

Cold plasma treatment to inhibit *Fusarium graminearum* growth

A protocol for the use of cold plasma treatment to inhibit *in vitro* growth of *Fusarium graminearum*.

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Abstract

1 Cold plasma is an ionised gas that can be used to control a range of microorganisms. In this
2 study, a protocol was developed for cold plasma treatment of *in vitro* growth of *Fusarium*
3 *graminearum*, a global mycotoxic species generally associated with cereal grain. Four *F.*
4 *graminearum* isolates on potato dextrose agar (PDA) were treated with cold plasma for 70 s
5 from a distance of 21 cm in a closed environment, and their radial colony growth was measured.
6 To consider whether cold plasma modified the culture media, non-inoculated half-strength
7 PDA plates were also treated with cold plasma prior to inoculation with the *F. graminearum*
8 isolates. Similarly, to determine if a rise in temperature during the treatment impacted the
9 growth of the isolates or the culture media itself, the plates were treated with dry heat before
10 and after inoculation with the isolates. Treatment of *F. graminearum* isolates with cold plasma
11 inhibited their growth and was not associated with the culture media or the rise in temperature
12 during the treatment. Optical emission spectroscopy of cold plasma identified reactive (ionised)
13 species of argon, nitrogen, hydrogen, oxygen, copper, and carbon with the highest number of
14 peaks produced for argon. These results demonstrate that cold plasma can significantly reduce
15 the *in vitro* growth of *F. graminearum* isolates when treated in a closed environment and
16 suggest there is potential to control the *in vivo* growth of *F. graminearum*.

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17 **Introduction**

18 *Fusarium graminearum* is a devastating fungal pathogen associated with fusarium head blight
19 (FHB) in wheat and other cereal crops. *F. graminearum* chlamydospores and perithecia survive
20 in host crop residues or stubble and serve as the primary source of inoculum (Wegulo, 2012).
21 Ascospores or conidia are released from the infected stubble under warm wet conditions, and
22 dispersed to nearby wheat plants by wind or rain splash, where they germinate and infect
23 susceptible wheat heads, initiating the development of FHB (Wegulo, 2012). An integrated
24 management approach to reduce inoculum levels through crop rotation and fungicide
25 application during or before anthesis (approx. GS 59 - 65) is the primary strategy to mitigate
26 disease development (Blandino et al., 2012; Wegulo et al., 2011). However, the integrated
27 management strategy's efficacy depends on many factors such as cultivar resistance to the
28 pathogen, weather conditions, and the type of fungicide used (Blandino et al., 2012; Edwards,
29 2004). Furthermore, if the disease infection occurs at later stages of grain development, grain
30 may appear normal and potentially carry the pathogen to storage (Osborne & Stein, 2007)
31 intensifying the need for an effective and reliable treatment.

32 Mycotoxins, such as trichothecenes, are also produced by *F. graminearum*. These are toxic
33 secondary metabolites, and the consumption of products from mycotoxin-contaminated cereal
34 grain causes serious health issues in humans and animals (Foroud & Eudes, 2009).
35 Trichothecenes are mainly produced during crop maturation or when the grain are stored under
36 high humidity and temperature (Hope et al., 2005). Once the mycotoxins are established in
37 cereal grain, they are difficult to eliminate due to their chemical and thermal stability (Generotti
38 et al., 2015). Cold plasma could potentially provide a solution for managing postharvest
39 problems of *F. graminearum* in cereal grain.

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40 Cold plasma is an ionised form of gas, which contains reactive oxygen species (ROS), reactive
41 nitrogen species (RNS), electrons, and other free radicals. ROS and RNS are known to damage
42 microbial cells through the oxidation of cytoplasmic membranes, lipids, proteins, and DNA
43 strands (Bourke et al., 2017; Misra et al., 2019), which induce membrane perforation and other
44 changes in the hyphal surface, thereby impacting growth (Avramidis et al., 2010; Simoncicova
45 et al., 2018). The potential of cold plasma to reduce seed-borne fungal contamination in wheat,
46 oats, corn, and other grain was first reported by Selcuk et al. (2008). They demonstrated that
47 up to a 99 % reduction in fungal load of *Aspergillus* and *Penicillium* species on wheat grain
48 was achievable following cold plasma treatment for 15 min; however, disinfection was
49 dependent on the gas used to generate the plasma and the surface of the treated grain, amongst
50 other factors. Other authors have reported similar findings with different grain commodities,
51 including rice (Kang et al., 2015; Ochi et al., 2017) and barley (Los et al., 2018).

52 A problem with most of the research in this field is that published studies have used widely
53 different and often custom-built plasma generation systems, resulting in a diverse range of
54 treatments that make comparisons difficult and identification of optimal conditions almost
55 impossible (Sarangapani et al., 2018). The effectiveness of cold plasma can be influenced by
56 factors such as the electrical voltage used to generate the plasma and the duration of treatment
57 of the commodity with the plasma (Braşoveanu et al., 2015). Therefore, it is essential to define
58 each commodity's treatment conditions or microbe of interest based on the cold plasma
59 equipment being used.

60 This study developed optimised methods for cold plasma treatment of *in vitro* cultures of *F.*
61 *graminearum*. Experiments were also designed to test the hypothesis that cold plasma inhibits
62 the *in vitro* growth of *F. graminearum* cultures isolated from postharvest wheat and barley.

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63 **Materials and Methods**

64 *Fungal cultures*

65 Four *F. graminearum* isolates, WAC11354, WAC11387, WAC11490 and WAC12336, were
66 provided by the Department of Primary Industries and Regional Development (DPIRD) in
67 Western Australia. Of these isolates, two were isolated from wheat grain, one from barley grain
68 and one from wheat stubble (Table 1). Two isolates, WAC11354 and WAC11490, were
69 associated with FHB disease in wheat (previously determined by DPIRD). The isolates were
70 grown on half-strength Potato Dextrose Agar (PDA [Difco] powder 19.5 g + Agar [Difco] 9.5
71 g + deionised water 1 L, referred to as ½ PDA for all experiments), in the dark at 25±1°C.
72 Disks of 3 mm taken from the edge of actively growing, four-day-old cultures were aseptically
73 transferred to 10 mL of ½ PDA in 90 mm Petri plates immediately before cold plasma
74 treatment.

75 **Table 1** *Fusarium graminearum* isolates used for all the treatments, their host species,
76 collection year and the place of origin.

WAC ^{ab} number	Host species	Collection year	Isolated from
WAC11354	<i>Triticum aestivum</i> (Wheat)	2005	Grain
WAC11387	<i>T. aestivum</i>	2004	Grain
WAC11490	<i>T. aestivum</i>	2004	Stubble
WAC12336	<i>Hordeum vulgare</i> (Barley)	2004	Grain

77 ^a WAC-WA Culture Collection numbers allocated by the Department of Primary Industries and
78 Regional Development Herbarium.

79 ^b Locality of origin of all the isolates- Wellstead, Western Australia.

80 *Cold plasma Equipment*

81 A Blown-Arc Pro Series plasma surface treater (Enercon, Wisconsin, US) was used for all
82 experiments. The Blown-Arc Pro produces a gliding arc plasma with a maximum output

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83 voltage of 3.4 KV_{RMS} at an output frequency range of 20-60 kHz. The plasma is generated by
84 supplying compressed atmospheric air at 70-90 psi through high voltage electrodes.

85 *Optical emission spectroscopy and temperature measurement*

86 A fibre optic spectrometer (Avantes AvaSpec-2048-8 with AvaSpec v. 8.3.1.0 software) was
87 used to monitor the reactive oxygen and nitrogen species produced during plasma treatment. A
88 400 nm fibre optic cable was placed at a distance of 21 cm from the plasma emission point to
89 detect the spectra of reactive species, as previously described by Siddique et al. (2019). The
90 Blown-Arc Pro emission spectra were measured at the centre and edge of the plasma flame and
91 then combined to produce a final spectrum for each environment.

92 The temperature during treatment was recorded every 15 s by placing a four-inch food-grade
93 stainless steel probe attached to a Hobo UX120-006 4-Channel Analog Logger (Temperature
94 Sensor TMC6-HC) at the level of the Petri plate surface.

95 *Optimised cold plasma treatment of F. graminearum*

96 Following a series of preliminary trials (data not shown), the Blown-Arc Pro was optimised to
97 treat the *F. graminearum* isolates for a duration of 70 s from a distance of 21 cm inside a closed
98 box which was referred as a closed environment treatment, with the Petri plate lids removed.
99 All plates were sealed with plastic food wrap (Glad® Wrap) and incubated at 25±1°C in the
100 dark after treatment. The radial colony growth was measured every 24 hr by recording the
101 cultures' diameter on the two perpendicular axes for up to five days or until the untreated
102 control cultures reached the edge of the plate. The radial colony growth was then calculated as

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103 the average growth rate over the five days. There were ten replicate plates for each isolate, and
104 the entire trial was conducted twice.

105 *Effect of media treated with cold plasma on F. graminearum growth*

106 To determine whether cold plasma treatment modified the culture media and consequently
107 inhibited culture growth, non-inoculated ½ PDA was also treated with cold plasma with the
108 Petri plate lids removed, using the optimised conditions described above. Treated PDA was
109 then aseptically inoculated with disks of actively growing cultures of all four isolates and radial
110 colony growth was measured as described above. There were ten replicates, and this trial was
111 conducted twice.

112 *Effect of heat generated during cold plasma treatment on F. graminearum growth*

113 While conducting the preliminary trials, the cold plasma produced by the Blown-Arc Pro was
114 observed to increase the temperature at the surface of the PDA plate to no more than 120 °C in
115 the closed environment treatment (data not shown). Therefore to eliminate any possible impact
116 of temperature on culture growth or directly on PDA during treatment, two dry heat trials were
117 conducted. In the first trial, disks of 3 mm taken from the edge of actively growing, four-day-
118 old cultures were aseptically transferred to 10 mL of ½ PDA in 90 mm Petri plates as described
119 above, sealed with plastic food wrap, and placed in an oven at 120±2 °C for 80 s. In the second
120 trial, non-inoculated ½ PDA plates were placed in the oven as described above and then
121 aseptically inoculated. Controls in both trials were kept at room temperature. All cultures were
122 incubated at 25±1 °C in the dark, and radial colony growth was measured as described above.
123 There were ten replicate plates for each isolate in each trial, and both trials were conducted
124 twice.

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125 *Statistical analysis*

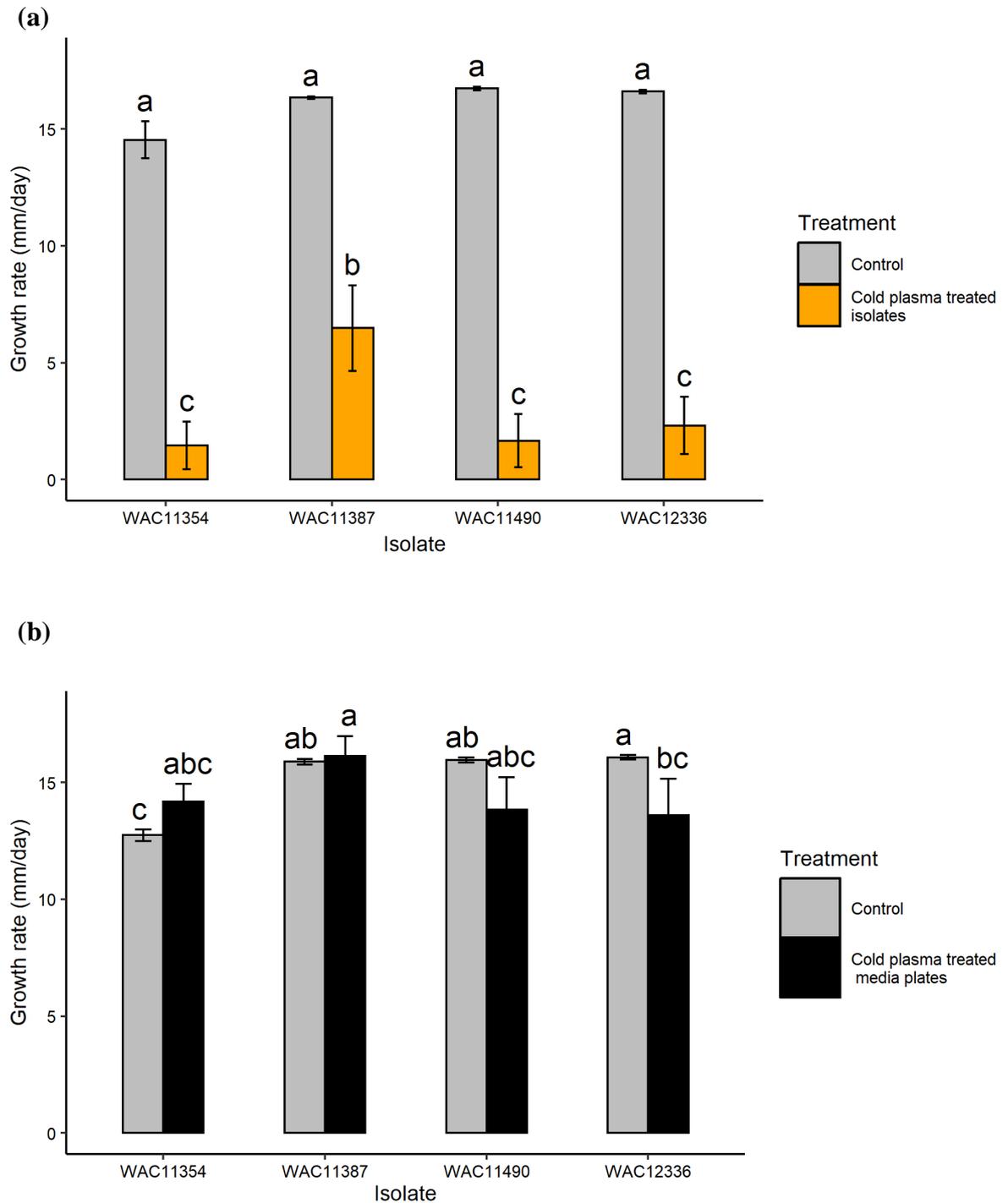
126 All data were expressed as means and standard errors of all replicates. The repeated measure
127 ANOVA showed no significant difference between the repeated trials; therefore, all means of
128 the repeated trials were combined. The data were analysed using One-way Analysis of
129 Variance (ANOVA) and compared by Fisher's Least Significant Difference (LSD) in RStudio
130 (version 3.5.2). The level of significance was considered below 0.05. The results were plotted
131 using ggplot packages in R.

132 **Results**

133 *Impact of cold plasma on F. graminearum growth rates*

134 The application of cold plasma to *in vitro* cultures of *F. graminearum* for 70 s from a distance
135 of 21 cm significantly ($P < 0.001$) reduced the growth rate of all isolates in a closed
136 environment, although WAC11387 was less affected than the other isolates (Fig. 1a). Further,
137 the treatment of ½ PDA with cold plasma for 70 s before inoculation showed no noticeable (P
138 > 0.2) impact on the media, such that it subsequently influenced *F. graminearum* growth rates.
139 However, the treatment had a significant interaction ($P = 0.01$, Fig. 1b) with the isolates. At
140 the individual isolate level, the isolate WAC12336 had a significantly lower growth rate on
141 treated ½ PDA plates compared to untreated plates, whereas the growth rate of the other three
142 isolates did not change significantly from the controls.

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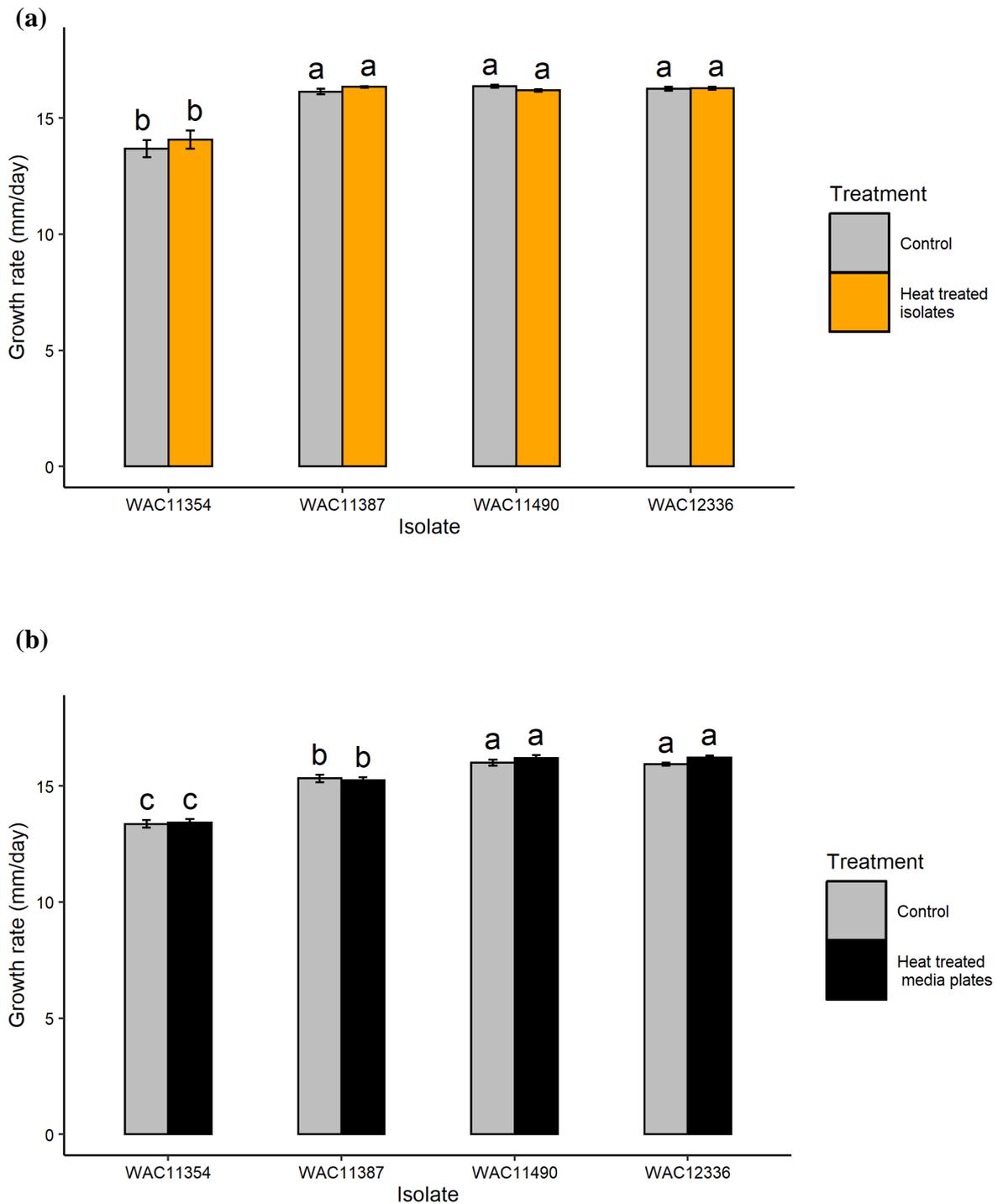
143 **Fig. 1** Cold plasma treatments of four *Fusarium graminearum* isolates for 70 s at a distance of
144 21 cm from the plasma emission point in a closed environment with the Blown-Arc Pro. The
145 graphs represent a) cold plasma treatment of inoculated plates and b) cold plasma treatment of
146 half-strength Potato Dextrose Agar plates that were then immediately inoculated. Bars =

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147 standard errors of the mean (n = 20). Means with the same letter are not significantly different
148 according to Fisher's Least Significant Difference test.

149 Dry heat treatment of plates inoculated with *F. graminearum* isolates (Fig. 2a) also had no
150 significant (P = 0.497) influence on culture growth rate. Similarly, when the PDA was treated
151 with dry heat and inoculated immediately after the treatment, no significant (P = 0.167) impact
152 of the treatment was observed for the isolates grown on the treated plates (Fig. 2b). Therefore,
153 the rise in temperature during cold plasma treatment did not influence the growth rate of *F.*
154 *graminearum*.

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155 **Fig. 2** Dry heat treatments of four *Fusarium graminearum* isolates performed at 120 °C for 80
156 s. The graphs represent a) dry heat treatment of inoculated plates and b) dry heat treatment of
157 half-strength Potato Dextrose Agar plates that were immediately inoculated. Bars = standard
158 errors of the mean (n = 20). Means with the same letter are not significantly different according
159 to Fisher's Least Significant Difference test.

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160 *Optical emission spectroscopy of plasma generated by the Blown-Arc Pro*

161 The Blown-Arc Pro produced a total of 28 peaks for different ionised molecules. At a selected
162 noise/signal level of 15, the peak number and the intensity of the peaks produced by the Blown-
163 Arc Pro were highest for argon (Ar) (Fig. 3). High numbers and intensities of peaks were also
164 observed for ionised nitrogen (N), oxygen (O), copper (Cu), and carbon (C II). Other molecules
165 detected by the emission spectra were ionised carbon dioxide (CO₂⁺), carbon monoxide anion
166 (CO⁺), dihydrogen (H₂), and amino radical (NH₂), dinitrogen (N₂), N₂⁺, hydroxide (OH), and
167 oxygen molecules (O₂), cyclopropatriene (C₃) and nitrogen dioxide (NO₂).

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174 **Discussion**

175 This study has demonstrated that cold plasma treatment can significantly reduce the *in vitro*
176 growth rate of *F. graminearum* colonies. This finding was consistent with Na et al. (2013),
177 who demonstrated that *in vitro* cold plasma treatment of 60 s or longer completely inhibited
178 the spore germination and hyphal growth of *F. graminearum*. In contrast to Na et al. (2013),
179 the current study also demonstrated that the temperature generated during treatment and the
180 impact of cold plasma on the culture medium did not influence culture growth. Conclusively,
181 the inhibition of *F. graminearum* growth was a result of cold plasma treatment.

182 The closed treatment environment chosen for the current study seems to be commonly practised
183 for cold plasma application to different microorganisms and commodities. The use of cold
184 plasma in a closed or sealed environment has been demonstrated to successfully treat various
185 bacterial and fungal pathogens (Los et al., 2018; Los et al., 2020; Misra et al., 2014; Ouf et al.,
186 2015; Selcuk et al., 2008) and food commodities (Ouf et al., 2015; Selcuk et al., 2008). It was
187 suggested by Siddique et al. (2019) that there is higher efficacy of the treatment in a closed
188 environment due to the higher intensity of ionised molecules observed when compared to an
189 open environment. The ionised molecules are likely dissipated into the air during an open
190 environment treatment. Interestingly, Siddique et al. (2019), using a similar type of gliding arc
191 plasma generator (Enercon Dyne-A-Mite), only observed three peaks during cold plasma
192 treatment in the closed environment. In contrast, in the current study, the Blown-Arc Pro
193 produced 28 peaks, indicating distinct differences between similar forms of cold plasma. The
194 properties of the cold plasma produced by different machines should be considered when
195 developing a pathogen treatment protocol.

196 Currently, the grain industry prominently relies on foliar fungicides to manage the pathogen in
197 the field (Wegulo et al., 2015) in addition to other management practices. However, the
198 efficacy of these fungicides highly depends upon the timing of the fungicide application,

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199 coverage, crop cultivars and virulence of isolates (Mesterházy et al., 2003). Becher et al. (2010)
200 reported that the *F. graminearum* strains can adapt to these fungicides, potentially developing
201 completely new isolates with stronger virulence. The emission spectroscopy in the current
202 study showed that cold plasma produced by the Blown-Arc Pro contains various ionised
203 molecules of N, O and C which could potentially be responsible for the inhibition of *F.*
204 *graminearum* growth. Though the mode of action of cold plasma yet requires further
205 investigation, other authors have found that plasma reactive species changes the hyphae surface
206 and spores' structure thus reducing the fungi's ability to grow (Kang et al., 2015; Simoncicova
207 et al., 2018). It also kills pathogens by damaging microbial cell structures such as cell
208 membranes, lipids, proteins, and DNA strand via ROS and RNS (Bourke et al., 2017). Due to
209 its multimodal and non-specific action mechanism, it is unlikely that the pathogen will adapt
210 or develop resistance against the treatment. It is also evident from the study that the efficacy of
211 the treatment is consistent against multiple isolates of *F. graminearum*. Therefore, cold plasma
212 could provide an advantageous solution for the global grains industry.

213 A possible application of cold plasma for the grains industry could be treating seed grain to
214 prevent seedling blight caused by *F. graminearum*. Seedling blight primarily originates from
215 planting *Fusarium* infected seed which carries the pathogen but are asymptomatic (Jones,
216 1999). Gilbert et al. (2003) demonstrated that the infected seed impacts seed germination up
217 to 75 % possibly by overtaking and killing the seed at germination, thus ultimately reducing
218 crop productivity. Under warm and humid conditions, the fungus from infected seed could
219 produce prithecia and wind-spread ascospores, and increase the FHB severity (Gilbert et al.,
220 2003). Apart from fungicide seed treatment, currently no other treatments are available to
221 manage *F. graminearum* infected seed. Similar to foliar fungicides, as stated above, the
222 efficacy of fungicide seed treatment depends on the inoculum level present on the seed and the
223 environmental conditions to develop seedling blight (Hysing & Wiik, 2014). A potential use

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224 of cold plasma treatment could be to reduce the fungal load present on the infected seed, which
225 would be investigated in future.

226 In conclusion, this study confirmed that cold plasma could significantly reduce the *in vitro*
227 growth of *F. graminearum* in a closed environment. Cold plasma does not influence the
228 pathogen growth due to its impact on culture media or rise in the temperature during the
229 treatments. The emission spectroscopy showed the presence of different ionised molecules in
230 the cold plasma generated during the treatment. However, the study also indicated that the
231 intensity of these ionised molecules could differ with the type of machine used to generate the
232 plasma. For this reason, it is essential to standardise the cold plasma treatment conditions, to
233 effectively control plant pathogens before beginning trials on an industrial scale.

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Author contributions

All authors contributed to the study's conception and design. Material preparation, data collection and analysis were performed by Maninder Kaur. The first draft of the manuscript was written by Maninder Kaur and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Declarations

Conflicts of interest

The authors declare no conflicts of interest.

Availability of data and material (data transparency)

The current study data are not publicly available due to the technology's commercial interest but are available from the corresponding author on reasonable request.

Code availability

All data analysis was performed using Rstudio (version 3.5.2) and the codes available upon request.

Ethical approval

This article does not contain any studies with human participants or animals performed by any of the authors.

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References:

- Avramidis G, Stüwe B, Wascher R, Bellmann M, Wieneke S, von Tiedemann A, Viöl W (2010). Fungicidal effects of an atmospheric pressure gas discharge and degradation mechanisms. *Surface and Coatings Technology* 205:S405-S408. <https://doi.org/10.1016/j.surfcoat.2010.08.141>
- Becher R, Hettwer U, Karlovsky P, Deising HB, Wirsler SGR (2010). Adaptation of *Fusarium graminearum* to tebuconazole yielded descendants diverging for levels of fitness, fungicide resistance, virulence, and mycotoxin production. *Phytopathology* 100(5):444-453. <https://doi.org/10.1094/PHYTO-100-5-0444>
- Blandino M, Haidukowski M, Pascale M, Plizzari L, Scudellari D, Reyneri A (2012). Integrated strategies for the control of *Fusarium* head blight and deoxynivalenol contamination in winter wheat. *Field Crops Research* 133:139-149. <https://doi.org/10.1016/j.fcr.2012.04.004>
- Bourke P, Ziuzina D, Han L, Cullen PJ, Gilmore BF (2017). Microbiological interactions with cold plasma. *Journal of Applied Microbiology* 123(2):308-324. <https://doi.org/10.1111/jam.13429>
- Braşoveanu M, Nemţanu M, Surdu-Bob C, Karaca G, Erper I (2015). Effect of glow discharge plasma on germination and fungal load of some cereal seeds. *Romanian Reports in Physics* 67(2):617-624.
- Edwards SG (2004). Influence of agricultural practices on *Fusarium* infection of cereals and subsequent contamination of grain by trichothecene mycotoxins. *Toxicology Letters* 153(1):29-35. <https://doi.org/10.1016/j.toxlet.2004.04.022>
- Foroud NA, Eudes F (2009). Trichothecenes in cereal grains. *International Journal of Molecular Sciences* 10(1):147-173. <https://doi.org/10.3390/ijms10010147>

Cold plasma treatment to inhibit *Fusarium graminearum* growth

Generotti S, Cirlini M, Malachova A, Sulyok M, Berthiller F, Dall'Asta C, & Suman M (2015).

Deoxynivalenol & Deoxynivalenol-3-Glucoside Mitigation through Bakery Production

Strategies: Effective Experimental Design within Industrial Rusk-Making Technology.

Toxins (Basel) 7(8):2773-2790. <https://doi.org/10.3390/toxins7082773>

Gilbert, J., Woods, S. M., Conner, R. L., Fernandez, M. R., & McLaren, D. (2003). Role of

spring wheat seed infested with *Fusarium graminearum* in spread and development of

fusarium head blight and effects on agronomic performance. Canadian Journal of Plant

Pathology, 25(1), 73-81. <https://doi.org/10.1080/07060660309507051>

Hope R, Aldred D, Magan N (2005). Comparison of environmental profiles for growth and

deoxynivalenol production by *Fusarium culmorum* and *F. graminearum* on wheat grain.

Letters in Applied Microbiology 40(4):295-300. [https://doi.org/10.1111/j.1472-](https://doi.org/10.1111/j.1472-765X.2005.01674.x)

[765X.2005.01674.x](https://doi.org/10.1111/j.1472-765X.2005.01674.x)

Hysing, S.-C., & Wiik, L. (2014). *Fusarium* seedling blight of wheat and oats: effects of

infection level and fungicide seed treatments on agronomic characters. Acta

Agriculturae Scandinavica, Section B — Soil & Plant Science, 64(6), 537-546.

<https://doi.org/10.1080/09064710.2014.929731>

Jones, R. (1999). Seedling blight development and control in spring wheat damaged by

Fusarium graminearum group 2. Plant Disease, 83(11), 1013-1018.

<https://doi.org/10.1094/PDIS.1999.83.11.1013>

Kang MH, Pengkit A, Choi K, Jeon SS, Choi HW, Shin DB, Choi EH, Uhm HS, Park G (2015).

Differential inactivation of fungal spores in water and on seeds by ozone and arc

discharge plasma. PLoS One 10(9):e0139263.

<https://doi.org/10.1371/journal.pone.0139263>

Los A, Ziuzina D, Akkermans S, Boehm D, Cullen PJ, Van Impe J, Bourke P (2018).

Improving microbiological safety and quality characteristics of wheat and barley by

Cold plasma treatment to inhibit *Fusarium graminearum* growth

high voltage atmospheric cold plasma closed processing. *Food Research International* 106:509-521. <https://doi.org/10.1016/j.foodres.2018.01.009>

Los A, Ziuzina D, Boehm D, Bourke P (2020). Effects of cold plasma on wheat grain microbiome and antimicrobial efficacy against challenge pathogens and their resistance. *International Journal of Food Microbiology* 335:108889. <https://doi.org/10.1016/j.ijfoodmicro.2020.108889>

Mesterházy Á, Bartók T, Lamper C (2003). Influence of wheat cultivar, species of *Fusarium*, and isolate aggressiveness on the efficacy of fungicides for control of Fusarium head blight. *Plant Disease* 87(9):1107-1115. <https://doi.org/10.1094/PDIS.2003.87.9.1107>

Misra N, Yadav B, Roopesh M, Jo C (2019). Cold plasma for effective fungal and mycotoxin control in foods: mechanisms, inactivation effects, and applications. *Comprehensive Reviews in Food Science and Food Safety* 18(1):106-120. <https://doi.org/10.1111/1541-4337.12398>

Misra NN, Patil S, Moiseev T, Bourke P, Mosnier JP, Keener KM, Cullen PJ (2014). In-package atmospheric pressure cold plasma treatment of strawberries. *Journal of Food Engineering* 125:131-138. <https://doi.org/10.1016/j.jfoodeng.2013.10.023>

Na YH, Park G, Choi EH, Uhm HS (2013). Effects of the physical parameters of a microwave plasma jet on the inactivation of fungal spores. *Thin Solid Films* 547:125-131. <https://doi.org/10.1016/j.tsf.2013.04.055>

Ochi A, Konishi H, Ando S, Sato K, Yokoyama K, Tsushima S, Yoshida S, Morikawa T, Kaneko T, Takahashi H (2017). Management of bakanae and bacterial seedling blight diseases in nurseries by irradiating rice seeds with atmospheric plasma. *Plant Pathology* 66(1):67-76. <https://doi.org/10.1111/ppa.12555>

Cold plasma treatment to inhibit *Fusarium graminearum* growth

Osborne LE, Stein JM (2007). Epidemiology of *Fusarium* head blight on small-grain cereals.

International Journal of Food Microbiology 119(1-2):103-108.

<https://doi.org/10.1016/j.ijfoodmicro.2007.07.032>

Ouf SA, Basher AH, Mohamed AA (2015). Inhibitory effect of double atmospheric pressure

argon cold plasma on spores and mycotoxin production of *Aspergillus niger*

contaminating date palm fruits. Journal of the Science of Food and Agriculture

95(15):3204-3210. <https://doi.org/10.1002/jsfa.7060>

Sarangapani C, Patange A, Bourke P, Keener K, Cullen P (2018). Recent advances in the

application of cold plasma technology in foods. Annual review of food science and

technology 9:609-629. <https://doi.org/10.1146/annurev-food-030117-012517>

Selcuk M, Oksuz L, Basaran P (2008). Decontamination of grains and legumes infected with

Aspergillus spp and *Penicillium spp* by cold plasma treatment. Bioresource Technology

99(11):5104-5109. <https://doi.org/10.1016/j.biortech.2007.09.076>

Siddique SS, St J Hardy GE, Bayliss KL (2019). Cold plasma as a novel treatment to reduce

the *in vitro* growth and germination of *Colletotrichum species*. Plant Pathology

68(7):1361-1368. <https://doi.org/10.1111/ppa.13059>

Simoncicova J, Kalinakova B, Kovacik D, Medvecká V, Lakatos B, Krystofova S, Hoppanova

L, Paluskova V, Hudecova D, Durina P, Zahoranova A (2018). Cold plasma treatment

triggers antioxidative defense system and induces changes in hyphal surface and

subcellular structures of *Aspergillus flavus*. Applied Microbiology and Biotechnology

102(15):6647-6658. <https://doi.org/10.1007/s00253-018-9118-y>

Wegulo SN (2012). Factors influencing deoxynivalenol accumulation in small grain cereals.

Toxins (Basel) 4(11):1157-1180. <https://doi.org/10.3390/toxins4111157>

Cold plasma treatment to inhibit *Fusarium graminearum* growth

Wegulo SN, Baenziger PS, Hernandez Nopsa J, Bockus WW, Hallen-Adams H (2015).

Management of Fusarium head blight of wheat and barley. *Crop Protection* 73:100-107.

<https://doi.org/10.1016/j.cropro.2015.02.025>

Wegulo SN, Bockus WW, Nopsa JH, De Wolf ED, Eskridge KM, Peiris KH, Dowell FE (2011).

Effects of integrating cultivar resistance and fungicide application on Fusarium head blight and deoxynivalenol in winter wheat. *Plant Disease* 95(5):554-560.

<https://doi.org/10.1094/PDIS-07-10-0495>