Touch receptor end-organ innervation and function requires sensory neuron expression of the transcription factor Meis2.

Simon Desiderio, [2, §] Fred Schwaller, [3, §] Kevin Tartour, [1] Kiran Padmanabhan, [1] Gary R. Lewin, [3] Patrick Carroll, [2, #] and Frédéric Marmigère, [1, #, *]

[1] IGFL, UMR 5242 CNRS/ENS Lyon
32/34 Avenue Tony Garnier
69007 Lyon, France.

[2] Institute for Neurosciences of Montpellier, University of Montpellier, INSERM U 1298, Montpellier, France.

[3] Max-Delbrück Centre for Molecular Medicine, Department of Neuroscience, Robert-Rössle Str. 10, 13125, Berlin-Buch, Germany.

[§, #] These authors contributed equally

[*] Corresponding author

Short title: Meis2 in DRG sensory neurons

Abstract:

Touch sensation is primarily encoded by mechanoreceptors, called Low-Threshold Mechanoreceptors (LTMRs), with their cell bodies in the Dorsal Root Ganglia. Because of their great diversity in terms of molecular signature, terminal endings morphology and electrophysiological properties, mirroring the complexity of tactile experience, LTMRs are a model of choice to study the molecular cues differentially controlling neuronal diversification. While the transcriptional codes that define different LTMR subtypes have been extensively studied, the molecular players that participate in their late maturation and in particular in the striking diversity of their end-organ morphological specialization are largely unknown. Here we identified the TALE homeodomain transcription factor Meis2 as a key regulator of LTMRs target-field innervation. *Meis2* is specifically expressed in cutaneous LTMRs and its expression depends on target-derived signals. While LTMRs lacking *Meis2* survived and are normally specified, their end-organ innervations, electrophysiological properties and transcriptome are differentially and markedly affected, resulting in impaired sensory-evoked behavioral responses. These data establish *Meis2* as a major transcriptional regulator controlling the orderly formation of sensory neurons innervating peripheral end-organs required for light touch.

Key words:

Dorsal root ganglion, Meis2, sensory neurons, ASD, transcription factor, TALE homeodomain.

Introduction

Tactile stimuli like brush, light pressure, or roughness engage highly specialized and diverse arrays of mechanoreceptors in both the hairy and glabrous skin (*1–7*). Somatosensory perception via these mechanoreceptors involves primary sensory neurons whose cell bodies reside within Dorsal Root Ganglia (DRG) and cranial sensory ganglia. These sensory neurons within the DRG can be broadly classified as nociceptors, mechanoreceptors or proprioceptors and each group is characterized by the expression of specific combination of genes, have distinctive physiological properties and projections within the spinal cord and periphery (*8–10*).

Cutaneous mechanoreceptors or Low Threshold Mechanoreceptors (LMTRs) exhibit a variety of specialized terminal endings in the hairy and glabrous skin with strikingly unique morphologies (*1–4*, *6*, *7*, *11*, *12*). LTMRs projecting to the glabrous skin innervate Merkel cell complexes or Meissner corpuscles at the dermal-epidermal border. Those innervating Merkel cells in the glabrous or hairy skin have large thickly myelinated axons (Aβ-fibers) and are characterized as slowly-adapting mechanoreceptors responding to skin movement and static displacement (also referred to as Aβ-SAIs). On the other hand, Meissner corpuscles are mechanoreceptors which are only sensitive to skin movement or vibration (rapidly-adapting mechanoreceptors) and are referred to as Aβ-RAs. LTMRs innervating hair follicles in the hairy skin can form lanceolate endings or circumferential endings. Virtually all mechanoreceptors innervating hairs show rapidly-adapting properties and respond only to hair movement but not to static displacement (*4*). LTMRs with large myelinated axons innervating hairy skin are characterized as Aβ-RAs, and a specialized population of slowly conducting myelinated fibers called D-hair mechanoreceptors (or Aδ-RAs) also form lanceolate endings on small hairs. D-hair mechanoreceptors are most sensitive to low velocity stroking, have large receptive fields and are directionally tuned (*7*, *13*). A small number of LTMRs in the hairy skin are not activated by hair movement but show properties of rapidly-adapting mechanoreceptors (*14*). These were originally characterized as so-called field receptors (*14*, *15*) and were recently shown to form circumferential endings around hair follicles (*16*). LTMRs tuned to high frequency vibration are called Aβ-RAII and innervate Pacinian corpuscles deep in the skin or on the bone (*12*). Ruffini endings that are thought to be innervated by stretch sensitive mechanoreceptors (Aβ-SAII) remain poorly characterized in mice (*3*). Recent advances in combining single cell transcriptomic, and deep RNA sequencing with genetic tracing have tremendously extended the classical subtypes repertoire and clustered at least 20 different subtypes of LTM neurons (*5*, *11*, *16–18*).

Cracking the transcriptional codes supporting sensory neurons identity and diversification has been the object of tremendous efforts in the last decades (*5*, *8*, *9*, *11*, *17*, *18*). For instance, the functions of specification factors or terminal selectors, like Maf, Shox2, Runx3, Pea3 and ER81 have been functionally implicated in LTMR segregation (*8*, *9*, *19–22*). Whereas the specific function of adhesion molecules in shaping the assembly of touch circuitry is being unraveled (*23*), the transcriptional control of target cell innervation within the skin and of the establishment of specialized peripheral end-organ complexes is less understood. Recent advances in the understanding of Autism Spectrum Disorder (ASD) suggest that centrally affected neurons in ASD and LTMRs share transcriptional programs regulating late neuronal differentiation (*24*, *25*). Genes known to be involved in ASD are expressed by LTMRs, mouse models for ASD exhibit sensory deficits, and DRG specific ablation of some of these genes causes ASD-like behaviors (*24*, *25*). Meis2 is another TF in which mutations in humans are associated with severe ASD (*26*), and is also specifically expressed in LTMRs (*11*, *17*, *18*). It belongs to a highly conserved homeodomain family containing three members in mammals, Meis1, Meis2 and Meis3 (*27*, *28*), and *Meis1* is necessary for target-field innervation of sympathetic peripheral neurons (*29*). Since ASD is a highly genetically and phenotypically heterogeneous neurodevelopmental disorder encompassing deficiencies in neurogenesis, neurite outgrowth, synaptogenesis and synaptic plasticity (*30*, *31*), we wondered if Meis2 could also be a pertinent regulator of late sensory neurons differentiation.

Here, we show that Meis2 regulates the innervation of specialized cutaneous end-organs important for LTMRs function. We confirmed that *Meis2* expression is restricted to LTMR subclasses at late developmental stages compatible with functions in specification and/or target-field innervation. We generated mice carrying an inactive *Meis2* gene in post-mitotic sensory neurons. While these animals are healthy and viable and do not exhibit any neuronal loss, they display severe significant tactile sensory defects in electro-physiological and behavioral assays. Consistent with these findings we identified major morphological alterations in LTMR end-organ structures in *Meis2* null sensory neurons. Finally, transcriptomic analysis at late embryonic stages showed dysregulation of synapse and neuronal projection-related genes that underpin these functional and behavioral phenotypes.

Results:

***Meis2* is expressed by cutaneous LTMRs.**

We analyzed *Meis2* expression using *in situ* hybridization (*ISH*) at various developmental stages in both mouse and chick lumbar DRG, combined with well-established molecular markers of sensory neuron subclasses (Figures 1A; Figures 1 Supplementary 1 and 2). In mouse, *Meis2* mRNA was first detected at embryonic day (E) 11.5 in a restricted group of large DRG neurons. This restricted expression pattern was maintained at E14.5, E18.5 and adult stages (Figure 1A). In chick, *Meis2* was expressed in most DRG neurons at Hamburger-Hamilton stage (HH) 24, but later becomes restricted to a well-defined subpopulation in the ventro-lateral part of the DRG where LTMRs and proprioceptors are located(*32*) (Figure 1 Supplementary 2A). In both species, *Meis2*-positive cells also expressed the pan-neuronal marker Islet1, indicating that they are post-mitotic neurons. In chick, we estimated that *Meis2*-positive cells represented about 15% of of Islet1-positive DRG neurons at HH29 and HH36 respectively, suggesting a stable expression in given neuronal populations during embryonic development. Double *ISH* for *Meis2* and *Ntrk2*, *Ntrk3* or *c-Ret* mRNAs in E14.5 and E18.5 mouse embryonic DRG (Figure 1 Supplementary 1A and B) showed a large co-expression in *Ntrk2*- and *Ntrk3*-positive neurons confirming that *Meis2*-positive neurons belong to the LTMR and proprioceptive subpopulations. Finally, double *ISH* for *Meis2* and *c-Ret* in E14.5 mouse DRG showed that virtually all large *c-Ret*-positive neurons representing part of the LTMR pool co-expressed *Meis2* at this stage before the emergence of the small nociceptive *Ret*-positive population. Similar results were found in chick at HH29 (Figure 1 Supplementary 2B).

In mouse, comparison of *Meis2* mRNA expression to *Ntrk1*, a well-established marker for early nociceptive and thermo-sensitive neurons, showed that only few *Meis2*-positive neurons co-expressed *Ntrk1* at E11.5 and E18.5 (Figure 1 Supplementary 1C). In chick HH29 embryos, *Meis2* expression was fully excluded from the *Ntrk1* subpopulation (Figure 1 Supplementary 2C). In adult mouse DRG, comparison of *Meis2* mRNA to Ntrk1, Calca and TrpV1 immunostaining confirmed that *Meis2*-expressing neurons are largely excluded from the nociceptive and thermo-sensitive populations of DRG neurons. Instead, a large proportion of *Meis2*-positive neurons co-expressed *Nefh*, a marker for large myelinated neurons including LTMR and proprioceptors, and *Pvalb*, a specific marker for proprioceptors (Figure 1 Supplementary 1D). Finally, *Meis2* expression in LTMRs projecting to the skin was confirmed by retrograde-tracing experiments using Cholera toxin B subunit (CTB) coupled with a fluorochrome injected into hind paw pads of P5 newborn mice. Analyses of CTB expression in lumbar DRG three days later at P8 showed that many retrogradely labelled sensory neurons were also immuno-positive for Meis2, Maf, Ntrk2 and Ntrk3 (Figure 1B).

Altogether, our results on Meis2 co-localization with Nefh, Ntrk2, Ntrk3, Ret, Pvalb and Maf at different embryonic and postnatal stages are consistent with previous report on restricted Meis2 expression to the Aβ-field, Aβ-SA1 and Aβ-RA subclasses of LTMR neurons and proprioceptive neurons (*11*, *17*, *18*, *33*). The relatively lower co-incidence of Meis2 and Ntrk2 expressions compared to Ntrk3 is consistent with Meis2 being excluded from the Aδ-LTMRs (D-hair mechanoreceptors). The lack of co-expression with Ntrk1 and TrpV1 also confirmed Meis2 exclusion from peptidergic and non-peptidergic subpopulations.

Target-derived signals are necessary to maintain *Meis2* expression.

The requirement for extrinsic signals provided by limb mesenchyme and muscles for proprioceptor and LTMR development has been documented (*11*, *21*, *33–37*). To test the influence of target-derived signals on *Meis2* expression in sensory neurons, limb buds were unilaterally ablated in HH18 chick embryos. Embryos were harvested at HH27 and HH36, before and after ventro-lateral neurons are lost respectively (*38–40*) (Figure 1C and D; Figure 1 Supplementary 2D). In HH36 embryos, about 65% of Meis2-positive neurons were lost on the ablated side compared to the contralateral side (Figure 1C). This is consistent with a 30% loss of all sensory DRG neurons represented by the pan-neuronal marker Islet1, and the 50% and 65% loss of Ntrk2 and Ntrk3 positive VL-neurons respectively (Figure 1 Supplementary 2D). The number of Ntrk2-positive DL neurons were not significantly affected. In HH27 embryos, while no significant loss of Islet1-positive neurons was detected following limb ablation, about 40% of Meis2-positive neurons were lost, and remaining *Meis2*-positive neurons expressed very low levels of *Meis2* mRNAs (Figure 1D).

These results indicate that target-derived signals are necessary for the maintenance but not the induction of Meis2 expression in sensory neurons.

***Meis2* gene inactivation in post-mitotic sensory neurons induces severe behavioral defects.**

We next asked whether *Meis2* inactivation would induce changes in LTMR structure and function. We generated a conditional mouse mutant strain for *Meis2* (*Meis2*$^{LoxP/LoxP}$) in which the first coding exon for the homeodomain was flanked by *LoxP* sites (Figure 1 Supplementary 3A). To validate the use of our strain, we first cross the *Meis2*$^{LoxP/LoxP}$ mice with the *Wnt1*^{Cre} strain. This crossing efficiently inactivated *Meis2* in the neural crest, and *Wnt1*^{Cre}::*Meis2*$^{LoxP/LoxP}$ new-born pups exhibited a cleft palate as previously reported in another conditional *Meis2* mouse strain (*41*) (Figure 1 Supplementary 3B). They were however not viable, precluding functional and anatomical analyses at adult stages. To bypass this neural crest phenotype and to more specifically address *Meis2* function in post-mitotic neurons, we crossed our *Meis2* strain with the Isl1$^{Cre/+}$ and focused our analysis on the

Isl1$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ strain. Mutant pups were viable, appeared healthy and displayed a normal palate, allowing sensory behavior investigations.

We monitored tactile evoked behaviors in adult WT, *Isl1*$^{Cre/+}$ and *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ mice, using stimuli applied to both glabrous and hairy skin. We used von Frey filaments to apply a series of low forces ranging from 0.008 to 1.4g to the hind paw and found the frequency of withdrawal responses to be significantly decreased in *Isl1*$^{+/Cre}$*::Meis2*$^{LoxP/LoxP}$ mice compared to control WT and *Isl1*$^{+/Cre}$ mice between 0.16 to 0.6g (Figure 1E), indicating that mutant mice are less responsive to light touch. No differences were observed between *WT* and *Isl1*$^{+/Cre}$ mice for any of the stimuli used. Behaviors evoked from stimulation of the glabrous skin were next assessed using the dynamic touch assay "cotton swab" test (*42*). Here, responses were significantly decreased in Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice compared to control WT and Isl1$^{+/Cre}$ littermates (Figure 1F). In the von Frey test, the thresholds for paw withdrawal were similar between all genotypes when using filaments exerting forces ranging from 1 to 1.4g, which likely reflects the activation of mechanical nociception suggesting that *Meis2* gene inactivation did not affect nociceptor function. We also used the hot plate assay to assess noxious heat evoked behaviors and found no difference in jump latencies between WT, *Isl1*$^{Cre/+}$ and *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ mice (Figure 1G). Finally, we compared the sensitivity of mice to stimuli applied to the hairy skin using the sticky tape test. Placing sticky tape on the back skin evoked attempts to remove the stimulus in a defined time window and we found that such bouts of behavior were significantly reduced in *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ mice compared to WT and *Isl1*$^{Cre/+}$ control mice (Figure 1H). Finally, although *Meis2* and *Isl1* are both expressed by spinal motor neurons and proprioceptors (*43–45*), we did not observe obvious motor deficits in *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ mice. Thus, in a catwalk analysis we found no differences in any of the gait parameters measured between WT and mutant mice (Figure 1 Supplementary Table 1).

Overall, these behavior analyses indicate that *Meis2* gene inactivation specifically affects light touch sensation both in the glabrous and the hairy skin while sparing other somatosensory behaviors. The impaired behavioral response to light touch in *Meis2* mutant suggested that *Meis2* gene activity is necessary for the anatomical and functional maturation of LTMRs.

Meis2 is dispensable for LTMR specification and survival.

To investigate whether *Meis2* gene inactivation interfered with LTMR survival during embryonic development, we performed histological analysis of the *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ and *Wnt1*^{Cre}*::Meis2*$^{LoxP/LoxP}$ strains (Figure 2). There was no difference in the size of the DRGs between E16.5 WT and *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ embryos as well as in the number of Ntrk2 and Ntrk3-positive neurons (Figure 2A) suggesting no cell loss. In E18.5 embryonic DRGs, the number of LTMR and

proprioceptors identified as positive for Ntrk2, Ntrk3, Ret and Maf were unchanged following *Meis2* inactivation (Figure 2C). Consistent with the lack of *Meis2* expression in nociceptors, the number of Ntrk1-positive neurons was also unaffected (Figure 2C). Finally, quantification of DRG neuron populations at P0 in *Wnt1*^{Cre}*::Meis2*$^{LoxP/LoxP}$ mice showed similar results with no differences in the number of Ntrk2 and Ntrk3-positive neurons (Figure 2B). At this stage, phosphoCreb (pCreb) expression in Ntrk2 and Ntrk3-positive neurons was similar in WT and mutants (Figure 2 Supplementary 2), suggesting that Ntrk signaling is not affected. Altogether, these results show that *Meis2* is dispensable for LTMR and proprioceptor survival and specification during embryogenesis.

***Meis2* is necessary for normal end-organ innervation.**

To better understand the molecular changes underlying tactile defects in Meis2 mutant mice, we performed RNAseq analysis on DRGs dissected form WT, Isl1$^{Cre/+}$ and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ E18.5 embryos.

For all analyses, consistent with the changes measured for *Meis2* and *Isl1* genes (Figure 3 Supplementary 1), only DEGs with of minimal fold change of 20% and a *p* value lower than *0.05* were considered. Analyses of the dataset (n=3; $p<0.05$; Figure 3; Figure 3 Supplementary 1; Figure 3 Supplementary Table 1) identified 51 differentially expressed genes (DEGs) in Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ DRGs that were significantly different from both WT and Isl1$^{Cre/+}$ control DRGs, and 488 DEGs in Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ DRGs that were significantly different from either WT or Isl1$^{Cre/+}$ DRGs (Figure 3A and B). Gene ontology analysis for the 488 DEGs revealed significant relevant hits with many terms associated with neuronal projections and functions (Figure 3C and D; Figure 3 Supplementary Table 2). These include subsets for the GO term associated with synapse, dendrite and axons and more specifically with GABAergic synapses, dendritic shaft or postsynaptic membrane. Interestingly, a significant association with the GO term Cadherin point at the protocadherin family in which several members were down-regulated. Finally, comparing these genes to scRNAseq analysis in adult DRG neurons (*17*) showed that their expression profiles in DRG sensory subtypes largely overlap with Meis2-expressing subtypes (Figure 3 Supplementary 2), in particular for down-regulated genes such as the synaptic or dendritic proteins Syndig1, Calb2, Pde4a, Col13a1, Fstl4, Sorcs3 or Syt17. These molecular analyses strongly support a role of *Meis2* in regulating embryonic target-field innervation. We thus investigated this hypothesis, and in P0 *Wnt1*^{Cre}*::Meis2*$^{LoxP/LoxP}$, Nefh staining in the hind paws showed strong innervation deficits as reflected by a paucity of neurofilament-positive myelinated branches in both the glabrous and hairy skin (Figure 3E).

***Meis2* gene is necessary for SA-LTMR morphology and function only in the glabrous skin.**

LTMRs form specialized sensory endings in a variety of end-organs specialized to shape the mechanoreceptor properties. We used the *Isl1*$^{+/Cre}$*::Meis2*$^{LoxP/LoxP}$ mice to assess the effects of late loss of *Meis2* on LTMR structure and function, and to investigate if post mitotic *Meis2* inactivation impacts terminal morphologies and physiological properties of LTMRs. We made recordings from single mechanoreceptors and probed their responses to defined mechanical stimuli in adult WT, *Isl1*$^{+/Cre}$ and *Isl1*$^{+/Cre}$*::Meis2*$^{LoxP/LoxP}$ mice using *ex vivo* skin nerve preparations as previously described (*12*, *13*, *46*).

We recorded single myelinated afferents in the saphenous nerve which innervates the hairy skin of the foot or from the tibial nerve that innervates the glabrous skin of the foot (*12*, *13*). In control nerves all the single units (n=78) with conduction velocities in the Aβ-fiber range (>10 m/s) could be easily classified as either a rapidly-adapting mechanoreceptor or a slowly-adapting mechanoreceptor (RA-LTMR or SA-LTMRs respectively), using a set of standard quantitative mechanical stimuli. However, in the *Isl1*$^{+/Cre}$*::Meis2*$^{LoxP/LoxP}$ mice about 10 and 18% of Aβ fibers in the hairy and glabrous skin respectively could not be reliably activated by any of the quantitative mechanical stimuli used. Sensory neurons that could not be activated by our standard array of mechanical stimuli, but could still be activated by rapid manual application of force with a glass rod were classified as so called "Tap" units (Figure 4A). Such "tap" units have been found in several mice with deficits in sensory mechano-transduction (*46*, *47*).

In both the glabrous and the hairy skin, Merkel cells are innervated by Slowly-Adapting Mechanoreceptor type I (SAI-LTMR) neurons responding to both static skin indentation and moving stimuli such as vibration. In the glabrous skin, Merkel cells form clusters in the basal layer of the epidermis, and in the hairy skin, similar clusters of Merkel cells called touch-domes are located at the bulge region of guard hairs. Histological analysis indicated that in the forepaw glabrous skin of *Isl1*$^{+/Cre}$*::Meis2*$^{LoxP/LoxP}$ adult mice, the number of Merkel cells contacted by Nefh-positive fibers was strongly decreased compared to WT (Figure 4B and E). However, in contrast to the glabrous skin, Merkel cell innervation by Nefh-positive fibers appeared largely unaffected in the hairy skin of *Isl1*$^{Cre/+}$*::Meis2*$^{LoxP/LoxP}$ mice (Figure 4D and E). Whole mount analysis of CK8-positive Merkel cells in the hairy back skin of E18.5 embryos showed that the overall number of touch domes and of Merkel cells per touch dome were unchanged in mutant animals compared to WT (Figure 4C).

We made recordings from SA-LTMRs from both glabrous and hairy skin, but decided to pool the data as there was an insufficient sample size from either skin area alone. We reasoned that electrophysiological recordings would pick up primarily receptors that had successfully innervated Merkel cells and miss those fibers that had failed to innervate end-organs and would likely not be

activated by mechanical stimuli. In this mixed sample of SA-LTMRs the mean vibration threshold was significantly elevated in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$mice, but it was clear that many fibers in this sample had mechanical thresholds similar to those in the wild type (Figure 4F). The response of the same SA-LTMRs to a 25 Hz sinusoidal stimulus was unchanged in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$ mice compared to controls (Figure 4F). The response of these fibers to ramp stimuli of increasing velocities or to increasing amplitudes of ramp and hold stimuli were also not significantly different in mutant mice compared to controls (Figure 4F). Finally, consistent with the lack of neuronal loss in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$, the number of recorded fibers were identical in WT and *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$. Altogether, these data indicate that *Meis2* is necessary for Merkel cells innervation in the glabrous, but not in the hairy skin. In addition, electrophysiological recording indicate that amongst SA-LTMRs, there was a light loss of sensitivity that could be associated with poor innervation of Merkel cells in the glabrous skin.

***Meis2* is necessary for RA-LTMRs structure and function.**

In the glabrous skin, Meissner corpuscles are located in the dermal papillae and are innervated by Rapidly-Adapting type LTMR (RA-LTMR) that detect small-amplitude skin vibrations <80 Hz.

Histological analysis of the glabrous skin showed that Nefh-positive innervation of the Meissner corpuscles was strongly disorganized (Figure 5A, Figure 5 Supplementary video 1-2) with decrease complexity of the Nefh^{+} fibers within the corpuscle as shown by quantification of the average number of time that fibers crossed the midline of the terminal structure. However recordings from RA-LTMRs innervating these structures in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$ animals showed largely normal physiological properties (Figure 5B). Thus RA-LTMRs recorded from *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$ displayed normal vibration sensitivity in terms of absolute threshold and their ability to follow 25 sinusoids. There was a tendency for RA-LTMRs in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$ mutant mice to fire fewer action potentials to sinusoids and to the ramp phase of a series 2 second duration ramp and hold stimuli, but these differences were not statistically significant (Figure 5B).

In the hairy skin, RA-LTMRs form longitudinal lanceolate endings parallel to the hair shaft of guard and awl/auchene hairs and respond to hair deflection only during hair movement, but not during maintained displacement (*4*). Similar to Meissner corpuscles, they are tuned to frequencies between 10 and 50 Hz (*12*). Whole mount analysis of Nefh-positive fibers in the adult back skin showed an overall decrease in the innervation density of hairs in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$ animals compared to WT (Figure 6A). Our analysis revealed significant decreases in both the number of plexus branch points and in the number of innervated hair follicles (Figure 6B).

Consistent with the hypo-innervation of hair follicles in *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$, we observed robust deficits in the mechanosensitivity of RA-LTMRs in the hairy skin (Figure 6C and Figure 4 Supplementary 1B). Thus, we needed sinusoids of significantly larger amplitudes to evoke the first (threshold) spike in RA-LTMRs. We therefore measured the total number of spikes evoked by a sinusoid stimulus (25 Hz) of gradually increasing amplitude. Again, RA-LTMRs fired considerably less in *Isl1*$^{+/Cre}$::*Meis2*$^{LoxP/LoxP}$ mutant than in control mice. This finding was confirmed using a series of vibration steps of increasing amplitudes again demonstrating decreased firing in response to 25 Hz vibration stimuli (Figure 6C). Thus, the functional deficits in RA-LTMRs correlate well with the defects in LTMR cutaneous projections we observed in *Isl1*$^{+/Cre}$::*Meis2*$^{LoxP/LoxP}$ mutant mice.

Finally, D-Hair mechanoreceptors or Aδ–LTMRs are the most sensitive skin mechanoreceptors with very large receptive fields (*7*, *13*, *48*). They form lanceolate endings, are thinly myelinated and are activated by movement of the smaller zigzag hairs (*4*). Consistent with the lack of Meis2 expression in this population reported by single-cell RNA-seq databases, Aδ fibers D-hair in the hairy skin showed similar vibration responses in WT and *Isl1*$^{+/Cre}$::*Meis2*$^{LoxP/LoxP}$ mice (Figure 6 Supplementary 1).

Discussion:

The function of the Meis family of TFs in post-mitotic neurons has only been marginally addressed (*29*, *49*, *50*). Here, we showed that *Meis2* is selectively expressed by subpopulations of early post-mitotic cutaneous LTMR and proprioceptive neurons during development of both mouse and chick, highlighting the conserved Meis2 expression across vertebrate species in those neurons. Our results on Meis2 expression are in agreement with previous combined single cell RNAseq (scRNAseq) analysis and genetic tracing reporting Meis2 in proprioceptive neurons, Aβ field-LTMR, Aβ-SA1-LTMR, and Aβ-RA-LTMR, but not in in C-LTMR, Aδ-LTMR, peptidergic and non-peptidergic neurons (*11*, *17*, *18*). We unambiguously demonstrate that *Meis2* differentially regulates target-field innervation and function of post-mitotic LTMR neurons. *Meis2* inactivation in post-mitotic sensory neurons modified their embryonic transcriptomic profile and differentially impaired adult LTMR projections and functions without affecting their survival and molecular subtype identity.

The morphological and functional phenotypes we report following specific *Meis2* gene inactivation in post-mitotic sensory neurons are consistent with its expression pattern, and ultimately, both defective morphological and electrophysiological responses result in specifically impaired behavioral responses to light touch mechanical stimuli. In these mutants, the decreased innervation of Merkel cells in the glabrous skin and the decreased sensitivity in SA-LTMR electrophysiological responses to mechanical stimuli is consistent with *Meis2* being expressed by Aβ-SA1-LTMR neurons. Interestingly, *Meis2* gene inactivation compromises Merkel cells innervation and electrophysiological responses in the glabrous skin but not in touch domes of the hairy skin where innervation appeared unchanged. This difference supports previous work suggesting that the primary afferents innervating Merkel cells in the glabrous and the hairy skin maybe different (*51*, *52*). Whereas Merkel cells of the glabrous skin are exclusively contacted by large Ntrk3/Nefh-positive Aβ afferents, neonatal mouse touch domes receive innervation of two types of neuronal populations, a Ret/Ntrk1-positive one that depend on Ntrk1 for survival and innervation, and another Ntrk3/Nefh-positive that do not depend on Ntrk1 signaling during development (*51*). However, the functional significance of these different innervations is unknown. Denervation in rat also pointed at differences between Merkel cells of the glabrous and the hairy skin. Following denervation, Merkel cells of the touch dome almost fully disappear, whereas in the footpad, Merkel cells developed normally (*53*). Thus, to our best knowledge, Meis2 may represent the first TF which inactivation differentially affects Merkel cells innervation in the glabrous skin and hairy touch domes.

Because touch domes innervation and Aδ fibers D-hair vibration responses were unaffected in *Isl1*$^{+/Cre}$*::Meis2*$^{LoxP/LoxP}$ mice, we postulate that the innervation defects we observed in the hairy skin is

supported by defects in lanceolate endings with RA-LTMR electrophysiological properties. However, the increased number of "tap" units both in the hairy and glabrous skin is compatible with wider deficits also including Aβ-field LTMRs peripheral projections. Similarly, although the severely disorganized Meissner corpuscle architecture did not result in significant consequences on RAM fibers electrophysiological responses in the glabrous skin, it is possible that the large increase in the number of "tap units" within Aβ fibers of the glabrous skin represent Meissner corpuscles whose normal electrophysiological responses are abolished. In agreement, challenging sensory responses in the glabrous skin with either von Frey filament application or cotton swab stroking clearly showed a dramatic loss of mechanical sensitivity specifically within the range of gentle touch neurons. Recent work reported that the von Frey filaments test performed within low forces and challenging light touch sensation could distinguish Merkel cells from Meissner corpuscles dysfunctions. Mice depleted of Merkel cells performed normally on this test while mice mutated at the *Ntrk2* locus with Meissner corpuscle innervation deficits were less sensitive in response to filament within the 0.02-0.6 g range (*54*). Thus, our result in the von Frey filament test likely reflects aberrant functioning of the RAM LTMR-Meissner corpuscle complex. Finally, the unaltered D-hair fibers electrophysiological responses, the normal noxious responses in the hot plate setting and the normal responses to high forces Von Frey filaments are consistent with the absence of *Meis2* expression in Aδ-LTMR, peptidergic and non-peptidergic neurons. Surprisingly, although *Meis2* is expressed in proprioceptive neurons (*17*, *18*, *33*), their function appeared not to be affected as seen by normal gait behavior in catwalk analysis. This is in agreement with studies in which HoxC8 inactivation, a classical Meis TF co-factor expressed by proprioceptive neurons from E11.5 to postnatal stages, affected neither proprioceptive neurons early molecular identity nor their survival (*33*).

Understanding the transcriptional programs controlling each step in the generation of a given fully differentiated and specified neuron is an extensive research field in developmental neurobiology. Basic studies in model organisms led to a functional classification of TFs. Proneural TFs such as *Neurogenins* control the expression of generic pan-neuronal genes and are able to reprogram nearly any cell type into immature neurons (*55*, *56*). Terminal selectors are TFs mastering the initiation and maintenance of terminal identity programs through direct regulation of neuron type-specific effector genes critical for neuronal identity and function such as genes involved in neurotransmitters synthesis and transport, ion channels, receptors, synaptic connectivity or neuropeptide content (*57*, *58*). The proneural function of *Ngn1* and *Ngn2* genes in neural crest cells, the precursors of DRG sensory neurons is well demonstrated (*9*, *59*), and several terminal selector genes shaping the different DRG sensory subpopulations have also been clearly identified including for cutaneous LTMRs (*8*, *9*, *20*, *52*, *60–63*). *Maf*, *Runx3*, *Shox2*, *ER81* and *Pea3* are part of this regulatory

transcriptional network regulating cutaneous LTMR neurons diversification through intermingled crossed activation and/or repression of subclasses specific effector genes (*64*).

In humans, at least 17 different mutations in the *Meis2* gene have been associated with neurodevelopmental delay and ASD (*26*, *65*), emphasizing its essential function in neuronal differentiation. *Meis2* function in late differentiation of post-mitotic peripheral sensory neurons adds to the wide actions of this TF in the developing and adult nervous system in number of regions of the mouse nervous system. Its expression both in dividing neural progenitors, in immature neurons and in discrete populations of mature neurons (*49*, *66–73*) argues for diverse functions ranging from regulation of neuroblasts cell-cycle exit, to cell-fate decision, neurogenesis, neuronal specification, neurites outgrowth, synaptogenesis and maintenance of mature neurons. In DRG LTMRs, *Meis2* fulfils some but not all of the criteria defining terminal selector genes. Its inactivation in neural crest cells does not affect sensory neurons generation nor pan-neuronal features, clearly excluding it from a proneural TF function. Although *Meis2* expression is continuously maintained in defined sensory neurons subtypes starting from early post-mitotic neurons throughout life, its expression is not restricted to a unique neuronal identity and its early or late inactivation in either sensory neurons progenitors or post-mitotic neurons does not influence neuronal subtypes identity nor survival as seen by the normal numbers of Ntrk2, Ntrk3 or c-Ret positive neurons. However, our transcriptomic analyses strongly support that in LTMRs, Meis2 regulates other types of terminal effector genes such as genes participating in neurotransmitters machinery specification and/or recognition, establishment and maintenance of physical interactions between LTMRs and their peripheral targets. Our RNAseq and gene ontology analyses reveal alterations in pathways needed for shaping synapses in general and GABAergic and glutamatergic synapses in particular. Several GABA(A) Receptor Subunits (GABRA1, GABRA4, GABRR1), the GABA synthesis enzyme GAD1, and the K-Cl cotransporters SLC12A5 associated to GABAergic neurotransmission, but also the glutamate receptor subunits GRID1 and GRIK3, are down or up-regulated in Meis2 mutants. Thus, the gene list associated with the GOterm "synapse" strongly links LTMR neurotransmitter type of synapses, their dysfunction in mouse model of ASD (*24*, *25*) and the association of *Meis2* with ASD (*26*, *65*, *74*). Interestingly, Meis2 inactivation seems to interfere with the embryonic expression of many members of the protocadherin family, and the protocadherin γ cluster (Pcdhg) in particular has recently been highlighted as essential for building central and peripheral LTMRs innervation and synapses and establish proper peripheral target-field innervation and touch sensation (*23*).

In conclusion, this study reveals a novel function for the Meis2 transcription factor in selectively regulating target field innervation of LTMR neurons. More broadly, it opens new perspectives to molecularly understand how Meis2 is linked to ASD. Together with studies on Meis2 function in the

SVZ where it is necessary to maintain the neurogenic effect of Pax6 in neural progenitors and is later expressed in their mature progenies (*50*), our results raise the possibility that this TF sets up a lineage specific platform on which various specific co-factors in turn participate in additional and/or subsequent steps of the neuronal differentiation program. Whether loss of such central functions underlies its role in autism and ASDs remains to be explored.

Acknowledgements:

Stéphanie Ventéo for help with ChTx experiments, Anne-Laure Bonnefont for cat walk experiments, al staff at animal house and in particular Flora for great help at the animal facility. Yves Dusabyinema, Benjamin Gillet and Sandrine Hughes at the IGFL sequencing platform (PSI) for Illumina sequencing.

Contribution:

Design of experiments, data analysis and writing: GL, PC, FM

Electrophysiology experiments: GL and FS

Histology and behavior: SD and FM

Transcriptomic analysis: KT, KP and FM

Figure legends:

Figure 1: *Meis2* is expressed in subclasses of DRG cutaneous mechanoreceptive neurons in mouse embryos (A-B). A) *ISH* for *Meis2* mRNA showed expression in a subpopulation of DRG sensory neurons at embryonic stages E11.5, E14.5 and E18.5, at P0 and at adult stages. Dashed lines delineate the DRG. Scale bar=50µm. B) IF for Meis2 (red) and c-Maf, Ntrk2 or Ntrk3 (blue) at P7 following injection of cholera toxin B (CTB in green) in the skin of new-born pups. Note that $Meis2^{+}/CTB^{+}$ retro-traced sensory neurons co-expressed c-Maf, Ntrk2 or Ntrk3 (arrows). Scale bar=50µm. We estimated that 30.5 ± 3.5% (mean ± SEM; n = 3) of Meis2-positive neurons co-expressed Ntrk2, and that 39.5 ± 5.4% co-expressed Ntrk3. Conversely, Meis2 was co-expressed in 53.6 ± 9.4% of Ntrk2-positive neurons, and in 78.5 ± 5.0% of Ntrk3-positive neurons. ***Meis2* expression depends on target-derived signals (C-D).** C) Representative images of *Meis2* mRNA expression (blue or pseudo-colored in red) and islet1 (green) in DRGs of Hamburger-Hamilton stage (HH) 36 chick embryos on the ablated and contralateral sides. Box plots showing the number of $Islet1^{+}/Meis2^{+}$ DRG neurons per section at stage HH36 following limb bud ablation. For Islet1-positive neurons, the contralateral side was taken as 100%. For Meis2, values represent the percentage of $Meis2^{+}$ over $Islet1^{+}$ neurons. D) Representative images of *Meis2* mRNA expression (blue or pseudo-colored in red) and islet1 (green) in DRGs of HH29 chick embryos on the ablated and contralateral sides. Box plots showing the quantification of $Islet1^{+}/Meis2^{+}$ neurons number per section at stage HH29 on the contralateral and ablated sides. Arrow heads point at remaining Meis2-positive VL neurons. Dashed lines encircle the DRGs. ***$p\leq0.005$ *** $p\leq0.0005$; *ns*= not significant following Student t-test. n=3 chick embryos. Scale bar=100µm. **Altered touch perception in *Meis2* mutant mice (E-H).** E) Box plots showing the responses following application of Von Frey filaments of different forces. $Isl1^{+/Cre}$::$Meis2^{LoxP/LoxP}$ mice exhibited a significantly reduced sensitivity to the 0.16, 0.4 and 0.6 g Von Frey filaments but not to higher forces filaments compared to WT and $Isl1^{+/CRE}$ littermates. * $p\leq0.05$; ** $p\leq0.005$; *** $p\leq0.001$ following Kruskal-Wallis statistical analysis. F) Box plots showing the dynamic touch responses when the hind paw palms of individual mice were stroked with a tapered cotton-swab. Analysis showed that $Isl1^{+/Cre}$::$Meis2^{LoxP/LoxP}$ mice were less responsive to the stimulus than WT and $Isl1^{+/Cre}$ littermates. *** $p\leq0.0001$ following a one-way Anova statistical analysis. G) Box plots indicating that the latency to the first signs of aversive behavior in the hot plate test is similar in all groups of mice. WT, n=19; $Isl1^{+/Cre}$, n=16; $Isl1^{+/Cre}$::$Meis2^{LoxP/LoxP}$, n=9. H) Box plots showing the number of bouts when a sticky paper tape was applied on the back skin of mice. Analysis indicated a significant decrease in the number of bouts in $Isl1^{+/Cre}$::$Meis2^{LoxP/LoxP}$ mice compared to WT and $Isl1^{+/Cre}$ littermates. * $p\leq0.05$ following a one-way Anova statistical analysis.

Figure 2: *Meis2* is necessary for LTM neurons peripheral projections but dispensable for their survival and specification. A) Box plots showing that the DRGs volumes along the rostro-caudal axis are similar in E16.5 WT and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ embryos. IF for Ntrk2 or Ntrk3 (red) and Islet1 (blue) and box plots analysis indicating that the percentage of Ntrk2^{+} and Ntrk3^{+} neurons are not affected in E16.5 Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$. Dashed lines encircle the DRGs. n=4; *n.s.* = not significant. Scale bar = 20µm. B) Representatives images showing IF for Ntrk2 or Ntrk3 (green) with Pvalb or Maf (red) in P0 WT and Wnt1^{Cre}::Meis2$^{LoxP/LoxP}$ DRGs. Box plots showing that the number of Ntrk2^{+} and Ntrk3^{+} neurons are unchanged in P0 WT and Wnt1^{Cre}::Meis2$^{LoxP/LoxP}$ DRGs. n=3, *n.s.* = not significant. Scale bar = 20µm. C) Representative images showing IF for Ntrk1 (red), Ret, Ntrk2, Ntrk3 and Maf in E18.5 WT and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ DRGs. Box plots showing that the number of Ret^{+}, Ntrk2^{+}, Ntrk3^{+} and Maf^{+} LTM neurons and of Ntrk1^{+} nociceptive neurons are similar in E18.5 WT and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ DRGs. n=3; *n.s.* = not significant. Scale bar = 100µm.

Figure 3: *Meis2* inactivation dysregulates genes linked to neuronal projections and synaptogenesis. **A-** Venn diagram comparing the number of DEGs between each genotype (n=3; $p<0.05$). This comparison identified 51 DEGs genes that were differentially expressed compared to both control genotypes (WT or *Isl1*$^{+/CRE}$ embryos), and a total of 488 genes that were differentially expressed in *Meis2* mutant compared to either WT or *Isl1*$^{+/CRE}$ embryos. **B-** Heat maps for the 488 DEGs compared to both control genotypes showed that about half of the DEGs were either up- (257) or down- (231) regulated in *Meis2* mutant. Names of genes that are shown in F are labeled in red (up-regulated) or blue (down-regulated). Members of the protocadherin family are indicated. **C-** Gene ontology analysis for the 488 DEGs using David and the full RNAseq gene list as background. Graphs showing the result of gene ontology analysis using David. The upper graph shows the -log10(*p* value) associated with selected significant ($p<0.05$) GO terms or KEGG_PATHWAY terms. The lower graph shows the fold enrichment for the same selected GO terms or KEGG_PATHWAY terms. Only terms associated with a minimum of 5 genes were considered. Bars in red indicated terms for which the heat maps are shown in D. **D-** Heat maps showing the DEGs related to previous GO terms (Red bars in C). E- Representative images showing a strong overall deficits in Nfeh^{+} (red) sensory projections in the hairy and the glabrous skin of P0 Wnt1^{Cre}::Meis2$^{LoxP/LoxP}$ neonates forepaw compared to WT littermates. Dashed lines delineate the hair follicle and the epidermis. Scale bar = 50µm.

Figure 4: *Meis2* gene inactivation compromised Merkel cells innervation in the glabrous skin and increased SAM vibration threshold. A) Percentage of tap units among Aβ fibers in the hairy and glabrous skin detected by electrophysiological recording in the nerve-skin preparation. Note that

Tap-units are only present in both the hairy and glabrous skin of adult Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice but not in WT and Isl1$^{Cre/+}$ littermates. B) Confocal images of Nefh^{+} innervation (green) of CK8^{+} Merkel cells (red) in the forepaw glabrous skin of WT and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ adult mice. Dotted white squares indicate the close-up on CK8^{+} Merkel cells. Note the lack of Nefh^{+} fibers innervating Merkel cells in mutant mice. Scale bar = 10µm. C) Representative images of whole mount staining for CK8 in the hairy back skin of adult WT and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ E18.5 embryos showing no difference in the number of touch dome between genotypes. Box plots show the number of touch domes per surface area and the number of Merkel cells per guard hair. No significant differences were found between both genotype in Mann-Whitney test. n=5 (WT) and 4 (Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$). D) Confocal images of Nefh^{+} innervation (green) and CK8^{+} Merkel cells (red) of guard hairs in the hairy back skin of WT and Isl1$^{Cre/+}$::Meis2$^{LoxP/LoxP}$ adult mice. Dotted white squares indicate the close-up on CK8^{+} Merkel cells with apparently normal Nefh^{+} innervation. Scale bar = 10µm. E) Box plot indicating the percentage of Merkel cells in contact with Nfeh^{+} fibers in the forepaw glabrous and in the back hairy skin of adult WT and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice. n=4; * $p \leq 0.05$ in Mann-Whitney test. F) In the hairy and glabrous skins, SAMs in Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice (n = 22 from 6 mice) had significantly increased vibration threshold compared to WT mice (n = 29 from 6 mice), but normal firing activity to a 25-Hz vibration. Trace shows the stimulation and red square indicate the time frame during when the number of spikes was calculated. G) SAM response to a ramp of 50 Hz vibration with increasing amplitude are similar in WT, Isl1$^{+/Cre}$ and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice. SAM responses to ramp stimuli and their static force responses were also identical in the different genotypes. Fibers from WT and Isl1$^{+/Cre}$ mice (n=5) displayed similar responses.* $p \leq 0.05$; ** $p \leq 0.005$.

Figure 5: *Meis2* gene inactivation affects Meissner corpuscles morphology but not RAM fibers electrophysiological responses in the glabrous skin. A) Representative images showing S100β^{+} Meissner corpuscles (red) and their innervation by Nefh^{+} fibers (green) in the glabrous skin of WT and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ adult mice. Scale bar=10µm. The box plot shows the average number of time that Nefh+ fibers crossed the midline of the Meissner corpuscles. Dashed blue line indicate the Meissner corpuscle midline. Blue arrow heads indicate sites where Nefh^{+} fibers cross this midline. B) RAMs of the glabrous skin exhibited similar vibration threshold and firing activity to a 25-Hz vibration in WT (n = 16 from 4 mice) and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice (n = 21 from 6 mice). Glabrous RAMs showed a non-significant decrease in firing activity to a ramp of 50 Hz vibration with increasing amplitude in Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ compared to WT littermates, but their response to ramp stimuli was similar in both genotypes. Traces indicate the type of stimulation and red squares the time frame during which the number of spikes was calculated. *** $p \leq 0.001$.

Figure 6: *Meis2* gene inactivation affects hair follicle innervation and RAM fibers electrophysiological responses in the hairy skin. A) Representative images of whole mount immunostaining for Nfeh^{+} sensory projections in the hairy skin of adult WT and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ embryos. Scale bar=100µm. B) Box plots showing the quantification for the number of branch points in the innervation network and for the number of innervated hair follicles. n=3; * *p≤0.05*. C) RAMs in the hairy skin of Isl1^{Cre}::Meis2$^{LoxP/LoxP}$ mice (n = 24 from 3 mice) exhibited significantly increased vibration threshold and reduced firing activity to a 25-Hz vibration compared to WT mice (n = 20 from 3 mice). Traces show the stimulation and red square indicate the time frame when the numbers of spikes were calculated. RAMs in the hairy skin of Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice also showed a reduced firing activity in response to a ramp of 50 Hz vibration with increasing amplitude compared to WT and Isl1$^{+/Cre}$ animals. Fibers recorded from Isl1$^{+/Cre}$ mice (n=5) showed similar responses than those recorded from WT mice. * *p≤0.05*; ** *p≤0.005*.

Figure 1 supplementary 1: *Meis2* mRNA expression in LTM neurons of mouse DRG. A) Double *ISH* for *Meis2* (blue) and *Ntrk2*, *Ntrk3* or *c-Ret* (red) showed that *Meis2* mRNA partly colocalises with mRNA for *Ntrk2*, *Ntrk3* and *c-Ret* in E14.5 mouse embryos. Arrows point at double positive neurons. Arrowheads point at only Meis2^{+} neurons. Stars indicate Meis2^{-}/Ntrk2^{+} or Ntrk3^{+} neurons. Note that all large c-Ret-positive neurons (pseudo-color in green) express *Meis2* mRNA. Scale bar=25µm. We estimated that 16.0 ± 1.2% (mean ± SEM; n = 3) of Meis2-positive neurons co-expressed Ntrk2 and that 43.7 ± 2.1% co-expressed Ntrk3. Conversely, Meis2 was co-expressed in 55.9 ± 3.1% of Ntrk2-positive neurons and in 79.1 ± 3.4% of Ntrk3-positive neurons. B) Double ISH for *Meis2* (blue) and *Ntrk2* or *Ntrk3* (red) on E18.5 embryos. Arrows point at double positive neurons. Arrowheads point at only Meis2^{+} neurons. Stars indicate Meis2^{-}/Ntrk2^{+} or Ntrk3^{+} neurons. Scale bar=25µm. We estimated that 28.7 ± 2.5 of Meis2-positive neurons co-expressed Ntrk2 and that 57.8 ± 5.1 co-expressed Ntrk3. Conversely, Meis2 was co-expressed in 52.1 ± 4.5 % of Ntrk2-positive neurons and 63.8 ± 5.1 ± 3.4% of Ntrk3-positive neurons. C) Combined *ISH* for *Meis2* mRNA with IF for Ntrk1 (red) and Islet1 (green) showed that *Meis2* is expressed by Islet1-positive post-mitotic neurons and is mostly excluded from the Ntrk1-positive subpopulation of DRG sensory neurons at E11.5 and E18.5. Arrowheads point at Meis2^{+}/Ntrk1^{-} neurons, arrows point at Meis2^{+}/Ntrk1^{+} neurons. Note that the level of *Meis2* mRNA expression in Ntrk1^{+}/Meis2^{+} neurons is very low at the limit of detection. Dashed lines delineate the DRG. Scale bar=50µm. D) *ISH* for *Meis2* (blue) combined with IF against Ntrk1, Calca, Trpv1, Pvalb (red) and Islet1 or Nefh (green) in adult mouse lumbar DRG. Arrows point at Meis2^{+} neurons. Arrowheads point at Ntrk1^{+}, Calca^{+}, Trpv1^{+} or Pvalb^{+} neurons. Stars indicate neurons that are both positive for Meis2 and Ntrk1, Calca, Trpv1 or Pvalb. Graphs showing the percentage of Meis2^{+} neurons showing immunoreactivity for Ntrk1, Trpv1, Calca, Pvalb and Nefh,

and the percentage of Isl1$^+$, Ntrk1$^+$, Calca$^+$, Trpv1$^+$, Pvalb$^+$ or Nefh$^+$ neurons co-expressing Meis2. Scale bar=50µm.

Figure 1 Supplementary 2: *Meis2* is expressed in a subset of chick ventro-lateral DRG sensory neurons during embryogenesis. A) Developmental expression of *Meis2* visualized by *ISH* in chick DRG at HH24, HH29 and HH36. Dashed lines delineate the DRGs split into the ventro-lateral (VL) and dorso-medial (DM) parts. At early stages after DRG condensation, *Meis2* showed a broad expression in chick DRGs neurons. As differentiation progresses (HH29 and HH36), *Meis2* expression became progressively restricted to the VL population of sensory neurons which represents the Ntrk2$^+$ and Ntrk3$^+$ populations of mechano- and proprioceptive neurons. Meis2-positive cells represented 16.4 ± 1.1 and 14.4 ± 1.2% (mean ± SEM; n = 3) of Islet1-positive DRG neurons at HH29 and HH36 respectively, B) Combined *ISH* for *Meis2* (blue) with IF against Islet1 (green) and Ntrk2 or Ntrk3 (red) in chick lumbar DRGs at HH29 showed that Meis2 expression is shared between the Ntrk2$^+$ and Ntrk3$^+$ subpopulations of sensory neurons. Arrowheads point at Meis2$^+$/Islet1$^+$/Ntrk2$^-$ and Meis2$^+$/Islet1$^+$/Ntrk3$^-$ neurons; arrows point at Meis2$^+$/Islet1$^+$/Ntrk2$^+$ and Meis2$^+$/Islet1$^+$/Ntrk3$^+$ neurons; stars indicate Meis2$^-$/Islet1$^+$/Ntrk2$^+$ and Meis2$^-$/Islet1$^+$/Ntrk3$^+$ neurons. We estimated that 38.4 ± *7.7* and 41.7 ± *3.0*% (mean ± *SEM*; n = 3) of *Meis2*-positive neurons co-expressed *Ntrk2* or *Ntrk3* respectively, and inversely, that 62.1 ± *3.8* and 45.1 ± *4.7*% (mean ± *SEM*; n = 3) of *Ntrk2*- and *Ntrk3*-positive neurons co-expressed *Meis2* mRNA respectively. C) Combined *ISH* for *Meis2* (blue) with IF against Islet1 (green) and Ntrk1 (red) in chick lumbar DRGs at HH29 showed that Meis2 is excluded from the Ntrk1$^+$ population of sensory neurons. Arrowheads point at Meis2$^+$/Islet1$^+$/Ntrk1$^-$ neurons. Enlargement is indicated by a dashed square. D) Representative images showing immunostaining for Ntrk2 or Ntrk3 (red) and Islet1 (green) on HH36 chick embryos DRG sections following limb ablation at HH18. Dashed lines delineate the DRGs. Arrows point at Ntrk2-positive VL neurons. Arrow heads point at Ntrk2-positive DL neurons. C) Box plots showing the percentage of Ntrk2 and Ntrk3 VL neurons and of Ntrk2 DL neurons. *** $p \leq 0.0005$; *ns*= not significant following Student t-test. n=3 chick embryos.

Figure 1 Supplementary 3: Mice with a conditional deletion of *Meis2* gene in neural crest derivatives (Wnt1^{Cre}) exhibited cleft-palate and died at birth. A) Targeting vector used for the generation of a conditional knockout mouse strain for Meis2 (Meis2$^{LoxP/LoxP}$). The Exon 8 (first coding exon of the DNA binding Homeodomain) is flanked by LoxP sites allowing deletion of Meis2 DNA binding domain. B) Representative images showing that new-born Wnt1^{Cre}::Meis2$^{LoxP/LoxP}$ pups mutant pups were smaller in size at birth compared to WT littermates, were unable to stand on their

legs and exhibited a cleft palate phenotype. Representatives images showing coronal sections of new-born WT and Wnt1^{Cre}::Meis2$^{LoxP/LoxP}$ heads stained by eosin/hematoxylin treatment. Black arrows indicate the cleft palate.

Figure 1 Supplementary Table 1: Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ adult mice exhibit normal locomotion. Table recapitulating different Catwalk two-paw analysis parameters in 3 months old female mice. Several recordings were performed for each mice. Only sequence when mice showed a constant and straight locomotion with an average speed between 25 and 55 cm/sec were selected for analysis. Student-t-test analysis showed no significant differences for any of the parameters.

Figure 2 Supplementary 1: *Meis2* gene inactivation does not affect phospho-Creb expression. Representative images showing phosphor-Creb (pCreb in green), Ntrk2 or Ntrk3 (red) and Isl1 (blue) expression in WT and Isl$^{1+/Cre}$::Meis2$^{LoxP/LoxP}$ E16.5 DRG embryos.

Figure 3 Supplementary Table 1: Table showing the results of the bulk RNAseq analysis.

Figure 3 Supplementary Table 2: Table showing the results of the GO-terms analysis performed with David.

Figure 3 Supplementary 1: A) Volcano plots showing differentially expressed genes (DEGs) in red including genes with of minimal fold change of 20% (n=3 ; $p<0.05$). Volcano plots show the comparison between the 3 genotypes (WT, *Isl1*$^{Cre/+}$and *Isl1*$^{Cre/+}$::*Meis2*$^{LoxP/LoxP}$. Plots reporting *Meis2* and *Isl1* mRNA expression are in blue and green respectively. B) Graphs showing the individual number of reads for *Meis2* and *Isl1* genes in each genotype. n=3; *p* values are indicated.

Figure 3 Supplementary 2: Comparison of DEGs in Meis2 mutant embryonic DRG neurons with their expression in the different adult DRG sensory neurons subtypes. A) Heat map showing the expression of down- (A) and up- (B) regulated genes in the different adult DRG sensory neurons subtype according to Usoskin et al.(*17*). Red frames indicate Meis2-expressing subpopulations.

Figure 5 Supplementary video 1: Meissner corpuscle in WT. 3D visualization of Meissner corpuscles in adult WT glabrous skin visualized by IF against Nefh and S100β.

Figure 5 Supplementary video 2: Meissner corpuscles in Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$. 3D visualization of Meissner corpuscles in adult Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ glabrous skin visualized by IF against Nefh and S100β.

Figure 5 Supplementary 1: Normal electrophysiological responses of D-Hair following *Meis2* gene inactivation. A) D-Hair of the hairy skin exhibited similar vibration threshold and firing activity to a 25-Hz vibration in WT (n=8 from 3 mice) and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ mice (n = 10 from 3 mice). B) D-Hair showed similar responses to a ramp of 50 Hz vibration with increasing amplitude and to a ramp stimuli in WT and Isl1$^{+/Cre}$::Meis2$^{LoxP/LoxP}$ animals. Traces showed the type of stimulation and red squares indicate the time frame during when the number of spikes was calculated.

Animals.

All procedures involving animals and their care were conducted according to European Parliament Directive 2010/63/EU and the 22 September 2010 Council on the protection of animals, and were approved by the French Ministry of research (APAFIS#17869-2018112914501928 v2, June the 4th fo 2020).

Mice strains: Wnt1^{CRE} and Islet1$^{+/CRE}$ mice were previously described(*75*, *76*). We previously reported on the conditional mutant strain for Meis2 (Meis2$^{LoxP/LoxP}$) used in the present study(*74*). To generate this strain, exon 8 of the Meis2 gene was flanked by the LoxP recognition elements for the Cre recombinase at the ITL (Ingenious Targeting Laboratory, NY, USA) using standard homologous recombination technology in mouse embryonic stem cells. FLP-FRT recombination was used to remove the neomycin selection cassette and the Meis2$^{LoxP/LoxP}$ mutant mice were backcrossed for at least 8 generations onto the C57BL/6 background before use. Primers used to genotype the different strains were: Meis2 sense 5'-TGT TGG GAT CTG GTG ACT TG-3'; Meis2 antisense 5'-ACT TCA TGG GCT CCT CAC AG-3'; CRE sense 5'-TGC CAG GAT CAG GGT TAA AG-3'; CRE antisense 5'-GCT TGC ATG ATC TCC GGT AT-3'. Mice were kept in an animal facility and gestational stages were determined according to the date of the vaginal plug.

For retro-tracing experiments, newborn pups were anesthetized on ice and cholera toxin B coupled to Cholera Toxin Subunit B conjugated with Alexa Fluo 488 (Thermofisher) was injected using a glass micropipette in several points of the glabrous and hairy forepaw. Mice were sacrificed 7 days after injection and L4 to L6 DRGs were collected for analysis.

For behavioral assays, skin-nerve preparation and electrophysiological recording, sex-matched 12-weeks old mutant and WT littermates mice were used.

Chick: Fertilized eggs were incubated at 37°C in a humidified incubator. For limb ablation experiments, eggs were opened on the third day of incubation (embryonic day 3, stage 17/18) (*77*) and the right hind limb bud was surgically removed as previously reported (*38*). Eggs were closed with tape and further grown in the incubator for 4 (HH27) or 7 (HH36) additional days before collection.

Tissue preparation.

Mouse and chick embryos were collected at different stages, fixed in 4% paraformaldehyde/PBS overnight at 4°C and incubated overnight at 4°C for cryopreservation in increasing sucrose/PBS solutions (10 to 30% sucrose). After snap freezing in TissueTek, embryos were sectioned at 14-µm thickness and stored at -20°C until use.

Cloning of mouse and chick Meis2 and probes preparation.

For preparation of digoxigenin- and fluorescein- labeled probes, RNA from whole mouse or chick embryos was extracted using Absolutely RNATM Nanoprep kit (Stratagene) following manufacturer's instruction. Reverse transcription (RT) was carried out 10 minutes at 65°C followed by 1 hour at 42°C and 15 minutes at 70°C in 20 µl reactions containing 0.5 mM dNTP each, 10 mM DTT, 0.5 µg oligod(T)15 (Promega) and 200 U of Super Script II RT (Gibco BRL Life Technologies). A 1206 bp long and a 1201 bp long Meis2 fragments were amplified from mouse and gallus cDNA respectively using the following primers: mMeis2 forward: 5'-ATGGCGCAAAGGTACGATGAGCT-3'; mMeis2 reverse: 5'-TTACATATAGTGCCACTGCCCATC-3'; gMeis2 forward: 5'-ATGGCGCAAAGGTACGATGAG-3'; gMeis2 reverse: 5'-TTACATGTAGTGCCATTGCCCAT-3'. PCR were conducted in 50 µl reactions containing 10% RT product, 200 µM each dNTP, 10 pmol of each primer (MWG-Biotech AG), 3 mM MgCl2, 6% DMSO and 2.5 U of Herculase hotstart DNA polymerase (Stratagene). cDNA was denatured 10 minutes at 98°C and amplified for 35 cycles in a three steps program as following: 1 minute denaturation at 98°C, 1 minute annealing at annealing temperature and then 1.5 minutes polymerization at 72°C. PCR products were separated on 2% agarose gels containing ethidium bromide. Bands at the expected size were excised, DNA was extracted, and the fragment was cloned into pCR4Blunt-TOPO vector (Invitrogen) and confirmed by sequencing. Other probes used for ISH have been described elsewhere (*29*).

***In situ* hybridization (ISH).**

Before hybridization, slides were air dried for 2-3 hours at room temperature. Plasmids containing probes were used to synthesize digoxigenin-labeled or fluorescein-labeled antisense riboprobes according to supplier's instructions (Roche) and purified by LiCl precipitation. Sections were hybridized overnight at 70°C with a solution containing 0.19 M NaCl, 10 mM Tris (pH 7.2), 5 mM NaH2PO4*2H2O/Na2HPO4 (pH 6.8), 50 mM EDTA, 50% formamide, 10% dextran sulphate, 1 mg/ml yeast tRNA, 1XDenhardt solution and 100 to 200 ng/ml of probe. Sections were then washed four times for 20 minutes at 65°C in 0.4X SSC pH 7.5, 50% formamide, 0.1% Tween 20 and three times for 20 minutes at room temperature in 0.1 M maleic acid, 0.15 M NaCl and 0.1% Tween 20 (pH 7.5). Sections were blocked 1 hour at room temperature in presence of 20% goat serum and 2% blocking agent (Roche) prior to incubation overnight with AP-conjugated anti-DIG-Fab-fragments (Roche, 1:2000). After extensive washing, hybridized riboprobes were revealed by performing a NBT/BCIP reaction in 0.1 M Tris HCl pH 9.5, 100 mM NaCl, 50 mM MgCl2 and 0.1% Tween 20.

For double *in situ* hybridization, the procedure was the same except that hybridization was conducted by incubation with 100-200 ng/ml of one digoxigenin-labeled probe and 100-200 ng/ml of

one fluorescein-labeled probe. Fluorescein-labeled probe was first revealed after overnight incubation with AP-conjugated anti-fluorescein-Fab-fragment (Roche, 1:2000) and further incubation with Fast Red tablets in 0.1 M Tris HCl pH 8.5, 100 mM NaCl, 50 mM MgCl2 and 0.1% Tween 20. Pictures of fluorescein alone were taken after mounting in glycerol/PBS (1:9). To reveal the digoxigenin-labeled probe, sections were unmounted, washed extensively in PBS and Alkalin Phosphatase was inhibited by incubation in a solution of 0.1M glycin pH2.2, 0.2% Tween 20 for 30 minutes at room temperature. After extensive washing in PBS, digoxigenin-labeled-probe was revealed as described using the AP-conjugated anti-DIG-Fab-fragments (Roche, 1:2000) and the NBT/BCIP reaction. Sections were mounted again in glycerol/PBS (1:9) and pictures of both fluorescein and digoxigenin were taken. For removing the Fast Red staining, sections were unmounted again, washed extensively in PBS and incubated in increasing solutions of ethanol/PBS solutions (20-100% ethanol). After extensive washing in PBS, sections were mounted in glycerol/PBS (1:9) and pictures of the digoxigenin staining alone. Wide field microscopy (Leica DMRB, Germany) was only used for ISH and ISH combined with immunochemistry.

Immunochemistry.

Immunochemistry was performed as previously described (*29*). *In situ* hybridized sections or new sections were washed 3x10 min with PBS, blocked with 4% normal goat serum, 1% bovine serum albumin and 0.1% Triton X100 in PBS and incubated overnight at 4°C with primary antibodies. After washing 3x10 min with PBS incubation occurred for 2-4 hours with secondary species and isotype-specific fluorescent antibodies (Alexa Fluor Secondary Antibodies, Molecular Probes). After repeated washing with PBS, slides were mounted in Glycerol/PBS (9/1) or Mowiol. Picture were taken using a confocal microscope (Leica SP5-SMD, Germany). Confocal images are presented as maximal projections.

The following antibodies were used for immunochemistry: mouse-anti-islet1 39.4D used for mouse and chick (diluted 1:100, Developmental Studies Hybridoma Bank); TrkB (1/2000; R and D Systems); TrkC (1/1000; R and D Systems); CGRP (1/500; Sigma-Aldrich); TrkA (1/500; Millipore); rabbit anti-parvalbumin antibody (1:500, Swant); guinea pig anti-calcitonin gene-related peptide (CGRP) antibody (1:500, Peninsula Laboratories); Nfh (Sigma N4142, rabbit 1:1000), c-maf (generous gift of C. Birchmeier, MDC, 1/10000), TrpV1 (Sigma V2764, rabbit 1:1000), S100β (Sigma S2532, mouse 1:1000), Phospho-CREB (Cell Signaling, 87G3, 1/200, Germany); Goat anti-c Ret (R and D Systems, Cat#AF482, 1/100) and Meis2 (Sigma Aldrich, WH0004212M1 or Abcam ab244267, 1/500). The chick TrkB and C antibodies were a generous gift from LF Reichardt, UCSF and have been previously reported.

Whole-mount immunohistochemistry.

Whole-mount immunohistochemistry of adult mice back hairy skin was performed a described elsewhere (*78*). Briefly, mice were euthanized by CO_2 asphyxiation. The back skin was shaved and cleaned with commercial hair remover. The back skin was removed, carefully handled with curved forceps and fixated in 4% PFA at 4°C for 20'. The tissue was then washed with PBS containing 0.3% Triton X-100 (PBST) every 30 minutes for 3 to 5 hours and kept overnight in the washing solution. The next day, the skin was incubated for 5 days with primary antibodies diluted in PBST containing 5% donkey serum and 20% DMSO. The skin was washed the following day 8 to 10 times over a day before being incubated with secondary antibodies diluted in PBST containing 5% donkey serum and 20% DMSO. The skin was then washed every 30 minutes for 6 to 8 hours before being dehydrated in successive baths of 25%, 50%, 75% and 100% methanol. They were then incubated overnight in a 1:2 mixture of benzyl alcohol and benzyl benzoate before being mounted and sealed into chambers filled with the same medium.

Hematoxylin-Eosin staining.

As previously described(*29*), air dried frozen sections were washed in water then stained with hematoxylin for 1 min at room temperature and washed extensively with water. After dehydration in PBS/alcohol (70%), slides were stained with eosin for 30 sec at room temperature. After serial wash in water, sections were dehydrated in PBS solutions with increasing alcohol concentration (50%, 75%, 95%, and 100%), mounted and observed with a microscope (Leica DMRB, Germany).

RNA-sequencing and analysis.

DRGs were dissected from E18.5 mouse embryos, collected in lysis buffer and stored at -80°C until RNA extraction with RNeasy extraction kit (Qiagen). After mRNA purification using the NEBNext® Poly(A) mRNA Magnetic Isolation (NEB), libraries were prepared with the CORALL mRNA-Seq Library Prep Kits with UDIs (Lexogen) following manufacturer's recommendations. After a qPCR assay to determine the optimal PCR cycle number for endpoint PCR, 14 PCR cycles were completed to finalize the library preparation. Quantitation and quality assessment of each library were performed using Qubit 4.0 (HS DNA kit, Thermofisher) and 4150 Tapestation analyzer (D5000 ScreenTape kit, Agilent). Indexed libraries were sequenced in an equimolar manner on NextSeq 500 Illumina sequencer. Sequencing conditions were as follow: denatured libraries were loaded on a HighOutput flowcell kit v2 and sequenced in single-end 84pb reads. Data were extracted and processed following Illumina recommendations. After a quality check of the fastq files with FastQC, UMI sequences were extracted with UMI tools (version 1.1.2) (*79*) default parameters followed by STAR alignement (version 2.7.10) (*80*) on mm10 genome and removal of PCR duplicate with UMI tools, default

parameters. Uniquely mapped sequences from the STAR output files (bam format) were then used for further analysis. HT-seq count (version 0.6) (*81*) was used to aggregate read count per gene followed by differential gene expression analysis with Limma voom on Galaxy (version 3.50.1) (*82*). Data are available under the following accession codes: GSE223788. Genes with more than 1.2-fold differential expression and p-value <0.05 were used for gene ontology analysis (https://david.ncifcrf.gov) using the list of expressed genes in our experiment as background.

Mouse skin-nerve preparation and sensory afferent recordings

Cutaneous sensory fiber recordings were performed using the ex vivo skin-nerve preparation as previously described (*12*). Mice were euthanized by CO2 inhalation for 2–4 min followed by cervical dislocation. Three different preparations were performed in separate experiments using different paw regions: the saphenous nerve innervating the hairy hind paw skin; the tibial nerve innervating the glabrous hind paw skin; and the medial and ulnar nerves innervating the forepaw glabrous skin. In all preparations, the hairy skin of the limb was shaved and the skin and nerve were dissected free and transferred to the recording chamber, where muscle, bone and tendon tissues were removed from the skin to improve recording quality. The recording chamber was perfused with a 32 °C synthetic interstitial fluid: 123 mM NaCl, 3.5 mM KCl, 0.7 mM MgSO4, 1.7 mM NaH2PO4, 2.0 mM CaCl2, 9.5 mM sodium gluconate, 5.5 mM glucose, 7.5 mM sucrose and 10 mM 4-(2-hydroxyethyl)-1-piperazine-ethanesulfonic acid (Hepes), pH 7.4. The skin was pinned out and stretched, such that the outside of the skin could be stimulated using stimulator probes. The peripheral nerve was fed through to an adjacent chamber in mineral oil, where fine filaments were teased from the nerve and placed on a silver-wire recording electrode.

The receptive fields of individual mechanoreceptors were identified by mechanically probing the surface of the skin with a blunt glass rod or blunt forceps. Analog output from a Neurolog amplifier was filtered and digitized using the Powerlab 4/30 system and Labchart 7.1 software (AD instruments). Spike-histogram extension for Labchart 7.1 was used to sort spikes of individual units. Electrical stimuli (1 Hz, square pulses of 50–500 ms) were delivered to single-unit receptive fields to measure conduction velocity and enable classification as C-fibers (velocity <1.2 m s−1), Aδ-fibers (1.2–10 m s−1) or Aβ-fibers (>10 m s−1). Mechanical stimulation of the receptive fields of neurons were performed using a piezo actuator (Physik Instrumente, catalog no. P-841.60) and a double-ended Nanomotor (Kleindiek Nanotechnik, catalog no. MM-NM3108) connected to a force measurement device (Kleindiek Nanotechnik, catalog no. PL-FMS-LS). Calibrated force measurements were acquired simultaneously using the Powerlab system and Labchart software during the experiment.

As different fiber types have different stimulus-tuning properties, different mechanical stimuli protocols were used based on the unit type. Low-threshold Aβ-fibers (RAMs and SAMs) and Aδ-fiber D-hairs were stimulated with the piezo actuator with three vibration stimuli (5 Hz, 25 Hz and 50 Hz, distortions introduced by the in-series force sensor precluded using frequencies >50 Hz) with increasing amplitude over six steps (peak-to-peak amplitudes of ~6–65 mN; 20 cycles per step), and a dynamic stimulus sequence with four ramp-and-hold waveforms with varying probe deflection velocities (3 s duration; 0.075, 0.15, 0.45 and 1.5 mm s−1; average amplitude 100 mN). Aβ-fiber SAMs and RAMs were classified by the presence or absence of firing during the static phase of a ramp-and-hold stimulus, respectively, as previously described. Single units were additionally stimulated with a series of five static mechanical stimuli with ramp-and-hold waveforms of increasing amplitude (3 s duration; ranging from ~10 mN to 260 mN). Low-threshold SAMs, high-threshold Aδ-fibers and C-fibers were also stimulated using the nanomotor with five ramp-and-hold stimuli with increasing amplitudes.

Behavioral assays.

Von Frey paw withdrawal test: Mice were placed on an elevated wire mesh grid into PVC chambers. Before the test, mice were habituated to the device for one hour for two consecutive days. On the day test, mice were placed in the chamber one hour before Von Frey filaments application. The test was performed as previously described (*54*). During the test, withdrawal response following Von Frey filament application on the palm of the left hind paw was measured. Starting with the lowest force, each filament ranging from 0.008 g to 1.4 g was applied ten times in a row with a break of 30 seconds following the fifth application. The number of paw withdrawals for each filament was counted.

Hot Plate test: Before starting the test, mice were habituated to the experimentation room for at least five minutes. Mice were individually placed on the hot plate set up at 53°C and removed at the first signs of aversive behavior (paw licking or shaking). The time to this first stimulus was recorded. A 30 sec cut off was applied to avoid skin damages. After 5 min recovery in their home cage, the test was repeated three times for each mouse and averaged. Data are shown as the average of these three measurements.

Sticky tape test: A two cm^2 of laboratory tape was placed on the upper back skin of mice just before they were placed on an elevated wire mesh grid into PVC chambers. The number of tape-directed reactions were then counted during 5 min. Considered responses were body shaking like a “wet-dog”, hindlimb scratching directed to the tape, trying to reach the tape with the snout and grooming of the neck with forepaws.

Dynamic touch test: Mice were placed in the same conditions as described above for the Von Frey paw withdrawal test. Sensitivity to dynamic touch was performed by stroking hind paws with a tapered cotton-swab in a heel to toe direction. The stimulation was repeated 10 times by alternating left and right hind paws and the number of paw withdrawals was counted.

Gait analysis: Gait was analyzed using the Catwalk™ system (Noldus Information Technology, Netherlands) in a dark room with minimized light emission from the computer screen. Mice were allowed to voluntarily cross a 100-cm-long, 5-cm-wide walkway with a glass platform illuminated by green fluorescent light. An illuminated image is produced when a mouse paw touches the glass floor through dispersion of the green light, and footprints were captured by a high-speed camera placed under the glass floor. Data were analyzed using the CatWalk™ XT 10.1 software. For each mouse, several recordings were performed until at least 3 runs met criteria defined by a minimum of three consecutive complete step cycles of all four paws without stopping or hesitation and within the range of 25 to 50 $cm.s^{-1}$. Data are reported as the average of at least three runs per mouse.

References:

1. V. E. Abraira, D. D. Ginty, The sensory neurons of touch. *Neuron*. **79** (2013), pp. 618–639.
2. P. Delmas, J. Hao, L. Rodat-Despoix, Molecular mechanisms of mechanotransduction in mammalian sensory neurons. *Nat. Rev. Neurosci.* **12** (2011), pp. 139–153.
3. A. Handler, D. D. Ginty, The mechanosensory neurons of touch and their mechanisms of activation. *Nat. Rev. Neurosci.* **22** (2021), pp. 521–537.
4. S. G. Lechner, G. R. Lewin, Hairy sensation. *Physiology*. **28** (2013), pp. 142–150.
5. H. Wu, C. Petitpré, P. Fontanet, A. Sharma, C. Bellardita, R. M. Quadros, P. R. Jannig, Y. Wang, J. A. Heimel, K. K. Y. Cheung, S. Wanderoy, Y. Xuan, K. Meletis, J. Ruas, C. B. Gurumurthy, O. Kiehn, S. Hadjab, F. Lallemend, Distinct subtypes of proprioceptive dorsal root ganglion neurons regulate adaptive proprioception in mice. *Nat. Commun.* **12** (2021), doi:10.1038/s41467-021-21173-9.
6. A. Zimmerman, L. Bai, D. D. Ginty, The gentle touch receptors of mammalian skin. *Science (80-.).* **346** (2014), pp. 950–954.
7. L. Li, M. Rutlin, V. E. Abraira, C. Cassidy, L. Kus, S. Gong, M. P. Jankowski, W. Luo, N. Heintz, H. R. Koerber, C. J. Woodbury, D. D. Ginty, The functional organization of cutaneous low-threshold mechanosensory neurons. *Cell.* **147**, 1615–1627 (2011).
8. F. Lallemend, P. Ernfors, Molecular interactions underlying the specification of sensory neurons. *Trends Neurosci.* **35** (2012), pp. 373–381.
9. F. Marmigère, P. Ernfors, *Specification and connectivity of neuronal subtypes in the sensory lineage* (Nat Rev Neurosci, 2007; https://pubmed-ncbi-nlm-nih-gov.insb.bib.cnrs.fr/17237804/), vol. 8.
10. S. Vermeiren, E. J. Bellefroid, S. Desiderio, Vertebrate Sensory Ganglia: Common and Divergent Features of the Transcriptional Programs Generating Their Functional Specialization. *Front. Cell Dev. Biol.* **8** (2020), , doi:10.3389/fcell.2020.587699.
11. N. Sharma, K. Flaherty, K. Lezgiyeva, D. E. Wagner, A. M. Klein, D. D. Ginty, The emergence of transcriptional identity in somatosensory neurons. *Nature*. **577**, 392–398 (2020).
12. F. Schwaller, V. Bégay, G. García-García, F. J. Taberner, R. Moshourab, B. McDonald, T. Docter, J. Kühnemund, J. Ojeda-Alonso, R. Paricio-Montesinos, S. G. Lechner, J. F. A. Poulet, J. M. Millan, G. R. Lewin, USH2A is a Meissner's corpuscle protein necessary for normal vibration sensing in mice and humans. *Nat. Neurosci.* **24**, 74–81 (2021).
13. J. Walcher, J. Ojeda-Alonso, J. Haseleu, M. K. Oosthuizen, A. H. Rowe, N. C. Bennett, G. R. Lewin, Specialized mechanoreceptor systems in rodent glabrous skin. *J. Physiol.* **596**, 4995–5016 (2018).
14. G. R. Lewin, S. B. McMahon, Physiological properties of primary sensory neurons appropriately and inappropriately innervating skin in the adult rat. *J. Neurophysiol.* **66**, 1205–1217 (1991).
15. P. R. Burgess, K. W. Horch, Specific regeneration of cutaneous fibers in the cat. *J. Neurophysiol.* **36**, 101–114 (1973).
16. L. Bai, B. P. Lehnert, J. Liu, N. L. Neubarth, T. L. Dickendesher, P. H. Nwe, C. Cassidy, C. J. Woodbury, D. D. Ginty, Genetic Identification of an Expansive Mechanoreceptor Sensitive to Skin Stroking. *Cell*. **163**, 1783–1795 (2015).
17. D. Usoskin, A. Furlan, S. Islam, H. Abdo, P. Lönnerberg, D. Lou, J. Hjerling-Leffler, J. Haeggström, O. Kharchenko, P. V. Kharchenko, S. Linnarsson, P. Ernfors, Unbiased classification of sensory neuron types by large-scale single-cell RNA sequencing. *Nat. Neurosci.* **18**, 145–153 (2015).
18. Y. Zheng, P. Liu, L. Bai, J. S. Trimmer, B. P. Bean, D. D. Ginty, Deep Sequencing of Somatosensory Neurons Reveals Molecular Determinants of Intrinsic Physiological Properties. *Neuron*. **103**, 598-616.e7 (2019).
19. A. Scott, H. Hasegawa, K. Sakurai, A. Yaron, J. Cobb, F. Wang, Transcription factor short stature homeobox 2 is required for proper development of tropomyosin-related kinase B-expressing mechanosensory neurons. *J. Neurosci.* **31**, 6741–6749 (2011).
20. H. Abdo, L. Li, F. Lallemend, I. Bachy, X. J. Xu, F. L. Rice, P. Ernfors, Dependence on the transcription factor Shox2 for specification of sensory neurons conveying discriminative touch. *Eur. J. Neurosci.* **34**, 1529–1541 (2011).
21. S. Arber, D. R. Ladle, J. H. Lin, E. Frank, T. M. Jessell, ETS gene Er81 controls the formation of functional connections between group Ia sensory afferents and motor neurons. *Cell.* **101**, 485–498 (2000).
22. J. Hu, T. Huang, T. Li, Z. Guo, L. Cheng, C-Maf is required for the development of dorsal horn laminae III/IV neurons and mechanoreceptive DRG axon projections. *J. Neurosci.* **32**, 5362–5373 (2012).
23. S. Meltzer, K. C. Boulanger, A. M. Chirila, E. Osei-Asante, M. DeLisle, Q. Zhang, B. T. Kalish, A. Tasnim, E. L. Huey, L. C. Fuller, E. K. Flaherty, T. Maniatis, A. M. Garrett, J. A. Weiner, D. D. Ginty, γ-Protocadherins

control synapse formation and peripheral branching of touch sensory neurons. *Neuron* (2023), doi:10.1016/J.NEURON.2023.03.012.

24. L. L. L. Orefice, A. L. L. Zimmerman, A. M. M. Chirila, S. J. J. Sleboda, J. P. P. Head, D. D. D. Ginty, Peripheral Mechanosensory Neuron Dysfunction Underlies Tactile and Behavioral Deficits in Mouse Models of ASDs. *Cell*. **166**, 299–313 (2016).
25. L. L. Orefice, J. R. Mosko, D. T. Morency, M. F. Wells, A. Tasnim, S. M. Mozeika, M. Ye, A. M. Chirila, A. J. Emanuel, G. Rankin, R. M. Fame, M. K. Lehtinen, G. Feng, D. D. Ginty, Targeting Peripheral Somatosensory Neurons to Improve Tactile-Related Phenotypes in ASD Models. *Cell*. **178**, 867-886.e24 (2019).
26. A. Gangfuß, G. Yigit, J. Altmüller, P. Nürnberg, J. C. Czeschik, B. Wollnik, N. Bögershausen, P. Burfeind, D. Wieczorek, F. Kaiser, A. Roos, H. Kölbel, U. Schara-Schmidt, A. Kuechler, Intellectual disability associated with craniofacial dysmorphism, cleft palate, and congenital heart defect due to a de novo MEIS2 mutation: A clinical longitudinal study. *Am. J. Med. Genet. Part A*. **185**, 1216–1221 (2021).
27. D. Geerts, N. Schilderink, G. Jorritsma, R. Versteeg, "The role of the MEIS homeobox genes in neuroblastoma" in *Cancer Letters* (Elsevier Ireland Ltd, 2003), vol. 197, pp. 87–92.
28. E. Longobardi, D. Penkov, D. Mateos, G. De Florian, M. Torres, F. Blasi, Biochemistry of the tale transcription factors PREP, MEIS, and PBX in vertebrates. *Dev. Dyn.* **243** (2014), pp. 59–75.
29. F. Bouilloux, J. Thireau, S. Ventéo, C. Farah, S. Karam, Y. Dauvilliers, J. Valmier, N. G. Copeland, N. A. Jenkins, S. Richard, F. Marmigère, Loss of the transcription factor Meis1 prevents sympathetic neurons target-field innervation and increases susceptibility to sudden cardiac death. *Elife*. **5** (2016), doi:10.7554/eLife.11627.
30. J. Gilbert, H. Y. Man, Fundamental elements in autism: From neurogenesis and neurite growth to synaptic plasticity. *Front. Cell. Neurosci.* **11** (2017), , doi:10.3389/fncel.2017.00359.
31. K. McFadden, N. J. Minshew, Evidence for dysregulation of axonal growth and guidance in the etiology of ASD. *Front. Hum. Neurosci.* **7** (2013), , doi:10.3389/fnhum.2013.00671.
32. J. T. Rifkin, V. J. Todd, L. W. Anderson, F. Lefcort, Dynamic expression of neurotrophin receptors during sensory neuron genesis and differentiation. *Dev. Biol.* **227**, 465–480 (2000).
33. M. M. Shin, C. Catela, J. Dasen, Intrinsic control of neuronal diversity and synaptic specificity in a proprioceptive circuit. *Elife*. **9**, 1–27 (2020).
34. T. D. Patel, I. Kramer, J. Kucera, V. Niederkofler, T. M. Jessell, S. Arber, W. D. Snider, Peripheral NT3 signaling is required for ETS protein expression and central patterning of proprioceptive sensory afferents. *Neuron*. **38**, 403–416 (2003).
35. J. C. de Nooij, S. Doobar, T. M. Jessell, Etv1 Inactivation Reveals Proprioceptor Subclasses that Reflect the Level of NT3 Expression in Muscle Targets. *Neuron*. **77**, 1055–1068 (2013).
36. S. Poliak, A. L. Norovich, M. Yamagata, J. R. Sanes, T. M. Jessell, Muscle-type Identity of Proprioceptors Specified by Spatially Restricted Signals from Limb Mesenchyme. *Cell*. **164**, 512–525 (2016).
37. Y. Wang, H. Wu, P. Zelenin, P. Fontanet, S. Wanderoy, C. Petitpré, G. Comai, C. Bellardita, Y. Xue-Franzén, R. E. Huettl, A. B. Huber, S. Tajbakhsh, O. Kiehn, P. Ernfors, T. G. Deliagina, F. Lallemend, S. Hadjab, Muscle-selective RUNX3 dependence of sensorimotor circuit development. *Dev.* **146** (2019), doi:10.1242/dev.181750.
38. R. A. Oakley, A. S. Garner, T. H. Large, E. Frank, Muscle sensory neurons require neurotrophin-3 from peripheral tissues during the period of normal cell death. *Development*. **121**, 1341–1350 (1995).
39. R. A. Oakley, F. B. Lefcort, D. O. Clary, L. F. Reichardt, D. Prevette, R. W. Oppenheim, E. Frank, Neurotrophin-3 promotes the differentiation of muscle spindle afferents in the absence of peripheral targets. *J. Neurosci.* **17**, 4262–4274 (1997).
40. J. Calderó, D. Prevette, X. Mei, R. A. Oakley, L. Li, C. Milligan, L. Houenou, M. Burek, R. W. Oppenheim, Peripheral target regulation of the development and survival of spinal sensory and motor neurons in the chick embryo. *J. Neurosci.* **18**, 356–370 (1998).
41. O. Machon, J. Masek, O. Machonova, S. Krauss, Z. Kozmik, Meis2 is essential for cranial and cardiac neural crest development. *BMC Dev. Biol.* **15** (2015), doi:10.1186/s12861-015-0093-6.
42. S. Bourane, K. S. Grossmann, O. Britz, A. Dalet, M. G. Del Barrio, F. J. Stam, L. Garcia-Campmany, S. Koch, M. Goulding, Identification of a spinal circuit for light touch and fine motor control. *Cell*. **160**, 503–515 (2015).
43. C. Catela, M. M. Shin, D. H. Lee, J. P. Liu, J. S. Dasen, Hox Proteins Coordinate Motor Neuron Differentiation and Connectivity Programs through Ret/Gfrα Genes. *Cell Rep.* **14**, 1901–1915 (2016).
44. J. S. Dasen, B. C. Tice, S. Brenner-Morton, T. M. Jessell, A Hox regulatory network establishes motor neuron pool identity and target-muscle connectivity. *Cell*. **123**, 477–491 (2005).

45. J. Ericson, S. Thor, T. Edlund, T. M. Jessell, T. Yamada, Early stages of motor neuron differentiation revealed by expression of homeobox gene Islet-1. *Science (80-.).* **256**, 1555–1560 (1992).
46. C. Wetzel, J. Hu, D. Riethmacher, A. Benckendorff, L. Harder, A. Eilers, R. Moshourab, A. Kozlenkov, D. Labuz, O. Caspani, B. Erdmann, H. Machelska, P. A. Heppenstall, G. R. Lewin, A stomatin-domain protein essential for touch sensation in the mouse. *Nature*. **445**, 206–209 (2007).
47. S. S. Ranade, S. H. Woo, A. E. Dubin, R. A. Moshourab, C. Wetzel, M. Petrus, J. Mathur, V. Bégay, B. Coste, J. Mainquist, A. J. Wilson, A. G. Francisco, K. Reddy, Z. Qiu, J. N. Wood, G. R. Lewin, A. Patapoutian, Piezo2 is the major transducer of mechanical forces for touch sensation in mice. *Nature*. **516**, 121–125 (2014).
48. J. B. Shin, C. Martinez-Salgado, P. A. Heppenstall, G. R. Lewin, A T-type calcium channel required for normal function of a mammalian mechanoreceptor. *Nat. Neurosci.* **6**, 724–730 (2003).
49. M. Jakovcevski, H. Ruan, E. Y. Shen, A. Dincer, B. Javidfar, Q. Ma, C. J. Peter, I. Cheung, A. C. Mitchell, Y. Jiang, C. L. Lin, V. Pothula, A. Francis Stewart, P. Ernst, W. D. Yao, S. Akbarian, Neuronal Kmt2a/Mll1 histone methyltransferase is essential for prefrontal synaptic plasticity and working memory. *J. Neurosci.* **35**, 5097–5108 (2015).
50. Z. Agoston, P. Heine, M. S. Brill, B. M. Grebbin, A. C. Hau, W. Kallenborn-Gerhardt, J. Schramm, M. Götz, D. Schulte, Meis2 is a Pax6 co-factor in neurogenesis and dopaminergic periglomerular fate specification in the adult olfactory bulb. *Dev.* **141**, 28–38 (2014).
51. J. Niu, A. Vysochan, W. Luo, Dual Innervation of neonatal Merkel cells in mouse touch domes. *PLoS One*. **9** (2014), doi:10.1371/journal.pone.0092027.
52. W. Olson, P. Dong, M. Fleming, W. Luo, The specification and wiring of mammalian cutaneous low-threshold mechanoreceptors. *Wiley Interdiscip. Rev. Dev. Biol.* **5**, 389–404 (2016).
53. L. R. Mills, C. A. Nurse, J. Diamond, The neural dependency of Merkel cell development in the rat: The touch domes and foot pads contrasted. *Dev. Biol.* **136**, 61–74 (1989).
54. N. L. Neubarth, A. J. Emanuel, Y. Liu, M. W. Springel, A. Handler, Q. Zhang, B. P. Lehnert, C. Guo, L. L. Orefice, A. Abdelaziz, M. M. DeLisle, M. Iskols, J. Rhyins, S. J. Kim, S. J. Cattel, W. Regehr, C. D. Harvey, J. Drugowitsch, D. D. Ginty, Meissner corpuscles and their spatially intermingled afferents underlie gentle touch perception. *Science (80-.).* **368** (2020), doi:10.1126/science.abb2751.
55. F. Guillemot, B. A. Hassan, Beyond proneural: emerging functions and regulations of proneural proteins. *Curr. Opin. Neurobiol.* **42** (2017), pp. 93–101.
56. N. E. Baker, N. L. Brown, All in the family: Proneural bHLH genes and neuronal diversity. *Dev.* **145** (2018), , doi:10.1242/dev.159426.
57. O. Hobert, "Terminal Selectors of Neuronal Identity" in *Current Topics in Developmental Biology* (Academic Press Inc., 2016), vol. 116, pp. 455–475.
58. O. Hobert, P. Kratsios, Neuronal identity control by terminal selectors in worms, flies, and chordates. *Curr. Opin. Neurobiol.* **56** (2019), pp. 97–105.
59. Q. Ma, C. Fode, F. Guillemot, D. J. Anderson, NEUROGENIN1 and NEUROGENIN2 control two distinct waves of neurogenesis in developing dorsal root ganglia. *Genes Dev.* **13**, 1717–1728 (1999).
60. H. Wende, S. G. Lechner, C. Cheret, S. Bourane, M. E. Kolanczyk, A. Pattyn, K. Reuter, F. L. Munier, P. Carroll, G. R. Lewin, C. Birchmeier, The transcription factor c-Maf controls touch receptor development and function. *Science*. **335**, 1373–6 (2012).
61. M. Yoshikawa, Y. Murakami, K. Senzaki, T. Masuda, S. Ozaki, Y. Ito, T. Shiga, Coexpression of Runx1 and Runx3 in mechanoreceptive dorsal root ganglion neurons. *Dev. Neurobiol.* **73**, 469–479 (2013).
62. J. H. Lin, T. Saito, D. J. Anderson, C. Lance-Jones, T. M. Jessell, S. Arber, Functionally related motor neuron pool and muscle sensory afferent subtypes defined by coordinate ETS gene expression. *Cell.* **95**, 393–407 (1998).
63. K. ichi Inoue, S. Ozaki, T. Shiga, K. Ito, T. Masuda, N. Okado, T. Iseda, S. Kawaguchi, M. Ogawa, S. C. Bae, N. Yamashita, S. Itohara, N. Kudo, Y. Ito, Runx3 controls the axonal projection of proprioceptive dorsal root ganglion neurons. *Nat. Neurosci.* **5**, 946–954 (2002).
64. F. Marmigère, P. Carroll, Neurotrophin signalling and transcription programmes interactions in the development of somatosensory neurons. *Handb. Exp. Pharmacol.* **220**, 329–353 (2014).
65. A. Giliberti, A. Currò, F. T. Papa, E. Frullanti, F. Ariani, G. Coriolani, S. Grosso, A. Renieri, F. Mari, MEIS2 gene is responsible for intellectual disability, cardiac defects and a distinct facial phenotype. *Eur. J. Med. Genet.* **63** (2020), doi:10.1016/j.ejmg.2019.01.017.
66. H. Toresson, A. M. De Urquiza, C. Fagerström, T. Perlmann, K. Campbell, Retinoids are produced by glia in the lateral ganglionic eminence and regulate striatal neuron differentiation. *Development*. **126**, 1317–1326 (1999).

67. H. Toresson, M. Parmar, K. Campbell, Expression of Meis and Pbx genes and their protein products in the developing telencephalon: Implications for regional differentiation. *Mech. Dev.* **94**, 183–187 (2000).
68. I. Chang, M. Parrilla, Expression patterns of homeobox genes in the mouse vomeronasal organ at postnatal stages. *Gene Expr. Patterns*. **21**, 69–80 (2016).
69. S. Frazer, J. Prados, M. Niquille, C. Cadilhac, F. Markopoulos, L. Gomez, U. Tomasello, L. Telley, A. Holtmaat, D. Jabaudon, A. Dayer, Transcriptomic and anatomic parcellation of 5-HT3A R expressing cortical interneuron subtypes revealed by single-cell RNA sequencing. *Nat. Commun.* **8** (2017), doi:10.1038/ncomms14219.
70. K. M. Bumsted-O'Brien, A. Hendrickson, S. Haverkamp, R. Ashery-Padan, D. Schulte, Expression of the homeodomain transcription factor Meis2 in the embryonic and postnatal retina. *J. Comp. Neurol.* **505**, 58–72 (2007).
71. W. Yan, M. A. Laboulaye, N. M. Tran, I. E. Whitney, I. Benhar, J. R. Sanes, Mouse Retinal Cell Atlas: Molecular Identification of over Sixty Amacrine Cell Types. *J. Neurosci.* **40**, 5177–5195 (2020).
72. L. Yang, Z. Su, Z. Wang, Z. Li, Z. Shang, H. Du, G. Liu, D. Qi, Z. Yang, Z. Xu, Z. Zhang, Transcriptional profiling reveals the transcription factor networks regulating the survival of striatal neurons. *Cell Death Dis.* **12**, 262 (2021).
73. Z. J. Allen, R. R. Waclaw, M. C. Colbert, K. Campbell, Molecular identity of olfactory bulb interneurons: Transcriptional codes of periglomerular neuron subtypes. *J. Mol. Histol.* **38**, 517–525 (2007).
74. J. Roussel, R. Larcher, P. Sicard, P. Bideaux, S. Richard, F. Marmigère, J. Thireau, The autism-associated Meis2 gene is necessary for cardiac baroreflex regulation in mice. **12** (2022) (available at https://pubmed-ncbi-nlm-nih-gov.insb.bib.cnrs.fr/36418415/).
75. L. Yang, C. L. Cai, L. Lin, Y. Qyang, C. Chung, R. M. Monteiro, C. L. Mummery, G. I. Fishman, A. Cogen, S. Evans, Isl1 Cre reveals a common Bmp pathway in heart and limb development. *Development*. **133**, 1575–1585 (2006).
76. A. E. Lewis, H. N. Vasudevan, A. K. O'Neill, P. Soriano, J. O. Bush, The widely used Wnt1-Cre transgene causes developmental phenotypes by ectopic activation of Wnt signaling. *Dev. Biol.* **379**, 229–234 (2013).
77. V. Hamburger, H. L. Hamilton, A series of normal stages in the development of the chick embryo. *Dev. Dyn.* **195**, 231–272 (1992).
78. H. Chang, Y. Wang, H. Wu, J. Nathans, Flat mount imaging of mouse skin and its application to the analysis of hair follicle patterning and sensory axon morphology. *J. Vis. Exp.* (2014), doi:10.3791/51749.
79. T. Smith, A. Heger, I. Sudbery, UMI-tools: modeling sequencing errors in Unique Molecular Identifiers to improve quantification accuracy. *Genome Res.* **27**, 491–499 (2017).
80. A. Dobin, C. A. Davis, F. Schlesinger, J. Drenkow, C. Zaleski, S. Jha, P. Batut, M. Chaisson, T. R. Gingeras, STAR: ultrafast universal RNA-seq aligner. *Bioinformatics*. **29**, 15–21 (2013).
81. S. Anders, P. T. Pyl, W. Huber, HTSeq--a Python framework to work with high-throughput sequencing data. *Bioinformatics.* **31**, 166–169 (2015).
82. C. W. Law, Y. Chen, W. Shi, G. K. Smyth, voom: Precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol.* **15** (2014), doi:10.1186/GB-2014-15-2-R29.

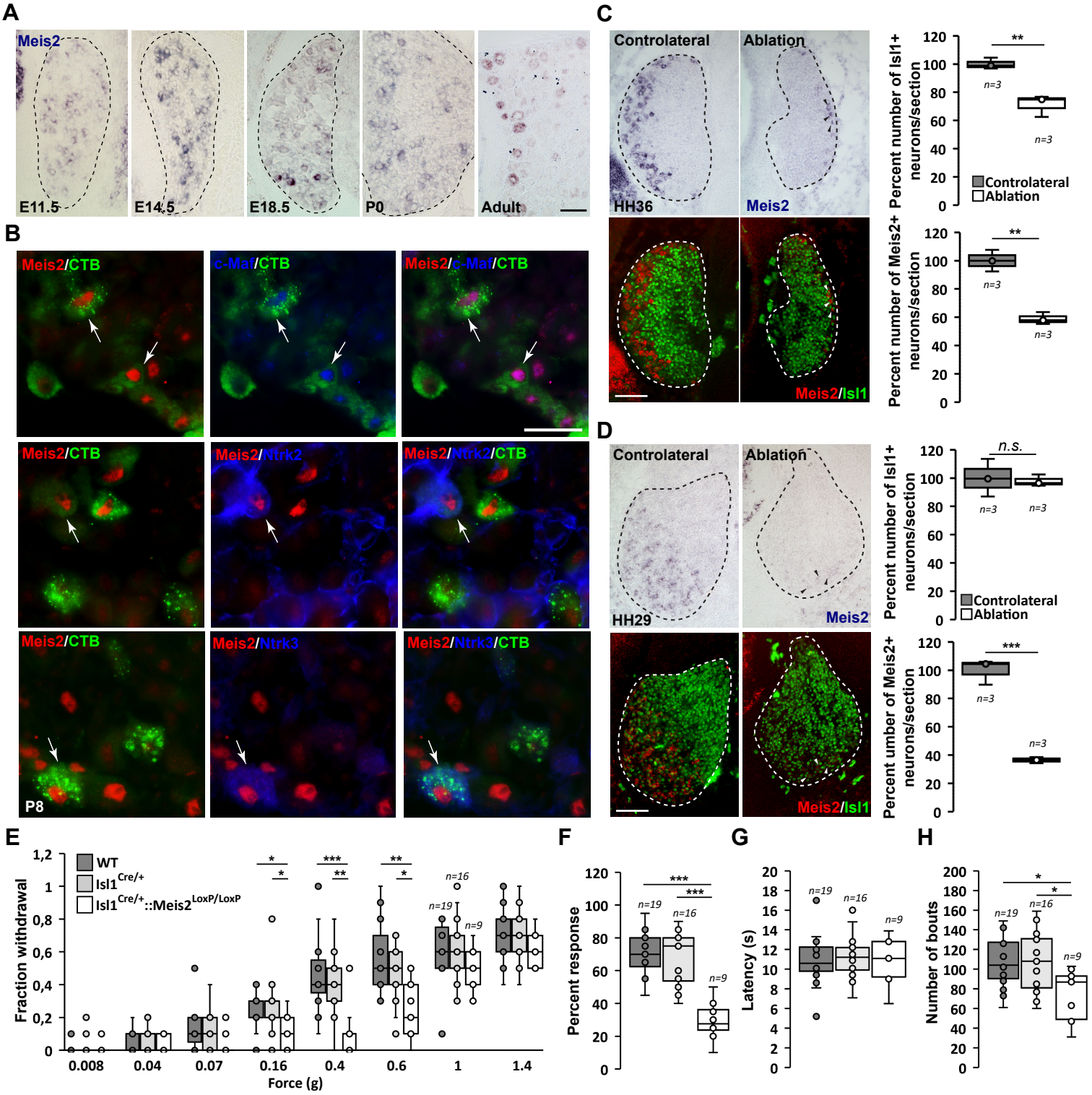
A
Meis2
E11.5
E14.5
E18.5
P0
Adult
B
Meis2/CTB
c-Maf/CTB
Meis2/c-Maf/CTB
Meis2/Ntrk2
Meis2/Ntrk2/CTB
Meis2/Ntrk3
Meis2/Ntrk3/CTB
P8
C
Controlateral
Ablation
HH36
Meis2
Meis2/Isl1
Percent number of Isl1+ neurons/section
Percent number of Meis2+ neurons/section
Controlateral
Ablation
n=3
D
HH29
n.s.
Percent umber of Meis2+ neurons/section
E
WT
Isl1Cre/+
Isl1Cre/+::Meis2LoxP/LoxP
Fraction withdrawal
Force (g)
0.008
0.04
0.07
0.16
0.4
0.6
1
1.4
n=19
n=16
n=9
F
Percent response
G
Latency (s)
H
Number of bouts

Figure 1

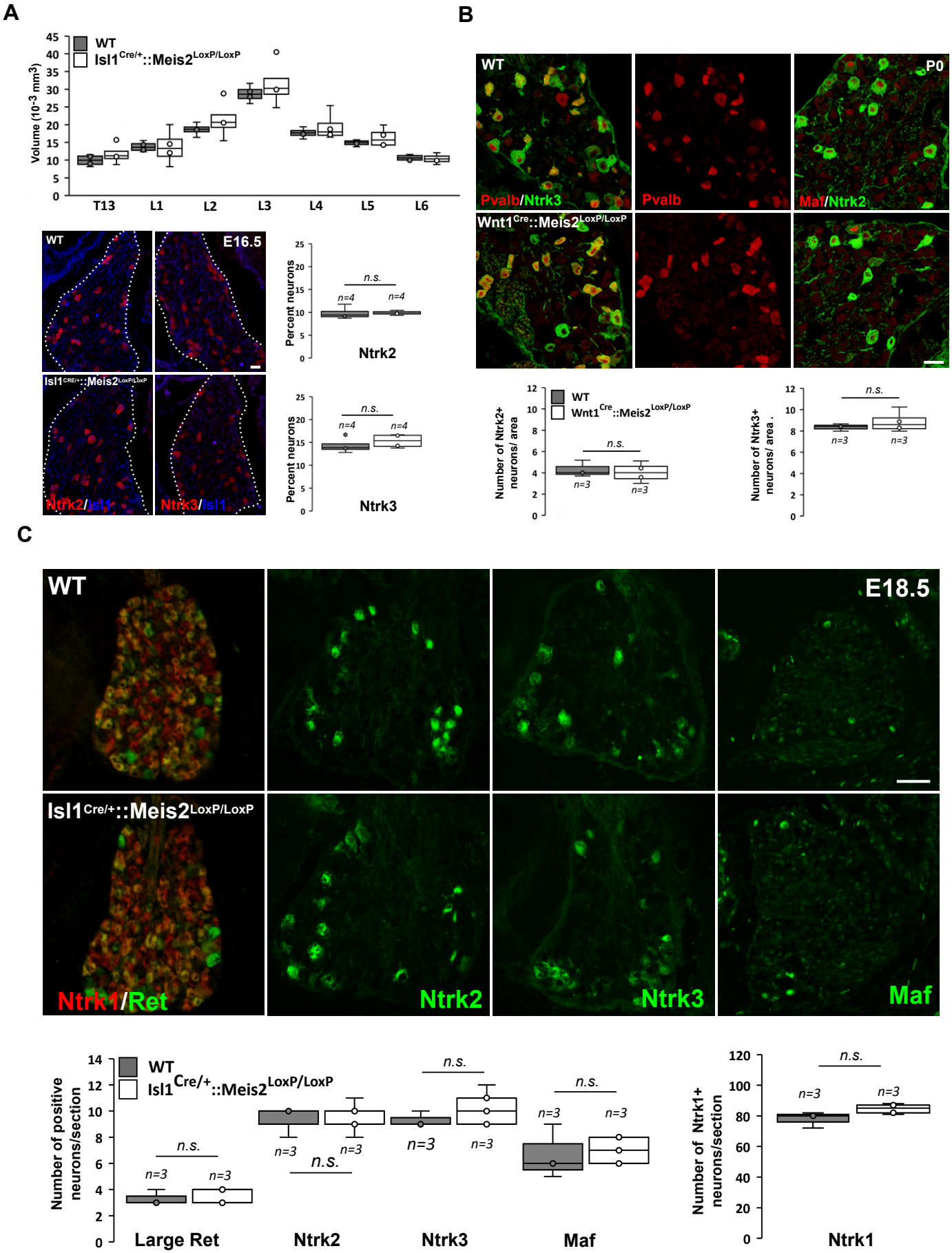
A
WT
Isl1Cre/+::Meis2LoxP/LoxP
Volume (10-3 mm3)
T13
L1
L2
L3
L4
L5
L6
WT
E16.5
Isl1CRE/+::Meis2LoxP/LoxP
Ntrk2/Isl1
Ntrk3/Isl1
Percent neurons
n.s.
n=4
Ntrk2
Ntrk3
B
WT
P0
Pvalb/Ntrk3
Pvalb
Maf/Ntrk2
Wnt1Cre::Meis2LoxP/LoxP
Number of Ntrk2+ neurons/ area
Number of Ntrk3+ neurons/ area .
n=3
C
WT
E18.5
Isl1Cre/+::Meis2LoxP/LoxP
Ntrk1/Ret
Ntrk2
Ntrk3
Maf
Number of positive neurons/section
Large Ret
Ntrk2
Ntrk3
Maf
Number of Ntrk1+ neurons/section
Ntrk1

Figure 2

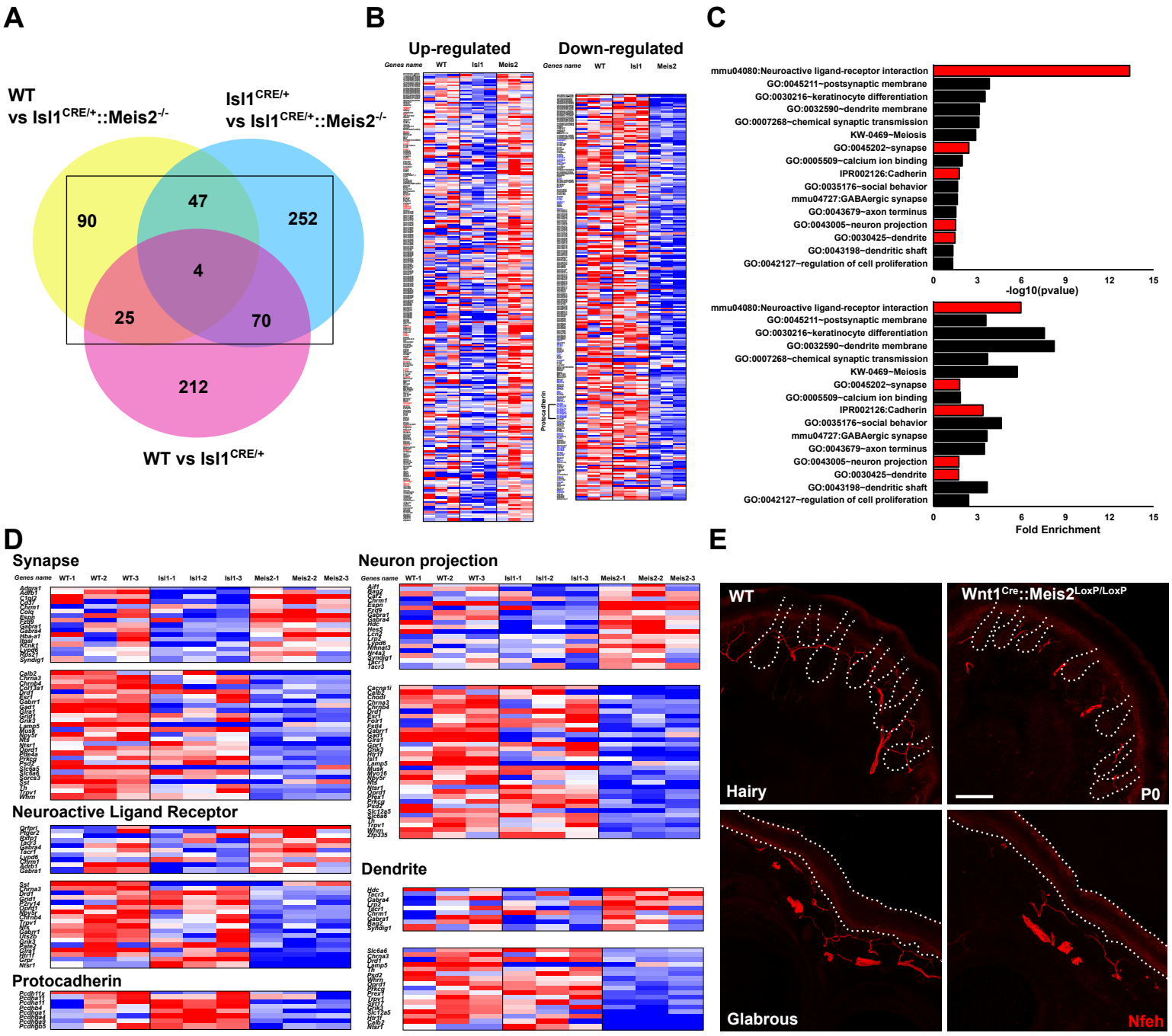
A
WT vs Isl1CRE/+::Meis2-/-
Isl1CRE/+ vs Isl1CRE/+::Meis2-/-
90
47
252
4
25
70
212
WT vs Isl1CRE/+
B
Up-regulated
Down-regulated
Genes name
WT
Isl1
Meis2
Protocadherin
C
mmu04080:Neuroactive ligand-receptor interaction
GO:0045211~postsynaptic membrane
GO:0030216~keratinocyte differentiation
GO:0032590~dendrite membrane
GO:0007268~chemical synaptic transmission
KW-0469~Meiosis
GO:0045202~synapse
GO:0005509~calcium ion binding
IPR002126:Cadherin
GO:0035176~social behavior
mmu04727:GABAergic synapse
GO:0043679~axon terminus
GO:0043005~neuron projection
GO:0030425~dendrite
GO:0043198~dendritic shaft
GO:0042127~regulation of cell proliferation
0 3 6 9 12 15
-log10(pvalue)
Fold Enrichment
D
Synapse
Neuroactive Ligand Receptor
Protocadherin
Neuron projection
Dendrite
WT-1 WT-2 WT-3 Isl1-1 Isl1-2 Isl1-3 Meis2-1 Meis2-2 Meis2-3
E
WT
Wnt1Cre::Meis2LoxP/LoxP
Hairy
P0
Glabrous
Nfeh

Figure 3

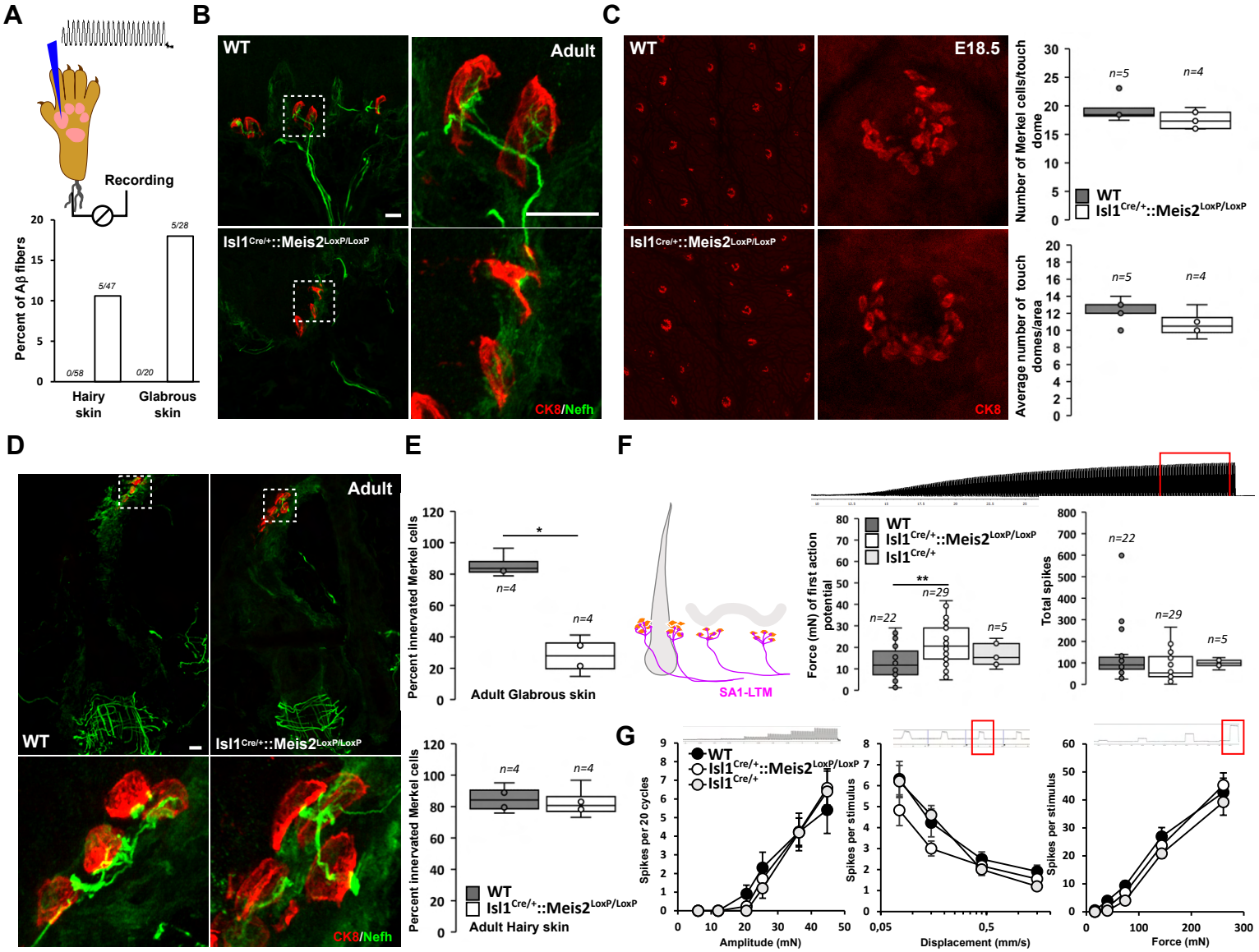
A
Recording
Percent of Aβ fibers
5/47
5/28
0/58
0/20
Hairy skin
Glabrous skin
B
WT
Adult
Isl1Cre/+::Meis2LoxP/LoxP
CK8/Nefh
C
WT
E18.5
Isl1Cre/+::Meis2LoxP/LoxP
CK8
Number of Merkel cells/touch dome
Average number of touch domes/area
n=5
n=4
WT
Isl1Cre/+::Meis2LoxP/LoxP
D
WT
Adult
Isl1Cre/+::Meis2LoxP/LoxP
CK8/Nefh
E
Percent innervated Merkel cells
Adult Glabrous skin
Adult Hairy skin
SA1-LTM
F
Force (mN) of first action potential
Total spikes
n=22
n=29
n=5
WT
Isl1Cre/+::Meis2LoxP/LoxP
Isl1Cre/+
G
Spikes per 20 cycles
Spikes per stimulus
Amplitude (mN)
Displacement (mm/s)
Force (mN)

Figure 4

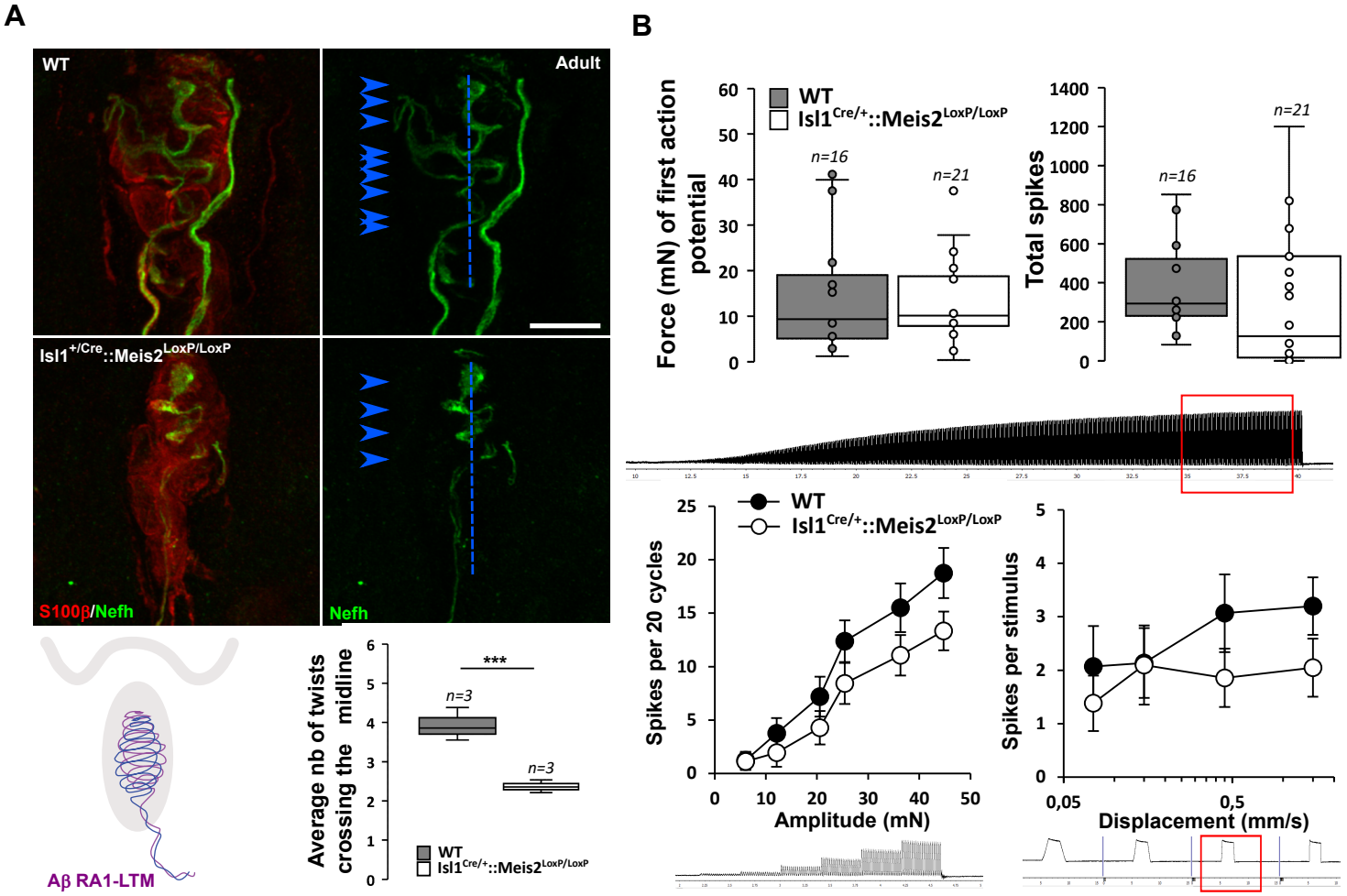
A
WT
Adult
Isl1+/Cre::Meis2LoxP/LoxP
S100β/Nefh
Nefh
Aβ RA1-LTM
Average nb of twists crossing the midline
***
n=3
n=3
WT
Isl1Cre/+::Meis2LoxP/LoxP
B
Force (mN) of first action potential
WT
Isl1Cre/+::Meis2LoxP/LoxP
n=16
n=21
Total spikes
n=16
n=21
Spikes per 20 cycles
Amplitude (mN)
Spikes per stimulus
Displacement (mm/s)

Figure 5

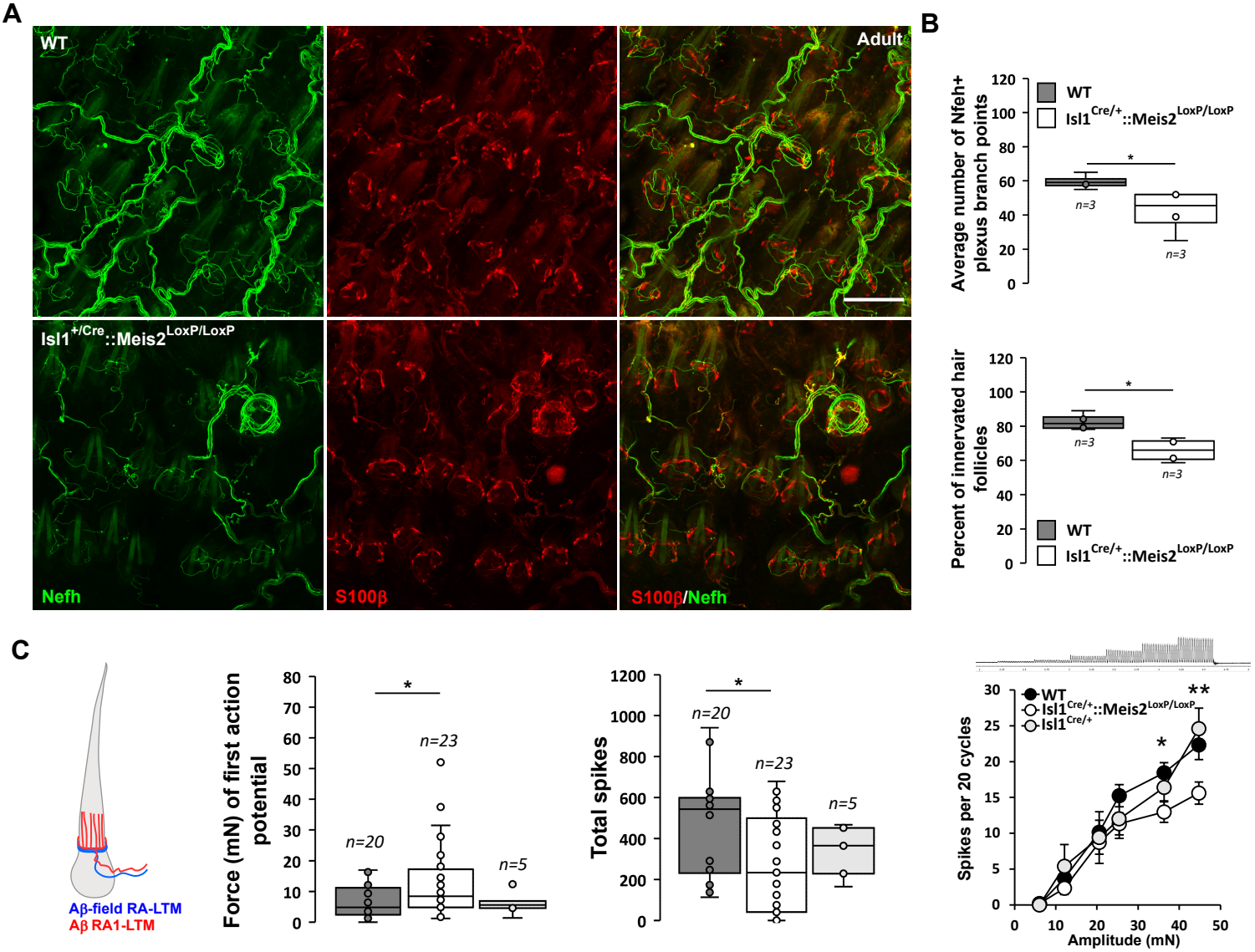
A
WT
Adult
Isl1+/Cre::Meis2LoxP/LoxP
Nefh
S100β
S100β/Nefh
B
Average number of Nfeh+ plexus branch points
WT
Isl1Cre/+::Meis2LoxP/LoxP
n=3
n=3
Percent of innervated hair follicles
n=3
n=3
C
Aβ-field RA-LTM
Aβ RA1-LTM
Force (mN) of first action potential
n=20
n=23
n=5
Total spikes
n=20
n=23
n=5
Spikes per 20 cycles
Amplitude (mN)
WT
Isl1Cre/+::Meis2LoxP/LoxP
Isl1Cre/+

Figure 6