# Calibration of crop phenology models: Going beyond recommendations

Wallach[1], Daniel; Palosuo[2], Taru; Thorburn[3], Peter; Mielenz[4], Henrike; Buis[5], Samuel; Hochman[3], Zvi; Gourdain[6], Emmanuelle; Andrianasolo[6], Fety; Dumont[7], Benjamin; Ferrise[8], Roberto; Gaiser[9], Thomas; Garcia[6], Cecile; Gayler[10], Sebastian; Hiremath[11], Santosh; Horan[3], Heidi; Hoogenboom[12,13], Gerrit; Jansson[14], Per-Erik; Jing[15], Qi; Justes[16], Eric; Kersebaum[17,18], Kurt-Christian; Launay[19], Marie; Lewan[20], Elisabet; Mequanint[10], Fasil; Moriondo[21], Marco; Nendel[17,18,22], Claas; Padovan[8], Gloria; Qian[15], Budong; Schütze[23], Niels; Seserman[17], Diana-Maria; Shelia[12,13], Vakhtang; Souissi[24], Amir; Specka[17], Xenia; Srivastava[9], Amit Kumar; Trombi[8], Giacomo; Weber[10], Tobias, K.D.; Weihermüller[25], Lutz; Wöhling[23,26], Thomas; Seidel, Sabine J.[9]

[1]INRAE, UMR AGIR, Castanet Tolosan, France. ORCID 0000-0003-3500-8179
[2]Natural Resources Institute Finland (Luke), Helsinki, Finland
[3]CSIRO Agriculture and Food, Brisbane, Queensland, Australia
[4]Institute for Crop and Soil Science, Federal Research Centre for cultivated Plants, Julius Kühn-Institut (JKI), Braunschweig, Germany
[5]INRAE, UMR 1114 EMMAH, Avignon, France
[6]ARVALIS - Institut du végétal Paris, France
[7]Plant Sciences & TERRA Teaching and Research Centre, Gembloux Agro-Bio Tech, University of Liege, Gembloux, Belgium
[8]Department of Agriculture, Food, Environment and Forestry (DAGRI), University of Florence, Italy
[9]Institute of Crop Science and Resource Conservation, University of Bonn, Germany
[10]Institute of Soil Science and Land Evaluation, Biogeophysics, University of Hohenheim, Stuttgart, Germany
[11]Aalto University School of Science, Espoo, Finland;
[12]Agricultural and Biological Engineering Department, University of Florida, Gainesville, Florida
[13]Institute for Sustainable Food Systems, University of Florida, Gainesville, Florida
[14]Royal Institute of Technology (KTH), Stockholm, Sweden
[15]Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, Ottawa, Canada
[16]CIRAD, UMR SYSTEM, Montpellier, France
[17]Leibniz Centre for Agricultural Landscape Research (ZALF), Müncheberg, Germany
[18]Global Change Research Institute CAS, Brno, Czech Republic
[19]INRAE, US 1116 AgroClim, Avignon, France
[20]Department of Soil and Environment, Swedish University of Agricultural Sciences (SLU), Uppsala, Sweden
[21]CNR-IBE, Firenze, Italy
[22] Institute of Biochemistry and Biology, University of Potsdam, Potsdam, Germany
[23]Institute of Hydrology and Meteorology, Chair of Hydrology, Technische Universität Dresden, Dresden, Germany
[24] Swift Current Research and Development Centre, Agriculture and Agri-Food Canada, Swift Current, Saskatchewan, Canada.
[25]Institute of Bio- and Geosciences - IBG-3, Agrosphere, Forschungszentrum Jülich GmbH, Jülich, Germany
[26]Lincoln Agritech Ltd., Hamilton, New Zealand

# Abstract

A major effect of environment on crops is through crop phenology, and therefore, the capacity to predict phenology as a function of soil, weather, and management is important. Mechanistic crop models are a major tool for such predictions. It has been shown that there is a large variability between predictions by different modeling groups for the same inputs, and therefore, a need for shared improvement of crop models. Two pathways to improvement are through improved understanding of the mechanisms of the modeled system, and through improved model parameterization. This article focuses on improving crop model parameters through improved calibration, specifically for prediction of crop phenology. A detailed calibration protocol is proposed, which covers all the steps in the calibration work-flow, namely choice of default parameter values, choice of objective function, choice of parameters to estimate from the data, calculation of optimal parameter values and diagnostics. For those aspects where knowledge of the model and target environments is required, the protocol gives detailed guidelines rather than strict instructions. The protocol includes documentation tables, to make the calibration process more transparent. The protocol was applied by 19 modeling groups to three data sets for wheat phenology. All groups first calibrated their model using their "usual" calibration approach. Evaluation was based on data from sites and years not represented in the training data. Compared to usual calibration, l calibration following the new protocol significantly reduced the error in predictions for the evaluation data, and reduced the variability between modeling groups by 22%.

# Key words

# Introduction

Plant phenology is a major aspect of plant response to environment, and a major determinant of plant response to climate change. This includes phenology of natural vegetation, which is a dominant aspect of plant ecology (Cleland et al., 2007) and has been shown to be affected by warming (Menzel et al., 2020; Piao et al., 2019; Stuble et al., 2021) and phenology of cultivated crops. For the latter, phenology must be taken into account for crop management (Sisheber et al., 2022), choice of cultivar or cultivar characteristics adapted to a particular region (Zhang et al., 2022) and for evaluating the impact of climate change on crop production (Rezaei et al., 2018). It is thus important to be able to predict phenology as a function of environment, in particular as a function of climate.

A number of mechanistic crop models have been developed, which include simulation of phenology, and such models are regularly used to evaluate management options (McNunn et al., 2019) or the effect of climate change on crops, including wheat (Asseng et al., 2013), rice, (Li et al., 2015),  maize (Bassu et al., 2014) and others. Such models are particularly important for taking into account an increasing diversity of combinations of weather events (Webber et al., 2020).

Mechanistic models in general, and models used to simulate crop phenology in particular (we will refer to such models as crop phenology models, though they are usually embedded within more general crop models), are based on our understanding of the processes and their inter-linkages that drive the evolution of the system. This conceptual understanding usually builds on detailed experiments that study specific aspects of the system (e.g. Brisson et al., 2003 for the crop model STICS). The set of model equations is referred to as "model structure" (Tao et al., 2018).

In addition to model structure, simulation requires values for all the model parameters. In essentially all uses of crop models, the model is first calibrated using observed data that is related to the target population for which predictions are required, for example observations for the specific variety of interest and/or for the particular set of growing environments of interest. Calibration is essentially universally necessary because mechanistic models are only approximations, without universally valid parameter values (Fath and Jorgensen, 2011; Wallach, 2011).

There are therefore two main tracks to improvement of crop phenology models. The first is through improvement of model structure through improved understanding of the underlying processes, and the second is through improvement of the model parameters. For a fixed data set, improvement of model parameters implies improvement of model calibration, and that is the topic here.

Crop models, like system models in general, are basically regression models, in that they predict outcomes of the system based on input variables. While calibration of regression models (usually referred to as parameter estimation in a statistical context) is a major topic in statistics, the application of statistical methods to system models is not straightforward. Among the difficulties are the fact that system models often have multiple output variables that can be compared to observed results (e.g., dates of heading and dates of flowering for crop phenology models) and

there are usually many parameters, often more than the number of data points available. While the details differ, these problems apply to essentially all system models, not only crop phenology models but also full crop models, hydrological models, ecology models  and models  in other fields. No doubt as a result, there are no widely accepted standard methods for calibration of system models. It has been found,  for example, that there is a wide diversity of calibration approaches and model outputs for crop phenology models furnished with identical data, even between modeling groups using the same model structure (Confalonieri et al., 2016; Wallach et al., 2021a, 2021b).

Because of the importance of calibration and the lack of standard approaches for calibration, there have been many studies published that make recommendations as to how to calibrate crop models or system models in other fields. One type of study is model-specific, and identifies the most important parameters to estimate for a particular model (Ahuja et al., 2011). Other studies have focused on the methodology of identifying the most important parameters through sensitivity analysis  (Khorashadi Zadeh et al., 2022), on the choice between frequentist and Bayesian paradigms (Gao et al., 2021), on the form of the objective function, or on the numerical algorithm for searching for the best parameter values (Rafiei et al., 2022). A recent study has emphasized the need to consider the full flow of the calibration exercise, including the choice of default parameter values, the choice of observed data to include in the objective function, the form of the objective function, the choice of parameters to estimate, and the choice of numerical algorithm for calculating the best parameter values, and proposed recommendations for each of those activities (Wallach et al., 2021c).

Recommendations for improved calibration are useful, but have important limitations. Not only do they generally concern only part of the calibration activity, but in addition they are generally not tested for a wide range of situations, to verify that they have general applicability, and they generally do not have any mechanism for ensuring that they are correctly followed. To date, recommendations have not resulted in a convergence of calibration practices for crop phenology models.

The purpose of this study is to propose, implement, and evaluate a calibration protocol for crop phenology models that does not have the above-mentioned limitations. Compared to usual recommendations, the protocol suggested here is more detailed, covers the full range of decisions involved in calibration, and includes documentation templates that help in correctly implementing the protocol and make the calibration strategy more transparent. To test the applicability of the protocol to a range of model structures, multiple modeling groups implemented the protocol, using multiple model structures. To test the applicability to differently structured data sets, each modeling group applied the protocol to three different wheat phenology data sets, with two quite different structures. The most important, and exacting, test of the proposed calibration protocol is its ability to  improve predictions of phenology for out-of-sample environments, compared to usual calibration practice. All of the modeling groups that applied the protocol in this study first used their usual method of calibration on the same data sets as those used here, in most cases as part of previous studies(Wallach et al., 2021a, 2021b). Thus, we were able to compare the predictive accuracy of usual calibration with calibration using the protocol proposed here. It is also important to reduce the uncertainty in phenology predictions, as measured by the variability between different modeling groups. Therefore, we also compared the

variability between groups for the case where each group uses their usual calibration approach or the protocol.

# Materials and Methods

## Data sets

The phenology stages targeted in this study are listed in **Error! Reference source not found.**, using the BBCH scale (Meier and Biologische Bundesanstalt fu□r Land- und Forstwirtschaft., 1997). Simulations were performed for three data sets. The two French data sets had a similar structure, but concerned two different winter wheat varieties. The third data set was from Australia, for a spring wheat variety. In each case, the data could be considered to come from a well-defined target population: conventionally managed wheat fields in the major wheat growing regions of France for the first two data sets, and of Australia for the third data set. The Australian data set included multiple planting dates from within the range of reasonable dates. Each data set was split into two subsets, one for calibration and one for evaluation. The two subsets had neither site nor year in common, so the evaluation is a rigorous test of how well a modeling group can simulate phenology for out-of-sample environments.

**Table 1**

**The BBCH development stages referred to in this study.**

| Growth stage | BBCH stage |
|---|---|
| Emergence | BBCH10 |
| Beginning of stem elongation | BBCH30 |
| Middle of heading | BBCH55 |
| Anthesis half way | BBCH65 |
| Fully ripe grain | BBCH90 |

The French data sets contained observations of stages BBCH30 and BBCH55. The Australian data set contained once weekly notations of BBCH stage in each environment. The Australian data were interpolated to give the date of each integer BBCH stage from the earliest that was observed to the latest, and these interpolated data were provided to the modeling groups. The interpolated data were provided to avoid each modeling group doing their own interpolation, which would add unwanted variability to the exercise. The number of environments (combinations of site and sowing date) and observations for each subset of data are shown in Table 2. More details about the French data set can be found in (Wallach et al., 2021a) and more details about the Australian data set in (Wallach et al., 2021b).

**Table 2.**
**Data for calibration and evaluation. The number of environments is the number of unique combinations of site and sowing date. There were no observations of BBCH10.**

| | | calibration | evaluation |
|---|---|---|---|

| France | Number of different environments | 14 | 8 |
|---|---|---|---|
| (both | Number of observations of BBCH30 | 14 | 8 |
| varieties) | Number of observations of BBCH55 | 14 | 8 |
| Australia | Number of different environments | 24 | 18 |
| | Number of observations of BBCH30 | 21 | 16 |
| | Number of observations of BBCH65 | 19 | 18 |
| | Number of observations of BBCH90 | 10 | 5 |

## Modeling groups

Nineteen modeling groups, using 16 different model structures (Supplementary Table S1), participated in this study, which was carried out within the Agricultural Model Intercomparison and Improvement Project (AgMIP; www.agmip.org). We refer to the simulated values as representing "modeling groups" rather than models since at least partly the results depend not only on model structure but also on decisions of the modeling group. The modeling groups are identified only by a code ("M1", "M2" etc.) without indicating which model structure they used, since it would be misleading to give the impression that the results are determined solely by model structure. The codes here refer to the same modeling groups as in Wallach et al. (2021a, 2021b).

## Evaluation

Our basic evaluation metric is the sum of squared errors (SSE) and the related quantities mean squared error (MSE) and root mean squared error (RMSE), where

$$SSE = \sum_i \sum_j \left( y_{ij} - \hat{y}_{ij} \right)^2$$
$$MSE = SSE / n \qquad (1)$$
$$RMSE = \sqrt{MSE}$$

The sum is over variables and environments. Here $y_{ij}$ is the observed value of variable $i$ for environment $j$, $\hat{y}_{ij}$ is the corresponding simulated value and $n$ is the number of terms in the sum. We also look at the decomposition of MSE as the sum of three terms, namely squared bias (bias²), a term related to the difference in standard deviations of the observed and simulated values (SDSD) and a term related to the correlation of observed and simulated vales (LCS) (Kobayashi, 2004).

In addition, we define two simple benchmark models. The first (the "naive" model) is simply the average number of days to each stage in the calibration data of each data set. This is used as the prediction model for all environments of that data set. The often used Nash Sutcliffe modeling efficiency is one minus the ratio of MSE of a model to MSE of the naive model. The naive model ignores all variability between environments, so it is a very low bar as a benchmark. We therefore also use a more sophisticated benchmark, the "onlyT" model, introduced in Wallach et al. (2021a). This benchmark model assumes that the sum of degree days above a threshold of 0°C from sowing to each stage is fixed for spring wheat. For winter wheat, a simple vernalization model is used, and then the fixed number of degree days applies after vernalization is completed

(van Bussel et al., 2015; Wallach et al., 2021a). Both benchmark models are quite easily parameterized based on calibration data, and then easily applied to new environments.

## Simulation exercise

The participants received input data (daily weather at the field, soil characteristics, management details and, where possible, initial conditions) for all environments of every data set. Also, the observed data from the calibration environments were provided to all participants. The participants were asked to use those data to calibrate their models, and then to simulate and report days after sowing to stages BBCH10, BBCH30, and BBCH55 for the French calibration and evaluation environments, and to stages BBCH10, BBCH30, BBCH65, and BBCH90 for the Australian calibration and evaluation environments. Days to emergence (BBCH10) was included to have an example of a variable for which there were no calibration data. The BBCH stages 30 and 55 requested for the French environments represent stages that are used for fertilizer decisions in France. The BBCH stages 30, 65, and 90 requested for the Australian environments represent major transitions that are explicitly simulated by many models.

Seventeen of the 19 participating modeling groups participated in previous phases of this project, and in that framework had already calibrated their model using their usual calibration approach (Wallach et al., 2021a, 2021b). The two remaining groups also calibrated their models using their usual approach before beginning to use the protocol proposed here. It is the results of the usual calibration method that are compared here to the results of using the proposed protocol. If any simulated results were missing from the usual calibration results, the corresponding results from the protocol calibration results were deleted, and vice versa. Thus, the results for the usual and protocol calibrations are comparable. At no time were the evaluation data shown to participants, neither in previous studies nor in the present study. To keep these data confidential, and thus, potentially useable in future studies, no graphs are presented here showing the evaluation data.

In the present study, participants were given the detailed protocol for model calibration and asked to implement it for the three data sets. The protocol was the same as described below, but the textual description was somewhat different. The team leaders used the protocol documentation generated by the modeling teams to identify problems and interact with the participants.

The protocol does not impose a specific software solution. However, several participants used trial and error in their usual approach and requested help in finding and implementing an automated search algorithm, since that is required for the protocol. To answer this need, the CroptimizR R package (Buis et al., 2021) was modified to do the protocol calculations, and many of the participants used this software.

In addition to the individual models, we report on two ensemble models, created by taking the mean (the e-mean model) or the median (the e-median model) of the simulated values. These ensemble models were calculated both for the usual and protocol calibration results.

## AICc and BIC

The protocol uses a model selection criterion to decide which parameters to estimate. The corrected Akaike Information Crition (AICc) and the Bayesian Information Criterion (BIC) are two different criteria that are often used for model selection (Chakrabarti and Ghosh, 2011). Both are based on model error, with a penalization term that increases with the number of estimated parameters. Assuming that model errors are normally distributed, the criteria are

$$AICc = n\ln(MSE) + 2p + \frac{2p(p+1)}{n-p-1} \tag{2}$$

$$BIC = n\ln(MSE) + p\ln(n)$$

where $n$ is the number of data points and $p$ is the number of calibrated parameters. These criteria are only used for comparing models calibrated using the same data, so any term that only involves $n$, the number of data points, has been dropped because it is the same for all models.

There have been comparisons between these criteria, but there does not seem to be one that systematically performs better than the other, for choosing the model that predicts best (Kuha, 2004). In applying the protocol here, participants were asked to perform the calculations twice, once using the AICc criterion and once using the BIC criterion to choose the parameters to estimate. In almost all cases, the two criteria led to exactly the same choice of parameters. In the few cases where the criteria led to different choices, the final models had very similar RMSE for the evaluation data, with a very slight advantage to BIC (Supplementary tables 22-23). All results shown here are based on the BIC criterion.

## The protocol

### Step 1. Choose default parameter values

The first step in the protocol is to define the default values for all parameters. In particular, the default values for cultivar specific parameters should be based on information from cultivars as similar as possible to the target cultivar. For phenology, one would want to have reasonable approximations to the cycle length for the cultivar in question, to photoperiod dependence and to vernalization requirements. The documentation for step 1 (see example in **Table 3Error! Reference source not found.**) specifies the cultivar which is used to provide default parameter values, and why that cultivar was chosen.

**Table 3**

**Example of protocol documentation for step 1, showing cultivar characteristics of the target population cultivar and of the cultivar that provides the default parameter values. This example is for the French data set and modeling group M21.**

| Variety | Characteristics |
|---|---|
| Variety of target population: Apache | A soft winter wheat. Stem elongation – semi-early. Heading – early. Vernalization requires 40 days where full vernalization occurs if daily average temperature is between 3°C and 10°C. There is no vernalization below -4°C or above 17°C. Otherwise there is a proportional reduction in vernalization effectiveness. |
| Default variety : Soissons | Soissons seems to be close to Apache in terms of vernalization requirements and earliness. |

## Step  2. Identify measured variables to be fit

List each of the observed variables and the corresponding simulated variables if any. In the simplest case, there is a simulated variable that corresponds directly to each measured variable. The documentation for step 2 is a table with one row for each observed variable (Table 4) .

**Table 4**
**Example of documentation of protocol step 2 with one row for each measured variable, showing the corresponding simulated variable. This example is for the French data set and modeling group M21.**

| Measured variable | Corresponding simulated value |
| --- | --- |
| Days to BBCH stage 30 | Days to end juvenile stage |
| Days to BBCH stage 55 | Days at maximum LAI |

## Step 3. Define the objective function

The protocol uses as objective function to be minimized SSE, where the sum over variables is over the variables from step 2 that have both observed and simulated values.  No choices are required here. Once step 2 has been finalized, the objective function follows.

## Step 4. Determine which parameters to estimate

The protocol distinguishes two categories of parameters to estimate: the obligatory parameters (those that will definitely be estimated) and the candidate parameters (those that will be tested, and only changed from the default value if the improvement in the fit to the calibration data is sufficiently large).

## Step 4a. Identify the obligatory parameters

The obligatory parameters are parameters that are nearly additive, i.e. such that changing the parameter has a similar effect for all environments. Usually, a parameter that represents degree days to a measured stage is a good choice as an obligatory parameter for phenology. Ideally, the number of obligatory parameters will be identical to the number of variables in the objective function. If one cannot identify a nearly additive parameter for every variable, there can be fewer nearly additive parameters than variables. There must not be more nearly additive parameters than variables. The documentation of step 4a (see Table 5**Error! Reference source not found.**) has one row for each obligatory parameter.

**Table 5**
**Example of documentation for protocol step 4a, showing obligatory parameters for the French data set, variety Apache for modeling group M21.**

| Name of obligatory parameter | explanation | Default value (lower,upper limits) |
| --- | --- | --- |
| stlevamf | Degree days sowing to end juvenile stage | 233 (150-400) |

| stamflax | Degree days sowing to maximum LAI | 354 (150-500) |
|---|---|---|

## Step 4b. Identify candidate parameters

The role of the candidate parameters is to reduce the variability between environments that remains after estimation of the obligatory parameters. It is the role of the modeler to identify the parameters that seem likely, when estimated, to explain a substantial part of the unexplained variability between environments, and to order them by amount of variability likely to be explained. **Error! Reference source not found.**The documentation for protocol step 4b has one row for each candidate parameter (Table 6).

**Table 6**
**Example of documentation for protocol step 4b, showing candidate parameters for the French data set, variety Apache for modeling group M21.**

| Candidate parameter (include units and a brief explanation) | Default value (Lower and upper bounds) |
|---|---|
| jvc (days): number of vernalizing days | 38 (25 – 60) |
| sensrsec (no unit): index of root sensitivity to drought (1=insensitive) | 0.5 (0 – 1) |
| belong (degree.day-1): parameter of the curve of coleoptile elongation | 0.012 (0.005 – 0.03) |
| JVCmini (days): minimum vernalizing days required | 7.0 (2 – 15) |
| stressdev (no unit): maximum phasic delay allowed due to stresses | 0 (0 – 1) |

Our definition of upper and lower bounds is that the modeler would be very surprised if the true best value for this target population were outside these bounds. This is admittedly a rather vague definition, but it is meant to translate the fact that while we do not know the true best value of each parameter, we do have some knowledge about a reasonable range.

## Step 5. Calculation of the optimal parameter values

The protocol prescribes using an algorithm based on the Nelder-Mead simplex, which is a simple robust derivative-free optimization algorithm, to minimize SSE. The optimization is done in several steps, first for the obligatory parameters and then for each candidate parameter. At each step, one estimates the previously chosen parameters plus the next candidate parameter. If this leads to a smaller value of the BIC criterion than the previous best model, the candidate parameter is added to the list of chosen parameters. If not, the candidate parameter returns to its default value and is not added to the list of parameters to estimate. The model with the lowest value of BIC is the final model. Table 7**Error! Reference source not found.** shows the documentation of protocol step 5, which has one row for each minimization step.

**Table 7**

**Example of documentation for protocol step 5, showing the calculations for the French data set, variety Apache, modeling group M21. The first line shows the optimization results for the obligatory parameters, then each subsequent line corresponds to a candidate parameter. In this example, the first candidate parameter (jvc) is accepted, and all the subsequent candidate parameters increase BIC, and are therefore, rejected. The model finally chosen (minimum BIC) has three estimated parameters.**

| Estimated.parameters | Initial.parameter.values | Final.values | Sum.of.squared. errors | BIC |
|---|---|---|---|---|
| stlevamf, stamflax | multiple | 227, 360 | 405 | 81.47 |
| **stlevamf, stamflax, jvc** | **227, 360, multiple** | **212, 367,  55.91** | **349** | **80.64** |
| stlevamf, stamflax, jvc, sensrsec | 212, 367, 55.91, multiple | 209, 367, 58.40, 0.057 | 322 | 81.71 |
| stlevamf, stamflax, jvc, belong | 212, 367, 55.91, multiple | 212, 367, 55.91, 0.012 | 349 | 83.97 |
| stlevamf, stamflax, jvc, jvcmini | 212, 367, 55.91, multiple | 197, 362, 55.28, 20.88 | 319 | 81.45 |
| stlevamf, stamflax, jvc, stressdev | 212, 367, 55.91, multiple | 212, 367, 55.91, 0.00 | 349 | 83.97 |

For the initial step, with more than one parameter to estimate, the protocol recommends using at least 20 starting values, chosen within the range of limits for each parameter. Latin hypercube sampling could be used to distribute the starting values within the space of possible values. When adding an additional candidate parameter, the protocol recommends starting from the parameter values that gave the previous lowest BIC value, and five different starting values for the new parameter.

## Step 6 Diagnostics

    a.  The protocol evaluation graph shows simulated versus observed values, with a different symbol for each variable (Figure 1).

**Figure 1**

**Graph for protocol documentation of step 6a, showing simulated and observed values of BBCH30 and BBCH55. The results are for the French data set, variety Apache, modeling group M21.**

    b.   Calculate MSE and its components for each variable (Table 8)

**Table 8**
**Example of documentation of protocol step 6b, showing the calculations for the French data set, variety Apache, modeling group M21. Mean squared error (MSE) is a sum of bias$^2$, SDSD, and LCS.**

|  | MSE (days$^2$) | bias$^2$ (days$^2$) | SDSD (days$^2$) | LCS (days$^2$) |
|---|---|---|---|---|
|  |  |  |  |  |
| BBCH30 | 19.64 | 0.25 | 5.93 | 13.46 |
| BBCH55 | 5.29 | 0.02 | 0.03 | 5.24 |

    c.   Compare results to benchmark models (Table 9).

**Table 9**
**Example of documentation for protocol step 6c, showing results for benchmark models for the French data set, variety Apache, modeling group M21. The benchmark "naive" assumes a constant number of days to each stage. The benchmark "onlyT" assumes a constant temperature sum to each stage.**

| | naive | onlyT | M21 |
|---|---|---|---|
| variable | RMSE (days) | RMSE (days) | RMSE (days) |
| BBCH30 | 12.5 | 8.4 | 3.1 |
| BBCH55 | 8.3 | 9.5 | 3.8 |

# Results

## Comparison of usual and protocol calibration for individual models

### Number of parameters and simulated values

There were substantial differences between the consequences of doing usual or protocol calibration. For the large majority of modeling groups, the number of estimated parameters in the final model was different between protocol and usual calibration (Supplementary Figure 1). **Table 10** shows that on average over modeling groups, protocol calibration led to fewer parameters (French data sets) or the same number of parameters (Australian data set) to be estimated in comparison to usual calibration. The modeling groups considered a larger number of parameters in the protocol calibration than in the usual calibration, but then rejected most of the candidates.

**Table 10**
**Number of estimated parameters in the final model after protocol or usual calibration, averaged over modeling groups.**

| Data set | Protocol calibration | | | | Usual calibration |
|---|---|---|---|---|---|
| | Additive parameters | Candidate parameters | Accepted candidate parameters | Total parameters | Total parameters |
| France Apache | 1.9 | 3.6 | 0.7 | 2.6 | 3.5 |
| France Bermude | 1.9 | 3.6 | 0.9 | 2.8 | 3.6 |
| Australia | 4.0 | 4.5 | 1.7 | 5.7 | 5.7 |

The differences between simulated values after usual and protocol calibration were small for BBCH10, for which there were no calibration data. For the other stages, the simulated values differed appreciably. The mean absolute difference between simulated values using usual or protocol calibration is shown in Table 11. Results by modeling group are shown in Supplementary Figure 2.

**Table 11**
**Mean over modeling groups and environments of the absolute difference in simulated values between usual calibration and protocol calibration for the calibration data.**

| French data set | stage | BBCH10 | BBCH30 | BBCH55 | |
|---|---|---|---|---|---|
| (Apache and | Mean | 0.8 | 4.5 | 3.1 | |

| Bermude) | absolute difference (days) | | | | |
|---|---|---|---|---|---|
| Australian data set | stage | BBCH10 | BBCH30 | BBCH65 | BBCH90 |
| | Mean absolute difference (days) | 1.8 | 7.9 | 7.9 | 8.5 |

## Goodness of fit and evaluation

Table 12 shows RMSE values for each data set, averaged over modeling groups, for usual and protocol calibration for the calibration and evaluation data. The *p* values for a one-sided paired *t*-test of the hypothesis that RMSE is larger for usual calibration than for protocol calibration are also shown. On average over stages, all three data sets have significantly larger RMSE values with usual calibration than with protocol calibration for the calibration data ($p<0.05$). For the evaluation data, p<0.01 for the two French data sets, but *p*=0.15 for the Australian data set. The table also shows the proportion of modeling groups where RMSE is larger for the usual calibration than for the protocol calibration. Looking at the averages over stages and then averaging over data sets, 75% of models have lower RMSE for protocol calibration than for usual calibration for the calibration data, and 66% for the evaluation data. Results for each modeling group are shown in Supplementary Figures 2-3 and Supplementary Tables 2-7.

Almost all modeling groups did better than the two benchmark models for all stages. Averaged over stages, for the evaluation data, only 2, 1, and 4 modeling groups using usual calibration did worse than the onlyT model for the French Apache, French Bermude, and Australian data sets, respectively. Using protocol calibration, 0, 1, and 1 modeling groups did worse than the onlyT model for the French Apache, French Bermude, and Australian data sets respectively.

**Table 12**
**RMSE averaged over modeling groups for each stage and averaged over stages for the calibration and evaluation data, using usual or protocol calibration, for each data set. The *p* value is for the test that RMSE using usual calibration is larger than RMSE using protocol calibration, and below that is the proportion of models for which RMSE using usual calibration is larger than RMSE using protocol calibration.**

| | | Calibration data | | | Evaluation data | | |
|---|---|---|---|---|---|---|---|
| | | Usual RMSE | protocol RMSE | *p*-value $RMSE_u > RMSE_p$ | usual RMSE | protocol RMSE | *p*-value $RMSE_u > RMSE_p$ |
| France Apache | BBCH30 | 7.7 | 6 | 0.007 10/16 | 6.7 | 6.2 | 0.20 9/16 |
| | BBCH55 | 5 | 4 | 0.02 11/17 | 7.3 | 5.8 | 0.005 13/17 |
| | average | 6.4 | 5 | 0.004 14/17 | 7 | 6 | 0.006 14/17 |
| France Bermude | BBCH30 | 7.1 | 5.4 | 0.05 11/16 | 8.8 | 7.5 | 0.07 9/16 |
| | BBCH55 | 6.8 | 5.7 | 0.04 11/17 | 6.9 | 6.1 | 0.08 11/17 |
| | average | 7 | 5.6 | 0.029 13/17 | 7.8 | 6.8 | 0.008 11/17 |
| Australia | BBCH30 | 13.3 | 12.0 | 0.09 9/17 | 15.1 | 14.8 | 0.38 8/17 |
| | BBCH65 | 11.4 | 9.0 | 0.06 14/19 | 11.1 | 11.4 | 0.60 10/19 |
| | BBCH90 | 11.9 | 9.9 | 0.096 11/18 | 9.0 | 6.2 | 0.20 12/18 |
| | average | 12.2 | 10.2 | 0.049 13/19 | 11.7 | 10.7 | 0.15 10/19 |

## Decomposition of MSE

The fraction of MSE due to each of bias², SDSD, and LCS is shown in

In every case, protocol calibration reduces the fraction of MSE due to bias. Bias is smaller for the calibration data than for the evaluation data. Details for each modeling group are shown in Supplementary Tables 9-23.

**Table 13**
**The decomposition of MSE on the average over modeling groups and over BBCH30 and BBCH55 for the French data sets and over BBCH30, BBCH65, and BBCH90 for the Australian data set.**

| Data set | calibration | Bias²/MSE | SDSD/MSE | LCS/MSE |
|---|---|---|---|---|
| Apache calibration data | usual | 0.15 | 0.18 | 0.67 |
| | protocol | 0.12 | 0.15 | 0.73 |
| Apache evaluation data | usual | 0.39 | 0.13 | 0.47 |
| | protocol | 0.35 | 0.13 | 0.52 |
| Bermude calibration data | usual | 0.23 | 0.19 | 0.58 |
| | protocol | 0.16 | 0.11 | 0.73 |
| Bermude evaluation data | usual | 0.38 | 0.15 | 0.47 |
| | protocol | 0.36 | 0.14 | 0.50 |
| Australian calibration data | usual | 0.23 | 0.27 | 0.52 |
| | protocol | 0.10 | 0.24 | 0.69 |
| Australian evaluation data | usual | 0.35 | 0.16 | 0.51 |
| | protocol | 0.28 | 0.16 | 0.59 |

The choice of usual or protocol calibration has little effect on the predictive accuracy of the ensemble models e-mean and e-median. Averaged over development stages (not including BBCH10) and over data sets, for the evaluation data, RMSE for e-median is respectively 5.7 and 5.8 days for usual and protocol calibration. The values for RMSE of e-mean are 6.1 and 6.2 days for usual and protocol calibration, respectively (Supplementary Tables 3, 5, 7, 8).

The variability between simulated results for individual modeling groups is shown in Table 14**Error! Reference source not found.**. The standard deviation is similar for usual and protocol calibration for BBCH10, for which there are no data for calibration, but is systematically smaller for protocol calibration for the other stages. Considering the average over stages other than BBCH10 and taking the average over data sets, protocol calibration reduced the standard deviation of simulated values by 31% for the calibration data and by 22% for the evaluation data.

**Table 14**
**Standard deviation of values simulated by modeling groups (days). The average is over stages without BBCH10.**

|  |  | Calibration data | | Evaluation data | |
| --- | --- | --- | --- | --- | --- |
|  |  | usual | protocol | usual | protocol |
| France Apache | BBCH10 | 4.2 | 4.1 | 4.8 | 5.2 |
|  | BBCH30 | 6.4 | 4.3 | 6.2 | 5.5 |
|  | BBCH55 | 4.5 | 3.0 | 6.3 | 3.7 |
|  | average | 5.4 | 3.6 | 6.2 | 4.6 |
| France Bermude | BBCH10 | 4.3 | 4.4 | 4.8 | 6.6 |
|  | BBCH30 | 6.9 | 4.3 | 6.7 | 6.2 |
|  | BBCH55 | 4.8 | 3.5 | 5.8 | 4.3 |
|  | average | 5.9 | 3.9 | 6.2 | 5.3 |
| Australia | BBCH10 | 8.6 | 7.5 | 9.6 | 8.0 |
|  | BBCH30 | 11.3 | 7.3 | 10.2 | 8.1 |
|  | BBCH65 | 10.3 | 7.4 | 8.2 | 7.1 |
|  | BBCH90 | 11.2 | 9.3 | 11.1 | 6.9 |
|  | average | 10.9 | 8.0 | 9.8 | 7.4 |

# Discussion

## The protocol

The proposed protocol follows the recommendations from Wallach et al. (2021c), but with additional details to extend, standardize, and document the implementation of those recommendations. The protocol covers all the steps involved in calibration, with detailed instructions for each step.

Step 1 default values. The step of choosing default parameter values is rarely if ever mentioned in discussions of system model calibration. It is however very important. In general, one only has enough data to estimate new values for a small fraction of all parameters. All other parameters will retain their default values, so it is important to choose appropriate default values. Furthermore, the protocol calculations work best if the default values of the parameters to be estimated are reasonably close to the new best values. The protocol provides guidelines for choosing default values or a default variety, based on earliness, photoperiod sensitivity, and vernalization requirements, and specifies the documentation to record and explain the choices made.

Step 2 variables to fit. Often this step is straightforward. One lists the observed variables and the corresponding simulated variables. In some cases, however, there may be stages for which simulations are required, and for which there are observations, but which are not specifically identified in the model. In the study here, for example, several models do not specifically simulate the stage BBCH30, which was observed in the French data set and which was to be simulated. Most models do, however, simulate an internal growth stage variable. One can then treat the internal growth stage that corresponds to BBCH30 as a new parameter, to be estimated. This is the recommendation of the protocol. This approach makes it possible for a much wider range of models to use all the data for calibration, than if only observed variables specifically simulated by the model were used.

Step 3 objective function. In discussing system models in hydrology, Hernandez-Suarez et al. (2021) distinguish two categories of objective functions, one based on traditional performance metrics and the second adapted to the specific aspects of the system that are of primary interest. This choice applies generally to system models, and here we opt for the first possibility. The objective function of the protocol is simply the total sum of squared errors, which is the objective function of ordinary least squares (OLS) regression and is often used in crop model calibration. Another major choice of the protocol is to include in the objective function all the observed variables that have a simulated equivalent, including variables that are not of primary interest. For example, in the study here, the Australian data set has observations of many stages. The protocol says to use all possible data, and in fact, most modeling groups did indeed use observations of other stages for calibration, in addition to data for the BBCH stages 30, 65, and 90, which are used for evaluation. A first reason for using all data is that often the same calibrated model will be used for several different objectives, so measured variables that are not of central interest in the current study may be important in future studies. Furthermore, using more variables makes the model a better representation of multiple aspects of the system dynamics, which is likely to improve all simulations.

The model with parameter values that minimize sum of squared errors has attractive optimal properties if the standard assumptions underlying OLS are satisfied (Seber and Wild, 1989). It has been argued that crop models probably do not fully satisfy these assumptions, not even the first assumption that the model is correctly specified, i.e. that there are parameter values such that the expectation of model error is zero (Wallach, 2011). However, even in this case, OLS has the property that the estimated parameter values tend, as the amount of data from the target population increases, toward the best parameter values (minimum SSE) for this model and target

population, (White, 1981). This is the justification for using OLS even if the model is misspecified.

The assumptions underlying the optimal properties of OLS also include homoscedasticity (equal variances of error for all variables) and independence between errors for all variables. Once again, these assumptions are probably not fully satisfied for crop phenology models. If problems of heteroscedasticity (unequal error variances) and non-independence are ignored, the calibration does not make optimal use of the data. One could correct the problem of heteroscedasticity by using weighted least squares, and the problem of non-independence by using generalized least squares (Seber and Wild, 1989), but in both cases that would require estimating additional parameters, related to the standard deviation or to the variance-covariance matrix of model error. The assumption here is that the problems of heteroscedasticity and non-independence are often not too important for crop phenology models, since the variables are all times to various development stages and thus, may have similar error variances, and different environments often have similar amounts of data, so taking non-independence into account would not greatly change the relative importance of different environments. The choice in the protocol is, therefore, to use OLS and to avoid estimating additional parameters related to variance and covariance of errors.

An alternative approach would be to do a Bayesian calculation, where one calculates the posterior parameter distribution rather than a single best parameter vector. However, this is more difficult computationally, and furthermore requires specification of the distribution of model error, including variances and covariances. Also, while in principle one could estimate any number or parameters, in practice, because of the large number of parameters, there is always a need to choose which parameters to include in the calculation.

Step 4 choice of parameters. Usual calibration approaches include various methods of choosing which parameters to estimate, including sensitivity analysis, expert opinion, and testing different options to find the best fit to the data (Wallach et al., 2021d). The protocol here combines expert opinion and a data-based criterion of choice.

The protocol identifies two different groups of parameters, the obligatory, nearly additive parameters and the candidate parameters. The role of the nearly additive parameters is to eliminate or nearly eliminate bias (i.e. make the average of observed and simulated values equal or nearly equal). If a model includes a constant and the constant is fit to the data using a least squares criterion, bias will be zero for the calibration data. One possible strategy then is simply to add one additive parameter for each output variable to each model, but this is often unacceptable, since it involves changing the model. The approach for the protocol is to identify existing model parameters that are almost additive, i.e. such that changing the parameter has a similar effect for all environments. Most crop models have parameters that represent degree days to different stages and these can be considered almost additive parameters since increasing degree days required to reach a stage will generally increase days to that stage by a similar amount for all environments. A few of the models in this study do not use a degree day formulation, but were able to choose other parameters that are almost additive. For example, one group chose as an almost additive parameter, the parameter that scales development rate against temperature. Reducing bias reduces MSE for the calibration data, since squared bias is one of the three terms that add up to MSE (Kobayashi and Salam, 2000). Once bias is nearly eliminated, one may already have a fairly reasonable fit to the data.

The number of almost additive parameters cannot be greater than the number of variables in the objective function, and each must be nearly additive for a different variable or combination of variables. Otherwise, the parameters would be very poorly estimated, or non-estimable if they are exactly additive. The protocol does allow fewer almost additive parameters than observed variables. In that case bias is only nearly eliminated on average over several variables, and not for each variable.

Certainly, one wants a model that does better than just eliminating bias. One also wants to explain as much as possible of the variability between environments. Some, but not all of this variability will already be explained using the default values of parameters other than the obligatory parameters. The candidate parameters are those parameters that the modeler considers as likely to explain an important part of the remaining unexplained variability between environments. Each parameter is associated with a particular process, so the choice here is of the process and then of the specific parameter in the equations describing that process. For example, in Table 6, the first and last candidate parameters are both associated with crop vernalization, but affect different aspects of that process. In the calculation steps, each candidate parameter is tested, and only those that lead to a reduction in BIC are retained for estimation. Otherwise, the parameter is kept at its default value. There should be only a limited number of candidate parameters, with those thought most important first, in order to reduce the risk of selection bias where unimportant parameters are chosen because of random error (Lukacs et al., 2010). The choice of almost additive and candidate parameters is the calibration step which requires the most detailed knowledge of the model.

Step 5 calculation. The algorithm chosen for searching the parameter space for the values that minimize SSE is the Nelder-Mead simplex algorithm. This is a derivative-free method, which is appropriate for crop models which may have multiple discontinuities. However,. the results are sensitive to starting values (Wang and Shoup, 2011), and there is no guarantee that the algorithm will converge to the global optimum. Therefore, the protocol calls for multiple starting points. The calculations take advantage of knowledge of the system to focus the search for best parameters on a limited portion of the parameter space, thus reducing calculation time. Specifically, when a new candidate parameter is estimated, it is expected that the optimal values of the other estimated parameters will often be similar to their previous optimal values. Therefore, the algorithm only samples multiple starting points for the new candidate parameter. The other estimated parameters have their previous optimal values as the starting point.

The upper and lower bounds for each parameter are used in two ways. First, the starting values for the optimization are sampled from within this range. Secondly, if the estimated value of a parameter is outside this range, this is a signal that the simulated dynamics will have unrealistic aspects. According to the protocol, the algorithm should not constrain the estimated parameter values to be within the prescribed upper and lower bounds. The reason is that it is important to know if the optimal value is outside this range. If it is, one should reexamine the model and the calibration.

Step 6 Diagnostics for simulation data. Plotting simulated versus observed values and comparing MSE with that of the naive model are common diagnostics used for crop models. The decomposition of MSE into separate contributions is also fairly common, but here this has a special importance. It is particularly the squared bias contribution to MSE that is of interest.

Since the protocol is designed to nearly eliminate bias, one would want to investigate further if this is not the case. A much less common diagnostic is the second benchmark model, the onlyT model. This is more realistic than the naive model, but is still simple enough to be easily calibrated. If MSE for a model is larger than for either of the benchmarks, one should reexamine the model, the data or the calibration.

## Goodness-of-fit and out-of-sample predictions

The protocol reduces RMSE averaged over predicted stages, for all cases (all three data sets, both calibration and evaluation data). The reduction is of 10-22% compared to RMSE for the usual calibration method. Not all individual models, though the majority, were improved. The improvement is less for the evaluation data than for the calibration data, but is still notable. The reduction in RMSE in almost all cases is statistically significant.

The evaluation data here are from environments similar to those of the calibration data. This has the important advantage of being a standard situation that does not introduce the additional complication of the degree of dissimilarity between the calibration and evaluation environments. However, it leaves open the question of how useful protocol calibration would be if the models were used to extrapolate to quite different conditions, such as projected future climate. Nonetheless, the improvement here for out-of-sample predictions is certainly encouraging.

Since the protocol specifically aims to reduce bias, one would expect squared bias to be a smaller fraction of MSE for protocol calibration than for usual calibration, and this is the case, both for the calibration data and the evaluation data.

In both usual and protocol methods, almost all models are better than both benchmarks. In general, the onlyT benchmark is better than the naive benchmark, but not always. It is useful to have both benchmarks.

## Ensemble results

Recently, many crop model studies have been based on ensembles of models (Jägermeyr et al., 2021). Many studies have found that the ensemble mean and median are good predictors, sometimes better than even the best individual model (Martre et al., 2015; Wallach et al., 2018). It has, thus, become quite common to base projections of climate change impact on crop production on the ensemble median (e.g. Asseng et al., 2019). The e-mean and e-median results here, compared to the individual modeling groups, are in keeping with previous results. The e-median model is among the better predictors though not the very best, and is somewhat better than e-mean. Using protocol calibration does not improve the e-mean or e-median models. The RMSE values for e-median for usual and protocol calibration, averaged over the evaluation data sets, are very similar, with the usual calibration e-median model being marginally better.

The variability among simulated values is a measure of uncertainty in impact assessments (Asseng et al., 2013). This variability arises from differences in model structure, but also from parameter uncertainty and uncertainty in climate projections (Tao et al., 2018) The variability due to the method of calibration has not been specifically studied. The comparison here between the variability after usual and protocol calibration indicates whether, and how much, variability between modeling groups can be reduced if all groups apply the same calibration procedure. For the calibration environments, when all modeling groups use the protocol, the standard deviation is reduced by over 30% compared to the case where each group uses its usual method. That is,

even for the relatively simple case of crop phenology models, calibration approach accounts for over 30% of the variability in simulated results for the calibration data. For the evaluation data the reduction in variability is less but still substantial (on average 22%). It seems that using a standard calibration approach would be an effective way of substantially reducing variability of crop phenology model simulations.

## Conclusions

We have shown that it is possible to define a detailed protocol for calibration of crop phenology models that is applicable to a wide range of wheat models and to data sets with different structures. The protocol is designed to provide strict instructions where possible and clear guidelines where input from the modeling team is required. While the application here is to wheat phenology models, the same protocol could undoubtedly be used more generally for phenology models of other crops.

Comparison with usual calibration practices shows that, on average over modeling groups, the protocol leads to a better fit to the calibration data and to a better fit to out-of-sample data. Use of the protocol would be advantageous not just for individual modeling studies, but also for studies based on ensembles of models, including projections of climate impact. In particular, we have shown that if all modeling groups use the protocol, between-model variability can be substantially reduced, thus reducing the uncertainty of projections.

Models of crop phenology are in general relatively simple, compared to models that simulate not only phenology, but also crop growth and soil processes. It seems likely that having a standardized protocol for calibration would be even more important for these more complex models than for simpler models. Defining a protocol for more complex models would introduce new problems compared to the study here, in particular because of the number of output variables. However, the protocol here would be a useful starting point for treating more complex situations.

## References

Ahuja, L. (Lajpat), Ma, L., American Society of Agronomy., Crop Science Society of America., Soil Science Society of America., 2011. Methods of introducing system models into agricultural research. American Society of Agronomy.

Asseng, S., Ewert, F., Rosenzweig, C., Jones, J.W., Hatfield, J.L., Ruane, A.C., Boote, K.J., Thorburn, P.J., Rötter, R.P., Cammarano, D., Brisson, N., Basso, B., Martre, P., Aggarwal, P.K., Angulo, C., Bertuzzi, P., Biernath, C., Challinor, A.J., Doltra, J., Gayler, S., Goldberg, R., Grant, R., Heng, L., Hooker, J., Hunt, L.A., Ingwersen, J., Izaurralde, R.C., Kersebaum, K.C., Müller, C., Naresh Kumar, S., Nendel, C., O'Leary, G., Olesen, J.E., Osborne, T.M., Palosuo, T., Priesack, E., Ripoche, D., Semenov, M.A., Shcherbak, I., Steduto, P., Stöckle, C., Stratonovitch, P., Streck, T., Supit, I., Tao, F., Travasso, M., Waha, K., Wallach, D., White, J.W., Williams, J.R., Wolf, J., 2013. Uncertainty in simulating wheat yields under climate change. Nat. Clim. Chang. 3, 827–832. https://doi.org/10.1038/nclimate1916

Asseng, S., Martre, P., Maiorano, A., Rötter, R.P., O'Leary, G.J., Fitzgerald, G.J., Girousse, C., Motzo, R., Giunta, F., Babar, M.A., Reynolds, M.P., Kheir, A.M.S., Thorburn, P.J., Waha, K., Ruane, A.C., Aggarwal, P.K., Ahmed, M., Balkovič, J., Basso, B., Biernath, C., Bindi, M., Cammarano, D., Challinor, A.J., De Sanctis, G., Dumont, B., Eyshi Rezaei, E., Fereres, E., Ferrise, R., Garcia-Vila, M., Gayler, S., Gao, Y., Horan, H., Hoogenboom, G., Izaurralde, R.C., Jabloun, M., Jones, C.D., Kassie, B.T., Kersebaum, K.-C., Klein, C., Koehler, A., Liu, B., Minoli, S., Montesino San Martin, M., Müller, C.,

Naresh Kumar, S., Nendel, C., Olesen, J.E., Palosuo, T., Porter, J.R., Priesack, E., Ripoche, D., Semenov, M.A., Stöckle, C., Stratonovitch, P., Streck, T., Supit, I., Tao, F., Van der Velde, M., Wallach, D., Wang, E., Webber, H., Wolf, J., Xiao, L., Zhang, Z., Zhao, Z., Zhu, Y., Ewert, F., 2019. Climate change impact and adaptation for wheat protein. Glob. Chang. Biol. 25, 155–173. https://doi.org/10.1111/gcb.14481

Bassu, S., Brisson, N., Durand, J.-L., Boote, K., Lizaso, J., Jones, J.W., Rosenzweig, C., Ruane, A.C., Adam, M., Baron, C., Basso, B., Biernath, C., Boogaard, H., Conijn, S., Corbeels, M., Deryng, D., De Sanctis, G., Gayler, S., Grassini, P., Hatfield, J., Hoek, S., Izaurralde, C., Jongschaap, R., Kemanian, A.R., Kersebaum, K.C., Kim, S.-H., Kumar, N.S., Makowski, D., Müller, C., Nendel, C., Priesack, E., Pravia, M.V., Sau, F., Shcherbak, I., Tao, F., Teixeira, E., Timlin, D., Waha, K., 2014. How do various maize crop models vary in their responses to climate change factors? Glob. Chang. Biol. 20, 2301–20. https://doi.org/10.1111/gcb.12520

Brisson, N., Gary, C., Justes, E., Roche, R., Mary, B., Ripoche, D., Zimmer, D., Sierra, J., Bertuzzi, P., Burger, P., Bussière, F., Cabidoche, Y.M., Cellier, P., Debaeke, P., Gaudillère, J.P., Hénault, C., Maraux, F., Seguin, B., Sinoquet, H., 2003. An overview of the crop model STICS. Eur. J. Agron. 18, 309–332.

Buis, S., Lecharpentier, P., Vezy, R., tvailhere, GinerM, Holzworth, D., 2021. SticsRPacks/CroptimizR: v0.4.0. https://doi.org/10.5281/ZENODO.5121194

Chakrabarti, A., Ghosh, J.K., 2011. AIC, BIC and Recent Advances in Model Selection. Philos. Stat. 583–605. https://doi.org/10.1016/B978-0-444-51862-0.50018-6

Cleland, E.E., Chuine, I., Menzel, A., Mooney, H.A., Schwartz, M.D., 2007. Shifting plant phenology in response to global change. Trends Ecol. Evol. 22, 357–365. https://doi.org/10.1016/j.tree.2007.04.003

Confalonieri, R., Orlando, F., Paleari, L., Stella, T., Gilardelli, C., Movedi, E., Pagani, V., Cappelli, G., Vertemara, A., Alberti, L., Alberti, P., Atanassiu, S., Bonaiti, M., Cappelletti, G., Ceruti, M., Confalonieri, A., Corgatelli, G., Corti, P., Dell'Oro, M., Ghidoni, A., Lamarta, A., Maghini, A., Mambretti, M., Manchia, A., Massoni, G., Mutti, P., Pariani, S., Pasini, D., Pesenti, A., Pizzamiglio, G., Ravasio, A., Rea, A., Santorsola, D., Serafini, G., Slavazza, M., Acutis, M., 2016. Uncertainty in crop model predictions: What is the role of users? Environ. Model. Softw. 81, 165–173. https://doi.org/10.1016/j.envsoft.2016.04.009

Fath, B., Jorgensen, S.E., 2011. Fundamentals of ecological modelling: Applications in environmental management and research. 4th edition. Elsevier, Amsterdam.

Gao, Y., Wallach, D., Hasegawa, T., Tang, L., Zhang, R., Asseng, S., Kahveci, T., Liu, L., He, J., Hoogenboom, G., 2021. Evaluation of crop model prediction and uncertainty using Bayesian parameter estimation and Bayesian model averaging. Agric. For. Meteorol. 311, 108686. https://doi.org/10.1016/J.AGRFORMET.2021.108686

Hernandez-Suarez, J.S., Nejadhashemi, A.P., Deb, K., 2021. A novel multi-objective model calibration method for ecohydrological applications. Environ. Model. Softw. 144, 105161. https://doi.org/10.1016/J.ENVSOFT.2021.105161

Jägermeyr, J., Müller, C., Ruane, A.C., Elliott, J., Balkovic, J., Castillo, O., Faye, B., Foster, I., Folberth, C., Franke, J.A., Fuchs, K., Guarin, J.R., Heinke, J., Hoogenboom, G., Iizumi, T., Jain, A.K., Kelly, D., Khabarov, N., Lange, S., Lin, T.-S., Liu, W., Mialyk, O., Minoli, S., Moyer, E.J., Okada, M., Phillips, M., Porter, C., Rabin, S.S., Scheer, C., Schneider, J.M., Schyns, J.F., Skalsky, R., Smerald, A., Stella, T., Stephens, H., Webber, H., Zabel, F., Rosenzweig, C., 2021. Climate impacts on global agriculture emerge earlier in new generation of climate and crop models. Nat. Food 2, 873–885. https://doi.org/10.1038/s43016-021-00400-y

Khorashadi Zadeh, F., Nossent, J., Woldegiorgis, B.T., Bauwens, W., Van Griensven, A., 2022. A fast and effective parameterization of water quality models. Environ. Model. Softw. 149, 105331. https://doi.org/10.1016/J.ENVSOFT.2022.105331

Kobayashi, K., 2004. Comments on another way of partitioning mean squared deviation proposed by Gauch et al. (2003). With reply. Agron. J. 96, 1206–1207.

Kobayashi, K., Salam, M.U., 2000. Comparing simulated and measured values using mean squared deviation and its components. Agron. J. 92, 345–352.

Kuha, J., 2004. AIC and BIC: Comparisons of Assumptions and Performance. Sociol. Methods Res. 33, 188–229. https://doi.org/10.1177/0049124103262065

Li, T., Hasegawa, T., Yin, X., Zhu, Y., Boote, K., Adam, M., Bregaglio, S., Buis, S., Confalonieri, R., Fumoto, T., Gaydon, D., Marcaida, M., Nakagawa, H., Oriol, P., Ruane, A.C., Ruget, F., Singh, B.-, Singh, U., Tang, L., Tao, F., Wilkens, P., Yoshida, H., Zhang, Z., Bouman, B., 2015. Uncertainties in predicting rice yield by current crop models under a wide range of climatic conditions. Glob. Chang. Biol. 21, 1328–41. https://doi.org/10.1111/gcb.12758

Lukacs, P.M., Burnham, K.P., Anderson, D.R., 2010. Model selection bias and Freedman's paradox. Ann. Inst. Stat. Math. 62, 117–125. https://doi.org/10.1007/s10463-009-0234-4

Martre, P., Wallach, D., Asseng, S., Ewert, F., Jones, J.W., Rötter, R.P., Boote, K.J., Ruane, A.C., Thorburn, P.J., Cammarano, D., Hatfield, J.L., Rosenzweig, C., Aggarwal, P.K., Angulo, C., Basso, B., Bertuzzi, P., Biernath, C., Brisson, N., Challinor, A.J., Doltra, J., Gayler, S., Goldberg, R., Grant, R.F., Heng, L., Hooker, J., Hunt, L.A., Ingwersen, J., Izaurralde, R.C., Kersebaum, K.C., Müller, C., Kumar, S.N., Nendel, C., O'leary, G., Olesen, J.E., Osborne, T.M., Palosuo, T., Priesack, E., Ripoche, D., Semenov, M.A., Shcherbak, I., Steduto, P., Stöckle, C.O., Stratonovitch, P., Streck, T., Supit, I., Tao, F., Travasso, M., Waha, K., White, J.W., Wolf, J., 2015. Multimodel ensembles of wheat growth: many models are better than one. Glob. Chang. Biol. 21, 911–25. https://doi.org/10.1111/gcb.12768

McNunn, G., Heaton, E., Archontoulis, S., Licht, M., VanLoocke, A., 2019. Using a Crop Modeling Framework for Precision Cost-Benefit Analysis of Variable Seeding and Nitrogen Application Rates. Front. Sustain. Food Syst. 3, 108. https://doi.org/10.3389/fsufs.2019.00108

Meier, U., Biologische Bundesanstalt für Land- und Forstwirtschaft., 1997. Growth stages of mono- and dicotyledonous plants: BBCH-Monograph. Blackwell Wissenschafts-Verlag.

Menzel, A., Yuan, Y., Matiu, M., Sparks, T., Scheifinger, H., Gehrig, R., Estrella, N., 2020. Climate change fingerprints in recent European plant phenology. Glob. Chang. Biol. 26, 2599–2612. https://doi.org/10.1111/gcb.15000

Piao, S., Liu, Q., Chen, A., Janssens, I.A., Fu, Y., Dai, J., Liu, L., Lian, X., Shen, M., Zhu, X., 2019. Plant phenology and global climate change: current progresses and challenges. Glob. Chang. Biol. gcb.14619. https://doi.org/10.1111/gcb.14619

Rafiei, V., Nejadhashemi, A.P., Mushtaq, S., Bailey, R.T., An-Vo, D.-A., 2022. An improved calibration technique to address high dimensionality and non-linearity in integrated groundwater and surface water models. Environ. Model. Softw. 149, 105312. https://doi.org/10.1016/J.ENVSOFT.2022.105312

Rezaei, E.E., Siebert, S., Hüging, H., Ewert, F., 2018. Climate change effect on wheat phenology depends on cultivar change. Sci. Rep. 8, 4891. https://doi.org/10.1038/s41598-018-23101-2

Seber, G.A.F., Wild, C.J., 1989. Nonlinear regression. Wiley, New York.

Sisheber, B., Marshall, M., Mengistu, D., Nelson, A., 2022. Tracking crop phenology in a highly dynamic landscape with knowledge-based Landsat–MODIS data fusion. Int. J. Appl. Earth Obs. Geoinf. 106, 102670. https://doi.org/10.1016/J.JAG.2021.102670

Stuble, K.L., Bennion, L.D., Kuebbing, S.E., 2021. Plant phenological responses to experimental warming – A synthesis. Glob. Chang. Biol. gcb.15685. https://doi.org/10.1111/gcb.15685

Tao, F., Rötter, R.P., Palosuo, T., Gregorio Hernández Díaz-Ambrona, C., Mínguez, M.I., Semenov, M.A., Kersebaum, K.C., Nendel, C., Specka, X., Hoffmann, H., Ewert, F., Dambreville, A., Martre, P., Rodríguez, L., Ruiz-Ramos, M., Gaiser, T., Höhn, J.G., Salo, T., Ferrise, R., Bindi, M., Cammarano, D., Schulman, A.H., 2018. Contribution of crop model structure, parameters and climate projections to

uncertainty in climate change impact assessments. Glob. Chang. Biol. 24, 1291–1307. https://doi.org/10.1111/gcb.14019

van Bussel, L.G.J., Stehfest, E., Siebert, S., Müller, C., Ewert, F., 2015. Simulation of the phenological development of wheat and maize at the global scale. Glob. Ecol. Biogeogr. 24, 1018–1029. https://doi.org/10.1111/geb.12351

Wallach, D., 2011. Crop model calibration: A statistical perspective. Agron. J. 103, 1144–1151.

Wallach, D., Martre, P., Liu, B., Asseng, S., Ewert, F., Thorburn, P.J., van Ittersum, M., Aggarwal, P.K., Ahmed, M., Basso, B., Biernath, C., Cammarano, D., Challinor, A.J., De Sanctis, G., Dumont, B., Eyshi Rezaei, E., Fereres, E., Fitzgerald, G.J., Gao, Y., Garcia-Vila, M., Gayler, S., Girousse, C., Hoogenboom, G., Horan, H., Izaurralde, R.C., Jones, C.D., Kassie, B.T., Kersebaum, K.C., Klein, C., Koehler, A.-K., Maiorano, A., Minoli, S., Müller, C., Naresh Kumar, S., Nendel, C., O'Leary, G.J., Palosuo, T., Priesack, E., Ripoche, D., Rötter, R.P., Semenov, M.A., Stöckle, C., Stratonovitch, P., Streck, T., Supit, I., Tao, F., Wolf, J., Zhang, Z., 2018. Multimodel ensembles improve predictions of crop-environment-management interactions. Glob. Chang. Biol. 24, 5072–5083. https://doi.org/10.1111/gcb.14411

Wallach, D., Palosuo, T., Thorburn, P., Gourdain, E., Asseng, S., Basso, B., Buis, S., Crout, N., Dibari, C., Dumont, B., Ferrise, R., Gaiser, T., Garcia, C., Gayler, S., Ghahramani, A., Hochman, Z., Hoek, S., Hoogenboom, G., Horan, H., Huang, M., Jabloun, M., Jing, Q., Justes, E., Kersebaum, K.C., Klosterhalfen, A., Launay, M., Luo, Q., Maestrini, B., Mielenz, H., Moriondo, M., Nariman Zadeh, H., Olesen, J.E., Poyda, A., Priesack, E., Pullens, J.W.M., Qian, B., Schütze, N., Shelia, V., Souissi, A., Specka, X., Srivastava, A.K., Stella, T., Streck, T., Trombi, G., Wallor, E., Wang, J., Weber, T.K.D., Weihermüller, L., de Wit, A., Wöhling, T., Xiao, L., Zhao, C., Zhu, Y., Seidel, S.J., 2021a. How well do crop modeling groups predict wheat phenology, given calibration data from the target population? Eur. J. Agron. 124, 126195. https://doi.org/https://doi.org/10.1016/j.eja.2020.126195

Wallach, D., Palosuo, T., Thorburn, P., Hochman, Z., Andrianasolo, F., Asseng, S., Basso, B., Buis, S., Crout, N., Dumont, B., Ferrise, R., Gaiser, T., Gayler, S., Hiremath, S., Hoek, S., Horan, H., Hoogenboom, G., Huang, M., Jabloun, M., Jansson, P.-E., Jing, Q., Justes, E., Kersebaum, K.C., Launay, M., Lewan, E., Luo, Q., Maestrini, B., Moriondo, M., Olesen, J.E., Padovan, G., Poyda, A., Priesack, E., Pullens, J.W.M., Qian, B., Schütze, N., Shelia, V., Souissi, A., Specka, X., Kumar Srivastava, A., Stella, T., Streck, T., Trombi, G., Wallor, E., Wang, J., Weber, T.K.D., Weihermüller, L., de Wit, A., Wöhling, T., Xiao, L., Zhao, C., Zhu, Y., Seidel, S.J., 2021b. Multi-model evaluation of phenology prediction for wheat in Australia. Agric. For. Meteorol. 298–299, 108289. https://doi.org/10.1016/J.AGRFORMET.2020.108289

Wallach, D., Palosuo, T., Thorburn, P., Hochman, Z., Gourdain, E., Andrianasolo, F., Asseng, S., Basso, B., Buis, S., Crout, N., Dibari, C., Dumont, B., Ferrise, R., Gaiser, T., Garcia, C., Gayler, S., Ghahramani, A., Hiremath, S., Hoek, S., Horan, H., Hoogenboom, G., Huang, M., Jabloun, M., Jansson, P.-E., Jing, Q., Justes, E., Kersebaum, K.C., Klosterhalfen, A., Launay, M., Lewan, E., Luo, Q., Maestrini, B., Mielenz, H., Moriondo, M., Nariman Zadeh, H., Padovan, G., Olesen, J.E., Poyda, A., Priesack, E., Pullens, J.W.M., Qian, B., Schütze, N., Shelia, V., Souissi, A., Specka, X., Srivastava, A.K., Stella, T., Streck, T., Trombi, G., Wallor, E., Wang, J., Weber, T.K.D., Weihermüller, L., de Wit, A., Wöhling, T., Xiao, L., Zhao, C., Zhu, Y., Seidel, S.J., 2021c. The chaos in calibrating crop models: Lessons learned from a multi-model calibration exercise. Environ. Model. Softw. 145, 105206. https://doi.org/10.1016/J.ENVSOFT.2021.105206

Wang, P.C., Shoup, T.E., 2011. Parameter sensitivity study of the Nelder–Mead Simplex Method. Adv. Eng. Softw. 42, 529–533. https://doi.org/10.1016/J.ADVENGSOFT.2011.04.004

Webber, H., Lischeid, G., Sommer, M., Finger, R., Nendel, C., Gaiser, T., Ewert, F., 2020. No perfect storm for crop yield failure in Germany. Environ. Res. Lett. 15. https://doi.org/10.1088/1748-9326/aba2a4

White, H., 1981. Consequences and detection of misspecified nonlinear regression models. J. Am. Stat.

Assoc. 374, 419–433.

Zhang, L., Zhang, Z., Tao, F., Luo, Y., Zhang, J., Cao, J., 2022. Adapting to climate change precisely through cultivars renewal for rice production across China: When, where, and what cultivars will be required? Agric. For. Meteorol. 316, 108856. https://doi.org/10.1016/J.AGRFORMET.2022.108856