

Comprehensive analysis of gene regulatory dynamics, fitness landscape, and population evolution during sexual reproduction

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Abstract

The fitness landscape is a critical concept in evolutionary biology and genetics that depicts fitness in the genotype space and visualizes the 2 relationship between genotype and fitness. However, the fitness landscape is challenging to characterize because the quantitative relationships 3 between genotype and phenotype and their association to fitness has not been comprehensively well described. To address this challenge, we adopted gene regulatory networks to determine gene expression dynamics. We analyzed how phenotype and fitness are shaped by the genotype in two-gene networks. A two-by-two matrix provided the two-gene regulatory network in which a vector with two angle values (Θ) was introduced to characterize the genotype. Mapping from this angle vector to phenotypes allowed for the classification of steady-state expression patterns of genes into seven types. We then studied all possible fitness functions given by the Boolean output from the on/off expression of the 8 two genes. The possible fitness landscapes were obtained as a function of the genetic parameters Θ . Finally, the evolution of the population 9 distribution under sexual reproduction was investigated in the obtained landscape. We found that the distribution was restricted to a convex 10 region within the landscape, resulting in the branching of population distribution, including the speciation process.

Keywords: fitness landscape; gene regulatory network; sexual reproduction 12

Introduction

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he fitness landscape in evolutionary biology describes fitness as the height of the genotype space and intuitively visu-3 alizes the relationship between the genotype and fitness. The 4 fitness landscape provides a significantly simplified picture of 5 evolutionary biology and genetics and is relevant to study evolv-6 ability, evolutionary pathways, the effects of multiple mutations, 7 and speciation. 8

However, it is difficult to obtain suitable fitness landscapes. 9 In a fitness landscape, the genotype is represented by a set of 10 parameters that can be mapped onto the fitness. According 11 to the landscape, how fitness changes depends on the genetic 12 parameters that are prescribed. However, there are two funda-13 mental problems with this approach. It is necessary to determine 14 the genetic parameters that describe landscape axes. Genetic 15 information can be written as the DNA sequence and how con-16 tinuous parameters are derived from this genetic sequence is 17 not trivial because a slight change in the sequence could sig-18 nificantly change the phenotype. Therefore, the derivation of 19 continuous and measurable genetic parameters that create the fit-20 ness landscape must be addressed. Second, fitness is a function 21 of phenotype and is not computed from the genotype directly. 22 Usually, the expression of RNAs and proteins is a complex and 23 24 dynamic process that determines a resultant phenotype. In other words, the relationship between genotype and phenotype de-25 pends on complex gene expression dynamics. Fitness, then, is a 26

function of dynamically regulated phenotype.

We adopted gene regulatory networks (GRN) that describe 28 gene expression dynamics to address these questions about the 29 fitness landscape. First, we showed that the degree of activa-30 tion or inhibition in expression dynamics continuously defines 31 genotype parameters. Second, phenotypes, that is, protein ex-32 pression levels, are determined by gene expression dynamics; 33 whereas, genomes provide the gene regulatory network. Thus, a 34 complex genotype-phenotype relationship was obtained. Then, 35 fitness was defined by the phenotypes, and thus, represented 36 as a function of the introduced genetic parameters. In fact, the 37 evolution of GRN has been studied(Glass and Kauffman (1973); 38 Mjolsness et al. (1991); Salazar-Ciudad et al. (2001, 2000); Kaneko 39 (2006)) extensively with respect to the robustness or the phe-40 notypic plasticity(Martin and Wagner (2009); Wagner (2013); 41 Azevedo et al. (2006); Glass and Kauffman (1973); Mjolsness et al. 42 (1991); Salazar-Ciudad et al. (2001, 2000); Kaneko (2006); Okubo 43 and Kaneko (2021a,b); Kaneko (2007); Swain et al. (2002); Ou 44 et al. (2008); Furusawa et al. (2005); Ayroles et al. (2015); Cubillos 45 et al. (2014); Chapal et al. (2019); Miller et al. (2015); Kaneko and 46 Kikuchi (2020); Nagata and Kikuchi (2020)). 47

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While there are extensive studies of evolution in the fitness 48 landscape(Soyer and Bonhoeffer (2006); Neyfakh et al. (2006); Ho 49 and Zhang (2016); Orlenko et al. (2016); Yubero et al. (2017); Fried-50 lander et al. (2017); Cuypers et al. (2017); Orlenko et al. (2017); 51 Schiffman and Ralph (2022); Hether and Hohenlohe (2014)), 52 the relationship between the global fitness landscape and GRN 53

2 Comprehensive analysis of gene networks

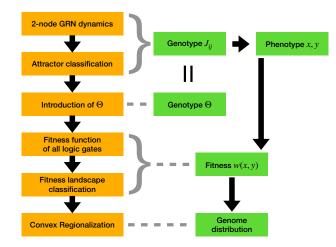


Figure 1 A flow chart in this paper to obtain the genotype– phenotype relationship and fitness landscape.

structures remains uncharacterized. In particular, Heather and
Hohenlohe (Hether and Hohenlohe (2014)) classified GRN dynamics into six cases. However, the global change in gene expression dynamics due to GRN changes and the classification
of the fitness landscape need to be further investigated by comprehensively considering all classes of possible GRNs and their
relationship to phenotypes and fitness.

Generally, GRNs and their dynamic interactions with many 8 regulatory genes are too complex to analyze. Here, we consider 9 a GRN with two genes, which provides a straightforward and 10 basic system to study comprehensively the dynamics that cre-11 ate a genotype-phenotype relationship. Here, two-node GRNs 12 are represented by 2×2 matrices. By using on/off expression 13 dynamics, we could evaluate all the possible GRN structures by 14 introducing two-dimensional parameters specifying the matrix. 15 16 Still, there can be many types of fitness functions for given expressions; in most theoretical studies, however, a specific fit-17 ness function is selected depending on the purpose of the study. 18 Here, we studied all possible fitness functions that depended 19 on the expression of the two genes (i.e., Boolean functions). The 20 possible fitness functions were limited to 16 types, all of which 21 were investigated to obtain the possible fitness landscape. Once 22 the fitness landscape was obtained, the genome distribution 23 of the GRN parameter was computed. The distribution was 24 concentrated on a higher-fitness region; however, the robust-25 ness against recombination or sexual reproduction (Martin and 26 Wagner (2009); Azevedo et al. (2006); Kim and Fernandes (2009); 27 Okubo and Kaneko (2021a); Omholt et al. (2000)) will impose 28 further restrictions on the genome distribution, as offspring from 29 the parent selected from the high-fitness region could fall into 30 a non-fitted region. Hence, it was crucial to determine how 31 stability against recombination shaped the genome distribution 32 depending on the fitness landscape. 33

This paper consists of two parts. The first part analyzes the dynamics of the two-gene network and the second part discusses the convex regionalization of the population by sexual reproduction based on the fitness landscape. (See Fig.1 for the flow used in this study).

In the first part, the phenotype and fitness are obtained from
 the genotype in a two-gene network. Because the two-gene
 network has only a few possible steady states, we classified these

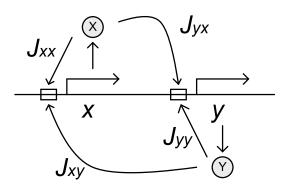


Figure 2 Regulatory network of two genes. The arrows represent the transcribed regions and the boxes represent the promoter regions. The mRNA is transcribed from each of the X- and Y-transcribed regions, from which proteins X and Y are synthesized. These proteins bind to the promoter regions of the two genes and regulate their transcription. The translation is assumed to occur very quickly and is therefore, omitted from this figure. Because proteins X and Y regulate genes *x* and *y*, there are four interactions. The magnitudes of these interactions are indicated by J_{xx} , J_{xy} , J_{yy} , and J_{yy}

steady states by introducing the angle vector $\Theta = (\theta_x, \theta_y)$ from the 2×2 gene regulation matrix. Θ provides the characteristic parameters of the genotype. Next, we considered fitness as a function of the on/off expression patterns to describe the fitness landscape as a function of Θ .

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The second part discusses the evolution of population distribution by sexual reproduction within all possible fitness landscapes. In particular, when the high-fitness region consisted of two disjointed parts, we found that the speciation of the two groups occurred by sexual recombination, whereas convex regionalization from the non-concave fitted region was also demonstrated. Finally, we discuss the relationship between the fitness landscape and the convex regionalization of epistasis, sexual reproduction, and speciation.

Dynamics of the two-gene regulatory network and fitness landscape

Two-gene regulatory network model

Our two-gene regulatory networks model assumed that there 59 were two genes *X* and *Y*. Let *x* and *y* be the concentrations of 60 proteins transcribed and translated from each gene, respectively 61 Fig.2. Proteins X and Y are produced from the transcription of 62 genes X and Y, respectively, and bind to the promoter regions of 63 X and Y (represented by the squares on the line in Fig.2. When 64 proteins X and Y bind to a gene promoter, they either promote or 65 inhibit transcription of that gene. When protein X regulates the 66 transcription of gene X, the degree of regulation is represented 67 by J_{xx} . Similarly, the regulation of gene X by Y is represented by 68 J_{xy} , regulation of gene Y by X by J_{yx} , and regulation of gene Y 69 by *Y* by J_{yy} . These are collectively denoted by J_{ij} . The value of J_{ij} 70 can take any real value; thus, when J_{ij} is positive, transcription 71

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is promoted, and when J_{ij} is negative, it is inhibited. 1

When a protein binds to a promoter, gene transcription is 2 promoted or inhibited. For simplicity, we assumed that a gene is 3 transcribed when the sum of the effects of molecular regulation exceeds a threshold value. The activation function was given by 5 the sigmoid function $f[x] = \frac{1}{1 + \exp[-\beta x]}$. The degree by which 6 gene *m* expression is regulated by protein N transcribed by gene 7 are reported by n as J_{mn} , with the dynamics of the expression 8 level and protein concentration of X and Y represented by 9

$$\dot{x} = f[J_{xx}(x - \xi) + J_{xy}(y - \xi)] - x$$
(1)

$$\dot{y} = f[J_{yx}(x-\xi) + J_{yy}(y-\xi)] - y$$
 (2)

-x and -y on the right-hand side of the second term represent 10 protein X and Y degradation, respectively. Here, the expres-11 sion threshold was set at 0.5, to make the expression and non-12 expression states symmetric for later fitness function simplicity. 13 β of the sigmoid function was set to 100 to make the Hill function 14 $(f[x] = \frac{x^n}{K^n + x^n})$ sharp. 15

Attractors of gene-expression dynamics 16

Considering the relationship between genotype and phenotype, 17 we focused on where the expression levels *x* and *y* converged 18 to a fixed point or cycle. We investigated how phenotype (x, y)19 was determined depending on the genotype, $J_{ij}(i, j = x, y)$. 20

To investigate the fixed point, we first obtained the nullclines 21 of *x* and *y*, respectively, which were curves that satisfied $\dot{x} = 0$ or 22 $\dot{y} = 0$ in Eq.(2). The nullclines were represented by the equation: 23

$$x = f[J_{xx}(x - \xi) + J_{xy}(y - \xi)]$$
(3)

$$y = f[J_{yx}(x - \xi) + J_{yy}(y - \xi)],$$
 (4)

which are illustrated in Fig.3. Since $\beta >> 1$, f[x] approaches 24 a step function, so that x = f[x] is satisfied either at $x \approx 1$ or 25 $x \approx 0$, as well as $x \approx \zeta = 0.5$. In this paper, for simplicity, these 26 $x \approx 1$ and $x \approx 0$ states are written as just x = 1 and x = 027 considering $\beta >> 1$. 28

Classification of two-gene expression dynamics by at-29 tractors 30

For given gene regulatory networks, phenotypes determined by 31 attractors in gene expression dynamics can be organized into 32 seven classes (Fig.4). The attractor types, their numbers, and 33 their configurations are classified based on the value of J_{ij} as 34 follows: 35

36	1. Equal expression level of x, y (S, symmetric or synchro-
37	nized)
38	x = 1, y = 1, or $x = 0, y = 0$ is the fixed-point attractor
39	depending on the initial conditions. We referred to this as

- dynamics-type-S. 40
- Different expression level of x, y (A, antagonistic) 41

x = 1, y = 0 or x = 1, y = 0 is the fixed-point attractor 42 depending on the initial conditions. We referred to this 43 dynamic as type-D. 44

3. Same or different expression level of x, y (Q, quad) 45

All possible four cases with x = 0 or 1, y = 0 or 1 give 46

- the fixed-point attractor depending on the initial conditions 47
- and were referred to as dynamics-type-Q. 48

- 4. Intermediate expression level of *x* (Cx, continuous for *x*) $x = \alpha, y = 0$ or $x = \alpha, y = 1$ is the fixed-point attractor, 50 where $0 < \alpha < 1$, depending on the initial conditions and 51 were defined as dynamics-type-Cx. 52
- 5. Intermediate expression level of *x* (Cy, intermediate for *y*) $x = 0, y = \alpha$ or $x = 1, y = \alpha$ is the fixed-point attractor, where $0 < \alpha < 1$, depending on the initial conditions. We defined this a dynamics-type-Cy.
- 6. Half expression level for *x*, *y* (H, half) x = 0.5 and y = 0.5 is the fixed-point attractors that were observed all initial conditions were defined as dynamicstype-H.
- 7. Periodic expression *x*, *y* (P, periodic) When the limit cycle was obtained in all initial conditions, we referred to this as dynamics-type-P.

Introduction of Θ

We were able to classify steady states that corresponded to phe-65 notypes and introduced a vector that characterized the genotype. 66 Originally, the genotype in the model was represented by a J_{ii} in 67 the 2×2 real matrix. However, the GRN that adopted a sigmoid 68 function only uses two states, 0, 1, beside 0.5, which reduced the 69 dimension of the genotype. This two-dimensional parameter 70 represented the genotype's declination angle, Θ . 71

We then focused on the shape of the nullcline in Fig.3. The 72 shape was determined by the direction of the line segment that 73 passed through point (0.5, 0.5), which determined the attractor. 74 Therefore, the change in the dynamics were due to the change 75 in J_{ii} and could be specified in the direction of normal vectors 76 of the J_{ij} row vectors. When examining the direction of this line 77 segment, one of the normal vectors of this line segment (Fig.3) 78 pointed in the same direction as the row vector of J_{ij} , that is, 79 (J_{xx}, J_{xy}) or (J_{yx}, J_{yy}) . Here, only the direction of the row vectors 80 mattered because the magnitude of the vectors was related to 81 the shape of the nullcline. Hence, the shape of the nullcline was 82 described by the angles of the $J_{ii}(\Theta)$ row vectors. In other words, 83 as the angle between a line extended from the origin and the 84 row vector, (J_{xx}, J_{xy}) or (J_{yx}, J_{yy}) , we could take advantage of 85 the symmetry between *x* and *y* to define the counterclockwise 86 angle between the positive part of the x-axis and (J_{xx}, J_{xy}) as θ_x , and the counterclockwise angle between the positive part of the 88 y-axis and (J_{yx}, J_{yy}) as θ_y . They were defined by 89

$$\theta_x = \arctan[\frac{J_{xy}}{J_{xx}}] \tag{5}$$

$$\theta_y = \arctan[\frac{-J_{yx}}{J_{yy}}].$$
(6)

The dynamics and attractors were determined by using $\Theta =$ (θ_x, θ_y)

Θ torus and attractor classification

Using θ_x and θ_y , we could explicitly classify the attractors of the 93 teo-gene regulatory dynamics (Fig.4). This section introduces 94 the θ torus, which had a finite range of θ_x and θ_y and specified 95 each attractor type. 96

First, the behavior of the gene expression dynamics was de-97 termined by the direction of the J_{ij} , Θ row vector. Here, the 98 phenotype was visualized by introducing the Θ torus with θ_x 99 on the horizontal axis and θ_y on the vertical axis, where θ_x and 100

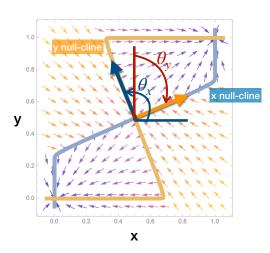


Figure 3 Nullcline normal vector. First, the nullclines were obtained from $\dot{x} = \dot{y} = 0$ in Eq.(1) and Eq.(2). The blue line represents the nullcline for x and orange that for y. The intersection of these two null lines was a fixed point. The shape of the nullcline was characterized by normal vectors that indicate the directions of the nullclines around (0.5,0.5) and are denoted as (J_{xx}, J_{xy}) or (J_{yx}, J_{yy}) . Furthermore, the vector field around the fixed point assisted in determining whether the fixed point was a stable fixed point (i.e., when the vector field around the fixed point was directed toward the fixed point from any direction) or an unstable fixed point (i.e., when the vector field around the fixed point is directed away from it). In this figure, (0,0), (0.5,0.5), and (1,1) are nullcline intersections that are fixed points, whereas only (0,0) and (1,1) are stable fixed point.

 θ_{ν} were cyclic with mod 2π , as in Fig.4. Therefore, Θ torus rep-1 resented the attractor types. Each of the types (S, D, Q, Cy, Cx, 2 H, and P) was characterized by the nullclines shown in Fig.4A. 3 Note that Θ determined the directions in the nullcline near point 4 (0.5, 0.5). The attractor determined by the two nullclines could 5 change depending on the relative positions of θ_x and θ_y . Then, 6 S, D, Q, Cy, Cx, H, and P were classified depending on (θ_x, θ_y) 8 as presented in Fig.4B. For the derivation of the boundaries for 9 each classification, see Supplementary Material S1. This diagram shows the relationship from Θ to the attractors in (*x*, *y*) and the 10 one between genotype and phenotype. 11

12 Defining the fitness function

We obtained the fitness landscape for Θ torus by first defining the 13 fitness. Here, the phenotype was determined by the stationary 14 15 expression levels of x and y. Here, we consider the case that the fitness takas maximal either at each expression level 0 or 16 1 (i.e., the fitness depends monotonically on the combination 17 of each expression level, so that the fitness takes maximum at 18 0 or 1). Hence we focused on the states with values of 0 or 19 1; therefore, the fitness was described by a combination of the 20 four-state input of (x, y) = (0, 0), (0, 1), (1, 0), (1, 1) and their 21 two-state output. We assumed that binary fitness (fitted or non-22 fitted) was based on the binary phenotype. Hence, the fitness 23 function was described by a Boolean (logical) function with two 24 binary inputs and one binary output. In this case, $2^4 = 16$ 25 possible fitness degree functions existed, which were reduced 26 to five fitness functions using the symmetry of the model and 27

identifying logical operations. (See Supplementary Material S2).

As for the fitness function w(x, y), we introduced a continuous function to satisfy the Boolean function such that w(x, y) for x, y = (0, 1) was either 0 or 1, and determined that the simplest form for w(x, y) had intermediate values between 0 and 1. We characterized four typical fitness functions here. Other cases including NEUTRAL are described in Supplementary Material S3.

AND: requires expression of both genes This function used a maximum value of 1 when both *x* and *y* were set to 1.

$$w(x,y) = xy \tag{7}$$

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This corresponded to the case where x and y expression was required for survival (e.g., the formation of complexes by the proteins X and Y).

X ONLY: requires only X, whereas Y expression is neutral In this function, fitness referred only to x and thus, if x was 1, fitness returned a maximum value of 1.

$$w(x,y) = x \tag{8}$$

This corresponded to the selection pressure affecting only the expression level of *x* (e.g., protein X is functionally dominant or protein Y had a neutral function). When the fitness depended only on the expression of *y*, it presented similarly to that of w(x,y) = y.

XOR: requires the expression of only X **or** Y When (x, y) was (1,0) or (0,1), the fitness had a maximum value of 1. By contrast, when (x, y) was (0,0) or (1,1), the fitness had a minimum value of 0. As the simplest form,

$$w(x,y) = |x-y|.$$
 (9)

This function required that the expression of only one gene, x or y, was necessary for survival, but fitness was lost if both genes were expressed (e.g., switching the two pathways by the inputs). ⁴⁴

OR: requires the expression of either one of the two genes If at least one of *x*, *y* is 1, then the fitness had a maximum value of 1.

$$w(x,y) = 1 - (1 - x)(1 - y) = x + y - xy$$
(10)

This corresponded to the case in which the expression of x or y 47 was needed for survival (e.g., both X and Y proteins have similar 48 functions). 49

Classification of the fitness landscape

We next explored the fitness landscape of genotype spaces that 51 depended on the fitness function represented by Θ torus. Here, 52 for a given Θ , we obtained the attractor of (x, y) for a fixed initial 53 expression $(x_0, y_0) = (0.05, 0.5) ((x_0, y_0) = (0.18, 0.18)$ for the 54 XOR. Depending on the fitness function, we defined (AND, X 55 ONLY, XOR, *etc.*) to obtain the fitness landscape for Θ , as shown 56 in the diagram presented in Fig.4B, which contains information 57 about the complex genotype-phenotype relationship. 58

We defined four typical landscapes (ONE RECTANGLE, 59 TWO RECTANGLES, L-SHAPE, and ONE BAND in Fig.5), as well as additional landscapes that are described in Supplementary Material S3. 62

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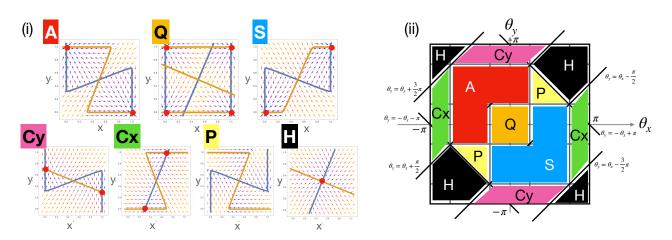


Figure 4 Fixed point classification of the Θ torus. (i)Each type of dynamics and nullclines in the two-gene regulatory network is represented. The Θ values for each type in the figure are $(-\frac{3}{8}\pi, \frac{3}{8}\pi)$ for (A), $(-\frac{1}{8}\pi, -\frac{1}{8}\pi)$ for (Q), $(\frac{3}{8}\pi, -\frac{5}{8}\pi)$ for (S), $(-\frac{3}{8}\pi, \frac{7}{8}\pi)$ for (Cy), $(\frac{7}{8}\pi, -\frac{3}{8}\pi)$ for (Cx), $(\frac{3}{8}\pi, \frac{3}{8}\pi)$ for (P), and $(\frac{7}{8}\pi, \frac{7}{8}\pi)$ for (H). (ii) Classification of each dynamic category was based on (θ_x, θ_y) . The boundaries between the categories in the figure are $\theta_y = \theta_x \pm \frac{3}{2}\pi$, $\theta_y = \theta_x \pm \frac{\pi}{2}$, $\theta_y = -\theta_x \pm \pi$, $\theta_y = \pm \frac{3}{4}\pi$, $\theta_y = \pm \frac{1}{4}\pi$, $\theta_x = \pm \frac{3}{4}\pi$, and $\theta_x = \pm \frac{1}{4}\pi$, respectively.

ONE RECTANGLE (seen in AND): Fig.5(i) presents an example of the "rectangle" landscape, where the region of maximum fitness is in a single rectangle. This landscape is mainly observed in the AND fitness function. The high-fitness region occurs only in the parts of classes S and Q of the AND fitness because the optimal fitness was achieved for only (x, y) = (1, 1). In the Θ space for S in Fig.5(i) the maximum fitness is determined by the initial expression (x_0, y_0) .

TWO RECTANGLES (seen in X ONLY): This fitness landscape 9 was composed of two rectangular high-fitness regions (Fig.5(ii)) 10 where x = 1 and y values were y = 1 for one region and y = 011 for the other. The fitness maximum was X ONLY when a stable 12 fixed point satisfied x = 1. The dynamics that satisfy this con-13 dition were either S or D and the Θ space for S and A in Fig.5(i) 14 reached the maximum fitness, which was determined by the 15 initial expression $(x_0, y_0) = (0.05, 0.5)$. Q achieved maximum 16 fitness, but all maximum fitness regions were connected. 17

L-SHAPE (seen in XOR or OR): The region of maximum fitness 18 was distributed in an L-shape, as shown in Fig.5(iii) with the 19 XOR fitness function. The shape of this landscape was an in-20 verted L, which we collectively referred to as an L shape. This 21 landscape was obtained when the fitness function was either an 22 XOR or an OR. When using the XOR or OR fitness functions, the 23 initial condition (x_0, y_0) existed in $0 \le x_0 \le 1$ and $0 \le y_0 \le 1$, 24 except for the unstable fixed-point singularity. We found that 25 the region of L-SHAPE had the maximum fitness for all initial 26 conditions tested. Therefore, the fitness was always maximum 27 in the dynamics-type-A region because the stable fixed points of 28 dynamics-type-A were only (1,0) and (0,1), respectively. The L-29 shaped area was robust against noise in the expression or initial 30 conditions with XOR or OR fitness. 31

³² **ONE BAND (seen in OR or X ONLY):** The maximum fitness re-³³ gion was extended to the entire range of θ_x or θ_y as a band for the ³⁴ OR fitness function, as shown in Fig.5(iv). This band extended ³⁵ over the boundary of the Θ torus in a single direction and there-³⁶ fore, the expression level of one *x*, *y* was neutral in this band. In OR, fitness was maximized where x = 1 or y = 1, which was 37 when a stable fixed point was reached and the dynamic types S, 38 D, Q, Cy, and Cx were satisfied. In particular, in dynamics-type-39 D, the fitness of the OR was always maximally independent of 40 the initial condition because only the stable fixed points were 41 (1,0) and (0,1), and there were no other steady states. In S, Q, Cy, 42 and Cx, the maximum fitness regions were determined by the 43 initial expression (x_0, y_0) . Depending on the initial expression, 44 this type of landscape was also observed in the X ONLY fitness 45 landscape.

Genotype distribution is restricted by evolution with sexual reproduction

Thus far, we have investigated the shape and features of the fitness landscape by comprehensively calculating the correlations between genotype, phenotype, and fitness in a two-gene network. By introducing Θ as a genotype parameter, we were able to examine the phenotype and fitness for a given genotype and obtain the global structure of the fitness landscape. Once we have information on the global fitness landscape, we can predict the gene distribution and evolvability of the population during sexual reproduction and mutation. Thus, we investigated how the population distribution of the gene parameter Θ changed through evolution, with or without sexual reproduction.

In particular, we focused on how the population distribution was restricted to a convex set of maximum fitness regions by evolution during sexual reproduction (i.e., convex regionalization). We introduced asexual and sexual reproduction into this model and numerically evolved the population distribution in these conditions for each typical fitness landscape.

Definition of mutation and recombination

Before discussing convex regionalization, we defined the procedure for genetic evolution. We considered two inheritance modes: asexual and sexual reproduction processes. In asexual reproduction, GRN J^{new} in the next generation was chosen from a set of J^{fit} with high fitness in the population. For mutations, the network adjacency matrix for the next generation, J^{new} , was

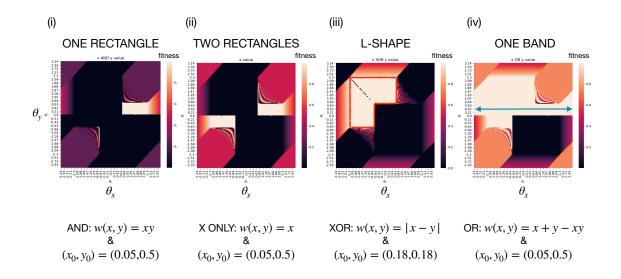


Figure 5 Four typical landscapes. The fitness value is represented by a color plotted as a function of (θ_x, θ_y) . (i) A single rectangular region has maximum fitness. (ii) The maximum fitness region was divided into two parts. (iii) Even when the initial conditions were changed, the genotypes on the red line surrounding the region achieved maximum fitness. (iv) The arrowed band extends in the horizontal direction. If θ_y is fixed in this range, θ_x will be maximally fit for any value (neutral for *x*).

changed by adding a random value generated by a normal distribution with mean 0 and variance σ . Note that the mutation was not introduced to Θ but to J^{new} . This was because Θ was an abstract characteristic value and not the actual value for simplicity. In sexual reproduction, two individuals, J^{fit1} and J^{fit2} , were selected from the set J^{fit} to generate high fitness. The GRN in the next generation was then defined by row-wise mixing of the two highly fitted individuals. Therefore,

$$\begin{pmatrix} J_{xx}^{\text{fit1}} & J_{xy}^{\text{fit1}} \\ J_{yx}^{\text{fit2}} & J_{yy}^{\text{fit2}} \end{pmatrix} \text{ or } \begin{pmatrix} J_{xx}^{\text{fit2}} & J_{xy}^{\text{fit2}} \\ J_{yx}^{\text{fit1}} & J_{yy}^{\text{fit1}} \end{pmatrix}$$

yields J^{new}. This corresponded to the recombination of the pro-1 moter or enhancer regions. The evolutionary change in regula-2 tory regions is much faster than in the gene that encodes for the 3 protein itself (Luscombe et al. (2004)). Because proteins can bind to gene promotor regions, it is appropriate to use row-by-row 5 mixing of the GRN adjacency matrix *J* as a model for sexual 6 reproduction with free recombination. In the Θ torus, sexual re-7 production could be expressed as parents with J^{fit1} : $(\theta_x^{\text{fit1}}, \theta_y^{\text{fit1}})$ 8 and J^{fit2} : $(\theta_x^{\text{fit2}}, \theta_y^{\text{fit2}})$, and with $\theta_x^{\text{fit1}} \neq \theta_x^{\text{fit2}}$ and $\theta_y^{\text{fit1}} \neq \theta_y^{\text{fit2}}$. Ad-9 ditionally, suppose a rectangle with each side parallel to the θ_x 10 or θ_{y} axis (Fig.6). By mixing the row vectors of the adjacency 11 matrices with sexual reproduction, the children from $(\theta_x^{\text{fit1}}, \theta_v^{\text{fit1}})$ 12 and $(\theta_x^{\text{fit2}}, \theta_y^{\text{fit2}})$ could be represented by $(\theta_x^{\text{fit1}}, \theta_y^{\text{fit2}})$ or $(\theta_x^{\text{fit2}}, \theta_y^{\text{fit1}})$. 13 This corresponded to the vertex of the other side of the diagonal 14 compared to the parent in the rectangle of Fig.6. Mutations in 15 sexual reproduction were introduced in the same way as asexual 16 reproduction, in which a normal distribution with mean 0 and 17 variance σ were added to J^{new} . Thus, sexual reproduction with 18 a slight mutation could create a population of genotypes on the 19 vertices of the rectangle. 20

Convex regionalization of sexually reproducing populations from a non-convex fitted area

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For each fitness landscape category, we used simulations to examine changes in the population distribution due to sexual reproduction.

Simulation method: We ran simulations for 130 generations with 26 a mutation size σ of 0.01 or 0.1, and a population size of 100. First, 27 we set the initial expression to (x_0, y_0) . The initial expression 28 (x_0, y_0) was fixed and did not change throughout the subsequent 29 simulations. Each element of J_{ij} in the 0th generation was given 30 a uniform random number in the interval [-1,+1]. The dynamics 31 of the GRN were calculated using Eq.(1) and Eq.(2) for 100 time 32 steps. The average of each *x*, *y* in the last ten steps was used 33 as the phenotype (expression). Fitness was calculated from the 34 phenotype using the function defined in the previous section. 35

TWO RECTANGLES fitness landscape: The difference in the 36 distribution between the asexually and sexually reproducing 37 populations in the simulation is shown in Fig.7. During asex-38 ual reproduction, the population remained distributed in both 39 maximum-fitness regions. In contrast, the evolution during sex-40 ual reproduction was concentrated only on one of the two fitted 41 rectangles. These results can be explained as follows. The TWO 42 RECTANGLES fitness landscape had two regions with equal 43 maximum fitness values. If a population was distributed across 44 both, the offspring from the parent of the two rectangles fall onto 45 the non-fitted region and the sexually reproducing population 46 concentrated in one of the two regions was selected. Thus, the 47 population distribution was in a convex region and sustained. 48 In contrast, during asexual reproduction, the population was 49 distributed across the two regions. Therefore, evolution of sex-50 ual reproduction could induce speciation. Here, the offspring 51 from the two regions were less fit, resulting in hybrid sterility. 52

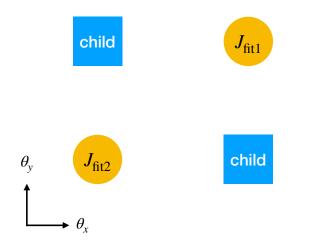


Figure 6 Transfer of genetic parameters (θ_x, θ_y) to children through recombination. Sexual reproduction involves mixing the row vectors of the adjacency matrix. The vector of children from the parents $(\theta_x^{fit1}, \theta_y^{fit1})$ and $(\theta_x^{fit2}, \theta_y^{fit2})$ were randomly chosen from θ_x^{fit1} or θ_x^{fit1} , and θ_y^{fit1} or θ_y^{fit1} . Graphically, two vertices can fall to a child on another diagonal from the parent rectangular axis.

Following the definition of speciation by hybrid sterility, it can
 be concluded that speciation occurred in this case.

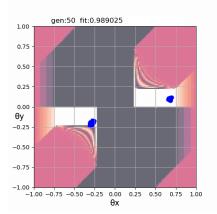
L-SHAPE fitness landscape: The difference in population dis-3 tribution between asexual and sexual reproduction was also ob-4 served in the L-SHAPE fitness landscape, as shown in Fig.8(iii). 5 During asexual reproduction, the genotype population was spread over the entire L-shaped area of the landscape; how-7 ever, during sexual reproduction, the population was biased 8 in one direction of the L-SHAPE. Here, the L-SHAPE fitness 9 landscape (Fig.9) extended vertically and horizontally in two 10 directions, but the offspring from the parent between these two 11 directions (IV of Fig.9) were less fit as a result of the genetic 12 change in Fig.6. 13

Hence, the offspring produced from sexual reproduction be-14 tween the two branches of the L-SHAPE landscape (II and III in 15 Fig.9) shrank into a rectangular region, either vertically (I and III 16 in Fig.9) or horizontally (I and II in Fig.9). Such convex region-17 alization of the population distribution did not occur during 18 asexual reproduction. This convex regionalization was similar 19 to speciation in the Bateson-Dobzhansky-Muller model (Bateson 20 (1909); Dobzhansky (1936, 1937); Muller (1940, 1942)), which is 21 supposed to be an L-shaped fitness landscape. However, we 22 found that some individuals from the common square area (I 23 in Fig.9) of the two edges maintained high fitness and were not 24 reproductively isolated, indicating that complete speciation did 25 not occur. Only some of the two rectangular regions (II and III 26 in Fig.9) were not fit, as an offspring could be located in IV in 27 Fig.9. However, this convex regionalization was achieved so that 28 the population in the L-SHAPE region was not allowed under 29 sexual reproduction. 30

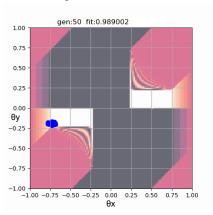
31 Discussion

32 This study analyzed the genotype–phenotype relationship across

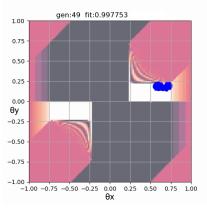
³³ all two-gene regulatory networks and obtained the fitness land-



(a) asexual reproduction

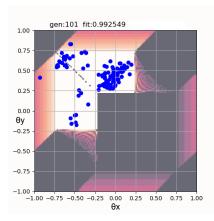


(b) sexual reproduction (right region is selected)

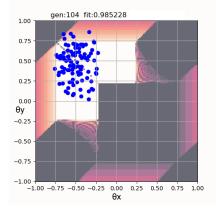


(c) sexual reproduction (right region is selected)

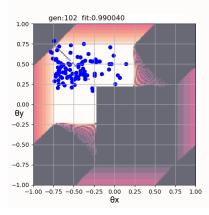
Figure 7 Comparison of the population distributions in two rectangular fitness landscapes with X ONLY(w(x, y) = x) fitness (a)during asexual reproduction (mutation only) or during sexual reproduction, where the population is branched into two cases, (b) and (c), which differ in each run of the evolution simulation. The initial expression was chosen as $(x_0, y_0) = (0.05, 0.5)$. The mutation rate was set to 0.01. "gen:" number is the generation number and "fit:" number is the average fitness of the population.



(a) asexual reproduction



(b) sexual reproduction (case of vertical convex regionalization)



(c) sexual reproduction (case of horizontal convex regionalization)

Figure 8 Comparison of the population distributions in two rectangular fitness landscapes with X ONLY(w(x, y) = x) fitness during (a) asexual reproduction (mutation only) and (b,c) sexual reproduction. The population branched into two cases, (b) and (c), which differ in each run of the evolution simulation. The initial expression was chosen as $(x_0, y_0) =$ (0.18, 0.18). The mutation rate was set at 0.1. "gen:" number is the generation number and "fit:" number is the average fitness of the population.

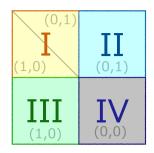


Figure 9 Partition of L-SHAPE fitness landscape. Regions I, II, and III support the highest fitness, while fitness is lost in region IV. Values with brackets like (1,0) shows the stable expression levels of *x* and *y*.

scapes according to possible Boolean fitness functions. Characteristic landscapes including TWO RECTANGLES with the two optimal regions, L-SHAPE and ONE BAND, were obtained. As a result of evolution during sexual reproduction, the population becomes restricted to the convex region, leading in speciation.

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To characterize the genetic changes in the gene regulatory matrix, a pair of continuous parameters $\Theta = (\theta_x, \theta_y)$ was introduced and the relationship between genotype and phenotype was characterized to examine how these relationships are associated with fitness and genome distribution. Several studies have discussed how the genotype-phenotype-fitness relationship affects GRN dynamics(Hether and Hohenlohe (2014); Friedlander et al. (2017); Schiffman and Ralph (2022)). However, these analyses were restricted to a specific level. For instance, genotype to phenotype in Hether and Hohenlohe (2014)), while a complete analysis of GRN dynamics has not been performed. We conducted a comprehensive analysis of the genotype-phenotypefitness-distribution relationship. Furthermore, our results improve our understanding of how sexual reproduction changes population distribution or leads to speciation-like events in the 20 fitness landscape, which requires a global landscape structure. 21

This study demonstrated that sexual reproduction limits populations only in a restricted, convex set in the maximum fitness regions (convex regionalization). The population was restricted to horizontal or vertical rectangular regions in the L-SHAPE landscape. In the TWO RECTANGLES landscape, the population was limited to one of the two separate high-fitness regions, which led to speciation. In the L-SHAPE landscape, complete speciation was not achieved, but convex regionalization led to two distinct population distributions, as in speciation.

When discussing the effects of multiple gene interactions, 31 epistasis is often adopted. Epistasis is defined as a nonlinear 32 change in fitness with multiple mutations. When the fitness 33 change is lower or higher than the addition of changes in mul-34 tiple mutations, it is called negative or positive epistasis, re-35 spectively. Epistasis is applied to local fitness changes, which 36 benefits the study of the effects of relatively small mutations. In 37 contrast, the fitness landscape provides global information on 38 the fitness. This study showed that such information is essential 39 for studying changes in population distribution, robustness of 40 sexual reproduction, and speciation. 41

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The method and results presented in this study can be used to solve other network related problems. First, the genotype 2 parameter Θ may be used for evolution in other Boolean net-3 work systems such as machine learning and social or ecological networks(Raimundo et al. (2018); Saavedra et al. (2007); Shizuka 5 and McDonald (2015); Sinha et al. (2022); Gordon (2014)). It can 6 also be applied to a system that interacts with a steep sigmoid 7 function. Here, the threshold $\zeta = 0.5$ was used for simplicity and symmetry, but even if the threshold for each genes is changed, 9 10 Θ can be used because the dynamics equivalent to this study were obtained by the transformation of variables, even though 11 the classification of dynamics was more complex. In addition, Θ 12 can be extended to systems with more than two (N) genes. In 13 this case, the Θ space had N(N-1) dimensions, which made it 14 more difficult to obtain a global fitness landscape. However, the 15 fixed points and their stability can be evaluated according to the 16 value of Θ . The method of the present study can be applied by 17 maintaining some Θ values within a certain range. For instance, 18 the present results can be extended to a system of multiple pairs 19 of corresponding genes. In general, by introducing network 20 modules (network motifs Alon (2019)), the application of this 21 method is straightforward. 22

23 In conclusion, our global analysis of GRNs based on Θ val-24 ues and the characterized fitness landscape contributes to the comprehensive understanding of GRN evolution, particularly 25 convex regionalization associated with sexual reproduction and 26 resultant speciation. 27

Acknowledgments 28

The authors would like to thank Tetsuhiro Hatakeyama, Yuichi 29

Wakamoto, Naoki Irie, Akira Sasaki, and Shuji Ishihara for 30

their stimulating discussions and Hideki Innan and Takahiro 31

Sakamoto for providing information about genetics. 32

Funding 33

This research was partially supported by a Grant-in-Aid for 34

Scientific Research (A) 431 (20H00123) and a Grant-in-Aid for 35

Scientific Research on Innovative Areas (17H06386) from the 36

Ministry of Education, Culture, Sports, Science, and Technology 37

(MEXT) of Japan. 38

Conflicts of interest 39

The authors declare that they have no conflicts of interest. 40

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