Evolutionary processes, not environmental drivers, determine the resilience of natural populations

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Abstract

Escalating climatic and anthropogenic pressures expose ecosystems worldwide to increasingly frequent disturbances. Yet, our ability to forecast the responses of natural populations to global change is impeded by a limited understanding of how exposure to environmental change shapes population resilience. Here, we test the association between local regimes of environmental change and the resilience to disturbance of 2,242 natural populations across 369 terrestrial and aquatic animal and plant species. Unexpectedly, we find that recent abiotic disturbance regimes (<50 years) do not predict the inherent ability of populations to resist and recover from disturbances. Instead, population resilience carries a strong evolutionary signal, with survival and developmental investments playing a key role in shaping population responses to disturbance. Contrary to assumptions that exposure to more frequent disturbances confers a greater ability to cope with future global change, population resilience relies on evolutionary processes that have shaped species characteristics over many generations.

Introduction

Predicting the outcomes of ongoing shifts in the timing and severity of environmental variability challenges ecologists to identify the processes underlying the capacity for populations to resist and recover from disturbances[1,2] (*i.e.*, their resilience[3]). However, half a century since Holling first defined resilience in ecological systems[3], we still do not know whether and how past environmental regimes shape the resilience of extant species[4]. Resolving this knowledge gap is pivotal for identifying those species most vulnerable to future increases in the frequency of disturbance regimes[5,6] and for designing effective ecosystem management strategies[7].

Resilience is far from a neglected concept[8,9], but approaches used to evaluate resilience often overlook its short-term nature[10]. Classic life-history theory argues that organisms operate under a strong trade-off coordinating their biological investment across survival and reproduction[11,12]. Indeed, considerable focus has been applied towards using the vital rates of survival, progression (*e.g.*, growth, development), retrogression (*e.g.*, shrinkage[13], rejuvenation[14]), and reproduction, to describe how these trade-offs then shape population performance in stochastic environments, characterised by frequent disturbances[15–19]. Yet, attention has still largely remained focused on the use of stochastic derivatives of long-term (*i.e.,* asymptotic) population characteristics (such as stochastic population growth rate, λ_s) as measures of population performance in the face of recurrent disturbances. Instead, disturbances, defined here as external (a)biotic forces with the capacity to modify the structural composition of natural populations, can displace populations into a transient (*i.e.*, short-term) phase during which their dynamics can vary considerably from their asymptotic trajectories[20] (Fig. 1A). Amid this transient phase, characteristics that inhibit short-term declines in population growth, or those promoting a short-term elevation are likely more beneficial than those maximising long-term performance[21].

Within stochastic environments, with populations exposed to repeated disturbances, it

is logical that natural selection would act upon heritable traits enhancing their overall

resilience[22,23]: increasing *resistance* (*i.e*., a population's ability to avoid a decline in size after

disturbance) and/or *compensation* (*i.e*., ability to increase in size after disturbance), and/or

shortening *time of recovery* (henceforth recovery; *i.e*., the time needed for converging back to

a stationary equilibrium after disturbance) following disturbances[24,25] (Fig. 1A). Long-term

measures of population performance cannot, therefore, reveal how environmental stochasticity

shapes the resilience of populations to disturbances[10]. Alternatively, metrics of short-term (*i.e.,*

transient) population dynamics, describing how the dynamics of populations within a transient

state can be expected to change relative to their long-term characteristics[26,27], can provide

greater insight into the resilience of natural populations[25,28,29]. These transient metrics thus

represent a unique opportunity for exploring the responses of natural populations to

disturbances (their resilience)[24,25], and how these might have been influenced by past-

disturbance regimes.

Here, to better understand the mechanisms underpinning resilience, we provide a global

assessment of the environmental and phylogenetic determinants of population resilience.

Specifically, we quantify the inherent capacity for resistance, compensation, and recovery of

2,242 natural populations across 61 animal, 305 plant, and 3 algae species from global

demographic databases[30,31] (Fig. 1B & Supplementary Table S1). Building on the expectation

that past disturbance regimes have shaped current resilience[23,32], we anticipate that (H1a)

exposure to a higher disturbance frequency will select for faster recovery, whilst (H1b)

exposure to broader spectra in environmental stochasticity selects for increased resistance and

reduced compensation. With the vital rates of survival, progression, retrogression, and

reproduction describing how trade-offs in individual-level fitness translates into population-

level characteristics[33], we expect that (H2) the effects of environmental stochasticity on

population resilience will manifest through the underlying vital rates of populations.

Results and Discussion

It is often assumed that past periodic exposure to disturbance will promote the resilience of

ecological systems towards future disturbances[4]. Here, however, using phylogenetically-

corrected partial least squares regression (pPLS) we found no relationship among the resilience

attributes of populations (*i.e.*, resistance, compensation, and recovery; Fig. 1A) and their

recent-past exposure to varying degrees of environmental stochasticity (Fig. 2). We used six

transient metrics to quantify resistance (first-step attenuation, $\underline{\rho}$ & maximal attenuation, $\underline{\rho}_{max}$),

compensation (reactivity, $\overline{\rho}$ & maximal amplification, $\overline{\rho}_{max}$), and recovery (damping ratio, ρ

& period of oscillation, ψ) in each population following disturbance[25,27,34] (Fig. 1A). Next, we

quantified the exposure of these populations to environmental stochasticity using measures of

mean thermal range (m), as well as the spectral frequency and autocorrelation of temperature

(β_T & a_T) and precipitation regimes (β_P & a_P) they experienced during the 50-years preceding

the beginning of each study (Supplementary S4). Overall, no measure of environmental

stochasticity constituted a strong predictor of variation in resistance, compensation, or recovery

($r^2 <0.001$ in all cases; Fig. 2).

The limited association between exposure to environmental stochasticity and the

resilience attributes of natural populations is further evident in phylogenetically-corrected

Pearson's tests of correlation between our transient metrics and our estimates of environmental

stochasticity ($|r| <0.015$ in all cases; Table 1). Importantly, this finding is insensitive to

phylogenetic imputation[35] (Fig. S3) and the length of time used to quantify recent-past

exposure to environmental stochasticity (Fig. S6 & S7). Our finding also persists regardless of

mean life expectancy (η_e), with both long- and short-lived populations displaying an equally

negligible influence of recent-past exposure to environmental stochasticity on their resilience

attributes (Fig. S5). This similarity occurs over equivalent 50-year intervals, even though short-

lived populations ($\eta_e \leq 10$ years) are likely to have had a larger number of generations exposed

to local stochasticity, thereby offering greater opportunity for adaptive change.

Resistance and compensation, but not recovery, are however, determined by selection

pressures shaping their energetic investments across somatic maintenance/development, and

reproduction. Using pPLS across our 2,242 populations, we evaluated the relationship between

our six measures of demographic resilience (Fig. 1A) and their sensitivities to each of the vital

rates of survival (σ), progression (γ), retrogression (τ), and fecundity (φ; Fig. 3). These vital

rate sensitivities reflect how much each transient metric would change following alterations in

each vital rate[34]. Thus, these sensitivities highlight how investments into any one vital rate

influence a populations' capacity to resist, compensate, and recover following a disturbance,

and provide a measure of the absolute importance of each vital rate in shaping demographic

resilience. We focused on sensitivities here, rather than elasticities (proportional

sensitivities[36]), as they provide a closer representation of selection gradients across populations

possessing traits with negative values or no natural zero level[37]. Patterns across the vital rate

sensitivities of each transient characteristic corroborate recent evidence of an association

between population resistance and compensation[38], revealing that the resistance and

compensation attributes of populations are constrained by individual-level investments on

survival, progression, retrogression, and reproduction, whilst there is no evidence for

demographic selection pressures on the attribute of recovery (Fig. 3 & Table 2). Again, these

findings are insensitive to phylogenetic imputation (Fig. S4).

Using estimates of phylogenetic signal (Pagel's λ[39]; not to be confused with long-term

population growth rate, λ) we further demonstrate a strong phylogenetic signal (Pagel's $\lambda >$

0.94 & p <0.001) across the transient metrics of resistance (first-timestep attenuation &

maximal attenuation) and compensation (reactivity & maximal amplification), and their

corresponding vital rate sensitivities (Table 3). Phylogenetic signal reflects the proportion of

variation in a trait that can be explained by species' shared evolutionary history, and so

represents the extent to which evolution constraints the expression of that trait[39,40] (here the

ability of a natural population to respond to disturbances). That resistance, compensation, and

recovery have evolved regardless of whether populations endure frequent disturbances or not

is indicative of indirect selection and may undermine the capacity for many populations to

persist with future increases in environmental stochasticity. Indirect selection occurs when

selective forces operate simultaneously upon multiple traits[22,41,42]. When population

characteristics rely on similar underlying mechanisms, such as enhanced individual survival,

directional selection can promote covariance across a combination of trait characteristics[22].

Accordingly, non-adaptive and maladaptive trait characteristics can appear contrary to

expected selection pressures due to responses driven by selective forces acting upon other trait

characteristics[42].

Caution is necessary, however, when interpreting observations regarding the selection

pressures maintained by environmental stochasticity. Our exploration into the environmental

drivers of demographic resilience focuses only on terrestrial populations. Physiological

adaptations, such as thermoregulation, that serve to broaden abiotic tolerance, are more

prevalent across terrestrial taxa, whilst the ability to seek out tolerable microclimates further

reduces the susceptibility of terrestrial populations to environmental stochasticity[43]. Marine

species often inhabit conditions close to their physiological limits, rendering them more

sensitive to abiotic shifts than terrestrial species[43,44]. More importantly, though, here we

quantify past exposure to environmental stochasticity using maximum exposure legacies of

100-years. It is plausible, however, that deep-time environmental regimes would offer greater

predictive potential. Indeed, deep-time studies have demonstrated how the latitudinal diversity

gradient, a prominent pattern underlying contemporary ecological understanding, has only

persisted during the past 30 million years[45]. Whilst environmental stochasticity is known to

influence population dynamics[46], its observable effects on population characteristics can

remain negligible until compounded by external factors such as changing habitat

configurations[47]. Any direct impacts of stochasticity on the resilience of natural populations

may, therefore, become more detectable with time.

Despite the potential influence of historical climate legacies on the resilience of

populations, we illustrate how our observations here persist even when explicitly accounting

for lifespan and legacy time. Our findings illustrate that through evolutionary time, adaptation

plays an important role in the development of resilience traits. Adaptation is the accumulation

of beneficial, heritable characteristics within a population over multiple generations[48].

Therefore, populations exposed to specific conditions over extended timeframes, or those that

turn over multiple generations within a short period of time, are more likely to display trait

characteristics adapted to those conditions[49–51]. To evaluate the implications of differing

exposure periods on our observations, we first repeated our pPLS analyses exploring the

relationship between demographic resilience and environmental stochasticity focusing on only

populations of short-lived species (Supplementary S6). We determined population longevity

using estimates of mean life expectancy (η_e), with populations of $\eta_e \leq$10-years defined as short-

lived (n = 1,606 populations). Next, we repeated our analyses using only populations for which

100-year abiotic legacies could be sourced (Supplementary S7). Neither approach improved

our ability to predict the demographic resilience attributes of natural populations using

measures of environmental stochasticity (Figs. S5, S6 & S7).

Conclusions

Climate shifts and anthropogenic activities are impacting upon the periodicity of phenological

drivers[52,53], and reducing the return times between severe disturbance events[54,55]. Natural

populations worldwide are thus being exposed to increasingly stochastic environments, with

many facing imminent collapse[56]. Here, we illustrate, however, that the past exposure of natural

populations to environmental stochasticity does not limit, nor guarantee, their resilience

towards future disturbances. [1,2]Resilience is a complex and heavily debated concept[9,24,57]. Yet,

the realisation that population resilience is mediated by the adaptation of species over

evolutionary timeframes, not the conditions experienced by populations following their

dispersal, is not trivial. Rather, this understanding enables attention to be better focused

towards identifying populations with high adaptive potential, whilst maintaining the high

genetic diversity necessary to enhance the resilience of natural populations.

Sourcing climate records at the necessary temporal and spatial resolution for

quantifying deep-time environmental legacies presents a considerable challenge[58,59]. Thus, the

susceptibility, and resilience, of species to future global change is more often inferred from

their exposure to more contemporary conditions[60–62]. It is necessary, therefore, to highlight the

limitations associated with using the recent exposure of populations to environmental

stochasticity to predict their continued resilience. Instead, we argue that selective forces acting

upon other adaptive population characteristics have indirectly refined the ability for natural

populations to resist, compensate, and recover from disturbances. Selection pressures constrain

how individuals allocate finite resources across survival, somatic development, and

reproduction, thus mediating the capacity for populations to exploit and prevail within their

local environments[12,22]. Over time, these selective forces have moulded the resilience attributes

of populations, but not necessarily in response to the current disturbance regimes they endure.

Methods

Demographic data extraction & transient indices

To evaluate the selection pressures on the resilience attributes of natural populations, we

extracted Matrix Population Models (MPMs) from the open-source COMPADRE Plant Matrix

Database[31] (v. 5.0.1) and COMADRE Animal Matrix Database[30] (v. 3.0.1). MPMs are discrete

time, structured population models where the lifecycle is categorised into discrete state classes

(*i.e.*, age, size, and/or developmental stages)[34]. Combined, COMPADRE and COMADRE

contain over 12,000 MPMs from more than 1,100 animal and plant species. However, here we

only retained MPMs satisfying the following six criteria to test our hypotheses: (1) MPMs

reflecting the demographic characteristics of individual populations recorded across a single

time period (*Individual* MPMs). We did, however, select MPMs consisting of demographic

information averaged across multiple populations and/or time periods (*mean* MPMs), for

populations for which no individual matrices were available (330 populations after applying

the additional criteria below); (2) MPMs based on annual surveys to ensure all subsequent

metrics obtained reflected identical units of time, thus allowing for their comparability; (3)

MPMs representing wild, un-manipulated populations, to guarantee investigating the selection

pressures underpinning the resilience of natural populations and the possibility to link their

dynamics to their local environmental regimes; (4) MPMs comprised of three or more life

stages, as lower dimension MPMs typically lack the necessary resolution for estimating vital

rates[63] and transient dynamics[64]; (5) MPMs from populations with known latitude and

longitude information to allow us to link their demographic properties to local environmental

regimes; and finally (6) MPMs describing full life cycles (e.g., no missing data on survival,

progression, retrogression, and reproduction) to ensure the correct calculation of vital rates and

transient metrics. Following these criteria, we retained 3,890 MPMs corresponding with 3,204

populations across 438 plant species, 665 populations across 112 animal species, and 21

populations across six algae species (Supplementary Table S1).

We further refined our list of MPMs according to their transient, asymptotic, and

species-specific properties. All MPMs were tested for irreducibility (*i.e.,* all life cycle stages

are either directly, or indirectly connected to one another), ergodicity (*i.e.,* asymptotic

dynamics are independent of the initial population structure), and primitivity (*i.e.,* MPMs

consist of non-negative elements)[34]. We carried out these tests using the corresponding

isErgodic, *isIrreducible*, and *isPrimitive*, diagnostic functions from the R package

'*popdemo*'[65]. A total of 1,203 reducible, imprimitive, and/or non-ergodic MPMs were excluded

from further analyses on the basis that they represent untenable life cycles that defy logical

biological processes[66]. MPMs with population growth rates $\lambda>2$, indicating that the population

is projected to increase two-fold or more every year, were also rejected as they represent

unlikely realisations of population performance in our experience. Equally, MPMs from highly

migratory (e.g., > 1,000 km) species were discarded, since their vital rate schedules are unlikely

to be mostly shaped by the environment in which they were measured. We also note here that,

across our initial population sample, the vital rate of clonality (κ) was rare, with only 140

populations across 37 plant species, and two populations from one animal species

(*Amphimedon compressa*[67]) explicitly exhibiting this demographic process. Thus, to focus our

analyses on common demographic currencies, we excluded all populations exhibiting clonality.

Overall, our strict selection criteria resulted in a final sample of 2,242 MPMs, corresponding

with 369 species: 402 populations from 61 animal species, 1,830 populations from 305 plant

species, and 10 populations from three species of algae (Supplementary Table S1).

For each retained MPM, we calculated six transient metrics quantifying each

population's potential for demographic resistance (first-timestep attenuation, $\underline{\rho}$ & and maximal

attenuation, $\underline{\rho}_{max}$), compensation (reactivity, $\overline{\rho}$ & maximal amplification, $\overline{\rho}_{max}$), and recovery

(damping ratio, ρ & period of oscillation, ψ), following a disturbance[25] (Fig. 1). Firstly, with

estimates of transient dynamics known to be contingent on the reproductive strategies of

populations, it was necessary to convert all post-reproductive matrices into a pre-reproductive

format by adjusting patterns of reproduction to include a measure of adult survival[68]. All MPMs

were then standardised to separate their transient and asymptotic properties by dividing each

matrix element by the MPM's dominant eigenvalue, λ[26,34]. Following standardisation, from

each MPM we obtained estimates of reactivity and first-timestep attenuation as the upper and

lower bounds returned by the function *reac*, and calculated maximal amplification and maximal

attenuation indices using their respective functions *maxamp* and *maxatt*, all within the R

package '*popdemo*'[65]. Meanwhile, we calculated the damping ratio of each MPM using

eigen.analysis from the '*popbio*' package[69]. Finally, we calculated the period of oscillation (ψ)

for each MPM using its subdominant eigenvalue (λ_2) (equation 1)[34].

$$\psi = \frac{2\pi}{\theta_2} \qquad \text{Where: } \theta_2 = \tan^{-1}\left(\frac{I(\lambda_2)}{R(\lambda_2)}\right) \tag{1}$$

To explore how the fitness components of individuals mediate the selection gradients

placed on demographic resilience by environmental variability, we calculated the sensitivity of

each transient metric towards the vital rates of survival (σ), progression (γ), retrogression (τ),

and fecundity (φ). For each MPM, we first estimated all vital rate sensitivities from their

element-level constituents. Individual elements a_{ij} within the MPM **A** typically describe

combinations of multiple vital rates[70]. Subsequently, calculating the sensitivity of each

transient metric (s_x) with respect to underlying vital rates requires the decomposition of

element-level sensitivities into their vital rate components[70]. Briefly, this decomposition

requires the estimation of stage-specific survival probabilities (σ_j) for each MPM. These

estimates of σ_j are then used to determine the proportion of each matrix element a_{ij}

corresponding with survival (σ), progression (γ), retrogression (τ), and fecundity (φ)[70].

Accordingly, we initially calculated the sensitivity of each transient metric at the matrix

element-level (s_{ij}). The sensitivities of the damping ratio (ρ) and period of oscillation (ψ) were

determined using equations 2 & 3, with s_{ij}^1 and s_{ij}^2 as the matrix element-level sensitivity

matrices of the dominant and subdominant eigenvalues, respectively[34].

$$s_{\rho_{ij}} = \frac{1}{|\lambda_2|}\left(s_{ij}^1 - \frac{\rho}{|\lambda_2|}\left[R(s_{ij}^2) + I(s_{ij}^2)\right]\right) \tag{2}$$

$$s_{\psi_{ij}} = \frac{-2\pi}{\theta_2|\lambda_2|^2}\left[I(s_{ij}^2) - R(s_{ij}^2)\right] \tag{3}$$

The sensitivities of reactivity, amplification, and attenuation (s_{ij}) with respect to

element a_{ij} were then estimated as the magnitude of change (δ) in each transient metric (x)

following a small change (here 0.01) in a_{ij} (Equation 4)[71].

$$s_{ij} = \frac{\delta x}{\delta a_{ij}} = \frac{x_{Perturbed} - x_{Original}}{a_{ij}^{Perturbed} - a_{ij}^{Original}} \tag{4}$$

The distributions of each transient metric and its corresponding vital rate sensitivities

were checked prior to subsequent regression analyses. To ensure normality across each

distribution, outliers, defined as values outside the 95% confidence intervals of the distribution,

were omitted, with the remaining estimates then transformed if necessary. For each transient

metric, power transformations (y^x) were used to achieve approximate normality using the Box-

Cox transformation functions of the R package '*caret*'[72] to estimate *x*. The distributions of

damping ratio, period of oscillation, reactivity, and maximal amplification raised negative *x*

values, and so their transformations took the form *1/y^{|x|}*. Inverse and log transformations were

also necessary for several of the vital rate sensitivity variables (See Table S2 for further details).

Phylogenetic correction

Evaluating the selection pressures exerted on attributes of demographic resilience across

multiple species requires an explicit consideration for how traits are expected to covary due to

phylogenetic relationships[40,73,74]. To account for such relationships in our analyses, we

constructed a population-within-species level phylogenetic tree using phylogenetic data

extracted from the Open tree of Life for the animal, plant, and algae species across our

population sample (OTL[75]; Supplementary S3). Our approach here also allowed us to

accommodate studies that included multiple, separate populations for the same species (*see*

below). Firstly, the scientific names of each species associated with our extracted MPMs were

checked against current taxonomy records using the R package '*taxize*'[76]. Next, we extracted

information regarding the taxonomic classification and phylogeny of each species from the

OTL database with the R package '*rotl*'[77]. Subsequently, using the '*ape*'[78] and '*phytools*'

packages[79], this phylogenetic information was used to construct a species-level phylogenetic

tree corresponding with the 369 unique species within our MPM list.

Beyond accounting for phylogenetic signals in trait variance-covariance across our

population sample, it was necessary to ensure that our phylogenetic tree reflected the influence

of spatial signals in the development of traits within species. Thus, we expanded our

phylogenetic tree by adding branch tips to incorporate multiple population entries per species

(*sensu*[80]), generating a population-level tree comprising our full sample of 2,242 populations

(Supplementary S3). Finally, we computed the branch lengths for our phylogenetic tree using

the function *compute.brlen* in the R package '*ape*'[78]. These branch lengths were estimated

using Grafen's arbitrary branch lengths[81], assuming a Brownian motion model with the

variance between species directly proportional to time since divergence[82]. Importantly, we

constrained branch lengths between populations of the same species to approximately zero

(0.0000001) under the assumption of negligible phylogenetic distance between species

replicate populations.

Quantifying environmental variability

To investigate the role of environmental selection pressures on the resistance, compensation,

and recovery components of resilience in natural populations, we used a pPLS regression

exploring the association between transient characteristics and metrics of environmental

variability. We quantified the magnitude and frequency of environmental variation to which

each population was exposed using the GPS location information extracted with each MPM

from COMPADRE & COMADRE[30,31] (Supplementary S4). Since temperature and

precipitation rates are universal drivers of biological community assembly across terrestrial

environments[83], we selected data describing temporal trends in thermal and precipitation

regimes as a measure of the environmental variability experienced by each population.

Crucially, however, with precipitation not directly influencing marine environments (although

see[84]), we excluded marine populations (29 populations from six animal species, and 10

populations from three algal species) from this portion of our analyses.

We quantified environmental variance through the metrics of autocorrelation, abiotic

range, and frequency spectrum using long-term temperature and precipitation records sourced

from the CHELSA climate database[85]. For each population, we extracted monthly records of

maximum and minimum temperatures (°C) and mean precipitation rates (kg m^{-2})

corresponding with the specific period during which the population was surveyed as detailed

in COMPADRE and COMADRE, plus an additional 50-years prior to the onset of censusing

to account for environmental legacy effects[86]. We condensed maximum and minimum

temperature readings into monthly estimates of mean temperatures and thermal range. Next, as

a gauge of disturbance magnitude (m), we estimated the mean thermal range experienced by

each population across their associated temporal records. We then arranged our monthly mean

temperature and precipitation estimates into time series depicting the temporal environmental

regimes to which each population was exposed. Using the '*colorednoise*' package[87], we

calculated the autocorrelation of each temperature (a_T) and precipitation (a_P) time series as a

measure of environmental predictability[88].

The colour of environmental variation is depicted on a red to blue colour scale, from

lower to higher frequencies, respectively[88]. The frequency spectrum of a time series is

expressed by its spectral exponent (β), which is calculated as the negative slope of the linear

regression between the log spectral density and log frequency of the time series[89]. We

calculated the frequency spectrum of each precipitation (β_P) and temperature (β_T) time series

as an indicator of the colour of environmental variation experience by each population[89]. The

spectral exponent for each time series was estimated using the spectrum function from the R

package '*stats*'[90].

Partial Least Squares Regression

We utilised a phylogenetically corrected Partial Least Squares regression (pPLS) framework

to test our hypothesis that the resistance, compensatory, and recovery of natural populations

correspond with gradients in environmental variability and evaluate how this is mediated by

fitness investments. Using a pPLS, we evaluated the relationship between our estimates of

population resilience, and both associated environmental variability regimes and their vital rate

sensitivities. The pPLS technique is considered a more powerful comparative tool than other

available multivariate regression methods[91], as it simultaneously condenses the variation

among numerous predictors whilst maximising the variance explained among response

variables. Subsequently, we investigated the selection pressures on the resistance,

compensation, and recovery attributes of natural populations, and therefore the capacity for

environmental legacies, and vital rate characteristics, to serve as predictors of resilience

attributes.

We first applied a phylogenetically-corrected Pearson's test of correlation and pPLS to

analyse the correlation between environmental variability and transient demographic

characteristics. This approach enabled us to test for covariation between the transient

characteristics of populations and gradients in their exposure to environmental variability.

pPLS tests were carried out for each transient measure with the predictor variable set comprised

of our five metrics of environmental variability. From each test, we then extracted component

scores and loadings, which describe the arrangement of the environmental predictor variables

within a multivariate space. We also obtained the percentage variance (*%var*) among the

predictors explained by each regression component and the proportion of variance in the

transient response variable explained by each component (r^2) to estimate the strength of any

association between environmental variability and the transient dynamics of our population

sample.

Phylogenetically corrected correlation tests and pPLS analyses were used to examine

for patterns between each transient characteristics and its associated vital rate sensitivities.

Again, test coefficients (r), component scores, loadings, *%var*, and r^2 values were calculated

to quantify the influence of the fitness components of survival, progression, retrogression, and

reproduction towards the transient characteristics of natural populations. All pPLS analyses

were conducted using the *'pls'* R package[92], with modifications included to ensure our analyses

accounted for any evolutionary covariance between the sensitivity patterns and transient

characteristics of different populations[79,93,94].

We carried out all pPLS analyses using only complete entries, omitting populations

missing estimates for any one variable. To provide further clarity regarding any patterns we

observed between our measures of population resilience, their environmental legacies, and their

vital-rate sensitivities, we repeated each analysis twice. During these repeated tests, we first

evaluated whether considering the life expectancies of populations influenced any observed

patterns (Supplementary S6). Within a given time period, long-lived species will likely

experience fewer generations than shorter-lived species diminishing the relative impact of

existing selection pressures on their trait characteristics[95]. Thus, it was necessary to ensure that

the inclusion of long-lived species within our population sample did not limit our capacity for

exploring environmental selection pressures. We categorised each population within our

sample as either long- or short-lived according to their associated mean life expectancy (η_e),

calculated from each extracted MPM using the R package '*IPMpack*'[96]. Next, we repeated our

pPLS analyses using only 'short-lived' populations for which $\eta_e \leq 10$ years (n = 1606

populations). This threshold was selected as a balance between maximising the number of

generations experienced by populations during the multi-year time series used in calculating

environmental legacies and maximising our sample size.

Understanding patterns in the responses of natural populations to environmental change

is often impeded by gaps in available data linking population characteristics to abiotic

regimes[97]. To ensure that missing data was not obscuring any correlation between our measures

of population resilience and environmental stochasticity we repeated our analyses using

phylogenetic imputation to estimate any demographic measures of resilience missing across

our population sample[35] (Supplementary S5). We calculated the phylogenetic signal (Pagel's

λ[39]) of each transient and sensitivity variable using the *phylosig* function from the '*phytools*'

package[79]. Pagel's λ exists on the scale $0 < \lambda > 1$, with 0 indicating traits have evolved

independently of phylogeny, and 1 representing a high phylogenetic signal[39]. For any variable

exhibiting a strong phylogenetic signal (*i.e.,* Pagel's $\lambda \geq 0.65$) that differs significantly from

zero ($p < 0.05$), we imputed all missing values assuming a Brownian motion evolutionary

model, before repeating our pPLS analyses.

Acknowledgments

Funding for this research was provided by a DTP Natural Environment Research Council

Scholarship to JC, a NERC IRF (NE/M018458/1) to RS-G, a Ramon Areces Foundation

Postdoctoral Fellowship to PC, and Winifred Violet Scott Estate funding to MB. The authors

would also like to acknowledge the University of Leeds High Performance Computing (ARC)

team who maintain the facilities used for the extraction of extensive climate records from the

CHELSA database (https://chelsa-climate.org/).

Author Contributions

JC, RS-G, MB, and PC conceived the ideas and methodology for the project. JC, PC, and RS-

G carried out data extraction and analysis. JC led the writing of the manuscript, with all authors

contributing critically to the writing and giving final approval for publication.

Data Accessibility

All Matrix Projection Models used within this study are available from the COMADRE and COMPADRE demographic databases (https://compadre-db.com/). Equally, all taxonomic information used in the construction of our phylogenetic trees can be sourced from the Open Tree of Life (https://tree.opentreeoflife.org/), and all temperature, and precipitation records used in this study can be downloaded from the CHELSA climate database (https://chelsa-climate.org/). Finally, the R code used for all the data extraction and analyses presented throughout this manuscript have been uploaded to GitHub and will be made publicly available following publication.

References

1. Standish, R. J. *et al.* Resilience in ecology: Abstraction, distraction, or where the action is? *Biological Conservation* **177**, 43–51 (2014).

2. Angeler, D. G. & Allen, C. R. Quantifying resilience. *Journal of Applied Ecology* **53**, 617–624 (2016).

3. Holling, C. S. Resilience and stability of ecological systems. *Annual Review of Ecology and Systematics* **4**, 1–23 (1973).

4. Walker, B. H. Resilience: what it is and is not. *Ecology and Society* **25**, 1–3 (2020).

5. Gaillard, J. C. Vulnerability, capacity and resilience: Perspectives for climate and development policy. *Journal of International Development* **22**, 218–232 (2010).

6. McLeod, E., Shaver, E. C., Beger, M., Koss, J. & Grimsditch, G. Using resilience assessments to inform the management and conservation of coral reef ecosystems. *Journal of Environmental Management* **277**, 1–8 (2021).

7.   Pressey, R. L., Cabeza, M., Watts, M. E., Cowling, R. M. & Wilson, K. A.

Conservation planning in a changing world. *Trends in Ecology and Evolution* **22**, 583–

592 (2007).

8.   Capdevila, P. *et al.* Reconciling resilience across ecological systems, species and

subdisciplines. *Journal of Ecology* **109**, 3102–3113 (2021).

9.   Kéfi, S. *et al.* Advancing our understanding of ecological stability. *Ecology Letters* **22**,

1349–1356 (2019).

10. Cant, J., Salguero-Gómez, R. & Beger, M. Transient demographic approaches can

drastically expand the toolbox of coral reef science. *bioRxiv* 2021.04.08.438926 (2021)

doi:10.1101/2021.04.08.438926.

11. Stearns, S. C. *The Evolution of Life Histories*. (Oxford University Press, 1992).

12. Stearns, S. C. Trade-Offs in Life-History Evolution. *Functional Ecology* **3**, 259–268

(1989).

13. Salguero-Gómez, R. & Casper, B. B. Keeping plant shrinkage in the demographic

loop. *Journal of Ecology* **98**, 312–323 (2010).

14. Salguero-Gómez, R., Shefferson, R. P. & Hutchings, M. J. Plants do not count... or do

they? New perspectives on the universality of senescence. *Journal of Ecology* **101**,

545–554 (2013).

15. Boyce, M. S. *et al.* Demography in an increasingly variable world. *Trends in Ecology*

and Evolution **21**, 141–148 (2006).

16. Jonzén, N., Pople, T., Knape, J. & Sköld, M. Stochastic demography and population

dynamics in the red kangaroo Macropus rufus. *Journal of Animal Ecology* **79**, 109–116

(2010).

17.   Tuljapurkar, S. D. Population dynamics in variable environments. II. Correlated

environments, sensitivity analysis and dynamics. *Theoretical Population Biology* **21**,

114–140 (1982).

18.   Tuljapurkar, S. D. & Orzack, S. H. Population dynamics in variable environments I.

Long-run growth rates and extinction. *Theoretical Population Biology* **18**, 314–342

(1980).

19.   Tuljapurkar, S., Gaillard, J. M. & Coulson, T. From stochastic environments to life

histories and back. *Philosophical Transactions of the Royal Society B* **364**, 1499–1509

(2009).

20.   Hastings, A. Transient dynamics and persistence of ecological systems. *Ecology*

Letters **4**, 215–220 (2001).

21.   McDonald, J. L., Stott, I., Townley, S. & Hodgson, D. J. Transients drive the

demographic dynamics of plant populations in variable environments. *Journal of*

Ecology **104**, 306–314 (2016).

22.   Lande, R. & Arnold, S. J. The measurement of selection on correlated characters.

Evolution (N Y) **37**, 1210–1226 (1983).

23.   Boyd, P. W. *et al.* Biological responses to environmental heterogeneity under future

ocean conditions. *Glob Chang Biol* **22**, 2633–2650 (2016).

24.   Hodgson, D., McDonald, J. L. & Hosken, D. J. What do you mean, "resilient"? *Trends*

in Ecology and Evolution **30**, 503–506 (2015).

25.   Capdevila, P., Stott, I., Beger, M. & Salguero-Gómez, R. Towards a comparative

framework of demographic resilience. *Trends in Ecology & Evolution* **35**, 776–786

(2020).

26.   Koons, D. N., Grand, J. B., Zinner, B. & Rockwell, R. F. Transient population

509   dynamics: Relations to life history and initial population state. *Ecological Modelling*

510   **185**, 283–297 (2005).

511   27.   Stott, I., Townley, S. & Hodgson, D. A framework for studying transient dynamics of

512   population projection matrix models. *Ecology Letters* **14**, 959–970 (2011).

513   28.   Hastings, A. Transients: The key to long-term ecological understanding? *Trends in*

514   *Ecology and Evolution* **19**, 39–45 (2004).

515   29.   Hastings, A. *et al.* Transient phenomena in ecology. *Science (1979)* **361**, 1–9 (2018).

516   30.   Salguero-Gómez, R. *et al.* COMADRE: A global data base of animal demography.

517   *Journal of Animal Ecology* **85**, 371–384 (2016).

518   31.   Salguero-Gómez, R. *et al.* The COMPADRE Plant Matrix Database: An open online

519   repository for plant demography. *Journal of Ecology* **103**, 202–218 (2015).

520   32.   Rivest, E. B., Comeau, S. & Cornwall, C. E. The role of natural variability in shaping

521   the response of coral reef organisms to climate change. *Current Climate Change*

522   *Reports* **3**, 271–281 (2017).

523   33.   Metcalf, C. J. E. & Pavard, S. Why evolutionary biologists should be demographers.

524   *Trends in Ecology and Evolution* **22**, 205–212 (2007).

525   34.   Caswell, H. *Matrix population models: Construction, analysis and interpretation*.

526   (Oxford University Press Inc, 2001).

527   35.   James, T. D., Salguero-Gómez, R., Jones, O. R., Childs, D. Z. & Beckerman, A. P.

528   Bridging gaps in demographic analysis with phylogenetic imputation. *Conservation*

529   *Biology* **35**, 1210–1221 (2021).

530   36.   de Kroon, H., van Groenendael, J. & Ehrlén, J. Elasticities: a review of methods and

531   model limitations. *Ecology* **81**, 607–618 (2000).

532   37.   van Tienderen, P. H. Elasticities and the link between demographic and evolutionary

533   dynamics. *Ecology* **81**, 666–679 (2000).

534   38.   Capdevila, P. *et al.* Life history mediates the trade-offs among different components of

535   demographic resilience. *Ecology Letters* **00**, 1–14 (2022).

536   39.   Pagel, M. Inferring the historical patterns of biological evolution. *Nature* **401**, 877–884

537   (1999).

538   40.   Freckleton, R. P., Harvey, P. H. & Pagel, M. Phylogenetic analysis and comparative

539   data: A test and review of evidence. *American Naturalist* **160**, 712–726 (2002).

540   41.   Conner, J. K. How strong is natural selection? *Trends in Ecology & Evolution* **16**, 215–

541   217 (2001).

542   42.   Lande, R. Quantitative genetic analysis of multivariate evolution, applied to

543   brain:body size allometry. *Evolution (N Y)* **33**, 402–416 (1979).

544   43.   Blowes, S. A. *et al.* The geography of biodiversity change in marine and terrestrial

545   assemblages. *Science (1979)* **366**, 339–345 (2019).

546   44.   Paniw, M., Ozgul, A. & Salguero-Gómez, R. Interactive life-history traits predict

547   sensitivity of plants and animals to temporal autocorrelation. *Ecology Letters* **21**, 275–

548   286 (2018).

549   45.   Mannion, P. D., Upchurch, P., Benson, R. B. J. & Goswami, A. The latitudinal

550   biodiversity gradient through deep time. *Trends in Ecology and Evolution* **29**, 42–50

551   (2014).

552   46.   Lande, R., Engen, S. & Sther, B. E. *Stochastic population dynamics in ecology and*

553   *conservation*. (Oxford University Press, 2003).

554   47.   Fraterrigo, J. M., Pearson, S. M. & Turner, M. G. Joint effects of habitat configuration

555   and temporal stochasticity on population dynamics. *Landscape Ecology* **24**, 863–877

556   (2009).

557   48.   Gregory, T. R. Understanding natural selection: Essential concepts and common

558   misconceptions. *Evolution: Education and Outreach* **2**, 156–175 (2009).

559   49.   Bell, G. Evolutionary rescue and the limits of adaptation. *Philosophical Transactions*

560   *of the Royal Society B: Biological Sciences* **368**, 1–6 (2013).

561   50.   Bell, G. & Gonzalez, A. Adaptation and evolutionary rescue in metapopulations

562   experiencing environmental deterioration. *Science (1979)* **332**, 1327–1330 (2011).

563   51.   Compagnoni, A. *et al.* Herbaceous perennial plants with short generation time have

564   stronger responses to climate anomalies than those with longer generation time. *Nature*

565   *Communications* **12**, 1–8 (2021).

566   52.   Paniw, M., Maag, N., Cozzi, G., Clutton-Brock, T. & Ozgul, A. Life history responses

567   of meerkats to seasonal changes in extreme environments. *Science (1979)* **363**, 631–

568   635 (2019).

569   53.   Cordes, L. S. *et al.* Contrasting effects of climate change on seasonal survival of a

570   hibernating mammal. *Proc Natl Acad Sci U S A* **117**, 18119–18126 (2020).

571   54.   Thornton, P. K., Ericksen, P. J., Herrero, M. & Challinor, A. J. Climate variability and

572   vulnerability to climate change: A review. *Global Change Biology* **20**, 3313–3328

573   (2014).

574   55.   Hughes, T. P. *et al.* Spatial and temporal patterns of mass bleaching of corals in the

575   Anthropocene. *Science (1979)* **359**, 80–83 (2018).

576   56.   Lenton, T. M. *et al.* Climate tipping points — too risky to bet against. *Nature* **575**,

577   592–595 (2019).

578   57.   Ingrisch, J. & Bahn, M. Towards a comparable quantification of resilience. *Trends in*

579   *Ecology and Evolution* **33**, 251–259 (2018).

580   58.   Dixon, A. M., Forster, P. M. & Beger, M. Coral conservation requires ecological

581   climate-change vulnerability assessments. *Frontiers in Ecology and the Environment*

582   **19**, 243–250 (2021).

583   59.   Kwiatkowski, L., Halloran, P. R., Mumby, P. J. & Stephenson, D. B. What spatial

584   scales are believable for climate model projections of sea surface temperature? *Climate*

585   *Dynamics* **43**, 1483–1496 (2014).

586   60.   Foden, W. B. *et al.* Climate change vulnerability assessment of species. *Wiley*

587   *Interdisciplinary Reviews: Climate Change* **10**, 1–36 (2019).

588   61.   Willis, K. J., Jeffers, E. S. & Tovar, C. What makes a terrestrial ecosystem resilient?

589   *Science (1979)* **359**, 988–989 (2018).

590   62.   Darling, E. S. & Côté, I. M. Seeking resilience in marine ecosystems. *Science (1979)*

591   **359**, 986–987 (2018).

592   63.   Salguero-Gómez, R. & Plotkin, J. B. Matrix dimensions bias demographic inferences:

593   Implications for comparative plant demography. *American Naturalist* **176**, 710–722

594   (2010).

595   64.   Tenhumberg, B., Tyre, A. J. & Rebarber, R. Model complexity affects transient

596   population dynamics following a dispersal event: a case study with pea aphids.

597   *Ecology* **90**, 1878–1890 (2009).

598   65.   Stott, I., Hodgson, D. J. & Townley, S. Popdemo: An R package for population

599   demography using projection matrix analysis. *Methods in Ecology and Evolution* **3**,

600   797–802 (2012).

601   66.   Stott, I., Townley, S., Carslake, D. & Hodgson, D. J. On reducibility and ergodicity of

602   population projection matrix models. *Methods in Ecology and Evolution* **1**, 242–252

603   (2010).

67.   Mercado-Molina, A. E., Sabat, A. M. & Yoshioka, P. M. Demography of the

demosponge Amphimedon compressa: Evaluation of the importance of sexual versus

asexual recruitment to its population dynamics. *Journal of Experimental Marine*

Biology and Ecology **407**, 355–362 (2011).

68.   Jelbert, K. *et al.* Demographic amplification is a predictor of invasiveness among

plants. *Nature Communications* **10**, 1–6 (2019).

69.   Stubben, C. & Milligan, B. Estimating and analyzing demographic models using the

popbio package in R. *Journal of Statistical Software* **22**, 1–23 (2007).

70.   Franco, M. & Silvertown, J. A comparative demography of plants based upon

elasticities of vital rates. *Ecology* **85**, 531–538 (2004).

71.   Morris, W. F. & Doak, D. F. *Quantitative conservation biology: Theory and practice*

of population viability analysis. (Sinauer Associates, Inc., 2002).

72.   Kuhn, M. caret: Classification and regression training. (2020).

73.   Freckleton, R. P. The seven deadly sins of comparative analysis. *Journal of*

Evolutionary Biology **22**, 1367–1375 (2009).

74.   Revell, L. J. Phylogenetic signal and linear regression on species data. *Methods in*

Ecology and Evolution **1**, 319–329 (2010).

75.   Hinchliff, C. E. *et al.* Synthesis of phylogeny and taxonomy into a comprehensive tree

of life. *Proceedings of the National Academy of Sciences* **112**, 12764–12769 (2015).

76.   Chamberlain, S. *et al.* taxize: Taxonomic information from around the web. (2020).

77.   Michonneau, F., Brown, J. & Winter, D. rotl: An R package to interact with the Open

Tree of Life data. *Methods in Ecology and Evolution* **7**, 1–17 (2016).

78.   Paradis, E. & Schliep, K. ape 5.0: an environment for modern phylogenetics and

evolutionary analyses in R. *Bioinformatics* **35**, (2018).

79.   Revell, L. J. phytools: An R package for phylogenetic comparative biology (and other

629   things). *Methods in Ecology and Evolution* **3**, 217–223 (2012).

630   80.   Freckleton, R. P. & Jetz, W. Space versus phylogeny: Disentangling phylogenetic and

631   spatial signals in comparative data. *Proceedings of the Royal Society B: Biological*

632   *Sciences* **276**, 21–30 (2009).

633   81.   Grafen, A. The phylogenetic regression. *Philosophical Transactions of the Royal*

634   *society of London. Series B. Biological Sciences* **326**, 119–157. (1989).

635   82.   Revell, L. J., Harmon, L. J. & Collar, D. C. Phylogenetic signal, evolutionary process,

636   and rate. *Syst Biol* **57**, 591–601 (2008).

637   83.   Howard, C., Flather, C. H. & Stephens, P. A. A global assessment of the drivers of

638   threatened terrestrial species richness. *Nature Communications* **11**, 1–10 (2020).

639   84.   Haapkylä, J. *et al.* Seasonal rainfall and runoff promote coral disease on an inshore

640   reef. *PLoS ONE* **6**, (2011).

641   85.   Karger, D. N. *et al.* Climatologies at high resolution for the earth's land surface areas.

642   *Scientific Data* **4**, 1–20 (2017).

643   86.   Evers, S. *et al.* Lagged and dormant-season climate better predict plant vital rates than

644   climate during the growing season. *Global Change Biology* **27**, 1927–1941 (2021).

645   87.   Pilowsky, J. colorednoise: Simulate temporally autocorrelated populations. (2019).

646   88.   Ruokolainen, L., Lindén, A., Kaitala, V. & Fowler, M. S. Ecological and evolutionary

647   dynamics under coloured environmental variation. *Trends in Ecology and Evolution*

648   **24**, 555–563 (2009).

649   89.   Vasseur, D. A. & Yodzis, P. The color of environmental noise. *Ecology* **85**, 1146–1152

650   (2004).

651   90.   R Core Team. R: A language and environment for statistical computing. (2019).

652   91.   Carrascal, L. M., Galván, I. & Gordo, O. Partial least squares regression as an

653   alternative to current regression methods used in ecology. *Oikos* **118**, 681–690 (2009).

654   92.   Mevik, B., Wehrens, R. & Hovde Liland, K. pls: Partial least squares and principal

655   components regression. (2019).

656   93.   Adams, D. C. & Felice, R. N. Assessing trait covariation and morphological

657   integration on phylogenies using evolutionary covariance matrices. *PLoS ONE* **9**,

658   (2014).

659   94.   Revell, L. J. Size-correction and principal components for interspecific comparative

660   studies. *Evolution (N Y)* **63**, 3258–3268 (2009).

661   95.   Robert, A., Sarrazin, F., Couvet, D. & Legendre, S. Releasing adults versus young in

662   reintroductions: Interactions between demography and genetics. *Conservation Biology*

663   **18**, 1078–1087 (2004).

664   96.   Metcalf, C. J. E., McMahon, S. M., Salguero-Gómez, R. & Jongejans, E. IPMpack: An

665   R package for integral projection models. *Methods in Ecology and Evolution* **4**, 195–

666   200 (2013).

667   97.   Conde, D. A. *et al.* Data gaps and opportunities for comparative and conservation

668   biology. *PNAS* **116**, 9658–9664 (2019).

Table 1. The resilience attributes of resistance (green), compensation (blue), and recovery (orange) of natural populations do not correlate with their relative exposure to environmental stochasticity. Using phylogenetically-corrected Pearson's tests of correlation, we explored the association between transient metrics of demographic resistance (first-timestep attenuation, $\underline{\rho}$ & and maximal attenuation, $\underline{\rho}_{max}$), compensation (reactivity, $\overline{\rho}$ & maximal amplification, $\overline{\rho}_{max}$), and recovery (damping ratio, ρ & period of oscillation, ψ; Fig. 1A) and five metrics of environmental stochasticity: temperature frequency spectrum (β_T), temperature autocorrelation (a_T), thermal range (m), precipitation frequency spectrum (β_P), and precipitation autocorrelation (a_P). Correlation displayed using Pearson's correlation coefficient (r).

Transient metric	Resilience attribute	β_T	a_T	m	β_P	a_P
$\underline{\rho}$	Resistance	-0.0106	0.0060	-0.0094	-0.0024	0.0042
$\underline{\rho}_{max}$		-0.0071	0.0031	-0.0121	-0.0093	0.0068
$\overline{\rho}$	Compensation	-0.0026	0.0007	0.0070	-0.0080	0.0041
$\overline{\rho}_{max}$		-0.0016	<0.0001	0.0069	-0.0026	0.0009
ρ	Recovery	0.0012	-0.0007	0.0026	-0.0124	0.0147
ψ		-0.0071	0.0133	0.0052	0.0037	-0.0070

Table 2. Variation across resistance (green) and compensation (blue) of natural populations corresponds with the energetic investments of their individuals, but characteristics of recovery (orange) do not. Using phylogenetically-corrected Pearson's tests of correlation, we explored the association between the transient metrics of resistance (first-timestep attenuation, $\underline{\rho}$ & and maximal attenuation, $\underline{\rho}_{max}$), compensation (reactivity, $\overline{\rho}$ & maximal amplification, $\overline{\rho}_{max}$), and recovery (damping ratio, ρ & period of oscillation, ψ; Fig. 1A) and their sensitivities to the vital rates of survival (σ), progression (γ), retrogression (τ), and reproduction (ϕ). Correlation displayed using Pearson's correlation coefficient (r), with the colour gradation highlighting the strength of the absolute correlation between each transient metric and its vital rate sensitivities, with darker shades representing stronger associations.

Transient metric	Resilience attribute	Survival σ	Progression γ	Retrogression τ	Reproduction ϕ
$\underline{\rho}$	Resistance	-0.70	-0.07	0.28	-0.39
$\underline{\rho}_{max}$	Resistance	-0.17	0.07	0.31	-0.34
$\overline{\rho}$	Compensation	-0.11	-0.49	0.39	-0.04
$\overline{\rho}_{max}$	Compensation	-0.02	-0.49	-0.13	-0.14
ρ	Recovery	-0.06	-0.06	0.11	-0.01
ψ	Recovery	<0.01	0.06	-0.08	0.08

Table 3. Variation in resistance (green), compensation (blue), and recovery (orange) is
strongly predicted by phylogenetic association. However, whilst a strong phylogenetic
signal is also evident across the vital rate sensitivities of measures of resistance and
compensation, there is a negligible phylogenetic signal across the vital rate sensitivities of
measures of recovery. To quantify the strength of statistical non-independence in the
resilience attributes of natural populations due to species' common ancestry, we estimated the
phylogenetic signal (Pagel's λ[39]) across our transient metrics of demographic resistance (first-
timestep attenuation, $\underline{\rho}$ & and maximal attenuation, $\underline{\rho}_{max}$), compensation (reactivity, $\overline{\rho}$ &
maximal amplification, $\overline{\rho}_{max}$), and recovery (damping ratio, ρ & period of oscillation, ψ; Fig.
1A), as well as their sensitivities to the vital rates of survival (σ), progression (γ), retrogression
(τ), and reproduction (ϕ). Pagel's λ[39] ranges between 0, indicating that traits have evolved
independently of phylogeny, and 1, representing a high phylogenetic signal. Colour gradation
highlights the likelihood that the phylogenetic signal observed across each transient metric and
its vital rate sensitivities differs significantly from 0 ($p < 0.05$), with darker shades representing
stronger signals.

Transient metric	Resilience attribute	Pagel's λ	*Survival* σ	*Progression* γ	*Retrogression* τ	*Reproduction* ϕ
$\underline{\rho}$	Resistance	0.997 $p<0.001$	0.997 $p<0.001$	0.989 $p<0.001$	0.967 $p<0.001$	0.985 $p<0.001$
$\underline{\rho}_{max}$		0.997 $p<0.001$	0.993 $p<0.001$	0.985 $p<0.001$	0.952 $p<0.001$	0.989 $p<0.001$
$\overline{\rho}$	Compensation	0.998 $p<0.001$	0.990 $p<0.001$	0.999 $p<0.001$	0.995 $p<0.001$	0.996 $p<0.001$
$\overline{\rho}_{max}$		0.998 $p<0.001$	0.988 $p<0.001$	0.998 $p<0.001$	0.946 $p<0.001$	0.996 $p<0.001$
ρ	Recovery	0.996 $p<0.001$	0.003 $p=0.344$	<0.001 $p=0.686$	0.005 $p=0.094$	<0.001 $p>0.999$
ψ		0.992 $p<0.001$	<0.001 $p>0.999$	0.179 $p<0.001$	<0.001 $p>0.999$	<0.001 $p>0.999$

 Figure captions

Figure 1. Demographic resistance, compensation, and time of recovery across 2,242

natural populations show how population resilience corresponds with exposure to

environmental stochasticity. (A) Within stochastic environments, the dynamics of

populations can vary considerably from long-term (*i.e*., asymptotic) expectations. Under stable

stationary conditions, populations display asymptotic growth trajectories whereby the size of a

population changes at a constant rate (λ). However, following a disturbance (*), populations

can enter a transient state during which their growth rate can differ significantly from

asymptotic expectations. The duration and form of this transient phase depends, in addition to

key moments of disturbances (*e.g.*, magnitude, frequency, *etc.*), on a population's resilience

attributes of resistance, compensation, and time to recovery. Here, *resistance* is the ability for

a population to avoid a decline in size following a disturbance, whereas *compensation* is the

extent to which a population may increase in size following a disturbance. Meanwhile, time of

recovery (henceforth *recovery*) is the duration needed for a population to converge back to its

stationary equilibrium following a disturbance. Transient demographic theory offers the

opportunity to quantify the short-term dynamics of natural populations, thus unlocking the

potential for macroecological studies exploring patterns and plausible mechanism of

demographic resilience worldwide. Transient increases in population size (N) can be evaluated

using metrics of population reactivity ($\overline{\rho}$; increase in N within a one-time step following a

disturbance) and maximal amplification ($\overline{\rho}_{max}$; maximum increase in N during the transient

period). Equally, the magnitude of transient declines in population size can be assessed using

the metrics of first-step attenuation ($\underline{\rho}$; decrease in N within a one-time step following a

disturbance) and maximal attenuation ($\underline{\rho}_{max}$; maximum decrease in N during transient period).

Finally, the damping ratio (ρ; rate of convergence back to stationary stability), and period of

oscillation (ψ; time between corresponding phases of the largest oscillatory cycle in N) of

populations offer insights into their capacity to go back to a stationary stable equilibrium. **(B)** We quantified the aforementioned transient metrics of demographic resilience for 2,242 populations from 369 species for whom Matrix Population Models (MPMs) are available from the global COMPADRE[25] (Blue; plants & algae) and COMADRE[24] databases (Orange; animals). Point size illustrates the relative length of time over which each population was surveyed to parameterise the MPMs deposited within COMPADRE and COMADRE.

Figure 2. Variation across metrics of demographic resilience: resistance (green), compensation (blue), and recovery (orange) does not correspond with patterns in the exposure of populations to environmental stochasticity. Scores and loadings of a phylogenetically-weighted Partial Least Squares (pPLS) regression analysis exploring the correlation between patterns in the variation of the six transient metrics of **(A)** first-step attenuation ($\underline{\rho}$), **(B)** maximal attenuation ($\underline{\rho}_{max}$), **(C)** reactivity ($\overline{\rho}$), **(D)** maximal amplification ($\overline{\rho}_{max}$), **(E)** damping ratio (ρ), and **(F)** period of oscillation (ψ), and the five metrics of environmental stochasticity: temperature frequency spectrum (β_T), temperature autocorrelation (a_T), thermal range/magnitude (m), precipitation frequency spectrum (β_P), and precipitation autocorrelation (a_P). The component scores along each axis display the percentage variance in the environmental stochasticity variables captured by each component, with the first two components alone explaining >50% of the variance across all models. A gradation in point colour is then used to reflect patterns in the relative magnitude of each transient metric recorded from each population, with darker shades indicating higher estimates. Insert barplots are the standardised regression coefficients (b) highlighting the relative weighting of each abiotic variable in the overall capacity of each pPLS model to explain variation in a given transient metric (r^2).

Figure 3. The resilience attributes of resistance (green), compensation (blue), and recovery (orange), in natural populations are determined by the relative energetic investments of their individuals. Scores and loadings of a phylogenetically weighted Partial Least Squares regression analysis exploring the sensitivity patterns of the six transient metrics of **(A)** first-step attenuation ($\underline{\rho}$), **(B)** maximal attenuation ($\underline{\rho}_{max}$), **(C)** reactivity ($\overline{\rho}$), **(D)** maximal amplification ($\overline{\rho}_{max}$), **(E)** damping ratio (ρ), and **(F)** period of oscillation (ψ), towards the vital rates of survival (σ), progression (γ), retrogression (τ), and reproduction (ϕ). The component scores along each axis display the percentage variance in the environmental stochasticity variables captured by each component, with the first two components alone explaining >47% of the variance across all models. A gradation in point colour is then used to reflect patterns in the relative magnitude of each transient metric recorded from each population, with darker shades indicating higher estimates. Insert barplots are the standardised regression coefficients (*b*) highlighting the relative weighting of each vital rate in the overall capacity of each pPLS model to explain variation in a given transient metric (r^2).

777 **Figure 2.**

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 Figure 3.