Hippocampal place cell remapping occurs with memory storage of aversive experiences

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ABSTRACT. Aversive stimuli can cause hippocampal place cells to remap their firing fields, but

it is not known whether remapping plays a role in storing memories of aversive experiences.

Here we addressed this question by performing *in-vivo* calcium imaging of CA1 place cells in

freely behaving rats (n=14). Rats were first trained to prefer a short path over a long path for

obtaining food reward, then trained to avoid the short path by delivering a mild footshock.

Remapping was assessed by comparing place cell population vector similarity before

acquisition versus after extinction of avoidance. Some rats received shock after systemic

injections of the amnestic drug scopolamine at a dose (1 mg/kg) that impaired avoidance

learning but spared spatial tuning and shock-evoked responses of CA1 neurons. Place cells

remapped significantly more following remembered than forgotten shocks (drug-free versus

scopolamine conditions); shock-induced remapping did not cause place fields to migrate toward

or away from the shocked location and was similarly prevalent in cells that were responsive

versus non-responsive to shocks. When rats were exposed to a neutral barrier rather than

aversive shock, place cells remapped significantly less in response to the barrier. We conclude

that place cell remapping occurs in response to events that are remembered rather than merely

perceived and forgotten, suggesting that reorganization of hippocampal population codes may

play a role in storing memories for aversive events.

INTRODUCTION

The rodent hippocampus contains place cells that fire selectively during visits to specific locations in space (O'Keefe and Dostrovsky 1971). When place cells were first discovered, they were hypothesized to encode "cognitive maps" that store stable long-lasting memories for the spatial geometry of familiar environments (O'Keefe and Nadel 1978). Supporting this view, electrophysiology studies showed that place cells can retain stable positional tuning across repeated visits to the same environment (Lever et al. 2002). However, methodological limitations of single-unit electrophysiology made it challenging to consistently record the same population of place cells across long time spans, and thus to investigate the duration of time over which place cell tuning properties remained stable.

Recent advances in *in-vivo* calcium imaging have made it possible to perform longitudinal studies of place cell tuning over days or weeks of experience. Such studies suggest that place cells in mice (Ziv et al. 2013; Cai et al. 2016; Kinsky et al. 2018; Keinath, Mosser, and Brandon 2022) and to a lesser extent in rats (Wirtshafter and Disterhoft 2022) can change their spatial tuning properties (that is, "remap") with the mere passage time (see also (Mankin et al. 2012). This phenomenon suggests that place cells may not simply encode spatial maps of environmental geometry, but also encode mnemonic information about accumulated past experiences that have occurred within an environment (Leutgeb et al. 2005; Colgin, Moser, and Moser 2008; Mankin et al. 2012; Ziv et al. 2013; Cai et al. 2016; Sanders, Wilson, and Gershman 2020). If so, then the rate at which place cells remap over time within a given environment may be proportional to the rate at which an animal stores new episodic memories of events that are linked to that environment.

If the rate of place remapping reflects the rate of hippocampal memory storage, then motivationally significant events (such as encounters with appetitive or aversive stimuli) that are preferentially stored to episodic memory might be expected to accelerate the rate of place cell

remapping within environments where such events occur. Consistent with this view, studies

have shown that place cells remap in response to behaviorally significant events (Breese,

Hampson, and Deadwyler 1989; Bostock, Muller, and Kubie 1991; Hollup et al. 2001; Colgin,

Moser, and Moser 2008; Dupret et al. 2010; Alvernhe, Save, and Poucet 2011; Mamad et al.

2017). Numerous studies have shown that when an aversive stimulus is encountered in an

environment, place cells respond by remapping their firing fields in that environment (Moita et al.

2003; Moita et al. 2004; Wang et al. 2012; Wang et al. 2015; Kim et al. 2015; Mamad et al.

2019; Schuette et al. 2020; Ormond, Serka, and Johansen 2023), but it is not well understood

whether such remapping plays a role in storing memories of aversive experiences.

Here we investigated the role of place cell remapping in memory by using the MiniLFOV

(Guo et al. 2023), a recently developed large-field-of-view version of the UCLA Miniscope (Cai

et al. 2016; Aharoni and Hoogland 2019), to image population activity of CA1 place cells in

freely behaving rats as they acquired and extinguished a spatial avoidance learning task. The

task was performed on a rectangular maze where rats ran stereotyped linear trajectories,

allowing the speed and direction of running behavior to be closely matched during sessions

conducted before acquisition versus after extinction of avoidance learning. In agreement with

prior studies, we observed that avoidance learning induced place cell remapping in an

environment where footshock was encountered, as compared with control conditions where: 1)

rats encountered a neutral barrier instead of an aversive footshock, or 2) rats received

footshock after injections of the muscarinic ACh receptor (mAChR) antagonist scopolamine at a

dose (1 mg/kg) which disrupted long-term retention of avoidance without acutely impairing

immediate avoidance or spatial tuning of place cells. Remapping after avoidance learning was

similarly prevalent in subpopulations of shock responsive and non-responsive place cells, and in

contrast with some prior findings (Mamad et al. 2019), remapping did not alter the spatial

distribution of place fields on the maze (that is, there was no net migration of place fields toward

or away from the shocked location). These findings add to a large body of evidence showing

that place cell remapping occurs in response to aversive events, and also provide new evidence

that remapping occurs preferentially under conditions where aversive encounters are

remembered rather than forgotten, consistent with the hypothesis that place cell remapping may

play a role in storing memories of aversive encounters.

83 RESULTS

Male (n=7) and female (n=8) Long-Evans rats underwent survival surgery to inject 1.2 uL

of AAV9-Syn-GCamp7s (*AddGene*) in the hippocampal CA1 layer; ~2 weeks later, a 1.8 mm

diameter GRIN lens stack (~0.5 pitch) was unilaterally implanted to image CA1 neurons. A

baseplate was attached to the skull for mounting the miniscope, and then rats began a regimen

of running one 15 min session per 48 h on a 250x125 cm rectangular maze (Fig. 1A). Rats

earned 20 mg chocolate pellets by alternating between two rewarded corners. On each trial,

rats were free to choose a direct short path (250 cm) or an indirect long path (500 cm) to reach

the next reward. During early sessions, rats learned to prefer the short over the long path (Fig.

1C,D). Upon reaching criterion for short path preference (>2 rewards/min, and twice as many

short paths to long paths), rats began receiving drug-free shock, scopolamine shock, or barrier

training sessions (Fig. 1B).

Acquisition and extinction of avoidance learning

During shock training sessions (drug-free or scopolamine), rats performed the standard

alternation task for 10 min. Then a 1.0 mA scrambled current was switched on to electrify grid

bars in a 50 cm segment of the short path's center (Fig. 1A). Identical grid bars spanned the full

length of the long and short paths, so there were no local cues to indicate the shock zone's

location. After encountering the shock zone 1-2 times, rats subsequently avoided the short path

during the final 5 min of the session (Fig. 1E). In the scopolamine shock condition, lens-

implanted rats received 1.0 mg/kg scopolamine via intraperitoneal injection 30 min prior to the

start of the shock session. In the drug-free condition, lens-implanted rats received no injection

prior to the start of the shock session. To control for effects of the injection procedure, a

separate cohort of rats without lens implants was given injections of saline or scopolamine prior

to shock training; in animals without lens implants, scopolamine-injected rats showed impaired

avoidance learning compared with saline-injected rats that was similar to the impairment seen

between scopolamine-injected and drug-free (non-injected) rats with lens implants (Fig. 1-figure

supplement 1, panels B and C), indicating that the injection procedure itself was not responsible

for effects induced by scopolamine. In the barrier control condition, the maze was not

electrified; instead, a clear plexiglass barrier was placed in the center of the short path, forcing

the rat to take the long path during the final 5 min of the session.

During the first 10m of training on scopolamine (before shock delivery), 6/10 rats (3M,

3F) fell below criterion for preferring the short path (compared with 0/14 drug-free rats). These

rats earned a median of 5 short path rewards/min during the drug-free session given 48 h

before training, but only 1.9 short path rewards/min during the first 10 min (before shock) of their

training session on scopolamine (Wilcoxon test, p=.0098). Thus, prior to shock delivery,

scopolamine impaired expression of the rats' learned preference for taking the short path (Fig.

1E, middle).

During the training session, rats earned fewer rewards/min on the short path after than

before grid electrification (Fig. 1F) in both the drug-free (n=14, signed-rank test, p=1.2e^{-4}) and

scopolamine (n=10, signed-rank test, p=.004) conditions. Rats in the barrier condition (n=6)

were likewise forced to stop taking the short path after it was blocked by the barrier (Fig. 1F, left

panel). Rats in the drug-free shock condition showed long-term retention of avoidance (Fig. 1G,

left), earning fewer rewards/min on the short path 48 h after than 48 h before training (Fig. 1H,

Wilcoxon test, p=3.7e^{-4}). By contrast, most rats trained on scopolamine showed similar short

path reward rates during drug-free sessions given 48 h before and after training (Fig. 1H,

Figure 1. Disruption of aversive avoidance acquisition by scopolamine. A) Overhead view of the rectangular maze. B) Order of treatment for 5 subgroups of rats in the study. The number of sessions in ACQ and EXT boxes varied per rat, depending on how many sessions it took individual subjects to reach behavioral criteria. C) Example trajectories from sessions 1, 3, & 5 during initial acquisition of short path preference. D) Acquisition curves for short path preference; criterion was reached on session 0. E) Example maze trajectories from training sessions. F) Left: Rewards/min earned on the short path during the first 10 min (pre10) versus last 5 min (post5) of training sessions; symbols indicate whether each rat received its first (shk1) or second (shk2) shock during a session. Right: Box plots show post-shock reduction in short path rewards/min (post5/pre10). G) Example maze trajectories 48 h after training. H) Left: Rewards/min earned on the short path during drug-free sessions given 48 h before (-48h) vs. after (+48h) training. Right: Box plots show 48 h retention of shock avoidance (+48h / -48h). I) Example maze trajectories 144 h after training. J) Left: Extinction curves over the first 3 post-extinction sessions; number of rewards earned on the short path is measured as a percentage of baseline (48 h before training; thin lines: individual rats, thick lines: per session median). Right: Cumulative distributions show percentage of rats in each training condition meeting extinction criterion (≥2 rewards/min on the short path, and at least twice as many short path rewards as long path rewards) at 1, 2, 3, or 4-days after training. *p<.05, **p<.01, ****p<.0001.

Wilcoxon test, p=.125), and thus failed to exhibit long-term retention of avoidance (Fig. 1G,

middle). When tested 48 h after training, rats shocked on scopolamine showed less avoidance

of the short path than rats shocked drug-free (rank sum test: p=.0235; Fig. 1H). Hence, pre-

training scopolamine injections impaired aversive learning, consistent with prior studies (Decker,

Tran, and McGaugh 1990; Anagnostaras et al. 1999; Green et al. 2005).

Between- versus within-session place cell stability

A large-field-of view version of the UCLA Miniscope ("MiniLFOV"; (Guo et al. 2023) was

used to image neurons in the hippocampal CA1 layer through the implanted GRIN lens during

maze sessions (Fig. 2A). Although the GRIN lens caused some flattening and compression of

stratum oriens (Fig. 2-figure supplement 1), this did not appear to cause significant disruptions

of CA1 activity since spatial tuning properties of place cells were stable and intact (see below).

Calcium activity was analyzed only during traversals of the short path, since the long

path was undersampled after rats had learned to prefer the short path. Analysis was further

restricted to *beeline trials* during which the rat ran directly from one reward zone to the other on

the short path, without any pause or change in direction (Fig. 2B; Video 1); this assured that

imaging data came from periods of active locomotion when place cells exhibit reliable spatial

tuning, and that the rats' behavior during imaging was similar across all experimental sessions

and conditions. To further control for possible confounding effects of behavior differences

between sessions (see Fig. 2-figure supplement 2), beeline trials were randomly subsampled

using an algorithm (see Methods) that minimized trial count and running speed differences

between the drug-free and scopolamine conditions (Fig. 2C). Spatial tuning was then analyzed

and compared during these behaviorally homogeneous trials. To analyze spatial tuning, the

short path was subdivided into 23 spatial bins (each 10.8 cm wide). Two spatial tuning curves

168
Figure 2. *Between- versus within-session population coding.* A) *Upper left*: Illustration of rat wearing MiniLFOV.
Upper right: Cell contours identified during pre-training (green) and training (red) sessions in one rat; regions of overlap
between contours that recurred in both sessions appear yellow. *Lower left*: Target position of GRIN lens over the CA1
layer. *Lower right*: Fluorescence image of lens position from the example rat. B) *Top*: Rastergram shows normalized
calcium fluorescence traces of place cells (one per row, sorted by preferred firing location and direction) during several
traversals of the short path during an example session. *Bottom*: Rat's position (black) with running epochs colored by
direction of travel ("LR", left to right, in blue; "RL", right to left, in red). C) Individual rat data (lines & symbols) and
session means (bars) for the number of subsampled beeline trials (left) and median beeline running speed (right) during
pre-training (*pre*) and training (*trn*) sessions; subsampled beeline trial counts were lower for *trn* than *pre* sessions
because only trials from the first 10 min of *trn* (prior to shock delivery) were included; running speeds were lower for *trn*
than *pre* sessions because scopolamine (SCP) reduced running speeds, resulting in preferential subsampling of slower
beeline trials from drug-free (DF) sessions by the algorithm that minimized running speed differences between training
conditions (see Methods). D) Bar/line graphs show number of place cells imaged per rat (left) and percentage of all
imaged cells classified as place cells (right) during *pre* and *trn* sessions. E) Pie graphs show percentages of place cells
imaged per condition ('n' gives total number summed over rats) that were spatially tuned in the LR only, RL only, or
both LR&RL running directions. F) Tuning curve properties of place cells imaged during *pre* and *trn* sessions. G) Place
cell recurrence ratios (RR) between *pre* and *trn* sessions. H) *Top*: Diagram shows timeline for *pre* and *trn* sessions
given 48 h apart. *Bottom:* Tuning curve heatmaps for recurring place cells (from all rats combined, co-sorted by peak
locations from the *trn1* session) that were spatially tuned in the LR (top) or RL (bottom) running directions; separate
heatmaps are shown for *pre*, *trn* part one (*trn1*), and *trn* part two (*trn2*) sessions. I) Between- (B) and within- (W) session
population vector correlation matrices are shown for DF and SCP shock training conditions; middle bar graph shows
median place tuning stability (S) for each rat (lines & symbols) and mean over rats (bars) for B and W heatmap pairs.
Asterisks in 'C' and 'D' denote significance for main effect of DF vs. SCP or uncorrected t-test comparing *pre* vs. *trn*
sessions; asterisks in 'E' and 'G' denote significance for uncorrected t-tests. †p<.1; *p<.05; **p<.01; ***p<. 001.

(one per running direction) were derived for each neuron. Cell activity was measured in units of

active frames per second (Af/s), defined as the mean number of imaging frames per second

during which a neuron generated at least one inferred spike. A neuron was classified as a place

cell if its LR or RL tuning curve (or both) met defined criteria for minimum Af/s, spatial selectivity,

and spatial stability within a session (see Methods). About 1/3 of place cell tuning curves met

criteria for spatial selectivity only in the LR direction, 1/3 only in the RL direction, and 1/3 in both

directions (Fig. 2E). It should be noted that since we used a ubiquitous synapsin promoter to

drive expression of GcaMP7s, some neurons classified as place cells might have been CA1

interneurons rather than pyramidal cells.

To analyze acute effects of scopolamine on CA1 place cell activity, data from drug-free

or scopolamine shock sessions was compared against data from pre-training baseline sessions

(always drug-free) given 48 h earlier. The drug-free condition only included rats (n=9) that

retained avoidance of the short path 48 h after shock training, whereas the scopolamine

condition only included rats (n=7) that failed to avoid the short path 48 h after shock training

(indicating that the drug had blocked avoidance learning). A 2x2 ANOVA on the number of place

cells detected per session (Fig. 2D, left) found no effect of training condition ($F_{1,14}$=.03, p=.87) or

session ($F_{1,14}$=.014, p=.92), and no interaction ($F_{1,14}$=2.1, p=.17). A similar ANOVA on the

percentage of imaged cells per session that were classified as place cells (Fig. 2D, right) found

no effect of training condition ($F_{1,14}$=.07, p=.79), but place cell percentages were lower during

training than pre-training sessions ($F_{1,14}$=5.1, p=.04); this reduction was greater in the

scopolamine than drug-free condition, resulting in a marginal interaction effect ($F_{1,14}$=4.4,

p=.0539). An analysis of how spatial tuning varied with the size and location of cell contours

revealed that contours of non-place cells tended to be slightly but significantly smaller (p<.01)

and nearer to the lens center (p<.05) than contours of place cells (Fig. 2-figure supplement 3A).

For tuning curves that met criteria for spatial selectivity in each session, four tuning

properties were analyzed: peak Af/s rate, out-of-field Af/s rate, tuning width, and spatial

information content (see Methods). For each rat and each session, median values of these

tuning properties were taken over all LR and RL tuning curves that met spatial selectivity criteria

(Fig. 2-figure supplement 4); 2x2 mixed ANOVAs were then performed on the resulting rate

medians for each of the four tuning properties (Fig. 2F). There were large main effects of

session (pre-training vs. training) for all four tuning properties, which were driven by

correspondingly large differences in behavior sampling and running speed (Fig. 2C). However,

there were no main effects of training condition (drug-free vs. scopolamine) or interactions

between training condition and session (Fig. 2F), indicating that none of the four tuning

properties differed during training sessions given drug-free versus on scopolamine.

The *place field recurrence ratio* was defined as $RR = N_R/N_T$, where N_R is the number of

place fields that recurred during the pre-training and training sessions, and N_T is the total

number of recurring and non-recurring place fields detected during both sessions combined

(see Methods). Mean RR values were lower (independent $t_{14} = 2.15, p = .0497$) during

scopolamine (40.7±8.8%) than drug-free (50.7±3.9%) training sessions, consistent with the

finding reported above that the proportion of neurons classified as place cells during the training

session was lower on scopolamine than drug-free (Fig. 2D, right). It was observed that contours

of recurring place cells tended to be slightly but significantly smaller (p<.01) and nearer to the

lens center (p<.01) than contours of non-recurring place cells (see panels B,C of Fig. 2-figure

supplement 3).

To investigate how scopolamine affected population coding by recurring place cells, two

heatmaps (one per running direction, denoted H_{LR}^{pre} and H_{RL}^{pre}) were created from pre-training

tuning curves of recurring place cells (Fig. 2H; Fig. 2-figure supplement 5). Additionally, two

heatmaps per running direction were created from data collected during the first (H_{LR}^{trn1}, H_{RL}^{trn1})

and second (H_{LR}^{trn2}, H_{RL}^{trn2}) half of each training session's subsampled beeline trials, all of which

occurred prior to shock delivery. Between-session population vector correlation matrices (one

per running direction) were derived for each rat (Fig. 2-figure supplement 5) with matrix entries

given by $\rho_{i,j}^{btwn} = R(H_i^{pre}, H_j^{trn1})$, where H_i^{pre} and H_j^{trn1} denote population vectors in columns i

and j of the pre-training and trn1 heatmaps, respectively, and R is the Pearson correlation

coefficient. Within-session population vector correlation matrices were derived in a similar

manner with matrix entries given by $\rho_{i,j}^{wthn} = R(H_i^{trn1}, H_j^{trn2})$, where H_i^{trn1} and H_j^{trn2} denote

population vectors in columns i and j of the trn1 and trn2 heatmaps, respectively.

When population vector correlation matrices were averaged together across running

directions and rats (Fig. 2I), high mean ρ^{wthn} values were observed along diagonals of within-

session correlation matrices for both the drug-free and scopolamine conditions. By contrast,

lower mean ρ^{btwn} values were observed along diagonals of between-session correlation

matrices, in accordance with prior studies showing that place cell population vectors become

decorrelated over time (Mankin et al. 2012; Ziv et al. 2013; Cai et al. 2016; Keinath, Mosser,

and Brandon 2022). To quantify this decorrelation, a place tuning stability score (S) was

computed as $S=median(\rho_1^{LR} - \rho_{off}^{LR}, \rho_2^{LR} - \rho_{off}^{LR}, ..., \rho_K^{LR} - \rho_{off}^{LR}, \rho_1^{RL} - \rho_{off}^{RL}, \rho_2^{RL} - \rho_{off}^{RL}, ..., \rho_K^{RL} -$

$\rho_{off}^{RL})$, where ρ_{off}^{LR} and ρ_{off}^{RL} are mean off-diagonal ρ values and ρ_i^{LR} and ρ_i^{RL} are individual peri-

diagonal ρ values for bins in the rat's LR and RL correlation matrices (see Methods). A 2x2

mixed ANOVA on S scores with training condition (drug-free vs. scopolamine) as an

independent factor and time interval (between vs. within session) as a repeated factor found a

marginal main effect of training condition ($F_{1,14}=3.5$, p=.08), but there was a large main effect of

time interval ($F_{1,14}=32.0$, p=5.9e^{-5}) and a significant interaction ($F_{1,14}=5.7$, p=.0312). Uncorrected

post-hoc comparisons confirmed that this was because scopolamine injections reduced

between- ($t_{14}=3.2$, $S_{DF}<S_{SCP}$ p=.0066) but not within- ($t_{14}=.2$, $S_{DF}<S_{SCP}$ p=.84) session population

vector correlations (Fig. 2I, center). Hence, even though scopolamine did not acutely disrupt

spatial tuning of CA1 neurons (Fig. 2F), the drug reduced the percentage of recurring place cells

and degraded the between-session (but not within-session) similarity of recurring place cell

population vectors (Fig. 2H,I). Similar disruption of between- but not within-session place cell

stability has been previously reported in aged rats (Barnes et al., 1997).

Disruption of between-session place cell stability by scopolamine was similar in male

and female rats (Fig. 2-figure supplement 6). Cross-session stability of population vectors was

completely abolished in control analyses (see Methods) where inferred spikes were circularly

shifted against position tracking data during each individual beeline trial of the pre-training

reference session (Fig. 2-figure supplement 7). When analyses of Fig. 2 were repeated on

scopolamine rats (n=2) that retained avoidance despite the injection, population vector similarity

was not degraded between pre-training and training sessions, suggesting that impairment of

between-session place tuning stability was related to impairment of avoidance learning (Fig. 2-

figure supplement 8). When the analyses were repeated on drug-free rats (n=2) that failed to

retain avoidance despite being trained drug free, between- and within-session population vector

similarity were indistinguishable from drug rats that retained avoidance (Fig. 2-figure

supplement 8), implying that failure to acquire avoidance in these rats was not attributable to

any noticeable deficiency in the fidelity of the hippocampal place code during training.

Induction of place cell remapping by aversive learning

Training-induced place cell remapping was analyzed in the same subset of rats shown in

Fig. 2. To measure remapping, post-extinction CA1 population vectors were compared against

the 48 h pre-acquisition baseline session. For the drug-free shock condition, post-extinction

population vectors were sampled from the first session after shock training during which the rat

met criterion (>2 rewards/min, and twice as many short paths to long paths) for extinction of

short path avoidance (Fig. 3A, left); this was the earliest post-training session with adequate

sampling of the short path. For the scopolamine shock condition, post-extinction population

vectors were sampled from the session given 8 d (that is, 4 sessions including the training

session) after the pre-training session (Fig. 3A, center); this matched the modal intersession

interval for the drug-free condition, so that confounds would not arise from measuring

remapping across different time intervals for the drug-free versus scopolamine conditions.

Likewise, for the barrier control condition, post-extinction population vectors were also sampled

8 d after the pre-training session; one barrier training rat became startled due to external factors

and stopped running early in the 8 d post-training session, so the 6 d session was used instead

(Fig. 3A, right).

To control for potential confounds from between-session behavior differences, beeline

trials were subsampled using an algorithm (see Methods) that equalized the number of trials

from pre- and post-extinction sessions in each rat (Fig. 3B, left), while also minimizing running

speed differences between sessions and training conditions (Fig. 3B). Spatial tuning was

analyzed during subsampled trials, and a 3x2 mixed ANOVA on the number of place cells

detected during each session (Fig. 3C, left) yielded no effect of training condition ($F_{2,19}$=.43,

p=.66) or session ($F_{1,19}$=.01, p=.92), and no interaction effect ($F_{2,19}$=1.57, p=.24). A similar

ANOVA on the percentage of imaged cells per session classified as place cells also yielded no

effect of training condition ($F_{2,19}$=.43, p=.66) or session ($F_{1,19}$=.01, p=.92), and no significant

effects (training condition: $F_{2,19}$=.14, p=.87; session: $F_{1,19}$=.56, p=.46; interaction: $F_{2,19}$=1.28,

p=.3). The place field recurrence ratio (RR) between pre- and post-extinction sessions also did

not differ among training conditions (Fig. 3C, right; one-way ANOVA: $F_{2,19} = .65, p = .53$).

To investigate whether shock-induced remapping occurred, a pair of heatmaps (pre and

post) was created from tuning curves of recurring place cells in each running direction (LR and

RL) for each training condition (Fig. 3D). As in prior calcium imaging studies (Ziv et al. 2013),

place fields tended to be more densely concentrated near the ends of the short path than in the

middle (Fig. 3E). Heatmap pairs were used to derive two population vector correlation matrices

per rat, P_{LR} and P_{RL} (Fig. 3-figure supplement 1). When P_{LR} and P_{RL} were averaged across rats,

a gap of low correlation appeared in the center of the diagonal for the drug-free shock condition

(Fig. 3F, left) which was not seen in the scopolamine shock (Fig. 3F, middle) or barrier (Fig. 3F,

right) conditions, suggesting that drug-free shock training induced more remapping of the short

path than scopolamine shock or barrier training, in accordance with prior studies (Moita et al.

2003; Moita et al. 2004; Wang et al. 2012; Mamad et al. 2019; Schuette et al. 2020; Ormond,

Serka, and Johansen 2023).

To statistically analyze this remapping effect, a place tuning stability score S was derived

for each rat by the methods described above. Boxplots in Fig. 3H show each rat's median and

range of S values from all peridiagonal bins in P_{LR} and P_{RL} combined. Bar graphs (Fig. 3H, left)

for mean stability scores, $\underline{S}$, for each training condition differed significantly between drug-free,

scopolamine, and barrier conditions (one-way ANOVA: $F_{2,19} = 9.7, p = .0012$); uncorrected post-

hoc t-tests confirmed that this was because $\underline{S}$ was lower in the drug-free shock than

scopolamine shock ($t_{14} = 4.3, p = .0007$) or barrier ($t_{13} = 2.2, p = .0498$) conditions, which

differed only marginally from one another ($t_{11} = 1.98, p = .0834$). Hence, drug-free avoidance

training and extinction induced more place cell remapping than the scopolamine shock or barrier

training conditions; this remapping occurred at all locations on the short path but a 3x3 repeated

measures ANOVA with training condition (drug-free shock, scopolamine shock, barrier) and

track zone (left safe, center shock, right safe) as factors suggested that remapping was most

pronounced in the center zone where shocks occurred than at L and R ends of the track where

no shocks occurred ($F_{2,40} = 3.0, p = .0596$). Induction of remapping by avoidance training and

extinction did not differ in male versus female rats, nor did the impairment of remapping by

scopolamine (Fig. 3-figure supplement 2). When the analyses of Fig. 3 were repeated on

scopolamine rats (n=2) that successfully retained avoidance despite the injection, the drug was

less effective at inducing place cell remapping (Fig. 3-figure supplement 3), suggesting (but not

proving) that impairment of remapping was causally related to impairment of avoidance

learning. When the analyses were repeated on drug-free rats (n=2) that failed to retain

avoidance despite being trained drug free (Fig. 3-figure supplement 2), one rat showed less and

Figure 3. Remapping of place cells is induced by avoidance learning. A) Top: bar graphs show distributions of intersession intervals between pre-training (pre) and post-extinction (post) session pairs included in the analysis. Bottom: Diagram shows timeline for pre and post sessions. B) Left: Number of subsampled beeline trials did not differ significantly by training condition or session. Right: Running speeds did not differ significantly by training condition; there was a marginal difference between pre vs. post running speeds in the barrier (but not drug-free or scopolamine shock) condition. C) Left: Total number of imaged place cells (recurring and non-recurring) did not differ significantly by training condition; there was a marginal difference between pre vs. post place cell counts in the barrier (but not drug-free or scopolamine shock) condition. Middle: Percentage of all imaged cells (recurring and non-recurring) per session that were classified as place cells. Right: Between-session place cell recurrence ratios did not differ by training condition. D) Heatmap pairs show pre and post tuning curves for recurring place cells that were spatially tuned in the LR (top row) and RL (bottom row) running direction; both heatmaps in each pair are co-sorted by peak locations from the pre session. Pie charts show the proportion of recurring place cells (total number given at top) for which tuning curves were included in LR and RL heatmap pairs. E) Kolmogorov–Smirnov tests show that distributions of place field peaks locations (LR and RL combined) were unchanged between pre and post sessions. F) Mean population vector correlation matrices for the three training conditions. G) Decoding accuracy index (DAI) at each position on the track for each of the three training conditions; '*' indicates locations where decoding was significantly ($p < .05$) less accurate for the drug-free condition than the other two conditions. H) Left column: Bar graphs show means and standard errors of place tuning stability scores (S); results for L, C, & R track zones are shown in top, middle, and bottom rows, respectively. Middle columns: Boxplots show median and range of template-selected population vector correlation bins in each rat. Right column: Templates used to select peri-diagonal correlation values from the L, C, & R track zones. †$p < .1$; *$p < .05$; **$p < .01$; ***$p < .001$.

the other showed more place tuning stability across the entire length of the track when

compared against rats that retained avoidance.

Kolmogorov–Smirnov tests found that place field density distributions (accumulated over

all rats) did not change significantly between the pre versus post sessions in any training

condition, including the drug-free shock condition (Fig. 3E, left). Hence, remapping did not

cause place fields to migrate toward or away from the shock zone (Fig. 3-figure supplement 4),

in contrast with prior reports that remapping can cause place fields to migrate toward locations

where aversive stimuli are encountered (Mamad et al. 2019). All place cell remapping effects

and differences among training conditions were abolished when remapping analyses were

performed on shuffled data (Fig. 3-figure supplement 5).

To analyze the extent of remapping caused by aversive learning, the rat's position on the

track was decoded from place cell calcium activity (see Methods). The decoder was trained on

data from one session (pre- or post-extinction) and tested on data from the other session. The

decoding error in a given position bin was measured as the median of the absolute difference

(in cm) between the rat's actual versus predicted position over all visits to that bin. Wilcoxon

signed-rank tests compared decoding errors obtained with shuffled versus non-shuffled

population vectors; a *decoding accuracy index* (DAI) was then derived for each bin, with DAI=0

corresponding to p=.01 for more accurate decoding from unshuffled than shuffled population

vectors (see Methods). When mean DAI values were compared among training conditions,

significant differences were detected only in two bins at the center of the track (in the shock

zone) where decoding in the drug-free shock condition was significantly less accurate (p<.05)

than in scopolamine shock and barrier training conditions combined (asterisks in Fig. 3D,

bottom). In accordance with results from population vector correlation analysis, these decoding

results support the conclusion that drug-free shocks induced more remapping of the center zone

than shocks while on scopolamine or from barrier training.

Figure 4. Shock-evoked responses of place cells. A) Left column: Frequency distribution of shock responsive place cell percentages in each rat. Middle column: Population-averaged Af/s rates (from all rats combined) for shock responsive versus shock responsive place cells. RIght column: Spatial distribution of place field peak locations during the training session, prior to grid electrification; x-axis shows distance traveled between the start (left) and end (right) of the short path in the current running direction (LR or RL). B) Heatmaps (LR and RL tuning curves combined) are plotted for shock responsive place cells in the drug-free (top) versus scopolamine (bottom) conditions during pre- (left) and post-(right) training sessions (rows are co-sorted on peaks from the pre-training session, columns are sorted so that the rat's running direction goes from left to right for both LR and RL tuning curves); P matrices show pre- vs. post-extinction population vector correlations for shocks given drug-free (top) or on scopolamine (bottom); line graphs show that signed p-values (y-axis, log scale) fall below .01 at center track positions where drug-free shocks induced more remapping than shocks on scopolamine. C) Same as 'B' but for shock non-responsive place cells. ****p<.0001.

Shock-evoked responses of place cells

Shock-induced place cell remapping has been observed in prior studies (Moita et al. 2003; Moita et al. 2004; Wang et al. 2012; Schuette et al. 2020), but the cellular and synaptic mechanisms of such remapping are not well understood. Individual CA1 neurons can form new place fields at locations where they are artificially stimulated or inhibited (Bittner et al. 2015; Geiller et al. 2022), and since some CA1 place cells are naturally responsive to shocks (Moita et al. 2003; Moita et al. 2004), it is possible that individual CA1 neurons might gain or lose place fields at locations where they respond to shocks. If so, then this might partly account for remapping at the population level. If shock-evoked responses of CA1 neurons are involved in remapping, then blockade of remapping by scopolamine could be related to effects of the drug upon these shock-evoked responses. To investigate this, each place cell imaged during a training session was classified as shock responsive if its Af/s rate was significantly higher (p<.01) during entries to the electrified than non-electrified shock zone; otherwise, it was classified as non-responsive to shocks (see Methods).

The mean percentage of shock responsive cells per rat (Fig. 4A, left column) was 35.5±2.3% for drug-free training and 29.6±3.3% for scopolamine training; these percentages did not differ significantly (independent t_{14} =1.58; p=.13). Pre-shock baseline Af/s rates (Fig. 4A, middle column) were higher for shock responsive than non-responsive place cells in both the drug-free (t_{1931} =9.3; p<.0001) and scopolamine (t_{1034} =4.34; p<.0001) conditions, which was attributable to the fact that in both conditions, place field peaks for shock responsive and non-responsive cells tended to be segregated on the 'approach' and 'departure' sides of the shock zone, respectively (Fig. 4A, right column). Thus, as rats approached the shock zone in either running direction, shock responsive cells were more likely than non-responsive cells to be active in their place fields just before shock zone entry.

To compare remapping by shock responsive versus non-responsive CA1 neurons,

analyses were performed on place cells that recurred during pre- and post-extinction sessions

and were also active during the training session (so that each cell could be classified as

responsive or non-responsive to shocks). Pre- and post-extinction heatmaps were derived from

tuning curves of shock responsive (Fig. 4B) and non-responsive (Fig. 4C) place cells. LR and

RL tuning curves were combined together in each heatmap, with RL (but not LR) tuning curves

reflected on the horizontal so that the rat's running direction was oriented from left to right in all

rows; hence, columns to the left or right of center in each heatmap denoted locations where rats

were approaching or departing the shock zone, respectively. From these heatmaps, shock

responsive and non-responsive population vector correlation matrices were derived for the drug-

free (P_{shk+}^{DF}, P_{shk-}^{DF}) and scopolamine (P_{shk+}^{SCOP}, P_{shk-}^{SCOP}) conditions. A gap of low population vector

correlation values in the center of P_{shk+}^{DF} but not P_{shk+}^{SCOP} (Fig. 4B) and of P_{shk-}^{DF} but not P_{shk-}^{SCOP} (Fig.

4C) indicated that drug-free shocks induced remapping of the center zone in both shock

responsive and non-responsive place cell populations, whereas scopolamine shocks failed to

induce remapping in either population. Confirming this, a series of independent t-tests was

performed on clusters of peri-diagonal bins arrayed along diagonals of P_{shk+}^{DF} versus P_{shk+}^{SCOP}, and

P_{shk-}^{DF} versus P_{shk-}^{SCOP} (see Methods); resulting significance values (line graphs in Figs. 4B,C)

dipped well below p<.01 in the center region of the track (but not at the ends), indicating that

drug-free shocks caused shock responsive and non-responsive place cells to remap the center

zone more than scopolamine shocks.

DISCUSSION

Place cell remapping has been shown to occur at locations where animals experience

aversive events such as shocks, predator odors, or attacks (Moita et al. 2003; Moita et al. 2004;

Wang et al. 2012; Wang et al. 2015; Kim et al. 2015; Mamad et al. 2019; Schuette et al. 2020;

Ormond, Serka, and Johansen 2023), but it is not well understood whether such remapping

plays a functional role in storing memories for aversive encounters. Here we investigated this

question by using a large-field-of-view version of the UCLA Miniscope (Guo et al. 2023) to

image population activity of CA1 place cells in rats as they acquired and extinguished a spatial

avoidance learning task. We found that significantly more place cell remapping was induced

under experimental conditions where aversive encounters were remembered (drug-free shocks)

than under control conditions where they were forgotten (shock training on scopolamine), or

where a neutral rather than aversive path-blocking stimulus was encountered (barrier training).

While these results do not definitively prove that place cell remapping is causally necessary for

storing memories of aversive encounters, they provide correlational evidence that remapping

occurs selectively under conditions where a stimulus occurs and is subsequently remembered

rather than forgotten.

Spatial distribution of place fields

Hippocampal place fields are sometimes observed to be preferentially concentrated at

locations where rodents spend large amounts of time, such as the two rewarded locations on

our rectangular maze (Fig. 3E). This makes sense from an information theoretic perspective,

since it is efficient for frequently occurring stimuli to be overrepresented (and conversely for

rarely occurring stimuli to be underrepresented) among preferred stimuli of tuned neurons in a

population code. In accordance with this principle, when animals learn to obtain reward at

specific locations (and thus to spend more time at those locations), a form of remapping occurs

whereby place fields migrate toward rewarded locations (Breese, Hampson, and Deadwyler

1989; Hollup et al. 2001; Dupret et al. 2010; Mamad et al. 2017).

Less is known about how aversive learning affects the spatial distribution of place fields,

but based purely upon information theoretic considerations, one might expect that place fields

should migrate away from (rather than toward) aversively reinforced locations as animals shift

their behavior to spend less time there. Contradicting this intuition, there is evidence to suggest

that place fields migrate toward (rather than away from) aversively reinforced locations (Mamad

et al. 2019). Possibly, overrepresentation of aversively reinforced locations increases the

likelihood that place cells encoding such locations will become active during "replay events" that

occur during sleep (Girardeau, Inema, and Buzsáki 2017), or during waking as the animal

deliberates over whether to approach a remote location where danger has been encountered

(Wu et al. 2017; Ormond, Serka, and Johansen 2023). This may help animals to avoid

dangerous places by anticipating in advance the threats that await them there.

We analyzed place cell remapping after rats first learned to avoid a shocked location,

then extinguished avoidance and resumed spending time at the formerly shocked location. In

agreement with prior findings (Moita et al. 2003; Moita et al. 2004; Wang et al. 2012; Schuette et

al. 2020), we observed that place cell population vectors were less correlated (and thus showed

more remapping) in rats that underwent acquisition and extinction of avoidance than in control

rats that encountered a neutral barrier or failed to retain avoidance after being trained under the

influence of scopolamine (Fig. 3F-H). In contrast with (Mamad et al. 2019), place cells did not

shift their center of mass toward the site of the aversive stimulus; instead, the spatial distribution

of place fields remained unchanged by remapping (Fig. 3E), indicating that place fields shifted

toward and away from the shocked location in equal proportions.

The discrepancy in place field migration findings might be related to our use of calcium

imaging rather than electrophysiology to record neural activity; mChARs can regulate calcium

entry into cells via a wide range of mechanisms (Brown 2018) and thus scopolamine may have

different effects on spike detection from calcium traces versus electrophysiological recordings.

The discrepancy might also be a consequence of using different aversive stimuli (shock versus

noxious odor), different degrees of fear acquisition and extinction during the task, or differences

in the spatial layout of the maze. Rats in our experiment only received 1 or 2 self-initiated

shocks during the shock sessions, thus a stronger conditioning paradigm might be necessary to

induce place field migration. Another possible source of discrepancy could be different methods

for normalizing speed and occupancy across experimental conditions. Aversive events cause

lasting behavioral changes that alter the manner in which an animal samples its spatial

environment (including reduced movement speed and avoidance of locations where aversive

stimuli have been encountered). To control for confounding effects of these learning-induced

behavior changes upon measures of place cell remapping, studies rely upon subsampling

methods to normalize speed and occupancy distributions from experimental sessions conducted

before versus after encounters with aversive events (Moita et al. 2003; Moita et al. 2004; Wang

et al. 2012; Schuette et al. 2020). Here, we minimized confounds of behavioral sampling by

restricting data analysis to "beeline trials" during which rats ran highly stereotyped trajectories

on a linear track, so that behavior was matched as closely as possible between sessions

conducted before acquisition versus after extinction of avoidance learning. Using this approach,

we replicated prior findings that aversive learning induces place cell remapping (Moita et al.

2003; Moita et al. 2004; Wang et al. 2012; Wang et al. 2015; Kim et al. 2015; Mamad et al.

2019; Schuette et al. 2020; Ormond, Serka, and Johansen 2023) but found no change in the

spatial distribution of place fields.

Acute effects of scopolamine

Significantly more place cell remapping occurred when aversive encounters were

remembered (as assessed by 48 h avoidance expression) after drug-free training than when

they were forgotten after training on scopolamine. This suggests that place cell remapping might

play a functional role in storing memories of aversive encounters, but potential confounds must

be considered. A trivial explanation for our finding that scopolamine impaired both avoidance

learning and place cell remapping could be that, at the dose given, the drug impaired rats'

awareness of their surroundings and their ability to perceive shock stimulus during the training

session. This interpretation is not well supported by the data for several reasons. First,

scopolamine did not acutely affect median running speeds during beeline trials (Fig. 2C),

consistent with previous demonstrations that 1 mg/kg scopolamine does not impair locomotion

in rodents (Sun, Unnithan, and French 2021; Anagnostaras et al. 1999). Second, scopolamine

did not acutely impair immediate avoidance of the shocked location during training (Fig. 1F),

consistent with prior evidence that mAChRs are not necessary for immediate shock avoidance

(Decker, Tran, and McGaugh 1990). Third, under the influence of scopolamine, place cells

retained stable within-session spatial tuning (Fig. 2F-I) and exhibited shock-evoked responses

similar to those observed in the drug-free condition (Fig 4A). By contrast, a recent calcium

imaging study in mice (Sun, Unnithan, and French 2021) reported that tuning properties of place

cells on a linear track were acutely disrupted by systemic scopolamine injections at the same

dosage used here (1 mg/kg) (but see also Brazhnik et al. 2004). More consistent with our

current findings, electrophysiology studies in rats have reported that systemic ACh antagonists

alter temporal firing properties of place cells, such as their synchronization with hippocampal

theta rhythm, while largely preserving their spatial firing properties (Douchamps et al. 2013;

Newman et al. 2017). Conversely, cholinergic activation of the hippocampus acutely alters EEG

states by promoting theta oscillations and suppressing sharp-wave ripples (Zylla et al. 2013;

Vandecasteele et al. 2014; Hunt et al. 2018; Ma et al. 2020; Jarzebowski et al. 2021).

Therefore, it is possible that theta oscillations and sharp waves were acutely altered by

scopolamine during our experiments, but these signals were not detectable using our calcium

imaging methods, so it was not possible to analyze these phenomena in the current study.

Although rats in our study were still able to locomote through the environment and keep

track of their own location while on scopolamine, some acute effects of the drug upon behavior

and place cell activity were observed. For example, while on scopolamine, rats showed a

significant reduction of their previously learned preference for the short path (Fig. 1F) prior to

any delivery of shocks. This suggests that scopolamine may have affected the animals'

motivation to seek reward on the maze or impaired recall of previously learned strategies for

finding the reward, consistent with prior evidence that scopolamine can impair navigational

learning and decision making (Nieto-Escámez, Sánchez-Santed, and Bruin 2002; Vales and

Stuchlik 2005; Huang et al. 2011; Svoboda, Popelikova, and Stuchlik 2017; Rashid and Ahmed

2019; Hales et al. 2020). Although place cells continued to exhibit spatial tuning while rats were

on scopolamine, between-session place cell recurrence probabilities were reduced (Fig. 2G)

and population vectors showed degraded between-session similarity with drug-free maze visits

on prior days (Fig. 2H,I); interestingly similar impairment of between-session place cell stability

has been reported in aged rats (Barnes et al. 1997).

By selectively degrading between-session (but not within-session) stability of place cells,

scopolamine may have prevented CA1 from retrieving accurate memory representations of the

familiar maze environment during the training session. If so, then this could help to explain why

rats acutely stopped preferring the short path while on scopolamine; expressing the learned

preference for the short path may depend upon recalling stored maze representations that were

previously active when the preference was learned. It may also help to explain why scopolamine

prevented the aversive shock experience from inducing long-term remapping of place cells,

since failure to recall previously stored maze representations during training may have shielded

those representations from modification by the aversive experience, and thus from remapping.

Effects of scopolamine on memory storage and retrieval

Many studies have shown that ACh receptor antagonist drugs can impair hippocampal

memory processing in animal models (Decker, Tran, and McGaugh 1990; Anagnostaras et al.

1999; Atri et al. 2004; Hasselmo and McGaughy 2004; Green et al. 2005; Hasselmo 2006;

Huang et al. 2011; Dannenberg, Young, and Hasselmo 2017; Solari and Hangya 2018).

Conversely, drugs that upregulate ACh neurotransmission can facilitate learning and memory

(Barten and Albright 2008; Digby, Shirey, and Conn 2010; Ragozzino et al. 2012) and are

commonly prescribed to treat aging-related amnesia and dementia in human patients

(Lombardo and Maskos 2015). Here, rats that received avoidance training on scopolamine were

impaired when tested for retention 48 h later (Fig. 1H), consistent with prior studies showing that

mAChRs are necessary for acquisition of aversive learning (Decker, Tran, and McGaugh 1990;

Anagnostaras, Maren, and Fanselow 1995; Anagnostaras et al. 1999; Wallenstein and Vago

2001). In conjunction with blocking avoidance learning, scopolamine also blocked place cell

remapping between pre-training and post-extinction sessions (Fig. 3D-H). Prior studies have

similarly shown that scopolamine interferes with the ability of place cells to form and update

cognitive maps across repeated visits to the same environment (Douchamps et al. 2013), but to

our knowledge, the present study is the first to show that impairment of memory for an episodic

event (in this case an aversive shock) by scopolamine is accompanied by disruption of place

cell remapping that normally occurs in response to such an event.

Since rats both acquired and extinguished avoidance behavior during the timespan over

which remapping was measured, it was not possible to fully dissociate whether place cell

remapping in drug-free rats was driven mainly by acquisition, extinction, or both. Rats trained on

scopolamine did not undergo extinction (since they failed to exhibit long-term retention of

avoidance behavior in the first place), so if remapping in drug-free rats was driven mainly by

extinction, then scopolamine may have impaired remapping not by disrupting learning during the

training session but instead by depriving rats of the extinction experience that would otherwise

have caused remapping to occur after training. Prior evidence indicates that acquisition and

extinction of conditioned fear can both contribute to place cell remapping (Moita et al. 2004;

Wang et al. 2012; Wang et al. 2015), so it is likely that impairment of remapping by pre-training

injections of scopolamine was at least partly caused by disruption of experience-dependent

changes to the hippocampal population code during the training session.

Manipulations that impair ACh neurotransmission have been found to selectively impair

storage but not retrieval in certain memory tasks, such as remembering semantic information in

humans (Atri et al. 2004; Hasselmo and McGaughy 2004) and fear learning in rodents (Decker,

Tran, and McGaugh 1990; Huang et al. 2011). Hippocampal synaptic plasticity is likewise

thought to be more important for storage than retrieval of hippocampal-dependent memory

tasks (Martin, Grimwood, and Morris 2000). Hippocampal plasticity is sensitive to mAChR

antagonists (Drever, Riedel, and Platt 2011) and has been implicated in place cell remapping

(Dragoi, Harris, and Buzsáki 2003), so it is possible that in our study, scopolamine impaired

avoidance learning and place cell remapping by interfering with synaptic plasticity in the

hippocampus, and thus preventing the storage of memories for the aversive shock experience.

Some CA1 place cells are excited by shocks (Moita et al. 2003; Moita et al. 2004), and it has

been shown that individual CA1 neurons can form new place fields at locations where they are

artificially excited or inhibited (Bittner et al. 2015; Schoenenberger, O'Neill, and Csicsvari 2016;

Geiller et al. 2022; McKenzie et al. 2021). This raises the possibility that during acquisition of

fear learning, shock-evoked responses of CA1 neurons might trigger synaptic plasticity that

leads to place cell remapping. If scopolamine disrupted shock-evoked hippocampal plasticity, it

likely did so by impairing mechanisms downstream from neural responses to shock, since we

found that scopolamine had no effect on shock-evoked responses of CA1 place cells and

shock-induced remapping was similarly prevalent in subpopulations of shock responsive and

non-responsive place cells.

Since the drug was given systemically, scopolamine may have interfered with plasticity

not only in the hippocampus but also in other brain regions that contribute to the storage of fear

memories, such as the amygdala, where associations between predictive cues and aversive

events are thought to be stored by synaptic plasticity that is both triggered by noxious stimuli

(Blair et al. 2001; Grewe et al. 2017; d'Aquin et al. 2022) and susceptible to cholinergic

modulation (Izquierdo et al. 1992; Jiang et al. 2016). Regardless of whether scopolamine

disrupted place cell remapping via its effects upon synaptic plasticity or other mechanisms (such

as retrieval of previously stored maze responsentations; see above), or whether the drug acted

upon pharmacological targets inside versus outside of the hippocampus (or both), our findings

indicate that the drug disrupted shock-induced place cell remapping and thus support the idea

that reorganization of the hippocampal place code occurs in conjunction with hippocampal

storage of episodic memories for aversive events. Where and how scopolamine acts in the brain

to impair remapping is a worthwhile question for further research that could help to elucidate the

broader role of acetylcholine in memory storage and retrieval.

Implications for brain disease and aging

If remapping is a mechanism for memory, then disruption of remapping by systemic

cholinergic antagonists may serve as a useful animal model for memory loss accompanied by

degeneration of cholinergic signaling in dementia (Wenk, Bigl, and Meyer 1980; Arendt et al.

1983; Hardy 2006; Schliebs and Arendt 2011). Degradation of cholinergic signaling also occurs

with alcohol abuse disorder (Arendt 1994; Miller and Kamens 2020). By further investigating

how the cholinergic neuromodulatory system interacts with the hippocampus, we may develop a

deeper understanding of how memory systems are impacted by a variety of neurodegenerative

insults and make progress toward improved therapies for memory diseases.

Like humans, rodents show memory deficits with aging (C. A. Barnes 1979; C. A.

Barnes, Nadel, and Honig 1980). Interestingly, prior studies have shown that place cell

population dynamics in aged rats are altered in ways similar to those observed here under

scopolamine. For example, we found that scopolamine degraded between- but not within-

session stability of the CA1 population code, which has also been observed in aged rats (Carol

A. Barnes et al. 1997). This is in contrast to other brain diseases, such as epilepsy, which

degrades both within- and between-session place coding (Shuman et al. 2020). Additionally, we

found that in rats that showed impaired avoidance learning after pre-training injections of

scopolamine, the drug concurrently prevented CA1 place cells from remapping during learning

and extinction (Figs. 3,4). Place cells in aged rodents have likewise been reported to show

deficits in experience-dependent modification of their tuning properties (Oler and Markus 2000;

I.A Wilson et al. 2003; Iain A Wilson et al. 2004; Tanila et al. 2018). These parallels between the

effects of aging and scopolamine upon place cells suggest that mAChR blockade and aging

may produce similar memory deficits by causing similar impairments to hippocampal population

stability. Further study is warranted to investigate whether such impairments of hippocampal

population coding can be reversed using anti-amnestic drugs, such as those that upregulate

ACh transmission or other novel compounds. By identifying interventions that can improve the

long-term stability of hippocampal population coding in animal models, it may be possible to

make progress toward improved therapies for memory deficits associated with aging and

neurodegenerative disease.

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METHODS

All experimental procedures were approved by the Chancellor's Animal Research Committee of the University of California, Los Angeles, in accordance with the US National Institutes of Health (NIH) guidelines, protocol #2017-038. Raw data files and analysis code are openly available at https://github.com/tadblair/tadblair/tree/Blair_et_al.

Subjects and Apparatus

15 Long Evans rats (8F, 7M) acquired from Charles River at 3 months of age were used in this experiment. Subjects were singly-housed within a temperature and humidity controlled vivarium on a 12 hour reverse light-cycle. Beginning at 4 months of age, rats were reduced 85% of their ad lib weight (450-550 g) through limited daily feeding, and then underwent virus injection and GRIN lens implantation surgery (see below). After 5d recovery from surgery, rats began a regimen of behavior and recording sessions on the rectangular maze. The maze was constructed from a 4" wide track composed from metal grid bars along the full length of the long and short paths. The track was elevated 20" from the floor. There were extramaze cues in the surrounding environment (doors, shelving, checkerboard poster on the wall) but no local cues on the maze to indicate the shock zone's location. During shock training sessions (drug-free or scopolamine), a 1.0 mA current from a grid scrambler (Med Associates Corporation, Fairfax, VT) was switched on to electrify grid bars in a 50 cm segment of the short path's center (Fig. 1A). The exact duration of the rat's exposure to shock varied with the length of time it spent in contact with the electrified bars as it ran across or retreated from the shock zone, but was typically very brief (on the order of 1 s or less).

Fig. 1B shows the order in which experimental manipulations were given to different groups of rats. 6 rats (3 male, 3 female) received drug-free shock training as their first manipulation after reaching criterion for preferring the short path. Of these, 5 rats (3 male, 2

female) then received extinction sessions and ended the experiment. The remaining female rat

was mistakenly given shock at the start of the drug-free training session (rather than after 10

min), and was thus dropped from imaging analysis of the drug-free shock condition, but after

extinction, this rat subsequently received scopolamine shock training and was thus included in

analysis for that condition (see below). 4 rats (2 male, 2 female) were given their drug-free

shock sessions after previously receiving barrier training as their first manipulation (so these

rats were still naive to shock during drug-free shock avoidance training). 3 rats (1 male, 2

female) were given their drug-free shock session after previously receiving a scopolamine shock

session (followed by extinction) as their first manipulation, and 2 rats (1 male, 1 female) were

given their drug-free shock session after previously receiving both barrier training and a

scopolamine shock session (followed by extinction). Among rats that were given scopolamine

shock sessions, 5 (2 males, 3 females) had not previously experienced shock at the time of their

scopolamine shock avoidance training session, whereas 5 rats (3 males, 2 females) had

previously received drug-free shock avoidance training followed by extinction. To compare

imaging results across training conditions, rats from the drug-free condition were only included if

they ran <60% as many trials during their first post-extinction session as during their final pre-

training session (indicating that they had successfully acquired short path avoidance), whereas

rats from the scopolamine shock condition were only included if they ran >60% as many trials

during their first post-extinction session as during their final pre-training session (indicating that

scopolamine had successfully blocked avoidance learning).

Surgical procedures

Subjects were given two survival surgeries prior to behavior training in order to record

fluorescent calcium activity from hippocampal CA1 cells. During the first surgery, rats were

anesthetized with 5% isoflurane at 2.5 L/min of oxygen, then maintained at 2-2.5% isoflurane

while a craniotomy was made above the dorsal hippocampus. Next, 1.2 uL of AAV9-Syn-

GCamp7s (AddGene) was injected at .12uL/min just below the dorsal CA1 pyramidal layer (-3.6

AP, 2.5 ML, 2.6 DV) via a 10uL Nanofil syringe (World Precision Instruments) mounted in a

Quintessential Stereotaxic Injector (Stoelting) controlled by a Motorized Lab Standard Stereotax

(Harvard Apparatus). Left or right hemisphere was balanced across all animals. One week later,

the rat was again anesthetized and 4 skull screws were implanted to provide stable hold for the

subsequent implant. The viral craniotomy was reopened to a diameter of 1.8 mm, and cortical

tissue and corpus callosal fibers above the hippocampus were aspirated away using a 27 and

30 gauge blunt needle. Following this aspiration, and assuring no bleeding persisted in the

craniotomy, a 1.8mm diameter Gradient Refractive INdex lens ("GRIN lens", Edmund Optics)

was implanted over the hippocampus and cemented in place with methacrylate bone cement

(Simplex-P, Stryker Orthopaedics). The dorsal surface of the skull and the bone screws were

cemented with the GRIN lens to ensure stability of the implant, while the dorsal surface of the

implanted lens was left exposed.

Two to three weeks later, rats were again placed under anesthesia in order to cement a

3D printed baseplate above the lens. First a second GRIN lens was optically glued (Norland

Optical Adhesive 68, Edmund Optics) to the surface of the implanted lens and cured with UV

light. The pitch of each GRIN lens is .25, so implanting 2 together provides a .5 pitch. This half

pitch provides translation of the image at the bottom surface of the lenses to the top while

maintaining the focal point below the lens. This relay implant enables access to tissue deep

below the skull surface. Magnification of the image is performed by the miniscope. The

miniscope was placed securely in the baseplate and then mounted to the stereotax to visualize

the calcium fluorescence and tissue. The baseplate was then cemented in place above the relay

lenses at the proper focal plane and allowed to cure. Once rats had been baseplated, they were

placed on food restriction to reach a goal weight of 85% *ad lib* weight and then began

behavioral training. Time between the beginning of the surgical procedures and the start of

behavior training was typically 6-8 weeks.

Scopolamine Injections

800   On some training days rats were injected intraperitoneally with vehicle (0.9% sterile

801 saline) or scopolamine (1 mg/kg dissolved at a concentration of 1 mg/ml) prior to shock

802 sessions. After the injection, rats were returned to their home cage for 30 min (located on a cart

803 in the experiment room) for 30 min prior to starting the session.

Histology

At the end of the experiment, rats were anesthetized with isoflurane, intraperitoneally

injected with 1 mL of pentobarbital, then transcardially perfused with 100 mL of 0.01M PBS

followed by 200 mL of 4% paraformaldehyde in 0.01M PBS to fix the brain tissue. Brains were

sectioned at 40 μm thickness on a cryostat (Leica), mounted on gelatin prepared slides, then

imaged on a confocal microscope (Zeiss) to confirm GFP expression and GRIN lens placement

(see example in Fig. 2A).

MiniLFOV calcium imaging system

To record calcium activity during unrestrained behavior, we utilized a large field of view

version of the Miniscope imaging system, "MiniLFOV" (Guo et al. 2023). This open-source

epifluorescence imaging camera weighs 13.9 g with a 3.6 x 2.3 mm field of view while

maintaining a 2.5 um resolution at the center of view and 4.4 um at the periphery. The

MiniLFOV can record at 22 Hz, has a 5MP CMOS imaging sensor, an electrowetting lens for

digitally setting the focal plane, and a modular lens configuration to enable a longer working

distance (either 1.8 mm, used here, or 3.5 mm). The system is 20x more sensitive than previous

v3 Miniscopes, and twice as sensitive as current v4 Miniscopes. Power, communication, and

image data are packed into a flexible 50 Ω coaxial cable (CW2040-3650SR, Cooner Wire) using

power-over-coax filtering and a serializer/deserializer pair for bi-directional control

communication with I^2C protocol and uni-directional high bandwidth data streaming. The

MiniLFOV device interfaces with UCLA open-source Miniscope DAQ Software to stream,

visualize and record neural dynamics and head orientation data. This DAQ platform and

software allow for excitation intensity adjustment, focus change, image sensor gain selection,

and frame rate setting. See the MiniLFOV website (github.com/Aharoni-Lab/Miniscope-LFOV)

and methods paper (Guo et al. 2023) for further details and printable part files.

Trace extraction and spike inference

To accelerate image processing, each session's video stack was cropped to a rectangle

that excluded edge regions containing no neural activity in any session from that animal. The

stack was also temporally downsampled to every other frame, yielding an effective sample rate

of ~11 Hz. Non-rigid motion correction was applied to remove residual non-uniform tissue

motion artifacts (Pnevmatikakis and Giovannucci 2017). Source extraction was then performed

using the python implementation of the Calcium Imaging Analysis package, '*CaImAN*'

(Giovannucci et al. 2019), yielding individual spatial contours and demixed temporal

fluorescence traces for each detected neuron; parameters used for source extraction can be

found in scripts provided at https://github.com/tadblair/tadblair/tree/Blair_et_al. Deconvolved

spikes were derived from denoised fluorescence traces by CaImAN 's 'deconvolveCa' function

using a second order autoregressive model with automated estimation of baseline and

convolution kernel parameters. To match cell contours across sessions, spatial contour weights

from CaImAn were thresholded to 50% of their peak value to generate binary pixel masks which

were then analyzed using *CellReg* (Sheintuch et al. 2017) for matching contours across all

sessions included in the analysis for each rat.

Behavior tracking

A webcam mounted in the behavior room tracked a red LED located on the top of the

miniscope and this video was saved alongside the calcium imaging via the miniscope software

with synchronized frame timestamps. These behavior video files were initially processed by

custom python code, where all the session videos were concatenated together into one tiff

stack, downsampled to 15 frames per second, the median of the stack was subtracted from

each image, and finally they were all rescaled to the original 8-bit range to yield the same

maximum and minimum values before subtraction. Background subtracted behavior videos

were then processed in MATLAB. The rat's position in each frame was determined using the

location of the red LED on the camera. Extracted positions were then rescaled using an inverted

model of the empirically measured camera distortion and convert the pixel position to

centimeters according to the maze size. Positional information was then interpolated to the

timestamps of the calcium imaging video.

Behavioral performance measures and criteria

During the initial phase of training where rats learn to prefer the short path over the long

path on the maze, rats were required to reach a criterion of earning at least 2 rewards per

minute by taking the short path and earning at least twice as many rewards on the short versus

long path during a single 15 min session. After reaching this criterion, rats were given avoidance

training in the next session as explained in Results. After a single avoidance training session,

rats received standard (non-shock) sessions until avoidance behavior was extinguished. Rats

were considered to have extinguished avoidance behavior on the first post-training day that they

recovered to the original criterion (defined above) for preferring the short path.

48 h retention of short path avoidance was quantified by the ratio $A = N_{+48}/N_{-48}$, where

N_{+48} and N_{-48} are the number of rewards earned on the short path during the first session after

and before the training session, respectively. Hence, lower values of A indicated better 48 h

retention of short path avoidance. In the drug-free shock condition, an individual rat was

considered to show failed 48 h retention of short path avoidance if $A>0.6$, which was the cutoff

for the top quartile of A values in all rats (n=11) from which imaging was successfully obtained

during the drug-free shock condition (of the 14 rats that received drug-free shock training, one

was omitted from imaging analysis because shocks were mistakenly delivered at the start of the

session, and two were omitted because they failed to ever reach criterion for extinction of

avoidance behavior after multiple post-training sessions). In the scopolamine shock condition,

an individual rat was considered to show successful 48 h retention of short path avoidance if

$A<0.31$, which was the cutoff for the bottom quartile of A values in all rats (n=9) from which

imaging was successfully obtained during the scopolamine shock condition (of the 10 rats that

received scopolamine shocks, one was omitted from imaging analysis because the GRIN lens

shifted and imaging was lost prior to the avoidance training session).

Behavioral downsampling

Since behavior on the maze was altered by training and drug administration (Fig. 1),

imaging analyses only included data from beeline trials during which rats ran from one end of

the track to the other with no change in direction (a beeline trial was defined as a continuous

traversal of the short path throughout which the rat's 1D position along the track changed

monotonically in one direction). This assured that rats were behaving as similarly as possible

during analyzed data segments from all sessions. Beeline trials from each session were then

further downsampled prior to imaging analyses, to control for potential effects of behavior or

sampling differences on the outcome of imaging analyses.

For comparisons of pre-training versus training sessions (Fig. 2C-H) in the drug-free

versus scopolamine shock conditions, it was necessary to correct for two imbalances in

behavior between the drug-free versus scopolamine conditions (Fig. 2-figure supplement 2,

Panel A): 1) rats ran more slowly (and thus also ran fewer beeline trials) during training sessions

on scopolamine than drug-free, and 2) rats ran more pre-training trials per session in the

scopolamine than the drug free condition. To correct for imbalance #1, we computed that the

mean number of beeline trials (in the LR and RL directions combined) during training in the

scopolamine condition (23.85 trials) was 64% of mean beeline trials in the drug-free condition

(37.22 trials); since running speed was also slower for scopolamine training, we sorted each

rat's drug free training beeline trials in order of running speed, and omitted the fastest 36% of

trials from each running direction (rounded to the nearest integer). This simultaneously

equalized beeline trial counts and running speeds during training sessions between the drug-

free versus scopolamine conditions. To correct for imbalance #2, we computed that the mean

number of beeline trials during pre-training (in the LR and RL directions combined) in the drug-

free condition (53.78 trials) was 71% of mean beeline trials in the drug-free condition (76.0

trials); we omitted the last 29% of trials from each running direction in the pre-training session.

This equalized trial counts across training conditions without significantly affecting mean running

speeds. Also, since omitted pre-training trials were removed from the end of the session, the

effective duration of pre-training sessions was reduced from 15 min down to a shorter duration

that was more similar to the 10 min pre-shock period during which beeline trials were sampled

from the training session. After these behavioral downsampling measures had been taken, rats

in both conditions still ran fewer beeline trials during the analyzed portion of the training session

(the first 10 min prior to shock delivery) than during the included portion of the pre-training

session, as a consequence of the fact that scopolamine rats ran fewer trials in the training

session and this low trial count had to be matched in the drug-free group. But the difference in

trial counts between pre-training and training sessions, though large, was similar for the drug-

free and scopolamine training conditions, and was therefore assumed not to be a confound in

analyses of the training condition effects (Figs. 2C-H).

For comparisons of pre-training versus post-extinction sessions (Fig. 3), it was

necessary to correct for two imbalances in behavior between the drug-free versus scopolamine

conditions (Fig. 2-figure supplement 2): 1) rats in the drug-free shock condition tended to run

fewer laps at lower speeds during the post than pre session (presumably because they still had

some residual fear of the short path even after reaching the extinction criterion) whereas rats in

the barrier condition tended to run more laps at higher speeds during the post than the pre

session (presumably because they continued to acquire a preference for the short path without

being interrupted by avoidance learning), and 2) during both the pre and post sessions, rats in

the scopolamine shock condition ran more short path beeline trials than rats in the other two

conditions. Imbalance #1 was corrected in two steps. The first step was to take the difference

between the number of pre and post session trials for each rat in the drug free condition:

$N_{diff}^{DF} = N_{pre}^{DF} - N_{post}^{DF}$. Trials from the pre session were then sorted by speed, and the fastest

N_{diff}^{DF} trials were removed from the pre session, which equalized the number of pre and post

session beeline trials while also making their mean running speeds much more similar, so that

there was no longer a significant running speed difference (Fig. 3B, right). The second step was

to take the difference between the number of pre and post session trials for each rat in the

barrier condition: $N_{diff}^{BAR} = N_{post}^{BAR} - N_{pre}^{BAR}$. Trials from the post session were then sorted by speed,

and the fastest N_{diff}^{BAR} trials were removed from the post session, which equalized the number of

pre and post session beeline trials while also making their mean running speeds much more

similar, so that there was no longer a significant running speed difference (Fig. 3B, right). Then

an additional 20% of the fastest trials were removed from both sessions to render the mean

number of beeline trials per session similar across the drug-free and barrier conditions. To

correct for imbalance #2, the absolute difference was taken between the number of pre and post

session trials for each rat in the drug free condition: $N_{diff}^{SCOP} = |N_{pre}^{SCOP} - N_{post}^{SCOP}|$; trials from the

scopolamine shock condition were then sorted by running speed, and the number of pre and

post trials was equalized by removing fastest N_{diff}^{SCOP} trials from whichever session had fewer

trials in a given rat. Then an additional 25% of the fastest trials were removed from both

sessions to render the mean number of beeline trials per session similar across the drug-free

and scopolamine shock conditions.

Spatial tuning properties

To analyze acute effects of scopolamine on spatial tuning properties, we measured each

peak Af/s rate, out-of-field Af/s rate, tuning width, and spatial information content from place cell

tuning curves. Peak Af/s rate was defined as the maximum Af/s rate observed for any of the 23

bins in the tuning curve. Out-of-field Af/s rate was defined as the mean rate in the subset of

spatial bins that had rates <50% of the peak Af/s rate. Tuning width was computed as $W \times N_{50}$,

where W=10.8 cm is the width if each bin and N_{50} is the number of bins that had rates >50% of

the peak Af/s rate. Spatial mutual information content was measured in units of bits per active

frame (bits/Af), which was computed as (Skaggs et al. 1993):

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$$-r_{\underline{B}}log_2(r_{\underline{B}}) + \sum_{i=1}^{B} r_i p_i log_2(r_i)$$

961 where B is the set of spatial bins with non-zero mean Af/s rates, r_i is the mean Af/s rate in

962 spatial bin i, p_i is the probability that the rat was occupying bin i, and $r_{\underline{B}}$ is the mean Af/s rate

963 over all bins in the set B.

Selection of place cells

Stable place cells were identified in each imaging session by randomly splitting the

session's beeline trials (separately for each running direction) into two halves, and then

computing two spatial tuning curves, one for each randomly chosen half of the beeline trials. A

Pearson correlation coefficient (R) between these two tuning curves was then computed, and its

p-value was recorded. A cell was classified as a place cell in a given session if the median p-

value for R was <.01 over 200 iterations of random beeline splits in either of the two running

directions. To be classified as a place cell, a neuron also had to generate an average of at least

one inferred spike per beeline trial, and was not permitted to fire at >70% of its peak inferred

spike rate over more than 50% of the track (this was done to exclude likely interneurons, and

thereby restrict analysis to putative pyramidal neurons).

Recurring place fields

A recurring place field was defined as an LR or RL tuning curve that: 1) met the spatial

selectivity criteria described above during at least one of the two sessions, and 2) contained

active frames (that is, generated at least one inferred spike) during subsampled beeline trials in

the specified running direction from both sessions. Place cell tuning curves were excluded from

the between-session heatmap pair if they contained *non-recurring place fields*, defined as those

that: 1) met spatial selectivity criteria in during one of the two sessions, but 2) generated no

inferred spikes (and thus went completely undetected) during subsampled beeline trials in the

specified running direction from the other session.

987   Shock responses

To analyze shock-evoked responses of CA1 place cells, we identified each frame during

which the rat made an entry into the electrified shock zone, and noted in which direction the rat

was running when shock onset began. For each entry into the shock zone, a significant shock-

evoked response occurred when the count of active frames in a 2 s window after entry into the

electrified shock zone exceeded the confidence limit (p<.01) for a Poisson fit to the baseline

distribution of active frame counts from prior visits (during the first 10 min of the session) to

unelectrified shock zone in the same running direction. Most rats experienced the shock in both

running directions before they began to avoid the short path, but a few rats only experienced the

shock in one direction, and in a small number of rats, more than two shocks were received

before the rat began avoiding the short path. To equalize statistical power for detecting shock

responses across all rats (regardless of the shocks they received), a neuron was classified as

shock responsive only if it exhibited a significant shock response during the rat's first or second

visit (or both) to the electrified shock zone; later visits to the shock zone were not considered, so

that place cells would not have a higher likelihood of being classified as shock responsive in rats

that made more visits to the shock zone.

To compute population averaged responses of place cells to shocks, only the first

significant shock-evoked response of each shock responsive place cell (from either the first or

second shock encounter, whichever yielded the first significant response) was included in the

average, so that each cell's contribution to the average always came from a single significant

shock response trial, and therefore did not vary depending upon the number of shocks different

animals received. For shock responsive place cells, the population average included only the

first shock encounter (which by definition failed to elicit a significant shock response), so that

again, each cell's contribution to the average came from a single trial.

Distributions of tuning curve peak locations (Fig, 7A, right) for shock responsive place

cells included only tuning curve peaks for the running direction in which the rat received its first

significant shock-evoked response (from either the first or second shock encounter, whichever

yielded the first significant response). Distributions of tuning curve peak locations for shock

responsive place cells included tuning curve peaks from both running directions if the rat

received its first and second shock in different directions, but if the first and second shock were

both received in the same running direction, then only the tuning curve peak for that direction

was included in the distribution of peak locations.

To assess whether a place cell's tuning curve peak was on the approaching versus

departing side of the shock zone, the 23-bins of the tuning curve were reflected across the

middle (bin 13) for RL tuning curves, but not for LR tuning curves, so that for both running

directions, tuning curve peaks in bins 1-12 were on the approaching side of the shock zone, and

peaks in bins 14-23 were on the departing side of the shock zone. For tuning curves with peaks

in the center (bin 13), an additional step was taken to measure the percentage of the area under

the tuning curve on either side of center. If the area in bins 1-12 was greater than in bins 14-23,

then the tuning curve peak was considered to be on the approaching side, otherwise, it was

considered to be on the departing side.

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Position decoding

For decoding analysis, frame positions in one imaging session were predicted using

place cell heatmaps derived from a different session. For each analyzed pair of pre- and post-

sessions, decoding was performed using memoryless Bayesian inference (Davidson,

Kloosterman, and Wilson 2009; Zhang et al. 1998) with place maps constructed from each

running direction. Post-training positions were predicted from pre-training heatmaps (pre→post),

and vice versa (post→pre), taking the maximally likely position bin at each frame, and results

from both directions were averaged together to obtain the results in Fig. 3G. A subsampling

procedure was used to equalize the number of place cells used for each running direction (LR

and RL, 100 subsample iterations), so that any observed differences in decoding performance

were not attributable to differences in the sizes of the population vectors (that is, the number of

recurring place cells) that were used for pre-training versus cross-training decoding.

The *decoding error* (DE) in each frame was computed as the absolute error (in cm)

between the rat's actual versus predicted position in that frame, DE=abs(x_{act}-x_{pred}). The *shuffled*

decoding error (DE$_{shuffle}$) was derived in the same manner, except that place cells in the

population vector were randomly permuted against their place maps before deriving the

predicted position, x_{pred}. Shuffled decoding error was computed multiple times per frame using

different random place cell permutations (n=2000 shuffles). The decoding error in a given

position bin was measured as the median of the absolute difference (in cm) between the rat's

actual versus predicted position over all visits to that bin. The non-shuffled decoding error was

subtracted from the median DE$_{shuffle}$ to obtain the *accuracy gain* in each frame,

G=median(DE$_{shuffle}$)-DE, which measured by how many cm the decoding error was reduced

when position was predicted from non-shuffled rather than shuffled population vectors. The

median value of *G* was derived for all frames in each actual position bin, *x*, yielding an accuracy

gain measurement at each position on the track, *G*(*x*)=median(G_{x1}, G_{x2}, … G_{xn}). Accuracy gain

was computed separately at each position in each running direction (LR and RL) and each

decoding direction (pre→post and post→pre) to obtain four different G(*x*) values per session

pair: $G_{pre \to post}^{LR}(x)$, $G_{pre \to post}^{RL}(x)$, $G_{post \to pre}^{LR}(x)$, and $G_{post \to pre}^{RL}(x)$. For statistical analyses, each of

these four values was treated as an independent observation at each position. Accuracy gains

tended to be greater near the ends of the track than in the middle, but this was mainly a

consequence of bias in the G(*x*) accuracy measure, rather than better decoding at the ends of

the track, because greater reductions of G(*x*) were attainable near the ends of the track, where

x_{act}-x_{pred} could span the entire track length, than in the middle, where x_{act}-x_{pred} could only span

half the track length at most. To eliminate this positional bias, we used Wilcoxon signed-rank

tests (rather than subtraction) to test whether the median error difference, $DE_{shuffle}$-DE, differed

significantly from zero (see Methods). P-values from the Wilcoxon tests provided a measure of

decoding performance that was unbiased by track position, but these P-values were non-

normally distributed, and thus could not be analyzed to test for interaction effects using

parametric statistics. We therefore took the negative log of the Wilcoxon P value and multiplied

by the sign of *G* to derive a *decoding accuracy index*, DAI = $-\log_{10}$ (P) x sign(*G*) - 2, which was

both normally distributed and unbiased by track position. Subtracting 2 at the end of the

expression was done so that DAI = 0 would correspond to p=.01 for more significant decoding

from unshuffled than shuffled population vectors, since $\log_{10}$ .01=2.

SUPPLEMENTARY FIGURES

Figure 1-figure supplement 1. Avoidance learning in male versus female and scopolamine versus saline
injected rats. *A*) Top: Mann-Whitney U tests found that 48 hour retention of avoidance behavior did not differ for males
versus females in the drug-free or scopolamine shock conditions. Bottom: Median beeline running speeds per session.
B,C) Seven rats without lens implants were trained to criterion for preferring the short path on the maze, then given a
shock avoidance session. On the day of shock avoidance training rats either received injections of saline (n=3) or
scopolamine (n=4) 30 min prior to the session. After shock avoidance training rats were given standard sessions every
48 h (with no injection) until they reached criterion for extinction of avoidance. Rats then received another avoidance
training session preceded by an injection that differed from the one they had received before (i.e., rats previously
injected with saline were injected with scopolamine and vice versa). One of the four rats injected with saline prior to the
second shock session experienced a health issue and was removed from the study prior to being given a shock session.
Graphs in (B) show number of short laps per minute during 5 sessions (2 prior to shock training session, shock training
session, 2 after shock training session) for rats injected with saline and scopolamine; a 2x2 mixed ANOVA (rightmost
graph) was performed with drug condition (saline versus scopolamine) as an independent factor and training condition
(pre versus post) as a repeated factor; shock session includes only data from the 10' pre-shock period whereas all
other days include data from the full 15' session. There was a significant interaction ($F_{1,11}=7.6$, p=.0186) indicating that
saline injected rats exhibited more post-training avoidance (that is, greater reduction in short laps/min) than
scopolamine-injected rats. Post-hoc independent t-tests (Bonferroni corrected, indicated by asterisks) found that rats
in the two drug conditions ran similar numbers of short laps/min before avoidance training, but saline rats ran
significantly fewer short laps/min after training. Scopolamine rats ran significantly fewer short laps/min prior to shock
on the training day (paired $t_5=8.7$, p=.00034), replicating the finding from the main text that scopolamine rats were
impaired at expressing their previously learned preference for the short path. Graphs in (C) are the same as in (A) but
show the ratio of short laps to all laps run during each session. A significant interaction ($F_{1,11}=7.6$, p=.0186) again
indicated that saline injected rats exhibited more post-training avoidance (that is, greater reduction in proportion of short
laps) than scopolamine-injected rats. Scopolamine rats chose the short path significantly less than the long path on the
training day (paired $t_5=5.4$, p=.003), replicating the finding from the main text that scopolamine rats were impaired at
expressing their previously learned preference for the short path. *p<.05 for Bonferroni correct post-hoc comparisons.

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Figure 2-figure supplement 1. *GRIN lens placements and cell contours for imaged neurons in CA1.* The left image in
each pair shows a photomicrograph of the GRIN lens placement for one of the rats in the study; arrows point to the
CA1 layer, scale bar = 1 mm. The right image in each pair shows cell contours (green) identified by CNMFe
superimposed over cropped images of the MiniLFOV's view through the 1.8 mm GRIN lens; the image shown for each
rat was obtained from the day of the rat's drug-free shock training session. Figure shows results from 14 rats that were
included in imaging data analysis (one rat that was included in the behavior analysis but dropped from image analysis
is not shown).

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Figure 2-figure supplement 2. *Beeline trial counts and running speeds prior to downsampling.* To control for any possible confounding effects of behavior upon imaging results, analyses presented in the main text were performed on behavioral data that was downsampled to equalize the number of beeline trials and median running speed across imaging sessions (see Methods). Graphs plotted here show the rats' behavior prior to downsampling beeline trials in the drug-free shock condition for rats that retained avoidance learning (n=9), scopolamine shock condition for rats that failed to retain avoidance learning (n=7), and barrier training condition (n=6). *A*) Number of beeline trials during pre-training (pre; data from full 15' session), training (trn; data from 10' pre-shock portion of session), and post-extinction (pst; data from full 15' session) sessions prior to downsampling; a 3x3 mixed ANOVA yielded a marginal main effect of training condition (drug free shock, scopolamine shock, barrier: $F_{2,19}$=3.5, p=.0509), a significant main effect of session (pre, trn, pst: $F_{2,38}$=51.5, p=1.49e^{-11}), and a significant interaction ($F_{4,38}$=13.9, p=4.37e^{-7}). In all training conditions there were fewer trials during 'trn' sessions because data only came from the first 10 min (prior to shock or barrier introduction) of the training sessions, compared with 15 min of data for other session types. For the drug-free shock condition, the mean number of beeline trials is marginally lower during 'pst' than 'pre' sessions (paired t_8=2.1, p=.0676; uncorrected) because despite having reached the extinction criteria during the 'pst' session, rats persisted in showing some avoidance of the short path and thus earned fewer short path rewards during the 'pst' session. *B*) Median beeline running speed during per session prior to downsampling; a 3x3 mixed ANOVA yielded a marginal main effect of training condition (drug free shock, scopolamine shock, barrier: $F_{2,19}$=3.07, p=.0698), a significant main effect of session (pre, trn, pst: $F_{2,38}$=5.08, p=.0111), and a significant interaction ($F_{4,38}$=20.8, p=3.74e^{-9}). Uncorrected post-hoc paired t-tests found that for the drug-free shock condition, beeline running speed was significantly lower during 'pst' than 'pre' (t_8=4.4, p=.0022) or 'trn' (t_8=4.7, p=.0015) sessions; this was because despite having reached the avoidance extinction criteria during the 'pst' session, rats persisted in running more a bit slowly on the short path after avoidance extinction. In addition, during 'trn' sessions given on scopolamine, rats ran more slowly during 'trn' sessions than during 'pre' (t_6=5.5, p=.0015) or 'pst' (t_6=5.2, p=.0019) sessions (both of which were given drug free) because the drug acutely reduced running speeds on the maze, as reported in the main text.

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Figure 2-figure supplement 3. *Relationship between place tuning versus size and eccentricity of imaged cell contours.*
A) Scatter plot shows contour area (in pixels, x-axis) versus eccentricity of contour from center of the GRIN lens (in
pixels, y-axis) for all place cells and non-place cells imaged during the pre-training session for the drug-free and
scopolamine shock conditions; histograms on perimeter show distributions of values plotted on each axis. Independent
t-test results show that non-place cells were slightly but significantly smaller and less eccentric from the lens center
than place cells, possibly because residual post-correction motion artifact may have caused the smallest cell contours
to jitter occasionally out of their ROI and therefore meet the criterion for spatial tuning stability slightly less often. B)
Same as 'A' except graphs show contour area and eccentricity for all place cells imaged during the pre-training session
prior to drug-free shocks; place cells that recurred during the subsequent shock session are shown in blue whereas
place cells that did not recur are shown in orange. Independent t-test results show that non-recurring cells were slightly
but significantly larger and more eccentric than recurring place cells, possibly because compared with contours near
the center of the lens, between-session overlap of contours at eccentric locations (near the edge of the lens) was
slightly more sensitive to small between-session changes in the mounted position of the miniscope camera. B) Same
as 'B' except data is shown for place cells imaged during the pre-training session prior to shocks on scopolamine (rather
than drug free); as in the drug free condition, non-recurring cells were slightly but significantly larger and more eccentric
than recurring place cells; hence the effect of eccentricity on recurrence probability was similar on the drug-free and
scopolamine conditions. Confirming this, independent t-tests found no significant difference between the drug-free
versus scopolamine conditions in the size of recurring (t_{1860}=.18, p=.86) or non-recurring (t_{882}=.02, p=.98) place cells,
nor in the eccentricity of recurring (t_{1860}=1.62, p=.11) or non-recurring (t_{882}=1.01, p=.31) place cells.

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Figure 2-figure supplement 4. *Distributions of place cell tuning properties.* Histograms at top compare pre-training
and training session distributions of firing properties measured from tuning curves that were spatially tuned in the LR
(top row, blue) or RL (second from top row, red) direction during one or both sessions. Boxplots below histograms show
each rat's median and range of tuning property values for LR and RL tuning curves combined; boxplot medians for
each rat are the same values plotted for each rat in Fig. 2F of the Results. *A*) *Peak Af/s rates.* From the histograms, it
can be seen that scopolamine acutely reduces the proportion of tuning curves peaks that are <5 Af/s while having little
effect on the frequency of peaks that are >5 Af/s. *B*) *Out-of-field Af/s rates.* Similar to peak Af/s rates, scopolamine
acutely reduced the incidence of low out-of-field rates (<1 Af/s), while having little effect on the incidence of higher rates
(>1 Af/s). *C*) *Field widths.* Scopolamine slightly narrows the width of place fields; since the field width is defined as the
number of spatial bins that exceed 50% of the peak rate, this slight narrowing of place fields may be related to the fact
that peak Af/s rates are higher on scopolamine (see 'A'). *D*) *Spatial information.* Distributions of bits/Af showed little
change on scopolamine.

1169

Figure 2-figure supplement 5. *Pre-training and training session data from individual rats.* As in Fig. 2H, heatmaps are
shown for the pre, trn1, and trn2 sessions, with between- (pre vs trn1) and within- (trn1 vs trn2) session correlation
matrices below heatmaps. For brevity, LR and RL tuning curves are co-sorted within heatmaps so that cells meeting
spatial selectivity criteria in both running directions contribute two rows to each heatmap (one per direction), whereas
cells meeting selectivity criteria in only one direction contribute only row to each heatmap. *A1*) Rats (n=9) from the
drug-free shock condition that showed 48 h avoidance retention. *A2*) Rats (n=2) from the drug-free shock condition that
failed to show 48 h avoidance retention. *B1*) Rats (n=7) from the scopolamine condition that failed to show 48 h
avoidance retention. *B2*) Rats (n=2) from the scopolamine shock condition that showed 48 h avoidance retention.

A

B

C

Figure 2-figure supplement 6. *Between and within-session place coding in male versus female rats.* Statistical analyses reported in Fig. 2 were performed to compare results from male versus female rats. 2x2 ANOVAs were performed with sex (M,F) and drug (drug-free, scopolamine) as independent factors. A) There were no significant effects of sex upon the number of place cells imaged per rat (analysis shown for training session), recurrence ratio (RR), or spatial tuning properties (peak rate, out-of-field rate, field width, spatial information); main effect of sex is shown above each graph, however sex-by-drug interactions also were not significant (not shown). B) As reported in the main text, there was no significant main effect of drug upon within-session population vector correlations, however there was a significant main effect of sex arising from the fact that females exhibited lower within-session population vector correlations than males in both drug conditions; this was the only significant effect of sex that was found in any of the analyses. C) As reported in the main text, there was a significant main effect of drug upon between-session population vector correlations, however there was no significant main effect of sex and no sex-by-drug interaction.

1211
Figure 2-figure supplement 7. *Shuffled analysis of between- and within-session population coding.* Results shown
are similar to those plotted in Fig. 2, except that prior to analysis, the deconvolved spike train from every beeline trial
was circularly shifted by a random amount against position tracking data from the trial. A) Bar/line graphs show number
(left) and percentage (right) of cells per rat that were classified as place cells after the random circular shift; the number
of place cells was greatly reduced from that shown in Fig.2. There was a significant tendency for more shuffled trains
to meet place criteria in *trn* than *pre* sessions, attributable to the fact that all detections of place cells in shuffled data
were artifactual and the likelihood of artifactual detection was higher for *trn* than *pre* sessions because fewer trials were
analyzed in *trn* sessions (from which only the first 10 min pre-shock were analyzed) than *pre* sessions (from which all
trials were analyzed across the 15 min session). B) Pie graphs show percentages of "place cells" detected in shuffled
data ('n' gives total number summed over rats) that were spatially tuned in the LR only, RL only, or both LR&RL running
directions. C) Tuning curve properties of "place cells" detected in shuffled data; tuning property differences between *trn*
and *pre* sessions are attributable to differences in behavior sampling. D) Recurrence ratios (RR) between *pre* and *trn*
sessions for "place cells" detected in shuffled data. E-F) Results plotted in panels E-F are from analyses of shuffled
trains performed on cells that were classified as place cells based on their unshuffled trains (the same population of
place cells analyzed in Fig. 2). E) *Top*: Diagram shows timeline for *pre* and *trn* sessions given 48 h apart. *Bottom:*
Tuning curve heatmaps of shuffled trains for recurring place cells (from all rats combined, co-sorted by peak locations
from the *trn1* session) whose unshuffled trains were spatially tuned in the LR (top) or RL (bottom) running directions;
separate heatmaps are shown for *pre*, *trn* part one (*trn1*), and *trn* part two (*trn2*) sessions. F) Between- (B) and within-
(W) session population vector correlation matrices are shown for DF and SCP shock training conditions; middle bar
graph shows median place tuning stability (S) derived from shuffled trains for each rat (lines & symbols) and mean over
rats (bars) for B and W heatmap pairs. p<.1; *p<.05; **p<.01; ***p<. 001. Higher correlation values were uniformly
distributed throughout the within than between session correlation matrices, indicating that baseline firing rates tended
to be better correlated within than between sessions.

Figure 2-figure supplement 8. *Pre-training and training session results from drug-free (DF) rats that failed to retain avoidance and scopolamine (SCP) rats that successfully retained avoidance.* Panels A-G are similar to those shown in panels C-I of Fig. 2 in Results, except here, results are shown for DF rats that failed to retain avoidance (n=2) and SCP rats that successfully retained avoidance (n=2) 48 h after training. The sample size of rats was not sufficient to perform ANOVAs, so independent t-tests were performed to compare values included in each graph bar against values in corresponding bars from Fig. 2 in Results. The only comparison to beat p<.05 (uncorrected) between-session place tuning stability (S) in the SCP condition ('*' and up arrow over center bar graph in panel G; compare with panel I of Fig. 2 in Results). Hence, in rats for which SCP failed to block avoidance, SCP also failed to impair between-session stability of place cell population vectors. Further supporting this conclusion, a marginally significant correlation was observed (see regression line in panel H, upper right) between 48 h avoidance retention scores and between-session S values in the SCP condition (n=11 rats, combining those that retained and forgot the avoidance response). Hence, lower between-session place tuning stability was associated with more severely impaired retention of avoidance. No such correlations were observed in the DF condition or for within-session stability scores.

Figure 3-figure supplement 1. *Pre-training and post- extinction session data from individual rats.* As in Fig. 3, heatmaps are shown for pre-training and post-extinction sessions, along with pre vs post population vector correlation matrices. *A1*) Rats (n=9) from the drug-free shock condition that showed 48 h avoidance retention. *A2*) Rats (n=2) from the drug-free shock condition that failed to show 48 h avoidance retention. *B1*) Rats (n=7) from the scopolamine condition that failed to show 48 h avoidance retention. *B2*) Rats (n=2) from the scopolamine shock condition that showed 48 h avoidance retention. *C*) Rats (n=6) from the barrier training control condition.

Figure 3-figure supplement 2. *Remapping in male versus female rats.* Statistical analyses shown in Fig. 3 were performed to compare results from male versus female rats. 3x2 ANOVAs were performed with sex (M,F) and training condition (DF,SCP,BAR) as independent factors. A) The change in the number of imaged place cells between pre and post sessions (measured as the number of post session cells over the number of pre session cells) did not differ for males versus females; hence, neither sex showed a tendency to gain or lose more place cells than the other after avoidance acquisition and extinction. B) Place cell recurrence rates (RR) between the pre and post sessions did not differ for males versus females. C) Males and females showed similar learning-induced remapping in the center of the short path (black bars) and similar blockade of this remapping by scopolamine (pink bars).

Figure 3-figure supplement 3. *Pre-training and post-extinction results from drug-free (DF) rats that failed to retain avoidance and scopolamine (SCP) rats that successfully retained avoidance.* Panels A-E are similar to Fig. 3 panels B,C,D,F,G, respectively, in Results. Here, results are shown for DF rats that failed to retain avoidance (n=2) and SCP rats that successfully retained avoidance (n=2) 48 h after training. The sample size of rats was not sufficient to perform ANOVAs, so independent t-tests were performed to compare values included in each graph bar against values included in corresponding bars from Fig. 3. The number of beeline trials per session was equalized between sessions in each rat, but not between training conditions; beeline trial counts were marginally lower in SCP rats that retained avoidance responses (panel A, left graph) compared to those in Fig. 3 that failed to retain avoidance; this was because SCP rats that retained avoidance ran fewer beeline trials in the post extinction session (probably because they were expressing residual fear despite reaching the avoidance extinction criterion), resulting in a smaller equalized number of trials. Of the two DF rats that failed to retain avoidance, one appeared to show remapping and the other did not (see individual rat heatmaps in Fig. 3-figure supplement 1, panel A2). Panel E shows that in the male DF rat that failed to retain avoidance or show remapping, place tuning stability (S) was significantly larger in the unsafe C zone and also in the safe R (but not L) zone compared with the mean from Fig. 3 for DF rats that retained avoidance (**p<.01, ***p<.001). Of the two SCP rats that retained avoidance, one appeared to show remapping of the unsafe center zone and the other did not (see individual rat heatmaps in Fig. 3-figure supplement 1, panel B2). Panel E shows that in the male SCP rat that retained avoidance and showed remapping, S was significantly lower in the unsafe C zone (**p<.01 with down arrow), consistent with remapping of the shock zone.

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Figure 3-figure supplement 4. *Place field shifts induced by remapping.* A) Graphs show distributions of the
distances (in units of spatial bins) by which remapping caused recurring place cells (from all rats included in the
analyses of Fig 3) to shift their place fields toward (negative shift values) versus away from (positive shift values) the
center of the short path where shock was delivered. In each graph, inset p-value shows the significance for a z-test
comparison between zero and the mean of the distribution. The distributions did not differ significantly from zero in
the drug-free, scopolamine, or barrier training conditions, indicating that place cells showed no bias for shifting toward
or away from the shock or barrier location in any condition. B) Graphs show distributions of the absolute distance (in
units of spatial bins) by which remapping caused recurring place cells to shift their place fields before versus after
avoidance learning. Inset p-values show results of a ranksum comparison between each distribution's median shift
value versus the medians of the distributions for the other two conditions. The median shift distance was significantly
larger in the drug-free condition than in the scopolamine or barrier conditions, consistent with findings reported in the
main text showing that remapping occurred preferentially in the drug-free condition (since larger shifts of place field
centers are consistent with more place cell remapping). Median shift distances did not differ significantly between the
scopolamine and barrier conditions, confirming that remapping was less prevalent in these two conditions than in the
drug-free condition.

Figure 3-figure supplement 5. *Shuffled analysis of remapping.* Results plotted in panels A-D were performed as described for corresponding results in Fig. 3, except that prior to analysis, the deconvolved spike train from every beeline trial was circularly shifted by a random amount against position tracking data from the trial. A) Heatmap pairs show pre and post tuning curves for recurring place cells (same population of cells as in Fig. 3 which were classified as place cell based on their unshuffled trains) that were spatially tuned in the LR (top row) and RL (bottom row) running direction; both heatmaps in each pair are co-sorted by peak locations of the shuffled tuning curve from the pre session. B) Distributions of shuffled tuning curve peak locations (LR and RL combined) are no longer concentrated preferentially at the ends of the track. C) Mean population vector correlation matrices calculated from shuffled tuning curves for the three training conditions; uniformly distributed positive values (greater than zero) reflect a tendency for cells to have similar baseline firing rates across sessions. D) Left column: Graphs show scatter plots of each rat's place tuning stability scores (S); means did not differ significantly from zero. Results for L, C, & R track zones are shown top, middle, and bottom rows, respectively. Middle columns: Boxplots show median and range of template-selected population vector correlation bins in each rat. Right column: Templates used to select peri-diagonal correlation values from the L, C, & R track zones.

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1413 **Video 1** Video demonstrating behavior and calcium imaging recorded from one rat performing linear alternation along
1414 the short path of the maze A) position of the rat (red dot) B) Motion corrected video of calcium fluorescence C)
1415 Processed demixed/denoised CaImAn output (Spatial contours modulated by denoised calcium trace) D) Right half
1416 shows calcium activity of 290 identified neurons in the video, sorted by their preferred firing direction (teal/pink) and
1417 location along the track. Current time highlighted in the center gray region.

REFERENCES

Aharoni, Daniel, and Tycho M. Hoogland. 2019. "Circuit Investigations With Open-Source Miniaturized Microscopes: Past, Present and Future." *Frontiers in Cellular Neuroscience* 13: 141. doi:10.3389/fncel.2019.00141.

Alvernhe, Alice, Etienne Save, and Bruno Poucet. 2011. "Local Remapping of Place Cell Firing in the Tolman Detour Task." *European Journal of Neuroscience* 33 (9): 1696–1705. doi:10.1111/j.1460-9568.2011.07653.x.

Anagnostaras, Stephan G., Stephen Maren, and Michael S. Fanselow. 1995. "Scopolamine Selectively Disrupts the Acquisition of Contextual Fear Conditioning in Rats." *Neurobiology of Learning and Memory* 64 (3): 191–194. doi:10.1006/nlme.1995.0001.

Anagnostaras, Stephan G, Stephen Maren, Jennifer R Sage, Stacy Goodrich, and Michael S Fanselow. 1999. "Scopolamine and Pavlovian Fear Conditioning in Rats: Dose-Effect Analysis." *Neuropsychopharmacology* 21 (6): 731–744. doi:10.1016/s0893-133x(99)00083-4.

Arendt, T. 1994. "Impairment in Memory Function and Neurodegenerative Changes in the Cholinergic Basal Forebrain System Induced by Chronic Intake of Ethanol," 173–187. doi:10.1007/978-3-7091-9350-1_13.

Arendt, T., V. Bigl, A. Arendt, and A. Tennstedt. 1983. "Loss of Neurons in the Nucleus Basalis of Meynert in Alzheimer's Disease, Paralysis Agitans and Korsakoff's Disease." *Acta Neuropathologica* 61 (2): 101–108. doi:10.1007/bf00697388.

Atri, Alireza, Seth Sherman, Kenneth A. Norman, Brenda A. Kirchhoff, Marlene M. Nicolas, Michael D. Greicius, Steven C. Cramer, Hans C. Breiter, Michael E. Hasselmo, and Chantal E. Stern. 2004. "Blockade of Central Cholinergic Receptors Impairs New Learning and Increases Proactive Interference in a Word Paired-Associate Memory Task." *Behavioral Neuroscience* 118 (1): 223–236. doi:10.1037/0735-7044.118.1.223.

Barnes, C. A. 1979. "Memory Deficits Associated with Senescence: A Neurophysiological and Behavioral Study in the Rat." *Journal of Comparative and Physiological Psychology* 93 (1): 74–104. doi:10.1037/h0077579.

Barnes, C. A., L. Nadel, and Werner K. Honig. 1980. "Spatial Memory Deficit in Senescent Rats." *Canadian Journal of Psychology/Revue Canadienne de Psychologie* 34 (1): 29–39. doi:10.1037/h0081022.

Barnes, Carol A., Matthew S. Suster, Jiemin Shen, and Bruce L. McNaughton. 1997. "Multistability of Cognitive Maps in the Hippocampus of Old Rats." *Nature* 388 (6639): 272–275. doi:10.1038/40859.

Barten, Donna M., and Charles F. Albright. 2008. "Therapeutic Strategies for Alzheimer's Disease." *Molecular Neurobiology* 37 (2–3): 171–186. doi:10.1007/s12035-008-8031-2.

Bittner, Katie C, Christine Grienberger, Sachin P Vaidya, Aaron D Milstein, John J Macklin, Junghyup Suh, Susumu Tonegawa, and Jeffrey C Magee. 2015. "Conjunctive Input Processing Drives Feature Selectivity in Hippocampal CA1 Neurons." *Nature Neuroscience* 18 (8): 1133–1142. doi:10.1038/nn.4062.

Blair, Hugh T., Glenn E. Schafe, Elizabeth P. Bauer, Sarina M. Rodrigues, and Joseph E. LeDoux. 2001. "Synaptic Plasticity in the Lateral Amygdala: A Cellular Hypothesis of Fear Conditioning." *Learning & Memory* 8 (5): 229–242. doi:10.1101/lm.30901.

Bostock, Elizabeth, Robert U. Muller, and John L. Kubie. 1991. "Experience-dependent Modifications of Hippocampal Place Cell Firing." *Hippocampus* 1 (2): 193–205. doi:10.1002/hipo.450010207.

Brazhnik, Elena, Ramie Borgnis, Robert U. Muller, and Steven E. Fox. 2004. "The Effects on Place Cells of Local Scopolamine Dialysis Are Mimicked by a Mixture of Two Specific Muscarinic Antagonists." *The Journal of Neuroscience* 24 (42): 9313–9323. doi:10.1523/jneurosci.1618-04.2004.

Breese, CR, RE Hampson, and SA Deadwyler. 1989. "Hippocampal Place Cells: Stereotypy and Plasticity." *The Journal of Neuroscience* 9 (4): 1097–1111. doi:10.1523/jneurosci.09-04-01097.1989.

Brown, David A. 2018. "Regulation of Neural Ion Channels by Muscarinic Receptors." *Neuropharmacology* 136 (Pt C): 383–400. doi:10.1016/j.neuropharm.2017.11.024.

Cai, Denise J., Daniel Aharoni, Tristan Shuman, Justin Shobe, Jeremy Biane, Weilin Song, Brandon Wei, et al. 2016. "A Shared Neural Ensemble Links Distinct Contextual Memories Encoded Close in Time." *Nature* 534 (7605): 115–118. doi:10.1038/nature17955.

Colgin, Laura Lee, Edvard I. Moser, and May-Britt Moser. 2008. "Understanding Memory through Hippocampal Remapping." *Trends in Neurosciences* 31 (9): 469–477. doi:10.1016/j.tins.2008.06.008.

Dannenberg, Holger, Kimberly Young, and Michael Hasselmo. 2017. "Modulation of Hippocampal Circuits by Muscarinic and Nicotinic Receptors." *Frontiers in Neural Circuits* 11: 102. doi:10.3389/fncir.2017.00102.

d'Aquin, Simon, Andras Szonyi, Mathias Mahn, Sabine Krabbe, Jan Gründemann, and Andreas Lüthi. 2022. "Compartmentalized Dendritic Plasticity during Associative Learning." *Science* 376 (6590): eabf7052. doi:10.1126/science.abf7052.

Davidson, Thomas J., Fabian Kloosterman, and Matthew A. Wilson. 2009. "Hippocampal Replay of Extended Experience." *Neuron* 63 (4): 497–507. doi:10.1016/j.neuron.2009.07.027.

Decker, Michael W., Thuy Tran, and James L. McGaugh. 1990. "A Comparison of the Effects of Scopolamine and Diazepam on Acquisition and Retention of Inhibitory Avoidance in Mice." *Psychopharmacology* 100 (4): 515–521. doi:10.1007/bf02244005.

Digby, Gregory J., Jana K. Shirey, and P. Jeffrey Conn. 2010. "Allosteric Activators of Muscarinic Receptors as Novel Approaches for Treatment of CNS Disorders." *Molecular BioSystems* 6 (8): 1345–1354. doi:10.1039/c002938f.

Douchamps, Vincent, Ali Jeewajee, Pam Blundell, Neil Burgess, and Colin Lever. 2013. "Evidence for Encoding versus Retrieval Scheduling in the Hippocampus by Theta Phase and Acetylcholine." *The Journal of Neuroscience* 33 (20): 8689–8704. doi:10.1523/jneurosci.4483-12.2013.

Dragoi, George, Kenneth D Harris, and György Buzsáki. 2003. "Place Representation within Hippocampal Networks Is Modified by Long-Term Potentiation." *Neuron* 39 (5): 843–853. doi:10.1016/s0896-6273(03)00465-3.

Drever, Benjamin D., Gernot Riedel, and Bettina Platt. 2011. "The Cholinergic System and Hippocampal Plasticity." *Behavioural Brain Research* 221 (2): 505–514. doi:10.1016/j.bbr.2010.11.037.

Dupret, David, Joseph O'Neill, Barty Pleydell-Bouverie, and Jozsef Csicsvari. 2010. "The Reorganization and Reactivation of Hippocampal Maps Predict Spatial Memory Performance." *Nature Neuroscience* 13 (8): 995–1002. doi:10.1038/nn.2599.

Geiller, Tristan, Sadra Sadeh, Sebastian V. Rolotti, Heike Blockus, Bert Vancura, Adrian Negrean, Andrew J. Murray, et al. 2022. "Local Circuit Amplification of Spatial Selectivity in the Hippocampus." *Nature* 601 (7891): 105–109. doi:10.1038/s41586-021-04169-9.

Giovannucci, Andrea, Johannes Friedrich, Pat Gunn, Jérémie Kalfon, Brandon L Brown, Sue Ann Koay, Jiannis Taxidis, et al. 2019. "CaImAn an Open Source Tool for Scalable Calcium Imaging Data Analysis." *ELife* 8: e38173. doi:10.7554/elife.38173.

Girardeau, Gabrielle, Ingrid Inema, and György Buzsáki. 2017. "Reactivations of Emotional Memory in the Hippocampus–Amygdala System during Sleep." *Nature Neuroscience* 20 (11): 1634–1642. doi:10.1038/nn.4637.

Green, Amity, Kathryn A. Ellis, Julia Ellis, Cali F. Bartholomeusz, Susan Ilic, Rodney J. Croft, K. Luan Phan, and Pradeep J. Nathan. 2005. "Muscarinic and Nicotinic Receptor Modulation of Object and Spatial N-Back Working Memory in Humans." *Pharmacology Biochemistry and Behavior* 81 (3): 575–584. doi:10.1016/j.pbb.2005.04.010.

Grewe, Benjamin F., Jan Gründemann, Lacey J. Kitch, Jerome A. Lecoq, Jones G. Parker, Jesse D. Marshall, Margaret C. Larkin, et al. 2017. "Neural Ensemble Dynamics Underlying a Long-Term Associative Memory." *Nature* 543 (7647): 670–675. doi:10.1038/nature21682.

Guo, Changliang, Garrett J. Blair, Megha Sehgal, Federico N. Sangiuliano Jimka, Arash Bellafard, Alcino J. Silva, Peyman Golshani, Michele A. Basso, Hugh Tad Blair, and Daniel Aharoni. 2023. "Miniscope-LFOV: A Large-Field-of-View, Single-Cell-Resolution, Miniature Microscope for Wired and Wire-Free Imaging of Neural Dynamics in Freely Behaving Animals." *Science Advances* 9 (16): eadg3918. doi:10.1126/sciadv.adg3918.

Hales, C. A., J. M. Bartlett, R. Arban, B. Hengerer, and E. S. J. Robinson. 2020. "Role of the Medial Prefrontal Cortex in the Effects of Rapid Acting Antidepressants on Decision-Making Biases in Rodents." *Neuropsychopharmacology* 45 (13): 2278–2288. doi:10.1038/s41386-020-00797-3.

Hardy, John. 2006. "A Hundred Years of Alzheimer's Disease Research." *Neuron* 52 (1): 3–13. doi:10.1016/j.neuron.2006.09.016.

Hasselmo, Michael E. 2006. "The Role of Acetylcholine in Learning and Memory." *Current Opinion in Neurobiology* 16 (6): 710–715. doi:10.1016/j.conb.2006.09.002.

Hasselmo, Michael E., and Jill McGaughy. 2004. "High Acetylcholine Levels Set Circuit Dynamics for Attention and Encoding and Low Acetylcholine Levels Set Dynamics for Consolidation." *Progress in Brain Research* 145: 207–231. doi:10.1016/s0079-6123(03)45015-2.

Hollup, Stig A, Sturla Molden, James G Donnett, May-Britt Moser, and Edvard I Moser. 2001. "Accumulation of Hippocampal Place Fields at the Goal Location in an Annular Watermaze Task." *The Journal of Neuroscience* 21 (5): 1635–1644. doi:10.1523/jneurosci.21-05-01635.2001.

Huang, Zhen-Bo, Hao Wang, Xiu-Rong Rao, Gui-Fa Zhong, Wen-Hui Hu, and Guo-Qing Sheng. 2011. "Different Effects of Scopolamine on the Retrieval of Spatial Memory and Fear Memory." *Behavioural Brain Research* 221 (2): 604–609. doi:10.1016/j.bbr.2010.05.032.

Hunt, David L., Daniele Linaro, Bailu Si, Sandro Romani, and Nelson Spruston. 2018. "A Novel Pyramidal Cell Type Promotes Sharp-Wave Synchronization in the Hippocampus." *Nature Neuroscience* 21 (7): 985–995. doi:10.1038/s41593-018-0172-7.

Izquierdo, Ivan, Claudio da Cunha, Renata Rosat, Diana Jerusalinsky, Maria Beatriz C. Ferreira, and Jorge H. Medina. 1992. "Neurotransmitter Receptors Involved in Post-Training Memory Processing by the Amygdala, Medial Septum, and Hippocampus of the Rat." *Behavioral and Neural Biology* 58 (1): 16–26. doi:10.1016/0163-1047(92)90847-w.

Jarzebowski, Przemyslaw, Clara S Tang, Ole Paulsen, and Y Audrey Hay. 2021. "Impaired Spatial Learning and Suppression of Sharp Wave Ripples by Cholinergic Activation at the Goal Location." *ELife* 10: e65998. doi:10.7554/elife.65998.

Jiang, Li, Srikanya Kundu, James D. Lederman, Gretchen Y. López-Hernández, Elizabeth C. Ballinger, Shaohua Wang, David A. Talmage, and Lorna W. Role. 2016. "Cholinergic Signaling Controls Conditioned Fear Behaviors and Enhances Plasticity of Cortical-Amygdala Circuits." *Neuron* 90 (5): 1057–1070. doi:10.1016/j.neuron.2016.04.028.

Keinath, Alexandra T., Coralie-Anne Mosser, and Mark P. Brandon. 2022. "The Representation of Context in Mouse Hippocampus Is Preserved despite Neural Drift." *Nature Communications* 13 (1): 2415. doi:10.1038/s41467-022-30198-7.

Kim, Eun Joo, Mijeong Park, Mi-Seon Kong, Sang Geon Park, Jeiwon Cho, and Jeansok J. Kim. 2015. "Alterations of Hippocampal Place Cells in Foraging Rats Facing a 'Predatory' Threat." *Current Biology* 25 (10): 1362–1367. doi:10.1016/j.cub.2015.03.048.

Kinsky, Nathaniel R., David W. Sullivan, William Mau, Michael E. Hasselmo, and Howard B. Eichenbaum. 2018. "Hippocampal Place Fields Maintain a Coherent and Flexible Map across Long Timescales." *Current Biology* 28 (22): 3578-3588.e6. doi:10.1016/j.cub.2018.09.037.

Leutgeb, Stefan, Jill K. Leutgeb, Carol A. Barnes, Edvard I. Moser, Bruce L. McNaughton, and May-Britt Moser. 2005. "Independent Codes for Spatial and Episodic Memory in Hippocampal Neuronal Ensembles." *Science* 309 (5734): 619–623. doi:10.1126/science.1114037.

Lever, Colin, Tom Wills, Francesca Cacucci, Neil Burgess, and John O'Keefe. 2002. "Long-Term Plasticity in Hippocampal Place-Cell Representation of Environmental Geometry." *Nature* 416 (6876): 90–94. doi:10.1038/416090a.

Lombardo, Sylvia, and Uwe Maskos. 2015. "Role of the Nicotinic Acetylcholine Receptor in Alzheimer's Disease Pathology and Treatment." *Neuropharmacology* 96 (Pt B): 255–262. doi:10.1016/j.neuropharm.2014.11.018.

Mamad, Omar, Beshoy Agayby, Lars Stumpp, Richard B. Reilly, and Marian Tsanov. 2019. "Extrafield Activity Shifts the Place Field Center of Mass to Encode Aversive Experience." *ENeuro* 6 (2): ENEURO.0423-17.2019. doi:10.1523/eneuro.0423-17.2019.

Mamad, Omar, Lars Stumpp, Harold M. McNamara, Charu Ramakrishnan, Karl Deisseroth, Richard B. Reilly, and Marian Tsanov. 2017. "Place Field Assembly Distribution Encodes Preferred Locations." *PLoS Biology* 15 (9): e2002365. doi:10.1371/journal.pbio.2002365.

Mankin, Emily A., Fraser T. Sparks, Begum Slayyeh, Robert J. Sutherland, Stefan Leutgeb, and Jill K. Leutgeb. 2012. "Neuronal Code for Extended Time in the Hippocampus." *Proceedings of the National Academy of Sciences* 109 (47): 19462–19467. doi:10.1073/pnas.1214107109.

Martin, S. J., P. D. Grimwood, and R. G. M. Morris. 2000. "Synaptic Plasticity and Memory: An Evaluation of the Hypothesis." *The Neuroscientist* 23 (1): 649–711. doi:10.1146/annurev.neuro.23.1.649.

Ma, Xiaoyu, Yiyao Zhang, Lina Wang, Na Li, Edi Barkai, Xiaohui Zhang, Longnian Lin, and Jiamin Xu. 2020. "The Firing of Theta State-Related Septal Cholinergic Neurons Disrupt Hippocampal Ripple Oscillations via Muscarinic Receptors." *The Journal of Neuroscience* 40 (18): 3591–3603. doi:10.1523/jneurosci.1568-19.2020.

McKenzie, Sam, Roman Huszár, Daniel F. English, Kanghwan Kim, Fletcher Christensen, Euisik Yoon, and György Buzsáki. 2021. "Preexisting Hippocampal Network Dynamics Constrain Optogenetically Induced Place Fields." *Neuron* 109 (6): 1040-1054.e7. doi:10.1016/j.neuron.2021.01.011.

Miller, C.N., and H.M. Kamens. 2020. "The Role of Nicotinic Acetylcholine Receptors in Alcohol-Related Behaviors." *Brain Research Bulletin* 163: 135–142. doi:10.1016/j.brainresbull.2020.07.017.

Moita, Marta A.P., Svetlana Rosis, Yu Zhou, Joseph E. LeDoux, and Hugh T. Blair. 2003. "Hippocampal Place Cells Acquire Location-Specific Responses to the Conditioned Stimulus during Auditory Fear Conditioning." *Neuron* 37 (3): 485–497. doi:10.1016/s0896-6273(03)00033-3.

Moita, Marta A. P., Svetlana Rosis, Yu Zhou, Joseph E. LeDoux, and Hugh T. Blair. 2004. "Putting Fear in Its Place: Remapping of Hippocampal Place Cells during Fear Conditioning." *The Journal of Neuroscience* 24 (31): 7015–7023. doi:10.1523/jneurosci.5492-03.2004.

Newman, Ehren L., Sarah Jo C. Venditto, Jason R. Climer, Elijah A. Petter, Shea N. Gillet, and Sam Levy. 2017. "Precise Spike Timing Dynamics of Hippocampal Place Cell Activity Sensitive to Cholinergic Disruption." *Hippocampus* 27 (10): 1069–1082. doi:10.1002/hipo.22753.

Nieto-Escámez, Francisco A, Fernando Sánchez-Santed, and Jan P.C de Bruin. 2002. "Cholinergic Receptor Blockade in Prefrontal Cortex and Lesions of the Nucleus Basalis: Implications for Allocentric and Egocentric Spatial Memory in Rats." *Behavioural Brain Research* 134 (1–2): 93–112. doi:10.1016/s0166-4328(01)00458-2.

O'Keefe, John, and J. Dostrovsky. 1971. "The Hippocampus as a Spatial Map. Preliminary Evidence from Unit Activity in the Freely-Moving Rat." *Brain Research* 34 (1): 171–175. doi:10.1016/0006-8993(71)90358-1.

O'Keefe, John, and Lynn Nadel. 1978. "The Hippocampus as a Cognitive Map." *Oxford University Press*.

Oler, Jonathan A., and Etan J. Markus. 2000. "Age-related Deficits in the Ability to Encode Contextual Change: A Place Cell Analysis." *Hippocampus* 10 (3): 338–350. doi:10.1002/1098-1063(2000)10:3<338::aid-hipo14>3.0.co;2-y.

Ormond, Jake, Simon A. Serka, and Joshua P. Johansen. 2023. "Enhanced Reactivation of Remapping Place Cells during Aversive Learning." *The Journal of Neuroscience* 43 (12): 2153–2167. doi:10.1523/jneurosci.1450-22.2022.

Pnevmatikakis, Eftychios A., and Andrea Giovannucci. 2017. "NoRMCorre: An Online Algorithm for Piecewise Rigid Motion Correction of Calcium Imaging Data." *Journal of Neuroscience Methods* 291: 83–94. doi:10.1016/j.jneumeth.2017.07.031.

Ragozzino, Michael E., Sonja Artis, Amritha Singh, Trevor M. Twose, Joseph E. Beck, and William S. Messer. 2012. "The Selective M1 Muscarinic Cholinergic Agonist CDD-0102A Enhances Working Memory and Cognitive Flexibility." *Journal of Pharmacology and Experimental Therapeutics* 340 (3): 588–594. doi:10.1124/jpet.111.187625.

Rashid, Habiba, and Touqeer Ahmed. 2019. "Muscarinic Activity in Hippocampus and Entorhinal Cortex Is Crucial for Spatial and Fear Memory Retrieval." *Pharmacological Reports* 71 (3): 449–456. doi:10.1016/j.pharep.2019.02.004.

Sanders, Honi, Matthew A Wilson, and Samuel J Gershman. 2020. "Hippocampal Remapping as Hidden State Inference." *ELife* 9: e51140. doi:10.7554/elife.51140.

Schliebs, Reinhard, and Thomas Arendt. 2011. "The Cholinergic System in Aging and Neuronal Degeneration." *Behavioural Brain Research* 221 (2): 555–563. doi:10.1016/j.bbr.2010.11.058.

Schoenenberger, Philipp, Joseph O'Neill, and Jozsef Csicsvari. 2016. "Activity-Dependent Plasticity of Hippocampal Place Maps." *Nature Communications* 7 (1): 11824. doi:10.1038/ncomms11824.

Schuette, Peter J., Fernando M.C.V. Reis, Sandra Maesta-Pereira, Meghmik Chakerian, Anita Torossian, Garrett Blair, Weisheng Wang, et al. 2020. "Long-Term Characterization of Hippocampal Remapping during Contextual Fear Acquisition and Extinction." *Journal of Neuroscience* 40 (43): 8329–8342. doi:10.1523/jneurosci.1022-20.2020.

Sheintuch, Liron, Alon Rubin, Noa Brande-Eilat, Nitzan Geva, Noa Sadeh, Or Pinchasof, and Yaniv Ziv. 2017. "Tracking the Same Neurons across Multiple Days in Ca2+ Imaging Data." *Cell Reports* 21 (4): 1102–1115. doi:10.1016/j.celrep.2017.10.013.

Shuman, Tristan, Daniel Aharoni, Denise J. Cai, Christopher R. Lee, Spyridon Chavlis, Lucia Page-Harley, Lauren M. Vetere, et al. 2020. "Breakdown of Spatial Coding and Interneuron Synchronization in Epileptic Mice." *Nature Neuroscience* 23 (2): 229–238. doi:10.1038/s41593-019-0559-0.

Skaggs, William E., Bruce L. McNaughton, Katalin M. Gothard, and Etan J. Markus. 1993. "An Information-Theoretic Approach for Deciphering the Hippocampal Code." *Advances in Neural Information Processing Systems*.

Solari, Nicola, and Balázs Hangya. 2018. "Cholinergic Modulation of Spatial Learning, Memory and Navigation." *European Journal of Neuroscience* 48 (5): 2199–2230. doi:10.1111/ejn.14089.

Sun, Dechuan, Ranjith Rajasekharan Unnithan, and Chris French. 2021. "Scopolamine Impairs Spatial Information Recorded With 'Miniscope' Calcium Imaging in Hippocampal Place Cells." *Frontiers in Neuroscience* 15: 640350. doi:10.3389/fnins.2021.640350.

Svoboda, Jan, Anna Popelikova, and Ales Stuchlik. 2017. "Drugs Interfering with Muscarinic Acetylcholine Receptors and Their Effects on Place Navigation." *Frontiers in Psychiatry* 8: 215. doi:10.3389/fpsyt.2017.00215.

Tanila, H., S. Ku, F. Kloosterman, and M. A. Wilson. 2018. "Characteristics of CA1 Place Fields in a Complex Maze with Multiple Choice Points." *Hippocampus* 28 (2): 81–96. doi:10.1002/hipo.22810.

Vales, Karel, and Ales Stuchlik. 2005. "Central Muscarinic Blockade Interferes with Retrieval and Reacquisition of Active Allothetic Place Avoidance despite Spatial Pretraining." *Behavioural Brain Research* 161 (2): 238–244. doi:10.1016/j.bbr.2005.02.012.

Vandecasteele, Marie, Viktor Varga, Antal Berényi, Edit Papp, Péter Barthó, Laurent Venance, Tamás F. Freund, and György Buzsáki. 2014. "Optogenetic Activation of Septal Cholinergic Neurons Suppresses Sharp Wave Ripples and Enhances Theta Oscillations in the Hippocampus." *Proceedings of the National Academy of Sciences* 111 (37): 13535–13540. doi:10.1073/pnas.1411233111.

Wallenstein, Gene V., and David R. Vago. 2001. "Intrahippocampal Scopolamine Impairs Both Acquisition and Consolidation of Contextual Fear Conditioning." *Neurobiology of Learning and Memory* 75 (3): 245–252. doi:10.1006/nlme.2001.4005.

Wang, Melissa E., Ellen G. Wann, Robin K. Yuan, Manuel M. Ramos Álvarez, Squire M. Stead, and Isabel A. Muzzio. 2012. "Long-
Term Stabilization of Place Cell Remapping Produced by a Fearful Experience." *The Journal of Neuroscience* 32 (45):
15802–15814. doi:10.1523/jneurosci.0480-12.2012.
Wang, Melissa E., Robin K. Yuan, Alexander T. Keinath, Manuel M. Ramos Álvarez, and Isabel A. Muzzio. 2015. "Extinction of
Learned Fear Induces Hippocampal Place Cell Remapping." *The Journal of Neuroscience* 35 (24): 9122–9136.
doi:10.1523/jneurosci.4477-14.2015.
Wenk, Hartmut, Volker Bigl, and Ursula Meyer. 1980. "Cholinergic Projections from Magnocellular Nuclei of the Basal Forebrain to
Cortical Areas in Rats." *Brain Research Reviews* 2 (1–3): 295–316. doi:10.1016/0165-0173(80)90011-9.
Wilson, I.A, S Ikonen, R.W McMahan, M Gallagher, H Eichenbaum, and H Tanila. 2003. "Place Cell Rigidity Correlates with
Impaired Spatial Learning in Aged Rats." *Neurobiology of Aging* 24 (2): 297–305. doi:10.1016/s0197-4580(02)00080-5.
Wilson, Iain A, Sami Ikonen, Irina Gureviciene, Robert W McMahan, Michela Gallagher, Howard Eichenbaum, and Heikki Tanila.
2004. "Cognitive Aging and the Hippocampus: How Old Rats Represent New Environments." *Journal of Neuroscience* 24
(15): 3870–3878. doi:10.1523/jneurosci.5205-03.2004.
Wirtshafter, Hannah S, and John F Disterhoft. 2022. "In Vivo Multi-Day Calcium Imaging of CA1 Hippocampus in Freely Moving
Rats Reveals a High Preponderance of Place Cells with Consistent Place Fields." *The Journal of Neuroscience* 42 (22):
4538–4554. doi:10.1523/jneurosci.1750-21.2022.
Wu, Chun-Ting, Daniel Haggerty, Caleb Kemere, and Daoyun Ji. 2017. "Hippocampal Awake Replay in Fear Memory Retrieval."
Nature Neuroscience 20 (4): 571–580. doi:10.1038/nn.4507.
Zhang, Kechen, Iris Ginzburg, Bruce L. McNaughton, and Terrence J. Sejnowski. 1998. "Interpreting Neuronal Population Activity by
Reconstruction: Unified Framework With Application to Hippocampal Place Cells." *Journal of Neurophysiology* 79 (2):
1017–1044. doi:10.1152/jn.1998.79.2.1017.
Ziv, Yaniv, Laurie D Burns, Eric D Cocker, Elizabeth O Hamel, Kunal K Ghosh, Lacey J Kitch, Abbas El Gamal, and Mark J
Schnitzer. 2013. "Long-Term Dynamics of CA1 Hippocampal Place Codes." *Nature Neuroscience* 16 (3): 264–266.
doi:10.1038/nn.3329.
Zylla, Maura M., Xiaomin Zhang, Susanne Reichinnek, Andreas Draguhn, and Martin Both. 2013. "Cholinergic Plasticity of
Oscillating Neuronal Assemblies in Mouse Hippocampal Slices." *PLoS ONE* 8 (11): e80718.
doi:10.1371/journal.pone.0080718.
1649