The right inferior frontal gyrus as pivotal node and effective regulator of the

basal ganglia-thalamocortical response inhibition circuit

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Abstract

The involvement of specific basal ganglia-thalamocortical circuits in response inhibition has

been extensively mapped in the last few decades. However, the pivotal brain nodes and

directed casual regulation within this inhibitory circuit in humans remains controversial. Here,

we capitalize on recent progress in robust and biologically plausible directed causal modelling

(DCM-PEB) and a large fMRI response inhibition dataset (n=218) to determine key nodes,

their causal regulation and modulation via biological variables (sex) and inhibitory

performance in the inhibitory control circuit encompassing the right inferior frontal gyrus

(rIFG), caudate nucleus (rCau), globus pallidum (rGP) and thalamus (rThal).The entire neural

circuit exhibited high intrinsic connectivity and an increasing rIFG inflow and its causal

regulation over the rCau and rThal during response inhibition. In addition, sex and

behavioral performance influenced the architecture of the regulatory circuits such that

women displayed increased rThal self-inhibition and decreased rThal to GP

modulation, while better inhibitory performance was associated with stronger rThal to

rIFG communication. Furthermore, control analyses did not reveal a similar key

communication in a left lateralized model. Together these findings indicate a pivotal role

of the rIFG as input and causal regulator of subcortical response inhibition nodes.

Keywords: response inhibition, basal ganglia, thalamus, inferior frontal gyrus,

effective connectivity, DCM, cognitive control, sex difference

Introduction

Animal models and human neuroimaging studies convergently demonstrated that inhibitory

control critically relies on highly specific basal ganglia-thalamocortical circuits (Alexander et

al., 1986, 1991; Alexander and Crutcher, 1990; Aron et al., 2007; Jahfari et al., 2019; Morein-

Zamir and Robbins, 2015; Pfeifer et al., 2022; Schall and Godlove, 2012; Stuphorn, 2015;

Verbruggen and Logan, 2009; Wei and Wang, 2016). Dysregulations in this circuit have been

implicated in disorders characterized by inhibitory control deficits, including addiction

(Klugah-Brown et al., 2020; Morein-Zamir and Robbins, 2015; Zhou et al., 2018), attention

deficit/hyperactivity (ADHD, Morein-Zamir et al., 2014; Sonuga-Barke, 2005), schizophrenia

(Camchong et al., 2006; Feng et al., 2018; Mamah et al., 2007) and Parkinson Disorder

(DeLong and Wichmann, 2015; Obeso et al., 2000). The key nodes within this response

inhibition circuitry have been extensively mapped with convergent evidence suggesting

critical contributions from the pre-supplementary motor area (pre-SMA) and lateral prefrontal

cortex (lPFC), particularly inferior frontal gyrus (IFG, Aron et al., 2003; Dambacher et al.,

2014; Hampshire et al., 2010; Maizey et al., 2020; Schaum et al., 2021; Verbruggen and

Logan, 2008; Zhang et al., 2017) as well as striatal regions, in particular the caudate and

putamen (Eagle et al., 2011; Ghahremani et al., 2012; Hampton et al., 2017; Kelly et al.,

2004; Ott and Nieder, 2019; Robertson et al., 2015; Robbins, 2007).

Anatomical and neurochemical studies further suggest that response inhibitory control

within this circuitry is modulated by dopaminergic and noradrenergic signaling (Bari et al.,

2011; Ghahremani et al., 2012; Li et al., 2020; Pfeifer et al., 2022; Rae et al., 2016; Robertson

et al., 2015). Dopamine receptor availability in the fronto-striatal circuits is significantly

related to inhibition-related neural responses (Ghahremani et al., 2012; Pfeifer et al., 2022)

and dopamine receptor availability in the lPFC modulates motor control via downstream

regulatory projections to the striatum (Ott and Nieder, 2019; Vijayraghavan et al., 2016).

Enhanced norepinephrine signaling facilitates response inhibition via modulation of the IFG

and its connections with the striatum (Chamberlain et al., 2009; Rae et al., 2016), while the

dorsal striatum represents an important locus of dopaminergic control of response inhibition

(Ghahremani et al., 2012; Robertson et al., 2015) and the IFG plays an important role in top-

down control of the basal ganglia regions (Buschman and Miller, 2014; Hampshire et al.,

2010; Jahfari et al., 2012; Kim, 2014; Puiu et al., 2020; Renteria et al., 2018; Schaum et al.,

2020; Tops and Boksem, 2011). In the basal ganglia-thalamocortical model of response

inhibition (Alexander et al., 1986, 1991; Alexander and Crutcher, 1990) the thalamus relays

information between the basal ganglia and cortex (Collins et al., 2018; Haber and Mcfarland,

2001; Haber and Calzavara, 2009; McFarland and Haber, 2002) - thus facilitating response

inhibition and performance monitoring (Bosch-Bouju et al., 2013; Huang et al., 2018;

Saalmann and Kastner, 2015; Tanaka and Kunimatsu, 2011) - via dense reciprocal

connections with the basal ganglia and PFC (Guillery, 1995; Phillips et al., 2021; Xiao et al.,

2009; Tanaka and Kunimatsu, 2011).

Convergent evidence from human lesion studies and neuroimaging meta-analyses

demonstrates a right-lateralized inhibitory control network encompassing the right IFG

(rIFG), right caudate nucleus (rCau), right globus pallidum (rGP) and right thalamus (rThal)

(Aron et al., 2003; Chevrier et al., 2007; Garavan et al., 1999; Hung et al., 2018; Jahfari et al.,

2011; Thompson et al., 2021). However, while extensive research has highlighted the critical

role of these regions within a right-lateralized inhibitory control circuitry, the causal

information flow and critical contribution of single nodes within this network have not been

determined.

We therefore capitalized on a novel dynamic causal modelling (DCM) approach based

on a priori specification of biologically and anatomically plausible models which allows

estimation of directed causal influences between nodes and their modulation by changing task

demands (Friston et al., 2003; Stephan et al., 2010) in the largest sample to-date (n=218).

DCM further allows comparison of modulatory effective connectivity strength across

different experimental conditions using Bayesian contrasts (Dijkstra et al., 2017) and in

combination with the recently developed Parametrical Empirical Bayes (PEB) hierarchical

framework (DCM-PEB method) allows modeling of both commonalities and differences in

effective connectivity between subjects e.g. to determine the neurobiological basis of sex and

behavioral performance variations (Friston et al., 2016; Zeidman et al., 2019a; Zeidman et al.,

2019b).

To determine the causal information flow and critical nodes in the basal ganglia-

thalamocortical circuits and whether these are modulated by biological factors (i.e. sex) and

show functional relevance in terms of associations with performance we capitalized on DCM-

PEB in combination with functional magnetic resonance imaging (fMRI) data collected in a

large sample of healthy individuals (n=218) during a well-established response inhibition

paradigm (emotional Go/NoGo task, see also Zhuang et al., 2021). To unravel the key nodes

and causal influences within the inhibitory control network, we firstly estimated the effective

connectivity between and within key regions involved in response inhibitory control within

the rIFG-rCau-rGP-rThal functional circuit (right lateralized model) and secondly estimated

sex differences and behavioral performance effects on connectivity parameters. To validate

the hemispheric asymmetry of the inhibitory control network, an identical model of nodes

was tested in the left hemisphere (left lateralized model).

Given convergent evidence on a pivotal role of the right IFG in mediating top-down

cortical-subcortical control during response inhibition (Aron et al., 2003; Dambacher et al.,

2014; Hampshire et al., 2010; Maizey et al., 2020), we predicted a greater modulatory effect

on rIFG and its directed connectivity to both rCau and rThal in the NoGo compared to Go

condition. Additionally, based on previous findings we expected a modulation of the key

pathways by biological (i.e. sex, Li et al., 2006; Ribeiro et al., 2021; Sjoberg and Cole, 2018)

and performance variations (Chang et al., 2020; Jahfari et al., 2011; Wei and Wang, 2016; Xu

et al., 2016) with better response inhibition being associated with stronger causal regulation in

the inhibition circuitry. Finally, we hypothesized a different causal structure for the left and

right models given the hemispheric asymmetry in the inhibitory network (Aron et al., 2003;

Chevrier et al., 2007; Hung et al., 2018; Jahfari et al., 2011; Thompson et al., 2021).

Results

Behavioral Results

The two-way repeated-measures ANOVA on accuracy found a significant main effect of

inhibition ($F(1,115)=21.73$, $p<0.001$, $\eta_p^2=0.16$), with a higher accuracy for Go compared to

No Go trials (Go trials: mean± SEM=98.47% ±0.31, No Go trials: mean ±SEM=70.34%

±1.44, Cohen's d=2.48). No sex-differences were found for accuracy or reaction times

(ps>0.18).

BOLD Activation (GLM) Analysis

Examination of domain general inhibition (contrast: NoGo>Go) revealed a widespread

fronto-parietal cortical and thalamo-striatal subcortical network including the IFG, striatal,

pallidal and thalamic regions (**Figure 1 and Table 1**) during response inhibition. Group-level

peaks in the rIFG, rCau, rGP and rThal were selected as centers of the ROIs for model testing

(**Figure 2a**). No significant sex difference were observed in BOLD activation.

Figure 1. Brain activation maps for general response inhibition on whole

brain level (contrast: NoGo > Go; p < 0.05 FWE, peak level). FWE, family-

wise error; L, left; R, right.

Table 1. Activation and peak values for key regions included in the right model

Regions	Cluster K	Coordinates			t value
		X	Y	Z	
rIFG	611	51	12	18	21.40
rCau	144	15	-3	15	13.61
rGP	63	21	3	9	12.43
rThal	340	15	-6	12	14.30

Note: These clusters survived from the overlay between image masks of corresponding

regions defined by Human Brainnetome Atlas ([84]) and group level brain activation

maps (peak level, $p_{FWE} < 0.05$) and thus served as regions of interest combined with

the individual peak location search on the individual level. FWE, family-wise error;

Cau, caudate nucleus; GP, global pallidum; IFG, inferior frontal gyrus; r, right; Thal,

thalamus.

Causal Connectivity (DCM) Analysis

For the A matrix, the diagonal cells represent self-connection which are unitless log scaling

parameters and were multiplied with the default value of -0.5Hz (Zeidman et al., 2019a).

Positive values indicate increased self-inhibition due to task condition and decreased

responsivity to the inputs from the other regions of the network, while negative values

indicate decreased self-inhibition and increased responsivity to the inputs from other nodes of

the network (Zeidman et al., 2019a). Our findings revealed negative self-inhibition values for

the rIFG, rCau and rThal but a positive value for the rGP (**Figure 2b, 2f**), indicating that the

GP increased self-connection while the other nodes increased interaction with other nodes in

the network.

Figure 2. Location of regions included in the right model and group-level

connectivity parameters. (a) Location of regions included in the right model.

The A matrix: intrinsic connectivity across all experimental conditions (b, f).

The B matrix: modulatory effect on effective connectivity between regions

and self-inhibitions from NoGo (c, g) and Go condition (d, h). The C matrix:

Driving inputs in ROIs in the NoGo and Go condition (e, i). Values in

matrices reflect the connectivity parameters. Parameters with stronger

evidence (posterior probability > 95%) are presented and subthreshold

parameters are marked with "n.s.".

For the off-diagonal cells in the A matrix, the values (in Hz) reflect the rate of change in

the activity of the target region caused by the source region per second. Positive values reflect

excitatory effects while negative values indicate inhibitory effects. In the forward direction

(e.g. rIFG‑rThal‑rGP‑rCau‑rIFG), we found a significant negative connectivity from rIFG to

rThal and positive connectivity from rThal to rGP as well as rCau to rIFG. In the backward

direction (e.g. rIFG‑rCau‑rGP‑rThal‑rIFG), rIFG exhibited a negative inhibitory influence

onto rCau, alongside an excitatory connection from rCau to rGP and rGP to rThal (**Figure 2b,**

2f). Although the connectivity from rThal to rIFG was not significant, a weak evidence

(posterior probability=57%) for this connection was observed with a more lenient threshold.

Values in the B matrix represent the rate of change, in Hz, in the connectivity from

source area to target area induced by the experimental conditions (Zeidman et al., 2019a).

During inhibitory control (NoGo condition) the rIFG exerted a negative influence onto the

rCau and rThal whereas the rGP exerted a negative influence on the rCau (**Figure 2c, 2g**). In

addition, we found negative self-inhibition values in both rCau and rThal respectively. During

the Go condition a negative influence of the rIFG on both rCau and rThal was observed

(**Figure 2d, 2h**), while the positive influence was observed from the rGP to rCau and from

rThal to rIFG. Moreover, we found a positive self-inhibition value in rIFG and a negative

value in rCau. A Bayesian contrast (NoGo > Go) allowed us to compare the connectivity

strength modulation during the different experimental conditions and revealed a very strong

evidence (posterior probability >99%) that the causal influence of the rIFG to both, the rCau

and rThal was stronger during inhibitory control (NoGo vs Go condition). This reflects that

response inhibition critically requires a causal top-down cortical-subcortical regulation via the

right IFG. We additionally found a very strong evidence (posterior probability >99%) for a

considerably stronger inhibitory connectivity from rGP to rCau in the NoGo compared to Go

condition.

The C matrix represents the rate of change in neural response of one brain region due to

the driving input from an experimental condition (Zeidman et al., 2019a). During inhibitory

control (NoGo) all regions (rIFG, rCau, rGP and rThal) exhibited excitatory driving input

while during the Go condition only the rIFG exhibited excitatory input (**Figure 2e, 2i**).

Bayesian contrasts directly comparing the conditions (NoGo > Go) demonstrated an

increasing driving input specifically in the rIFG during engagement of cognitive control

(NoGo > Go condition) with a 100% posterior probability.

Sex Differences in Connectivity Parameters

Examining sex effects on intrinsic connectivity showed a negative influence from rThal to

rGP in female compared to male subjects across all experimental conditions (**Figure 3a**). For

the modulatory effects on connectivity, we found a greater self-inhibition in rThal in female

than male subjects in the NoGo condition (**Figure 3b**). This suggests that for female subjects,

rThal exhibits reduced sensitivity to inputs from the other regions of the selected network

during response inhibition.

Figure 3. Sex effect on connectivity parameters in terms of A matrix and B

matrix. (a) For intrinsic connectivity in A matrix, female subjects showed a

more negative influence from rThal to rGP compared to male subjects. (b) In

the NoGo condition, there is a greater self-inhibition in rThal in female than

male subjects in terms of B matrix. Parameters with stronger evidence

(posterior probability > 95%) are presented.

Brain Behavior Associations: Inhibitory Behavioral Performance and Connectivity

Parameters

Examining associations between inhibitory performance on the behavioral level (NoGo

performance) and connectivity parameters revealed very strong evidence (posterior

probability > 99%) that NoGo accuracy was positively associated with the directed

connectivity from rThal to rIFG.

DCM Analyses in the Left Hemisphere

To further validate the hemispheric asymmetry of the inhibitory control network, an identical

model for the left hemisphere including lIFG, lCau, lGP and lThal was tested. In contrast to

the right model, no directed influences from IFG to subcortical regions were observed in

terms of matrix A in the left model, and the hemispheric models different in terms of

inhibition induced connectivity changes and differed in terms of the driving inputs. The

different causal structure in the left and right model indicated a hemispheric asymmetry in the

inhibition network (details see **Supplementary Materials Figure S3**). Additional Bayesian

analyses confirmed the lack of a robust cortical-subcortical pathway in the left hemisphere

(**Supplementary Material**).

Discussion

We capitalized on a combination of recent progress in biologically plausible causal

hierarchical modelling (DCM-PEB) and a comparably large fMRI response inhibition dataset

to determine causal information flow and key nodes within the extensively described basal

ganglia-thalamocortical response inhibition circuits (Alexander et al., 1986, 1991; Alexander

and Crutcher, 1990; Aron et al., 2007; Jahfari et al., 2019; Morein-Zamir and Robbins, 2015;

Pfeifer et al., 2022; Schall and Godlove, 2012; Stuphorn, 2015; Verbruggen and Logan, 2009;

Wei and Wang, 2016). Our neurocomputational model successfully validated a right-

lateralized inhibitory control causal circuit and the best model showed significant intrinsic

connectivity within this functional loop and captured an increasing causal influence of the

cortical rIFG node on both the rCau and rThal as well as from the rGP to the rCau during

inhibition. Direct comparison between different experimental conditions (e.g. NoGo and Go)

revealed enhanced input into rIFG in terms of matrix C and increased connectivity from rIFG

to rCau and rThal in the NoGo compared to the Go condition in terms of matrix B, suggesting

a higher engagement of causal top-down cortical-to-subcortical control via the rIFG during

inhibitory control. Although no sex differences were observed in inhibitory performance or

BOLD activation, females exhibited decreased intrinsic connectivity from rThal to rGP and

increased self-inhibition in rThal during the NoGo condition as compared to males. This

indicates that similar behavioral performance in response inhibition might be mediated by

different brain processes in men and women, particularly in thalamic loops. Moreover, a

higher NoGo response accuracy was associated with stronger causal information flow from

the rThal to rIFG in the NoGo condition, suggesting a particular behavioral inhibitory

relevance of this pathway. Finally, our findings showed different left and right model

structures, suggesting a hemispheric asymmetry in the inhibitory control network and

confirming a critical role of the rIFG in implementing response inhibition. Together these

findings identified a pivotal role of the rIFG and its effective connectivity with the rCau/rThal

within the basal ganglia-thalamocortical circuit during response inhibition.

Causal modelling successfully determined a right lateralized inhibitory control causal

circuit encompassing the rIFG, rCau, rGP and rThal (Aron et al., 2003; Chevrier et al., 2007;

Hung et al., 2018; Jahfari et al., 2011; Thompson et al., 2021). In terms of the A matrix, a

significant rIFG-rCau-rGP-rThal loop was observed with rIFG exhibiting a negative influence

onto rThal, alongside a positive information flow from from rThal to rGP and rCau to rIFG in

the forward direction. In the backward direction, we found significant negative connectivity

from rIFG to rCau and positive connectivity from rCau to rGP as well as rGP to rThal. A

more lenient threshold additionally revealed rThal to rIFG connections (posterior probability

= 57%). Importantly, accounting for behavioral task context revealed a significant positive

modulatory effect on rIFG in both NoGo and Go condition in terms of matrix C which was

considerably stronger during response inhibition. The direct driving inputs into the rIFG are

in line with its role in top-down target detection and attentional control in the context of

response inhibition (Hampshire et al., 2010; Krämer et al., 2013) and indicate that the rIFG

represents the key regulator of other nodes. In line with this hypothesis the best model in

terms of matrix B revealed strong evidence for causal effective connectivity from the rIFG to

both rCau and rThal during response inhibition (posterior probability >95%). This inhibitory

pathway is consistent with previous reports on negative coupling between the rIFG and

striatal regions during behavior control (Behan et al., 2015; Diekhof and Gruber, 2010).

Notably, direct comparison using Bayesian contrast revealed a very strong evidence (posterior

probability >99%) for increased modulatory connectivity from rIFG to rCau and rThal in the

NoGo condition compared to the Go condition, suggesting the rIFG driven engagement of

cortical-to-subcortical top-down control during response inhibition. Previous animal models

and human neuroimaging meta-analyses have consistently identified the rIFG, as a key region

implicated in dopaminergic and noradrenergic modulated inhibitory regulation (Bari et al.,

2011; Hauber, 2010; Ott and Nieder, 2019; Pfeifer et al., 2022; Terra et al., 2020;

Vijayraghavan et al., 2016; Zhukovsky et al., 2021) in particular during motor control and

inhibition (Aron et al., 2003; Chamberlain and Sahakian, 2007; Puiu et al., 2020; Xu et al.,

2016), while both, fronto-striatal and fronto-thalamic projections have been extensively

involved in response inhibition (Ahissar and Oram, 2015; Bosch-Bouju et al., 2013;

Marzinzik et al., 2008; Phillips et al., 2021; Schmitt et al., 2017; Sommer, 2003; Tanaka and

Kunimatsu, 2011).

In addition to the cortical-subcortical pathways significant excitatory connectivity was

observed from the rGP to rCau during the Go condition and switched to inhibitory

connectivity when response inhibition was required during the NoGo condition. Direct

comparison confirmed a considerably stronger inhibitory influence of the rGP on the rCau

during response inhibition (posterior probability >99%), suggesting that communication

between basal ganglia nodes is crucial for context-appropriate behavioral response control.

The involvement of this pathway is in line with extensive neurophysiological evidence

showing that GABA inhibitory projections from the external segment of the GP to the

striatum play an essential role in cancelling a planned response when it is inappropriate

(Mallet et al., 2016; Wei and Wang, 2016) (but see also subthalamic nucleus to substantia

nigra pars reticulata pathways in Hikosaka et al., 2006; Mallet et al., 2016).

With respect to sex differences we observed that females exhibited decreased intrinsic

connectivity from rThal to rGP and increased modulation by NoGo condition on self-

inhibition in rThal compared to male subjects in the absence of performance differences.

While previous findings on sex differences in response inhibition remained inconsistent

(Chung et al., 2020; Gaillard et al., 2020; Gaillard et al., 2021; Li et al., 2006; Ribeiro et al.,

2021; Sjoberg and Cole, 2018) the present findings suggest that our model was sensitive to

biological variables and that separable information processes may underly response inhibition

in men and women (see also Chung et al., 2020; Li et al., 2006). The functional relevance of

the identified pathways was further underscored by a significant association between response

inhibition performance and the causal influence from the rThal to rIFG in the NoGo condition

demonstrating that this pathway involved in motor inhibition critically mediates behavioral

success during inhibition (Wei and Wang, 2016).

Finally, our modelling tests confirmed a hemispheric asymmetry and support the critical

role of right IFG circuit in response inhibition (Hung et al., 2018; Jahfari et al., 2011; Maizey

et al., 2020). The different causal structures suggest a strong cortical-subcortical intrinsic

connectivity and rIFG control on the right side. The left model revealed a different causal

structure and null hypothesis tests showed moderate evidence for the difference between

NoGo and Go condition's modulatory effects on effective connectivity from lIFG to lCau and

to rThal (e.g. lIFG to lCau: Bayes factor = 5.47; lIFG to lThal: Bayes factor = 8.20).

Response inhibition impairments have been observed in several disorders and

identification of the rIFG as critical input and top-down regulator for response inhibition

opens new targets for regional or connectivity-based neuromodulation such as real-time

neurofeedback which has been established for these regions (Li et al., 2019; Weiss et al.,

2022; Zhao et al., 2019). For instance, rIFG and response inhibition deficits have been

determined in ADHD (Clark et al., 2007; Morein-Zamir et al., 2014) and targeting the rIFG in

ADHD may be a promising treatment.

There are several limitations in the current study. First, in line with our main aim we did

not account for emotional valence in the DCM model which may affect response

inhibition(Schimmack and Derryberry, 2005). Second, we focused on specific nodes that

were based on established basal ganglia-thalamocortical circuits proposed by Alexander

(Alexander et al., 1986, 1991; Alexander and Crutcher, 1990) (see also neuroimaging meta-

analysis (Hung et al., 2018). Other regions such as the STN (Aron et al., 2016; Aron and

Poldrack, 2006; Chen et al., 2020) could be integrated in future studies.

In conclusion, our findings demonstrated a critical role of the rIFG as well as top-down

cortical-subcortical control from the rIFG to rCau and rThal in response inhibition. The nodes

and pathways of the model were sensitive to biological and performance variations. The

nodes and pathways may represent promising targets to improve response inhibition in mental

disorders.

Materials and Methods

Participants

N=250 healthy right-handed participants were enrolled in the current study and underwent a

validated Go/NoGo fMRI paradigm. The data has been previously used to examine undirected

functional connectivity within domain-general and emotion-specific inhibitory brain systems

(Zhuang et al., 2021) and was part of larger neuroimaging project examining pain empathy

(Li et al., 2018; Zhou et al., 2020), emotional face memory (Liu et al., 2022) and mirror

neuron processing (Xu et al., 2022). After quality assessment n=218 subjects were included

(104 males, **Supplementary Materials**). The study was approved by the local ethics

committee and in accordance with the latest version of the Declaration of Helsinki.

Response Inhibition Paradigm

A validated mixed event-related block design linguistic emotional Go/NoGo fMRI paradigm

was employed (Goldstein et al., 2007; Protopopescu et al., 2005, details see Zhuang et al.,

2021). Participants were required to make responses as accurately and quickly as possible

based on orthographical cues, i.e. words were presented in normal or italic font. For normal

font words subjects were instructed to perform a button-press (Go trials), while inhibiting

their response to words presented in italic font (NoGo trials). Positive, negative and neutral

words were included, however, given that the present study aimed to examine the causal

influence within the general inhibition network and to increase statistical power in this respect

the different emotional contexts were not further accounted for in the DCM analysis. Stimuli

were presented in 2 runs and each run included 12 blocks (6 blocks: Go; 6 blocks: NoGo).

Each Go block encompassed 18 normal font words (100% Go trials) while each NoGo block

encompassed 12 normal font words (66.7% Go trials) and 6 italicized font words (33.3%

NoGo trials). Further details in (Zhuang et al., 2021) and **Supplementary Materials**.

Behavioral Data Analysis

In our previous study we demonstrated that subjects exhibited more errors during inhibitory

control (i.e., NoGo>Go) as well as faster responses in positive Go contexts and lower

accuracy in positive NoGo contexts (Zhuang et al., 2021). Given that sex-differences were

examined in the DCM model the present analyses additionally examined sex-differences on

accuracy and reaction times (**Supplementary Materials)**. Given age-related effects on

inhibition (Rey-Mermet et al., 2018; Rubia et al., 2007) age was included as covariate.

MRI Data Acquisition and Preprocessing

MRI data were collected on a 3T MRI system using standard sequences and were initially

preprocessed using validated protocols in SPM 12 (details see **Supplementary Materials**)

GLM Analysis

An event-related general linear model (GLM) was established in SPM12. To examine domain

general inhibitory control (irrespective of emotional context) the overarching inhibitory

control contrast was modelled (e.g. all NoGo>all Go trials) and convolved with the canonical

384 hemodynamic response function (HRF). Six head motion parameters were included in the

385 design matrix to control movement-related artifacts and a high-pass filter (1/128Hz) was

386 applied to remove low frequency components. The contrast of interest (contrast: NoGo>Go)

387 was created and subjected to one-sample t-test at the second level. In line with previous

388 studies (Aron et al., 2003; Chevrier et al., 2007; Hung et al., 2018; Jahfari et al., 2011;

389 Thompson et al., 2021), group-level (contrast: NoGo>Go) peaks in the IFG, Cau, GP and Thal

390 within the identified general inhibition network were then used to define individual-specific

391 regions of interest (ROIs) for the DCM analysis. Additionally, a two-sample t-test was

392 conducted (contrast: NoGo>Go) to examine sex-dependent effects on the response inhibition

393 network. Analyses were corrected for multiple comparisons using a conservative peak-level

394 threshold on the whole brain level (p<0.05 family-wise error, FWE).

395 **Dynamic Causal Modeling and Node Definition**

396 A DCM analysis was employed to determine directed causal influences according to the

397 circuitry model proposed by Alexander et al. (Alexander et al., 1986, 1991; Alexander and

398 Crutcher, 1990). The DCM approach allows to construct a realistic neuronal model of

399 interacting regions and to predict the underlying neuronal activity from the measured

400 hemodynamic response (Friston et al., 2003; Stephan et al., 2007). To this end directed causal

401 influences between the key regions including IFG, Cau, GP and Thal in the basal ganglia-

402 thalamocortical loop and their modulation via experimental manipulations (engagement of

403 motor inhibitory control) were examined. In line with previous neuroimaging studies and

404 meta-analyses demonstrating a right-lateralized inhibition model (right model) encompassing

the rIFG, rCau, rGP, rThal (Aron et al., 2003; Chevrier et al., 2007; Hung et al., 2018; Jahfari

et al., 2011; Thompson et al., 2021) our main hypothesis testing focused on the right

lateralized network. To further validate the hemispheric asymmetry of the inhibitory control

network an identical model was tested for the left hemisphere including the lIFG, lCau, lGP,

and lThal. In line with previous studies, we combined atlas-based masks (Human

Brainnetome Atlas, Fan et al., 2016) with group-level and individual level activity maps to

generate the corresponding nodes (Fernández-Espejo et al., 2015; Holmes et al., 2021; Qiao et

al., 2020; Van Overwalle et al., 2020).

Model Specification and Estimation

A two-step DCM analysis was performed using the DCM-parametric empirical Bayes (PEB)

approach (Zeidman et al., 2019a; Zeidman et al., 2019b). On the first-level, time-series from

four ROIs (rIFG, rCau, rGP, rThal) were extracted. A full DCM model was specified for each

subject and all connectivity parameters in both forward (e.g. rIFG-rThal-rGP-rCau-rIFG)

and backward (e.g. rIFG-rCau-rGP-rThal-rIFG) directions were estimated. We estimated

three key DCM parameters: (1) the A matrix reflecting all connections including forward and

backward connectivity between ROIs and self-inhibitions in each ROI, (2) the B matrix

representing modulatory effects of Go and NoGo condition on all connections, (3) the C

matrix representing the driving inputs into ROIs from Go and NoGo conditions separately.

Given that all inputs in the model were mean-centered, intrinsic connectivity in the A matrix

indicates mean effective connectivity independent of all experimental conditions. The model

was estimated using Variational Laplace (Friston et al., 2007). Further details **Supplementary**

Material. At the second (group) level, we constructed a PEB model over the first-level

estimated parameters. In accordance with previous studies (Bencivenga et al., 2021;

Rupprechter et al., 2020), we evaluated the explained variance by the model on the individual

level - higher values reflect better model inversion (Zeidman et al., 2019a) – and then we only

included subjects with >10% of explained variance in the PEB model. A total of 118 subjects

(56 males, age: mean ± SEM = 21.57 ± 0.21 years) were included for further analyses. The

differences on behavioral performance were examined between the excluded and included

subjects and no significant differences were found (all ps ≥ 0.23, for details see

Supplementary Material), suggesting no evidence of biased selection.

The primary aim of the present study was to establish a causal neurobiological model for

response inhibition and to determine the interaction between key players in this circuitry. To

evaluate the model three PEB analyses were carried out separately for A, B and C matrices.

Separate analyses examined sex and performance variations (details see **Supplementary**

Material).

Next, to identify the model that best represented our data, Bayesian Model Reduction

(BMR) was performed to compare the free energy of the full model with numerous reduced

models for which specific parameters were "switched off" (Friston et al., 2016). An automatic

greedy search procedure (iterative procedure) was employed to facilitate an efficient

comparison of thousands of models. In this procedure parameters which do not contribute to

free energy were pruned away. Next, Bayesian Model Average (BMA), performing a

weighted average of the parameters of each model, was calculated over the 256 models

obtained from the final iteration (Friston et al., 2016).

Finally, to compare the effective connection strength, especially the cortical-subcortical

connectivity and driving inputs into each region from different experimental conditions

(NoGo and Go condition), Bayesian contrasts (Dijkstra et al., 2017) were computed over

parameters from the B and C matrices. Group-level estimated parameters were thresholded at

posterior probability > 95% (indicating strong evidence, Kass and Raftery, 1995) based on

free energy.

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Author contributions

Qian Zhuang: Formal analysis; Investigation; Writing-original draft. **Lei Qiao**: Formal analysis; Validation. **Lei Xu**: Project administration. **Shuxia Yao**: Conceptualization; Supervision**. Shuaiyu Chen:** Formal analysis**. Xiaoxiao Zheng, Jialin Li, Meina Fu and Keshuang Li**: Project administration. **Deniz Vatansever and Stefania Ferraro:** Validation. **Keith M. Kendrick and Benjamin Becker:** Conceptualization; Funding acquisition; Project administration; Resources; Supervision; Validation; Writing.

496 **Declaration of Conflicting Interests**

497 The authors declared no conflicts of interest with their research, authorship or the publication

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References

Ahissar, E., and Oram, T. (2015). Thalamic relay or cortico-thalamic processing? Old

question, new answers. Cerebral Cortex *25*, 845-848. 10.1093/cercor/bht296.

Alexander, G.E., and Crutcher, M.D. (1990). Functional architecture of basal ganglia circuits:

neural substrates of parallel processing. Trends in neurosciences *13*, 266-271. 10.1016/0166-

2236(90)90107-l.

Alexander, G.E., Crutcher, M.D., and DeLong, M.R. (1991). Basal ganglia-thalamocortical

circuits: parallel substrates for motor, oculomotor,"prefrontal" and "limbic" functions.

Progress in brain research *85*, 119-146. 10.1016/S0079-6123(08)62678-3

Alexander, G.E., DeLong, M.R., and Strick, P.L. (1986). Parallel organization of functionally

segregated circuits linking basal ganglia and cortex. Annual review of neuroscience *9*, 357-

381. 10.1146/annurev.ne.09.030186.002041.

Aron, A.R., Durston, S., Eagle, D.M., Logan, G.D., Stinear, C.M., and Stuphorn, V. (2007).

Converging evidence for a fronto-basal-ganglia network for inhibitory control of action and

cognition. Journal of Neuroscience *27*, 11860-11864. 10.1523/JNEUROSCI.3644-07.2007.

10.1523/JNEUROSCI.3644-07.2007.

Aron, A.R., Fletcher, P.C., Bullmore, E.T., Sahakian, B.J., and Robbins, T.W. (2003). Stop-

signal inhibition disrupted by damage to right inferior frontal gyrus in humans. Nature

neuroscience *6*, 115-116. 10.1038/nn1003.

Aron, A.R., Herz, D.M., Brown, P., Forstmann, B.U., and Zaghloul, K. (2016).

Frontosubthalamic circuits for control of action and cognition. Journal of Neuroscience *36*,

11489-11495. 10.1523/JNEUROSCI.2348-16.2016.

Aron, A.R., and Poldrack, R.A. (2006). Cortical and subcortical contributions to stop signal

response inhibition: role of the subthalamic nucleus. Journal of Neuroscience *26*, 2424-2433.

10.1523/JNEUROSCI.4682-05.2006.

Bari, A., Mar, A.C., Theobald, D.E., Elands, S.A., Oganya, K.C., Eagle, D.M., and Robbins,

T.W. (2011). Prefrontal and monoaminergic contributions to stop-signal task performance in

rats. Journal of Neuroscience *31*, 9254-9263. 10.1523/JNEUROSCI.1543-11.2011.

Behan, B., Stone, A., and Garavan, H. (2015). Right prefrontal and ventral striatum

interactions underlying impulsive choice and impulsive responding. Human brain mapping

36, 187-198. 10.1002/hbm.22621.

Bencivenga, F., Sulpizio, V., Tullo, M.G., and Galati, G. (2021). Assessing the effective

connectivity of premotor areas during real vs imagined grasping: a DCM-PEB approach.

NeuroImage *230*, 117806. 10.1016/j.neuroimage.2021.117806.

Bosch-Bouju, C., Hyland, B.I., and Parr-Brownlie, L.C. (2013). Motor thalamus integration

of cortical, cerebellar and basal ganglia information: implications for normal and

parkinsonian conditions. Frontiers in computational neuroscience *7*, 163.

10.3389/fncom.2013.00163.

Buschman, T.J., and Miller, E.K. (2014). Goal-direction and top-down control. Philosophical

Transactions of the Royal Society B: Biological Sciences *369*, 20130471.

10.1098/rstb.2013.0471.

Camchong, J., Dyckman, K.A., Chapman, C.E., Yanasak, N.E., and McDowell, J.E. (2006).

Basal ganglia-thalamocortical circuitry disruptions in schizophrenia during delayed response

tasks. Biological psychiatry *60*, 235-241. 10.1016/j.biopsych.2005.11.014.

Chamberlain, S.R., Hampshire, A., Müller, U., Rubia, K., Del Campo, N., Craig, K.,

Regenthal, R., Suckling, J., Roiser, J.P., and Grant, J.E. (2009). Atomoxetine modulates right

inferior frontal activation during inhibitory control: a pharmacological functional magnetic

resonance imaging study. Biological psychiatry *65*, 550-555.

10.1016/j.biopsych.2008.10.014.

Chamberlain, S.R., and Sahakian, B.J. (2007). The neuropsychiatry of impulsivity. Current

opinion in psychiatry *20*, 255-261. 10.1097/YCO.0b013e3280ba4989.

Chang, J., Hu, J., Li, C.-S.R., and Yu, R. (2020). Neural correlates of enhanced response

inhibition in the aftermath of stress. Neuroimage *204*, 116212.

10.1016/j.neuroimage.2019.116212.

Chen, W., de Hemptinne, C., Miller, A.M., Leibbrand, M., Little, S.J., Lim, D.A., Larson,

P.S., and Starr, P.A. (2020). Prefrontal-subthalamic hyperdirect pathway modulates

movement inhibition in humans. Neuron *106*, 579-588. e573. 10.1016/j.neuron.2020.02.012.

Chevrier, A.D., Noseworthy, M.D., and Schachar, R. (2007). Dissociation of response

inhibition and performance monitoring in the stop signal task using event-related fMRI.

Human brain mapping *28*, 1347-1358. 10.1002/hbm.20355.

Chung, Y.S., Calhoun, V., and Stevens, M.C. (2020). Adolescent sex differences in cortico-

subcortical functional connectivity during response inhibition. Cognitive, Affective, &

Behavioral Neuroscience *20*, 1-18. 10.3758/s13415-019-00718-y.

Clark, L., Blackwell, A.D., Aron, A.R., Turner, D.C., Dowson, J., Robbins, T.W., and

Sahakian, B.J. (2007). Association between response inhibition and working memory in adult

ADHD: a link to right frontal cortex pathology? Biological psychiatry *61*, 1395-1401.

10.1016/j.biopsych.2006.07.020.

Collins, D.P., Anastasiades, P.G., Marlin, J.J., and Carter, A.G. (2018). Reciprocal circuits

linking the prefrontal cortex with dorsal and ventral thalamic nuclei. Neuron *98*, 366-379.

e364. 10.1016/j.neuron.2018.03.024.

Dambacher, F., Sack, A.T., Lobbestael, J., Arntz, A., Brugman, S., and Schuhmann, T.

(2014). A network approach to response inhibition: dissociating functional connectivity of

neural components involved in action restraint and action cancellation. European Journal of

Neuroscience *39*, 821-831. 10.1111/ejn.12425.

DeLong, M.R., and Wichmann, T. (2015). Basal ganglia circuits as targets for

neuromodulation in Parkinson disease. JAMA neurology *72*, 1354-1360.

10.1001/jamaneurol.2015.2397.

Diekhof, E.K., and Gruber, O. (2010). When desire collides with reason: functional

interactions between anteroventral prefrontal cortex and nucleus accumbens underlie the

human ability to resist impulsive desires. Journal of Neuroscience *30*, 1488-1493.

10.1523/JNEUROSCI.4690-09.2010.

Dijkstra, N., Zeidman, P., Ondobaka, S., van Gerven, M.A., and Friston, K. (2017). Distinct

top-down and bottom-up brain connectivity during visual perception and imagery. Scientific

reports *7*, 1-9. 10.1038/s41598-017-05888-8.

Eagle, D.M., Wong, J.C., Allan, M.E., Mar, A.C., Theobald, D.E., and Robbins, T.W. (2011).

Contrasting roles for dopamine D1 and D2 receptor subtypes in the dorsomedial striatum but

not the nucleus accumbens core during behavioral inhibition in the stop-signal task in rats.

Journal of Neuroscience *31*, 7349-7356. 10.1523/JNEUROSCI.6182-10.2011.

Fan, L., Li, H., Zhuo, J., Zhang, Y., Wang, J., Chen, L., Yang, Z., Chu, C., Xie, S., and Laird,

A.R. (2016). The human brainnetome atlas: a new brain atlas based on connectional

architecture. Cerebral cortex *26*, 3508-3526. 10.1093/cercor/bhw157.

Feng, C., Becker, B., Huang, W., Wu, X., Eickhoff, S.B., and Chen, T. (2018). Neural

substrates of the emotion-word and emotional counting Stroop tasks in healthy and clinical

populations: A meta-analysis of functional brain imaging studies. NeuroImage *173*, 258-274.

10.1016/j.neuroimage.2018.02.023.

Fernández-Espejo, D., Rossit, S., and Owen, A.M. (2015). A thalamocortical mechanism for

the absence of overt motor behavior in covertly aware patients. JAMA neurology *72*, 1442-

1450. 10.1001/jamaneurol.2015.2614.

Friston, K., Mattout, J., Trujillo-Barreto, N., Ashburner, J., and Penny, W. (2007). Variational

free energy and the Laplace approximation. Neuroimage *34*, 220-234.

10.1016/j.neuroimage.2006.08.035.

Friston, K.J., Harrison, L., and Penny, W. (2003). Dynamic causal modelling. Neuroimage

19, 1273-1302. 10.1016/s1053-8119(03)00202-7.

Friston, K.J., Litvak, V., Oswal, A., Razi, A., Stephan, K.E., Van Wijk, B.C., Ziegler, G., and

Zeidman, P. (2016). Bayesian model reduction and empirical Bayes for group (DCM) studies.

Neuroimage *128*, 413-431. 10.1016/j.neuroimage.2015.11.015.

Gaillard, A., Fehring, D.J., and Rossell, S.L. (2021). A systematic review and meta-analysis

of behavioural sex differences in executive control. European Journal of Neuroscience *53*,

519-542. 10.1111/ejn.14946.

Gaillard, A., Rossell, S.L., Carruthers, S.P., Sumner, P.J., Michie, P.T., Woods, W., Neill, E.,

Phillipou, A., Toh, W.L., and Hughes, M.E. (2020). Greater activation of the response

inhibition network in females compared to males during stop signal task performance.

Behavioural brain research *386*, 112586. 10.1016/j.bbr.2020.112586.

10.1073/pnas.96.14.8301.

Garavan, H., Ross, T., and Stein, E. (1999). Right hemispheric dominance of inhibitory

control: an event-related functional MRI study. Proceedings of the National Academy of

Sciences *96*, 8301-8306. 10.1073/pnas.96.14.8301.

Ghahremani, D.G., Lee, B., Robertson, C.L., Tabibnia, G., Morgan, A.T., De Shetler, N.,

Brown, A.K., Monterosso, J.R., Aron, A.R., and Mandelkern, M.A. (2012). Striatal dopamine

D2/D3 receptors mediate response inhibition and related activity in frontostriatal neural

circuitry in humans. Journal of Neuroscience *32*, 7316-7324. 10.1523/JNEUROSCI.4284-

11.2012.

Goldstein, M., Brendel, G., Tuescher, O., Pan, H., Epstein, J., Beutel, M., Yang, Y., Thomas,

K., Levy, K., and Silverman, M. (2007). Neural substrates of the interaction of emotional

stimulus processing and motor inhibitory control: an emotional linguistic go/no-go fMRI

study. Neuroimage *36*, 1026-1040. 10.1016/j.neuroimage.2007.01.056.

Guillery, R. (1995). Anatomical evidence concerning the role of the thalamus in

corticocortical communication: a brief review. Journal of anatomy *187*, 583.

Haber, S., and Mcfarland, N.R. (2001). The place of the thalamus in frontal cortical-basal

ganglia circuits. The Neuroscientist *7*, 315-324. 10.1177/107385840100700408.

Haber, S.N., and Calzavara, R. (2009). The cortico-basal ganglia integrative network: the role

of the thalamus. Brain research bulletin *78*, 69-74. 10.1016/j.brainresbull.2008.09.013.

Hampshire, A., Chamberlain, S.R., Monti, M.M., Duncan, J., and Owen, A.M. (2010). The

role of the right inferior frontal gyrus: inhibition and attentional control. Neuroimage *50*,

1313-1319. 10.1016/j.neuroimage.2009.12.109.

Hampton, W.H., Alm, K.H., Venkatraman, V., Nugiel, T., and Olson, I.R. (2017). Dissociable

frontostriatal white matter connectivity underlies reward and motor impulsivity. Neuroimage

150, 336-343. 10.1016/j.neuroimage.2017.02.021.

Hauber, W. (2010). Dopamine release in the prefrontal cortex and striatum: temporal and

behavioural aspects. Pharmacopsychiatry *43*, S32-S41. 10.1055/s-0030-1248300.

Hikosaka, O., Nakamura, K., and Nakahara, H. (2006). Basal ganglia orient eyes to reward.

Journal of neurophysiology *95*, 567-584. 10.1152/jn.00458.2005.

Holmes, E., Zeidman, P., Friston, K.J., and Griffiths, T.D. (2021). Difficulties with speech-in-

noise perception related to fundamental grouping processes in auditory cortex. Cerebral

Cortex *31*, 1582-1596. 10.1093/cercor/bhaa311.

Huang, A.S., Mitchell, J.A., Haber, S.N., Alia-Klein, N., and Goldstein, R.Z. (2018). The

thalamus in drug addiction: from rodents to humans. Philosophical Transactions of the Royal

Society B: Biological Sciences *373*, 20170028. 10.1098/rstb.2017.0028.

Hung, Y., Gaillard, S.L., Yarmak, P., and Arsalidou, M. (2018). Dissociations of cognitive

inhibition, response inhibition, and emotional interference: Voxelwise ALE meta-analyses of

fMRI studies. Human brain mapping *39*, 4065-4082. 10.1002/hbm.24232.

Jahfari, S., Ridderinkhof, K.R., Collins, A.G., Knapen, T., Waldorp, L.J., and Frank, M.J.

(2019). Cross-task contributions of frontobasal ganglia circuitry in response inhibition and

conflict-induced slowing. Cerebral Cortex *29*, 1969-1983. 10.1093/cercor/bhy076.

Jahfari, S., Verbruggen, F., Frank, M.J., Waldorp, L.J., Colzato, L., Ridderinkhof, K.R., and

Forstmann, B.U. (2012). How preparation changes the need for top–down control of the basal

ganglia when inhibiting premature actions. Journal of Neuroscience *32*, 10870-10878.

10.1523/JNEUROSCI.0902-12.2012.

Jahfari, S., Waldorp, L., van den Wildenberg, W.P., Scholte, H.S., Ridderinkhof, K.R., and

Forstmann, B.U. (2011). Effective connectivity reveals important roles for both the

hyperdirect (fronto-subthalamic) and the indirect (fronto-striatal-pallidal) fronto-basal ganglia

pathways during response inhibition. Journal of Neuroscience *31*, 6891-6899.

10.1523/JNEUROSCI.5253-10.2011.

Kass, R.E., and Raftery, A.E. (1995). Bayes factors. Journal of the american statistical

association *90*, 773-795.

Kelly, A.C., Hester, R., Murphy, K., Javitt, D.C., Foxe, J.J., and Garavan, H. (2004).

Prefrontal-subcortical dissociations underlying inhibitory control revealed by event-related

fMRI. European Journal of Neuroscience *19*, 3105-3112. 10.1111/j.0953-816X.2004.03429.x.

Kim, H. (2014). Involvement of the dorsal and ventral attention networks in oddball stimulus

processing: A meta-analysis. Human brain mapping *35*, 2265-2284. 10.1002/hbm.22326.

Klugah-Brown, B., Di, X., Zweerings, J., Mathiak, K., Becker, B., and Biswal, B. (2020).

Common and separable neural alterations in substance use disorders: A coordinate-based

meta-analyses of functional neuroimaging studies in humans. Human brain mapping *41*,

4459-4477. 10.1002/hbm.25085.

Krämer, U.M., Solbakk, A.-K., Funderud, I., Løvstad, M., Endestad, T., and Knight, R.T.

(2013). The role of the lateral prefrontal cortex in inhibitory motor control. Cortex *49*, 837-

849. 10.1016/j.cortex.2012.05.003.

Li, C.-s.R., Huang, C., Constable, R.T., and Sinha, R. (2006). Gender differences in the neural

correlates of response inhibition during a stop signal task. Neuroimage *32*, 1918-1929.

10.1016/j.neuroimage.2006.05.017.

Li, J., Xu, L., Zheng, X., Fu, M., Zhou, F., Xu, X., Ma, X., Li, K., Kendrick, K.M., and

Becker, B. (2018). Common and dissociable contributions of alexithymia and autism to

domain-specific interoceptive dysregulations–a dimensional neuroimaging approach.

bioRxiv, 432971. 10.1159/000495122.

Li, J., Yang, X., Zhou, F., Liu, C., Wei, Z., Xin, F., Daumann, B., Daumann, J., Kendrick,

K.M., and Becker, B. (2020). Modafinil enhances cognitive, but not emotional conflict

processing via enhanced inferior frontal gyrus activation and its communication with the

dorsomedial prefrontal cortex. Neuropsychopharmacology *45*, 1026-1033. 10.1038/s41386-

020-0625-z.

Li, K., Jiang, Y., Gong, Y., Zhao, W., Zhao, Z., Liu, X., Kendrick, K.M., Zhu, C., and

Becker, B. (2019). Functional near-infrared spectroscopy-informed neurofeedback: regional-

specific modulation of lateral orbitofrontal activation and cognitive flexibility.

Neurophotonics *6*, 025011. 10.1117/1.NPh.6.2.025011.

Liu, X., Zhou, X., Zeng, Y., Li, J., Zhao, W., Xu, L., Zheng, X., Fu, M., Yao, S., and

Cannistraci, C.V. (2022). Medial prefrontal and occipito-temporal activity at encoding

determines enhanced recognition of threatening faces after 1.5 years. Brain Structure and

Function, 1-18. 10.1007/s00429-022-02462-5.

Maizey, L., Evans, C.J., Muhlert, N., Verbruggen, F., Chambers, C.D., and Allen, C.P.

(2020). Cortical and subcortical functional specificity associated with response inhibition.

NeuroImage *220*, 117110. 10.1016/j.neuroimage.2020.117110.

Mallet, N., Schmidt, R., Leventhal, D., Chen, F., Amer, N., Boraud, T., and Berke, J.D.

(2016). Arkypallidal cells send a stop signal to striatum. Neuron *89*, 308-316.

10.1016/j.neuron.2015.12.017.

Mamah, D., Wang, L., Barch, D., de Erausquin, G.A., Gado, M., and Csernansky, J.G.

(2007). Structural analysis of the basal ganglia in schizophrenia. Schizophrenia research *89*,

59-71. 10.1016/j.schres.2006.08.031.

Marzinzik, F., Wahl, M., Schneider, G.-H., Kupsch, A., Curio, G., and Klostermann, F.

(2008). The human thalamus is crucially involved in executive control operations. Journal of

Cognitive Neuroscience *20*, 1903-1914. 10.1162/jocn.2008.20124.

McFarland, N.R., and Haber, S.N. (2002). Thalamic relay nuclei of the basal ganglia form

both reciprocal and nonreciprocal cortical connections, linking multiple frontal cortical areas.

Journal of Neuroscience *22*, 8117-8132. 10.1523/JNEUROSCI.22-18-08117.2002.

Morein-Zamir, S., and Robbins, T.W. (2015). Fronto-striatal circuits in response-inhibition:

Relevance to addiction. Brain research *1628*, 117-129. 10.1016/j.brainres.2014.09.012.

Morein-Zamir, S., Dodds, C., van Hartevelt, T.J., Schwarzkopf, W., Sahakian, B., Müller, U.,

and Robbins, T. (2014). Hypoactivation in right inferior frontal cortex is specifically

associated with motor response inhibition in adult ADHD. Human brain mapping *35*, 5141-

5152. 10.1002/hbm.22539.

Obeso, J.A., Rodriguez-Oroz, M.C., Rodriguez, M., Lanciego, J.L., Artieda, J., Gonzalo, N.,

and Olanow, C.W. (2000). Pathophysiology of the basal ganglia in Parkinson's disease.

Trends in neurosciences *23*, S8-S19. 10.1016/s1471-1931(00)00028-8.

Ott, T., and Nieder, A. (2019). Dopamine and cognitive control in prefrontal cortex. Trends in

cognitive sciences *23*, 213-234. 10.1016/j.tics.2018.12.006.

Pfeifer, P., Sebastian, A., Buchholz, H.G., Kaller, C.P., Gründer, G., Fehr, C.,

Schreckenberger, M., and Tüscher, O. (2022). Prefrontal and striatal dopamine D2/D3

receptors correlate with fMRI BOLD activation during stopping. Brain imaging and behavior

16, 186-198. 10.1007/s11682-021-00491-y.

Phillips, J.M., Kambi, N.A., Redinbaugh, M.J., Mohanta, S., and Saalmann, Y.B. (2021).

Disentangling the influences of multiple thalamic nuclei on prefrontal cortex and cognitive

control. Neuroscience & Biobehavioral Reviews *128*, 487-510.

10.1016/j.neubiorev.2021.06.042.

Protopopescu, X., Pan, H., Altemus, M., Tuescher, O., Polanecsky, M., McEwen, B.,

Silbersweig, D., and Stern, E. (2005). Orbitofrontal cortex activity related to emotional

processing changes across the menstrual cycle. Proceedings of the National Academy of

Sciences *102*, 16060-16065. 10.1073/pnas.0502818102.

Puiu, A.A., Wudarczyk, O., Kohls, G., Bzdok, D., Herpertz-Dahlmann, B., and Konrad, K.

(2020). Meta-analytic evidence for a joint neural mechanism underlying response inhibition

and state anger. Human brain mapping *41*, 3147-3160. 10.1002/hbm.25004.

Qiao, L., Xu, M., Luo, X., Zhang, L., Li, H., and Chen, A. (2020). Flexible adjustment of the

effective connectivity between the fronto-parietal and visual regions supports cognitive

flexibility. NeuroImage *220*, 117158. 10.1016/j.neuroimage.2020.117158.

Rae, C.L., Nombela, C., Rodríguez, P.V., Ye, Z., Hughes, L.E., Jones, P.S., Ham, T.,

Rittman, T., Coyle-Gilchrist, I., and Regenthal, R. (2016). Atomoxetine restores the response

inhibition network in Parkinson's disease. Brain *139*, 2235-2248. 10.1093/brain/aww138.

Renteria, R., Baltz, E.T., and Gremel, C.M. (2018). Chronic alcohol exposure disrupts top-

down control over basal ganglia action selection to produce habits. Nature communications *9*,

1-11. 10.1038/s41467-017-02615-9.

Rey-Mermet, A., Gade, M., and Oberauer, K. (2018). Should we stop thinking about

inhibition? Searching for individual and age differences in inhibition ability. Journal of

Experimental Psychology: Learning, Memory, and Cognition *44*, 501. 10.1037/xlm0000450.

Ribeiro, F., Cavaglia, R., and Rato, J.R. (2021). Sex differences in response inhibition in

young children. Cognitive Development *58*, 101047. 10.1016/J.COGDEV.2021.101047.

Robbins, T.W. (2007). Shifting and stopping: fronto-striatal substrates, neurochemical

modulation and clinical implications. Philosophical Transactions of the Royal Society B:

Biological Sciences *362*, 917-932. 10.1098/rstb.2007.2097.

Robertson, C.L., Ishibashi, K., Mandelkern, M.A., Brown, A.K., Ghahremani, D.G., Sabb, F.,

Bilder, R., Cannon, T., Borg, J., and London, E.D. (2015). Striatal D1-and D2-type dopamine

receptors are linked to motor response inhibition in human subjects. Journal of Neuroscience

35, 5990-5997. 10.1523/JNEUROSCI.4850-14.2015.

Rubia, K., Smith, A.B., Taylor, E., and Brammer, M. (2007). Linear age-correlated functional

development of right inferior fronto-striato-cerebellar networks during response inhibition

and anterior cingulate during error-related processes. Human brain mapping *28*, 1163-1177.

10.1002/hbm.20347.

Rupprechter, S., Romaniuk, L., Series, P., Hirose, Y., Hawkins, E., Sandu, A.-L., Waiter,

G.D., McNeil, C.J., Shen, X., and Harris, M.A. (2020). Blunted medial prefrontal cortico-

limbic reward-related effective connectivity and depression. Brain *143*, 1946-1956.

10.1093/brain/awaa106.

Saalmann, Y.B., and Kastner, S. (2015). The cognitive thalamus. Frontiers in systems

neuroscience *9*, 39. 10.3389/fnsys.2015.00039.

Schall, J.D., and Godlove, D.C. (2012). Current advances and pressing problems in studies of

stopping. Current opinion in neurobiology *22*, 1012-1021. 10.1016/j.conb.2012.06.002.

Schaum, M., Pinzuti, E., Sebastian, A., Lieb, K., Fries, P., Mobascher, A., Jung, P., Wibral,

M., and Tüscher, O. (2020). Cortical network mechanisms of response inhibition. biorxiv.

10.1101/2020.02.09.940841.

Schaum, M., Pinzuti, E., Sebastian, A., Lieb, K., Fries, P., Mobascher, A., Jung, P., Wibral,

M., and Tüscher, O. (2021). Right inferior frontal gyrus implements motor inhibitory control

via beta-band oscillations in humans. ELife *10*, e61679. 10.7554/eLife.61679.

Schimmack, U., and Derryberry, D.E. (2005). Attentional interference effects of emotional

pictures: threat, negativity, or arousal? Emotion *5*, 55. 10.1037/1528-3542.5.1.55.

Schmitt, L.I., Wimmer, R.D., Nakajima, M., Happ, M., Mofakham, S., and Halassa, M.M.

(2017). Thalamic amplification of cortical connectivity sustains attentional control. Nature

545, 219-223. 10.1038/nature22073.

Sjoberg, E.A., and Cole, G.G. (2018). Sex differences on the Go/No-Go test of inhibition.

Archives of Sexual Behavior *47*, 537-542. 10.1007/s10508-017-1010-9.

Sommer, M.A. (2003). The role of the thalamus in motor control. Current opinion in

neurobiology *13*, 663-670. 10.1016/j.conb.2003.10.014.

Sonuga-Barke, E.J. (2005). Causal models of attention-deficit/hyperactivity disorder: from

common simple deficits to multiple developmental pathways. Biological psychiatry *57*, 1231-

1238. 10.1016/j.biopsych.2004.09.008.

Stephan, K.E., Harrison, L.M., Kiebel, S.J., David, O., Penny, W.D., and Friston, K.J. (2007).

Dynamic causal models of neural system dynamics: current state and future extensions.

Journal of biosciences *32*, 129-144. 10.1007/s12038-007-0012-5.

Stephan, K.E., Penny, W.D., Moran, R.J., den Ouden, H.E., Daunizeau, J., and Friston, K.J.

(2010). Ten simple rules for dynamic causal modeling. Neuroimage *49*, 3099-3109.

10.1016/j.neuroimage.2009.11.015.

Stuphorn, V. (2015). Neural mechanisms of response inhibition. Current opinion in

behavioral sciences *1*, 64-71. 10.1016/j.cobeha.2014.10.009.

Tanaka, M., and Kunimatsu, J. (2011). Contribution of the central thalamus to the generation

of volitional saccades. European Journal of Neuroscience *33*, 2046-2057. 10.1111/j.1460-

9568.2011.07699.x.

Terra, H., Bruinsma, B., de Kloet, S.F., van der Roest, M., Pattij, T., and Mansvelder, H.D.

(2020). Prefrontal cortical projection neurons targeting dorsomedial striatum control

behavioral inhibition. Current Biology *30*, 4188-4200. e4185. 10.1016/j.cub.2020.08.031.

Thompson, A., Schel, M.A., and Steinbeis, N. (2021). Changes in BOLD variability are

linked to the development of variable response inhibition. NeuroImage *228*, 117691.

10.1016/j.neuroimage.2020.117691.

Tops, M., and Boksem, M.A. (2011). A potential role of the inferior frontal gyrus and anterior

insula in cognitive control, brain rhythms, and event-related potentials. Frontiers in

psychology *2*, 330. 10.3389/fpsyg.2011.00330.

Van Overwalle, F., Van de Steen, F., van Dun, K., and Heleven, E. (2020). Connectivity

between the cerebrum and cerebellum during social and non-social sequencing using dynamic

causal modelling. NeuroImage *206*, 116326. 10.1016/j.neuroimage.2019.116326.

Verbruggen, F., and Logan, G.D. (2008). Response inhibition in the stop-signal paradigm.

Trends in cognitive sciences *12*, 418-424. 10.1016/j.tics.2008.07.005.

Verbruggen, F., and Logan, G.D. (2009). Models of response inhibition in the stop-signal and

stop-change paradigms. Neuroscience & Biobehavioral Reviews *33*, 647-661.

10.1016/j.neubiorev.2008.08.014.

Vijayraghavan, S., Major, A.J., and Everling, S. (2016). Dopamine D1 and D2 receptors make

dissociable contributions to dorsolateral prefrontal cortical regulation of rule-guided

oculomotor behavior. Cell reports *16*, 805-816. 10.1016/j.celrep.2016.06.031.

Wei, W., and Wang, X.-J. (2016). Inhibitory control in the cortico-basal ganglia-

thalamocortical loop: complex regulation and interplay with memory and decision processes.

Neuron *92*, 1093-1105. 10.1016/j.neuron.2016.10.031.

Weiss, F., Zhang, J., Aslan, A., Kirsch, P., and Gerchen, M.F. (2022). Feasibility of training

the dorsolateral prefrontal-striatal network by real-time fMRI neurofeedback. Scientific

reports *12*, 1-10. 10.1038/s41598-022-05675-0.

Xiao, D., Zikopoulos, B., and Barbas, H. (2009). Laminar and modular organization of

prefrontal projections to multiple thalamic nuclei. Neuroscience *161*, 1067-1081.

10.1016/j.neuroscience.2009.04.034.

Xu, B., Sandrini, M., Wang, W.T., Smith, J.F., Sarlls, J.E., Awosika, O., Butman, J.A.,

Horwitz, B., and Cohen, L.G. (2016). PreSMA stimulation changes task-free functional

connectivity in the fronto-basal-ganglia that correlates with response inhibition efficiency.

Human brain mapping *37*, 3236-3249. 10.1002/hbm.23236.

Xu, L., Zheng, X., Yao, S., Li, J., Fu, M., Li, K., Zhao, W., Li, H., Becker, B., and Kendrick,

K.M. (2022). The mirror neuron system compensates for amygdala dysfunction-associated

social deficits in individuals with higher autistic traits. NeuroImage *251*, 119010.

10.1016/j.neuroimage.2022.119010.

Zeidman, P., Jafarian, A., Corbin, N., Seghier, M.L., Razi, A., Price, C.J., and Friston, K.J.

(2019a). A guide to group effective connectivity analysis, part 1: First level analysis with

DCM for fMRI. Neuroimage *200*, 174-190. 10.1016/j.neuroimage.2019.06.031.

Zeidman, P., Jafarian, A., Seghier, M.L., Litvak, V., Cagnan, H., Price, C.J., and Friston, K.J.

(2019b). A guide to group effective connectivity analysis, part 2: Second level analysis with

PEB. Neuroimage *200*, 12-25. 10.1016/j.neuroimage.2019.06.032.

Zhang, R., Geng, X., and Lee, T. (2017). Large-scale functional neural network correlates of

response inhibition: an fMRI meta-analysis. Brain Structure and Function *222*, 3973-3990.

10.1007/s00429-017-1443-x.

Zhao, Z., Yao, S., Li, K., Sindermann, C., Zhou, F., Zhao, W., Li, J., Lührs, M., Goebel, R.,

Kendrick, K.M., and Becker, B. (2019). Real-time functional connectivity-informed

neurofeedback of amygdala-frontal pathways reduces anxiety. Psychotherapy and

psychosomatics *88*, 5-15. 10.1159/000496057.

Zhou, F., Li, J., Zhao, W., Xu, L., Zheng, X., Fu, M., Yao, S., Kendrick, K.M., Wager, T.D.,

and Becker, B. (2020). Empathic pain evoked by sensory and emotional-communicative cues

share common and process-specific neural representations. Elife *9*, e56929.

10.7554/eLife.56929.

Zhou, F., Zimmermann, K., Xin, F., Scheele, D., Dau, W., Banger, M., Weber, B.,

Hurlemann, R., Kendrick, K.M., and Becker, B. (2018). Shifted balance of dorsal versus

ventral striatal communication with frontal reward and regulatory regions in cannabis-

dependent males. Human brain mapping *39*, 5062-5073. 10.1002/hbm.24345.

Zhuang, Q., Xu, L., Zhou, F., Yao, S., Zheng, X., Zhou, X., Li, J., Xu, X., Fu, M., Li, K.,

Vatansever, D., Kendrick, K.M., and Becker, B. (2021). Segregating domain-general from

emotional context-specific inhibitory control systems-ventral striatum and orbitofrontal cortex

serve as emotion-cognition integration hubs. NeuroImage *238*, 118269.

10.1016/j.neuroimage.2021.118269.

Zhukovsky, P., Morein-Zamir, S., Ziauddeen, H., Fernandez-Egea, E., Meng, C., Regenthal,

R., Sahakian, B.J., Bullmore, E.T., Robbins, T.W., and Dalley, J.W. (2021). Prefrontal Cortex

Activation and Stopping Performance Underlie the Beneficial Effects of Atomoxetine on

Response Inhibition in Healthy Volunteers and Those With Cocaine Use Disorder. Biological

Psychiatry: Cognitive Neuroscience and Neuroimaging. 10.1016/j.bpsc.2021.08.010.