Privacy-preserving integration of multiple institutional data for single-cell type identification with scPrivacy

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Abstract

The rapid accumulation of large-scale single-cell RNA-seq datasets from multiple institutions presents remarkable opportunities for automatically cell annotations through integrative analyses. However, the privacy issue existed but being ignored, since we are limited to access all the reference datasets distributed in different institutions globally due to the prohibited data transmission across institutions by data regulation laws. To this end, we present *scPrivacy*, which is the first and generalized automatically single-cell type identification prototype to facilitate single cell annotations in a data privacy-preserving collaboration manner. We evaluated *scPrivacy* on a comprehensive set of publicly available benchmark datasets for single-cell type identification to stimulate the scenario that the reference datasets are rapidly generated and distributed in multiple institutions, while they are prohibited to be integrated directly or exposed to each other due to the data privacy regulations, demonstrating its effectiveness and time efficiency for privacy-preserving integration of multiple institutional datasets in single cell annotations.

Introduction

Single-cell transcriptomics is indispensable for understanding cellular mechanisms of complex tissues and organisms[1-6]. As single-cell technologies rapidly developed over recent years, its experimental throughput increased substantially, allowing to profile increasingly complex and diverse samples, and accumulating vast numbers of datasets over time. Integrative analyses of such large-scale datasets originating from various

samples, different platforms and different institutions globally, offer unprecedented opportunities to establish a comprehensive picture of cell landscape. To this end, various community generated large-scale atlas-level single cell reference data, such as the Human Cell Atlas (HCA[7]), Human Tumor Atlas Network[8], BRAIN Initiative Cell Census Network[9] , Human Lung Atlas[10], Human Gut Atlas[11], Human BioMolecular Atlas Program (HuBMAP[12]), The Tabula Sapiens[13], hECA[14] etc., and recently great achievements has been made in the building of pan-tissue single-cell transcriptome atlases covering more than a million cells, including 500 cell types,across more than 30 human tissues from 68 donors[13, 15-18].These references data facilitate the automatically cell type annotations in a supervised way without prior marker gene annotations[19-24]. It is obviously that integrating more reference datasets or combining these atlas-level data will improve the cell type annotations [19, 20, 24, 25], and various integration methods for single cell annotations have been presented[19, 26, 27, 28]. However, all these existing integration methods require to access the relevant reference datasets directly, which may be unavailable due to the data privacy and security issues. Currently, the privacy and political issues towards omics data transmission and sharing across different institutions or countries are gradually attracting attentions. On the one hand, countries around the world are strengthening laws to protect data privacy and security by prohibition of certain data transition across countries or organizations. Such regulations include the General Data Protection Regulation (GDPR[29]) implemented by the European Union, the Health Insurance Portability and Accountability Act (HIPPA[30]) and Health Information Technology for Economic and Clinical Health Act (HITECH[31]) enacted by U.S. and etc. On the other hand, single-cell reference datasets are generated and accumulated rapidly in different institutions around the world. It is a great demand to integrate all these institutional data globally to facilitate the establishment of the comprehensive picture of human cell reference map. However, these institutions may be required to protect data privacy and security and prohibit certain data transmission across organizations and countries by data regulation laws (Table 1). As a result, there exist an inevitable contradiction between the rapidly accumulated single cell reference data and the privacy issue among data sharing and integration. Current references integrating strategies failed to address these issues, hindered by legal restrictions on data sharing[32]. Moreover, current integrating methods ignore the problem of privacy towards single cell sequencing data, as scRNA-seq datasets of human are sensitive and likely to contain sufficient sequencing depth to call genetic variants[33].

To this end, taking the single-cell transcriptome data as an initial study, we propose *scPrivacy*, an efficient, flexible and extendable automatically single-cell type

identification prototype and a proof-of-concept study to facilitate single cell annotations in a data privacy-preserving collaboration manner, by integrating multiple references single cell transcriptome data distributed in different institutions using an federated learning based deep metric learning framework. Federated learning is a collaborative paradigm in privacy-preserving computing community that enables the institutions collaboratively train a model while keeping the data in local institutions[34, 35]. We summarized existing privacy-preserving methods with their distint characteristics and explained the necessity to perform privacy-preserving computing for large scale single cell data using federated learning framework (Table 2). In addition, in our previous studies[25, 27], metric learning was also proven to be effective for single-cell type annotation. Briefly, the basic idea of *scPrivacy* is to make each institution train their models locally and aggregate encrypted models parameters for all institutions to avoid putting raw data of all institutions together directly. We evaluated *scPrivacy* on a comprehensive set of 27 publicly available benchmark datasets for single cell type identification to stimulate the scenario that the reference datasets are rapidly generated and accumulated from multiple institutions, while they are prohibited to be integrated directly or exposed to each other due to the data privacy regulations, and demonstrated its effectiveness and time efficiency for privacy-preserving integration of multiple institutional datasets.

Table 1. Single cell atlases and their corresponding data regulation laws

Atlas	Data regulation laws
Human Cell Atlas[7]	GDPR
Expression Atlas[36]	EMBL Internal Policy for Data Protection
Human Tumor Atlas Network[8]	NIH Genomic Data Sharing Policy
BRAIN Initiative Cell Census Network[9]	NIH Genomic Data Sharing Policy
Human Lung Atlas[10]	All U.S. Federal, state and local laws and regulations
Human Gut Atlas[11]	All U.S. Federal, state and local laws and regulations
HuBMAP[12]	HuBMAP External Data Sharing Policy
The Tabula Sapiens[13]	The Tabula Sapiens Privacy Policy including GDPR

Table 2. Current privacy-preserving methods and their properties

	Cleartext	Software Guard Extensions	Homomorphic Encryption	Secure Multi-Party Computation	Federated learning
Risk for calling genetic variants?	Yes	No	No	No	No
Encrypted raw data?	No	No	Yes	Yes	No
Practicable for handling large scale single cell reference data?	Yes	No	No	No	Yes
Applicable to data transmission regulation laws?	No	No	No	No	Yes
Performance compared to cleartext based integration?	/	Identical	Identical	Identical	Upper bound

Results

Overview of *scPrivacy*

scPrivacy is an efficient, flexible and extentable automatically single-cell type identification prototype to facilitate single cell annotations in a data privacy-perserving collaboration manner, by integrating multiple references single cell transcriptome data distributed in different institutions using an federated learning based deep metric learning framework. *scPrivacy* can effectively integrate information from multiple references while keeping each reference in local institutions, so as to solve the problem of data privacy protection. In particular, each institution trains its model on its local dataset and sends encrypted model parameters to server and then the server aggregates the parameters and sends back the aggregated model to institutions iteratively in training process. Specifically, *scPrivacy* comprises two main steps: model learning and cell assignment (Fig. 1 and see Methods).

In the model learning stage (Fig. 1a), *scPrivacy* trains a federated deep metric learning model on multiple institutional datasets in a data privacy-preserving manner. For an individual institution, deep metric learning (DML) is applied to learn an

optimal measurement fitting the relationship among cells in the reference dataset, and the N-pair loss[37] is used as the loss function for model training. With DML, cells belong to the same type became more similar and cells belong to different types became more dissimilar. Then, *scPrivacy* extends DML to a federated learning framework by aggregating model parameters of institutions to construct an aggregated model (Fig. 1a and see Methods), which fully utilized the information contained in multiple institutional datasets to train the aggregated model while avoiding integrating datasets physically. In addition, *scPrivacy* can utilize the complementary information from different institutional datasets to boost the cell assignment performance, while also avoid the over correction of batch effect, as proven in previous study[37]. In the cell assignment stage (Fig. 1b and see Methods), the query dataset is first transformed by the federated model to the same embedding space as that of the transformed institutional datasets. Then, the transformed query dataset is assigned to proper cell types by comparing with cell type landmarks of transformed institutional datasets. Specifically, for each transformed institutional dataset, *scPrivacy* carries out a cell search by measuring the similarity between the transformed query cells and cell type landmarks of the transformed institutional datasets. Finally, the query cells are assigned to the proper cell type with the highest similarity among all landmarks of the transformed institutional datasets.

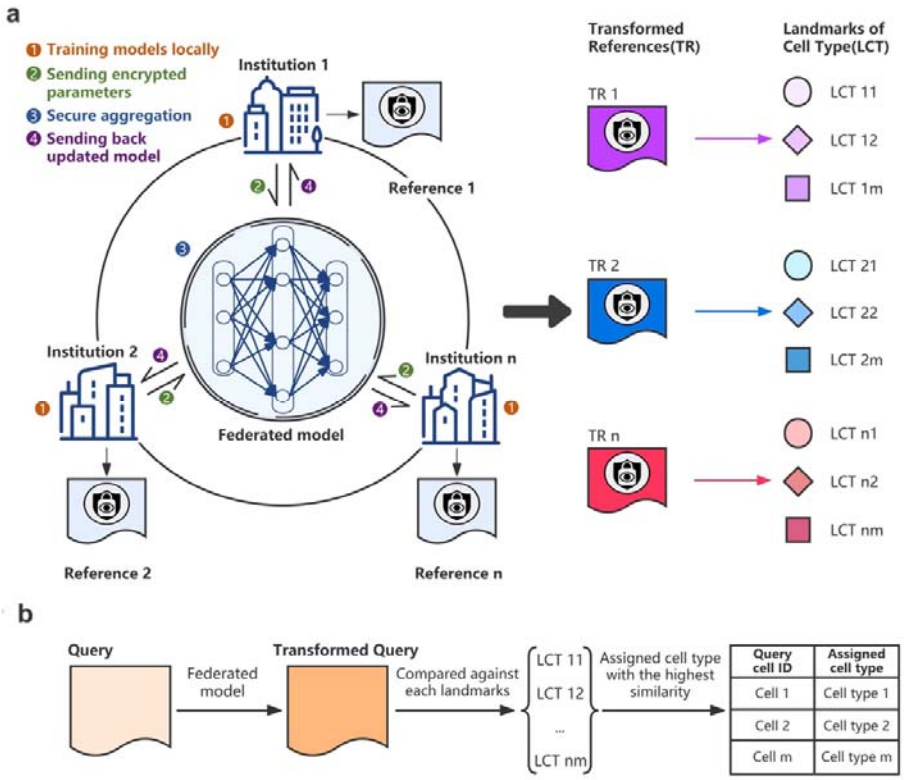

Fig. 1 | The *scPrivacy* workflow. a. The model learning process of *scPrivacy*. The federated model was trained with four steps: (1) Training models locally; (2) Sending encrypted parameters; (3) Secure aggregation; (4) Sending back updated model. Then cell type landmarks will be calculated for each transformed reference. **b.** The cell assignment process of *scPrivacy*. The federated model is utilized to transform the query cells. Then, the transformed query cells are compared against cell type landmarks of transformed institutional datasets, and the predicted cell type with the highest similarity among all cell type landmarks is obtained.

Benchmarking *scPrivacy* from multiple institution with single institution

We firstly benchmarked *scPrivacy* from multiple institution with that of *scPrivacy* from single institution to prove the benefits and necessity of integrating multiple institutional datasets. To this end, we collected 27 datasets (Supplementary Table S1) from four studies of three tissues to simulate the data collaboration scenario among institutions: one study on the brain, one study on the pancreas and two studies on peripheral blood mononuclear cells (PBMCs)[38, 39]. For brain tissue, the study contained four brain datasets[40, 41] with different sources. For pancreas tissue, the study

contained four commonly used pancreas datasets[42-45]. For PBMCs, the first study[39] contained 12 datasets from 12 different sequencing platforms ("PBMC-Mereu"), and the second study[38] contained 7 datasets from 7 different sequencing platforms ("PBMC-Ding"). In this case, each dataset of a tissue is simulated as an institution. For the strategy of *scPrivacy* from single institution, each dataset among multiple datasets was simulated as the dataset in an individual institution to train *scPrivacy* and the rest datasets were used as query datasets. For the strategy of *scPrivacy* from multiple institutions, each dataset among the multiple datasets was used as the query, and the others datasets were used to simulate distributed multiple institutional datasets and they are "virtually" integrated to train *scPrivacy*. It should be noted that in all benchmark scenarios in our study, macro-F1 score was used as the evaluation metric and only query cell types included in multiple institutional datasets were calculated. As shown in Fig. 2a, and Supplementary Table S2, it can be clearly seen that *scPrivacy* from multiple institutions generally obtained great improvement compared with that of *scPrivacy* from single institution, demonstrating the importance of integrating multiple institutional datasets. To further analyze the results, we compared macro-F1 score on 'PBMC-Mereu' and 'PBMC-Ding' studies in terms of each cell type, as the two studies shared several common cell types. The results showed that *scPrivacy* achieved a better performance in almost all cell types, further demonstrating the benefits and necessity to integrate multiple institutional datasets (Fig. 2b and Supplementary Table S3).

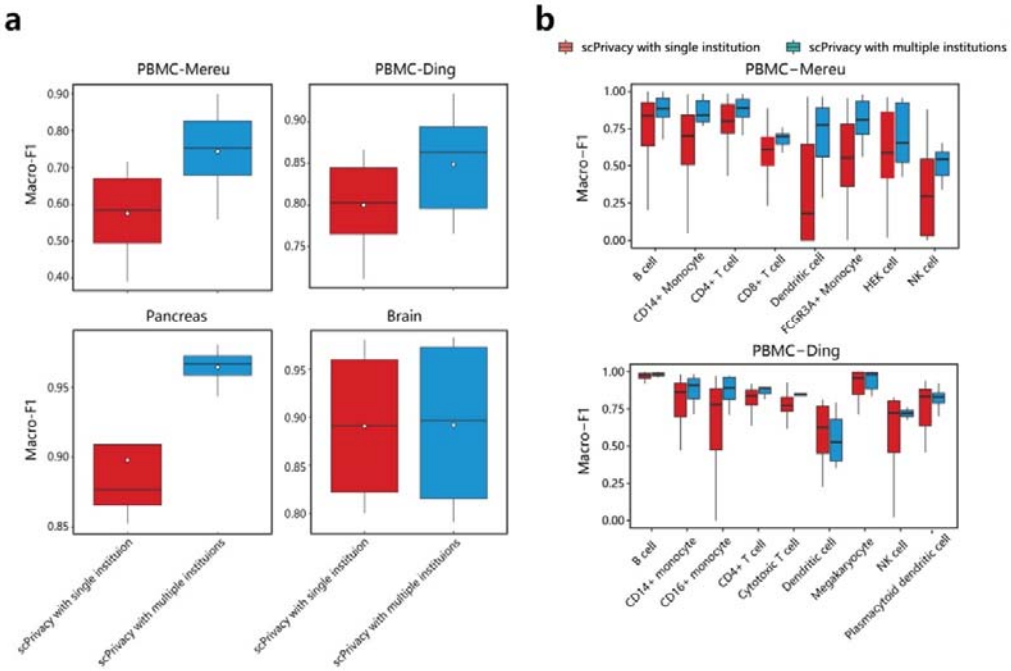

Figure 2. Benchmarking *scPrivacy* with single institution and multiple institutions. **a.** The macro-F1 scores of *scPrivacy* with single institution and multiple institutions for "PBMC-Mereu", "PBMC-Ding", "Brain" and "Pancreas" studies, respectively. The white diamond represents the mean value. **b.** The macro-F1 of each cell type for *scPrivacy* with single institution and multiple institutions on "PBMC-Ding" and "PBMC-Mereu" studies.

Benchmarking *scPrivacy* with non-privacy-preserving multiple reference integrating methods

Then we benchmarked *scPrivacy* with existing non-privacy-preserving multi-reference based single cell type identification methods, including scmap-cluster[26], SingleR[19], Seurat v3[46] and mtSC[47]. In this study, Seurat v3 applied a data-level integration strategy; scmap-cluster and SingleR applied a decision-level integration strategy, and mtSC applied an algorithm-level integration strategy (See Methods), which represent the three mainstream integrative analysis strategies for single cell assignment. The above 27 datasets in 4 studies are used as the benchmark datasets here. In this case, each dataset among the multiple datasets was treated as the query, and the others were used to simulate multiple institutional datasets. It should be noted that *scPrivacy* integrated multiple institutional datasets in a data privacy-preserving

manner while other multi-reference based methods accessed all datasets to integrate them directly. The results are shown in Fig. 3a and Supplementary Table S4. We can see that *scPrivacy* achieved comparable performance to mtSC and performed better than the other methods, while it was trained in a data protection manner. Furthermore, as shown in Fig. 3b and Supplementary Table S5, we compared the macro-F1 of each cell type on 'PBMC-Ding' and 'PBMC-Mereu' studies, and achieved the consistent conclusion that *scPrivacy* achieved the comparable best performance in all four studies as those of mtSC. As a conclusion, *scPrivacy* is able to achieve a comparable best performance to integrate multiple institutional datasets, while keeping the integration in a data privacy-preserving manner.

***scPrivacy* consumes much less time than most multiple reference integrating methods**

As the amount and size of scRNA datasets increasing rapidly, consuming time is an important concern for single cell integration and annotation. Due to the distributed properties of federated learning, the training time of *scPrivacy* only depends on the max training time among the institutions datasets while the training time of other multi-reference based methods are the sum of training time of all institution datasets. Thus, *scPrivacy* obtains a high scalability to deal with the large-scale data integration and model training, which serves a very challenging issue in the building of large-scale atlas level single cell reference data. The following training time and query time comparisons further proved this point (Fig. 3c, Supplementary Table S6 and S7): (1) *scPrivacy* consumes much less time than Seurat v3 and mtSC in the training process. More importantly, the time consumed by *scPrivacy* does not increase exponentially or linearly, since it only depends on the max training time among institutions datasets, indicating its potential ability to handle large-scale datasets. (2) For querying process, *scPrivac*y is also very fast (<1 min for 9000 query cells) and consumes much less time than all the other methods except for scmap-cluster, since scmap-cluster is the simplest method while expense of accuracy substantially.

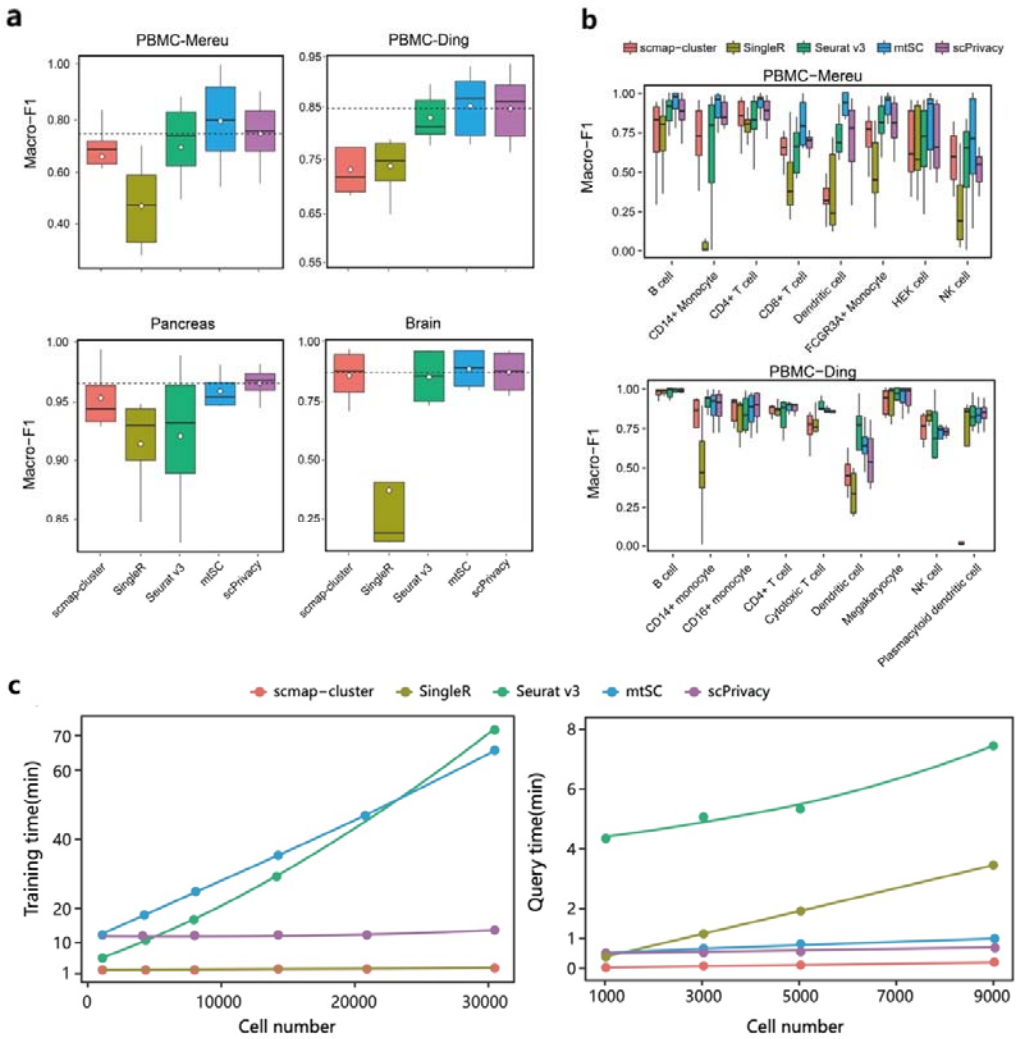

Figure 3. Benchmarking *scPrivacy* with non-privacy-preserving multiple reference integrating methods. **a.** The macro-F1 scores of *scPrivacy* and other existing non-privacy-preserving multiple reference integrating methods on "PBMC-Mereu", "PBMC-Ding", "Brain" and "Pancreas" studies, respectively. **b.** The macro-F1 of each cell type for *scPrivacy* and other existing non-privacy-preserving multiple reference integrating methods on 'PBMC-Ding' and 'PBMC-Mereu' studies. **c.** Training and query time of *scPrivacy* and other existing non-privacy-preserving multiple reference integrating methods. Solid lines are loess regression fitting (span = 2), implemented with R function geom smooth().

***scPrivacy* performs increasingly better as the number of institution reference**

datasets increases

We finally investigated the impact of the number of institutions on the performance of *scPrivacy*. We considered each dataset in "PBMC-Ding" as an institution dataset to simulate the scenario. In this test, *scPrivacy* was trained with different numbers of institution datasets, and then the corresponding macro-F1 score was calculated to show the trend of the performance as the number of institution datasets increased. Specifically, each time, we randomly selected one dataset from the 7 datasets as the query dataset and selected 1 to 6 datasets without replacement from the remaining datasets as the institutional reference datasets. This process was repeated 5 times to reduce randomness. As shown in Fig. 4 and Supplementary Table S8, we can see that *scPrivacy* generally performs increasingly better as the number of institutions increases. Specifically, if the performance is relatively lower in the beginning, the improvement becomes more evident as the number of institution datasets increases. Such an improvement obtained by increasing institution datasets is very important for developing an effect and robust cell annotation system in the era of explosive growth of single-cell datasets. Together with the fast training of *scPrivacy*, it is expected to integrate a growing number of institution datasets to obtain an increasingly better cell annotation in a privacy-preserving manner.

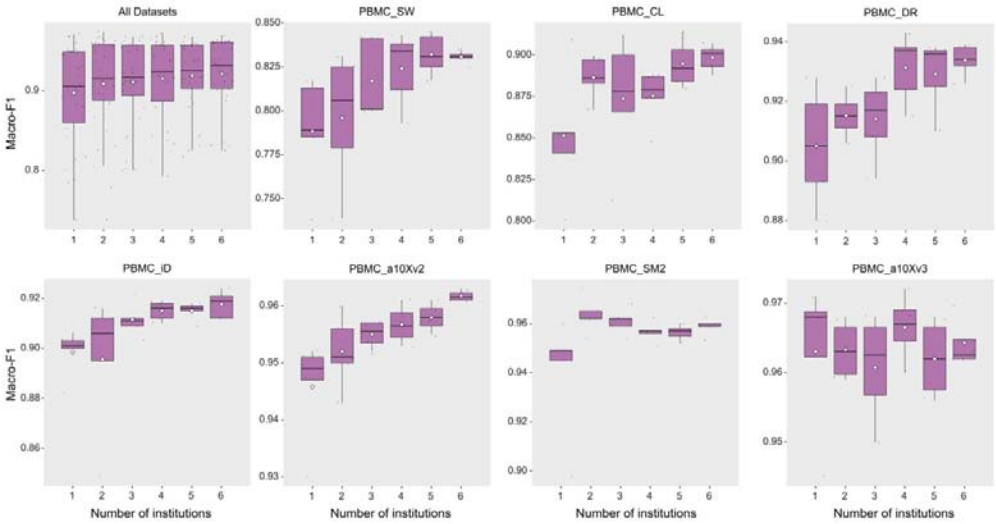

Figure 4. The performance of *scPrivacy* as the number of institution datasets increases. We considered each dataset in "PBMC-Ding" as an institution dataset to simulate the scenario, and showed the macro-F1 scores for overall datasets and each dataset (as query dataset) respectively. The white diamond represents the mean value.

Discussion

In this study, we present the first and generalized federated deep metric learning-based single cell type identification prototype *scPrivacy* to facilitate single cell annotations by integrating multiple institutional datasets in a privacy-preserving manner. As scRNA-seq datasets grow exponentially, multiple institutional datasets can be integrated to build a more comprehensive, effective and robust cell annotation system. Traditional multi-reference based methods faced the problem of data privacy protection. *scPrivacy* solves this issue by federated learning. Specifically, *scPrivacy* trained each institution dataset locally and aggregated encrypted model parameters for all institutions instead of putting raw data of all institutions together to train a model. We evaluated *scPrivacy* on a comprehensive set of publicly available benchmark datasets for single cell type identification to stimulate the scenario that the reference datasets are rapidly generated and distributed in multiple institutions, while they are prohibited to be integrated directly or exposed to each other due to the data privacy regulations, and demonstrated its effectiveness for privacy-perserving integration of multiple institutional datasets. In addition, we proved that *scPrivacy* is time efficient and performs increasingly better as the number of institution datasets increases, which is of great potential utility when more and more institution datasets have been accumulated in the future.

In addition to scRNA-seq technology, other single cell omics, such as scATAC-seq and etc., are developing rapidly and vast numbers of datasets will accumulate in the future. Integrating these multi-omics datasets has the potential to provide a more comprehensive picture of basic biological processes. However, privacy issue is still an unavoidable problem for data sharing and integration. *scPrivacy* can be easily extended to other single cell omics for privacy-preserving integration. In addition, the privacy-preserving computing technologies are evolved rapidly, although federated learning is a suitable solution for large-scale privacy-preserving data integration, it still needs a central server to aggregate model parameters from clients. Recently, blockchain-based federated learning, such as swarm learning[48, 49], is developed which doesn't need the central server, however, its efficiency is waiting to be further evaluated. Nevertheless, a privacy-preserving integration of multiple institutional data globally to build a more comprehensive cell landscape is a challenging issue that must be faced under the global data sharing protection and regulations. We therefore call for attention to such problems and efficient privacy-preserving system for cell annotations are needed to be carefully designed.

Methods

Benchmark data collection. We evaluated *scPrivacy* on 27 reference based single-cell type identification benchmark datasets which were curated from four studies including three tissues: peripheral blood mononuclear cells (PBMCs)[38, 39], the brain[40, 41] and the pancreas[42-45] (Supplementary Table S1). For all datasets, cell types with less than 10 cells were removed since they do not contain enough information and are unreliable for subsequent assignment. Cells labeled "alpha.contaminated", "beta.contaminated", "gamma.contaminated" and "delta.contaminated" in the dataset generated from Xin et al.[45] were removed because they likely corresponded to cells of lower quality. Cells labeled "not applicable" in the dataset generated by Segerstolpe et al.[44] were removed. "L6b", "Pvalb", "Sst" and "Vip" cell types in the dataset generated from Tasic et al.[41] were retained to match the names of cell types in the other three brain datasets[40]. For "PBMC-Mereu"[39], 12 datasets were used and Smart-seq2-based dataset was excluded, which was too small (Supplementary Table S1).

Data preprocessing. The data preprocessing step of *scPrivacy* consists of three parts: cell quality control, rare cell type filtering and gene expression profile formatting. *scPrivacy* evaluates the cell quality on strict criteria. In particular, the number of genes detected requires >500, the number of unique molecular identifiers induced requires >1500, and the percentage of mitochondrial genes detected requires <10% among all genes. Only cells satisfying all three criteria are reserved. Then, all the datasets were normalized by scaling to 10,000 and then with log(counts+1). Finally, all the datasets are processed into an identical format, i.e., expression profiles with the union of the genes in all institution datasets. The column of the gene in query dataset will be filled with zeros if the gene is not in the gene union of the institution datasets.

Model learning of *scPrivacy*. In the model learning stage, *scPrivacy* utilizes federated deep metric learning algorithms to train the federated model on multiple institutional datasets in data privacy protection manner. For a single institution, we use deep metric learning (DML) as the training algorithm and N-pair loss[37] is used as the loss function. DML is applied to learn an optimal measurement fitting the relationship among cells in the reference dataset. With DML, cells with the same label become more similar and cells with different labels become more dissimilar. The implementation of DML in *scPrivacy* is the same as the counterpart of mtSC. The detailed description of DML and N-pair loss can be found in mtSC[47].

scPrivacy extends DML to a federated learning framework. Define N institutions $\{F_1, ...F_N\}$ and their respective data $\{D_1, ...D_N\}$. A federated learning framework allows institutions to collaboratively train a model and no institution F_i expose its data D_i to others.

As shown in Fig. 1, in *scPrivacy*, institutions datasets learn a federated model collaboratively with the help of a server. To train a federated model, our training process can be divided into the following four steps[50]:

•Step 1: Each institution trains its model on its own dataset with DML;

•Step 2: Institutions encrypt their own model parameters and send encrypted parameters to server;

•Step 3: Server performs secure aggregation without obtaining information from any institutions;

•Step 4: Server sends back the aggregated results to institutions and institutions update their models with the decrypted parameters.

In our implementation of *scPrivacy*, we use the Crypten, a ML framework built on PyTorch[51] to implement encryption of model parameters and their operations in secure aggregation. At the central server, we adapted widely-used FedAvg[52] algorithm to aggregate model parameters.

Model parameters of *scPrivacy*. The neural network model was implemented with PyTorch. We use Adam optimizer to update parameters of the network via backpropagation. The learning rate was set to 0.0005. The number of training epochs was set to 300. The L2 regularization rate was set to 0.05.

Query cells assignment. First, the query dataset was scaled to 10,000 and normalized with log(counts+1). The column of the gene will be filled with zeros if the gene is not in the gene union of the institution datasets. Next, the query cells were transformed by the federated model to the same embedding space as that of the transformed institution datasets. Then, the transformed query dataset was assigned to proper cell types by comparing with cell type landmarks of transformed institution datasets. Specifically, for each transformed institution dataset, *scPrivacy* carried out a cell search by measuring similarity between transformed query cells and cell type landmarks of the transformed institution datasets. Pearson correlation coefficient was adopted to calculate similarity as in our previous study[23]. Finally, the query cells obtained the predicted cell types with the highest similarity among all landmarks of the transformed institution datasets.

Benchmarking existing tools. To evaluate the performance of *scPrivacy*, four classical non-privacy-preserving data-integration methods were compared: Seurat v3[24], scmap-cluster[20], SingleR[19] and mtSC[25]. Seurat v3 applied a data-level integration strategy, where multiple datasets are integrated into one dataset directly; scmap-cluster and SingleR applied a decision-level integration method, where the final assignment results are ensembled by individual assignment results; and mtSC applied an algorithm-level integration strategy, where efficient algorithms are designed for model integration, while keeping datasets separately. However, all these methods are needed to access the individual institutional data directly. For scmap-cluster, 'threshold = 0' was set to not assign query cells to 'unassigned'. For SingleR, as the fine-tuning process of SingleR is extremely time consuming, 'fine.tune = FALSE' was set. For Seurat v3, all parameters were the defaults. For mtSC, all parameters were the defaults. In all tests, Seurat v3, scmap-cluster and SingleR and were trained and tested with CPU Intel Xeon Platinum 8165 2.3-3.7GHz. The deep learning based methods mtSC and *scPrivacy* were trained with GPU 1080Ti and tested with the same CPU as the other methods.

Evaluation criteria. The macro-F1 score was used as evaluation criteria. First, the precision and recall of each cell type were calculated. Then, the macro-F1 score was calculated as listed below:

$$\text{macro} - \text{F1} = \frac{1}{N}\sum_{i=1}^{N}\frac{2 * Precision_i * Recall_i}{Precision_i + Recall_i}$$

in which N denotes the number of cell types in a dataset and $Precision_i$ and $Recall_i$ are the precision and recall of the i-th cell type in the dataset.

Data availability

The 27 single cell reference based single-cell type identification benchmark datasets were curated from four studies including three tissues: Peripheral blood mononuclear cells(PBMCs)[38, 39], the brain[40, 41] and the pancreas[42-45] (Supplementary Table S1). The four pancreas datasets[42-45] and one of brain datasets[41] was collected in previous work of scmap[20] (https://hemberg-lab.github.io/scRNA.seq.datasets), and the other three brain datasets and seven datasets in "PBMC-Ding"[38] were curated from the following benchmark study[53] (https://doi.org/10.5281/zenodo.3357167). The 12 datasets in "PBMC-Mereu"[39] were from GSE133549, and the corresponding RData file can be downloaded in https://www.dropbox.com/s/i8mwmyymchx8mn8/sce.all_classified.technologies.RDa

ta?dl=0. All these datasets were converted into Bioconductor SingleCellExperiment (http://bioconductor.org/packages/SingleCellExperiment)class objects.

Code availability

scPrivacy is developed as python package for simulations, which is available at https://github.com/bm2-lab/scPrivacy.

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Author contributions

Q.L, Q.Y. and L.X.F. proposed the idea. S.Q.C implemented models and analyzed data. B.D. and C.T curated the benchmark datasets. B.D, S.G.W, Y.C.G and S.L.F contributed by visualized and analyzed results. Q.L, S.Q.C and B.D wrote the manuscript with assistance from other authors.

Competing interests

The authors declare no competing interests.

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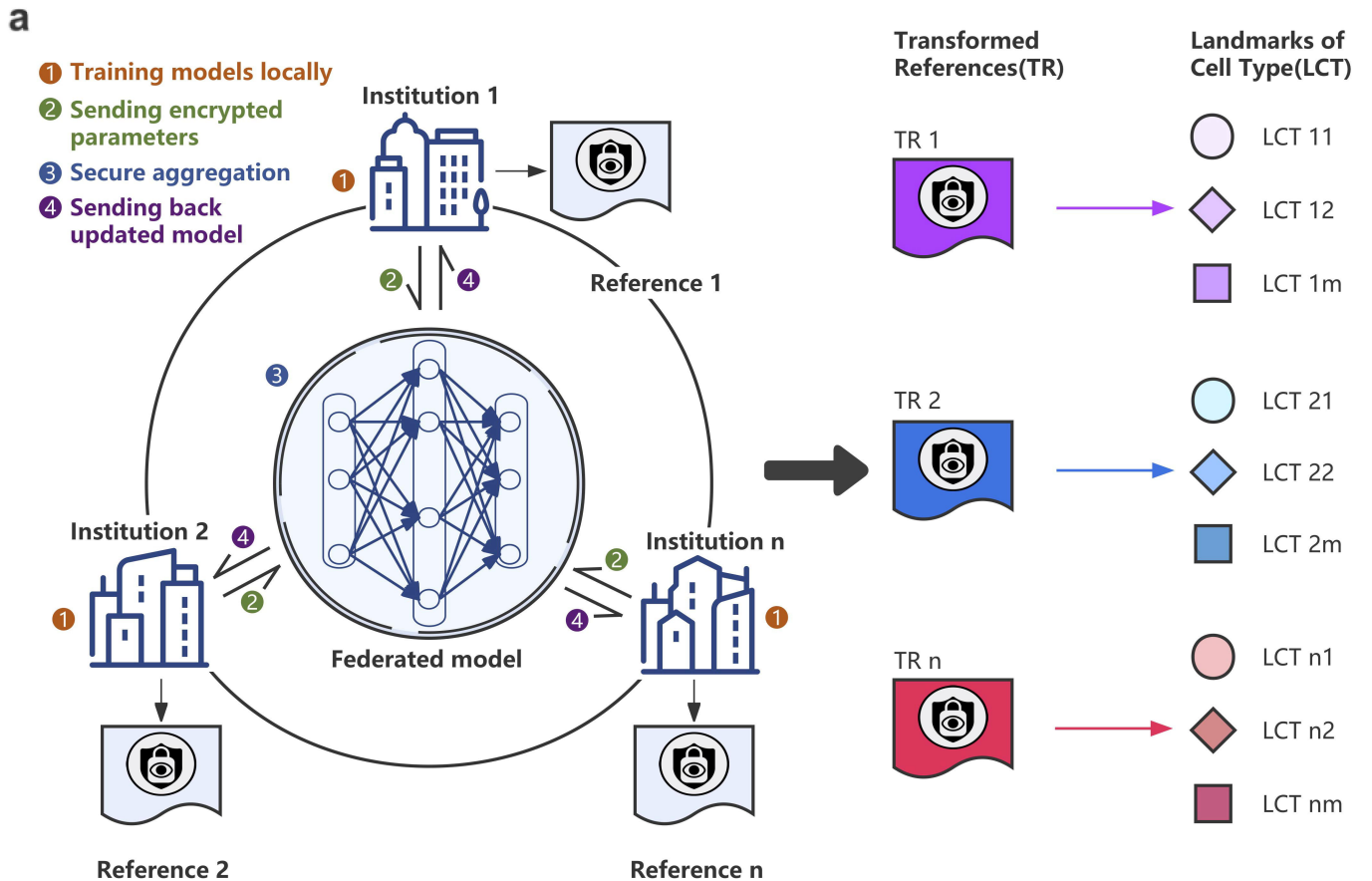
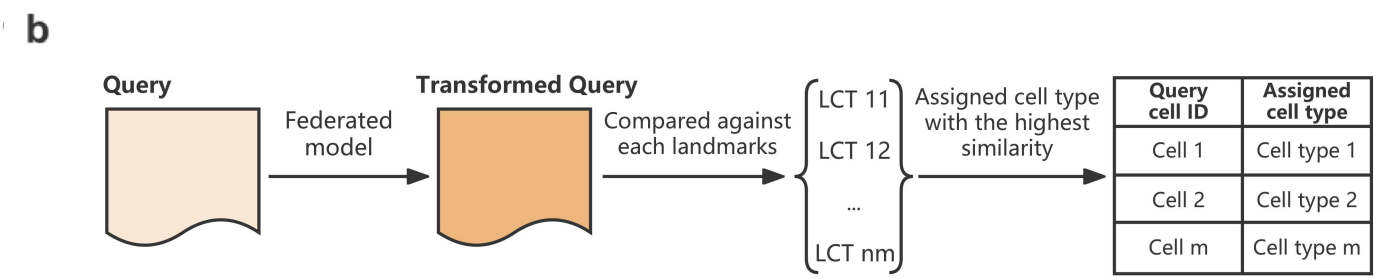
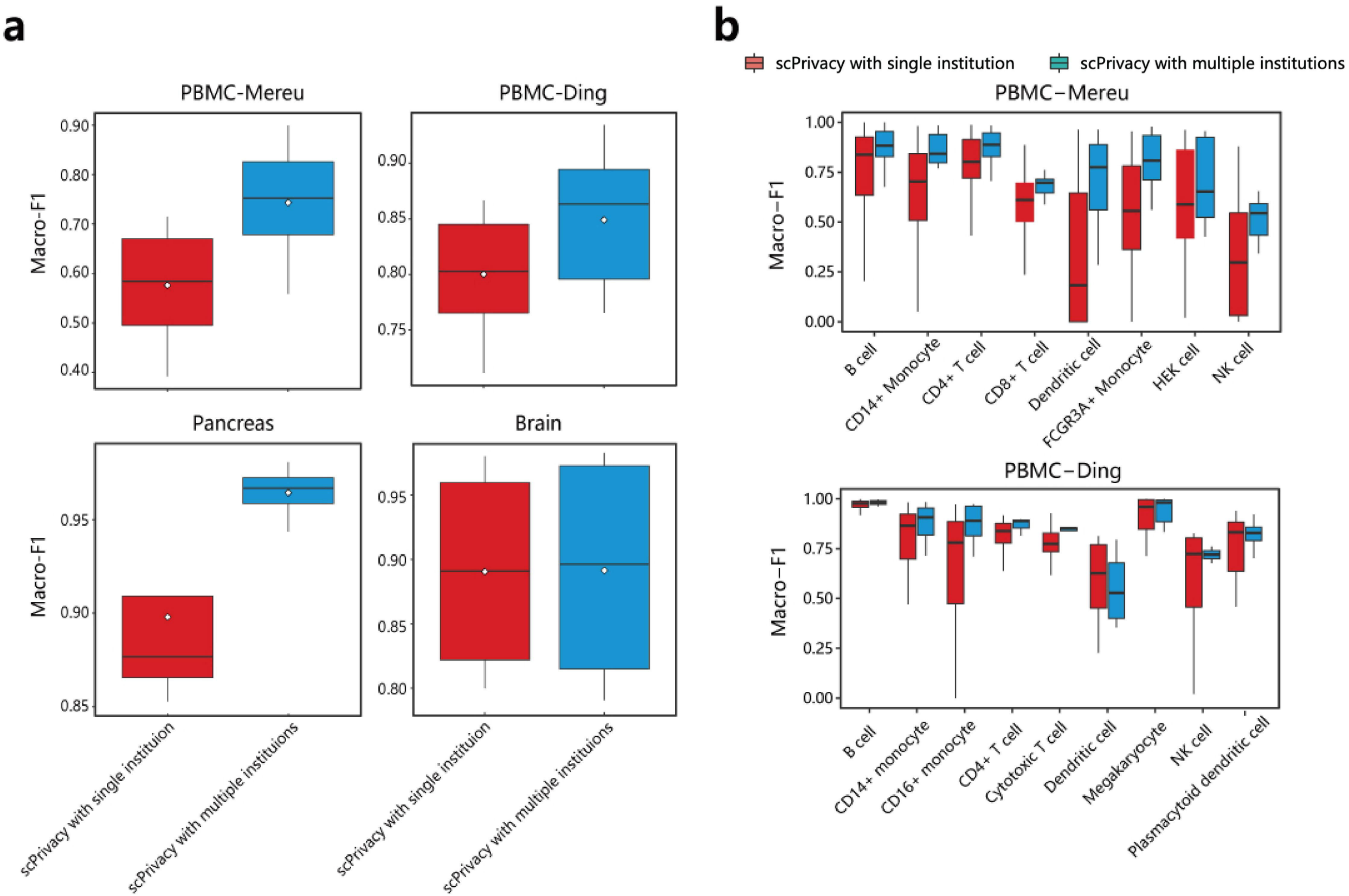
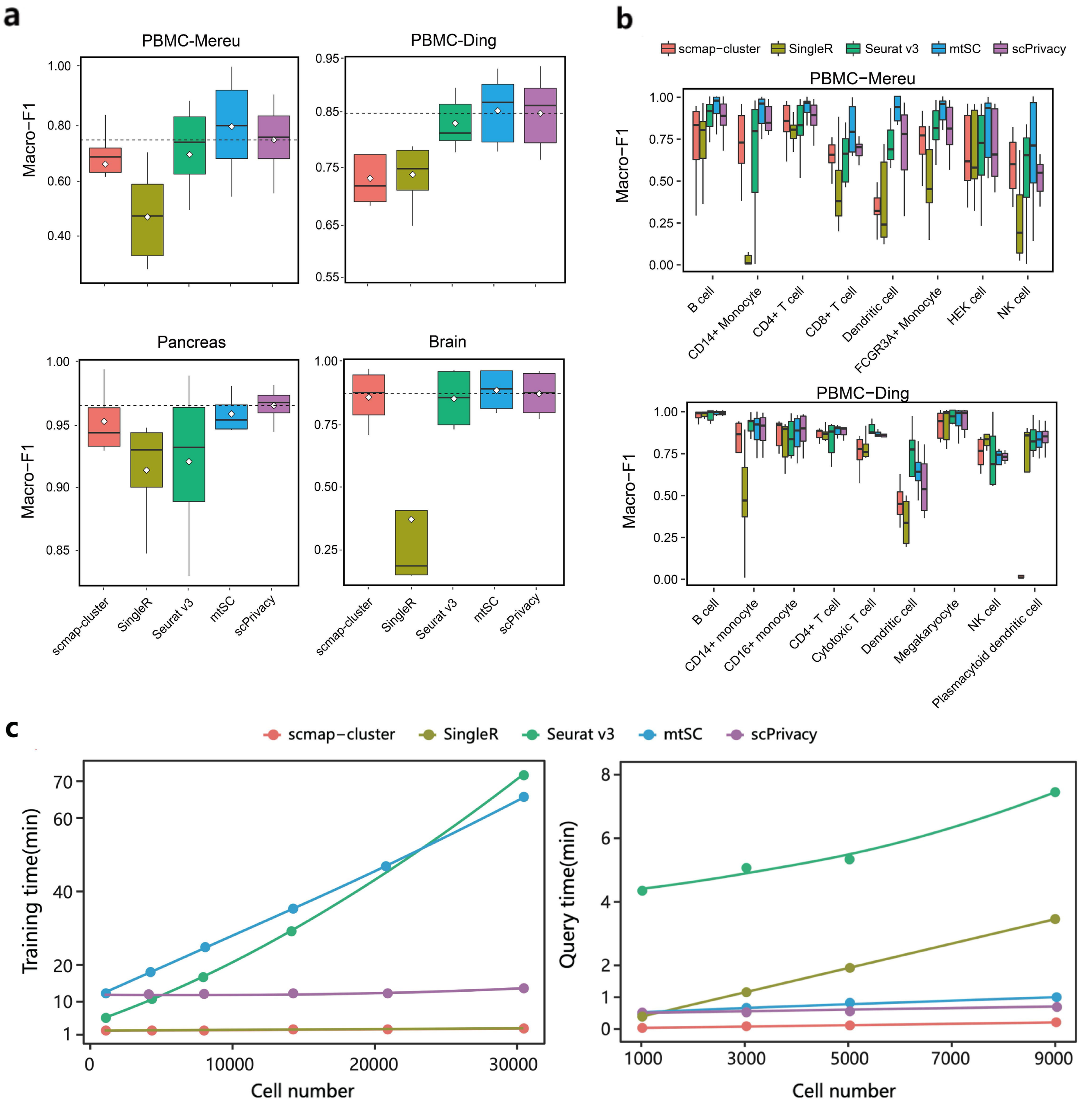
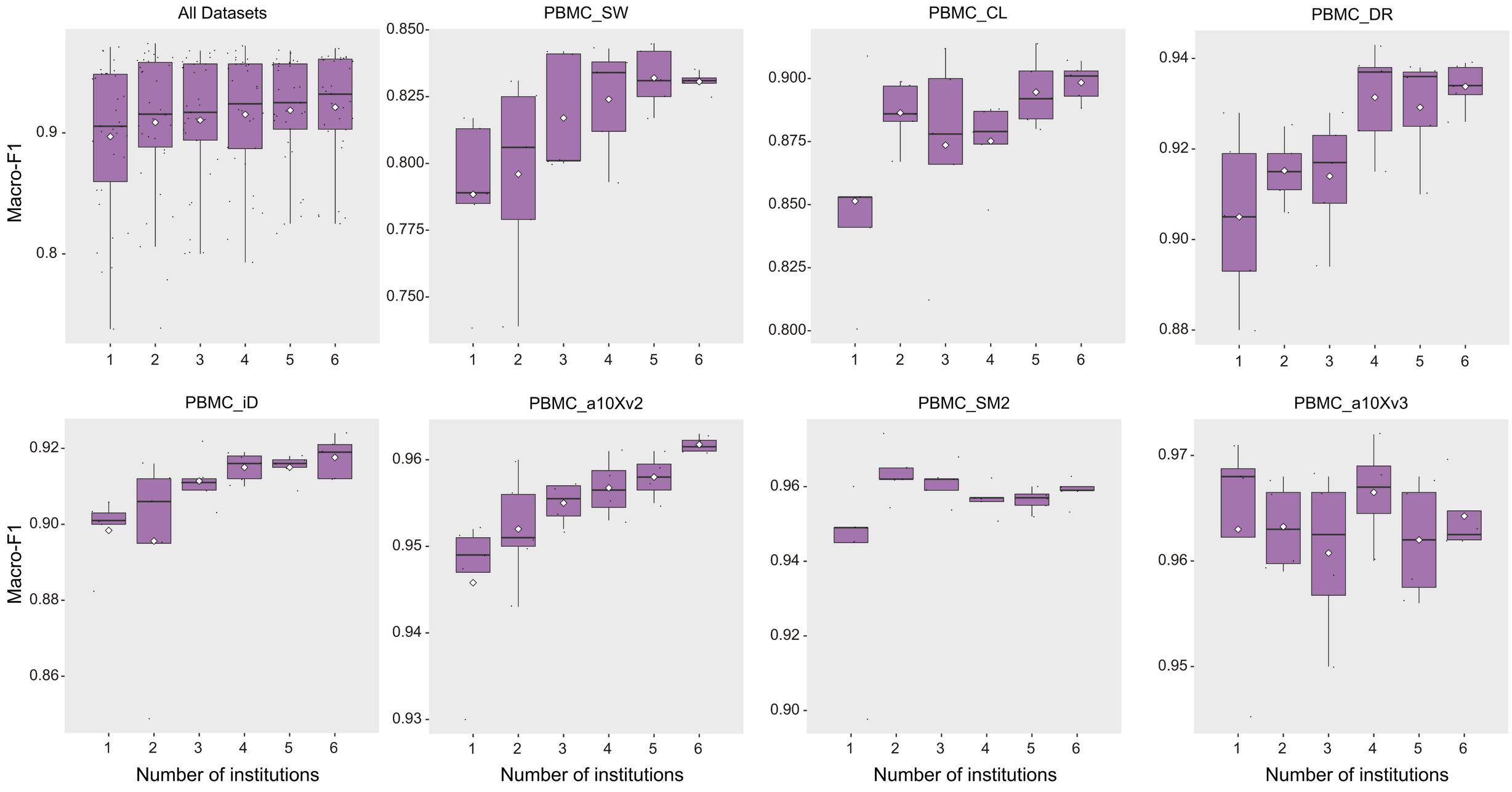

a
1 Training models locally
2 Sending encrypted parameters
3 Secure aggregation
4 Sending back updated model
Institution 1
Reference 1
Institution 2
Reference 2
Institution n
Reference n
Federated model
Transformed References(TR)
Landmarks of Cell Type(LCT)
TR 1
LCT 11
LCT 12
LCT 1m
TR 2
LCT 21
LCT 22
LCT 2m
TR n
LCT n1
LCT n2
LCT nm


b
Query
Federated model
Transformed Query
Compared against each landmarks
LCT 11
LCT 12
...
LCT nm
Assigned cell type with the highest similarity
Query cell ID
Assigned cell type
Cell 1
Cell type 1
Cell 2
Cell type 2
Cell m
Cell type m


a
PBMC-Mereu
PBMC-Ding
Pancreas
Brain
Macro-F1
scPrivacy with single instituion
scPrivacy with multiple instituions
b
scPrivacy with single institution
scPrivacy with multiple institutions
PBMC−Mereu
Macro−F1
B cell
CD14+ Monocyte
CD4+ T cell
CD8+ T cell
Dendritic cell
FCGR3A+ Monocyte
HEK cell
NK cell
PBMC−Ding
CD14+ monocyte
CD16+ monocyte
Cytotoxic T cell
Megakaryocyte
Plasmacytoid dendritic cell


a
PBMC-Mereu
PBMC-Ding
Pancreas
Brain
Macro−F1
scmap-cluster
SingleR
Seurat v3
mtSC
scPrivacy
b
scmap−cluster
SingleR
Seurat v3
mtSC
scPrivacy
PBMC−Mereu
PBMC−Ding
Macro−F1
B cell
CD14+ Monocyte
CD4+ T cell
CD8+ T cell
Dendritic cell
FCGR3A+ Monocyte
HEK cell
NK cell
CD14+ monocyte
CD16+ monocyte
Cytotoxic T cell
Megakaryocyte
Plasmacytoid dendritic cell
c
scmap−cluster
SingleR
Seurat v3
mtSC
scPrivacy
Training time(min)
Query time(min)
Cell number


All Datasets
PBMC_SW
PBMC_CL
PBMC_DR
PBMC_iD
PBMC_a10Xv2
PBMC_SM2
PBMC_a10Xv3
Macro-F1
Number of institutions