From the ear flap to the nerve: A complete computer model of the peripheral auditory system

Ondrej Tichacek[1], Pavel Mistrík[2], and Pavel Jungwirth[1]

[1] *Institute of Organic Chemistry and Biochemistry of the Czech Academy of Sciences, Flemingovo náměstí 542/2, 160 00 Praha 6, Czech Republic*
[2] *MED-EL, Fürstenweg 77a, A-6020 Innsbruck, Austria*

July 22, 2022

Abstract

Computer models of the individual components of the peripheral auditory system – the outer, middle, and inner ears and the auditory nerve – have been developed in the past, with varying level of detail, breadth, and faithfulness of the underlying parameters. Here, we move modeling of the ear to the next level by presenting a complete, physiologically justified, bottom-up computer model based on up-to-date experimental data that integrates all these parts together in a seamless way. Such a model is instrumental for advancing quantitative understanding of the mechanism of hearing, as well as for devising next generation hearing aids and cochlear implants.

Proper understanding of the mechanism of hearing and especially the cochlear function is important for the advancement of the compensation of hearing loss, including the design and setup of hearing aids and cochlear implants. While some psychophysical properties of the human cochlea have been thoroughly studied and a sufficient amount of data is available (Robles and Ruggero 2001), others are hardly accessible due to the invasive nature of the measurements. In these cases, data may be obtained, at least to a certain extent, from animal models. Importantly, computer modeling can provide insight to particular problems, where experiment is not feasible, and help to establish a detailed quantitative understanding of the system as a whole.

The mammalian peripheral auditory system consists of the outer, middle, and inner ears (the latter including the cochlea) and the auditory nerve leading to the cochlear nucleus, where in turn the central auditory nervous system (CNS) starts. Its main function is to detect the sound and convert it to neural activity to be processed by the CNS. Considering the immense range of sound frequencies of 20 Hz up to 20 kHz and intensities from 0 to ∼100 dB sound pressure level (SPL), the function of the human auditory periphery is enabled by highly specialized multi-cellular and intracellular structures, often unique to the auditory system, that are responsible for key mechanisms such as mechanosensitivity, amplification, and adaptation. Besides that, there are many other processes on the peripheral auditory pathway that enable the mechanism of hearing (see Fig. 1), all of which need to be included in the computer model in order to obtain an accurate description of the physiological reality.

Computer modelling of the auditory periphery has been a busy field in the past decades, see for instance (Bruce 2006; Heil et al. 2011; Mammano and Nobili 1993; Meddis 1986; Mistrík, Mullaley,

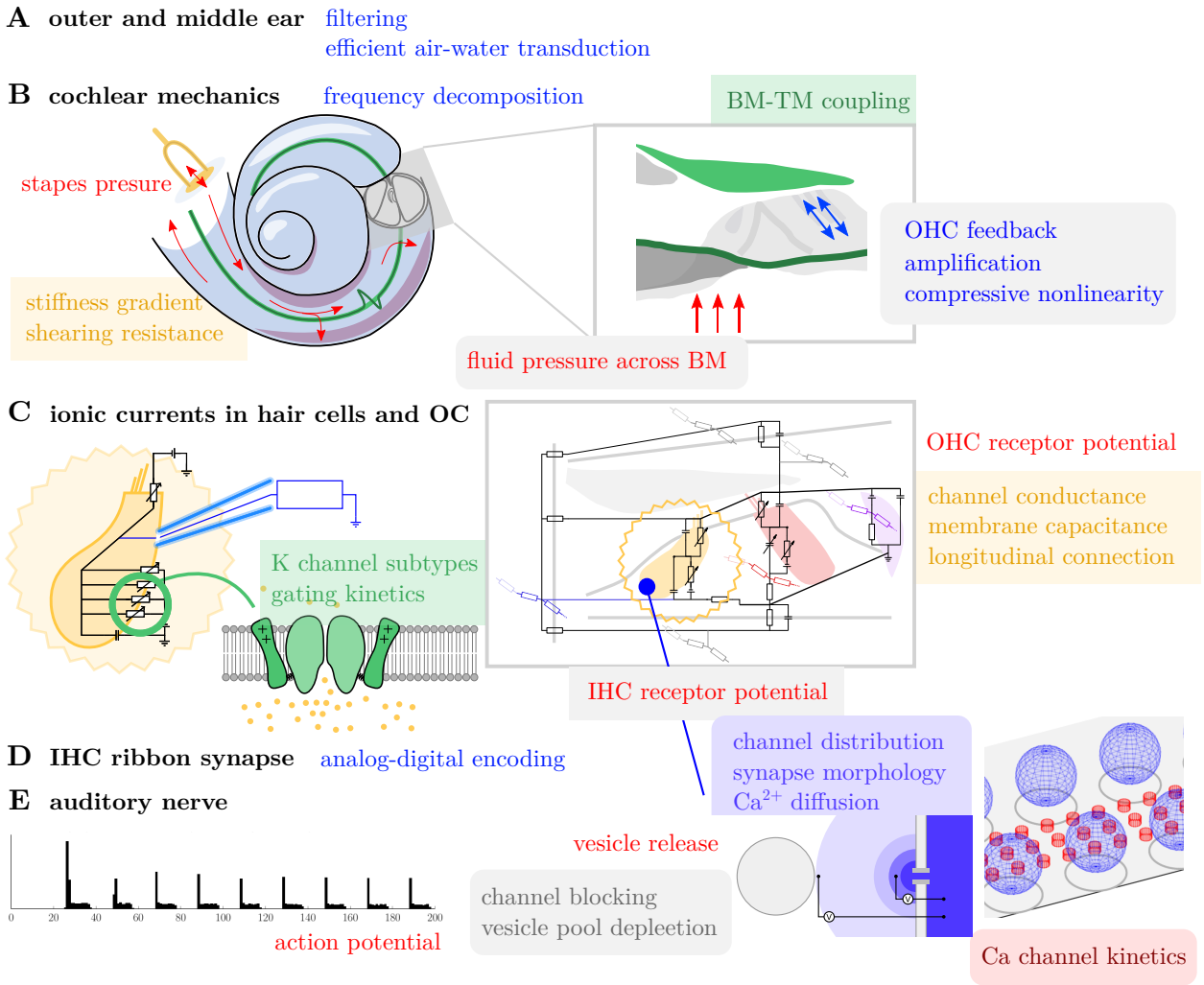

Figure 1: Model blocks and the mechanism of hearing. **A**: Acoustic waves pass through the outer ear and middle ear, which effectively conveys the vibration from air to the cochlear fluids. **B**: Pressure oscillations in the cochlear fluids are spatially decomposed across the basilar membrane (BM), coupled to the tectorial membrane (TM) depending on their frequency in the first step to sound encoding. **C**: Motion of the organ of Corti (OC) and subsequent deflection of the stereocilia in the sensory cells opens ion channels and induces transduction of ionic current. **D**: Change of the receptor potential in the inner hair cells (IHC) triggers release of synaptic vesicles and thus evoked action potential in the afferent nerve fibers finalizes the passage from analog to "digital" signal (**E**). In the outer hair cells (OHC), the receptor potential causes conformation change of a voltage-sensitive protein prestin which translates to macroscopic contraction and elongation of the cell and a consequent force acting on the organ of Corti. When in phase, this force can act as an amplifier of the cochlear mechanics, being responsible for a sharper tuning and increased level detection range through the compressive nonlinearity.

et al. 2008; Pascal et al. 1998; Strelioff 1973; Sumner et al. 2002; Verhulst, Altoè, et al. 2018; Verhulst, Dau, et al. 2012; Vetešník and Gummer 2012). Most of these models, however, focus only on individual parts of the auditory periphery and mostly have not been updated to reflect the most recent experimental findings. Here, we have overcome these limitations in two ways: First, by creating a framework of models, each of which being more advanced than its predecessor in the literature and, second, by organically connecting these models into a single system. Tackling computationally expensive tasks by efficient implementation has allowed us to avoid compromises concerning the level of description of the individual components of the peripheral auditory system.

In this paper, we thus present a physiologically well-justified computational model, with its core being a complete cochlear partitioning with a detailed description of electrical excitability of individual hair cells. We couple it with models of outer and middle ears, a model of the cochlear mechanics, a conceptually novel model of the ribbon synapse of the inner hair cell, and with a model of the auditory nerve. The present complete model of the peripheral auditory system is constructed in a "bottom-up" way, based on the appropriate laws of physics and physiologically relevant measurable properties of its building blocks, avoiding thus "black-box" heuristic approaches. Where human data are not available, animal data have been used instead, coming exclusively from mammalian species such as the guinea pig.

Results

From the ear flap to the nerve, the sound waves are conveyed, processed, and encoded by several mutually interconnected and cooperating biological systems. The present model is designed in the same way – as an aggregate of coupled models of the outer ear, middle ear, the cochlear mechanics, the ionic currents within the cochlea and the hair cells, the ribbon synapse, and the auditory nerve. In the following sections, key novel aspects of the individual models and their interconnections are described, while an exhaustive description is provided in the methods section and in the supplementary information.

Acoustic transduction through outer and middle ears

The outer ear transfer characteristics are implemented in the present model via a set of independent band-pass filters as proposed by Meddis (2011) and calibrated to experimental data of Mehrgardt and Mellert (1977), see Fig. 2A. Following Pascal et al. (1998) the middle ear is represented as a lumped-element model of selected structures — namely the eardrum, the middle ear cavities, malleus, incus, stapes, and the cochlea. Here, we recalibrated the model to recent experimental tympanic membrane pressure-stapes velocity data by Chien et al. (2009), as demonstrated in Fig. 2B.

The model can closely reproduce the experimental data up to ∼10 kHz (outer ear) and ∼8 kHz (middle ear). For higher frequencies, individual experimental datasets are mutually less consistent or even missing, which complicates accurate validation of the model in the very high frequency range.

Cochlear mechanics

The first step of sound encoding occurs as the basilar membrane spatially decomposes the sound wave based on its frequency components. This is enabled primarily by the gradual change in its stiffness and other mechanical properties along the tonotopy axis, i.e., from the base to the apex of the basilar membrane.

Here we employ a model based on that by Vetešník and Gummer (2012). A major step forward in the present work is an advanced description of the cochlear amplifier employing a direct feedback from the subsequent model of the electric currents within the cochlea (see below). The amplification with respect to a passive model can reach (depending on the stimulus frequency) ∼45 dB SPL (Fig. 2D), while maintaining the zero crossing condition (Supplementary Fig. S-1). The compression ratio of the input/output curves of the BM displacement (Fig. 2F) is comparable with psychoacoustic experimental data (Plack and Oxenham 2000). We have also reparametrized the model to reproduce

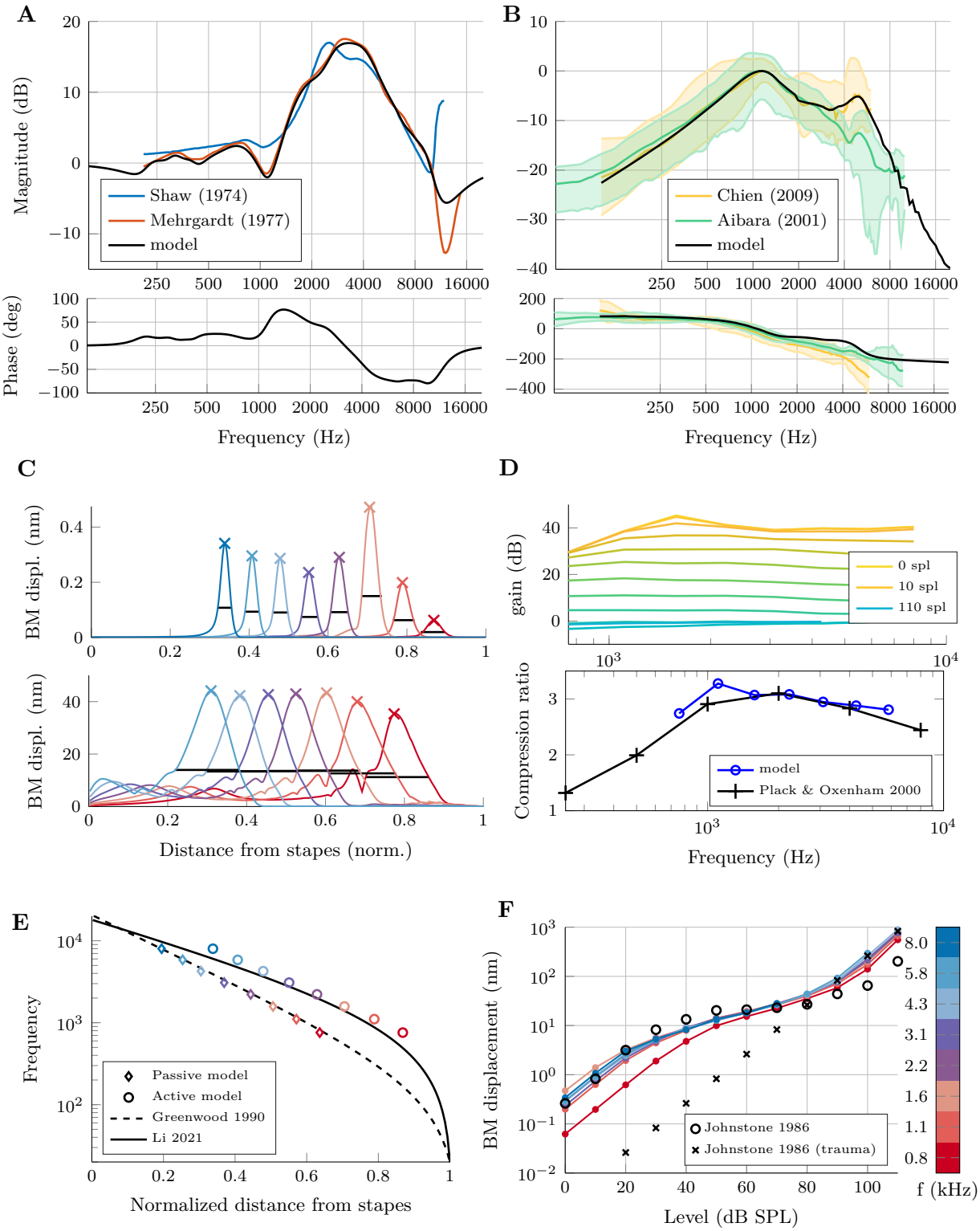

Figure 2: Mechanics of the outer and middle ears, and the cochlea: A: Pressure-pressure transfer function of the outer ear model compared to experimental data by Shaw (1974), and Mehrgardt and Mellert (1977). Experimental phase shifts are not available. **B**: Pressure-velocity transfer function of the middle ear model with experimental data by Chien et al. (2009) and Aibara et al. (2001). **C-F**: Basilar membrane response as calculated by the model; **C**: Traveling wave envelopes for of BM oscillations for pure tones of 0.8 to 8 kHz at 0 dB SPL (top) and 80 dB SPL (bottom). **D**: Measured relative gain of the active vs. passive model (top) and the mean compression ratio between 50 an 80 dB SPL (bottom) compared to psychoacoustic experimental data by Plack and Oxenham (2000). **E**: characteristic frequency (CF) as a function of the position along the BM as reproduced by the model (in both active and passive modes) compared to experimentally based CF curves by Greenwood (1990) and Li et al. (2021). **F**: Input-output curves of BM displacement vs. sound level for range of frequencies from 0.8 to 8 kHz compared to experimental data by Johnstone et al. (1986).

the human tonotopic maps Greenwood (1990) and Li et al. (2021) (Fig. 2E), which is crucial for correct assessment of tonotopy specialization of hair cells.

Cochlea as an electrical circuit

In this section, we provide the description of the cochlear model in terms of a large-scale electrical circuit. In particular, we summarize the updates to the circuit design compared to earlier models in the literature, focusing on the present advanced description of the hair cells. These improvements concern primarily the implementation of (i) the conductance of the stereocilia through the mechanosensitive channels, (ii) the voltage-dependent conductance of the basolateral membranes dominated by the potassium channels, (iii) potassium channel sub-types with different gating kinetics, and (iv) the non-linear part of the capacitance of the cell membranes of the outer hair cell (OHC).

The modelling approach adopted here follows a general concept of a lumped three-dimensional model, first introduced by Strelioff (1973) for the analysis of the intracochlear distribution of electrical potentials. Such approach was used later also for modelling of a cochlea implanted with a multielectrode stimulating array (Suesserman and Spelman 1993) and of a three-dimensional current flow and its effect on the amplification of sound (Mistrík, Mullaley, et al. 2008), as well as for modelling of reduced OHC electromotility associated with connexin-related forms of deafness (Mistrík and Ashmore 2010).

In the present model, the cochlea is approximated by interconnected electrical circuits representing the radial flow of an electric current at each cross-section, as well as the longitudinal current flow between cross-sections. Compared to previous models (Mistrík and Ashmore 2010), the number of longitudinally connected cross-sections was increased from 300 to 3000, corresponding to a typical number of inner hair cells (IHCs) in the human cochlea (the number of OHCs being about four times larger). In this way, each cross-section accounts for the radial current flow through each row of hair cells in the human organ of Corti. An equivalent electrical circuit of a single cross-section of the cochlear partition is presented in Figure 3A. Given experimentally accessible input parameters, such as the characteristic conductances and capacitances of the cells of interest and electrical/electrochemical potentials, the model provides currents and transmembrane potentials at each part of the system (Supplementary Fig. S-3). Its principal advantages are revealed upon coupling with a model of cochlear mechanics that simulates the motion of the basilar membrane and the stereocilia deflection. Namely, these motions can be directly translated into conductance changes of the hair cell stereocilia and, consequently, into evoked currents and potentials throughout the system. In this way, receptor potentials in the hair cells, generated in response to arbitrary sound stimuli, can be realistically estimated (Figures 3B-C, 4D, Supplementary Figures S-2, S-3, and S-4).

Parametrization of the electrical properties was adopted from Strelioff (1973) regarding the conductivity of fluids in the scala vestibuli (SV), scala media (SM), scala tympani (ST), and spiral limbus (SL) and from Mistrík, Mullaley, et al. (2008) concerning the organ of Corti (OC), being based on resistive and capacitive properties of the cellular constituents. The numerical values of electric elements were updated to reflect the latest experimental data (Johnson 2015; Johnson, Beurg, et al. 2011), particularly for the OHC and IHC specialization along the tonotopic axis. The calibration was aided by a supplementary model of an isolated hair cell, reproducing in-vitro electrophysiological experiments (Fig. 3E-J). Such an approach is necessary as experiments with

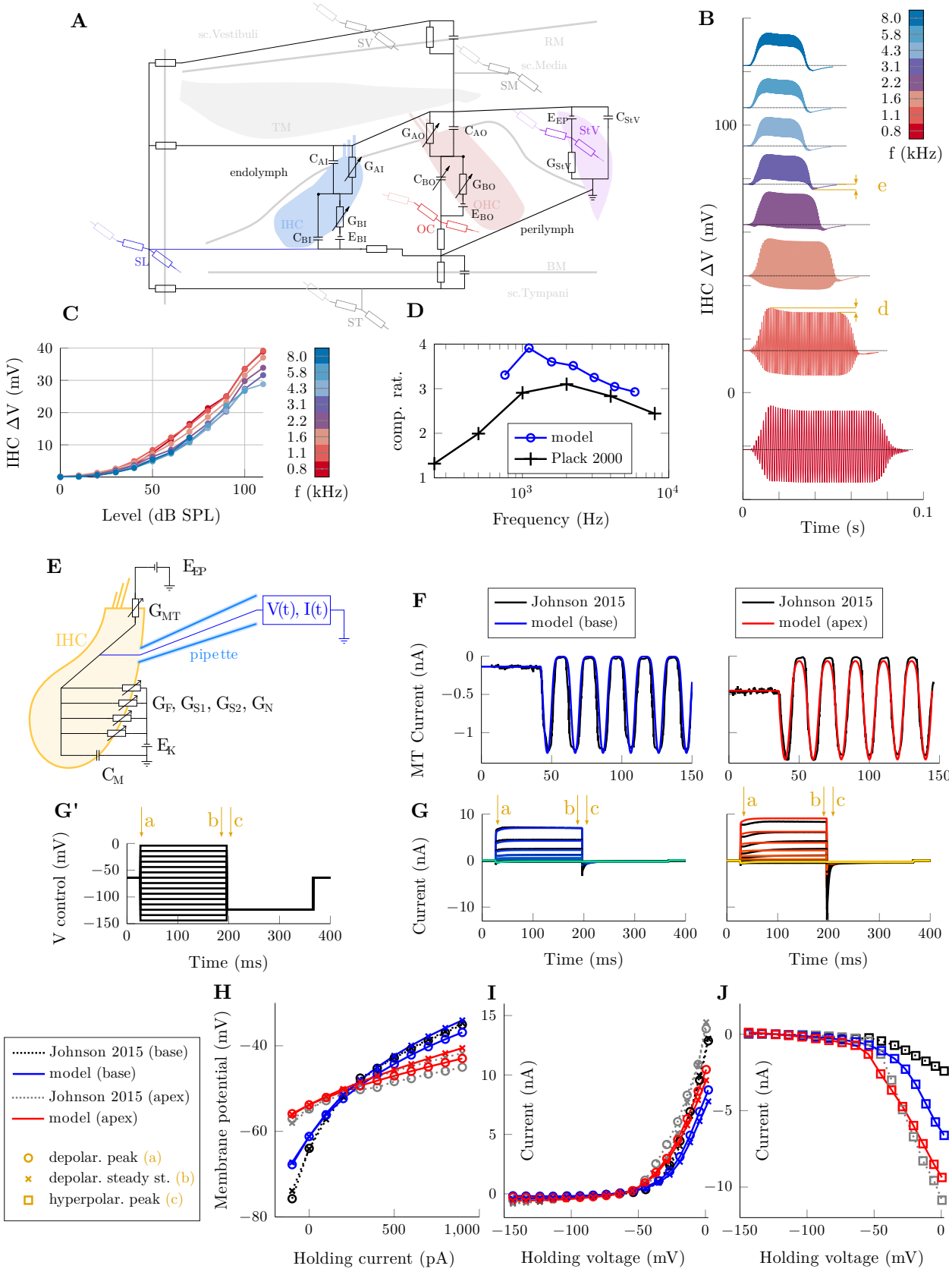

Figure 3: Ionic currents within the cochlea: A: An equivalent electrical circuit of a cochlear cross section. Electrical properties of the apical and basolateral parts of inner and outer hair cell membranes are described by the corresponding resistors (conductance G) and capacitors (C). The reversal potential arising from the difference between intracellular and extracellular potassium concentrations is represented by a voltage source (battery). Each IHC is accounted for individually, while four OHCs in a cross section are represented by one OHC with scaled values of their G and C elements. Stria vascularis (StV) is represented by a corresponding resistor and capacitor, and the presence of the endocochlear potential at scala media

(Caption continues next page.)

Figure 3: *(Continued from previous page.)* (SM) by a battery. Other cellular structures in the organ of Corti and spiral limbus (SL) are represented by resistors. The Reisner's and basilar membranes separating the scala vestibuli (SV), media, and tympani (ST) are represented by a resistor and a capacitor. Each of the three scalae, the spiral limbus, organ of Corti (OC) and the stria vascularis are longitudinally connected to other cross sections via resistors. Some of the elements are variable; the resistors G_{AI} and G_{AO} are modulated by the stereocilia deflection, G_{BI} and G_{BO} are regulated by voltage and the kinetics of the ion channels they represent, and the value of C_{BO} includes the nonlinear capacitance, which is voltage dependent. Moreover, the hair cell basolateral resistors are composed of four independent resistors representing different subtypes of expressed potassium channels, see panel E.

B-D: IHC receptor potential (ΔV) as computed by the full 3D-circuit model in response to pure tones of varying frequency and level: **B**: Waveforms of IHC ΔV at the maximal magnitude cross-section at 70 dB SPL exhibit expected properties such as AC/DC ratio decrease with stimulus frequency. The peak value occurring shortly after the stimulus onset is larger than the steady-state maximum (indicated by "d" in the figure) and similarly, the potential short after the stimulus end is hyperpolarized compared to the resting state (indicated by "e"). Both of these effects are caused by the non-instantaneous response of the channels to the change of the membrane potential (i.e., the channel kinetics). See supplementary figure S-4 for more details on the effect of the stimulus onset. **C**: Input-Output curves of the peak IHC receptor potential exhibit a compressive nonlinearity similarly to the BM displacement in the 40 to 80 dB SPL range. Another compression occurs for high sound levels due to the displacement-induced conductance saturation of the mechanosensitive ion channels in the stereocilia tips. Consequently, as can be seen from the figure, the receptor potential is almost linear with respect to the the sound level in dB SPL from about 40 dB. The receptor potential is diminished for higher frequencies due to a low pass filtering of the RC circuit (i.e., the membrane plus channel system), see also panel C. **D**: The mean compression ratio between 50 an 80 dB SPL compared to psychoacoustic experimental data by Plack and Oxenham (2000), is increased compared to the BM displacement (Fig. 2).

E-J: Model of an isolated hair cell. **E**: Diagram of the model in the whole-cell patch-clamp configuration. **F**: The mechano-electrical transduction (MET) current elicited by stereocilia displacement in the isolated cell model configuration compared to experimental data measured by Johnson (2015) with stereocilia displaced using a fluid jet. **G**: Example of a voltage-clamp protocol (depolarizing and hyperpolarizing voltage steps of different magnitude (G')), simulation and experiment by Johnson (2015). The arrows **a**, **b**, **c** denote features of the response to the current clamp (**H**) and voltage clamp (**I**, **J**) protocols used by (Johnson 2015) to differentiate between basal and apical cells. **E**: Peak (a) and steady state (b) voltage in response to a step in control current. **F**: Peak (a) and steady state (b) current in response to voltage step. **G**: Amplitude of tail currents (c) in response to a hyperpolarizing step.

isolated cell cannot be correctly reproduced solely by a model of the whole OC, and similarly, in-vivo characteristics of hair cells (e.g. receptor potentials) are affected by other sub-strcutures of OC and thus cannot be reliably estimated with models of isolated hair cells.

The non-linearity and voltage-dependence of membrane resistances due to the presence of voltage-gated ion channels was implemented together with the potassium channel gating kinetics that accounts for a non-instantaneous response of the population of the channels. Channel kinetics were modeled similarly as in Lopez-Poveda and Eustaquio-Martín (2006), but with the model being extended to include different potassium channel subtypes ($\mathrm{K_f}$, $\mathrm{K_n}$, and multiple variants of $\mathrm{K_s}$, their distributions varying along the tonotopy axis) and calibrated to match the recent experimental data (Johnson 2015), see Fig. 3F-J. The nonlinear capacitance of the OHC basolateral membrane, which is critical for the amplification mechanism driven by the cochlear electromotor prestin, (Dallos and Fakler 2002; Santos-Sacchi and Tan 2020) was also implemented (see Methods).

Due to the non-instantaneous hair cell channel kinetics as implemented by our model the stimulus envelope (e.g. the slope of the onset) affects the later response – a short (steep) onset causes temporarily higher depolarization magnitude than a long onset. A short vs. long offset of a stimulus has a similar effect; see Fig. 3B and Supplementary Fig. S-4. The effect mimics adaptation of the neural response caused by stockpiling/depletion of the readily-releasable vesicles by the ribbon synapse. Since depolarization eventually triggers synaptic release events while hyperpolarization attenuates them, the hair cell channel kinetics can be partially responsible for the observed adaptation.

Mechano-electrical transduction

Deflection of the IHC and OHC stereocilia opens MET channels and increases the instantaneous conductance of the hair cells. Analogously, in the present model, the electrical circuit is modulated by the motion of the basilar and tectorial membranes. In contrast to Mistrík, Mullaley, et al. (2008) where OHCs are coupled to the BM displacement, in the present model behavior OHCs is governed by explicitly calculated deflections of stereocilia which is physiologically more realistic. The MET transduction parameters have been updated to reflect experimental data by Jia et al. (2007) and Johnson (2015).

Cochlear amplifier

Nobili and Mammano (1996) investigated the nonlinear cochlear mechanics and introduced an "active amplifier" term in their model. This term was theoretically derived from the response of prestin to the OHC receptor potential, but the potential itself was not explicitly calculated. Instead, the active force term was made proportional to the stereocilia deflection, saturating with increasing intensity, and calibrated so that it mostly cancels the shearing resistance. The same approach was adopted also in the later model by Vetešník and Gummer (2012).

In order to make the model more realistic and to simplify it from a theoretical point of view, we replaced the indirect estimation of the OHC receptor potential by an exact computation from the electrical model of cochlear partition. The main advantage of this approach is the direct inclusion of effects produced by the present complex and fine-grained electrical model, in particular, the voltage sensitivity and dynamics of the ion channels, tonotopic gradients in OHC conductance and capacitance, as well as the nonlinear OHC capacitance. Consequently, there are three main sources of nonlinear behaviour in the present amplifier model – the active force as a function of the receptor potential, the voltage-dependent conductance of basolateral K^+ channels, and the MET conductance governed by the stereocilia deflection.

Increasing progressively the amplification level of the model allowed us to investigate the amplification mechanism (see Fig. 4A-C). For amplification to occur, the force has to be in phase with the BM velocity. Due to the phase roll-off of the motion of BM, this happens at a slightly different tonotopic location than what would correspond to the characteristic frequency of the passive vibration. When the BM velocity and the amplification force are in opposite phases, attenuation occurs instead of amplification, which leads to sharpening of the response (Fig. 4A vs. B and also Fig. 2C). While in the original model by Vetešník and Gummer (2012) the amplification force is by definition in phase with the stereocilia deflection, in the present model the membrane capacitance together with the kinetics of the basolateral K^+ channels induce a frequency and amplitude-dependent phase lag. Consequently, the tonotopic map of the active model is shifted

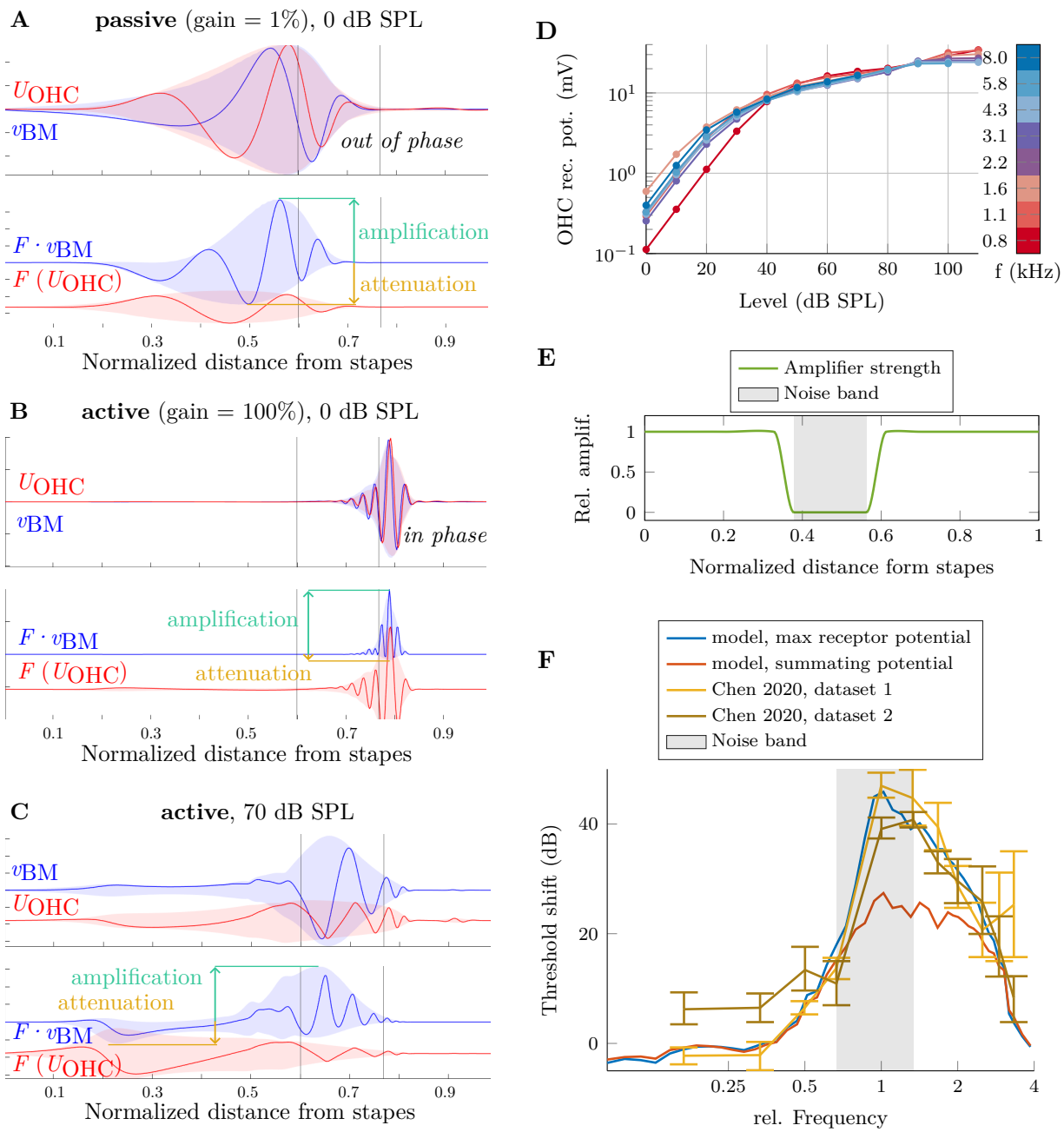

Figure 4: The mechanism and effect of the cochlear amplifier. A-C: The OHC receptor potential U_{OHC}, the BM velocity v_{BM}, the amplification force F and the amplification measure, calculated as $F \cdot v_{\mathrm{BM}}$, all along the normalized distance from the stapes. The curves show a value at a single time point, with colored areas showing the min-max envelope throughout the simulation. All three panels show a simulation of pure tone of 1 kHz, at 0 dB SPL for (A) passive and (B) active cochlea and at 70 dB SPL for an active cochlea (C). With increasing gain, the characteristic position for a given frequency moves towards the apex, where the BM velocity and the amplifier force are correctly in phase for amplification to occur. **D**: The magnitude of the OHC receptor potential, crucial for proper function of the cochlear amplifier, is not dramatically reduced for high frequencies, as might be expected due to the low-pass filtering properties of OHC cell membrane. Instead OHC specialization along the tonotopy axis largely cancels the filtering effect, rendering the OHC receptor potential magnitude similar across frequencies. **E-F**: Modelling noise induced hearing loss – effect of the cochlear amplifier: We have exposed a model cochlea to a 1.6–3.2 kHz single octave 95 dB SPL band-noise, measuring the extent of BM oscillations for band-edge frequencies, thus determining the damage extent along the tonotopy axis, resulting in a locally reduced function of the cochlear amplifier (E). Then we performed simulations of 0.2–9.3 kHz pure tones in both normal and damaged cochlei, recording the IHC receptor potential. The figure (F) shows a comparison of the peak IHC receptor potential and estimated summating potential "damaged-to-control shift" to the experimental CAP threshold shift in mice by Chen et al. (2020) normalized to the central frequency of the noise band.

towards the apex compared both to a passive model and to a model with implicitly calculated OHC potential, see Fig. 2C,E and 4B vs. C, which can explain the shift in experimental tonotopic maps (Greenwood 1990; Li et al. 2021).

We modelled also the amplification in a cochlea with locally impaired amplification ability. Aiming to reproduce experimental measurements of Chen et al. (2020), who measured compound action potentials (CAP) in mice after long-term exposure to high-level octave band noises leading to severe noise-induced hearing loss, we selectively turned off the OHC amplifier and measured the IHC response in terms of the receptor potential. It turns out, that the experimental CAP threshold shift can largely be explained in our model by damage to the OHC only (IHCs being unaffected), including the spread of threshold shift outside the noise band; see Fig. 4E-F.

Response to soft sounds

Sounds near the hearing threshold inflict a peak displacement of the BM in the sub-nanometer level and the consequential MET probability change is very small correspondingly (0.7 % for 1 nm displacement). Considering the number of MET channels in a hair cell (∼300 based on the single channel conductance, Beurg et al. (2021) and Géléoc et al. (1997)), using the deterministic approach traditionally employed in models of hair cells (Dierich et al. 2020; Lopez-Poveda and Eustaquio-Martín 2006; Mistrík and Ashmore 2010), soft sounds would evoke an opening of less than one channel. In reality, the channels open stochastically and only their mean open count can be considered as being continuous. While the approximation coming from the continuous representation of the conductance is negligible for high-level stimuli, it can play a significant role for low-level ones. We demonstrate here that in such cases in reality at certain random time instances the IHC depolarize significantly more than in a classical deterministic model, while at other instances it does not depolarize at all. On average, however, the mean MET open probability/conductance as well as the value of the receptor potential are identical. Considering that some of the elemental processes of the IHC signal transduction ($Ca_V1.3$ open probability, vesicle release calcium threshold from the Ribbon synapse, etc.) are nonlinear in nature (Beutner et al. 2001; Zampini, Johnson, Franz, Lawrence, et al. 2010), this stochastic effect may be significant for the detection of signals close to the hearing threshold. It may also be the first instance, when the level is (partially) encoded as rate, such as is the case in the auditory nerve. We have implemented this phenomenon into the preprocessing of the IHC synapse model by solving again an isolated IHC circuit, while including the stochastic nature of the channels. Sample results can be seen in the Supplementary Figure S-5.

Ribbon synapse

Excitation of the afferent type I auditory nerve fibers is governed by several consecutive steps – change of the IHC receptor potential, opening of $Ca_V1.3$ channels in the IHC basolateral membrane, increase in the local Ca^{2+} concentration in the vicinity of the synapse, and ultimately a neurotransmitter release from fusing vesicles into the synaptic cleft.

The ribbon synapse holds synaptic vesicles close to the active zone, encoding the stimulus level in the rate of neurotransmitter release. The $Ca_V1.3$ ion channels are expressed nearby the ribbon synapse, indicating a strong local effect of each channel. During the maturation period, channels are found progressively closer to the ribbon synapse (P. F. Vincent et al. 2018), suggesting that precise encoding requires co-localization of the ion channels and the ribbon synapse. It has been shown that the ribbon synapse itself can vary in size and shape (Moser et al. 2006), and it is thought

that the morphology of the ribbon synapse plays an important role for its function (L.-Y. Wang and Augustine 2015). Specialization of the ribbon synapse and their long-term maturation has been confirmed recently by Niwa et al. (2021), who resolved differences in EPSC properties in pillar vs modiolar subgroups. Since the structure of the synapse can be experimentally resolved using transmission electron microscopy and super-resolution light microscopy (STED), see, e.g., Rutherford (2015), we designed the present model to reflect the spatial structure of the ribbon synapse.

The model is defined by positions of readily releasable vesicles and of individual $Ca_V1.3$ channels. While the vesicle positions are experimentally directly accessible, resolving individual ion channels is currently not possible. However, the localization of ion channels with respect to the synaptic ribbon can already be measured (P. F. Vincent et al. 2018), and thus the function of the synapse may be studied with respect to variables like mean channel-vesicle distance. An example of a possible configuration of the ribbon synapse model can be seen in the Fig. 5C-E.

The voltage sensitive $Ca_V1.3$ channels play an important role in regulating the vesicle exocytosis and have been, therefore, the topic of intensive research. The crucial voltage-dependent steady state open probability has been reported repeatedly (Johnson and Marcotti 2008; Ohn et al. 2016; Zampini, Johnson, Franz, Knipper, et al. 2014; Zampini, Johnson, Franz, Lawrence, et al. 2010). While the results differ in absolute values of the curves (see Fig. M-3), they all agree that the open probability is very low close to the resting membrane potential of the IHC. Considering the total number of channels per active zone (80–600), it is clear that a release of a synaptic vesicle can be triggered by the opening of one or only a few channels (e.g. Magistretti et al. (2015)). Thus, compared to other models, each channel is explicitly calculated here.

Elementary properties of isolated $Ca_V1.3$ channels have been investigated using patch-clamp techniques (Zampini, Johnson, Franz, Knipper, et al. 2014), yielding not only the voltage-dependent steady state open probabilities, but also the mean open/close durations. This allows us to express the open probability of the channel as a transfer rate within a two state Markov model.

Finally, one should take into account that that the channel activity can be temporarily blocked. Recent experimental findings indicate that this may be caused by protons released into the synaptic cleft alongside the glutamate neurotransmitter, particularly during multi-vesicular release events (P. F. Vincent et al. 2018).

Here, we describe the model of the ribbon synapse as a chain of events. After a channel opens, Ca^{2+} ions enter the cell following the concentration and voltage gradient. As a first approximation, the ionic current follows the Ohm's law, the voltage being described by the Nernst equation. However, the current can be calculated more precisely by the Goldman–Hodgkin–Katz (GHK) equation, as is done in the present model. In addition, the immediate intracellular concentration in the channel vicinity is taken into account, with the extracellular concentration considered constant.

The Ca^{2+} concentration in the vicinity of a neurotransmitter vesicle is estimated considering all individual channels within the synaptic domain using a point-source diffusion model with space restricted by the membrane. The concentration at an arbitrary distance from the group of channels can be calculated analytically from the ionic current through each channel using the experimentally accessible diffusion coefficient of Ca^{2+} ions in cytoplasm. Compared to previous models, no ad hoc time-constants or any other calibration parameters are needed. In this way, the present model can predict differences in behavior of ribbon synapses of different spatial conformations.

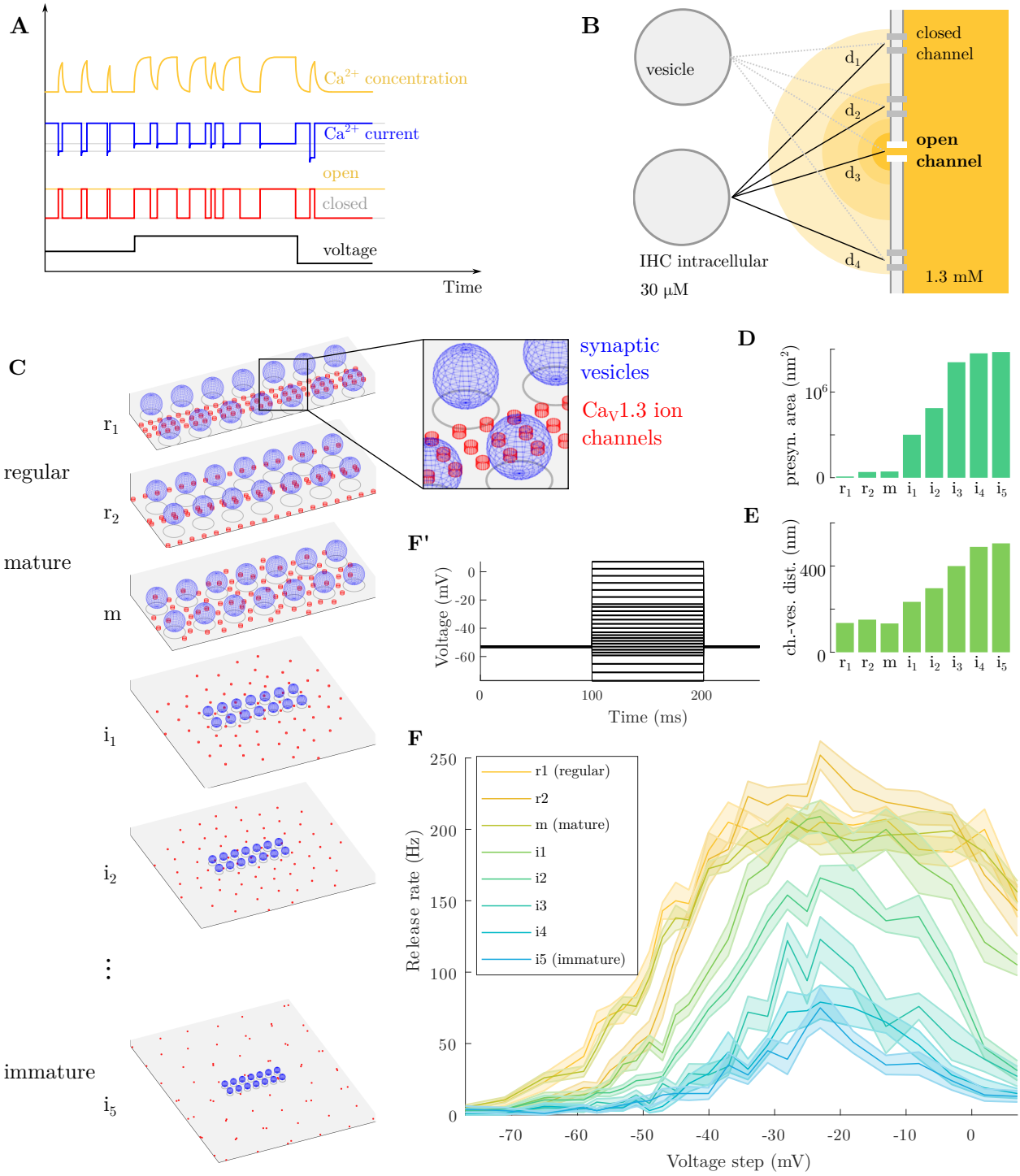

Figure 5: Effect of the morphology of the ribbon synapse. **A**: Illustration of a single $Ca_V1.3$ stochastic opening. Depolarization increases the channel open probability, opening of the channel causes ionic current and concentration increase in its vicinity. **B**: Cumulative effect of many ion channels has to be taken into account, considering their positions. **C**: Sample models of the ribbon synapse, from a regular organization of the $Ca_V1.3$ ion channels (r_1, r_2), through tightly-packed unorganized channels (m), up to spread-out configuration (i_1-i_5). **D**, **E**: Quantitative description of the channel distribution in terms of the presynaptic area (D) and the mean channel–vesicle distance (E). **F**: Average synaptic vesicle release rate for different channel configurations as a function of the peak voltage step in (F').

The molecular mechanism behind Ca^{2+} governed synaptic vesicle exocytosis is still not fully understood, although considerable progress has been made in recent years, for example identifying Otoferlin as the Ca^{2+} sensor for both vesicle fusion and vesicle pool replenishment (Michalski et al. 2017). From a macroscopic point of view, the calcium-mediated exocytosis exhibits a highly positively cooperative influence of calcium, with the Hill's coefficient for calcium dependent vesicle release from the Ribbon synapse being around five (Beutner et al. 2001; Jarsky et al. 2010), which is reflected in the present model.

Once a vesicle is released, its place is not immediately occupied by a new one, hence a vesicle pool depletion must be taken into account. Following Meddis (1986) and Sumner et al. (2002), we model the ribbon synapse as pools of free (i.e., ready to be released), re-processed, released, and freshly manufactured vesicles. In comparison to previous models, the release probability is calculated here individually for each free vesicle, depending on the respective local Ca^{2+} concentration. Additionally, each vesicle release is usually governed by only a small group of adjacent ion channels, compared to the previously published models, where individual channels have not been considered (instead, channels were modelled as a large population in a mean way). However, the number of "controlling" ion channels per vesicle is not a model parameter. Instead, the contribution of each channel is taken into account and, therefore, depending on the localization of calcium ion channels with respect to the ribbon synapse or depending on the stimulus level, the vesicle release can be caused both by a nearby channel or by a cumulative effect of multiple more distant channels. Finally, all vesicles from a single ribbon synapse released into the synaptic cleft are considered to be equal and the neurotransmitter re-uptake, vesicle factory, and reprocessing stores are joined into a single pool.

We have used the model to illustrate the effect of the post-natal maturation of the ribbon synapse (described by P. F. Vincent et al. (2018)) on its function. First, we have created different configurations of the ribbon synapse, having the same number of channels, but different presynaptic area and channel-vesicle distance distribution (Fig. 5C-E). Then, we simulated the response of the synapse to a voltage step in transmembrane potential and measured the mean vesicle release rate (Fig. 5F). Our results confirm that the "mature synapse", i.e. the synapse with close colocalization of the vesicles and $Ca_V1.3$ ion channels, has significantly increased sensitivity (especially close to the resting potential of the hair cell) compared to the "immature synapse" with high effective presynaptic area.

Auditory Nerve Model

An action potential is generated in the auditory nerve when a neurotransmitter is released from a vesicle at the ribbon synapse. However, when two such events take place immediately after each other, a new action potential may not occur after the second event. This is because a short ($\sim$1 ms) recovery interval (refractory period) is required. Also, an action potential may not occur if the amount of neurotransmitter released is too low and the resulting post-synaptic current does not reach the required threshold value. To account for all these effects, we include an up-to-date implementation of the Hodgkin–Huxley model of the auditory nerve by Bruce (2009) as the last stage in the system.

The integrated model

The present approach to the computer model of the peripheral auditory system allowed us to extend the focus from a single cross-section to an array of 3000 coupled cross-sections (each

containing a single IHC and four joined OHCs), explicitly calculating the response of each of them. Furthermore, for each IHC tens of individual ribbon synapses can be modelled individually. Being formulated and numerically solved in the time domain, the model can simulate the response to an arbitrary acoustic signal. As an illustration of the different model outputs, we show here the responses to a single word "greasy", and a noise-band bursts, see Fig. 6.

Thanks to an efficient implementation, we are able to simulate hundreds of different ∼100 ms signals in an over-night run on a high-end personal computer, so in general, the work with the present model can be realized in an interactive way without the need for high performance computing systems. Importantly, the modular implementation allows for easy extensions of the computer program, both in terms of new features and specific analyses.

Discussion

The present model was built upon a foundation of older models (Bruce 2009; Lopez-Poveda and Eustaquio-Martín 2006; Mammano and Nobili 1993; Meddis 1986; Mistrík, Mullaley, et al. 2008; Pascal et al. 1998; Strelioff 1973; Sumner et al. 2002; Vetešník and Gummer 2012), with other ones being developed in parallel (e.g. Heil et al. (2011), Motallebzadeh et al. (2018), Sasmal and Grosh (2019), and Verhulst, Altoè, et al. (2018)). Here, we briefly discuss these models putting the present one in the context of the existing literature, focusing in particular on the cochlear mechanics and ion flows, as well as on the ribbon synapse.

In the field of modelling of the cochlear mechanics several approaches have been employed in parallel, which differ by the level of detail of description (and the corresponding computational costs), ranging from digital filter-bank models to finite element 3-dimensional models (Motallebzadeh et al. 2018; Sasmal and Grosh 2019). In the present study, we have built upon an older model of intermediate complexity originated by Mammano and Nobili (1993) and Nobili and Mammano (1996) who showed that such a model, where the cochlear amplifier is explicitly based in the OHCs, can reproduce a wide range of phenomena, such as the two-tone suppression. The model is 2-dimensional, taking into account the height of the cochlear duct, which has been recently been shown to be important for a proper BM response (Altoè and Shera 2020). Vetešník and Gummer (2012) improved the original model, calibrated it to the human cochlea, and along with Vencovský et al. (2019) employed it to the study of distortion products and otoacoustic emissions. In this context it is important to stress that the modelling approach presented in this study is not bound to a particular model of cochlear mechanics (or any other parts of the peripheral auditory system).

As far as ionic flows in the cochlea are concerned, the original model by Strelioff (1973) included all three scalae, but the organ of Corti was oversimplified to a single resistor and a single voltage source per cross-section. Later, Mistrík, Mullaley, et al. (2008) adopted the general idea of a three-dimensional electric model of the entire cochlea, allowing to investigate the ionic flow not only within a single cross-section, but also in-between them along the tonotopy axis. Focusing on the organ of Corti they were also able model the ionic currents through both inner and outer hair cells. However, the basolateral hair cell conductance was considered to be constant (i.e., voltage independent), the channel kinetics were not accounted for, and the channel subtypes with different activation properties were not resolved, in contrast to what has been done for a single IHC by Lopez-Poveda and Eustaquio-Martín (2006) and later by Dierich et al. (2020). The tonotopic gradient in the properties of the hair cells was not included in a more recent model by Mistrík and

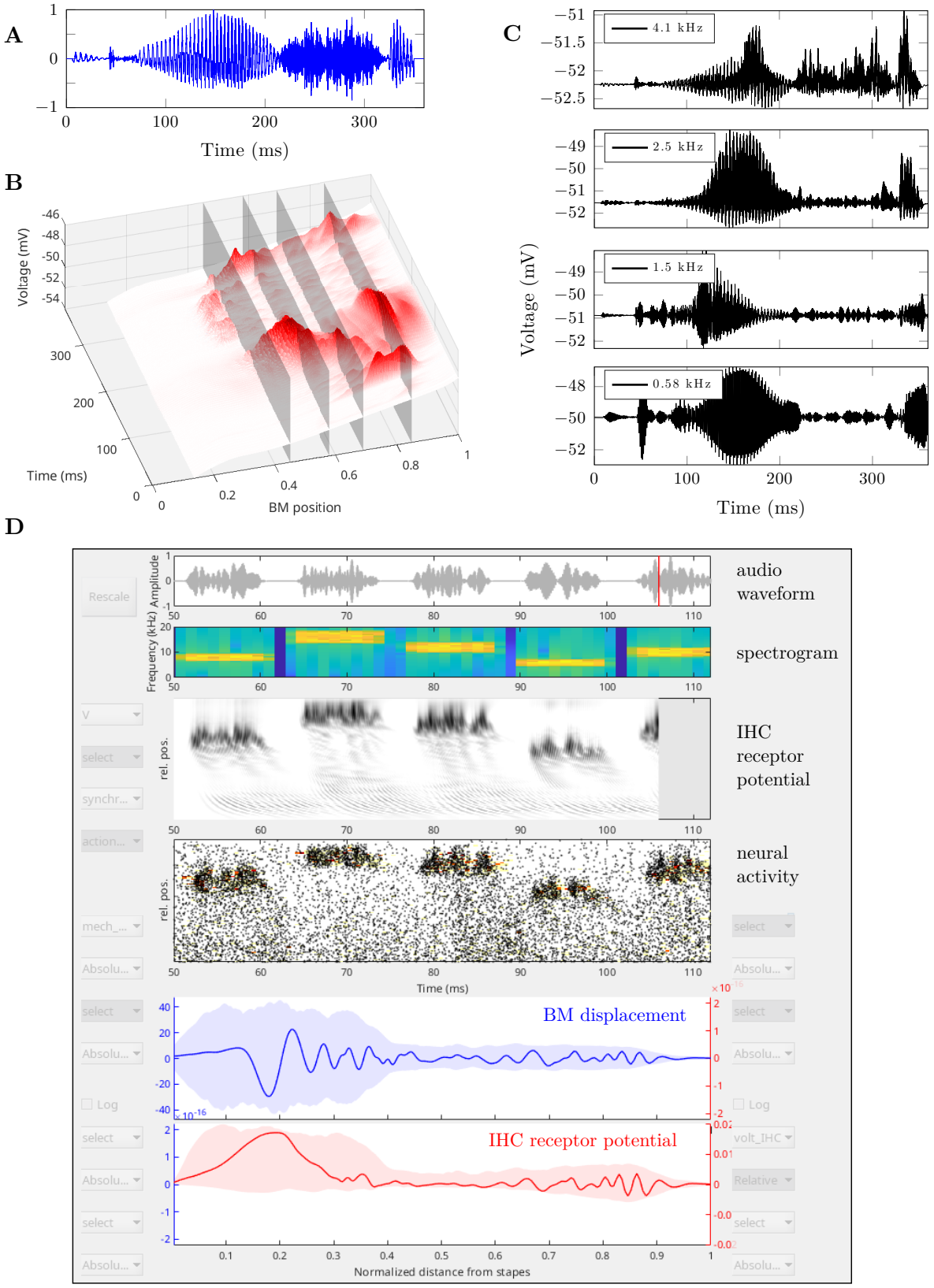

Figure 6: Output analysis and the graphical interface: **A-C**: Example model output of a simulation of person saying the word "greasy". **A**: Stimulus waveform. **B**: IHC basolateral voltage as a function of time along the tonotopy axis. **C**: Cuts of the three-dimensional graphs in B showing the response at specific characteristic frequency locations. **D**: Snapshot from the graphical user interface (GUI) allowing the user to visualize any calculated variable on the fly as well as after the simulation has been completed. Videos of different simulations are available as part of the supplementary material available online at `doi.org/10.6084/m9.figshare.20348271`.

Ashmore (2010) either. Moreover, the nonlinear capacitance, specific to outer hair cells, has not been included in any of these models. The present approach overcomes these limitations and models each of the 3000 cochlear partitions with a separate set of parameters.

For the IHC synapse, it was Meddis (1986) who laid grounds to the modeling approach and his concept of synaptic neurotransmitter "stores" has been in varying forms adopted also in more recent studies including the present one. Later, Sumner et al. (2002) introduced stochasticity of the synaptic vesicle release, which served as inspiration for the present work. While Sumner et al. (2002) were pioneers at aiming to provide a model of the whole auditory periphery, their model components were rather simplified (this concerns, in particular, the filter-based model of cochlear mechanics). Additionally, the calcium currents and concentrations in the vicinity of the ribbon synapse were not modelled in sufficient detail, and the model was calibrated only to a single tonotopic location of 18 kHz. In contrast, the present model is tonotopically resolved and detailed enough to be able to capture even subtle effects, such as the post-natal maturation of the ribbon synapse.

In conclusion, we have built a complete computational model of the human peripheral auditory pathway from the outer ear to the afferent auditory nerve. While most of the cochlear models previously published target only a specific part of the auditory pathway, here we present a complete model of the peripheral auditory system built in a bottom-up way in a physically and physiologically well justified way and with parameters reflecting up-to-date experiments. On the one side, the present model represents a research tool that allows to analyze the effects of the chosen parameter values and investigate the consequences of changes thereof. In this way, it provides a deep quantitative insight into the mechanism of hearing. On the other side, various forms of hearing impairment can be readily implemented and their effects investigated. As one such example for the cochlear model, effects connexin mutations on hearing impairment can be modelled in significantly more detail than attempted previously (Mistrík and Ashmore 2010). Along similar lines, a straightforward extension lies in the application of the present model to investigate the functionality of a cochlear implant. This will allow to model for example the combined effect of using the implant together with hearing aids, which will be instrumental in developing new simulation and fitting strategies for cochlear implants.

Acknowledgements

O.T. acknowledges the Faculty of Mathematics and Physics of the Charles University (Prague, Czech Republic) where he is enrolled as a PhD student and the International Max Planck Research School for "Many-Particle Systems in Structured Environments" (Dresden, Germany) for support. **P.M.** is an employee of Med-El company, which manufactures hearing prosthesis. **P.J.** acknowledges support from the European Regional Development Fund (project ChemBioDrug no. CZ.02.1.01/0.0/0.0/16_019/0000729).

Methods

Outer ear filtering

The outer ear transfer characteristics are implemented via a set of independent band-pass filters as proposed by Meddis (2011). Specifically, we used $N = 11$ fourth-order Butterworth band-pass filters, that transform the sound pressure p_0 to the pressure at the tympanic membrane p_{tymp}:

$$p_{\text{tymp}}(t) = p_0(t) + \sum_{n=1}^{N} g_n \cdot \texttt{butter}(p_0(t), l_n, h_n) \tag{1}$$

where g_n, l_n, and h_n are parameters of the filter – gain, low- and high-cutoff frequencies. Note that even though this design is purely additive, moderate attenuation at certain frequency bands can be achieved because of phase shift introduced by the filters, see Figure 2A.

Middle ear lumped-element model

The middle ear is represented as a lumped-element model of selected structures — namely the eardrum, the middle ear cavities, malleus, incus, stapes, the cochlea — and by their connections. The design has been taken directly from Pascal et al. (1998). For simplicity, we chose to omit the non-linearities in the response caused by the acoustic reflex and the influence of the angular ligament on the maximum stapes displacement, which appear for very high-level stimuli.

The mechanical model can be translated into an equivalent electrical representationand be driven by a voltage V_0 and the response can be observed in the form of voltage across the 'cochlea' resistor, V_{cochlea}. The values of V_0 and V_{cochlea} can be related to the tympanic membrane pressure p_{tymp} and the oval window pressure p_{cochlea} as

$$V_{\text{tymp}}(t) := K_{\text{tymp}} p_{\text{tymp}}(t), \qquad p_{\text{cochlea}}(t) := K_{\text{cochlea}} V_{\text{cochlea}}(t). \tag{2}$$

Since the model is linear in respect to the driving voltage (and therefore the acoustic pressure), the conversion constant K_{tymp} can be set to 1 V/Pa. The scaling constant K_{cochlea} [Pa/V] needs to be calibrated for the model to yield correct oval window pressure levels.

The electrical circuit representing the middle ear can be translated to a mathematical description using the Modified Nodal Analysis (MNA). This method results in a differential-algebraic equation (DAE) of the form

$$\begin{bmatrix} \boldsymbol{C} & 0 & 0 \\ 0 & \boldsymbol{L} & 0 \\ 0 & 0 & 0 \end{bmatrix} \frac{d}{dt} \begin{bmatrix} v \\ i_L \\ i_V \end{bmatrix} + \begin{bmatrix} \boldsymbol{G} & \boldsymbol{A}_L & \boldsymbol{A}_V \\ -\boldsymbol{A}_L' & 0 & 0 \\ \boldsymbol{A}_V' & 0 & 0 \end{bmatrix} \begin{bmatrix} v \\ i_L \\ i_V \end{bmatrix} = \begin{bmatrix} -\boldsymbol{A}_I I \\ 0 \\ E \end{bmatrix}, \tag{3}$$

where $\boldsymbol{C}$, $\boldsymbol{L}$ and $\boldsymbol{G}$ are matrices defined by the capacitances, inductances and conductances of the circuit elements, $\boldsymbol{A}_L$, $\boldsymbol{A}_V$, and $\boldsymbol{A}_I$ are defined by their connectivities, $I = 0$ and $E = (V_{\text{tymp}}, 0, \ldots, 0)$ are current and voltage sources and v, i_L and i_V are the node voltages and currents through inductors and voltage sources. The equation can be solved in the time domain as well as the frequency domain by employing the Fourier transform. Here, we used routines based on the symbolic circuit solver by

Cheever (2018) to assemble the matrices and solve the system in the time domain using standard DAE integrators (MATLAB 2021).

In their original paper Pascal et al. (1998) shown that such a model can be calibrated to reproduce an experimental sound pressure-intracochlear fluid pressure transfer function of a single individual. Since the cochlear mechanics model we employ in the later stage requires the stapes acceleration as an input, we recalibrated the model to a recent experimental tympanic membrane pressure-stapes velocity average data by Chien et al. (2009), see Figure 2B.

Cochlear Mechanics

Cochlear mechanics is governed by two partial differential equations for functions $\xi(t,x)$ and $\eta(t,x)$ that represent the displacement of the basilar membrane and the deflection of OHC stereocilia. The variable x here and in all future occurrences describes the position, along the longitudinal axis of an "uncoiled" cochlea, normalized so that $x=0$ at the base and $x=1$ at the apex; the variable t means time in all occasions.

$$m\,\frac{\partial^2\xi}{\partial t^2} + h\,\frac{\partial\xi}{\partial t} - \left[\frac{\partial}{\partial x}\,s\,\frac{\partial}{\partial x}\right]\frac{\partial\xi}{\partial t} + k\xi = F_{\text{stapes}} + F_{\text{BM}} - F_{\text{ampl}}(V_{\text{OHC}}) \tag{4}$$

$$\frac{\partial^2\eta}{\partial t^2} + \gamma_{\text{tect}}\,\frac{\partial\eta}{\partial t} + \omega_{\text{tect}}^2\eta = -\sin\theta\,\frac{\partial^2\xi}{\partial t^2}\,. \tag{5}$$

The equations are parameterized by the mechanical properties of the BM such as the mass $m(x)$, damping $h(x)$, shearing resistance $\partial_x s(x)\partial_x$, and stiffness $k(x)$, and tuning characteristics of the TM – $\gamma_{\text{tect}}(x)$, $\omega_{\text{tect}}^2(x)$, $\theta(x)$. The model is driven by the stapes motion $\sigma(t)$ (computed by the model of middle ear) via a fluid coupling between the stapes displacement and the BM displacement

$$F_{\text{stapes}} = -G_{\text{stapes}}\,\frac{\mathrm{d}^2\sigma}{\mathrm{d}t^2} \tag{6}$$

modeled by the Green's function $G_{\text{stapes}}(x)$. Individual BM segments are also connected via fluid coupling

$$F_{\text{BM}} = -\int_0^1 G_{\text{BM}}(x,\bar{x})\,\frac{\partial^2\xi(t,\bar{x})}{\partial t^2}\,\mathrm{d}\bar{x}, \tag{7}$$

where $G_{\text{BM}}(x,\bar{x})$ is the Green's function. Here we refer the reader to the original paper by Vetešník and Gummer (2012) for detailed description of the parameters.

The differential equations are coupled to the model of ionic currents through the cochlea by regulating the conductance of MET channels (described later) and via the OHC transmembrane voltage V_{OHC}, that induces an active force on the system resulting in amplification. The amplifier force F_{ampl} is explicitly computed (for each position x) via the differential equation

$$\frac{\mathrm{d}F_{\text{ampl}}}{\mathrm{d}t} + \tau F_{\text{ampl}} = \frac{\mathrm{d}V_{\text{OHC}}}{\mathrm{d}t}\,, \tag{8}$$

which is a high-pass filter defined by the time constant τ. The force F_{ampl} needs to be zero at steady state, which is fulfilled for this equation by choosing a proper initial conditions. The filtering inherently involves a magnitude change and phase shift, however, for small enough τ, the filtering affects only very low frequency components, leaving the physiologically relevant unchanged (see Supplementary Fig. S-6).

Due to the novel description employing directly the OHC receptor potential, there is a (nonlinear) feedback loop between the model of cochlear mechanics and the ionic currents. Consequently, the partial differential equation 4 has to be solved simultaneously with the differential-algebraic equation 12.

Channel dynamics

Measuring the basolateral K^+ currents in isolated guinea-pig IHCs, Kros and Crawford (1990) determined that each of the channel subtypes exhibits kinetics with two closed states (C_1, C_2) and one open state (O) governed by voltage dependent transfer rates α, β, γ, and η

$$C_1 \underset{\eta}{\overset{\alpha}{\rightleftharpoons}} C_2 \underset{\gamma}{\overset{\beta}{\rightleftharpoons}} O. \tag{9}$$

Considering a population of the channels, the stochastic dynamics of the channels can be described in a mean way by differential equations, see Lopez-Poveda and Eustaquio-Martín (2006) for derivation and details. They show that open state probability $O(t, V)$ of the channels can be equivalently described by a second-order differential equation

$$\tau_1 \tau_2 \frac{d^2 O}{dt^2} + (\tau_1 + \tau_2) \frac{dO}{dt} - O_\infty = 0 \tag{10}$$

where $\tau_1(V)$ and $\tau_2(V)$ are voltage dependent time constants and $O_\infty(V)$ is voltage dependent steady-state open probability. The time constants τ_1, τ_2 and the steady state open probability O_∞ are directly related to the transfer rates from Eq. 9, while being experimentally accessible. We have applied the methodology for both IHCs and OHCs; an independent set of these nonlinear differential equations, one per channel subtype and per hair cell, is inherently coupled with the system 12 describing the ionic currents through the cochlea, as the instantaneous channel conductance is proportional to the open probability:

$$G(t, V) = G_{\max} O(t, V). \tag{11}$$

This creates yet another nonlinear feedback loop in the system, and therefore the Eq. 10 has to be solved concurrently with the Eq. 12.

Model of a single hair cell

Electrophysiological experiments are usually performed on isolated hair cells. The hair bundles are then bathed in the same extracellular solution as the rest of the cell, eliminating the endocochlear potential. In order to compare directly to in-vitro experiments, we implemented a model of an isolated hair cell that can simulate whole-cell patch clamp recordings as well as response to hair bundle stimulation.

The model (Fig. 3E) is designed in a similar fashion as the model of a cochlear cross-section. It does not include the endocochlear potential, but is driven as in a standard patch-clamp experiment by a current or voltage pulses. Due to it's computational simplicity, it can be used for parameter estimation using multivariate optimizers (we have used differential evolution). Results of optimization can be seen in the Figure 3F-J.

Equivalent circuit of the cochlea

In addition to resistors and capacitors representing the conductive properties of cellular membranes expressing ion channels or prestin, the circuit is composed also of voltage sources accounting for actively maintained electric fields and ionic concentration gradients across the cellular membrane through the Nernst equation. Since the primary interest is in the current flow through the sensory cells of the cochlea, the inner and outer hair cells are implemented in the most detail (apical and basolateral membranes). An equivalent electrical circuit of a single cross-section of the cochlear partition can be seen in the Figure 3A. The outer hair cells are represented at each cross-section by a single element with its values scaled up accordingly. Since it is assumed that the outer hair cells within one cross-section are independent and identical, this does not change the results.

The construction of the equivalent electrical circuit representing the whole cochlear partition is concluded by a longitudinal connection of the partial "cross-section" circuits. As seen in the Figure 3A, the cross-section circuits are linked together by groups of resistors, representing corresponding sections of conductive paths in the scalae (SV, SM, ST), and in the stria vascularis (StV), spiral limbus (SL) and organ of Corti (OC). In the full circuit, there is one cross-section sub-circuit for every row of hair cells (single IHC and four OHCs), resulting in approximately 3000 sections for the human cochlea. For some experiments, due to computational requirements, some of these sections were lumped together while adjusting the values of the electric elements. This can be done without significant loss in accuracy; one only has to consider the loss of spatial resolution along the longitudinal axis of the cochlea.

Since the circuit only contains passive linear elements (resistors and capacitors) plus only independent voltage sources are considered, the modified nodal analysis (MNA) can be used for constructing the circuit equations. The resulting system of differential equations can be written as

$$\mathbf{C}(\vec{y})\frac{\mathrm{d}\vec{y}}{\mathrm{d}t} = -\mathbf{A}(\vec{y},t)\vec{y} + \vec{z} =: \vec{f}(\vec{y},t), \tag{12}$$

where the unknown vector $\vec{y}$ holds the voltage at nodes of the circuit and current through voltage sources, the matrix $\mathbf{A}(\vec{y},t)$ is a time-, voltage-, and current-dependent block matrix defined by the interconnections of resistors and voltage sources, $\mathbf{C}(\vec{y})$ is a singular matrix defined by the capacitors, the vector $\vec{z}$ contains voltage and current sources. Since the matrix $\mathbf{C}(\vec{y})$ is singular, the system 12 is in fact a differential-algebraic equation (DAE), and consequently a suitable numerical method must be employed.

Let us now describe the equation and its connection to the cochlear model in more detail. In the following, let n denote the number of nodes (except for the ground node *zero*) in the circuit and m the number of independent voltage sources.

The matrix $\mathbf{A}(\vec{y},t)$ is generally constructed as a block matrix

$$\mathbf{A} = \begin{pmatrix} \mathbf{G} & \mathbf{B} \\ \mathbf{E} & \mathbf{D} \end{pmatrix} \tag{13}$$

where $\mathbf{G} \in \mathbb{R}^{n,n}$ holds the conductances of the circuit elements between the nodes, matrices $\mathbf{B} \in \mathbb{R}^{n,m}$ and $\mathbf{E} \in \mathbb{R}^{m,n}$ are determined by connection of the voltage sources, and $\mathbf{D} \in \mathbb{R}^{m,m}$ is zero if only independent voltage sources are considered.

The vector $\vec{y} = (\vec{v}, \vec{j})^\mathsf{T} \in \mathbb{R}^{m+n}$ contains unknown quantities — voltages at nodes $\vec{v} \in \mathbb{R}^n$ and currents through voltage sources $\vec{j} \in \mathbb{R}^m$. The vector $\vec{z} = (\vec{i}, \vec{e})^\mathsf{T} \in \mathbb{R}^{m+n}$ is defined by current and voltage sources values — the sum of all current sources at each node defines the vector $\vec{i} \in \mathbb{R}^n$, the electromotive force of each voltage source defines the vector $\vec{e} \in \mathbb{R}^m$.

An equivalent electrical circuit consisting of k cross-section each containing n nodes and m voltage sources results in a system of $k \times (m+n)$ unknowns. In the current design, $(m+n) = 17$, and k can vary from 600 up to 3000. Consequently, the computational requirements are rather high. Since a significant portion of the resources was spent on creating the time-dependent matrix $\mathbf{A}(t)$ during the construction of the Jacobian matrix or while computing the right-hand side of the (12), an alternative to the classic MNA procedure was employed. It exploits the fact that only the submatrix $\mathbf{G}$ of the matrix $\mathbf{A}$ is time and voltage dependent and, moreover, that $\mathbf{G}$ can be expressed as

$$\mathbf{G}(t) = \mathbf{G}_{\text{const}} + \mathbf{G}_{\text{displ}}(\vec{\xi}(t)) + \mathbf{G}_{\text{defl}}(\vec{\eta}(t)) + \mathbf{G}_{\text{volt}}(\vec{v}), \tag{14}$$

where $\vec{\xi}$ is the BM displacement, $\vec{\eta}$ is the deflection of the stereocilia in OHCs. Moreover, the voltage-dependent part of $\mathbf{G}$ can be further decomposed to

$$\mathbf{G}_{\text{volt}}(\vec{v}) = \mathbf{G}_{\text{volt, IHC}}(\vec{V}_{\text{in}}^{\text{IHC}}, \vec{V}_{\text{ex}}^{\text{IHC}}) + \mathbf{G}_{\text{volt, OHC}}(\vec{V}_{\text{in}}^{\text{OHC}}, \vec{V}_{\text{ex}}^{\text{OHC}}), \tag{15}$$

where $\vec{V}_{\text{in}}^{\text{IHC}}$, $\vec{V}_{\text{ex}}^{\text{IHC}}$, $\vec{V}_{\text{in}}^{\text{OHC}}$, $\vec{V}_{\text{ex}}^{\text{OHC}}$ are IHC and OHC intra- and extracellular voltages.

The Jacobian matrix of the DAE is defined as

$$\mathbf{J} := \frac{\partial \vec{f}(\vec{y}, t)}{\partial \vec{y}} \tag{16}$$

where $\vec{f}(\vec{x}, t)$ is defined by the system

$$\boldsymbol{C}\frac{\mathrm{d}\vec{y}}{\mathrm{d}t} = -\boldsymbol{A}(t, \vec{y})\vec{y} + \vec{z}(t, \vec{y}) = \vec{f}(\vec{x}, t).$$

The Jacobian matrix is necessary for most DAE solvers, including the employed matlab ode15s.

If $\boldsymbol{A}(t, \vec{y}) = \boldsymbol{A}(t)$, then $\boldsymbol{J} = -\vec{A}(t) + \frac{\mathrm{d}\vec{z}(t,\vec{y})}{\mathrm{d}\vec{y}}$, otherwise product rule must be used

$$\boldsymbol{J} = -\boldsymbol{A}(t, y) - \frac{\mathrm{d}\boldsymbol{A}(t, \vec{y})}{\mathrm{d}\vec{y}}\vec{y} + \frac{\mathrm{d}\vec{z}(t, \vec{y})}{\mathrm{d}\vec{y}}$$

Note that $\vec{h}(\vec{y}) = -\boldsymbol{A}\vec{y}$ is matrix times vector, i.e.

$$h_i = -A_{i,1}y_1 - A_{i,2}y_2 - ... - A_{i,n}y_n = -\sum_j^n A_{i,j}y_j$$

and therefore

$$\left(\frac{\mathrm{d}\vec{h}}{\mathrm{d}\vec{y}}\right)_{i,k} = \frac{\mathrm{d}h_i}{\mathrm{d}y_k} = -A_{i,k} - \sum_j^n \frac{\mathrm{d}A_{i,j}}{\mathrm{d}y_k}\, y_j = -A_{i,k} - \frac{\mathrm{d}A_{i,\bullet}}{\mathrm{d}y_k}\,\vec{y}$$

where $A_{i,\bullet}$ is a i-th row vector.

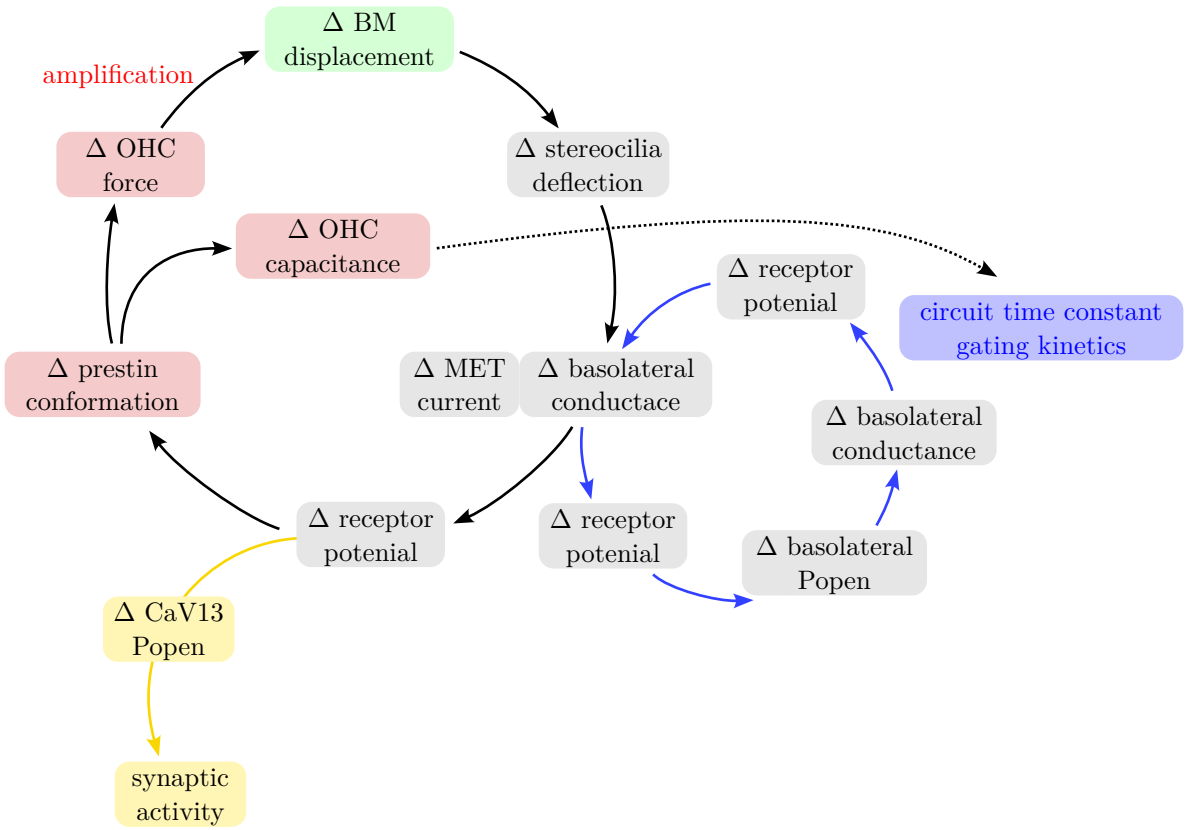

Figure M-1: Feedback loops in the model of cochlear mechanics and ionic currents.

Closed feedback loop in the system of models

Deflection of the OHC stereocilia η changes the instantaneous conductance of the MET channels $G_{\mathrm{MET}}^{\mathrm{OHC}}$ changing the potential across the apical, and also the basolateral OHC membrane V. The voltage gated basolateral channels adjust their open-state probability O and conductance $G_{\mathrm{B}}^{\mathrm{OHC}}$, pushing the voltage change partially back towards the equilibrium. At the same time, the voltage gated prestin protein changes its conformation and inflicts a force on the basilar membrane. If this happens in phase with the BM oscillation, it results in amplification of the motion of the coupled BM–TM system. Finally, the simultaneous motion of the BM and TM induces a deflection of the OHC stereocilia, thus closing the loop. The feedback is visualized schematically in the Figure M-1 and also through the Jacobian matrix of the system in the figure M-2.

Ribbon Synapse

The ribbon synapse is defined by the positions of readily releasable (membrane-tethered) vesicles and the $\mathrm{Ca_V}1.3$ ion channels. Vesicles attached to the ribbon further away from the membrane and cytosolic vesicles are currently not considered. In the present configuration of the model, there are 14 readily releasable vesicles distributed in two uniform rows (Khimich et al. (2005) states 16–30 ready-to-release docked vesicles), covering an area of approx. 120×400 nm (approx. in line with Moser et al. (2006)). The vesicles in the model are considered identical spheres with a radius of 20 nm based on our measurements of electron micrograph (circular cross-sections with radius $r = 16 \pm 2$ nm from Frank et al. (2010), elliptical cross-sections with $a = 25 \pm 4$ nm, $b = 17 \pm 2$ nm from Rutherford (2015)). The vesicles are located 5 nm from the plasma membrane, again estimated

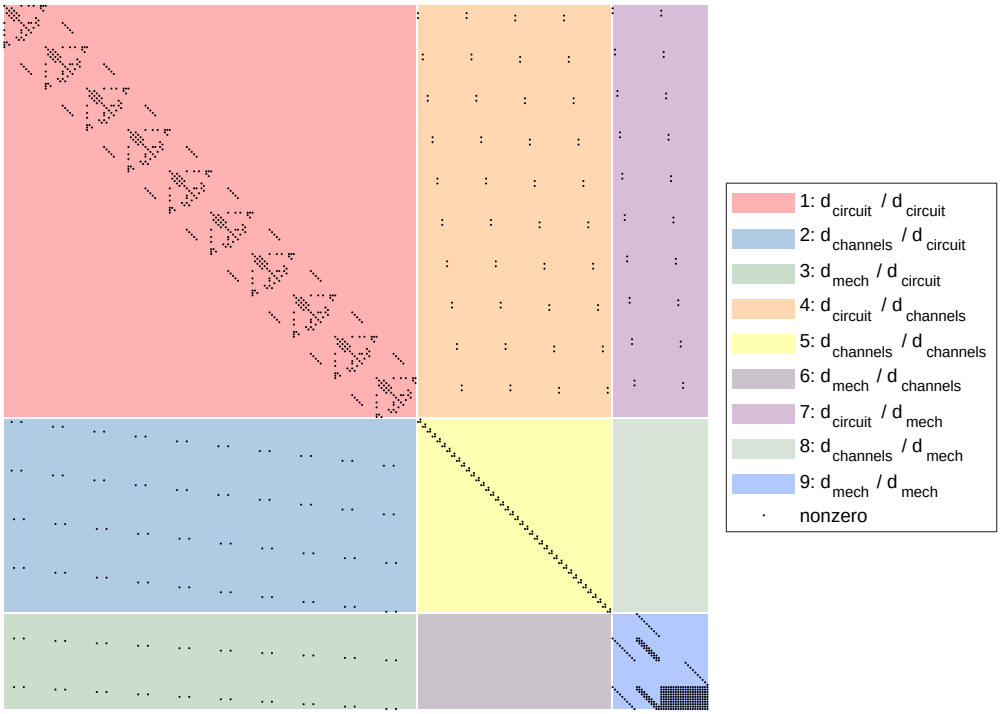

Figure M-2: The interaction between the model blocks visualized via the Jacobian matrix. Internally, the model is composed of three blocks: the electrical circuit, the gating model of the basolateral IHC and OHC ion channels, and the mechanical model, each composed of multiple equations. When the i-th equation depends on the j-th component of the solution, the derivative $\partial f_i/\partial y_j$ is generally nonzero, and therefore, the $J_{i,j}$ element of the jacobian matrix is nonzero. In the figure, nonzero elements are displayed as black dots. The matrix is shaded with colors corresponding to different classes of the dependence. The diagonal blocks represent internal dependence within the circuit, the channels and the mechanical models. The off-diagonal blocks instead symbolize the feedback between the models. The off-diagonal blue block (2) represents the voltage dependence of the basolateral IHC/OHC channels, the green block (3) shows the voltage-dependent OHC amplifier, the orange block (4) represents the basolateral IHC/OHC conductance change upon the opening of the channels, and finally the purple block (7) represents conductance change of the MET channels induced by vibrations of the BM and deflection of the stereocilia. Note that for visualization purposes, the number of cross-sections (corresponding to the size of the system) was reduced to 10. Otherwise, the fine structure of the nonzero elements would be hidden due to the image resolution.

from the micrographs. Note that the number of vesicles, their size and spatial distribution are intended as model parameters.

The $Ca_V1.3$ ion channels are represented by their positions on the membrane surface. The distribution of the ion channels in the active zone of the synapse is an input parameter of the model. For an ease of use, we have implemented a simple Monte Carlo simulation controlling the spread of the ion channels, producing random configurations from wide-spread-out to closely-packed (simulating synapse maturation) by changing just two parameters.

The number of ion channels in a IHC can be estimated experimentally via non-stationary fluctuation analysis of Ca^{2+} tail currents. For the mammalian IHC Brandt (2005) gives about 1700 Ca^{2+} channels, similar numbers have also been reported later (P. F. Y. Vincent et al. 2014; Wong et al. 2013). The channels are mostly expressed around the active zones (about 5–30 per IHC, Moser et al. (2006)). However, Magistretti et al. (2015) argues that the total number is in fact higher, when accounting for inactivated channels. They give 6400 channels (middle turn) and 1152 in about 16 active zones (basal turn). Along these lines, Zampini, Johnson, Franz, Lawrence, et al. (2010)

gives 9600 channels (P5-P10 mouse, apex). Zampini, Johnson, Franz, Knipper, et al. (2014) gives about 16000 channels in about 21 active zones (adult gerbil, middle turn 2 kHz). A single channel conductance is estimated by Zampini, Johnson, Franz, Knipper, et al. (2014) to 15 pS.

The $\mathrm{Ca_V}1.3$ channels are known to be voltage-sensitive, the open probability is usually expressed in terms of a Boltzmann function

$$P_{\text{open}} = \left(1 + \exp\left(-\frac{V - H}{S}\right)\right)^{-1}.$$

The open probability can be accessed indirectly by measuring the whole cell Ca2+ conductance or by analyzing open and close distributions of single channel measurements. Several reported Boltzmann curves can be found in Fig. M-3 as well as the curve used in the present model. It has been suggested that the parametrization of the $\mathrm{Ca_V}1.3$ open probability can change among individual active zones, displaying a pillar–modiolar gradient (Ohn et al. 2016), yielding a possible extension to this model.

Channels in the synapse model can be either closed or open, corresponding to two Markov states

$$\text{(closed)} \quad \underset{k^-}{\overset{k^+}{\rightleftharpoons}} \quad \text{(open)}.$$

The transfer rates k^+ and k^- can be expressed as

$$k^+ = k_0^+ e^{-\alpha V}, \qquad k^- = k_0^- e^{-\beta V}$$

where α (unit V^{-1}) and k_0^+ (unit s^{-1}) are control parameters and

$$\beta = \alpha + 1/S \qquad \text{and} \qquad k_0^- = k_0^+ \cdot \exp(H \cdot (\beta - \alpha))$$

Then the probability transfer matrix is

$$Q = \begin{pmatrix} 1 - k^+\,\mathrm{d}t & k^-\,\mathrm{d}t \\ k^+\,\mathrm{d}t & 1 - k^-\,\mathrm{d}t \end{pmatrix}$$

where $\mathrm{d}t$ is the simulation time step.

The control parameters α and k_0^+ then control the timescale in which the transfers occur (the kinetics of the process). Their values can be obtained by comparing the mean open and closed times of the Markov process with experimental data. We have calibrated the rates to reflect the mean open and closed times of single channel patch-clamp measurements, as reported by Zampini, Johnson, Franz, Knipper, et al. (2014).

At any point in the space in the vicinity of the active zone, the Ca^{2+} concentration C is calculated as a sum of the background concentration C_{bg} and the partial concentrations C_i caused by elementary current through the i-th ion channel. The background (bulk) endolymph calcium concentration ranges from 17 µM (base) to 41 µM (apex) (Salt et al. 1989). For each location of interest and each ion channel, the concentration C_i is calculated via the single-point source diffusion equation

$$\frac{\partial C}{\partial t} = D\nabla^2 C(t, \vec{r}) + I(t)\delta(\vec{r}),$$

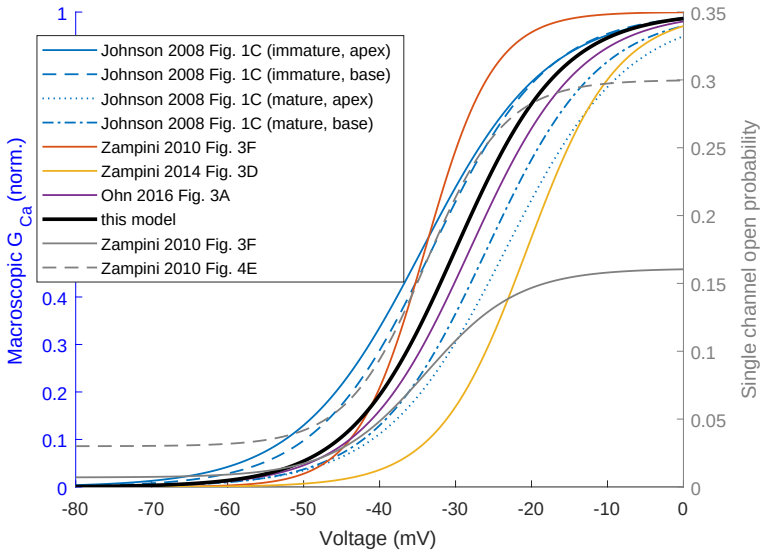

Figure M-3: $Ca_V1.3$ normalized conductance and open probabilities used in this model compared to available experimental data by Johnson and Marcotti (2008), Ohn et al. (2016), Zampini, Johnson, Franz, Knipper, et al. (2014), and Zampini, Johnson, Franz, Lawrence, et al. (2010).

where r is the distance from the ion channel, D is the diffusion coefficient, I current through the channel and δ is the Dirac delta function. We solve this equation analytically (using propagation integrals) for each time step, as described in Gillespie (2020). The main advantage of this method is that since the solution is exact, it does not require any spatial discretization and the time steps can be arbitrarily large, as long as the current through the channel can be considered constant.

For each vesicle, we are taking into account all $Ca_V1.3$ channels within the active zone (and the distance from each channel.

The calcium current through individual channels can be calculated via the Goldman–Hodgkin–Katz equation

$$\Phi = P_{\mathrm{Ca}} z^2 \frac{VF^2}{RT} \frac{C_{\mathrm{in}} - C_{\mathrm{out}} \exp(-zVF/RT)}{1 - \exp(-zVF/RT)},$$

where V is the receptor potential, P_{Ca} is permeability, R is the gas constant, F is the Faraday constant, $z = 2$ is the calcium cation valence, C_{in} is the intracellular and $C_{\mathrm{out}} = 1.3$ mM the extracellular calcium concentration (assumed constant along the tonotopy axis) (Salt et al. 1989).

It is unclear in which distance from the channel should one calculate the effective calcium reversal potential for that channel. Zampini, Johnson, Franz, Knipper, et al. (2014) measured macroscopic Ca^{2+} currents in middle-coil IHC bathed in high Na^+, 5 mM Ca^{2+} solution in a voltage range -70 to 0 mV and estimated the calcium reversal potential to 28 ± 2 mV. Having performed analogous simulations, we estimated the distance of 20 nm to yield similar reversal potentials for sustained Ca^{2+} currents.

The probability of vesicle release is proportional to the calcium concentration at the calcium sensor, saturating for high concentration values, which is achieved for simplicity through a Boltzmann function. To achieve adaptation properties, a depleteable vesicle pool model is introduced following Meddis (1986) and Sumner et al. (2002). In this model, the vesicles can exist in three states – free (readily available for release) denoted q, released into the synaptic cleft c and being reprocessed by the ribbon synapse w. Considering k being the calcium concentration dependent target release rate,

r being the re-uptake rate of vesicles from the cleft and l the loss rate, x the rate of replenishing the free pool and y the rate of manufacture, the model can be described mathematically as

$$\begin{aligned}\frac{\mathrm{d}q(t)}{\mathrm{d}t} &= N(w(t), x) + N(M - q(t), y) - N(q(t), k(C(t))),\\ \frac{\mathrm{d}c(t)}{\mathrm{d}t} &= N(q(t), k(C(t))) - lc(t) - rc(t),\\ \frac{\mathrm{d}w(t)}{\mathrm{d}t} &= rc(t) - N(w(t), x).\end{aligned}$$

The function $N(n, \varrho)$ is a stochastic function governing the neurotransmitter transfer of n vesicles between the stores, with a mean release rate of ϱ for each single vesicle.

In contrast to the original models by Meddis (1986) and Sumner et al. (2002), our model contains discrete locations for readily releasable vesicles. Therefore, internally, we don't work with a variable q describing a number of readily releasable vesicles, but rather with an array of release sites, for which only valid state transitions may occur (e.g. only an empty release site may be replenished).

Following a multivesicular release, the $\mathrm{Ca_V}1.3$ channel can be temporarily blocked by protons, released from the vesicle to the synaptic cleft (P. F. Vincent et al. 2018). Therefore, a third "blocked" state is added to the two Markov states. However, since we assume the un-blocking of a channel is not affected by external conditions (such as the receptor potential), we can use the Gillespie algorithm, sampling a "blocked duration" randomly upon the blocking of the channel. The probability for the channel to be blocked is assumed to be a Boltzmann function of the normalized proton concentration in the cleft C_+, where $C_+ = 1$ is the concentration recorded immediately upon a single vesicle release event. The half activation and slope of the Boltzmann function remain calibration parameters, the values 3 and 0.3 yield a reasonable agreement with experiment (P. F. Vincent et al. 2018).

Implementation notes

The model is implemented in MATLAB 2021 optionally employing the Signal Processing toolbox for parts of the analysis and Parallel Computing toolbox for an improved performance. The in-house implementation of the Modified Nodal Analysis (MNA) is based on the symbolic MNA solver SCAM (Cheever 2018). The GNU parallel program (Tange 2018) was used for launching independent simulations in large-scale analysis of the model parameters.

It is worth noting, that our MATLAB implementation allows automatic construction of the final 3D electrical circuit from a single table describing the radial and longitudinal connectivity between electrical elements representing selected cochlear structures. Therefore, the equivalent circuit can be easily modified for studying cochleas of different species or different forms of hearing impairments. An implementation of known deafness mutations (Dror and Avraham 2010; Petit et al. 2001) or changes in aging cochlea such as the reduced number of sensory cells (J. Wang and Puel 2020) are easily implemented.

References

Aibara, Ryuichi et al. (Feb. 2001). "Human middle-ear sound transfer function and cochlear input impedance". In: *Hearing Research* 152.1-2, pp. 100–109. DOI: 10.1016/s0378-5955(00)00240-9.

Altoè, Alessandro and Christopher A. Shera (Nov. 2020). "The cochlear ear horn: geometric origin of tonotopic variations in auditory signal processing". In: *Scientific Reports* 10.1. DOI: 10.1038/s41598-020-77042-w.

Beurg, Maryline et al. (Aug. 2021). "The speed of the hair cell mechanotransducer channel revealed by fluctuation analysis". In: *Journal of General Physiology* 153.10. DOI: 10.1085/jgp.202112959.

Beutner, Dirk et al. (Mar. 2001). "Calcium Dependence of Exocytosis and Endocytosis at the Cochlear Inner Hair Cell Afferent Synapse". In: *Neuron* 29.3, pp. 681–690. DOI: 10.1016/s0896-6273(01)00243-4.

Brandt, A. (Dec. 2005). "Few CaV1.3 Channels Regulate the Exocytosis of a Synaptic Vesicle at the Hair Cell Ribbon Synapse". In: *Journal of Neuroscience* 25.50, pp. 11577–11585. DOI: 10.1523/jneurosci.3411-05.2005.

Bruce, Ian C. (Dec. 2006). "Implementation Issues in Approximate Methods for Stochastic Hodgkin–Huxley Models". In: *Annals of Biomedical Engineering* 35.2, pp. 315–318. DOI: 10.1007/s10439-006-9174-9.

— (Jan. 2009). "Evaluation of Stochastic Differential Equation Approximation of Ion Channel Gating Models". In: *Annals of Biomedical Engineering* 37.4, pp. 824–838. DOI: 10.1007/s10439-009-9635-z.

Cheever, Erik (2018). *SCAM (Symbolic Circuit Analysis with MATLAB)*. URL: https://github.com/echeever/scam.

Chen, G.D. et al. (Mar. 2020). "Novel oral multifunctional antioxidant prevents noise-induced hearing loss and hair cell loss". In: *Hearing Research* 388, p. 107880. DOI: 10.1016/j.heares.2019.107880.

Chien, Wade et al. (Mar. 2009). "Measurements of stapes velocity in live human ears". In: *Hearing Research* 249.1-2, pp. 54–61. DOI: 10.1016/j.heares.2008.11.011.

Dallos, Peter and Bernd Fakler (Feb. 2002). "Prestin, a new type of motor protein". In: *Nature Reviews Molecular Cell Biology* 3.2, pp. 104–111. DOI: 10.1038/nrm730.

Dierich, Marlen et al. (July 2020). "Optimized Tuning of Auditory Inner Hair Cells to Encode Complex Sound through Synergistic Activity of Six Independent K+ Current Entities". In: *Cell Reports* 32.1, p. 107869. DOI: 10.1016/j.celrep.2020.107869.

Dror, Amiel A. and Karen B. Avraham (Oct. 2010). "Hearing Impairment: A Panoply of Genes and Functions". In: *Neuron* 68.2, pp. 293–308. DOI: 10.1016/j.neuron.2010.10.011.

Frank, Thomas et al. (Nov. 2010). "Bassoon and the Synaptic Ribbon Organize Ca2+ Channels and Vesicles to Add Release Sites and Promote Refilling". In: *Neuron* 68.4, pp. 724–738. DOI: 10.1016/j.neuron.2010.10.027.

Géléoc, G. S. G. et al. (Apr. 1997). "A quantitative comparison of mechanoelectrical transduction in vestibular and auditory hair cells of neonatal mice". In: *Proceedings of the Royal Society of London. Series B: Biological Sciences* 264.1381, pp. 611–621. DOI: 10.1098/rspb.1997.0087.

Gillespie, Dirk (June 2020). "Simulating diffusion from a cluster of point sources using propagation integrals". In: *European Biophysics Journal* 49.5, pp. 385–393. DOI: 10.1007/s00249-020-01438-9.

Greenwood, Donald D. (June 1990). "A cochlear frequency-position function for several species—29 years later". In: *The Journal of the Acoustical Society of America* 87.6, pp. 2592–2605. DOI: 10.1121/1.399052.

Heil, P. et al. (Oct. 2011). "An Improved Model for the Rate-Level Functions of Auditory-Nerve Fibers". In: *Journal of Neuroscience* 31.43, pp. 15424–15437. DOI: 10.1523/jneurosci.1638-11.2011.

Jarsky, T. et al. (Sept. 2010). "Nanodomain Control of Exocytosis Is Responsible for the Signaling Capability of a Retinal Ribbon Synapse". In: *Journal of Neuroscience* 30.36, pp. 11885–11895. DOI: 10.1523/jneurosci.1415-10.2010.

Jia, S. et al. (Jan. 2007). "Mechanoelectric Transduction of Adult Inner Hair Cells". In: *Journal of Neuroscience* 27.5, pp. 1006–1014. DOI: 10.1523/jneurosci.5452-06.2007.

Johnson, Stuart L. (Nov. 2015). "Membrane properties specialize mammalian inner hair cells for frequency or intensity encoding". In: *eLife* 4. DOI: 10.7554/elife.08177.

Johnson, Stuart L., Maryline Beurg, et al. (June 2011). "Prestin-Driven Cochlear Amplification Is Not Limited by the Outer Hair Cell Membrane Time Constant". In: *Neuron* 70.6, pp. 1143–1154. DOI: 10.1016/j.neuron.2011.04.024.

Johnson, Stuart L. and Walter Marcotti (Feb. 2008). "Biophysical properties of CaV1.3 calcium channels in gerbil inner hair cells". In: *The Journal of Physiology* 586.4, pp. 1029–1042. DOI: 10.1113/jphysiol.2007.145219.

Johnstone, B.M. et al. (Jan. 1986). "Basilar membrane measurements and the travelling wave". In: *Hearing Research* 22.1-3, pp. 147–153. DOI: 10.1016/0378-5955(86)90090-0.

Khimich, Darina et al. (Apr. 2005). "Hair cell synaptic ribbons are essential for synchronous auditory signalling". In: *Nature* 434.7035, pp. 889–894. DOI: 10.1038/nature03418.

Kros, C J and A C Crawford (Feb. 1990). "Potassium currents in inner hair cells isolated from the guinea-pig cochlea." In: *The Journal of Physiology* 421.1, pp. 263–291. DOI: 10.1113/jphysiol.1990.sp017944.

Li, Hao et al. (Feb. 2021). "Three-dimensional tonotopic mapping of the human cochlea based on synchrotron radiation phase-contrast imaging". In: *Scientific Reports* 11.1. DOI: 10.1038/s41598-021-83225-w.

Lopez-Poveda, Enrique A. and Almudena Eustaquio-Martín (May 2006). "A Biophysical Model of the Inner Hair Cell: The Contribution of Potassium Currents to Peripheral Auditory Compression". In: *Journal of the Association for Research in Otolaryngology* 7.3, pp. 218–235. DOI: 10.1007/s10162-006-0037-8.

Magistretti, Jacopo et al. (Apr. 2015). "Elementary properties of Ca2+ channels and their influence on multivesicular release and phase-locking at auditory hair cell ribbon synapses". In: *Frontiers in Cellular Neuroscience* 9. DOI: 10.3389/fncel.2015.00123.

Mammano, F. and R. Nobili (June 1993). "Biophysics of the cochlea: Linear approximation". In: *The Journal of the Acoustical Society of America* 93.6, pp. 3320–3332. ISSN: 0001-4966. DOI: 10.1121/1.405716.

MATLAB (2021). *version 9.11.0.1809720 (R2021b) Update 1*. Natick, Massachusetts: The MathWorks Inc.

Meddis, Ray (Mar. 1986). *Simulation of mechanical to neural transduction in the auditory receptor*. DOI: 10.1121/1.393460.

— (2011). *MAP*. URL: https://github.com/rmeddis/MAP.

Mehrgardt, S. and V. Mellert (June 1977). *Transformation characteristics of the external human ear*. DOI: 10.1121/1.381470.

Michalski, Nicolas et al. (Nov. 2017). "Otoferlin acts as a Ca2+ sensor for vesicle fusion and vesicle pool replenishment at auditory hair cell ribbon synapses". In: *eLife* 6. DOI: 10.7554/elife.31013.

Mistrík, Pavel and Jonathan F. Ashmore (July 2010). "Reduced Electromotility of Outer Hair Cells Associated with Connexin-Related Forms of Deafness: An In silico Study of a Cochlear Network Mechanism". In: *Journal of the Association for Research in Otolaryngology* 11.4, pp. 559–571. DOI: 10.1007/s10162-010-0226-3.

Mistrík, Pavel, Chris Mullaley, et al. (Aug. 2008). "Three-dimensional current flow in a large-scale model of the cochlea and the mechanism of amplification of sound". In: *Journal of The Royal Society Interface* 6.32, pp. 279–291. DOI: 10.1098/rsif.2008.0201.

Moser, Tobias et al. (Aug. 2006). "Hair cell ribbon synapses". In: *Cell and Tissue Research* 326.2, pp. 347–359. DOI: 10.1007/s00441-006-0276-3.

Motallebzadeh, Hamid et al. (May 2018). "Cochlear amplification and tuning depend on the cellular arrangement within the organ of Corti". In: *Proceedings of the National Academy of Sciences* 115.22, pp. 5762–5767. DOI: 10.1073/pnas.1720979115.

Niwa, Mamiko et al. (June 2021). "Functional subgroups of cochlear inner hair cell ribbon synapses differently modulate their EPSC properties in response to stimulation". In: *Journal of Neurophysiology* 125.6, pp. 2461–2479. DOI: 10.1152/jn.00452.2020.

Nobili, R. and F. Mammano (Apr. 1996). "Biophysics of the cochlea II: Stationary nonlinear phenomenology". In: *The Journal of the Acoustical Society of America* 99.4, pp. 2244–2255. DOI: 10.1121/1.415412.

Ohn, Tzu-Lun et al. (July 2016). "Hair cells use active zones with different voltage dependence of Ca^{2+} influx to decompose sounds into complementary neural codes". In: *Proceedings of the National Academy of Sciences* 113.32. DOI: 10.1073/pnas.1605737113.

Pascal, Jérôme et al. (Sept. 1998). "Linear and nonlinear model of the human middle ear". In: vol. 104. 3. Acoustical Society of America (ASA), pp. 1509–1516. DOI: 10.1121/1.424363.

Petit, Christine et al. (Dec. 2001). "Molecular Genetics of Hearing Loss". In: *Annual Review of Genetics* 35.1, pp. 589–645. DOI: 10.1146/annurev.genet.35.102401.091224.

Plack, Christopher J. and Andrew J. Oxenham (Jan. 2000). "Basilar-membrane nonlinearity estimated by pulsation threshold". In: *The Journal of the Acoustical Society of America* 107.1, pp. 501–507. DOI: 10.1121/1.428318.

Robles, Luis and Mario A. Ruggero (July 2001). "Mechanics of the Mammalian Cochlea". In: *Physiological Reviews* 81.3, pp. 1305–1352. DOI: 10.1152/physrev.2001.81.3.1305.

Rutherford, Mark A. (Mar. 2015). "Resolving the structure of inner ear ribbon synapses with STED microscopy". In: *Synapse* 69.5, pp. 242–255. DOI: 10.1002/syn.21812.

Salt, Alec N. et al. (Nov. 1989). "Calcium gradients in inner ear endolymph". In: *American Journal of Otolaryngology* 10.6, pp. 371–375. DOI: 10.1016/0196-0709(89)90030-6.

Santos-Sacchi, Joseph and Winston Tan (Apr. 2020). "Complex nonlinear capacitance in outer hair cell macro-patches: effects of membrane tension". In: *Scientific Reports* 10.1. DOI: 10.1038/s41598-020-63201-6.

Sasmal, Aritra and Karl Grosh (June 2019). "Unified cochlear model for low- and high-frequency mammalian hearing". In: *Proceedings of the National Academy of Sciences* 116.28, pp. 13983–13988. DOI: 10.1073/pnas.1900695116.

Shaw, E. A. G. (Dec. 1974). "Transformation of sound pressure level from the free field to the eardrum in the horizontal plane". In: *The Journal of the Acoustical Society of America* 56.6, pp. 1848–1861. DOI: 10.1121/1.1903522.

Strelioff, David (Sept. 1973). *A computer simulation of the generation and distribution of cochlear potentials*. DOI: 10.1121/1.1913642.

Suesserman, M.F. and F.A. Spelman (Mar. 1993). "Lumped-parameter model for in vivo cochlear stimulation". In: *IEEE Transactions on Biomedical Engineering* 40.3, pp. 237–245. DOI: 10.1109/10.216407.

Sumner, Christian J. et al. (2002). "A revised model of the inner-hair cell and auditory-nerve complex". In: *The Journal of the Acoustical Society of America* 111.5, p. 2178. DOI: 10.1121/1.1453451.

Tange, Ole (Apr. 2018). *GNU Parallel 2018*. Ole Tange. DOI: 10.5281/zenodo.1146014. URL: https://doi.org/10.5281/zenodo.1146014.

Vencovský, Václav et al. (May 2019). "The influence of distributed source regions in the formation of the nonlinear distortion component of cubic distortion-product otoacoustic emissions". In: *The Journal of the Acoustical Society of America* 145.5, pp. 2909–2931. DOI: 10.1121/1.5100611.

Verhulst, Sarah, Alessandro Altoè, et al. (Mar. 2018). "Computational modeling of the human auditory periphery: Auditory-nerve responses, evoked potentials and hearing loss". In: *Hearing Research* 360, pp. 55–75. DOI: 10.1016/j.heares.2017.12.018.

Verhulst, Sarah, Torsten Dau, et al. (Dec. 2012). "Nonlinear time-domain cochlear model for transient stimulation and human otoacoustic emission". In: *The Journal of the Acoustical Society of America* 132.6, pp. 3842–3848. DOI: 10.1121/1.4763989.

Vetešník, Aleš and Anthony W. Gummer (May 2012). "Transmission of cochlear distortion products as slow waves: A comparison of experimental and model data". In: *The Journal of the Acoustical Society of America* 131.5, pp. 3914–3934. DOI: 10.1121/1.3699207.

Vincent, P. F. Y. et al. (Aug. 2014). "Exocytotic Machineries of Vestibular Type I and Cochlear Ribbon Synapses Display Similar Intrinsic Otoferlin-Dependent Ca2+ Sensitivity But a Different Coupling to Ca2+ Channels". In: *Journal of Neuroscience* 34.33, pp. 10853–10869. DOI: 10.1523/jneurosci.0947-14.2014.

Vincent, Philippe F.Y. et al. (Dec. 2018). "Clustered Ca2+ Channels Are Blocked by Synaptic Vesicle Proton Release at Mammalian Auditory Ribbon Synapses". In: *Cell Reports* 25.12, 3451–3464.e3. DOI: 10.1016/j.celrep.2018.11.072.

Wang, Jing and Jean-Luc Puel (Jan. 2020). "Presbycusis: An Update on Cochlear Mechanisms and Therapies". In: *Journal of Clinical Medicine* 9.1, p. 218. DOI: 10.3390/jcm9010218.

Wang, Lu-Yang and George J. Augustine (Jan. 2015). "Presynaptic nanodomains: a tale of two synapses". In: *Frontiers in Cellular Neuroscience* 8. DOI: 10.3389/fncel.2014.00455.

Wong, A. B. et al. (June 2013). "Concurrent Maturation of Inner Hair Cell Synaptic Ca2+ Influx and Auditory Nerve Spontaneous Activity around Hearing Onset in Mice". In: *Journal of Neuroscience* 33.26, pp. 10661–10666. DOI: 10.1523/jneurosci.1215-13.2013.

Zampini, Valeria, Stuart L. Johnson, Christoph Franz, Marlies Knipper, et al. (Nov. 2014). "Fine Tuning of CaV1.3 Ca2+ Channel Properties in Adult Inner Hair Cells Positioned in the Most Sensitive Region of the Gerbil Cochlea". In: *PLoS ONE* 9.11. Ed. by Bernd Sokolowski, e113750. DOI: 10.1371/journal.pone.0113750.

Zampini, Valeria, Stuart L. Johnson, Christoph Franz, Neil D. Lawrence, et al. (Jan. 2010). "Elementary properties of CaV1.3 Ca2+ channels expressed in mouse cochlear inner hair cells". In: *The Journal of Physiology* 588.1, pp. 187–199. DOI: 10.1113/jphysiol.2009.181917.

From the ear flap to the nerve: A complete computer model of the peripheral auditory system **supplementary figures**

Ondrej Tichacek[1], Pavel Mistrík[2], and Pavel Jungwirth[1]

[1] *Institute of Organic Chemistry and Biochemistry of the Czech Academy of Sciences, Flemingovo náměstí 542/2, 160 00 Praha 6, Czech Republic*
[2] *MED-EL, Fürstenweg 77a, A-6020 Innsbruck, Austria*

July 22, 2022

This document contains supplementary figures for the article "From the ear flap to the nerve: A complete computer model of the peripheral auditory system". Figure S-1 shows that the model fulfills the zero-crossing condition in response to click stimuli of different sound levels. The figures S-2, S-3, and S-4 contain additional simulation results from the model of the ionic currents within the organ of Corti. The figure S-5 illustrates the difference between classic and stochastic description of mechanosensitive ion channels in the inner hair cells. The figure S-6 verifies, that the high-pass differential equation used for the cochlear amplifier does not affect the results for physiologically relevant frequencies.

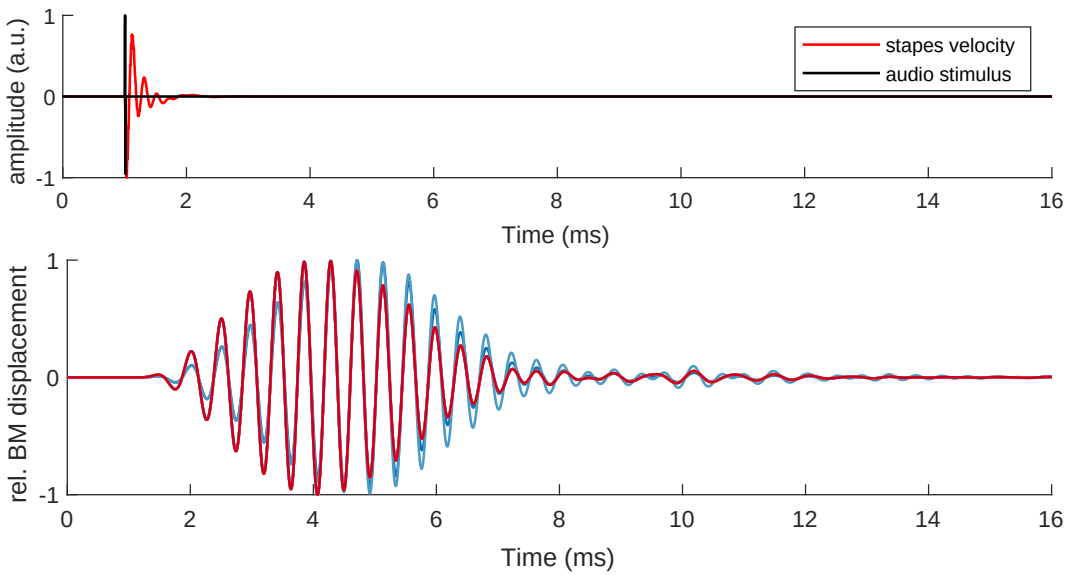

Figure S-1: Response of the electro-mechanical model to a click (22 us) of different levels from 0 to 80 dB SPL. **Top:** normalized stimulus waveform with the calculated stapes velocity. **Bottom:** normalized BM displacement at a characteristic position of 1 kHz (as per Li et al. (2021)). While the relative response changes slightly with amplitude, the zero crossing points remain virtually identical, which is consistent with experimental observations.

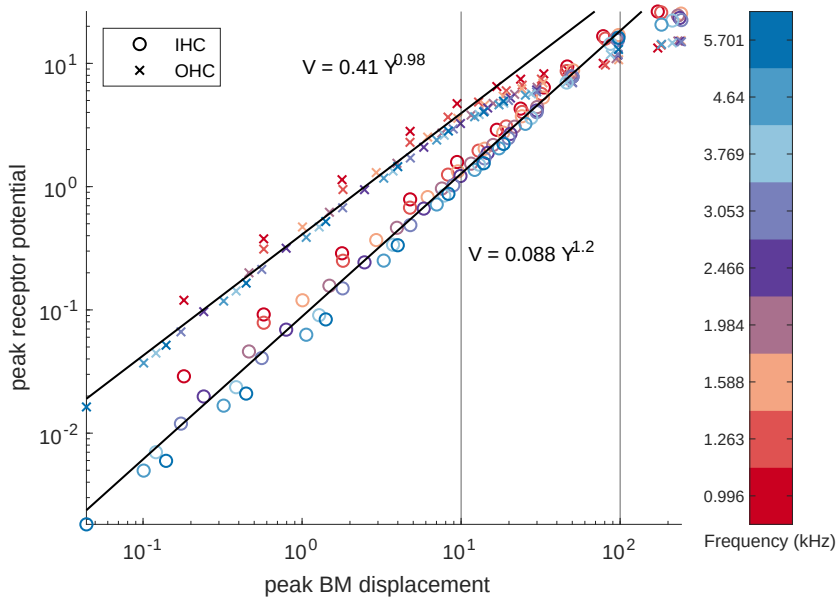

Figure S-2: Peak IHC and OHC receptor potential as a function of peak BM displacement (both at the best frequency location) extracted from a set of simulations of pure tones (1–5.7 kHz, 0–100 dB SPL).

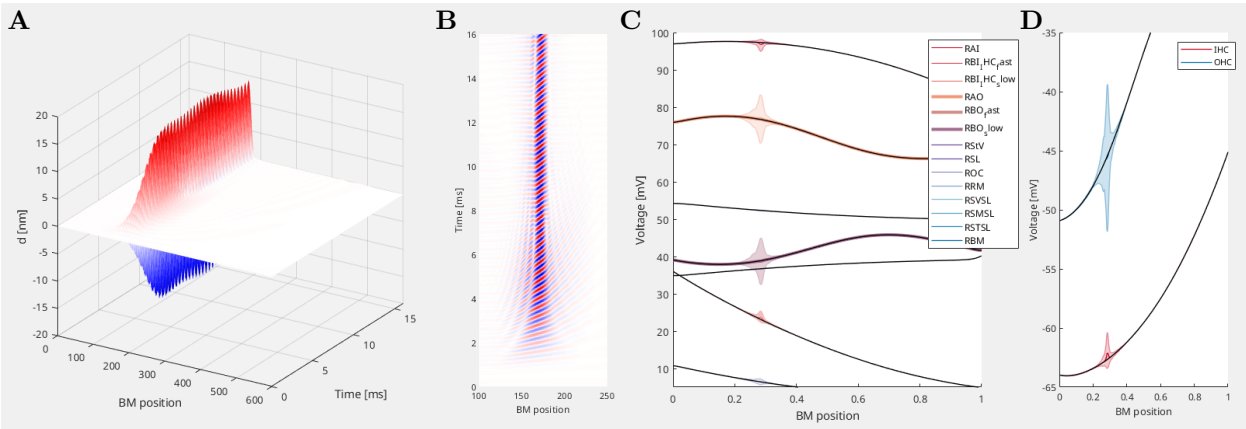

Figure S-3: Sample results of the mechanoelectrical model with a pure tone of 2466 Hz and 40 dB SPL. **A-B**: BM displacement as a function of time and position along the BM. The top view (panel B) shows sharpening of the response short after the stimulus onset. **C-D**: Voltage profiles across resistors in the system (C) and voltage across the basolateral IHC and OHC membrane (D). Steady state values in black, changes of sound evoked potentials in colored patches.

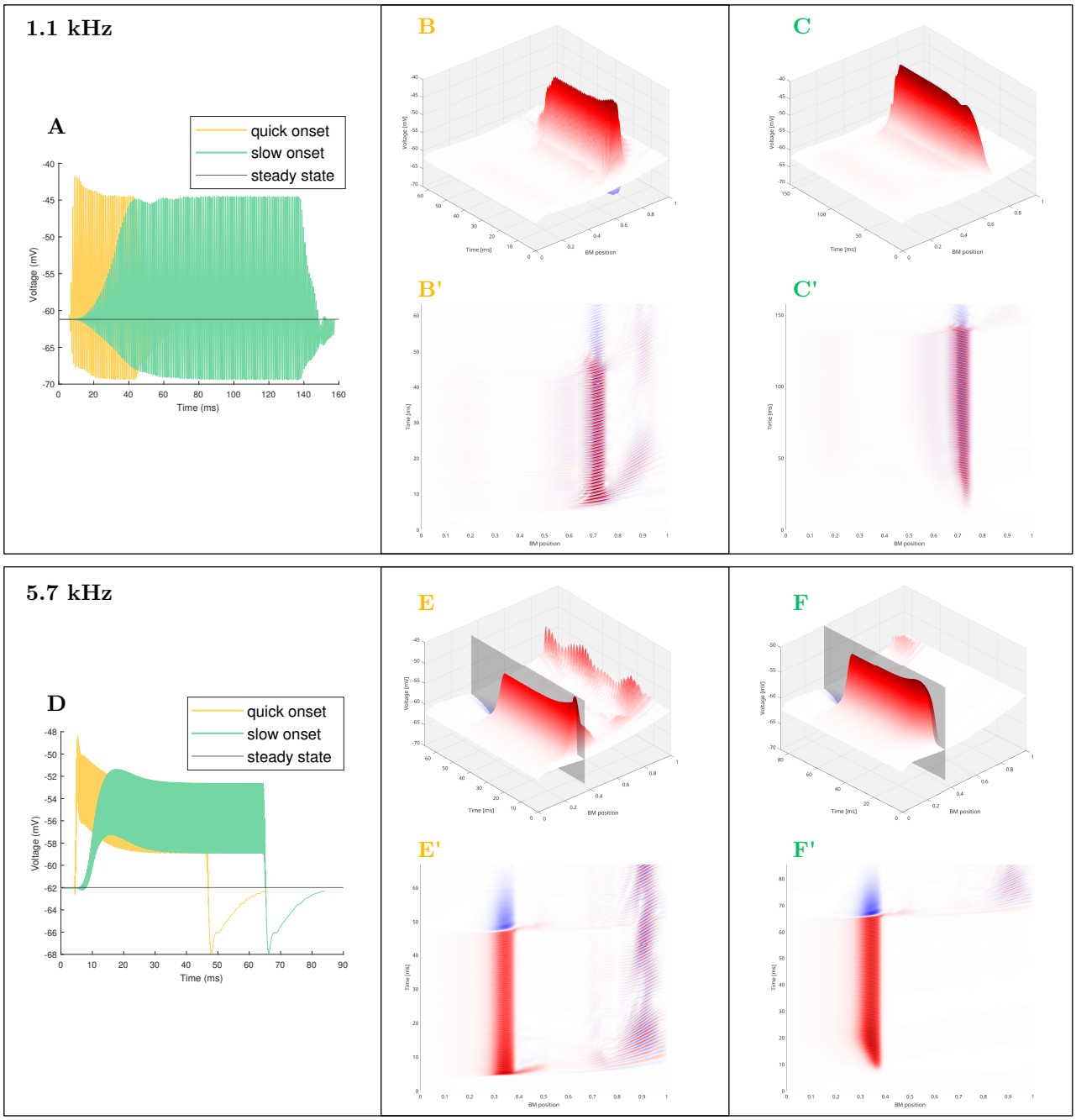

Figure S-4: The effect of stimulus onset slope: IHC basolateral transmembrane voltage from four simulations, 5701 and 1123 Hz, 50 dB SPL using a slow and quick onset. **A,D**: maximal cross-section, **B,C,E,F**: three-dimensional graph of the voltage, **B',C',E',F'**: "top view".

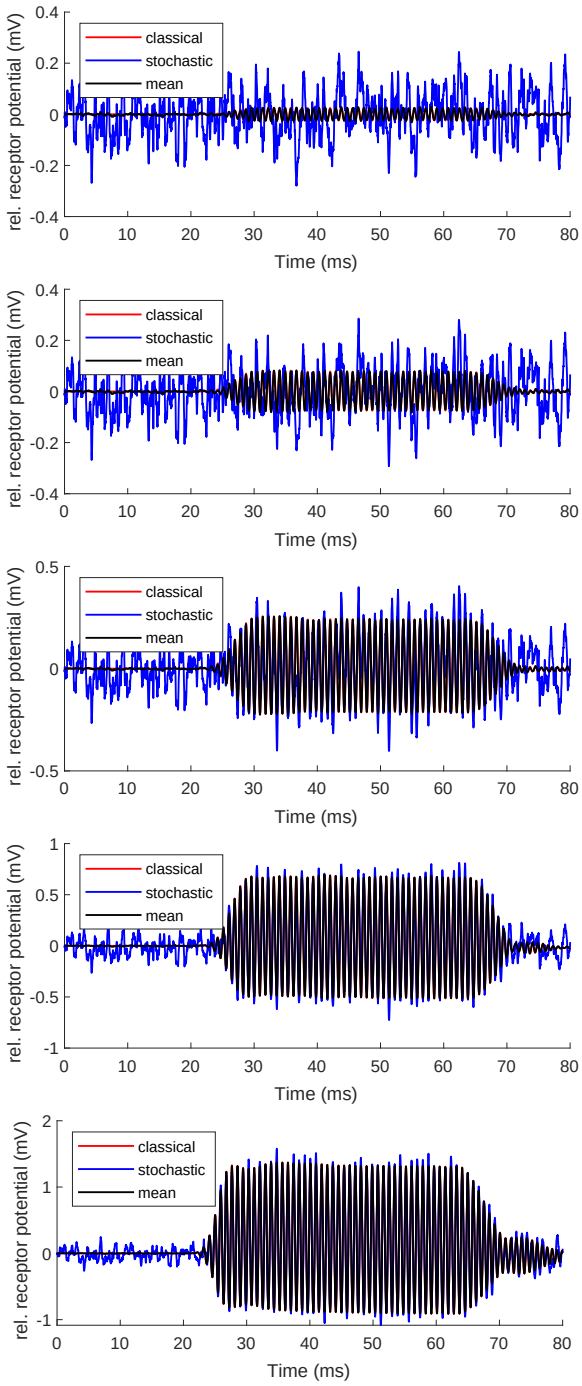

Figure S-5: Comparison of the classical and stochastic approach: IHC receptor potential for a 1.1 kHz pure tone of 0, 10, 20, 30, and 40 dB SPL. For low-level stimuli (up to 20-30 dB SPL), the potential of the stochastic model (blue) is significantly different from the continuous model (red) while its mean (1000 independent replicas, plotted black) is almost identical to the continuous model.

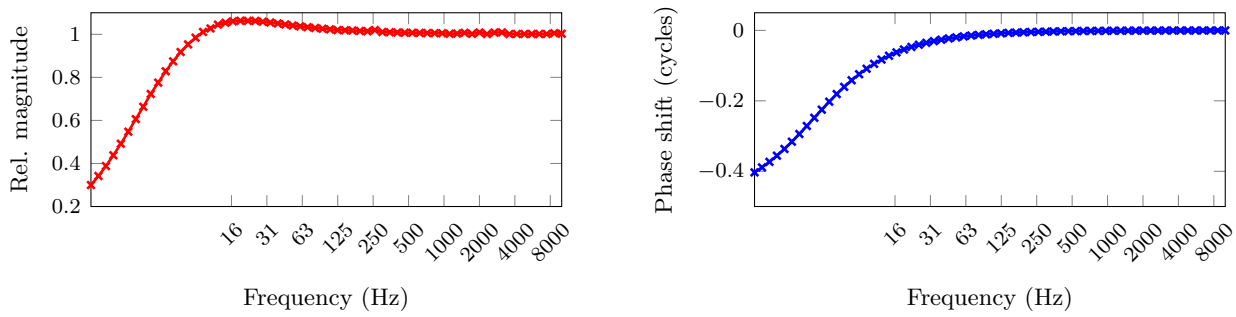

Figure S-6: Transfer function of the high pass filtering differential Equation 8. Above 100 Hz, the relative magnitude and phase of the force F_{ampl} is the same as the voltage V_{OHC}.