects (*e.g.* egg, nymph, pupa, adult). The number and interconnectedness of life stages via growth,

survival, and reproduction might have other complex and interesting influences on how fitness

differs between fluctuation architectures. The nonlinear response function of a vital rate can take

many different shapes depending on the more proximate developmental, physiological, or

behavioral mechanisms, from quadratic to self-excitatory threshold to polynomial functions (Ellis

& Post 2004; Stenseth *et al.* 2004; Nevoux *et al.* 2008). However, regardless of the life history

complexity or functional response shape, the joint effect of the two mechanisms we elucidated

here (consecutive patterns in fluctuations and Jensen's Inequality) must be in play if there is any

nonlinear fitness-related vital rate at any life history stage. If so, the effect should propagate

through the entire population dynamic, influence fitness, and drive apart fitness estimates in the

different architectures of environmental fluctuations as noise or unpredictability increases. What

would vary between systems is the magnitude of that effect depending on how nonlinear the vital

rates are, and how sensitive fitness is to those vital rates. Another open question is if interactions

between multiple nonlinear vital rates would amplify or dampen the effect of the mechanisms, to

exacerbate or lessen fitness differences between environmental fluctuation architectures.

Perturbation events or periods of harsh conditions are often stark enough to justify focusing on the
discrete intervals between them, as we did here, and as is common in disturbance ecology.
However, the amplitude, or magnitude, of a continuous environmental variable can itself fluctuate.
A prominent example is temperature variability, which is of particular interest in the context of
climate change (Post & Stenseth 1999; AghaKouchak *et al.* 2020). In the theoretical framework
we present (Fig. 1), variable amplitude can be simulated by allowing the magnitudes of subsequent
events to vary randomly or non-randomly, for example in the way events incur mortality or
influence the nonlinear vital rate response function following each event. Such analyses might
bridge the modelling philosophies that focus on refractory periods between events such as
temperature extremes and those that focus on continuous fluctuations in such variables.

Our general conclusion is that careful and appropriate treatments of the generating processes of
environmental fluctuations matter for estimating demographic fitness. In conjunction, while
nonlinear change in vital rates within seasons or disturbance phases might be arduous to measure
given the temporal sampling required, we argue that more often than not linearity is an assumption
made for simplicity, not inspired by nature. At least slight nonlinearity is likely a dominant feature
of many vital rates, and even if relatively weak, nonlinearity should unlock the synergistic role of
the mechanisms discussed here (consecutiveness of deviation patterns, and Jensen's Inequality) to
drive differences in fitness estimates depending on the architecture of an environment's temporal
fluctuations. Such analyses might significantly improve—as moving from constant to stochastic
assumptions did (Tuljapurkar *et al.* 2009)—our understanding of evolutionary demography based
on widespread stochastic assumptions.

Acknowledgements

We thank Tim Coulson, David Iles, John Jackson, Maja Kajin, Erik Kusch, Cathy Pfister, Roberto Rodríguez-Caro, Roberto Salguero-Gómez, Anna Vinton, and Tim Wootton for enriching discussions and feedback on drafts. We also thank members of the Coulson and Salguero-Gómez lab groups at the University of Oxford for providing a welcoming place to workshop our ideas, and for giving helpful feedback. JSP was supported by the European Union's Marie Skłodowska-Curie Individual Fellowship (101030973, "CyclesOfLife") and the National Science Foundation (OCE 1851489); AF was supported by the National Institutes of Health (U54 CA217376).

References

AghaKouchak, A., Chiang, F., Huning, L.S., Love, C.A., Mallakpour, I., Mazdiyasni, O., *et al.* (2020). Climate Extremes and Compound Hazards in a Warming World. *Annu. Rev. Earth Planet. Sci.*, 48, 519–548.

Altizer, S., Dobson, A., Hosseini, P., Hudson, P., Pascual, M. & Rohani, P. (2006). Seasonality and the dynamics of infectious diseases. *Ecol. Lett.*, 9, 467–484.

Boyce, M.S., Haridas, C.V., Lee, C.T., & the NCEAS Stochastic Demography Working Group. (2006). Demography in an increasingly variable world. *Trends Ecol. Evol.*, 21, 141–148.

Brown, C.J., Schoeman, D.S., Sydeman, W.J., Brander, K., Buckley, L.B., Burrows, M., *et al.* (2011). Quantitative approaches in climate change ecology. *Glob. Change Biol.*, 17, 3697–3713.

Bürkli, A. & Jokela, J. (2017). Increase in multiple paternity across the reproductive lifespan in a sperm-storing, hermaphroditic freshwater snail. *Mol. Ecol.*, 26, 5264–5278.

Caswell, H. (2001). Matrix population models: Construction, analysis, and interpretation. 2nd

edn sinauer associates. *Inc Sunderland MA*.

Caswell, H. (2008). Perturbation analysis of nonlinear matrix population models. *Demogr. Res.*,

18, 59–116.

Caswell, H. & Kaye, T.N. (2001). Stochastic demography and conservation of an endangered

perennial plant (Lomatium bradshawii) in a dynamic fire regime. In: *Advances in*

Ecological Research. Academic Press, pp. 1–51.

Caswell, H. & Trevisan, M.C. (1994). Sensitivity Analysis of Periodic Matrix Models. *Ecology*,

75, 1299–1303.

Chatfield, C. (2003). *The Analysis of Time Series: An Introduction, Sixth Edition*. 6th edn.

Chapman and Hall/CRC, New York.

Chevin, L.-M., Cotto, O. & Ashander, J. (2017). Stochastic Evolutionary Demography under a

Fluctuating Optimum Phenotype. *Am. Nat.*, 190, 786–802.

Childs, D.Z., Metcalf, C.J.E. & Rees, M. (2010). Evolutionary bet-hedging in the real world:

empirical evidence and challenges revealed by plants. *Proc. R. Soc. B Biol. Sci.*, 277,

3055–3064.

Cohen, D. (1966). Optimizing reproduction in a randomly varying environment. *J. Theor. Biol.*,

12, 119–129.

Cole, L.C. (1954). The Population Consequences of Life History Phenomena. *Q. Rev. Biol.*, 29,

103–137.

Crewe, P., Gratwick, R. & Grafen, A. (2018). Defining fitness in an uncertain world. *J. Math.*

Biol., 76, 1059–1099.

Davison, R., Stadman, M. & Jongejans, E. (2019). Stochastic effects contribute to population

fitness differences. *Ecol. Model.*, 408, 108760.

Denny, M.W., Hunt, L.J.H., Miller, L.P. & Harley, C.D.G. (2009). On the prediction of extreme

ecological events. *Ecol. Monogr.*, 79, 397–421.

Drake, J.M. (2005). Population effects of increased climate variation. *Proc. R. Soc. B Biol. Sci.*,

272, 1823–1827.

Dunn, P.O. & Møller, A.P. (2014). Changes in breeding phenology and population size of birds.

J. Anim. Ecol., 83, 729–739.

Easterling, D.R., Meehl, G.A., Parmesan, C., Changnon, S.A., Karl, T.R. & Mearns, L.O. (2000).

Climate Extremes: Observations, Modeling, and Impacts. *Science*, 289, 2068–2074.

Ehrlén, J., Morris, W.F., von Euler, T. & Dahlgren, J.P. (2016). Advancing environmentally

explicit structured population models of plants. *J. Ecol.*, 104, 292–305.

Ellis, A.M. & Post, E. (2004). Population response to climate change: linear vs. non-linear

modeling approaches. *BMC Ecol.*, 4, 2.

Emery, N.C. & La Rosa, R.J. (2019). The Effects of Temporal Variation on Fitness, Functional

Traits, and Species Distribution Patterns. *Integr. Comp. Biol.*, 59, 503–516.

Engen, S., Lande, R., Sæther, B. & Dobson, F.S. (2009). Reproductive Value and the Stochastic

Demography of Age-Structured Populations. *Am. Nat.*, 174, 795–804.

Engen, S., Sæther, B.-E., Armitage, K.B., Blumstein, D.T., Clutton-Brock, T.H., Dobson, F.S., *et*

al. (2013). Estimating the effect of temporally autocorrelated environments on the

demography of density-independent age-structured populations. *Methods Ecol. Evol.*, 4,

573–584.

Fieberg, J. & Ellner, S.P. (2001). Stochastic matrix models for conservation and management: a

comparative review of methods. *Ecol. Lett.*, 4, 244–266.

Fisher, R.A. (1958). *The genetical theory of natural selection*. Dover, New York.

Gadgil, M. & Bossert, W.H. (1970). Life Historical Consequences of Natural Selection. *Am.*

Nat., 104, 1–24.

Gonzalez-Andujar, J.L., Fernandez-Quintanilla, C. & Navarrete, L. (2006). Population Cycles

Produced by Delayed Density Dependence in an Annual Plant. *Am. Nat.*, 168, 318–322.

Hacket-Pain, A. & Bogdziewicz, M. (2021). Climate change and plant reproduction: trends and

drivers of mast seeding change. *Philos. Trans. R. Soc. B Biol. Sci.*, 376, 20200379.

Halley, J.M. (2005). Comparing aquatic and terrestrial variability: at what scale do ecologists

communicate? *Mar. Ecol. Prog. Ser.*, 304, 274–280.

Hayford, H.A., O'Donnell, M.J. & Carrington, E. (2018). Radio tracking detects behavioral

thermoregulation at a snail's pace. *J. Exp. Mar. Biol. Ecol.*, 499, 17–25.

Inouye, D.W. (2008). Effects of Climate Change on Phenology, Frost Damage, and Floral

Abundance of Montane Wildflowers. *Ecology*, 89, 353–362.

Jensen, J.L.W.V. (1906). Sur les fonctions convexes et les inégalités entre les valeurs moyennes.

Acta Math., 30, 175–193.

Keyfitz, N. (1968). *Introduction to the Mathematics of Population*.

Lande, R., Engen, P. of B.S., Engen, S., Sæther, B.-E. & Saether, P. of P.E.B.-E. (2003).

Stochastic Population Dynamics in Ecology and Conservation. Oxford University Press.

Lande, R., Engen, S. & Sæther, B.-E. (2017). Evolution of stochastic demography with life

history tradeoffs in density-dependent age-structured populations. *Proc. Natl. Acad. Sci.*,

114, 11582–11590.

Lande, R. & Orzack, S.H. (1988). Extinction dynamics of age-structured populations in a

fluctuating environment. *Proc. Natl. Acad. Sci.*, 85, 7418–7421.

Lawson, C.R., Vindenes, Y., Bailey, L. & van de Pol, M. (2015). Environmental variation and

population responses to global change. *Ecol. Lett.*, 18, 724–736.

Levins, R. (1968). Evolution in changing environments Princeton University Press. *Princet. N. J.*

Linderholm, H.W. (2006). Growing season changes in the last century. *Agric. For. Meteorol.*,

137, 1–14.

Lytle, D.A. (2001). Disturbance Regimes and Life-History Evolution. *Am. Nat.*, 157, 525–536.

Lytle, D.A., Bogan, M.T. & Finn, D.S. (2008). Evolution of aquatic insect behaviours across a

gradient of disturbance predictability. *Proc. R. Soc. B Biol. Sci.*, 275, 453–462.

MacArthur, R.H. (1962). Some generalized theorems of natural selection. *Proc. Natl. Acad. Sci.*

U. S. A., 48, 1893.

Marlon, J.R., Bartlein, P.J., Walsh, M.K., Harrison, S.P., Brown, K.J., Edwards, M.E., *et al.*

(2009). Wildfire responses to abrupt climate change in North America. *Proc. Natl. Acad.*

Sci., 106, 2519–2524.

Metcalf, C. j. e & Koons, D. n. (2007). Environmental uncertainty, autocorrelation and the

evolution of survival. *Proc. R. Soc. B Biol. Sci.*, 274, 2153–2160.

Morris, W.F., Pfister, C.A., Tuljapurkar, S., Haridas, C.V., Boggs, C.L., Boyce, M.S., *et al.*

(2008). Longevity Can Buffer Plant and Animal Populations Against Changing Climatic

Variability. *Ecology*, 89, 19–25.

Morris, W.F., Tuljapurkar, S., Haridas, C.V., Menges, E.S., Horvitz, C.C. & Pfister, C.A. (2006).

Sensitivity of the population growth rate to demographic variability within and between

phases of the disturbance cycle. *Ecol. Lett.*, 9, 1331–1341.

Nevoux, M., Barbraud, J.-C. & Barbraud, C. (2008). Nonlinear impact of climate on survival in a

migratory white stork population. *J. Anim. Ecol.*, 77, 1143–1152.

Noble, J.D., Collins, S.L., Hallmark, A.J., Maldonado, K., Wolf, B.O. & Newsome, S.D. (2019).

Foraging strategies of individual silky pocket mice over a boom–bust cycle in a

stochastic dryland ecosystem. *Oecologia*, 190, 569–578.

Nouvellet, P., Rasmussen, G.S.A., Macdonald, D.W. & Courchamp, F. (2012). Noisy clocks and

silent sunrises: measurement methods of daily activity pattern. *J. Zool.*, 286, 179–184.

Park, J.S. (2019). Cyclical environments drive variation in life-history strategies: a general

theory of cyclical phenology. *Proc. R. Soc. B*, 286, 20190214.

Park, J.S. & Wootton, J.T. (2021). Slower environmental cycles maintain greater life-history

variation within populations. *Ecol. Lett.*, 24, 2452–2463.

Pélisson, P.-F., Bel-Venner, M.-C., Rey, B., Burgevin, L., Martineau, F., Fourel, F., *et al.* (2012).

Contrasted breeding strategies in four sympatric sibling insect species: when a

proovigenic and capital breeder copes with a stochastic environment. *Funct. Ecol.*, 26,

198–206.

Pike, N., Tully, T., Haccou, P. & Ferrière, R. (2004). The effect of autocorrelation in

environmental variability on the persistence of populations: an experimental test. *Proc. R.*

Soc. Lond. B Biol. Sci., 271, 2143–2148.

Post, E. & Stenseth, N.Chr. (1999). Climatic Variability, Plant Phenology, and Northern

Ungulates. *Ecology*, 80, 1322–1339.

Priestley, M.B. (1981). *Spectral analysis and time series: probability and mathematical*

statistics. Academic Press.

Pyper, B.J. & Peterman, R.M. (1998). Comparison of methods to account for autocorrelation in

correlation analyses of fish data. *Can. J. Fish. Aquat. Sci.*, 55, 2127–2140.

Reuman, D.C., Costantino, R.F., Desharnais, R.A. & Cohen, J.E. (2008). Colour of

environmental noise affects the nonlinear dynamics of cycling, stage-structured

populations. *Ecol. Lett.*, 11, 820–830.

Ruel, J.J. & Ayres, M.P. (1999). Jensen's inequality predicts effects of environmental variation.

Trends Ecol. Evol., 14, 361–366.

Sabo, J.L. & Post, D.M. (2008). Quantifying Periodic, Stochastic, and Catastrophic

Environmental Variation. *Ecol. Monogr.*, 78, 19–40.

Schaffer, W.M. (1974). Optimal Reproductive Effort in Fluctuating Environments. *Am. Nat.*,

108, 783–790.

Schreiber, S.J. & Moore, J.L. (2018). The structured demography of open populations in

fluctuating environments. *Methods Ecol. Evol.*, 9, 1569–1580.

Shriver, R.K. (2016). Quantifying how short-term environmental variation leads to long-term

demographic responses to climate change. *J. Ecol.*, 104, 65–78.

Sletvold, N., Dahlgren, J.P., Øien, D.-I., Moen, A. & Ehrlén, J. (2013). Climate warming alters

effects of management on population viability of threatened species: results from a 30-

year experimental study on a rare orchid. *Glob. Change Biol.*, 19, 2729–2738.

Stearns, S.C. (1976). Life-History Tactics: A Review of the Ideas. *Q. Rev. Biol.*, 51, 3–47.

Steele, J.H. (1985). A comparison of terrestrial and marine ecological systems. *Nature*, 313,

355–358.

Stenseth, N.Chr., Chan, K.-S., Tavecchia, G., Coulson, T., Mysterud, A., Clutton-Brock, T., *et*

al. (2004). Modelling non–additive and nonlinear signals from climatic noise in

ecological time series: Soay sheep as an example. *Proc. R. Soc. Lond. B Biol. Sci.*, 271,

1985–1993.

Taylor, C., McCarthy, M.A. & Lindenmayer, D.B. (2014). Nonlinear Effects of Stand Age on

Fire Severity. *Conserv. Lett.*, 7, 355–370.

Tuljapurkar, S. (1985). Population dynamics in variable environments. VI. Cyclical

environments. *Theor. Popul. Biol.*, 28, 1–17.

Tuljapurkar, S. (2013). *Population Dynamics in Variable Environments*. Springer Science &

Business Media.

Tuljapurkar, S., Gaillard, J.-M. & Coulson, T. (2009). From stochastic environments to life

histories and back. *Philos. Trans. R. Soc. B Biol. Sci.*, 364, 1499–1509.

Tuljapurkar, S. & Haridas, C.V. (2006). Temporal autocorrelation and stochastic population

growth. *Ecol. Lett.*, 9, 327–337.

Tuljapurkar, S., Horvitz, C.C. & Pascarella, J.B. (2003). The Many Growth Rates and Elasticities

of Populations in Random Environments. *Am. Nat.*, 162, 489–502.

Tuljapurkar, S.D. (1982). Population dynamics in variable environments. III. Evolutionary

dynamics of r-selection. *Theor. Popul. Biol.*, 21, 141–165.

Vasseur, D.A. & Yodzis, P. (2004). The Color of Environmental Noise. *Ecology*, 85, 1146–1152.

Venable, D.L. (2007). Bet Hedging in a Guild of Desert Annuals. *Ecology*, 88, 1086–1090.

Walther, G.-R., Post, E., Convey, P., Menzel, A., Parmesan, C., Beebee, T.J.C., *et al.* (2002).

Ecological responses to recent climate change. *Nature*, 416, 389–395.

Westerling, A.L., Hidalgo, H.G., Cayan, D.R. & Swetnam, T.W. (2006). Warming and Earlier

Spring Increase Western U.S. Forest Wildfire Activity. *Science*, 313, 940–943.